

# Assessment of Iodine Status Using Dried Blood Spot Thyroglobulin: Development of Reference Material and Establishment of an International Reference Range in Iodine-Sufficient Children

Michael B. Zimmermann, Bruno de Benoist, Sandro Corigliano, Pieter L. Jooste, Luciano Molinari, Khairya Moosa, Eduardo A. Pretell, Zuhair Salman Al-Dallal, Yao Wei, Chen Zu-Pei, and Toni Torresani

Laboratory for Human Nutrition (M.B.Z.), Swiss Federal Institute of Technology, CH-8092 Zürich, Switzerland; Department of Nutrition for Health and Development (B.d.B.), World Health Organization, 1211 Geneva, Switzerland; Endocrine and Metabolism Unit (S.C., E.A.P.), High Altitude Research Institute, Cayetano Heredia Peruvian University, 430 Lima, Peru; Nutritional Intervention Research Unit (P.L.J.), Medical Research Council, Cape Town 7505, South Africa; Child Development Center (L.M.), University Children's Hospital, CH-8032 Zürich, Switzerland; Nutrition Department (K.M., Z.S.A.-D.), Ministry of Health, Manama, Bahrain; Dalian Center for Disease Control and Prevention (Y.W.), Dalian 116023, The People's Republic of China; Institute of Endocrinology (C.Z.-P.), Tianjin Medical University, Tianjin 300060, The People's Republic of China; and Department of Endocrinology (T.T.), Children's Hospital, The University of Zürich, CH-8032 Zurich, Switzerland

**Context:** Thyroglobulin (Tg) may be a valuable indicator of improving thyroid function in children after salt iodization. A recently developed Tg assay for use on dried whole blood spots (DBS) makes sampling practical, even in remote areas.

**Objective:** The study aim was to develop a reference standard for DBS-Tg, establish an international reference range for DBS-Tg in iodine-sufficient children, and test the standardized DBS-Tg assay in an intervention trial.

**Design, Participants, and Interventions:** Serum Tg reference material of the European Community Bureau of Reference (CRM-457) was adapted for DBS and its stability tested over 1 yr. DBS-Tg was determined in an international sample of 5- to 14-yr-old children (n = 700) who were euthyroid, anti-Tg antibody negative, and residing in areas of long-term iodine sufficiency. In a 10-month trial in iodine-deficient children, DBS-Tg and other indicators of iodine status were measured before and after introduction of iodized salt.

**Results:** Stability of the CRM-457 Tg reference standard on DBS over 1 yr of storage at  $-20$  and  $-50$  C was acceptable. In the international sample of children, the third and 97th percentiles of DBS-Tg were 4 and 40  $\mu\text{g/liter}$ , respectively. In the intervention, before introduction of iodized salt, median DBS-Tg was 49  $\mu\text{g/liter}$ , and more than two thirds of children had DBS-Tg values greater than 40  $\mu\text{g/liter}$ . After 5 and 10 months of iodized salt use, median DBS-Tg decreased to 13 and 8  $\mu\text{g/liter}$ , respectively, and only 7 and 3% of children, respectively, had values greater than 40  $\mu\text{g/liter}$ . DBS-Tg correlated well at baseline and 5 months with urinary iodine and thyroid volume.

**Conclusions:** The availability of reference material and an international reference range facilitates the use of DBS-Tg for monitoring of iodine nutrition in school-age children. (*J Clin Endocrinol Metab* 91: 4881–4887, 2006)

**D**ESPITE SIGNIFICANT GLOBAL progress against the iodine-deficiency disorders (IDDs), one in three school-age children remain iodine deficient (1). Iodine deficiency is the single most important preventable cause of mental retardation worldwide (2). Three measures, urinary iodine (UI), goiter rate, and serum TSH, are recommended for assessment of iodine nutrition in populations (2), but each has limitations. UI is an indicator of recent iodine intake but not thyroid function. Because thyroid size decreases only

slowly after iodine repletion, the goiter rate may remain high for several years after introduction of iodized salt (2, 3). TSH is a sensitive measure of iodine status only in the newborn period (2, 4). Thus, an additional indicator of thyroid function, sensitive to recent changes in iodine nutrition and applicable in children, would be valuable in monitoring iodine status in populations.

Thyroglobulin (Tg), a thyroid-specific protein that is a precursor in the synthesis of thyroid hormone, has no known physiological role outside the thyroid (5, 6). Stimulated by TSH, transcytosis of Tg-containing endosomes across the thyrocyte results in small amounts of Tg being released into the blood (7). If a sensitive assay is used, Tg can be detected in the serum of all healthy individuals (7, 8). In the absence of thyroid damage, the major determinants of serum Tg are thyroid cell mass and TSH stimulation (8). Thus, serum Tg is elevated in iodine-deficient areas due to TSH hyperstimulation and thyroid hyperplasia. In 1994, the World Health

First Published Online September 12, 2006

Abbreviations: BSA, Body surface area; CRM, Community Bureau of Reference; CV, coefficient of variation; Delfia, dissociation enhanced lanthanide fluorescent immunoassay; DTC, differentiated thyroid cancer; IDD, iodine-deficiency disorder; DBS, dried whole blood spots; Tg, thyroglobulin; Tg-Ab, Tg antibodies;  $T_{vol}$ , thyroid gland volume; UI, urinary iodine; WHO, World Health Organization.

JCEM is published monthly by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.

Organization (WHO) recommended using serum Tg to assess iodine nutrition and proposed that a median Tg concentration of less than 10  $\mu\text{g/liter}$  in a population indicated iodine sufficiency (9). However, data to support this Tg cut-off value were limited, and the recommendation was not included in the revised 2001 WHO guidelines (2).

We recently adapted and validated a widely used sandwich fluorimetric serum Tg assay for use on dried whole blood spots (DBS) (10). In an intervention study that measured DBS-Tg in children before and after the introduction of iodized salt, the assay was a sensitive indicator of improving thyroid function after iodine repletion. However, use of Tg for monitoring iodine status is limited by large interassay variability and lack of reference data for Tg in healthy, iodine-sufficient school-age children. The development of an international serum Tg reference standard [Community Bureau of Reference (CRM)-457] has led to the restandardization of many Tg assays and reduced intermethod variability (11, 12). When CRM-457 standardization is used, the normal serum Tg reference range in adults is approximately 3–40  $\mu\text{g/liter}$  (12).

Our study aims were: 1) to develop standard reference material for the DBS-Tg assay using the CRM-457 Tg reference preparation; 2) using this material, to establish an international reference range for DBS-Tg in iodine-sufficient children that could be used for monitoring iodine nutrition; and 3) to evaluate the standardized DBS-Tg assay and reference range in a longitudinal study of goitrous children before and after introduction of iodized salt.

### Subjects and Methods

As described previously (10), a two-site dissociation enhanced lanthanide fluorescent immunoassay (Delfia) serum Tg assay (PerkinElmer Life Sciences, Wallac, Turku, Finland) was adapted for DBS and validated in Swiss children. An advantage of two-site Tg assays is their lower cross-reactivity and improved specificity, compared with one-site assays (5).

#### *Development and stability testing of the DBS-Tg reference standard*

The lyophilized Tg reference preparation of the Community Bureau of Reference of the Commission of the European Communities (CRM-457) (11, 12) was provided by C. Profilis (BCR, Brussels, Belgium). The reference material was reconstituted by adding 1 ml distilled water to the ampule, mixing gently for 5 min, and again intermittently to maintain dissolution. The reconstituted standard was stored for no longer than 24 h at 4 C before use. Varying concentrations of the CRM-457 standard (1, 10, 100, 500, and 1000  $\mu\text{g/liter}$ ) were prepared using as the diluent the 0 standard from the Delfia kit (PerkinElmer Life Sciences).

Calibrators for the DBS-Tg assay were then prepared using both the CRM-457 reference standard and the calibration material provided in the Delfia kit. Whole blood obtained from the local blood bank was centrifuged at  $1529 \times g$  for 10 min at 18 C (Rotanta; Hettich Zentrifugen, Tuttlingen, Germany); the serum was removed, 0.9% NaCl was added, the solution rotated in a blood mixer until homogeneous, and then recentrifuged at  $1529 \times g$  for 10 min at 18 C, and the supernatant fluid was removed. This step was repeated three times. Calibration solutions prepared from the Delfia serum Tg assay kit and the CRM-457 reference preparation were then added to the washed erythrocytes, rotated in a blood mixer for 10 min, dropped onto filter paper (grade 903; Schleicher & Schuell, Einbeck, Germany), and air dried for 24 h at 20 C (10). The calibration curves were constructed by using duplicate measurements at 0, 1, 10, 100, 500, and 1000  $\mu\text{g/liter}$  and smoothed by fitting a third-order

polynomial with the use of a smoothed spline technique (Multicalc Program; PerkinElmer Life Sciences).

To assess the stability of the CRM-457 on DBS, a 1-yr stability study was done comparing the CRM-457 reference preparation on DBS with the Tg calibration material from the Delfia kit (prepared as described above) at concentrations of 100 and 20  $\mu\text{g/liter}$  and stored at 25, 4, –20, and –70 C. DBS-Tg concentrations were measured in duplicate every 2 wk for the first two months and at monthly intervals thereafter.

#### *Establishing an international reference range for DBS-Tg in iodine-sufficient children*

The subjects were healthy children living in areas of long-term iodine sufficiency (13) in South America, Central Europe, the Eastern Mediterranean, Africa, and the Western Pacific. Recruitment was from primary schools at the middle to lower socioeconomic level. The sample included children from five major ethnic groups: Lima, Peru (Hispanic); Zürich, Switzerland (white); Manama, Bahrain (Arabic); Cape Town, South Africa (black); and Dalian, China (Asian). Preanalytic exclusion criteria were: 1) age younger than 5 yr or older than 14 yr; 2) personal or immediate family history of thyroid disease; 3) cigarette smoking; or 4) pregnancy. Both cigarette smoking and pregnancy may be associated with higher serum Tg values (12). Postanalytic exclusion criteria were: 1) abnormal serum TSH and/or  $T_4$ ; and 2) detectable anti-Tg antibodies (Tg-Ab). With the relative precision for the 97th percentile for DBS-Tg specified at 3–5% of the total length of the 95% reference range, and the estimated SD of DBS-Tg taken as 2.1  $\mu\text{g/liter}$  (based on a small study in iodine-sufficient Swiss children), it was estimated that a sample size of approximately 500 children was required to obtain the required precision level (14). However, because of uncertainty on the variability of DBS-Tg in children due to the small amount of available data, roughly 700 children were finally enrolled.

Ethical committees approved the protocol at each local institution involved in the study. Informed written consent was obtained from the parents and oral assent from the participating children. Height and weight were measured using standard anthropometric technique (15). For the measurements, children removed their shoes, emptied their pockets, and wore light indoor clothing. Heights were recorded to the nearest mm and weights to the nearest 100 g. Pubertal staging was not done. Whole blood from a finger stick was spotted onto filter paper (grade 903; Schleicher & Schuell), allowed to dry at room temperature (~20 C), and stored at 4 C in sealed low-density polyethylene bags until analysis. Spot urine samples were collected and aliquots were stored at –20 C until analysis.

#### *Intervention study*

The study was done in a rural village in the Brikcha Commune, an area of endemic goiter in northern Morocco (16). Per-capita salt intakes in school-age children in this region are 7–12 g/d (16). A local cooperative supplies nearly all salt in this region; the salt is produced in drying ponds using water from a salty spring. Although Morocco legislated mandatory salt iodization in 1997, it is estimated that only approximately 45% of the population has access to iodized salt (Chaouki, N., personal communication, 2002). Due to financial constraints, this small local cooperative had not yet begun iodization. To prepare the iodized salt, iodine was added as reagent-grade potassium iodate (Sigma & Aldrich, Buchs, Switzerland) at a level of 25  $\mu\text{g}$  iodine per gram of salt, using an electric rotating drum mixer (ELTE 650; Engelsmann, Ludwigshafen, Germany) as described previously (16).

The subjects were primary school children, aged 5–14 yr ( $n = 86$ ). Informed oral consent was obtained from parents of the children and oral assent from the children. The Swiss Federal Institute of Technology in Zürich and the Ministry of Health in Rabat gave ethical approval for the study. At baseline, height and weight were measured using standard anthropometric technique (15). Whole blood from a finger stick was spotted directly onto filter paper (grade 903; Schleicher & Schuell), allowed to dry at room temperature (~20 C), and stored at 4 C in sealed low-density polyethylene bags until analysis. To check for the influence of sampling on the DBS-Tg assay, whole blood was obtained from a subsample of children ( $n = 20$ ; mean  $\pm$  SD age,  $9.2 \pm 2.1$  yr) by both

**TABLE 1.** Age, gender ratio, anthropometric measurements, and UI concentration in an international sample of 5- to 14-yr-old children from areas of long-term iodine sufficiency and with normal thyroid function, by site and combined

Site	n	Age (yr) <sup>a</sup>	Male to female ratio	Height (cm) <sup>b</sup>	Weight (kg) <sup>b</sup>	UI (μg/liter) <sup>a</sup>
Bahrain	142	10.1 (5.9–14.1)	1.03	137 ± 11	33.5 ± 12.5	177 (43–701)
Peru	125	10.0 (5.0–12.0)	1.66	133 ± 11	32.4 ± 9.2	161 (15–860)
South Africa	127	9.7 (6.0–13.1)	1.02	135 ± 14	34.7 ± 13.9	266 (38–758)
China	230	9.0 (6.0–12.0)	1.05	138 ± 13	32.5 ± 11.1	234 (0–672)
Switzerland	86	10.0 (7.0–14.0)	1.53	143 ± 11	36.4 ± 9.8	130 (6–390)
Total	710	9.6 (5.0–14.1)	1.18	137 ± 13	33.5 ± 11.6	198 (0–860)

<sup>a</sup> Data shown as median (range).<sup>b</sup> Data shown as mean ± SD.

venipuncture of a forearm vein and finger prick. The blood from venipuncture was collected, and then spotted onto filter paper. Spot urine samples were collected, and aliquots were stored at –20 C until analysis. Thyroid gland volume ( $T_{vol}$ ) was measured using a portable Aloka SSD-500 Echocamera (Aloka, Mure, Japan) with a high-resolution 7.5-MHz linear transducer (17). Each household was then provided 2 kg salt to supply all household needs at the beginning of each month for 10 months. Salt aliquots were taken ( $n = 6$ ) after each mixing for determination of iodine content. The salt was dispensed directly to the head of the household from a central supply at the local health center, and it was explained to the participating families that the new salt should be used for all cooking and food preparation as well as at the table. At 5 and 10 months, the baseline measurements were repeated.

### Laboratory analyses

UI was measured using the Pino modification of the Sandell-Kolthoff reaction (18). At UI concentrations of 47 and 79 μg/liter, the interrater coefficient of variation (CV) of this assay at the Human Nutrition Laboratory at the Swiss Federal Institute of Technology, Zürich, our laboratory, is 10.3 and 12.7%, respectively. The limit of detection is 2 μg/liter; samples below this limit were assigned a value of 0. Salt iodine concentration was measured using a modification of the Sandell-Kolthoff reaction, after dissolution of salt aliquots in distilled water. At iodine concentrations of 30 μg/g salt, the interrater CV of this assay at the Human Nutrition Laboratory is 7%.  $T_{vol}$  was calculated using the method of Brunn *et al.* (19). M.B.Z. performed all ultrasound measurements during the study. To estimate intraobserver variability, duplicate  $T_{vol}$  measurements were done in 10 children at the 5- and 10-month time points; the mean (SD) variability was 3.9 (2.1)%. Goiter was defined using sex- and body surface area (BSA)-specific reference criteria for  $T_{vol}$  (17). DBS-Tg analyses were done in Zürich at the Protein hormone Laboratory at the University Children's Hospital (10). DBS were analyzed for TSH (Delfia NeoTSH) (20) and total  $T_4$  (Delfia Neonatal  $T_4$  kit, PerkinElmer Life Sciences). Normal reference values are whole blood TSH, 0.2–3.7 mU/liter; and serum total  $T_4$ , 65–165 nmol/liter. Tg-Ab were measured by RIA (RIA Tg-Ab; RSR, Cardiff, UK) adapted in our laboratory for measurement on DBS. For the Tg-Ab assay, between- and within-assay CV is 10.1 and 2.5%, respectively ( $n = 145$ ). Tg-Ab status was classified as: detectable, Tg-Ab greater than 0.3 U/ml; and elevated, Tg-Ab greater than 10 U/ml.

**TABLE 2.** Whole blood TSH, serum total  $T_4$  (TT4), and anti-Tg-Ab in an international sample of 5- to 14-yr-old euthyroid children from areas of long-term iodine sufficiency, by site and combined

Site	n	TSH (mU/liter) blood <sup>a</sup>	TT4 (pmol/liter) <sup>b</sup>	Tg-Ab (U/ml) <sup>a</sup>
Bahrain	142	1.1 (0.3–3.6)	102 ± 17	0.3 (0.3–6.1)
Peru	125	1.0 (0.5–3.4)	97 ± 16	0.3 (0.3–7.8)
South Africa	127	0.9 (0.5–2.8)	108 ± 22	0.3 (0.3–2.9)
China	230	1.4 (0.4–3.7)	114 ± 19	0.3 (0.3–9.0)
Switzerland	86	0.6 (0.2–1.2)	90 ± 22	0.3 (0.3–7.3)
Total	710	1.1 (0.2–3.7)	105 ± 21	0.3 (0.3–9.0)

<sup>a</sup> Data shown as median (range).<sup>b</sup> Data shown as mean ± SD.

### Statistical analyses

Data processing and statistics were done using SPLUS-2000 (Insightful Corp., Seattle, WA) and Excel (Enterprise Edition, Microsoft, Seattle, WA). Distribution of Tg in the sample exhibited a long-tailed-positive skewness and kurtosis. Log transformation removed most of the skewness and kurtosis, leaving a nearly Gaussian distribution at all ages and BSAs, for both sexes. Transformed data were used to calculate percentiles based on the Gaussian distribution, which were then transformed back to the linear scale. ANOVA was used to test differences between the six sites modeling age, BSA, and sex dependence. Simultaneous 95% confidence intervals for the differences in DBS-Tg between sites were done using the Tukey method of multiple comparisons (21). For the ANOVA, data from 12 subjects (two from Bahrain, three from China, one from Peru, and six from South Africa) of 710 were excluded as outliers in the relation of DBS-Tg to age and would possibly have had an undue influence on the calculations. In the intervention study, one-way ANOVA was done to compare changes in urinary iodine, TSH,  $T_4$ , Tg, Tg-Ab, and thyroid volume. Tukey's test was used for *post hoc* comparisons. Variables not normally distributed were logarithmically transformed before analysis. To evaluate the relation between Tg and other IDD indicators, Pearson's correlation coefficients were calculated as well as linear regression models with Tg as the dependent variable. For the regression, to reduce the influence of age and sex on thyroid volume, SD scores were obtained using current reference values (17) as previously described (10). Significance was set at  $P < 0.05$ .

### Results

#### Stability of the CRM-457 Tg reference standard on DBS

Regression analysis of the results of the 1-yr stability study showed that the slopes for both CRM and Delfia material were not significantly different from 0. The intercepts for the 100- and 20-μg/liter samples were: 109.7 and 17.0 μg/liter (Delfia) and 115.1 and 18.4 μg/liter (CRM), respectively. The stability study showed that both reference materials (CRM-457 and kit calibrator) were stable in DBS for 1 yr when stored at least at –20 C. At 4 C, both preparations were stable for 2 months. At higher storage temperatures, Tg degraded sig-

**TABLE 3.** DBS Tg on the linear scale and log(10) transformed, by site, showing the median and 97th percentiles in an international sample of 5- to 14-yr-old euthyroid children from areas of long-term iodine sufficiency, by site

Site	n	Tg ( $\mu\text{g}/\text{liter}$ )		log(10) Tg	
		Median (25th, 75th percentile)	97th percentile	Median (25th, 75th percentile)	97th percentile
Bahrain	140	19.3 (12.6, 27.6)	48.9	1.28 (1.10, 1.44)	1.69
Peru	124	11.6 (7.0, 19.2)	36.1	1.06 (0.85, 1.28)	1.56
South Africa	121	18.4 (13.2, 26.0)	40.7	1.27 (1.12, 1.42)	1.61
China	227	13.3 (8.9, 21.2)	35.6	1.12 (0.95, 1.33)	1.55
Switzerland	86	11.2 (7.0, 15.9)	24.8	1.05 (0.85, 1.20)	1.39
Total	698	14.5 (9.4, 22.7)	40.2	1.16 (0.97, 1.36)	1.61

nificantly. Recovery of CRM-457 material when measured against the Delfia reference was 87%. There was excellent correlation between the CRM-457 and Delfia results in the international sample of iodine sufficient children (described below) ( $r = 0.967$ ,  $P < 0.0001$ ).

#### Reference range for DBS-Tg in iodine-sufficient children

Table 1 shows the age, gender ratio, anthropometric measurements, and UI concentration in the study sample by site and combined. All sites were iodine sufficient as defined by a median UI between 100 and 300  $\mu\text{g}/\text{liter}$  (1). Overall, mean age  $\pm$  SD was 9.6 (5.0–14.1), with the whole sample nearly equally divided between boys and girls. Table 2 shows the whole blood TSH, serum total  $T_4$ , and anti-Tg-Ab, in the study sample, by site and combined.

Table 3 shows DBS-Tg on the linear scale and log(10) transformed, by site, showing the median and 97th percentiles. Pooling the data, the median (third, 97th percentiles) log(10) DBS-Tg was 1.16 (0.57, 1.61). Transformed back to the linear scale, the median (third, 97th percentiles) for DBS-Tg was 14.5 (3.7, 40.3). The nonparametric 95% confidence intervals for the median, third, and 97th percentiles were 13.4, 15.5; 3.1, 4.3; and 37.8, 45.6, respectively. There were complex and significant differences for DBS-Tg between sites in the means and slope with respect to age but no significant gender differences at any of the sites. The differences in DBS-Tg between sites after controlling for age are shown in Fig. 1.

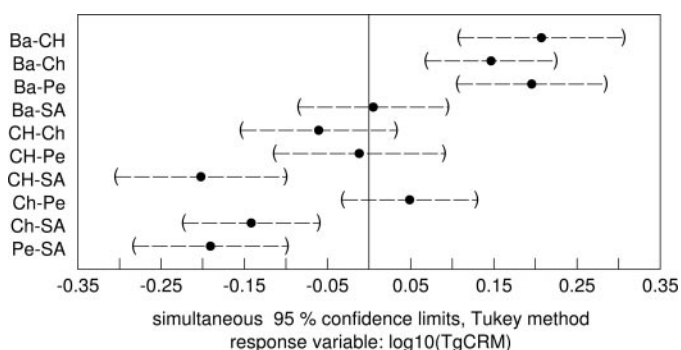
There was no clear relationship between DBS-Tg and UI; within a country, the correlation coefficients ranged between  $-0.14$  (Bahrain) and  $0.084$  (China), and none were statisti-

cally significant. The correlation coefficients of DBS-Tg and TSH, within country, were: Bahrain (0.077, NS); Peru (0.035, NS); South Africa (0.21,  $P = 0.02$ ); China (0.18,  $P = 0.007$ ); and Switzerland (0.34;  $P = 0.001$ ); and for the pooled sample (0.13,  $P = 0.0004$ ). The differences between countries were statistically significant in level but not slope (no significant interaction). The correlation coefficients of DBS-Tg and total  $T_4$ , within country, were: Bahrain (0.071, NS); Peru (0.30,  $P = 0.0006$ ); South Africa (0.13, NS); China (0.14, NS); and Switzerland (0.072; NS); and for the pooled sample (0.15,  $P < 0.0001$ ). The differences between countries were statistically significant in level but not in slope (no significant interaction).

#### Intervention study

Of the 86 children who began the study, 83 completed it; three children moved away or were absent from school on the measurement days. Characteristics of the children at baseline were as follows: mean  $\pm$  SD age was  $10.6 \pm 2.4$  yr; the gender ratio was 40 females to 43 males; mean  $\pm$  SD weight was  $31.2 \pm 9.9$  kg; mean  $\pm$  SD height was  $1.34 \pm 0.14$  m. In the monitoring aliquots of iodized salt taken at the monthly mixings, the mean iodine concentration ( $\pm$  SD) was  $22.8 \pm 5.1$   $\mu\text{g}/\text{g}$ . As shown in Table 4, median UI at 5 and 10 months was significantly increased, compared with baseline ( $P < 0.001$ ). At 10 months, median UI had increased to near the WHO/International Council for the Control of Iodine Deficiency Disorders cutoff value (100  $\mu\text{g}/\text{liter}$ ) for risk of iodine deficiency (2). Mean  $T_{\text{vol}}$  and median TSH at 10 months were significantly decreased, compared with baseline ( $P < 0.05$ ). Median (range) DBS-Tg was 49 (1–862)  $\mu\text{g}/\text{liter}$  at baseline and fell rapidly after introduction of iodized salt to 13 (1–208) and 8 (1–95)  $\mu\text{g}/\text{liter}$  at 5 and 10 months ( $P < 0.001$ ). Tg-Abs were not measured in the children. Using the 97th percentile of the reference range for DBS-Tg reported above, 68, 7, and 3% of children had elevated DBS-Tg concentrations at baseline and 5 and 10 months, respectively.

DBS-Tg correlated well with the other major response variables during the study. At baseline and at 5 months, DBS-Tg was negatively correlated with urinary iodine [correlation coefficients  $-0.41$  ( $P < 0.001$ ) and  $-0.19$  ( $P < 0.05$ ), respectively] and was positively correlated with thyroid volume [correlation coefficients  $0.47$  ( $P < 0.001$ ) and  $0.18$  ( $P < 0.05$ ), respectively]. DBS-Tg was significantly correlated with TSH at baseline only [correlation coefficient  $0.32$  ( $P < 0.001$ )]. The regression of DBS-Tg on urinary iodine, TSH,  $T_4$ , and thyroid volume was done at each time point. The regression



**FIG. 1.** Simultaneous 95% confidence intervals for the differences in DBS-Tg between sites, using the Tukey method of multiple comparisons (17). The differences are adjusted for age and are significant if the confidence interval does not include zero. Ba, Bahrain; Pe, Peru; SA, South Africa; Ch, China; CH, Switzerland.

**TABLE 4.** UI concentration, T<sub>vol</sub>, and goiter rate measured using ultrasound, serum total T<sub>4</sub> (TT4), whole blood TSH, and DBS-Tg in Moroccan schoolchildren (n = 83) before (0 months) and 5 and 10 months after introduction of iodized salt

	Baseline	5 months	12 months
UI ( $\mu\text{g/liter}$ ) <sup>1,3</sup>	12 (2–70) <sup>a</sup>	74 (2–239) <sup>b</sup>	104 (22–1784) <sup>b</sup>
T <sub>vol</sub> (ml) <sup>2,3</sup>	8.3 $\pm$ 3.5 <sup>a</sup>	7.8 $\pm$ 3.1 <sup>a</sup>	6.9 $\pm$ 2.2 <sup>b</sup>
TT4 (nmol/liter) <sup>2</sup>	98 $\pm$ 18	111 $\pm$ 21	101 $\pm$ 19
TSH (mU/liter blood) <sup>1,3</sup>	1.3 (0.3–7.4) <sup>a</sup>	0.7 (0.3–3.0) <sup>b</sup>	0.6 (0.2–4.4) <sup>b</sup>
DBS-Tg ( $\mu\text{g/liter}$ ) <sup>1,3</sup>	49 (1–862) <sup>a</sup>	13 (1–208) <sup>b</sup>	8 (1–95) <sup>c</sup>

Values in the same row with *different superscript letters* are significantly different,  $P < 0.05$  (Tukey's test for *post hoc* comparisons).

<sup>1</sup> Data shown as median (range).

<sup>2</sup>  $P < 0.0001$  (ANOVA).

<sup>3</sup> Data shown as mean  $\pm$  SD.

of UI and thyroid volume on DBS-Tg was significant ( $P < 0.001$ ) at baseline and at 5 months ( $P < 0.01$ ). Comparing the DBS-Tg assay on the parallel venipuncture and finger-stick samples (n = 20), geometric mean (range) DBS-Tg was 8.4 (1.8–94.9) and 7.8 (2.1–115.7)  $\mu\text{g/liter}$  (NS).

### Discussion

In our international sample of iodine-sufficient children, there were no significant gender differences in DBS-Tg. In adults, serum Tg levels are slightly higher in females, but this is not clinically informative, and Tg reference ranges for adults are not gender specific (12). Data on the effects of chronological age on serum Tg are scarce. In healthy term infants, serum Tg increases in the first few days after birth, presumably due to the postnatal surge in TSH (22). Serum Tg then decreases by 50% over the next few months and then more slowly declines throughout childhood to reach adult levels at puberty (23). In our sample, although there were statistically significant differences in DBS-Tg between younger and older children, these differences were small and varied by site. Overall, our data suggest age-, site-, or gender-adjusted reference ranges for DBS-Tg are unnecessary for children in the age range of 5–14 yr. We would therefore recommend use of a single reference range for screening and monitoring of iodine nutrition in this age group. Based on our data (Table 3), the DBS-Tg reference interval for iodine-sufficient, Tg-Ab-negative, euthyroid school-age children, using CRM-457-standardization, is 4–40  $\mu\text{g/liter}$ .

This proposed reference range is nearly the same as the usual adult reference range for serum Tg when CRM-457 standardization is used, *i.e.* approximately 3–40  $\mu\text{g/liter}$  (12). The close similarity of the child and adult reference ranges for Tg parallels that of serum concentrations of TSH and T<sub>4</sub>: although age-specific reference limits for TSH and T<sub>4</sub> are needed for young children, by age 5 yr, the child to adult ratio for TSH and T<sub>4</sub> are approximately 1 (24). The long-term intraindividual variability in serum Tg in adults is low (CV ~14%); there appears to be no significant diurnal or seasonal variability in serum Tg concentrations in moderately iodine-deficient populations (22), but this has not been studied in severe iodine deficiency. Using a different serum Tg assay, Vanderschueren-Lodeweyckx (25) proposed normal mean values and ranges for serum Tg in children aged 1–10 and 11–20 yr of 35 (2–65) and 18 (2–36)  $\mu\text{g/liter}$ , respectively. Previous studies measuring serum Tg in children from iodine-deficient areas have reported median values ranging

from 27–214  $\mu\text{g/liter}$  (26–29). In Chinese adults in regions of mild iodine deficiency and iodine excess, serum Tg values were in the range of 6.0–11.7 ng/ml, with no significant differences between regions (30). Further investigation in other iodine-deficient populations to determine whether DBS-Tg is elevated relative to the degree of iodine deficiency would be useful.

Even with CRM-457 standardization, there is significant technical variability in serum Tg assays, presumably due to epitope specificity differences that cause interassay biases independent of standardization (12). The primary clinical use of serum Tg measurements is a marker for patients carrying a diagnosis of differentiated thyroid cancer (DTC). Interassay variability has precluded the use of serial serum Tg measurements by different laboratories for following individuals after surgery for DTC (12). However, for the purposes of diagnostic testing, *i.e.* distinguishing iodine-deficient from iodine-sufficient populations using a normal reference interval, assay bias and imprecision goals need not be as stringent as for serial measurements for DTC follow-up. Therefore, although use of our DBS CRM Tg standard will not eliminate DBS-Tg interassay variability, it may facilitate the calibration of assays and allow the interchangeable use of different DBS Tg assays to characterize iodine status in a population. More research is needed on this issue.

A potential limitation to the use of a DBS-Tg assay for IDD monitoring is interference from Tg-Ab. Even trace amounts of Tg-Ab in a specimen may interfere with Tg measurement, and this is a common source of Tg assay error in adults followed up for thyroid cancer (12). Detectable Tg-Ab are found in approximately 10% of the general adult population (31). It is unclear how prevalent Tg-Ab are in iodine-deficient children or whether they are precipitated by iodine prophylaxis (32). Several studies (33, 34) reported high prevalences, ranging from 7 to 69%, in iodine-deficient children and during iodine prophylaxis. In contrast, data from our group and others (35–37) suggest elevated Tg antibodies in IDD-affected children are rare (0–2% prevalence). In our international sample of children, only 25 (3.3% of the total sample) had detectable (2.4%) or elevated (0.9%) Tg-Ab. These data suggest screening for Tg-Ab may not be necessary when using a DBS-Tg assay in children to classify population iodine status. Further research is needed to address the question of whether simultaneous measurement of anti-Tg antibodies is necessary.

Cross-sectional studies have reported a negative correla-

tion between Tg and UI and positive correlation between Tg and thyroid volume and TSH (26, 38, 39). Intervention studies suggested that Tg is a more sensitive indicator than TSH or T<sub>4</sub> in measuring response to iodized oil (26–28) and potassium iodide (29). In the present study, before introduction of iodized salt, median DBS-Tg was high (49  $\mu\text{g/liter}$ ), and more than two thirds of children had DBS-Tg values greater than 40  $\mu\text{g/liter}$ . After 5 months of iodized salt use, the median had decreased to 13  $\mu\text{g/liter}$ , well within the normal range, and only 7% of children had a value greater than 40  $\mu\text{g/liter}$ . During the intervention study, DBS-Tg correlated well at both baseline and 5-month time points with UI and thyroid volume, the two other major response variables used to measure impact of iodized salt in children. Thus, the data suggest that Tg, used in conjunction with UI to measure recent iodine intake and thyroid volume to assess long-term anatomic response, may be a useful biological indicator for monitoring thyroid function in children after introduction of iodized salt. The DBS assay makes sampling practical even in remote areas.

### Acknowledgments

We thank S. Schoeman and E. Strydom (Medical Research Council, Cape Town, South Africa); I. Aeberli, S. Renggli, C. Zeder, N. Hurrell, and C. Mini (Swiss Federal Institute of Technology, Zürich, Switzerland); Toni Trachsel (Uster, Switzerland); K. Bagchi (WHO Eastern Mediterranean Regional Office); M. Tello and M. Ramirez (Lima, Peru); and the teachers and children in the participating schools.

Received June 26, 2006. Accepted September 5, 2006.

Address all correspondence and requests for reprints to: Michael B. Zimmermann, M.D., Laboratory for Human Nutrition, Swiss Federal Institute of Technology, LfV E19, Schmelzbergstrasse 7, CH-8092 Zürich, Switzerland. E-mail: michael.zimmermann@ilw.agrl.ethz.ch.

This work was supported by the Swiss Federal Institute of Technology (Zürich, Switzerland), the World Health Organization (Geneva, Switzerland), PerkinElmer Life Sciences (Wallac, Turku, Finland), Merck KGaA (Darmstadt, Germany), and the Medical Research Council (Cape Town, South Africa). Each of the authors made substantial contributions to the study design, data collection, and data analyses as well as the writing and/or editing of the paper.

Disclosure statement: The authors have nothing to disclose. None of the authors has a personal or financial interest in the companies or organizations sponsoring this research, including advisory board affiliations.

### References

- de Benoist B, Andersson M, Takkouche B, Egli I 2003 Prevalence of iodine deficiency worldwide. *Lancet* 362:1859–1860
- WHO, UNICEF, ICCIDD 2001 Assessment of iodine deficiency disorders and monitoring their elimination: a guide for programme managers. 2nd ed. Geneva: World Health Organization/NHD/01.1
- Zimmermann MB, Hess SY, Adou P, Torresani T, Wegmüller R, Hurrell RF 2003 Thyroid size and goiter prevalence after introduction of iodized salt: a 5-year prospective study using ultrasonography in schoolchildren in Côte d'Ivoire. *Am J Clin Nutr* 77:663–667
- Zimmermann MB, Aeberli I, Torresani T, Bürgi H 2005 Increasing the iodine concentration in the Swiss iodized salt program markedly improves iodine status in pregnant women and children: a 5-yr prospective national study. *Am J Clin Nutr* 88:388–392
- Tórrrens JL, Burch HB 2001 Serum thyroglobulin measurement. *Endocrinol Metab Clin North Am* 30:429–467
- De Vijlder JJM, Ris-Stalpers C, Vulsma T 1999 On the origin of circulating thyroglobulin. *Eur J Endocrinol* 140:7–8
- Dunn JT, Dunn AD 2000 Thyroglobulin: chemistry, biosynthesis and proteolysis. In: Braverman LE, Utiger RD, eds. *Werner and Ingbar's the thyroid: a fundamental and clinical text*. 8th ed. Philadelphia: Lippincott, Williams and Wilkins
- Spencer CA 1995 Thyroglobulin measurement: techniques, clinical benefits, and pitfalls. *Endocrinol Metab Clin North Am* 24:841–863
- WHO/ICCIDD/UNICEF 1994 Indicators for assessing iodine deficiency disorders and their control through salt iodization. Geneva: World Health Organization/NUT/94.6
- Zimmermann MB, Moretti D, Chaouki N, Torresani T 2003 Development of a dried whole blood spot thyroglobulin assay and its evaluation as an indicator of thyroid status in goitrous children receiving iodized salt. *Am J Clin Nutr* 77:1453–1458
- Feldt-Rasmussen U, Profilis C, Colinet E, Black E, Bornet H, Bourdoux P, Carayon P, Ericsson UB, Koutras DA, Lamas de Leon L, DeNayer P, Pacini F, Palumbo G, Santos A, Schlumberger M, Seidel C, Van Herle AJ, DeVijlder JJ 1996 Human thyroglobulin reference material (CRM 457). 2nd part: physicochemical characterization and certification. *Ann Biol Clin (Paris)* 54:343–348
- 2003 Laboratory medicine practice guidelines. Thyroglobulin (Tg) measurement. In: Demers LM, Spencer CA, eds. *Laboratory support for the diagnosis and monitoring of thyroid disease*. *Thyroid* 13:57–67
- Current IDD Status Database International Council for the Control of Iodine Deficiency Disorders. Available at: <http://www.icidd.org> (accessed June 2006)
- Armitage P, Berry G, Matthews JNS 2002 *Statistical methods in medical research*. 4th ed. Oxford, UK: Blackwell Science; 397–399
- WHO 1995 Physical status: the use and interpretation of anthropometry. WHO Technical report series no. 854. Geneva: World Health Organization
- Zimmermann MB, Zeder C, Chaouki N, Saad A, Torresani T, Hurrell RF 2003 Dual fortification of salt with iodine and microencapsulated iron: a randomized, double blind, controlled trial in Moroccan schoolchildren. *Am J Clin Nutr* 77:425–432
- Zimmermann MB, Hess SY, Molinari L, De Benoist B, Delange F, Braverman LE, Fujieda K, Ito Y, Jooste PL, Moosa K, Pearce EN, Pretell EA, Shishiba Y 2004 New reference values for thyroid volume by ultrasound in iodine-sufficient schoolchildren: a WHO/NHD Iodine Deficiency Study Group Report. *Am J Clin Nutr* 79:231–237
- Pino S, Fang SL, Braverman LE 1996 Ammonium persulfate: a safe alternative oxidizing reagent for measuring urinary iodine. *Clin Chem* 42:239–243
- Brunn J, Block U, Ruf G, Bos I, Kunze WP, Scriba PC 1981 Volume measurement of the thyroid gland using real-time sonography. *Deutsch Med Wochenschrift* 106:1338–1340
- Torresani T, Scherz R 1986 Thyroid screening of neonates without use of radioactivity: evaluation of time-resolved fluoroimmunoassay of thyrotropin. *Clin Chem* 32:1013–1016
- Hsu JC 1996 *Multiple comparisons: theory and methods*. London: Chapman and Hall
- Spencer CA 2000 Thyroglobulin. In: Braverman LE, Utiger RD, eds. *Werner and Ingbar's the thyroid: a fundamental and clinical text*. 8th ed. Philadelphia: Lippincott, Williams and Wilkins; 402–413
- Penny R, Spencer CA, Frasier SD, Nicoloff JT 1983 Thyroid-stimulating hormone and thyroglobulin levels decrease with chronological age in children and adolescents. *J Clin Endocrinol Metab* 56:177–180
- 2003 Laboratory medicine practice guidelines. Preanalytic factors. In: Demers LM, Spencer CA, eds. *Laboratory support for the diagnosis and monitoring of thyroid disease*. *Thyroid* 13:6–18
- Vanderschueren-Lodeweyckx M 1996 Thyroid function tests. In: Ranke M, ed. *Diagnostics of endocrine function in children and adolescents*. 2nd ed. Mannheim, Germany: J&J Verlag
- Missler U, Gutekunst R, Wood W 1994 Thyroglobulin is a more sensitive indicator of iodine deficiency than thyrotropin: development and evaluation of dry blood spot assays for thyrotropin and thyroglobulin in iodine-deficient geographical areas. *Eur J Clin Chem Clin Biochem* 32:137–143
- Van den Briel T, West CE, Hautvast J, Vulsma T, de Vijlder JM, Ategbro EA 2001 Serum thyroglobulin and urinary iodine concentration are the most appropriate indicators of iodine status and thyroid function under conditions of increasing iodine supply in schoolchildren in Benin. *J Nutr* 131:2701–2706
- Benmiloud M, Chauoki ML, Gutekunst R, Teichert M, Wood WG, Dunn JT 1994 Oral iodised oil for correcting iodine deficiency: optimal dosing and outcome indicator selection. *J Clin Endocrinol Metab* 79:20–24
- Todd CH, Dunn JT 1998 Intermittent oral administration of potassium iodide solution for the correction of iodine deficiency. *Am J Clin Nutr* 67:1279–1283
- Teng W, Shan Z, Teng X, Guan H, Li Y, Teng D, Jin Y, Yu X, Fan C, Chong W, Yang F, Dai H, Yu Y, Li J, Chen Y, Zhao D, Shi X, Hu F, Mao J, Gu X, Yang R, Tong Y, Wang W, Gao T, Li C 2006 Effect of iodine intake on thyroid diseases in China. *N Engl J Med* 354:2783–2793
- Spencer CA, Takeuchi M, Kazarosyan M, Wang CC, Guttler RB, Singer PA, Fatemi S, LoPresti JS, Nicoloff JT 1998 Serum thyroglobulin autoantibodies: prevalence, influence on serum thyroglobulin measurement, and prognostic significance in patients with differentiated thyroid carcinoma. *J Clin Endocrinol Metab* 83:1121–1127

32. Delange F 1998 Risks and benefits of iodine supplementation. *Lancet* 351: 923–924
33. Zonenberg A, Kinalska I, Zarzycki W, Telejko B 1994 Incidence of thyroid autoantibodies in endemic goiter. *Horm Metab Res* 26:238–242
34. Premawardhana LD, Parkes AB, Smyth PP, Wijeyaratne CN, Jayasinghe A, de Silva DG, Lazarus JH 2000 Increased prevalence of thyroglobulin antibodies in Sri Lankan schoolgirls—is iodine the cause? *Eur J Endocrinol* 143: 185–188
35. Loviselli A, Velluzzi F, Mossa P, Cambosu MA, Secci G, Atzeni F, Taberlet A, Balestrieri A, Martino E, Grasso L, Songini M, Bottazzo GF, Mariotti S; Sardinian Schoolchildren Study Group 2001 The Sardinian Autoimmunity Study: 3. Studies on circulating antithyroid antibodies in Sardinian schoolchildren: relationship to goiter prevalence and thyroid function. *Thyroid* 11: 849–857
36. Marwaha RK, Tandon N, Nandita AKK, Gupta N, Verma K, Kochupillai N 2000 Hashimoto's thyroiditis: countrywide screening of goitrous healthy young girls in postiodization phase in India. *J Clin Endocrinol Metab* 85:3798–3802
37. Zimmermann MB, Moretti D, Chaouki N, Torresani T 2003 Introduction of iodized salt to severely iodine-deficient children does not provoke thyroid autoimmunity: a 1-year prospective trial in northern Morocco. *Thyroid* 13: 199–203
38. Knudsen N, Bülow I, Jorgenson T, Perril H, Ovesen L, Laurberg P 2001 Serum thyroglobulin—a sensitive marker of thyroid abnormalities and iodine deficiency in epidemiologic studies. *J Clin Endocrinol Metab* 86:3599–3603
39. Fenzi GF, Ceccarelli C, Macchia E, Monzani F, Bartalena L, Giani C, Ceccarelli P, Lippi F, Baschieri L, Pinchera A 1985 Reciprocal changes of serum thyroglobulin and TSH in residents of a moderate endemic goitre area. *Clin Endocrinol (Oxf)* 23:115–122

JCEM is published monthly by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.