ORIGINAL ARTICLE

Association between *CYP19A1* polymorphisms and sex hormones in postmenopausal Japanese women

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In postmenopausal women, extraovarian sex hormone production plays an important role in hormone-related diseases, such as breast and endometrial cancers. Aromatase, an enzyme encoded by *CYP19A1*, is a key enzyme involved in estrogen biosynthesis. The impact of *CYP19A1* polymorphisms on serum sex hormone levels in the Japanese population has never been investigated. This study enrolled 100 postmenopausal Japanese women found to be without cancer. Twenty-five *CYP19A1* loci were identified, and measurements were conducted on serum levels of sex hormones; lifestyle data were collected, namely estrone (E1), estradiol (E2), testosterone and sex hormone-binding globulin (SHBG). We conducted a cross-sectional analysis to evaluate the impact of *CYP19A1* haplotype on serum sex hormone levels. We found that subjects with BMI $\ge 25 \text{ kg/m}^2$ showed a significant difference in circulating testosterone levels (0.29 ± 0.19 , P=0.050). Neither age nor the amount of physical exercise or drinking habits showed any effect on hormone levels. We identified seven haplotype blocks in *CYP19A1* by LD analysis. Estrone levels were associated with rs4441215 (SNP11). Testosterone levels were strongly associated with rs752760 (SNP24) and rs2445768 (SNP25) and weakly associated with SNP 1, SNP11 and SNP14 as well. We found that polymorphisms in *CYP19A1* influence sex hormone levels in Japanese postmenopausal women.

Journal of Human Genetics (2009) 54, 78-85; doi:10.1038/jhg.2008.11; published online 16 January 2009

Keywords: CYP19A1; polymorphism; aromatase; sex hormone; lifestyle; endometrial cancer

INTRODUCTION

An association has been recognized between sex hormones and the risk of breast, ovarian and endometrial cancers, particularly in postmenopausal women. Earlier studies have shown a clear association between the circulating levels of estrogen and testosterone in postmenopausal women and the risk of these cancers.^{1–3} These findings highlight the importance of identifying factors that define hormonal levels in postmenopausal women.

On reaching menopause, the ovaries cease to produce endogenous estrogens, at which point extragonadal sites such as adipose tissues become a major alternative source.^{4–6} Estrogen production occurs through the catalytic activity of the aromatase enzyme, which is encoded by *CYP19A1*. On the assumption that certain *CYP19A1* polymorphisms might affect estrogen levels, several studies have explored the potential impact of polymorphisms in *CYP19A1* on the risk of several types of cancer in Caucasian populations.^{7–9} More recently, Haiman *et al.*¹⁰ examined 103 single nucleotide polymorphisms (SNPs) in more than 3000 cancer-free subjects of mainly

European descent (93%) and found an association between haplotypetagging SNPs in the 5' region and circulating levels of estrogen in *CYP19A1*. Dunning *et al.*¹¹ examined two loci in *CYP19A1* and found a significant association between these two loci and sex hormone levels. Although hormone-related cancer incidence is gradually increasing in Asian populations, including the Japanese,¹² information regarding the effect of *CYP19A1* SNPs on estrogen levels is still strongly lacking.

In this study, we examined the association between the *CYP19A1* polymorphisms in conjunction with environmental elements and serum hormone levels in postmenopausal Japanese women.

MATERIALS AND METHODS

Subjects

This study enrolled 100 cancer-free naturally postmenopausal women through the Hospital-based Epidemiologic Research Program at Aichi Cancer Center (HERPACC) between December 2005 and June 2006 as a part of the Japan Multi-institutional Collaborative Cohort Study (J-MICC).¹³ Subjects visited ACC for more intensive examination after testing positive in a cancer screening

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Received 3 October 2008; revised 6 November 2008; accepted 7 December 2008; published online 16 January 2009

test, but were eventually found to not have cancer. All subjects consented to blood sampling for the measurement of hormone levels and genetic assay. Subjects currently receiving any kind of hormonal therapy were excluded. In the HERPACC, all first-visit outpatients at ACC Hospital (ACCH) are asked to fill out a self-administered questionnaire and provide a 14-ml blood sample after the written informed consent is obtained. All questionnaires are then checked by trained interviewers for incomplete responses. Details of the HERPACC study are described elsewhere.¹⁴ This study was approved by the ethics committee at ACC.

Measurement of hormone levels

Hormone level analyses were carried out on serum samples stored at -80 °C. Levels of estradiol, estrone, testosterone and sex hormone-binding globulin (SHBG) were measured by SRL Inc. (Tokyo, Japan). Estrone levels were measured by radioimmunoassay (Teikokuzoukiseiyaku Medical, Tokyo, Japan), estradiol and testosterone levels by electrochemiluminescent immunoassay (Roche Diagnostics, Tokyo, Japan) and SHBG levels by immunoradiometric assay (Siemens Medical Solutions Diagnostics, Tokyo, Japan).

SNP selection, genotyping and analysis for linkage disequilibrium (LD)

The SNP loci examined were basically selected by SNP Browser ver. 3.5 (Applied Biosystems, Foster City, CA, USA)¹⁵ on the basis of the HapMap database for the Japanese in Tokyo (build 36).^{16,17} Selection criteria for each locus were (1) minor allele frequency (MAF) greater than 30% and (2) haplotype R^2 -value greater than 0.95. Of 169 candidate SNPs, 82 were excluded on the basis of MAF criteria. Of the remaining 87 SNPs, the 25 SNPs that were able to tag the other 62 SNPs owing to strong LD were finally selected.

DNA was extracted from the buffy coat fraction in each subject using a Blood Mini Kit (Qiagen KK, Tokyo, Japan) and analyzed using the polymerase chain reaction (PCR) TaqMan method¹⁸ with the 7500 fast real-time PCR System (Applied Biosystems). The probes used were specifically designed for rs12148604, rs2899473, rs12900487, rs1865803, rs10459592, rs12591359, rs767199, rs16964221, rs12908960, rs7172156, rs4545755, rs11636403, rs3889391, rs2414101, rs17647707, rs17647719, rs1902586, rs936306, rs999480, rs2470152, rs3751591, rs1004982, rs1902585, rs752760 and rs2445768. Genotyping was conducted in duplicate in cases where accordance with the Hardy–Weinberg equilibrium (HWE) was violated.

LD was evaluated by means of LD coefficients (D'). Haplotype blocks were defined using 95% confidence intervals of D', where the upper and lower boundaries for strong LD were set as 0.98 and 0.60, as proposed by Gabriel *et al.*¹⁹ Minimum R^2 -value for strong LD was set as 0.30. Haplotype alleles and their frequencies were estimated using SNPAlyze (Dynacom, Yokohama, Japan) on the basis of multiple SNPs obtained with the expectation–maximization algorithm.

Exposure data

Consumption of types of alcoholic beverages (Japanese sake, beer, shochu, whiskey and wine) was determined with reference to the average number of drinks per day, which was then converted into a Japanese sake (rice wine) equivalent.²⁰ The drinking habit was further classified into the four categories of never, former, moderate and heavy drinking. Heavy drinkers were those currently drinking alcoholic beverages 5 or more days per week at 46 g (two 'go') or more per day, whereas moderate drinkers were those currently consuming less frequently 5 days per week. Former drinkers were defined as those who had quit drinking at least 1 year before the survey. Body mass index (BMI) was calculated as self-reported weight (kg) divided by the square of self-reported height (m). A validation study showed the correlation coefficient for BMI to be 0.92.²¹

The questionnaire also enquired about regular physical exercise. Subjects were asked to report the frequency and intensity of recreational exercise. In this study, subjects who reported a frequency of at least once a month were defined as those who exercised.

Statistical analysis

Intergroup comparison was conducted using one-way ANOVA followed by pairwise comparison across groups. All statistical analyses were performed using Stata Ver. 10 (Stata Corp., College Station, TX, USA). A *P*-value of < 0.05 was considered as statistically significant, and a value between 0.05 and 0.1 was considered as marginally significant.

RESULTS

Table 1 shows hormone and SHBG levels at baseline. We found significantly higher levels of circulating testosterone (P=0.050) in subjects with BMI \ge 25 kg/m² than in subjects with BMI < 25. In contrast, no differences were observed for estrone, estradiol or SHBG levels. We found no significant differences across groups in levels examined according to age, amount of physical exercise or drinking.

Figure 1 shows the location of CYP19A1 on chromosome 15 with the distribution of 25 SNPs examined (a), and the genetic structure of CYP19A1 with untranslated first exons (b). SNP23 and SNP17 violated HWE, and interpretation for these loci should therefore be approached with caution. Table 2 shows hormone and SHBG levels according to the 25 SNPs examined and Table 3 displays a detailed analysis for loci showing significant association in Table 2. Estrone levels were significantly different in SNP1 (rs12148604) located in the 3'-flanking region of CYP19A1. Estrone levels rose with an increased number of T alleles at this locus in any of the models. Further, the C allele of SNP14 (rs2414101) located in intron 1 showed a significant difference in estrone levels in recessive and codominant models. No SNP showed a significant association with a change in estradiol levels. SHBG levels showed a significant difference with the G allele of SNP 11 (rs11636403) in intron 1 in all three models. Testosterone levels were significantly different with the A allele of SNP24 (rs752760) in exon 1 and T allele of SNP25 (rs2445768) in the 5' near-gene region. SNP1, SNP11 and SNP14 also showed weak but significant association with testosterone levels.

The results of the LD analysis are shown in Figure 2. We identified seven haplotype blocks. Table 4 lists haplotypes on the basis of selected SNPs in each block along with their frequencies, as well as associations between hormone and SHBG levels and haplotypes in each block. Results showed a statistically significant difference in testosterone levels in block 7 (P=0.0066). The haplotype G-T-C showed relatively smaller values compared with others. Blocks 1 and 5 showed marginally significant associations with regard to estrone levels (P=0.0795 and 0.0995, respectively). In contrast, this haplotype block-based analysis revealed no remarkable findings with regard to estradiol and SHBG levels.

DISCUSSION

In this study, we identified seven haplotype blocks in *CYP19A1* by assessing 25 tag SNPs. We further examined the possible association between haplotypes/genotypes and the levels of sex hormones and SHBG in postmenopausal Japanese women. Results showed that estrone levels were significantly affected by SNP1 (rs12148604) in the 3' region and by SNP14 (rs11632903) in the region between untranslated exons I.2 and I.6. These loci were weakly but significantly associated with testosterone levels. Moreover, testosterone levels were markedly different across haplotypes in block 7 as well as with SNP24 and SNP25 (rs752760, rs2445768) located in the 5' region of *CYP19A1*. Estrone and testosterone levels were higher in subjects with higher BMI, particularly with regard to testosterone.

We found a significant association between estrone levels and SNP1 genotype located in the 3'UTR. This finding is consistent with earlier studies that showed an association between polymorphisms in the 3'UTR (rs10046) and estrone and estradiol levels.^{10,11,22} According to the HapMap database, SNP1 (rs12148604) and rs10046 are in LD in

Table 1 Characteristics and serum concentrations of hormones and SHBG

Characteristics	n=100	Estrone ($pgmI^{-1}$)	Estradiol ($pgml^{-1}$) SHBG ($nmol l^{-1}$)		Testosterone (ng ml $^{-1}$)	
Age (years)						
50–55	2	125.0 ± 55.0	13.5 ± 4.9	74.1 ± 16.9	0.44 ± 0.40	
56–60	38	100.0 ± 39.8	10.7 ± 2.1	71.7±28.2	0.22 ± 0.14	
61–65	25	111.5 ± 47.7	10.5 ± 0.8	85.3±32.8	0.19 ± 0.067	
66–70	15	91.3±34.8	11.7 ± 2.3	89.4±38.0	0.22 ± 0.083	
71–75	9	93.9±23.5	11.1 ± 2.1	112.3±82.7	0.23 ± 0.092	
76–80	8	99.1±41.2	14.8 ± 11.1	84.2±17.3	0.27 ± 0.092	
P-values ^a		(<i>P</i> =0.71)	(<i>P</i> =0.07)	(<i>P</i> =0.11)	(<i>P</i> =0.06)	
<i>BMI</i> ^b						
<25 kg/m ²	88	99.5±38.9	11.1 ± 3.8	85.2±39.7	0.21 ± 0.11	
$25 \text{kg/m}^2 \leqslant$	11	111.1 ± 48.8	11.8 ± 2.0	66.9±26.9	0.29 ± 0.19	
<i>P</i> -values ^a		(<i>P</i> =0.2564)	(<i>P</i> =0.568)	(<i>P</i> =0.143)	(<i>P</i> =0.050)	
Physical exercise						
Light exercise						
Yes	73	102.3 ± 42.3	11.1±3.9	82.4±41.1	0.22 ± 0.12	
No	27	97.7±32.7	11.6±2.8	83.6±33.0	0.23 ± 0.11	
<i>P</i> -values ^a		(<i>P</i> =0.58)	(<i>P</i> =0.21)	(<i>P</i> =0.67)	(<i>P</i> =0.27)	
Moderate exercise						
Yes	73	99.6 ± 45.8	11.1 ± 2.0	85.0±42.1	0.23 ± 0.13	
No	27	101.6 ± 37.8	11.3 ± 4.1	76.3 ± 28.1	0.22 ± 0.11	
P-values ^a		(<i>P</i> =0.68)	(<i>P</i> =0.76)	(<i>P</i> =0.32)	(<i>P</i> =0.54)	
Heavy exercise						
Yes	1	99.5±0	10±0	73±0	0.18 ± 0	
No	99	101.1 ± 40.1	11.3±3.7	82.8±39.1	0.22 ± 0.12	
P-values ^a		(<i>P</i> =0.97)	(<i>P</i> =0.49)	(<i>P</i> =0.80)	(<i>P</i> =0.74)	
Drinking habit						
Current	32	94.1 ± 35.8	10.9 ± 1.7	80.8±30.1	0.23 ± 0.17	
Former	1	61.4 ± 0	10±0	86.3±0	0.15 ± 0	
Never	65	104.7 ± 42.0	11.5 ± 4.4	83.1±43.5	0.21 ± 0.1	
Unknown	2	105 ± 14.8	10.5 ± 0.7	97.3±18.0	0.21 ± 0.1	
P-values ^a		(<i>P</i> =0.154)	(<i>P</i> =0.917)	(<i>P</i> =0.70)	(<i>P</i> =0.924)	

^a*P*-values were test for equality by ANOVA test. ^bOne subject was excluded because of the lack of BMI information.





four ethnicities: JPT (Japanese individuals from Tokyo), HCB (Han Chinese individuals from Beijing), CEU (American individuals from Utah with northern and western European ancestry) and YRI (Yoruba individuals from Ibadan and Nigeria). Therefore, our results for SNP1,

in combination with those of other studies, suggest the presence of a functional region in the 3'UTR of CYP19A1, which influences hormone levels among postmenopausal women of various ethnic backgrounds.

Table 2 Concentrations of hormones and SHBG according to tagSNPs

			Location in			Estrone (pgml ⁻¹)	Estradiol (pg ml ⁻¹)	SHBG (nmpl I−1)	Testosterone (ng ml-1)
SNP ID	dbSNP rs numb	er	chromosome	Genotype	n=100	(P-value) ^a	(P-value) ^a	(P-value) ^a	(P-value) ^a
SNP 1	rs12148604	3′UTR	49288696	CC	27	86.7±23.6	10.6 ± 2.1	80.8±35.1	0.19 ± 0.072
				CT	50	102.4 ± 40.3	11.7 ± 4.7	83.0±44.3	0.23 ± 0.13
				TT	23	115.0 ± 49.2	11.1 ± 1.9	84.2±31.1	0.22 ± 0.13
						(<i>P</i> =0.04)	(<i>P</i> =0.435)	(<i>P</i> =0.953)	(<i>P</i> =0.311)
SNP 2	rs2899473	int4	49306365	CC	16	114.0 ± 52.2	11 ± 1.8	81.7±32.7	0.22 ± 0.16
				CT	49	102.6 ± 41.3	11.1 ± 1.9	85.2 ± 44.1	0.22 ± 0.097
				TT	35	93.0±29.3	11.6 ± 5.6	79.6±34.3	0.22 ± 0.13
						(<i>P</i> =0.205)	(<i>P</i> =0.819)	(<i>P</i> =0.803)	(<i>P</i> =0.996)
SNP 3	rs12900487	int3	49310451	CC	55	106.4 ± 47.2	11.1 ± 2.2	83.7±44.3	0.23 ± 0.14
				CT	40	96.5±27.0	11.6 ± 5.2	82.9±32.0	0.21 ± 0.093
				TT	5	79.3±31.0	10.2 ± 0.4	70.0±29.0	0.21 ± 0.047
						(<i>P</i> =0.225)	(<i>P</i> =0.653)	(<i>P</i> =0.752)	(<i>P</i> =0.801)
SNP 4	rs1865803	int2	49318306	CC	9	89.3±31.6	10.2 ± 0.4	69.9 ± 30.3	0.19 ± 0.069
				CT	44	98.3±27.5	11.7 ± 5.0	84.7±31.2	0.23 ± 0.12
				TT	47	105.9 ± 49.8	11.0 ± 2.2	83.2±46.4	0.22 ± 0.12
						(<i>P</i> =0.436)	(<i>P</i> =0.442)	(<i>P</i> =0.580)	(<i>P</i> =0.690)
SNP 5	rs10459592	int1	49323433	GG	16	113.7 ± 52.9	10.8 ± 1.8	84.6±35.1	0.22 ± 0.16
				GT	48	102.2 ± 41.4	11.1 ± 1.8	85.3±43.5	0.22 ± 0.098
				TT	36	94.0±29.5	11.7 ± 5.6	78.4±34.5	0.22 ± 0.13
						(<i>P</i> =0.2513)	(<i>P</i> =0.682)	(<i>P</i> =0.712)	(<i>P</i> =0.987)
SNP 6	rs12591359	int1	49326660	AA	46	100.1 ± 32.6	11.7 ± 5.1	81.4±32.5	0.22 ± 0.12
				AG	43	96.8±40.4	10.8 ± 1.3	87.0±46.4	0.22 ± 0.093
				GG	11	121.8 ± 60.0	11.2 ± 2.3	71.1±31.4	0.23 ± 0.19
						(<i>P</i> =0.1742)	(<i>P</i> =0.525)	(<i>P</i> =0.469)	(<i>P</i> =0.907)
SNP 7	rs767199	int1	49327679	AA	17	111.4 ± 51.7	10.9±1.8	79.4±33.0	0.22 ± 0.15
				AG	46	103.6±41.8	11.2 ± 1.9	85.8±44.5	0.23 ± 0.099
				GG	37	93.1±29.5	11.5 ± 5.5	80.3±34.3	0.21 ± 0.13
						(P=0.251)	(P=0.869)	(P=0.761)	(P=0.936)
SNP 8	rs16964220	int1	49330674	AA	10	89.3+29.8	10.3+0.5	68 4 + 28 9	0.2+0.067
0.11 0	1010501220		10000071	AG	44	98 9 + 27 6	117+50	86 5 + 31 8	0.22 ± 0.12
				GG	46	105.6 ± 50.3	11.0+2.2	82.1+46.3	0.22 ± 0.12
					10	(P=0.454)	(P=0.474)	(P=0.416)	(<i>P</i> =0.789)
SNP 9	rs2008691	int1	49332423	AA	46	105.6 ± 50.3	11.0+2.2	82.1+46.3	0.22 ± 0.13
0.11 5	102000001		19002 120	AG	44	98 9 + 27 6	11.7 + 5.0	86 5 + 31 7	0.23+0.12
				GG	10	89.3+29.8	10.3 ± 0.5	68 4 + 28 9	0.2+0.067
					10	(P=0.454)	(P=0.474)	(P=0.416)	(<i>P</i> =0.789)
SNP 10	rs11856927	int1	49333152	GG	18	1094 ± 508	11 1 + 1 8	80 6 + 32 4	0.22 ± 0.15
0111 10	1311000327		15000102	GT	50	102.6 ± 40.7	11.1 = 1.0	80 5 + 42 9	0.21 + 0.099
				TT	32	93 9 + 30 7	11.2 ± 2.3 11.4 ± 5.7	80 2 + 36 5	0.22±0.13
					52	(<i>P</i> =0.394)	(<i>P</i> =0.958)	(<i>P</i> =0.835)	(<i>P</i> -0.850)
SNP 11	rs4441215	int1	49336036	CC	38	(7 = 0.03 +) 102 2 + 32 1	112+20	78 9 + 26 1	0 23 + 0 11
0111 11	131111210		19000000	CG	45	1039+490	11.2 ± 2.0	783+339	0.22 ± 0.11
				66	17	91 1 + 26 4	11.3 ± 4.5 11.1 ± 2.7	102.7 ± 64.2	0.19+0.083
				uu	17	(<i>P</i> =0.520)	(<i>P</i> _0.978)	(<i>P</i> =0.065)	(P = 0.341)
SNP 12	rc/5/5755	int1	19336336	ΔΔ	17	(1 = 0.520)	109+18	79 4 + 33 0	(7 = 0.3 + 1) 0.22 + 0.15
0111 12	13-10-107 00	IIIII	+3330330	AG	16	1027 ± 422	11.2+1.9	85 2 + 11 7	0.22 ± 0.15
				CC C	37	Q1 3 + 29 1	11.2 ± 1.5 11.5 ± 5.5	81 1 + 34 2	0.22 ± 0.33
				uu	57	(P=0.320)	$(P_{-0.849})$	(<i>P</i> _0 835)	(P-0.936)
SND 13	rc3880301	int1	10315711	٨٨	11	110 3 + 30 0	(1 = 0.043)	80 4 + 20 3	(7 = 0.000)
2141 12	13000000000		+5545714	ΔG	11	106 3 + 48 1	11. 4 ± 2.2	79 0 + 30 0	0.20 ± 0.17 0.22 + 0.10
				66	40	100.3 ± 40.1	11.1 ± 1.9	7 5.0 ± 30.0	0.22 ± 0.10
				99	44	שט.4 ± ∠0.4 (<i>P</i> _∩ מימו)	11.4 ± 0.1 (P_0 014)	07.0±40.3 (P-0 608)	0.21 ± 0.12 (P=0 515)
	rc11632002	int1	10351633	CC	20	(1 - 0.223)	(1 - 0.914) 11 $1 \pm 5 = 1$	(1 - 0.000)	(1 - 0.010)
SIF 14	1211025202	11111	49001000	CT CT	39	90.1 ± 20.1	11.4±3.4	03.1 ± 43.1	0.20 ± 0.000
					40	100.7 ± 47.3	11.U±1.8	73.3 ± 29.4	0.23 ± 0.13
				11	15	112.3 ± 30.5	11.5 ± 2.3	87.3±29.9	0.25 ± 0.15
						(P=0.077)	(<i>r</i> =0.863)	(<i>r</i> =0.208)	(P=0.224)

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Table 2 Continued

			Location in			Estrone (pg ml ⁻¹)	Estradiol (pg ml ⁻¹)	SHBG (nmpl I−1)	Testosterone (ng ml-1)
SNP ID	dbSNP rs number		chromosome	Genotype	n=100	(P-value) ^a	(P-value) ^a	(P <i>-value</i>) ^a	(P-value) ^a
SNP 15	rs17647707	int1	49355312	GG	60	106.7±44.8	11.1±2.1	86.0±44.4	0.22±0.13
				GT	37	92.8±29.6	11.6 ± 5.4	79.4±28.6	0.22 ± 0.091
				TT	3	90.2±33.5	10.7 ± 0.6	56.6±22.5	0.21 ± 0.061
						(<i>P</i> =0.227)	(<i>P</i> =0.798)	(<i>P</i> =0.360)	(<i>P</i> =0.950)
SNP 16	rs17647719	int1	49355496	AA	51	106.0 ± 47.4	11.0 ± 2.1	85.2±46.3	0.22 ± 0.12
				AG	43	94.8±30.2	11.6 ± 5.1	81.7±29.4	0.22 ± 0.12
				GG	6	103.8 ± 26.5	10.5 ± 0.5	67.9±30.8	0.21 ± 0.061
						(<i>P</i> =0.397)	(<i>P</i> =0.647)	(<i>P</i> =0.578)	(<i>P</i> =0.814)
SNP 17	rs1902586	int1	49358145	AA	8	101.4 ± 25.3	10.4 ± 0.5	75.6±30.2	0.21 ± 0.067
				AG	55	95.6±37.5	11.6 ± 4.6	82.3±43.6	0.23 ± 0.14
				GG	37	109.1 ± 44.9	11.0 ± 2.2	84.8±33.5	0.21 ± 0.097
						(<i>P</i> =0.28)	(<i>P</i> =0.573)	(<i>P</i> =0.829)	(<i>P</i> =0.740)
SNP 18	rs936306	int1	49366890	CC	33	109.7 ± 47.4	11±23	80.0±30.0	0.21 ± 0.1
				СТ	54	97.3±36.9	11.6 ± 4.6	84.8±45.5	0.22 ± 0.14
				TT	13	94.6±28.3	10.4 ± 0.7	80.5±30.0	0.21 ± 0.063
						(<i>P</i> =0.307)	(<i>P</i> =0.483)	(<i>P</i> =0.840)	(<i>P</i> =0.792)
SNP 19	rs999480	int1	49375152	AA	3	90.2±33.5	10.7 ± 0.6	56.6±22.5	0.21 ± 0.061
				GA	42	94.1±30.6	11.6 ± 5.1	78.4±29.3	0.23 ± 0.13
				GG	55	107.0±45.6	11.0 ± 2.1	87.4 ± 45.1	0.21 ± 0.12
						(<i>P</i> =0.261)	(<i>P</i> =0.699)	(<i>P</i> =0.266)	(<i>P</i> =0.892)
SNP 20	rs2470152	int1	49382264	CC	27	98.1±32.8	11.3 ± 2.1	72.8±23.5	0.23 ± 0.13
				TC	54	106.7 ± 46.0	11.2 ± 4.5	83.2±33.8	0.21 ± 0.095
				TT	19	89.3±26.4	11.4 ± 2.8	95.1 ± 62.1	0.22 ± 0.17
						(<i>P</i> =0.236)	(<i>P</i> =0.989)	(P=0.159)	(<i>P</i> =0.745)
SNP 21	rs3751591	int1	49394002	CC	4	102.3 ± 19.0	10.5 ± 0.6	71.4 ± 21.1	0.19 ± 0.053
				CT	36	95.6 ± 35.0	11.9 ± 5.6	78.4±31.1	0.23 ± 0.13
				TT	60	104.3 ± 43.5	10.9 ± 1.9	86.0±43.7	0.21 ± 0.12
						(<i>P</i> =0.589)	(<i>P</i> =0.426)	(<i>P</i> =0.549)	(<i>P</i> =0.626)
SNP 22	rs1004982	int1	49401103	AA	41	105.7 ± 45.2	11.1 ± 2.2	83.4±33.9	0.21 ± 0.092
				GA	53	96.9±36.6	11.5 ± 4.6	83.7±43.5	0.23 ± 0.13
				GG	6	105.7 ± 28.5	10.2 ± 0.4	68.9 ± 29.4	0.21 ± 0.15
						(P=0.551)	(P=0.671)	(P=0.673)	(P=0.895)
SNP 23	rs1902585	int1	49401198	CC	15	95.9 ± 15.9	11 ± 1.7	87.9±38.2	0.24 ± 0.11
				CG	48	106.6 ± 45.1	11.0 ± 1.9	86.4 ± 46.4	0.23 ± 0.13
				GG	37	96.0 ± 39.1	11.8 ± 5.5	75.8 ± 26.7	0.19 ± 0.095
						(P=0.415)	(P=0.584)	(P=0.400)	(P=0.178)
SNP 24	rs752760	exon1	49418771	CC	16	100.1+23.0	11 4+2 2	86.3+37.5	0.27 ± 0.16
0.0.2.	107 027 00	0,0112	10110771	CT	49	104.2+44.4	10.9 ± 1.7	85.8+46.4	0.22 ± 0.11
				TT	35	97.1 + 39.8	11.8+5.7	76.7 + 26.4	0.19 ± 0.097
						(P=0.726)	(P=0.526)	(P=0.536)	(P=0.071)
SNP 25	rs2445768	5' near gene	49422672	AA	26	100.8 ± 24.8	11.1 ± 2.0	81.8±36.1	0.27 ± 0.17
2 20		2 mour Solito		AC	46	103.7 ± 45.4	10.9 ± 1.7	87.3+47.1	0.21 ± 0.090
				CC	28	97.0+42.4	12+6.3	75.9+23.8	0.18 ± 0.086
					20	(P=0.785)	(P=0.439)	(P=0.471)	(P=0.015)
						(1 -0.700)	() =0.1007	(7 = 0. 17 1)	()=0.010)

^aANOVA test was used.

Regarding estrone levels, Cai *et al.*²³ recently reported that the association between CYP19A1 SNPs (rs28566535, rs730154 and rs936306) were associated with estrone levels in the Chinese population. They found that the region of these loci is close to untranslated exon I.4 and includes a specificity protein-1-binding site, (γ)-interferon-activating sequence and glucocorticoid response element, which are essential for aromatase expression from I.4. However, we did not find any association with any of the loci we examined. Moreover, we found SNP14 (rs11632903) for estrone levels. The

inconsistency may be explained partly by the difference of baseline estrone levels between our population (mean: 101.1 pg ml^{-1}) and the Chinese population (16.8 pg ml^{-1}). This might reflect genetic differences in other enzymes involved in estrogen synthesis between the two populations. Meanwhile, replication studies in other Asian populations are required.

We also found an association between testosterone levels and SNP24 (rs752760) and SNP25 (rs2445768) located in the 5' noncoding region of *CYP19A1*. Several studies, conducted primarily in

Table 3	Detailed	analyses of	of selected	loci for	estrone,	SHBG and	teststerone levels
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				Estrone			SHBG		Testosterone		
SNP	rs number	Minor allele	Dominant	Recessive	Cocominant	Dominant	Recessive	Cocominant	Dominant	Recessive	Cocominant
SNP1	rs12148604	Т	0.0018	0.0065	0.0014	0.6797	0.771	0.9064	0.0336	0.8101	0.0932
SNP11	rs4441215	G	0.1072	0.1072	0.2645	0.0009	0.0009	0.0039	0.0487	0.0487	0.1122
SNP14	rs11632903	С	0.0924	0.0016	0.0055	0.4805	0.0413	0.0413	0.1558	0.0176	0.0479
SNP24	rs752760	Т	0.8854	0.3042	0.5212	0.5692	0.1115	0.2821	0.005	0.013	0.0046
SNP25	rs2445768	А	0.9598	0.3662	0.6109	0.8444	0.1231	0.217	0.0001	0.005	0.0002

Figure 2 LD plot across the *CYP19A1* loci. Cells feature pairwise D' values. Rectangles filled with red, pink and white correspond to LD magnitude of |D'| > 0.9, $0.9 > |D'| \ge 0.5$ and |D'| < 0.5, respectively. Groups of rectangles outlined in red represent haplotype blocks as estimated by SNPAlyze (Dynacom, Yokohama, Japan). The color reproduction of this figure is available on the html full text version of the manuscript.

Caucasian subjects, have examined the association between testosterone levels and *CYP19A1* polymorphisms, although to our knowledge, none have found any association with 5'UTR loci.^{10,24,25} SNP24 is known to locate in the region containing the placenta-specific promoter (I.1 and I.2a). Considering that the subjects in this study were postmenopausal women, there is thus far no biological evidence supporting the possible contribution of promoters to testosterone levels. These findings, however, suggest the presence of functional SNPs around SNP24 and SNP25, which lead to differences in testosterone levels specific to the Japanese population. A weak but significant association between testosterone levels and SNP1, SNP11 and SNP14 is also suggestive of other functional regions. These findings warrant further studies into the underlying cause of these differences.

Although recent advances in genotyping have enabled an examination of the association between specific phenotypes, the study of hormone levels in postmenopausal women and potential loci by whole-genome SNP typing, as carried out in this study, is costly and susceptible to low statistical power. It is, therefore, more feasible to conduct association studies examining targeted genes on the basis of known biological mechanisms with tag SNPs. Haiman *et al.*¹⁰ recently conducted a tag SNP-based large scale cross-sectional study in postmenopausal Caucasian women and found a significant association between specific haplotype blocks and hormone levels. Our study is the first to apply a tag SNP-based approach to examine the impact of *CYP19A1* SNPs on hormone levels in postmenopausal Japanese women. Although the function of loci showing a significant association requires further clarification, our study showed the effectiveness of this tag SNP-based approach in examining hormone levels in postmenopausal women.

In premenopausal women, the main source of estrogen is estradiol produced in ovarian granulose cells during the follicular phase, although fibroblasts in adipose tissue expressing aromatase are an alternate source. After menopause, however, adipose tissue becomes the major source of estrogen, and estrogenically weak estrone is produced from adrenal androstenedione.^{26,27} Moreover, at least half of this peripherally produced estrone is eventually converted to estradiol in extraovarian tissue.²⁸ Testosterone is the precursor of estradiol produced in the ovary by the cholesterol metabolism pathway. It accumulates after menopause with the loss of the ovarian function. Our observation that the estrone and testosterone levels are higher in women with higher BMI than in those with lower BMI is consistent with the current knowledge of the metabolism of hormones in postmenopausal women. Studies conducted in other populations have also shown consistently increased hormone levels in accordance with higher BMI.^{26,29–31} Epidemiologically, obesity is a risk factor of hormone-dependent carcinoma, given the findings that the incidence of breast, endometrial and ovarian cancer is increased in postmenopausal women, along with increased levels of estrogen production in adipose tissues.^{32–35} On this basis, identifying those CYP19A1 loci that influence hormone levels may be an effective indicator of patients susceptible to hormone-related cancers.

Table 4 Haplotype frequencies according to haplotypes in seven blocks

	Haplotype number						n	Frequencies	Estrone (pg m l^{-1})	Estradiol ($pgml^{-1}$)	SHBG (nmol I^{-1})	Testosterone (ngml-1)
Block 1		SNP1	SNP 2									
	1	С	Т				102	0.5100	94.6±33.6	11.1 ± 3.7	82.2±39.7	0.21 ± 0.11
	2	Т	С				79	0.3950	107.8 ± 45.9	11.1 ± 1.9	84.3±39.8	0.22 ± 0.12
	3	Т	Т				17	0.0850	111.3 ± 39.5	12.8 ± 7.7	80.1±30.6	0.26 ± 0.16
	4	С	С				2	0.0100	79.3±14.6	10±0	66.3±33.2	0.20 ± 0.007
	(P-value) ^a								(<i>P</i> =0.0795)	(<i>P</i> =0.3054)	(<i>P</i> =0.9013)	(P=0.4071)
Block 2		SNP3	SNP4									
	1	С	Т				138	0.6900	103.5 ± 43.8	11.2 ± 3.3	83.7±41.9	0.22 ± 0.12
	2	Т	С				50	0.2500	93.0±28.0	11.3 ± 4.6	80.3±31.2	0.21 ± 0.085
	3	С	С				12	0.0600	106.8±28.9	11.1 ± 1.9	80.9±32.4	0.26 ± 0.18
									(<i>P</i> =0.2492)	(<i>P</i> =0.9885)	(<i>P</i> =0.8556)	(<i>P</i> =0.5050)
Block 3		SNP5	SNP6	SNP7								
	1	Т	А	G			114	0.5700	97.1±34.7	11.4 ± 4.5	82.6±38.5	0.22 ± 0.12
	2	G	G	А			59	0.2950	105.8 ± 49.3	10.8 ± 1.6	84.5±43.0	0.22 ± 0.13
	3	G	А	А			18	0.0900	110.2 ± 36.6	11.7 ± 2.4	83.5±29.1	0.22 ± 0.092
	4	Т	G	G			3	0.0150	90.3 ± 51.8	11.3 ± 2.3	52.9 ± 16.0	0.2 ± 0.087
	5	Т	G	А			3	0.0150	109.6±23.6	12.7 ± 2.5	53.5±14.8	0.27 ± 0.061
	6	G	А	G			3	0.0150	106.0 ± 36.4	10±0	104.2±37.8	0.15 ± 0.046
									(<i>P</i> =0.6567)	(<i>P</i> =0.8445)	(<i>P</i> =0.4819)	(<i>P</i> =0.9133)
Block 4		SNP8	SNP9									
	1	G	A				136	0.6800	103.5 ± 44.0	11.3 ± 3.4	83.5±41.9	0.22 ± 0.12
	2	А	G				64	0.3200	95.9±28.2	11.3 ± 4.2	80.9±31.6	0.22 ± 0.11
									(<i>P</i> =0.2121)	(<i>P</i> =0.9881)	(<i>P</i> =0.6503)	(<i>P</i> =0.9314)
Block 5		SNP13	SNP14	1								
	1	G	С				123	0.6150	96.4±37.1	11.3 ± 4.4	84.1±43.6	0.21 ± 0.10
	2	А	Т				65	0.3250	108.7±45.3	11.2 ± 2.0	79.6±29.7	0.23 ± 0.13
	3	G	Т				10	0.0500	114.3 ± 24.6	11.4 ± 2.1	83.2±32.8	0.28 ± 0.18
	others						2	0.0100				
								1.0000	(<i>P</i> =0.0995)	(<i>P</i> =0.9134)	(<i>P</i> =0.8591)	(<i>P</i> =0.1826)
Block 6		SNP15	5 SNP16	5 SNP1	7 SNF	P18 SNP	19					
	1	G	A	G	С	G	118	0.5900	104.2±43.3	11.3 ± 3.6	81.7±37.6	0.22 ± 0.11
	2	Т	G	А	Т	А	42	0.2100	92.9±29.5	11.5 ± 5.0	75.9±28.8	0.22 ± 0.087
	3	G	А	А	Т	G	12	0.0600	101.0 ± 54.6	10.6 ± 1.0	102.4 ± 75.3	0.19 ± 0.087
	4	G	G	А	Т	G	10	0.0500	114.3 ± 24.6	11.4 ± 2.1	83.2±32.8	0.28 ± 0.18
	5	G	A	G	т	G	8	0.0400	89.9±27.1	10.6 ± 1.0	100.6 ± 38.5	0.19 ± 0.07
	6	G	А	A	т	A	4	0.0200	97.1 ± 46.5	12.5 ± 3.1	55.9 ± 10.6	0.33 ± 0.29
	Others	-			-		6	0.0300				
							0	0.0000	(P=0.6063)	(<i>P</i> =0.9200)	(<i>P</i> =0.1335)	(P=0.1120)
Block 7		SNP23	SNP24	1 SNP2	5				(, , , , , , , , , , , , , , , , , , ,	(, 0,5200)	(, , , , , , , , , , , , , , , , , , ,	(, 01120)
2.0017	1	G	T	C	-		102	0.5100	100 0 + 43 5	11.5+4.8	81.1+36.4	0 20 + 0 088
	2	C	C	A			78	0.3900	102 5 + 37 0	11.0+1.8	87 0 + 43 0	0 24 + 0 12
	3	G	т	A			17	0.0850	100.2 + 28.3	10.8+1.6	76 8 + 34 7	0.25 ± 0.12
	4	G	C	Δ			-, 2	0.0150	105.6 + 52.8	133 + 35	60.5 ± 26.7	0.37 + 0.32
		J	<u> </u>				5	0.0100	(<i>P</i> _0 9752)	(P=0.5317)	(<i>P</i> _0 4797)	(P = 0.066)
									(1 -0.57 52)	(1 -0.0017)	(1 - 0.77 37)	(1 - 0.0000)

^a*P*-values were tested for equality by the ANOVA test.

But, there are some limitations to this study. In this study, we aimed to identify CYP19A1 loci that showing an association between hormone levels. Significant findings between hormone levels and marker loci do not warrant functional importance of the loci. Therefore, a lack of functional validation is one limitation of the study. The moderate number of cases in this study could be another limitation. Accumulation of evidence from the Asian population is strongly needed.

In conclusion, to the best of our knowledge, this is the first study to examine the association between tag SNPs in *CYP19A1* and sex hormone levels in postmenopausal Japanese women. Results showed an association between estrone and testosterone levels and some *CYP19A1* loci. These associations might be potential susceptibility

markers of hormone-dependent diseases in postmenopausal women. A further study of these associations to determine potential clinical applications is warranted.

ACKNOWLEDGEMENTS

We are grateful to the assistant staff at the Division of Epidemiology and Prevention at Aichi Cancer Center Research Institute for their support. This study was supported by a grant-in-aid for Scientific Research on Cancer Epidemiology in a Special Priority Area (C) (17015052) from the Ministry of Education, Science, Sports, Culture and Technology of Japan; a grant-in-aid for Scientific Research (C) from the Ministry of Education, Science, Sports, Culture and Technology of Japan; a grant-in-aid for the Third Term Comprehensive 10-Year Strategy for Cancer Control from the Ministry of Health, Labour and Welfare of Japan; and by a grant-in-aid for Cancer Research from the Ministry of Health, Labour and Welfare of Japan (19-19).

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