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Association between *PARP-1* V762A polymorphism and cancer susceptibility: a meta-analysis

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Abstract

Background—Poly(ADP-ribose) polymerase-1 (PARP-1 catalyzes poly(ADP-ribosyl)ation to various proteins involved in many cellular processes, including DNA damage detection and repair, and cell proliferation and death. PARP-1 has been implicated in human carcinogenesis, but the association between the most-studied *PARP-1* V762A polymorphism (rs1136410) and risk of various cancers was reported with inconclusive results.

Aims—To assess the association between the PARP-1 V762A polymorphism and cancer risk.

Methods—A meta-analysis of 21 studies with 12027 cancer patients and 14106 cancer-free controls was conducted to evaluate the strength of the association using odds ratio (OR) with 95% confidence interval (CI).

Results—Overall, no significant association was found between the *PARP-1* V762A polymorphism and cancer risk. In the stratified analyses, however, it was found that the variant A allele of the *PARP-1* V762A polymorphism was associated with an increased risk of cancer among Asian populations (VA+AA vs.VV: OR = 1.11, 95% CI: 1.01-1.23; $P_{heterogeneity} = 0.210$) but a decreased risk of cancer (VA+AA vs.VV: OR =0.89, 95% CI: 0.80-1.00; $P_{heterogeneity} = 0.004$), among Caucasian populations, especially for glioma risk (OR = 0.79, 95% CI: 0.69-0.90; $P_{heterogeneity} = 0.800$).

Conclusions—This meta-analysis found evidence for an association of the *PARP-1* V 762A polymorphism with increased risk of cancer among Asians but decreased risk of cancer among Caucasians, particularly of glioma. Further well designed studies with large sample sizes of different ethnic populations and different cancer types are warranted to confirm these findings.

Keywords

DNA repair; Case-control study; Meta-analysis; Polymorphism; Susceptibility

Introduction

Genomic DNA damage in mammalian cells can be caused by numerous physical and chemical agents, of which some are endogenous products, such as reactive oxygen species generated by normal cellular metabolism processes, while others are exogenous insults, such as ultraviolet right, ionizing radiation, and genotoxic chemicals. Such DNA damage, if left unrepaired or repaired incorrectly, may induce mutations and genomic instability that lead to cellular malignant transformation and tumorgenesis [Hoeijmakers 2001; Jackson and Bartek 2009]. Nevertheless, human cells have developed a set of complex DNA repair systems that

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safeguard the integrity of the genome to minimize the consequences of detrimental mutations [Hoeijmakers 2001]. At least four main repair pathways are involved in the removal of DNA damage: base excision repair (BER), nucleotide excision repair (NER), mismatch repair (MMR), and double-strand breaks repair (DSBR) [Decordier, et al. 2010; Hoeijmakers 2001]. Inherited defects in DNA damage response and repair pathways predispose to the development of malignancies [Hoeijmakers 2001]. Indeed, several human hereditary diseases, which are characterized by severe developmental problems and /or a predisposition to cancer, have been found to be linked to deficiencies in DNA repair [Friedberg 2008; Wiesmuller, et al. 2002].

Poly(ADP-ribose) polymerase-1 (PARP-1), also known as ADPRT, is a major member of the PARP family that contains 18 distinct proteins in humans [Kim, et al. 2005]. Human PARP-1 protein (113KDa) is a nuclear protein comprised of three functional domains (Figure 1). The N-terminal DNA-binding domain (DBD) harbors two zinc-finger motifs that are important for the binding of PARP1 to DNA-strand breaks. A third zinc finger was found to be important for coupling damage-induced changes in the DBD to alterations in PARP-1 catalytic activity [Rouleau, et al.2010]. In the central auto-modification domain, specific glutamate and lysine residues serve as potential acceptor sites for poly(ADP-ribosyl) ation. This domain also comprises a BRCA1 C-terminus motif, via which PARP-1 mediates protein-protein interactions. The third functional domain is a C-terminal catalytic domain containing a 'PARP-signature' motif, a highly evolutionally conservative sequence that forms the active site [Ame, et al. 2004; Kim, et al. 2005; Rouleau, et al. 2010].

Extensive studies of PARP-1 to date have implicated PARP-1 in a variety of cellular processes, all of which involve DNA-related transactions [Rouleau, et al. 2010] (Figure 1). Most importantly, PARP-1 functions as a DNA damage sensor in the DNA repair process [Kim, et al. 2005]. In general, in response to DNA damage generated either exogenously or endogenously, PARP-1 binds to sites of DNA damage through its DNA-binding domain, which activates the poly(ADP-ribosyl)ation of target proteins involved in numerous biological events. A growing number of evidence has shown that PARP-1 physically and functionally interacts with various proteins involved in multiple DNA repair pathways, including the BER, SSB, and DSBR pathways, and may recruit the repair proteins to sites of DNA damage (e.g., XRCC1 in BER, DNA-dependent protein kinase in DSBR repair), thereby plays a critical role in DNA damage repair [Burkle 2001; Kim, et al. 2005; Masson, et al. 1998; Ruscetti, et al. 1998]. In addition, PARP-1 is also involved in many other molecular and cellular processes, such as modulation of gene transcription, chromatin remodeling, maintaining telomeres and genomic stability, as well as cell proliferation and death [Gurung, et al. 2010; Masutani, et al. 2003; Masutani, et al. 2005; Tong, et al. 2001]. Accumulated evidence from the results of animal and cellular studies indicates that deficiency of *PARP-1* leads to DNA repair defects, genomic instability, failure of induction of cell death, and modulation of gene transcription, thereby contributing to carcinogenesis [Masutani, et al. 2003; Masutani, et al. 2005].

The human *PARP-1* gene is located on chromosome 1q41-42, consists of 23 exons, and spans 47.3 kb. To date, at least 439 single nucleotide polymorphisms (SNPs), including 17 non-synonymous SNPs (nsSNPs) in the *PARP-1* gene, have been reported, of which only one coding nsSNP (rs1136410, V762A) is common [minor allele frequency (MAF) >0.05] (http://www.ncbi.nlm.nih.gov/SNP) and most frequently studied for its association with cancer risk. *PARP-1* V762A is a base T to C transition at codon 762 in exon 17, which results in the substitution of alanine for valine in the catalytic domain of PARP-1, and *PARP-1* V762A polymorphism was reported to be associated with an altered activity of PARP-1 [Lockett, et al. 2004; Wang, et al. 2007].

To date, a number of molecular epidemiological studies have been conducted to evaluate the effect of the *PARP-1* V762A polymorphism on risk of many cancer types, including cancers of the brain [Liu, et al. 2009; McKean-Cowdin, et al. 2009; Rajaraman, et al. 2010; Yosunkaya, et al.2010], stomach [Miao, et al. 2006; Zhang, et al. 2009], breasts [Cao, et al. 2007; Haiman, et al. 2008; Smith, et al. 2008; Zhai, et al. 2006; Zhang, et al. 2006], colorectum [Brevik, et al. 2010; Stern, et al. 2009; Stern, et al. 2007], bladder [Figueroa, et al. 2007; Huang, et al. 2007; Wang, et al. 2010; Wu, et al. 2006], prostate [Gao, et al. 2010; Lockett, et al. 2004], head and neck [Li, et al. 2007], lung [Choi, et al. 2003; Landi, et al. 2006; Zhang, et al. 2005], esophagus [Hao, et al. 2004], and skin [Li, et al. 2006] as well as other types of cancer [Chiang, et al. 2008; Jin, et al. 2010; Rajaraman, et al. 2010; Shen, et al. 2006]. However, until now, those studies that investigated associations between the *PARP-1* V762A polymorphism and cancer risk have yielded inconsistent results. Considering the potential important role of *PARP-1* V762A polymorphism with overall cancer risk as well as the risk of specific type of cancers in different ethnic populations.

Materials and Methods

Literature search

For this meta-analysis, we searched for relevant studies by using PubMed software to search Medline (U.S. National Library of Medicine) to identify candidate studies. The latest search was performed on May 12, 2011. The following terms were jointly used for the search: '*PARP-1*' or '*ADPRT*', 'polymorphism or variant', 'cancer', 'tumor', or 'carcinoma'. In addition, additional studies were also identified by a manual search for the reference cited in the retrieved studies and review articles. Search results were limited to studies published in English.

Inclusion and exclusion criteria

All human-associated studies were included, if they met the following criteria: (1) evaluation of the *PARP-1* V762A polymorphism (rs1136410) and cancer risk, (2) case-control, nested case-control, or cohort study, (3) the study reported an odds ratio (OR) with 95% confidence interval (95% CI) or data for their calculation, and (4) the study was published in English, (5) Genotype distribution of control populations must be consistent with Hardy-Weinberg equilibrium (HWE). Exclusion criteria were: (1) duplicate data, (2) abstract, case report, comment, review and editorial, (3) no sufficient genotyping data were provided, (4) the outcome was benign tumors, precancerous lesions, and adenomas, (5) family-based study, (6) genotype distribution of control subjects was inconsistent with Hardy–Weinberg equilibrium (HWE). If studies had overlapped subjects, only the one with largest sample size was included in the final analysis.

Data extraction

Two investigators independently reviewed the articles and extracted the data from all eligible publications according to the criteria listed above. Disagreements were resolved by discussion between the two investigators. The following information was recorded for each study: first author, year of publication, country or region of origin, ethnicity, cancer type, number of cases and controls, number of cases and controls by genotype, source of control group (population-based or hospital-based), genotyping methods, minor allele frequency in controls. For studies including subjects of different ethnic descents, data were extracted separately for each ethnicity, which was categorized as Caucasian, Asian, and the mixed descendants (admixture of different ethnic groups).

Statistical analysis

Genotypic frequency for the PARP-1 V762A polymorphism was tested for deviation from HWE in control subjects using the chi-square goodness of fit, and a P value < 0.05 was considered significant. Odds ratios (OR) and corresponding 95% confidence interval (95% CI) were used to estimate the association between the PARP-1 V762A polymorphism and cancer risk. We estimated the risk for variant homozygous AA and heterogeneous VA genotypes, compared with the wild-type homozygous VV genotype, and then for the (VA + AA) versus (vs.) VV and AA versus (VA + VV), assuming recessive and dominant effect models, respectively. The statistical heterogeneity among studies was assessed with the Qtest and \hat{I} statistics [Lau, et al. 1997]. The heterogeneity across studies was considered significant when a *P* value < 0.1 for the Q statistic [Lau, et al. 1997]. If there was no significant heterogeneity, the fixed-effects model was used to estimate the summary OR and 95% CI; otherwise, the random-effects model was used [DerSimonian and Laird 1986]. To explore potential sources of heterogeneity across studies, we did stratification and metaregression analyses. Stratified analyses were conducted by ethnicity (Caucasian, Asian, and mixed), type of cancers and specific type of cancers in different ethnic populations (if one cancer type contains less than three studies, it was merged into the 'other cancers' group), source of controls (hospital-based and population-based), and genotyping methods. Publication bias was evaluated with the funnel plot and Begg's and Egger's tests [Begg and Mazumdar 1994; Egger, et al. 1997]. A P value < 0.05 was used as an indication for the presence of potential publication bias. Sensitivity analyses were conducted by removing one study at a time to assess the influence of individual studies on the pooled ORs. All analyses were conducted using Review Manager (v.5.0; Oxford, England) and Stata software (version 8.2; StataCorp LP, College Station, TX, USA). All the P values were two-sided.

Results

Characteristics of studies

The flowchart of study selection for this meta-analysis is presented in Figure 2. A total of 52 published records up to May 12, 2011 were identified, of which 29 were considered potentially eligible for inclusion in this meta-analysis and were retrieved in full texts. Among these, eight studies were excluded: three studies had the controls with genotype distributions inconsistent with HWE (P < 0.050) [Miao, et al. 2006; Yosunkaya, et al. 2010; Zhang, et al. 2009], two were duplicate publications [Huang, et al. 2007; Stern, et al. 2009], one was a family-based study [Brevik, et al. 2010], and two provided no genotyping data [Choi, et al. 2003; Haiman, et al. 2008]. Finally, 21 articles were selected in the metaanalysis. In addition, the studies investigating multiple types of cancers [Rajaraman, et al. 2010] or multiple ethnicities[Lockett, et al. 2004; Smith, et al. 2008] were separated into multiple studies in the subgroup analysis. One study [Huang, et al. 2007; Liu, et al. 2009] that only provided the total number of variant genotypes (VA + AA and VV) was included in the analysis for the dominant model but not for other genetic models. Main characteristics of the included studies are presented in Table 1. Overall, 21 publications including 12027 cases and 14106 controls were available for this meta-analysis. All studies were case-control studies, including two studies for glioma, one study for multiple types of cancer (adult meningioma, glioma, and acoustic neuroma), four studies for breast cancer, three studies for bladder cancer, and eleven studies for other types of cancers such as lung cancer, gastric cancer, melanoma, prostate cancer. Among 21 studies included in this meta-analysis, 11 studies were conducted in Caucasians, seven studies in Asians, three studies in the mixed ethnic populations (including Caucasians and African-Americans). The distributions of genotypes in the controls were all in agreement with HWE.

Quantitative synthesis

We included 12027 cancer patients and 14106 control subjects in this meta-analysis. We found that the frequency of the variant allele (A) of the *PARP-1* V762A polymorphism was significantly higher in Asians (0.423, 95% CI: 0.403 - 0.443) than in Caucasians (0.166, 95% CI: 0.154-0.178) (Figure 3). Overall, no significant association was found between the *PARP-1* V762A polymorphism and cancer susceptibility (Table 2, Figure 4). Individuals carrying the A allele did not have an altered cancer risk, compared with individuals with the wild-type VV homozygous genotype (VA vs. VV: OR = 0.97, 95% CI: 0.90-1.06, *P*_{heterogeneity} = 0.010; AA vs. VV: OR = 0.99, 95% CI: 0.84-1.17; *P*_{heterogeneity} = 0.010). Similarly, no significant association with the risk of cancer was found in either a dominant model (VA+AA vs. VV: OR = 0.96, 95% CI: 0.88-1.05, *P*_{heterogeneity} = 0.0003) or a recessive model (AA vs. VA+VV: OR =1.03, 95% CI: 0.93 -1.14, *P*_{heterogeneity} = 0.100) (Table 2).

In the stratified analysis by ethnicity, a significantly increased cancer risk was found among Asians in the dominant model (VA+AA vs. VV: OR = 1.11, 95% CI: 1.01-1.23, $P_{\text{heterogeneity}} = 0.210$) (Table 2, Figure 5). In contrast, the *PARP-1* V762A polymorphism was significantly associated with a decreased cancer risk among Caucasians in the dominant model (VA+AA vs. VV: OR =0.89, 95% CI: 0.80-1.00, $P_{\text{heterogeneity}} = 0.004$) (Table 2, Figure 6). No significant associations were detected in all genetic models in the mixed ethnicity population group. No significant associations were found when the data were stratified by the source of control subjects and by genotyping methods (Table 2).

Further stratified analyses by specific type of cancers and ethnicity were performed. In Caucasians, the VA heterozygous genotype was found to be associated with significantly decreased risk of glioma compared with the common VV homozygous genotype (VA vs. AA: OR = 0.81, 95% CI: 0.70-0.95, $P_{heterogeneity} = 0.670$). A significant main effect was also observed in the dominant model for glioma (VA+AA vs. VV: OR = 0.79, 95% CI: 0.69-0.90, $P_{heterogeneity} = 0.800$). However, no significant association was found in ether breast cancer or other cancers (each type of cancers having less than three studies was merged into the 'other cancers' group) (Table 3). We were not able to further evaluate the effect of the *PARP-1* V762A polymorphism on cancer risk by specific type of cancers in Asian and in the mixed ethnicity populations because only one studies of specific type of cancers were included in these two groups.

Evaluation of heterogeneity

In the present study, heterogeneity across studies was observed in overall comparisons. Therefore, we assessed the source of heterogeneity for the dominant model (VA+AA vs. VV) by ethnicity, type of cancers, source of controls, and genotyping methods. We found that ethnicity (P = 0.044) and type of cancers (P = 0.038), but not source of controls (P = 0.903) and genotyping techniques (P = 0.146), contributed to substantial heterogeneity.

Sensitivity analysis

In the sensitivity analysis, the influence of each study on the pooled OR was examined by repeating the meta-analysis while omitting each study, one at a time. Such a leave-one-out sensitivity analysis indicated that no single study influenced the pooled ORs qualitatively (**data not shown**), which indicted the stability of our overall result.

Publication bias

Begger's funnel plot and Egger's test were performed to evaluate the publication bias of the publication bias of literature on cancer. The shape of the funnel plot seemed asymmetrical

(Figure 7), which was confirmed by an Egger's test (t = -0.74, P = 0.465 for VA+AA vs. VV).

Discussion

Carcinogenesis is a multi-step process involving aberrations in a variety of cellular processes including DNA damage detection and repair, genome maintenance, cell-cycle control, proliferation, differentiation, and cell death [Kim, et al. 2005]. PARP-1 catalyzes poly(ADP-ribosyl)ation, which was functionally involved in all of these processes, suggesting a possible connection between PARP-1 function and development of cancer [Kim, et al. 2005; Masutani, et al. 2003]. A body of evidence shows that PARP-1 plays a critical role in human carcinogenesis [Bhatia, et al. 1990a; Bhatia, et al. 1990b; Bieche, et al. 1996; Lyn, et al. 1993; Masutani, et al. 2005; Prasad, et al. 1990; Shiokawa, et al. 2005]. It has been found that *PARP-1* knockout mice (*PARP-1^{-/-}*) treated with either alkylating agents or γ -irradiation displayed high genomic instability, high frequencies of chromosome aberrations, and shortened telomere compared with the wild-type mice [d'Adda di Fagagna, et al. 1999; de Murcia, et al. 1997; Samper, et al. 2001]. PARP-1-/-mice also showed an increased susceptibility to chemically-induced tumorigenesis in the liver, colon, and lung [Nozaki, et al. 2003; Tsutsumi, et al. 2001], although in certain situations, PARP-1 may also facilitate the growth of tumor cells [Martin-Oliva, et al. 2004] . For example, Martin-Olva et al. found that PARP-1^{-/-} mice displayed a strikingly reduced susceptibility to skin carcinogenesis [Martin-Oliva, et al. 2004]. Dysregulation of PARP-1 expression has been found in a variety of human cancers, such as breast cancer, head and neck cancer, melanoma, and colorectal cancer [Goncalves, et al. 2010; Nosho, et al. 2006; Staibano, et al. 2005]. Furthermore, reduced PARP-1 activity in human peripheral blood lymphocytes has been observed in patients with cancers of the breasts, colon, lung, larynx, and prostate as well as glioma [Barton, et al. 2009; Lockett, et al. 2004; Pero, et al. 1990; Rajaee-Behbahani, et al. 2002]. Therefore, it is biologically plausible that the PARP-1 V762A polymorphism residing in the catalytic domain of the PARP-1 enzyme, which causes an amino acid change from Val to Ala, may alter the enzymatic activity of PARP-1 and thereby contribute to cancer susceptibility.

In the present meta-analysis with 12027 cancer cases and 14106 controls, no significant association between the *PARP-1* V762A polymorphism and overall risk of cancer was found. However, in the subgroup analyses by ethnicity, the variant 762A allele was found to be associated with an increased risk of cancer among Asian populations but a decreased risk of cancer, particularly glioma, among Caucasian populations,. These findings indicate that the *PARP-1* V762A polymorphism may play a role in cancer development, at least in some ethnic groups or some specific cancer types. However, it is still unclear whether the same polymorphism may have the same effect across different types of cancer, though evidence has shown that a few SNPs, such as rs401681 in *CLPTM1L*, are associated with risk of many cancer types including basal cell carcinoma and cancers of the lung, bladder, prostate, and cervix [Rafnar, et al. 2009]. Therefore, our results should be interpreted cautiously. Nevertheless, our findings are interesting and worthy of further investigation.

If significant heterogeneity is present, pooled summary estimates from such meta-analyses are hard to interpret. In our meta-analysis, obvious heterogeneity across studies was observed in the overall comparison and some subgroup analyses. We then used the meta-regression analysis to explore the sources of heterogeneity. We found that ethnicity and type of cancers might contribute to the potential heterogeneity, suggesting that the *PARP-1* V762A polymorphism may modulate cancer risk differently depending on ethnicity and type of cancers. Indeed, in the stratified analysis by ethnicity, the effect of the variant A allele of the *PAPR-1* V762A polymorphism appeared to be associated with cancer risk more likely in

Asians than in Caucasians. When stratifying by type of cancers, we found that the variant A allele of this polymorphism was associated with a decreased risk for glioma in Caucasians but not for other types of cancer in the dominant model, and no heterogeneity was found across the studies.

In our meta-analysis, we found that the minor allele frequencies of the *PARP-1* V762A polymorphism exhibited striking racial differences. The A allele frequency of Asians was significantly higher (42.3%) than that of Caucasians (16.6%). Similarly, in the HapMap database, the frequency of the minor A allele among Asians (45%) is also higher than that of European (15%) (http://hapmap.ncbi.nlm.nih.gov/). To date, no association study between the *PARP-1* V762A polymorphism and glioma risk has been reported in Asians. Therefore, further studies are needed to explore the role of the *PARP-1* V762A polymorphism in the development of different type of cancers including glioma in different ethnic populations.

Some potential limitations in this meta-analysis should be considered. Firstly, only papers published in English were included in this meta-analysis, and this could have introduced bias in our results. Secondly, obvious heterogeneity across studies, which might result from different ethnicities and types of cancer, existed in overall comparisons and also some subgroup analyses. Although various genotyping techniques were used in those studies used in our meta-analysis, including polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP), TaqMan, and MassARRAY assays, it did not contribute to the observed heterogeneity across studies (P = 0.146). Thirdly, the small sample sizes in some subgroup analyses may have limited statistical power. Furthermore, due to limited studies included in this meta-analysis, we were unable to perform further subgroup analyses, such as by specific type of cancers in Asians. Fourthly, our results were based on unadjusted OR estimates, because not all studies included in this meta-analysis provided adjusted ORs or the ORs were not adjusted by the same potential confounders, such as age, sex, ethnicity, and other environmental factors. A more precise analysis should be conducted if individual data were available. Fifthly, despite the confirmatory findings from this meta-analysis, we can not rule out the possibility that the current significant results might be due to chance, because many factors, such as those we mentioned above and potential publication bias that may exist but was undetectable in this meta-analysis, could have an impact on our analysis. Therefore, these results should be interpreted with caution and confirmed from additional analysis with more published studies in the future. Finally, evaluation of the potential genegene and gene-environment interaction effects on the risk of cancer was limited because of lacking the original data of the reviewed studies.

In conclusion, our meta-analysis with a total of 12027 cases and 14106 controls suggests that the variant A allele of *PARP-1* V763A polymorphism may increase the risk of cancer in Asian populations but decrease the risk of cancer, especially for the risk of glioma, in Caucasian populations. Further well designed studies with large sample sizes of different ethnic populations and different cancer types are warranted. Furthermore, the effects of gene-gene and gene-environment interactions on the risk of cancer should also be taken into account.

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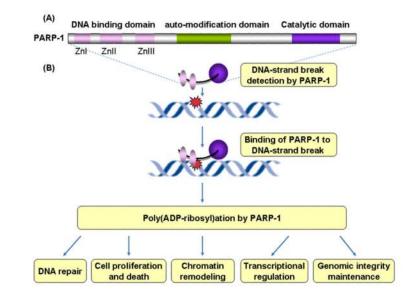


Figure 1.

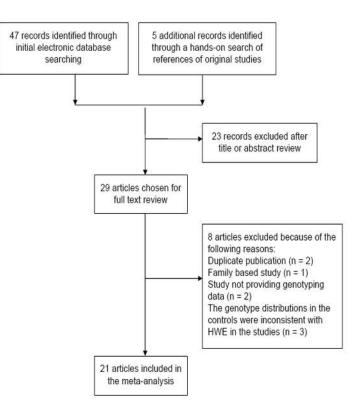


Figure 2.

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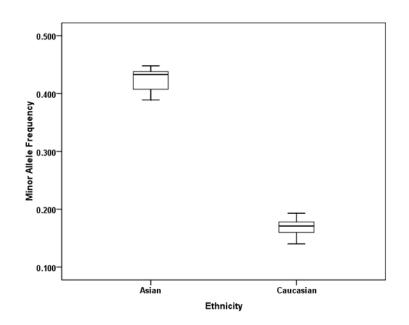


Figure 3.

Study	Cancer*	Cases	Controls	Odds Ratio	Odds Ratio
				M-H, Random, 95% CI	M-H, Random, 95% Cl
Cao2007	Breast	83	100	0.71 [0.36, 1.41]	
Chiang2008	Thyroid	283	469	1.28 [0.93, 1.75]	
Figueroa2007	Bladder	1138	1131	1.28 [1.06, 1.55]	
Gao2010	Prostate	453	119	0.90 [0.58, 1.38]	
Hao2004	Esophagus	414	479	1.25 [0.94, 1.65]	+
Jin2010	NHL	573	721	0.90 [0.71, 1.14]	
Landi2006	Lung	292	307	0.90 [0.64, 1.28]	
Li 2006	Melanoma	602	603	0.82 [0.64, 1.05]	
Li2007	Head and neck	830	854	0.78 [0.63, 0.97]	
Liu2009	Glioma	372	365	0.72 [0.53, 0.98]	
Lockett2004	Prostate	488	524	1.27 [0.96, 1.69]	+
McKean-Cowdin2009	Glioblastoma	987	1935	0.79 [0.67, 0.94]	
Rajaraman2010	Multiple	526	464	0.77 [0.59, 1.02]	
Shen2006	NHL	455	535	0.86 [0.66, 1.13]	-+
Smith2008	Breast	366	469	0.79 [0.58, 1.09]	
Stern 2007	Colorectum	307	1173	1.11 [0.84, 1.45]	- - -
Wang2010	Bladder	234	253	1.09 [0.74, 1.60]	
Wu2006	Bladder	606	595	0.90 [0.70, 1.15]	
Zhai2006	Breast	302	639	0.90 [0.67, 1.21]	-+-
Zhang 2005	Lung	1000	1000	1.26 [1.05, 1.52]	
Zhang 2006	Breast	1716	1371	1.03 [0.88, 1.20]	+
Total (95% CI)		12027	14106	0.96 [0.88, 1.05]	•

Figure 4.

Study	Cases	Controls	Odds Ratio M-H, Fixed, 95% Cl	Odds Ratio M-H, Fixed, 95% Cl
Chiang2008	283	469	1.28 [0.93, 1.75]	 • -
Hao2004	414	479	1.25 [0.94, 1.65]	+
Jin2010	573	721	0.90 [0.71, 1.14]	
Stern 2007	307	1173	1.11 [0.84, 1.45]	
Wang2010	234	253	1.09 [0.74, 1.60]	- <u>-</u> -
Zhai2006	302	639	0.90 [0.67, 1.21]	
Zhang 2005	1000	1000	1.26 [1.05, 1.52]	
Total (95% CI)	3113	4734	1.11 [1.01, 1.23]	•
Heterogeneity: Chi ² = 8.	40, df = 6 (P = 0.21); l² = 29%	H0.2	
Test for overall effect: Z	= 2.11 (P = 0.04)		0.2	. 0.0 1 2 0

Figure 5.

Study	Cases	Controls	N	Odds Ratio I-H, Random, 95% Cl	Odds Ratio M-H, Random, 95% CI
Cao2007	83	100		0.71 [0.36, 1.41]	*1
Figueroa2007	1138	1131		1.28 [1.06, 1.55]	
Gao2010	453	119		0.90 [0.58, 1.38]	
Landi2006	292	307		0.90 [0.64, 1.28]	
Li 2006	602	603		0.82 [0.64, 1.05]	
Li2007	830	854		0.78 [0.63, 0.97]	17.7.7.7.7
Liu2009	372	365		0.72 [0.53, 0.98]	
Lockett2004	438	427		1.18 [0.88, 1.59]	
McKean-Cowdin2009	987	1935		0.79 [0.67, 0.94]	111111
Rajaraman2010	526	464		0.77 [0.59, 1.02]	
Smith2008	314	397		0.72 [0.52, 1.00]	
Wu2006	606	595		0.90 [0.70, 1.15]	
Zhang 2006	. 1716	1371		1.03 [0.88, 1.20]	+
Total (95% CI)	8357	8668		0.89 [0.80, 1.00]	•
Heterogeneity: Tau ² = 0.02;	Chi ² = 29.34, df =	12 (P = 0.004);	l² = 59%	H	0.5 1 2 5
Test for overall effect: Z = 2	.01 (P = 0.04)			0.2	0.5 1 2 5

Figure 6.

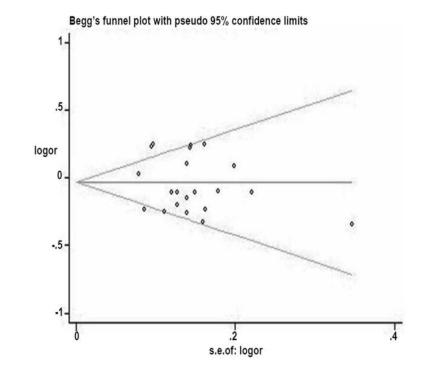


Figure 7.

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Table 1

Characteristics of studies included in the meta-an analysis

First author	Year	Country	Ethnicity ^a	Types of cancer b	OR (95%CI) ^C (VA +AA vs. VV)	Sample size (Cases /Controls)	Source of controls	Genotyping method ^g
Lockett	2004	America	Mixed	Prostate Cancer	1.27 (1.11 - 1.46) ^d	488 / 524	hospital-based	Mass ARRAY
Hao	2004	China	Asian	ESCC	1.25 (1.07 - 1.46) ^d	414/479	hospital-based	PCR-RFLP
Zhang	2005	China	Asian	Lung Cancer	1.68 (1.27 - 2.23) ^d	1000 / 1000	hospital-based	PCR-RFLP
Zhang	2006	America	Caucasian	Breast Cancer	1.03 (0.96 - 1.11) ^d	1716 / 1371	population-based	TaqMan
Zhai	2006	China	Asian	Breast Cancer	0.87~(0.64 - 1.19) e	302 / 639	hospital-based	PCR-RFLP
Li	2006	America	Caucasian	Cutaneous Melanoma	0.94 (0.68 - 1.30) ^d	602 / 603	hospital-based	PCR-RFLP
Shen	2006	America	Multiple	THN	0.87 (0.66 - 1.15) ^d	455 / 535	population-based	TaqMan
Wu	2006	America	Caucasian	Bladder Cancer	0.89 (0.68 - 1.15) ^d	606 / 595	hospital-based	TaqMan
Landi	2006	Multiple regions	Caucasian	Lung Cancer	0.90 (0.75 - 1.08) ^d	292 / 307	hospital-based	APEX
Li	2007	America	Caucasian	SCCHN	0.79 (0.63 - 1.00) ^e	830 / 854	hospital-based	PCR-RFLP
Cao	2007	France	Caucasian	Breast Cancer	0.71 (0.48 - 1.06) ^d	83 / 100	hospital-based	Sequence
Figueroa	2007	Spain	Caucasian	Bladder Cancer	1.24 (1.02 - 1.51) ^d	1138 / 1131	hospital-based	TaqMan
Stern	2007	Singapore	Asian	Colorectal Cancer	1.1 (0.8 - 1.4) ^e	307 / 1173	population-based	TaqMan
Chiang	2008	Taiwan	Asian	Thyroid Carcinoma	1.28 (1.04 - 1.56) ^d	283 / 469	hospital-based	TaqMan
Smith	2008	America	Multiple	Breast Cancer	0.79 (0.66 - 0.96) ^d	366 / 469	hospital-based	MassARRAY
McKean-Cowdin	2009	America	Caucasian	Glioblastoma	0.80 (0.67 - 0.95) ^e	987 / 1935	hospital-based and population-based	MassARRAY
Liu	2009	America	Caucasian	Glioma	0.71 (0.52 - 0.97) ^e	372 / 365	population-based	MassARRAY
Jin	2010	Korea	Asian	THN	0.90 (071 - 1.04) <i>e</i> Glioma: 0.86 (0.63 - 1.17) <i>e</i>	573 / 721	population-based	PCR-HRM
Rajaraman	2010	America	Caucasian	Multiple Cancers	Meningioma: 0.67 (0.42 - 1.07) e Acoustic neuroma: 0.53 (0.28 - 1.00) e	526/464	hospital-based	TaqMan
Wang	2010	China	Asian	Bladder Cancer	1.08 (0.89 - 1.04) ^d	234 / 253	hospital-based	PCR-RFLP
Gao	2010	America	Caucasian	Prostate Cancer	0.90 (0.82 - 0.99) ^d	453 / 119	hospital-based	Sequence
^a Ethnicity ^a : Mixed: mixed ethnicities.	mixed et	thnicities.						

b cancer types: ESCC: Esophageal Squamous Cell Carcinoma; NHL: Non-Hodgkin lymphoma; SCCHN: Squamous Cell Carcinoma of the Head; and Neck; Multiple cancers: Meningioma, Glioma, and Acoustic neuroma.

 $c_{VA + AA v.s. VV: Dominant model.}$

 $d_{\rm U}$ nadjusted odds ratio (OR) and 95% Confidence Interval (CI).

^eAdjusted OR (95 CI %).

iPLEX SpectroCHIP bioarrays; APEX: polymorphism was analyzed together for a given sample by a microarray technique based on the arrayed primer extension principle; PCR-RFLP: Polymerase Chain Reaction-restriction Fragment Length Polymorphism; PCR-HRM: PCR cycling and high resolution melting (HRM) analysis was performed on the Rotor-Gene 6000TM. genotyping method: MassARRAY: Genotyping was performed using the Sequenom MassARRAY iPLEXTM platform2. MassARRAY Workstation version 3.3 software was used to process and analyze

Meta-analysis for the association between the PARP-1 V762/A polymorphism and cancer risk

لامشمالهم	No. of subjects		VA vs. VV		AA vs. VV		$VA + AA v_S$, VV^b		AA vs. VA +VV ^c	
v ariables	Cases / Controls	'n	OR (95% CI)	pd	OR (95% CI)	pd	OR (95% CI)	pd	OR (95% CI)	p d
Total	12027 / 14106	21	0.97 (0.90-1.06)	0.010	0.99 (0.84-1.17)	0.010	0.96 (0.88-1.05)	0.0003	1.03 (0.93-1.14)	0.100
Ethnicities										
Asian	3113 / 4734	٢	1.09 (0.98-1.21)	0.560	1.16 (0.95-1.41)	0.070	1.11 (1.01-1.23)	0.210	1.12 (0.99-1.26)	0.230
Caucasian	8357 / 8668	13	0.91 (0.81-1.02)	0.009	0.84 (0.70-1.02)	0.280	$0.89\ (0.80-1.00)$	0.004	0.86 (0.71-1.04)	0.350
$Mixed^{e}$	557 / 704	ю	0.93 (0.71-1.21)	0.250	0.66 (0.26-1.69)		0.91 (0.70-1.18)	0.240	1.12 (0.99-1.26)	ı
Types of cancers										
Glioma	1699 / 2764	3	0.81 (0.70-0.95)	0.670	0.69 (0.45-1.07)	0.610	0.79 (0.69-0.90)	0.800	0.73 (0.48-1.13)	0.580
Breast cancer	2467 / 2579	4	0.96 (0.85-1.10)	0.330	$0.90\ (0.68-1.18)$	0.850	0.96 (0.84-1.08)	0.370	0.92 (0.71-1.19)	0.860
Bladder cancer	1978 / 1979	б	$1.10\ (0.84-1.44)$	0.060	0.99(0.70-1.40)	0.850	1.09 (0.86-1.39)	0.080	$0.96\ (0.69-1.33)$	0.820
Other cancers f	5883 / 7712	13	0.98 (0.91-1.07)	0.100	1.05 (0.83-1.34)	0.006	0.98 (0.86-1.11)	0.004	1.04 (0.86-1.27)	0.040
Sources of controls										
Population-based	3423 / 4165	5	1.00 (0.89-1.12)	0.600	0.94 (0.77-1.14)	0.520	0.95 (0.86-1.05)	0.190	0.95 (0.79-1.13)	0.670
Hospital-based	7617 / 8006	15	0.99 (0.89-1.10)	0.010	1.03 (0.83-1.29)	0.010	0.99 (0.88-1.12)	0.001	1.03 (085-1.24)	0.060
Other ^g	987 / 1935	-								ı
Genotyping methods										
PCR-RFLP	3382 / 3828	9	0.98 (0.83-1.16)	0.050	1.06 (0.79-1.43)	0.020	0.020 1.00 (0.82-1.21)	0.005	1.05 (0.84-1.33)	0.080
Other h	8645 / 10278	15	0.96 (0.83-1.11)	0.150	0.97 (0.88-1.07)	0.030	0.95 (0.85-1.05)	0.004	0.96 (0.84-1.11)	0.300
^a Number of comparisons.	ns.									
$b_{VA} + AA vs VV$: Dominant model	minant model									
$c_{AA vs VV + VA: Recessive model}$	essive model									
d value of Q-test for heterogeneity test. The random-effects model was used when P-value for heterogeneity test < 0.1; otherwise, the fixed-effects model was used.	leterogeneity test. The	randc	om-effects model wa	ts used w	/hen P-value for het	erogeneit	y test < 0.1; otherw	ise, the fiv	ced-effects model w	as used.

Genet Epidemiol. Author manuscript; available in PMC 2013 January 01.

 e^{θ} Mixed: one comparison between the AA genotype and the VV genotype, and one comparison for the recessive model (AA vs.VA/VV).

 $^{\mathcal{B}}$ Other: Controls of this study (McKean-Cowdin, et al.) were from both hospital-based controls and population-based controls.

 $f_{\rm o}$ ther cancers: each type of cancer which contains less than three studies was merged into the 'other cancers' group.

 h Other The genotyping methods included TaqMan, MassARRAY assays, Sequence, ant the others.

Yu et al.

Table 3

Meta-analysis for the association between the PARP-I V762/A polymorphism and the risk of different types of cancers in Caucasians

:	No. of subjects	1	VA vs. VV		AA vs. VV		$VA + AA vs. VV^b$	$q\Lambda_{1}$	AA vs. VA +VV ^c	νVc
Variables	Cases / Controls	_p u	OR (95% CI)	P^{d}	OR (95% CI)	p d	OR (95% CI) Pd	p d	OR (95% CI)	P^{d}
Types of cancers		15								
Glioma	1699 / 2764	ю	0.81 (0.70-0.95)	0.670	0.69 (0.45-1.07)	0.610	$0.81 \ (0.70 - 0.95) 0.670 0.69 \ (0.45 - 1.07) 0.610 0.79 \ (0.69 - 0.90) 0.800 0.73 \ (0.48 - 1.13) 0.580 0.580 0.73 \ (0.48 - 1.13) 0.580 0.580 0.73 \ (0.48 - 1.13) 0.580 0.$	0.800	0.73 (0.48-1.13)	0.580
Breast cancer	2113 / 1868	б	0.86 (0.63-1.17)	060.0	0.91 (0.63-1.32)	0.660	$0.86\ (0.63-1.17) 0.090 0.91\ (0.63-1.32) 0.660 0.95\ (0.83-1.09) 0.110 0.92\ (0.64-1.32) 0.670$	0.110	0.92 (0.64-1.32)	0.670
Other cancers e	4545 / 4964	6	0.93 (0.79-1.08) 0.020 0.86 (0.66-1.11) 0.170 0.91 (0.78-1.07) 0.008 0.87 (0.67-1.13) 0.200	0.020	0.86 (0.66-1.11)	0.170	0.91 (0.78-1.07)	0.008	0.87 (0.67-1.13)	0.200
^a Number of comparisons.	sons.									
bVA /AA vs VV: Dominant model	minant model									
c AA vs VV /VA: Recessive model	cessive model									
$\frac{d}{d}$ value of 0-test fir heteroreneity test. The random-effects model was used when B value for heteroreneity test ≤ 0.1 , otherwise, the fixed-effects model was used	r heterogeneity test	The rai	ndom-effects model	9311 36/M	d when <i>P</i> -value for	heteroae	meity test < 0.1: oth	amica th	o fived effects mo	يا مس اما

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 e Other cancers: each type of cancer which contains less than three studies was merged into the 'other cancers' group.