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Association of Dietary Patterns With Risk of Colorectal Cancer Subtypes Classified by *Fusobacterium nucleatum* in Tumor Tissue

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IMPORTANCE Fusobacterium nucleatum appears to play a role in colorectal carcinogenesis through suppression of the hosts' immune response to tumor. Evidence also suggests that diet influences intestinal *F* nucleatum. However, the role of *F* nucleatum in mediating the relationship between diet and the risk of colorectal cancer is unknown.

OBJECTIVE To test the hypothesis that the associations of prudent diets (rich in whole grains and dietary fiber) and Western diets (rich in red and processed meat, refined grains, and desserts) with colorectal cancer risk may differ according to the presence of *F* nucleatum in tumor tissue.

DESIGN, SETTING, AND PARTICIPANTS A prospective cohort study was conducted using data from the Nurses' Health Study (June 1, 1980, to June 1, 2012) and the Health Professionals Follow-up Study (June 1, 1986, to June 1, 2012) on a total of 121700 US female nurses and 51529 US male health professionals aged 30 to 55 years and 40 to 75 years, respectively (both predominantly white individuals), at enrollment. Data analysis was performed from March 15, 2015, to August 10, 2016.

EXPOSURES Prudent and Western diets.

MAIN OUTCOMES AND MEASURES Incidence of colorectal carcinoma subclassified by *F nucleatum* status in tumor tissue, determined by quantitative polymerase chain reaction.

RESULTS Of the 173 229 individuals considered for the study, 137 217 were included in the analysis, 47 449 were male (34.6%), and mean (SD) baseline age for men was 54.0 (9.8) years and for women, 46.3 (7.2) years. A total of 1019 incident colon and rectal cancer cases with available *F nucleatum* data were documented over 26 to 32 years of follow-up, encompassing 3 643 562 person-years. The association of prudent diet with colorectal cancer significantly differed by tissue *F nucleatum* status (P = .01 for heterogeneity); prudent diet score was associated with a lower risk of *F nucleatum*-positive cancers (P = .003 for trend; multivariable hazard ratio of 0.43; 95% CI, 0.25-0.72, for the highest vs the lowest prudent score quartile) but not with *F nucleatum*-negative cancers (P = .47 for trend, the corresponding multivariable hazard ratio of 0.95; 95% CI, 0.77-1.17). There was no significant heterogeneity between the subgroups in relation to Western dietary pattern scores.

CONCLUSIONS AND RELEVANCE Prudent diets rich in whole grains and dietary fiber are associated with a lower risk for *F nucleatum*-positive colorectal cancer but not *F nucleatum*-negative cancer, supporting a potential role for intestinal microbiota in mediating the association between diet and colorectal neoplasms.

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Corresponding Author: Shuji Ogino, MD, PhD, Division of MPE Molecular Pathological Epidemiology, Brigham and Women's Hospital and Harvard Medical School, 450 Brookline Ave, DFCI Room SM1036, Boston, MA 02215 (shuji_ogino@dfci.harvard.edu). ccumulating evidence suggests that the human gut microbiome is linked to colorectal cancer development.¹⁻⁴ *Fusobacterium nucleatum* has been found to be enriched in colorectal cancer tissue relative to normal adjacent colonic tissue and is detected at higher levels in stool among individuals with colorectal cancer compared with those without cancer.^{1,5-10} Recent experimental data suggest that *F nucleatum* may contribute to colorectal carcinogenesis through modulation of host immunity and activation of pathways associated with cellular proliferation.^{9,11,12} Furthermore, a higher amount of *F nucleatum* in colorectal cancer tissue has been linked to shorter survival, proximal tumor location, and specific tumor molecular features, such as highlevel CpG island methylator phenotype and microsatellite instability.¹³⁻¹⁵

Prudent dietary patterns-rich in fruits, vegetables, and whole grains-have been associated with a lower risk of colorectal cancer and adenoma¹⁶⁻²¹ as reviewed in a recent systematic meta-analysis.²² In contrast, Western dietary patternsdominated by red and processed meats-have been linked with colorectal carcinogenesis.^{16,22} Although mechanisms underlying these diet-cancer associations remain unclear, it is postulated that the gut microbiota may play a mediating role.²³ Recently, in a dietary intervention study, stool F nucleatum levels markedly increased after participants were switched from a prudent-style, high-fiber, low-fat diet to a low-fiber, high-fat diet.²⁴ In addition, accumulating data suggest that low fiber consumption and high meat intake may be associated with altered bacterial and metagenomic profiles as well as an inflammatory phenotype determined by serum levels of metabolites.25-28

Based on these findings, we hypothesized that the inverse association between prudent diets and risk of colorectal cancer might be more evident for a cancer subgroup enriched with tissue *F nucleatum* than for a subgroup without detectable tissue *F nucleatum*. To test this hypothesis, we used 2 US nationwide, prospective cohort studies: the Nurses' Health Study (NHS) (June 1, 1980, to June 1, 2012) and the Health Professional Follow-up Study (HPFS). These 2 studies offered a unique opportunity to integrate prospectively collected, regularly updated dietary intake data with tissue microbial features in incident colorectal cancers that occurred over long-term follow-up.

Methods

Study Population

We used data drawn from 2 ongoing prospective cohort studies, the NHS and the HPFS. The NHS began in 1976 among 121 700 US female nurses aged 30 to 55 years at enrollment. The HPFS began in 1986 among 51 529 US male health professionals aged 40 to 75 years at enrollment. In both cohorts, participants have returned questionnaires every 2 years, with follow-up rates exceeding 90%, to provide information about lifestyle and dietary factors, medication use, and diagnoses of colorectal cancer and other diseases. The institutional review board at the Brigham and Women's Hospital and Harvard **Question** Does the association between prudent diets (rich in whole grains and dietary fiber) and risk of colorectal cancer vary by presence of the bacterial species *Fusobacterium nucleatum* in tumor tissue?

Findings In this cohort study of 137 217 adults, the association of a prudent diet with colorectal cancer was more evident for a cancer subgroup enriched with tumor *F nucleatum* than a subgroup without detectable tumor *F nucleatum*.

Meaning There may be a potential role for intestinal microbiota, such as *F nucleatum*, in mediating the complex association between diet and the development of colorectal cancer.

T.H. Chan School of Public Health approved this study, and informed consent was obtained from all participants. The study was conducted from June 1, 1980, to June 1, 2012.

Of 173 229 individuals considered for the study, a total of 137 217 individuals (47 449 men and 89 768 women) were included in this analysis. We excluded participants with implausibly high or low caloric intakes (ie, <600 or >3500 kcal/d for women and <800 or >4200 kcal/d for men), missing dietary pattern data, or those with a history of ulcerative colitis or cancer (except for nonmelanoma skin cancer) before baseline (1980 for the NHS and 1986 for the HPFS) (eMethods in the Supplement).

Assessment of Diet

Participants reported average food intake over the preceding year (of each questionnaire return) through semiquantitative food frequency questionnaires, which have been previously validated and described.²⁹ Total nutrient intake was calculated by summing intakes from all foods and adjusted for total energy intake by the residual method. As previously described, total dietary fiber was calculated according to methods from the Association of Official Analytic Chemists.³⁰ For this analysis, we used information from food frequency questionnaires administered in the following years: 1980, 1984, 1986, 1990, 1994, 1998, 2002, 2006, and 2010 for the NHS and 1986, 1990, 1994, 1998, 2002, 2006, and 2010 for the HPFS.

Assessment of Colorectal Cancer Cases

In both cohorts, incident cases of colorectal cancer were reported by participants through the 2012 follow-up for the HPFS and NHS. We identified and confirmed lethal colorectal cancer cases through information from various sources including next of kin, the National Death Index, death certificates, and medical records. A study physician (including J.A.M. and C.S.F.), blinded to exposure information, reviewed records and extracted data on histologic type, anatomical location, and stage. The cohort study groups attempted to collect formalin-fixed, paraffin-embedded (FFPE) tissue specimens from hospitals throughout the United States as previously detailed.⁹ Cases with available tissue data (n = 1019) for the present study were similar to those without tissue data (n = 2241) regarding patient and clinical characteristics (eMethods in the Supplement).

F nucleatum Analysis

We extracted DNA from colorectal cancer tissue obtained from sections of FFPE tumor blocks (QIAamp DNA FFPE tissue kits; Qiagen). We performed a real-time polymerase chain reaction (PCR) assay using custom TaqMan primer/probe sets (Applied Biosystems) for the *nusG* gene of *F nucleatum*.⁹ The interassay coefficient of variation of cycle threshold values from each of 5 selected specimens in 5 different batches was less than 1% for all targets in the validation study.¹⁴ *Fusobacterium nucleatum* positivity was defined as a detectable level of *F nucleatum* nucleatum DNA within 45 PCR cycles, and *F nucleatum* negativity was defined as an undetectable level with a proper amplification of human reference gene *SLCO2A1* (HGNC: 10955).

Statistical Analysis

All statistical tests were 2-sided. To account for multiple testing for the 2 primary hypotheses (related to prudent and Western dietary scores) associated with the 2 tumor subtype variables, we adjusted the 2-sided α level to .01 (approximately .05/4) by simple Bonferroni correction in our primary and secondary analysis.

Two maximally uncorrelated dietary patterns—one named *prudent* and another named *Western*—were derived by principal component analysis, as previously described and validated with good reproducibility.^{16,31} Factor loadings were derived based on the correlations between food groups and the 2 derived factors. Each participant was assigned a factor score, determined by adding the reported frequencies of food group intakes weighted by the factor loadings. These factor scores were then standardized to have a mean (SD) of 0 (1). To capture long-term habitual consumption, we calculated the cumulative mean of the prudent (or Western) dietary pattern scores from preceding food frequency questionnaires up to each questionnaire cycle. Then, the cumulative average score was categorized into sex-specific quartiles and used as the primary exposure variable.

Using Cox proportional hazards regression models, we computed hazard ratios (HRs) to examine the association of the prudent or Western dietary score with incidence of colorectal cancer. To test for trend with the Wald test, participants were assigned to the median score of their sex-specific dietary pattern quartile, and then this variable was entered into the models as a continuous term. The covariates included in the multivariable models are described in the Table and the eMethods in the Supplement. All analyses were adjusted for total caloric intake (kilocalories per day) and stratified by age (in months), year of questionnaire return, and sex (in the analysis using combined cohorts). In multivariable analysis, we adjusted for potential confounders, including body mass index (calculated as weight in kilograms divided by height in meters squared), pack-years of smoking (never, 0-4 pack-years, 5-19 pack-years, 20-39 pack-years, or >40 pack-years), family history of colorectal cancer in any first-degree relative (yes or no), previous lower gastrointestinal endoscopy (yes or no), postmenopausal hormone use (for women only: never, past, or current), physical activity (quintiles of metabolicequivalent task hours per week), and regular use of aspirin or nonsteroidal anti-inflammatory agents (≥ 2 tablets per week: yes or no).

To examine whether the association between dietary patterns and incidence of colorectal cancer subgroups differed according to tissue F nucleatum status, we used Cox proportional hazards regression models with a duplication method for competing risks data. As our primary hypothesis testing, we tested for heterogeneity by using a likelihood ratio test, comparing a model that allows for separate associations of dietary patterns and risk of cancer subgroups according to F nucleatum status with a model that assumes a common association.³² In secondary analyses, we examined heterogeneity of the associations with cancer subgroups in relation to dominant factor loadings for the prudent dietary pattern using cumulative average intakes of fruits, vegetables, legumes, and whole grains as well as energy-adjusted intakes of fat, fiber, and protein, all of which were categorized into quartiles. We used SAS software, version 9.3 (SAS Institute Inc) for all statistical analyses. Data analysis was performed from March 15, 2015, to August 10, 2016.

Results

Of the 137 217 individuals included in the analysis, 47 449 were male (34.6%); mean (SD) baseline age for men was 54.0 (9.8) years and for women, 46.3 (7.2) years. Two major, uncorrelated dietary patterns were identified by factor analysis. The prudent dietary pattern was characterized by high intake of vegetables, fruits, whole grains, and legumes, and the Western dietary pattern was characterized by red and processed meats, refined grains, and desserts (eTable 1 in the Supplement). Consistent with prior analyses, ¹⁶ participants with high prudent scores in the HPFS and NHS tended to smoke less, exercise more, and have greater rates of lower gastrointestinal endoscopy, whereas Western pattern scores were associated with behaviors typically considered unhealthy (eTable 2 in the Supplement).

After 26 years (in HPFS) and 32 years (in NHS) of follow-up encompassing 3643562 person-years, we documented 1019 incident colorectal cancers with available data on tissue F nucleatum status. Among these cancer cases, there were 125 (12.3%) F nucleatum-positive tumors and 894 (87.7%) F nucleatum-negative tumors. We examined the association of prudent and Western dietary pattern scores with the incidence of overall colorectal cancer. Western dietary pattern scores showed a trend toward associations with overall risk of colorectal cancer in the HPFS (eTable 3 in the Supplement) and the combined cohort (Table); however, statistical significance was not reached with the adjusted a level of .01. We did not observe significant heterogeneity in the associations of the dietary scores with colorectal cancer risk between the 2 cohorts ($P \ge .21$). To maximize statistical power, we used the combined cohort for further analyses.

We then tested our primary hypothesis that the association of prudent and Western diets with colorectal cancer incidence might differ according to the presence of *F* nucleatum in tumor tissue. Notably, the association between prudent dietary pattern and risk of colorectal cancer significantly differed by tumor *F* nucleatum status (P = .01 for heterogeneity)

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					P Value	
Characteristic	Quartile 1	Quartile 2	Quartile 3	Quartile 4	Trend ^b	Heterogeneity ^c
Prudent Dietary Pattern						
Overall colorectal cancer						
Person-years	913 569	907 676	912 395	909 922	NA	NA
No. of cases (n = 1019), No. (%)	250 (24.5)	248 (24.3)	268 (26.3)	253 (24.8)		NA
Age-adjusted HR (95% CI) ^d	1 [Reference]	0.93 (0.77-1.11)	0.90 (0.75-1.08)	0.79 (0.65-0.95)	.01	NA
Multivariable HR (95% CI) ^e	1 [Reference]	0.95 (0.80-1.14)	0.95 (0.79-1.14)	0.85 (0.69-1.03)	.08	NA
F nucleatum-positive colorectal cancer						
No. of cases (n = 125), No. (%)	43 (34.4)	26 (20.8)	34 (27.2)	22 (17.6)	NA	NA
Age-adjusted HR (95% CI) ^d	1 [Reference]	0.54 (0.33-0.89)	0.67 (0.42-1.05)	0.40 (0.24-0.67)	<.001	NA
Multivariable HR (95% CI) ^e	1 [Reference]	0.56 (0.34-0.92)	0.70 (0.44-1.10)	0.43 (0.25-0.72)	.003	NA
F nucleatum-negative colorectal cancer					NA	.01
No. of cases (n = 894), No. (%)	207 (23.2)	222 (24.8)	234 (26.2)	231 (25.8)	NA	NA
Age-adjusted HR (95% CI) ^d	1 [Reference]	1.01 (0.83-1.22)	0.96 (0.79-1.16)	0.88 (0.72-1.08)	.15	NA
Multivariable HR (95% CI) ^e	1 [Reference]	1.04 (0.86-1.26)	1.00 (0.83-1.22)	0.95 (0.77-1.17)	.47	NA
Western Dietary Pattern						
Overall colorectal cancer						
Person-years	910 656	910 525	910 465	911 916	NA	NA
No. of cases (n = 1019), No. (%)	244 (23.9)	275 (27.0)	243 (23.8)	257 (25.2)	NA	NA
Age-adjusted HR (95% CI) ^d	1 [Reference]	1.24 (1.04-1.48)	1.21 (1.00-1.46)	1.46 (1.18-1.82)	.001	NA
Multivariable HR (95% CI) ^e	1 [Reference]	1.19 (1.00-1.43)	1.12 (0.92-1.36)	1.29 (1.03-1.62)	.05	NA
F nucleatum-positive colorectal cancer						NA
No. of cases (n = 125), No. (%)	25 (20.0)	33 (26.4)	33 (26.4)	34 (27.2)	NA	NA
Age-adjusted HR (95% CI) ^d	1 [Reference]	1.42 (0.84-2.40)	1.59 (0.94-2.69)	1.92 (1.12-3.29)	.01	NA
Multivariable HR (95% CI) ^e	1 [Reference]	1.37 (0.81-2.31)	1.49 (0.88-2.53)	1.69 (0.98-2.90)	.05	NA
F nucleatum-negative colorectal cancer					NA	.23
No. of cases (n = 894), No. (%)	219 (24.5)	242 (27.1)	210 (23.5)	223 (24.9)	NA	NA
Age-adjusted HR (95% CI) ^d	1 [Reference]	1.25 (1.03-1.50)	1.16 (0.95-1.42)	1.42 (1.13-1.78)	.006	NA
Multivariable HR (95% CI) ^e	1 [Reference]	1.20 (0.99-1.44)	1.08 (0.88-1.33)	1.25 (0.99-1.58)	.12	NA

Table. Hazard Ratios of Incident Colorectal Cancer, Overall and by Fusobacterium nucleatum Status^a

Abbreviations: HR, hazard ratio; NA, not applicable.

^a According to prudent or Western dietary score quartiles in the combined cohort of the Health Professionals Follow-up Study (1986-2012) and the Nurses' Health Study (1980-2012).

^b Tests for trend were conducted using the median value of each quartile category as a continuous variable.

^c We tested for heterogeneity by using a likelihood ratio test comparing a model that allows separate associations for the 2 colorectal cancer subgroups (ie, *F nucleatum*-positive and *F nucleatum*-negative subgroups) with a model that assumes a common association.

^d Stratified by age, calendar year, and sex and adjusted for total caloric intake (kilocalories per day).

^e Stratified as listed in the above footnote and additionally adjusted for family history of colorectal cancer in any first-degree relative, history of previous endoscopy, pack-years of smoking (never, 0-4, 5-19, 20-39, or ≥40), body mass index, physical activity (metabolic-equivalent task hours per week), and regular aspirin or nonsteroidal anti-inflammatory drug use (≥2 tablets/wk).

(Table). We found a significant inverse association of prudent dietary scores with *F nucleatum*-positive cancer risk (P = .003 for trend) but not with *F nucleatum*-negative cancer risk (P = .47 for trend). Comparing participants in the highest prudent dietary score quartile with those in the lowest quartile, the multivariable HR for *F nucleatum*-positive tumors was 0.43 (95% CI, 0.25-0.72); in contrast, the corresponding HR for *F nucleatum*-negative tumors was 0.95 (95% CI, 0.77-1.17). We found similar differential associations by *F nucleatum* status in men (HPFS) and women (NHS), although statistical power was limited (eTable 4 in the Supplement). In addition, although statistical power was limited, we found similar results when lev-

els of *F nucleatum* were categorized as low or high on the basis of the median cutoff point among *F nucleatum*-positive cases as performed in our previous analyses (eTable 5 in the Supplement).⁹ Because we observed that the fraction of colorectal cancers enriched with *F nucleatum* gradually decreased from cecum to rectum,³³ we conducted exploratory analyses stratified by tumor location (eTable 6 in the Supplement). The differential association of prudent diet score with colorectal cancer by tissue *F nucleatum* status appeared to be consistent in both proximal and distal cancer strata.

When we examined the association of the Western dietary pattern with colorectal cancer subgroups according to tumor *F* nucleatum status, although Western dietary pattern scores appeared to be more strongly associated with *F* nucleatum-positive cancer risk, there was no significant heterogeneity between the subgroups (P = .23 for heterogeneity) (Table).

In a secondary analysis, we sought to determine whether specific food groups might explain the observed differential associations between prudent dietary patterns and risk of colorectal cancer according to *F nucleatum* status. We examined the top 4 dominantly contributing food groups to the prudent diet pattern (vegetables, fruits, legumes, and whole grains) in relation to the risk of colorectal cancer according to *F nucleatum* status (eTable 7 in the Supplement). We observed no significant heterogeneity (with the adjusted a of .01).

Finally, to further determine whether any specific macronutrient components of the prudent dietary pattern might explain the observed differential associations according to F nucleatum status, we explored associations of fiber, fat, and protein intake with colorectal cancer subgroups (eTable 8 in the Supplement). There appeared to be heterogeneity in the differential association of fiber intake with cancer subgroups classified by F nucleatum status (P = .02 for heterogeneity), similar to the findings for prudent dietary pattern scores. Comparing participants in the highest quartile of fiber intake (>26 g/d for men and >19 g/d for women) with those in the lowest quartile (<18 g/d for men and <13 g/d for women), the multivariable HR for F nucleatum-positive tumors was 0.54 (95% CI, 0.32-0.92); in contrast, the corresponding HR for F nucleatum-negative tumors was 1.13 (95% CI, 0.92-1.40). In further exploratory analyses, we found that intakes of cereal-derived fiber might be differentially associated with colorectal cancer according to F nucleatum status (*P* = .01 for heterogeneity) (eTable 9 in the Supplement). We did not observe such heterogeneity for fat or protein.

Discussion

In the 2 US nationwide prospective cohorts, we found that participants with higher long-term prudent dietary pattern scores were associated with a lower risk of F nucleatumpositive colorectal cancers but not F nucleatum-negative cancers. Our data also suggest that higher intakes of dietary fiber, one of the components of the prudent diet, may be associated with a lower risk of F nucleatum-positive colorectal cancer but not F nucleatum-negative cancer. These findings support the hypothesis that the possible cancerpreventive effects of prudent diets rich in dietary fiber may be mediated by modulation of specific species in the gut microbiota and subsequent alteration of the amount of F nucleatum in local colonic tissue. To our knowledge, our study represents the first to examine the intersection of diet and incidence of colorectal cancer subgroups according to microbial status in human tumor tissue.

The potential role of diet in modulating the risk of a variety of diseases, including colorectal cancer, has been widely recognized.^{23,34} According to the World Cancer Research Fund and American Institute for Cancer Research, foods with fiber including whole grains are one of the strongest factors linked to decreasing the risk of colorectal cancer.³⁵ However, there has been considerable heterogeneity in the epidemiologic data associating prudent dietary patterns and the major components of the prudent diet with colorectal cancer.³⁶ Our results here suggest that the inconsistency in the association of prudent dietary patterns (and components of the diet) with lower colorectal cancer risk may be in part attributable to differential associations with cancer subgroups according to *F nucleatum* in tumor tissue. In addition, given recent findings between increasing amounts of *F nucleatum* DNA in colorectal cancer tissue and worsened survival,¹⁴ our data lend additional support to the promotion of healthy diets to reduce mortality from colorectal cancer.

The precise mechanism by which prudent diets rich in dietary fiber may lower F nucleatum-enriched cancer incidence remains unclear. Accumulating evidence suggests that longterm dietary fiber intake has a profound effect on the gut microbiome, specifically through promotion of microbial diversity and by lowering levels of inflammatory metabolites.^{25,37-40} A recent study showed that a 2-week feeding intervention switching rural-dwelling South Africans from a high-fiber, low-fat diet to a low-fiber, high-fat diet was associated with an increase in F nucleatum measured by PCR in the stool.²⁴ In addition, some have hypothesized that the variation observed in F nucleatum levels in colorectal cancers collected from Spain, Vietnam, Japan, and the United States may be attributable to differences in dietary practices in these countries.^{5,41} Furthermore, in a cross-sectional study, participants with advanced adenoma were associated with lower dietary fiber intakes as well as distinct fecal microbiome communities compared with healthy controls.⁴² It is plausible that an abundance of microbiota-accessible carbohydrates from prudent diets may influence bacterial fermentation of dietary fiber, resulting in altered levels of short-chain fatty acids. These changes may alter pH, increase transit time of gut contents, or lead to differences in local immune surveillance, which are less hospitable for nonnative species, such as *F* nucleatum, to establish themselves in the colonic niche and potentiate colorectal carcinogenesis.^{24,25,43,44} Taken together, these data provide evidence of substantial influences of diet on the gut microbiome, which may in turn influence tumorigenesis.

There are several strengths in this study. First, our dietary data were prospectively collected and have been well validated.²⁹ Second, our data were detailed and updated such that we could examine long-term effects of overall dietary patterns, specific food groups, and macronutrients in relation to colorectal cancer risk. Third, we collected detailed data on multiple potential confounders, although residual confounding cannot be excluded. Finally, our molecular pathological epidemiology (MPE) research⁴⁵ provides refined risk estimates for specific cancer subgroups, such as *F* nucleatum-positive cancer, and thereby offers insights into pathogenesis and causality. Molecular subtyping in the MPE approach can gather pathogenetically similar cases, and thus can enhance statistical inference (even with a relatively small number of cases).⁴⁶ The present study represents emerging unique microbial MPE research⁴⁷ in which the microbial feature in tumor tissue can serve as a pathogenic signature.

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Limitations

We acknowledge limitations of this study. First, this study was observational, and residual confounding may be an issue. Nevertheless, adjustment for a variety of known risk factors for colorectal cancer showed no substantial effect on the results. Second, the diet data were derived from food frequency questionnaires and subject to measurement errors. Nonetheless, studies have shown that food frequency questionnaires can better capture long-term dietary intakes than detailed diet diaries in a limited period.⁴⁸ Third, with the use of FFPE tissue specimens, routine histopathologic procedures might have influenced performance characteristics of our PCR assay to detect F nucleatum. Nonetheless, we conducted a rigorous validation study that showed high precision of our PCR assay to detect F nucleatum.9 Moreover, our assay has previously been shown to have high specificity for *F nucleatum*.⁶ Fourth, we could not collect FFPE blocks from all colorectal cancer cases in the cohorts; nonetheless, cases with available tissue were generally similar to those without tissue with regard to patient characteristics. Fifth, because our participants were all health professionals and most were white, generalizability of the findings to other populations needs to be examined in future studies.

Conclusions

This study has shown that a prudent diet is associated with a lower risk of *F* nucleatum-positive colorectal cancer but not *F* nucleatum-negative cancer. Our data generate new hypotheses about how the intestinal microbiota may mediate the association between diet and colorectal neoplasms. Further studies are needed to confirm these findings and determine the potential utility of characterization of *F* nucleatum in colonic mucosa, tumor, or stool as a biomarker for personalized nutritional, probiotic, or antibiotic interventions. In addition, our findings underscore the importance of future large-scale, prospective studies that examine the gut microbiota to understand the complex intersection of diet, the gut microbiome, and carcinogenesis.⁴⁹

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REFERENCES

1. Ahn J, Sinha R, Pei Z, et al. Human gut microbiome and risk for colorectal cancer. *J Natl Cancer Inst*. 2013;105(24):1907-1911.

2. Dejea CM, Wick EC, Hechenbleikner EM, et al. Microbiota organization is a distinct feature of proximal colorectal cancers. *Proc Natl Acad Sci U S A*. 2014;111(51):18321-18326.

3. Nakatsu G, Li X, Zhou H, et al. Gut mucosal microbiome across stages of colorectal carcinogenesis. *Nat Commun.* 2015;6:8727.

4. Flemer B, Lynch DB, Brown JMR, et al. Tumour-associated and non-tumour-associated microbiota in colorectal cancer [published online March 18, 2016]. *Gut.* doi:10.1136/gutjnl-2015-309595 5. Kostic AD, Gevers D, Pedamallu CS, et al. Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. *Genome Res.* 2012;22(2):292-298.

6. Castellarin M, Warren RL, Freeman JD, et al. *Fusobacterium nucleatum* infection is prevalent in human colorectal carcinoma. *Genome Res.* 2012;22 (2):299-306.

7. McCoy AN, Araújo-Pérez F, Azcárate-Peril A, Yeh JJ, Sandler RS, Keku TO. *Fusobacterium* is associated with colorectal adenomas. *PLoS One*. 2013;8(1):e53653.

8. Warren RL, Freeman DJ, Pleasance S, et al. Co-occurrence of anaerobic bacteria in colorectal carcinomas. *Microbiome*. 2013;1(1):16.

9. Mima K, Sukawa Y, Nishihara R, et al. *Fusobacterium nucleatum* and T cells in colorectal carcinoma. *JAMA Oncol.* 2015;1(5):653-661.

 Sinha R, Ahn J, Sampson JN, et al. Fecal microbiota, fecal metabolome, and colorectal cancer interrelations. *PLoS One*. 2016;11(3):e0152126.

11. Rubinstein MR, Wang X, Liu W, Hao Y, Cai G, Han YW. Fusobacterium nucleatum promotes colorectal carcinogenesis by modulating E-cadherin/ β -catenin signaling via its FadA adhesin. *Cell Host Microbe*. 2013;14(2):195-206.

12. Gur C, Ibrahim Y, Isaacson B, et al. Binding of the Fap2 protein of *Fusobacterium nucleatum* to human inhibitory receptor TIGIT protects tumors from immune cell attack. *Immunity*. 2015;42(2): 344-355.

13. Ito M, Kanno S, Nosho K, et al. Association of *Fusobacterium nucleatum* with clinical and molecular features in colorectal serrated pathway. *Int J Cancer*. 2015;137(6):1258-1268.

14. Mima K, Nishihara R, Qian ZR, et al. *Fusobacterium nucleatum* in colorectal carcinoma tissue and patient prognosis. *Gut*. 2016;65(12): 1973-1980.

15. Tahara T, Yamamoto E, Suzuki H, et al. *Fusobacterium* in colonic flora and molecular features of colorectal carcinoma. *Cancer Res.* 2014; 74(5):1311-1318.

16. Fung T, Hu FB, Fuchs C, et al. Major dietary patterns and the risk of colorectal cancer in women. *Arch Intern Med.* 2003;163(3):309-314.

17. Terry P, Hu FB, Hansen H, Wolk A. Prospective study of major dietary patterns and colorectal cancer risk in women. *Am J Epidemiol*. 2001;154 (12):1143-1149.

18. Kim MK, Sasaki S, Otani T, Tsugane S; Japan Public Health Center-based Prospective Study Group. Dietary patterns and subsequent colorectal cancer risk by subsite: a prospective cohort study. *Int J Cancer*. 2005;115(5):790-798.

19. Cottet V, Bonithon-Kopp C, Kronborg O, et al; European Cancer Prevention Organisation Study Group. Dietary patterns and the risk of colorectal adenoma recurrence in a European intervention trial. *Eur J Cancer Prev.* 2005;14(1):21-29.

20. Flood A, Rastogi T, Wirfält E, et al. Dietary patterns as identified by factor analysis and

colorectal cancer among middle-aged Americans. *Am J Clin Nutr.* 2008;88(1):176-184.

21. Mizoue T, Yamaji T, Tabata S, et al. Dietary patterns and colorectal adenomas in Japanese men: the Self-Defense Forces Health Study. *Am J Epidemiol.* 2005;161(4):338-345.

22. Magalhães B, Peleteiro B, Lunet N. Dietary patterns and colorectal cancer: systematic review and meta-analysis. *Eur J Cancer Prev*. 2012;21(1):15-23.

23. Song M, Garrett WS, Chan AT. Nutrients, foods, and colorectal cancer prevention. *Gastroenterology*. 2015;148(6):1244-60.e16.

24. O'Keefe SJD, Li JV, Lahti L, et al. Fat, fibre and cancer risk in African Americans and rural Africans. *Nat Commun.* 2015;6:6342.

25. Sonnenburg ED, Smits SA, Tikhonov M, Higginbottom SK, Wingreen NS, Sonnenburg JL. Diet-induced extinctions in the gut microbiota compound over generations. *Nature*. 2016;529 (7585):212-215.

26. Cotillard A, Kennedy SP, Kong LC, et al; ANR MicroObes Consortium. Dietary intervention impact on gut microbial gene richness. *Nature*. 2013;500(7464):585-588.

27. Le Chatelier E, Nielsen T, Qin J, et al; MetaHIT Consortium. Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013; 500(7464):541-546.

28. Koeth RA, Wang Z, Levison BS, et al. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med*. 2013; 19(5):576-585.

29. Rimm EB, Giovannucci EL, Stampfer MJ, Colditz GA, Litin LB, Willett WC. Reproducibility and validity of an expanded self-administered semiquantitative food frequency questionnaire among male health professionals. *Am J Epidemiol*. 1992;135(10):1114-1126.

30. Ananthakrishnan AN, Khalili H, Konijeti GG, et al. A prospective study of long-term intake of dietary fiber and risk of Crohn's disease and ulcerative colitis. *Gastroenterology*. 2013;145(5): 970-977.

31. Hu FB, Rimm EB, Stampfer MJ, Ascherio A, Spiegelman D, Willett WC. Prospective study of major dietary patterns and risk of coronary heart disease in men. *Am J Clin Nutr.* 2000;72(4):912-921.

32. Wang M, Spiegelman D, Kuchiba A, et al. Statistical methods for studying disease subtype heterogeneity. *Stat Med*. 2016;35(5):782-800.

33. Mima K, Cao Y, Chan AT, et al. *Fusobacterium nucleatum* in colorectal carcinoma tissue according to tumor location. *Clin Transl Gastroenterol*. 2016;7 (11):e200.

34. Tuddenham S, Sears CL. The intestinal microbiome and health. *Curr Opin Infect Dis*. 2015; 28(5):464-470.

35. World Cancer Research Fund/American Institute for Cancer Research. Continuous update project: keeping the science current. Colorectal Cancer 2011 Report: food, nutrition, physical activity, and the prevention of colorectal cancer. http://wcrf.org/int/research-we-fund/our-cancer -prevention-recommendations. Accessed April 21, 2016.

36. Aune D, Chan DSM, Lau R, et al. Dietary fibre, whole grains, and risk of colorectal cancer: systematic review and dose-response meta-analysis of prospective studies. *BMJ*. 2011; 343:d6617.

37. Wu GD, Chen J, Hoffmann C, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science*. 2011;334(6052):105-108.

38. Filippis FD, Pellegrini N, Vannini L, et al. High-level adherence to a Mediterranean diet beneficially impacts the gut microbiota and associated metabolome. *Gut*. 2016;65(11):1812-1821.

39. Ou J, Carbonero F, Zoetendal EG, et al. Diet, microbiota, and microbial metabolites in colon cancer risk in rural Africans and African Americans. *Am J Clin Nutr.* 2013;98(1):111-120.

40. Claesson MJ, Jeffery IB, Conde S, et al. Gut microbiota composition correlates with diet and health in the elderly. *Nature*. 2012;488(7410):178-184.

41. Nosho K, Sukawa Y, Adachi Y, et al. Association of *Fusobacterium nucleatum* with immunity and molecular alterations in colorectal cancer. *World J Gastroenterol.* 2016;22(2):557-566.

42. Chen H-M, Yu Y-N, Wang J-L, et al. Decreased dietary fiber intake and structural alteration of gut microbiota in patients with advanced colorectal adenoma. *Am J Clin Nutr.* 2013;97(5):1044-1052.

43. Garrett WS. Cancer and the microbiota. *Science*. 2015;348(6230):80-86.

44. Smith PM, Howitt MR, Panikov N, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science*. 2013;341(6145):569-573.

45. Ogino S, Chan AT, Fuchs CS, Giovannucci E. Molecular pathological epidemiology of colorectal neoplasia: an emerging transdisciplinary and interdisciplinary field. *Gut*. 2011;60(3):397-411.

46. Ogino S, Nishihara R, VanderWeele TJ, et al. The role of molecular pathological epidemiology in the study of neoplastic and non-neoplastic diseases in the era of precision medicine. *Epidemiology*. 2016;27(4):602-611.

47. Hamada T, Keum N, Nishihara R, Ogino S. Molecular pathological epidemiology: new developing frontiers of big data science to study etiologies and pathogenesis [published online October 13, 2016]. *J Gastroenterol*.

48. Willett W. *Nutritional Epidemiology*. New York, NY: Oxford University Press; 2012.

49. Fu BC, Randolph TW, Lim U, et al. Characterization of the gut microbiome in epidemiologic studies: the multiethnic cohort experience. *Ann Epidemiol.* 2016;26(5):373-379.