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# Association of FOXO3A Locus with Extreme Longevity in the Southern Italian Centenarian Study

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Title: Association of FOXO3A Locus with Extreme Longevity in the Southern Italian Centenarian Study

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## **ABSTRACT**

A number of potential candidate genes in a variety of biological pathways have been associated with longevity in model organisms. Many of these genes have human homologues and thus have potential to provide insights into human longevity. Recently, several studies suggested that FOXO3A functions as a key bridge of various signaling pathways that influence aging and longevity. Interestingly, Willcox BJ and colleagues identified several variants which displayed significant genotype-gender interaction in male human longevity. In particular, a nested case-control study was performed in an ethnic Japanese population in Hawaii and five candidate longevity genes were chosen based on links to the insulin-IGF1-signaling pathway. In the Willcox study, the investigated genetic variations (rs2802292, rs2764264, and rs13217795) within the FOXO3A gene were significantly associated with longevity in male centenarians.

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We validated the association of FOXO3A polymorphisms with extreme longevity in males <u>from the Southern</u> Italian Centenarian Study. Particularly, rs2802288, a proxy of rs2802292, showed the best allelic association (MAF=0.49, p=0.003, OR=1.51, 95% CI: 1.15-1.98). Furthermore we undertook a meta-analysis in order to explore the significance of rs2802292 association with longevity by combining the association results of the current study and the findings coming from <u>the Willcox BJ et al.</u> investigation.

Our data point to a key role of FOXO3A in human longevity and confirm the feasibility of the identification of such genes with centenarian-controls studies. Moreover, we hypothesize the susceptibility to <a href="the-longevity">the-longevity</a> phenotype may well be the result of complex interactions involving genes, environmental factors but also gender.

# INTRODUCTION

processes and toxin degradation <sup>7,8</sup>.

Aging is a multifactorial and complex process regulated by stochastic interactions (random damage to vital molecules), extrinsic interventions (such as diet and caloric restriction) and intrinsic/genetic alterations. A number of centenarian (people aged 100 years old or older) studies exploit the strong selection of favourable genotypes in exceptionally aged individuals to study candidate genes and to perform genome wide analyses. Replications of observed associations of genotypes with longevity, coupled with functional studies to define mechanisms whereby specific genotypes influence lifespan associated phenotypes, are essential to delineate true positive genetic findings. Such findings could potentially lead to preventive and therapeutic interventions for several age-related diseases that cause significant morbidity and mortality among older people. Numerous genes have been identified that are either positively or negatively selected in the centenarian population as a consequence of a demographic selection 1, but no consistent replications have been observed in independent population, with the exception of Apolipoprotein, E (APOE) 2.3. In addition, animal studies have provided insights into the types of genes that can be involved in the regulation of lifespan. The first longevity mutant to be identified was the C. Elegans gene PAX2 age-1 4 that encodes phosphatidylinositol 3-kinase (PI3K) 5, which has a key role in a signalling pathway that is homologous to the mammalian insulin- IGF1 (insulin-like growth factor 1) pathway. Genetic studies in C. Elegans have demonstrated that activation of the PIK3/Akt pathway by IGF1 suppresses activity of DAF-16 forkhead transcription factor, the nematode ortholog of mammalian FOXO proteins. FOXO transcription factors belong to the Forkhead family of proteins, a family characterized by a conserved DNA binding domain (Forkhead box) for phosphorylation by the survival kinase

A moderate reduction in the intake of <u>calories</u> [also known as caloric restriction (CR)] is extremely effective in delaying <u>age-related decline</u> and increasing longevity in organisms ranging from yeast to mammals <sup>9,10</sup>. <u>In this</u>

Akt 6. Particularly, in mammals the FOXO subgroup (FOXO1, FOXO3, FOXO4 e FOXO6) promote the

expression of numerous downstream genes that mediate stress resistance, innate immunity, metabolic

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regard, Bonkowski and colleagues observed a striking difference in the response of growth hormone (GH) receptor/GH-binding protein knockout (GHRKO) and normal mice to an identical regimen of CR <sup>11</sup>. When male and female longevity data were analyzed separately, it became evident that the effects of CR on maximal longevity were sexually dimorphic in GHRKO mice but not in normal mice.

To be noted, susceptibility to many common diseases, <u>well</u> as longevity, may well be the result of complex interactions involving genes, environmental factors and, intriguingly, gender.

In <u>this</u> scenario, *FOXO* is a key component of <u>the insulin pathway</u>, free radical <u>production</u> and human longevity.

In the prospective population based Leiden 85 plus Study (866 females and 379 males, aged 85 years or more), the effect of genetic variants in FOXO1A and FOXO3A were analyzed on metabolic profiles, fertility, fecundity, age-related diseases (i.e. diabetes, myocardial infarction, etc) and mortality 12. In detail, FOXO3A haplotypes showed no association with longevity and other investigated variables. One possible drawback of this finding is total absence of gender stratification. In fact, the hormonal environment as well as tissue specific gene expression is known to differ significantly between genders in vertebrates. Hormones may differentially affect gene expression in somatic tissues, thus leading to the gender specific susceptibility to a complex phenotype, as longevity 11. At this regard, Willcox BJ et al. 13 performed a nested case-control study of five candidate longevity genes (ADIPOQ, FOXO1A, FOXO3A, SIRT1 and COQ7) with links to the insulin/IGF signaling pathway (IIS). This nested case-control study was conducted in an ethnic Japanese population from the Island of Oahu (Hawaii) with 213 male longevity "cases" and 402 male "average-lived" controls, and found three SNPs associated with the "longevity" phenotype in the locus of FOXO3A gene: rs2764264 (p=0.0002), rs13217795 (p=0.0006) and rs2802292 (p<0.0001) 13. The aim of our study was to support the potential role of FOXO3A as a key component that influence longevity with a case-control study. In this regard, we screened 480 long-living "cases" (281 males and 199 females) and 335 young controls (195 males, 140 females) from an isolated and homogeneous population of Southern Jtaly (Southern Italian Centenarian Study-SICS) with Illumina BeadChip 300K and we focused on FOXO3A gene. As previously

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separately. We performed a powerful method for correcting for stratification of our population. Lastly, we exploited a meta-analysis approach to pool Willcox's findings in order to find evidence for statistical association for SNP rs2802292.

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## **RESULTS**

We recruited and phenotyped 281 male longevity "cases" (mean age 95 years) and 195 male "controls" (mean age 33 years) as part of <a href="mailto:the\_SICS">the\_SICS</a>. Subsequently, we genotyped these subjects (Table 1) using the Illumina 300k SNPs mapping BeadChip.

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After a preliminary phase of quality control (see methods section), we looked for evidence of population heterogeneity on a set of 258 males centenarians (cases) and 178 males controls by applying the Principal Component Analysis (PCA) as implemented in EigenSoft 2.0 package  $^{14}$ . The estimation of the over-dispersion of trend test statistics ( $\lambda$ ) was 1.05, decreasing to 1.027 after correction for the first 4 significant principal components: this indicates a small or null confounding effect of population structure on our association results. (Fig.1)

In order to test for associations between SNPs and the longevity phenotype, we compared allele and genotype frequencies between cases and controls by means of allelic and genotypic association tests (Table 2, Table S1). Results from all analyses were very similar for significantly associated SNPs whether or not adjustments for population structure were applied (Table S2). None of the analyzed SNPs showed deviations from the Hardy Weinberg Equilibrium (p-HWD<0.05) or significant differences in missing data fractions between cases and controls (p<0.05). All SNPs showed a genotyping call rate > 98.8%.

We also performed logistic regression analyses assuming <u>an additive model</u> and by comparing homozygous minor *versus* homozygous major alleles on a set of SNPs located on the *FOXO3A* locus (Table 2). In order to confirm the null-influence of population structure on our results we also performed association tests correcting along the top 4 principal components by means of logistic regression assuming an additive effect of allele dosage on the same SNP set.

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We first focused on rs2802288, showing Linkage Disequilibrium (LD) with rs2802292 (r²=1) in the HapMap CEU data panel <sup>15</sup>. Particularly, allelic association tests highlighted significant association for rs2802288 in males (MAF=0.49, p=0.003, OR=1.51, 95% CI: 1.15-1.98). Moreover, this association was also confirmed by logistic regression under <u>an\_additive model</u> (p=0.0026, OR=1.53, 95% CI: 1.16-2.03) and by comparing homozygous minor *versus* homozygous major alleles (p=0.0023, OR=2.44, 95% CI: 1.37-4.34). (Table 2)

Successively, we imputed missing SNPs by means of Impute <sup>16</sup> software to better represent the 6q21 region.

Rather, interestingly, the imputed rs2802292 (MAF=0.47) that was previously indicated by Willcox BJ *et al.* <sup>13</sup> as the best association, showed r2 =0.79 with rs2802288 (MAF=0.49), similarly to that observed in the HapMap CEU panel <sup>15</sup>. In fact, our imputation analysis confirmed that rs2802292, unrepresented on *the BeadChip*, was responsible for the strongest association effect in this region (MAF=0.468, p=0.0028; OR: 1.53; CI 95%: 1.16-2.03, allelic association test). Additionally, we confirmed this significant association by logistic regression under additive model (p=0.0022; OR: 1.57; CI 95%: 1.17-2.10) and by comparing homozygous minor *versus* 

To evaluate a possible joint effect of FOXO3A gene and the environmental exposure (such as smoke, etc) or FOXO3A and the non –genetic attributes (such as clinical-cognitive variables), we performed a covariates analysis. No significant association of genotypes with cognitive variables (Barthel score, cognitive score, anxiety, depression) or clinical phenotypes (diabetes mellitus, glaucoma, high blood pressure, kidney diseases, osteoporosis, smoking history, cancer) was detected. Then, we can hypothesize that FOXO3A have an effect on longevity independently of the tested variables. Subsequently, we generated a LD plot with the software HaploView v4.0 <sup>17</sup>. LD blocks are delineated by black lines and defined using the Four Gamete Rule, a variant on the algorithm described in previously published study <sup>18</sup>, as implemented in HaploView software <sup>19</sup>.

homozygous major alleles (p= 0.0019; OR: 2.58; CI 95%: 1.42-4.70). (Table 2)

haplotype (Fig 2, Block 1; Table 3) comprising 3 SNPs carrying the minor allele (A) for rs2802288,and defining boundaries of a locus affected by a potential causative mutation for the extreme longevity phenotype. As can be observed in Figure 2, the LD-plot defines a locus that includes the 5' untranslated regions and the first

Using this analysis, multimarker association tests showed a slightly significant association (p= 0.03) for a 24kb

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coding region of the investigated gene. Subsequently, in order to confirm the gender-specificity of the FOXO3A association, we also studied 199 female longevity "cases" (mean age 98 years) and 140 female "controls" (mean age 31 years) drawn from the same population (88% power to detect the association). No differences in terms of allele frequencies between centenarians and controls have been detected for rs2802288 in female individuals (176 centenarians and 113 controls that pass the quality control filtering criteria) (MAF=0.48, p= 0.3975, OR= 0.86, 95% CI: 0.62-1.21) (Table 2). The absence of association in the female gender has been further confirmed by logistic regression assuming an additive model (p=0.4034, OR=0.87, 95% CI: 0.62-1.21) and by comparing homozygous minor versus homozygous major alleles (p= 0.4048, OR=0.75, 95% CI: 0.39-1.47) (Table 2).

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Furthermore, we undertook a meta-analysis in order to explore the significance of <a href="mailto:the-rs2802292">the-rs2802292</a> association with longevity by combining the association results of the current study and the findings coming from <a href="mailto:the-willcox">the-willcox</a> BJ et al. investigation. We combined the logarithm of the OR estimates and 95% CI for homozygous minor vs homozygous major alleles from the current analysis (OR=2.44, 95% CI: 1.37-4.34) and from Willcox BJ et al. study (OR=2.75, 95% CI: 1.51-5.02) to obtain a summary OR using fixed <sup>20</sup> and Der Simonian and Laird random effects models assume that the effect of <a href="mailto:a-risk-allele">a-risk-allele</a> has the same value in each dataset while random effects models assume that the risk allele effects for each study vary around some overall average effect <sup>22</sup>. The inconsistency metric I<sup>2</sup> <sup>23</sup> was 0%, while the Q statistic <sup>24</sup> was not statistically significant (p=0.88) indicating a null between-study heterogeneity evidence. Since no evidence of between-study heterogeneity have been proven, fixed effects <sup>20</sup> and Der Simonian and Laird random effects <sup>21</sup> coincide, generating identical summary point estimates (OR= 2.66, 95% CI: 1.74-4.07, p=0.0001) (Fig 3).

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# DISCUSSION

Research into ageing has been rejuvenated by the recent discovery of SNPs in candidate genes that can extend lifespan of laboratory model organisms. Thus, these polymorphisms can also delay, ameliorate or even abolish the impact of many ageing-related diseases, including cardiovascular disease, neurodegeneration and

cancer. FOXO transcription factors have been implicated in regulating different cellular functions, <u>such</u> as differentiation, metabolism, proliferation, and survival <sup>7,25</sup>.

At this respect, in the Leiden Prospective Study the overall and individual haplotype frequencies were not different between the elderly and young control group for the FOXO3A locus 12. However, possible limitations of the study is in the absence of gender stratification. In fact, due to the profound effect of this pathway on the reproductive/hormonal system, that differs between males and females, it is possible that variants that affect genes of this pathway are gender specific enriched/depleted as the populations age. Hence, variants that influence longevity in males may not affect female longevity and vice-versa.

Recently, Willcox BJ and colleagues demonstrated an association with exceptional longevity and FOXO3A gene in an ethnic Japanese population in Hawaii, focusing on male individuals only <sup>13</sup> Although there was no population stratification, effect observed in this study a possible limitation in most. Stratification is the result of imbalance of ethnic background between cases and controls. It is usually modest when the recruitment is consistent among cases and controls, and for populations that didn't suffer of recent immigration. In the present study, we extrapolated the FOXO3A locus from our genome wide association data on a part of the SICS. Particularly, we genotyped (with BeadChip 300k, Illumina) 281 SIC male nonagenarians and 195 SIC male controls. It was encouraging to report the association between rs2802292 at FOXO3A locus and longevity phenotype was convincingly and independently confirmed in our SICS homozygous minor versus homozygous major alleles (p= 0.0019; OR: 2.58; CI 95%: 1.42-4.70). Moreover, to avoid a possible drawback of population structure we demonstrated a small or null confounding effect of population structure on our association results, thus excluding type I errors (Fig. 1, Table S2). Our aim was to replicate rs2802292 in male longevity, and to explore the FOXO3A locus in males and females to generate eventual new candidate SNPs for longevity and to clarify the gender interaction on the FOXO3A locus and longevity. Replication efforts require adequate sample size and our male population was sufficient to achieve >90% power to detect an allele with 40% allele frequency and a factor of 1.5 effect, as previously reported in Willcox paper for rs2802292 26.

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Meta-analysis represents a well-established method to summarize results and draw conclusions from different studies for a set of common hypothesis, therefore we applied this approach in order to combine our association results for the SNP rs2802292 with the findings coming from Willcox BJ *et al.* study and to achieve more statistical power. The null-significance of the Q-statistics <sup>24</sup> (p=0.88) and the inconsistency metric <sup>23</sup> value (I<sup>2</sup>=0%) confirmed the absence of between-study heterogeneity (Table 4). In absence of heterogeneity between the two datasets we obtained identical estimates of the ORs and the 95% CIs for homozygous minor vs homozygous major alleles from both fixed <sup>20</sup> and random effects <sup>21</sup> models (OR= 2.66, 95% CI: 1.74-4.07, p=0.0001) (Fig 3).

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To be noted, this is the first convincing replication of an association with longevity after the APOE association

As stressed by National Cancer Institute-National Human Genome Research Institute (NCI-NHGRI) Working Group on Replication in Association Studies <sup>27</sup>, validation of the postulated gene-investigated phenotype association in a population different from that of the previous study is of great value.

It is also established that mitochondrial Sirtuin, 3 interacts with and under specific cellular conditions regulates the activity of *FOXO3A*. Additionally, SIRT3 regulates a series of essential intracellular processes that defend the cell against multiple types of cellular damage, including oxidative damage <sup>28</sup>. A rise in the level of reactive oxygen species (ROS) has two important effects: it can damage proteins, lipids, and DNA, leading to cell death or it can trigger the activation of specific physiologic signalling pathway <sup>29</sup>. Moreover, a GH-IGF1 pathway has been implicated in determination of longevity in a variety of species <sup>10,30</sup>.

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Genetic variants that are either positively or negatively selected as population ages impact on survival demographic selection). In this regard, FOXO3A gene SNP rs2802292 should have a protective role.

These observations imply that the FOXO3A gene SNP rs2802292, or SNPs in LD, have a protective role by partially increasing the ability of other proteins to activate FOXO3A, or by increasing FOXO3A activity on downstream targets. Re-sequencing of the FOXO3A locus will clarify the nature of the genetic variation that is tracked by rs2802292. It is possible that the variation does not need to affect coding genes, since coding genes from less than one-third of the evolutionary conserved genome <sup>26</sup>. Furthermore, future population

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studies on diseases of aging will clarify if *FOXO3A* associate across diseases, as for the longevity gene *APOE* and Alzheimer 's / cardiovascular diseases.

To be noted, from the data that we obtained in our population, the association is strictly linked with malegender. Taking in consideration these new evidences, we would suggest a gender specific analysis for the longevity phenotype. Poleted: C

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To conclude, the discovery and replication of a convincing association of *FOXO3A* locus contributes to the hypothesis that the impact of the IGF-I/insulin pathway on longevity is a property that has been evolutionarily conserved throughout the animal kingdom.

# **MATERIALS and METHODS**

#### Subject features

This case/control study was carried on as a part of the Southern Italian Centenarian Study (SICS). The SIC study (2002) has assembled a large cohort of 600 DNA samples of nonagenarians/centenarians (age range 90-109 years) and 800 DNA samples of young controls (age range 18-48 years) from a isolated region of Southern Italy east of Naples with a high prevalence of long lived and healthy and characterized by a high level of endogamy <sup>31</sup>. All participants for the current study was well-characterized, in fact detailed phenotypic information about self reported health history, cognitive and physical functions measured through Blessed memory tests and Barthel scores of activity of daily living, as well as demographic and exposure to common risk factors such as alcohol and smoking were collected. For the aim of the present investigation, 281 long-lived men (age range 90-108 years) and 195 controls (age range 18-48 years) were genotyped (Table 1).

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The subjects were enrolled by an Italian subsidiary (Associazione Longevità) of Elixir Pharmaceuticals (former Centagenetix Inc). All subjects donated blood samples for DNA study and gave written informed consent to the study. The study was conducted in accordance with the ethical principles that have their origins in the Declaration of Helsinki.

Genotyping: Illumina BeadChip

Genotyping was carried out using the Infinium II Assay-HumanHap BeadChip 317K-duo (Illumina) using standard protocol. In fact, Illumina's Infinium II Whole-Genome Genotyping Assay is designed to interrogate a large number of single nucleotide polymorphisms (SNPs) at unlimited levels of loci multiplexing. HumanHap 317-Duo workflow can be divided in three main segments: (1) sample preparation, (2) sample fragmentation and hybridization, (3) extension, staining, and scanning.

The BeadChps are imaged using a two-color confocal laser system with 0.8-µm resolution. The bead intensities are extracted, and genotypes are calculated using an Illumina-supplied cluster file, which is based on a set of reference samples. The normalization algorithm adjusts for nominal offset, cross-talk, and intensity variations observed in the two-color channels. The data for each BeadChip is self-normalized using information contained within the array. This normalization algorithm removes outliers, adjusts for channel-dependent background and global intensity differences, and also scales the data. The X and Y color channels undergo an affine coordinate transformation to make the data appear as canonical as possible with the homozygotes lying along the transformed *x*- and *y*-axes. All genotypes will be evaluated using a quantitative quality score called GenCall score. A GenCall score ranges from 0 to 1 and reflects the proximity within a cluster plot of the intensities of that genotype to the centroid of the nearest cluster.

#### Statistical analysis

**Quality control:** Individuals showing genotyping rate <93% and SNPs with genotyping rate <95% have been excluded from the dataset. We screened for contaminations and evaluated relationships among individuals using Identity By Descent (IBD) estimations: samples showing extreme heterozygosis and related individuals have been removed. X chromosome data have been used to check for the discordances in terms of gender assignment. SNPs with MAF < 0.01 and markers deviating from the Hardy Weinberg Equilibrium (HWE) in the control population (p-HWD < 0.001) have been excluded from the analysis.

**Population structure**: In order to assess the absence of population stratification we applied the Principal Component Analysis (PCA) approach, a method that can capture subtle and extensive variations due to both ethnical, experimental and technical features <sup>14</sup>. All autosomal SNPs and individuals that passed the quality control filters (299772 SNPs, 258 centenarians, 178 controls) have been used as input to the EigenSoft 2.0 <sup>14</sup>

software using the default parameters except for the number of outliers removal iterations that was set to 0 to obtain estimate for all subjects.

**Imputation:** To impute SNPs from multimarker tags we used Impute v0.5.0 <sup>16</sup>.

**Association tests:** We evaluated the association of markers with longevity phenotype by means of allelic association tests (1d.f.) and applied Pearson  $\chi^2$  test (2 d.f.) and Fisher exact test (if cell counts were < 5) in order to compare genotype frequencies between cases and controls. We also performed logistic regression under an additive model and logistic regression comparing homozygous minor vs homozygous major alleles. Correction for population stratification have been performed using logistic regression under additive model adjusting along the top 4 statistical significant principal components. Allele frequencies, genotype counts, OR and 95% confidence intervals (95% CI) have been also estimated for each SNP.

**Meta-analysis**: We used the OR and 95% CI for homozygous minor vs homozygous major alleles from the current analysis and from Willcox BJ *et al.* study to calculate the natural logarithms of the OR (Log-OR) and its standard error (SE). The Log-OR estimates were combined to obtain a summary OR following fixed  $^{20}$  and Der Simonian and Laird random effects  $^{21}$  models using inverse variance calculations. Between dataset heterogeneity was identified using the  $^{12}$  metric for inconsistency  $^{23}$  and its statistical significance was assessed by means of the  $\chi^2$  distributed Q statistic  $^{24}$  (the number of d.f. is given by k-1, where k is the number of analyzed datasets). All calculations have been performed using R v2.7.1 software.

**Haplotype analyses:** We used HaploView 4.0  $^{17}$  for blocks definition, haplotypes estimation and association tests. We defined haplotype blocks using the Four Gamete Rule, a variant on the algorithm described in Wang study  $^{18}$  and estimated haplotypes using the default accelerated EM algorithm  $^{17}$ . Differences in haplotype frequencies between centenarians and controls have been tested by means of  $\chi^2$  tests, excluding from the analysis haplotypes with a frequency < 0.01.

**Covariate analyses**: Associations between clinical risk factors, cognitive variables with genotypes have been tested among nonagenarians by means of logistic and linear regression using R v2.7.1 software (http://www.R-project.org).

All statistical analyses have been performed using PLINK v1.04 32 software except when specified.

#### **AKNOWLEDGMENTS:**

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## **TABLE TITLE:**

Table 1 : Age (years) distribution by gender and by centenarian/control phenotype

Table 2: FOXO3A locus in Jong-living individuals and controls.

Table 3: Haplotype association test of block 1 using HaploView.

Deleted: Table 4: Meta-analysis for rs2802292 combining Okinawa study and SICS ¶

Deleted: male

## FIGURE LEGENDS:

**Figure 1. Population structure of centenarians (red) and control subjects (black).** Each scatter plot shows the first two principal components that were estimated using genotype data for more than 300K SNPs in centenarians and control subjects using the program EigenSoft 2.0. The two populations are ethnically identical as shown in the right scatter plot.

Figure 2. Pattern of LD of FOXO3A region displayed using D' with HaploView. LD displays were generated using the D' colour scheme. The different shades of red indicate different D' value (0<D'<1). Haplotype blocks were generated using the Four Gamete Rule as implemented in HaploView software. Boxed in green is rs2802288, boxed in blue are the SNPs explored by Willcox BJ *et al.* 

Figure 3. Meta-analysis of rs2802292 variant in the current investigation and in Willcox study.

Each study is represented by the estimate of the homozygous minor vs homozygous major alleles OR and its 95% CI (circle). Summary effects by random effects (rhombus) and fixed effects (rectangle) calculations are also shown.

Table 1 : Age (years) distribution by gender and by centenarian/control phenotype

		Males			Females	1
Phenotype	n	Mean ±SD	Min-Max	n	Mean ±SD	Min-Max
Centenarians	281	$94.96 \pm 2.97$	90-108	199	$98.05 \pm 3.25$	90-109
Controls	195	$33.53 \pm 7.33$	18-48	140	$31.39 \pm 7.31$	18-48
Footnote: n= nu	mber of	individuals; SD= s	standard deviat	ion; Mir	n= minimum age;	

Table 2: FOXO3A locus in long-living individuals and controls

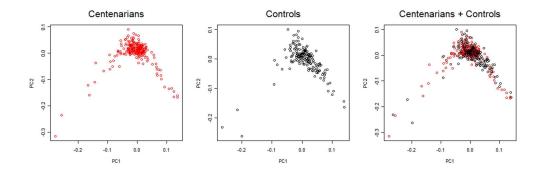
SNP												
<u></u>	<u>gender</u>		MAF		<u>Allelic</u>	<u>Allelic</u>		<u>Additive</u>		<u>Homozygous</u>		
		Overall	Cases	Controls	OR (95%CI)	р	OR (95%CI)	p	OR (95% CI)	p		
rs9486902	M	0.2	0.22	0.18	1.27(0.90-1.80)	0.1659	1.30 (0.91-1.85)	0.1524	0.92 (0.30-2.80)	0.8786		
137400702	F	0.23	0.21	0.27	0.69 (0.47-1.02)	0.0634	0.67 (0.45-1.01)	0.0553	0.52 (0.16-1.70)	0.2800		
rs2802288	M	0.49	0.53	0.43	1.51 (1.15-1.98)	0.003	1.53 (1.16-2.03)	0.0026	2.44 (1.38-4.34)	0.0023		
182002200	F	0.48	0.47	0.5	0.86 (0.62-1.21)	0.3975	0.87 (0.62-1.21)	0.4034	0.75 (0.39-1.47)	0.4048		
rs10499051	M	0.12	0.14	0.1	1.57 (1.02-2.41)	0.04	1.56 (1.01-2.41)	0.0409	NA	0.9982		
1810499031	F	0.12	0.11	0.12	0.95 (0.57-1.60)	0.8497	0.95 (0.56-1.61)	0.8474	0.32 (0.03-3.62)	0.3598		
rs2802292*†	M	0.47	0.51	0.41	1.53 (1.16-2.03)	0.0028	1.57 (1.17-2.10)	0.0022	2.58 (1.42-4.70)	0.0019		
132002272	F	NA	NA	NA	NA	NA	NA	NA	NA	NA		
rs13220810	M	0.2	0.18	0.22	0.79 (0.563-1.11)	0.1682	0.79 (0.56-1.10)	0.1653	0.80 (0.29-2.20)	0.6619		
1313220010	F	0.22	0.23	0.2	1.17 (0.78-1.77)	0.4482	1.17 (0.78-1.78)	0.4471	1.12 (0.35-3.57)	0.8441		
rs2764264 <b>†</b>	M	0.4	0.43	0.36	1.37 (1.04-1.81)	0.0267	1.40 (1.05-1.86)	0.0227	1.99 (1.07-3.70)	0.0285		
	F	0.41	0.4	0.42	0.93 (0.66-1.30)	0.6575	0.92 (0.65-1.31)	0.6553	1.03 (0.49-2.18)	0.9312		
rs7341233	M	0.17	0.17	0.15	1.16 (0.80-1.67)	0.4364	1.15 (0.80-1.64)	0.4512	2.14 (0.67-6.79)	0.1963		
	F	0.14	0.16	0.12	1.51 (0.92-2.48)	0.1047	1.51 (0.92-2.49)	0.1070	3.61 (0.41-31.41)	0.2454		
rs17598747	M	0.15	0.15	0.15	1.05 (0.71-1.53)	0.8172	1.04 (0.72-1.50)	0.8237	1.72 (0.53-5.60)	0.3683		
	F	0.12	0.13	0.1	1.44 (0.84-2.46)	0.1825	1.43 (0.84-2.43)	0.1911	2.79 (0.31-25.35)	0.3626		
rs9285397	M	0.22	0.24	0.2	1.29 (0.93-1.80)	0.1274	1.34 (0.94-1.90)	0.1054	1.76 (0.53-5.88)	0.3561		
137203377	F	0.26	0.24	0.31	0.71 (0.49-1.03)	0.0699	0.69 (0.47-1.02)	0.0637	0.71 (0.26-1.99)	0.5190		
rs12202209	M	0.15	0.15	0.15	1.07 (0.73-1.56)	0.739	1.06 (0.74-1.53)	0.7479	1.74 (0.53-5.60)	0.359		
rs12202209	F	0.12	0.13	0.1	1.44 (0.84-2.46)	0.1825	1.43 (0.84-2.43)	0.1911	2.79 (0.31-25.35)	0.3626		
rs9480866	M	0.21	0.22	0.19	1.24 (0.87 1.75)	0.206	1.27 (0.89-1.80)	0.1881	1.71 (0.52-5.71)	0.3781		
	F	0.24	0.21	0.29	0.67 (0.46-0.99)	0.0412	0.65 (0.44-0.97)	0.0370	0.51 (0.17-1.53)	0.2279		
rs12207868	M	0.15	0.15	0.15	1.06 (0.72 1.54)	0.7751	1.05 (0.73-1.52)	0.7829	1.72 (0.53-5.60)	0.3683		
	F	0.12	0.13	0.1	1.43 (0.84-2.44)	0.1905	1.42 (0.83-2.41)	0.1992	2.77 (0.30-25.16)	0.3662		
rs13217795 <b>*</b> †	M	0.35	0.38	0.3	1.42 (1.06 1.90)	0.0176	1.44 (1.07-1.945)	0.0159	2.30 (1.14-4.64)	0.0196		
1010211170	F	NA	NA	NA	NA	NA	NA	NA	NA	NA		
rs2153960	F	0.4	0.38	0.42	0.83 (0.59-1.18)	0.3009	0.83 (0.59-1.18)	0.2979	0.83 (0.39-1.73)	0.6107		
152133700	M	0.38	0.41	0.34	1.31 (0.99 1.74)	0.0578	1.33 (1-1.78)	0.0521	1.87 (0.99-3.53)	0.0542		

	M	0.36	0.38	0.33	1.25 (0.94 1.67)	0.1187	1.26 (0.94-1.68)	0.1162	1.65 (0.87-3.12)	0.1222
rs3800229										
	F	0.38	0.36	0.41	0.81 (0.57-1.14)	0.2285	0.81 (0.57-1.15)	0.2310	0.75 (0.36-1.58)	0.4478
	M	0.36	0.38	0.33	1.25 (0.94 1.67)	0.118	1.26 (0.95-1.68)	0.1143	1.65 (0.87-3.12)	0.1222
rs1935949										
	F	0.38	0.36	0.41	0.81 (0.57-1.14)	0.2200	0.81 (0.57-1.14)	0.2222	0.77 (0.37-1.62)	0.4911

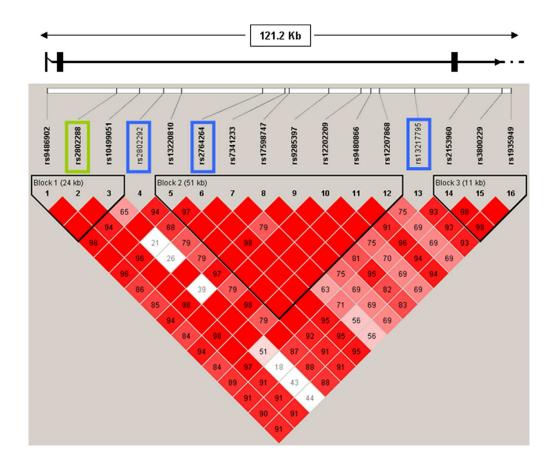
Footnotes: Gender = Gender-specific analysis (M = males, F = females), MAF Overall = minor allele frequency based on whole sample, MAF Cases = minor allele frequency in centenarians, MAF Controls = minor allele frequency in the control population; Odds Ratio (OR), 95% confidence intervals (95% C.I.) and p-value (p) respectively for allelic association test (Allelic), logistic regression under additive model (Additive) and logistic regression comparing homozygous minor (mm) vs homozygous major alleles (MM) (Homozygous); \* = imputed SNPs; † = SNPs explored by Willcox BJ *et al.* 

Table 3: Haplotype association test of block 1 using HaploView.

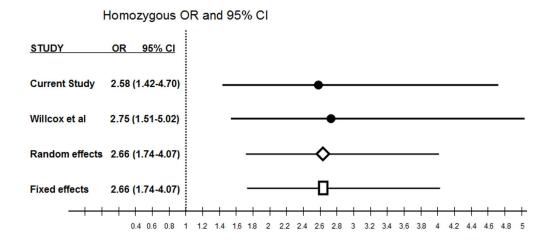
CGA         M         0.51         0.48         0.57         0.003           F         0.51         0.53         0.49         0.3683           TAA         M         0.2         0.21         0.17         0.1588           F         0.23         0.21         0.27         0.0634           CAA         F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           CAG         F         0.12         0.11         0.12         0.8689	CGA         F         0.51         0.53         0.49         0.3683           TAA         M         0.2         0.21         0.17         0.1588           F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           CAG         M         0.12         0.14         0.09         0.0381           CAG         F         0.12         0.11         0.12         0.8689	CGA         F         0.51         0.53         0.49         0.3683           TAA         M         0.2         0.21         0.17         0.1588           F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           CAA         F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           CAG         F         0.12         0.11         0.12         0.8689	Block 1	Gender	Frequency	Freq Cases	Freq Controls	p-value
F         0.51         0.53         0.49         0.3683           TAA         M         0.2         0.21         0.17         0.1588           F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           CAG         F         0.12         0.11         0.12         0.8689	F         0.51         0.53         0.49         0.3683           TAA         M         0.2         0.21         0.17         0.1588           F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           CAG         M         0.12         0.14         0.09         0.0381           CAG         F         0.12         0.11         0.12         0.8689	F         0.51         0.53         0.49         0.3683           TAA         M         0.2         0.21         0.17         0.1588           F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           CAG         M         0.12         0.14         0.09         0.0381           CAG         F         0.12         0.11         0.12         0.8689	CGA	M	0.51	0.48	0.57	0.003
TAA         F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	TAA         F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	TAA         F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	0.000	F	0.51	0.53	0.49	0.3683
F       0.23       0.21       0.27       0.0634         CAA       M       0.17       0.18       0.16       0.4968         F       0.13       0.15       0.11       0.2556         CAG       M       0.12       0.14       0.09       0.0381         F       0.12       0.11       0.12       0.8689	F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	F         0.23         0.21         0.27         0.0634           CAA         M         0.17         0.18         0.16         0.4968           F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	TAA	M	0.2	0.21	0.17	0.1588
CAA         F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	CAA         F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	CAA         F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689		F	0.23	0.21	0.27	0.0634
F         0.13         0.15         0.11         0.2556           M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	F         0.13         0.15         0.11         0.2556           CAG         M         0.12         0.14         0.09         0.0381           F         0.12         0.11         0.12         0.8689	CAA	М	0.17	0.18	0.16	0.4968
F         0.12         0.11         0.12         0.8689	CAG         F         0.12         0.11         0.12         0.8689	CAG         F         0.12         0.11         0.12         0.8689		F	0.13	0.15	0.11	0.2556
F 0.12 0.11 0.12 0.8689	F 0.12 0.11 0.12 0.8689	F 0.12 0.11 0.12 0.8689	CAG	М	0.12	0.14	0.09	0.0381
			<b>0.110</b> .	F	0.12	0.11	0.12	0.8689



180x60mm (600 x 600 DPI)



166x140mm (600 x 600 DPI)



179x87mm (600 x 600 DPI)