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## ATP-independent bioluminescent reporter variants to improve *in vivo* imaging

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### Abstract

Coelenterazine (CTZ)-utilizing marine luciferases and their derivatives have attracted significant attention because of their ATP-independency, fast enzymatic turnover, and high bioluminescence brightness. However, marine luciferases typically emit blue photons and their substrates, including CTZ and the recently developed diphenylterazine (DTZ), have poor water solubility, hindering their *in vivo* applications. Herein, we report a family of pyridyl CTZ and DTZ analogs that exhibit spectrally shifted emission and improved water solubility. Through directed evolution, we engineered a LumiLuc luciferase with broad substrate specificity. In the presence of corresponding pyridyl substrates (i.e., pyCTZ, 6pyDTZ, or 8pyDTZ), LumiLuc generates highly bright blue, teal, or yellow bioluminescence. We compared our LumiLuc-8pyDTZ pair with several benchmark reporters in a tumor xenograft mouse model. Our new pair, which does not need organic cosolvents for *in vivo* administration, surpasses other reporters by detecting early tumors. We further fused LumiLuc to a red fluorescent protein, resulting in a LumiScarlet reporter with further red-shifted emission and enhanced tissue penetration. LumiScarlet-8pyDTZ was comparable to Akaluc-AkaLumine, the brightest ATP-dependent luciferase-luciferin pair, for detecting cells in deep tissues of mice. In summary, we have engineered a new family of ATP-independent bioluminescent reporters, which will have broad applications because of their ATP-independency, excellent biocompatibility, and superior *in vivo* sensitivity.

### INTRODUCTION

In the past few decades, fluorescence imaging has evolved quickly and become a dominant visualization method for live-cell studies.<sup>1</sup> However, fluorescence imaging has several limitations, such as photobleaching, phototoxicity, and poor tissue penetration, largely due to the need for light excitation. Unlike fluorescence, bioluminescence produces photons *via* enzyme-catalyzed biochemical reactions in which luciferases oxidize their corresponding

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#### ASSOCIATED CONTENT

The Supporting Information is available free of charge via the Internet.

Experimental details, synthetic methods, compound characterization, and supporting figures, tables, and results (PDF).

#### NOTES

The University of Virginia filed a provisional patent application that is partially based on results described in the manuscript. H.-w.A. and H.-W.Y. are listed as coinventors.

small-molecule substrates (*a.k.a.*, luciferins) to generate excited-state emitters. As a result, bioluminescence signals glow essentially on dark background, leading to excellent signal-to-background ratios. Moreover, even though the spatiotemporal resolution of bioluminescence imaging (BLI) is usually worse than that of fluorescence imaging, the emitted photons can escape through several centimeters of tissue.<sup>2</sup> BLI is thus especially suited for diverse, noninvasive *in vivo* imaging applications.<sup>3-6</sup>

*Photinus pyralis* firefly luciferase (FLuc) and D-luciferin ( $\lambda_{\text{max}}$ : 563 nm) constitute the most widely used luciferase-luciferin pair for *in vivo* BLI. Recently, research has been performed to develop FLuc and D-luciferin derivatives for brighter and more red-shifted emission. In particular, an Akaluc-AkaLumine luciferase-luciferin pair with near-infrared (NIR) emission ( $\lambda_{\text{max}}$ : 650 nm) was reported for highly sensitive deep-tissue *in vivo* BLI.<sup>7</sup> Despite the progress, AkaLumine has been shown to induce cytotoxicity.<sup>8-10</sup> Moreover, FLuc, Akaluc, and other insect luciferases consume ATP for photon production; the bioluminescence reaction between FLuc and D-luciferin reduced the intracellular ATP-to-ADP ratio of live mammalian cells from > 40:1 to ~ 20:1,<sup>9</sup> suggesting metabolic disruption by all ATP-dependent luciferases. Because ATP is required for the activation of the luciferin substrates, this metabolic disruption issue cannot be addressed by simply improving insect luciferases and the corresponding substrates.

In contrast to insect luciferases, a large family of marine luciferases and photoproteins, such as *Renilla* luciferase (RLuc), *Gaussia* luciferase (GLuc), *Oplophorus* luciferase (OLuc), and aequorin, are ATP-independent and use coelenterazine (CTZ, Figure 1) as their native substrate for bioluminescence production.<sup>11</sup> The 19kDa catalytic domain of OLuc<sup>12</sup> was recently engineered into NanoLuc, which has a fast enzyme turnover and produces intense blue bioluminescence ( $\lambda_{\text{max}}$ : 456 nm) in the presence of a synthetic CTZ analog, furimazine (FRZ, Figure 1).<sup>13</sup> To expand the color palette, NanoLuc was further engineered into teLuc, which emits red-shifted photons ( $\lambda_{\text{max}}$ : 502 nm) when paired with a synthetic substrate diphenylterazine (DTZ, Figure 1).<sup>8</sup> Since biological tissues significantly absorb and scatter short-wavelength photons,<sup>14</sup> NanoLuc and teLuc have been fused to fluorescent proteins, resulting in Antares, Antares2, and enhanced Nano-Lanterns for further red-shifted emission *via* bioluminescence resonance energy transfer (BRET).<sup>8, 15, 16</sup> The water solubility of CTZ, FRZ, and DTZ is adequate for protein- and cell-based assays, because their solubility is already higher than typical substrate concentrations in these *in vitro* assays. However, *in vivo* applications of existing ATP-independent bioluminescent reporters are greatly hindered by the low solubility of CTZ, FRZ, and DTZ. Small animals, such as mice, can only tolerate a small injection volume. To enhance bioluminescence brightness by delivering more luciferin substrate, hydroxypropyl- $\beta$ -cyclodextrin, polyethylene glycols (PEGs), or other organic cosolvents are typically used to formulate CTZ, FRZ, or DTZ for *in vivo* administration.<sup>8, 15, 17, 18</sup> These formulation ingredients are not biologically inert and can cause irritation or biotoxicity. It is also practically difficult to intravenously (*i.v.*) inject these highly viscous solutions into small animals. Furthermore, it is still of great interest to further red-shift marine luciferases for enhanced tissue penetration.

In this work, we chemically modified CTZ and DTZ for spectrally shifted emission and enhanced water solubility. Concurrently, we engineered teLuc into a LumiLuc luciferase,

which is highly active toward the new substrates for intense blue, teal, and yellow emission. Moreover, by harnessing a recently reported high-quality red fluorescent protein, mScarlet-I, we developed a LumiScarlet reporter with significant emission longer than 600 nm. Our multipronged approach yielded a new family of ATP-independent bioluminescent reporters, which have improved biochemical and photophysical properties and are expected to have broad applications.

## RESULTS

### Design and synthesis of pyridyl CTZ and DTZ analogs with enhanced water solubility

Despite that recent studies have synthesized and tested a number of CTZ analogs with NanoLuc,<sup>19, 20</sup> the luciferase has not yet been optimized to pair with these new substrates and the water solubility issue of the substrates has not yet been tackled systematically. We sought to develop CTZ and DTZ analogs with improved water solubility by using the concept of bioisostere replacements in medicinal chemistry. Pyridine is considered a biocompatible *N*-heterocycle substituent for benzene with enhanced water solubility, because pyridine-containing molecules can be readily turned into pyridinium salts. Therefore, we designed a convergent synthetic route to prepare a series of CTZ and DTZ analogs with pyridyl isomer substitutions at the C-2, C-6 and C-8 positions of the imidazopyrazinone core (Scheme 1). Briefly, we first used Suzuki or Negishi cross-coupling reactions to regioselectively functionalize 2-amino-3,5-dibromopyrazine with either pyridyl, phenyl, or benzyl functional groups to give monosubstituted products (**1a-c**), which were subsequently derivatized via Suzuki cross-coupling reactions to afford disubstituted intermediates (**2a-f**, see structures and synthetic methods in the Supporting Information). In the second cross-coupling step, the XPhos-Pd-G2 catalyst was used to enhance reaction yields and minimize the protodeboronation of pyridyl boronic acids.<sup>21</sup> We further utilized an acid-catalyzed ring closing reaction<sup>22</sup> in dioxane to derive various pyridyl CTZ and DTZ analogs (**3a-f**, Table 1) from the disubstituted intermediates and corresponding  $\alpha$ -ketoacetals.

We next used turbidimetric solubility assays<sup>23</sup> to evaluate water solubility of these CTZ and DTZ analogs (Table 1 and Figure S1). Our newly synthesized pyridyl analogs enhanced the solubility by 4- to 14-fold from CTZ and DTZ. We also evaluated the autoluminescence and stability of these new analogs and they are comparable or even better than CTZ and FRZ (Figure S2). We further investigated their chemiluminescence, because the wavelength of bioluminescence is often related to the wavelength of substrate chemiluminescence, although the luciferase enzyme provides further electrostatic tuning which can reshape the emission (Figures S3). In addition, we evaluated the bioluminescence of these new substrates in the presence of several representative, ATP-independent luciferases such as RLuc8, NanoLuc, teLuc, and aequorin (Figures S4). Although each luciferase has different substrate preferences, the compound **3a** (pyCTZ) generated strong blue bioluminescence in the presence of each of these tested luciferases. When paired with aequorin, the bioluminescence intensity of pyCTZ was comparable to native CTZ, suggesting that pyCTZ may be directly used to replace CTZ for aequorin-based calcium sensing.<sup>24</sup> Furthermore, compared to DTZ, compounds **3c** (8pyDTZ), and **3f** were able to emit red-shifted

chemiluminescence and/or bioluminescence, while **3b** and **3d** (6pyDTZ) caused hypsochromic shift (Figure S3 and Table 1). Molecular mechanisms governing the spectral shift properties of these synthetic substrates remain to be investigated. Because **3c** showed the most red-shifted emission and red-shifted photons can penetrate through tissue better,<sup>25</sup> we selected **3c** (8pyDTZ) as our candidate substrate for further development of an optimized, red-shifted luciferase-luciferin pair.

### Directed evolution of the teLuc luciferase for improved brightness

teLuc was previously optimized for DTZ, a substrate with conjugated disubstitutions on the imidazopyrazinone core. 8pyDTZ exhibits ~ 30 nm red-shift but the emission of teLuc-8pyDTZ has been greatly attenuated compared to teLuc-DTZ. We next engineered teLuc for increased photon flux in the presence of 8pyDTZ. On the basis of a published apomanoKAZ structure<sup>26</sup> and our computational model,<sup>8</sup> we first introduced random mutations to residues 18 and 19 close to a putative substrate-binding pocket (Figure 2). After screening for improved mutants, we further randomized residues 27, 28, and 29 located deeper in the putative catalytic site. From the first two rounds of protein engineering, a teLuc-L18Q/S19A/V27L/S28T mutant was identified, to which we further introduced random mutations using error-prone PCR. After eight additional rounds of mutagenesis and screening, we derived a LumiLuc luciferase with 12 total mutations and ~ 5-fold enhancement of 8pyDTZ bioluminescence from teLuc (Figures 2 and S5).

The resultant LumiLuc-8pyDTZ pair has an emission peak at 525 nm. Its *in vitro* maximal photon emission rate ( $V_{\max}$ ) is ~ 60% and ~ 36% of NanoLuc-FRZ and teLuc-DTZ, respectively. The apparent Michaelis constant ( $K_M$ ) of LumiLuc-8pyDTZ was 4.6  $\mu\text{M}$ , lower than that of teLuc-DTZ or NanoLuc-FRZ (Figure S6a). This reduced  $K_M$  is practically beneficial, since LumiLuc-8pyDTZ would be relatively brighter when effective substrate concentrations are limited, such as in live cells (Figure S6b) and *in vivo*. Similar to NanoLuc and teLuc, the bioluminescence kinetics of LumiLuc is flash-type in phosphate buffer saline (PBS) and glow-type in a specially formulated assay buffer (Figure S6c).<sup>13</sup>

LumiLuc has broad substrate specificity. It improved the photon flux of **3a** (pyCTZ) and **3d** (6pyDTZ) from teLuc by ~ 120% and ~ 150%, respectively (Figure 2c). The directed evolution process to enhance photon flux of teLuc for 8pyDTZ did not preclude the luciferase from catalyzing other structurally relevant substrates. LumiLuc is capable of efficiently generating blue, teal, or yellow bioluminescence when paired with pyCTZ, 6pyDTZ or 8pyDTZ ( $\lambda_{\max}$ : 450, 476, and 525 nm, respectively; Figure 2d), thereby leading to a new family of ATP-independent bioluminescent reporters with water-soluble substrates.

### LumiLuc-8pyDTZ in cultured mammalian cells

We next evaluated LumiLuc-8pyDTZ in human embryonic kidney (HEK) 293T cells transiently expressing the luciferase (Figure S7). The LumiLuc-8pyDTZ pair produced ~ 3- to 5-fold more bioluminescence than teLuc-8pyDTZ at all tested substrate concentrations (Figure S8). Moreover, despite that LumiLuc-8pyDTZ is less bright than teLuc-DTZ at saturated substrate concentrations, LumiLuc-8pyDTZ is notably brighter than teLuc-DTZ at low substrate concentrations (from 6.25 to 25  $\mu\text{M}$ ; Figure 3a). Far-red emission at

wavelengths longer than 600 nm is more indicative of the *in vivo* performance of bioluminescent reporters, because mammalian tissue is more transparent in this spectral region.<sup>25</sup> To compare far-red emission intensities of bioluminescent reporters, we imaged HEK 293T cells in the presence of a 600–700 nm bandpass filter. At substrate concentrations from 6.25 to 100  $\mu$ M, LumiLuc-8pyDTZ consistently produces 1.6- to 3.9-fold higher photon flux than teLuc-DTZ (Figure 3b).

ATP-dependent luciferases, such as FLuc and Akaluc consumes one ATP molecule in each catalytic cycle, leading to metabolic disruption.<sup>9</sup> Instead, ATP-independent LumiLuc does not use ATP for catalysis. We monitored ATP/ADP ratios in live HEK 293T cells using PercevalHR, a previously reported fluorescent ATP/ADP biosensor.<sup>27</sup> No ATP perturbation was observed from 8pyDTZ-treated, LumiLuc-expressing cells (Figure S9).

### LumiLuc-8pyDTZ to track tumor growth in a mouse xenograft model

BLI has been a popular imaging modality for various animal models.<sup>14, 25</sup> The recently reported Akaluc-AkaLumine and Antares2-DTZ pairs are two benchmark reporters for *in vivo* BLI.<sup>7, 8</sup> We adapted a biologically-relevant tumor xenograft mouse model<sup>28</sup> to compare these bioluminescent reporters. We first generated cervical cancer HeLa cell lines stably expressing individual luciferases, including teLuc, Antares2, LumiLuc, and Akaluc (Figure S10). Next, we injected  $10^4$  or  $10^5$  luciferase-expressing HeLa cells into the left or right dorsolateral trapezius and thoracolumbar regions of immunodeficient NU/J mice (day 0) and monitored tumor growth over 4 weeks. Bioluminescence was quantified in days 1, 3, 5, 7, 14, and 28 after tail vein injection of corresponding substrates. AkaLumine-HCl was delivered at a dose of 1.5  $\mu$ mol per mouse. This dosage ( $\sim 75$  nmol/g), which is normalized against the body weights of mice, is identical to the previously reported dosage.<sup>7</sup> Moreover, when 3  $\mu$ mol of AkaLumine-HCl per mouse ( $\sim 150$  nmol/g) was used, we observed death for 2 out of 3 mice in our pilot experiment. 8pyDTZ was dissolved in normal saline to its saturation concentration and intravenously injected, resulting in a dose of 0.2  $\mu$ mol per mouse ( $\sim 10$  nmol/g). The LumiLuc-8pyDTZ pair showed detectable bioluminescence on day 1 at sites inoculated with  $10^4$  cells, and kept exhibiting  $\sim 3$ -fold higher photon flux over Akaluc-AkaLumine up to day 7 (Figures 4a and 4b). The signals for Akaluc-AkaLumine at sites inoculated  $10^4$  cells were not consistently higher than background until day 3. Furthermore, the *in vivo* brightness of LumiLuc-8pyDTZ is comparable to, if not higher than, the Antares2-DTZ pair (Figures 4 and S11), despite the fact that the majority of emitted photons from LumiLuc-8pyDTZ has not yet exceeded 600 nm. These data collectively support that LumiLuc-8pyDTZ is a superior bioluminescent reporter system for high-sensitivity *in vivo* BLI.

The bioluminescence of Akaluc-AkaLumine eventually surpassed LumiLuc-8pyDTZ from day 14 (Figure S11). In addition to differences in biodistribution and pharmacokinetic properties of AkaLumine and 8pyDTZ, we interpret that 8pyDTZ may be a limiting reagent in large tumors because AkaLumine-HCl could be delivered into mice at a much higher dose than 8pyDTZ due to the higher solubility of AkaLumine-HCl. Presumably, we may further enhance the *in vivo* performance of marine luciferases and their derivatives by further increasing the water solubility and thus the administration dosage of CTZ and DTZ analogs.

## Engineering of BRET-based LumiScarlet for deep-tissue BLI

mScarlet-I is a recently reported red fluorescent protein with high quantum yield and excellent performance as a Förster resonance energy transfer (FRET) acceptor.<sup>29</sup> We thus hypothesized that LumiLuc could be genetically fused to mScarlet-I for BRET, thereby red-shifting the emission of LumiLuc. We explored several fusion strategies between LumiLuc and mScarlet-I, constructed libraries by randomizing the linkers, and screened for mutants with high BRET efficiency (Figures S12a and S12b). We identified a mutant, namely LumiScarlet (Figure 5a), which is a fusion protein of mScarlet-I (residues 1–225) linked to the N-terminus of LumiLuc (residues 2–169) through a single-residue “Lys” linker.

High BRET efficiency was achieved with LumiScarlet in the presence of either pyCTZ, or 6pyDTZ, or 8pyDTZ (Figure 5b). In particular, because the emission spectrum of LumiLuc-8pyDTZ overlaps well with the excitation spectrum of mScarlet-I (Figure S12c), ~51% of the total emission of LumiScarlet, when paired with 8pyDTZ, was longer than 600 nm (Table S1).

We next compared our newly engineered LumiLuc-8pyDTZ and LumiScarlet-8pyDTZ with Akaluc-AkaLumine for deep-tissue BLI. We injected a million HeLa cells stably expressing corresponding luciferases into each of NU/J mice via tail vein and performed BLI imaging 4 h later. Immunodeficient mice were used here to minimize immune responses to HeLa cells, so that signals will be mostly from live cells trapped in the lungs. LumiScarlet gave ~3-fold higher detectable signals than LumiLuc under this condition (Figure 5c and 5d), even though the *in vitro* brightness of LumiScarlet is only ~70% of LumiLuc (Figure S12d). Moreover, the signals from LumiScarlet-8pyDTZ were comparable to the signals from Akaluc-AkaLumine.

We want to note that we observed some diffuse signals from areas other than the lungs. These signals were not caused by substrate background, as injection of 8pyDTZ into blank mice (Figure S13a) resulted in only weak background much lower than what we observed in Figure 5c. Luciferase activities were detectable in blood after tail vein injection of luciferase-expressing HeLa cells (Figure S13b), suggesting partial lysis of luciferase-labeled cells during cell injection. Also, in contrast to ATP-independent LumiLuc and LumiScarlet, ATP-dependent Akaluc is enzymatically inactive in serum because of relatively low ATP levels and further deactivation by serum components.<sup>9</sup> Thus, diffuse signals from LumiScarlet in Figure 5c were likely from the LumiScarlet luciferase released into blood.

Collectively, our deep-tissue BLI results confirm that red-shifted BRET-based LumiScarlet has better mammalian tissue penetration than LumiLuc. Moreover, LumiScarlet-8pyDTZ is a novel, ATP-independent bioluminescent reporter with exceptional deep-tissue BLI performance comparable to ATP-dependent Akaluc-AkaLumine.

## DISCUSSION

Conventionally, ATP-dependent bioluminescent reporters, such as FLuc and Akaluc, are considered to be more useful for *in vivo* BLI than ATP-independent marine luciferases, because the emission of ATP-dependent insect luciferases is often at the red end of the



visible spectrum where the mammalian tissue is relatively transparent. However, these insect luciferases require ATP and  $Mg^{2+}$  for bioluminescence. The ATP- and  $Mg^{2+}$ -dependency is sometimes problematic because ATP and  $Mg^{2+}$  levels may vary under different biological circumstances.<sup>30</sup> In particular, ATP-dependent luciferases are inactive in extracellular space and common biological fluids such as blood and urine, where ATP accessibility is limited.<sup>9</sup> Moreover, ATP-dependent luciferases consume ATP in bioluminescence reactions and may cause concerns such as metabolic disruption.<sup>9</sup> In contrast, most ATP-independent marine luciferases are enzymatically active in extracellular space and common biological fluids; they do not consume ATP for bioluminescence. Furthermore, some marine luciferase derivatives have fast catalytic turnover and thus give high photon flux. It is therefore not surprising that marine luciferase and their derivatives, such as NanoLuc and *Gaussia* luciferase, have been widely used for *in vitro* bioluminescence assays. Currently, the *in vivo* applications of marine luciferases are hindered by their blue emission and poor substrate water solubility. In this study, we combined chemical synthesis and protein engineering approaches to enhance ATP-independent marine luciferases for *in vivo* BLI by developing red-shifted colors and water-soluble substrates.

First, we prepared a series of pyridyl CTZ and DTZ analogs with diverse emission profiles. The water solubility of these synthetic analogs generally increased by ~10-fold from their ancestors. These substrate analogs can not only be paired with the new luciferases engineered here, but also existing ATP-independent reporters, such as RLuc and aequorin.

We further engineered a luciferase for the 8pyDTZ substrate via directed protein evolution. The resultant LumiLuc-8pyDTZ bioluminescent reporter system exhibited reduced  $K_M$  and red-shifted emission. These factors favored *in vivo* BLI. As a result, LumiLuc-8pyDTZ showed high sensitivity in a mouse xenograft model. In addition, LumiLuc-8pyDTZ did not perturb the intracellular ATP/ADP level, and 8pyDTZ could be dissolved up to ~2 mM in low-viscosity saline without using irritative and toxic organic cosolvent. Therefore, our effort enhanced not only the biocompatibility of bioluminescent reporters, but also reproducibility for intravenous injections.

Furthermore, we developed a BRET-based LumiScarlet reporter for further red-shifted emission. The emission of LumiLuc-8pyDTZ overlaps well with the excitation of mScarlet-I, an excellent red-emitting resonance energy transfer acceptor. LumiScarlet-8pyDTZ exhibited high brightness, significant emission longer than 600 nm, and excellent tissue penetration. LumiScarlet-8pyDTZ was comparable to NIR-emitting Akaluc-AkaLumine in a mouse model for deep-tissue BLI. Moreover, because LumiScarlet is enzymatically active in blood, we envision that it would be an excellent reporter for monitoring targets of interest in the blood of *in vivo* models.

LumiLuc is a luciferase with broad substrate specificity. When it was paired with different substrates, intense blue, teal, and yellow bioluminescence was generated. Subsequently, we gained different emission profiles from LumiScarlet in the presence of different substrates. We demonstrated the use of LumiScarlet-8pyDTZ for deep-tissue imaging. In addition, because the two emission peaks of LumiScarlet-pyCTZ or LumiScarlet-6pyDTZ are more separated than LumiScarlet-8pyDTZ, we envision that LumiScarlet-pyCTZ and

LumiScarlet-6pyDTZ would be useful for studying protein-protein interactions or constructing BRET-based biosensors.

In summary, we have developed several engineered luciferase-luciferin pairs that emit photons spanning an appreciable range in the visible spectrum. Our effort has greatly enhanced the biocompatibility and sensitivity of ATP-independent bioluminescent reporters for *in vivo* BLI. We expect that future studies will continuously increase the water-solubility of CTZ and DTZ analogs and red-shift the emission of marine luciferases. Subsequently, we expect that a large array of bioluminescent biosensors will be developed on the basis of these bright, ATP-independent bioluminescent reporters.<sup>31</sup> The new reporters and biosensors will further ease non-invasive imaging of freely moving animals, leading to new biological insights.

## METHODS

Experimental details are given in the Supporting Information.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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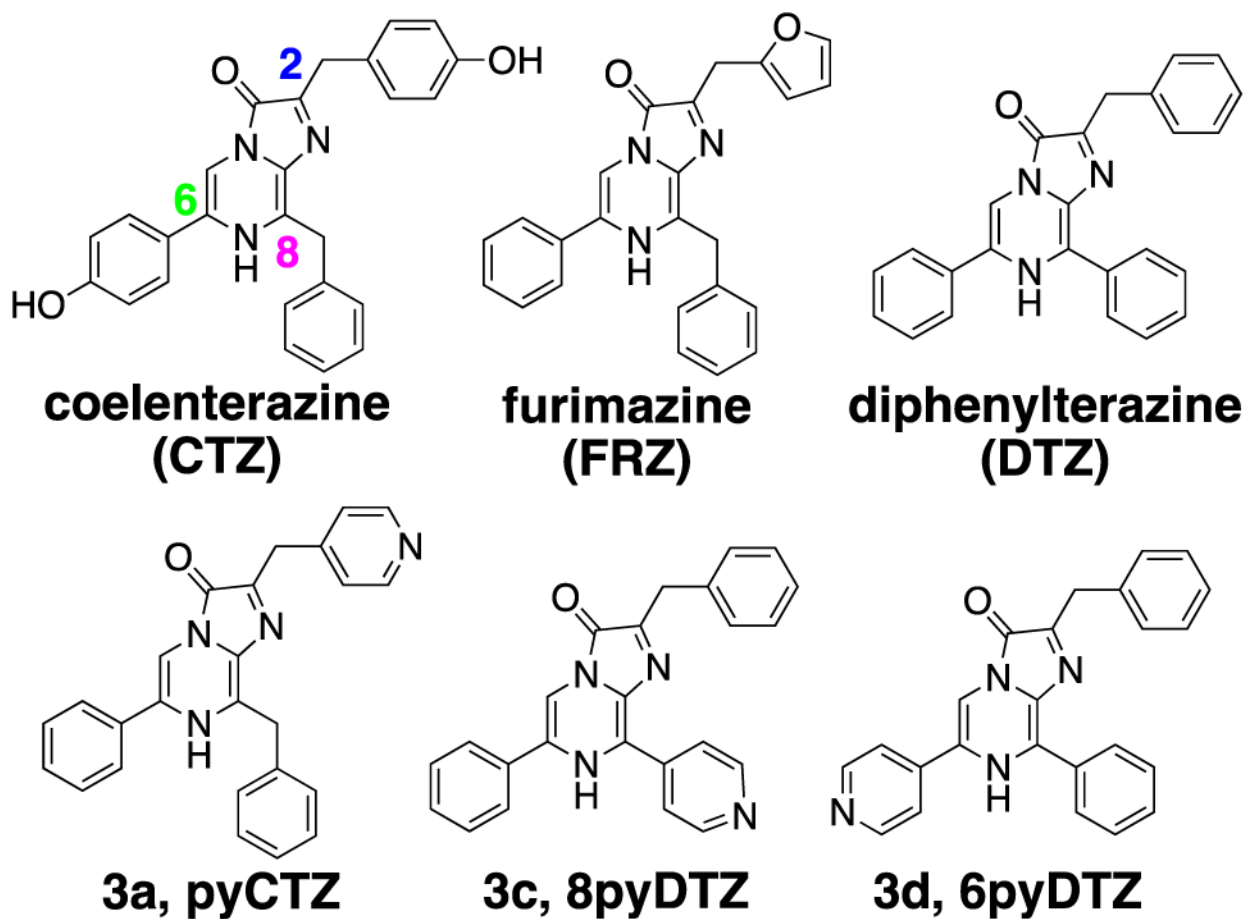
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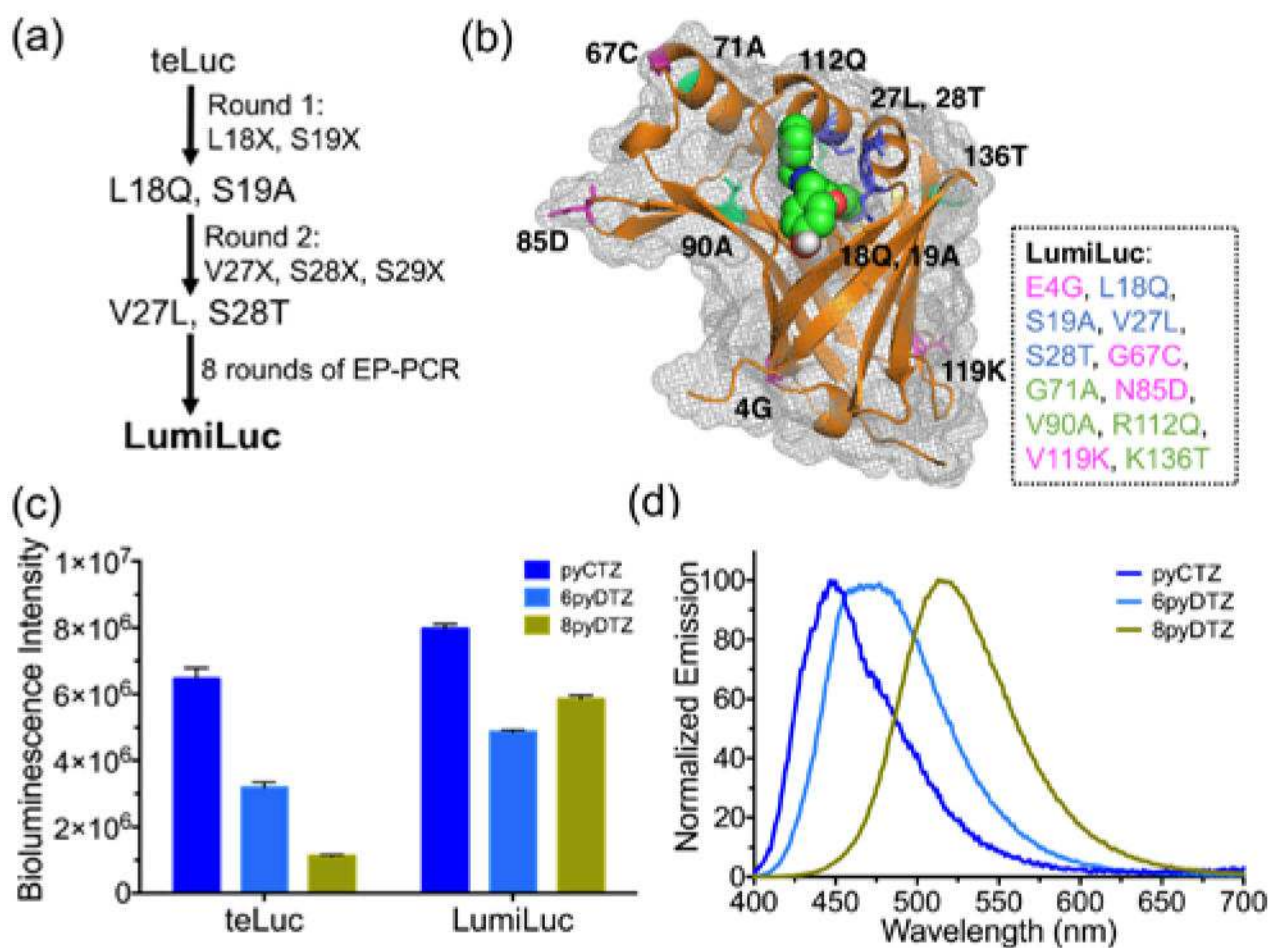


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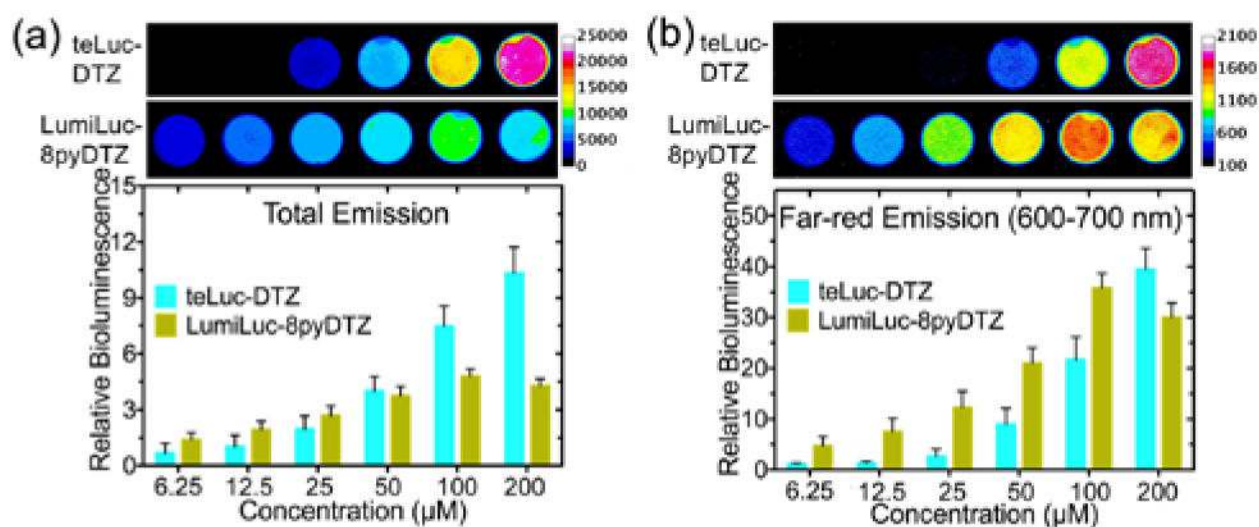


**Figure 1.** Chemical structures of coelenterazine (CTZ), furimazine (FRZ), diphenylterazine (DTZ), pyCTZ (**3a**), 8pyDTZ (**3c**) and 6pyDTZ (**3d**).

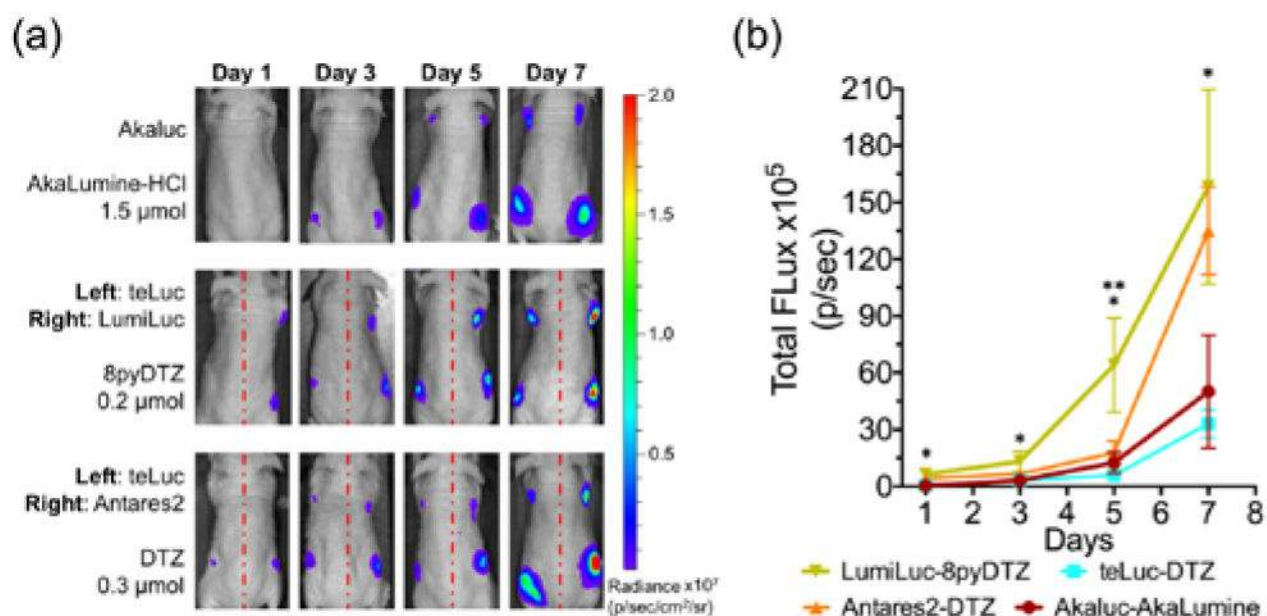


**Figure 2. Engineering of a LumiLuc luciferase.**

(a) Procedure to derive LumiLuc from teLuc. (b) Illustration of the putative substrate-binding site and LumiLuc mutations from teLuc. CTZ is shown as spheres and mutated residues are presented in sticks. (c) Bioluminescence of pyCTZ, 6pyDTZ, or 8pyDTZ in the presence of either teLuc or LumiLuc. (d) Normalized bioluminescence emission spectra of pyCTZ, 6pyDTZ, and 8pyDTZ in the presence of LumiLuc.



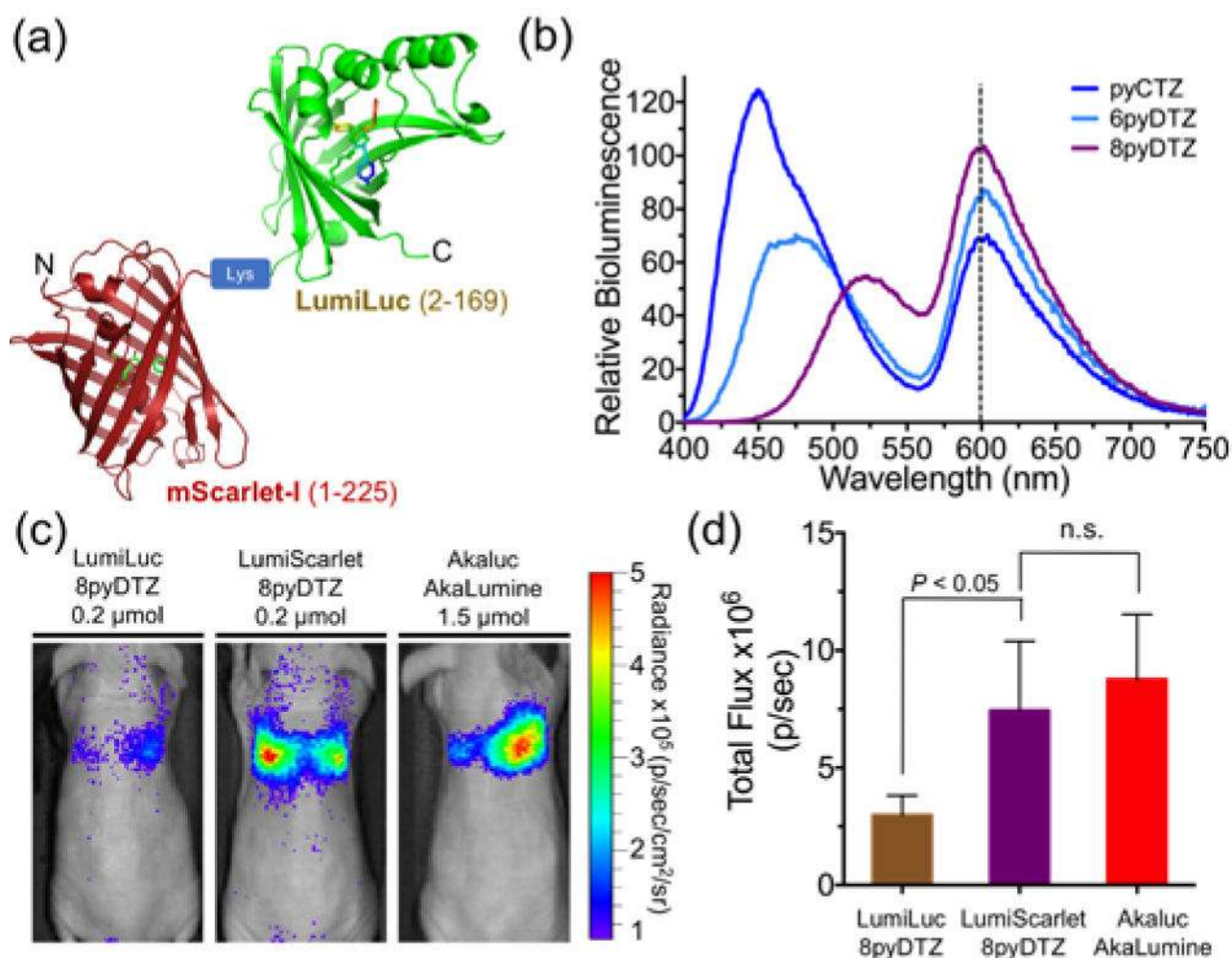
**Figure 3. Bioluminescence of teLuc- and LumiLuc-expressing HEK 293T cells.** Images were acquired (a) without or (b) with a 600-700 nm bandpass filter. Values for relative brightness were normalized to teLuc in the presence of 6.25  $\mu\text{M}$  DTZ.



**Figure 4. Tracking of tumor growth in a xenograft mouse model with various luciferase-luciferin pairs.**

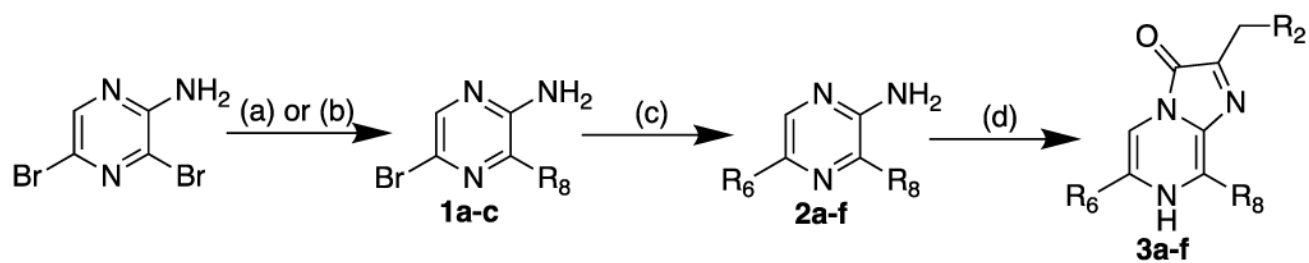
(a) BLI ( $n = 4$ ) on day 1, 3, 5, and day 7.  $10^4$  luciferase-expressing HeLa cells were injected to the left and right dorsolateral trapezius regions and  $10^5$  cells were injected to the left and right dorsolateral thoracolumbar regions of NU/J mice. For i.v. administration of substrates, AkaLumine-HCl ( $1.5 \mu\text{mol}/\text{mouse}$ ) and 8pyDTZ ( $0.2 \mu\text{mol}/\text{mouse}$ ) were dissolved in normal saline, and DTZ ( $0.3 \mu\text{mol}/\text{mouse}$ ) was formulated with a mixture of organic cosolvents. (b) Comparison of luciferase-luciferin pairs at tumor sites inoculated with  $10^4$  cells. (\* $p < 0.05$  for LumiLuc-8pyDTZ and teLuc-DTZ, and for LumiLuc-8pyDTZ and Akaluc-AkaLumine; \*\* $p < 0.05$  for LumiLuc-8pyDTZ and Antares2-DTZ).





**Figure 5. BRET-based LumiScarlet for deep tissue BLI.**

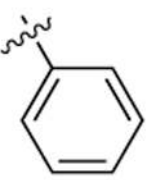
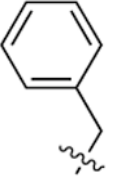
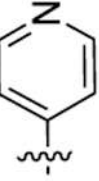
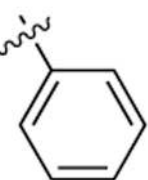

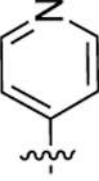
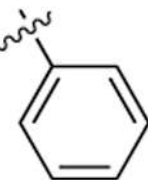
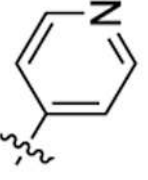
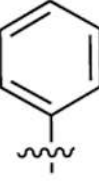
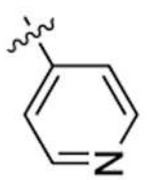

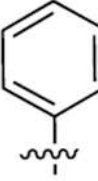
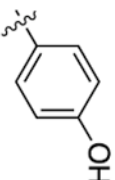
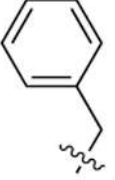
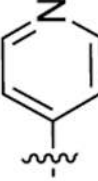
(a) Schematic diagram of LumiScarlet, a genetic fusion of mScarlet-I and LumiLuc. (b) Bioluminescence emission of LumiScarlet in the presence of pyCTZ, 6pyDTZ, or 8pyDTZ, showing significant emission longer than 600 nm. (c) Comparison of LumiLuc-8pyDTZ, LumiScarlet-8pyDTZ, and Akaluc-AkaLumine in NU/J mice (n = 4) at 4 h post i.v. injection of  $10^6$  luciferase-expressing HeLa cells. (d) Quantitative analysis of bioluminescence from the regions around the lungs in panel c (n.s.: not significant).

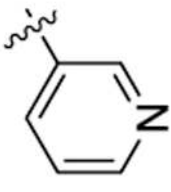
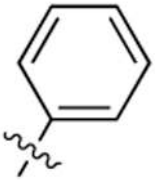
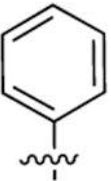
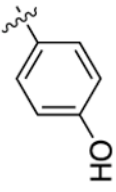
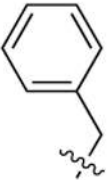
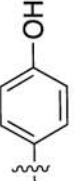
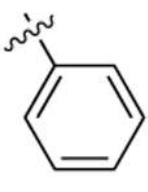
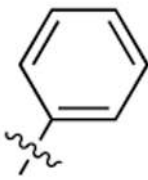
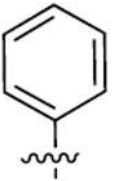


**Scheme 1. Synthesis of pyridyl CTZ and DTZ analogs.**

(a) Suzuki coupling:  $\text{Pd}(\text{PPh}_3)_4$ ,  $\text{Na}_2\text{CO}_3$ ,  $R_8\text{-B}(\text{OH})_2$ , and EtOH; (b) Negishi coupling:  $\text{PhCH}_2\text{MgCl}$ ,  $\text{ZnCl}_2$ ,  $(\text{PPh}_3)_2\text{PdCl}_2$ , and THF; (c) Suzuki coupling: XPhos-Pd-G2,  $\text{Na}_2\text{CO}_3$ ,  $R_6\text{-B}(\text{OH})_2$ , and EtOH; (d) Acid-catalyzed ring closing: corresponding  $\alpha$ -ketoacetal, HCl, and dioxane.

**Table 1.** Chemical and photoluminescence properties of synthetic pyridyl CTZ and DTZ analogs.

Compound	R <sub>6</sub>	R <sub>8</sub>	R <sub>2</sub>	Bioluminescence <sup>a</sup> λ <sub>max</sub> (nm)	Chemiluminescence <sup>b</sup> λ <sub>max</sub> (nm)	Water Solubility (μM)
3a				451	505	1416
3b				497	506	1813
3c				532	555	1711
3d				483	492	1736
3e				450	465	987

Compound	R <sub>6</sub>	R <sub>8</sub>	R <sub>2</sub>	Bioluminescence <sup>a</sup> λ <sub>max</sub> (nm)	Chemiluminescence <sup>b</sup> λ <sub>max</sub> (nm)	Water Solubility (μM)
3f				518	503	1562
CTZ				455	461	256
DTZ				502	510	131

Note:

<sup>a</sup>Determined with 1 nM teLuc in PBS;

<sup>b</sup>Triggered by peroxydicarbonate formed *in situ*.