INVITED REVIEW

Axonal damage: a key predictor of outcome in human CNS diseases

I. M. Medana¹ and M. M. Esiri^{2,3}

Departments of ¹Clinical Laboratory Sciences and ²Clinical Neurology, University of Oxford and ³Department of Neuropathology, Oxford Radcliffe NHS Trust, Oxford, UK

Summary

Axonal damage has recently been recognized to be a key predictor of outcome in a number of diverse human CNS diseases, including head and spinal cord trauma, metabolic encephalopathies, multiple sclerosis and other white-matter diseases (acute haemorrhagic leucoencephalitis, leucodystrophies and central pontine myelinolysis), infections [malaria, acquired immunodeficiency syndrome (AIDS) and infection with human lymphotropic virus type 1 (HTLV-I) causing HTLV-I- Correspondence to: Professor M. M. Esiri, Neuropathology Department, Radcliffe Infirmary, Oxford OX2 6HE, UK E-mail: margaret.esiri@clneuro.ox.ac.uk

associated myelopathy (HAM)/tropical spastic paraparesis (TSP)] and subcortical ischaemic damage. The evidence for axonal damage and, where available, its correlation with neurological outcome in each of these conditions is reviewed. We consider the possible pathogenetic mechanisms involved and how increasing understanding of these may lead to more effective therapeutic or preventive interventions.

Keywords: axonal damage; neuropathology; cerebral malaria; multiple sclerosis; acute haemorrhagic encephalitis; vascular dementia; HIV encephalitis

Abbreviations: AHLE = acute haemorrhagic leucoencephalitis; AIDS = acquired immunodeficiency syndrome; β -APP = β -amyloid precursor protein; DAI = diffuse axonal injury; HAM = HTLV-I-associated myelopathy; HIV = human immunodeficiency virus; HIVD = HIV dementia; HTLV-I = human lymphotropic virus type 1; ICP = increased intracranial pressure; TAI = traumatic axonal injury

Introduction

Axonal damage and its profound consequences for outcome in human brain and spinal cord injury and disease were first fully recognized in the context of head trauma. Severe diffuse axonal injury caused by mechanical shearing and stretch forces, inflicted particularly in rotational acceleration injuries (Gennarelli *et al.*, 1982), underlies the persistent vegetative state in subjects who survive severe head injury (Strich, 1956; Graham *et al.*, 1983; Adams *et al.*, 1989, 1999). It is also commonly present in those who succumb rapidly to such an injury (Gentleman *et al.*, 1993; Oehmichen *et al.*, 1998). At the other end of the scale of severity of traumatic injury, axonal tears were the lesions identified by Oppenheimer (1968) as likely to underlie transient loss of consciousness in those who suffer a mild head injury and die of something else.

Adoption of immunocytochemical staining for amyloid precursor protein (β -APP), a new technique that allows more

sensitive detection of damage to axons than previous methods have done, has opened the way for axonal damage to be investigated more systematically and in more detail in a greater variety of diseases than hitherto (Gentleman et al., 1993; Sherriff et al., 1994b; Blumbergs et al., 1995; McKenzie et al., 1996; Geddes et al., 1997). Such studies have shown that axon damage occurs in a wide range of CNS disorders. At the same time, imaging studies of the CNS in some of these disorders have reinforced recognition of the serious functional consequences of axonal damage (Matthews et al., 1998; Grimaud et al., 1999; Fisher et al., 2000; Paolillo et al., 2000; Pelletier et al., 2001). This recognition of the importance of axonal damage has implications both for diagnosis and treatment or prophylaxis of CNS damage in these diseases. In this review we summarize recent findings about axonal damage in a range of different CNS diseases,

and consider its importance for functional disability and recovery and for diagnosis and treatment.

Detecting damaged axons

The traditional stains for visualizing axons, the single, slender processes that emerge from neurons at the axon hillocks and extend with a relatively uniform calibre to the terminals up to a metre or more away, are silver stains that bind to the neurofilaments. With such stains, damaged axons appear swollen because of the interruption in their fast transport system and the proximal accumulation of organelles and fluid.

Immunocytochemical staining for β -APP more sensitively detects axons that have impaired fast axonal transport (Fig. 1). In normally functioning axons, the protein is transported in this way and never builds up to a concentration that allows its detection in tissue sections. However, axons that have this transport system disrupted rapidly accumulate β -APP proximal to the disrupted segment. This occurs before conventional morphological evidence of axonal damage (e.g. in the form of axonal end bulbs) develops, so the immunocytochemical method for detecting β -APP is more sensitive than routine histological methods for detecting axon damage. Other proteins transported by fast axonal transport also accumulate but antibodies to β -APP have been shown to be the most sensitive for detecting this type of damage (Grady et al., 1993; Gultekin and Smith, 1994; Ng et al., 1994; Sherriff et al., 1994a; Li et al., 1995; Pesini et al., 1999). Experimental animal studies of brain trauma have shown that some axonal damage is reversible but it is not known if axonal damage severe enough to be detected with β -APP immunoreactivity in humans is ever reversible. With regard to the timing of damage detectable in this way, the immunoreaction in damaged axons for B-APP becomes positive in head trauma 1-3 h after the insult (Sherriff et al., 1994b; McKenzie et al., 1996; Oehmichen et al., 1998) and remains positive for up to 1 month (Geddes et al., 2000). However, the possibility that degenerate axons survive in this state for longer is raised by the findings of Blumbergs and colleagues that β -APP immunoreactivity remained detectable 99 days after head trauma (Blumbergs et al., 1994). The distal parts of irreversibly damaged axons will undergo Wallerian degeneration, which can be detected by such traditional methods as the Marchi technique on tissue sections and in living subjects by neuroimaging with (Banati et al., 2000) or without (Simon et al., 2000) novel ligands. An estimate of the scale of long-previous irreversible axonal damage can be obtained by performing estimates of axon numbers, a relatively straightforward task using modern computerized image analysis facilities.

Head and spinal cord trauma *Head trauma*

Head trauma encompasses a broad spectrum of focal and diffuse pathologies. Focal brain injury is most often

associated with an impact to the head whereas diffuse brain injury results from inertial forces that are commonly produced by road-traffic accidents, falls from a height and, in some cases, from assaults and sports injuries (Gennarelli *et al.*, 1982; Geddes *et al.*, 2000). Focal brain injury may produce mass effects from haemorrhage, contusion or haematoma, which can induce herniation and brainstem compression. As a result, coma may not be immediate but may develop later. Diffuse brain injury has been demonstrated in animal models to induce an immediate and prolonged post-traumatic unconsciousness in the absence of mass lesions (Gennarelli *et al.*, 1982).

Damage to axons is almost a universal finding in cases of mild, moderate and severe head trauma (Gentleman *et al.*, 1995) with outcomes ranging from mild concussion (Jane *et al.*, 1985; Blumbergs *et al.*, 1994; Adams *et al.*, 2001) to profound coma and even the vegetative state (Kinney and Samuels, 1994; Fitzpatrick *et al.*, 1998; Adams *et al.*, 1999). Resultant traumatic axonal injury (TAI) may vary from small foci of axonal injury to the most severe form, diffuse TAI, originally termed diffuse axonal injury (DAI), in which there is widespread axonal injury throughout the brain, including the brainstem (Geddes *et al.*, 2000) (Fig. 1A).

Recently, Smith and colleagues have explored the anatomical origins of post-traumatic coma using a pig model of inertial brain injury induced by rotational acceleration of the head in the axial and coronal planes (Smith et al., 2000). They found that immediate and prolonged coma was produced only by head axial plane rotation, although DAI was produced by head rotation along both planes. However, extensive axonal damage in the brainstem was found only in the pigs injured by head rotation in the axial plane. Furthermore, the severity of coma was found to correlate with both the extent of axonal damage in the brainstem and the applied kinetic loading conditions. No relationship was found between coma and the extent of axonal damage in other brain regions. Thus, two major findings evolved from this work: injury to axons in the brainstem plays a major role in the induction of immediate post-traumatic coma, and DAI can occur without coma (Smith et al., 2000).

Even minor head injury is thought to result in structural damage to axons (Oppenheimer, 1968; Jane et al., 1985; Blumbergs et al., 1994). Minor degrees of axonal injury may provide an explanation for concussion, and for cognitive and behavioural sequelae in less severe head injury (Oppenheimer, 1968; Blumbergs et al., 1994). Work in animal models also supports this assertion (Povlishock et al., 1983; Jane et al., 1985). Brief concussive injury was produced in monkeys by acceleration-deceleration non-impact injury and the presence of degenerating axons was investigated using the Nauta and Fink-Heimer techniques (Jane et al., 1985). Degenerating axons were found predominantly in the brainstem but not all of the axonal projections in a given system were destroyed. This led the authors to suggest that the surviving neural elements would replace those that were degenerating, in particular the synaptic connections.

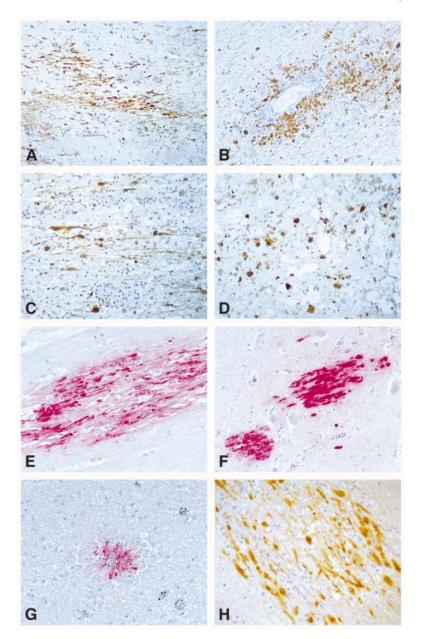


Fig. 1 Brain sections stained for β-amyloid precursor protein (β-APP) (brown immunostain in **A**–**D** and **H**, red immunostain in **E**–**G**) to visualize areas of axonal injury in patients who died with head injury (**A**), acute haemorrhagic leucoencephalitis (AHLE) (**B**), central pontine myelinolysis (CPM) (**C**–**D**), malaria (**E**–**G**) and HIV (**H**). (**A**) Patients who die with head injury often show waves of diffuse axonal staining. (**B**) AHLE is characterized by striking perivenous axon damage as shown here. (**C**–**D**) In CPM, axon damage is seen in the acutely demyelinated base of the pons and affects both transversely running fibres, seen in longitudinal section in **C**, and longitudinally running fibres, seen cut in transverse section in **D**. (**E**–**G**) Axonal damage is a common finding in patients with severe malaria (**E**–**F**). Focal patches of axonal injury are intensely immunoreactive for β-APP, are clearly demarcated, and consist of linear arrays of damaged axonal segments that often appear grossly swollen or club-shaped (**E**–**F**). Injured axons associated with a vessel containing sequestered malaria parasites and a haemorrhage (**G**). The extent and distribution of axonal injury are significantly higher in severe malaria patients with cerebral complications (**F**–**G**). (**H**) Disseminated and swollen axonal segments in cerebral white matter from a patient who died with HIV encephalitis.

However, it was Polvishock and colleagues, using a cat model, who suggested that the damage to axons may be reversible in minor head injury i.e. disruption of axons without physical shearing or tearing (Povlishock *et al.*, 1983). There have been only limited studies of minor head trauma in humans, as patients rarely die from their injuries. One study examined 20 patients with moderate disability after traumatic brain damage who survived for 1–47 years after injury (Adams et al., 2001). DAI was found in 30% of the patients but the diagnosis of axonal injury depended on the microglial reaction rather than the identification of acute abnormalities in axons because of the long period of survival in every case. Blumbergs and colleagues examined five cases of concussive head injury in humans who died of other causes (Blumbergs et al., 1994). Multifocal axonal injury in structures thought to be involved in consciousness and memory function was demonstrated using β-APP immunohistochemistry. In particular, there was involvement of the fornices, which are the major hippocampal projection pathways and are thought to be important in memory. Therefore the authors suggested that this may underlie some of the persisting memory deficits that occur in patients after concussion. The mechanisms of TAI have been investigated at the cellular level (reviewed in Maxwell et al., 1997) and will be discussed in more detail below.

Paediatric head trauma

Head injury in infancy and childhood is the single most common cause of death and permanent disability. The type of injury that results depends on the mechanism of trauma and the age of the patient. Young infants are particularly susceptible to head injury because of the thinness and pliability of the skull; the softness of the brain, which is composed primarily of neurons without dendritic connections; the paucity of the myelin sheaths of axons; the relative flatness of the skull base; the underdeveloped neck muscles supporting the large, heavy head; and the nature of the subarachnoid space, which is large in its extent but shallow in depth (reviewed by Case *et al.*, 2001). As the child becomes mobile, falls become the number one cause of accidental injury, frequently producing fractures and contusions (Zimmerman and Bilaniuk, 1994).

The most comprehensive neuropathological studies on the structural basis of clinical deficits in fatal paediatric brain damage have been made by Geddes and colleagues (Geddes et al., 2001a, b). These studies involved 53 cases of fatal nonaccidental head injury, which included β-APP immunohistochemistry for microscopic damage. The most important finding was a statistically significant pattern of age-related axonal damage. When axonal injury was present in children aged >1 year, the pattern was similar to that of DAI in adults. However, in the infant group, aged <1 year, DAI was rare whereas hypoxia was the predominant neuropathological finding. Axonal pathology appeared predominantly vascular in nature, associated with brain swelling and raised intracranial pressure. Anatomically, 31% showed evidence of localized axonal injury to the craniocervical junction or the cervical cord. This indicated that the craniocervical junction is vulnerable in infant head injury as a result of stretch injury from cervical hyperextension/flexion. The authors proposed that this damage could account for the observed apnoea, which could in turn lead to hypoxic brain damage. Two explanations for this finding given by this group were (i) the

unmyelinated axon of the immature cerebral hemispheres is relatively resistant to traumatic damage, and (ii) in shakingtype injuries the brain is not exposed to the forces necessary to produce DAI.

Spinal cord injuries

Spinal cord injury is often caused by mechanical depression from displacement of fractured vertebrae and/or discs extending into the spinal canal (Nashmi and Fehlings, 2001a, b). The injury often spans several segments and the lesion site is characterized by central cavitation and a subpial rim of surviving axons of small diameter with myelin disruption. However, the spinal cord is rarely totally transected even after severe spinal cord injury associated with complete paralysis (Nashmi and Fehlings, 2001a, b). The neurological deficits produced by spinal cord injury are predominantly caused by the loss of white matter, particularly the long tracts through which descending and ascending communications occur (Rosenberg *et al.*, 1999).

Immediately after fatal spinal cord injury in humans there is complete disruption of a proportion of axons and partial injury to the remaining axons. Injured axons show irregular varicosities, spheroids and β-APP immunoreactivity (Ahlgren and Olsson, 1996; reviewed in Nashmi and Fehlings, 2001b). Recently, Nashmi and Fehlings (2001b) reviewed the time course of events following spinal cord injury. Eight days to 5 weeks after injury, the injury site is characterized by axonal swelling, fragmentation and increased spacing between axons. Seven to eight weeks after injury, Wallerian degeneration becomes evident in dorsal columns above the injury and lateral columns below the injury, with debris-laden phagocytes intermingled with axonal and myelin debris. Invading macrophages remove the debris at the lesion and scar tissue forms due to astrogliosis. In the chronic stage, the lesions consist of glial-lined, multilocular cysts, nerve root regeneration and a subpial rim of preserved white matter.

The relationship between axonal sparing and neurological outcome has been studied extensively in rat, cat and ferret models of spinal cord injury (Eidelberg *et al.*, 1981; Fehlings and Tator, 1995). In these models, <12% of the normal number of axons were shown to be required for recovery of neurological function, such as restoration of locomotion. In human cases it was shown that there was no relationship between neurological function and the extent of preservation of white matter spinal cord tissue (reviewed in Nashmi and Fehlings, 2001*b*). However, retrograde tracing studies in a rat model of spinal cord injury showed that the integrity of certain spinal tracts, namely the rubrospinal, vestibulospinal and raphespinal tracts, correlated with neurological recovery (reviewed in Fehlings and Tator, 1995; Nashmi and Fehlings, 2001*b*).

Several mechanisms have been proposed to explain the dysfunction of surviving axons that traverse the site of spinal cord injury. In the case of experimental contusion injury, the white matter appears mostly intact. However, over the next 4 h pathology increases, suggesting that secondary injury mechanisms are involved in the loss of white matter (reviewed in Rosenberg *et al.*, 1999). Some of these secondary injury mechanisms include: altered activity of ion channels, abnormal myelination, disturbances in mito-chondrion function, excitotoxicity, changes in glial–axonal coupling, nitric oxide and anti-ganglioside antibodies (reviewed in Dumont *et al.*, 2001; Nashmi and Fehlings, 2001*b*).

Metabolic encephalopathies

There are various pathologies that may contribute to or be the cause of β -APP immunoreactivity in axons. This has implications for medicolegal practice and has inspired a number of groups to investigate the relationship between metabolic disruption and axonal injury. In this context it is particularly important to determine whether the patterns of axonal injury as a result of physical trauma and subsequent trauma-related mechanisms can be dissociated.

Several groups have investigated the role of hypoxia in the induction of axonal injury. Kaur and colleagues have suggested that cardiac arrest *per se* can mimic DAI (Kaur *et al.*, 1999). This is disputed by Dolinak and colleagues, who believe that proper sampling and due attention to brain swelling and increased intracranial pressure (ICP) allow the patterns to be distinguished (Dolinak *et al.*, 2000*a*, *b*). On the other hand, carbon monoxide poisoning, an example of histotoxic hypoxia that does not result in raised ICP, causes considerable damage in the white matter (Dolinak *et al.*, 2000*a*). However, the patterns of β -APP staining can be diverse, ranging from specific foci to a wide distribution.

Dolinak and colleagues have investigated the role of hypoglycaemia in the causation of axonal injury. In this study, extensive neuropathological examination was undertaken in 13 patients in whom coma was attributed to hypoglycaemia. Immunoreactivity for β -APP was present in 11 of the 13 cases. The amount of axonal injury could be attributed to the hypoglycaemia alone, although the amount and the distribution of the axonal injury were altered in the presence of raised ICP. In one case the distribution closely mimicked that seen in microscopic diffuse TAI (Dolinak *et al.*, 2000*b*).

Multiple sclerosis

Since the earliest descriptions of the pathology of multiple sclerosis, it has been characterized as a disease in which there is multifocal demyelination with relative preservation of axons. The relative axonal preservation serves to distinguish a plaque of demyelination from a white-matter infarct. The early investigators of multiple sclerosis were well aware of axonal damage in multiple sclerosis plaques (reviewed by Kornek and Lassmann, 1999) and of Wallerian degeneration secondary to this. However, relatively few attempts were

made to quantify axonal loss (Greenfield and King, 1936; Putnam, 1936) and relate it to clinical disability. Two recent developments have served to put axonal damage under greater scrutiny in multiple sclerosis. The first is the availability of the β -APP immunoreaction, which provides a much better means of detecting damaged axons in tissue sections, as mentioned above. The second is non-invasive MRI and spectroscopic methods that provide accurate means of assessing axon loss in living patients and allow longitudinal clinicopathological studies to be undertaken in multiple sclerosis for the first time. These latter studies have shown, on the one hand, that demvelination on its own does not account for chronic functional impairments, and on the other that magnetic resonance spectroscopy (MRS) and MRI measures of axonal loss show a strong correlation with disability in multiple sclerosis (Matthews et al., 1998; Grimaud et al., 1999; Fisher et al., 2000; Paolillo et al., 2000; Pelletier et al., 2001). In consequence comes the important realization that if disability is to be prevented we need to understand when, where and how axon damage arises in the disease.

There is now considerable consensus that axon damage occurs early in the course of multiple sclerosis. The two initial studies of β -APP immunoreactivity in multiple sclerosis showed that axonal damage occurs in acute plaques (Ferguson et al., 1997; Trapp et al., 1998) and this has been confirmed by later studies (Bitsch et al., 2000; Kornek et al., 2000; Kuhlmann et al., 2002). Acute plaques and the actively demyelinating borders of chronic active plaques display numerous β-APP-immunoreactive damaged axons, while chronic plaques contain few, if any (Ferguson et al., 1997; Kornek et al., 2000; Kuhlmann et al., 2002) (Fig. 2). The greatest abundance of damaged axons is seen in the acute plaques occurring in the early years of disease, as detected in biopsy specimens taken before the clinical diagnosis was certain (Kuhlmann et al., 2002). In several ways this early occurrence of axonal damage in multiple sclerosis is counterintuitive to established views of the disease. A primarily demyelinating disease might be expected to show demyelination followed by axon loss rather than simultaneous axon and myelin destruction. Furthermore, the initial relapsing-remitting pattern of disease in the early stages in most cases does not fit well with early axon damage, which is presumed to be largely irreversible. Axon loss occurring much later would have fitted better with secondary progression of disease. However an initial ability to repair at least some damage to axons and a normally substantial reserve followed by a progressive failure of repair would be compatible with the clinical course.

The overwhelming majority of damaged axons are found in or at the borders of plaques, and very few occur in normalappearing white matter. The damage to axons in plaques results in Wallerian degeneration, which is readily detectable as reduced axon density or total number in normal-appearing white matter (reduced by ~50%) as well as in plaques (reduced by 50–82%) (Mews *et al.*, 1998; Ganter *et al.*, 1999;

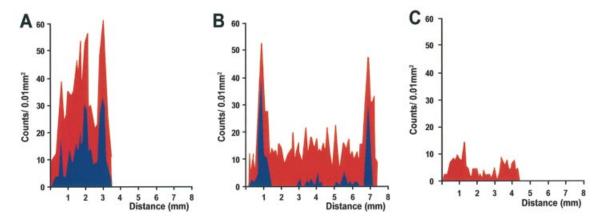


Fig. 2 Graphs illustrating the density of macrophages (blue) and APP⁺ axons (red) at distances from the centre of an acute plaque (**A**), a chronic active plaque (**B**) and a chronic inactive plaque (**C**) taken from autopsy material from cases of multiple sclerosis. (From an original figure published in Ferguson *et al.*, 1997. Reproduced with permission of Oxford University Press.)

Evangelou *et al.*, 2000*a*, 2001; Lovas *et al.*, 2000). That damage to axons in plaques is responsible for axon loss in normal-appearing white matter is supported by the finding that axon density in the corpus callosum strongly correlates with the volume of plaques in the cerebral hemispheres (Evangelou *et al.*, 2000*b*).

The question of how axon damage occurs in multiple sclerosis is harder to answer. Is it secondary to the loss of myelin or is it a direct effect of other changes, such as inflammation, that are taking place in acute plaques? Obviously, clarification of this point has important implications for trying to prevent disability in multiple sclerosis. If axon damage is secondary to demyelination, vigorous and early promotion of remyelination might be helpful, but if the damage is directly inflicted remyelination may have little effect. One interesting feature of the axonal loss in multiple sclerosis is that it is remarkably specific for small nerve fibres and spares large fibres, though whether this reflects selective damage of small fibres or selective inability of these fibres to recover from damage is not clear (Ganter et al., 1999; Lovas et al., 2000; Evangelou et al., 2001). That axonal damage has a reversible element in acute disease is suggested by the MRS observation of a temporary decrease in N-acetyl aspartate, a neuronal marker, in acute relapses of multiple sclerosis (Davie et al., 1994).

Inflammatory mediators are attractive candidates for axondamaging agents in acute multiple sclerosis plaques. Nitric oxide and its synthetic enzyme nitric oxide synthase, which is detectable in increased amounts in multiple sclerosis plaques (Bo *et al.*, 1994; Bagasra *et al.*, 1995; De Groot *et al.*, 1997; Bitsch *et al.*, 2000), has been shown to damage axons irreversibly *in vitro*, small fibres being particularly at risk (Smith *et al.*, 2001; Garthwaite *et al.*, 2002). Inducible nitric oxide synthase mRNA in multiple sclerosis plaques was found to correlate with axon density (Bitsch *et al.*, 2000). Matrix metalloproteinases, which are released in acute multiple sclerosis plaques (Anthony *et al.*, 1997; Gveric *et al.*, 2001; Leppert *et al.*, 2001; Lindberg *et al.*, 2001), are also known to be capable of damaging axons *in vitro* (Gijbels *et al.*, 1993; Chandler *et al.*, 1995). Autoimmune encephalitis in experimental animals appears to mimic the disability-related axonal loss seen in multiple sclerosis and may afford a convenient opportunity to study axon-damaging mechanisms of relevance to multiple sclerosis (Kornek *et al.*, 2000; Wujek *et al.*, 2002). More general aspects of the mechanics of axon damage are discussed below.

Other demyelinating diseases

Axon damage in other demyelinating diseases has been studied in much less detail than in multiple sclerosis but, as in that disease, it has long been recognized that axon loss occurs both in chronic diseases, such as the leucodystrophies (Schaumburg et al., 1975; Lake, 1997; Powers and Moser, 1998), and acute diseases, such as acute haemorrhagic leucoencephalitis (AHLE). In the latter para-infectious disease there is striking perivenous axon damage demonstrable with β -APP immunocytochemistry (Fig. 1B), whereas in the other acute para-infectious disease, acute perivenous encephalitis, very little axon damage is demonstrable (unpublished observations, Ghosh et al.). Axon damage is also a prominent feature of central pontine myelinolysis, another acute demyelinating condition with a high mortality rate seen in severely debilitated or alcoholic subjects who undergo rapid correction of hyponatraemia (N.Ghosh, G.C.De Luca, M.M.Esiri, unpublished observations) (Fig. 1C and D).

Infections *Malaria*

Impairment of consciousness and other signs of cerebral dysfunction are common complications of severe falciparum malaria. The majority of neurological complications are transient but a significant minority of patients develop sequelae (Newton *et al.*, 2000). The mechanism by which malaria infection induces severe but potentially reversible neurological dysfunction remains elusive. The malaria parasite invades and develops within erythrocytes, which sequester in the cerebral microvasculature by adhesion to specific endothelial receptors (Turner, 1997). However, it is not known how malaria parasites, which remain within the vascular space and do not infect brain cells, influence parenchymal brain function to induce coma and possibly death.

Cerebral white matter lesions have been associated with neurological complications in malaria infection (Davis et al., 1992; Kochar and Makkar, 1994; Dugbartey et al., 1998; Lewallen et al., 1999; Medana et al., 2002). In our recent pathological studies of cerebral malaria in Vietnamese adults (Medana et al., 2002) we have identified impairment of transport within axons as a possible cause of neurological dysfunction that has the potential either to resolve or to progress to irreversible damage. B-APP immunocytochemistry was performed on brain sections from the cortex, internal capsule, pons and cerebellum of 54 adult Vietnamese patients with falciparum malaria to determine whether defects in axonal transport would reflect cerebral impairment in this group. The extent and distribution of axonal damage was found to distinguish the groups of patients infected with Plasmodium falciparum with and without cerebral complications during life. This is the only finding, in our series, of positive quantitative associations with clinical manifestations of cerebral malaria with the exception of parasite sequestration. There were significant associations between axonal damage and important clinical and biochemical parameters, including lactate, CSF protein and Glasgow coma score. β-APP staining was also found in 'pure cerebral malaria' patients without other organ complications. However, the mechanisms of axonal injury remain elusive because (i) axonal injury could be found independently of oedema, haemorrhage and glial responses, (ii) there were no associations between axonal injury and ICP or systemic hypoglycaemia, (iii) there was no association with impairment of vital organ function, such as renal failure, jaundice and shock, and (iv) there was no exacerbation of axonal injury with increasing numbers of criteria of severity, which may imply that although these patients are increasingly ill and more likely to die, the extent of axonal injury within the CNS is determined independently or early in the disease (Medana et al., 2002). These findings suggest that axons are vulnerable to a broad range of cerebral insults that occur during falciparum malaria infection. Disruption in axonal transport may represent a final common pathway leading to neurological dysfunction in cerebral malaria (Medana et al., 2002).

These pathological findings are consistent with studies of cerebral malaria patients during life. Impairment of somatosensory discrimination and conduction has been found in patients with a history of cerebral malaria (Kochar and Makkar, 1994; Dugbartey *et al.*, 1998). Also, cotton wool spots have been found in retinas of patients with cerebral malaria (Davis *et al.*, 1992; Lewallen *et al.*, 1999). In both cases the findings are likely to be manifestations of axonal disruption or obstruction of axonal flow (McLeod *et al.*, 1977). Thus, discerning the mechanisms of axonal injury is likely to be an important step in understanding the specific neurological complications associated with cerebral malaria.

Kochar and Makkar (1994) recorded somatosensory evoked potentials by median nerve stimulation in 10 adult patients from India with cerebral malaria. They observed abnormalities in 80% of patients, of which the commonest problem was prolongation of central conduction time. It was not possible for the authors to elucidate the exact pathogenetic mechanisms responsible for the electrophysiological changes. However, they suggested that a phase exists in cerebral malaria which causes delay in the conduction of travelling impulses in the brain due to damage in the white matter and changes in volume conduction at the partition of geometric boundaries.

Dugbartey and colleagues also have investigated, using neuropsychological tests, neuropathological changes in paediatric patients with a history of cerebral malaria (Dugbartey et al., 1998). They pointed out that the corpus callosum is the largest neocortical fibre tract in the brain that facilitates efficient communication between the cerebral hemispheres. They reasoned that if cerebral malaria shows a predilection for cerebral white matter, interhemispheric transfer inefficiencies should be demonstrable. Bimanual tactile roughness discrimination was found to be significantly impaired in the cerebral malaria group. In contrast, intrahemispheric processing of tactile information was intact. This group suggested that inefficiency in the integrity of the callosal fibres could account for these findings, although damage to alternative subcortical pathways involved in transfer across the cerebral hemispheres could not be ruled out entirely.

Cotton wool spots have been found in the retinas of patients with cerebral malaria (Davis et al., 1992; Lewallen et al., 1999). The intense retinal whiteness of small cotton wool spots represents gross localized axonal distensions secondary to the cessation of axoplasmic flow (McLeod et al., 1977). Davis and colleagues found that cotton wool spots were observed much more frequently than retinal haemorrhages in adult Thai patients with severe malaria (Davis et al., 1992). Furthermore, all but one of the patients with cotton wool spots at presentation had an impairment of consciousness. In these cases the cotton wool spots were found in association with capillary non-perfusion. However, two comatose patients had neither cotton wool spots nor non-perfusion. The lack of cotton wool spots in these patients was thought to reflect the marked variability in the sequestered parasite biomass among patients with severe malaria as well as the variations within individuals and tissues. Cotton wool spots are less common in African children, occurring in 5% of those with cerebral malaria (Lewallen et al., 1999).

HIV

Neurological disease is a common occurrence in patients with acquired immunodeficiency syndrome (AIDS), causing clinical symptoms ranging from cognitive impairments, motor disturbances, behavioural changes, headache and peripheral neuropathy (reviewed by Lawrence and Major, 2002). It has been estimated that 10% of AIDS patients will have a CNS lesion as the first clinical manifestation and 40% will have some type of neurological complication during the course of the disease, and that >70% of AIDS autopsies will demonstrate neuropathological findings (reviewed by Gonzalez et al., 1998). A proportion of neurological disease will be associated with opportunistic infections as a result of immunodeficiency. However, in the absence of these infections 20-30% of patients will develop neurocognitive defects: human immunodeficiency virus (HIV)-1-associated cognitive motor complex, also termed HIV dementia (HIVD). At autopsy, the brains of patients with HIVD show numerous disseminated foci composed of microglia, macrophages and multinucleated giant cells (Gray et al., 1998), termed HIV encephalitis (Masliah et al., 1992). The presence of subclinical decline in cognitive performances in HIV patients before significant immunodeficiency is controversial. However, it has been shown that HIV is present in the CSF and brain during the early asymptomatic phases of HIV infection (An et al., 1997).

Although the neuropathological correlates of the clinical manifestations remain unclear, several candidates have been proposed, including viral load, myelin pallor and neuronal loss. As with cerebral malaria, an intriguing problem is that neurons are not the target of infection. In addition, there is some evidence supporting the reversibility of neurological complications. An example highlighted by Lawrence and Major (2002) is that some patients may improve after treatment and further deterioration may be delayed, suggesting that irreversible nerve cell loss is unlikely to be involved. It is likely that HIVD reflects neuronal dysfunction resulting from several pathological mechanisms, which will be discussed in more detail in later sections.

With the introduction of APP immunohistochemistry, widespread axonal injury in the white matter of AIDS patients and to a lesser degree in pre-AIDS cases has been revealed (An et al., 1997; Giometto et al., 1997; Raja et al., 1997; Gray et al., 1998; Adle-Biassette et al., 1999). APP+ axons have been found predominantly in the subcortical white matter, basal ganglia (An et al., 1997; Giometto et al., 1997; Raja et al., 1997; Gray et al., 1998; Adle-Biassette et al., 1999) and the brainstem, including pontocerebellar fibres (Gray et al., 1998; Adle-Biassette et al., 1999). APP+ axons are commonly found in association with microglia/ macrophages and multinucleated giant cells. These results have led some authors to hypothesize that areas of disturbed axonal injury in clinically active regions of the brain may contribute to the appearance of neuropsychological symptoms (Giometto et al., 1997).

Associations between axonal injury and other neuropathological findings have been made to try to define the possible mechanisms of axonal injury in HIV infection. Diffuse myelin pallor is a common autopsy finding in HIV infection and is more frequent in those who show features of encephalopathy (reviewed in Raja et al., 1997). In the majority of reports there is a parallel between APP+ axons and myelin pallor. However, axonal injury is also present in the absence of pallor. This led Raja and colleagues to suggest that β -APP staining is a more sensitive marker of some forms of white matter damage in HIV infection (Raja et al., 1997). The link between myelin pallor and axonal injury is less clear in some reports of HIV+ individuals without AIDS (An et al., 1997) and is clearly not involved in a featured case of AIDS with a relapsing course with neurological signs (Gray et al., 1998). In the former study, of 29 HIV-1+ asymptomatic patients, pallor was minimal in three cases and absent in the others. Axonal injury in the absence of myelin pathology has also been described in a patient who died after a relapsing course of neurological signs. In this case axonal damage was an extremely frequent finding without other neuropathological changes or evidence of productive HIV infection of the brain (Gray et al., 1998). Further, in a monkey model of HIV using simian immunodeficiency virus-infected macaques, significant increases in β -APP were found in the white matter in the absence of myelin pallor (Mankowski et al., 2002). From these results it is clear that axonal injury can be a primary manifestation of HIV infection in the brain.

The association of axonal injury with vessels has also been a topic of interest, and has produced variable findings. In the report by Raja and colleagues, APP+ foci showed an approximately perivascular distribution (Raja et al., 1997). Similar results were found in the case report of Gray and colleagues (Gray et al., 1998). In both cases the authors emphasized the potential role of systemic vascular-related factors in the pathogenesis of the axonal damage. In contrast, the study of HIV⁺ individuals without AIDS showed that the majority of β -APP did not colocalize with vessels (An *et al.*, 1997). Similar findings were made in another study by the same group (Giometto et al., 1997). In this study the authors made a distinction between β -APP⁺ structures—ballooned structures representing chronic axonal damage, and bundles of parallel formations representing acute axonal damage. They reported that, with few exceptions, acute axonal damage was not related to blood vessels.

Thus, although this is not conclusive, axonal injury remains an attractive candidate to explain the functional impairment in HIV⁺ patients and discerning the mechanisms of axonal injury is strongly warranted.

Human T lymphotropic virus type I (HTLV-I)

HIV is not the only viral disease that indirectly leads to axonal injury in the CNS. Another example is HTLV-I, which is associated with adult T-cell leukaemia and a chronic progressive disease of the spinal cord termed HTLV-I- associated myelopathy (HAM)/tropical spastic paraparesis (TSP) (Umehara *et al.*, 2000). Neuropathological studies have revealed infiltration of T lymphocytes and macrophages and increased expression of cytokines. However, demyelination and axonal loss have been described as the histological hallmarks of inflamed lesions of HAM/TSP and the pathological correlates of persistent disability that characterize spastic paraparesis and urinary disturbance.

How the CNS damage develops is unclear but, as in HIV, it is unlikely to be the result of direct infection with virus (Levin et al., 2002). Other findings similar to those with HIV are the association with myelin pallor and degeneration, although axonal injury can be found in the absence of myelin pathology. Furthermore, APP+ axons tend to be located in relation to blood vessels, particularly veins, with or without inflammatory cell infiltrates. Again, these findings have led to the conclusion that alteration in the blood-brain barrier and seepage of neurotoxins into the CNS might relate to the primary axonal changes in HAM/TSP (Umehara et al., 2000). Unlike HIV, the neurotoxic mechanisms are more clear, and evidence points to an autoimmune aetiology (Levin et al., 2002) involving autoantibodies and cytotoxic T cells. The role of autoimmunity in axonal injury will be discussed in a later section.

Subcortical ischaemic disease

Subcortical ischaemic disease is extremely common in the elderly. It was found in 78% of a recent community-acquired sample of 209 brains obtained from subjects dying aged >65 years (Esiri et al 2001). It is also a common substrate for vascular dementia (Esiri et al., 1997; Vinters et al., 2000; White et al., 2002). It takes the form of lacunes (small infarcts in subcortical grey and white matter) and multifocal areas of subtotal infarction or pallor of staining in myelin-stained sections of white matter associated with hyaline thickening of the walls of small arteries and arterioles and widening of perivascular spaces. A few cases with this pattern of pathology suffer from the inherited condition CADASIL (cerebral autosomal dominant arteriopathy with subcortical infarcts and leucoencephalopathy), in which, in addition to the features outlined above, there are granular periodic acid-Schiff (PAS)-positive deposits in the media of small subcortical arteries that are associated with an immunoreaction for the Notch 3 receptor (Joutel et al., 2000). People suffering from vascular dementia with this type of pathology usually show a neuropsychological profile of cognitive impairment more suggestive of subcortical than of cortical disease, and this impairment has been largely attributed to the demyelination and/or axon loss that occurs in the white matter (Skoog et al., 1996; Inzitari et al., 2000). There have been limited numbers of studies examining B-APP immunoreactivity as a marker of axon damage in vascular dementia, and these have described patchy axonal damage (Suenaga et al., 1994; Teahan et al., 2002). However, use of this marker is likely to underestimate the extent of axonal damage in this

disease because it only discloses relatively recently damaged axons while the clinical disease itself often lasts years. Therefore, as in multiple sclerosis, the extent of axon *depletion* is likely to be a better measure of axon damage over the course of the whole disease. Estimates of the severity of axon depletion in vascular dementia are, however, complicated by the fact that most cases of vascular dementia show some degree of Alzheimer-type pathology as well, and this disease can also be associated with axonal depletion, e.g. in the corpus callosum. The axonal depletion in Alzheimer's disease is thought to represent Wallerian degeneration secondary to neuronal loss from the association cortex (Weis *et al.*, 1991; Bozzali *et al.*, 2002).

Mechanisms of axonal injury

Selective axonal injury can occur in the absence of focally related somatic or dendritic alterations. Axons often extend for great distances from their cell bodies of origin, and are therefore susceptible to ischaemic or toxic damage in several different vascular territories, without the death of the parent neuron. Regardless of injury mode, axonal injury is associated with a broadly similar pattern of disruption, involving increased axonal membrane permeability, deleterious intracellular cascades and disturbance of axonal transport and possibly degeneration (Fig. 3). In most cases the injury mechanisms can occur in neuronal compartments other than the axon.

Increased axonal membrane permeability

Physical deformations (Ziv *et al.*, 1995; Rosenberg *et al.*, 1999; Wolf *et al.*, 2001), energy deficits (Fink *et al.*, 1994), acidosis (Hsu *et al.*, 2000) and neuroinflammatory disorders (Kornek *et al.*, 2001) are some of the causes of changes in the activity of ion channels leading to unphysiological increases in intra-axonal Na⁺ and Ca²⁺ (reviewed in LoPachin and Lehning, 1994). Some of the proposed mechanisms for the change in activity are increased Na⁺ channel permeability (Rosenberg *et al.*, 1999; Wolf *et al.*, 2001), decreased ATPase activity (Fink *et al.*, 1994; Tavalin *et al.*, 1997; Ahmed *et al.*, 2000), ectopic distribution of channels (Kornek *et al.*, 2001) and reversal of gradient-dependent ion channels (Stys, 1998; Wolf *et al.*, 2001).

Deleterious intracellular cascades

Changes at the axolemma are usually followed by excess Ca^{2+} influx into the axon. Ca^{2+} is pivotal in mediating axonal injury and degeneration. Mitochondrial perturbations are consistent with calcium overloading and opening of the mitochondrial membrane permeability transition (MPT) pore, which permeabilizes the mitochondrial membrane for molecules <1.5 kDa. The process leads to the uptake of water, mitochondrial swelling and ultimate mitochondrial rupture (reviewed in Büki *et al.*, 2000). Abnormal/pathological

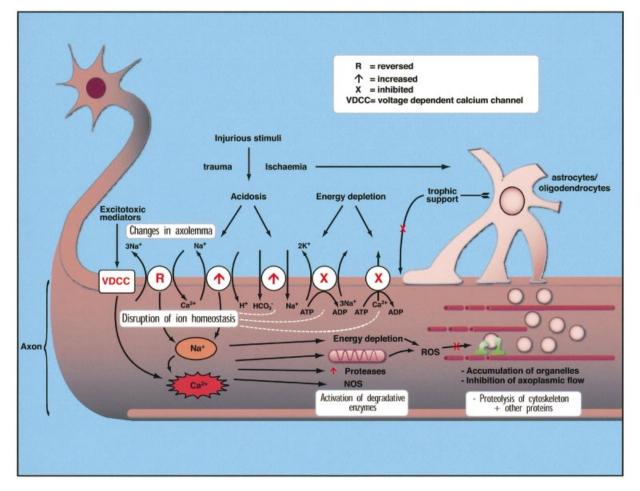


Fig. 3 Diagram illustrating mechanisms of axonal injury. Regardless of injury mode, axonal injury is associated with a broadly similar pattern of disruption: (i) increased axonal membrane permeability with excess Na^+ and/or Ca^{2+} influx into the axon; (ii) deleterious cascades involving activation of intracellular proteases, failure of the mitochondria and cytoskeleton breakdown; and (iii) disturbance of axonal transport and possibly degeneration. See text for more details.

mitochondria, indicative of some form of subcellular perturbation, are associated with cytochrome c release into the perimitochondrial domain. Cytochrome c release is associated with concomitant caspase activation in the axon (Büki *et al.*, 2000).

The activation of calcium-activated cysteine proteases and/ or caspases can continue to devastate intra-axonal cytoskeletal and organelle alterations, leading to the ultimate demise of the axon. Caspases can irreversibly cleave intraaxonal spectrin (Wang *et al.*, 1998) with ultimate destruction of the cytoskeleton, an event considered to be a major factor in the neuronal degeneration seen in various CNS disorders. Caspase-3 has also been reported to cleave calpastatin, an inhibitor of calpain that might further enhance the ongoing axonal demise (Wang *et al.*, 1998; Büki *et al.*, 2000). Calpain acts on several substrates, many of which are cytoskeletal proteins, such as neurofilament proteins, microtubule-associated proteins and spectrin (Saido *et al.*, 1994; Banik *et al.*, 1997; Springer *et al.*, 1997; Schumacher *et al.*, 1999).

Structural disorganization and/or dissolution of the cytoskeleton are likely to result in severe functional derangements in the axon and that would ultimately affect the neuron as a whole (Fitzpatrick *et al.*, 1998).

Indirect effects on axons

There are several indirect mechanisms that can lead to axonal injury. Neuronal–glial interactions are important in the maintenance of brain homeostasis and are vital for neuronal survival after brain injury. Death of glia or loss of function of glia, e.g. acidosis inhibiting glutamate transporters, could affect the outcome of injured axons (Liu *et al.*, 1999).

Glia and invading haematogenous cells may be a source of neurotoxic factors produced in response to infectious agents and/or damage to the neural tissue (Giulian *et al.*, 1993). Cytotoxic T cells (Manning *et al.*, 1987; Medana *et al.*, 2001; reviewed in Neumann *et al.*, 2002), macrophages and microglia have been implicated in the induction of axonal injury (Brück *et al.*, 1996; Bitsch *et al.*, 2000). Similarly, autoantibodies (Sloviter *et al.*, 1996; Rawes *et al.*, 1997; Genain *et al.*, 1999), metalloproteinases (Newman *et al.*, 2001) and other inflammatory mediators, such as tumour necrosis factor and nitric oxide, have also been implicated (Brück et al., 1996; Giovannoni et al., 1998).

Treatments

The question remains as to which processes associated with axonal injury constitute the best target for potential therapeutic intervention. The answer probably lies in the spatial and temporal progression of events. In addition, the interplay of multiple factors including Ca²⁺ buffering capacity, global versus selective nature of the injury, energy demands and production, duration of axonal injury and the resulting magnitude of the Ca2+ burden will influence outcome (LoPachin and Lehning, 1997). Potentially beneficial therapy would most likely act on a target that is activated before irreversible structural damage of the axon has occurred. Na+ channel blockers have been shown to improve functional outcome and reduce axonal pathology in models of spinal cord crush injury (Teng and Wrathall, 1997) and nitric oxideinduced damage (Kapoor et al., 2002), and may have application in treating diffuse axonal injury. Blockade of voltage-gated calcium channels has also been suggested as a potential therapeutic strategy for traumatic axonal injury.

Early phases of TAI involving calcium-activated neutral proteases, such as calpain, have been discussed as rational therapeutic targets. Several experimental studies have reported the potential efficacy of calpain inhibitors in traumatic brain injury (Schumacher *et al.*, 2000; Ray *et al.*, 2001). The mitochondrial permeability transition pore has also been suggested as an appropriate target for therapeutic intervention. This has been supported by the finding that the use of inhibitors of the mitochondrial permeability transition, which preserve mitochondria, also translate into significant axonal protection (Büki *et al.*, 2000).

Conclusion

This review has considered the evidence that axonal damage is of crucial importance in determining the outcome in a remarkably wide range of diseases from the acutely fatal (cerebral malaria, severe brain trauma, acute haemorrhagic leucoencephalitis) through those with an intermediate time of survival (HIV encephalopathy), to the long-standing (multiple sclerosis, vascular dementia). The severity of axon damage in these diseases helps to determine outcome in terms of death or disability. However, the question needs to be asked: is it the axon damage and loss itself that determines outcome, or something else that is closely linked to axon damage? Is it reasonable to attribute death in the acute diseases and disability in the chronic ones to axon damage per se? In the rapidly fatal diseases axon damage is widespread and includes axons in the brainstem that are necessary to maintain connections to and from nuclei whose function is essential for life, such as the respiratory and circulatory centres in the medulla. In young infants, for example, it is proposed that a mechanism of fatal non-accidental head

trauma involves a stretch injury at the craniocervical junction, resulting in apnoea from interference with respiratory function, hypoxic brain swelling and death as a consequence of brainstem compression. Some cases of immediately fatal head trauma in adult life are associated with traumatic separation of the pons and medulla (Lindenberg and Freytag, 1970; Simpson *et al.*, 1989), but lesser tears of axons in the brainstem are found commonly in those dying shortly after a head injury, particularly those involved in road-traffic accidents, though there may also be other lesions that account for death.

In fatal AHLE the degree of cerebral swelling is usually sufficient to account for the fatal outcome, though this does not seem to be true of cerebral malaria. In the more chronic diseases, the best studied of which is multiple sclerosis, the extent and distribution of axon loss correlates well with the clinical disability. Thus, the small fibres in the optic nerves which subserve colour vision are selectively lost and defective colour vision is the most prominent visual symptom in multiple sclerosis (Evangelou et al., 2001). Likewise, axon loss in the corticospinal tracts of the spinal cord well explains the common spastic paraparesis seen in multiple sclerosis (Ganter et al., 1999; Lovas et al., 2000). More detailed study of fatal neurological diseases associated with axonal injury and loss will be needed to clarify other factors that may be involved and that may be amenable to therapeutic interventions. All in all, the recognition of the importance of axonal damage in these diseases opens up new avenues for exploration in attempts to improve the management and treatment of these all-too-common conditions.

Acknowledgements

I.M.M. is supported by the Lloyd's of London Tercentenary Foundation and the Wellcome Trust. I.M.M. wishes to thank Drs N. Day, J. Farrar, C. Newton, G. Turner and N. White for helpful discussions.

References

Adams JH, Doyle D, Ford I, Gennarelli TA, Graham DI, McLellan DR. Diffuse axonal injury in head injury: definition, diagnosis and grading. Histopathology 1989; 15: 49–59.

Adams JH, Jennett B, McLellan DR, Murray LS, Graham DI. The neuropathology of the vegetative state after head injury. J Clin Pathol 1999; 52: 804–6.

Adams JH, Graham DI, Jennett B. The structural basis of moderate disability after traumatic brain damage. J Neurol Neurosurg Psychiatry 2001; 71: 521–4.

Adle-Biassette H, Chretien F, Wingertsmann L, Hery C, Ereau T, Scaravilli F, et al. Neuronal apoptosis does not correlate with dementia in HIV infection but is related to microglial activation and axonal damage. Neuropathol Appl Neurobiol 1999; 25: 123–33.

Ahlgren S, Li GL, Olsson Y. Accumulation of beta-amyloid precursor protein and ubiquitin in axons after spinal cord trauma in

526 I. M. Medana and M. M. Esiri

humans: immunohistochemical observations on autopsy material. Acta Neuropathol (Berl) 1996; 92: 49–55.

Ahmed SM, Rzigalinski BA, Willoughby KA, Sitterding HA, Ellis EF. Stretch-induced injury alters mitochondrial membrane potential and cellular ATP in cultured astrocytes and neurons. J Neurochem 2000; 74: 1951–60.

An SF, Giometto B, Groves M, Miller RF, Beckett AA, Gray F, et al. Axonal damage revealed by accumulation of beta-APP in HIV-positive individuals without AIDS. J Neuropathol Exp Neurol 1997; 56: 1262–8.

Anthony DC, Ferguson B, Matyzak MK, Miller KM, Esiri MM, Perry VH. Differential matrix metalloproteinase expression in cases of multiple sclerosis and stroke. Neuropathol Appl Neurobiol 1997; 23: 406–15.

Bagasra O, Michaels FH, Zheng YM, Bobroski LE, Spitsin SV, Fu ZF, et al. Activation of the inducible form of nitric oxide synthase in the brains of patients with multiple sclerosis. Proc Natl Acad Sci USA 1995; 92: 12041–5.

Banati RB, Newcombe J, Gunn RN, Cagnin A, Turkheimer F, Heppner F, et al. The peripheral benzodiazepine binding site in the brain in multiple sclerosis: quantitative in vivo imaging of microglia as a measure of disease activity. Brain 2000; 123: 2321–37.

Banik NL, Matzelle DC, Gantt-Wilford G, Osborne A, Hogan EL. Increased calpain content and progressive degradation of neurofilament protein in spinal cord injury. Brain Res 1997; 752: 301–6.

Bitsch A, Schuchardt J, Bunkowski S, Kuhlmann T, Bruck W. Acute axonal injury in multiple sclerosis. Correlation with demyelination and inflammation. Brain 2000; 123: 1174–83.

Blumbergs PC, Scott G, Manavis J, Wainwright H, Simpson DA, McLean AJ. Staining of amyloid precursor protein to study axonal damage in mild head injury. Lancet 1994; 344: 1055–6.

Blumbergs PC, Scott G, Manavis J, Wainwright H, Simpson DA, McLean AJ. Topography of axonal injury as defined by amyloid precursor protein and the sector scoring method in mild and severe closed head injury. J Neurotrauma 1995; 12: 565–72.

Bo L, Dawson TM, Wesselingh S, Mork S, Choi S, Kong PA, et al. Induction of nitric oxide synthase in demyelinating regions of multiple sclerosis brains. Ann Neurol 1994; 36: 778–86.

Bozzali M, Falini A, Franceschi M, Cercignani M, Zuffi M, Scotti G, et al. White matter damage in Alzheimer's disease assessed in vivo using diffusion tensor magnetic resonance imaging. J Neurol Neurosurg Psychiatry 2002; 72: 742–6.

Brück W, Sommermeier N, Bergmann M, Zettl U, Goebel HH, Kretzschmar H, et al. Macrophages in multiple sclerosis. Immunobiology 1996; 195: 588–600.

Büki A, Okonkwo DO, Wang KK, Povlishock JT. Cytochrome c release and caspase activation in traumatic axonal injury. J Neurosci 2000; 20: 2825–34.

Case ME, Graham MA, Handy TC, Jentzen JM, Monteleone JA. Position paper on fatal abusive head injuries in infants and young children. Am J Forensic Med Pathol 2001; 22: 112–22.

Chandler S, Coates R, Gearing A, Lury J, Wells G, Bone E. Matrix

metalloproteinases degrade myelin basic protein. Neurosci Lett 1995; 201: 223-6.

Davie CA, Hawkins CP, Barker GJ, Brennan A, Tofts PS, Miller DH, et al. Serial proton magnetic resonance spectroscopy in acute multiple sclerosis lesions. Brain 1994; 117: 49–58.

Davis TM, Suputtamongkol Y, Spencer JL, Ford S, Chienkul N, Schulenburg WE, et al. Measures of capillary permeability in acute falciparum malaria: relation to severity of infection and treatment. Clin Infect Dis 1992; 15: 256–66.

De Groot CJ, Langeveld CH, Jongenelen CA, Montagne L, Van Der Valk P, Dijkstra CD. Establishment of human adult astrocyte cultures derived from postmortem multiple sclerosis and control brain and spinal cord regions: immunophenotypical and functional characterization. J Neurosci Res 1997; 49: 342–54.

Dolinak D, Smith C, Graham DI. Hypoglycaemia is a cause of axonal injury. Neuropathol Appl Neurobiol 2000a; 26: 448–53.

Dolinak D, Smith C, Graham DI. Global hypoxia per se is an unusual cause of axonal injury. Acta Neuropathol (Berl) 2000b; 100: 553–60.

Dugbartey AT, Spellacy FJ, Dugbartey MT. Somatosensory discrimination deficits following pediatric cerebral malaria. Am J Trop Med Hyg 1998; 59: 393–6.

Dumont RJ, Okonkwo DO, Verma S, Hurlbert RJ, Boulos PT, Ellegala DB, Dumont AS. Acute spinal cord injury, part I: pathophysiologic mechanisms. Clin Neuropharmacol 2001; 24: 254–64.

Eidelberg E, Story JL, Walden JG, Meyer BL. Anatomical correlates of return of locomotor function after partial spinal cord lesions in cats. Exp Brain Res 1981; 42: 81–8.

Esiri MM, Wilcock GK, Morris JH. Neuropathological assessment of the lesions of significance in vascular dementia. J Neurol Neurosurg Psychiatry 1997; 63: 749–55.

Esiri MM, Mathews F, Brayne C, Ince PG for CFAS. Pathological correlates of late-onset dementia in a multicentre, community-based population in England and Wales. Lancet 2001; 357: 169–75.

Evangelou N, Esiri MM, Smith S, Palace J, Matthews PM. Quantitative pathological evidence for axonal loss in normal appearing white matter in multiple sclerosis. Ann Neurol 2000a; 47: 391–5.

Evangelou N, Konz D, Esiri MM, Smith S, Palace J, Matthews PM. Regional axonal loss in the corpus callosum correlates with cerebral white matter lesion volume and distribution in multiple sclerosis. Brain 2000b; 123: 1845–9.

Evangelou N, Konz D, Esiri MM, Smith S, Palace J, Matthews PM. Size-selective neuronal changes in the anterior optic pathways suggest a differential susceptibility to injury in multiple sclerosis. Brain 2001; 124: 1813–20.

Fehlings MG, Tator CH. The relationships among the severity of spinal cord injury, residual neurological function, axon counts, and counts of retrogradely labeled neurons after experimental spinal cord injury. Exp Neurol 1995; 132: 220–8.

Ferguson B, Matyszak MK, Esiri MM, Perry VH. Axonal damage in acute multiple sclerosis lesions. Brain 1997; 120: 393–9.

Fink DJ, Datta S, Mata M. Isoform specific reductions in Na+,K(+)-ATPase catalytic (alpha) subunits in the nerve of rats with streptozotocin-induced diabetes. J Neurochem 1994; 63: 1782–6.

Fisher E, Rudick RA, Cutter G, Baier M, Miller D, Weinstock-Guttman B, et al. Relationship between brain atrophy and disability: an 8-year follow-up study of multiple sclerosis patients. Mult Scler 2000; 6: 373–7.

Fitzpatrick MO, Dewar D, Teasdale GM, Graham DI. The neuronal cytoskeleton in acute brain injury. [Review]. Br J Neurosurg 1998; 12: 313–7.

Ganter P, Prince C, Esiri MM. Spinal cord axonal loss in multiple sclerosis: a post-mortem study. Neuropathol Appl Neurobiol 1999; 25: 459–67.

Garthwaite G, Goodwin DA, Batchelor AM, Leeming K, Garthwaite J. Nitric oxide toxicity in CNS white matter: an *in vitro* study using rat optic nerve. Neuroscience 2002; 109: 145–55.

Geddes JF, Vowles GH, Beer TW, Ellison DW. The diagnosis of diffuse axonal injury: implications for forensic practice. Neuropathol Appl Neurobiol 1997; 23: 339–47.

Geddes JF, Whitwell HL, Graham DI. Traumatic or diffuse axonal injury? Author's response. Neuropathol Appl Neurobiol 2000; 26: 491.

Geddes JF, Hackshaw AK, Vowles GH, Nickols CD, Whitwell HL. Neuropathology of inflicted head injury in children. I. Patterns of brain damage. Brain 2001a; 124: 1290–8.

Geddes JF, Vowles GH, Hackshaw AK, Nickols CD, Scott IS, Whitwell HL. Neuropathology of inflicted head injury in children. II. Microscopic brain injury in infants. Brain 2001b; 124: 1299–306.

Genain CP, Cannella B, Hauser SL, Raine CS. Identification of autoantibodies associated with myelin damage in multiple sclerosis. Nat Med 1999; 5: 170–5.

Gennarelli TA, Thibault LE, Adams JH, Graham DI, Thompson CJ, Marcincin RP. Diffuse axonal injury and traumatic coma in the primate. Ann Neurol 1982; 12: 564–74.

Gentleman SM, Nash MJ, Sweeting CJ, Graham DI, Roberts GW. Beta-amyloid precursor protein (beta APP) as a marker for axonal injury after head injury. Neurosci Lett 1993; 160: 139–44.

Gentleman SM, Roberts GW, Gennarelli TA, Maxwell WL, Adams JH, Kerr S, et al. Axonal injury: a universal consequence of fatal closed head injury? Acta Neuropathol (Berl) 1995; 89: 537–43.

Gijbels K, Proost P, Masure S, Carton H, Billiau A, Opdenakker G. Gelatinase B is present in the cerebrospinal fluid during experimental autoimmune encephalomyelitis and cleaves myelin basic protein. J Neurosci Res 1993; 36: 432–40.

Giometto B, An SF, Groves M, Scaravilli T, Geddes JF, Miller R, et al. Accumulation of beta-amyloid precursor protein in HIV encephalitis: relationship with neuropsychological abnormalities. Ann Neurol 1997; 42: 34–40.

Giovannoni G, Miller RF, Heales SJ, Land JM, Harrison MJ, Thompson EJ. Elevated cerebrospinal fluid and serum nitrate and nitrite levels in patients with central nervous system complications of HIV-1 infection: a correlation with blood-brain barrier dysfunction. J Neurol Sci 1998; 156: 53-8.

Giulian D, Vaca K, Corpuz M. Brain glia release factors with opposing actions upon neuronal survival. J Neurosci 1993; 13: 29–37.

Gonzalez RG, Ruiz A, Tracey I, McConnell J. Structural, functional, and molecular neuroimaging in AIDS. In: Gendelman H, Lipton SA, Epstein L, Swindells S, editors. The neurology of AIDS. New York: Chapman and Hall; 1998. p. 333–52.

Grady MS, McLaughlin MR, Christman CW, Valadka AB, Fligner CL, Povlishock JT. The use of antibodies targeted against the neurofilament subunits for the detection of diffuse axonal injury in humans. J Neuropathol Exp Neurol 1993; 52: 143–52.

Graham DI, McLellan D, Adams JH, Doyle D, Kerr A, Murray LS. The neuropathology of the vegetative state and severe disability after non-missile head injury. Acta Neurochir (Wien) Suppl 1983; 32: 65–7.

Gray F, Belec L, Chretien F, Dubreuil-Lemaire ML, Ricolfi F, Wingertsmann L, et al. Acute, relapsing brain oedema with diffuse blood-brain barrier alteration and axonal damage in the acquired immunodeficiency syndrome. Neuropathol Appl Neurobiol 1998; 24: 209–16.

Greenfield JG, King LS. Observations on the histopathology of the cerebral lesions in disseminated sclerosis. Brain 1936; 59: 445–58.

Grimaud J, Barker GJ, Wang L, Lai M, MacManus DG, Webb SL, et al. Correlation of magnetic resonance imaging parameters with clinical disability in multiple sclerosis: a preliminary study. J Neurol 1999; 246: 961–7.

Gultekin SH, Smith TW. Diffuse axonal injury in craniocerebral trauma. A comparative histologic and immunohistochemical study. Arch Pathol Lab Med 1994; 118: 168–71.

Gveric D, Hanemaaijer R, Newcombe J, van Lent NA, Sier CF, Cuzner ML. Plasminogen activators in multiple sclerosis lesions: implications for the inflammatory response and axonal damage. Brain 2001; 124: 1978–88.

Hsu KS, Liang YC, Huang CC. Influence of an extracellular acidosis on excitatory synaptic transmission and long-term potentiation in the CA1 region of rat hippocampal slices. J Neurosci Res 2000; 62: 403–15.

Inzitari D, Romanelli M, Pantoni L. Leukoaraiosis and cognitive impairment. In: O'Brien J, Ames D, Burns A, editor. Dementia. 2nd ed. London: Arnold; 2000. p. 635–53.

Jane JA, Steward O, Gennarelli T. Axonal degeneration induced by experimental noninvasive minor head injury. J Neurosurg 1985; 62: 96–100.

Joutel A, Andreux F, Gaulis S, Domenga V, Cecillon M, Battail N, et al. The ectodomain of the Notch3 receptor accumulates within the cerebrovasculature of CADASIL patients. J Clin Invest 2000; 105: 597–605.

Kapoor R, Davies M, Baker P, Hall SM, Smith KJ. Blockers of sodium and calcium entry protect axons from nitric oxide-mediated degeneration. Ann Neurol 2002; DOI10.102/ana.10443.

528 I. M. Medana and M. M. Esiri

Kaur B, Rutty GN, Timperley WR. The possible role of hypoxia in the formation of axonal bulbs. J Clin Pathol 1999; 52: 203–9.

Kinney HC, Samuels MA. Neuropathology of the persistent vegetative state. [Review]. J Neuropathol Exp Neurol 1994; 53: 548–58.

Kochar DK, Makkar RK. Somatosensory evoked potentials in cerebral malaria. A preliminary study. Electromyogr Clin Neurophysiol 1994; 34: 301–7.

Kornek B, Lassmann H. Axonal pathology in multiple sclerosis. A historical note. Brain Pathol 1999; 9: 651–6.

Kornek B, Storch MK, Weissert R, Wallstroem E, Stefferl A, Olsson T, et al. Multiple sclerosis and chronic autoimmune encephalomyelitis: a comparative quantitative study of axonal injury in active, inactive, and remyelinated lesions. Am J Pathol 2000; 157: 267–76.

Kornek B, Storch MK, Bauer J, Djamshidian A, Weissert R, Wallstroem E, et al. Distribution of a calcium channel subunit in dystrophic axons in multiple sclerosis and experimental autoimmune encephalomyelitis. Brain 2001; 124: 1114–24.

Kuhlmann T, Lingfeld G, Bitsch A, Schuchardt J, Bruck W. Acute axonal damage in multiple sclerosis is most extensive in early disease stages and decreases over time. Brain 2002; 125: 2202–12.

Lake B. Lysosomal and peroxisomal disorders. In: Graham DI, Lantos PL, editors. Greenfield's neuropathology, Vol. 1. London: Arnold; 1997. p. 658–753.

Lawrence DM, Major EO. HIV-1 and the brain: connections between HIV-1-associated dementia, neuropathology and neuroimmunology. [Review]. Microbes Infect 2002; 4: 301–8.

Leppert D, Lindberg RL, Kappos L, Leib SL. Matrix metalloproteinases: multifunctional effectors of inflammation in multiple sclerosis and bacterial meningitis. [Review]. Brain Res Brain Res Rev 2001; 36: 249–57.

Levin MC, Lee SM, Kalume F, Morcos Y, Dohan FC, Hasty KA, et al. Autoimmunity due to molecular mimicry as a cause of neurological disease. Nat Med 2002; 8: 509–13.

Lewallen S, Harding SP, Ajewole J, Schulenburg WE, Molyneux ME, Marsh K, et al. A review of the spectrum of clinical ocular fundus findings in P. falciparum malaria in African children with a proposed classification and grading system. Trans R Soc Trop Med Hyg 1999; 93: 619–22.

Li GL, Farooque M, Holtz A, Olsson Y. Changes of beta-amyloid precursor protein after compression trauma to the spinal cord: an experimental study in the rat using immunohistochemistry. J Neurotrauma 1995; 12: 269–77.

Lindberg RL, De Groot CJ, Montagne L, Freitag P, van der Valk P, Kappos L, et al. The expression profile of matrix metalloproteinases (MMPs) and their inhibitors (TIMPs) in lesions and normal appearing white matter of multiple sclerosis. Brain 2001; 124: 1743–53.

Lindenberg R, Freytag E. Brainstem lesions characteristic of traumatic hyperextension of the head. Arch Pathol 1970; 90: 509–15.

Liu D, Smith CL, Barone FC, Ellison JA, Lysko PG, Li K, et al.

Astrocytic demise precedes delayed neuronal death in focal ischemic rat brain. Brain Res Mol Brain Res 1999; 68: 29-41.

LoPachin RMJ, Lehning EJ. Acrylamide-induced distal axon degeneration: a proposed mechanism of action. [Review]. Neurotoxicology 1994; 15: 247–59.

LoPachin RM Jr, Lehning EJ. Mechanism of calcium entry during axon injury and degeneration. [Review]. Toxicol Appl Pharmacol 1997; 143: 233–44.

Lovas G, Szilagyi N, Majtenyi K, Palkovits M, Komoly S. Axonal changes in chronic demyelinated cervical spinal cord plaques. Brain 2000; 123: 308–17.

Mankowski JL, Queen SE, Tarwater PM, Fox KJ, Perry VH. Accumulation of beta-amyloid precursor protein in axons correlates with CNS expression of SIV gp41. J Neuropathol Exp Neurol 2002; 61: 85–90.

Manning PT, Johnson EM Jr, Wilcox CL, Palmatier MA, Russell JH. MHC-specific cytotoxic T lymphocyte killing of dissociated sympathetic neuronal cultures. Am J Pathol 1987; 128: 395–409.

Masliah E, Achim CL, Ge N, DeTeresa R, Terry RD, Wiley CA. Spectrum of human immunodeficiency virus-associated neocortical damage. Ann Neurol 1992; 32: 321–9.

Matthews PM, De Stefano N, Narayanan S, Francis GS, Wolinsky JS, Antel JP, et al. Putting magnetic resonance spectroscopy studies in context: axonal damage and disability in multiple sclerosis. [Review]. Semin Neurol 1998; 18: 327–36.

Maxwell WL, Povlishock JT, Graham DL. A mechanistic analysis of nondisruptive axonal injury: a review. [Review]. J Neurotrauma 1997; 14: 419–40.

McKenzie KJ, McLellan DR, Gentleman SM, Maxwell WL, Gennarelli TA, Graham DI. Is beta-APP a marker of axonal damage in short-surviving head injury? Acta Neuropathol (Berl) 1996; 92: 608–13.

McLeod D, Marshall J, Kohner EM, Bird AC. The role of axoplasmic transport in the pathogenesis of retinal cotton-wool spots. Br J Ophthalmol 1977; 61: 177–91.

Medana IM, Martinic M, Wekerle H, Neumann H. Transection of major histocompatibility complex class I-induced neurites by cytotoxic T lymphocytes. Am J Pathol 2001; 159: 809–15.

Medana I, Day NP, Hien TT, Mai NT, Bethell D, Phu NH, et al. Axonal injury in cerebral malaria. Am J Pathol 2002; 160: 655–66.

Mews I, Bergmann M, Bunkowski S, Gullotta F, Bruck W. Oligodendrocyte and axon pathology in clinically silent multiple sclerosis lesions. Mult Scler 1998; 4: 55–62.

Nashmi R, Fehlings MG. Changes in axonal physiology and morphology after chronic compressive injury of the rat thoracic spinal cord. Neuroscience 2001a; 104: 235–51.

Nashmi R, Fehlings MG. Mechanisms of axonal dysfunction after spinal cord injury: with an emphasis on the role of voltage-gated potassium channels. Brain Res Brain Res Rev [Review]. 2001b; 38: 165–91.

Neumann H, Medana IM, Bauer J, Lassmann H. Cytotoxic T lymphocytes in autoimmune and degenerative CNS diseases. Trends Neurosci 2002; 25: 313–9.

Neuropathology Group of the Medical Research Council Cognitive Function and Ageing Study(MRC CFAS). Neuropathological assessment of the lesions of significance in vascular dementia. J Neurol Neurosurg Psychiatry 1997; 63: 749–53.

Newman TA, Woolley ST, Hughes PM, Sibson NR, Anthony DC, Perry VH. T-cell- and macrophage-mediated axon damage in the absence of a CNS-specific immune response: involvement of metalloproteinases. Brain 2001; 124: 2203–14.

Newton CR, Hien TT, White N. Cerebral malaria. [Review]. J Neurol Neurosurg Psychiatry 2000; 69: 433–41

Ng HK, Mahaliyana RD, Poon WS. The pathological spectrum of diffuse axonal injury in blunt head trauma: assessment with axon and myelin strains. Clin Neurol Neurosurg 1994; 96: 24–31.

Oehmichen M, Meissner C, Schmidt V, Pedal I, Konig HG, Saternus KS. Axonal injury–a diagnostic tool in forensic neuropathology? A review. Forensic Sci Int 1998; 95: 67–83.

Oppenheimer DR. Microscopic lesions in the brain following head injury. J Neurol Neurosurg Psychiatry 1968; 31: 299–306.

Paolillo A, Pozzilli C, Gasperini C, Giugni E, Mainero C, Giuliani S, et al. Brain atrophy in relapsing–remitting multiple sclerosis: relationship with 'black holes', disease duration and clinical disability. J Neurol Sci 2000; 174: 85–91.

Pelletier J, Suchet L, Witjas T, Habib M, Guttmann CR, Salamon G, et al. A longitudinal study of callosal atrophy and interhemispheric dysfunction in relapsing-remitting multiple sclerosis. Arch Neurol 2001; 58: 105–11.

Pesini P, Kopp J, Wong H, Walsh JH, Grant G, Hokfelt T. An immunohistochemical marker for Wallerian degeneration of fibers in the central and peripheral nervous system. Brain Res 1999; 828: 41–59.

Povlishock JT, Becker DP, Cheng CL, Vaughan GW. Axonal change in minor head injury. J Neuropathol Exp Neurol 1983; 42: 225–42.

Powers JM, Moser HW. Peroxisomal disorders: genotype, phenotype, major neuropathologic lesions, and pathogenesis. [Review]. Brain Pathol 1998; 8: 101–20.

Putnam TJ. Studies in multiple sclerosis. Arch Neurol Psychiat 1936; 35: 1289–308.

Raja F, Sherriff FE, Morris CS, Bridges LR, Esiri MM. Cerebral white matter damage in HIV infection demonstrated using betaamyloid precursor protein immunoreactivity. Acta Neuropathol (Berl) 1997; 93: 184–9.

Rawes JA, Calabrese VP, Khan OA, DeVries GH. Antibodies to the axolemma-enriched fraction in the cerebrospinal fluid and serum of patients with multiple sclerosis and other neurological diseases. Mult Scler 1997; 3: 363–9.

Ray SK, Matzelle DD, Wilford GG, Hogan EL, Banik NL. Cell death in spinal cord injury (SCI) requires de novo protein synthesis. Calpain inhibitor E-64-d provides neuroprotection in SCI lesion and penumbra. Ann NY Acad Sci 2001; 939: 436–49.

Rosenberg LJ, Teng YD, Wrathall JR. Effects of the sodium channel blocker tetrodotoxin on acute white matter pathology after experimental contusive spinal cord injury. J Neurosci 1999; 19: 6122–33.

Saido TC, Sorimachi H, Suzuki K. Calpain: new perspectives in molecular diversity and physiological-pathological involvement. [Review]. FASEB J 1994; 8: 814–22.

Schaumburg HH, Powers JM, Raine CS, Suzuki K, Richardson EP Jr. Adrenoleukodystrophy. A clinical and pathological study of 17 cases. Arch Neurol 1975; 32: 577–91.

Schumacher PA, Eubanks JH, Fehlings MG. Increased calpain Imediated proteolysis, and preferential loss of dephosphorylated NF200, following traumatic spinal cord injury. Neuroscience 1999; 91: 733–44.

Schumacher PA, Siman RG, Fehlings MG. Pretreatment with calpain inhibitor CEP-4143 inhibits calpain I activation and cytoskeletal degradation, improves neurological function, and enhances axonal survival after traumatic spinal cord injury. J Neurochem 2000; 74: 1646–55.

Sherriff FE, Bridges LR, Gentleman SM, Sivaloganathan S, Wilson S. Markers of axonal injury in post mortem human brain. Acta Neuropathol (Berl) 1994a; 88: 433–9.

Sherriff FE, Bridges LR, Sivaloganathan S. Early detection of axonal injury after human head trauma using immunocytochemistry for beta-amyloid precursor protein. Acta Neuropathol (Berl) 1994b; 87: 55–62.

Simon JH, Kinkel RP, Jacobs L, Bub L, Simonian N. A Wallerian degeneration pattern in patients at risk for MS. Neurology 2000; 54: 1155–60.

Simpson DA, Blumbergs PC, Cooter RD, Kilminster M, McLean AJ, Scott G. Pontomedullary tears and other gross brainstem injuries after vehicular accidents. J Trauma 1989; 29: 1519–25.

Skoog I, Berg S, Johansson B, Palmertz B, Andreasson LA. The influence of white matter lesions on neuropsychological functioning in demented and non-demented 85-year-olds. Acta Neurol Scand 1996; 93: 142–8.

Sloviter RS, Dean E, Sollas AL, Goodman JH. Apoptosis and necrosis induced in different hippocampal neuron populations by repetitive perforant path stimulation in the rat. J Comp Neurol 1996; 366: 516–33.

Smith DH, Nonaka M, Miller R, Leoni M, Chen XH, Alsop D, et al. Immediate coma following inertial brain injury dependent on axonal damage in the brainstem. J Neurosurg 2000; 93: 315–22.

Smith KJ, Kapoor R, Hall SM, Davies M. Electrically active axons degenerate when exposed to nitric oxide. Ann Neurol 2001; 49: 470–6.

Springer JE, Azbill RD, Kennedy SE, George J, Geddes JW. Rapid calpain I activation and cytoskeletal protein degradation following traumatic spinal cord injury: attenuation with riluzole pretreatment. J Neurochem 1997; 69: 1592–600.

Strich SSJ. Diffuse degeneration of the cerebral white matter in severe dementia following hand injury. J Neurol Neurosurg Psychiatry 1956; 19: 163–85.

Stys PK. Anoxic and ischemic injury of myelinated axons in CNS white matter: from mechanistic concepts to therapeutics. [Review]. J Cereb Blood Flow Metab 1998; 18: 2–25.

Suenaga T, Ohnishi K, Nishimura M, Nakamura S, Akiguchi I,

Kimura J. Bundles of amyloid precursor protein-immunoreactive axons in human cerebrovascular white matter lesions. Acta Neuropathol (Berl) 1994; 87: 450–5.

Tavalin SJ, Ellis EF, Satin LS. Inhibition of the electrogenic Na pump underlies delayed depolarization of cortical neurons after mechanical injury or glutamate. J Neurophysiol 1997; 77: 632–8.

Teahan O, Slade JY, Perry RH, Ballard CG, Kalaria RN. White matter pathology in Alzheimer's disease and vascular dementia: quantification by amyloid-beta precursor protein immunocytochemistry [abstract]. Neuropathol Appl Neurobiol 2002; 28: 168.

Teng YD, Wrathall JR. Local blockade of sodium channels by tetrodotoxin ameliorates tissue loss and long-term functional deficits resulting from experimental spinal cord injury. J Neurosci 1997; 17: 4359–66.

Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mork S, Bo L. Axonal transection in the lesions of multiple sclerosis. New Engl J Med 1998; 338: 278–85.

Turner G. Cerebral malaria. [Review]. Brain Pathol 1997; 7: 569-82.

Umehara F, Abe M, Koreeda Y, Izumo S, Osame M. Axonal damage revealed by accumulation of beta-amyloid precursor protein in HTLV-I-associated myelopathy. J Neurol Sci 2000; 176: 95–101.

Vinters HV, Ellis WG, Zarow C, Zaias BW, Jagust WJ, Mack WJ, et al. Neuropathologic substrates of ischemic vascular dementia. [Review]. J Neuropathol Exp Neurol 2000; 59: 931–45.

Wang KK, Posmantur R, Nadimpalli R, Nath R, Mohan P, Nixon RA, et al. Caspase-mediated fragmentation of calpain inhibitor protein calpastatin during apoptosis. Arch Biochem Biophys 1998; 356: 187–96.

Weis S, Jellinger K, Wenger E. Morphometry of the corpus callosum in normal aging and Alzheimer's disease. J Neural Transm Suppl 1991; 33: 35–8.

White L, Petrovich H, Hardman J, Nelson J, Davis DG, Ross G-W, et al. Cerebrovascular pathology and impaired cognitive function in autopsied Honolulu-Asia aging study participants. Ann NY Acad Sci 2002; 977: 9–23.

Wolf JA, Stys PK, Lusardi T, Meaney D, Smith DH. Traumatic axonal injury induces calcium influx modulated by tetrodotoxinsensitive sodium channels. J Neurosci 2001; 21: 1923–30.

Wujek JR, Bjartmar C, Richer E, Ransohoff RM, Yu M, Tuohy VK, et al. Axon loss in the spinal cord determines permanent neurological disability in an animal model of multiple sclerosis. J Neuropathol Exp Neurol 2002; 61: 23–32.

Zimmerman RA, Bilaniuk LT. Pediatric head trauma. [Review]. Neuroimaging Clin N Am 1994; 4: 349–66.

Ziv NE, Spira ME. Axotomy induces a transient and localized elevation of the free intracellular calcium concentration to the millimolar range. J Neurophysiol 1995; 74: 2625–37.

Received July 24, 2002. Revised September 20, 2002. Accepted September 21, 2002