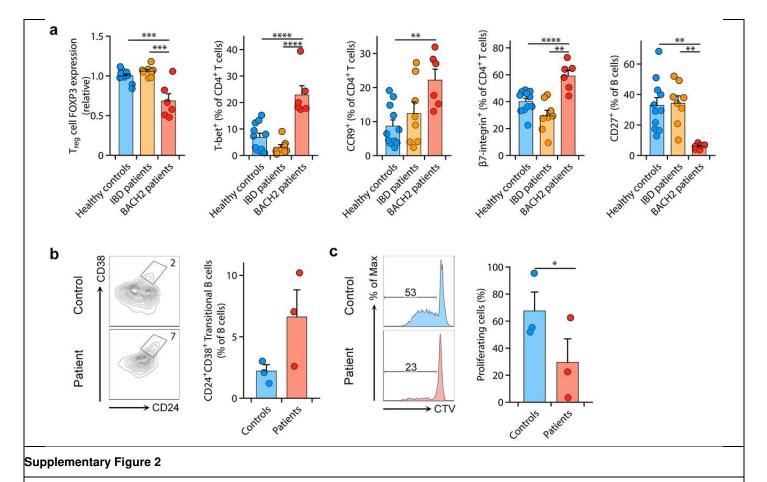


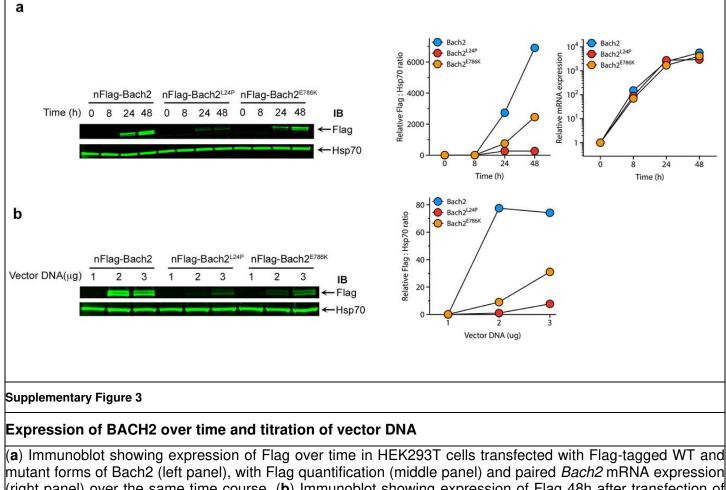
Sanger sequencing chromatograms for the two families.

Shown are the Sanger sequencing chromatograms for the two *BACH2* mutations (A.II.1 – left; B.II.1 and B.III.2 – right) and unaffected family members (Family A on the left, family B on the right). For each individual the two alleles of the sequenced region and base positions are shown above the sequencing chromatograms. Subject A.II.1 had a heterozygous T to C mutation at coding position 71 whereas patients B.II.1 and B.III.2 were heterozygous for G to A base substitutions at position 2362.

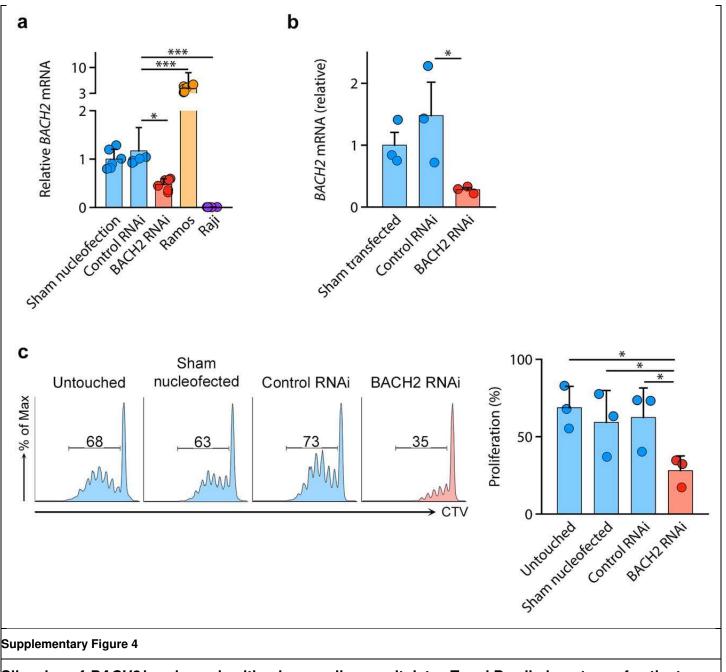


Additional phenotyping of patient cells.

(a) Expression of FOXP3, T-bet, CCR9 and β 7-integrin in CD4⁺ T cells (left four panels, respectively) and frequency of memory B cells (right panel) in peripheral blood of healthy (n=11) and disease controls (n=8), compared to the patients with BACH2 mutations. (b) CD38⁺CD24⁺ transitional B cells²⁸ in patients compared to controls. Shown are representative flow cytometry plots (left) and cumulative data (right) from all patients and matched controls. Patients had elevated proportions of CD38⁺CD24⁺ transitional B cells compared to controls but the observed difference does not reach statistical significance. (c) Proliferation (Cell Trace Violet (CTV) dilution) of primary patient CD4⁺ T cells in response to anti-CD3 and anti-CD28. Shown in b-c are representative FACS plots and cumulative data. Bars show mean ± sem throughout. *p<0.05 **p<0.01 ***p<0.001 ****p<0.001 by ANOVA (a) and t-test (c).

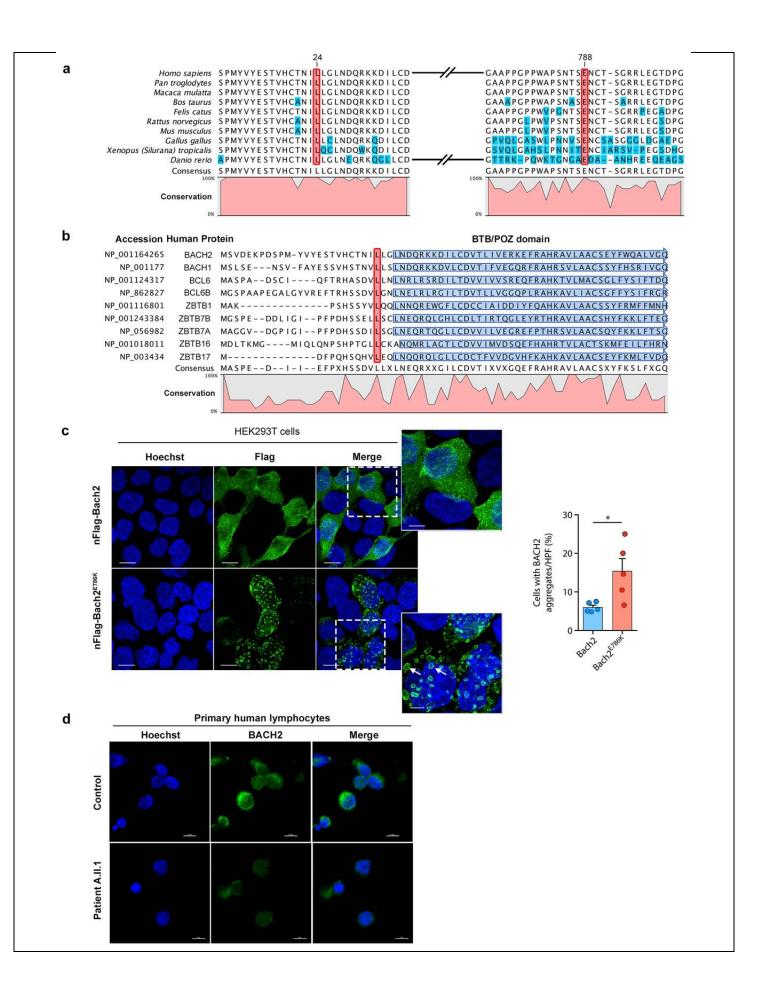


(right panel) over the same time course. (**b**) Immunoblot showing expression of Flag 48h after transfection of HEK293T cells with titrated doses of Flag-tagged WT and mutant *Bach2*-expressing vector (left panel). Flag quantification is shown in the right panel. (**a** and **b**) show representative examples of n=2 experiments.



Silencing of BACH2 in primary healthy donor cells recapitulates T and B cell phenotype of patients

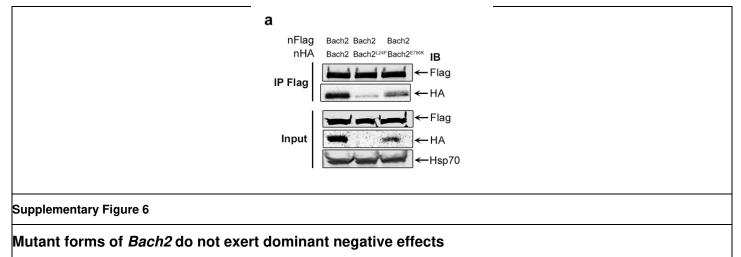
(**a** and **b**) BACH2 mRNA expression in primary healthy donor CD4⁺ T cells (**a**) and naïve B cells (**b**) transfected with control RNAi or RNAi specific for BACH2. Shown are mean \pm sem from n=6 (**a**) and n=3 (**b**) experiments; high (Ramos cell line) and low (Raji cell line) controls for BACH2 expression are additionally shown in **a**. (**c**) Proliferation (Cell Trace Violet (CTV) dilution) of primary healthy control T cells transfected with control RNAi or RNAi specific for BACH2 (n=3 from two independent experiments). Shown are representative flow cytometry examples and cumulative mean \pm sem from multiple experiments. *p<0.05 **p<0.01 ***p<0.001 by ANOVA.



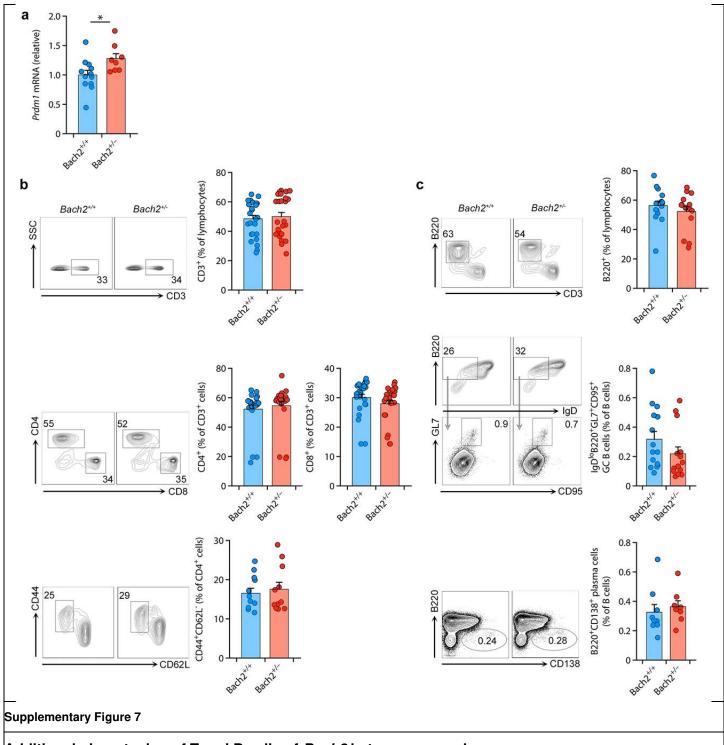
Supplementary Figure 5

BACH2 coding mutations affect conserved amino acids.

(a) Conservation of both mutated residues across species. (b) Conservation of N-terminus mutation across other BTB/POZ family members. Highlighted in red are the residues mutated in patients, light blue represents non-conserved residues and purple the location of the BTB/POZ domains. Note that the BTB/POZ domain is from residues 27-133 in human BACH2 but that the first α -helix extends from residues 18-34 of the protein. (c) Confocal imaging of HEK293T cells transfected with Flag-tagged murine WT or Bach2^{E786K} and stained for Flag (green) and Hoechst (blue). Insets show enlarged images of single cells and localization of cytoplasmic Flag aggregates (white arrows). Images are representative from 5 independent experiments. The bar graph (right) shows quantification (mean ± sem) of the number of cells containing aggregates per high power field (HPF) from n=5 experiments. White scale bar represents 10 μ m in sections and 5 μ m in insets. (d) Confocal images of primary lymphocytes from healthy control and patient A.II.1 stained for BACH2 (green) and Hoechst (blue). Scale bars: 5 μ m. *p<0.05 by t-test.

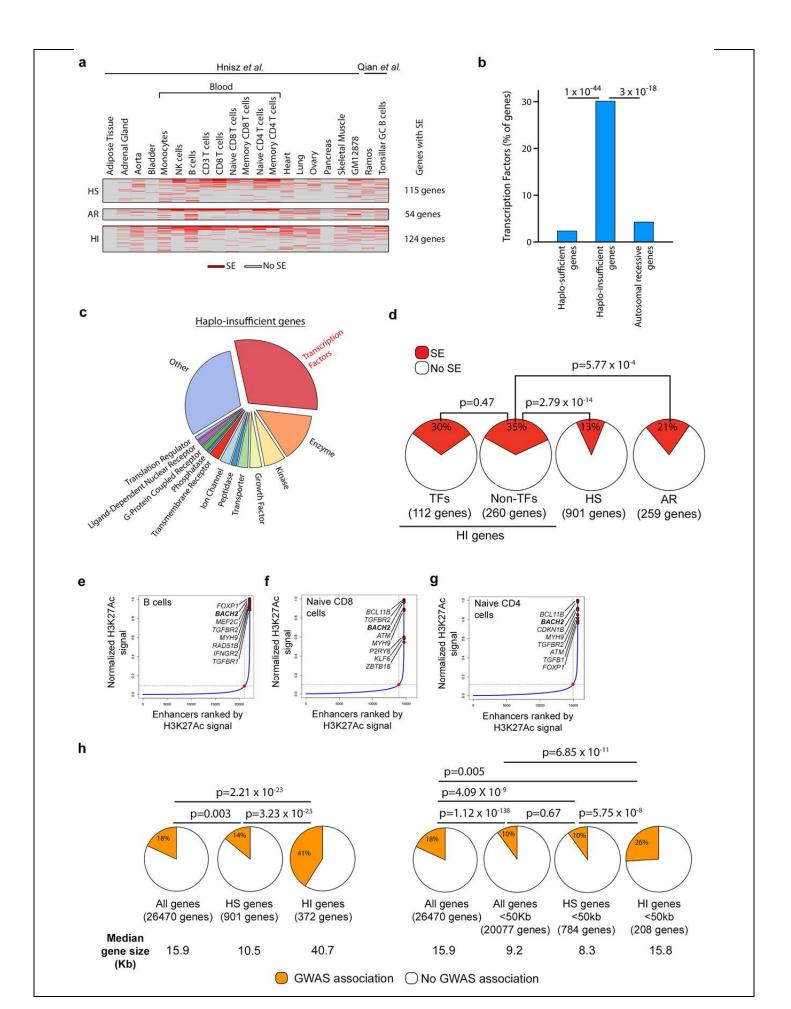


(a) Co-immunoprecipitation of Flag-tagged WT with HA-tagged mutant forms of murine *Bach2* transfected into HEK293T cells at 1:1 ratio. Shown is a representative example from n=3 independent experiments.



Additional phenotyping of T and B cells of *Bach2* heterozygous mice

(a) Expression of *Prdm1* mRNA in B cells of *Bach2*^{+/+} and *Bach2*^{+/-} mice. (b) Proportions of CD3⁺ T cells (top panels), CD4⁺ and CD8⁺ T cells (middle panels) and effector CD4⁺ T cells (CD44⁺CD62L⁻; lower panels) in *Bach2*^{+/-} mice compared to *Bach2*^{+/+} littermates. (c) B cell subsets in unimmunized heterozygous mice, showing total B cells (top panels), germinal center (GC) B cells (middle panels) and plasma cells (bottom panels). Data in **b** and **c** show representative flow cytometry plots with bar graphs depicting mean \pm sem values from a minimum of n=8 mice per group.



Supplementary Figure 8

Superenhancer (SE) and GWS association of haploinsufficient genes

(a) Tissue distribution of genes with SE structure. Shown are only the genes with SE structure in the haplosufficient (HS, top panel), autosomal recessive (AR, middle panel) and haploinsufficient (HI, bottom panel) lists. The presence (red fill) or absence (grey fill) of an SE structure in each tissue is indicated. Each row represents a single gene. Tissue types are shown in columns. Indicated in the "overall" column, is the number of genes with SE architecture in at least one tissue type (see also **supplementary Table 3**). Source data are indicated. (**b-c**) Percentage of genes among HS, HI and AR genes that are transcription factors (**b**) and pie-chart showing function of genes causing HI diseases (**c**). Source data for **b** and **c** are from Qiagen Ingenuity Pathway Analysis. (**d**) Percentage of SE-regulated genes. (**e-g**) Ranked order of H3K27Ac-loaded enhancers in human B cells (**e**), naïve CD8⁺ (**f**) and naïve CD4⁺ (**g**) T cells. Indicated are the relative positions, ranked according to signal intensity (higher = greater signal intensity), of the top 8 genes from the HI gene list in those cells. Source data from²⁹. (**h**) Percentage of genome-wide association study (GWAS)-associated genes in HS and HI genes compared to all genes, with (right) and without (left) correction for size. Shown are median gene sizes for each gene category. All p-values are from exact Fisher tests.

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