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Behavioural, ecological, and evolutionary aspects of diversity in frog colour patterns

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1 **Behavioural, ecological, and evolutionary aspects of diversity**
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6 **in frog colour patterns**
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25
26
27 **ABSTRACT**
28

29 The role of colours and colour patterns in behavioural ecology has been extensively
30 studied in a variety of contexts and taxa, while almost overlooked in many others. For
31
32 decades anurans have been the focus of research on acoustic signalling due to the
33
34 prominence of vocalisations in their communication. Much less attention has been paid
35
36 to the enormous diversity of colours, colour patterns, and other types of putative visual
37
38 signals exhibited by frogs. With the exception of some anecdotal observations and
39
40 studies, the link between colour patterns and the behavioural and evolutionary ecology
41
42 of anurans had not been addressed until approximately two decades ago. Since then,
43
44 there has been ever-increasing interest in studying how colouration is tied to different
45
46 aspects of frog behaviour, ecology and evolution. Here I review the literature on three
47
48 different contexts in which frog colouration has been recently studied: predator–prey
49
50 interactions, intraspecific communication, and habitat use; and I highlight those aspects
51
52 that make frogs an excellent, yet understudied, group to examine the role of colour in the
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1 evolution of anti-predation strategies and animal communication systems. Further, I
2 argue that in addition to natural-history observations, more experiments are needed in
3 order to elucidate the functions of anuran colouration and the selective pressures
4 involved in its diversity. To conclude, I encourage researchers to strengthen current
5 experimental approaches, and suggest future directions that may broaden our current
6 understanding of the adaptive value of anuran colour pattern diversity.

7
8 *Key words:* colouration, predator–prey interactions, visual communication, sexual
9 selection, conflict resolution, space use.

10 11 CONTENTS

12 I. Introduction

13 II. Predator–prey interactions

14 (1) Camouflage

15 (2) Aposematism and mimicry

16 (a) The puzzle of polymorphic warning signals

17 (b) Honesty in warning signals

18 (3) Conspicuous colouration revealed through movement or behaviour

19 III. Intraspecific communication

20 (1) Mate preferences and assortative mating

21 (a) Colours can attract both mates and predators

22 (b) Sexual dichromatism

23 (2) Intrasexual competition and conflict resolution

24 IV. Habitat selection and space use

25 V. Future directions

- 1
2
3 1 VI. Conclusions
4
5 2 VII. Acknowledgements
6
7 3 VIII. References
8
9
10 4

11 **I. INTRODUCTION**

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13
14 6 *“An extensive survey of the organic world thus leads us to the conclusion that colour is*
15
16 7 *by no means so unimportant or inconstant a character as at first sight it appears to be;*
17
18 8 *and the more we examine it the more convinced we shall become that it must serve some*
19
20 9 *purpose in nature, and that besides charming us by its diversity and beauty it must be*
21
22 10 *well worthy of our attentive study, and have many secrets to unfold to us.” – A. R.*
23
24
25 11 Wallace (1877, p. 643)

26
27 12 Signals are supposed to evolve so that the signal-to-noise ratio (the contrast
28
29 13 between the signal and the background noise) is maximised (Bradbury & Vehrencamp,
30
31 14 2011; Endler, 1992, 1993*a*), while signal degradation is minimised (Endler, 1992).
32
33
34 15 Selection also tends to favour signals with a high efficacy not only in terms of their
35
36 16 transmission and detection, but also in their ability to elicit a response in the receiver
37
38 17 that increases the sender’s fitness (Guilford & Dawkins, 1991) while maintaining the
39
40 18 receiver’s fitness unaffected. An exception to this is deceptive signals, which deliver
41
42 19 incorrect information about the signaller and thus benefit the sender at the expense of the
43
44 20 receiver (Mokkonen & Lindstedt, 2015; Wiley, 1983, 1994).

45
46
47 21 Different modalities of communication entail diverse advantages and constraints
48
49 22 on the signals involved (Bradbury & Vehrencamp, 2011). Acoustic signals can be
50
51 23 advantageous over long distances because sound waves can travel for longer through
52
53 24 either air or water without degrading compared to, for example, light. This is particularly
54
55 25 useful for nocturnal animals given the low or non-existent light levels at night (Bradbury
56
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1 & Vehrencamp, 2011). Chemical or olfactory signals are also good in low-light
2 scenarios, but they rely heavily on the characteristics of the medium in which they are
3 transmitted (Bradbury & Vehrencamp, 2011). Visual signals, on the other hand, work
4 well over short to medium distances provided there is a lack of physical obstacles
5 between the sender and the receiver, suitable contrast with the background, and a
6 minimum of environmental light (Endler, 1992, 1993a); they can have different shapes
7 and sizes, and may convey information either on their own (static signals; e.g. diverse
8 and conspicuous colour patterns in bird plumage; Andersson, 1994) or when
9 accompanied or enhanced by repeated movements or displays (dynamic signals; e.g. the
10 extension of a coloured dewlap in combination with head bobbing in *Anolis* lizards;
11 Losos & Chu, 1998).

12 The role of colour patterns as visual signals in animal behaviour and ecology has
13 been studied extensively in a variety of contexts and taxa, but has been neglected in
14 others. For example, for decades anurans have been the focus of studies on acoustic
15 signalling due to the prominence of vocalisations in their communication system. Not
16 only has it been demonstrated that frogs emit calls with different functions (Gerhardt &
17 Huber, 2002), but they have also been shown to have outstanding sensory abilities that
18 allow them to be both physiologically (Capranica & Moffat, 1983) and behaviourally
19 (Amézquita *et al.*, 2011) tuned to the characteristics of their own species-specific
20 acoustic signals. By contrast, the link between colouration and the behavioural and
21 evolutionary ecology of anurans had not been properly addressed until approximately
22 two decades ago. Previous studies focused mostly on the inheritance of colour patterns
23 (Blouin, 1989; Davison, 1963; Fogleman, Corn & Pettus, 1980; Resnick & Jameson,
24 1963); but see Nevo, 1973, for an early study on the selective pressures involved in the
25 maintenance of colour polymorphism in cricket frogs). Recently, more attention has

1
2
3 1 been paid to the enormous diversity of colours, colour patterns, and other types of visual
4
5 2 signals displayed by frogs (e.g. Hödl & Amézquita, 2001), and how those signals are
6
7 3 tied to different aspects of frog ecology and behaviour. Here I aim to review the
8
9 4 diversity of frog colouration in relation to behaviour and ecology. I do so whilst
10
11 5 focusing on those cases in which colour patterns are visual signals on their own,
12
13 6 describing how these signals are currently thought to function in the context of anti-
14
15 7 predation strategies, intraspecific communication and habitat use. Lastly, I suggest
16
17 8 future directions within each context that might fill some of the current gaps in frog
18
19 9 colouration research; and highlight the need to strengthen and broaden the current
20
21 10 experimental approaches in order to widen our understanding of the adaptive value of
22
23 11 diversity in frog colouration, and to identify the candidate selective pressures that might
24
25 12 be shaping such diversity.
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32 **II. PREDATOR–PREY INTERACTIONS**

33
34 15 Colouration may have an enormous effect on animals' fitness because of its adaptive
35
36 16 function as an interspecific signal in the context of predation, among others. While some
37
38 17 animals gain protection from predators by blending with their surroundings
39
40 18 (camouflage; Edmunds, 1974), many species also have conspicuous colour patterns that
41
42 19 warn predators about their unprofitability (Ruxton, Sherratt & Speed, 2004). The latter
43
44 20 strategy is referred to as aposematism (Poulton, 1890).
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49 **(1) Camouflage**

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51
52 23 Camouflage involves a series of strategies that prevent prey from being detected or
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54 24 recognised by predators (Edmunds, 1974; Stevens & Merilaita, 2009). Such strategies
55
56 25 include, for instance, crypsis and masquerade. Common types of crypsis are background
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1
2
3 1 matching, whereby an animal resembles its background colouration and thus avoids
4
5 2 detection (Endler, 1988), and disruptive colouration, which involves markings that make
6
7 3 it difficult for the predator to distinguish the outline or shape of the prey (Cott, 1940;
8
9 4 Thayer, 1909). Masquerade, whereby animals resemble an uninteresting object in their
10
11 5 surroundings (i.e. a rock, a leaf, a stick, etc.), prevents prey recognition instead
12
13 6 (Skelhorn *et al.*, 2010). Both crypsis and masquerade ultimately deprive the predator of
14
15 7 key information about the prey and, therefore, constitute a form of deception (Caro,
16
17 8 2014; Morkkonen & Lindstedt, 2015). Camouflage is a widespread anti-predator strategy
18
19 9 among anurans, which is reflected in the prevalence of earthy colours such as different
20
21 10 shades of green, brown and grey in many species (Wells, 2007). It can be effective on its
22
23 11 own, as in young *Pristimantis zeuctotylus* whose colour patterns resemble those of the
24
25 12 mossy substrate (Fig. 1A), or *Hyla japonica*, which changes its dorsal colour to match
26
27 13 that of the background (Choi & Jang, 2014); but can also be enhanced with particular
28
29 14 behaviours. Individuals of *Craugastor fitzingeri*, for example, become completely
30
31 15 immobile after every jump and can also hide their head in the leaf litter; experiments
32
33 16 with human observers who knew exactly where to search but still could not easily locate
34
35 17 individuals have demonstrated that the detection of these frogs can be extremely difficult
36
37 18 for predators (Cooper, Caldwell & Vitt, 2008).

38
39
40
41
42 19 Colour pattern polymorphisms [the simultaneous occurrence of two or more forms
43
44 20 within a population with the rarest form occurring at frequencies higher than those
45
46 21 expected by mutation pressure (Ford, 1945)] are very common among cryptically
47
48 22 coloured species, and frogs are no exception (Hoffman & Blouin, 2000; Wells, 2007).
49
50 23 For example, the frequencies of colour patterns (grey, green, and red) in two species of
51
52 24 *Acris* (*A. gryllus* and *A. crepitans*) are correlated locally with variations in substrate
53
54 25 colour (Nevo, 1973). Remarkably, these species show seasonal variation in the
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1
2
3 1 frequencies of each colour pattern, suggesting that a morph that is, for example,
4
5 2 favoured in spring when everything is green, will not be favoured in the autumn, when
6
7 3 the background vegetation will be more red. Likewise, in *Dendropsopus* (formerly *Hyla*)
8
9 4 *labialis*, a frog species with an extensive latitudinal and altitudinal distribution, there are
10
11 5 at least five distinct morphs whose frequency seems to be correlated with the
12
13 6 predominant background at each location. For instance, individuals with spotted colour
14
15 7 patterns are more common in populations at high elevations, where the background
16
17 8 vegetation is dominated by mosses (Amézquita, 1999). This suggests that colour
18
19 9 polymorphism in this species may have evolved as a form of crypsis (Amézquita, 1999).
20
21 10 Interestingly, colour patterns also seem to be related to body size, so that green-
22
23 11 dominated morphs are smaller than brown ones (Amézquita, 1999), pointing at a
24
25 12 potential link between colouration and life-history traits that has been surprisingly
26
27 13 understudied in anurans. A long-term study on *Eleutherodactylus coqui* in Puerto Rico
28
29 14 revealed the existence of 21 distinct pattern morphs whose frequencies also differ among
30
31 15 populations and are correlated with the background colouration (Woolbright & Stewart,
32
33 16 2008). Individuals with longitudinal dorsal stripes were most common in grassland
34
35 17 areas, whereas individuals with spots and bars were more common in the forest
36
37 18 (Woolbright & Stewart, 2008). Likewise, females of *Rhinoderma darwinii*, which are
38
39 19 mostly brown, are found on brown substrates, whereas males, which can be either green
40
41 20 or brown, are distributed across brown, green and brown–green backgrounds (Bourke,
42
43 21 Busse & Bakker, 2011).

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45
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48
49 22 In general, the association between body and background colouration suggests that
50
51 23 frogs try to reduce predation risk through crypsis. Cryptic polymorphic species may
52
53 24 have an advantage over monomorphic ones if they are exposed to predators that prey
54
55 25 disproportionately on the most common phenotypes (apostatic selection), because the
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1 fitness of each morph will be inversely related to its frequency in the population (Allen
2 & Greenwood, 1988; Bond, 2007; Bond & Kamil, 1998; Endler, 1991a). Thus, by being
3 polymorphic the *per capita* predation risk of a given species might be reduced, as the
4 abundance of any of its morphs would be low compared to a monomorphic species
5 (Endler, 1991a). Despite the prevalence of colour pattern polymorphisms among
6 cryptically coloured anurans, the mechanisms that allow for their maintenance remain
7 rather obscure. According to Hoffman & Blouin (2000), there is evidence that variation
8 in colour patterns can be inherited and strong indications that predation could indeed be
9 the selective pressure behind such diversity, but with a few exceptions (Morey, 1990;
10 Tordoff, 1980; Wente & Phillips, 2005) most evidence to date remains merely
11 correlational.

12 Although there are many frog examples illustrating the benefits of background
13 matching, evidence describing the function of disruptive colouration or masquerade is
14 very limited and mostly anecdotal. Indeed, despite the fact that it has been suggested to
15 be common among anurans (Wells, 2007), disruptive colouration has not been purposely
16 examined experimentally (Rudh & Qvarnström, 2013). Examples of what might be
17 disruptive colouration are markings such as light dorsal stripes, and blotches or spots
18 that hinder definition of the body shape or break up the limbs (Wells, 2007), as well as
19 lateral lines that cross the eyes while confounding their shape (Amat, Wollenberg &
20 Vences, 2013; Toledo & Haddad, 2009). The best-known illustration of masquerade, on
21 the other hand, is probably found in species living in forests, which can presumably trick
22 predators by looking like dead leaves (Duellman & Trueb, 1994), such as some species
23 of the genus *Rhinella* (Fig. 1B–D).

24 Attempts to demonstrate the adaptive value of any of these forms of camouflage,
25 or to test specifically for the role of predators as the selective pressure behind this

1
2
3 1 diversity in protective colour patterns (i.e. through apostatic selection), could benefit
4
5 2 from the use of well-established protocols employed in other systems. For example,
6
7 3 experiments with both human and avian predators foraging on either artificial prey
8
9
10 4 items, or virtual prey on computer screens have demonstrated that certain colour patterns
11
12 5 increase prey survival, *via* background matching or disruptive colouration, implicating
13
14 6 the role of visual predators as a selection agent on the evolution of diverse protective
15
16 7 colouration (Allen & Clarke, 1968; Allen & Greenwood, 1988; Bond & Kamil, 2006;
17
18 8 Fraser *et al.*, 2007; Karpestam, Merilaita & Forsman, 2013, 2014). An alternative
19
20 9 approach to tackle similar kinds of questions, which has been widely used for research
21
22 10 on aposematism (see Section II.2), is to deploy artificial prey in the field and document
23
24 11 and compare attack rates on different morphs by natural visual predators (Cuthill *et al.*,
25
26 12 2005; Farallo & Forstner, 2012; Valkonen *et al.*, 2011). Studies involving actual frogs
27
28 13 would also be highly informative, as they can account for decisions made by the
29
30 14 individual frogs themselves. For instance, in an elegant experiment investigating
31
32 15 whether individuals of two different morphs of *Hyla regilla* expressed colour pattern-
33
34 16 mediated microhabitat selection, chemical cues of a snake predator were used to
35
36 17 evaluate how predator presence affected the frogs' choice (Wente & Phillips, 2005). The
37
38 18 authors found that, in the presence of predator cues, both green and brown frogs
39
40 19 preferred a substrate that matched their own colour. In the absence of predator cues,
41
42 20 however, only green frogs exhibited a significant preference for a matching background,
43
44 21 which suggests a possible genetically linked association between phenotype (i.e. dorsal
45
46 22 colouration) and behaviour (i.e. preference for a matching background) (Wente &
47
48 23 Phillips, 2005).

54 24 Finally, the combination of these kinds of experiments with methods that allow for
55
56 25 detailed analyses of colour patterns while accounting for predator perception (Endler,
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3 1978, 1984, 1990, 2012; Endler & Mielke, 2005; Kemp *et al.*, 2015; Osorio &
4
5 Srinivasan, 1991; Renoult, Kelber & Schaeffer, 2015; Voroyeb & Osorio, 1998;
6
7 Voroyeb *et al.*, 1998) opens a broad range of possibilities to study the evolution of
8
9
10 4 camouflage strategies in anurans. For instance, a field experiment where dummies with
11
12 5 different colour patterns are deployed on different backgrounds to evaluate how attack
13
14 6 rates differ among groups could be complemented with a survey of the actual frogs in
15
16 7 their natural habitat, recording information on the exact spot where each individual is
17
18 8 found. If not only this information is collected, but also standardised photographs are
19
20 9 taken of both the individual frog and its microhabitat, similarities between the frog
21
22 10 colour patterns and its background could be measured, and the accuracy of camouflage
23
24 11 quantified.
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30 **(2) Aposematism and mimicry**

31
32 14 Aposematism is an anti-predator strategy through which prey warn predators about their
33
34 15 unprofitability (presence of toxins or physical defences such as spines or irritant hairs)
35
36 16 by means of specific colour patterns that act as warning signals (Cott, 1940; Poulton,
37
38 17 1890; Rojas, Valkonen & Nokelainen, 2015*b*; Ruxton *et al.*, 2004). This strategy works
39
40 18 in such a way that predators learn the association between the warning signals and the
41
42 19 unprofitability of the prey, and subsequently avoid them (Endler, 1991*a*; Endler &
43
44 20 Mappes, 2004; Ruxton *et al.*, 2004).

45
46
47 21 Several species of frogs exhibit conspicuous colouration and have a wide array of
48
49 22 skin toxins (Wells, 2007), such as various species of *Mantella* (Fig. 2A,B) and the
50
51 23 ‘Tomato frogs’ (genus *Dyscophus*, Microhylidae; Fig. 2C) from Madagascar (Garraffo
52
53 24 *et al.*, 1993*a*); the Corroboree frogs (*Pseudophryne corroboree*; Fig. 2D) and other
54
55 25 myobatrachids from Australia (Daly *et al.*, 1990); *Brachycephalus ephippium* from
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2
3 1 Brazil (Fig. 2E) (Sebben *et al.*, 1986); and numerous species of Bufonids in the genera
4
5 2 *Melanophryniscus* (Fig. 2F) (Garraffo *et al.*, 1993b; Grant *et al.*, 2012) and *Atelopus*
6
7 3 (Fig. 3A,B; Kim, Kim & Yotsu-Yamashita, 2003) from South and Central America.
8
9
10 4 However, probably the best-known example of aposematic frogs are the dart poison
11
12 5 frogs (Dendrobatidae; Fig. 4) (Stynoski, Shelton & Stynoski, 2014). The varied toxins
13
14 6 found in this Neotropical frog family (Daly *et al.*, 2002, 1994; Daly & Myers, 1967;
15
16 7 Myers & Daly, 1976, 1983) are sequestered from their specialised diet (Saporito *et al.*,
17
18 8 2007a, 2004), which consists mainly of ants, termites, mites and other arthropods found
19
20 9 in the leaf litter (Darst *et al.*, 2005; Toft, 1995). Toxins vary noticeably within the family
21
22 10 in composition, amount, and power, but most are lipophilic alkaloids (Saporito *et al.*,
23
24 11 2012). One dendrobatid species, *Phyllobates terribilis* (Fig. 4D), has the most potent
25
26 12 non-proteolytic (alkaloid) toxin among vertebrates, batrachotoxin (Myers, Daly &
27
28 13 Malkin, 1978). Each of these golden yellow or metallic orange frogs can have up to 1.2
29
30 14 mg of toxin which, if it comes into contact with an open wound, could potentially be
31
32 15 lethal to humans in a dose as low as 200 µg (Myers *et al.*, 1978).

36 16 During the last 15 years, various studies have demonstrated an evolutionary link
37
38 17 between colouration and toxicity in dendrobatid frogs (Darst *et al.*, 2005; Santos,
39
40 18 Coloma & Cannatella, 2003; Summers, 2003; Summers & Clough, 2001), suggesting
41
42 19 that bright colouration has evolved independently at least three times. Diet
43
44 20 specialisation, which is in turn linked with higher levels of toxicity (Darst *et al.*, 2005),
45
46 21 might have itself evolved independently at least two, but possibly three times (Santos *et*
47
48 22 *al.*, 2003; Vences *et al.*, 2003). The combination of bright colours and high toxicity in
49
50 23 these frogs has traditionally been put forward as an example of aposematism (Myers &
51
52 24 Daly, 1983; Pough *et al.*, 2001), even though the first experimental attempts to show
53
54 25 predator aversion of colourful dendrobatids only took place a few years ago (Saporito *et*
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1
2
3 1 *al.*, 2007b). While it has been suggested that some crabs, snakes, beetle larvae and
4
5 2 spiders feed on dendrobatid tadpoles (Gray & Christy, 2000; Stynoski *et al.*, 2014), and
6
7 3 some seemingly toxin-resistant snakes might feed on juveniles (Myers *et al.*, 1978), the
8
9 4 major predators of adult poison frogs are still not known with certainty, presumably due
10
11 5 to the frogs' success at deterring predators. As indicated by anecdotal observations and
12
13 6 experiments in the field, ants, *Paraponera clavata* (Fritz, Rand & Depamphilis, 1981),
14
15 7 and spiders, *Cupiennius coccineus* (Szelistowski, 1985) and *Sericopelma rubronitens*
16
17 8 (Gray, Kaiser & Green, 2010), reject them as prey; but there are also some accounts of
18
19 9 fish (Santos & Cannatella, 2011), snake (Fig. 5A) (Lenger, Berkey & Dugas, 2014;
20
21 10 Ringler, Ursprung & Hödl, 2010), and spider (T. Larsen, personal communication; Fig.
22
23 11 5B) predators. Experiments with frog clay models (see below), suggest that poison frogs
24
25 12 could be subject to attack by birds and crabs. These results are in agreement with at least
26
27 13 one observation of a crab feeding on an individual of *Oophaga histrionica* in the Chocó
28
29 14 region of Colombia (A. Vélez & S. Körting, personal communication; Fig. 5C), and two
30
31 15 observations of adult rufous motmots (*Baryphthengus martii*) consuming one individual
32
33 16 of *D. auratus* with no apparent negative effects (Master, 1999) or feeding individuals of
34
35 17 *O. pumilio* to their offspring (Alvarado, Alvarez & Saporito, 2013) (Fig. 5D). Additional
36
37 18 evidence obtained in studies incorporating taxon-specific vision modelling (Crothers &
38
39 19 Cummings, 2013; Dreher, Cummings & Pröhl, 2015; Maan & Cummings, 2012)
40
41 20 indicate that the colour patterns of poison frogs are indeed likely to be designed to signal
42
43 21 primarily to birds and crabs. However, it is important to note that both these vision
44
45 22 models and the experiments with clay models are unable to assess the importance of
46
47 23 predators such as snakes, which do not rely predominantly on visual cues for prey
48
49 24 detection, but use mostly olfactory, thermal or movement cues instead (Saviola,
50
51 25 McKenzie & Chiszar, 2012).

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3 1 Further support for the role of poison frog colour patterns as an anti-predator
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5 2 strategy has been obtained in recent studies. Various field experiments have shown that
6
7 3 colourful models representing local frogs are usually less frequently attacked than dull
8
9 4 models, models representing novel morphs, or familiar models placed on novel
10
11 5 backgrounds, at least for colours resembling the morphs of *Oophaga pumilio* in Costa
12
13 6 Rica (Hegna *et al.*, 2011; Saporito *et al.*, 2007b; Stuart, Dappen & Losin, 2012) and
14
15 7 *Dendrobates tinctorius* in French Guiana (Noonan & Comeault, 2009; Rojas, Rautiala &
16
17 8 Mappes, 2014b). Interestingly, an experiment carried out in Isla Colón (Panamá) showed
18
19 9 that the local, green morph of *O. pumilio* was attacked at significantly higher frequency
20
21 10 than the foreign, red morph from the mainland (Hegna, Saporito & Donnelly, 2013).
22
23 11 According to the authors, this result suggests that red might be a more efficient predator-
24
25 12 deterrent warning signal regardless of what the local signal is. Finally, movement has
26
27 13 been shown to affect attack rates on clay models of different colours (Paluh, Hantak &
28
29 14 Saporito, 2014). In a study comparing attack rates on stationary brown and red clay
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31 15 models to those on moving models of the same colours, Paluh *et al.* (2014) demonstrated
32
33 16 that not only was bird predation significantly higher on moving brown frog models, but
34
35 17 also significantly lower on moving red frog models. These findings provide evidence of
36
37 18 the significance of prey movement for visual predators, and highlight the importance of
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39 19 incorporating elements that offer more representative measures of predation in the wild
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41 20 into clay model experiments.
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47 21 In addition to aposematism *per se*, studies demonstrate that both Batesian (a
48
49 22 palatable species mimicking the colouration of a defended one; Bates, 1862) and
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51 23 Müllerian (two defended species sharing similar colour patterns; Müller, 1878) mimicry
52
53 24 exist among frog species. In Ecuador, the aposematic frogs *Ameerega (Epipedobates)*
54
55 25 *bilinguis* (Fig. 4B) and *A. parvula* occur parapatrically and serve as models to their non-
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1 defended mimic, *Allobates zaparo*, which adopts the corresponding colouration of its
2 model at each locality (Darst & Cummings, 2006; Darst, Cummings & Cannatella,
3 2006). Likewise, *Allobates femoralis* (Fig. 6A, right), an Amazonian species with a wide
4 geographic distribution and great interpopulational variation in the colouration of both
5 their inguinal and axillary patches, has been thought to be a Batesian mimic of
6 *Ameerega hahneli* (Fig. 6A, left) (Amézquita *et al.*, 2009). *Ranitomeya imitator* from
7 Peru, on the other hand, is by far the best-known example of a Müllerian mimetic
8 radiation in amphibians (Symula, Schulte & Summers, 2001; Twomey, Vestergaard &
9 Summers, 2014; Twomey *et al.*, 2013). This species (Fig. 6H–K) has diverged in both
10 colour pattern and brightness among populations to resemble the colour patterns of its
11 putative defended models: *R. variabilis* (Fig. 6D, highland morph; Fig. 6G, lowland
12 morph), *R. summersi* (Fig. 6E), and *R. fantastica* (Fig. 6F) (Yeager *et al.*, 2012; but see
13 Chouteau *et al.*, 2011). Recent experiments with chickens have demonstrated that
14 models and mimics in this complex might indeed share the costs of predator learning
15 (Stuckert, Venegas & Summers, 2014b). Furthermore, a comparison between the
16 alkaloid profiles of mimics and models has confirmed that all co-mimics possess
17 chemical defences (Stuckert *et al.*, 2014a). Reciprocal learned avoidance by predators
18 and possession of secondary defences by all the species in the mimetic complex are two
19 fundamental assumptions of Müllerian mimicry (Müller, 1878). Among frogs, however,
20 these assumptions have been tested and confirmed only for the *R. imitator* complex thus
21 far. Another case of Müllerian mimicry was recently proposed, without further
22 experimental support, where the leptodactylid *Leptodactylus lineatus* (Fig. 6C)
23 previously thought not to be chemically defended, resembles the colouration of the
24 dendrobatid *Ameerega picta* (Fig. 6B) (Prates *et al.*, 2012). Also, phylogenetic analyses
25 used to study the evolution of colour patterns in Malagassy poison frogs (genus

1
2
3 1 *Mantella*) suggest that the convergence in colouration between *M. madagascariensis*
4
5 2 and *M. baroni* (Fig. 2A), which occur sympatrically, may represent another case of
6
7 3 Müllerian mimicry (Schaefer, Vences & Veith, 2002). This hypothesis, however, has not
8
9
10 4 been tested either. Therefore, given the great variability in toxicity and colour patterns
11
12 5 within anurans, and the co-occurrence of toxic with nontoxic species in wide
13
14 6 geographical ranges, it seems likely that even more examples of both types of mimicry
15
16 7 are waiting to be uncovered.

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21 9 (a) *The puzzle of polymorphic warning signals*

22
23 10 Warning signal variability may reduce the ability of predators to learn and retain
24
25 11 the association between colour patterns and distastefulness (Exnerová *et al.*, 2006;
26
27 12 Greenwood, Wood & Batchelor, 1981; Mallet & Joron, 1999). As a result of stabilising
28
29 13 selection, it is expected that aposematic prey will have little to no variation in their
30
31 14 warning signals (Darst & Cummings, 2006; Endler, 1988; Endler & Mappes, 2004;
32
33 15 Joron & Mallet, 1998). Hence, polymorphisms should be selected against in aposematic
34
35 16 species (Endler & Mappes, 2004). However, there are several cases of aposematic
36
37 17 species exhibiting colour polymorphisms in nature, for example ladybirds (O'Donald &
38
39 18 Majerus, 1984; Ueno, Sato & Tsuchida, 1998), beetles (Borer *et al.*, 2010) and moths
40
41 19 (Hegna, Galarza & Mappes, 2015). In anurans, despite many aposematic species
42
43 20 showing geographic variation in colouration (Amézquita *et al.*, 2013; Brusa *et al.*, 2013;
44
45 21 Chouteau *et al.*, 2011; Hoogmoed & Avila-Pires, 2012; Lötters *et al.*, 1997; Myers &
46
47 22 Daly, 1983; Noonan & Gaucher, 2006; Wollenberg *et al.*, 2008), within-population
48
49 23 variability in warning signals has been reported only for *Oophaga pumilio* (Fig. 7A–D)
50
51 24 (Maan & Cummings, 2012), *O. histrionica* (Amézquita *et al.*, 2013), *Dendrobates*
52
53 25 *tinctorius* (Rojas & Endler, 2013) (Fig. 7E–H) and *Melanophryniscus rubriventris* (Fig.

1
2
3 1 3C) (Bonansea & Vaira, 2012). Nevertheless, given that levels of variation have not
4
5 2 been investigated in many species, it is highly likely that more examples are yet to be
6
7 3 discovered.

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9
10 4 Although there are several instances of polymorphic aposematic populations, the
11
12 5 mechanisms allowing warning signal polymorphisms to persist are not yet fully
13
14 6 understood. Recent approaches suggest a number of possible explanations: (1) an
15
16 7 interaction between natural and sexual selection (Crothers & Cummings, 2013;
17
18 8 Cummings & Crothers, 2013; Nokelainen *et al.*, 2012); for example, in the aposematic
19
20 9 wood tiger moth (*Parasemia plantaginis*), where males can be either yellow or white,
21
22 10 yellow individuals have been found to be better protected from predators, whereas under
23
24 11 certain circumstances whites have a higher mating success (Gordon *et al.*, 2015;
25
26 12 Nokelainen *et al.*, 2012). In *O. pumilio* from Solarte Island (in the Bocas del Toro
27
28 13 archipelago), the stabilising selection exerted by predators to keep colour patterns
29
30 14 uniform is most likely counteracted by directional sexual selection favouring brighter
31
32 15 males, which are preferred by females (Maan & Cummings, 2009). (2) Spatio-temporal
33
34 16 variation in selection (Endler & Rojas, 2009; Galarza *et al.*, 2014; Nokelainen *et al.*,
35
36 17 2014), for instance in the composition of local predator communities, which may
37
38 18 generate a geographic mosaic of selection throughout the distribution range of a species.
39
40 19 (3) A link between colour patterns and a fitness-related trait (or group of traits), either
41
42 20 behavioural or physiological. Examples of this are differential activity patterns and
43
44 21 investment in immune defences according to colour, as in wood tiger moths, or the
45
46 22 association between movement type and colour patterns in the dyeing poison frog
47
48 23 (*Dendrobates tinctorius*) (Nokelainen, Lindstedt & Mappes, 2013; Rojas,
49
50 24 Devillechabrolle & Endler, 2014a; Rojas, Gordon & Mappes, 2015a). (4) Relaxed
51
52 25 selection towards warning signals or predator generalisation, which involves predator
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3 1 avoidance of one signal and the expansion of this aversion towards other signals similar
4
5 2 enough to the one learnt (Amézquita *et al.*, 2013; Darst *et al.*, 2006; Richards-Zawacki,
6
7 3 Yeager & Bart, 2013). (5) Non-adaptive forces such as hybridisation among geographic
8
9
10 4 variants or drift (Gray & McKinnon, 2007; Medina *et al.*, 2013; Thompson, 1984).
11
12 5 Whatever the mechanism, for polymorphisms to be maintained there should be no
13
14 6 differences in fitness among the morphs.
15

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19 8 *(b) Honesty in warning signals*

20
21 9 The ‘handicap principle’ (Zahavi, 1975) suggests that selection should favour
22
23 10 signals that provide reliable information about an individual’s quality. These signals
24
25 11 must represent costs for the signaller, so that they are unaffordable for individuals whose
26
27 12 quality is lower. Although suggested originally for sexual signals, honest signals have
28
29 13 the potential to evolve in predator–prey relationships as well (Guilford & Dawkins,
30
31 14 1993).
32
33

34 15 For aposematic individuals, it is likely that warning signals are reliable indicators
35
36 16 of prey’s unprofitability (e.g. toxicity), as only well-defended prey can afford the costs
37
38 17 of being easily detectable by predators (Sherratt, 2002). However, detectability is not
39
40 18 necessarily the only cost aposematic species may incur. Theoretical studies have shown
41
42 19 that simultaneous investment in pigments and chemical defences may trade off within
43
44 20 the same individual (Blount *et al.*, 2009). Thus, warning signals are likely to be honest
45
46 21 signals of the quality (or quantity) of chemical defences, as only well-defended
47
48 22 individuals could profitably tolerate the costs of strong warning signals.
49
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51 23 A positive correlation between warning signals and toxicity has been found
52
53 24 across species of dart poison frogs in a study where phylogenetic constraints were taken
54
55 25 into account, and where both colouration and toxicity were considered ‘either/or’ traits
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1 (Summers & Clough, 2001). However, quantitative approaches to whether poison frogs'
2 warning signals are honest or not have yielded mixed evidence. For instance, among
3 different populations of *Oophaga pumilio* colouration has been suggested to be an
4 honest indicator of toxicity, such that the brightest populations are also the most toxic
5 (Maan & Cummings, 2012). However, a negative correlation between these two traits
6 has also been found, both between and within species. The conspicuous species
7 *Ameerega (Epipedobates) bilinguis* has a lower toxicity level than its less-conspicuous
8 counterpart, *Ameerega (Epipedobates) parvulus* (Darst *et al.*, 2006). Notably,
9 experiments with chicks revealed that both increased conspicuousness and increased
10 toxicity are equally effective when it comes to predator deterrence (Darst *et al.*, 2006).
11 Likewise, in different populations of *Oophaga granulifera* conspicuous colouration and
12 toxicity are inversely related, such that the most conspicuous populations show the
13 lowest levels of toxicity. In fact, these populations, where individuals are bright red, lack
14 four of the alkaloids present in populations with the less conspicuous yellow or green
15 colouration (Wang, 2011). According to Blount *et al.* (2009), this scenario is plausible
16 when resource availability is high, as suggested for the seven-spot ladybird, *Coccinella*
17 *septempunctata* (Blount *et al.*, 2012). However, a recent theoretical approach found that,
18 at equilibrium, a negative correlation between conspicuousness and defence strength
19 does not hold (Holen & Svanungsen, 2012). This is because, if a trade-off between the
20 resources invested in warning signals and chemical defences is assumed, the benefit of
21 more conspicuous signals is a low risk of prey being attacked after detection, whereas
22 the cost is a reduced defence level, which will bring decreased probabilities of survival
23 upon attack (Holen & Svanungsen, 2012). Toxins such as those found in dendrobatid,
24 mantellid or bufonid frogs are believed to be mainly sequestered from their diet (Clark *et*
25 *al.*, 2005; Daly *et al.*, 1997; Hantak *et al.*, 2013), which consists primarily of leaf-litter

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2
3 1 arthropods. This means that, as suggested by previous studies (Maan & Cummings,
4
5 2 2012; Wang, 2011), variation in toxicity levels among populations of the same species
6
7 3 might be due to variation in prey availability. However, this does not necessarily explain
8
9 4 why some species (or individuals within the same population) are more toxic, or more
10
11 5 conspicuous, than others.

12
13
14 6 Finding correlations, either positive or negative, between warning signals and the
15
16 7 quality or quantity of chemical defences in the species mentioned above has been very
17
18 8 informative on the dynamics of predator–prey relationships in anurans. However,
19
20 9 empirical studies tend to miss one (or more) of the key elements about honesty, such as
21
22 10 evidence that predators really pay attention to variation in toxicity and/or see (and care
23
24 11 about) the difference among signals. Often only a correlation between toxicity and
25
26 12 colouration is found, and the rest is assumed. Therefore, as Summers *et al.* (2015) state
27
28 13 in a recent review, one key question is how to differentiate between quantitative honesty
29
30 14 and other ways in which conspicuousness and toxicity may be correlated without
31
32 15 involving honest signalling.

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34
35
36 16 The next steps thus should also include observations and experiments leading to a
37
38 17 better understanding of the costs associated both with toxin sequestration and storage,
39
40 18 and with the production of warning signals (i.e. pigments). For example, in species with
41
42 19 variable colour patterns, it could be tested whether in cohorts of individuals raised from
43
44 20 larvae in the same conditions, juvenile colour patterns are correlated with life-history
45
46 21 traits such as size and time to metamorphosis. Additionally, juveniles could be raised in
47
48 22 semi-natural enclosures differing in diet restrictions to see whether resource availability
49
50 23 affects colouration, or toxicity, or both. A primer on this is a recent study on
51
52 24 *Dendrobates auratus*, where the effects of rearing tadpoles on either a high-food or low-
53
54 25 food diet were tested on body size and luminance at metamorphosis (Flores *et al.*, 2013).
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1 The authors found that in metamorphs raised on a high-food diet body size and
2 luminance were negatively correlated, whereas this correlation was positive in froglets
3 reared on a low-food diet. According to Flores *et al.* (2013), these findings suggest
4 either a trade-off in resource allocation, or developmental plasticity aimed at minimising
5 predation risk at the most vulnerable (early) stages. Life-history trade-offs in relation to
6 frog colouration may therefore be more common than usually thought, especially if the
7 resource-allocation hypothesis holds.

8

9 **(3) Conspicuous colouration revealed through movement or behaviour**

10 Besides the very well-studied active visual displays that some frog species perform
11 with their fore and hind limbs (i.e. arm waving, foot flagging, etc.), some of which
12 expose coloured patches (reviewed in Hödl & Amézquita, 2001), several anuran species
13 reveal coloured parts of their body through movement or specific positions (Hödl &
14 Amézquita, 2001). Some species that use camouflage as their primary defence against
15 predators have a secondary ‘hidden’ conspicuous colouration which is revealed under
16 threat (i.e. deimatic displays; Edmunds, 1974; Umbers, Lehtonen & Mappes, 2015).
17 Species in the genera *Phyllomedusa* and *Agalychnis*, for example, have this kind of
18 colouration on their flanks (Fig. 8A). Their markings display combinations of yellow,
19 orange, or purple with black stripes, and might serve as a warning signal to their
20 irritating skin secretions (Wells, 2007, and references therein) and strong smell (B.
21 Rojas, personal observations) when handled. The anti-predator function of these colour
22 patterns warrants experimental testing. Species of the genera *Physalaemus*, *Pleurodema*
23 and *Edalorhina* exhibit what resemble eyespots in the lower part of their dorsum (Fig.
24 8B). These markings are better seen when the frogs hide their head in the substrate,
25 leaving visible only the posterior part of the body (Lenzi-Mattos *et al.*, 2005; Martins,

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2
3 1 1989). Fire toads of the genus *Bombina* expose their conspicuous ventral colouration by
4
5 2 raising the legs and arching their body ('unken reflex') when threatened (Bajger, 1980).
6
7 3 This behaviour is also common among South American frogs of the genus
8
9 4 *Melanophryniscus* (Fig. 8C–E) (Caorsi *et al.*, 2014; Grant *et al.*, 2012; Santos & Grant,
10
11 5 2011). Finally, the conspicuous and very distinctive ventral colouration exhibited by
12
13 6 some species of *Atelopus* (Fig. 3B) has also the potential to deter predators, especially
14
15 7 given the highly toxic tetrodotoxins present in the genus (Fuhrman, Fuhrman & Mosher,
16
17 8 1969; Kim *et al.*, 2003), but this hypothesis also requires in-depth examination.
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19 9

10 **III. INTRASPECIFIC COMMUNICATION**

11 **(1) Mate preferences and assortative mating**

12 Bright colours can be important signals in mate choice (Andersson, 1994), particularly,
13
14 13 but not exclusively, for diurnal species that use a wide range of visual signals during
15
16 14 courtship (e.g. poison frogs: Hödl & Amézquita, 2001; Zimmermann & Zimmermann,
17
18 15 1988). For example, females of different taxa prefer to mate with more colourful or
19
20 16 brighter individuals (Bajer *et al.*, 2010; Gomez *et al.*, 2009; Maan *et al.*, 2004). In some
21
22 17 cases, not only the colour but also a variety of different criteria such as the number, size
23
24 18 or shape of patches or spots are important in mate choice (Petrie, Halliday & Sanders,
25
26 19 1991; Pincemy, Dobson & Jouventin, 2009). Although in many cases the explanations
27
28 20 for mate preferences remain obscure, it has often been found that they are associated
29
30 21 with the indirect benefits of 'good genes' (Cutrera, Fanjul & Zenuto, 2012; Hamilton &
31
32 22 Zuk, 1982; Milinski & Bakker, 1990; Stange & Ronacher, 2012). Female preferences
33
34 23 can exert directional selection on male traits that provide reliable information about their
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36 24 quality, because high-quality mates presumably lead to higher quality offspring, *via*
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38 25 increased fitness [through, for example, parasite resistance (Barber *et al.*, 2001; Demuth,
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3 1 Naidu & Mydlarz, 2012; Horak *et al.*, 2001; Milinski & Bakker, 1990) or protection
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5 2 from predators (Lancaster, Hipsley & Sinervo, 2009; Sheldon *et al.*, 2003)]. In spite of
6
7 3 this, there are hardly any studies either on the influence of colouration on mate choice or
8
9 4 on the fitness consequences of such choices in anuran species.

10
11 5 Some nocturnal frog species are known to pay attention to colours when choosing
12
13 6 a mate, given their ability to see some colour cues under low light (Gomez *et al.*, 2010).
14
15 7 Using video playbacks of frogs emitting identical calls but differing in the colour and
16
17 8 brightness of their vocal sac, Gómez *et al.* (2009) demonstrated that female *Hyla*
18
19 9 *arborea* are more attracted to males with a more colourful sac and prominent lateral
20
21 10 stripe, which may together boost male conspicuousness. Furthermore, given that
22
23 11 carotenoid production is costly, the authors hypothesised that carotenoid-based colours
24
25 12 in the vocal sac might convey information about male quality (Gomez *et al.*, 2009).
26
27 13 Similarly, female *Scaphiopus couchii* prefer brighter males; in this species, both colour
28
29 14 and patterning of males are reliable indicators of body size, which may in turn be an
30
31 15 indicator of male quality (Vasquez & Pfennig, 2007). Finally, when presented with two
32
33 16 models differing in colour pattern but emitting identical calls, female *Hyla squirella*
34
35 17 seem to be more attracted to males with large lateral stripes (Taylor *et al.*, 2011).

36
37 18 Among diurnal frogs, the only species in which visual mate choice has been
38
39 19 studied extensively to date is *Oophaga pumilio*. Females in this species have been
40
41 20 shown to prefer males with higher dorsal brightness (Maan & Cummings, 2009).
42
43 21 Although ventral colouration could be expected to be more important than dorsal
44
45 22 patterns for intraspecific communication given that most interactions occur while frogs
46
47 23 are facing each other (Siddiqi *et al.*, 2004), this and other studies support the idea that
48
49 24 male dorsal colouration is the most relevant trait on which females base their
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51 25 preferences (Maan & Cummings, 2008; Summers *et al.*, 1999). *Oophaga pumilio* has
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2
3 1 also been the focus of several studies because of its extensive interpopulation variation
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5 2 in colour patterns, especially in the Bocas del Toro Archipelago in Panama (Fig. 7A–D).
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7 3 Here, at least 15 different morphs have been identified among different islands (Myers
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9 4 & Daly, 1983; Siddiqi *et al.*, 2004), but, with the exception of one population at Isla
10
11 5 Bastimentos, populations of this species are known to be monomorphic. Understanding
12
13 6 the origin of such geographic diversity in colour pattern has proved challenging,
14
15 7 although there are strong indications that, besides natural selection, sexual selection may
16
17 8 play a role in its maintenance (Crothers & Cummings, 2013; Cummings & Crothers,
18
19 9 2013; Maan & Cummings, 2009; Summers *et al.*, 1997).

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22
23 10 A possible mechanism by which sexual selection can promote the maintenance of
24
25 11 geographic variants is through assortative mating, the non-random pairing of two
26
27 12 individuals on the basis of a shared trait (Kondrashov & Shpak, 1998); be it
28
29 13 morphological, ecological, or behavioural. Assortative mating has been broadly
30
31 14 documented in a wide variety of organisms, and can be either positive (matings between
32
33 15 individuals with the same phenotype) or negative (matings between different
34
35 16 phenotypes). Colours seem to play a prominent role in the occurrence of assortative
36
37 17 mating in taxa such as fish (Maan & Seehausen, 2011; Seehausen & van Alphen, 1998;
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39 18 Terai & Okada, 2011), birds (Andersson, Ornborg & Andersson, 1998), butterflies
40
41 19 (Melo *et al.*, 2009), and frogs. In a set of choice experiments with *O. pumilio* in the
42
43 20 laboratory, females presented with males from two different populations (i.e. two
44
45 21 different colour morphs) were more likely to spend time with the male from their own
46
47 22 population (Reynolds & Fitzpatrick, 2007; Summers *et al.*, 1999). Another study, with
48
49 23 four populations, found that female mating preferences are influenced by male colours
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51 24 in such a way that native males were favoured, but that this preference relies heavily on
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53 25 the phenotype of the second male that is presented in the experiment (Maan &
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3 1 Cummings, 2008). Recent work inferring patterns of colour-based mating in *O. pumilio*
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5 2 from wild pedigrees indicates that the assortative mating found in the laboratory does
6
7 3 not necessarily translate completely to the field, where choices are actually made
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9
10 4 (Richards-Zawacki, Wang & Summers, 2012). It would be interesting to determine
11
12 5 whether these preference patterns apply to other dart poison frog species with
13
14 6 comparable geographic variation in colour patterns (Hoogmoed & Avila-Pires, 2012;
15
16 7 Lötters *et al.*, 2007; Silverstone, 1975). Likewise, it would be intriguing to determine
17
18 8 whether colours play a role in mate choice in completely unrelated taxa such as species
19
20 9 of the genus *Mantella*. This group of Malagassy frogs seems a striking example of
21
22 10 evolutionary convergence with dendrobatids not only because of their conspicuous
23
24 11 colouration (Schaefer *et al.*, 2002) and skin alkaloids (Garraffo *et al.*, 1993a), but also
25
26 12 because of their complex social and reproductive behaviour (Heying, 2001). Other
27
28 13 suitable candidates for this kind of investigation are species of the genus
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30 14 *Melanophryniscus*, such as *M. rubriventris* (Bonansea & Vaira, 2012).
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36 16 (a) *Colours can attract both mates and predators*

37
38 17 One problem that animals face when they have conspicuous colour patterns is that they
39
40 18 may be providing information about their presence and location to both wanted and
41
42 19 unwanted receivers. While aposematic species have evolved distastefulness as a way to
43
44 20 deter predators, unprotected organisms must find a way to efficiently attract conspecifics
45
46 21 without attracting too much attention from predators or sneakers (Endler, 1978, 1980).
47
48 22 This problem might be overcome by using 'private communication channels' so that
49
50 23 conspicuousness is higher towards conspecifics than it is towards predators (Endler,
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52 24 1978, 1980, 1991b; Schaefer, 2010), which would imply that the visual sensitivity of
53
54 25 conspecifics is better 'tuned' to the characteristics of their visual signals than is the
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1 predators' sensitivity (Endler, 1991*b*). Alternatively, there could be spatio-temporal
2 constraints on the use of conspicuous signals, for example in species with explosive
3 breeding (i.e. breeding activity limited to only one or a few days, often following the
4 first rains of the season; Wells, 2007, and references therein).

5 There are various examples of spatio-temporal niche partitioning in frogs, most of
6 which relate to their acoustic signals in complex assemblages (Amézquita *et al.*, 2011;
7 Check, Bogart & Lougheed, 2003; Duellman & Pyles, 1983; Lüddecke *et al.*, 2000).
8 Recent studies, however, have shown that such partitioning could also be related to
9 colouration, and occur at an intraspecific level. The toad *Bufo luetkeni* breeds at the very
10 beginning of the rainy season in Costa Rica. Males of this species, which are otherwise
11 dully coloured, become lemon yellow when they are ready to breed, and combine this
12 conspicuous signal with distinct calls to create a multimodal display that attracts
13 females. Females, as well as recently mated males, are dull brown, which suggests that
14 this temporal change in colouration may be influenced by sexual selection, and restricted
15 in time presumably due to increased risk of predation when conspicuous (Doucet &
16 Mennill, 2010). In *Rana temporaria*, males in breeding aggregations display brighter
17 throats (Fig. 9A), whereas male moor frogs (*Rana arvalis*) turn blue (Fig. 9B). Such
18 colour changes have been suggested to function as a sex-recognition cue to prevent
19 mismating attempts during the intense scramble competition common among explosive
20 breeders (Sztatecsny *et al.*, 2012, 2010).

21

22 (b) Sexual dichromatism

23 The changes in colouration described above occur in just one sex, and are therefore
24 considered examples of sexual dichromatism. Also, because they occur only
25 temporarily, during breeding, they are considered to be 'dynamic'. This type of colour

1 change has been studied most thoroughly in the context of frog behaviour or ecology.
2
3 However, due to its visibility for only a short period of time – ranging from a few hours
4
5 to a few weeks – its occurrence is thought to be underestimated, i.e. it probably is more
6
7 common than we are aware of in anuran taxa (Bell & Zamudio, 2012). There are also
8
9 cases of ontogenetic colour change, where males differ from females in colouration from
10
11 early life stages (Bell & Zamudio, 2012). For example, female African reed frogs of the
12
13 genus *Hyperolius* change their colour patterns when attaining maturity, whereas males
14
15 keep their juvenile colours (Veith *et al.*, 2009).
16
17 In one of the populations of the Bocas del Toro archipelago, the aposematic frog *O.*
18
19 *pumilio* is known to be sexually dimorphic in terms of brightness, a non-chromatic
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21 component of colour patterns, with adult males as the brighter sex (Maan & Cummings,
22
23 2009). This is most likely due to directional sexual selection (Maan & Cummings,
24
25 2009). Male *Dendrobates tinctorius* from a highly polymorphic population in French
26
27 Guiana, on the other hand, are overall yellower than females. This has been suggested to
28
29 be the product of a synergy between sexual selection in the form of parental care, and
30
31 the potential increased predation risk for males during the prolonged exposure involved
32
33 in tadpole transport (Rojas & Endler, 2013). Despite these examples, sexual
34
35 dichromatism is still surprisingly understudied considering how widespread it seems to
36
37 be among anurans (Bell & Zamudio, 2012).
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48 **(2) Intrasexual competition and conflict resolution**

49 Intraspecific aggression has been widely documented and can occur in different contexts
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51 (Lorenz, 1966; Riechert, 1998). Individuals can behave aggressively against
52
53 conspecifics when they threaten the survival of their offspring (Sommer, 1987), when
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55 they threaten access to potential mates (Andersson, 1994), or when they are defending a
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1 space that holds resources essential for their survival (i.e. food or shelter) or
2 reproduction. The latter case is normally referred to as territoriality (Kaufmann, 1983;
3 Maher & Lott, 1995). However, aggression *via* escalated conflict is costly in terms of
4 energy and confers the risk of being injured (Maynard Smith & Harper, 2005).
5 Therefore, under specific circumstances, the resolution of conflicts *via* the exchange of
6 signals is expected to be favoured over physical aggression as an evolutionarily stable
7 strategy (Maynard Smith, 1982; Maynard Smith & Harper, 2005). In fact, the mitigating
8 effect of signalling during agonistic interactions has empirical support (Logue *et al.*,
9 2010).

10 During a conflict, contestants should assess the probability of defeat by comparing
11 their own fighting abilities with those of their opponent (Riechert, 1998). This means
12 that both individuals are at the same time senders and receivers, as they exchange
13 information about status, relative fighting ability, and relative resource value. Such
14 information could be given and obtained on the basis of behavioural traits such as the
15 rate at which displays are repeated (Johnstone, 1997). For example male jacky dragons
16 (*Amphibolurus muricatus*) respond to video playbacks of conspecifics displaying at
17 varying time intervals with aggressive push-ups (Ord & Evans, 2003). Alternatively,
18 information can be contained in intrinsic characteristics of an individual such as colours,
19 size, condition, or the size or complexity of weaponry such as horns or antlers (Bradbury
20 & Vehrencamp, 2011).

21 Colouration is a reliable signal of status and an important determinant of conflict
22 resolution for several species of insects, fish, lizards, and birds (López, Martin &
23 Cuadrado, 2004; Morimoto, Yamaguchi & Ueda, 2005; Pryke & Andersson, 2003a;
24 Stuart-Fox & Johnston, 2005). Not only the colour, but also the size and shape of visual
25 signals may provide information about fighting abilities during agonistic encounters.

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3 1 There is much evidence that more colourful males, and sometimes females, tend to
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5 2 acquire and retain higher status within a group of individuals, whereas dull-coloured
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7 3 individuals are often subordinate. For example, redder male firemouth cichlids
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9 4 (*Cichlasoma meeki*) are more likely to win contests than duller ones (Evans & Norris,
10
11 5 1996); and sand lizards (*Lacerta agilis*) with more colourful badges are more likely to
12
13 6 initiate and win fights (Olsson, 1994); Similar trends have been found in African red-
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15 7 shouldered widowbirds (*Euplectes axillaris*), where individuals with redder patches are
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17 8 more likely to acquire territories and outcompete rivals (Pryke & Andersson, 2003b).

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20 9 If colour pattern is an honest signal of an individual's ability to defend its territory,
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22 10 one may also expect brighter or more colourful individuals to win contests more often
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24 11 than the dull ones, and thus be more successful in acquiring and maintaining high-
25
26 12 quality territories. As predicted, we find many examples of this throughout different
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28 13 taxa. For instance, territorial Augrabies flat lizard (*Platysaurus broadleyi*) males have a
29
30 14 lower ultraviolet reflectance than floaters (Whiting *et al.*, 2006), and the size of the red
31
32 15 spots in the wings of male rubyspot damselflies (*Hetaerina americana*) promotes
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34 16 successful defence of a territory (Grether, 1996).

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38 17 Nearly all anurans defend small, short-term individual spaces during the breeding
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40 18 season, as they confer a particular advantage for the propagation of mating signals or
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42 19 better access to mates (Wells, 2007, and references therein). Dendrobatid frogs seem to
43
44 20 be a remarkable exception to this generalisation, as all of the species studied to date
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46 21 exhibit some degree of long-term territoriality (Pröhl, 2005; but see Born *et al.*, 2010).
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48 22 They defend multi-purpose territories that aid in mate attraction (Wells, 2007) and
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50 23 subsequent breeding. Defended resources within each territory vary from species to
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52 24 species (Pröhl, 2005), but are most often related to their elaborate parental care.
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3 1 Even though most frogs rely on acoustic signals for communication, resident male
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5 2 dendrobatids generally advertise territory ownership to intruders with a combination of
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7 3 acoustic and visual signals (Hödl & Amézquita, 2001; Zimmermann & Zimmermann,
8
9 4 1988), and in some cases physical combats can occur until the intruder is chased away,
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11 5 or gains the territory over the resident. For example, according to Wells (1980),
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13 6 territorial male *Mannophryne* (= *Colostethus*) *trinitatis* turn black when calling while
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15 7 non-calling males remain brown. Most interestingly, black males seem to react
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17 8 aggressively only towards other black males, and often engage in wrestling. At the end
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19 9 of the encounters, one of the males changes his colour back to brown, which presumably
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21 10 decreases the probability of being attacked by a black male (Wells, 1980). A recent
22
23 11 study in laboratory conditions showed that *O. pumilio* males call and approach brighter
24
25 12 males more frequently than duller ones. Furthermore, a male's own brightness also
26
27 13 predicts his own behaviour, such that brighter males approach stimulus frogs faster and
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29 14 call more towards other bright individuals (Crothers, Gering & Cummings, 2011). A
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31 15 field study on males of the same species showed that the most aggressive males from
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33 16 eight sampled populations were the ones with the most conspicuous colouration (Rudh,
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35 17 Breed & Qvarnstrom, 2013).

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40 18 While these links between colouration and behaviour seem to be reported more
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42 19 frequently, the mechanisms explaining how they appear and persist warrant in-depth
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44 20 examination. Correlational selection, which may favour certain combinations of traits
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46 21 (such as behaviour and phenotype) expressed at the same time in an individual without
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48 22 affecting the expression of each trait on its own (Brodie, 1992; Endler, 1986; Sinervo &
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50 23 Svensson, 2002), may be one possibility. Pleiotropy and gene linkage have also been
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52 24 advocated as possible explanations for the higher levels of dominance of Gouldian
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1 finches (*Erythrura gouldiae*) bearing a red head, even when their head colour was
2 manipulated to look like the subordinate morphs (Pryke & Griffith, 2006).

3 Some species need acoustic and visual signals together (like the inflation–deflation
4 of the vocal sac) in order to trigger agonistic behaviour (de Luna, Hödl & Amézquita,
5 2010; Narins, Hödl & Grabul, 2003). Thus, the importance of visual signals and the
6 potential role of colouration and colour patterns in the agonistic interactions of species
7 that emit very soft sounds (*Dendrobates leucomelas*, *D. truncatus* and *D. auratus*;
8 Erdtmann & Amézquita, 2009), or lack an acoustic advertisement call (*Dendrobates*
9 *tinctorius*; Born *et al.*, 2010; B. Rojas, personal observations) demands further
10 investigation.

11 As shown above, both male territorial behaviour and male–male aggression have
12 been well documented in dendrobatoids (Pröhl, 2005). Aggression between females, on
13 the other hand, has been poorly studied. Wells (1980) reported aggression between
14 females of *Mannophryne trinitatis*. A resident female challenges an intruder by
15 acquiring an upright posture, and pulsating her bright yellow throat. Female
16 *Dendrobates auratus* behave aggressively by chasing each other or wrestling in the
17 presence of calling males (Summers, 1989). Apart from these studies, no others have
18 been conducted on the role of colours in the agonistic behaviour of female frogs, and
19 reports of its occurrence remain mostly a matter of side observations in male mating
20 behaviour studies (Summers, 1992; Wells, 1978). Female aggression occurs in at least
21 four colourful species: *O. pumilio*, *D. auratus*, *D. leucomelas* (Meuche, Linsenmair &
22 Proehl, 2011; Summers, 1989, 1992; Wells, 1978) and *D. tinctorius* (B. Rojas, personal
23 observations). Given that females do not vocalise at all, their means of acquiring,
24 defending, and maintaining resources are still obscure. Hence, species with territorial

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3 1 females constitute an excellent target for examining the importance of colour patterns
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5 2 during agonistic interactions.
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9 10 4 **IV. HABITAT SELECTION AND SPACE USE**

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12 5 The distribution of individuals of a species within a particular habitat might be explained
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14 6 by the location of resources or anti-predator refuges, the suitability of different
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16 7 microhabitats, or the preferences of conspecific individuals (Alcock, 2001). There are at
17
18 8 least two ways in which colours or colour patterns can be associated with habitat
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20 9 selection and space use. First, animals might choose a habitat that provides the best
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22 10 blending opportunities in order to minimise detection by predators (Ruxton *et al.*, 2004).
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24 11 This can be achieved, for example, either by background matching (Endler, 1984;
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26 12 Pellissier *et al.*, 2011), or by mixing with groups of species with similar colouration
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28 13 (Munday, Eyre & Jones, 2003). Second, animals can select microhabitats that increase
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30 14 their own conspicuousness or the conspicuousness of their sexual displays (Endler,
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32 15 1993*b*; Heindl & Winkler, 2003; Théry, 2001; Théry & Endler, 2001). Section II.1
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34 16 discussed how some species with cryptic colouration may select the habitat that best
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36 17 matches their colours in order to avoid being detected by predators. By contrast, in this
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38 18 section I will address implications other than background matching of colour-mediated
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40 19 space use and habitat selection.
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45 20 As discussed above, aposematic species rely on their conspicuousness to teach
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47 21 predators about their unprofitability which should lead to monomorphism of the warning
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49 22 signal. However, as explained previously there are many polymorphic (and polytypic)
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51 23 aposematic species. It is possible that these species benefit from colour pattern-mediated
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53 24 habitat selection as a mechanism to maintain colour pattern variability. In such cases,
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55 25 each morph could choose a habitat that maximises its conspicuousness. Alternatively, if
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1 conspicuousness differed among morphs, less-protected individuals could benefit from
2 choosing microhabitats that provide extra protection from predators, for example hiding
3 places. The former seems to be the case for males of *O. pumilio*, which seem to select
4 vocalisation perches that maximise their conspicuousness (Pröhl & Ostrowski, 2011).
5 Likewise, males from more conspicuously coloured populations of *O. granulifera* (Fig.
6 4G) tend to call from more exposed perches than populations with duller males (Willink
7 *et al.*, 2013).

8 Aposematic colour patterns may be associated with aspects of space use other than
9 habitat selection. For aposematism to work, prey density must be high (Mappes, Marples
10 & Endler, 2005; Ruxton *et al.*, 2004; Speed, 2000), or at least above a certain threshold
11 (Endler & Rojas, 2009), so that the frequency of encounters between the predator and
12 the defended prey enhances predator learning. Therefore, aposematic organisms are
13 assumed to benefit from spatial aggregations (Alatalo & Mappes, 1996; Lindström *et al.*,
14 2001; Mappes & Alatalo, 1997; Riipi *et al.*, 2001), and at least in some taxa it has been
15 demonstrated that gregariousness appeared after warning colouration (Beltrán *et al.*,
16 2007; Sillén-Tullberg, 1988). This does not necessarily occur in all aposematic species.
17 While colourful dendrobatids are also highly toxic, their strong territorial behaviour
18 makes them unlikely to be aggregated. However, one might predict that the territories of
19 conspicuous species are more aggregated than those of dull-coloured species.

20 Another way to approach the same conceptual problem would be to conduct
21 studies comparing populations of colourful species that differ in abundance. There is
22 good theoretical and increasing experimental support in favour of the idea that spatial
23 variation in selective pressures (i.e. predation) might be a relevant factor shaping the
24 evolution of aposematic signals (Chouteau & Angers, 2011; Nokelainen *et al.*, 2014;
25 Valkonen *et al.*, 2012), and that the emergence of novel signals can be a frequency-

1 dependent process (e.g. Endler & Mappes, 2004; Endler & Rojas, 2009). To date,
2 however, there is only one empirical study examining how the spatial variation in the
3 frequency of different warning signals may enable the emergence of novel signal types
4 (Comeault & Noonan, 2011). Using clay models Comeault & Noonan (2011) compared
5 the attack rate on different morphs of *Dendrobates tinctorius* in two populations
6 differing in the abundance of the local morph. This study provided evidence that
7 selective pressures affecting the survival rate of a 'protected' morph vary in accordance
8 with its relative abundance in the population. In other words, an aposematic signal that is
9 successful at high densities might not be so when the densities are low.

10

11 **V. FUTURE DIRECTIONS**

12 Even though the diversity of colours seen in anurans has been documented for decades,
13 only relatively recent studies have directly assessed its function and adaptive
14 significance (reviewed in Rudh & Qvarnström, 2013). Specifically, greater emphasis has
15 been placed on examining the role that colours and other visual signals play in frog
16 behaviour and ecology, especially in relation to protection from predators and mating.
17 Likewise, current research has increasingly explored the synergy between different
18 sensory modalities during mate attraction and male–male competition. Altogether, this
19 indicates that frogs are excellent subjects for research on the evolution of colour-related
20 anti-predation strategies and intraspecific communication systems. Future research
21 should focus on a variety of key issues listed below.

22 First, experimental approaches could be used to test the efficacy of eyespots and flash
23 colours as signals addressed to potential predators. Among diurnal frogs, predation
24 seems to be the selective pressure that stimulates the most interest among researchers
25 studying colour pattern diversity, but most studies have been carried out in one taxon

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3 1 only (Neotropical dart poison frogs) even though there is a great array of anuran taxa
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5 2 that possess diverse colour patterns (i.e. Mantellidae, Myobatrachidae, Bufonidae,
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7 3 Brachycephalidae, etc.). Furthermore, many recent studies have used clay models;
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9 4 although these studies represent a significant step forward in the study of warning
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11 5 signals, they are biased towards an overestimation of deaths, i.e. ignoring the fact that an
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13 6 attacked individual may survive. Additionally, research with clay models does not
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15 7 account for predators like snakes, which most likely would not attack the models, or for
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17 8 any behavioural correlates that act in concert with the warning signal. Future research
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19 9 could thus benefit from complementary studies where the survival of individuals bearing
20
21 10 different colour patterns is tracked, for example, using capture–mark–recapture methods
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23 11 in semi-natural or wild conditions. Also, given their dissimilar distribution ranges, it
24
25 12 would be especially interesting to compare the role of natural and sexual selection in the
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27 13 colour diversification of dendrobatids to that of mantellids, which show remarkable
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29 14 signs of evolutionary convergence in other aspects of their biology.

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32 15 Second, the role of colour patterns is relatively unexplored in mate choice and other
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34 16 aspects of sexual selection such as intrasexual competition, especially in highly
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36 17 territorial species. Studies on how nocturnal species use information conveyed in visual
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38 18 signals and why they may have retained colour vision and coloured traits whose
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40 19 expression may be costly, could refine our understanding of the function and importance
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42 20 of multimodal communication, which has proven increasingly prominent in anurans
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44 21 (Starnberger, Preininger & Hödl, 2014). Beyond the actual mate preferences, there is not
45
46 22 much evidence to date supporting the function of colours as reliable indicators of good
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48 23 genes in frogs. It is unknown, for example, whether females obtain indirect benefits
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50 24 when mating with brighter males or males with certain colour patterns and whether that
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52 25 enhances the survival of their offspring.
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3 1 Third, there is a surprisingly large gap in studies linking colour patterns and life history,
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5 2 especially in aposematic species. It would be very exciting to know what the costs and
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7 3 constraints are that compromise signal efficacy, and whether colours are honest signals
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9 4 for all receivers (i.e. both conspecifics and heterospecifics), or whether they are instead
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11 5 deceptive.

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14 6 Finally, we must consider other selective pressures besides mate choice and predation in
15
16 7 order to investigate how variation in colour patterns can originate and be maintained.

17
18 8 Future research should thus focus on exploring more aspects of the behaviour and
19
20 9 ecology of the studied species in order to have a better understanding of the function and
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22 10 evolution of the great diversity in colour patterns that exists among anurans today.
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27 12 **VI. CONCLUSIONS**

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30 13 (1) Animals use different modalities of communication depending on both intrinsic and
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32 14 extrinsic factors. Frogs are widely known for their acoustic signals, which are
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34 15 particularly useful for nocturnal animals. Recently, also the use of visual signals in frog
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36 16 communication has become a focus of research.

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39 17 (2) The first approaches to studying variation in frog colouration dealt mostly with the
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41 18 inheritance of colour patterns. However, not many studies have addressed directly the
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43 19 selective pressures involved in the maintenance of such variation.

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45 20 (3) Dorsal colour patterns can be directly related to predator avoidance. Some frogs
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47 21 blend with their surroundings in different ways such as matching the colouration of their
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49 22 background, exhibiting colour patterns that make it difficult to distinguish their outline
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51 23 the shape, or by resembling an inanimate object. By contrast, other species of frogs
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53 24 display conspicuous colour patterns which are often coupled with some form of
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55 25 unprofitability. Such colour patterns are thought to aid in predator education. Bright
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1 colouration displayed in other parts of the body such as the flanks or ventral side have
2 been much less studied, and their role in predator deterrence is yet to be tested.

3 (4) Cryptically coloured frogs are often polymorphic, which may be favourable as a
4 strategy to reduce the *per capita* predation risk. The mechanisms that maintain such
5 polymorphisms are still poorly understood, but predation seems to be a good candidate.
6 Aposematic species, on the other hand, are not expected to be polymorphic because
7 intrapopulational variation in warning signals might interfere with predator avoidance
8 learning. Nonetheless, a few aposematic species exhibit colour pattern polymorphisms.
9 Recent research suggests that such polymorphisms could be maintained *via* an
10 interaction between predation and sexual selection, spatio-temporal variation in
11 selection, an association between fitness-related traits and colour patterns, predator
12 generalisation, hybridisation among variants or genetic drift.

13 (5) Experiments in the field with frog models displaying conspicuous colouration have
14 shown the effectiveness of some colour patterns in predator deterrance. With a few
15 exceptions, local morphs tend to be less attacked than novel ones, and moving models
16 exhibiting conspicuous colouration receive fewer attacks than stationary ones.

17 (6) In natural frog populations some palatable species mimic the coloration of defended
18 species, obtaining benefits against predation without investing in the production or
19 sequestration of defensive chemicals (Batesian mimicry). Two (or more) chemically
20 defended species can also share similar colour patterns, thus sharing the costs of
21 predator education (Müllerian mimicry). Only a handful of studies have been able to
22 demonstrate the occurrence of either type of mimicry in anurans, and they have focused
23 only on one taxonomic group.

24 (7) Aposematic individuals are likely to display warning signals that are reliable
25 indicators of their unprofitability, as only defended prey can afford the costs of being

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3 1 detectable and of having the pigments required for strong warning signals. To
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5 2 demonstrate signal honesty, showing a positive correlation between warning signals and
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7 3 toxicity is not enough. It is necessary to differentiate between quantitative honesty and
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9 4 other ways in which colour patterns and chemical defences can be correlated without
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11 5 involving honest signalling.

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14 6 (8) Colours can have an important role in mate choice, particularly for diurnal species.

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16 7 Interestingly, some species of nocturnal frogs have been shown to pay attention to
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18 8 different components of male colour patterns. This has also been studied extensively in
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20 9 one diurnal species. However, the fitness consequences of female preferences regarding
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22 10 male colouration remain poorly understood.

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25 11 (9) In certain cases, colour patterns may provide information about the social status or
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27 12 fighting abilities of an individual during agonistic encounters. Although this has been
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29 13 shown for the males of some species, there is increasing evidence that colours might
30
31 14 play a role in conflict resolution also among females.

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34 15 (10) Besides background matching, habitat selection and space use may also be related
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36 16 to colouration such that different morphs of a species choose a habitat that boosts their
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38 17 conspicuousness. Alternatively, in accordance with the assumed benefits of aggregation
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40 18 of aposematic species, the territories of aposematic frogs might be clumped compared to
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42 19 those of cryptic species. This hypothesis warrants further examination.

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45 20 (11) The detailed study of colour pattern variation, particularly with an experimental
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47 21 approach, should facilitate the formulation and testing of hypotheses on the evolution of
48
49 22 predator–prey interactions, mating preferences and communication systems, and provide
50
51 23 valuable knowledge on the mechanisms promoting and maintaining signal diversity.

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60

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11 VIII. REFERENCES

- 12 ALATALO, R. V. & MAPPES, J. (1996). Tracking the evolution of warning signals. *Nature*
13 **382**, 708–710.
- 14 ALCOCK, J. (2001). *Animal behavior: An evolutionary approach*. Seventh Edition.
- 15 ALLEN, J. A. & CLARKE, B. (1968). Evidence for apostatic selection by wild passerines.
16 *Nature* **220**, 501–502.
- 17 ALLEN, J. A. & GREENWOOD, J. J. D. (1988). Frequency-dependent selection by
18 predators. *Philosophical Transactions of the Royal Society of London B* **319**,
19 485–503.
- 20 ALVARADO, J. B., ALVAREZ, A. & SAPORITO, R. A. (2013). *Oophaga pumilio*
21 (Strawberry Poison Frog), *Baryphthengus martii* (Rufous Motmot). Predator
22 prey interactions. *Herpetological Review* **44**, 298.
- 23 AMAT, F., WOLLENBERG, K. C. & VENCES, M. (2013). Correlates of eye colour and
24 pattern in mantellid frogs. *Salamandra* **49**, 7–17.

- 1
2
3 1 AMÉZQUITA, A. (1999). Color pattern, elevation and body size in the high-Andean frog
4
5 2 *Hyla labialis*. *Revista de la Academia Colombiana de Ciencias Exactas Físicas y*
6
7 3 *Naturales* **XXIII**, 231–238.
- 9
10 4 AMÉZQUITA, A., CASTRO, L., ARIAS, M., GONZÁLEZ, M. & ESQUIVEL, C. (2013). Field
11
12 5 but not lab paradigms support generalisation by predators of aposematic
13
14 6 polymorphic prey: the *Oophaga histrionica* complex. *Evolutionary Ecology* **27**,
15
16 7 769–782.
- 18
19 8 AMÉZQUITA, A., FLECHAS, S. V., LIMA, A. P., GASSER, H. & HÖDL, W. (2011). Acoustic
20
21 9 interference and recognition space within a complex assemblage of dendrobatid
22
23 10 frogs. *Proceedings of the National Academy of Sciences* **108**, 17058–17063.
- 25
26 11 AMÉZQUITA, A., LIMA, A. P., JEHL, R., CASTELLANOS, L., RAMOS, O., CRAWFORD, A. J.,
27
28 12 GASSER, H. & HOEDL, W. (2009). Calls, colours, shape, and genes: a multi-trait
29
30 13 approach to the study of geographic variation in the Amazonian frog *Allobates*
31
32 14 *femorales*. *Biological Journal of the Linnean Society* **98**, 826–838.
- 34
35 15 ANDERSSON, M. (1994). *Sexual Selection*. Princeton University Press, Princeton, New
36
37 16 Jersey.
- 38
39 17 ANDERSSON, S., ORNBORG, J. & ANDERSSON, M. (1998). Ultraviolet sexual dimorphism
40
41 18 and assortative mating in blue tits. *Proceedings of the Royal Society of London B*
42
43 19 **265**, 445–450.
- 45
46 20 BAJER, K., MOLNAR, O., TOEROEK, J. & HERCZEG, G. (2010). Female European green
47
48 21 lizards (*Lacerta viridis*) prefer males with high ultraviolet throat reflectance.
49
50 22 *Behavioral Ecology and Sociobiology* **64**, 2007–2014.
- 52
53 23 BAJGER, J. (1980). Diversity of defensive responses in populations of fire toads
54
55 24 (*Bombina bombina* and *Bombina variegata*). *Herpetologica* **36**, 133–137.

- 1
2
3 1 BARBER, I., ARNOTT, S. A., BRAITHWAITE, V. A., ANDREW, J. & HUNTINGFORD, F. A.
4
5 2 (2001). Indirect fitness consequences of mate choice in sticklebacks: offspring of
6
7 3 brighter males grow slowly but resist parasitic infections. *Proceedings of the*
8
9 4 *Royal Society of London B* **268**, 71–76.
- 10
11 5 BATES, H. W. (1862). Contributions to an insect fauna of the Amazon Valley.
12
13 6 Lepidoptera: Heliconidae. 23. *Transactions of the Linnean Society of London* **23**,
14
15 7 495–566.
- 16
17 8 BELL, R. C. & ZAMUDIO, K. R. (2012). Sexual dichromatism in frogs: natural selection,
18
19 9 sexual selection and unexpected diversity. *Proceedings of the Royal Society B*
20
21 10 **279**, 4687–4693.
- 22
23 11 BELTRÁN, M., JIGGINS, C. D., BROWER, A. V. Z., BERMINGHAM, E. & MALLETT, J. (2007).
24
25 12 Do pollen feeding, pupal-mating and larval gregariousness have a single origin in
26
27 13 *Heliconius* butterflies? Inferences from multilocus DNA sequence data.
28
29 14 *Biological Journal of the Linnean Society* **92**, 221–239.
- 30
31 15 BLOUIN, M. S. (1989). Inheritance of a naturally-occurring color polymorphism in the
32
33 16 ornate chorus frog, *Pseudacris ornata*. *Copeia*, 1056–1059.
- 34
35 17 BLOUNT, J. D., ROWLAND, H. M., DRIJFHOUT, F. P., ENDLER, J. A., INGER, R., SLOGGETT,
36
37 18 J. J., HURST, G. D. D., HODGSON, D. J. & SPEED, M. P. (2012). How the ladybird
38
39 19 got its spots: effects of resource limitation on the honesty of aposematic signals.
40
41 20 *Functional Ecology* **26**, 334–342.
- 42
43 21 BLOUNT, J. D., SPEED, M. P., RUXTON, G. D. & STEPHENS, P. A. (2009). Warning
44
45 22 displays may function as honest signals of toxicity. *Proceedings of the Royal*
46
47 23 *Society B* **276**, 871–877.
- 48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 BONANSEA, M. I. & VAIRA, M. (2012). Geographic and intrapopulational variation in
4
5 2 colour and patterns of an aposematic toad, *Melanophryniscus rubriventris*
6
7 3 (Amphibia, Anura, Bufonidae). *Amphibia-Reptilia* **33**, 11–24.
8
9
10 4 BOND, A. B. (2007). The evolution of color polymorphism: Crypticity searching images,
11
12 5 and apostatic selection. *Annual Review of Ecology Evolution and Systematics* **38**,
13
14 6 489–514.
15
16 7 BOND, A. B. & KAMIL, A. C. (1998). Apostatic selection by blue jays produces balanced
17
18 8 polymorphism in virtual prey. *Nature* **395**, 594–596.
19
20 9 BOND, A. B. & KAMIL, A. C. (2006). Spatial heterogeneity, predator cognition, and the
21
22 10 evolution of color polymorphism in virtual prey. *Proceedings of the National*
23
24 11 *Academy of Sciences* **103**, 3214–3219.
25
26
27 12 BORER, M., VAN NOORT, T., RAHIER, M. & NAISBIT, R. E. (2010). Positive frequency-
28
29 13 dependent selection on warning color in alpine leaf beetles *Evolution* **64**, 3629–
30
31 14 3633.
32
33
34 15 BORN, M., BONGERS, F., POELMAN, E. H. & STERCK, F. J. (2010). Dry-season retreat and
35
36 16 dietary shift of the dart-poison frog *Dendrobates tinctorius* (Anura:
37
38 17 *Dendrobatidae*). *Phyllomedusa* **9**, 37–52.
39
40
41 18 BOURKE, J., BUSSE, K. & BAKKER, T. C. M. (2011). Sex differences in polymorphic body
42
43 19 coloration and dorsal pattern in Darwin's frogs (*Rhinoderma darwinii*).
44
45 20 *Herpetological Journal* **21**, 227–234.
46
47
48 21 BRADBURY, J. W. & VEHCAMP, S. L. (2011). *Principles of Animal Communication*,
49
50 22 2nd Edition edition. Sinauer Associates Inc. , Sunderland, Massachusetts.
51
52 23 BRODIE, E. D. (1992). Correlational selection for color pattern and antipredator behavior
53
54 24 in the garter snake *Thamnophis ordinoides*. *Evolution* **46**, 1284–1298.
55
56
57
58
59
60

- 1 BRUSA, O., BELLATI, A., MEUCHE, I., MUNDY, N. I. & PROHL, H. (2013). Divergent
2 evolution in the polymorphic granular poison-dart frog, *Oophaga granulifera*:
3 genetics, coloration, advertisement calls and morphology. *Journal of*
4 *Biogeography* **40**, 394–408.
- 5 CAORSI, V. Z., COLOMBO, P., FREIRE, M. D., AMARAL, I. B., ZANK, C. & GRANT, T.
6 (2014). Natural history, coloration pattern and conservation status of the
7 threatened South Brazilian red bellied toad, *Melanophryniscus macrogranulosus*
8 Braun, 1973 (Anura, Bufonidae). *Herpetology Notes* **7**, 585–598.
- 9 CAPRANICA, R. R. & MOFFAT, A. J. M. (1983). Neurobehavioral correlates of sound
10 communication in anurans. In *Advances in Vertebrate Neuroethology* (ed. J. P.
11 Ewert, R. R. Capranica and D. J. Ingle), pp. 701–730. Plenum Press, New York.
- 12 CARO, T. (2014). Antipredator deception in terrestrial vertebrates. *Current Zoology* **60**,
13 16–25.
- 14 CHECK, A., BOGART, J. P. & LOUGHEED, S. C. (2003). Mating signal partitioning in
15 multi-species assemblages: a null model test using frogs. *Ecology Letters* **6**, 235–
16 247.
- 17 CHOI, N. & JANG, Y. (2014). Background matching by means of dorsal color change in
18 treefrog populations (*Hyla japonica*). *Journal of Experimental Zoology A:*
19 *Ecological Genetics and Physiology* **321**, 108–118.
- 20 CHOUTEAU, M. & ANGERS, B. (2011). The role of predators in maintaining the
21 geographic organization of aposematic signals. *American Naturalist* **178**, 810–
22 817.
- 23 CHOUTEAU, M., SUMMERS, K., MORALES, V. & ANGERS, B. (2011). Advergence in
24 Mullerian mimicry: the case of the poison dart frogs of Northern Peru revisited.
25 *Biology Letters* **7**, 796–800.

- 1
2
3 1 CLARK, V. C., RAXWORTHY, C. J., RAKOTOMALALA, V., SIERWALD, P. & FISHER, B. L.
4
5 2 (2005). Convergent evolution of chemical defense in poison frogs and arthropod
6
7 3 prey between Madagascar and the Neotropics. *Proceedings of the National*
8
9 4 *Academy of Sciences* **102**, 11617–11622.
- 10
11 5 COMEAULT, A. A. & NOONAN, B. P. (2011). Spatial variation in the fitness of divergent
12
13 6 aposematic phenotypes of the poison frog, *Dendrobates tinctorius*. *Journal of*
14
15 7 *Evolutionary Biology* **24**, 1374–1379.
- 16
17 8 COOPER, W. E., JR., CALDWELL, J. P. & VITT, L. J. (2008). Effective crypsis and its
18
19 9 maintenance by immobility in *Craugastor* frogs. *Copeia*, 527–532.
- 20
21 10 COTT, H. B. (1940). *Adaptive Colouration in Animals*. Methuen, London.
- 22
23 11 CROTHERS, L., GERING, E. & CUMMINGS, M. (2011). Aposematic signal variation
24
25 12 predicts male-male interactions in a polymorphic poison frog *Evolution* **65**, 599–
26
27 13 605.
- 28
29 14 CROTHERS, L. R. & CUMMINGS, M. E. (2013). Warning signal brightness variation:
30
31 15 sexual selection may work under the radar of natural selection in populations of a
32
33 16 polytypic poison frog. *American Naturalist* **181**, E116–E124.
- 34
35 17 CUMMINGS, M. E. & CROTHERS, L. R. (2013). Interacting selection diversifies warning
36
37 18 signals in a polytypic frog: an examination with the strawberry poison frog.
38
39 19 *Evolutionary Ecology* **27**, 693–710.
- 40
41 20 CUTHILL, I. C., STEVENS, M., SHEPPARD, J., MADDOCKS, T., PARRAGA, C. A. &
42
43 21 TROSCIANKO, T. S. (2005). Disruptive coloration and background pattern
44
45 22 matching. *Nature* **434**, 72–74.
- 46
47 23 CUTRERA, A. P., FANJUL, M. S. & ZENUTO, R. R. (2012). Females prefer good genes:
48
49 24 MHC-associated mate choice in wild and captive tuco-tucos. *Animal Behaviour*
50
51 25 **83**, 847–856.
- 52
53
54
55
56
57
58
59
60

- 1 DALY, J. W., GARRAFFO, H. M., HALL, G. S. E. & COVER, J. F., JR. (1997). Absence of
2 skin alkaloids in captive-raised Madagascan mantelline frogs (*Mantella*) and
3 sequestration of dietary alkaloids. *Toxicon* **35**, 1131–1135.
- 4 DALY, J. W., GARRAFFO, H. M., PANNELL, L. K., SPANDE, T. F., SEVERINI, C. &
5 ERSPAMER, V. (1990). Alkaloids from Australian frogs (Myobatrachidae) –
6 Pseudophrynamines and pumiliotoxins. *Journal of Natural Products* **53**, 407–
7 421.
- 8 DALY, J. W., KANEKO, T., WILHAM, J., GARRAFFO, H. M., SPANDE, T. F., ESPINOSA, A. &
9 DONNELLY, M. A. (2002). Bioactive alkaloids of frog skin: Combinatorial
10 bioprospecting reveals that pumiliotoxins have an arthropod source. *Proceedings*
11 *of the National Academy of Sciences* **99**, 13996–14001.
- 12 DALY, J. W. & MYERS, C. W. (1967). Toxicity of Panamanian poison frogs
13 (*Dendrobates*) – Some biological and chemical aspects. *Science* **156**, 970–&.
- 14 DALY, J. W., SECUNDA, S. I., GARRAFFO, H. M., SPANDE, T. F., WISNIESKI, A. & COVER,
15 J. F., JR. (1994). An uptake system for dietary alkaloids in poison frogs
16 (*Dendrobatidae*). *Toxicon* **32**, 657–663.
- 17 DARST, C. R. & CUMMINGS, M. E. (2006). Predator learning favours mimicry of a less-
18 toxic model in poison frogs. *Nature* **440**, 208–211.
- 19 DARST, C. R., CUMMINGS, M. E. & CANNATELLA, D. C. (2006). A mechanism for
20 diversity in warning signals: Conspicuousness versus toxicity in poison frogs.
21 *Proceedings of the National Academy of Sciences* **103**, 5852–5857.
- 22 DARST, C. R., MENENDEZ-GUERRERO, P. A., COLOMA, L. A. & CANNATELLA, D. C.
23 (2005). Evolution of dietary specialization and chemical defense in poison frogs
24 (*Dendrobatidae*): A comparative analysis. *American Naturalist* **165**, 56–69.

- 1
2
3 1 DAVISON, J. (1963). Gene action mechanisms in determination of color and pattern in
4
5 2 frog (*Rana pipiens*). *Science* **141**, 648–&.
- 6
7 3 DE LUNA, A. G., HÖDL, W. & AMÉZQUITA, A. (2010). Colour, size and movement as
8
9 4 visual subcomponents in multimodal communication by the frog *Allobates*
10
11 5 *femorialis*. *Animal Behaviour* **79**, 739–745.
- 12
13 6 DEMUTH, J. P., NAIDU, A. & MYDLARZ, L. D. (2012). Sex, war, and disease: the role of
14
15 7 parasite infection on weapon development and mating success in a horned beetle
16
17 8 (*Gnatocerus cornutus*). *PLoS One* **7**, e28690.
- 18
19 9 DOUCET, S. M. & MENNILL, D. J. (2010). Dynamic sexual dichromatism in an
20
21 10 explosively breeding Neotropical toad. *Biology Letters* **6**, 63–66.
- 22
23 11 DREHER, C. E., CUMMINGS, M. E. & PRÖHL, H. (2015). An analysis of predator selection
24
25 12 to affect aposematic coloration in a poison frog species *PLoS One* **10**, e0130571.
- 26
27 13 DUELLMAN, W. E. & PYLES, R. E. (1983). Acoustic resource partitioning in anuran
28
29 14 communities. *Copeia* **1983**, 639–649.
- 30
31 15 DUELLMAN, W. E. & TRUEB, L. (1994). *Biology of Amphibians*. McGraw Hill Publ. Co.,
32
33 16 New York.
- 34
35 17 EDMUNDS, M. (1974). *Defence in Animals: A Survey of Antipredator Defences*.
36
37 18 Longman, New York.
- 38
39 19 ENDLER, J. A. (1978). A predator's view of animal colour patterns. *Evolutionary Biology*
40
41 20 **11**, 319–364.
- 42
43 21 ENDLER, J. A. (1980). Natural selection on color patterns in *Poecilia reticulata*.
44
45 22 *Evolution* **34**, 76–91.
- 46
47 23 ENDLER, J. A. (1984). Progressive background matching in moths, and a quantitative
48
49 24 measure of crypsis *Biological Journal of the Linnean Society* **22**, 187–231.
- 50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 ENDLER, J. A. (1986). *Natural selection in the wild*. Princeton University Press,
4
5 2 Princeton.
6
7 3 ENDLER, J. A. (1988). Frequency-dependent predation, crypsis and aposematic
8
9 4 coloration *Philosophical Transactions of the Royal Society of London B* **319**,
10
11 5 505–523.
12
13 6 ENDLER, J. A. (1990). On the measurement and classification of colour in studies of
14
15 7 animal colour patterns. *Biological Journal of the Linnean Society* **41**, 315–352.
16
17 8 ENDLER, J. A. (1991a). Interactions between predators and prey. In *Behavioural*
18
19 9 *Ecology. An evolutionary approach*. (ed. J. R. Krebs and N. B. Davies), pp. 169–
20
21 10 196. Cambridge University Press, Cambridge.
22
23 11 ENDLER, J. A. (1991b). Variation in the appearance of guppy color patterns to guppies
24
25 12 and their predators under different visual conditions. *Vision Research* **31**, 587–
26
27 13 608.
28
29 14 ENDLER, J. A. (1992). Signals, signal condition and the direction of evolution *American*
30
31 15 *Naturalist* **139**, S125–S153.
32
33 16 ENDLER, J. A. (1993a). Some general comments on the evolution and design of animal
34
35 17 communication systems. *Philosophical Transactions of the Royal Society of*
36
37 18 *London B* **340**, 215–225.
38
39 19 ENDLER, J. A. (1993b). The color of light in forests and its implications. *Ecological*
40
41 20 *Monographs* **63**, 1–27.
42
43 21 ENDLER, J. A. (2012). A framework for analysing colour pattern geometry: adjacent
44
45 22 colours. *Biological Journal of the Linnean Society* **107**, 233–253.
46
47 23 ENDLER, J. A. & MAPPES, J. (2004). Predator mixes and the conspicuousness of
48
49 24 aposematic signals. *American Naturalist* **163**, 532–547.
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 ENDLER, J. A. & MIELKE, P. W. (2005). Comparing entire colour patterns as birds see
4
5 2 them. *Biological Journal of the Linnean Society* **86**, 405–431.
6
7 3 ENDLER, J. A. & ROJAS, B. (2009). The spatial pattern of natural selection when selection
8
9 4 depends on experience. *American Naturalist* **173**, E62–E78.
10
11 5 ERDTMANN, L. & AMÉZQUITA, A. (2009). Differential evolution of advertisement call
12
13 6 traits in dart-poison frogs (Anura: Dendrobatidae). *Ethology* **115**, 801–811.
14
15 7 EVANS, M. R. & NORRIS, K. (1996). The importance of carotenoids in signaling during
16
17 8 aggressive interactions between male firemouth cichlids (*Cichlasoma meeki*).
18
19 9 *Behavioral Ecology* **7**, 1–6.
20
21 10 EXNEROVÁ, A., SVÁDOVÁ, K., STYS, P., BARCALOVÁ, S., LANDOVÁ, E., PROKOPOVÁ,
22
23 11 M., FUCHS, R. & SOCHA, R. (2006). Importance of colour in the reaction of
24
25 12 passerine predators to aposematic prey: experiments with mutants of *Pyrrhocoris*
26
27 13 *apterus* (Heteroptera). *Biological Journal of the Linnean Society* **88**, 143–153.
28
29 14 FARALLO, V. R. & FORSTNER, M. R. J. (2012). Predation and the maintenance of color
30
31 15 polymorphism in a habitat specialist squamate. *PLoS One* **7**, e30316.
32
33 16 FLORES, E. E., STEVENS, M., MOORE, A. J. & BLOUNT, J. D. (2013). Diet, development
34
35 17 and the optimization of warning signals in post-metamorphic green and black
36
37 18 poison frogs. *Functional Ecology* **27**, 816–829.
38
39 19 FOGLEMAN, J. C., CORN, P. S. & PETTUS, D. (1980). The genetic-basis of a dorsal color
40
41 20 polymorphism in *Rana pipiens*. *Journal of Heredity* **71**, 439–440.
42
43 21 FORD, E. B. (1945). Polymorphism. *Biological Reviews of the Cambridge Philosophical*
44
45 22 *Society* **20**, 73–88.
46
47 23 FRASER, S., CALLAHAN, A., KLASSEN, D. & SHERRATT, T. N. (2007). Empirical tests of
48
49 24 the role of disruptive coloration in reducing detectability. *Proceedings of the*
50
51 25 *Royal Society B* **274**, 1325–1331.
52
53
54
55
56
57
58
59
60

- 1
2
3 1 FRITZ, G., RAND, A. S. & DEPAMPHILIS, C. W. (1981). The aposematically colored frog,
4
5 2 *Dendrobates pumilio*, is distasteful to the large, predatory ant, *Paraponera*
6
7 3 *clavata*. *Biotropica* **13**, 158–159.
8
9
10 4 FUHRMAN, F. A., FUHRMAN, G. J. & MOSHER, H. S. (1969). Toxin from skin of frogs of
11
12 5 the genus *Atelopus*: differentiation from dendrobatid toxins. *Science* **165**, 1376–
13
14 6 1377.
15
16 7 GALARZA, J. A., NOKELAINEN, O., ASHRAFI, R., HEGNA, R. H. & MAPPES, J. (2014).
17
18 8 Temporal relationship between genetic and warning signal variation in the
19
20 9 aposematic wood tiger moth (*Parasemia plantaginis*). *Molecular Ecology* **23**,
21
22 10 4939–4957.
23
24
25 11 GARRAFFO, H. M., CACERES, J., DALY, J. W., SPANDE, T. F., ANDRIAMAHARAVO, N. R. &
26
27 12 ANDRIANTSIFERANA, M. (1993a). Alkaloids in Madagascan frogs (*Mantella*).
28
29 13 Pumiliotoxins, indolizidines, quinolizidines, and pyrrolizidines. *Journal of*
30
31 14 *Natural Products* **56**, 1016–1038.
32
33
34 15 GARRAFFO, H. M., SPANDE, T. F., DALY, J. W., BALDESSARI, A. & GROS, E. G. (1993b).
35
36 16 Alkaloids from bufonid toads (*Melanophryniscus*) – Decahydroquinolines,
37
38 17 pumiliotoxins and homopumiliotoxins, indolizidines, pyrrolizidines, and
39
40 18 quinolizidines. *Journal of Natural Products* **56**, 357–373.
41
42
43 19 GERHARDT, H. C. & HUBER, F. (2002). *Acoustic Communication in Insects and Anurans*.
44
45 20 *Common problems and diverse solutions*. . The University of Chicago Press,
46
47 21 Chicago.
48
49
50 22 GOMEZ, D., RICHARDSON, C., LENGAGNE, T., DEREX, M., PLENET, S., JOLY, P., LENA, J.-
51
52 23 P. & THÉRY, M. (2010). Support for a role of colour vision in mate choice in the
53
54 24 nocturnal European treefrog (*Hyla arborea*). *Behaviour* **147**, 1753–1768.
55
56
57
58
59
60

- 1
2
3 1 GOMEZ, D., RICHARDSON, C., LENGAGNE, T., PLENET, S., JOLY, P., LENA, J.-P. & THÉRY,
4
5 2 M. (2009). The role of nocturnal vision in mate choice: females prefer
6
7 3 conspicuous males in the European tree frog (*Hyla arborea*). *Proceedings of the*
8
9 4 *Royal Society B* **276**, 2351–2358.
- 10
11 5 GORDON, S. P., KOKKO, H., ROJAS, B., NOKELAINEN, O. & MAPPES, J. (2015). Colour
12
13 6 polymorphism torn apart by opposing positive frequency-dependent selection,
14
15 7 yet maintained in space. *Journal of Animal Ecology* **84**, 1555–1564.
- 16
17 8 GRANT, T., COLOMBO, P., VERRASTRO, L. & SAPORITO, R. A. (2012). The occurrence of
18
19 9 defensive alkaloids in non-integumentary tissues of the Brazilian red-belly toad
20
21 10 *Melanophryniscus simplex* (Bufonidae). *Chemoecology* **22**, 169–178.
- 22
23 11 GRAY, H. M. & CHRISTY, J. H. (2000). Predation by the grapsid crab, *Armases angustum*
24
25 12 (Smith, 1870), on tadpoles of the green poison frog, *Dendrobates auratus* Girard,
26
27 13 1855. *Crustaceana* **73**, 1023–1025.
- 28
29 14 GRAY, H. M., KAISER, H. & GREEN, D. M. (2010). Does alkaloid sequestration protect
30
31 15 the green poison frog, *Dendrobates auratus*, from predator attacks? *Salamandra*
32
33 16 **46**, 235–238.
- 34
35 17 GRAY, S. M. & MCKINNON, J. S. (2007). Linking color polymorphism maintenance and
36
37 18 speciation. *Trends in Ecology & Evolution* **22**, 71–79.
- 38
39 19 GREENWOOD, J. J. D., WOOD, E. M. & BATCHELOR, S. (1981). Apostatic selection of
40
41 20 distasteful prey. *Heredity* **47**, 27–34.
- 42
43 21 GREYER, G. F. (1996). Intrasexual competition alone favors a sexually dimorphic
44
45 22 ornament in the rubyspot damselfly *Hetaerina americana*. *Evolution* **50**, 1949–
46
47 23 1957.
- 48
49 24 GUILFORD, T. & DAWKINS, M. S. (1991). Receiver psychology and the evolution of
50
51 25 animal signals. *Animal Behaviour* **42**, 1–14.
- 52
53
54
55
56
57
58
59
60

- 1
2
3 1 GUILFORD, T. & DAWKINS, M. S. (1993). Are warning colors handicaps? *Evolution* **47**,
4
5 2 400–416.
6
7 3 HAMILTON, W. D. & ZUK, M. (1982). Heritable true fitness and bright birds –a role for
8
9 4 parasites. *Science* **218**, 384–387.
10
11 5 HANTAK, M. M., GRANT, T., REINSCH, S., MCGINNITY, D., LORING, M., TOYOOKA, N. &
12
13 6 SAPORITO, R. A. (2013). Dietary alkaloid sequestration in a poison frog: an
14
15 7 experimental test of alkaloid uptake in *Melanophryniscus stelzneri* (Bufonidae).
16
17 8 *Journal of Chemical Ecology* **39**, 1400–1406.
18
19 9 HEGNA, R. H., GALARZA, J. A. & MAPPES, J. (2015). Global phylogeography and
20
21 10 geographical variation in warning coloration of the wood tiger moth (*Parasemia*
22
23 11 *plantaginis*). *Journal of Biogeography* **42**, 1669–1481.
24
25 12 HEGNA, R. H., SAPORITO, R. A. & DONNELLY, M. A. (2013). Not all colors are equal:
26
27 13 predation and color polytypism in the aposematic poison frog *Oophaga pumilio*.
28
29 14 *Evolutionary Ecology* **27**, 831–845.
30
31 15 HEGNA, R. H., SAPORITO, R. A., GEROW, K. G. & DONNELLY, M. A. (2011). Contrasting
32
33 16 colors of an aposematic poison frog do not affect predation. *Annales Zoologici*
34
35 17 *Fennici* **48**, 29–38.
36
37 18 HEINDL, M. & WINKLER, H. (2003). Interacting effects of ambient light and plumage
38
39 19 color patterns in displaying Wire-tailed Manakins (Aves, Pipridae). *Behavioral*
40
41 20 *Ecology and Sociobiology* **53**, 153–162.
42
43 21 HEYING, H. E. (2001). Social and reproductive behaviour in the Madagascan poison
44
45 22 frog, *Mantella laevis*, with comparisons to the dendrobatids. *Animal*
46
47 23 *Behaviour* **61**, 567–577.
48
49 24 HÖDL, W. & AMÉZQUITA, A. (2001). Visual signaling in anuran amphibians. In *Anuran*
50
51 25 *Communication*. (ed. M. J. Ryan), pp. 121–141.
52
53
54
55
56
57
58
59
60

- 1
2
3 1 HOFFMAN, E. A. & BLOUIN, M. S. (2000). A review of colour and pattern polymorphisms
4
5 2 in anurans. *Biological Journal of the Linnean Society* **70**, 633–665.
6
7 3 HOLEN, O. H. & SVENNUNGENSEN, T. O. (2012). Aposematism and the handicap principle.
8
9 4 *American Naturalist* **180**, 629–641.
10
11 5 HOOGMOED, M. & AVILA-PIRES, T. C. S. (2012). Inventory of color polymorphism in
12
13 6 populations of *Dendrobates galactonotus* (Anura: Dendrobatidae), a poison frog
14
15 7 endemic to Brazil. *Phyllomedusa* **11**, 95–115.
16
17 8 HORAK, P., OTS, I., VELLAU, H., SPOTTISWOODE, C. & MOLLER, A. P. (2001).
18
19 9 Carotenoid-based plumage coloration reflects hemoparasite infection and local
20
21 10 survival in breeding great tits. *Oecologia* **126**, 166–173.
22
23 11 JOHNSTONE, R. A. (1997). The evolution of animal signals. In *Behavioural ecology: an*
24
25 12 *evolutionary approach. Fourth edition.* (ed. J. R. D. N. B. Krebs), pp. 155–178.
26
27 13 Cambridge University Press, Cambridge.
28
29 14 JORON, M. & MALLET, J. (1998). Diversity in mimicry: paradox or paradigm? *Trends in*
30
31 15 *Ecology & Evolution* **13**, 461–463.
32
33 16 KARPESTAM, E., MERILAITA, S. & FORSMAN, A. (2013). Detection experiments with
34
35 17 humans implicate visual predation as a driver of colour polymorphism dynamics
36
37 18 in pygmy grasshoppers. *BMC Ecology* **13**.
38
39 19 KARPESTAM, E., MERILAITA, S. & FORSMAN, A. (2014). Natural levels of colour
40
41 20 polymorphism reduce performance of visual predators searching for
42
43 21 camouflaged prey. *Biological Journal of the Linnean Society* **112**, 546–555.
44
45 22 KAUFMANN, J. H. (1983). On the definitions and functions of dominance and
46
47 23 territoriality. *Biological Reviews of the Cambridge Philosophical Society* **58**, 1–
48
49 24 20.
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 KEMP, D. J., HERBERSTEIN, M. E., FLEISHMAN, L. J., ENDLER, J. A., BENNETT, A. T. D.,
4
5 2 DYER, A. G., HART, N. S., MARSHALL, J. & WHITING, M. J. (2015). An integrative
6
7 3 framework for the appraisal of coloration in nature. *American Naturalist* **185**,
8
9 4 705–724.
- 10
11 5 KIM, Y. H., KIM, Y. B. & YOTSU-YAMASHITA, M. (2003). Potent neurotoxins:
12
13 6 Tetrodotoxin, chiriquitoxin, and zetekitoxin from *Atelopus* frogs in Central
14
15 7 America. *Journal of Toxicology-Toxin Reviews* **22**, 521–532.
- 16
17 8 KONDRASHOV, A. S. & SHPAK, M. (1998). On the origin of species by means of
18
19 9 assortative mating. *Proceedings of the Royal Society of London B* **265**, 2273–
20
21 10 2278.
- 22
23 11 LANCASTER, L. T., HIPSLEY, C. A. & SINERVO, B. (2009). Female choice for optimal
24
25 12 combinations of multiple male display traits increases offspring survival.
26
27 13 *Behavioral Ecology* **20**, 993–999.
- 28
29 14 LENGER, D. R., BERKEY, J. K. & DUGAS, M. B. (2014). Predation on the toxic *Oophaga*
30
31 15 *pumilio* (Anura: Dendrobatidae) by *Rhadinaea decorata* (Squamata:
32
33 16 Collubridae) *Herpetology Notes* **7**, 83–84.
- 34
35 17 LENZI-MATTOS, R., M. M. ANTONIAZZI, M. M., HADDAD, C. F. B., TAMBOURGI, D. V.,
36
37 18 RODRIGUES, M. T. & JARED, C. (2005). The inguinal macroglands of the frog
38
39 19 *Physalaemus nattereri* (Leptodactylidae): structure, toxic secretion and
40
41 20 relationship with deimatic behaviour. *Journal of Zoology* **266**, 385–394.
- 42
43 21 LINDSTRÖM, L., ALATALO, R. V., LYYTINEN, A. & MAPPES, J. (2001). Strong
44
45 22 antiapostatic selection against novel rare aposematic prey. *Proceedings of the*
46
47 23 *National Academy of Sciences* **98**, 9181–9184.
- 48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 LOGUE, D. M., ABIOLA, I. O., RAINS, D., BAILEY, N. W., ZUK, M. & CADE, W. H. (2010).
4
5 2 Does signalling mitigate the cost of agonistic interactions? A test in a cricket that
6
7 3 has lost its song. *Proceedings of the Royal Society B* **277**, 2571–2575.
8
9
10 4 LÓPEZ, P., MARTIN, J. & CUADRADO, M. (2004). The role of lateral blue spots in
11
12 5 intrasexual relationships between male Iberian rock-lizards, *Lacerta monticola*.
13
14 6 *Ethology* **110**, 543–561.
15
16 7 LORENZ, K. (1966). *On Aggression*. Methuen & Co. Ltd. , London.
17
18 8 LOSOS, J. B. & CHU, L.-R. (1998). Examination of factors potentially affecting dewlap
19
20 9 size in Caribbean anoles. *Copeia* **1998**, 430–438.
21
22
23 10 LÖTTERS, S., CASTRO-HERRERA, F., KÖHLER, J. & RICHTER, R. (1997). Notes on the
24
25 11 distribution and color variation of poison frogs of the genus *Phylllobates* from
26
27 12 western Colombia (Anura: Dendrobatidae). *Revue Française d'Aquariologie* **24**,
28
29 13 55–58.
30
31
32 14 LÖTTERS, S., JUNGFER, K.-H., HENKEL, F. W. & SCHMIDT, W. (2007). *Poison frogs:*
33
34 15 *biology, species and captive husbandry*. Edition Chimaira.
35
36 16 LÜDDECKE, H., AMÉZQUITA, A., BERNAL, X. & GUZMAN, F. (2000). Partitioning of vocal
37
38 17 activity in a Neotropical highland-frog community. *Studies on Neotropical*
39
40 18 *Fauna and Environment* **35**, 185–194.
41
42
43 19 MAAN, M. E. & CUMMINGS, M. E. (2008). Female preferences for aposematic signal
44
45 20 components in a polymorphic poison frog. *Evolution* **62**, 2334–2345.
46
47 21 MAAN, M. E. & CUMMINGS, M. E. (2009). Sexual dimorphism and directional sexual
48
49 22 selection on aposematic signals in a poison frog. *Proceedings of the National*
50
51 23 *Academy of Sciences* **106**, 19072–19077.
52
53
54 24 MAAN, M. E. & CUMMINGS, M. E. (2012). Poison frog colors are honest signals of
55
56 25 toxicity, particularly for bird predators. *American Naturalist* **179**, E1–E14.
57
58
59
60

- 1
2
3 1 MAAN, M. E. & SEEHAUSEN, O. (2011). Ecology, sexual selection and speciation.
4
5 2 *Ecology Letters* **14**, 591–602.
6
7 3 MAAN, M. E., SEEHAUSEN, O., SODERBERG, L., JOHNSON, L., RIPMEESTER, E. A. P.,
8
9 4 MROSSO, H. D. J., TAYLOR, M. I., VAN DOOREN, T. J. M. & VAN ALPHEN, J. J. M.
10
11 5 (2004). Intraspecific sexual selection on a speciation trait, male coloration, in the
12
13 6 Lake Victoria cichlid *Pundamilia nyererei*. *Proceedings of the Royal Society of*
14
15 7 *London B* **271**, 2445–2452.
16
17 8 MAHER, C. R. & LOTT, D. F. (1995). Definitions of territoriality used in the study of
18
19 9 variation in vertebrate spacing systems. *Animal Behaviour* **49**, 1581–1597.
20
21 10 MALLET, J. & JORON, M. (1999). Evolution of diversity in warning color and mimicry:
22
23 11 Polymorphisms, shifting balance, and speciation. *Annual Review of Ecology and*
24
25 12 *Systematics* **30**, 201–233.
26
27 13 MAPPES, J. & ALATALO, R. V. (1997). Effects of novelty and gregariousness in survival
28
29 14 of aposematic prey. *Behavioral Ecology* **8**, 174–177.
30
31 15 MAPPES, J., MARPLES, N. & ENDLER, J. A. (2005). The complex business of survival by
32
33 16 aposematism. *Trends in Ecology & Evolution* **20**, 598–603.
34
35 17 MARTINS, M. (1989). Deimatic Behavior in *Pleurodema brachyops*. *Journal of*
36
37 18 *Herpetology* **23**, 305–307.
38
39 19 MASTER, T. L. (1999). Predation by Rufous Motmot on black-and-green poison dart
40
41 20 frog. *Wilson Bulletin* **111**, 439–440.
42
43 21 MAYNARD SMITH, J. (1982). *Evolution and the Theory of Games*. Cambridge University
44
45 22 Press, Cambridge, UK.
46
47 23 MAYNARD SMITH, J. & HARPER, D. G. C. (2005). *Animal Signals*. Oxford University
48
49 24 Press, Oxford, UK.
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 MEDINA, I., WANG, I. J., SALAZAR, C. & AMÉZQUITA, A. (2013). Hybridization promotes
4
5 2 color polymorphism in the aposematic harlequin poison frog, *Oophaga*
6
7 3 *histrionica*. *Ecology and Evolution* **3**, 4388–4400.
8
9
10 4 MELO, M. C., SALAZAR, C., JIGGINS, C. D. & LINARES, M. (2009). Assortative mating
11
12 5 preferences among hybrids offers a route to hybrid speciation. *Evolution* **63**,
13
14 6 1660–1665.
15
16 7 MEUCHE, I., LINSENMAIR, K. E. & PROEHL, H. (2011). Female Territoriality in the
17
18 8 Strawberry Poison Frog (*Oophaga pumilio*). *Copeia* **2011**, 351–356.
19
20 9 MILINSKI, M. & BAKKER, T. C. M. (1990). Female sticklebacks use male coloration in
21
22 10 mate choice and hence avoid parasitized males. *Nature* **344**, 330–333.
23
24 11 MOKKONEN, M. & LINDSTEDT, C. (2015). The evolutionary ecology of deception.
25
26 12 *Biological Reviews* DOI: 10.1111/brv.12208.
27
28 13 MOREY, S. R. (1990). Microhabitat selection and predation in the Pacific treefrog,
29
30 14 *Pseudacris regilla*. *Journal of Herpetology* **24**, 292–296.
31
32 15 MORIMOTO, G., YAMAGUCHI, N. & UEDA, K. (2005). Plumage color as a status signal in
33
34 16 male-male interaction in the red-flanked bushrobin, *Tarsiger cyanurus*. *Journal*
35
36 17 *of Ethology* **24**, 261–266.
37
38 18 MÜLLER, F. (1878). Ueber die Vortheile der Mimicry bei Schmetterlingen *Zoologischer*
39
40 19 *Anzeiger* **1**, 54–55.
41
42 20 MUNDAY, P. L., EYRE, P. J. & JONES, G. P. (2003). Ecological mechanisms for
43
44 21 coexistence of colour polymorphism in a coral-reef fish: an experimental
45
46 22 evaluation. *Oecologia* **137**, 519–526.
47
48 23 MYERS, C. W. & DALY, J. W. (1976). Preliminary evaluation of skin toxins and
49
50 24 vocalizations in taxonomic and evolutionary studies of poison-dart frogs
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 (Dendrobatidae). *Bulletin of the American Museum of Natural History* **157**,
4
5 2 177–262.
6
7 3 MYERS, C. W. & DALY, J. W. (1983). Dart-poison frogs. *Scientific American* **248**, 96–
8
9 4 105.
10
11 5 MYERS, C. W., DALY, J. W. & MALKIN, B. (1978). A dangerously toxic new frog
12
13 6 (*Phyllobates*) used by Emberá indians of western Colombia, with discussion of
14
15 7 blowgun fabrication and dart poisoning. *Bulletin of the American Museum of*
16
17 8 *Natural History* **161**, 309–365.
18
19 9 NARINS, P. M., HÖDL, W. & GRABUL, D. S. (2003). Bimodal signal requisite for agonistic
20
21 10 behavior in a dart-poison frog, *Epipedobates femoralis*. *Proceedings of the*
22
23 11 *National Academy of Sciences* **100**, 577–580.
24
25 12 NEVO, E. (1973). Adaptive color polymorphism in cricket frogs. *Evolution* **27**, 353–367.
26
27 13 NOKELAINEN, O., HEGNA, R. H., REUDLER, J. H., LINDSTEDT, C. & MAPPES, J. (2012).
28
29 14 Trade-off between warning signal efficacy and mating success in the wood tiger
30
31 15 moth. *Proceedings of the Royal Society B* **279**, 257–265.
32
33 16 NOKELAINEN, O., LINDSTEDT, C. & MAPPES, J. (2013). Environment-mediated morph-
34
35 17 linked immune and life-history responses in the aposematic wood tiger moth.
36
37 18 *Journal of Animal Ecology* **82**, 653–662.
38
39 19 NOKELAINEN, O., VALKONEN, J., LINDSTEDT, C. & MAPPES, J. (2014). Changes in
40
41 20 predator community structure shifts the efficacy of two warning signals in
42
43 21 Arctiid moths. *Journal of Animal Ecology* **83**, 598–605.
44
45 22 NOONAN, B. P. & COMEAULT, A. A. (2009). The role of predator selection on
46
47 23 polymorphic aposematic poison frogs. *Biology Letters* **5**, 51–54.
48
49 24 NOONAN, B. P. & GAUCHER, P. (2006). Refugial isolation and secondary contact in the
50
51 25 dyeing poison frog *Dendrobates tinctorius*. *Molecular Ecology* **15**, 4425–4435.
52
53
54
55
56
57
58
59
60

- 1
2
3 1 O'DONALD, P. & MAJERUS, M. E. N. (1984). Polymorphism of melanic ladybirds
4
5 2 maintained by frequency-dependent sexual selection. *Biological Journal of the*
6
7 3 *Linnean Society* **23**, 101–111.
8
9
10 4 OLSSON, M. (1994). Nuptial coloration in the sand lizard, *Lacerta agilis*: an intra-
11
12 5 sexually selected cue to fighting ability. *Animal Behaviour* **48**, 607–613.
13
14 6 ORD, T. J. & EVANS, C. S. (2003). Display rate and opponent assessment in the Jacky
15
16 7 dragon (*Amphibolurus muricatus*): an experimental analysis. *Behaviour* **140**,
17
18 8 1495–1508.
19
20 9 OSORIO, D. & SRINIVASAN, M. V. (1991). Camouflage by edge enhancement in animal
21
22 10 coloration patterns and its implications for visual mechanisms. *Proceedings of*
23
24 11 *the Royal Society B* **244**, 81–85.
25
26
27 12 PALUH, D. J., HANTAK, M. M. & SAPORITO, R. A. (2014). A test of aposematism in the
28
29 13 dendrobatid poison frog *Oophaga pumilio*: the importance of movement in clay
30
31 14 model experiments. *Journal of Herpetology* **48**, 249–254.
32
33
34 15 PELLISSIER, L., WASSEF, J., BILAT, J., BRAZZOLA, G., BURI, P., COLLIARD, C., FOURNIER,
35
36 16 B., HAUSSER, J., YANNIC, G. & PERRIN, N. (2011). Adaptive colour
37
38 17 polymorphism of *Acrida ungarica* H. (Orthoptera: Acrididae) in a spatially
39
40 18 heterogeneous environment. *Acta Oecologica-International Journal of Ecology*
41
42 19 **37**, 93–98.
43
44
45 20 PETRIE, M., HALLIDAY, T. & SANDERS, C. (1991). Peahens prefer peacocks with
46
47 21 elaborate trains. *Animal Behaviour* **41**, 323–331.
48
49
50 22 PINCEMY, G., DOBSON, F. S. & JOUVENTIN, P. (2009). Experiments on colour ornaments
51
52 23 and mate choice in king penguins. *Animal Behaviour* **78**, 1247–1253.
53
54 24 POUGH, F. H., ANDREWS, R. M., CADLE, J. E., CRUMP, M. L., SAVITZKY, A. L. & WELLS,
55
56 25 K. D. (2001). *Herpetology*. Prentice Hall, Upper Saddle River, New Jersey.
57
58
59
60

- 1
2
3 1 POULTON, E. B. (1890). *The Colours of Animals: Their Meaning and Use*. Kegan Paul,
4
5 2 Trench, Trubner, London.
6
7 3 PRATES, I., ANTONIAZZI, M. M., SCIANI, J. M., PIMENTA, D. C., TOLEDO, L. F., HADDAD,
8
9 4 C. F. B. & JARED, C. (2012). Skin glands, poison and mimicry in dendrobatid and
10
11 5 leptodactylid amphibians. *Journal of Morphology* **273**, 279–290.
12
13 6 PRÖHL, H. (2005). Territorial behavior in dendrobatid frogs. *Journal of Herpetology* **39**,
14
15 7 354–365.
16
17 8 PRÖHL, H. & OSTROWSKI, T. (2011). Behavioural elements reflect phenotypic colour
18
19 9 divergence in a poison frog. *Evolutionary Ecology* **25**, 993–1015.
20
21 10 PRYKE, S. R. & ANDERSSON, S. (2003a). Carotenoid-based epaulettes reveal male
22
23 11 competitive ability: experiments with resident and floater red-shouldered
24
25 12 widowbirds. *Animal Behaviour* **66**, 217–224.
26
27 13 PRYKE, S. R. & ANDERSSON, S. (2003b). Carotenoid-based status signalling in red-
28
29 14 shouldered widowbirds (*Euplectes axillaris*): epaulet size and redness affect
30
31 15 captive and territorial competition. *Behavioral Ecology and Sociobiology* **53**,
32
33 16 393–401.
34
35 17 PRYKE, S. R. & GRIFFITH, S. C. (2006). Red dominates black: agonistic signalling among
36
37 18 head morphs in the colour polymorphic Gouldian finch. *Proceedings of the*
38
39 19 *Royal Society B* **273**, 949–957.
40
41 20 RENOULT, J. P., KELBER, A. & SCHAEFER, H. M. (2015). Colour spaces in ecology and
42
43 21 evolutionary biology. *Biological Reviews*, DOI: 10.1111/brv.12230.
44
45 22 RESNICK, L. E. & JAMESON, D. L. (1963). Color polymorphism in Pacific tree frogs.
46
47 23 *Science* **142**, 1081–&.
48
49 24 REYNOLDS, R. G. & FITZPATRICK, B. M. (2007). Assortative mating in poison-dart frogs
50
51 25 based on an ecologically important trait. *Evolution* **61**, 2253–2259.
52
53
54
55
56
57
58
59
60

- 1
2
3 1 RICHARDS-ZAWACKI, C. L., WANG, I. J. & SUMMERS, K. (2012). Mate choice and the
4
5 2 genetic basis for colour variation in a polymorphic dart frog: inferences from a
6
7 3 wild pedigree. *Molecular Ecology* **21**, 3879–3892.
8
9
10 4 RICHARDS-ZAWACKI, C. L., YEAGER, J. & BART, H. P. S. (2013). No evidence for
11
12 5 differential survival or predation between sympatric color morphs of an
13
14 6 aposematic poison frog. *Evolutionary Ecology* **27**, 783–795.
15
16 7 RIECHERT, S. E. (1998). Game theory and animal contests. In *Game Theory and Animal*
17
18 8 *Behavior*. (ed. L. A. R. H. K. Dugatkin), pp. 64–93. Oxford University Press,
19
20 9 Oxford.
21
22
23 10 RIIPI, M., ALATALO, R. V., LINDSTROM, L. & MAPPES, J. (2001). Multiple benefits of
24
25 11 gregariousness cover detectability costs in aposematic aggregations. *Nature* **413**,
26
27 12 512–514.
28
29
30 13 RINGLER, M., URSPRUNG, E. & HÖDL, W. (2010). Predation on *Allobates femoralis*
31
32 14 (Boulenger 1884; Anura: Aromobatidae) by the colubrid snake *Xenopholis*
33
34 15 *scalaris* (Wucherer 1861). *Herpetology Notes* **3**, 301–304.
35
36 16 ROJAS, B., DEVILLECHABROLLE, J. & ENDLER, J. A. (2014a). Paradox lost: variable
37
38 17 colour-pattern geometry is associated with differences in movement in
39
40 18 aposematic frogs. *Biology Letters* **10**, 20140193.
41
42
43 19 ROJAS, B. & ENDLER, J. A. (2013). Sexual dimorphism and intra-populational colour
44
45 20 pattern variation in the aposematic frog *Dendrobates tinctorius*. *Evolutionary*
46
47 21 *Ecology* **27**, 739–753.
48
49
50 22 ROJAS, B., GORDON, S. P. & MAPPES, J. (2015a). Frequency-dependent flight activity in
51
52 23 the colour polymorphic wood tiger moth. *Current Zoology* **61**, 765–772.
53
54
55
56
57
58
59
60

- 1
2
3 1 ROJAS, B., RAUTIALA, P. & MAPPE, J. (2014b). Differential detectability of polymorphic
4
5 2 warning signals under varying light environments *Behavioural Processes* **109**,
6
7 3 164–172.
8
9
10 4 ROJAS, B., VALKONEN, J. K. & NOKELAINEN, O. (2015b). Aposematism. *Current Biology*
11
12 5 **25**, R350–R351.
13
14 6 RUDH, A., BREED, M. F. & QVARNSTROM, A. (2013). Does aggression and explorative
15
16 7 behaviour decrease with lost warning coloration? *Biological Journal of the*
17
18 8 *Linnean Society* **108**, 116–126.
19
20 9 RUDH, A. & QVARNSTRÖM, A. (2013). Adaptive colouration in Amphibians. *Seminars in*
21
22 10 *Cell & Developmental Biology* **24**, 553–561.
23
24 11 RUXTON, G. D., SHERRATT, T. N. & SPEED, M. P. (2004). *Avoiding Attack: the*
25
26 12 *evolutionary ecology of crypsis, warning signals and mimicry*. Oxford University
27
28 13 Press, Oxford.
29
30 14 SANTOS, J. C. & CANNATELLA, D. C. (2011). Phenotypic integration emerges from
31
32 15 aposematism and scale in poison frogs. *Proceedings of the National Academy of*
33
34 16 *Sciences* **108**, 6175–6180.
35
36 17 SANTOS, J. C., COLOMA, L. A. & CANNATELLA, D. C. (2003). Multiple, recurring origins
37
38 18 of aposematism and diet specialization in poison frogs. *Proceedings of the*
39
40 19 *National Academy of Sciences* **100**, 12792–12797.
41
42 20 SANTOS, R. R. & GRANT, T. (2011). Diel pattern of migration in a poisonous toad from
43
44 21 Brazil and the evolution of chemical defenses in diurnal amphibians.
45
46 22 *Evolutionary Ecology* **25**, 249–258.
47
48 23 SAPORITO, R. A., DONNELLY, M. A., NORTON, R. A., GARRAFFO, H. M., SPANDE, T. F. &
49
50 24 DALY, J. W. (2007a). Oribatid mites as a major dietary source for alkaloids in
51
52 25 poison frogs. *Proceedings of the National Academy of Sciences* **104**, 8885–8890.
53
54
55
56
57
58
59
60

- 1
2
3 1 SAPORITO, R. A., DONNELLY, M. A., SPANDE, T. F. & GARRAFFO, H. M. (2012). A review
4
5 2 of chemical ecology in poison frogs. *Chemoecology* **22**, 159–168.
6
7 3 SAPORITO, R. A., GARRAFFO, H. M., DONNELLY, M. A., EDWARDS, A. L., LONGINO, J. T.
8
9 4 & DALY, J. W. (2004). Formicine ants: An arthropod source for the pumiliotoxin
10
11 5 alkaloids of dendrobatid poison frogs. *Proceedings of the National Academy of*
12
13 6 *Sciences* **101**, 8045–8050.
14
15
16 7 SAPORITO, R. A., ZUERCHER, R., ROBERTS, M., GEROW, K. G. & DONNELLY, M. A.
17
18 8 (2007b). Experimental evidence for aposematism in the dendrobatid poison frog
19
20 9 *Oophaga pumilio*. *Copeia* **2007**, 1006–1011.
21
22
23 10 SAVIOLA, A. J., MCKENZIE, V. J. & CHISZAR, D. (2012). Chemosensory responses to
24
25 11 chemical and visual stimuli in five species of colubrid snakes. *Acta*
26
27 12 *Herpetologica* **7**, 91–103.
28
29
30 13 SCHAEFER, H. C., VENCES, M. & VEITH, M. (2002). Molecular phylogeny of Malagasy
31
32 14 poison frogs, genus *Mantella* (Anura : Mantellidae): homoplastic evolution of
33
34 15 colour pattern in aposematic amphibians. *Organisms Diversity & Evolution* **2**,
35
36 16 97–105.
37
38
39 17 SCHAEFER, H. M. (2010). Visual communication: evolution, ecology, and functional
40
41 18 mechanisms. In *Animal Behaviour: Evolution and Mechanisms* (ed. P.
42
43 19 Kappeler), pp. 3–28.
44
45
46 20 SEBBEN, A., SCHWARTZ, C. A., VALENTE, D. & MENDES, E. G. (1986). A tetrodotoxin-
47
48 21 like substance found in the Brazilian frog *Brachycephalus ephippium*. *Toxicon*
49
50 22 **24**, 799–806.
51
52
53 23 SEEHAUSEN, O. & VAN ALPHEN, J. J. M. (1998). The effect of male coloration on female
54
55 24 mate choice in closely related Lake Victoria cichlids (*Haplochromis nyererei*
56
57 25 complex). *Behavioral Ecology and Sociobiology* **42**, 1–8.
58
59
60

- 1
2
3 1 SHELDON, B. C., ARPONEN, H., LAURILA, A., CROCHET, P. A. & MERILA, J. (2003). Sire
4
5 2 coloration influences offspring survival under predation risk in the moorfrog.
6
7 3 *Journal of Evolutionary Biology* **16**, 1288–1295.
8
9
10 4 SHERRATT, T. N. (2002). The coevolution of warning signals. *Proceedings of the Royal*
11
12 5 *Society of London B* **269**, 741–746.
13
14 6 SIDDIQI, A., CRONIN, T. W., LOEW, E. R., VOROBYEV, M. & SUMMERS, K. (2004).
15
16 7 Interspecific and intraspecific views of color signals in the strawberry poison
17
18 8 frog *Dendrobates pumilio*. *Journal of Experimental Biology* **207**, 2471–2485.
19
20 9 SILLÉN-TULLBERG, B. (1988). Evolution of gregariousness in aposematic butterfly
21
22 10 larvae: A phylogenetic analysis. *Evolution* **42**, 293–305.
23
24 11 SILVERSTONE, P. A. (1975). A revision of the poison-arrow frogs of the genus
25
26 12 *Dendrobates* Wagler. *Natural History Museum of Los Angeles County Science*
27
28 13 *Bulletin* **21**, 1–55.
29
30 14 SINERVO, B. & SVENSSON, E. (2002). Correlational selection and the evolution of
31
32 15 genomic architecture. *Heredity* **89**, 329–338.
33
34 16 SKELHORN, J., ROWLAND, H. M., SPEED, M. P. & RUXTON, G. D. (2010). Masquerade:
35
36 17 Camouflage Without Crypsis. *Science* **327**, 51.
37
38 18 SOMMER, V. (1987). Infanticide among free-ranging langurs (*Presbytis entellus*) at
39
40 19 Jodhpur (Rajasthan India) - Recent observations and a reconsideration of
41
42 20 hypotheses. *Primates* **28**, 163–197.
43
44 21 SPEED, M. P. (2000). Warning signals, receiver psychology and predator memory.
45
46 22 *Animal Behaviour* **60**, 269–278.
47
48 23 STANGE, N. & RONACHER, B. (2012). Grasshopper calling songs convey information
49
50 24 about condition and health of males. *Journal of Comparative Physiology A* **198**,
51
52 25 309–318.
53
54
55
56
57
58
59
60

- 1
2
3 1 STARNBERGER, I., PREININGER, D. & HÖDL, W. (2014). From uni- to multimodality:
4
5 2 towards an integrative view on anuran communication. *Journal of Comparative*
6
7 3 *Physiology A* **200**, 777–787.
8
9
10 4 STEVENS, M. & MERILAITA, S. (2009). Animal camouflage: current issues and new
11
12 5 perspectives. *Philosophical Transactions of the Royal Society B* **364**, 423–427.
13
14 6 STUART, Y. E., DAPPEN, N. & LOSIN, N. (2012). Inferring predator behavior from attack
15
16 7 rates on prey-replicas that differ in conspicuousness. *PLoS One* **7**, e40497.
17
18 8 STUART-FOX, D. M. & JOHNSTON, G. R. (2005). Experience overrides colour in lizard
19
20 9 contests. *Behaviour* **142**, 329–350.
21
22
23 10 STUCKERT, A. M. M., SAPORITO, R. A., VENEGAS, P. J. & SUMMERS, K. (2014a).
24
25 11 Alkaloid defenses of co-mimics in a putative Mullerian mimetic radiation. *BMC*
26
27 12 *Evolutionary Biology* **14**, 76.
28
29
30 13 STUCKERT, A. M. M., VENEGAS, P. J. & SUMMERS, K. (2014b). Experimental evidence
31
32 14 for predator learning and Müllerian mimicry in Peruvian poison frogs
33
34 15 (*Ranitomeya*, Dendrobatidae). *Evolutionary Ecology* **28**, 413–426.
35
36
37 16 STYNOSKI, J. L., SHELTON, G. & STYNOSKI, P. (2014). Maternally derived chemical
38
39 17 defences are an effective deterrent against some predators of poison frog
40
41 18 tadpoles (*Oophaga pumilio*). *Biology Letters* **10**, 20140187.
42
43
44 19 SUMMERS, K. (1989). Sexual selection and intra-female competition in the green poison-
45
46 20 dart frog, *Dendrobates auratus*. *Animal Behaviour* **37**, 797–805.
47
48 21 SUMMERS, K. (1992). Mating strategies in two species of dart-poison frogs: a
49
50 22 comparative study. *Animal Behaviour* **43**, 907–919.
51
52
53 23 SUMMERS, K. (2003). Convergent evolution of bright coloration and toxicity in frogs.
54
55 24 *Proceedings of the National Academy of Sciences* **100**, 12533–12534.
56
57
58
59
60

- 1
2
3 1 SUMMERS, K., BERMINGHAM, E., WEIGT, L. & MCCAFFERTY, S. (1997). Phenotypic and
4
5 2 genetic divergence in three species of dart-poison frogs with contrasting parental
6
7 3 behavior. *Journal of Heredity* **88**, 8–13.
8
9
10 4 SUMMERS, K. & CLOUGH, M. E. (2001). The evolution of coloration and toxicity in the
11
12 5 poison frog family (Dendrobatidae). *Proceedings of the National Academy of*
13
14 6 *Sciences* **98**, 6227–6232.
15
16 7 SUMMERS, K., SPEED, M. P., BLOUNT, J. D. & STUCKERT, A. M. M. (2015). Are
17
18 8 aposematic signals honest? A review. *Journal of Evolutionary Biology* **28**, 1583–
19
20 9 1599.
21
22
23 10 SUMMERS, K., SYMULA, R., CLOUGH, M. & CRONIN, T. (1999). Visual mate choice in
24
25 11 poison frogs. *Proceedings of the Royal Society of London B* **266**, 2141–2145.
26
27 12 SYMULA, R., SCHULTE, R. & SUMMERS, K. (2001). Molecular phylogenetic evidence for
28
29 13 a mimetic radiation in Peruvian poison frogs supports a Mullerian mimicry
30
31 14 hypothesis. *Proceedings of the Royal Society of London B* **268**, 2415–2421.
32
33 15 SZELISTOWSKI, W. A. (1985). Unpalatability of the poison arrow frog *Dendrobates*
34
35 16 *pumilio* to the ctenid spider *Cupiennius coccineus*. *Biotropica* **17**, 345–346.
36
37 17 SZTATECSNY, M., PREININGER, D., FREUDMANN, A., LORETTO, M.-C., MAIER, F. &
38
39 18 HÖDL, W. (2012). Don't get the blues: conspicuous nuptial colouration of male
40
41 19 moor frogs (*Rana arvalis*) supports visual mate recognition during scramble
42
43 20 competition in large breeding aggregations. *Behavioral Ecology and*
44
45 21 *Sociobiology* **66**, 1587–1593.
46
47 22 SZTATECSNY, M., STRONDL, C., BAIERL, A., RIES, C. & HÖDL, W. (2010). Chin up: are
48
49 23 the bright throats of male common frogs a condition-independent visual cue?
50
51 24 *Animal Behaviour* **79**, 779–786.
52
53
54
55
56
57
58
59
60

- 1
2
3 1 STYNOSKI, J. L., SCHULTE, L. M. & ROJAS, B. 2015. Poison frogs. *Current Biology* **25**,
4 R1026–R1028.
5
6
7 2
8 3 TAYLOR, R. C., KLEIN, B. A., STEIN, J. & RYAN, M. J. (2011). Multimodal signal
9 variation in space and time: how important is matching a signal with its signaler?
10 *Journal of Experimental Biology* **214**, 815–820.
11
12 5
13 6 TERAJ, Y. & OKADA, N. (2011). Speciation of Cichlid Fishes by Sensory Drive. In *From*
14 *Genes to Animal Behavior: Social Structures, Personalities, Communication by*
15 *Color. Primatology Monographs* (ed. M. K. S. W. A. InoueMurayama), pp. 311–
16 328.
17
18
19
20
21
22
23 10 THAYER, G. H. (1909). *Concealing Coloration in the Animal Kingdom. An Exposition of*
24 *the Laws of Disguise through Color and Pattern; Being a Summary of Abbott H.*
25 *Thayer's Discoveries* Macmillan, New York.
26
27
28
29
30 13 THÉRY, M. (2001). Forest light and its influence on habitat selection. *Plant Ecology* **153**,
31 251–261.
32
33
34 15 THÉRY, M. & ENDLER, J. A. (2001). Habitat selection, ambient light and colour patterns
35 in some lek-displaying birds. In *Nouragues: dynamics and plant-animal*
36 *interactions in a neotropical rainforest.*, vol. 80 (ed. F. Bongers, P. Charles-
37 Dominique, P.-M. Forget and M. Théry), pp. 161–166. Kluwer Academic
38 Publisher, Dordrecht, The Netherlands.
39
40
41
42
43
44
45 20 THOMPSON, V. (1984). Polymorphism under apostatic and aposematic selection.
46 *Heredity* **53**, 677–686.
47
48
49 22 TOFT, C. A. (1995). Evolution of diet specialization in poison-dart frogs
50 (Dendrobatidae). *Herpetologica* **51**, 202–216.
51
52
53
54 24 TOLEDO, L. F. & HADDAD, C. F. B. (2009). Colors and some morphological traits as
55 defensive mechanisms in anurans. *International Journal of Zoology* **2009**, 1–12.
56
57
58
59
60

- 1
2
3 1 TORDOFF, W. (1980). Selective predation of gray jays, *Perisoreus canadensis*, upon
4
5 2 boreal chorus frogs, *Pseudacris triseriata*. *Evolution* **34**, 1004–1008.
6
7 3 TWOMEY, E., VESTERGAARD, J. S. & SUMMERS, K. (2014). Reproductive isolation related
8
9 4 to mimetic divergence in the poison frog *Ranitomeya imitator*. *Nature*
10
11 5 *Communications* **5**, 4749.
12
13 6 TWOMEY, E., YEAGER, J., BROWN, J. L., MORALES, V., CUMMINGS, M. & SUMMERS, K.
14
15 7 (2013). Phenotypic and genetic divergence among poison frog populations in a
16
17 8 mimetic radiation. *PLoS One* **8**, e55443.
18
19 9 UENO, H., SATO, Y. & TSUCHIDA, K. (1998). Colour-associated mating success in a
20
21 10 polymorphic Ladybird Beetle, *Harmonia axyridis*. *Functional Ecology* **12**, 757–
22
23 11 761.
24
25 12 UMBERS, K. D. L., LEHTONEN, J. & MAPPES, J. (2015). Deimatic displays. *Current*
26
27 13 *Biology* **25**, R58–R59.
28
29 14 VALKONEN, J., NISKANEN, M., BJORKLUND, M. & MAPPES, J. (2011). Disruption or
30
31 15 aposematism? Significance of dorsal zigzag pattern of European vipers.
32
33 16 *Evolutionary Ecology* **25**, 1047–1063.
34
35 17 VALKONEN, J. K., NOKELAINEN, O., NISKANEN, M., KILPIMAA, J., BJORKLUND, M. &
36
37 18 MAPPES, J. (2012). Variation in predator species abundance can cause variable
38
39 19 selection pressure on warning signaling prey. *Ecology and Evolution* **2**, 1971–
40
41 20 1976.
42
43 21 VASQUEZ, T. & PFENNIG, K. S. (2007). Looking on the bright side: females prefer
44
45 22 coloration indicative of male size and condition in the sexually dichromatic
46
47 23 spadefoot toad, *Scaphiopus couchii*. *Behavioral Ecology and Sociobiology* **62**,
48
49 24 127–135.
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 1 VEITH, M., KOSUCH, J., ROEDEL, M.-O., HILLERS, A., SCHMITZ, A., BURGER, M. &
4
5 2 LÖTTERS, S. (2009). Multiple evolution of sexual dichromatism in African reed
6
7 3 frogs. *Molecular Phylogenetics and Evolution* **51**, 388–393.
8
9
10 4 VENCES, M., KOSUCH, J., BOISTEL, R., HADDAD, C. F. B., LA MARCA, E., LOTTTERS, S. &
11
12 5 VEITH, M. (2003). Convergent evolution of aposematic coloration in Neotropical
13
14 6 poison frogs: a molecular phylogenetic perspective. *Organisms Diversity &*
15
16 7 *Evolution* **3**, 215–226.
17
18 8 VOROBYEV, M. & OSORIO, D. (1998). Receptor noise as a determinant of colour
19
20 9 thresholds. *Proceedings of the Royal Society B* **265**, 351–358.
21
22
23 10 VOROBYEV, M., OSORIO, D., BENNETT, A. T. D., MARSHALL, N. J. & CUTHILL, I. C.
24
25 11 (1998). Tetrachromacy, oil droplets and bird plumage colours. *Journal of*
26
27 12 *Comparative Physiology A–Neuroethology Sensory Neural and Behavioral*
28
29 13 *Physiology* **183**, 621–633.
30
31
32 14 WALLACE, A. R. (1877). The colours of animals and plants. *American Naturalist* **11**,
33
34 15 641–662.
35
36 16 WANG, I. J. (2011). Inversely related aposematic traits: reduced conspicuousness evolves
37
38 17 with increased toxicity in a polymorphic poison-dart frog. *Evolution* **65**, 1637–
39
40 18 1649.
41
42
43 19 WELLS, K. D. (1978). Courtship and parental behavior in a Panamanian poison-arrow
44
45 20 frog (*Dendrobates auratus*). *Herpetologica* **34**, 148–155.
46
47 21 WELLS, K. D. (1980). Social behavior and communication of a dendrobatid frog
48
49 22 (*Colostethus trinitatis*). *Herpetologica* **36**, 189–199.
50
51
52 23 WELLS, K. D. (2007). *The Ecology and Behavior of Amphibians*. University of Chicago
53
54 24 Press, Chicago.
55
56
57
58
59
60

- 1 WENTE, W. H. & PHILLIPS, J. B. (2005). Microhabitat selection by the Pacific treefrog,
2 *Hyla regilla*. *Animal Behaviour* **70**, 279–287.
- 3 WHITING, M. J., STUART-FOX, D. M., O'CONNOR, D., FIRTH, D., BENNETT, N. C. &
4 BLOMBERG, S. P. (2006). Ultraviolet signals ultra-aggression in a lizard. *Animal*
5 *Behaviour* **72**, 353–363.
- 6 WILEY, R. H. (1983). The evolution of communication: information and manipulation.
7 In *Animal Behaviour Vol. 2: Communication* (ed. T. R. Halliday and P. B. J.
8 Slater), pp. 156–189. Blackwell Scientific Publications, Oxford.
- 9 WILEY, R. H. (1994). Errors, exaggeration, and deception in animal communication. In
10 *Behavioral Mechanisms in Ecology* (ed. L. A. Real), pp. 157–189. University of
11 Chicago Press, Chicago.
- 12 WILLINK, B., BRENES-MORA, E., BOLAÑOS, F. & PRÖHL, H. (2013). Not everything is
13 black and white: color and behavioral variation reveal a continuum between
14 cryptic and aposematic strategies in a polymorphic poison frog. *Evolution* **67**,
15 2783–2794.
- 16 WOLLENBERG, K. C., LÖTTERS, S., MORA-FERRER, C. & VEITH, M. (2008). Disentangling
17 composite colour patterns in a poison frog species. *Biological Journal of the*
18 *Linnean Society* **93**, 433–444.
- 19 WOOLBRIGHT, L. L. & STEWART, M. M. (2008). Spatial and temporal variation in color
20 pattern morphology in the tropical frog, *Eleutherodactylus coqui*. *Copeia*, 431–
21 437.
- 22 YEAGER, J., BROWN, J. L., MORALES, V., CUMMINGS, M. & SUMMERS, K. (2012). Testing
23 for selection on color and pattern in a mimetic radiation. *Current Zoology* **58**,
24 668–676.

- 1
2
3 1 ZAHAVI, A. (1975). Mate selection—a selection for a handicap. *Journal of Theoretical*
4
5 2 *Biology* **53**, 205–214.
6
7 3 ZIMMERMANN, H. & ZIMMERMANN, E. (1988). Etho-Taxonomie und zoogeographische
8
9 4 Artengruppenbildung bei Pfeilgiftfröschen (Anura: Dendrobatidae). *Salamandra*
10
11 5 **24**, 125–160.
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
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2 Figure legends

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4 **Fig. 1.** An example of background matching in a young *Pristimantis zeuctotylus* (A);
5 and examples of masquerading, where a female *Rhinella margaritifera* is shown to
6 resemble a dead leaf (B–D). Images C and D are for the same individual photographed
7 at different distances. Photo credits: Bibiana Rojas.

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9 **Fig. 2.** Aposematic frogs (A) *Mantella baroni*; (B) *Mantella aurantiaca*; (C) *Dyscophus*
10 *guineti*, known as the ‘Tomato frog’; (D) *Pseudophryne corroboree*; (E)
11 *Brachycephalus ephippium*; and (F) *Melanophryniscus rubriventris*. Photo credits: A–C,
12 Gerardo García; D, J.P. Lawrence; E, Taran Grant; F, Marcos Vaira.

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14 **Fig. 3.** *Atelopus* aff. *franciscus* (A) dorsal and (B) ventral colouration. (C) Geographic
15 variation in the dorsal and ventral colouration of *Melanophryniscus rubriventris*. Photo
16 credits: A,B, Bibiana Rojas; C, Marcos Vaira.

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18 **Fig. 4.** Dart poison frogs represent the best-known example of aposematism among
19 anurans. (A) *Adelphobates galactonotus*; (B) *Ameerega bilinguis*; (C) *Dendrobates*
20 *truncatus*; (D) *Dendrobates tinctorius*; (E) *Phyllobates terribilis*; (F) *Dendrobates*
21 *auratus*; (G) *Oophaga granulifera*; (H) *Phyllobates lugubris* and (I) *Ranitomeya*
22 *imitator*. Photo credits: A,C, Taran Grant; B,F,H,I, J.P. Lawrence; D,G, Bibiana Rojas;
23 E, Roberto Márquez.

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3 **Fig. 5.** Known predators of dendrobatid frogs: (A) the snake *Rhadinaea decorata*,
4 feeding on *Oophaga pumilio*; (B) wolf spider (Lycosidae) preying on *Ameerega*
5 *trivittata*; (C) crab holding an individual *Oophaga histrionica*; (D) rufous motmot
6 (*Baryphthengus martii*) taking an *O. pumilio* to its nest. Photo credits: A, Matthew
7 Dugas; B, Trond Larsen; C, Alejandro Vélez; D, Ralph Saporito.

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10 **Fig. 6.** Examples of mimicry among poison frog species. (A) *Ameerega hahneli* (left) is
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12 been suggested to be a Müllerian mimic of *Ameerega picta* (B). The best-known case of
13 a Müllerian mimicry system is that of *Ranitomeya imitator*. D–G are the model species:
14 *Ranitomeya variabilis*, highland morph; *R. summersi*, *R. fantastica*, *R. variabilis*
15 lowland morph. The different morphs of *R. imitator* are shown below: (H) spotted; (I)
16 banded; (J) varadero; and (K) striped. Photo credits: A, Jason L. Brown; B, Mauricio
17 Pacheco; C, Quentin Martínez; D–K Evan Twomey.

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20 **Fig. 7.** Examples of geographic variation in the colour patterns of *Oophaga pumilio* (A–
21 D); and intrapopulation variation in *Dendrobates tinctorius* (E–H). Photo credits: A–D,
22 J.P. Lawrence; E–H, Bibiana Rojas.

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25 the lower dorsum of *Edalorhina perezii* (B); and (C–E) the ‘unken reflex’ performed by
species of the genus *Melanophryniscus*, even in amplexus (D). All have been suggested
to function as deimatic displays. Photo credits: A,B, J.P. Lawrence; C,D, Marcos Vaira;
E, Taran Grant.

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3 **Fig. 9.** Breeding aggregation of *Rana temporaria* (A) and a pair of the moor frog (*Rana*
4
5 *arvalis*) in amplexus (B). Note the bright throats and blue colouration of the males,
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8 which are thought to function as sex-recognition cues to prevent mismatings. Photo
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10 credits: Marc Sztatecsny.
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Fig. 1. An example of background matching in a young *Pristimantis zeuctotylus* (A); and examples of masquerading, where a female *Rhinella margaritifera* is shown to resemble a dead leaf (B–D). Images C and D are for the same individual photographed at different distances. Photo credits: Bibiana Rojas. 80x60mm (300 x 300 DPI)

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Fig. 2. Aposematic frogs (A) *Mantella baroni*; (B) *Mantella aurantiaca*; (C) *Discophus guineti*, known as the 'Tomato frog'; (D) *Pseudophryne corroborae*; (E) *Brachycephalus ephippium*; and (F) *Melanophryniscus rubriventris*. Photo credits: A–C, Gerardo García; D, J.P. Lawrence; E, Taran Grant; F, Marcos Vaira.
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Fig. 3. *Atelopopus* aff. *franciscus* (A) dorsal and (B) ventral colouration. (C) Geographic variation in the dorsal and ventral colouration of *Melanophryniscus rubriventris*. Photo credits: A,B, Bibiana Rojas; C, Marcos Vaira. 85x77mm (300 x 300 DPI)





Fig. 4. Dart poison frogs represent the best-known example of aposematism among anurans. (A) *Adelphobates galactonotus*; (B) *Ameerega bilinguis*; (C) *Dendrobates truncatus*; (D) *Dendrobates tinctorius*; (E) *Phyllobates terribilis*; (F) *Dendrobates auratus*; (G) *Oophaga granulifera*; (H) *Phyllobates lugubris* and (I) *Ranitomeya imitator*. Photo credits: A,C, Taran Grant; B,F,H,I, J.P. Lawrence; D,G, Bibiana Rojas; E, Roberto Márquez.
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Fig. 5. Known predators of dendrobatid frogs: (A) the snake *Rhadinæa decorata*, feeding on *O. pumilio*; (B) wolf spider (*Lycosidae*) preying on *Ameerega trivittata*; (C) crab holding an individual *Oophaga histrionica*; (D) rufous motmot (*Baryphthengus martii*) taking an *O. pumilio* to its nest. Photo credits: A, Matthew Dugas; B, Trond Larsen; C, Alejandro Vélez; D, Ralph Saporito.
104x81mm (300 x 300 DPI)



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107x59mm (300 x 300 DPI)



Fig. 7. Examples of geographic variation in the colour patterns of *Oophaga pumilio* (A–D); and intrapopulation variation in *Dendrobates tinctorius* (E–H). Photo credits: A–D, J.P. Lawrence; E–H, Bibiana Rojas.

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Fig. 9. Breeding aggregation of *Rana temporaria* (A) and a pair of the moor frog (*Rana arvalis*) in amplexus (B). Note the bright throats and blue colouration of the males, which are thought to function as sex-recognition cues to prevent mismatings. Photo credits: Marc Sztatecsny. 119x43mm (300 x 300 DPI)

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