

Review

Bench-to-bedside review: Endothelial cell dysfunction in severe sepsis: a role in organ dysfunction?

Benoît Vallet

Professor, Department of Anesthesiology and Intensive Care and Department of Pharmacology, University Hospital, Lille, France

Correspondence: Benoit Vallet, bvallet@chru-lille.fr

Published online: 6 January 2003

Critical Care 2003, **7**:130-138 (DOI 10.1186/cc1864)

This article is online at <http://ccforum.com/content/7/2/130>

© 2003 BioMed Central Ltd (Print ISSN 1364-8535; Online ISSN 1466-609X)

Abstract

During the past decade a unifying hypothesis has been developed to explain the vascular changes that occur in septic shock on the basis of the effect of inflammatory mediators on the vascular endothelium. The vascular endothelium plays a central role in the control of microvascular flow, and it has been proposed that widespread vascular endothelial activation, dysfunction and eventually injury occurs in septic shock, ultimately resulting in multiorgan failure. This has been characterized in various models of experimental septic shock. Now, direct and indirect evidence for endothelial cell alteration in humans during septic shock is emerging. The present review details recently published literature on this rapidly evolving topic.

Keywords coagulation, endothelial cell, monocyte, sepsis, shock, tissue factor, tissue oxygenation, tissue perfusion, vascular reactivity

The vascular endothelium regulates the flow of nutrient substances, diverse biologically active molecules and the blood cells themselves. This role of endothelium is achieved through the presence of membrane-bound receptors for numerous molecules, including proteins, lipid transporting particles, metabolites and hormones, as well as through specific junction proteins and receptors that govern cell-cell and cell-matrix interactions [1,2]. Endothelial dysfunction and/or injury with subendothelium exposure facilitates leucocyte and platelet aggregation, and aggravation of coagulopathy. Therefore, endothelial dysfunction and/or injury should favour impaired perfusion, tissue hypoxia and subsequent organ dysfunction.

The present review describes, within the context of sepsis, why altered endothelial properties may be suspected to be involved in organ failure (Table 1).

Endothelial injury

Endothelial injury describes a state in which microscopically visible endothelial cell (EC) shape change or injury can be identified, as well as defects in endothelial lining or elevated

soluble markers of endothelial injury [3]. Anatomical damage to the endothelium during septic shock has been assessed in several studies [4-6]. A single injection of bacterial lipopolysaccharide (LPS) has long been demonstrated to be a nonmechanical technique for removing endothelium [5]. In endotoxic rabbits, observations tend to demonstrate that EC surface modification occurs easily and rapidly [5,6], with ECs being detached from the internal elastic lamina with an indication of subendothelial oedema. As early as 15 min after LPS injection [7] cellular injuries are apparent, with nuclear vacuolization, cytoplasmic swelling and protrusion, cytoplasmic fragmentation, and various degrees of detachment of the endothelium from its underlying layer. This can also be observed 10 hours after the onset of sepsis in a caecal ligation and puncture rat model [8]. Proinflammatory cytokines increase permeability of the ECs, and this is manifested approximately 6 hours after inflammation is triggered and becomes maximal over 12-24 hours as the combination of cytokines exert potentiating effects [8,9]. Endothelial physical disruption allows inflammatory fluid and cells to shift from the blood into the interstitial space.

APC = activated protein C; EC = endothelial cell; eNOS = endothelial constitutive nitric oxide synthase; ICAM = intercellular adhesion molecule; ICU = intensive care unit; LPS = lipopolysaccharide; NO = nitric oxide; PC = protein C; PGI₂ = prostacyclin; TF = tissue factor; TFPI = tissue factor pathway inhibitor TM = thrombomodulin; vWF = von Willebrand factor.

Table 1**Physiology and pathophysiology of endothelial cells**

Properties of ECs	In sepsis
Surface area: 1–7 m ²	ECs become injured, prothrombotic and antifibrinolytic
Weight: 1 kg/70 kg body weight	They promote platelet adhesion
Number: 1–6 × 10 ¹³ cells	They promote leucocyte adhesion and inhibit vasodilation
They line vessels in every organ: 'gate keeping role'	
They favour vasodilatation	
They promote antithrombosis and profibrinolysis	
They inhibit platelet adhesion and leucocyte adhesion	

Shown are key endothelial cell (EC) functions that are altered in inflammation or sepsis.

Plasma levels of thrombomodulin (TM), intercellular adhesion molecule (ICAM)-1 and E-selectin may be measured in order to assess EC injury [10,11]. von Willebrand factor (vWF) and its propeptide can also be measured as circulating blood proteins to assess endothelial injury. It has been demonstrated that the half-life of mature vWF and that of its propeptide differ fourfold to fivefold [12]. The molar ratio of the propeptide to mature vWF can serve as a tool with which to assess the extent of EC injury and to distinguish between acute and chronic disease [13]. In patients with diabetes mellitus propeptide levels are only slightly elevated, whereas vWF levels are elevated twofold to threefold. In acute sepsis, both vWF and propeptide are elevated several fold. High levels of TM, ICAM-1 and vWF have been reported in several inflammatory diseases, sepsis and acute lung injury in patients with nonpulmonary sepsis, in which endothelial damage is thought to be important [11,14,15].

In a recent report, Mutunga *et al.* [16] developed a method for detecting circulating ECs that provides direct evidence of EC shedding in human sepsis. Blood samples were subsequently taken from 11 healthy volunteers, nine ventilated intensive care unit (ICU) control patients without sepsis, eight patients with sepsis but without shock, and 15 patients with septic shock. EC were identified by indirect immunofluorescence, using antibodies to vWF and the vascular endothelial growth factor receptor EGFR. vWF-positive EC counts per millilitre were significantly greater in patients with sepsis (16.1 ± 2.7 [mean \pm SEM]) and septic shock (30.1 ± 3.3) than in healthy (1.9 ± 0.5) or ICU control individuals (2.6 ± 0.6). EGFR-positive EC counts per ongoing EC lesions were also significantly higher in patients with sepsis (4.2 ± 1.1) and septic shock (10.4 ± 1.2) than in healthy (0.7 ± 0.3) or ICU control individuals (0.5 ± 0.2). Cell counts measured using anti-vWF antibody were consistently higher than those measured using anti-EGFR antibody, but correlation between the two counts was high ($r^2=0.93$). The number of circulating EGFR-positive ECs per millilitre was significantly higher in patients who died of septic shock than in survivors

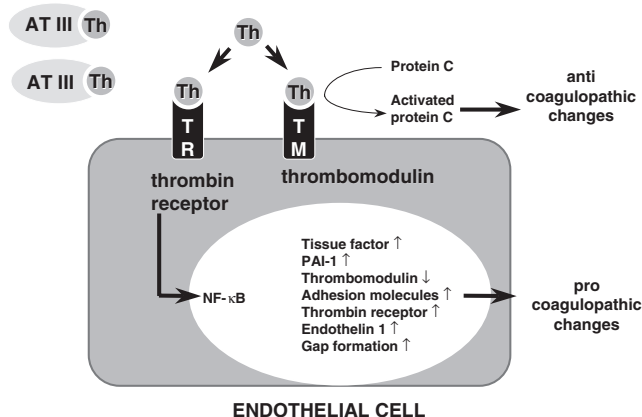
(12.0 ± 1.6 versus 7.1 ± 1.2 ; $P=0.026$). An increase in circulating ECs can therefore be identified during sepsis and septic shock. That study was among the first to support the hypothesis that endothelial damage occurs in human sepsis.

An important point is that EC injury is sustained over time. In an endotoxic rabbit model, we demonstrated that endothelium denudation is present at the level of the abdominal aorta as early as after several hours following injury and persisted for at least 5 days afterward [6,17]. After 21 days we observed that the endothelial surface had recovered. The de-endothelialized surface accounted for approximately 25% of the total surface. Similarly, in 12 human volunteers receiving 4 ng/kg *Escherichia coli* LPS by intravenous injection, Taylor and coworkers [18] showed that the immediate symptomatic inflammatory stage (0–8 hours after LPS injection) was followed after 12 hours by an asymptomatic noninflammatory stage (volunteers were back at work). The latter stage was characterized by decreased tumour necrosis factor, interleukin-10, thrombin–antithrombin and plasmin–antiplasmin complexes, and levels of TM peaked at 24 hours, suggesting ongoing EC lesions. Increased TM was associated with a level of tissue factor (TF) that was still increasing at 48 hours, suggesting risk for activated coagulation. Indeed, TF is the principal activator of the extrinsic coagulation pathway, and as such is responsible for an intravascular procoagulant state. Taylor and coworkers concluded that sustained injury to the vascular endothelium secondary to reperfusion of the microvasculature occurred in those asymptomatic individuals. In our endotoxic model, we also demonstrated that at 5 days the rabbits had maximal monocyte TF expression, which coincided with maximal endothelial injury [6,17]. This, together with altered coagulation modulation properties, may ultimately result in intravascular microthrombosis.

Endothelial injury associated abnormal coagulation and fibrinolysis

The outer membrane of ECs normally expresses various membrane-associated components with anticoagulant prop-

Figure 1



Thrombomodulin and protein C activation at the microcirculatory level. The endothelial cell surface thrombin (Th)-binding protein thrombomodulin (TM) is responsible for inhibition of thrombin activity. TM, when bound to Th, forms a potent protein C (PC) activator complex. Loss of TM and/or internalization results in Th–thrombin receptor (TR) interaction. Loss of TM and associated protein C activation represents the key event of decreased endothelial coagulation modulation ability and increased inflammation pathways. Adapted from Iba and coworkers [88]. ATIII, antithrombin III; NF-κ, nuclear factor-κB; PAI, plasminogen activator inhibitor.

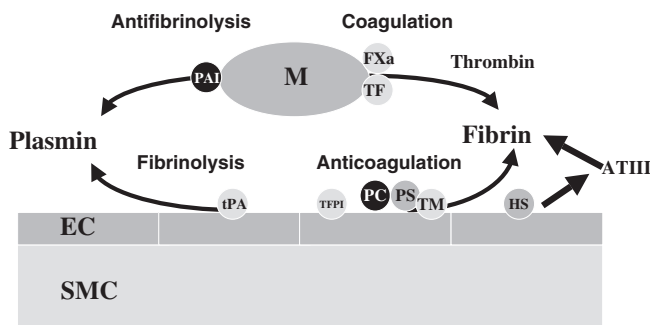
erties, among which are cell surface heparin-like molecules. These molecules accelerate inactivation of coagulation proteases by antithrombin and represent a TF pathway inhibitor (TFPI) reserve [19]. The EC surface thrombin-binding protein TM is responsible for inhibition of thrombin activity. TM, when bound to thrombin, forms a potent protein C (PC) activator complex (Fig. 1). Whereas unperturbed ECs confer anticoagulant properties (Fig. 2), exposure to inflammatory and/or septic stimuli rapidly lead to procoagulant behaviour (Figs 1 and 3). Moreover, the profibrinolytic property of ECs is blunted, because of decreased release of tissue plasminogen activator. This occurs within the context of increased release of plasminogen activator inhibitor-1. During sepsis the procoagulant activity of TF increases, with transcriptional upregulation of its expression on monocytes and ECs among other cell types, whereas levels of endothelium anticoagulant membrane components decrease, with internalization of TM [20] and release of inactive TM into the bloodstream (Fig. 3). Loss of TM and associated PC activation represents a key event, namely decreased endothelial coagulation modulation ability. Cleavage of TM by neutrophil elastase and other proteases certainly participate in the reduced expression of TM.

In severe meningococcal sepsis, Faust and coworkers [21] recently demonstrated that PC activation is impaired – a finding that is consistent with downregulation of the endothelial TM–endothelial PC receptor pathway. In 21 children (median age 41 months) with purpura fulminans (meningococcal sepsis), purpuric lesion skin biopsies exhibited decreased expression of endothelial TM and of the endothe-

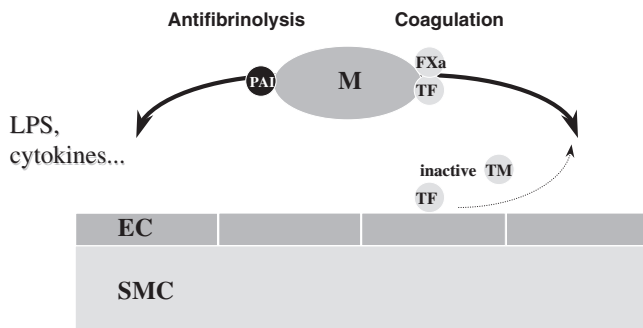
lial PC receptor as compared with control specimens, both in vessels with and in those without thrombosis. Plasma TM levels in the children with meningococcal sepsis (median 6.4 ng/l) were higher than those in the controls (median 3.6 ng/l; $P=0.002$). Plasma levels of PC antigen, protein S antigen and antithrombin antigen were lower than those in the controls. In two patients treated with unactivated PC concentrate, activated PC (APC) was undetectable at the time of admission, and plasma levels remained low.

Activation of coagulation concomitant with impaired fibrinolysis is associated with fibrin deposition, tissue ischaemia and tissue necrosis [22], and in critically ill patients with increased risk for death [23,24]. Conversely, inhibition of coagulation is associated with prevention of organ dysfunction [25,26]. Three therapeutic strategies that employ coagulation modulation – TFPI, antithrombin and APC – were recently proposed to reduce organ dysfunction and mortality in septic shock. It has clearly been shown in various animal models of septic shock that these treatments reduce organ dysfunction and mortality [27,28]. This was associated with a reduction in cytokine production [25,26,29]. With APC, it was further demonstrated that leucocyte–endothelial interactions were reduced [30]. Of note is the demonstration that APC was also able to improve fibrinolysis by inhibiting plasminogen activator inhibitor-1 [31]. Clinical phase II trials suggested that mortality might be reduced by using these coagulation modulators in critically ill septic patients [32–35]. Three phase III trials of antithrombin, TFPI and APC were subsequently performed and recently completed in large populations of patients with severe sepsis, the net effect being an overall lack of efficacy with antithrombin [36] and TFPI (unpublished results), and a 19.43% reduction in relative risk for death with APC [37].

Figure 2



Coagulation and fibrinolysis pathways. Unperturbed endothelial cells (ECs) provide anticoagulant (tissue factor pathway inhibitor [TFPI], protein C [PC], protein S [PS], thrombomodulin [TM], heparan sulphate [HS]) and fibrinolytic (tissue plasminogen activator [tPA]) properties. ATIII, antithrombin III; FXa, coagulation factor Xa; M, activated monocyte; PAI, plasminogen activator inhibitor; SMC, smooth muscle cell; TF, tissue factor.

Figure 3

Sepsis and coagulation–fibrinolysis pathways. Exposure to inflammatory and/or septic stimuli rapidly leads to procoagulant behaviour. The profibrinolytic property of endothelial cells (ECs) is blunted, due to decreased release of tissue plasminogen activator. This occurs in a context of increased plasminogen activator inhibitor (PAI)-1 release with antifibrinolysis. LPS, lipopolysaccharide; M, activated monocyte; SMC, smooth muscle cell; TF, tissue factor; TM, thrombomodulin.

Although ECs probably have an important role in disseminated intravascular coagulation, there is also some evidence favouring a major role for monocytes in the cellular mechanisms of coagulation activation. We recently assessed the relative impact of endothelial injury and monocyte activation on coagulation disorders in our rabbit endotoxemic shock model. L-arginine and the angiotensin-converting enzyme inhibitor perindopril were tested in that model for their demonstrated ability to treat endothelial injury [38,39]. We found that both L-arginine supplementation and perindopril could prevent septic-shock-associated deterioration in endothelium-dependent relaxation [40,41]. However, this preventive effect was not associated with any reduction in TF expression, suggesting that these two sepsis-associated abnormalities are not strictly linked. In a subsequent study [42] we used an anti-glycoprotein IIb/IIIa, which attenuated endotoxin-induced monocyte TF expression through decreased platelet activation. This was associated with marked reduction in endothelial injury, increased endothelium-derived relaxation and improved survival rates in the treated animals. Those findings suggest that monocyte activation and TF expression may be of importance in sepsis-associated injuries, and that coagulation activation may itself contribute to the EC injury observed during sepsis.

Endothelial injury, in turn, exacerbates sepsis-induced coagulation abnormalities. Indeed, release of endothelium-derived factors such as nitric oxide (NO) and prostacyclin (PGI₂) is impaired. Because NO and PGI₂ not only control vascular tone but also have antiadhesive and tissue plasminogen activator-like properties, loss of NO and PGI₂ release facilitates leucocyte and platelet aggregation, and aggravation of coagulopathy. Furthermore, when ECs generate adhesion molecules during endotoxaemia that bind leucocytes and

monocytes, they favour enhancement in local procoagulant reactions. The relationship between activation of innate immunity and coagulation is phylogenetically ancient [43,44]. Localized activation of the coagulation system, as with the innate immune response, serves to protect against a discrete traumatic injury [43]. However, generalized intravascular coagulation, as a generalized inflammatory response, is detrimental to the host, favouring widespread fibrin deposition and altered tissue perfusion.

Endothelial activation

As a prelude to their migration into tissues, monocytes and leucocytes must adhere to endothelium. Both adhesion and migration across endothelium are governed by the interaction of complementary adhesion molecules on the polymorphonuclear cells and endothelium [45]. The surface expression, adhesion avidity and surface modulation of these molecules are highly regulated by biological mediators such as cytokines. Local synthesis of platelet-activating factor and EC-derived cytokines such as interleukin-8, along with tumour necrosis factor and interleukin-1, are important in promoting neutrophil–EC interactions. ‘Endothelial activation’ refers to increased expression or release of endothelial adhesion molecules.

The first step in migration consists of a ‘rolling’ of leucocytes on endothelium, which involves the selectin family. Selectins are molecules expressed on leucocytes (L-selectin), or even on platelets (P-selectin) and on ECs (E-selectin); these act as receptors that permit loose binding, which in turn facilitates rolling. Selectins allow leucocytes to roll in the direction of flow into the proximity of activating signals exhibited by ECs.

The second step involves receptors from the integrin family (β_2 -integrin) and immunoglobulin-like receptors. These receptors allow leucocyte arrest and adhesion strengthening. Three heterodimers of β_2 -integrin are present on the outer cell membrane of activated leucocytes and are collectively termed the CD₁₁/CD₁₈ complex. Stimulation of ECs induces expression of cell surface adhesion molecules, which are members of the immunoglobulin superfamily. Monoclonal antibodies to these molecules have been shown to block leucocyte–EC interactions and to improve sepsis-associated organ dysfunction [46,47]. Endothelial adhesion molecules include ICAMs, endothelial leucocyte adhesion molecules (E-selectins), platelet EC adhesion molecules, and vascular cell adhesion molecules.

In the third step, activated leucocytes migrate to the borders of ECs to interact with ICAMs, endothelial leucocyte adhesion molecules, platelet EC adhesion molecules or vascular cell adhesion molecules (for review [48]). A large number of experimental studies have documented the consequences of inhibition of adhesion molecules. Inhibition of neutrophil adherence to the ECs exerts significant protective effects in these conditions [46,49].

Interestingly, decreased reactive hyperaemia (suggesting modified endothelial-derived relaxation) was also demonstrated to coexist with increased leucocyte aggregation and ICAM-1 levels [50]. It is also important to emphasize that recent evidence suggested that adhesion can occur independent of adhesion molecules in organs such as lung or liver. This led to the hypothesis that stimulus-induced increases in actin-containing stress fibres (such as LPS) at the cell periphery lead to decreased deformability, preventing neutrophils from trafficking through the capillary bed and therefore increasing their sequestration at sites of inflammation [51].

Sessler and coworkers [52] measured blood level of the adhesion molecule ICAM-1 as a potential marker of EC activation in septic adults and healthy volunteers. Those investigators established a relationship between increased ICAM-1 levels and consequences of sepsis (i.e. multiple organ failure and death). Watanabe and coworkers [53] prevented endotoxin shock in rabbits by administering a specific monoclonal antibody against CD₁₈ (integrin β_2). In a mouse lethal septic shock model, Xu and coworkers [54] observed that animals deficient in ICAM-1 were markedly protected against death. Whereas 80% of wild-type animals died within 48 hours after receiving 40 mg/kg LPS, more than 90% of ICAM-1-deficient animals survived for longer than 4 days. Interestingly, EC dysfunction was found to involve a CD₁₈-dependent neutrophil adherent mechanism. Consistently, Matsukawa and coworkers [55] recently provided evidence on the contributions of E-selectin and P-selectin to lethality in septic peritonitis. Mice that genetically lacked endothelial selectins were shown to be resistant. The experiments demonstrated that endothelial selectin mediated leucocyte rolling impacts on mouse survival by influencing the serum level of cytokines and by preventing renal dysfunction – a potential cause of death in that context.

Endothelial dysfunction

The term 'endothelial dysfunction' refers to decreased endothelial-dependent vascular relaxation or NO release, and decreased expression or activity of endothelial constitutive NO synthase (ecNOS). Endothelium-derived relaxation and/or production of endothelium-derived NO from the amino acid L-arginine by ecNOS may be used as an indicator of EC function. For example, the relaxing response of *in vitro* isolated vascular rings to picomolar concentrations of acetylcholine is dependent on the presence and integrity of ECs [56]. *In vivo* endothelial function can be determined by measurement of forearm blood flow responses to intra-arterial infusions of endothelium-dependent (i.e. acetylcholine) and endothelium-independent vasodilators (i.e. sodium nitropruside). Drugs are infused at a constant rate (1 ml/min) with an infusion pump. Forearm blood flow is recorded for 10 s at 15-s intervals during the last 3 min of the drug and saline infusion period using venous occlusion plethysmography combined with a rapid cuff inflator [57–59].

Abnormal endothelial-dependent vascular relaxation has been recognized in multiple sepsis conditions. Several investigations, including our own [17,60,61], have demonstrated attenuated acetylcholine-induced relaxation in vascular rings isolated from large arteries. Apart from anatomical injuries, such abnormalities observed in these vessels may result from several mechanisms: alteration in EC surface receptors; modified signal transduction pathways (receptor–ecNOS coupling); altered function and/or density of the ecNOS; changes in pathways that lead to release of NO; and/or changes in mechanisms that participate in subsequent degradation of NO.

In healthy volunteers, even brief exposure to endotoxin or certain cytokines impairs endothelium-dependent relaxation for many days [62,63]. This effect has been termed 'endothelial stunning'. After recovery from the acute insult, the endothelium may remain dysfunctional ('stunned') for a long period of time before full recovery. Hingorani and coworkers [59] also demonstrated that a mild inflammatory response, such as that generated by *Salmonella typhi* vaccine, is associated with temporary but profound dysfunction of arterial endothelium in both resistance and conduit vessels following application of both physical and pharmacological dilator stimuli. According to the concept of intrinsic metabolic regulation, vasodilatation in tissues with relatively high metabolic rates competes with sympathetic vasoconstrictor tone, thereby adjusting the balance between local tissue oxygen supply and demand. Although the nature of the oxygen-sensitive structures that act at the local tissue level is not completely understood, ECs in direct contact with blood have a number of properties that render them effective sensors. The endothelium and smooth muscle of arteries and arterioles appear to be coupled both structurally and functionally. Sensing involves local depolarization/hyperpolarization of the capillary EC, and communication is achieved by electronic spread via endothelium/smooth muscle cell–cell gap junctions [2,64]. During an hypoxic challenge, the ability of a tissue to extract oxygen – and to minimize shunting through areas with a high rate of perfusion relative to their oxygen uptake – may therefore be considered an integrative test of endothelial function and microcirculatory coordination [65].

We investigated the role of the endothelium in regulating the balance between oxygen demand and supply within an individual organ in an *in vivo* model of endothelial stripping in the dog hind limb [66]. The hind limb vascular endothelium was removed by injecting deoxycholate into the perfusing artery before ischaemic challenge. Deoxycholate – a detergent used to remove endothelium in *in vitro* studies – removes vascular endothelium within arteries, arterioles, capillaries and veins. It achieves this without causing apparent damage to either the vascular smooth muscle layer or the skeletal muscle parenchyma, as assessed by *in vitro* and *in vivo* studies of pharmacological vascular reactivity, tissue histology and electron microscopy. Hind limb oedema or capillary plugging by

endothelial fragments was not observed. During progressive limitation of oxygen supply to the limb, a profound and significant impairment in limb oxygen extraction ability (41.7% versus 81% in controls) at critical oxygen delivery (at which oxygen uptake begins to decrease) was observed. We concluded that this severe limitation in the increase in oxygen extraction capabilities during ischaemia suggested that vascular endothelium plays an important role in matching oxygen supply to demand.

In order to test the role of the endothelial-derived relaxing factors NO and PGI₂, we investigated, in a third group of dogs, the influence of a combination of *N*^G-nitro-L-arginine methyl ester (an inhibitor of NO synthesis) and indomethacin (an inhibitor of PGI₂ synthesis) [66]. In these dogs treated with indomethacin plus *N*^G-nitro-L-arginine methyl ester, the severity of the oxygen extraction defect was lower than that observed in the deoxycholate-treated dogs, suggesting that other mediators and/or mechanisms may be involved in microcirculatory control during hypoxia. As suggested above, one of these mediators or mechanisms could be related to hyperpolarization. Membrane potential is an important determinant of vascular smooth muscle tone through its influence on calcium influx via voltage-gated calcium channels. Hyperpolarization (as well as depolarization) has been shown to be a means of cell–cell communication in upstream vasodilatation and microcirculatory coordination [67]. It is important to emphasize that intercell coupling exclusively involves ECs. Interestingly, it was recently shown that sepsis, a situation that is characterized by impaired tissue perfusion and abnormal oxygen extraction, is associated with abnormal inter-EC coupling and reduction in the arteriolar conducted response [68].

An intra-organ defect in blood flow related to abnormal vascular reactivity, cell adhesion and coagulopathy may account for impaired organ oxygen regulation and function. If specific classes of microvessels must or must not be perfused to achieve efficient oxygen extraction during limitation in oxygen delivery, then impaired vascular reactivity and vessel injury might produce a pathological limitation in supply. In sepsis, the inflammatory response profoundly alters circulatory homeostasis, and this has been referred to as a 'malignant intravascular inflammation' that alters vasomotor tone and the distribution of blood flow among and within organs [69]. These mechanisms might coexist with other types of sepsis-associated cell dysfunction. For example, data suggest that endotoxin directly impairs oxygen uptake in ECs and indicate the importance of endothelium respiration in maintaining vascular homeostasis under conditions of sepsis [70].

Abnormal oxygen extraction is a key feature of severe sepsis and septic shock. In an experimental study in dogs, an ablated reactive hyperaemia was associated with endotoxaemia-induced impaired oxygen extraction at the level of the gastrointestinal tract [71]. Nevière and coworkers [72] showed that reactive hyperaemia is attenuated in critically ill

patients with septic shock, despite normal or elevated whole-body oxygen delivery. Proposed mechanisms to explain blunted hyperaemia in septic patients might include impaired vascular reactivity and/or microvascular obstruction that limits the number of recruitable capillaries. In critically ill patients with sepsis, it has been shown that decreased reactive hyperaemia coexists with increased leucocyte adhesion and increased release in surrogate markers of endothelium injury [50,73].

Thus, assessment of reactive hyperaemia might be used in the near future to evaluate the effects of treatments aimed at restoring endothelial function and tissue perfusion, such as coagulation modulators or leucocyte adhesion antagonists.

Conclusion

How do all of these altered properties contribute to altered perfusion and organ dysfunction? The combining effect of altered vascular relaxation, altered blood flow distribution, increased leucocyte adhesion and decreased coagulation modulation should significantly contribute to microcirculatory heterogeneity and lowered perfusion. Studies in the isolated perfused rabbit heart [74], autoperfused rat cremaster [75] and rat mesentery [76,77] suggested that these mechanisms are operative in the microvasculature. On an intravital microscopy extensor digitorum longus muscle model in rats with peritonitis [78], it was shown that sepsis is associated with a reduction in tissue perfused capillary density of up to 36%, increased perfusion heterogeneity and mean intercapillary distance, contributing to functional shunting. In another study [79], endotoxin administration resulted in a significant enhancement in leucocyte–EC interaction, as indicated by transiently increased number of leucocytes firmly attaching to the microvascular endothelium of arterioles and venules [79]. Microvascular injury and/or the appearance of greater heterogeneity of microvascular distribution of oxygen supply with respect to oxygen demand in endotoxin-treated animals is consistent with the observation that endotoxin impairs oxygen extraction [80–82]. The direct relationship between heterogeneity, decreased oxygen extraction and tissue acidosis was recently confirmed in a pig model of endotoxic shock [83].

Consistent with the hypothesis that alteration in endothelium plays a major role in the pathophysiology of sepsis, it was observed that chronic eNOS overexpression in the endothelium of mice resulted in resistance to LPS-induced hypotension, lung injury and death [84]. This observation was confirmed by another group of investigators, who used transgenic mice overexpressing adrenomedullin [85] – a vasodilating peptide that acts at least in part via an NO-dependent pathway. They demonstrated resistance of these animals to LPS-induced shock, and lesser declines in blood pressure and less severe organ damage than occurred in the control animals. It might therefore be of importance to favour eNOS expression and function during sepsis. The recent negative results obtained with therapeutic strategies aimed at blocking inducible NOS with the nonselective NOS inhibitor *N*^G-

monomethyl-L-arginine in human septic shock [86] further confirm the overall importance of favoring vessel dilatation. In contrast, positive results obtained with corticosteroids [87] and APC [37] suggest that improving haemodynamics while decreasing vasopressor agents (corticosteroids) and limiting coagulation activation are logical strategies that may greatly favour tissue perfusion and improved oxygen delivery.

Competing interests

None declared.

References

- Cines DB, Pollak ES, Buck CA, Loscalzo J, Zimmerman GA, McEver RP, Pober JS, Wick TM, Konkle BA, Schwartz BS, Barnathan ES, McCrae KR, Hug BA, Schmidt AM, Stern DM: **Endothelial cells in physiology and in the pathophysiology of vascular disorders.** *Blood* 1998, **91**:3527-3561.
- Sandow SL, Hill CE: **Incidence of myoendothelial gap junctions in the proximal and distal mesenteric arteries of the rat is suggestive of a role in endothelium-derived hyperpolarizing factor-mediated responses.** *Circ Res* 2000, **86**:341-346.
- Stefanec T: **Endothelial apoptosis: could it have a role in the pathogenesis and treatment of disease.** *Chest* 2000, **117**:841-854.
- Reidy MA, Schwartz SM: **Endothelial injury and regeneration. IV. Endotoxin: a nondegrading injury to aortic endothelium.** *Lab Invest* 1983, **48**:25-34.
- Reidy MA, Bowyer DE: **Scanning electron microscopy: morphology of aortic endothelium following injury by endotoxin and during subsequent repair.** *Atherosclerosis* 1977, **26**:319-328.
- Leclerc J, Pu Q, Corseaux D, Haddad E, Decoene C, Bordet R, Six I, Jude B, Vallet B: **A single endotoxin injection in the rabbit causes prolonged blood vessel dysfunction and a procoagulant state.** *Crit Care Med* 2000, **28**:3672-3678.
- Lee M, Schuessler G, Chien S: **Time dependent effects of endotoxin on the ultrastructure of the aortic endothelium.** *Artery* 1998, **15**:71-89.
- Wang P, Wood TJ, Zhou M, Ba ZF, Chaudry IH: **Inhibition of the biological activity of tumor necrosis factor maintains vascular endothelial cell function during hyperdynamic sepsis.** *J Trauma* 1996, **40**:694-701.
- Bauer PR: **Microvascular responses to sepsis: clinical significance.** *Pathophysiology* 2002, **8**:141-148.
- Bajaj MS, Tricomi SM: **Plasma levels of the three endothelial-specific proteins von Willebrand factor, tissue factor pathway inhibitor, and thrombomodulin do not predict the development of acute respiratory distress syndrome.** *Intensive Care Med* 1999, **25**:1259-1266.
- Kayal S, Jais JP, Aguin N, Chaudiere J, Labrousse J: **Elevated circulating E-selectin, intercellular adhesion molecule 1, and von Willebrand factor in patients with severe infection.** *Am J Respir Crit Care Med* 1998, **157**:776-784.
- Borchiellini A, Fijnvandraat K, ten Cate JW, Pajkt D, van Deventer SJ, Pasterkamp G, Meijer-Huizinga F, Zwart-Huinink L, Voorberg J, van Mourik JA: **Quantitative analysis of von Willebrand factor propeptide release in vivo: effect of experimental endotoxemia and administration of 1-deamino-8-D-arginine vasopressin in humans.** *Blood* 1996, **88**:2951-2958.
- Van Mourik JA, Boertjes R, Huisveld IA, Fijnvandraat K, Pajkt D, van Genderen PJ, Fijnheer R: **von Willebrand factor propeptide in vascular disorders: a tool to distinguish between acute and chronic endothelial cell perturbation.** *Blood* 1999, **94**:179-185.
- Blann AD, Babbs C, Neuberger JM: **Endothelial cell damage in primary biliary cirrhosis: influence of cholestasis and immunological mechanisms.** *Clin Exp Immunol* 1992, **90**:88-92.
- Rubin DB, Wiener-Kronish JP, Murray JF, Green DR, Turner J, Luce JM, Montgomery AB, Marks JD, Matthay MA: **Elevated von Willebrand factor antigen is an early plasma predictor of acute lung injury in nonpulmonary sepsis syndrome.** *J Clin Invest* 1990, **86**:474-480.
- Mutunga M, Fulton B, Bullock R, Batchelor A, Gascoigne A, Gillespie JI, Baudouin SV: **Circulating endothelial cells in patients with septic shock.** *Am J Respir Crit Care Med* 2001, **163**:195-200.
- Wiel E, Vallet B: **Vascular endothelial cell dysfunction in septic shock.** *Crit Care Med* 2001, **29**(suppl):S36-S41.
- Taylor FB, Haddad PA, Hack E, Chang AC, Peer GT, Morrissey JH, Li A, Allen R, Caa H, Kinasewitz GT: **Two-stage response to endotoxin infusion into normal human subjects: correlation of blood phagocyte luminescence with clinical and laboratory markers of the inflammatory, hemostatic response.** *Crit Care Med* 2001, **29**:326-334.
- Bombeli T, Mueller M, Haeberli A: **Anticoagulant properties of the vascular endothelium.** *Thromb Haemost* 1997, **77**:408-423.
- Rapaport S, Rao L: **Initiation and regulation of tissue factor-dependent blood coagulation.** *Arterioscler Thromb* 1992, **12**:1111-1121.
- Faust SN, Levin M, Harrison OB, Goldin RD, Lockhart MS, Kondaveeti S, Laszik Z, Esmon CT, Heyderman RS: **Dysfunction of endothelial protein C activation in severe meningococcal sepsis.** *N Engl J Med* 2001, **345**:408-416.
- Regoeczi E, Brain MC: **Organ distribution of fibrin in disseminated intravascular coagulation.** *Br J Haematol* 1969, **17**:73-81.
- Fourrier F, Chopin C, Goudemand J, Hendrycx S, Caron C, Rime A, Marey A, Lestavel P: **Septic shock, multiple organ failure, and disseminated intravascular coagulation. Compared patterns of antithrombin III, protein C, and protein S deficiencies.** *Chest* 1992, **101**:816-823.
- Gando S, Nakanishi Y, Tede I: **Cytokines and plasminogen activator inhibitor-1 in posttrauma disseminated intravascular coagulation: relationship to multiple organ dysfunction syndrome.** *Crit Care Med* 1995, **23**:1835-1842.
- Creasey AA, Chang AC, Feigen L, Wun TC, Taylor FB, Hinshaw LB: **Tissue factor pathway inhibitor reduces mortality from Escherichia coli septic shock.** *J Clin Invest* 1993, **91**:2850-2856.
- Kessler CM, Tang Z, Jacobs HM, Szymanski LM: **The suprapharmacologic dosing of antithrombin concentrate for Staphylococcus aureus-induced disseminated intravascular coagulation in guinea pigs: substantial reduction in mortality and morbidity.** *Blood* 1997, **89**:4393-4401.
- Carr C, Bild GS, Chang AC: **Recombinant E. coli-derived tissue factor pathway inhibitor reduces coagulopathic and lethal effects in the baboon gram-negative model of septic shock.** *Crit Care Med* 1994, **22**:126-137.
- Camerota AJ, Creasey AA, Patla V, Larkin VA, Fink MP: **Delayed treatment with recombinant human tissue factor pathway inhibitor improves survival in rabbits with gram-negative peritonitis.** *J Infect Dis* 1998, **177**:668-676.
- Grey ST, Tsuchida A, Hau H, Orthner CL, Salem HH, Hancock WW: **Selective inhibitory effects of the anticoagulant activated protein C on the responses of human mononuclear phagocytes to LPS, IFN-gamma, or phorbol ester.** *J Immunol* 1994, **153**:3664-3672.
- Grinnell BW, Hermann RB, Yan SB: **Human protein C inhibits selectin-mediated cell adhesion: role of unique fucosylated oligosaccharide.** *Glycobiology* 1994, **4**:221-225.
- Jackson CV, Bailey BD, Shetler TJ: **Pharmacological profile of recombinant, human activated protein C (LY203638) in a canine model of coronary artery thrombosis.** *J Pharmacol Exp Ther* 2000, **295**:967-971.
- Smith OP, White B, Vaughan D, Rafferty M, Claffey L, Lyons B, Casey W: **Use of protein-C concentrate, heparin, and haemodiafiltration in meningococcus-induced purpura fulminans.** *Lancet* 1997, **350**:1590-1593.
- Fourrier F, Chopin C, Huart JJ, Runge I, Caron C, Goudemand J: **Double-blind, placebo-controlled trial of antithrombin III concentrates in septic shock with disseminated intravascular coagulation.** *Chest* 1993, **104**:882-888.
- Baudo F, Caimi TM, de Cataldo F, Ravizza A, Arlati S, Casella G, Carugo D, Palareti G, Legnani C, Ridolfi L, Rossi R, D'Angelo A, Crippa L, Giudici D, Gallioli G, Wolfler A, Calori G: **Antithrombin III (ATIII) replacement therapy in patients with sepsis and/or postsurgical complications: a controlled double-blind, randomized, multicenter study.** *Intensive Care Med* 1998, **24**:336-342.

35. Eisele B, Lamy M, Thijs LG, Keinecke HO, Schuster HP, Matthias FR, Fourrier F, Heinrichs H, Delvos U: **Antithrombin III in patients with severe sepsis. A randomized, placebo-controlled, double-blind multicenter trial plus a meta-analysis on all randomized, placebo-controlled, double-blind trials with antithrombin III in severe sepsis.** *Intensive Care Med* 1998, **24**: 663-672.
36. Warren BL, Eid A, Singer P, Pillay SS, Carl P, Novak I, Chalupa P, Atherstone A, Penzes I, Kubler A, Knaub S, Keinecke HO, Heinrichs H, Schindel F, Juers M, Bone RC, Opal SM: **Caring for the critically ill patient. High-dose antithrombin III in severe sepsis: a randomized controlled trial.** *JAMA* 2001, **286**:1869-1878
37. Bernard GR, Vincent JL, Laterre PF, LaRosa SP, Dhainaut JF, Lopez-Rodriguez A, Steingrub JS, Garber GA, Helterbrand JD, Ely EW, Fisher CJ: **Efficacy and safety of recombinant human activated protein C for severe sepsis.** *N Engl J Med* 2001, **344**: 699-709.
38. Hamon M, Vallet B, Bauters C, Wernert N, McFadden EP, Lablanche JM, Dupuis B, Bertrand ME: **Long-term oral administration of L-arginine reduces intimal thickening and enhances neoendothelium-dependent acetylcholine-induced relaxation after arterial injury.** *Circulation* 1994, **90**:1357-1362.
39. Van Belle E, Vallet B, Auffray JL, Bauters C, Hamon M, McFadden EP, Lablanche JM, Dupuis B, Bertrand ME: **NO synthesis is involved in structural and functional effects of ACE inhibitors in injured arteries.** *Am J Physiol* 1996, **270**:H298-H305.
40. Wiel E, Pu Q, Corseaux D, Robin E, Bordet R, Lund N, Jude B, Vallet B: **Effect of L-arginine on endothelial injury and hemostasis in rabbit endotoxin shock.** *J Appl Physiol* 2000, **89**: 1811-1818.
41. Leclerc JF, Pu Q, Bordet R, Bauters C, Vallet B: **Endothelial cell dysfunction occurring during septic shock is prevented by angiotensin converting enzyme inhibitor [abstract].** *Am J Respir Crit Care Med* 2000, **157**:A101.
42. Pu Q, Wiel E, Corseaux D, Bordet R, Azrin MA, Ezekowitz MD, Lund N, Jude B, Vallet B: **Beneficial effect of glycoprotein IIb/IIIa inhibitor (AZ-1) on endothelium in *Escherichia coli* endotoxin-induced shock.** *Crit Care Med* 2001, **29**:1181-1188.
43. Opal SM: **Phylogenetic and functional relationships between coagulation and the innate immune response.** *Crit Care Med* 2000, **28**(suppl):S77-S80.
44. Opal SM, Palardy JE, Parejo NA, Creasey AA: **The activity of tissue factor pathway inhibitor in experimental models of superantigen-induced shock and polymicrobial intra-abdominal sepsis.** *Crit Care Med* 2001, **29**:13-17.
45. Springer T: **Traffic signals on endothelium for lymphocyte recirculation and leukocyte emigration.** *Ann Rev Physiol* 1995, **57**:827-872.
46. Gardinali M, Borrelli E, Chiara O, Lundberg C, Padelino P, Conciato L, Cafaro C, Lazzi S, Luzi P, Giomarelli PP, Agostoni A: **Inhibition of CD11b/CD18 complex prevents acute lung injury and reduces mortality after peritonitis in rabbits.** *Am J Respir Crit Care Med* 2000, **161**:1022-1029.
47. Kuebler WM, Borges J, Sckell A, Kuhnle EH, Bergh K, Messmer K, Goetz AE: **Role of L-selectin in leukocyte sequestration in lung capillaries in a rabbit model of endotoxemia.** *Am J Respir Crit Care Med* 2000, **161**:36-43.
48. Varani J, Ward P: **Mechanisms of endothelial cell injury in acute inflammation.** *Shock* 1994, **2**:311-319.
49. Ma X, Tsao P, Lefler A: **Antibody to CD-18 exerts endothelial cell and cardiac protective effects in myocardial ischemia and reperfusion.** *J Clin Invest* 1995, **88**:1237-1243.
50. Astiz ME, DeGent GE, Lin RY, Rackow EC: **Microvascular function and rheologic changes in hyperdynamic sepsis.** *Crit Care Med* 1995, **23**:265-271.
51. Doerschuk CM: **Leukocyte trafficking in alveoli and airway passages.** *Respir Res* 2000, **1**:136-140.
52. Sessler C, Windsor A, Schwartz M: **Circulating ICAM-1 is increased in septic shock.** *Am J Respir Crit Care Med* 1995, **151**:1420-1427.
53. Watanabe S, Mukaida N, Ikeda N, Akiyama M, Harada A, Nakanishi I, Nariuchi H, Watanabe Y, Matsushima K: **Prevention of endotoxin shock by an antibody against leukocyte integrin beta 2 through inhibiting production and action of TNF.** *Int Immunol* 1995, **7**:1037-1046.
54. Xu H, Gonzalo JA, St Pierre Y, Williams IR, Kupper TS, Cotran RS, Springer TA, Gutierrez-Ramos JC: **Leucocytosis and resistance to septic shock in intercellular adhesion molecule 1-deficient mice.** *J Exp Med* 1994, **180**:95-109.
55. Matsukawa A, Lukacs NW, Hogaboam CM, Knibbs RN, Bullard DC, Kunkel SL, Stoolman LM: **Mice genetically lacking endothelial selectins are resistant to the lethality in septic peritonitis.** *Exp Mol Pathol* 2002, **72**:68-76.
56. Furchgott R, Zawadzki J: **The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine.** *Nature* 1980, **288**:373-376.
57. Makimattila S, Virkamaki A, Groop PH, Cockcroft J, Utriainen T, Fagerudd J, Yki-Jarvinen H: **Chronic hyperglycemia impairs endothelial function and insulin sensitivity via different mechanisms in insulin-dependent diabetes mellitus.** *Circulation* 1996, **94**:1276-1282.
58. Vakkilainen J, Makimattila S, Seppala-Lindroos A, Vehkavaara S, Lahdenpera S, Groop PH, Taskinen MR, Yki-Jarvinen H: **Endothelial dysfunction in men with small LDL particles.** *Circulation* 2000, **102**:716-721.
59. Hingorani AD, Cross J, Kharbanda RK, Mullen MJ, Bhagat K, Taylor M, Donald AE, Palacios M, Griffin GE, Deanfield JE, MacAllister RJ, Vallance P: **Acute systemic inflammation impairs endothelium-dependent dilation in humans.** *Circulation* 2000, **102**:994-999.
60. Parker JL, Adams HR: **Selective inhibition of endothelium-dependent vasodilator capacity by *Escherichia coli* endotoxemia.** *Circ Res* 1993, **72**:539-551.
61. Umans JG, Wylam ME, Umans JG, Wylam ME, Samsel RW, Edwards J, Schumacker PT: **Effects of endotoxin in vivo on endothelial and smooth-muscle function in rabbit and rat aorta.** *Am Rev Respir Dis* 1993, **148**:1638-1645.
62. Bhagat K, Collier J, Vallance P: **Local venous responses to endotoxin in humans.** *Circulation* 1996, **94**:490-497.
63. Bhagat K, Moss R, Collier J, Vallance P: **Endothelial 'stunning' following a brief exposure to endotoxin: a mechanism to link infection and infarction?** *Cardiovasc Res* 1996, **32**:822-829.
64. Emerson GG, Segal SS: **Endothelial cell pathway for conduction of hyperpolarization and vasodilation along hamster feed artery.** *Circ Res* 2000, **86**:94-100.
65. Vallet B: **Endothelial cell dysfunction and abnormal tissue perfusion.** *Crit Care Med* 2002, **30**(suppl):S229-S234.
66. Curtis SE, Vallet B, Winn MJ, Caufield JB, King CE, Chapler CK, Cain SM: **Role of the vascular endothelium in O₂ extraction during progressive ischemia in canine skeletal muscle.** *J Appl Physiol* 1995, **79**:1351-1360.
67. Segal SS: **Microvascular recruitment in hamster striated muscle: role for conducted vasodilation.** *Am J Physiol* 1991, **261**:H180-H189.
68. Tyml K, Wang X, Lidington D, Ouellette Y: **Lipopolysaccharide reduces intercellular coupling in vitro and arteriolar conducted response in vivo.** *Am J Physiol* 2001, **281**:H1397-H1406.
69. Pinsky MR: **Regional blood flow distribution.** In *The Splanchnic Circulation: No Longer a Silent Partner*. Edited by Pinsky MR, Dhainaut JF, Artigas A. Berlin: Springer-Verlag; 1995:1-13.
70. Motterlini R, Kerger H, Green CJ, Winslow RM, Intaglietta M: **Depression of endothelial and smooth muscle cell oxygen consumption by endotoxin.** *Am J Physiol* 1998, **275**:H776-H782.
71. Nelson DP, Samsel RW, Wood LDH, Schumacker PT: **Pathological supply dependence of systemic and intestinal O₂ uptake during endotoxemia.** *J Appl Physiol* 1988, **64**:2410-2419.
72. Nevière R, Mathieu D, Chagnon JL, Lebleu N, Millien JP, Wattel F: **Skeletal muscle microvascular blood flow and oxygen transport in patients with severe sepsis.** *Am J Respir Crit Care Med* 1996, **153**:191-195.
73. Kirschenbaum LA, Astiz ME, Rackow EC, Saha DC, Lin R: **Microvascular response in patients with cardiogenic shock.** *Crit Care Med* 2000, **28**:1290-1294.
74. Smith R, Palmer R, Moncada S: **Coronary vasodilatation induced by endotoxin in the rabbit isolated perfused heart is nitric oxide-dependent and inhibited by dexamethasone.** *Br J Pharmacol* 1991, **140**:5-6.
75. Lubbe AS, Garrison RN, Cryer HM, Alsip NL, Harris PD: **EDRF as a possible mediator of sepsis-induced arteriolar dilatation in skeletal muscle.** *Am J Physiol* 1992, **262**:H880-H887.

76. Schneider F, Schott C, Stoclet JC, Julou-Schaeffer G: **L-arginine induces relaxation of small mesenteric arteries from endotoxin-treated rats.** *Eur J Pharmacol* 1992, **211**:269-272.
77. Wang P, Ba ZF, Chaudry IH: **Endothelium-dependent relaxation is depressed at the macro- and microcirculatory levels during sepsis.** *Am J Physiol* 1995, **269**:R988-R994.
78. Lam C, Tynl K, Martin C, Sibbald W: **Microvascular perfusion is impaired in a rat model of normotensive sepsis.** *J Clin Invest* 1994, **94**:2077-2083.
79. Hoffmann JN, Vollmar B, Inthorn D, Schilberg FW, Menger MD: **A chronic model for intravital microscopic study of microcirculatory disorders and leukocyte/endothelial cell interaction during normotensive endotoxemia.** *Shock* 1999, **12**:355-364.
80. Bredle DL, Samsel RW, Schumacker PT, Cain SM: **Critical O₂ delivery to skeletal muscle at high and low PO₂ in endotoxemic dogs.** *J Appl Physiol* 1989, **66**:2553-2558.
81. Bredle DL, Cain SM: **Systemic and muscle oxygen uptake/delivery after dopexamine infusion in endotoxemic dogs.** *Crit Care Med* 1991, **19**:198-204.
82. Vallet B, Lund N, Curtis SE, Kelly D, Cain SM: **Gut and muscle tissue PO₂ in endotoxemic dogs during shock and resuscitation.** *J Appl Physiol* 1994, **76**:793-800.
83. Tugtekin IF, Radermacher P, Theisen M, Matejovic M, Stehr A, Ploner F, Matura K, Ince C, Georgieff M, Trager K: **Increased ileal-mucosal-arterial PCO₂ gap is associated with impaired villus microcirculation in endotoxic pigs.** *Intensive Care Med* 2001, **27**:757-766.
84. Yamashita T, Kawashina S, Ohashi Y: **Resistance to endotoxin shock in transgenic mice overexpressing endothelial nitric oxide synthase.** *Circulation* 2000, **101**:931-937.
85. Shindo T, Kurihara H, Maemura K, Kurihara Y, Kuwaki T, Izumida T, Minamino N, Ju KH, Morita H, Oh-hashii Y, Kumada M, Kangawa K, Nagai R, Yazaki Y: **Hypotension and resistance to lipopolysaccharide-induced shock in transgenic mice overexpressing adrenomedullin in their vasculature.** *Circulation* 2000, **101**:2309-2316.
86. Grover R, Lopez A, Lorente J, Steingrub J, Bakker J, Willatts S: **Multi-center, randomized, placebo-controlled, double blind study of the nitric oxide synthase inhibitor 546C88: effect on survival in patients in septic shock [abstract].** *Crit Care Med* 1999, **27**:A33.
87. Annane D, Sebille V, Charpentier C, Bollaert PE, Francois B, Korach JM, Capellier G, Cohen Y, Azoulay E, Troche G, Chaumet-Riffaut P, Bellissant E: **Effect of treatment with low doses of hydrocortisone and fludrocortisone on mortality in patients with septic shock.** *JAMA* 2002, **288**:862-871.
88. Iba T, Kidokoro A, Yagi Y: **The role of the endothelium in changes in procoagulant activity in sepsis.** *J Am Coll Surg* 1998, **187**:321-329.