

## Beyond Low Plasma T<sub>3</sub>: Local Thyroid Hormone Metabolism during Inflammation and Infection

Anita Boelen, Joan Kwakkel, and Eric Fliers

Department of Endocrinology and Metabolism, Academic Medical Center, University of Amsterdam, 1105AZ Amsterdam, The Netherlands

Decreased serum thyroid hormone concentrations in severely ill patients were first reported in the 1970s, but the functional meaning of the observed changes in thyroid hormone levels, together known as nonthyroidal illness syndrome (NTIS), remains enigmatic. Although the common view was that NTIS results in overall down-regulation of metabolism in order to save energy, recent work has shown a more complex picture. NTIS comprises marked variation in transcriptional and translational activity of genes involved in thyroid hormone metabolism, ranging from inhibition to activation, dependent on the organ or tissue studied. Illness-induced changes in each of these organs appear to be very different during acute or chronic inflammation, adding an additional level of complexity. Organ- and timing-specific changes in the activity of thyroid hormone deiodinating enzymes (deiodinase types 1, 2, and 3) highlight deiodinases as proactive players in the response to illness, whereas the granulocyte is a novel and potentially important cell type involved in NTIS during bacterial infection. Although acute NTIS can be seen as an adaptive response to support the immune response, NTIS may turn disadvantageous when critical illness enters a chronic phase necessitating prolonged life support. For instance, changes in thyroid hormone metabolism in muscle during critical illness may be relevant for the pathogenesis of myopathy associated with prolonged ventilator dependence. This review focuses on NTIS as a timing-related and organ-specific response to illness, occurring independently from the decrease in serum thyroid hormone levels and potentially relevant for disease progression. (*Endocrine Reviews* 32: 670–693, 2011)

- I. Introduction
- II. The Central Component of the HPT Axis
  - A. Hypothalamic response during illness
  - B. Pituitary response during illness
- III. Differential Effects of Acute Inflammation on Thyroid Hormone Metabolism in Metabolic Organs
  - A. Hepatic thyroid hormone metabolism during acute illness
  - B. Muscle thyroid hormone metabolism during acute illness
  - C. Thyroid hormone metabolism in adipose tissue during acute illness
- IV. Differential Effects of Chronic Inflammation and Sepsis on Thyroid Hormone Action in Metabolic Organs
  - A. Hepatic thyroid hormone metabolism during chronic inflammation
  - B. Muscle thyroid hormone metabolism during chronic inflammation
  - C. Hepatic thyroid hormone metabolism during bacterial sepsis
  - D. Muscle thyroid hormone metabolism during bacterial sepsis
  - E. Thyroid hormone metabolism in adipose tissue during bacterial sepsis
  - F. Deiodinase expression in granulocytes
- V. Thyroid Hormone Metabolism in Metabolic Organs during Prolonged Critical Illness
  - A. Hepatic thyroid hormone metabolism during prolonged illness
  - B. Muscle thyroid hormone metabolism during prolonged illness
- VI. Endocrine Interventions in NTIS; Areas of Uncertainty
  - A. Endocrine interventions
  - B. Areas of uncertainty
- VII. Conclusion

### I. Introduction

It has been known for many years that profound changes in thyroid hormone metabolism occur during illness, collectively known as the nonthyroidal illness syndrome

ISSN Print 0021-972X ISSN Online 1945-7197

Printed in U.S.A.

Copyright © 2011 by The Endocrine Society

doi: 10.1210/er.2011-0007 Received February 16, 2011. Accepted July 1, 2011.

First Published Online July 26, 2011

Abbreviations: AP-1, Activator protein-1; D1, deiodinase type 1; D2, deiodinase type 2; D3, deiodinase type 3; D3KO, D3 knockout; GHRP, GH-releasing peptide; GSH, glutathione; HIF, hypoxia inducible factor; HPT, hypothalamus pituitary thyroid; ICU, intensive care unit; IFN, interferon; JNK, Jun N-terminal kinase; LPS, lipopolysaccharide; ME, malic enzyme; MPO, myeloperoxidase; NFκB, nuclear factor κB; NTIS, nonthyroidal illness syndrome; PMN, polymorphonuclear leukocyte; PVN, paraventricular nucleus; RXR, retinoid X receptor; Se, selenium; SRC-1, steroid receptor coactivator-1; T<sub>2</sub>, 3,3'-diiodothyronine; TG, triglyceride; TR, thyroid hormone receptor; TSHR, TSH receptor; UCP, uncoupling protein; WT, wild-type.

(NTIS), the sick euthyroid syndrome, or the low- $T_3$  syndrome. This condition is characterized by decreased serum  $T_3$  and, in severe illness, decreased serum  $T_4$ , increased serum  $rT_3$ , and no increase in serum TSH, reflecting a major change in negative feedback regulation (1). Although there was some doubt initially whether the changes in total thyroid hormone levels during NTIS reflect alterations in free thyroid hormone concentrations (1, 2), it is generally accepted now that free  $T_4$  and free  $T_3$  levels decrease during severe illness (3, 4).

NTIS can be seen as a useful adaptation of the body to counteract excessive catabolism observed during illness and may be viewed as part of the acute phase response (5), which is one of the major defense mechanisms of the body predominantly mediated by cytokines. Proinflammatory cytokines, especially IL-1 $\beta$ , IL-6, TNF $\alpha$ , and interferon  $\gamma$ , inhibit several genes involved in thyroid hormone metabolism *in vitro* (6–8). In an attempt to induce NTIS, several experimental studies have reported the effects of administration of cytokines *in vivo* in both humans and experimental animals (9–11), resulting in altered thyroid hormone metabolism exhibiting some, but not all, features of disease-related NTIS (Table 1). In these studies, cytokine administration induced a flu-like illness. Therefore, it cannot be excluded that the resulting illness, rather than the cytokines *per se*, accounts for the observed changes in thyroid hormone metabolism (9–12). At this stage, a variety of mechanisms including altered thyroid hormone secretion, transport, and clearance, are known to contribute to NTIS (13).

Because the combination of reduced serum  $T_3$  and  $T_4$  levels indicates poor prognosis in severely ill patients, several investigators have focused on the changes in plasma thyroid hormone levels. However, clinical studies aimed at restoring plasma  $T_3$  by  $T_3$  and/or  $T_4$  treatment were generally not beneficial, and were sometimes even harmful (1), indicating that restoring plasma  $T_3$  levels does not equal improved tissue and organ function.

Recent evidence has made it clear that thyroid hormone action at the tissue level during illness is not a simple reflection of serum thyroid hormone concentrations. Instead, NTIS has differential effects on local thyroid hormone metabolism in various organs that appear to occur

quite independently from decreased serum  $T_3$  and  $T_4$  concentrations. The net effect of these differential changes is probably a major determinant of thyroid hormone availability and therefore of thyroid hormone action at the tissue level. A better understanding of the mechanisms underlying altered thyroid hormone metabolism in specific target tissues during illness is mandatory to renew our appreciation of NTIS, with the ultimate aim of improving disease outcome in critically ill patients.

## II. The Central Component of the HPT Axis

The combination of low serum thyroid hormone and TSH levels suggests central down-regulation of the hypothalamus-pituitary-thyroid (HPT) axis, which was supported by the observation in human autopsy samples that TRH gene expression is decreased in the hypothalamic paraventricular nucleus (PVN) of patients with NTIS. Furthermore, the decrease in postmortem TRH mRNA expression in the PVN showed a positive correlation with premortem serum TSH, in keeping with hypothalamic down-regulation of the HPT axis setpoint in severe illness (14). Clinical studies showed that decreased TSH release as well as serum  $T_3$  and  $T_4$  in critically ill patients could be restored to a large extent by the administration of exogenous TRH in combination with GH-releasing peptide (GHRP)-2. This neuroendocrine effect coincided with decreased serum markers of catabolism and increased markers of anabolism (15), suggesting that hypothalamic down-regulation of the HPT axis in patients with prolonged critical illness is an unfavorable condition amenable to treatment. However, the experimental induction of NTIS in rodents resulted in simultaneous changes in hypothalamic, pituitary, and peripheral thyroid hormone metabolism (16), arguing against down-regulation of the HPT axis at the central level as the primary and main determinant of the complex picture of NTIS in the whole organism.

### A. Hypothalamic response during illness

#### 1. Thyroid hormone transport

Cellular entry of thyroid hormone is necessary for intracellular conversion of thyroid hormone and for its binding to the nuclear thyroid hormone receptor (TR). Two categories of thyroid hormone transporters have been described, the organic anion transporters and the amino acid transporters. The organic anion-transporting polypeptide family consists of a variety of homologous proteins, of which OATP1C1 is expressed in brain capillaries involved in the uptake of  $T_4$  across the blood-brain barrier. The expression levels of OATP14 mRNA and protein are inversely related to thyroid hormone availability (17).

**TABLE 1.** Illness-induced alterations in serum  $T_3$ ,  $T_4$ ,  $rT_3$ , and TSH in various species

Species	$T_3$	$T_4$	$rT_3$	TSH	Refs.
Human	↓ ↓	N or ↓	↑	N or ↓	1
Mouse	↓	↓ ↓	N or ↓	N	11, 32, 145
Rat	↓	↓	—	↓	31, 58
Rabbit	↓	↓	↓	N	174, 183

↓, Decreased; ↑, increased; N, not affected; —, not reported.

The thyroid hormone transporter MCT8, belonging to the family of the amino acid transporters, transports  $T_4$  and  $T_3$  and is expressed in neuron populations in many brain areas, including cortical regions, striatum, cerebellum, and hypothalamus (18, 19). Thyroid hormones are probably not involved in the regulation of MCT8 in these neurons because hypothalamic MCT8 expression is not altered by hypothyroidism (18).

MCT10, another member of the amino acid transporter family, preferentially transports  $T_3$  instead of  $T_4$ . MCT10 is expressed in kidney, liver, and muscle (20). Recent studies reported MCT10 mRNA expression in primary cultures of mouse astrocytes (21) and the human hypothalamus (22), but its functional role in the central nervous system is still unclear.

Only very sparse data are available on the effects of NTIS on thyroid hormone transporters in the hypothalamus. A study by Mebis *et al.* (23) in a rabbit model of critical illness reported unaltered hypothalamic MCT8 mRNA expression and clearly increased MCT10 and OATP1C1 expression. Because these changes in transporter expression were not related to increased local tissue  $T_3$  concentrations, their functional consequences remain unclear at present (23).

## 2. Deiodinase expression

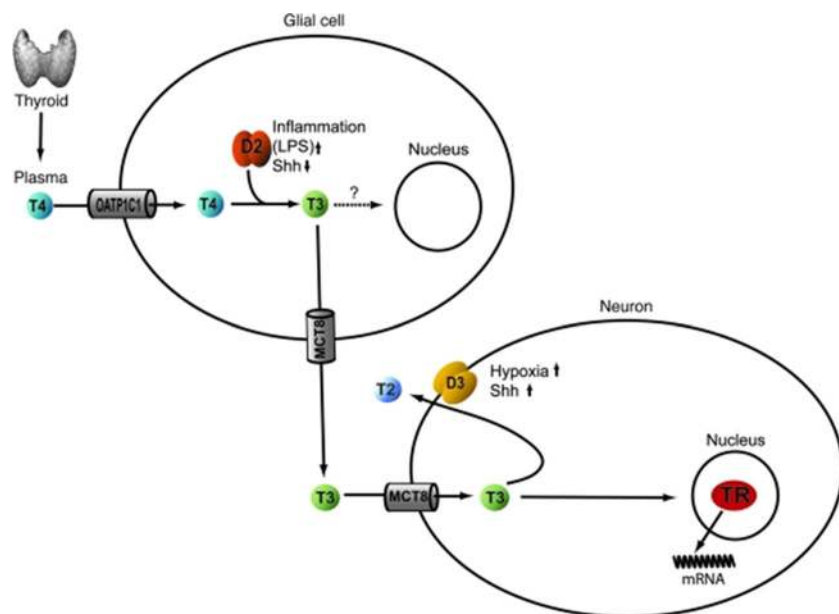
One of the major routes for metabolism of thyroid hormones is by deiodination through the iodothyronine deiodinases, together representing a selenocysteine-containing enzyme family consisting of three deiodinases, types 1 (D1), 2 (D2), and 3 (D3) (24). Both the inner (phenolic) ring and the outer (tyrosyl) ring of  $T_4$  can be deiodinated by the deiodinases, ultimately leading to the formation of 3,3'-diiodothyronine ( $T_2$ ). D1 is localized in the plasma membrane and is able to deiodinate both the inner and the outer ring of  $T_4$ . D1 expression is positively regulated by  $T_3$  and expressed in liver, kidney, thyroid, and pituitary (25, 26). D2 is localized in the endoplasmic reticulum and deiodinates  $T_4$  into the biologically active  $T_3$ . It is negatively regulated by thyroid hormone both pre- and post-transcriptionally because  $T_3$  down-regulates D2 mRNA expression (27), whereas  $T_4$  as well as  $rT_3$  (both substrates for D2) increase D2 ubiquitination and subsequent proteasomal degradation, resulting in decreased D2 activity (28). D3 is localized in the plasma membrane and can be viewed as the major thyroid hormone-inactivating enzyme because it catalyzes inner-ring deiodination of  $T_4$  and  $T_3$  exclusively, yielding the biologically inactive  $rT_3$  and  $rT_2$  (29).

Deiodinases are differentially expressed in several organs, including the brain, and their expression changes dramatically during NTIS (16). Both D2 and D3 are ex-

pressed in the human hypothalamus. D2 is mainly expressed in glial cells of the infundibular nucleus (IFN) and median eminence and in cells lining the third ventricle, whereas D3 expression is predominant in neurons of the PVN, supraoptic nucleus, and IFN (19). Studies in rodents have shown that hypothalamic D2 is a determinant of hypothalamic  $T_3$  production (30). In rodents, the administration of bacterial endotoxin [or lipopolysaccharide (LPS)], which is an established experimental model of acute NTIS, increases D2 mRNA expression (16) and enzyme activity in the hypothalamus, specifically in tanycytes lining the third ventricle (31). Increased D2 expression is not specific for the acute phase of illness because chronic inflammation in mice induces a short-lived increase of hypothalamic D2 expression as well (32), and hypothalamic D2 activity is up-regulated in mice with bacterial sepsis (33) and in rabbits with critical illness (23).

In contrast to the fasting-induced up-regulation of hypothalamic D2, which is related to the interplay between decreased serum leptin and increased serum corticosterone (34), the mechanism behind the induction of hypothalamic D2 during illness is still unclear. Remarkably, it appears to be independent of decreased serum thyroid hormone and increased corticosterone concentrations (35, 36). The dissociation between decreased serum thyroid hormone concentrations on the one hand and molecular changes in thyroid hormone metabolism at the tissue level on the other hand has been observed in many NTIS conditions (see *Sections III, IV and V*). Regarding the mechanism of hypothalamic D2 induction, activation of the nuclear factor  $\kappa B$  (NF $\kappa B$ ) pathway may be involved because the D2 promoter contains NF $\kappa B$ -responsive elements (31, 37). However, hypothalamic D2 activation precedes the hypothalamic up-regulation of NF $\kappa B$ , arguing against this notion (38). Moreover, inhibition of TNF $\alpha$  does not affect the LPS-induced stimulation of hypothalamic D2 despite reduced IL-6 levels, ruling out a major role for IL-6 or TNF $\alpha$ . A recent study showed that blocking TNF $\alpha$  does reduce the LPS-induced D2 response in a subpopulation of tanycytes located in the midportion of the third ventricle wall called  $\alpha$ -tanycytes. In these cells, LPS induces the expression of I $\kappa$ -B $\alpha$  (inhibitor of NF- $\kappa B$   $\alpha$ ), which is a sensitive marker for cytokine signaling through the NF $\kappa B$  pathway (38).

In contrast to hypothalamic D2, the expression of D3 decreases during both acute (39) and chronic inflammation (32), whereas hypothalamic D3 activity is unaltered, with a tendency to higher levels in rabbits with prolonged critical illness (23). Although  $T_3$  positively regulates D3 activity in brain (29), a causal role for decreased serum  $T_3$  in lowering hypothalamic D3 during inflammation can be excluded because serum  $T_3$  levels are similarly low in pair-



**Fig. 1.** Proposed model of thyroid hormone signaling in the brain during inflammation and hypoxic conditions. Three steps characterize this paracrine mechanism. First, the prohormone  $T_4$  enters the glial cells (astrocytes and tanycytes) via the thyroid hormone transporter OATP1C1. Subsequently,  $T_4$  is activated via D2, and the resultant  $T_3$  exits the glial cell compartment and enters adjacent neurons via the thyroid hormone transporter MCT8. Finally,  $T_3$  binds to neuronal TR and regulates transcriptional activity. LPS activates D2 transcription, and sonic hedgehog (Shh) promotes D2 inactivation via WSB-1-mediated ubiquitination; both hypoxia and Shh activate D3 gene transcription. [Adapted from B. C. Freitas *et al.*, Paracrine signaling by glial cell-derived triiodothyronine activates neuronal gene expression in the rodent brain and human cells. *J Clin Invest* 120:2206–2217, 2010 (50), with permission. © 2010, American Society for Clinical Investigation.]

fed control mice that do not display decreased hypothalamic D3 mRNA expression (32).

### 3. The role of the TR

$T_3$  is bound to the TR expressed in target tissues. TR are members of the nuclear receptor family and are encoded by two genes,  $TR\alpha$  and  $TR\beta$ . Due to alternative splicing and alternative promoter usage, the  $TR\alpha$  gene may give rise to six isoforms:  $TR\alpha 1$ ,  $TR\alpha 2$ ,  $TR\Delta\alpha 1$ ,  $TR\Delta\alpha 2$ , p43, and p28 (40). The  $TR\alpha 1$  isoform is the only *bona fide*  $TR\alpha$  because it has both a ligand binding and a DNA binding domain and modulates gene transcription (41–43).

The  $TR\beta$  gene encodes the  $TR\beta 1$  and  $TR\beta 2$  isoforms. In contrast to the  $TR\alpha$ ,  $TR\beta$  isoforms arise by alternative promoter usage exclusively, thereby differing in the N-terminal region (44). Both  $TR\beta 1$  and  $TR\beta 2$  bind  $T_3$  and are able to modulate gene transcription (40). TR are differentially expressed in various organs.  $TR\alpha 1$  is abundantly expressed in the brain (45), whereas the distribution of  $TR\beta$  in the brain is probably more restricted.  $TR\beta 2$  is expressed in a number of hypothalamic nuclei, including the human and rat PVN (46), and is assumed to be the predominant receptor involved in negative HPT axis feedback regulation (47).

Hypothalamic TR mRNA expression does not show dramatic changes during NTIS (23), but changes in specific TR isoform expression in PVN neurons have not been

carefully evaluated during NTIS. This could be of interest because the LPS-induced increase in hypothalamic D2 and decrease in hypothalamic D3 are exaggerated in  $TR\beta^{-/-}$  mice compared with wild-type (WT) mice (39), indicating that  $TR\beta$  modulates the hypothalamic deiodinase responses to acute inflammation. In line,  $TR\alpha^{0/0}$  mice do not display prominent changes in hypothalamic D2 and D3 responses to LPS (48).

### 4. Thyroid hormone action

Hypothalamic up-regulation of D2 and down-regulation of D3 during illness probably results in net increased local  $T_3$  production, which may explain the decreased TRH mRNA expression in the PVN as observed during illness both in animals and in humans (32, 46, 49). Recently, Bianco and colleagues (50) used an *in vitro* coculture system, confirming that glial D2 modulates  $T_3$  concentrations as well as neuronal D3 expression. The modulating role of LPS in these experiments reinforces the concept of increased hypothalamic  $T_3$  during

NTIS (schematically depicted in Fig. 1). Experiments in mice lacking  $TR\beta$  showed a less pronounced illness-induced decrease of hypothalamic TRH compared with WT, again supporting a role for  $T_3$  and  $TR\beta$  in TRH suppression during NTIS (39).

Very few studies have addressed hypothalamic  $T_3$  concentrations during NTIS directly. Mebis *et al.* (23) showed that prolonged illness in rabbits does not increase hypothalamic  $T_3$  tissue concentrations despite increased D2 expression, perhaps related to difficulties in selectively harvesting IFN and PVN samples for the assessment of  $T_3$  concentrations. In postmortem human hypothalamic specimens, however,  $T_3$  concentrations were even lower in NTIS patients compared with patients with acute death from trauma (51).

An additional factor in the pathogenesis of the illness-induced down-regulation of hypothalamic hypophysiotropic TRH neurons may be regulation by afferent pathways projecting from brainstem nuclei to the PVN. LPS is known to modulate autonomic centers in the lower brainstem, in turn activating hypophysiotropic CRH neurons in the PVN (52, 53). Like CRH neurons, hypophysiotropic TRH neurons receive neural input from the brainstem. However, because transection of these ascending pathways did not affect the LPS-induced decrease of TRH ex-

pression in the PVN, neural input from the brainstem appears to be relatively unimportant in this respect (54).

## B. Pituitary response during illness

### 1. Thyroid hormone transport

Few data have been reported on the expression and function of thyroid hormone transporters in the pituitary. A study in human anterior pituitary showed marked MCT8 immunostaining in the folliculostellate cells (55). Animal studies reported that the pituitary of MCT8-null mice is unable to sense adequately high serum  $T_3$ , although the pituitary of these mice seems to be euthyroid (56). The effect of NTIS on the pituitary expression of thyroid hormone transporters has not been reported yet.

### 2. Deiodinase expression

Both D1 and D2 are expressed in the rodent anterior pituitary, whereas the human pituitary expresses D2 and D3 (55). Although this may represent a species difference, another potentially important factor is that human studies rely on postmortem investigations. By definition, this implies some degree of NTIS, as well as a postmortem delay of at least a few hours. By contrast, the animal studies are performed in tissues harvested and processed immediately after acute death. The current view of negative feedback of thyroid hormone on pituitary TSH involves local D2-mediated conversion of  $T_4$  into  $T_3$ , which is subsequently bound by TR, finally resulting in repression of the TSH $\beta$  gene (55). The crucial role of pituitary D2 in TSH regulation is supported by impaired thyroid hormone feedback on TSH in D2 knockout mice (57). It is tempting to speculate that the mechanism for down-regulation of pituitary TSH $\beta$  during illness is similar to the repression of hypothalamic TRH during illness, *i.e.*, increased D2 expression with subsequent increased  $T_3$  production. In line with this assumption, proinflammatory cytokines increase D2 mRNA expression and activity in reagggregates of rat anterior pituitaries via activation of NF $\kappa$ B (5, 58). However, whereas hypothalamic D2 increases in various NTIS models studied so far, the pituitary D2 response to LPS shows a marked variation, depending on genetic background, species, and type of illness (58–60). Moreover, the LPS-induced D2 increase in the rat hypothalamus is independent of circulating thyroid hormone levels, whereas the increase in D2 activity in the anterior pituitary is absent when thyroid hormone levels are normal (35).

LPS treatment decreases pituitary D1. This response depends on proinflammatory cytokines because it is completely abolished in mice devoid of IL-12, IL-18, and the interferon  $\gamma$  receptor (59, 60), but not in TR $\beta^{-/-}$  mice (39). The LPS-induced decrease of pituitary D1 is difficult to reconcile with  $T_3$ -induced repression of the TSH $\beta$  gene.

### 3. The role of the TR

TR $\beta$ 2 protein is present in TSH-producing cells of the rat anterior pituitary (61, 62) and mediates the negative regulation of TSH $\beta$  mRNA expression by thyroid hormone (47). TR $\beta$ 2 is also implicated in the regulation of pituitary D1 by thyroid hormone (63). Acute inflammation decreases TR $\beta$ 2 mRNA expression in the pituitary (16). Because acute inflammation decreases pituitary D1 mRNA to the same extent in TR $\beta^{-/-}$  and WT mice (39), a role for TR $\beta$ 2 in the down-regulation of pituitary D1 during acute inflammation is unlikely. In line, the illness-induced decrease in pituitary D1 mRNA expression is abolished in mice lacking IL-12, IL-18, and interferon  $\gamma$ . Finally, pituitary D1 expression decreases to a larger extent in TR $\alpha^{0/0}$  mice upon LPS administration despite a similar acute phase response, pointing to a possible role for TR $\alpha$  in this respect (48).

### 4. Thyroid hormone action

One of the hallmarks of NTIS is the unresponsiveness of the pituitary to the low serum thyroid hormone levels, which implies that the physiological negative feedback mechanism of the HPT axis is altered in a major way during illness (13). Increased pituitary D2 mRNA expression may contribute to locally increased  $T_3$  concentrations, finally resulting in decreased pituitary TSH $\beta$  mRNA expression as has been observed in rodents. Alternatively, high  $T_3$  concentrations in the pituitary may result indirectly from increased D2 activity in hypothalamic tanycytes, increasing  $T_3$  concentrations in the portal capillary system, which may then suppress TSH $\beta$  mRNA expression in the anterior pituitary via the bloodstream (64). The assumption of elevated pituitary  $T_3$  concentrations during NTIS is, however, not in line with observations in human pituitary specimens showing low pituitary  $T_3$  concentrations in patients with NTIS compared with control patients who died from sudden trauma (51).

A role for the TR, and thus for  $T_3$ , is strongly supported by the observation that lacking the TR $\beta$  gene prevents the LPS-induced decrease of pituitary TSH $\beta$  mRNA (39). A role for D2 in altered TSH expression during illness is questionable, however, because LPS decreases TSH $\beta$  mRNA expression in the pituitary whereas D2 expression may decrease (16) or increase (39).

The administration of exogenous TSH to mice during LPS-induced NTIS attenuates the decline in serum  $T_4$  (65), strengthening the concept that central down-regulation of the HPT axis is one of the determinants of the decrease in serum thyroid hormone levels.

Recently, the existence of two novel glycoprotein subunits,  $\alpha$ 2 (GPA2) and  $\beta$ 5 (GPB5), was reported, assumed to heterodimerize into a glycoprotein hormone coined

“thyrostimulin.” Thyrostimulin activates the TSH receptor (TSHR) both *in vitro* and *in vivo* (66). Because GPA2 and GPB5 colocalize in the rat and human anterior pituitary where the TSHR is expressed, a paracrine role for thyrostimulin was suggested (67, 68). However, the putative biological role for the heterodimer *in vivo* is still a matter of debate (69–71). Because proinflammatory cytokines (TNF $\alpha$  and IL-1 $\beta$ ) regulate transcription of the GPB5 subunit via activation of NF $\kappa$ B in a murine pituitary corticotroph cell line, Suzuki *et al.* (72) postulated that thyrostimulin may play a role in the pathogenesis of NTIS. Subsequent *in vivo* studies in mice showed LPS-induced up-regulation of GPB5—but not GPA2—transcription in the pituitary and hypothalamus, whereas lacking the GPB5 subunit reduced LPS-induced suppression of TSHR mRNA in the same tissues. Although these observations confirmed a role for GPB5 in the regulation of TSHR expression during acute illness, this was not reflected in illness-induced alterations of serum thyroid hormone levels (73), again pointing to a dissociation between low serum thyroid hormone levels during NTIS and local thyroid hormone metabolism at the organ level.

### III. Differential Effects of Acute Inflammation on Thyroid Hormone Metabolism in Metabolic Organs

Although there is no question that illness profoundly affects the central part of the HPT axis, it is still unclear whether these hypothalamic and pituitary changes suffice to induce the marked decrease in serum thyroid hormone levels observed during NTIS. Interestingly, recent experimental studies have shown that down-regulation of the central part of the HPT axis observed during inflammation does not necessarily induce decreased thyroid hormone metabolism in key metabolic organs such as liver, muscle, and adipose tissue.

Local thyroid hormone metabolism during acute inflammation has not been studied extensively in humans. The effects of major surgery on the HPT axis have been investigated as a model for acute NTIS because major surgery induces a rapid inflammatory response characterized by activation of neutrophils and the release of a variety of proinflammatory cytokines (74–76). For example, patients undergoing cardiac surgery show significant alterations in serum T<sub>3</sub>, T<sub>4</sub>, and rT<sub>3</sub> concentrations and in T<sub>3</sub>/rT<sub>3</sub> and T<sub>3</sub>/T<sub>4</sub> ratios, suggestive of impaired thyroid hormone conversion (77). Nevertheless, animal studies are still needed to determine the effects of acute inflammation on thyroid hormone metabolism within metabolic organs. For this purpose, LPS can be used as an established animal model of acute NTIS. LPS elicits a strong inflam-

matory response characterized by the production of a variety of cytokines such as TNF $\alpha$ , IL-1, and IL-6 via the activation of signaling pathways like NF $\kappa$ B and activator protein-1 (AP-1) (78).

#### A. Hepatic thyroid hormone metabolism during acute illness

One of the key target organs of thyroid hormone is the liver, where it acts as a determinant of hepatic glucose and lipid metabolism. In liver, TR $\beta$ 1, TR $\alpha$ 1, and TR $\alpha$ 2 isoforms are abundantly expressed, and approximately 60% of liver T<sub>3</sub>-regulated genes are TR $\beta$ -dependent. Furthermore, both D1 and D3 are expressed in liver, although D3 is expressed only at very low levels in a healthy liver. For many years, liver D1 was thought to be the major source of plasma T<sub>3</sub> resulting from deiodination of T<sub>4</sub> (1). Because liver D1 expression decreases during acute inflammation in rodents, this was assumed to contribute significantly to the low serum T<sub>3</sub> levels observed during illness (16). Liver D1 regulation is primarily driven via the TR $\beta$  (79, 80), but only sparse *in vitro* work has been performed on the effects of cytokines on liver TR expression. TR binding capacity is decreased in the human hepatoma cell line HepG2 after stimulation with TNF $\alpha$ , IL-1, and IL-6 (81), whereas TR $\beta$  and TR $\alpha$  mRNA expression decrease in HepG2 cells upon stimulation with IL-1 $\beta$ . The IL-1 $\beta$ -induced decrease of TR $\beta$  is exclusively mediated by the NF $\kappa$ B pathway, whereas the IL-1 $\beta$ -induced decrease of TR $\alpha$  mRNA is abolished only by simultaneous inhibition of NF $\kappa$ B and AP-1 (82, 83). *In silico* analysis (84) revealed the presence of three NF $\kappa$ B-responsive elements in the TR $\beta$  promoter, which may explain the NF $\kappa$ B-dependent repression of the TR $\beta$  gene. The IL-1 $\beta$ -induced decrease of TR $\alpha$ 1 and TR $\alpha$ 2 expression is a direct effect of decreased promoter activity (83). The exact mechanism remains unknown, but may involve phosphorylation-dependent repression of the TR $\alpha$  promoter. In animal studies, liver TR $\alpha$  and TR $\beta$ 1 expression decreases after LPS administration (16, 85).

Because the D1 gene is activated via a TR/retinoid X receptor (RXR) heterodimer, decreased D1 expression during acute inflammation may result from reduced TR expression. Alternatively, the decrease in nuclear RXR $\alpha$  protein may play a role because this protein decreases after LPS administration due to rapid nuclear export via Jun N-terminal kinase (JNK) phosphorylation and subsequent proteasomal degradation (86). Because the IL-1 $\beta$ -induced decrease of D1 mRNA is not prevented by inhibition of JNK alone (82), it is unlikely that decreased RXR expression is the only factor responsible for the decrease in D1 mRNA observed during acute illness. In addition, because lacking TR $\beta$  does not prevent the illness-induced decrease in liver D1 mRNA either, a key role of TR $\beta$  in this respect

is unlikely as well (87). Surprisingly, experiments in  $TR\alpha^{0/0}$  mice showed that the down-regulation of liver D1 is partly mediated via the  $TR\alpha$  (88), probably resulting from diminished D1 promoter activity due to the effect of cytokines (89, 90).

An alternative mechanism for  $TR\beta$ -mediated repression of liver D1 during acute inflammation has been proposed by Yu *et al.* (91, 92), who showed both *in vivo* and *in vitro* that adding exogenous coactivator steroid receptor coactivator-1 (SRC-1) attenuates the illness-induced liver D1 decrease. These studies indicate that competition for limiting amounts of SRC-1, which is a shared coactivator for TR and inflammatory signaling pathways, is one of the mechanisms involved in the illness-induced D1 decrease (Fig. 2). Interestingly, restoration of liver D1 expression by exogenous SRC-1 in the study by Yu *et al.* (91) prevented the fall in serum thyroid hormone levels after LPS. Deiodinating enzymes require an endogenous intracellular thiol cofactor for catalytic activity. Glutathione (GSH) has been proposed to be one of these factors necessary for D1 activity *in vivo*. As a consequence, changes

in the intracellular concentrations of GSH may play a role in the regulation of D1 during NTIS. A recent study showed  $T_3$  production by D1 in intact cells to be suppressed by IL-6. The addition of N-acetyl-cysteine, an antioxidant that restores intracellular GSH levels, prevented the IL-6-induced inhibitory effect on D1-mediated  $T_3$  production. This suggests that the effect of IL-6 on D1 activity is mediated by intracellular GSH (93).

However, a dominant role for impaired type 1 deiodination in lowering serum  $T_3$  levels is not supported by the observation that illness-induced alterations in serum  $T_3$  and  $T_4$  levels are similar between WT and  $TR\beta^{-/-}$  mice despite marked differences in liver D1 mRNA expression and activity (87).

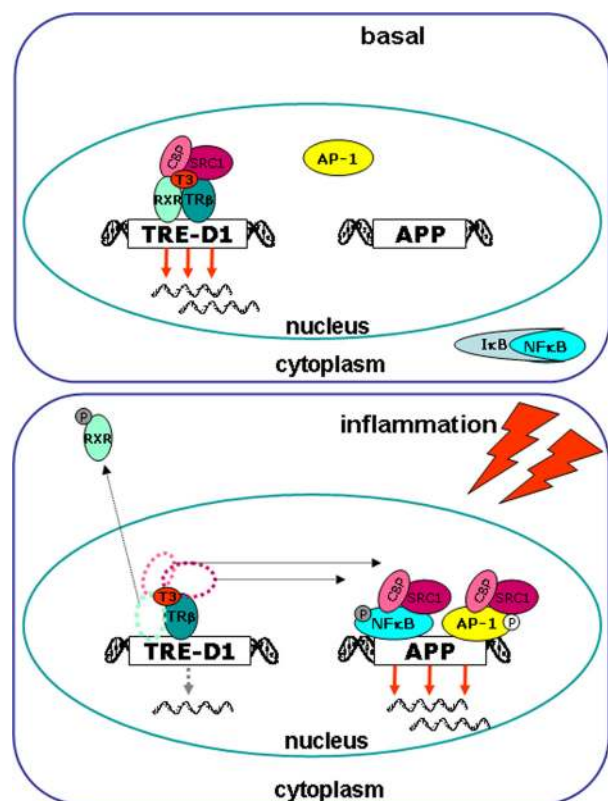
Similar to hypothalamic D3, liver D3 decreases during acute inflammation (94). This may be mediated via  $TR\beta$  because the LPS-induced decrease in hepatic D3 is abolished in  $TR\beta^{-/-}$  mice and aggravated in  $TR\alpha^{0/0}$  mice (88). These studies suggest a dominant role of  $TR\beta$  in the illness-induced regulation of liver D3 because  $TR\alpha^{0/0}$  mice have more sensitive  $TR\beta$  signaling (80). Involvement of  $TR\beta$  in the regulation of hepatic D3 suggests a role for  $T_3$ . However, the time course of the changes in serum  $T_3$  and liver D3 expression after LPS administration does not support this (94).

To date, no *in vitro* studies reporting decreased D3 expression upon cytokine administration have been published. By contrast, TGF- $\beta$  stimulates D3 expression in primary human fibroblasts, and this effect is abolished by inhibition of the MAPK ERK and p38 (95).

The decrease in liver D3 does not appear to contribute to changes in serum thyroid hormone levels because illness-induced alterations in serum  $T_3$  and  $T_4$  are similar in  $TR\beta^{-/-}$  and WT mice despite differences in liver D3 expression (87). This was recently supported by a similar decrease of serum thyroid hormone levels in *Streptococcus pneumoniae*-infected D3 knockout (D3KO) mice compared with WT mice (33).

Liver MCT8 and MCT10 transporter expression has not been studied extensively. Mebis *et al.* (96) studied MCT8 and MCT10 expression in postmortem liver biopsy samples of prolonged critically ill patients compared with patients after elective abdominal surgery. They observed lower liver MCT8 mRNA expression during acute surgical stress compared with prolonged illness, whereas liver MCT10 expression did not differ between the groups. For obvious reasons, it was not possible to compare hepatic transporter expression in both groups to healthy controls, which makes the study difficult to interpret (96).

The combination of low liver D1 and low liver D3 should theoretically result in only moderate changes in liver  $T_3$  content. It is tempting to speculate that low liver



**Fig. 2.** Schematic model of the proposed mechanisms involved in illness-induced down-regulation of liver D1 expression. In this model, limited amounts of SRC-1, a coactivator for both  $NF\kappa B$  and  $TR\beta$ , decrease liver D1 mRNA expression as a result of competition between cytokine-induced  $NF\kappa B$  and the  $TR\beta$ /D1 complex for SRC-1. Note the role of decreased nuclear  $RXR\alpha$  protein by LPS-induced rapid nuclear export via JNK-phosphorylation and subsequent proteasomal degradation. APP, Acute phase protein; TRE, thyroid hormone responsive element.

D3 activity during the acute state of illness compensates for reduced production of  $T_3$  by D1 and thereby helps to maintain organ function. To our knowledge, no human or animal studies have been performed reporting liver thyroid hormone concentrations during the acute state of inflammation. As an indirect measure of local  $T_3$  availability, the determination of  $T_3$ -regulated gene expression in liver during acute illness is an interesting option because it reflects local thyroid status.  $T_3$  is known to stimulate a number of genes, *e.g.*, liver malic enzyme (ME) (97), Spot 14 (98), and cholesterol 7 $\alpha$ -hydroxylase (Cyp7a) (99). It has been shown that LPS results in a rapid decrease of liver Cyp7 $\alpha$  mRNA expression in Syrian hamsters (100) and liver ME and Spot 14 mRNA expression in mice (85). These observations are in keeping with reduced  $T_3$  action in the liver during acute inflammation, possibly reflecting decreased  $T_3$  liver content. However, several caveats should be taken into account: liver TR expression decreases during the acute phase of illness (16, 85), making it impossible with this approach to discriminate between reduced  $T_3$  action and reduced  $T_3$  content. Furthermore, the kinetics of the observed decreases in  $T_3$ -regulated gene expression are in disagreement with low liver deiodinase activity levels, making a key role for diminished  $T_3$  availability less likely. Future studies reporting liver  $T_3$  content during acute inflammation are needed.

### B. Muscle thyroid hormone metabolism during acute illness

Thyroid hormones are key players in skeletal muscle function because a variety of genes expressed in muscle are positively or negatively regulated by  $T_3$ . Skeletal muscles consist of a mixture of muscle fiber types. The type I fibers are innervated by slow motor neurons and found in low-twitch muscles. These fibers are rich in mitochondria and rely on oxidative phosphorylation for ATP generation. Lipids are their preferred substrate, whereas glycogen content is low. By contrast, type II fibers are active during high-frequency contraction and in need of high ATP turnover resulting from glycolysis (type IIA) or anaerobic glycolysis yielding lactate (type IIB). Most skeletal muscles contain both type I and II fibers. Muscle activation starts with the recruitment of type I fibers, which is followed by the activation of type II fibers. Type I fibers are highly dependent on thyroid hormone status, whereas type II fibers act relatively independent of thyroid hormones (101). The molecular machinery expressed in muscle required for thyroid action includes D2, D3, TR $\alpha$ 1 [the major TR isoform in muscle (102)], TR $\beta$ 1, three RXR isoforms, and the thyroid hormone transporters MCT8 and MCT10 (96, 103). Human skeletal muscle D2 was reported to be involved not only in local  $T_3$  production but also in periph-

eral production of  $T_3$  in the basal condition (104). However, this concept was recently challenged because D2 is expressed only at very low levels under euthyroid conditions in healthy persons (105). Interestingly, muscle D2 mRNA expression is modulated by fasting and by insulin serum levels (106), suggesting that muscle D2 is involved in energy metabolism. Several recent studies have reported alterations in muscle thyroid hormone metabolism during NTIS, both in acute and in more chronic models.

Acute inflammation in rodents increases D2 mRNA expression in skeletal muscle, whereas the effects on D2 activity are unknown at this stage (87). The increase in muscle D2 transcription is mirrored by a decrease in D3 mRNA and TR $\alpha$ 1 mRNA expression (88). TR expression has also been studied in the diaphragm, which is a ventilatory muscle with high energy demands, relying on lipid oxidation. A sublethal dose of LPS to mice profoundly decreases TR $\alpha$ 1 and TR $\beta$ 1 mRNA expression in the diaphragm in association with decreased expression of co-activators involved in nuclear receptor activation (107).

The role of TR isoforms in altered muscle D2 and D3 expression has been studied in TR knockout mice. The LPS-induced decrease of muscle D3 is attenuated in TR $\alpha^{0/0}$  mice, suggesting the involvement of TR $\alpha$ . The regulation of muscle D3 via TR $\alpha$  is supported by lower basal muscle D3 levels in TR $\alpha^{0/0}$  mice, but difficult to reconcile with the observation that LPS decreases TR $\alpha$  expression in forelimb but not in hindlimb muscle, whereas D3 is affected in both limbs. The LPS-induced increase of muscle D2 is more pronounced in TR $\beta^{-/-}$  mice, suggesting that TR $\beta$  and/or  $T_3$  partly suppress D2 induction during illness (80).

Comparison of MCT8 and MCT10 expression in muscle tissue of prolonged critically ill patients with patients after elective abdominal surgery showed remarkably low MCT8 mRNA expression during acute surgical stress compared with prolonged illness. By contrast, MCT10 expression was similar between the groups. Acute surgical stress patients displayed somewhat lower serum  $T_3$  levels compared with reference values. However, because MCT8 and MCT10 expression levels are unknown in muscle of healthy persons, no conclusion can be drawn at this stage about the possible role of these thyroid hormone transporters in altered serum concentrations of thyroid hormone during NTIS (96). The same authors also studied D2 mRNA expression and activity in skeletal muscle tissue of patients with acute surgical stress compared with a healthy control group. A significant decrease in serum  $T_3$  levels was observed during the surgical procedure, but D2 mRNA expression and activity in muscle were unaltered compared with controls (108).

In summary, D2 increases whereas D3 decreases in skeletal muscle during acute inflammation. The changes in



deiodinase expression coincide with decreased TR expression. The combination of increased D2 and decreased D3 is likely to result in increased local  $T_3$  availability. Two important questions that should be addressed are whether this assumption is supported by any significant changes in  $T_3$  target gene expression in muscle and whether the observed changes in thyroid hormone metabolism in muscle are clinically relevant.

Few data have been reported on expression levels of  $T_3$  target genes in muscle tissue during acute inflammation. Muscle uncoupling protein (UCP) 3, a  $T_3$  target gene, has been studied in a rat model of sepsis involving cecal ligation and puncture, which is an acute sepsis model characterized by a systemic inflammatory response. Both UCP3 mRNA and protein expression increased in skeletal muscles of rats 16 h after cecal ligation and puncture (109). Likewise, LPS administration to mice resulted in increased UCP3 mRNA expression in skeletal muscle, preceded by a rapid increase of the  $T_3$ -responsive transcription factor PGC-1 (110). Similarly, a study in humans showed increased UCP3 expression in skeletal muscle of NTIS patients, coinciding with increased D2 mRNA expression, although a variety of other  $T_3$  target genes (SERCA, MHC, and GLUT4) were unaltered (111). In contrast, GLUT4 expression decreased after LPS administration in the diaphragm of mice (107).

Taken together, these results suggest a selective increase in  $T_3$  signaling in skeletal muscle tissue during acute inflammation. A complicating factor is simultaneous downregulation of TR $\alpha$ 1 and TR $\beta$ 1 mRNA expression in muscle during acute inflammation, which is contradictory at first sight. However, whereas serum  $T_3$  contributes approximately 50% to TR occupancy in most tissues, TR saturation in D2-expressing tissues is much higher due to local generation of  $T_3$  (29). The TR heterodimer associates, in the absence of  $T_3$ , with corepressor proteins, which results in the repression of basal transcriptional activity (aporeceptor function) (112). Thus, decreased TR expression combined with increased production of local  $T_3$  by D2 is likely to result in near maximal occupancy of the TR. The resulting maximal activation of  $T_3$  target genes supports local activation of  $T_3$ -regulated genes.

The clinical relevance of increased muscle  $T_3$  signaling is unclear at present. Thyroid hormone promotes the shift from slow to fast muscle fiber types and regulates contractibility (101).  $T_3$  also increases the glycolytic and oxidative capacities of skeletal muscle and activates the insulin-activated glucose transporter GLUT4. An association between a mutation in the D2 gene (*Dio2*Thr92Ala) and insulin resistance has been established in healthy volunteers (113). This polymorphism is associated with diminished muscle D2 activity, sug-

gesting that reduced local  $T_3$  availability might be involved in insulin resistance (114). Of note, a recent case-control study and meta-analysis showed an association between the Thr92Ala polymorphism in the *Dio2* gene and type 2 diabetes, confirming earlier reports of an association with insulin resistance (115). Because acute inflammation results in hyperinsulinemia in rodents (116) and activation of the TLR4 by LPS in skeletal muscle induces a shift in substrate handling from lipids (preferred substrate type 1 fibers) to glycogen (preferred substrate type 2 fibers), a functional role for altered  $T_3$  availability in the adaptation of muscle metabolism and function during NTIS seems likely (107, 117).

Another potentially relevant aspect is the capacity of both  $T_3$  and  $T_2$  to increase mitochondrial activity via cytochrome c oxidase (118, 119). In addition,  $T_3$  is able to induce transcription of the mitochondrial genome via p43, a truncated isoform of the TR $\alpha$ , which is located in the mitochondrial matrix. Finally, stimulation of mitochondrial biogenesis probably involves genomic action of  $T_3$  via TR $\alpha$ 1 or TR $\beta$ 1 (120). This complex interaction between thyroid hormone and mitochondrial function makes it very difficult to define a role for  $T_3$  in skeletal muscle mitochondria during acute illness, without the availability of experimental data to discriminate between short-term and long-term effects, and between non-genomic and genomic effects. However, it is tempting to speculate at this stage that  $T_3$  is beneficial in improving muscle function in the acute state of illness by affecting a variety of target genes relevant for muscle metabolism and mechanical function.

### C. Thyroid hormone metabolism in adipose tissue during acute illness

Adipose tissue is a key metabolic organ that stores energy-rich fuels mainly as triglycerides (TG). In times of energy shortage, lipolysis is stimulated both via the central and autonomic nervous system (adrenaline and noradrenaline) and via endocrine routes. Acute inflammation stimulates lipolysis in adipose tissue while decreasing TG synthesis, explaining in part the increased serum free fatty acid levels observed during acute illness (121).

Thyroid hormone is one of the major determinants of lipid metabolism. For instance, thyrotoxicosis increases TG-derived fatty acid uptake in muscle and heart, whereas hypothyroidism increases TG-derived fatty acid uptake in white adipose tissue in association with increased lipoprotein lipase activity (122). White adipose tissue expresses multiple genes involved in thyroid hormone transport and metabolism, such as MCT8, D1, D3, TR $\alpha$ 1, TR $\beta$ 1, RXR $\gamma$ , and a variety of coactivators in-

volved in TR signaling (121), making a metabolic role for local thyroid hormone metabolism likely.

The effect of acute inflammation on thyroid hormone metabolism in adipose tissue has not been extensively studied. No data are available on illness-induced alterations in deiodinase expression. As for effects on TR, LPS administration to mice suppresses the mRNA expression in white adipose tissue of a variety of nuclear hormone receptors, including TR $\alpha$ 1 and TR $\beta$ 1. This is associated with a decrease in coactivators and reduced expression of ME and Spot 14, both T<sub>3</sub> target genes (121). In addition, LPS markedly decreases GLUT4 and phosphoenolpyruvate carboxykinase expression, reducing glycerol-3-phosphate production in adipose tissue, which is supposed to limit TG synthesis. Limiting TG synthesis in white adipose tissue may help to shuttle free fatty acids to the liver, promoting TG synthesis and very low-density lipoprotein production in liver necessary for an adequate acute phase response (123). Against this background, reduced T<sub>3</sub> action in white adipose tissue during acute illness might be a beneficial adaptation. Although one study suggests a link between altered thyroid hormone and lipid metabolism in adipose tissue during acute illness (121), the precise role of thyroid hormone has yet to be established.

#### IV. Differential Effects of Chronic Inflammation and Sepsis on Thyroid Hormone Action in Metabolic Organs

Chronic inflammation and sepsis are associated with markedly decreased plasma thyroid hormone levels. The severity of illness is reflected in the magnitude of the decrease in serum T<sub>3</sub>. In critical illness, defined as a clinical condition requiring prolonged support for failing vital organs (mechanical ventilation, need for inotropics), serum T<sub>3</sub> may become very low or even undetectable. In severely ill patients, serum T<sub>4</sub> decreases as well and is inversely correlated with mortality (1). Human studies in the intensive care unit (ICU) setting have shown that decreased serum thyroid hormone levels are associated with changes in deiodinase expression in liver and muscle. From recent animal experiments, the mechanism of these changes in muscle and liver appears to be very different from the changes in acute inflammation. Studying chronic inflammation in experimental animals requires a rigorous methodology because reduced food intake during illness interferes in a major way with thyroid hormone metabolism at the organ level (106, 124, 125). Interestingly, the molecular effects of chronic inflammation and sepsis on thyroid hormone metabolism appear to depend on the organ studied.

#### A. Hepatic thyroid hormone metabolism during chronic inflammation

The effects of chronic inflammation on hepatic thyroid hormone metabolism have been studied in rodents by inducing a sterile abscess with turpentine inoculation in a hindlimb. This experimental model resembles local tissue injury and inflammation and leads to serial activation of specific proinflammatory cytokines that are associated with decreased serum thyroid hormone levels and reduced food intake (32, 94, 126). In contrast to LPS-induced acute inflammation, liver D1 mRNA expression is unaffected in mice with chronic inflammation, whereas liver D3 mRNA expression and activity decrease in both models (94). A role for cytokines in decreasing liver D3 expression is unlikely because the decrease in liver D3 mRNA precedes the increase in liver IL-1 $\beta$  mRNA. Corticosteroids may be involved because dexamethasone acutely decreases D3 expression in chicken liver (127), whereas both acute and chronic inflammation increase serum corticosterone (39, 128). The role of diminished food intake on the observed alteration in liver deiodinase expression is unknown at present but may certainly interfere because it has been shown that food restriction in chickens increases liver D3, whereas liver D1 is not affected (129). The role of T<sub>3</sub> can be excluded because serum T<sub>3</sub> decreases to a similar extent in mice with chronic inflammation and pair-fed controls, whereas serum T<sub>4</sub> and rT<sub>3</sub> levels are lower during inflammation than in pair-fed controls (32). Apart from the effect on deiodinases in liver, no consistent changes in liver TR $\alpha$ 1, TR $\alpha$ 2, and TR $\beta$ 1 mRNA expression are reported in turpentine-treated rats (130).

In the clinical setting, Chamba *et al.* (131) studied hepatic TR $\alpha$ 1, TR $\alpha$ 2, and TR $\beta$ 1 and T<sub>3</sub> target gene expression in patients with a variety of chronic liver diseases and found no changes in TR or T<sub>3</sub> target gene expression despite reduced serum thyroid hormone levels, indicating a euthyroid condition in the liver of these patients. These findings are in line with the chronic inflammation mouse and rat models published to date and represent another example of a dissociation between serum thyroid hormone levels and thyroid hormone action at the organ level during NTIS.

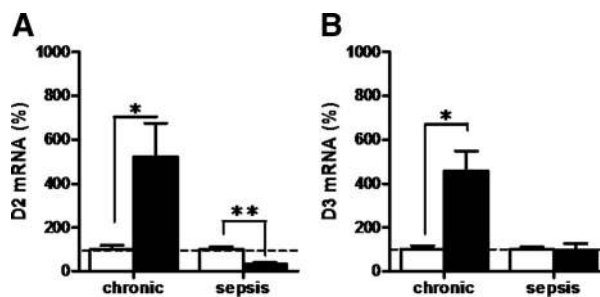
#### B. Muscle thyroid hormone metabolism during chronic inflammation

Although the animal model of chronic inflammation needs inoculation of turpentine in the hindlimb, which results in an abscess located between muscle and subcutis, the inflammatory response is not restricted to the injected muscle and can be characterized as a systemic acute phase response. In these mice, D2 mRNA expression and activity increase in the (noninjected) forelimb muscle. This phenomenon is unrelated to serum thyroid hormone levels,

proinflammatory cytokines, and activation of inflammatory pathways or D2-ubiquitinating factors. However, activation of the cAMP pathway does coincide with the simultaneous induction of D2 in muscle tissue of chronically inflamed mice (132), pointing to a role of cAMP in D2 induction as has been reported in brown adipose tissue (133). Unexpectedly, D3 mRNA expression and activity increase simultaneously with D2 (132). Dentice *et al.* (134) studied the effects of local muscle injury induced by a snake toxin on D2 mRNA expression and activity. Without any effect on serum thyroid hormone levels, muscle D2 appeared to increase within 1 wk and was followed by regeneration of muscle tissue. Interestingly, muscle regeneration was significantly impaired in *Dio2*<sup>-/-</sup> mice, although no morphological differences were observed early after injury between *Dio2*<sup>-/-</sup> and WT mice. Thus, injury-induced D2 induction in muscle is likely to be an adaptive response. Forkhead O3 (FoxO3), a forkhead box transcription factor involved in the regulation of a variety of cellular functions such as differentiation and proliferation, is probably important for D2 induction during differentiation (134). It is unknown at present whether FoxO3 also plays a role in turpentine-induced D2 induction in muscle. Recent studies reported D2 up-regulation in skeletal muscle by bile acids via the G protein-coupled receptor TGR5, followed by an increase in mitochondrial activity (135). In addition, activation of the peroxisome-proliferator activated receptor  $\gamma$ , which is an important metabolic regulator, increases D2 expression in skeletal muscle cells as well (136). The role of bile acids during chronic inflammation in muscle is not completely understood at present.

Muscle thyroid hormone metabolism has also been studied in patients with noninflammatory NTIS and in patients undergoing hip or knee replacement. Skeletal muscle biopsies in the NTIS group were taken within the first 48 h after ICU admission and in the control group during hip or knee surgery. Thus, the design of this study does not allow for a classification as either chronic or prolonged illness. In the NTIS patients, serum  $T_3$  was lower, whereas serum  $rT_3$ ,  $TNF\alpha$ , IL-6, IL-8, and IL-2R were higher despite similar  $NF\kappa B$  activation. Skeletal muscle displayed lower  $TR\alpha 1$ ,  $TR\beta 1$ ,  $RXR\gamma$ , and D2 mRNA expression compared with controls, whereas MCT8 expression did not differ (111), suggestive of decreased thyroid hormone action in this model.

In contrast to acute illness that leads to increased D2 and decreased D3 expression, chronic inflammation results in simultaneously increased D2 and D3 activity. This will inevitably result in a coordinated breakdown of both  $T_4$  and  $T_3$ , finally leading to increased intracellular  $T_2$  levels as depicted in Fig. 3 (137). Both  $T_3$  and  $T_2$  are known



**Fig. 3.** Muscle D2 (A) and D3 (B) mRNA expression during chronic inflammation (turpentine injection in the hindlimb) and sepsis (intranasal inoculation of *S. pneumoniae*). Control mice are pair-fed. Dark bars represent inflammation at 48 h, and open bars represent saline-treated controls. Bars represent mean values ( $n = 6$ ) and SEM of mRNA expression relative to HPRT, which is a housekeeping gene. The dotted line indicates expression in the control group set at 100%. Differences between specific groups are indicated by \*,  $P < 0.05$ ; and \*\*,  $P < 0.01$ . (Data summarized from Ref. 132.)

to regulate mitochondrial activity in skeletal muscle (118, 119), and the question is whether a shift toward local  $T_2$  and away from local  $T_3$  production might be relevant.

We have proposed that increased muscle  $T_3$  concentration might be beneficial during acute illness by supporting the illness-induced shift in substrate handling from lipids to glucose (107, 117) and by increasing mitochondrial activity (118, 119). Transition from the acute to chronic state might be accompanied by alterations in the metabolic state of skeletal muscles. The IL-1-induced increase in plasma lactate and lactate content in skeletal muscle probably lowers local pH (138).  $T_2$  activates cytochrome c oxidase most efficiently at pH 6.4, whereas for  $T_3$  the optimal pH is 7.4 (139). Thus, increased muscle  $T_2$  production during chronic inflammation may occur as an adaptation to altered muscle metabolism.

### C. Hepatic thyroid hormone metabolism during bacterial sepsis

Sepsis, a whole-body inflammatory state caused by bacterial infection, is a major complication in critically ill patients associated with poor outcome (140). Bacterial peritonitis and pneumonia are the two most common causes of sepsis (141, 142). It is well known that sepsis is associated with profound alterations in thyroid hormone serum levels in both humans and experimental animal models (143–145), closely resembling those observed in acute inflammation. Among the differences between experimental sepsis and LPS-induced acute inflammation are the magnitude of the inflammatory response and the severity of illness because sepsis is often lethal, whereas LPS-induced acute inflammation is a sublethal model. Sepsis is characterized by activation and deactivation of the systemic inflammatory response. Activation is reflected by the production of proinflammatory cytokines, increased endothelial activation, and activation of immune effector

cells like granulocytes, monocytes, and lymphocytes. The dampening of the inflammatory response, on the other hand, is characterized by the production of antiinflammatory cytokines like IL-10, IL-1 receptor antagonist, and by impaired up-regulation of granulocytes in response to pro-inflammatory signals. Uncontrolled activation of both pro- and antiinflammatory responses can lead to cell exhaustion, organ dysfunction, and death (146).

Liver thyroid hormone metabolism has not been studied extensively in sepsis patients. One study in critically ill children with sepsis or septic shock showed the expected profound alterations in serum thyroid hormone concentrations. Based on these changes, the authors suggested altered peripheral deiodination including an induction of liver D3 and inhibition of liver D1 as an explanation (147). However, this pattern was not confirmed by animal experimental studies performed in well-characterized sepsis models. Experimental sepsis can be induced by inoculation with *S. pneumoniae*, which is an important pathogen in a majority of clinical pneumonia cases (148). *S. pneumoniae* administration to mice induces severe bronchopneumonia and stimulates the production of TNF $\alpha$ , IL-1, IL-6, IL-10, and the chemokines (cytokine-induced neutrophilic chemoattractant) and macrophage inflammatory protein-2 (14, 149). In this sepsis model, serum thyroid hormone levels and TSH decrease in association with decreased expression of both liver D1 and D3 (33). Liver D3 mRNA and activity have also been studied in abdominal sepsis model induced by *Escherichia coli*, an enteric Gram-negative bacterium (150). Again, in this model liver D3 expression was decreased (151).

A requirement for appropriate D1 synthesis is the availability of selenium (Se). This trace element is critical to the function of D1, although studies have shown that Se deficiency only affects liver D1 activity and not thyroidal and pituitary D1. Despite the fact that plasma Se levels are decreased in severe illness, Se substitution to patients with severe sepsis failed to restore serum thyroid hormone levels (152).

The putative role of deiodinating enzymes in the observed alterations in serum thyroid hormone concentrations has been studied in D3 null (D3KO) mice. These mice display similar changes in thyroid hormone concentrations during sepsis as WT mice, which is a strong argument against a pivotal role for D3 in the pathogenesis of low serum thyroid hormone levels during sepsis. In addition, basal liver D1 activity is relatively low in D3KO mice and not responsive to sepsis, suggesting that changes in liver D1 are not critical either for the decrease in serum thyroid hormone levels (33).

In summary, liver thyroid hormone metabolism during experimental sepsis is characterized by decreased D1 and

D3 activity and accompanied by unaltered TR $\beta$ 1 expression. Because no information is present about liver T<sub>3</sub> content during sepsis, it is still puzzling whether the observed changes in liver deiodinase expression decrease local T<sub>3</sub> availability. Local T<sub>3</sub> availability may be determined both by serum thyroid hormone levels and by tissue deiodinase activities. The inhibition of hepatic D3 during sepsis might additionally affect local tissue T<sub>3</sub> concentrations (29), but it is unknown at present whether T<sub>3</sub>-regulated gene expression is affected in liver during sepsis.

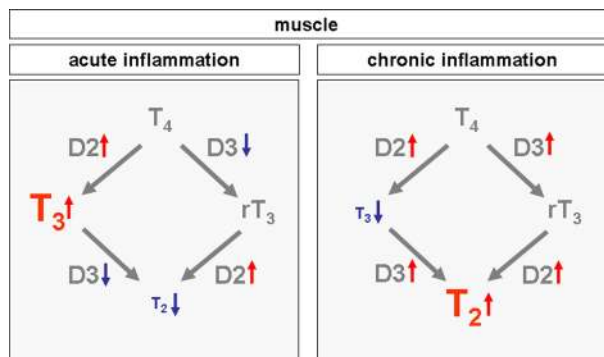
#### D. Muscle thyroid hormone metabolism during bacterial sepsis

In patients with septic shock, muscle thyroid hormone metabolism was compared with patients undergoing knee or hip replacement, representing acute surgical stress. Muscle D2 mRNA expression decreased during sepsis, whereas D2 activity was hardly detectable and did not differ from the control subjects undergoing surgery. By contrast, muscle D3 activity increased in septic patients compared with controls, possibly enhancing T<sub>3</sub> clearance in muscle. Furthermore, TR $\beta$ 1 mRNA as well as RXR $\gamma$  mRNA decreased during sepsis, whereas MCT8 expression did not change (103).

In animal models, muscle D2 expression decreased during *S. pneumoniae* infection, corresponding to the decrease reported in muscle tissue of septic patients (103). An effect of fasting in decreased D2 expression during sepsis cannot be excluded at this stage because D2 expression decreases by fasting and sepsis is associated with severely diminished food intake (106). During *S. pneumoniae* infection D3 mRNA expression was unchanged, whereas D3 activity tended to increase (132). These changes in skeletal muscles of septic mice were not related to serum T<sub>3</sub> levels and correlated negatively with muscle IL-1 $\beta$  mRNA expression. The signaling pathways of CREB, NF $\kappa$ B and ERK1/2 were not altered in muscles of septic mice compared with controls (132), implying involvement of noninflammatory mechanisms.

The combination of decreased D2 expression and unaltered or increased D3, observed in both human and experimental models for sepsis, may result in decreased T<sub>3</sub> bioavailability in muscle tissue. However, muscle UCP3 mRNA expression, which is a T<sub>3</sub> target gene, is unaltered in muscle tissue of septic patients compared with control patients (103). Thus, down-regulation of TR expression might be an adaptation to restore TR occupancy.

A shortage of T<sub>3</sub> may result in mitochondrial dysfunction, which is thought to play a role in the pathogenesis of sepsis (153, 154). Adding exogenous cytochrome c oxidase improves cardiomyocyte mitochondrial function in an animal model of sepsis (155). Furthermore, be-



**Fig. 4.** Schematic representation of the alterations in muscle deiodinase expression in acute inflammation (left, LPS administration) and chronic inflammation (right, turpentine-induced abscess) and the theoretical net result on local  $T_3$  and  $T_2$  concentrations. Note the increase in  $T_3$  in acute inflammation, and the increase in  $T_2$  in chronic inflammation.

cause thyroid hormone is an important regulator of muscle fiber type and contractility (101), changes in muscle deiodinase expression and, by inference, in local thyroid hormone availability might contribute to critical illness myopathy, which is frequently observed in ICU patients (156).

The complex discrepancies in muscle deiodinase expression between acute inflammation, chronic inflammation, and sepsis (Fig. 4) may relate to the severity of illness and support the concept that the net result of altered muscle deiodinase expression determines muscle function during specific stages of illness.

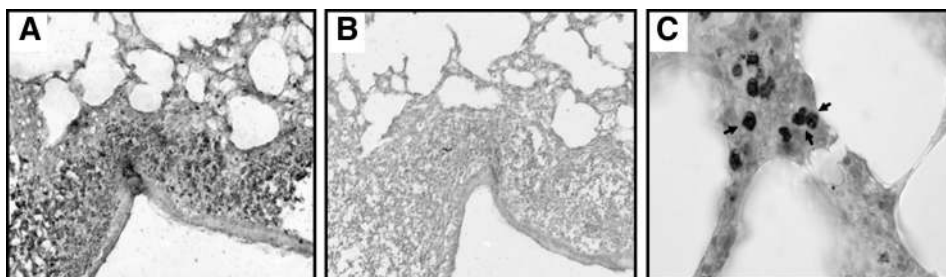
#### E. Thyroid hormone metabolism in adipose tissue during bacterial sepsis

Thyroid hormone metabolism has been studied in sc white adipose tissue of NTIS patients with septic shock within 5 d after admission to the ICU and compared with patients with acute surgical stress. White adipose tissue expresses MCT8,  $TR\beta_1$ ,  $TR\alpha_1$ ,  $RXR\alpha$ ,  $RXR\beta$ , and  $RXR\gamma$  and also D1, D2, and D3 (103). Sepsis appeared to be associated with moderately decreased MCT8,  $TR\beta_1$ ,  $TR\alpha_1$ , and  $RXR\gamma$  mRNA expression, whereas no differences were present in D1 and D2 mRNA expression. D1 and D3 activities were measured in adipose tissue samples

and showed no differences (103). Thus, septic shock seems to be associated with decreased  $T_3$  uptake and TR expression in adipose tissue. Surprisingly, UCP3, a  $T_3$  target gene, increased in adipose tissue during sepsis (103) suggesting increased  $T_3$  signaling, again showing the dissociation between  $T_3$  uptake and TR expression on one hand and  $T_3$  action on the other. Alternatively,  $T_3$  target gene expression might be affected by other genes. Because increased  $T_3$  signaling in adipose tissue during sepsis is opposite to decreased  $T_3$  signaling observed during nonlethal acute inflammation, the question remains whether the observed alterations are an active response to the septic state or rather represent a derangement of the tissue-specific response during inflammation, characteristic for sepsis.

#### F. Deiodinase expression in granulocytes

In addition to complex changes in deiodinase expression in muscle during illness, we recently reported strong D3 expression in infiltrating leukocytes during chemical and bacterial inflammation (94, 151). In this setting, D3 is highly expressed in polymorphonuclear leukocytes (PMN) infiltrating the abscess or the infected organ (Fig. 5). PMN are an important cell population involved in the innate immune response induced by bacterial pathogens because PMN are essential for adherence to and subsequent phagocytosis of bacteria. For this purpose, PMN contain bactericidal and tissue-toxic mechanisms such as the myeloperoxidase (MPO) system. During phagocytosis, invagination of the cell membrane occurs, which finally results in the incorporation of bacteria into the intracellular phagosome. Subsequently, an oxygen burst occurs together with a discharge of the content of cytoplasmic granules. MPO release into the phagosome then forms an effective antimicrobial system together with  $H_2O_2$  and iodide, chloride, bromide, or thiocyanate ions by oxidizing these ions (157). Oxidation of iodide results finally in the formation of hypoiodite, and the subsequent iodination of bacteria by hypoiodite promotes killing and degradation of the ingested bacteria. A clear relationship was observed between iodination of the



**Fig. 5.** An overview of a lung section showing *S. pneumoniae*-induced infiltrate after 48 h stained both with anti-D3 antibody (A) and preimmune serum (B). Note clearly stained D3-positive granulocytes in the infiltrated area that are shown in more detail in panel C (indicated by arrows). [Adapted from A. Boelen et al., Type 3 deiodinase is highly expressed in infiltrating neutrophilic granulocytes in response to acute bacterial infection. *Thyroid* 18:1095–1103, 2008 (151), with permission. © 2008, Mary Ann Liebert Inc.]

ingested bacteria and the bacterial killing capacity of PMN, suggesting that iodination is essential for their bacterial killing capacity (158).

Thyroid hormone metabolism in leukocytes was studied decades ago, when it appeared that leukocytes can take up and deiodinate  $T_4$ , thereby generating inorganic iodide and small quantities of  $T_3$  (159). Experimental induction of phagocytosis by leukocytes markedly enhanced  $T_4$  deiodination (160), and clinical observations of pulmonary uptake of  $^{131}\text{I}$ -labeled  $T_4$  during pneumonia (161) suggested binding of the isotope in the area of inflammation. If deiodination of thyroid hormone takes place at a higher rate in areas of bacterial infection, peripheral disposal of thyroid hormones should be accelerated. Indeed, an increased disposal rate of both  $T_4$  and  $T_3$  has been shown in patients with acute pulmonary bacterial infections (144, 162, 163).

We hypothesized that the induction of D3 in PMN during inflammation should not only inactivate  $T_4$  and  $T_3$  but also yield substantial amounts of iodide within the leukocyte. Therefore, D3 may contribute to the microbial killing capacity of the cell via MPO, finally resulting in the iodination of ingested pathogens. Our group then showed that D3KO mice have a much higher bacterial load in blood, lung, and spleen compared with WT upon infection with *S. pneumoniae* (Fig. 6) (33). Thus, the lack of D3 severely impaired the bacterial clearance capacity of the host, suggesting a completely novel and protective role for D3 in the defense against acute bacterial infection via iodine production (Fig. 7). D3-expressing granulocytes have subsequently been found in infected lungs, liver (151), and spinal cord (164), indicating that this phenomenon is highly characteristic of the innate immune response in general and is not limited to *S. pneumoniae*-induced sepsis.

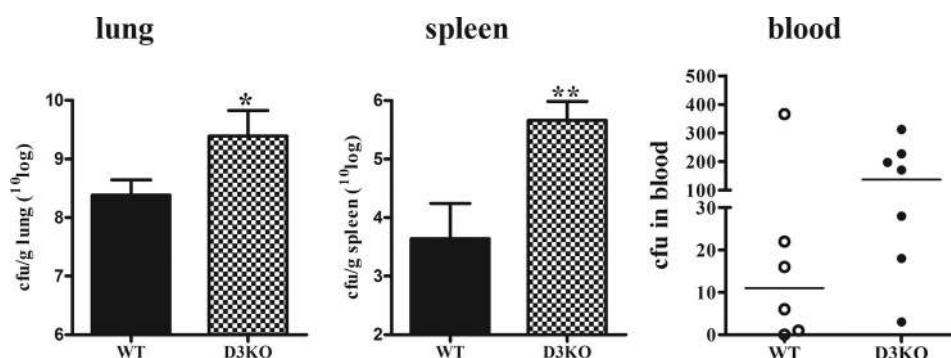
## V. Thyroid Hormone Metabolism in Metabolic Organs during Prolonged Critical Illness

Although during acute inflammation NTIS may represent an adaptive response, partly driven by the hypothalamus

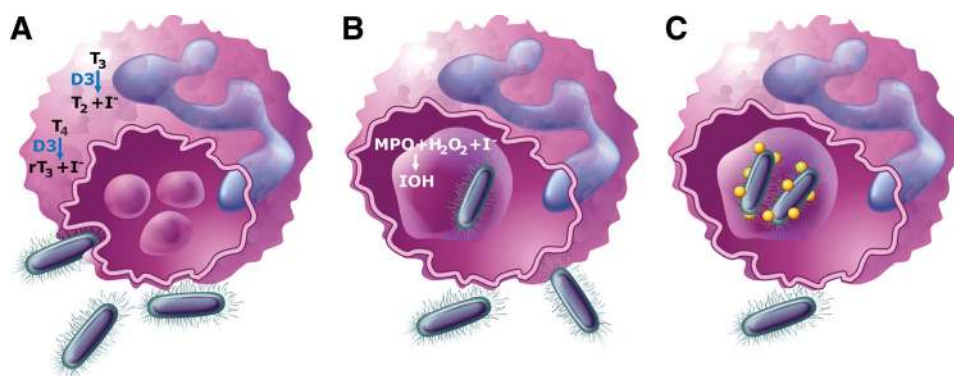
and aimed to support the immune response, NTIS might turn disadvantageous when critical illness enters a chronic phase necessitating prolonged mechanical ventilation, dialysis, and inotropic support. This perception of the neuroendocrine response to illness as a dynamic process with distinct features in the acute and chronic phase of critical illness was originally formulated by Van den Berghe (165). A relatively small number of studies have addressed changes in local thyroid hormone metabolism in patients with prolonged critical illness. It should be mentioned that some studies were based on samples obtained from critically ill patients shortly after death, introducing some degree of postmortem delay. To circumvent this bias and to be able to manipulate various confounding factors in a systematic way, a rabbit model of prolonged critical illness was developed in Leuven, displaying profound alterations in thyroid hormone metabolism while the serum thyroid hormone changes closely mimic those in critically ill ICU patients (166).

### A. Hepatic thyroid hormone metabolism during prolonged illness

Peeters *et al.* (167) studied thyroid hormone metabolism in deceased critically ill patients who had participated in a large, randomized, controlled study on intensive insulin treatment in the ICU setting. Liver biopsies were obtained within minutes after death. As expected in these patients, low serum  $T_3$ ,  $T_4$ , and TSH and increased serum  $rT_3$  levels were observed. In the liver biopsies, D1 activity was low and related to the cause of death: the lowest D1 activities were observed in patients who had died of cardiovascular collapse, and the highest D1 activities were observed in patients who had died of severe brain damage (167). Liver D1 activity was negatively correlated to serum  $T_4$  sulfate concentrations that were clearly increased during prolonged illness. This suggested that D1 plays a role in the illness-induced increase in serum  $T_4$  sulfate because liver sulfotransferase, which is an enzyme that catalyzes thyroid hormone sulfation, was unaltered (168). In con-



**Fig. 6.** Bacterial load in blood, lung, and spleen of WT mice (black bars) and D3KO mice (checkered bars) after *S. pneumoniae* infection. For lung and spleen, mean  $^{10}\log$  values of colony-forming units (cfu)/g  $\pm$  SEM are depicted. Differences between groups are indicated: \*,  $P = 0.05$ ; \*\*,  $P = 0.01$ . For blood, the number of cfu for each sample is depicted. [Adapted from A. Boelen *et al.*, Impaired bacterial clearance in type 3 deiodinase-deficient mice infected with *Streptococcus pneumoniae*. *Endocrinology* 150:1984–1990, 2009 (33), with permission. © 2009, The Endocrine Society.]



**Fig. 7.** Schematic overview of the proposed antibacterial action of type 3 deiodinase expressed in granulocytes. A, Deiodination of  $T_4$  and  $T_3$  at the cell surface and phagocytosis of bacteria. B, Formation of a phagosome by invagination of the cell membrane and generation of oxidized iodine compounds. C, Iodination of bacteria in the phagosome, followed by microbial killing.

trast to low liver D1, D3 was induced in liver of severely ill patients although no correlation was observed between activity levels and cause of death (167). Furthermore, liver D3 activity was positively correlated to serum  $rT_3$  concentrations and negatively to serum  $T_3$  levels (169).

Thyroid hormone transporter expression was also studied in these patients in comparison with liver biopsies obtained from living patients undergoing acute surgical stress. Liver MCT8 but not MCT10 mRNA expression increased during prolonged illness compared with acute surgical stress (96). Determination of  $TR\alpha$  and  $TR\beta$  mRNA expression in liver biopsies of patients with chronic liver disease before liver transplantation showed increased mRNA expression of both isoforms associated with low serum  $T_3$  and  $T_4$  levels (170). In postmortem liver biopsies from ICU patients, the  $TR\alpha_1/TR\alpha_2$  ratio increased with the severity of illness, whereas no relation was observed between liver D1 mRNA and the TR isoforms (171).

Because these studies are not suitable for dynamic observations, a rabbit model for prolonged critical illness was developed displaying severe, disease-induced, and feeding-resistant wasting; high mortality rate; limited spontaneous recovery; and a biphasic neuroendocrine response pattern. In addition, this model showed clinical, biochemical, and endocrine features resembling those observed in ICU patients (166, 172). Thyroid hormone levels in these rabbits change profoundly, with a persistent decrease in serum  $T_3$  and  $T_4$  within 3 d after injury without an increase in TSH (173). Liver D1 activity is clearly decreased in these rabbits and positively correlated with serum  $T_3$  as well as with  $T_3/rT_3$  ratio. Liver D3 activity tends to be up-regulated and to correlate negatively with serum  $T_3$  and the  $T_3/rT_3$  ratio (174). Liver MCT8 mRNA increases. The administration of exogenous thyroid hormones reduces MCT8 expression combined with an increase in serum thyroid hormone levels, suggesting

thyroid hormone-dependent regulation of MCT8 during illness (96).

In summary, prolonged illness results in profound alterations in liver thyroid hormone metabolism compared with acute and chronic inflammation. Thyroid hormone transport into the liver increases, whereas liver D1 decreases and liver D3 increases, the latter change perhaps resulting from tissue hypoxia (167). This might be mediated via the transcription factor hypoxia inducible factor (HIF)-1 $\alpha$  because an *in vitro* study showed that HIF-1 $\alpha$  interacts with the *DIO3* promoter, increasing D3 expression upon HIF-1 $\alpha$  activation. Using a rat model of cardiac failure, the simultaneous induction of HIF-1 $\alpha$  and D3 in the same myocardial area has been observed (175). This mechanism may be relevant for the ICU setting because tissue ischemia is frequently observed in critically ill patients.

Arem *et al.* (51) analyzed liver samples of NTIS patients and healthy controls and reported lower liver  $T_3$  and  $T_4$  concentrations in NTIS, indicating that the liver might be deficient in thyroid hormones during prolonged critical illness. This observation has been confirmed in an animal model because liver  $T_3$  levels were decreased in critically ill rabbits (176). Not much is known about  $T_3$ -regulated gene expression in liver during prolonged illness, but considering the fact that liver  $T_3$  levels are low,  $T_3$  target gene expression is probably reduced.

## B. Muscle thyroid hormone metabolism during prolonged illness

Prolonged critically ill patients often develop muscle dysfunction that is probably due to decreased mitochondrial activity and content (177). Muscle dysfunction is also associated with lower concentrations of energy-rich substrates and higher lactate content (153). Because thyroid hormones are involved in mitochondrial function and biogenesis, muscle thyroid hormone metabolism has been

studied during prolonged illness in sample biopsies of deceased critically ill patients. Muscle biopsies were taken within minutes after death and showed significant D3 activity levels, whereas D2 activity was undetectable. Muscle D3 activity was related to serum  $rT_3$  concentrations (169), and the serum  $rT_3/T_4$  ratio was higher in patients treated with inotropes (167). Mebis *et al.* (108) did report increased D2 activity in muscle tissue of prolonged critically ill patients compared with patients with acute surgical stress and to healthy volunteers. The samples were taken from the same cohort as reported by Peeters *et al.* (167), but the techniques and conditions were different (108). In contrast, muscle D2 and D3 activity were both undetectable in tissue of critically ill rabbits (174).

Muscle MCT8 mRNA expression increases significantly during prolonged illness compared with acute surgical stress, whereas MCT10 mRNA expression does not change. MCT8 expression correlates negatively to serum  $T_3$  and  $T_4$  concentrations (96). Muscle MCT8 and MCT10 mRNA expression was also studied in an animal model of prolonged illness, and the findings markedly differ from the human samples because MCT8 mRNA expression did not change, whereas MCT10 markedly increased. Treatment with thyroid hormones restored the illness-induced increase in MCT10 expression, suggesting that thyroid hormone transporter status is regulated by thyroid hormone levels during prolonged illness (96).

Of interest is whether high D2, high D3, and increased thyroid hormone transport in muscle of patients with prolonged critical illness results in substantially altered tissue thyroid hormone concentrations. Only one study has reported muscle  $T_3$  concentrations in postmortem tissue of patients with prolonged illness showing marked interindividual variation ranging from very low to extremely high values (51). No information is available at present about  $T_3$ -regulated gene expression in muscle tissue during prolonged illness. However, because muscle  $T_3$  content is supposed to be a determinant of muscle function, which may be severely impaired during prolonged illness,  $T_3$  concentrations may be assumed to be low. However, future studies will be needed to confirm or refute this assumption.

## VI. Endocrine Interventions in NTIS: Areas of Uncertainty

### A. Endocrine interventions

Because administration of thyroid hormone in various clinical settings associated with NTIS did not improve clinical outcome or organ function (178), whereas at the same time carrying the risk of overdosing, an interesting novel approach in light of the hypothalamic alterations in TRH expression is administration of TRH in combination with other hypothalamic peptides. Van den Berghe *et al.*

(179) showed that infusion of TRH alone or in combination with GH secretagogues to critically ill patients augmented the pulsatile TSH release that was associated with an increase in serum  $T_4$  and  $T_3$  levels. Prolonged administration of TRH and GHRP-2 to critically ill patients also showed that protein degradation was reduced in association with restoration of plasma thyroid hormone levels and reactivation of the blunted TSH secretion (15). Likewise, in the animal model of protracted critical illness, the administration of TRH in combination with GHRP-2 restored the illness-induced down-regulation of TSH shortly after administration of the hypothalamic peptides (166). Interestingly, infusion of TRH alone or in combination with GHRP-2 also restored the decrease in liver D1 decrease in relation to alterations in serum  $T_3$  and  $T_4$  (174). In mice, the administration of TRH during LPS-induced acute illness attenuated the decrease in serum  $T_4$  (65), confirming that stimulation of the pituitary during NTIS may partly restore the illness-induced decrease of serum thyroid hormone levels. These studies provide convincing evidence that the central part of the HPT axis is important in the pathogenesis of NTIS, while representing a possible treatment target. Treatment of NTIS with thyroid hormones in patients with prolonged illness has not been the focus of many recent clinical trials. Pingitore *et al.* (180) reported the effects of  $T_3$  infusion in NTIS patients with ischemic or nonischemic dilated cardiomyopathy and showed that short-term  $T_3$  therapy improved the neuroendocrine profile as well as ventricular performance. A recent systemic review evaluated the effects and risks of postoperative  $T_3$  treatment, analyzing 14 randomized clinical trials involving  $T_3$  administration in variable doses and durations, predominantly during cardiac surgery. Short duration postoperative  $T_3$  treatment appeared to increase cardiac index. However, mortality was not affected (181). The effects of two-dose regimens of thyroid hormone treatment have also been studied in prolonged critically ill rabbits. The low dose or substitution dose did not affect the illness-induced serum  $T_3$  decrease, whereas serum  $T_4$  was even further decreased upon treatment. Only a supraphysiological dose of thyroid hormone, aimed to target serum thyroid hormone levels within the range obtained earlier by TRH infusion, increased serum  $T_3$  and  $T_4$  levels and liver D1 activity within the normal range (176), which implicates enhanced degradation and/or excretion of thyroid hormone during prolonged illness. An additional study by the same authors showed that circulating  $T_3$  concentrations are causally related to liver D1 activity during prolonged illness, indicating that the illness-induced decrease of liver D1 is secondary to low serum thyroid hormone levels (182) in this particular model. Thyroid hormone treatment also restored illness-induced alterations



in liver MCT8 and muscle MCT10 mRNA expression. It is, however, unknown at present whether the various treatment strategies are beneficial in terms of clinical outcome (96).

### B. Areas of uncertainty

In summary, illness results in profound alterations in local thyroid hormone metabolism that are only partly related to changes in serum concentrations. Generally, acute illness is associated with reduced  $T_3$  action in liver and adipose tissue and increased  $T_3$  action in skeletal muscle. In contrast, chronic inflammation appears to be associated with unchanged  $T_3$  action in liver, whereas both D2 and D3 increase in skeletal muscle theoretically leading to increased  $T_2$  concentrations. Sepsis, a whole-body inflammatory state caused by bacterial infection and characterized by uncontrolled activation of the pro- and antiinflammatory responses, is associated with decreased muscle D2 activity and increased  $T_3$  action in adipose tissue. These differential changes are schematically presented in Fig. 8.

Experimental animal models are frequently used to study illness-induced alterations at the tissue level, and these studies have generally revealed similar changes in tissue thyroid hormone metabolism between humans and rodents. However, the serum characteristics of NTIS are not identical between humans and rodents (Table 1). This is a complicating factor in attempts to extrapolate data obtained in experimental animal studies to the human situation. In addition, the role of reduced food intake as a result of illness affects thyroid hormone metabolism. Therefore, the use of pair-fed controls is essential for an unbiased interpretation of animal experiments on NTIS. Furthermore, posttranscriptional regulation of deiodinase expression has been reported, resulting in a potentially uncertain relationship between deiodinase mRNA expression and activity. However, in most studies on NTIS, the

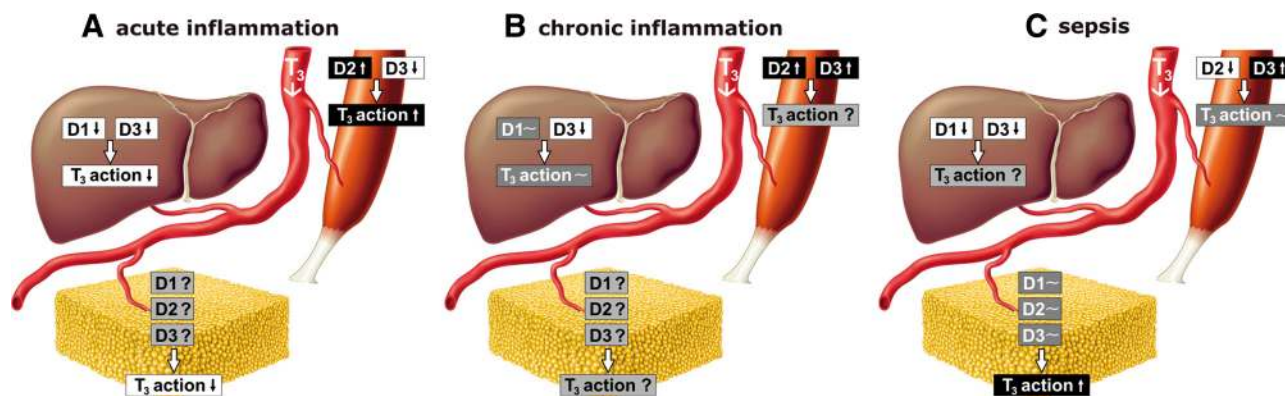
illness-induced alterations in deiodinase mRNA expression and enzyme activity are strictly correlated.

A number of key issues remain unsolved at present:

- Thyroid hormone availability should be assessed at the tissue level to establish the net result of the observed alterations in local deiodinase activities. The further development of liquid chromatography-mass spectrometry as a tool to assess various molecules in one tissue sample will probably facilitate these studies both in human and rodent tissue samples.
- The functional consequences of altered thyroid hormone metabolism in terms of organ or cellular function are largely unknown at present. Important and clinically relevant areas of NTIS research during sepsis and prolonged illness include mitochondrial function in skeletal muscle, bacterial killing capacity of infiltrating granulocytes during the innate immune response, and lipid metabolism in adipose tissue. We can only speculate at this stage that these future studies bear clinical relevance.
- Small but promising endocrine intervention studies in patients with protracted critical illness, involving treatment with neuropeptides and resulting in largely restored plasma thyroid hormone levels as well as improved metabolic parameters, should be expanded to larger studies involving more patients and longer treatment periods. These studies will help to establish whether or not such interventions are meaningful in terms of clinical outcome.

### VII. Conclusion

Current knowledge has completely altered the concept of NTIS. In the classic view, NTIS is a syndrome with lower plasma thyroid hormone concentration of unknown sig-



**Fig. 8.** Schematic representation of the differential expression of deiodinating enzymes and the resulting change in thyroid action in  $T_3$  target tissues during acute inflammation (A), chronic inflammation (B), and sepsis (C). The scheme summarizes experimental and human studies. *Black boxes* represent an increase, *white boxes* a decrease, and *gray boxes/white font* no change. *Gray boxes/black font* symbolize a lack of data.

nificance as its key phenomenon. Recent studies, however, have clearly shown that NTIS represents a profound and differential change in thyroid hormone physiology, both at the level of the HPT axis in terms of setpoint regulation and at the organ level in terms of local thyroid hormone metabolism. It should be interpreted in the context of type of illness and of the organ/tissue studied. Organ- and timing-specific changes in D1–D3 highlight deiodinases as proactive players in the response to illness. Furthermore, the granulocyte is proposed as a novel and important cell type involved in NTIS during bacterial infection. Finally, changes in thyroid hormone metabolism in muscle during critical illness may be relevant for the pathogenesis of respiratory failure.

## Acknowledgments

Address all correspondence and requests for reprints to: Anita Boelen, Ph.D., Department of Endocrinology and Metabolism, F5-165, Academic Medical Center, University of Amsterdam, Meibergdreef 9, 1105AZ Amsterdam, The Netherlands. E-mail: a.boelen@amc.uva.nl.

Disclosure Summary: The authors have nothing to disclose.

## References

1. Docter R, Krenning EP, de Jong M, Hennemann G 1993 The sick euthyroid syndrome: changes in thyroid hormone serum parameters and hormone metabolism. *Clin Endocrinol (Oxf)* 39:499–518
2. Kaptein EM, MacIntyre SS, Weiner JM, Spencer CA, Nicoloff JT 1981 Free thyroxine estimates in nonthyroidal illness: comparison of eight methods. *J Clin Endocrinol Metab* 52:1073–1077
3. Bello G, Pennisi MA, Montini L, Silva S, Maviglia R, Cavallaro F, Bianchi A, De Marinis L, Antonelli M 2009 Nonthyroidal illness syndrome and prolonged mechanical ventilation in patients admitted to the ICU. *Chest* 135:1448–1454
4. Gangemi EN, Garino F, Berchiolla P, Martinese M, Arecco F, Orlandi F, Stella M 2008 Low triiodothyronine serum levels as a predictor of poor prognosis in burn patients. *Burns* 34:817–824
5. Boelen A, Wiersinga WM, Kohrle J 2006 Contributions of cytokines to nonthyroidal illness. *Curr Opin Endocrinol Diabetes* 13:444–450
6. Sato K, Satoh T, Shizume K, Ozawa M, Han DC, Imamura H, Tsushima T, Demura H, Kanaji Y, Ito Y 1990 Inhibition of 125I organification and thyroid hormone release by interleukin-1, tumor necrosis factor- $\alpha$ , and interferon- $\gamma$  in human thyrocytes in suspension culture. *J Clin Endocrinol Metab* 70:1735–1743
7. Tominaga T, Yamashita S, Nagayama Y, Morita S, Yokoyama N, Izumi M, Nagataki S 1991 Interleukin 6 inhibits human thyroid peroxidase gene expression. *Acta Endocrinol (Copenh)* 124:290–294
8. Tang KT, Braverman LE, DeVito WJ 1995 Tumor necrosis factor- $\alpha$  and interferon- $\gamma$  modulate gene expression of type I 5'-deiodinase, thyroid peroxidase, and thyroglobulin in FRTL-5 rat thyroid cells. *Endocrinology* 136:881–888
9. van der Poll T, Romijn JA, Wiersinga WM, Sauerwein HP 1990 Tumor necrosis factor: a putative mediator of the sick euthyroid syndrome in man. *J Clin Endocrinol Metab* 71:1567–1572
10. Stouthard JM, van der Poll T, Endert E, Bakker PJ, Veenhof CH, Sauerwein HP, Romijn JA 1994 Effects of acute and chronic interleukin-6 administration on thyroid hormone metabolism in humans. *J Clin Endocrinol Metab* 79:1342–1346
11. Boelen A, Platvoet-ter Schiphorst MC, Bakker O, Wiersinga WM 1995 The role of cytokines in the lipopolysaccharide-induced sick euthyroid syndrome in mice. *J Endocrinol* 146:475–483
12. Hermus RM, Sweep CG, van der Meer MJ, Ross HA, Smals AG, Benraad TJ, Kloppenborg PW 1992 Continuous infusion of interleukin-1 $\beta$  induces a nonthyroidal illness syndrome in the rat. *Endocrinology* 131:2139–2146
13. Wiersinga WM 2005 Nonthyroidal illness. In: Braverman LE, Utiger RD, eds. *The thyroid*. 8th ed. Philadelphia: Lippincott; 246–263
14. Fliers E, Guldenaar SE, Wiersinga WM, Swaab DF 1997 Decreased hypothalamic thyrotropin-releasing hormone gene expression in patients with nonthyroidal illness. *J Clin Endocrinol Metab* 82:4032–4036
15. Van den Berghe G, Wouters P, Weekers F, Mohan S, Baxter RC, Veldhuis JD, Bowers CY, Bouillon R 1999 Reactivation of pituitary hormone release and metabolic improvement by infusion of growth hormone-releasing peptide and thyrotropin-releasing hormone in patients with protracted critical illness. *J Clin Endocrinol Metab* 84:1311–1323
16. Boelen A, Kwakkel J, Thijssen-Timmer DC, Alkemade A, Fliers E, Wiersinga WM 2004 Simultaneous changes in central and peripheral components of the hypothalamus-pituitary-thyroid axis in lipopolysaccharide-induced acute illness in mice. *J Endocrinol* 182:315–323
17. Sugiyama D, Kusahara H, Taniguchi H, Ishikawa S, Nozaki Y, Aburatani H, Sugiyama Y 2003 Functional characterization of rat brain-specific organic anion transporter (Oatp14) at the blood-brain barrier: high affinity transporter for thyroxine. *J Biol Chem* 278:43489–43495
18. Heuer H, Maier MK, Iden S, Mittag J, Friesema EC, Visser TJ, Bauer K 2005 The monocarboxylate transporter 8 linked to human psychomotor retardation is highly expressed in thyroid hormone-sensitive neuron populations. *Endocrinology* 146:1701–1706
19. Alkemade A, Friesema EC, Unmehopa UA, Fabriek BO, Kuiper GG, Leonard JL, Wiersinga WM, Swaab DF, Visser TJ, Fliers E 2005 Neuroanatomical pathways for thyroid hormone feedback in the human hypothalamus. *J Clin Endocrinol Metab* 90:4322–4334
20. Visser WE, Friesema EC, Visser TJ 2011 Minireview: thyroid hormone transporters: the knowns and the unknowns. *Mol Endocrinol* 25:1–14
21. Braun D, Kinne A, Bräuer AU, Sapin R, Klein MO, Köhrle J, Wirth EK, Schweizer U 2011 Developmental and cell type-specific expression of thyroid hormone transporters in the mouse brain and in primary brain cells. *Glia* 59:463–471
22. Alkemade A, Friesema EC, Kalsbeek A, Swaab DF, Visser

- TJ, Fliers E 2011 Expression of thyroid hormone transporters in the human hypothalamus. *J Clin Endocrinol Metab* 96:E967–E971
23. Mebis L, Debaveye Y, Ellger B, Derde S, Ververs EJ, Langouche L, Darras VM, Fliers E, Visser TJ, Van den Berghe G 2009 Changes in the central component of the hypothalamus-pituitary-thyroid axis in a rabbit model of prolonged critical illness. *Crit Care* 13:R147
  24. Köhrle J 2000 The selenoenzyme family of deiodinase isozymes controls local thyroid hormone availability. *Rev Endocr Metab Disord* 1:49–58
  25. Jakobs TC, Schmutzler C, Meissner J, Köhrle J 1997 The promoter of the human type I 5'-deiodinase gene—mapping of the transcription start site and identification of a DR+4 thyroid-hormone-responsive element. *Eur J Biochem* 247:288–297
  26. Toyoda N, Zavacki AM, Maia AL, Harney JW, Larsen PR 1995 A novel retinoid X receptor-independent thyroid hormone response element is present in the human type 1 deiodinase gene. *Mol Cell Biol* 15:5100–5112
  27. Burmeister LA, Pachucki J, St Germain DL 1997 Thyroid hormones inhibit type 2 iodothyronine deiodinase in the rat cerebral cortex by both pre- and posttranslational mechanisms. *Endocrinology* 138:5231–5237
  28. Sagar GD, Gereben B, Callebaut I, Mornon JP, Zeöld A, da Silva WS, Luongo C, Dentice M, Tente SM, Freitas BC, Harney JW, Zavacki AM, Bianco AC 2007 Ubiquitination-induced conformational change within the deiodinase dimer is a switch regulating enzyme activity. *Mol Cell Biol* 27:4774–4783
  29. Gereben B, Zeöld A, Dentice M, Salvatore D, Bianco AC 2008 Activation and inactivation of thyroid hormone by deiodinases: local action with general consequences. *Cell Mol Life Sci* 65:570–590
  30. Coppola A, Liu ZW, Andrews ZB, Paradis E, Roy MC, Friedman JM, Ricquier D, Richard D, Horvath TL, Gao XB, Diano S 2007 A central thermogenic-like mechanism in feeding regulation: an interplay between arcuate nucleus T3 and UCP2. *Cell Metab* 5:21–33
  31. Fekete C, Gereben B, Doleschall M, Harney JW, Dora JM, Bianco AC, Sarkar S, Liposits Z, Rand W, Emerson C, Kacs Kovics I, Larsen PR, Lechan RM 2004 Lipopolysaccharide induces type 2 iodothyronine deiodinase in the mediobasal hypothalamus: implications for the nonthyroidal illness syndrome. *Endocrinology* 145:1649–1655
  32. Boelen A, Kwakkel J, Wiersinga WM, Fliers E 2006 Chronic local inflammation in mice results in decreased TRH and type 3 deiodinase mRNA expression in the hypothalamic paraventricular nucleus independently of diminished food intake. *J Endocrinol* 191:707–714
  33. Boelen A, Kwakkel J, Wieland CW, St Germain DL, Fliers E, Hernandez A 2009 Impaired bacterial clearance in type 3 deiodinase-deficient mice infected with *Streptococcus pneumoniae*. *Endocrinology* 150:1984–1990
  34. Coppola A, Meli R, Diano S 2005 Inverse shift in circulating corticosterone and leptin levels elevates hypothalamic deiodinase type 2 in fasted rats. *Endocrinology* 146:2827–2833
  35. Fekete C, Sarkar S, Christoffolete MA, Emerson CH, Bianco AC, Lechan RM 2005 Bacterial lipopolysaccharide (LPS)-induced type 2 iodothyronine deiodinase (D2) activation in the mediobasal hypothalamus (MBH) is independent of the LPS-induced fall in serum thyroid hormone levels. *Brain Res* 1056:97–99
  36. Sánchez E, Singru PS, Fekete C, Lechan RM 2008 Induction of type 2 iodothyronine deiodinase in the mediobasal hypothalamus by bacterial lipopolysaccharide: role of corticosterone. *Endocrinology* 149:2484–2493
  37. Zeöld A, Doleschall M, Haffner MC, Capelo LP, Menyórt J, Liposits Z, da Silva WS, Bianco AC, Kacs Kovics I, Fekete C, Gereben B 2006 Characterization of the nuclear factor- $\kappa$ B responsiveness of the human dio2 gene. *Endocrinology* 147:4419–4429
  38. Sánchez E, Singru PS, Wittmann G, Nouriel SS, Barrett P, Fekete C, Lechan RM 2010 Contribution of TNF- $\alpha$  and nuclear factor- $\kappa$ B signaling to type 2 iodothyronine deiodinase activation in the mediobasal hypothalamus after lipopolysaccharide administration. *Endocrinology* 151:3827–3835
  39. Boelen A, Kwakkel J, Chassande O, Fliers E 2009 Thyroid hormone receptor  $\beta$  mediates acute illness-induced alterations in central thyroid hormone metabolism. *J Neuroendocrinol* 21:465–472
  40. Bassett JH, Harvey CB, Williams GR 2003 Mechanisms of thyroid hormone receptor-specific nuclear and extranuclear actions. *Mol Cell Endocrinol* 213:1–11
  41. Liu RT, Suzuki S, Miyamoto T, Takeda T, Ozata M, DeGroot LJ 1995 The dominant negative effect of thyroid hormone receptor splicing variant  $\alpha$  2 does not require binding to a thyroid response element. *Mol Endocrinol* 9:86–95
  42. Chassande O, Fraichard A, Gauthier K, Flamant F, Legrand C, Savatier P, Laudet V, Samarut J 1997 Identification of transcripts initiated from an internal promoter in the c-erbA  $\alpha$  locus that encode inhibitors of retinoic acid receptor- $\alpha$  and triiodothyronine receptor activities. *Mol Endocrinol* 11:1278–1290
  43. Yen PM 2001 Physiological and molecular basis of thyroid hormone action. *Physiol Rev* 81:1097–1142
  44. Wood WM, Dowding JM, Haugen BR, Bright TM, Gordon DF, Ridgway EC 1994 Structural and functional characterization of the genomic locus encoding the murine  $\beta$  2 thyroid hormone receptor. *Mol Endocrinol* 8:1605–1617
  45. Wallis K, Dudazy S, van Hogerlinden M, Nordström K, Mittag J, Vennström B 2010 The thyroid hormone receptor  $\alpha$ 1 protein is expressed in embryonic postmitotic neurons and persists in most adult neurons. *Mol Endocrinol* 24:1904–1916
  46. Fliers E, Alkemade A, Wiersinga WM, Swaab DF 2006 Hypothalamic thyroid hormone feedback in health and disease. *Prog Brain Res* 153:189–207
  47. Abel ED, Ahima RS, Boers ME, Elmquist JK, Wondisford FE 2001 Critical role for thyroid hormone receptor  $\beta$ 2 in the regulation of paraventricular thyrotropin-releasing hormone neurons. *J Clin Invest* 107:1017–1023
  48. Boelen A, Kwakkel GJ, Chassande O, Fliers E 2009 A possible role for the thyroid hormone receptor  $\alpha$  in inflammation-induced changes in pituitary thyroid hormone metabolism. *Thyroid* 19(s1):s68 (abstract ATA)
  49. Lechan RM, Fekete C 2004 Feedback regulation of thyrotropin-releasing hormone (TRH): mechanisms for the nonthyroidal illness syndrome. *J Endocrinol Invest* 27:105–119
  50. Freitas BC, Gereben B, Castillo M, Kalló I, Zeöld A, Egri

- P, Liposits Z, Zavacki AM, Maciel RM, Jo S, Singru P, Sanchez E, Lechan RM, Bianco AC 2010 Paracrine signaling by glial cell-derived triiodothyronine activates neuronal gene expression in the rodent brain and human cells. *J Clin Invest* 120:2206–2217
51. Arem R, Wiener GJ, Kaplan SG, Kim HS, Reichlin S, Kaplan MM 1993 Reduced tissue thyroid hormone levels in fatal illness. *Metabolism* 42:1102–1108
  52. Sagar SM, Price KJ, Kasting NW, Sharp FR 1995 Anatomic patterns of Fos immunostaining in rat brain following systemic endotoxin administration. *Brain Res Bull* 36:381–392
  53. Marvel FA, Chen CC, Badr N, Gaykema RP, Goehler LE 2004 Reversible inactivation of the dorsal vagal complex blocks lipopolysaccharide-induced social withdrawal and c-Fos expression in central autonomic nuclei. *Brain Behav Immun* 18:123–134
  54. Fekete C, Singru PS, Sarkar S, Rand WM, Lechan RM 2005 Ascending brainstem pathways are not involved in lipopolysaccharide-induced suppression of thyrotropin-releasing hormone gene expression in the hypothalamic paraventricular nucleus. *Endocrinology* 146:1357–1363
  55. Alkemade A, Friesema EC, Kuiper GG, Wiersinga WM, Swaab DF, Visser TJ, Fliers E 2006 Novel neuroanatomical pathways for thyroid hormone action in the human anterior pituitary. *Eur J Endocrinol* 154:491–500
  56. Trajkovic M, Visser TJ, Mittag J, Horn S, Lukas J, Darras VM, Raivich G, Bauer K, Heuer H 2007 Abnormal thyroid hormone metabolism in mice lacking the monocarboxylate transporter 8. *J Clin Invest* 117:627–635
  57. Schneider MJ, Fiering SN, Pallud SE, Parlow AF, St Germain DL, Galton VA 2001 Targeted disruption of the type 2 selenodeiodinase gene (DIO2) results in a phenotype of pituitary resistance to T4. *Mol Endocrinol* 15:2137–2148
  58. Baur A, Bauer K, Jarry H, Köhrle J 2000 Effects of proinflammatory cytokines on anterior pituitary 5'-deiodinase type I and type II. *J Endocrinol* 167:505–515
  59. Boelen A, Kwakkel J, Platvoet-ter Schiphorst M, Baur A, Köhrle J, Wiersinga WM 2004 Contribution of interleukin-12 to the pathogenesis of non-thyroidal illness. *Horm Metab Res* 36:101–106
  60. Boelen A, Kwakkel J, Platvoet-ter Schiphorst M, Mentrup B, Baur A, Köhrle J, Wiersinga WM 2004 Interleukin-18, a proinflammatory cytokine, contributes to the pathogenesis of non-thyroidal illness mainly via the central part of the hypothalamus-pituitary-thyroid axis. *Eur J Endocrinol* 151:497–502
  61. Li M, Boyages SC 1997 Expression of  $\beta 2$ -thyroid hormone receptor in euthyroid and hypothyroid rat pituitary gland: an in situ hybridization and immunocytochemical study. *Brain Res* 773:125–131
  62. Wood WM, Ocran KW, Gordon DF, Ridgway EC 1991 Isolation and characterization of mouse complementary DNAs encoding  $\alpha$  and  $\beta$  thyroid hormone receptors from thyrotrope cells: the mouse pituitary-specific  $\beta 2$  isoform differs at the amino terminus from the corresponding species from rat pituitary tumor cells. *Mol Endocrinol* 5:1049–1061
  63. Forrest D, Reh TA, Rüschi A 2002 Neurodevelopmental control by thyroid hormone receptors. *Curr Opin Neurobiol* 12:49–56
  64. Fekete C, Lechan RM 2007 Negative feedback regulation of hypophysiotropic thyrotropin-releasing hormone (TRH) synthesizing neurons: role of neuronal afferents and type 2 deiodinase. *Front Neuroendocrinol* 28:97–114
  65. Rocchi R, Kimura H, Tzou SC, Suzuki K, Rose NR, Pinchera A, Ladenson PW, Caturegli P 2007 Toll-like receptor-MyD88 and Fc receptor pathways of mast cells mediate the thyroid dysfunctions observed during nonthyroidal illness. *Proc Natl Acad Sci USA* 104:6019–6024
  66. Nakabayashi K, Matsumi H, Bhalla A, Bae J, Mosselman S, Hsu SY, Hsueh AJ 2002 Thyrostimulin, a heterodimer of two new human glycoprotein hormone subunits, activates the thyroid-stimulating hormone receptor. *J Clin Invest* 109:1445–1452
  67. Okada SL, Ellsworth JL, Durnam DM, Haugen HS, Hollo way JL, Kelley ML, Lewis KE, Ren H, Sheppard PO, Storey HM, Waggie KS, Wolf AC, Yao LY, Webster PJ 2006 A glycoprotein hormone expressed in corticotrophs exhibits unique binding properties on thyroid-stimulating hormone receptor. *Mol Endocrinol* 20:414–425
  68. Prummel MF, Brokken LJ, Meduri G, Misrahi M, Bakker O, Wiersinga WM 2000 Expression of the thyroid-stimulating hormone receptor in the folliculo-stellate cells of the human anterior pituitary. *J Clin Endocrinol Metab* 85:4347–4353
  69. Sudo S, Kuwabara Y, Park JI, Hsu SY, Hsueh AJ 2005 Heterodimeric fly glycoprotein hormone- $\alpha 2$  (GPA2) and glycoprotein hormone- $\beta 5$  (GPB5) activate fly leucine-rich repeat-containing G protein-coupled receptor-1 (DLGR1) and stimulation of human thyrotropin receptors by chimeric fly GPA2 and human GPB5. *Endocrinology* 146:3596–3604
  70. Okajima Y, Nagasaki H, Suzuki C, Suga H, Ozaki N, Arima H, Hamada Y, Civelli O, Oiso Y 2008 Biochemical roles of the oligosaccharide chains in thyrostimulin, a heterodimeric hormone of glycoprotein hormone subunits  $\alpha 2$  (GPA2) and  $\beta 5$  (GPB5). *Regul Pept* 148:62–67
  71. Dos Santos S, Bardet C, Bertrand S, Escriva H, Habert D, Querat B 2009 Distinct expression patterns of glycoprotein hormone- $\alpha 2$  and - $\beta 5$  in a basal chordate suggest independent developmental functions. *Endocrinology* 150:3815–3822
  72. Suzuki C, Nagasaki H, Okajima Y, Suga H, Ozaki N, Arima H, Iwasaki Y, Oiso Y 2009 Inflammatory cytokines regulate glycoprotein subunit  $\beta 5$  of thyrostimulin through nuclear factor- $\kappa B$ . *Endocrinology* 150:2237–2243
  73. van Zeijl CJ, Surovtseva OV, Wiersinga WM, Fliers E, Boelen A 2011 Acute inflammation increases pituitary and hypothalamic glycoprotein hormone subunit B5 (GPB5) mRNA expression in association with decreased TSH receptor mRNA expression in mice. *J Neuroendocrinol* 23:310–319
  74. Boelen A, Platvoet-ter Schiphorst MC, Wiersinga WM 1993 Association between serum interleukin-6 and serum 3,5,3'-triiodothyronine in nonthyroidal illness. *J Clin Endocrinol Metab* 77:1695–1699
  75. Hashimoto H, Igarashi N, Yachie A, Miyawaki T, Sato T 1994 The relationship between serum levels of interleukin-6 and thyroid hormone in children with acute respiratory infection. *J Clin Endocrinol Metab* 78:288–291
  76. Raja SG, Berg GA 2007 Outcomes of off-pump coronary artery bypass surgery: current best available evidence. *Indian Heart J* 59:15–27

77. Sabatino L, Cerillo AG, Ripoli A, Pilo A, Glauber M, Iervasi G 2002 Is the low tri-iodothyronine state a crucial factor in determining the outcome of coronary artery bypass patients? Evidence from a clinical pilot study. *J Endocrinol* 175:577–586
78. Pålsson-McDermott EM, O'Neill LA 2004 Signal transduction by the lipopolysaccharide receptor, Toll-like receptor-4. *Immunology* 113:153–162
79. Amma LL, Campos-Barros A, Wang Z, Vennström B, Forrest D 2001 Distinct tissue-specific roles for thyroid hormone receptors  $\beta$  and  $\alpha 1$  in regulation of type 1 deiodinase expression. *Mol Endocrinol* 15:467–475
80. Macchia PE, Takeuchi Y, Kawai T, Cua K, Gauthier K, Chassande O, Seo H, Hayashi Y, Samarut J, Murata Y, Weiss RE, Refetoff S 2001 Increased sensitivity to thyroid hormone in mice with complete deficiency of thyroid hormone receptor  $\alpha$ . *Proc Natl Acad Sci USA* 98:349–354
81. Wolf M, Hansen N, Greten H 1994 Interleukin  $1\beta$ , tumor necrosis factor- $\alpha$  and interleukin 6 decrease nuclear thyroid hormone receptor capacity in a liver cell line. *Eur J Endocrinol* 131:307–312
82. Kwakkel J, Wiersinga WM, Boelen A 2006 Differential involvement of nuclear factor- $\kappa B$  and activator protein-1 pathways in the interleukin- $1\beta$ -mediated decrease of deiodinase type 1 and thyroid hormone receptor  $\beta 1$  mRNA. *J Endocrinol* 189:37–44
83. Kwakkel J, Wiersinga WM, Boelen A 2007 Interleukin- $1\beta$  modulates endogenous thyroid hormone receptor  $\alpha$  gene transcription in liver cells. *J Endocrinol* 194:257–265
84. Schug J, Overton GC 1998 TESS: Transcription element search software on the WWW. Technical Report CBIL-TR-1997-1001-v0.0. 8th ed. Philadelphia: Computational Biology and Informatics Laboratory, School of Medicine University of Pennsylvania; 1–10
85. Beigneux AP, Moser AH, Shigenaga JK, Grunfeld C, Feingold KR 2003 Sick euthyroid syndrome is associated with decreased TR expression and DNA binding in mouse liver. *Am J Physiol Endocrinol Metab* 284:E228–E236
86. Beigneux AP, Moser AH, Shigenaga JK, Grunfeld C, Feingold KR 2000 The acute phase response is associated with retinoid X receptor repression in rodent liver. *J Biol Chem* 275:16390–16399
87. Kwakkel J, Chassande O, van Beeren HC, Wiersinga WM, Boelen A 2008 Lacking thyroid hormone receptor  $\beta$  gene does not influence alterations in peripheral thyroid hormone metabolism during acute illness. *J Endocrinol* 197: 151–158
88. Kwakkel J, Chassande O, van Beeren HC, Fliers E, Wiersinga WM, Boelen A 2010 Thyroid hormone receptor  $\alpha$  modulates lipopolysaccharide-induced changes in peripheral thyroid hormone metabolism. *Endocrinology* 151: 1959–1969
89. Jakobs TC, Mentrup B, Schmutzler C, Dreher I, Köhrle J 2002 Proinflammatory cytokines inhibit the expression and function of human type I 5'-deiodinase in HepG2 hepatocarcinoma cells. *Eur J Endocrinol* 146:559–566
90. Nagaya T, Fujieda M, Otsuka G, Yang JP, Okamoto T, Seo H 2000 A potential role of activated NF- $\kappa B$  in the pathogenesis of euthyroid sick syndrome. *J Clin Invest* 106:393–402
91. Yu J, Koenig RJ 2000 Regulation of hepatocyte thyroxine 5'-deiodinase by T3 and nuclear receptor coactivators as a model of the sick euthyroid syndrome. *J Biol Chem* 275: 38296–38301
92. Yu J, Koenig RJ 2006 Induction of type 1 iodothyronine deiodinase to prevent the nonthyroidal illness syndrome in mice. *Endocrinology* 147:3580–3585
93. Wajner SM, Goemann IM, Bueno AL, Larsen PR, Maia AL 2011 IL-6 promotes nonthyroidal illness syndrome by blocking thyroxine activation while promoting thyroid hormone inactivation in human cells. *J Clin Invest* 121: 1834–1845
94. Boelen A, Kwakkel J, Alkemade A, Renckens R, Kaptein E, Kuiper G, Wiersinga WM, Visser TJ 2005 Induction of type 3 deiodinase activity in inflammatory cells of mice with chronic local inflammation. *Endocrinology* 146:5128–5134
95. Huang SA, Mulcahey MA, Crescenzi A, Chung M, Kim BW, Barnes C, Kuijt W, Turano H, Harney J, Larsen PR 2005 Transforming growth factor- $\beta$  promotes inactivation of extracellular thyroid hormones via transcriptional stimulation of type 3 iodothyronine deiodinase. *Mol Endocrinol* 19:3126–3136
96. Mebis L, Paletta D, Debaveye Y, Ellger B, Langouche L, D'Hoore A, Darras VM, Visser TJ, Van den Berghe G 2009 Expression of thyroid hormone transporters during critical illness. *Eur J Endocrinol* 161:243–250
97. Dozin B, Magnuson MA, Nikodem VM 1986 Thyroid hormone regulation of malic enzyme synthesis. Dual tissue-specific control. *J Biol Chem* 261:10290–10292
98. Jump DB, Narayan P, Towle H, Oppenheimer JH 1984 Rapid effects of triiodothyronine on hepatic gene expression. Hybridization analysis of tissue-specific triiodothyronine regulation of mRNAs. *J Biol Chem* 259:2789–2797
99. Gullberg H, Rudling M, Forrest D, Angelin B, Vennström B 2000 Thyroid hormone receptor  $\beta$ -deficient mice show complete loss of the normal cholesterol 7 $\alpha$ -hydroxylase (CYP7A) response to thyroid hormone but display enhanced resistance to dietary cholesterol. *Mol Endocrinol* 14:1739–1749
100. Feingold KR, Spady DK, Pollock AS, Moser AH, Grunfeld C 1996 Endotoxin, TNF, and IL-1 decrease cholesterol 7  $\alpha$ -hydroxylase mRNA levels and activity. *J Lipid Res* 37: 223–228
101. Simonides WS, van Hardeveld C 2008 Thyroid hormone as a determinant of metabolic and contractile phenotype of skeletal muscle. *Thyroid* 18:205–216
102. Yu F, Göthe S, Wikström L, Forrest D, Vennström B, Larsson L 2000 Effects of thyroid hormone receptor gene disruption on myosin isoform expression in mouse skeletal muscles. *Am J Physiol Regul Integr Comp Physiol* 278: R1545–R1554
103. Rodriguez-Perez A, Palos-Paz F, Kaptein E, Visser TJ, Dominguez-Gerpe L, Alvarez-Escudero J, Lado-Abeal J 2008 Identification of molecular mechanisms related to nonthyroidal illness syndrome in skeletal muscle and adipose tissue from patients with septic shock. *Clin Endocrinol (Oxf)* 68:821–827
104. Maia AL, Kim BW, Huang SA, Harney JW, Larsen PR 2005 Type 2 iodothyronine deiodinase is the major source of plasma T3 in euthyroid humans. *J Clin Invest* 115: 2524–2533
105. Larsen PR 2009 Type 2 iodothyronine deiodinase in hu-

- man skeletal muscle: new insights into its physiological role and regulation. *J Clin Endocrinol Metab* 94:1893–1895
106. Heemstra KA, Soeters MR, Fliers E, Serlie MJ, Burggraaf J, van Doorn MB, van der Klaauw AA, Romijn JA, Smit JW, Corssmit EP, Visser TJ 2009 Type 2 iodothyronine deiodinase in skeletal muscle: effects of hypothyroidism and fasting. *J Clin Endocrinol Metab* 94:2144–2150
  107. Feingold KR, Moser A, Patzek SM, Shigenaga JK, Grunfeld C 2009 Infection decreases fatty acid oxidation and nuclear hormone receptors in the diaphragm. *J Lipid Res* 50:2055–2063
  108. Mebis L, Langouche L, Visser TJ, Van den Berghe G 2007 The type II iodothyronine deiodinase is up-regulated in skeletal muscle during prolonged critical illness. *J Clin Endocrinol Metab* 92:3330–3333
  109. Sun X, Wray C, Tian X, Hasselgren PO, Lu J 2003 Expression of uncoupling protein 3 is upregulated in skeletal muscle during sepsis. *Am J Physiol Endocrinol Metab* 285:E512–E520
  110. Yu XX, Barger JL, Boyer BB, Brand MD, Pan G, Adams SH 2000 Impact of endotoxin on UCP homolog mRNA abundance, thermoregulation, and mitochondrial proton leak kinetics. *Am J Physiol Endocrinol Metab* 279:E433–E446
  111. Lado-Abeal J, Romero A, Castro-Piedras I, Rodriguez-Perez A, Alvarez-Escudero J 2010 Thyroid hormone receptors are down-regulated in skeletal muscle of patients with non-thyroidal illness syndrome secondary to non-septic shock. *Eur J Endocrinol* 163:765–773
  112. Cheng SY, Leonard JL, Davis PJ 2010 Molecular aspects of thyroid hormone actions. *Endocr Rev* 31:139–170
  113. Mentuccia D, Proietti-Pannunzi L, Tanner K, Bacci V, Pollin TI, Poehlman ET, Shuldiner AR, Celi FS 2002 Association between a novel variant of the human type 2 deiodinase gene Thr92Ala and insulin resistance: evidence of interaction with the Trp64Arg variant of the  $\beta$ -3-adrenergic receptor. *Diabetes* 51:880–883
  114. Canani LH, Capp C, Dora JM, Meyer EL, Wagner MS, Harney JW, Larsen PR, Gross JL, Bianco AC, Maia AL 2005 The type 2 deiodinase A/G (Thr92Ala) polymorphism is associated with decreased enzyme velocity and increased insulin resistance in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab* 90:3472–3478
  115. Dora JM, Machado WE, Rheinheimer J, Crispim D, Maia AL 2010 Association of the type 2 deiodinase Thr92Ala polymorphism with type 2 diabetes: case-control study and meta-analysis. *Eur J Endocrinol* 163:427–434
  116. Cornell RP 1989 Hyperinsulinemia elicited by interleukin-1 and nonlethal endotoxemia in rats. *Circ Shock* 28:121–130
  117. Frisard MI, McMillan RP, Marchand J, Wahlberg KA, Wu Y, Voelker KA, Heilbronn L, Haynie K, Muoio B, Li L, Hulver MW 2010 Toll-like receptor 4 modulates skeletal muscle substrate metabolism. *Am J Physiol Endocrinol Metab* 298:E988–E998
  118. Harper ME, Seifert EL 2008 Thyroid hormone effects on mitochondrial energetics. *Thyroid* 18:145–156
  119. Moreno M, de Lange P, Lombardi A, Silvestri E, Lanni A, Goglia F 2008 Metabolic effects of thyroid hormone derivatives. *Thyroid* 18:239–253
  120. Wrutniak-Cabello C, Casas F, Cabello G 2001 Thyroid hormone action in mitochondria. *J Mol Endocrinol* 26:67–77
  121. Lu B, Moser AH, Shigenaga JK, Feingold KR, Grunfeld C 2006 Type II nuclear hormone receptors, coactivator, and target gene repression in adipose tissue in the acute-phase response. *J Lipid Res* 47:2179–2190
  122. Klieverik LP, Coomans CP, Endert E, Sauerwein HP, Havekes LM, Voshol PJ, Rensen PC, Romijn JA, Kalsbeek A, Fliers E 2009 Thyroid hormone effects on whole-body energy homeostasis and tissue-specific fatty acid uptake in vivo. *Endocrinology* 150:5639–5648
  123. Khovidhunkit W, Kim MS, Memon RA, Shigenaga JK, Moser AH, Feingold KR, Grunfeld C 2004 Effects of infection and inflammation on lipid and lipoprotein metabolism: mechanisms and consequences to the host. *J Lipid Res* 45:1169–1196
  124. O'Mara BA, Dittrich W, Lauterio TJ, St Germain DL 1993 Pretranslational regulation of type I 5'-deiodinase by thyroid hormones and in fasted and diabetic rats. *Endocrinology* 133:1715–1723
  125. Wartofsky L, Latham KR, Djuh YY, Burman KD 1981 Alterations in T3 and T4 receptor binding in fasting and diabetes mellitus. *Life Sci* 28:1683–1691
  126. Chopra IJ, Huang TS, Boado R, Solomon DH, Chua Teco GN 1987 Evidence against benefit from replacement doses of thyroid hormones in nonthyroidal illness (NTI): studies using turpentine oil-injected rat. *J Endocrinol Invest* 10:559–564
  127. Van der Geyten S, Buys N, Sanders JP, Decuyper E, Visser TJ, Kühn ER, Darras VM 1999 Acute pretranslational regulation of type III iodothyronine deiodinase by growth hormone and dexamethasone in chicken embryos. *Mol Cell Endocrinol* 147:49–56
  128. Turnbull AV, Smith GW, Lee S, Vale WW, Lee KF, Rivier C 1999 CRF type I receptor-deficient mice exhibit a pronounced pituitary-adrenal response to local inflammation. *Endocrinology* 140:1013–1017
  129. Gyorffy A, Sayed-Ahmed A, Zsarnovszky A, Frenyó VL, Decuyper E, Bartha T 2009 Effects of energy restriction on thyroid hormone metabolism in chickens. *Acta Vet Hung* 57:319–330
  130. Malik IA, Baumgartner BG, Naz N, Sheikh N, Moriconi F, Ramadori G 2010 Changes in gene expression of DOR and other thyroid hormone receptors in rat liver during acute-phase response. *Cell Tissue Res* 342:261–272
  131. Chamba A, Neuberger J, Strain A, Hopkins J, Sheppard MC, Franklyn JA 1996 Expression and function of thyroid hormone receptor variants in normal and chronically diseased human liver. *J Clin Endocrinol Metab* 81:360–367
  132. Kwakkel J, van Beeren HC, Ackermans MT, Platvoet-Ter Schiphorst MC, Fliers E, Wiersinga WM, Boelen A 2009 Skeletal muscle deiodinase type 2 regulation during illness in mice. *J Endocrinol* 203:263–270
  133. Silva JE, Larsen PR 1983 Adrenergic activation of triiodothyronine production in brown adipose tissue. *Nature* 305:712–713
  134. Dentice M, Marsili A, Ambrosio R, Guardiola O, Sibilio A, Paik JH, Minchiotti G, DePinho RA, Fenzi G, Larsen PR, Salvatore D 2010 The FoxO3/type 2 deiodinase pathway is required for normal mouse myogenesis and muscle regeneration. *J Clin Invest* 120:4021–4030
  135. Watanabe M, Houten SM, Matakaki C, Christoffolete MA,

- Kim BW, Sato H, Messaddeq N, Harney JW, Ezaki O, Kodama T, Schoonjans K, Bianco AC, Auwerx J 2006 Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. *Nature* 439:484–489
136. Grozovsky R, Ribich S, Rosene ML, Mulcahey MA, Huang SA, Patti ME, Bianco AC, Kim BW 2009 Type 2 deiodinase expression is induced by peroxisomal proliferator-activated receptor- $\gamma$  agonists in skeletal myocytes. *Endocrinology* 150:1976–1983
  137. Köhrle J 1999 Local activation and inactivation of thyroid hormones: the deiodinase family. *Mol Cell Endocrinol* 151:103–119
  138. Vary TC, O'Neill P, Cooney RN, Maish 3rd G, Shumate M 1999 Chronic infusion of interleukin 1 induces hyperlactatemia and altered regulation of lactate metabolism in skeletal muscle. *JPEN J Parenter Enteral Nutr* 23:213–217
  139. Goglia F, Lanni A, Barth J, Kadenbach B 1994 Interaction of diiodothyronines with isolated cytochrome c oxidase. *FEBS Lett* 346:295–298
  140. Dellinger RP, Levy MM, Carlet JM, Bion J, Parker MM, Jaeschke R, Reinhart K, Angus DC, Brun-Buisson C, Beale R, Calandra T, Dhainaut JF, Gerlach H, Harvey M, Marini JJ, Marshall J, Ranieri M, Ramsay G, Sevransky J, Thompson BT, Townsend S, Vender JS, Zimmerman JL, Vincent JL 2008 Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock: 2008. *Crit Care Med* 36:296–327
  141. Bernard GR, Vincent JL, Laterre PF, LaRosa SP, Dhainaut JF, Lopez-Rodriguez A, Steingrub JS, Garber GE, Helterbrand JD, Ely EW, Fisher Jr JC 2001 Efficacy and safety of recombinant human activated protein C for severe sepsis. *N Engl J Med* 344:699–709
  142. McClean KL, Sheehan GJ, Harding GK 1994 Intraabdominal infection: a review. *Clin Infect Dis* 19:100–116
  143. Richmand DA, Molitch ME, O'Donnell TF 1980 Altered thyroid hormone levels in bacterial sepsis: the role of nutritional adequacy. *Metabolism* 29:936–942
  144. Woeber KA 1971 Alterations in thyroid hormone economy during acute infection with *Diplococcus pneumoniae* in the rhesus monkey. *J Clin Invest* 50:378–387
  145. Burgi U, Feller C, Gerber AU 1986 Effects of an acute bacterial infection on serum thyroid hormones and nuclear triiodothyronine receptors in mice. *Endocrinology* 119:515–521
  146. Pinsky MR 2004 Dysregulation of the immune response in severe sepsis. *Am J Med Sci* 328:220–229
  147. den Brinker M, Dumas B, Visser TJ, Hop WC, Hazelzet JA, Festen DA, Hokken-Koelega AC, Joosten KF 2005 Thyroid function and outcome in children who survived meningococcal septic shock. *Intensive Care Med* 31:970–976
  148. Bernstein JM 1999 Treatment of community-acquired pneumonia—IDSA guidelines. *Infectious Diseases Society of America. Chest* 115:9S–13S
  149. Knapp S, Hareng L, Rijnneveld AW, Bresser P, van der Zee JS, Florquin S, Hartung T, van der Poll T 2004 Activation of neutrophils and inhibition of the proinflammatory cytokine response by endogenous granulocyte colony-stimulating factor in murine pneumococcal pneumonia. *J Infect Dis* 189:1506–1515
  150. Renckens R, Roelofs JJ, Florquin S, de Vos AF, Lijnen HR, van't Veer C, van der Poll T 2006 Matrix metalloproteinase-9 deficiency impairs host defense against abdominal sepsis. *J Immunol* 176:3735–3741
  151. Boelen A, Boersma J, Kwakkel J, Wieland CW, Renckens R, Visser TJ, Fliers E, Wiersinga WM 2008 Type 3 deiodinase is highly expressed in infiltrating neutrophilic granulocytes in response to acute bacterial infection. *Thyroid* 18:1095–1103
  152. Angstwurm MW, Schopohl J, Gaertner R 2004 Selenium substitution has no direct effect on thyroid hormone metabolism in critically ill patients. *Eur J Endocrinol* 151:47–54
  153. Fredriksson K, Hammarqvist F, Strigård K, Hultenby K, Ljungqvist O, Wernerman J, Rooyackers O 2006 Derangements in mitochondrial metabolism in intercostal and leg muscle of critically ill patients with sepsis-induced multiple organ failure. *Am J Physiol Endocrinol Metab* 291:E1044–E1050
  154. Zang Q, Maass DL, Tsai SJ, Horton JW 2007 Cardiac mitochondrial damage and inflammation responses in sepsis. *Surg Infect (Larchmt)* 8:41–54
  155. Levy RJ, Deutschman CS 2007 Cytochrome c oxidase dysfunction in sepsis. *Crit Care Med* 35:S468–S475
  156. Stevens RD, Dowdy DW, Michaels RK, Mendez-Tellez PA, Pronovost PJ, Needham DM 2007 Neuromuscular dysfunction acquired in critical illness: a systematic review. *Intensive Care Med* 33:1876–1891
  157. Klebanoff SJ 2005 Myeloperoxidase: friend and foe. *J Leukoc Biol* 77:598–625
  158. Klebanoff SJ 1967 Iodination of bacteria: a bactericidal mechanism. *J Exp Med* 126:1063–1078
  159. Siegel E, Sachs BA 1964 *In vitro* leukocyte uptake of 131-I labeled iodide, thyroxine and triiodothyronine, and its relation to thyroid function. *J Clin Endocrinol Metab* 24:313–318
  160. Woeber KA, Ingbar SH 1973 Metabolism of L-thyroxine by phagocytosing human leukocytes. *J Clin Invest* 52:1796–1803
  161. Adelberg HM, Siemsen JK, Jung RC, Nicoloff JT 1971 Scintigraphic detection of pulmonary bacterial infections with labeled thyroid hormones and pertechnetate. *Radiology* 99:141–146
  162. Gregerman RI, Solomon N 1967 Acceleration of thyroxine and triiodothyronine turnover during bacterial pulmonary infections and fever: implications for the functional state of the thyroid during stress and in senescence. *J Clin Endocrinol Metab* 27:93–105
  163. Kaptein EM, Robinson WJ, Grieb DA, Nicoloff JT 1982 Peripheral serum thyroxine, triiodothyronine and reverse triiodothyronine kinetics in the low thyroxine state of acute nonthyroidal illnesses. A noncompartmental analysis. *J Clin Invest* 69:526–535
  164. Boelen A, Mikita J, Boiziau C, Chassande O, Fliers E, Petry KG 2009 Type 3 deiodinase expression in inflammatory spinal cord lesions in rat experimental autoimmune encephalomyelitis. *Thyroid* 19:1401–1406
  165. Van den Berghe G 2001 The neuroendocrine response to stress is a dynamic process. *Best Pract Res Clin Endocrinol Metab* 15:405–419
  166. Weekers F, Van Herck E, Coopmans W, Michalaki M, Bowers CY, Veldhuis JD, Van den Berghe G 2002 A novel in vivo rabbit model of hypercatabolic critical illness re-

- veals a biphasic neuroendocrine stress response. *Endocrinology* 143:764–774
167. Peeters RP, Wouters PJ, Kaptein E, van Toor H, Visser TJ, Van den Berghe G 2003 Reduced activation and increased inactivation of thyroid hormone in tissues of critically ill patients. *J Clin Endocrinol Metab* 88:3202–3211
  168. Peeters RP, Kester MH, Wouters PJ, Kaptein E, van Toor H, Visser TJ, Van den Berghe G 2005 Increased thyroxine sulfate levels in critically ill patients as a result of a decreased hepatic type I deiodinase activity. *J Clin Endocrinol Metab* 90:6460–6465
  169. Peeters RP, Wouters PJ, van Toor H, Kaptein E, Visser TJ, Van den Berghe G 2005 Serum 3,3',5'-triiodothyronine (rT3) and 3,5,3'-triiodothyronine/rT3 are prognostic markers in critically ill patients and are associated with postmortem tissue deiodinase activities. *J Clin Endocrinol Metab* 90:4559–4565
  170. Williams GR, Franklyn JA, Neuberger JM, Sheppard MC 1989 Thyroid hormone receptor expression in the “sick euthyroid” syndrome. *Lancet* 2:1477–1481
  171. Thijssen-Timmer DC, Peeters RP, Wouters P, Weekers F, Visser TJ, Fliers E, Wiersinga WM, Bakker O, Van Den Berghe G 2007 Thyroid hormone receptor isoform expression in livers of critically ill patients. *Thyroid* 17:105–112
  172. Weekers F, Giulietti AP, Michalaki M, Coopmans W, Van Herck E, Mathieu C, Van den Berghe G 2003 Metabolic, endocrine, and immune effects of stress hyperglycemia in a rabbit model of prolonged critical illness. *Endocrinology* 144:5329–5338
  173. Weekers F, Michalaki M, Coopmans W, Van Herck E, Veldhuis JD, Darras VM, Van den Berghe G 2004 Endocrine and metabolic effects of growth hormone (GH) compared with GH-releasing peptide, thyrotropin-releasing hormone, and insulin infusion in a rabbit model of prolonged critical illness. *Endocrinology* 145:205–213
  174. Debaveye Y, Ellger B, Mebis L, Van Herck E, Coopmans W, Darras V, Van den Berghe G 2005 Tissue deiodinase activity during prolonged critical illness: effects of exogenous thyrotropin-releasing hormone and its combination with growth hormone-releasing peptide-2. *Endocrinology* 146:5604–5611
  175. Simonides WS, Mulcahey MA, Redout EM, Muller A, Zuidwijk MJ, Visser TJ, Wassen FW, Crescenzi A, da-Silva WS, Harney J, Engel FB, Obregon MJ, Larsen PR, Bianco AC, Huang SA 2008 Hypoxia-inducible factor induces local thyroid hormone inactivation during hypoxic-ischemic disease in rats. *J Clin Invest* 118:975–983
  176. Debaveye Y, Ellger B, Mebis L, Visser TJ, Darras VM, Van den Berghe G 2008 Effects of substitution and high-dose thyroid hormone therapy on deiodination, sulfoconjugation, and tissue thyroid hormone levels in prolonged critically ill rabbits. *Endocrinology* 149:4218–4228
  177. Vanhorebeek I, De Vos R, Mesotten D, Wouters PJ, De Wolf-Peters C, Van den Berghe G 2005 Protection of hepatocyte mitochondrial ultrastructure and function by strict blood glucose control with insulin in critically ill patients. *Lancet* 365:53–59
  178. Kaptein EM, Beale E, Chan LS 2009 Thyroid hormone therapy for obesity and nonthyroidal illnesses: a systematic review. *J Clin Endocrinol Metab* 94:3663–3675
  179. Van den Berghe G, de Zegher F, Bouillon R 1998 Clinical review 95: acute and prolonged critical illness as different neuroendocrine paradigms. *J Clin Endocrinol Metab* 83:1827–1834
  180. Pingitore A, Galli E, Barison A, Iervasi A, Scarlattini M, Nucci D, L'abbate A, Mariotti R, Iervasi G 2008 Acute effects of triiodothyronine (T3) replacement therapy in patients with chronic heart failure and low-T3 syndrome: a randomized, placebo-controlled study. *J Clin Endocrinol Metab* 93:1351–1358
  181. Kaptein EM, Sanchez A, Beale E, Chan LS 2010 Clinical review: thyroid hormone therapy for postoperative nonthyroidal illnesses: a systematic review and synthesis. *J Clin Endocrinol Metab* 95:4526–4534
  182. Debaveye Y, Ellger B, Mebis L, Darras VM, Van den Berghe G 2008 Regulation of tissue iodothyronine deiodinase activity in a model of prolonged critical illness. *Thyroid* 18:551–560
  183. Weekers F, Van den Berghe G 2004 Endocrine modifications and interventions during critical illness. *Proc Nutr Soc* 63:443–450