

## RESEARCH ARTICLE

# Bioaccumulation of heavy metals in some commercially important fishes from a tropical river estuary suggests higher potential health risk in children than adults

A. S. Shafiuddin Ahmed<sup>1</sup>, Sharmin Sultana<sup>2</sup>, Ahasan Habib<sup>1,3\*</sup>, Hadayet Ullah<sup>4</sup>, Najiah Musa<sup>3</sup>, M. Belal Hossain<sup>1</sup>, Md. Mahfujur Rahman<sup>5</sup>, Md. Shafiqul Islam Sarker<sup>6</sup>

**1** Department of Fisheries and Marine Science, Noakhali Science and Technology University, Noakhali, Bangladesh, **2** Department of Chemistry, National University, Gazipur, Bangladesh, **3** Faculty of Fisheries and Food Science, Universiti Malaysia Terengganu, Kuala Nerus, Terengganu, Malaysia, **4** Southern Seas Ecology Laboratories, School of Biological Sciences, The University of Adelaide, Adelaide, South Australia, SA, Australia, **5** Department of Statistics, Cumilla University, Cumilla, Bangladesh, **6** Forensic Science Laboratory, Rapid Action Battalions Headquarters, Dhaka, Bangladesh

\* [a.habib@umt.edu.my](mailto:a.habib@umt.edu.my)



## OPEN ACCESS

**Citation:** Ahmed ASS, Sultana S, Habib A, Ullah H, Musa N, Hossain MB, et al. (2019) Bioaccumulation of heavy metals in some commercially important fishes from a tropical river estuary suggests higher potential health risk in children than adults. PLoS ONE 14(10): e0219336. <https://doi.org/10.1371/journal.pone.0219336>

**Editor:** Zhenguang Yan, Chinese Research Academy of Environmental Sciences, CHINA

**Received:** June 19, 2019

**Accepted:** September 12, 2019

**Published:** October 17, 2019

**Copyright:** © 2019 Ahmed et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are with the manuscript.

**Funding:** Sampling was performed by self-funded. Heavy metal analysis were conducted and funded by Rapid Action Battalion Headquarter, Bangladesh. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

## Abstract

The Karnaphuli River estuary, located in southeast coast of Bangladesh, is largely exposed to heavy metal contamination as it receives a huge amount of untreated industrial effluents from the Chottagram City. This study aimed to assess the concentrations of five heavy metals (As, Pb, Cd, Cr and Cu) and their bioaccumulation status in six commercially important fishes, and also to evaluate the potential human health risk for local consumers. The hierarchy of the measured concentration level (mg/kg) of the metals was as follows: Pb (13.88) > Cu (12.10) > As (4.89) > Cr (3.36) > Cd (0.39). The Fulton's condition factor denoted that fishes were in better 'condition' and most of the species were in positive allometric growth. The bioaccumulation factors (BAFs) of the contaminants observed in the species were in the following orders: Cu (1971.42) > As (1042.93) > Pb (913.66) > Cr (864.99) > Cd (252.03), and among the specimens, demersal fish, *Apocryptes bato* appeared to be the most bioaccumulative organism. Estimated daily intake (EDI), target hazard quotient (THQ), hazard index (HI) and carcinogenic risk (CR) assessed for potential human health risk implications suggest that the values were within the acceptable threshold for both adults and children. However, calculated CR values indicated that both age groups were not far from the risk, and HI values demonstrated that children were nearly 6 times more susceptible to non-carcinogenic and carcinogenic health effects than adults.

## Introduction

Heavy metals pollution has become a major concern worldwide due to their toxicity, intrinsic persistence, non-biodegradable nature, and accumulative behaviors [1]. These metals differ from other toxic materials in a way that they are neither created nor destroyed by human. They are inert in the environment and are often considered to be conservative pollutants if left

undisturbed [2]. However, the rapid industrialization, urbanization, population growth, agricultural and other human activities have resulted in severe pollution by heavy metals globally, especially in developing countries [3]. Significant quantities of heavy metals from such activities are discharged into rivers, which can be strongly accumulated and biomagnified along water, sediment, and aquatic food chain, resulting in sublethal effects or death in local fish populations [4]. As fishes occupy higher trophic level in the food chain, they are considered one of the most common bioindicators for pollutants [5, 6]. Again, fishes are consumed by human as a major source of protein for many years. Thus, the human body is largely susceptible to enriched heavy metal concentration in fishes [7]. Consequently, an analysis of the levels of heavy metals in fish could be used to investigate anthropogenic impacts on ecosystem and human health.

Generally, bioaccumulation and biomagnification occur due to longstanding anthropogenic activities within a coastal ecosystem [8]. The accumulation of heavy metals in fish organs could also be driven by physiochemical and biological variables such as pH, temperature, hardness, exposure duration, feeding habits of species and habitat complexity [9]. While terrestrial species exhibit a strong pattern of biomagnification, marine and estuarine organisms show less clear pattern [10]. Condition factor, is one of the most common tools that is widely used to assess the life, reproduction and health conditions, as well as the life cycle of a fish species [11]. Along with that, condition factor also suggests the food availability and quality, breeding duration, and process for distinct populations [12, 13]. In addition, this tool indicates the status of fish health due to stress in the population within an ecosystem [14].

Heavy metals and metalloids, when occurring at higher concentrations, become severe poisons for all living organisms including human. For example, an excessive amount of Hg, As, Pb, and Cd elements could be detrimental to the living cells, and a prolonged exposure to the body can lead to illness or death [15]. Among the metals, Hg is the most toxic metal in our environment. Methyl mercury toxicity are inhibition of protein synthesis, microtubule disruption, increase of intracellular  $\text{Ca}^{2+}$  with disturbance of neurotransmitter function [16]. Prostatic proliferative lesions, lung cancer, bone fractures, kidney failures are associated with chronic exposure to Cd, even at a low concentration of ~ 1 mg/kg [17, 18]. Excessive Pb can have detrimental health effect on human [19] including nervous system disorders, mental retardation, skeletal hematopoietic function disorder and even death [20]. Cr was reported to have carcinogenic effects on human health [21, 22].

The Karnaphuli River estuary is one of the potential fish habitats along the southeast coast of Bay of Bengal known to be an important breeding, feeding and nursery ground for many aquatic species. At present, the ecosystem is receiving untreated effluents from several industries including textile crafts, dyeing industry and others as it passes through the industrial zone [23]. A number of studies have attempted to assess the contamination status from river and estuarine environment from Bangladesh [3, 24–27], from China [28], from Turkey [29]. However, to date there has been no proper investigation carried out on the potential human health risk due to heavy metal contamination in the fish species harvested and consumed from the Karnaphuli estuary. Therefore, this study aimed to fill this knowledge gap by assessing the concentration of heavy metals in some selected fish species and their bioaccumulation status, and the human health risk for local adult and children consumers.

## Materials and methods

### Ethical statement

Specimens from wild populations were collected from local fishermen. None of the sampled species were endangered or protected. No permit was required to conduct the present study. There were no ethical considerations linked to the experiment.

### Sampling

The study area (Karnaphuli River estuary) is located from 22.234008 N and 91.821105 E to 22.289695 N and 91.794403 E (Fig 1). A total of six commercial species (i.e. *Apocryptes bato* (Chewa), *Pampus chinensis* (Rup chanda), *Liza parsia* (Bata), *Mugil cephalus* (Flathead bata), *Hyporhamphus limbatus* (Ek Thuitta), and *Tenualosa toil* (Chandana ilish) were collected from fishermen for a period of seven months (February 2018 to August 2018) using seine net. The collected samples were carried in plastic ice container and immediately stored at  $-20^{\circ}\text{C}$ . Afterwards in the lab, total length (cm) and weights (gm) were measured carefully to the nearest 0.1 cm using a vernier caliper; total weight was determined with an electronic balance to 0.01 g accuracy. Muscles of each specimen were dissected with stainless steel scissors for further chemical analysis using inductively coupled plasma mass spectrometry (ICP-MS, Model: ELAN9000, Perkin-Elmer, Germany) for metal detection. Data were analysed statistically by fitting a straight line adopting the least square method.

### Chemical analysis procedures and accuracy

A 1.5 g of dissected muscle was used for subsequent analysis. The samples were dried in a Zirbus freeze drying machine (Model: VaCo 2, Germany, CFC free, condenser dimensions-

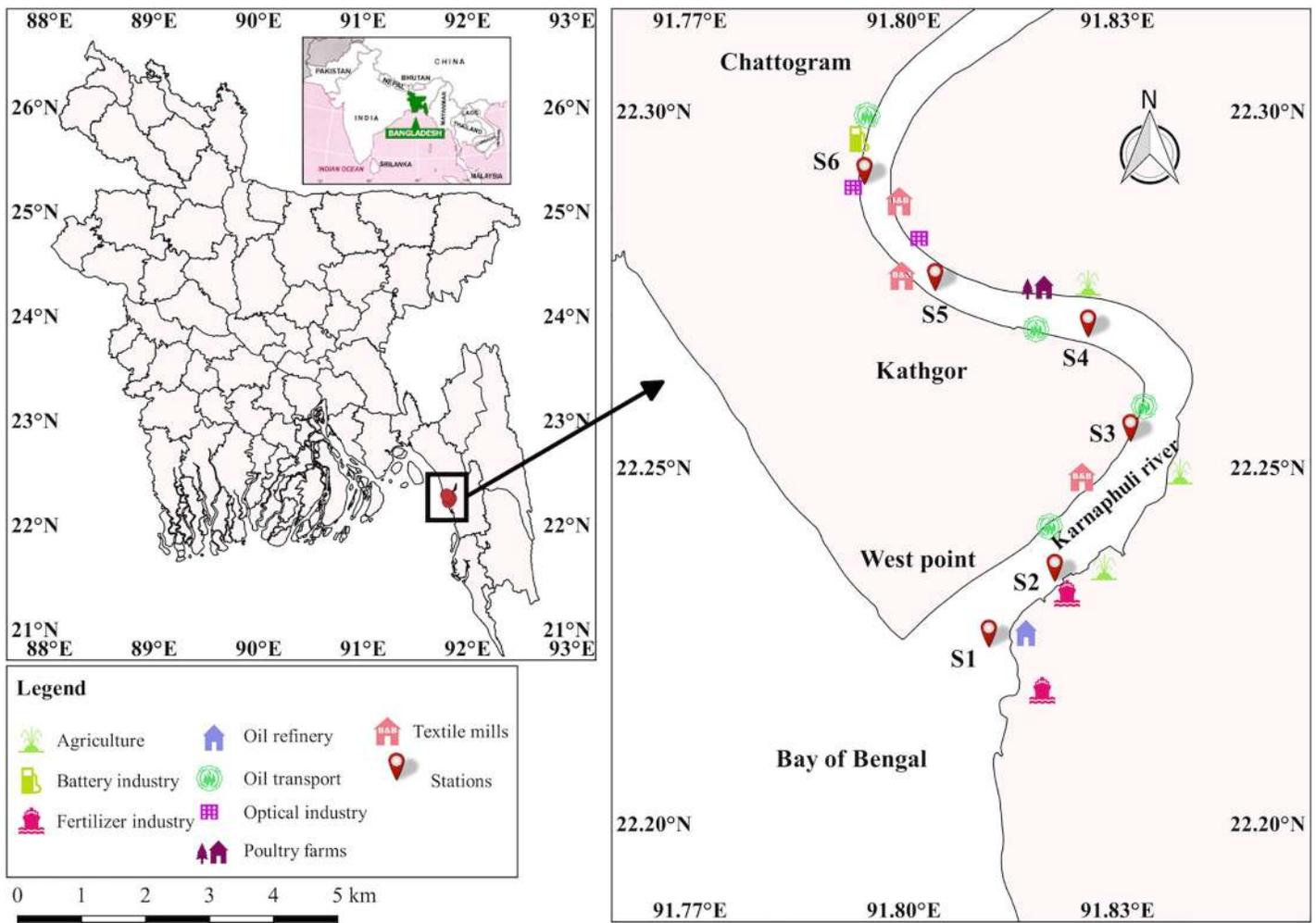


Fig 1. Six sampling locations in Karnaphuli River estuary and the source of heavy metal pollution. [Reprinted from ([www.wikipedia.org](http://www.wikipedia.org)) under a CC BY license, with permission from (Wikipedia), original copyright (2007)].

<https://doi.org/10.1371/journal.pone.0219336.g001>

Ø200×200 mm) for 24 h [30]. After cooling, 0.5 g of muscle samples were placed in 10ml of 100% HNO<sub>3</sub>. The solution was heated on an oil bath and added with 4 drops of H<sub>2</sub>O<sub>2</sub> repeatedly until the mixture turned clear. The solution was mineralized using microwave digester (model: WX-6000, China). After filtering, solution was diluted for 2h and kept in a 50 ml tezaron tube [3]. A standard reagent (Merck VI; Germany) was used to analyze the metal concentration in the targeted fish species, which was prepared from the same acid matrix [1]. Moreover, certified reference material (CRM 320, Merck KGaA, Germany) was employed for validation and accuracy check [3].

### Metal pollution index (MPI)

To assess the metal pollution, the metal pollution index (MPI) was adopted following [31] and [32]. The equation is as follows:

$$MPI = (CM_1 \times CM_2 \times CM_3 \times \dots \times CM_n)^{1/n}$$

where, CM<sub>1</sub> is the concentration value of first concerned metal, CM<sub>2</sub> is the concentration value of second concerned metal, CM<sub>3</sub> is the concentration of third concerned metal, CM<sub>n</sub> is the concentration of n<sup>th</sup> metal (mg/kg dry wt) in the tissue sample of a certain species.

### Statistical analysis

The mean and standard deviation of metal concentrations were calculated. The Kolmogorov-Smirnov, Shapiro-Wilk and Kruskal-Wallis tests were performed using SPSS 23. The Kolmogorov-Smirnov and Shapiro-Wilk tests were performed to check the normality of the data [33]. The Kruskal-Wallis test was carried out to identify significant variance of the targeted elements in the specimens of the studied area where  $p \leq 0.05$  was used as the cut-off for significance (confidence level in 95%). In addition, the Levene's test with the significant level at  $p \leq 0.05$  was adopted to determine the homogeneity of variances in terms of ANOVA tests. Correlation matrix (CM) and principal component analysis (PCA) were performed to determine the correlations and association between heavy metals in the studied fish species [34]. The correlation matrix used to analyze heavy metals could result in both positive and negative outcomes [35–38]. Hierarchical cluster analysis is one of the most widely used hierarchical algorithms which results in clusters in which variables or individuals are added in sequence considering hierarchy of the cluster [39]. Additionally, the similar groups of the studied elements within the sampling sites measured as a distance between the two closest members, and is conducted by Origin 9 software [3, 40]. Metals with similar properties were pooled in one or associated cluster while the dissimilar groups of elements were plotted in a separate cluster, and thus differentiated the contamination status of the samples [41–43].

### Bioaccumulation factor

Bioaccumulation factors (BAFs) were calculated as a ratio between the concentration level of biota (those in water) and the living environment of the specimens, and was expressed as follows [44–46]:

$$BAF = \frac{Cn_{Biota}}{Cn_{Water}}$$

where  $Cn_{Biota}$  is the concentration of metal in the tissues (mg/kg) and  $Cn_{Water}$  is the metal concentration in the aquatic environment (mg/l). BAF is categorized as follows: BAF < 1000: no

probability of accumulation;  $1000 < \text{BAF} < 5000$ : bioaccumulative;  $\text{BAF} > 5000$ : extremely bioaccumulative [47].

### Length-weight relationship and condition factor

The length-weight relationship of the fish samples were calculated using Fulton condition factor following the equation [48–51]:

$$Q = 100 \times W / L^3$$

where  $W$  is the total body weight of fish (gm),  $L$  is the total length of fish (cm). Fulton's  $Q$  is categorized as follows:  $Q = 1$ : Condition is poor,  $Q = 1.2$ : condition is moderate,  $Q \geq 1.40$ : condition is proportionally good [52]. The equation can be expressed by the following formula [12, 53]:

$$W = aL^b$$

The equation can be estimated using the least-square formula adopted with the logarithm form of the equation is shown as [54]:

$$\log W = \log a + b \log L$$

where 'a' is the calculated intercept of the regression line, and 'b' is the coefficient of that regression. The 'b' values signify the growth pattern of an organism which can be classified as follows:  $b < 3$ : negative allometric,  $b = 3$ : Isometric and  $b > 3$ : positive allometric [48, 55].

As Fulton's  $Q$  is substantially correlated with the length-weight relationship, exponent 'b' acts an identical role of determining the well-being of the organisms [48]. The deviation of the condition, further, depends on the food availability and the divergence of reproductive organ development [56].

### Human health risk

#### Estimated daily intake (EDI)

Estimated daily intake (EDI) was calculated by the following equation [22, 57, 58]:

$$EDI = \frac{(Cn \times IGr)}{Bwt}$$

where  $Cn$  is the concentration level of metal in the selected fish tissues (mg/kg dry-wt);  $IGr$  is the acceptable ingestion rate, which is 55.5 g/day for adults and 52.5 g/day for children [59, 60];  $Bwt$  is the body weight: 70 kg for adults and 15 kg for children [59].

#### Target hazard quotient (THQ) for non-carcinogenic risk assessment

THQ was estimated by the ratio of EDI and oral reference dose (RfD). RfDs of the different metals for example As, Pb, Cd, Cr, and Cu are 0.0003, 0.002, 0.001, 0.003 and 0.3, respectively [59, 61]. The value of ratio  $< 1$  implies a non-significant risk effects [62]. The THQ formula is expressed as follows [60, 63–65]:

$$\text{THQs} = \frac{E_d \times E_p \times \text{EDI}}{A_t \times \text{RfD}} \times 10^{-3}$$

Where  $E_d$  is exposure duration (65 years) (USEPA, 2008);  $E_p$  is exposure frequency (365 days/year) [21];  $A_t$  is the average time for the non-carcinogenic element ( $E_d \times E_p$ ).

## Hazard index (HI)

Hazard index (HI) was calculated for the multiple elements (Hg, As, Mn, and Cr) found in the fish samples and the equation is as follows [3, 19, 66, 67]:

$$HI = \sum_{i=k}^n THQs$$

where, THQs is the estimated risk value for individual metal [68]. When HI value is higher than 10, the non-carcinogenic risk effect is considered high for exposed consumers [69–71].

## Carcinogenic risk (CR)

To assess the probability of developing cancer over a lifetime, the carcinogenic risk is evaluated for the consequence of exposure to the substantial carcinogens [72, 73]. The acceptable range of the risk limit is  $10^{-6}$  to  $10^{-4}$  [74–77]. CRs higher than  $10^{-4}$  are likely to increase the probability of carcinogenic risk effect [78, 79]. The established equation to assess the CR is as follows [60, 61, 80, 81]:

$$CR = \frac{E_d \times E_p \times EDI \times CSf}{A_t} \times 10^{-3}$$

Where CSf is oral slope factor of particular carcinogen (mg/kg-day) [74]. Available CSf values (mg/kg-day) are: As (1.5), Pb (0.0085) and Cd (6.3) [74].

## Results

### The concentration of heavy metals and source identification

The average concentration of Pb, Cu, As, Cr, and Cd from the fish tissues were 13.88 mg/kg (range: 3.19–6.19 mg/kg); 12.10 mg/kg (range: 10.27–16.41 mg/kg); 4.89 mg/kg (range: 3.19–6.19 mg/kg); 3.36 mg/kg (range: 2.46–4.17 mg/kg), and 0.39 mg/kg (range: 0.21–0.74 mg/kg) respectively. Therefore, the hierarchy of the metal concentrations was: Pb > Cu > As > Cr > Cd (please see Table 1). From the results, Pb was 1.14, 2.8, 4.13, and 35.44 folds higher than Cu, As, Cr, and Cd respectively, and contributed 40% of all the elements in the study area. Along with that, Pb attributed a higher concentration of 15.73 mg/kg at S1, whereas Cd was accounted for lower concentration of 0.29 mg/kg on average observed at S1.

The evaluated MPIs ranged from 3.65 mg/kg to 4.70 mg/kg with the mean of 4.02 mg/kg (Table 1). Due to higher concentration level, the maximum MPI value (4.70 mg/kg) was corresponded to *A. bato*, followed by *P. chinensis* (4.2 mg/kg) *H. limbatus* (4.05 mg/kg), *T. toli* (3.75 mg/kg), *M. cephalus* (3.69 mg/kg), and *L. parsia* (3.65 mg/kg).

From the statistical results, Kolmogorov-Smirnov and Shapiro-Walk test revealed that the metals in the targeted fish species were non-normally distributed along the study area. The adopted Levene's tests showed that metals were non-homogenously distributed. Kruskal-Wallis test identified that the distribution of metal was significantly different ( $p \leq 0.05$ ) in the fish species along the sampling stations. From Table 2, among the species, *L. parsia* and *H. limbatus* exhibited significant relationship (regression line) with As only, where *T. toli* showed significant relationship with As and Pb ( $p \leq 0.05$ ). None of the other metals showed a significant linear relationship with the organisms. Among the species, *T. toli* showed the maximum response for Pb ( $R^2 = 99.5\%$ ), followed by *L. parsia* for As ( $R^2 = 83.7\%$ ).

Table 3 depicts the correlation matrix among the metals in different fish species. The most significant positive correlations were observed between As–Pb ( $r = 0.912$ ), As–Cr ( $r = 0.904$ ), Pb–Cr ( $r = 0.917$ ), Pb–Cu ( $r = 0.967$ ), and Cd–Cu ( $r = 0.911$ ) at  $p < 0.05$  level. Based on CM,

**Table 1. Concentration of heavy metals of different species and their feeding nature, length and weight and a comparison of other relevant studies along with various standard guideline values.**

Species	Feeding nature	Amounts	Length (cm)	Weight (gm)	Heavy metals (mean ± std) mg/kg					MPI	References
					As	Pb	Cd	Cr	Cu		
<i>A. bato</i>	Demersal	18	13.26±1.35	72.64±6.39	4.65±1.06	15.22±1.32	0.60±0.13	3.54±0.54	15.29±3.82	4.70	
<i>P. chinensis</i>	Demersal	18	6.96±0.90	166.58±21.68	5.03±0.86	14±1.79	0.44±0.14	3.59±0.55	13.10±2.49	4.29	
<i>L. parsia</i>	Demersal	18	7.34±1.17	25.20±3.12	4.36±0.93	13.98±1.93	0.34±0.09	3.30±0.40	9.50±1.16	3.65	
<i>M. cephalus</i>	Pelagic	18	27.17±2.30	711.44±111.03	4.89±0.48	12.70±1.72	0.31±0.09	3.14±0.36	11.48±1.27	3.69	
<i>H. limbatus</i>	Pelagic	18	10.97±0.80	26.77±4.39	5.14±0.86	13.77±1.54	0.35±0.08	3.52±0.48	12.52±1.40	4.05	
<i>T. toli</i>	Pelagic	18	30.68±2.54	646.82±41.16	5.26±0.49	13.61±0.82	0.31±0.07	3.11±0.54	10.72±1.47	3.75	
Water (mg/l)					0.006±0.003	0.017±0.006	0.002±0.001	0.006±0.002	0.006±0.001		
<b>Guidelines (mg/kg)</b>											
FAO					1	2.5	0.2	1	10		[82]
WHO					0.01	2	–	0.15	3		[83]
EU					–	0.1	0.05	1	3		[84]
Bangladesh (fish)					5	0.30	0.25	–	5.00		[85]
<b>Literature (mg/kg)</b>											
Coastal area, Bangladesh					0.08–13	0.07–0.63	0.03–0.09	0.15–2.2	1.3–14		[86]
Arasalar River, India					–	0.23	6.13	0.3	–		[87]
North east coast, India					0.64	–	0.33	–	3.9		[88]
Ganga River, India					–	3–6	0.1–2.9	–	10–100		[89]
Pearl River, China					–	8.64	8.55	8.73	2.48		[90]
Meiliang Bay, China					–	0.636	0.173	0.118	0.336		[91]
Iskenderun Bay, Turkey					–	0.09–6.95	0.01–4.16	0.07–6.46	0.04–5.43		[92]

<https://doi.org/10.1371/journal.pone.0219336.t001>

the rotated component plot, PCA was depicted in Fig 2. The PCA exhibited two factors PC 1 and PC 2, which resulted in the corresponded variance of 89.18% and 7.75% respectively. Both factors were responsible for cumulative variance of 96.93%. Cu and Pb were the most dominant elements in PC 1, corresponded with the loadings of 20.78% and 20.70% respectively. On the contrary, in PC 2, Cd and Cu showed the domination with the loadings of 77.81% and 22.19% respectively.

To execute hierarchical cluster dendrogram, the Ward-Linkage method was employed with Euclidean distance, which resulted in three distinct clusters, presented in Fig 3. Cluster 1 included As and Cr that could have been originated from anthropogenic activities like chemical industries. Pb, and Cu confined in cluster 2 could have been attributed from the textile organization, fertilization and oil droppings from the boats/ships in the study area. Lastly, Cd was included in cluster 3, and source of the Cd could have come from battery industry.

### Length-weight Relationship and condition factor evaluation

Higher ‘b’ value reflected the appetite state and reproductive organ development of the species [55]. The identified b value of *L. parsia* from the length weight relationship was close to 3, hence, it represented isometric growth pattern that was considered as ideal shape (Table 4). Meanwhile, among all, species, *P. chinensis* exhibited the highest positive allometric growth, which was 37 times higher, than average value and 14 folds, on average, higher than other species.

### Bioaccumulation (BAF) status of targeted species

The estimated BAFs are depicted in Fig 4. The BAFs were ranged from 110.53 for Cd observed in S4 to 3353.7 for Cu as well. The minimum value was found for *T. toli*, on the

Table 2. A regression analysis between metal distributions in the selected specimens.

Species and metals	Regression equation	Standard error	Pearson's r	P values	R <sup>2</sup> (%)
<i>A. bato</i>					
As	y = 8.66+0.99x	0.404	0.774	0.070	58.1
Pb	y = 11.88+0.09x	0.511	0.088	0.867	0.8
Cd	y = 13.44–0.31x	5.166	-0.029	0.955	0.01
Cr	y = 12.70+0.16x	1.25	0.063	0.904	0.4
Cu	y = 11.56+0.11x	0.168	0.313	0.545	9.8
<i>P. chinensis</i>					
As	y = 4.19 + 0.55x	0.449	0.522	0.287	27.3
Pb	y = 5.14 + 0.13x	0.243	0.257	0.621	6.6
Cd	y = 5.72 + 2.81x	3	0.425	0.4	18.1
Cr	y = 5.75 + 0.34x	0.8	0.205	0.695	4.2
Cu	y = 4.58 + 0.18x	0.157	0.502	0.309	25.3
<i>L. parsia</i>					
As	y = -0.96 + 0.73x	0.16	0.914	<b>0.01</b>	83.7
Pb	y = 7.79 + 0.84x	0.711	0.51	0.301	26.0
Cd	y = 0.12 + 0.03x	0.036	0.378	0.459	14.3
Cr	y = 1.86 + 0.2x	0.139	0.58	0.227	33.7
Cu	y = 4.24 + 0.72x	0.346	0.719	0.107	51.7
<i>M. cephalus</i>					
As	y = 2.08+0.11x	0.09	0.5	0.311	25.0
Pb	y = 23.98–0.42x	0.309	-0.557	-0.557	31.0
Cd	y = 0.87–0.02x	0.016	-0.547	0.26	30.0
Cr	y = 4.04–0.03x	0.075	-0.217	0.68	4.7
Cu	y = 16.95–0.21x	0.257	-0.364	0.477	13.3
<i>H. limbatus</i>					
As	y = -4.63+0.89x	0.309	0.821	<b>0.045</b>	67.5
Pb	y = 1.09+1.156x	0.778	0.596	0.211	35.6
Cd	y = 0.36–0.001x	0.048	-0.014	0.978	0.01
Cr	y = -0.6+0.38x	0.238	0.620	0.189	38.5
Cu	y = 2.54+0.91x	0.749	0.519	0.291	26.9
<i>T. toli</i>					
As	y = 0.48+ 0.16x	0.059	0.8	<b>0.05</b>	63.9
Pb	y = 3.74+0.32x	0.012	0.997	<b>1.11E-5</b>	99.5
Cd	y = 0.35–0.01x	0.013	-0.042	0.936	0.2
Cr	y = -1.22+ 0.14x	0.08	0.662	0.152	43.8
Cu	y = 0.37 + 0.34x	0.237	0.581	0.227	33.7

<https://doi.org/10.1371/journal.pone.0219336.t002>

other hand, *A. bato* showed maximum bioaccumulation result. Moreover, the mean BAFs of the metals were observed in the species as follows: Cu (1971.42) > As (1042.93) > Pb (913.66) > Cr (864.99) > Cd (252.03).

## Health risk evaluation

### Estimated daily intake (EDI)

The explored EDI of two concerned age groups, adults and children, is presented and summarized in Table 5. Adults and children showed comparably higher EDIs for demersal species than pelagic ones. In the consequence, high doses of demersal species were exposed to the

Table 3. Pearson’s correlation coefficient between heavy metal elements and matrix of PCA loadings.

Correlation analysis						Principal component analysis	
	As	Pb	Cd	Cr	Cu	PC 1	PC 2
As	1					0.432	-0.637
Pb	<b>0.912*</b>	1				0.465	-0.106
Cd	0.640	0.828	1			0.421	0.718
Cr	<b>0.904*</b>	<b>0.917*</b>	0.820	1		0.455	-0.160
Cu	0.840	<b>0.967*</b>	<b>0.911*</b>	0.890	1	0.463	0.205
Eigenvalues						4.459	0.387
% of Variance						89.18	7.75
Cumulative %						89.18	96.93

\* Significant at  $p \leq 0.05$

<https://doi.org/10.1371/journal.pone.0219336.t003>

consumers through consuming metal affected fish species as the food items. The EDIs for both groups were organized in the following order: Pb > Cu > As > Cr > Cd.

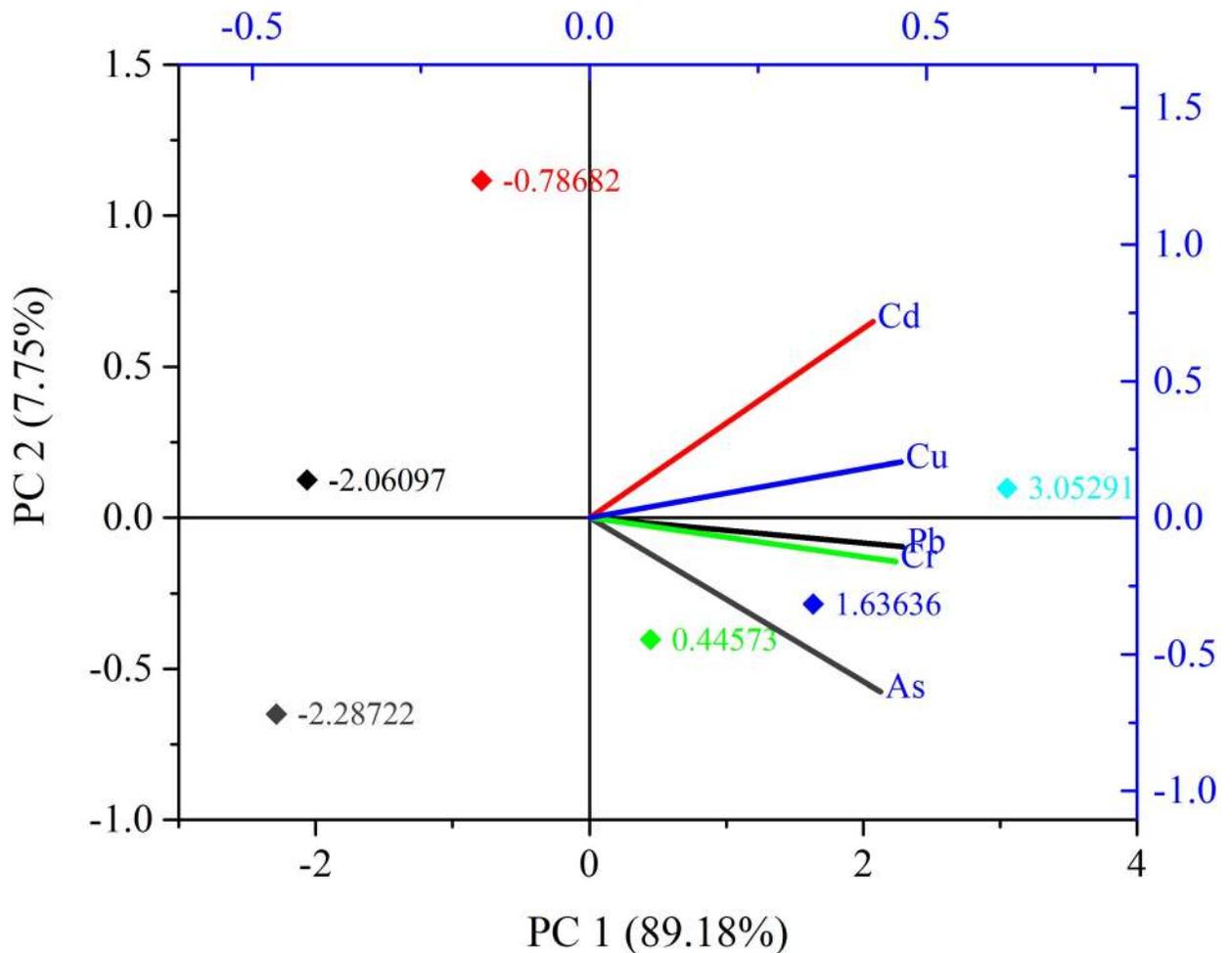


Fig 2. Loading plots of principal component analysis (PCA) of elements.

<https://doi.org/10.1371/journal.pone.0219336.g002>

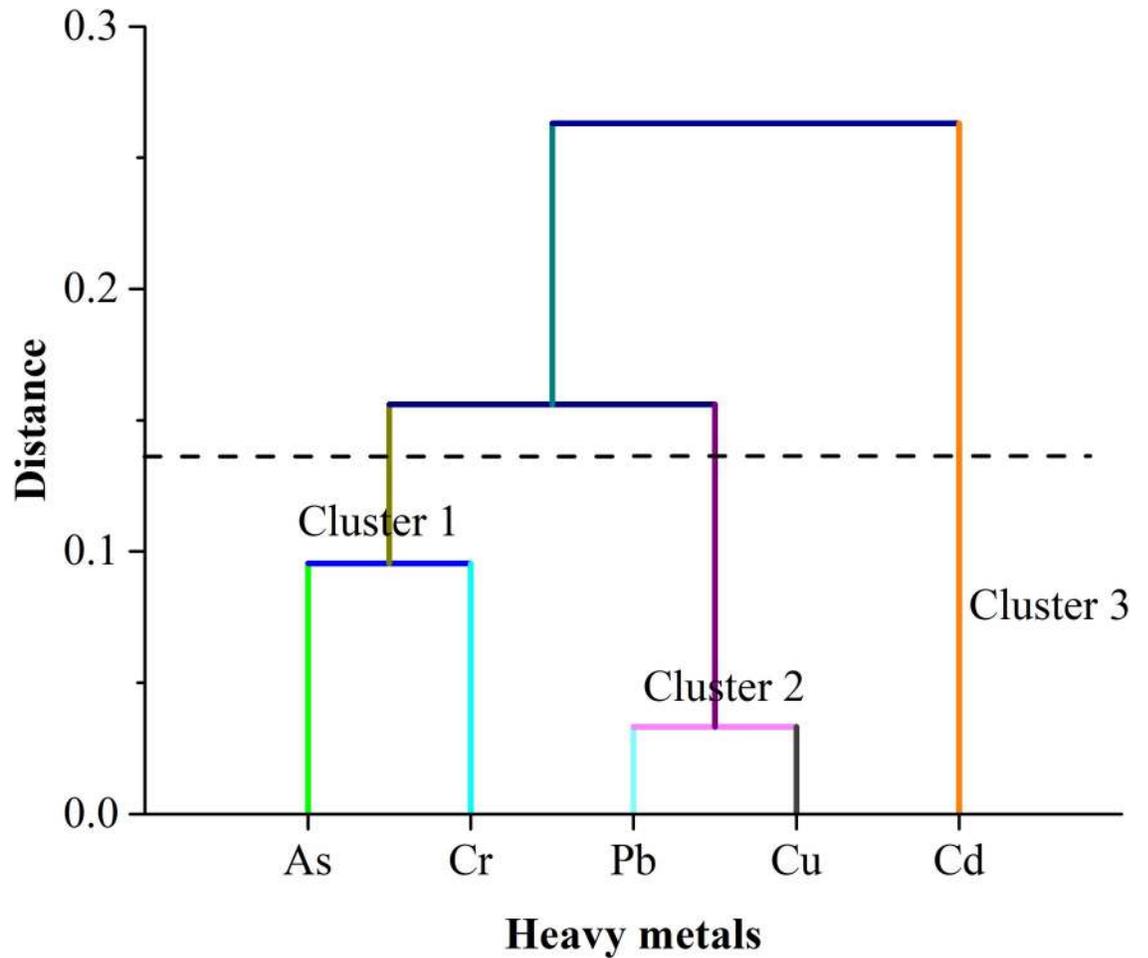


Fig 3. Hierarchical cluster (dendrogram) using Ward linkage method among the experimented metals in fish species.

<https://doi.org/10.1371/journal.pone.0219336.g003>

### Target hazard quotient (THQ) and Hazard Index (HI) for non-carcinogenic risk

The assessed target hazard quotient (THQ) for the studied fish species are displayed in Table 6. THQs in the adult group induced to As, Pb, Cd, Cr, and Cu were 0.016, 0.003, 3.0E-04, 0.001 and 2.38E-04, respectively, whereas for children the values were 0.097, 0.014, 0.001, 0.004 and 0.001, respectively. Moreover, the rank of the THQs of the elements was as follows: As > Pb > Cr > Cd > Cu. While, for the cumulative scenario of HI, children were 5.83 times more susceptible than adults. However, the investigated HI was not surpass the recommended limit (Table 6).

Table 4. Length-weight relationship, growth pattern and Fulton condition factor for the targeted fishes.

Species	a (intercept) ± SE	b (slope regression) ± SE	Group (Growth pattern)	W = aL <sup>b</sup>	Fulton's Q ± std	Fish Condition
<i>A. bato</i>	19.62 ± 16.63	3.99 ± 1.24	Positive allometric	W = 19.62 × L <sup>3.99</sup>	3.22 ± 0.78	Good
<i>P. chinensis</i>	7.23 ± 25.94	22.88 ± 3.7	Positive allometric	W = 7.23 × L <sup>22.88</sup>	51.77 ± 15.26	Good
<i>L. parsia</i>	7.13 ± 3.8	2.76 ± 0.51	Isometric	W = 7.13 × L <sup>2.46</sup>	7.10 ± 3.44	Good
<i>M. cephalus</i>	0.002 ± 0.0008	10.85 ± 4.98	Positive allometric	W = 0.002 × L <sup>10.85</sup>	3.56 ± 0.45	Good
<i>H. limbatus</i>	0.01 ± 0.002	2.43 ± 0.41	Negative	W = 0.01 × L <sup>2.43</sup>	2.03 ± 0.23	Good
<i>T. toli</i>	164.30 ± 60.97	15.72 ± 1.98	Positive allometric	W = 164 × L <sup>15.72</sup>	2.29 ± 0.46	Good

<https://doi.org/10.1371/journal.pone.0219336.t004>

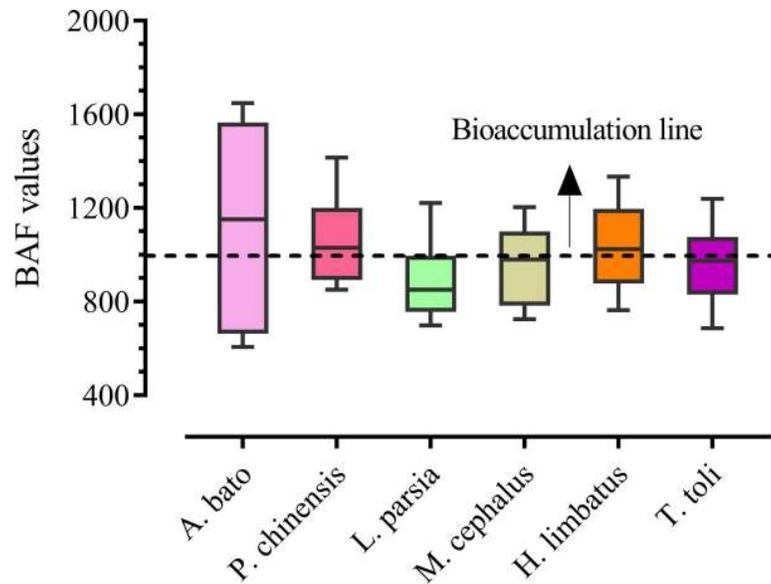


Fig 4. Bioaccumulation factor among the species that varied from particular metals and species.

<https://doi.org/10.1371/journal.pone.0219336.g004>

### Carcinogenic risk (CR)

Exposure of CR was estimated for a particular element and summarized in Table 6. The measured CR values of As, Pb and Cd were ranged from 5.14E-06- 1.35E-05, 8.48E-08- 1.79E-05 and 1.52E-06- 2.99E-06 respectively in adults and 2.29E-05- 1.35E-04, 3.78E-07- 7.99E-05 and 6.76E-06- 1.33E-05 in children. The results showed that children were exposed to higher CRs than adults. But, calculated CR values for both age groups were noted far from the risk as acceptable range is 10<sup>-6</sup> to 10<sup>-4</sup>.

### Discussion

#### Concentration of heavy metals and source identification

In the present study, *T. toli* had the highest concentration of As, whereas *A. bato* exhibited the maximum concentration for Pb, Cd and Cu, and *P. chinensis* had the highest concentration of Cr. The metal concentrations in the edible tissues were ranked in the following sequence: *A. bato* > *P. chinensis* > *H. limbatus* > *T. toli* > *M. cephalus* > *L. parsia*. Data from the previous literatures showed that metal concentrations in fish muscles varied widely depending on the location and species. The As and Pb concentrations in this study were higher than all other findings and recommended guidelines. Although Cd concentration was at the lower level comparing Northeast coast, Ganga River, and Pearl River, it surpassed all the guideline values

Table 5. A comparison between recommended daily allowance (RDA) and estimated daily intake (EDI) for adults and children.

Elements	Mean concentration (mg/kg)	RDA (mg/kg/person) [93]	EDIs (mg/day/person)	
			Adult	Child
As	4.89	0.15	0.005	0.029
Pb	13.88	0.25	0.011	0.049
Cd	0.39	0.07	0.001	0.001
Cr	3.36	0.23	0.003	0.012
Cu	12.10	35	0.010	0.042

<https://doi.org/10.1371/journal.pone.0219336.t005>

Table 6. Calculated THQ, HI and CR for the selected two aged groups.

Species	THQ (As)		THQ (Pb)		THQ (Cd)		THQ (Cr)		THQ (Cu)		HI		CR (As)		CR (Pb)		CR (Cd)	
	Ad	Ch	Ad	Ch	Ad	Ch	Ad	Ch	Ad	Ch								
<i>A. bato</i>	0.012	0.054	0.003	0.015	0.000	0.002	0.001	0.004	0.000	0.001	0.017	0.077	5.47E-06	2.44E-05	1.79E-05	8.00E-05	2.99E-06	1.30E-05
<i>P. chinensis</i>	0.013	0.059	0.003	0.014	0.000	0.002	0.001	0.004	0.000	0.001	0.018	0.08	5.93E-06	2.64E-05	9.35E-08	7.40E-05	2.18E-06	9.70E-06
<i>L. parsia</i>	0.011	0.051	0.003	0.014	0.000	0.001	0.001	0.004	0.000	0.001	0.016	0.071	5.14E-06	2.29E-05	9.33E-08	4.20E-07	1.68E-06	7.50E-06
<i>M. cephalus</i>	0.013	0.057	0.003	0.013	0.000	0.001	0.001	0.004	0.000	0.001	0.017	0.076	5.77E-06	2.57E-05	8.48E-08	3.80E-07	1.52E-06	6.80E-06
<i>H. limbatus</i>	0.013	0.06	0.003	0.014	0.000	0.001	0.001	0.004	0.000	0.001	0.018	0.08	6.06E-06	2.7E-05	9.2E-08	4.10E-07	1.72E-06	7.70E-06
<i>T. toli</i>	0.030	0.3	0.003	0.014	0.000	0.001	0.001	0.004	0.000	0.001	0.034	0.319	1.35E-05	1.35E-04	9.09E-08	4.00E-07	1.54E-06	6.90E-06
Mean	0.016	0.097	0.003	0.014	0.000	0.001	0.001	0.004	0.000	0.001	0.015	0.079	6.98E-06	4.36E-05	3.07E-06	2.58E-05	1.94E-06	8.64E-06

<https://doi.org/10.1371/journal.pone.0219336.t006>

along with other coastal environments, Meiliang Bay, Iskenderun Bay, Arasalar River and coastal area of Bangladesh. The results reported by [89] from the Ganga River were generally lower than present results, except Cu. The concentrations of Cd and Cu in the fish collected from the Pearl River were higher than present study. Different fish species of Bangladesh's coastal area have been reported to contain 0.08–13 As, 0.07–0.63 Pb, 0.03–0.09 Cd, 0.15–2.2 Cr and 1.3–14 Cu mg/kg dry weight of muscles [86], which were generally lower than the present findings. In this study, the demersal species had comparatively higher concentration level of metals than pelagic ones as they inhabit close to the bottom or sediment. The MPI values of 3.65 to 4.70 in the present study were much lower than those of Blackchin tilapia, *Sarotherodon melanotheron* and Silver catfish, *Chrysichthys nigrodigitatus* (8.1 to 17.76) from Okrika Estuary, Nigeria. This is most likely due to the oil bunkering and transportation activities along the study sites [94]. The findings of MPI in the present study were almost similar to that of *Rutilus rutilus* in Pluszne Lake [12]. However, the metal accumulation in fishes could be highly influenced by sampling locations and habitats [95, 96].

Previous studies have shown that heavy metals in the aquatic environment could come from different natural and anthropogenic sources [1, 3, 39, 97], where many factors influence their concentrations e.g., the original levels of rocks and parent materials, processes of soil formation, contamination by human activities, and other anthropogenic factors [98]. Generally, high correlations between specific heavy metals in the environment may reflect similar levels of contamination and/or release from the same sources of pollution [40]. In this study, strong positive correlations were found between Cd and Pb, Cr and Pb, Cr and Cd, and Cr and Cu indicating that they had the same source either natural or anthropogenic. The employed PCA revealed that the source of origin of the metal was anthropogenic. Pb, Cd, and Cu were the dominant compounds in PCA analysis due to their high loading scores. PCA also revealed the changes of geochemical composition and the justification of cluster analysis [99]. In the cluster analysis, Pb, Cd and Cu are grouped together in cluster 2, indicating that the anthropogenic sources of these heavy metals are closely related to the sediments of the study area. There are several manufacturing industries near the study area which largely use alloy, paints and poisonous chemicals containing As. The commercial uses of heavy metals in modern microelectronic and optical industries are notable sources of As intrusion in the aquatic environment [100]. Non-essential element, Pb originated from extreme agriculture, poultry farms, industries and textile mills might end up in the aquatic ecosystem [101], thus contributing to the metal in the study area. Thus the benthic feeders are to be greatly affected by the deposited Pb in the ecosystem [91]. Cd is typically found at a low concentration in the aquatic environment, however, indiscriminate use of phosphate fertilizer and industries are two primary sources of Cd introduction [102]. In the study area, nickel-cadmium battery manufacturing along with industries engaged with Cd metal incineration and production might increase the

Cd concentration level in the aquatic environment [103, 104]. Besides Cd and Pb, Cr is also widely introduced in the textile industries [105]. Near the bank of the Karnaphuli estuary, such commercial textile industries produce colour pigment and thus become a common contaminant for the aquatic ecosystem [106]. Notably, a considerable level of Cu become swelled up in the study area due to oil droppings from ships and boats, recurrent usage of antifouling paints and other boating interferes [107].

### Bioaccumulation (BAF) status of target species

The bioaccumulation potential of metals was assessed in the muscles of various fish species, and was found to vary from species to species. The hierarchy suggested that most of the species were tend to be bioaccumulative (BAF value close to 1000). *A. bato* exhibited the highest bioaccumulation in the studied area. The accumulation of the metal elements in an aquatic organism depends upon the classification of species, invasion pathways, metabolic characters of the sampled tissues and finally, the surrounding environmental condition in which the species live [108]. In our results, the BAFs of As, Cd, Cr, Pb and Cu were relatively higher than those of Pearl River estuary [90], where the BAFs in *Tilapia* were reported to be Cd > Cu > Pb > Cr. Such reports were mostly in line with our results. The fact is that, Cu is actively persistent in muscles due to being an essential element of living tissue [60, 109]. Notably, the bioaccumulation capability of Cd takes a long time to spare, thus making it relatively infirm [20].

### Human health risk evaluation

EDI, based on the oral reference dose (RfD) for an individual element [110], reflects the daily exposure to the toxic element, and is executed to avoid any harmful effect on human health [63]. The records of EDI of the people were compared with recommended daily allowance (RDA), provided by WHO, and introduced that, mean EDI values of the metals were still lower than RDAs. The values lower than RDA guidelines suggested a lower possible health effect of the elements to the consumers. However, it would not be wise to take it as a permanent measurement to reach a final conclusion [61, 63].

The <1 THQ values for both adult and children suggested that the adverse effect on human health might not occur. Similarly, the HI results also followed the THQ trend. Hence, there is no potential non-carcinogenic effect for the consumers due to intake of the fish species. Studies carried out by several authors in similar conditions were in line with our results [32, 69, 111–113]. In general, the assessment of THQ for human health risk evaluation has no dose-response relation of the examined elements [114]. However, human can dramatically suffer in the long run due to multiple simultaneous pollutants [77].

The CR values lower than  $10^{-6}$  indicates a negligible health risk whereas values in the range of  $10^{-6}$ – $10^{-4}$  are in the acceptable belt [63]. The CR values found in this study suggested an acceptable limit, and consumers are therefore less prone to carcinogenic risk. In fact, 90% of the carcinogenic risk is observed in the As contaminated aquatic food items. The inorganic state of As is more lethal than organic one [20, 115], and only 10% of the total As can be assessed as inorganic form [63]. The findings of the present study were in the acceptable range ( $10^{-6}$  to  $10^{-4}$ ) compared with Okogwu et al. [116], except for Cr which surpassed the CR limits. Again, in the Persian Gulf, consumers at the threshold limit for As were in concern of carcinogenic risk [117]. For this reason, carcinogenic risk should be given more attention due to intake of aquatic products, especially for the study area.

## Conclusions

Five heavy metals in the muscles of six fish species from the Karnaphuli River estuary were measured to investigate their potential sources, bioaccumulation rate and human health risk. Relatively high concentration of Cu was observed in *A. bato*. In most cases, the metal concentrations exceeded the recommended limits. The maximum metal accumulation was recorded in *A. bato*. *A. bato*, *P. chinensis* and *H. limbatus* were observed to be extreme bio-accumulative species. The mean bioaccumulation factors of the metals were observed in the studied species as follows: Cu > As > Pb > C > Cd.). Among the fishes studied, demersal fishes had higher levels of heavy metals compared with those fishes in the upper layers, suggesting higher level of heavy metals in the bottom water or sediment compared with the surface water. Accumulation of heavy metals in fish could be resulted from surface contact with the water, by breathing, and via the food chain. Heavy metals in the sediment enter the food chain through the feeding of benthic animals. Pearson correlation analysis showed strong positive correlations between Cd and Pb, Cr and Pb, Cr and Cd, and Cr and Cu indicating that they had the same source, either natural or anthropogenic. As per cluster analysis, Pb, Cd, and Cu, were grouped together, indicating the local anthropogenic sources. Existing textile mills, fertilizer factory, leather industry and agricultural activities in the catchment area of the river might be the sources of high concentration of Pb and Cd. Carcinogenic risk assessment suggested local consumers were free from the risk of cancer for the time being but they might be affected in future upon consumption of fish from studied region. Finally, it is found that children were six times more vulnerable to non-carcinogenic and carcinogenic risks than adults. Nonetheless, further study required ensuring the same conclusions are reached.

## Acknowledgments

The authors gratefully acknowledge the Rapid Action Battalion Headquarter, Bangladesh for providing necessary fund for heavy metal analyses and instrumental facilities. Comments from the editor and reviewers have significantly improved the manuscript.

## Author Contributions

**Conceptualization:** Ahasan Habib, M. Belal Hossain.

**Formal analysis:** A. S. Shafiuddin Ahmed, Md. Mahfujur Rahman.

**Funding acquisition:** Ahasan Habib, Najiah Musa.

**Investigation:** A. S. Shafiuddin Ahmed, Sharmin Sultana, Md. Shafiqul Islam Sarker.

**Methodology:** A. S. Shafiuddin Ahmed, Sharmin Sultana.

**Resources:** Hadayet Ullah, M. Belal Hossain, Md. Shafiqul Islam Sarker.

**Supervision:** Ahasan Habib.

**Writing – original draft:** A. S. Shafiuddin Ahmed, Sharmin Sultana.

**Writing – review & editing:** Ahasan Habib, Hadayet Ullah, Najiah Musa, M. Belal Hossain.

## References

1. Islam MS, Hossain MB, Matin A, Sarker MSI. Assessment of heavy metal pollution, distribution and source apportionment in the sediment from Feni River estuary, Bangladesh. *Chemosphere*. 2018; 202:25–32. <https://doi.org/10.1016/j.chemosphere.2018.03.077> <https://doi.org/10.1016/j.chemosphere.2018.03.077> PMID: 29554504

2. Wilcock D. River and inland water environments. In: Nath B, Hens L, Compton P, Devuyt D (Eds), environmental management in practice (volume 3), Routledge, New York. 1999. p.328
3. Hossain MB, Ahmed ASS, Sarker MSI. Human health risks of Hg, As, Mn, and Cr through consumption of fish, Ticto barb (*Puntius ticto*) from a tropical river, Bangladesh. *Environmental Science and Pollution Research*. 2018; 25:31727–31736. <https://doi.org/10.1007/s11356-018-3158-9> <https://doi.org/10.1007/s11356-018-3158-9> PMID: 30209769
4. Xu S, Tao S. Coregionalization analysis of heavy metals in the surface soil of Inner Mongolia. *Science of the total environment*. 2004; 320:73–87. [https://doi.org/10.1016/S0048-9697\(03\)00450-9](https://doi.org/10.1016/S0048-9697(03)00450-9) PMID: 14987928
5. Idriss A, Ahmad A. Heavy metal concentrations in fishes from Juru River, estimation of the health risk. *Bulletin of Environmental Contamination and Toxicology*. 2015; 94:204–208. <https://doi.org/10.1007/s00128-014-1452-x> <https://doi.org/10.1007/s00128-014-1452-x> PMID: 25564001
6. Authman MM, Zaki MS, Khallaf EA, Abbas HH. Use of fish as bio-indicator of the effects of heavy metals pollution. *Journal of Aquaculture Research & Development*. 2015; 6:1–13. <http://dx.doi.org/10.4172/2155-9546.1000328>
7. Ali H, Khan E. Bioaccumulation of non-essential hazardous heavy metals and metalloids in freshwater fish. Risk to human health. *Environmental Chemistry Letters*. 2018; 16:903–917. <https://doi.org/10.1007/s10311-018-0734-7>
8. Bayen S, Wurl O, Karupiah S, Sivasothi N, Lee HK, Obbard JP. Persistent organic pollutants in mangrove food webs in Singapore. *Chemosphere*. 2005; 61:303–313. <https://doi.org/10.1016/j.chemosphere.2005.02.097> <https://doi.org/10.1016/j.chemosphere.2005.02.097> PMID: 16182847
9. Zeitoun MM, Mehana E. Impact of water pollution with heavy metals on fish health: overview and updates. *Global Veterinaria*. 2014; 12:219–231. DOI: <https://doi.org/10.5829/idosi.gv.2014.12.02.82219>
10. Kelly BC, Ikononou MG, Blair JD, Morin AE, Gobas FA. Food web-specific biomagnification of persistent organic pollutants. *Science*. 2007; 317:236–239. DOI: <https://doi.org/10.1126/science.1138275> PMID: 17626882
11. Siddique M, Arshad A, Amin S. Length-weight and length-length relationships of two tropical fish *Secutor megalolepis* (Mochizuki & Hayashi, 1989) and *Rhabdamia gracilis* (Bleeker, 1856) from Sabah, Malaysia. *Journal of Applied Ichthyology*. 2015; 31:574–575. <https://doi.org/10.1111/jai.12752>
12. Luczynska J, Paszczyk B, Luczynski MJ. Fish as a bioindicator of heavy metals pollution in aquatic ecosystem of Pluszne Lake, Poland, and risk assessment for consumer's health. *Ecotoxicology and Environmental Safety*. 2018; 153:60–67. <https://doi.org/10.1016/j.ecoenv.2018.01.057> <https://doi.org/10.1016/j.ecoenv.2018.01.057> PMID: 29407739
13. Zamri Z, Arshad A, Amin SN, Rahman MA, Al Khayat JA. Sex ratio, gonad development and fecundity of *Miyakella nepa* (Crustacea, Stomatopoda) of Pantai Remis coastal waters of Malaysia. *Journal of Environmental Biology*. 2016; 37:677–683 PMID: 28779726
14. Parente T, Hauser-Davis R. The use of fish biomarkers in the evaluation of water pollution. *Pollution and Fish Health in Tropical Ecosystems*. 2013:164–181. <https://doi.org/10.1201/b16298-8>
15. Azaman F, Juahir H, Yunus K, Azid A, Kamarudin MKA, Toriman ME, et al. Heavy metal in fish: Analysis and human health—a review. *Jurnal Teknologi*. 2015; 77:61–69. <https://doi.org/10.11113/jt.v77.4182>
16. Castro-González M, Méndez-Armenta M. Heavy metals: Implications associated to fish consumption. *Environmental Toxicology and Pharmacology*. 2008; 26:263–271. <https://doi.org/10.1016/j.etap.2008.06.001> <https://doi.org/10.1016/j.etap.2008.06.001> PMID: 21791373
17. Rahman MS, Hossain MB, Babu SOF, Rahman M, Ahmed AS, Jolly Y, et al. Source of metal contamination in sediment, their ecological risk, and phytoremediation ability of the studied mangrove plants in ship breaking area, Bangladesh. *Marine Pollution Bulletin*. 2019; 141:137–146. <https://doi.org/10.1016/j.marpolbul.2019.02.032> <https://doi.org/10.1016/j.marpolbul.2019.02.032> PMID: 30955718
18. Rahman MA, Mustafa MG, Barman BK. Impacts of aquaculture extension activities on female fish farmers in different areas of Bangladesh. *Bangladesh Journal of Zoology*. 2012; 39:213–221. <https://doi.org/10.3329/bjz.v39i2.10591>
19. Saha N, Mollah M, Alam M, Rahman MS. Seasonal investigation of heavy metals in marine fishes captured from the Bay of Bengal and the implications for human health risk assessment. *Food Control*. 2016a; 70:110–118. <https://doi.org/10.1016/j.foodcont.2016.05.040>
20. Zhong W, Zhang Y, Wu Z, Yang R, Chen X, Yang J, et al. Health risk assessment of heavy metals in freshwater fish in the central and eastern North China. *Ecotoxicology and Environmental Safety*. 2018; 157:343–349. <https://doi.org/10.1016/j.ecoenv.2018.03.048> <https://doi.org/10.1016/j.ecoenv.2018.03.048> PMID: 29627419

21. Ahmed MK, Baki MA, Islam MS, Kundu GK, Habibullah-Al-Mamun M, Sarkar SK, et al. Human health risk assessment of heavy metals in tropical fish and shellfish collected from the river Buriganga, Bangladesh. *Environmental Science and Pollution Research*. 2015; 22:15880–15890. <https://doi.org/10.1007/s11356-015-4813-z> <https://doi.org/10.1007/s11356-015-4813-z> PMID: 26044144
22. Varol M, Kaya GK, Alp A. Heavy metal and arsenic concentrations in rainbow trout (*Oncorhynchus mykiss*) farmed in a dam reservoir on the Firat (Euphrates) River: Risk-based consumption advisories. *Science of the Total Environment*. 2017; 599:1288–1296. <https://doi.org/10.1016/j.scitotenv.2017.05.052> <https://doi.org/10.1016/j.scitotenv.2017.05.052> PMID: 28525936
23. Dey S, Das J, Manchur M. Studies on heavy metal pollution of Karnaphuli river, Chittagong, Bangladesh. *IOSR Journal of Environmental Science, Toxicology and Food Technology*. 2015; 9:79–83. DOI: <https://doi.org/10.9790/2402-09817983>
24. Islam MM, Rahman SL, Ahmed SU, Haque MKI. Biochemical characteristics and accumulation of heavy metals in fishes, water and sediments of the river Buriganga and Shitalakhya of Bangladesh. *Journal of Asian Scientific Research*. 2014; 4:270.
25. Ahmed MK, Baki MA, Kundu GK, Islam MS, Islam MM, Hossain MM. Human health risks from heavy metals in fish of Buriganga river, Bangladesh. *SpringerPlus*. 2016; 5:1697. <https://doi.org/10.1186/s40064-016-3357-0> <https://doi.org/10.1186/s40064-016-3357-0> PMID: 27757369
26. Rahman MS, Molla AH, Saha N, Rahman A. Study on heavy metals levels and its risk assessment in some edible fishes from Bangshi River, Savar, Dhaka, Bangladesh. *Food Chemistry*. 2012; 134:1847–1854. <https://doi.org/10.1016/j.foodchem.2012.03.099> <https://doi.org/10.1016/j.foodchem.2012.03.099> PMID: 23442629
27. Amin MN, Begum A, Mondal MK. Trace element concentrations present in five species of freshwater fish of Bangladesh. *Bangladesh Journal of Scientific and Industrial Research*. 2011; 46:27–32. <https://doi.org/10.3329/bjsir.v46i1.8101>
28. Xie W, Chen K, Zhu X, Nie X, Zheng G, Pan D, et al. Evaluation of heavy metal contents in water and fishes collected from the waterway in Pearl River Delta in south China. *Journal of Agro-Environment Science*. 2010; 29:1917–1923.
29. Ciftci N, Ay O, Karayakar F, Cicik B, Erdem C. Effects of zinc and cadmium on condition factor, hepatosomatic and gonadosomatic index of *Oreochromis niloticus*. *Fresenius Environmental Bulletin*. 2015; 24:3871–3874.
30. Hossein KP, Takhsha M, Aein JK, Aghshenas A. Assessment level of heavy metals (Pb, Cd, Hg) in four fish species of Persian Gulf (Bushehr-Iran). *International Journal of Advanced Technology & Engineering Research*. 2014; 4:7–11.
31. Usero J, González-Regalado E, Gracia I. Trace metals in the bivalve mollusc *Chamelea gallina* from the Atlantic coast of southern Spain. *Oceanographic Literature Review*. 1996; 10:1058. [https://doi.org/10.1016/0025-326x\(95\)00209-6](https://doi.org/10.1016/0025-326x(95)00209-6)
32. Abdel-Khalek AA, Elhaddad E, Mamdouh S, Marie M- AS. Assessment of metal pollution around sabal drainage in River Nile and its impacts on bioaccumulation level, metals correlation and human risk hazard using *Oreochromis niloticus* as a bioindicator. *Turkish Journal of Fisheries and Aquatic Sciences*. 2016; 16:227–239. [https://doi.org/10.4194/1303-2712-v16\\_2\\_02](https://doi.org/10.4194/1303-2712-v16_2_02)
33. Keshavarzi B, Hassanaghaei M, Moore F, Mehr MR, Soltanian S, Lahijanzadeh AR, et al. Heavy metal contamination and health risk assessment in three commercial fish species in the Persian Gulf. *Marine Pollution Bulletin*. 2018; 129:245–252. <https://doi.org/10.1016/j.marpolbul.2018.02.032> <https://doi.org/10.1016/j.marpolbul.2018.02.032> PMID: 29680544
34. Liu Q, Liao Y, Shou L. Concentration and potential health risk of heavy metals in seafoods collected from Sanmen Bay and its adjacent areas, China. *Marine Pollution Bulletin*. 2018; 131:356–364. <https://doi.org/10.1016/j.marpolbul.2018.04.041> <https://doi.org/10.1016/j.marpolbul.2018.04.041> PMID: 29886958
35. Monsefrad F, Imanpour Namin J, Heidary S. Concentration of heavy and toxic metals Cu, Zn, Cd, Pb and Hg in liver and muscles of *Rutilus frisii kutum* during spawning season with respect to growth parameters. *Iranian Journal of Fisheries Sciences*. 2012; 11:825–839.
36. Yi YJ, Zhang SH. Heavy metal (Cd, Cr, Cu, Hg, Pb, Zn) concentrations in seven fish species in relation to fish size and location along the Yangtze River. *Environmental Science and Pollution Research*. 2012; 19:3989–3996. <https://doi.org/10.1007/s11356-012-0840-1> <https://doi.org/10.1007/s11356-012-0840-1> PMID: 22382698
37. Tekin-Özan S, Aktan N. Relationship of heavy metals in water, sediment and tissues with total length, weight and seasons of *Cyprinus carpio* L., 1758 from Işikli Lake (Turkey). *Pakistan Journal of Zoology*. 2012; 44:1405–1416.
38. Merciai R, Guasch H, Kumar A, Sabater S, García-Berthou E. Trace metal concentration and fish size: Variation among fish species in a Mediterranean river. *Ecotoxicology and Environmental Safety*. 2014;

- 107:154–161. <https://doi.org/10.1016/j.ecoenv.2014.05.006> <https://doi.org/10.1016/j.ecoenv.2014.05.006> PMID: 24946163
39. Varol M, Şen B. Assessment of nutrient and heavy metal contamination in surface water and sediments of the upper Tigris River, Turkey. *Catena*. 2012; 92:1–10. <https://doi.org/10.1016/j.catena.2011.11.011>
  40. Li J, He M, Han W, Gu Y. Analysis and assessment on heavy metal sources in the coastal soils developed from alluvial deposits using multivariate statistical methods. *Journal of Hazardous Materials*. 2009; 164:976–981. <https://doi.org/10.1016/j.jhazmat.2008.08.112> <https://doi.org/10.1016/j.jhazmat.2008.08.112> PMID: 18976857
  41. Yang Z, Wang Y, Shen Z, Niu J, Tang Z. Distribution and speciation of heavy metals in sediments from the mainstream, tributaries, and lakes of the Yangtze River catchment of Wuhan, China. *Journal of Hazardous Materials*. 2009; 166:1186–1194. <https://doi.org/10.1016/j.jhazmat.2008.12.034> <https://doi.org/10.1016/j.jhazmat.2008.12.034> PMID: 19179000
  42. Sundaray SK, Nayak BB, Lin S, Bhatta D. Geochemical speciation and risk assessment of heavy metals in the river estuarine sediments—a case study: Mahanadi basin, India. *Journal of Hazardous Materials*. 2011; 186:1837–1846. <https://doi.org/10.1016/j.jhazmat.2010.12.081> <https://doi.org/10.1016/j.jhazmat.2010.12.081> PMID: 21247687
  43. Chung C-Y, Chen J-J, Lee C-G, Chiu C-Y, Lai W-L, Liao S-W. Integrated estuary management for diffused sediment pollution in Dapeng Bay and neighboring rivers (Taiwan). *Environmental Monitoring and Assessment*. 2011; 173:499–517. <https://doi.org/10.1007/s10661-010-1401-z> <https://doi.org/10.1007/s10661-010-1401-z> PMID: 20195746
  44. Wang Q, Chen M, Shan G, Chen P, Cui S, Yi S, et al. Bioaccumulation and biomagnification of emerging bisphenol analogues in aquatic organisms from Taihu Lake, China. *Science of The Total Environment*. 2017; 598:814–820. <https://doi.org/10.1016/j.scitotenv.2017.04.167> <https://doi.org/10.1016/j.scitotenv.2017.04.167> PMID: 28458198
  45. Qin D, Jiang H, Bai S, Tang S, Mou Z. Determination of 28 trace elements in three farmed cyprinid fish species from Northeast China. *Food Control*. 2015; 50:1–8. <https://doi.org/10.1016/j.foodcont.2014.08.016>
  46. Zhang L, Shi Z, Jiang Z, Zhang J, Wang F, Huang X. Distribution and bioaccumulation of heavy metals in marine organisms in east and west Guangdong coastal regions, South China. *Marine Pollution Bulletin*. 2015; 101:930–937. <https://doi.org/10.1016/j.marpolbul.2015.10.041> <https://doi.org/10.1016/j.marpolbul.2015.10.041> PMID: 26506025
  47. Arnot JA, Gobas FA. A review of bioconcentration factor (BCF) and bioaccumulation factor (BAF) assessments for organic chemicals in aquatic organisms. *Environmental Reviews*. 2006; 14:257–297. <https://doi.org/10.1139/a06-005>
  48. Sekitar PKA, Hamid M, Mansor M, Nor SAM. Length-weight relationship and condition factor of fish populations in Temengor reservoir: Indication of environmental health. *Sains Malaysiana*. 2015; 44:61–66.
  49. Luczynska J, Luczynski MJ, Paszczyk B, Tonska E. Concentration of mercury in muscles of predatory and non-predatory fish from lake Pluszne (Poland). *Journal of Veterinary Research*. 2016; 60:43–47. DOI: <https://doi.org/10.1515/jvetres-2016-0007>
  50. Froese R. Cube law, condition factor and weight–length relationships: history, meta-analysis and recommendations. *Journal of Applied Ichthyology*. 2006; 22:241–453. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
  51. Jin S, Yan X, Zhang H, Fan W. Weight–length relationships and Fulton’s condition factors of skipjack tuna (*Katsuwonus pelamis*) in the western and central Pacific Ocean. *PeerJ*. 2015; 3:e758. <https://doi.org/10.7717/peerj.758> <https://doi.org/10.7717/peerj.758> PMID: 25699207
  52. C B A B. Condition Factor, K, for Salmonid Fish, *Fisheries Notes*. 1998.
  53. Datta SN, Kaur VI, Dhawan A, Jassal G. Estimation of length-weight relationship and condition factor of spotted snakehead *Channa punctata* (Bloch) under different feeding regimes. *SpringerPlus*. 2013; 2(1):436.
  54. Ricker W. Linear regressions in fishery research. *Journal of the fisheries board of Canada*. 1973; 30(3):409–34.
  55. Pervin MR, Mortuza MG. Notes on length-weight relationship and condition factor of fresh water fish, *Labeo boga* (Hamilton)(Cypriniformes: Cyprinidae). *University Journal of Zoology, Rajshahi University*. 2008; 27:97–98
  56. Gupta B, Sarkar U, Bhardwaj S, Pal A. Condition factor, length–weight and length–length relationships of an endangered fish *Ompok pabda* (Hamilton 1822) (Siluriformes: Siluridae) from the River Gomti, a tributary of the River Ganga, India. *Journal of Applied Ichthyology*. 2011; 27:962–964. doi: <https://doi.org/10.1111/j.1439-0426.2010.01625.x>

57. USEPA. Guideline for Assessing Chemical Contaminant Data for Use in Fish Advisories, Vol. I: Fish Sampling and Analysis. Third Edition. Office of Water. U.S. Environmental Protection Agency. Washington, DC: Document No. EPA 823-B-00-007. November 2000.; 2000a.
58. Wei Y, Zhang J, Zhang D, Tu T, Luo L. Metal concentrations in various fish organs of different fish species from Poyang Lake, China. *Ecotoxicology and Environmental Safety*. 2014; 104:182–188. <https://doi.org/10.1016/j.ecoenv.2014.03.001> <https://doi.org/10.1016/j.ecoenv.2014.03.001> PMID: 24681447
59. USEPA. Integrated risk information system, United States Environmental Protection Agency, Washington, DC, USA. <https://www.epa.gov/iris>. 2008. Accessed on 25 September 2018.
60. Traina A, Bono G, Bonsignore M, Falco F, Giuga M, Quinci EM, et al. Heavy metals concentrations in some commercially key species from Sicilian coasts (Mediterranean Sea): Potential human health risk estimation. *Ecotoxicology and Environmental Safety*. 2019; 168:466–478. <https://doi.org/10.1016/j.ecoenv.2018.10.056> <https://doi.org/10.1016/j.ecoenv.2018.10.056> PMID: 30419523
61. Vu CT, Lin C, Yeh G, Villanueva MC. Bioaccumulation and potential sources of heavy metal contamination in fish species in Taiwan: assessment and possible human health implications. *Environmental Science and Pollution Research*. 2017; 24:19422–19434. <https://doi.org/10.1007/s11356-017-9590-4> <https://doi.org/10.1007/s11356-017-9590-4> PMID: 28677040
62. Abtahi M, Fakhri Y, Oliveri Conti G, Keramati H, Zandsalimi Y, Bahmani Z, et al. Heavy metals (As, Cr, Pb, Cd and Ni) concentrations in rice (*Oryza sativa*) from Iran and associated risk assessment: a systematic review. *Toxin Reviews*. 2017; 36:331–341. <https://doi.org/10.1080/15569543.2017.1354307>
63. Baki MA, Hossain MM, Akter J, Quraishi SB, Shojib MFH, Ullah AA, et al. Concentration of heavy metals in seafood (fishes, shrimp, lobster and crabs) and human health assessment in Saint Martin Island, Bangladesh. *Ecotoxicology and Environmental Safety*. 2018; 159:153–163. <https://doi.org/10.1016/j.ecoenv.2018.04.035> <https://doi.org/10.1016/j.ecoenv.2018.04.035> PMID: 29747150
64. Rahmani J, Fakhri Y, Shahsavani A, Bahmani Z, Urbina MA, Chirumbolo S, et al. A systematic review and meta-analysis of metal concentrations in canned tuna fish in Iran and human health risk assessment. *Food and Chemical Toxicology*. 2018; 118:753–765. <https://doi.org/10.1016/j.fct.2018.06.023> <https://doi.org/10.1016/j.fct.2018.06.023> PMID: 29913231
65. Heshmati A, Ghadimi S, Khaneghah AM, Barba FJ, Lorenzo JM, Nazemi F, et al. Risk assessment of benzene in food samples of Iran's market. *Food and Chemical Toxicology*. 2018; 114:278–284. <https://doi.org/10.1016/j.fct.2018.02.043> <https://doi.org/10.1016/j.fct.2018.02.043> PMID: 29471007
66. Yi Y, Yang Z, Zhang S. Ecological risk assessment of heavy metals in sediment and human health risk assessment of heavy metals in fishes in the middle and lower reaches of the Yangtze River basin. *Environmental Pollution*. 2011; 159:2575–2585. <https://doi.org/10.1016/j.envpol.2011.06.011> <https://doi.org/10.1016/j.envpol.2011.06.011> PMID: 21752504
67. Xiao C-C, Chen M-J, Mei F-B, Fang X, Huang T-R, Li J-L, et al. Influencing factors and health risk assessment of microcystins in the Yongjiang river (China) by Monte Carlo simulation. *PeerJ*. 2018; 6: e5955. <https://doi.org/10.7717/peerj.5955> PMID: 30479903
68. USEPA. Region III Risk-Based Concentration Table: Technical background information. pp 227. 2006.
69. Dadar M, Adel M, Nasrollahzadeh Saravi H, Fakhri Y. Trace element concentration and its risk assessment in common kilka (*Clupeonella cultriventris caspia* Bordin, 1904) from southern basin of Caspian Sea. *Toxin Reviews*. 2017; 36:222–227. <https://doi.org/10.1080/15569543.2016.1274762>
70. Lei M, Tie B-q, Song Z-g, Liao B-H, Lepo JE, Huang Y-z. Heavy metal pollution and potential health risk assessment of white rice around mine areas in Hunan Province, China. *Food Security*. 2015; 7:45–54. <https://doi.org/10.1007/s12571-014-0414-9>
71. Fakhri Y, Mohseni-Bandpei A, Oliveri Conti G, Keramati H, Zandsalimi Y, Amanidaz N, et al. Health risk assessment induced by chloroform content of the drinking water in Iran: systematic review. *Toxin Reviews*. 2017; 36:342–351. <https://doi.org/10.1080/15569543.2017.1370601>
72. Fantke P, Friedrich R, Jolliet O. Health impact and damage cost assessment of pesticides in Europe. *Environment International*. 2012; 49:9–17. <https://doi.org/10.1016/j.envint.2012.08.001> <https://doi.org/10.1016/j.envint.2012.08.001> PMID: 22940502
73. Pepper I, Gerba C, Brusseau M. *Environmental and pollution science (pollution science series)*. Academic Press. 2012:212–232.
74. USEPA. Risk-based concentration table. United States Environmental Protection Agency, Washington, DC. 2000b.
75. FAO. The State of the World Fisheries and Aquaculture. Opportunities and Challenges. FAO Fisheries and Aquaculture Dept. Rome, Italy, pp 243. 2014.

76. Yin S, Feng C, Li Y, Yin L, Shen Z. Heavy metal pollution in the surface water of the Yangtze Estuary: a 5-year follow-up study. *Chemosphere*. 2015; 138:718–725. <https://doi.org/10.1016/j.chemosphere.2015.07.060> <https://doi.org/10.1016/j.chemosphere.2015.07.060> PMID: 26256308
77. Li PH, Kong SF, Geng CM, Han B, Lu B, Sun RF, et al. Assessing the hazardous risks of vehicle inspection workers' exposure to particulate heavy metals in their work places. *Aerosol and Air Quality Research*. 2013; 13:255–265. doi: <https://doi.org/10.4209/aaqr.2012.04.0087>
78. Hu B, Jia X, Hu J, Xu D, Xia F, Li Y. Assessment of heavy metal pollution and health risks in the soil-plant-human system in the Yangtze river delta, China. *International Journal of Environmental Research and Public Health*. 2017; 14:1042. <https://doi.org/10.3390/ijerph14091042>
79. USEPA. Integrated Risk Information System (IRIS); United States Environmental Protection Agency: Washington, DC, USA. Available online: <http://www.epa.gov/ncea/iris/index.html> (Accessed on 15 September 2018). 2010.
80. Cao S, Duan X, Zhao X, Ma J, Dong T, Huang N, et al. Health risks from the exposure of children to As, Se, Pb and other heavy metals near the largest coking plant in China. *Science of The Total Environment*. 2014; 472:1001–1009. <https://doi.org/10.1016/j.scitotenv.2013.11.124> <https://doi.org/10.1016/j.scitotenv.2013.11.124> PMID: 24345860
81. Vieira C, Morais S, Ramos S, Delerue-Matos C, Oliveira M. Mercury, cadmium, lead and arsenic levels in three pelagic fish species from the Atlantic Ocean: intra-and inter-specific variability and human health risks for consumption. *Food and Chemical Toxicology*. 2011; 49:923–932. <https://doi.org/10.1016/j.fct.2010.12.016> <https://doi.org/10.1016/j.fct.2010.12.016> PMID: 21193008
82. Nauen CE. Compilation of legal limits for hazardous substances in fish and fishery products. FAO Fisheries Circular (FAO) no 764. 1983.
83. WHO. Guideline for drinking water quality, 2nd edition. Recommendation. World Health Organization General. 1:30–113. 1985.
84. EU. Commission Regulation as Regards Heavy metals, Directive, 2001/22/EC, No: 466. 2001.
85. MOFL. Bangladesh Gazette, Bangladesh Ministry of Fisheries and Livestock, SRO no. 233/Ayen. 2014.
86. Raknuzzaman M, Ahmed MK, Islam MS, Habibullah-Al-Mamun M, Tokumura M, Sekine M, et al. Trace metal contamination in commercial fish and crustaceans collected from coastal area of Bangladesh and health risk assessment. *Environmental Science and Pollution Research*. 2016; 23:17298–17310. <https://doi.org/10.1007/s11356-016-6918-4> <https://doi.org/10.1007/s11356-016-6918-4> PMID: 27225006
87. Lakshmanasenthil S, Vinothkumar T, Ajithkumar TT, Marudhupandi T, Veetil DK, Ganeshamurthy R, et al. Harmful metals concentration in sediments and fishes of biologically important estuary, Bay of Bengal. *Journal of Environmental Health Science and Engineering*. 2013; 11:33. <https://doi.org/10.1186/2052-336X-11-33> PMID: 24355110
88. Kumar B, Sajwan K, Mukherjee D. Distribution of heavy metals in valuable coastal fishes from North East Coast of India. *Turkish Journal of Fisheries and Aquatic Sciences*. 2012; 12:81–88. DOI: [https://doi.org/10.4194/1303-2712-v12\\_1\\_10](https://doi.org/10.4194/1303-2712-v12_1_10)
89. Mitra A, Chowdhury R, Banerjee K. Concentrations of some heavy metals in commercially important finfish and shellfish of the River Ganga. *Environmental Monitoring and Assessment*. 2012; 184:2219–2230. <https://doi.org/10.1007/s10661-011-2111-x> <https://doi.org/10.1007/s10661-011-2111-x> PMID: 21660552
90. Kwok C, Liang Y, Wang H, Dong Y, Leung S, Wong MH. Bioaccumulation of heavy metals in fish and Ardeid at Pearl River Estuary, China. *Ecotoxicology and Environmental Safety*. 2014; 106:62–67. <https://doi.org/10.1016/j.ecoenv.2014.04.016> PMID: 24836879
91. Rajeshkumar S, Li X. Bioaccumulation of heavy metals in fish species from the Meiliang Bay, Taihu Lake, China. *Toxicology Reports*. 2018; 5:288–295. <https://doi.org/10.1016/j.toxrep.2018.01.007> <https://doi.org/10.1016/j.toxrep.2018.01.007> PMID: 29511642
92. Türkmen A, Türkmen M, Tepe Y, Akyurt I. Heavy metals in three commercially valuable fish species from Iskenderun Bay, Northern East Mediterranean Sea, Turkey. *Food Chemistry*. 2005; 91:167–172. <https://doi.org/10.1016/j.foodchem.2004.08.008>
93. WHO. World Health Organization technical report series. Evaluation of certain food additives and contaminants. Fifty-third report of the joint FAO/WHO expert committee on food additives (JECFA)', World Health Organization, Geneva, Switzerland. 2000. <http://www.Who.Int/foodsafety/publications/jecfa-reports/en/>. Accessed on 26 March 2019.
94. Ogunola OS, Onada OA, Falaye AE. Ecological Risk Evaluation of Biological and Geochemical Trace Metals in Okrika Estuary. *International Journal of Environmental Research*. 2017; 11:149–173. <https://doi.org/10.1007/s41742-017-0016-4>

95. Rejomon G, Nair M, Joseph T. Trace metal dynamics in fishes from the southwest coast of India. *Environmental Monitoring and Assessment*. 2010; 167:243–255. <https://doi.org/10.1007/s10661-009-1046-y> <https://doi.org/10.1007/s10661-009-1046-y> PMID: 19543989
96. Weber P, Behr ER, Knorr CDL, Vendruscolo DS, Flores EM, Dressler VL, et al. Metals in the water, sediment, and tissues of two fish species from different trophic levels in a subtropical Brazilian river. *Microchemical Journal*. 2013; 106:61–66. <https://doi.org/10.1016/j.microc.2012.05.004>
97. Bing H, Wu Y, Sun Z, Yao S. Historical trends of heavy metal contamination and their sources in lacustrine sediment from Xijiu Lake, Taihu Lake Catchment, China. *Journal of Environmental Sciences*. 2011; 23:1671–1678. [https://doi.org/10.1016/S1001-0742\(10\)60593-1](https://doi.org/10.1016/S1001-0742(10)60593-1)
98. Li S, Zhang Q. Geochemistry of the upper Han River basin, China, 1: spatial distribution of major ion compositions and their controlling factors. *Applied Geochemistry*. 2008; 23:3535–3544. <https://doi.org/10.1016/j.apgeochem.2008.08.012>
99. Liu H, Liu G, Yuan Z, Ge M, Wang S, Liu Y, et al. Occurrence, potential health risk of heavy metals in aquatic organisms from Laizhou Bay, China. *Marine Pollution Bulletin*. 2019; 140:388–394. <https://doi.org/10.1016/j.marpolbul.2019.01.067> <https://doi.org/10.1016/j.marpolbul.2019.01.067> PMID: 30803658
100. WHO. *Guideline for Drinking Water Quality, Recommendation vol. 1*, WHO, Geneva. p.130. 2002.
101. Rashed M. Monitoring of environmental heavy metals in fish from Nasser Lake. *Environment International*. 2001; 27:27–33. [https://doi.org/10.1016/S0160-4120\(01\)00050-2](https://doi.org/10.1016/S0160-4120(01)00050-2) PMID: 11488387
102. SS M. *Forms of cadmium in soils of Western Australia*. University of Western Australia. 1993.
103. Bustueva KA, Revich BA, Bezpalko LE. Cadmium in the environment of three Russian cities and in human hair and urine. *Archives of Environmental Health: An International Journal*. 1994; 49:284–288. <https://doi.org/10.1080/00039896.1994.9937481>
104. Bennet-Chambers M, Davies P, Knott B. Cadmium in aquatic ecosystems in Western Australia: A legacy of nutrient-deficient soils. *Journal of Environmental Management*. 1999; 57:283–295. <https://doi.org/10.1006/jema.1999.0304>
105. Sanyal T, Kaviraj A, Saha S. Deposition of chromium in aquatic ecosystem from effluents of handloom textile industries in Ranaghat–Fulia region of West Bengal, India. *Journal of Advanced Research*. 2015; 6:995–1002. <https://doi.org/10.1016/j.jare.2014.12.002> PMID: 26644938
106. Manzoor S, Shah MH, Shaheen N, Khalique A, Jaffar M. Multivariate analysis of trace metals in textile effluents in relation to soil and groundwater. *Journal of Hazardous Materials*. 2006; 137:31–37. <https://doi.org/10.1016/j.jhazmat.2006.01.077> <https://doi.org/10.1016/j.jhazmat.2006.01.077> PMID: 16600476
107. Sivaperumal P, Sankar T, Nair PV. Heavy metal concentrations in fish, shellfish and fish products from internal markets of India vis-a-vis international standards. *Food Chemistry*. 2007; 102(3):612–620. <https://doi.org/10.1016/j.foodchem.2006.05.041>
108. Ozmen M, Ayas Z, Güngördü A, Ekmekci GF, Yerli S. Ecotoxicological assessment of water pollution in Sariyar Dam Lake, Turkey. *Ecotoxicology and Environmental Safety*. 2008; 70:163–173. <https://doi.org/10.1016/j.ecoenv.2007.05.011> <https://doi.org/10.1016/j.ecoenv.2007.05.011> PMID: 17582495
109. ATSDR. *Agency for Toxic Substances and Disease Registry, Toxicological Profile for Zinc*. U.S. Department of Health and Human Services, Public Health Service, Atlanta; 2005.
110. USEPA. *Region 9, Regional Screening Level (RSL) Summary Table (TR = 1E-6, HQ = 1.0)*. 2014. <https://www.epa.gov/region9/superfund/prg.htm>
111. Zafarzadeh A, Bay A, Fakhri Y, Keramati H, Hosseini Pouya R. Heavy metal (Pb, Cu, Zn, and Cd) concentrations in the water and muscle of common carp (*Cyprinus carpio*) fish and associated non-carcinogenic risk assessment: Alagol wetland in the Golestan, Iran. *Toxin Reviews*. 2018; 37(2):154–160. <https://doi.org/10.1080/15569543.2017.1386684>
112. Tvermoes BE, Banducci AM, Devlin KD, Kerger BD, Abramson MM, Bebenek IG, et al. Screening level health risk assessment of selected metals in apple juice sold in the United States. *Food and Chemical Toxicology*. 2014; 71:42–50. <https://doi.org/10.1016/j.fct.2014.05.015> <https://doi.org/10.1016/j.fct.2014.05.015> PMID: 24882758
113. Saha N, Mollah M, Alam M, Rahman MS. Seasonal investigation of heavy metals in marine fishes captured from the Bay of Bengal and the implications for human health risk assessment. *Food Control*. 2016b; 70:110–118. <https://doi.org/10.1016/j.foodcont.2016.05.040>
114. USEPA. *Risk Assessment Guideline for Superfund Volume 1 Human Evaluation Manual (Part A)* 291; 1989.
115. Kalantzi I, Pergantis S, Black K, Shimmield T, Papageorgiou N, Tsapakis M, et al. Metals in tissues of seabass and seabream reared in sites with oxic and anoxic substrata and risk assessment for

- consumers. *Food Chemistry*. 2016; 194:659–670. <https://doi.org/10.1016/j.foodchem.2015.08.072> <https://doi.org/10.1016/j.foodchem.2015.08.072> PMID: 26471605
116. Okogwu OI, Nwonumara GN, Okoh FA. Evaluating Heavy Metals Pollution and Exposure Risk Through the Consumption of Four Commercially Important Fish Species and Water from Cross River Ecosystem, Nigeria. *Bulletin of Environmental Contamination and Toxicology*. 2019; 102:867–872. <https://doi.org/10.1007/s00128-019-02610-4> <https://doi.org/10.1007/s00128-019-02610-4> PMID: 30989284
117. Fakhri Y, Saha N, Miri A, Baghaei M, Roomiani L, Ghaderpoori M, et al. Metal concentrations in fillet and gill of parrotfish (*Scarus ghobban*) from the Persian Gulf and implications for human health. *Food and Chemical Toxicology*. 2018; 118:348–354. <https://doi.org/10.1016/j.fct.2018.05.041> <https://doi.org/10.1016/j.fct.2018.05.041> PMID: 29782897