



Bioaerosol Concentrations and Size Distributions during the Autumn and Winter Seasons in an Industrial City of Central China

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ABSTRACT

The ambient bioaerosols were measured in PM_{2.5} and PM₁₀ samples taken in Huangshi City, Hubei Province, China, during autumn and winter from November 2017 to February 2018. Both the bioaerosol number concentration and size distribution (0.37–16 μm) were obtained by direct fluorescent staining coupled with microscopic imaging. The bioaerosol number concentrations ranged from 0.05 to 3.4 # cm⁻³ for PM_{2.5} and from 0.17 to 5.7 # cm⁻³ for PM₁₀, with averages of 0.90 # cm⁻³ and 1.9 # cm⁻³, respectively. In terms of particle number, the bioaerosols were dominated by fine particles (0.37–2.5 μm in diameter), with a larger proportion of submicron than supermicron particles. Assuming a unit density of 1 g cm⁻³ and a spherical shape for the particles, the mass abundances of the bioaerosols were estimated to be 2.4 ± 1.9% and 4.8 ± 3.2% of the PM_{2.5} and PM₁₀, respectively, as measured by a nearby compliance monitor. Higher bioaerosol concentrations were observed in winter than autumn and on polluted than non-polluted days. During heavily polluted conditions, bioaerosols in the PM_{2.5} and PM₁₀ were enriched by 6 and 3.7 times, respectively, compared to non-polluted days and contributed up to 15% of the PM₁₀ mass. Rainfall and snowfall appeared to lower the bioaerosol levels. As enhanced emission controls on combustion and dust sources decrease PM levels in China, the bioaerosol fraction in measured PM concentrations will likely increase.

Keywords: Primary biological aerosol particles (PBAP); PM_{2.5}; PM₁₀; Size distribution; Air quality; Fluorescence microscopy.

INTRODUCTION

Bioaerosols, also referred to as primary biological aerosol particles (PBAPs), include fur fibers, dandruff, skin fragments, plant fragments, pollen, spores, bacteria, algae, fungi, and viruses ranging in diameters from tens of nanometers to millimeters (Jaenicke, 2005). Bioaerosols are important because some PBAPs cause human, plant, and animal diseases (Douwes *et al.*, 2003; Griffin, 2007) while

others act as cloud condensation nuclei affecting cloud formation and precipitation (Hauspie and Pagezy, 2002; Heidi *et al.*, 2003; Group, 2004; Rosenfeld *et al.*, 2008). Recent studies suggest diverse bioaerosol concentrations and size distributions in both indoor and outdoor environments while calling for a better understanding of how bioaerosols co-vary with other air pollutants, especially PM_{2.5} and PM₁₀ (airborne particulate matter [PM] with aerodynamic diameter < 2.5 and < 10 μm, respectively), and their effects on climate and health (Wei *et al.*, 2016; Huang *et al.*, 2017; Xie *et al.*, 2018; Zhai *et al.*, 2018).

Poor air quality in China, particularly elevated PM_{2.5} during cold seasons, is often attributed to extensive coal combustion for heating coupled with unfavorable weather patterns (Cao *et al.*, 2012; Huang *et al.*, 2014; Fu and

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Chen, 2017). Although PM_{2.5} chemical compositions have been studied and used for source apportionment (Henry *et al.*, 1984; Watson *et al.*, 2008, 2016; Cheng *et al.*, 2018), bioaerosol contributions to PM_{2.5} are rarely assessed. Several recent studies examined the spatial and temporal variability of bioaerosol concentrations, size distributions, and speciation in China (Cao *et al.*, 2014; Gao *et al.*, 2015; Li *et al.*, 2015; Dong *et al.*, 2016; Wei *et al.*, 2016). Higher bioaerosol concentrations were reported on polluted than on non-polluted days. Xie *et al.* (2018) found a positive relationship between air quality index (AQI) and bioaerosol concentrations. Most of these studies were conducted in heavily populated megacities of northern China, such as Beijing, Xi'an, and Qingdao (Cao *et al.*, 2014; Wei *et al.*, 2016; Dong *et al.*, 2016; Xie *et al.*, 2018). Bioaerosol characteristics may differ in smaller cities and in central-southern China, owing to different vegetation, land use, and climate conditions.

Huangshi, a medium-size city with an area of 4,583 km² and a population of ~2.5 million, is located ~90 km southwest of Wuhan in central China. It features a subtropical continental monsoon climate, with a mean annual temperature of 17°C and precipitation of ~1400 mm. The diverse vegetation coupled with mild temperatures and high relative humidity facilitates bioaerosol production. Huangshi is an important industrial source of raw materials with high energy consumption and the accompanying environmental pollution (Liu *et al.*, 2017). Affected by local industries, particularly coal-based ore smelters, and intrusion of regional polluted air, poor air quality often occurs during autumn and winter. Average PM_{2.5} concentrations for the two seasons have exceeded 100 µg m⁻³ with ~20% from organic carbon (OC) (Zhan *et al.*, 2017). Based on the EC-tracer analysis, Zhan *et al.* (2017) concluded that a substantial fraction of OC originated from bioaerosols and/or secondary organic carbon (SOC), but they were not able to distinguish between the two. Average OC in PM₁₀ was ~5 µg m⁻³ higher than in PM_{2.5} (Zhan *et al.*, 2017) and the difference may be due to bioaerosols such as fungal spores, pollen, and vegetative detritus (Edgerton *et al.*, 2009).

Chen *et al.* (2019) demonstrated the practicality of using a direct-staining (DS) technique, coupled with epifluorescence microscopy (FM), to quantify bioaerosols collected on filters. The DS-FM method measures bioaerosol number concentration and size distribution, from which the bioaerosol mass can be estimated. Compared to culture-based methods, fluorescence methods detect bioaerosols in viable, non-viable, and viable but non-culturable states (Li and Huang, 2006; Chi and Li, 2007; Li *et al.*, 2011). DS-FM is also more cost-effective than online monitoring with fluorescence analyzers, such as the Waveband Integrated Bioaerosol Sensor (WIBS) and Ultraviolet Aerodynamic Particle Sizer (UV-APS). This study applies the DS-FM method to PM_{2.5} and PM₁₀ filter samples collected in Huangshi. The concentrations and size distributions of bioaerosols are examined. Results from this work further our understanding of bioaerosol contributions to PM and offer insights to the regional air pollution management.

METHODS

Sampling Site and Methods

The sampling site (30°12'35.71"N, 115°01'30.75"E) was on the rooftop of a five-story building (about 15 m AGL) at the Hebei Polytechnic University (HBPU) campus in central Huangshi (Fig. 1). This site is surrounded by trees, greenbelts, and a stadium and is ~400 m from major highways with no nearby industrial activities. Ambient PM_{2.5} and PM₁₀ samples were acquired daily between November 4, 2017, and February 10, 2018, using a pair of MiniVol samplers (Airmetrics, Springfield, OR, USA) equipped with size-selective inlet sampling at ~5 L min⁻¹. Sampling started at 13:00 LST and lasted for 5–8 hours. Particles were collected onto 47-mm diameter black polycarbonate filters (0.2 µm pore size, PCTE; Whatman, Little Chalfont, UK) as these provide the lowest background for DS-FM.

Hourly PM_{2.5} and PM₁₀ mass concentrations were measured using a Tapered Element Oscillating Microbalance (1405 TEOM™; Thermal Fisher Scientific, Waltham, MA, USA) located ~80 m from the HBPU site (see Fig. 1), as part of the Ministry of Ecology and Environment's compliance network to determine Air Quality Index (AQI). According to China's "Technical Regulation on Ambient Air Quality Index (HJ633-2012)," the AQI value from PM_{2.5} is used to categorize air pollution. One of the six air quality levels—good (0–50), moderate (51–100), lightly polluted (101–150), moderately polluted (151–200), heavily polluted (201–300), and severely polluted (> 300)—was assigned to each monitoring day. Days with AQI values below 100 (good and moderate) were considered as non-polluted days.

Local meteorological parameters such as wind speed and precipitation were obtained from the China Weather Network (<http://www.weather.com.cn/weather1d/101200601.shtml>).

Bioaerosol Particle Detection

Bioaerosols were measured using a fluorescence microscope after staining with 4',6-diamidino-2-phenylindole dihydrochloride (DAPI) following the DS-FM protocol as described in Chen *et al.* (2019). Briefly, a 13-mm diameter disc was removed from each polycarbonate filter sample and placed with the deposit-side up onto a drop of DAPI working solution (20 µg mL⁻¹). It was incubated at room temperature in the dark for 20 minutes while the stain permeated through the filter to interact with the bioaerosols (Griffin *et al.*, 2001; Prussin *et al.*, 2015). A coverslip was then mounted on the stained sample with a water-soluble, anti-fading adhesive (Mounting Medium; Solarbio, Beijing, China) and stored below 4°C before epifluorescence investigation. DS-FM differs from the extraction-staining fluorescence microscopy (ES-FM) method (Cao *et al.*, 2014; Dong *et al.*, 2016; Wei *et al.*, 2016; Xie *et al.*, 2018) in that the latter stains particles in a liquid after they are extracted from filter samples. The stained particles are then re-deposited onto polycarbonate filters for FM analysis.

The prepared sample slides were examined on a fluorescence microscope (DM2500; Leica, Germany) equipped with a Charge Coupled Device (CCD) camera

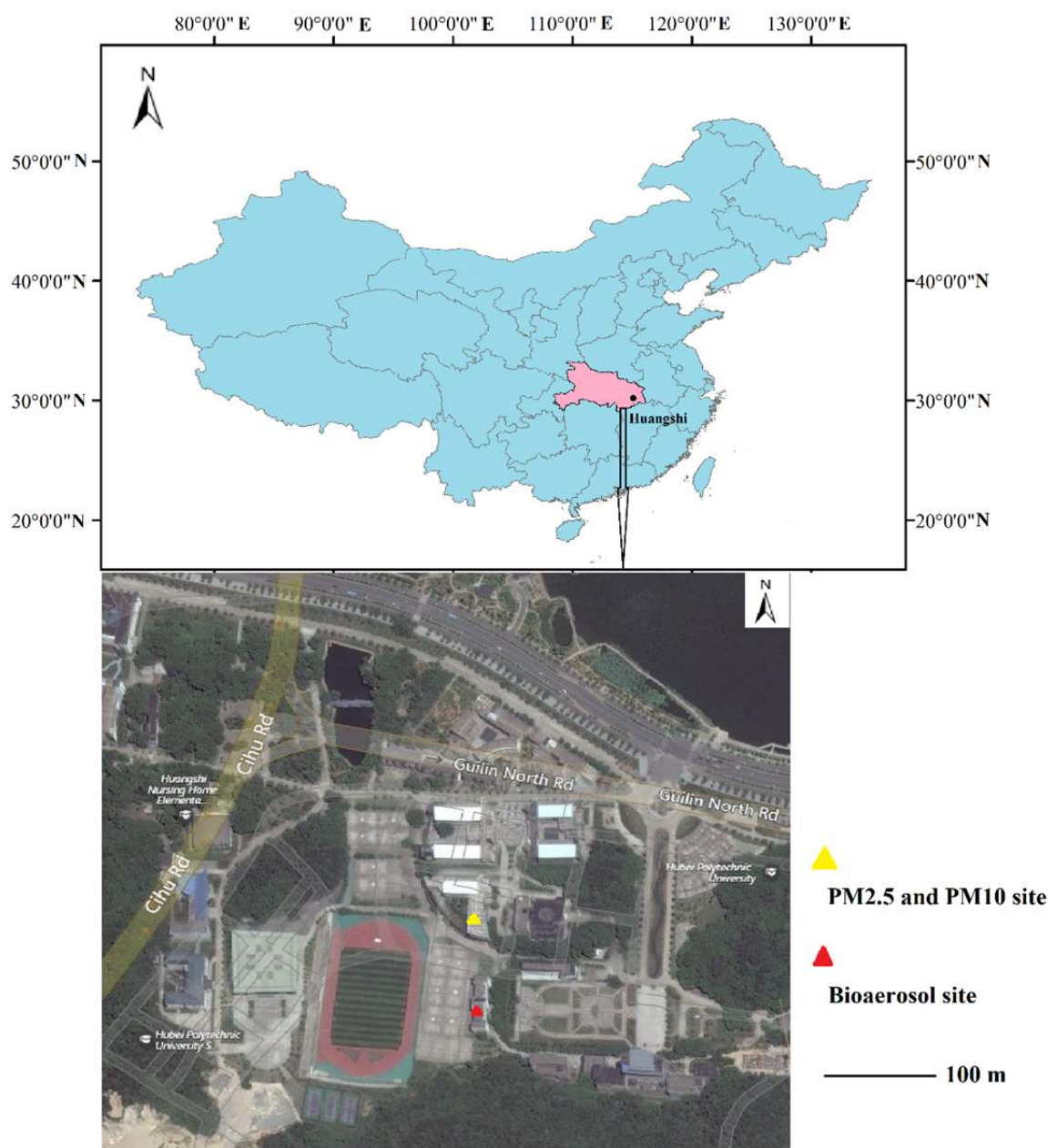


Fig. 1. Bioaerosol monitoring site and the nearby (~80 m) air quality monitoring station at the Hubei Polytechnic University (HBPU), Huangshi, China.

(DFC450 C; Leica, Germany) and a filter cube containing a 350/50 band pass (BP) excitation filter, a 400 nm dichromatic mirror, and a 460/50 BP emission filter. The DAPI-DNA coupling produces a bright blue fluorescence at ~460 nm when excited with 365 nm light (Porter and Feig, 1980). Under a 400× magnification, about 30 images of view ($0.218 \times 0.163 \text{ mm}^2$, 2560×1920 pixels) were captured along a filter dissection to represent the entire 13-mm diameter deposit area.

Controls were prepared from a subset of exposed filters in the same way as the samples except for staining with DAPI. Fluorescing particles were not detected on the controls (see examples in Fig. S1 of the supplementary information). Though non-biological particles such as those containing

polycyclic aromatic hydrocarbons (PAHs) auto-fluoresce in the emission region of 440–470 nm (Pan, 2015), the auto-fluorescence may be too weak to be distinguished from the background of polycarbonate filter.

Quantification of Bioaerosol Concentration and Size Distribution

Using the ImageJ[®] software, the total number and area of bioaerosol particles on each fluorescence image were calculated automatically. The images were converted to binary (black-and-white) pictures with the fluorescent particles, a surrogate for bioaerosols, in black against a white background. The thresholding between particles and background was accomplished with the “Triangle” algorithm

(Zack *et al.*, 1977; Seo *et al.*, 2014). Particles smaller than 15 pixels were excluded to suppress false positives. ImageJ[®] reported the particle number in the image and the projected area (A_p) of each individual particle, from which the equivalent projected area diameter ($D_{eq,A}$) was calculated:

$$D_{eq,A} = 2 \left(\frac{A_p}{\pi} \right)^{1/2} \quad (1)$$

The cut-off $D_{eq,A}$ was 0.37 μm , corresponding to 15 pixels, which should be sufficient to detect most bacteria, fungal spores, and other major mass-contributing bioaerosol classes. The automatic bioaerosol counts were verified by manual counting for selected samples. Bioaerosol number concentration ($\# \text{cm}^{-2}$) was determined from the average particle counts over the 30 fluorescence images taken for each sample, divided by the image area ($0.218 \times 0.163 \text{ mm}^2$), with the standard error representing the uncertainty.

To estimate bioaerosol mass, particle volume (V) and density (ρ) are required. The first-order approximation assumes spherical particles, thus:

$$V = \frac{\pi}{6} (D_{eq,A})^3 \quad (2)$$

Particle density may depend on the bioaerosol type, but an assumption of 1 g cm^{-3} (Matthias-Maser and Jaenicke, 2000; Chow *et al.*, 2015) was used in this study for all bioaerosol particles. The bioaerosol number and mass concentrations were converted to $\# \text{cm}^{-3}$ and $\mu\text{g m}^{-3}$, respectively, using the MiniVol sampling time and flow rate. Errors from the bioaerosol count and flow rate were propagated to yield the measurement uncertainty. Excel 2013 and SPSS 19.0 software were used to statistically analyze the experimental data.

RESULTS AND DISCUSSION

Bioaerosol Concentrations and Size Distribution

A total of 51 pairs of $\text{PM}_{2.5}/\text{PM}_{10}$ samples were collected. Bioaerosol number concentrations ranged from 0.05 to $3.37 \# \text{cm}^{-3}$ for $\text{PM}_{2.5}$ and from 0.17 to $5.73 \# \text{cm}^{-3}$ for PM_{10} (Fig. 2), with averages of $0.90 \# \text{cm}^{-3}$ and $1.86 \# \text{cm}^{-3}$, respectively. Taking November as autumn and December to January as winter, the concentrations varied significantly by season with higher values found in winter ($1.04 \# \text{cm}^{-3}$ for $\text{PM}_{2.5}$ and $2.16 \# \text{cm}^{-3}$ for PM_{10}) than during autumn ($0.60 \# \text{cm}^{-3}$ for $\text{PM}_{2.5}$ and $1.25 \# \text{cm}^{-3}$ for PM_{10}). Xie *et al.* (2018) and Dong *et al.* (2016) observed similar seasonal differences in Xi'an and Qingdao, China, respectively. They attributed the high winter bioaerosol levels to stagnant weather conditions, high PM levels, and hazy days. In Huangshi, severe haze pollution was also observed more frequently in winter.

With respect to the particle size-number distribution, bioaerosols were dominated by fine particles 0.37–2.5 μm in diameter (Fig. 3). Image analysis showed that coarse particles ($D_{eq,A} > 2.5 \mu\text{m}$) accounted for 2.6% and 6.5% of bioaerosol numbers in $\text{PM}_{2.5}$ and PM_{10} samples, respectively. The small fraction of coarse particles in $\text{PM}_{2.5}$ samples reflects the imperfect MiniVol size-selective inlet that passes some coarse particles. Bioaerosols with diameters smaller than 1 μm were classified as prokaryotes (e.g., bacteria) and viruses, while supermicrometer particles were classified as eukaryotes (e.g., fungal spores, pollen, and organic debris) (Mayol *et al.*, 2014). The proportions of prokaryotes were 68.7% among $\text{PM}_{2.5}$ and 59.5% among PM_{10} bioaerosols, and the proportions of eukaryotes were lower with 31.3% among $\text{PM}_{2.5}$ and 40.5% among PM_{10} bioaerosols.

If determined from the difference of bioaerosol concentrations between PM_{10} and $\text{PM}_{2.5}$ samples (i.e., 1.86 minus $0.90 \# \text{cm}^{-3}$), coarse bioaerosols would account for $0.96 \# \text{cm}^{-3}$ or 51.6% of the PM_{10} bioaerosol numbers,

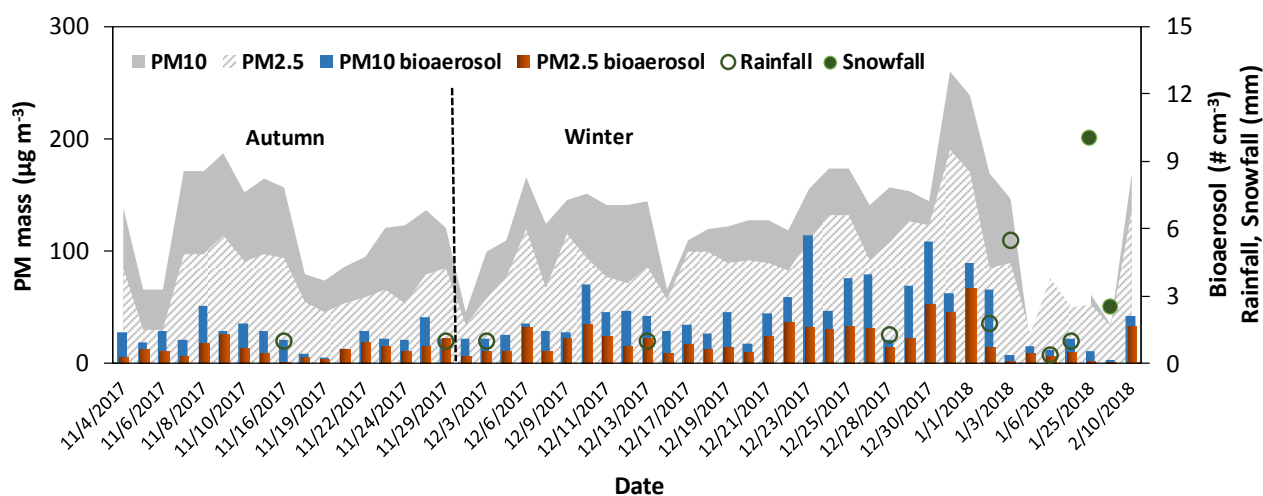


Fig. 2. Day-to-day variability in PM_{10} and $\text{PM}_{2.5}$ mass (from TEOM compliance monitors) and bioaerosol number concentrations from November 4, 2017, to February 10, 2018, in Huangshi, China. $\text{PM}_{2.5}$ concentration exceeded PM_{10} on January 5 and 7, 2018, likely due to interference from heavy rain.

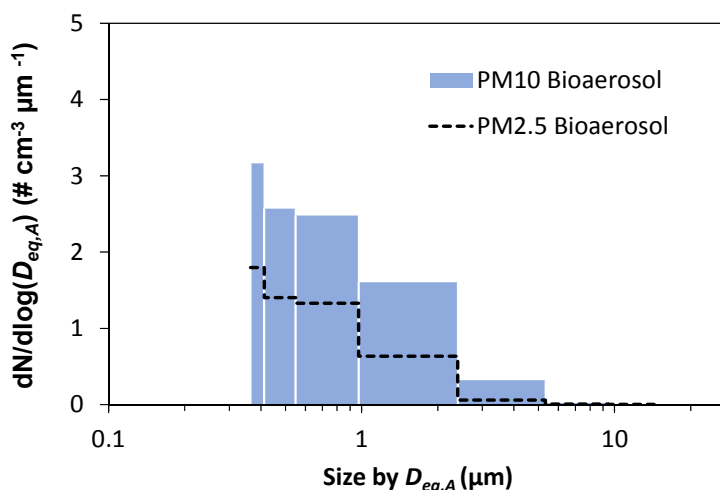


Fig. 3. Average number-size distribution of bioaerosol samples measured by DS-FM. Particles were counted by 7 size bins (0.37–0.42, 0.42–0.56, 0.56–1, 1–2.5, 2.5–5.6, 5.6–10, and 10–16 μm) and averaged over all samples.

much larger than the 6.5% number fraction inferred solely from the image analysis of PM_{10} samples. The contrast suggests that substantial fine bioaerosol particles might have been attached to other coarse particles (e.g., fugitive dust) and only captured in PM_{10} samples. Similar phenomena were reported during dust episodes (Yeo and Kim, 2002; Hallar et al., 2011).

The reconstructed bioaerosol mass from Eq. (2) exhibits a dominant contribution from supermicrometer particles (Fig. 4). As shown in Fig. 2, $\text{PM}_{2.5}$ mass ranged from 27 to 190 $\mu\text{g m}^{-3}$ with an average of 85.7 $\mu\text{g m}^{-3}$ while PM_{10} mass ranged from 34 to 260 $\mu\text{g m}^{-3}$ with an average of 129 $\mu\text{g m}^{-3}$. The bioaerosol component, on average, accounted for 2.1 $\mu\text{g m}^{-3}$ ($2.4 \pm 1.9\%$) of $\text{PM}_{2.5}$ mass and 6.3 $\mu\text{g m}^{-3}$ ($4.8 \pm 3.2\%$) of PM_{10} mass with the highest mass fractions being 10% for $\text{PM}_{2.5}$ (December 13, 2017) and 14% for PM_{10} (December 18, 2017).

The summed mass of bioaerosols with $D_{eq,A} < 2.5 \mu\text{m}$ on PM_{10} sample images only accounts for $1.8 \pm 1.0\%$ of $\text{PM}_{2.5}$ mass, lower than the $2.4 \pm 1.9\%$ reconstructed from $\text{PM}_{2.5}$ sample images by combining bioaerosols of all sizes. On the other hand, coarse bioaerosols $> 2.5 \mu\text{m}$ $D_{eq,A}$ would account for $13.5 \pm 15.5\%$ of $\text{PM}_{2.5-10}$ determined from the difference of PM_{10} and $\text{PM}_{2.5}$ mass. The bioaerosol components were more substantial in coarse than fine PM.

Relationships of Bioaerosol, PM, and Meteorological Conditions

Temperature, relative humidity, wind speed, and ambient PM may influence bioaerosol concentrations. $\text{PM}_{2.5}$ and PM_{10} mass were found to significantly correlate with the bioaerosol numbers (Table 1), consistent with previous research (Haas et al., 2013; Alghamdi et al., 2014; Xie et al., 2018). This suggests that bioaerosols and PM are impacted by similar factors. Coarse particles can act as carriers for microorganisms, and the increasing quantity of PM offers more surface area on which microbes can adhere (Jeon et al., 2011; Xie et al., 2018). On the other hand, ultrafine particles ($< 100 \text{ nm}$), including soot particles (30–

40 nm), may adhere to bioaerosols.

Daily mean temperatures varied from 0 to 23°C and daily mean wind speeds varied from 0.84 to 2.88 m s^{-1} throughout the monitoring period. Neither of them was correlated with bioaerosol levels. Relative humidity was negatively correlated with bioaerosol concentrations ($p < 0.05$). High relative humidity mainly occurred on rainy and snowy days (Fig. 2), while bioaerosol concentrations during and after the precipitation decreased due to scavenging (Li et al., 2017; Xie et al., 2018).

For example, through a prolonged rainfall from January 1 to January 3, 2018, $\text{PM}_{2.5}$ concentrations were reduced from 170 to 89 $\mu\text{g m}^{-3}$ and PM_{10} concentrations decreased from 239 to 147 $\mu\text{g m}^{-3}$ while bioaerosol number concentrations decreased from 3.37 to 0.10 $\# \text{ cm}^{-3}$ in $\text{PM}_{2.5}$ and from 4.48 to 0.41 $\# \text{ cm}^{-3}$ in PM_{10} . A similar situation was observed for the January 25–26, 2018 snowfall period with low bioaerosol number concentrations (i.e., 0.05–0.09 $\# \text{ cm}^{-3}$ for $\text{PM}_{2.5}$ and 0.17–0.52 $\# \text{ cm}^{-3}$ for PM_{10}) observed after the snow. By contrast, some studies reported higher bioaerosol levels after precipitation, particularly during the spring-summer blooming seasons (Huffman et al., 2013; Kang et al., 2015; Rathnayake et al., 2017).

Bioaerosol versus Air Quality Levels

Based on the AQI, there were 3 good days, 14 moderate days, 27 lightly polluted days, 5 moderately polluted days, and 2 heavily polluted days during the sampling period. Table 2 presents bioaerosol number and PM mass concentrations by AQI level. Average bioaerosol concentrations for non-polluted days were $0.47 \pm 0.24 \# \text{ cm}^{-3}$ for $\text{PM}_{2.5}$ and $1.02 \pm 0.53 \# \text{ cm}^{-3}$ for PM_{10} and were significantly different from those for the heavily polluted days (t -test: $p = 0.002$). Average bioaerosol concentrations for moderately and heavily polluted days were 3.6–6 times higher for $\text{PM}_{2.5}$ and 3.3–3.7 times higher for PM_{10} as compared with non-polluted days. There was no appreciable difference in the bioaerosol size distributions among different air quality levels.

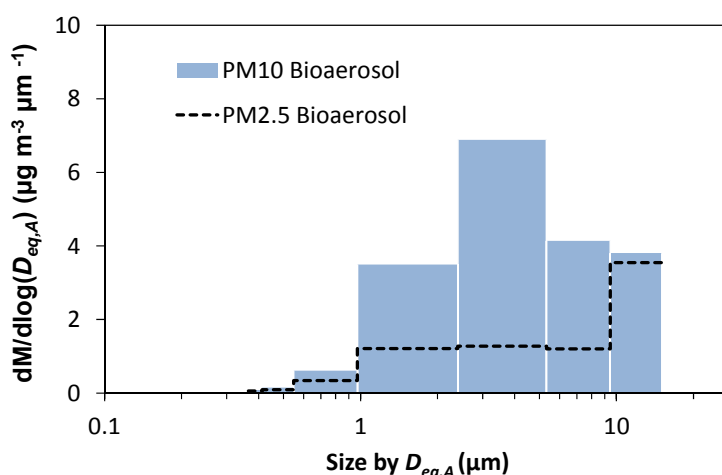


Fig. 4. Average mass-size distribution of PM_{2.5} and PM₁₀ bioaerosols by DS-FM. Particle mass (M) was calculated for 7 size bins between 0.37 and 16 µm in diameter (0.37–0.42, 0.42–0.56, 0.56–1, 1–2.5, 2.5–5.6, 5.6–10, and 10–16 µm) and averaged over all samples.

Table 1. Spearman correlation coefficients between the bioaerosol concentrations and environmental factors for all sampling days.

	PM _{2.5} ^c	Temperature ^c	RH ^f	Wind Speed ^g
PM _{2.5} Bioaerosols ^a	0.627**	0.180	−0.463**	0.124
	PM ₁₀ ^d	Temperature ^c	RH ^f	Wind Speed ^g
PM ₁₀ Bioaerosols ^b	0.582**	0.178	−0.434*	0.094

^a bioaerosols of PM_{2.5} (# cm^{−3}).

^b bioaerosols of PM₁₀ (# cm^{−3}).

^c PM_{2.5} mass (µg m^{−3}).

^d PM₁₀ mass (µg m^{−3}).

^e The daily mean temperature (°C).

^f The daily mean relative humidity (%).

^g The daily mean wind speed (m sec^{−1}).

** p < 0.01 (2-tailed).

* p < 0.05 (2-tailed).

Table 2. Summary of bioaerosol and PM mass concentrations by air quality level in Huangshi, China (November 4, 2017–February 10, 2018).

Air quality level ^a	N ^b	PM _{2.5} (µg m ^{−3})	PM ₁₀ (µg m ^{−3})	PM _{2.5} Bioaerosol (# cm ^{−3})	PM ₁₀ Bioaerosol (# cm ^{−3})
Good	3	31 ± 4	32 ± 15	0.26 ± 0.19	0.70 ± 0.49
Moderate	14	54 ± 12	92 ± 27	0.52 ± 0.23	1.09 ± 0.53
Lightly polluted	27	96 ± 18	140 ± 26	0.88 ± 0.51	1.96 ± 1.16
Moderately polluted	5	127 ± 5	162 ± 13	1.71 ± 0.54	3.36 ± 1.40
Heavily polluted	2	180 ± 14	250 ± 15	2.84 ± 0.75	3.81 ± 0.95

^a Air quality levels were categorized by AQI: Good (0–50), Moderate (51–100), Lightly polluted (101–150), Moderately polluted (151–200), Heavily polluted (201–300), and Severely polluted (> 300). ^b Number of days for each air quality level.

AQI values were consistently above 100 and exceeded 200 (i.e., polluted) during the December 21, 2017–January 1, 2018 period with PM₁₀ bioaerosol numbers > 2.25 # cm^{−3}, except on December 28, 2017 (1.08 # cm^{−3}), when precipitation of ~1.3 mm occurred (Fig. 2). Elevated bioaerosol concentrations (2.6 # cm^{−3} for PM_{2.5} and 5.4 # cm^{−3} for PM₁₀), PM_{2.5} (123 µg m^{−3}), and PM₁₀ (144 µg m^{−3}) returned on December 30, 2017, right after the precipitation. The haze episode on December 30, 2017 covered part of Hubei Province (e.g., Wuhan and Huangshi) while also affecting nearby Anhui

and Jiangshu Provinces. Back trajectory analysis (Fig. S2) reveals possible transport of polluted air masses from northern China. Bioaerosols contributed to ~10% of PM₁₀ mass during the episode.

In an urban environment such as Huangshi, conditions that engender high PM episodes, such as stagnation and shallow boundary layers, might also lead to the accumulation of bioaerosols, while precipitation would lower bioaerosol and other PM components simultaneously. It is noteworthy that bioaerosol concentrations did not always coincide with

the deteriorating air quality. The highest PM₁₀ bioaerosol concentration, 5.73 # cm⁻³, was found on December 23, 2017, with an AQI of 144 (i.e., lightly polluted). Xie *et al.* (2018) reported the highest bioaerosol levels under the moderately polluted condition, whereas Wei *et al.* (2016) found that upon the occurrence of haze the number concentration of fluorescent particles ascended briefly, and then started to decrease as the haze progressed over time during a short monitoring period. The differences in source and formation mechanism between secondary aerosol and bioaerosols may explain their distinct temporal trends during the pollution episodes.

Table 3 compares the bioaerosol concentrations of polluted days. The average PM₁₀ bioaerosol concentration in this study was 3–4 times higher than those measured in Beijing, Xi'an, and Qingdao. The Xi'an and Qingdao bioaerosols were measured using the ES-FM method with DAPI stain. ES-FM likely yields lower bioaerosol counts than DS-FM because of particle losses during the wash-off and re-deposition steps (Chen *et al.*, 2019). Beijing's bioaerosols were quantified by UV-APS, though real-time auto-fluorescence methods including UV-APS and WIBS are known to underestimate bioaerosol counts as not every bioaerosol particle fluoresces under the experimental conditions (Huffman *et al.*, 2010; Després *et al.*, 2012). Different analytical methods partly explain the higher bioaerosol concentrations measured in this study. Other factors include different climate conditions among these cities.

CONCLUSIONS

The DS-FM method was applied to quantify not only the bioaerosol number concentration but also the size distribution. Higher concentrations were observed in winter than autumn and on polluted than non-polluted days. However, these levels did not peak at the climax of haze episodes, implying different sources and/or subtle interactions between the bioaerosols and the PM. The results also confirm the important role of precipitation in removing bioaerosols. The bioaerosol number concentration was higher in the PM_{2.5} fraction, but the mass concentration was higher in the PM_{2.5-10} fraction. Bioaerosols contributed substantially to the mass of airborne particles in Huangshi, especially on polluted days. As a result of enhanced emission controls that have been reducing PM from combustion and dust sources in China, bioaerosols are expected to constitute a growing fraction of measured PM concentrations. Investigations of bioaerosol contributions to air pollution should be continued and extended to other regions of China.

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Table 3. Comparison of bioaerosol concentrations for different cities in China during polluted periods.

City	Period	Bioaerosol type	Polluted period criteria	Bioaerosols of		Analytical method	Reference
				polluted period (# cm ⁻³)	non-polluted period (# cm ⁻³)		
Beijing, China	Dec. 2013–Mar. 2014	> 0.5 μm bio-PM	AQI > 150	0.5	0.07	UV-APS ^a	Wei <i>et al.</i> (2016)
Qingdao, China	Oct. 2013–Aug. 2014	Total bioaerosol	Unknown	0.709	0.655	DAPI ^b and fluorescence microscope	Dong <i>et al.</i> (2016)
Xi'an, China	Apr. 2016–Feb. 2017	Total bioaerosol	AQI > 100; atmospheric visibility < 10 km; RH < 90%	0.612	0.215	DAPI ^b and fluorescence microscope	Xie <i>et al.</i> (2018)
Huangshi, China	Nov. 2017–Feb. 2018	PM ₁₀ bioaerosol	AQI > 100	2.28	1.02	DAPI ^b and fluorescence microscope	This study

^a Ultraviolet Aerodynamic Particle Sizer.

^b 4',6-diamidino-2-phenylindole dihydrochloride.

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SUPPLEMENTARY MATERIAL

Supplementary data associated with this article can be found in the online version at <http://www.aaqr.org>.

REFERENCES

- Alghamdi, M.A., Shamy, M., Redal, M.A., Khoder, M., Awad, A.H. and Elserougy, S. (2014). Microorganisms associated particulate matter: A preliminary study. *Sci. Total Environ.* 479–480: 109–116.
- Bauer, H., Claeys, M., Vermeylen, R., Schueller, E., Weinke, G., Berger, A. and Puxbaum, H. (2008). Arabitol and mannitol as tracers for the quantification of airborne fungal spores. *Atmos. Environ.* 42: 588–593.
- Burge, H.A. (1995). Bioaerosol investigations. In *Bioaerosols*, Burge, H.A. (Ed.), Lewis Publishers, Boca Raton, FL, USA, p. 13.
- Buser, M.D., Parnell, C.B., Shaw, B.W. and Lacey, R.E. (2007). Particulate matter sampler errors due to the interaction of particle size and sampler performance characteristics: Ambient PM_{2.5} samplers. *Trans. ASABE* 50: 241–254.
- Cao, C., Jiang, W., Wang, B., Fang, J., Lang, J., Tian, G., Jiang, J. and Zhu, T.F. (2014). Inhalable microorganisms in Beijing's PM_{2.5} and PM₁₀ pollutants during a severe smog event. *Environ. Sci. Technol.* 48: 1499–1507.
- Cao, J.J., Shen, Z.X., Chow, J.C., Watson, J.G., Lee, S.C., Tie, X.X. and Han, Y.M. (2012). Winter and summer PM_{2.5} chemical compositions in fourteen Chinese Cities. *J. Air Waste Manage. Assoc.* 6: 1214–1226.
- Chen, L.W.A. and Cao, J.J. (2018). PM_{2.5} source apportionment using a hybrid environmental receptor model. *Environ. Sci. Technol.* 52: 6357–6369.
- Chen, L.W.A., Zhang, M., Liu, T., Fortier, K., Chow, J.C., Kolberg, R., Cao, J.J., Lin, G., Patel, T.Y., Cruz, P., Buttner, M.P. and Watson, J.G. (2019). Evaluation of epifluorescence methods for quantifying bioaerosols in fine and coarse particulate air pollution. *Atmos. Environ.* Submitted.
- Chi, M.C. and Li, C.S. (2007). Fluorochrome in monitoring atmospheric bioaerosols and correlations with meteorological factors and air pollutants. *Aerosol Sci. Technol.* 41: 672–678.
- Chow, J.C., Yang, X.F., Wang, X.L., Kohl, S.D., Hurbain, P.R., Chen, L.W.A. and Watson, J.G. (2015). Characterization of ambient PM₁₀ bioaerosols in a California agricultural town. *Aerosol Air Qual. Res.* 15: 1433–1447.
- Cox, C.S. and Wathes, C.M. (1995). *Bioaerosol handbook*. Lewis Publishers, Boca Raton, FL, USA.
- Després, V., Huffman, J.A., Burrows, S.M., Hoose, C., Safatov, A., Buryak, G., Fröhlich-Nowoisky, J., Elbert, W., Andreae, M., Poeschl, U. and Jaenicke, R. (2012). Primary biological aerosol particles in the atmosphere: A review. *Tellus B* 64: 15598.
- Dong, L.J., Qi, J.H., Shao, C.C., Zhong, X., Gao, D.M., Cao, W.W., Gao, J.W., Bai, R., Long, G.Y. and Chu, C.C. (2016). Concentration and size distribution of total airborne microbes in hazy and foggy weather. *Sci. Total Environ.* 541: 1011–1018.
- Douwes, J., Thorne, P., Pearce, N. and Heederik, D. (2003). Bioaerosol health effects and exposure assessment: Progress and prospects. *Ann. Occup. Hyg.* 47: 187–200.
- Edgerton, E.S., Casuccio, G.S., Saylor, R.D., Lersch, T.L., Hartsell, B.E., Jansen, J.J. and Hansen, D.A. (2009). Measurements of OC and EC in coarse particulate matter in the southeastern United States. *J. Air Waste Manage. Assoc.* 59: 78–90.
- Fröhlich-Nowoisky, J., Kampf, C.J., Weber, B., Huffman, J.A., Pöhlker, C., Andreae, M.O., Lang-Yona, N., Burrows, S.M., Gunthe, S.S., Elbert, W. and Su, H. (2016). Bioaerosols in the Earth system: Climate, health, and ecosystem interactions. *Atmos. Res.* 182: 346–376.
- Fu, H.B. and Chen, J.M. (2017). Formation, features and controlling strategies of severe haze-fog pollutions in China. *Sci. Total Environ.* 578: 121–138.
- Gao, M., Jia, R.Z., Qiu, T.L., Han, M.L., Song, Y. and Wang, X.M. (2015). Seasonal size distribution of airborne culturable bacteria and fungi and preliminary estimation of their deposition in human lungs during non-haze and haze days. *Atmos. Environ.* 118: 203–210.
- Griffin, D.W. (2007). Atmospheric movement of microorganisms in clouds of desert dust and implications for human health. *Clin. Microbiol. Rev.* 20: 459–477.
- Group, A.E. (2004). New Directions: The role of bioaerosols in atmospheric chemistry and physics. *Atmos. Environ.* 38: 1231–1232.
- Haas, D., Galler, H., Luxner, J., Zarfel, G., Buzina, W., Friedl, H., Marth, E., Habib, J. and Reinthaler, F.F. (2013). The concentrations of culturable microorganisms in relation to particulate matter in urban air. *Atmos. Environ.* 65: 215–222.
- Hallar, A., Chirokova, G., McCubbin, I., Painter, T.H., Wiedinmyer, C. and Dodson, C. (2011). Atmospheric bioaerosols transported via dust storms in the western United States. *Geophys. Res. Lett.* 38: L17801.
- Hara, K. and Zhang, D.Z. (2012). Bacterial abundance and viability in long-range transported dust. *Atmos. Environ.* 47: 20–25.
- Hauspie, R.C. and Pagezy, H. (2002). The contribution of bacteria and fungal spores to the organic carbon content of cloud water, precipitation and aerosols. *Atmos. Res.* 64: 109–119.
- Heidi, B., Heinrich, G., Regina, H., Anne, K., Georg, R., Franziska, Z. and Hans, P. (2003). Airborne bacteria as cloud condensation nuclei. *J. Geophys. Res.* 108: 1919–1964.
- Henry, R.C., Lewis, C.W., Hopke, P.K. and Williamson, H.J. (1984). Review of receptor model fundamentals. *Atmos. Environ.* 18: 1507–1515.
- Huang, H.L., Lee, M.K., and Shih, H.W. (2017). Assessment of indoor bioaerosols in public spaces by

- real-time measured airborne particles. *Aerosol Air Qual. Res.* 17: 2276–2288.
- Huffman, J.A., Treutlein, B. and Pöschl, U. (2010). Fluorescent biological aerosol particle concentrations and size distributions measured with an Ultraviolet Aerodynamic Particle Sizer (UV-APS) in Central Europe. *Atmos. Chem. Phys.* 10: 3215–3233.
- Huffman, J.A., Prenni, A.J., DeMott, P.J., Pöhlker, C., Mason, R.H., Robinson, N.H., Fröhlich-Nowoisky, J., Tobo, Y., Després, V.R., Garcia, E. and Gochis, D.J. (2013). High concentrations of biological aerosol particles and ice nuclei during and after rain. *Atmos. Chem. Phys.* 13: 6151–6164.
- Jaenicke, R. (2005). Abundance of cellular material and proteins in the atmosphere. *Science* 308: 73.
- Jeon, E.M., Kim, H.J., Jung, K., Kim, J.H., Kim, M.Y., Kim, Y.P. and Ka, J.O. (2011). Impact of Asian dust events on airborne bacterial community assessed by molecular analyses. *Atmos. Environ.* 45: 4313–4321.
- Johnson, D.L., Pearce, T.A. and Esmen, N.A. (1999). The effect of phosphate buffer on aerosol size distribution of nebulized *Bacillus subtilis* and *Pseudomonas fluorescens* bacteria. *Aerosol Sci. Technol.* 30: 202–210.
- Kang, S.M., Heo, K.J. and Lee, B.U. (2015). Why does rain increase the concentrations of environmental bioaerosols during monsoon. *Aerosol Air Qual. Res.* 15: 2320–2324.
- Li, C.S. and Huang, T.Y. (2006). Fluorochrome in monitoring indoor bioaerosols. *Aerosol Sci. Technol.* 40: 237–241.
- Li, M.F., Qi, J.H., Zhang, H.D., Huang, S., Li, L. and Gao, D.M. (2011). Concentration and size distribution of bioaerosols in an outdoor environment in the Qingdao coastal region. *Sci. Total Environ.* 409: 3812–3819.
- Li, Y.P., Fu, H.L., Wang, W., Liu, J., Meng, Q.L. and Wang, W.K. (2015). Characteristics of bacterial and fungal aerosols during the autumn haze days in Xi'an, China. *Atmos. Environ.* 122: 439–447.
- Li, Y.P., Lu, R., Li, W.X., Xie, Z.S. and Song, Y. (2017). Concentrations and size distributions of viable bioaerosols under various weather conditions in a typical semiarid city of Northwest China. *J. Aerosol Sci.* 106: 83–92.
- Liu, H.X., Zheng, J.R., Qu, C.K., Zhang, J.Q., Wang, Y.K., Zhan, C.L., Yao, R.Z. and Cao, J.J. (2017). Characteristics and source analysis of water-soluble inorganic ions in PM₁₀ in a typical mining city, Central China. *Atmosphere* 8: 74.
- Marty, A.N., DeLeon-Rodriguez, N., Waters, S., Ziemba, L., Anderson, B., Bergin, M., Konstantinidis, K. and Nenes, A. (2017). Understanding the link between meteorology and speciated abundance of bioaerosols in an urban environment using collocated flow cytometry and real-time autofluorescence measurements. 19th EGU General Assembly, EGU2017, proceedings from the conference held 23–28 April, 2017 in Vienna, Austria, p. 9555.
- Masiello, C.A. (2004). New directions in black carbon organic geochemistry. *Mar. Chem.* 92: 201–213.
- Matthias-Maser, S. and Jaenicke, R. (2000). The size distribution of primary biological aerosol particles in the multiphase atmosphere. *Aerobiologia* 16: 207–210.
- Mayol, E., Jiménez, M.A., Herndl, G.J., Duarte, C.M. and Arrieta, J.M. (2014). Resolving the abundance and air-sea fluxes of airborne microorganisms in the North Atlantic Ocean. *Front. Microbiol.* 5: 557.
- Pan, Y.L. (2015). Detection and characterization of biological and other organic-carbon aerosol particles in atmosphere using fluorescence. *J. Quant. Spectrosc. Radiat. Transfer* 150: 12–35.
- Perrino, C. and Marcovecchio, F. (2016). A new method for assessing the contribution of primary biological atmospheric particles to the mass concentration of the atmospheric aerosol. *Environ. Int.* 87: 108–115.
- Porter, K.G. and Feig, Y.S. (1980). The use of DAPI for identifying and counting aquatic microflora. *Limnol. Oceanogr.* 25: 943–948.
- Prussin, A.J. II, Garcia, E.B. and Marr, L.C. (2015). Total concentrations of virus and bacteria in indoor and outdoor air. *Environ. Sci. Technol. Lett.* 2: 84–88.
- Rathnayake, C.M., Metwali, N., Jayarathne, T., Kettler, J., Huang, Y., Thorne, P.S., O'Shaughnessy, P.T. and Stone, E.A. (2017). Influence of rain on the abundance of bioaerosols in fine and coarse particles. *Atmos. Chem. Phys.* 17: 2459–2475.
- Rosenfeld, D., Lohmann, U., Raga, G.B., O'Dowd, C.D., Kulmala, M., Fuzzi, S., Reissell, A. and Andreae, M.O. (2008). Flood or drought: How do aerosols affect precipitation? *Science* 321: 1309–1311.
- Saari, S., Niemi, J.V., Rönkkö, T., Kuuluvainen, H., Järvinen, A., Pirjola, L., Aurela, M., Hillamo, R. and Keskinen, J. (2015). Seasonal and diurnal variations of fluorescent bioaerosol concentration and size distribution in the urban environment. *Aerosol Air Qual. Res.* 15: 572–581.
- Seo, I., Jha, B.K., Lim, J.G., Suh, S.I., Suh, M.H. and Baek, W.K. (2014). Identification of lysosomotropic compounds based on the distribution and size of lysosomes. *Biochem. Biophys. Res. Commun.* 450: 189–194.
- Tong, Y. and Lighthart, B. (1999). Diurnal distribution of total and culturable atmospheric bacteria at a rural site. *Aerosol Sci. Technol.* 30: 246–254.
- Watson, J.G., Chen, L.W.A., Chow, J.C., Doraiswamy, P. and Lowenthal, D.H. (2008). Source apportionment: Findings from the US supersites program. *J. Air Waste Manage. Assoc.* 58: 265–288.
- Watson, J.G., Chow, J.C., Chen, L.W.A., Engling, G. and Wang, X.L. (2016). Source apportionment: Principles and methods. In *Airborne particulate matter: Sources, atmospheric processes and health*, Harrison, R.M., Hester, R.E. and Querol, X. (Eds.), The Royal Society of Chemistry, London, UK, pp. 72–125.
- Wei, K., Zou, Z.L., Zheng, Y.H., Li, J., Shen, F.X., Wu, C.Y., Wu, Y.S., Hu, M. and Yao, M.S. (2016). Ambient bioaerosol particle dynamics observed during haze and sunny days in Beijing. *Sci. Total Environ.* 550: 751–759.
- Xie, C.Q., Shen, F.X. and Yao, M.S. (2011). A novel method for measuring the charge distribution of airborne microbes. *Aerobiologia* 27: 135–145.

- Xie, Z.S., Li, Y.P., Lu, R., Li, W.X., Fan, C.L., Liu, P.X., Wang, J.L. and Wang, W.K. (2018). Characteristics of total airborne microbes at various air quality levels. *J. Aerosol Sci.* 116: 57–65.
- Yeo, H.G. and Kim, J.H. (2002). SPM and fungal spores in the ambient air of west Korea during the Asian dust (Yellow sand) period. *Atmos. Environ.* 36: 5437–5442.
- Zack, G.W., Rogers, W.E. and Latt, S.A. (1977). Automatic measurement of sister chromatid exchange frequency. *J. Histochem. Cytochem.* 25: 741–753.
- Zhai, Y., Li, X., Wang, T., Wang, B., Li, C. and Zeng, G. (2018). A review on airborne microorganisms in particulate matters: Composition, characteristics and influence factors. *Environ. Int.* 113: 74–90.
- Zhan, C.L., Zhang, J.Q., Zheng, J.R., Yao, R.Z., Wang, P., Liu, H.X., Xiao, W.S., Liu, X.L. and Cao, J.J. (2017). Characterization of carbonaceous fractions in PM_{2.5} and PM₁₀ over a typical industrial city in central China. *Environ. Sci. Pollut. Res.* doi: 10.1007/s11356-017-9970-9.

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