

Bioengineering of metal-organic frameworks for nanomedicine

Liu, Yuan; Zhao, Yanli; Chen, Xiaoyuan

2019

Liu, Y., Zhao, Y., & Chen, X. (2019). Bioengineering of Metal-organic Frameworks for Nanomedicine. *Theranostics*, 9(11), 3122-3133. doi:10.7150/thno.31918

<https://hdl.handle.net/10356/89741>

<https://doi.org/10.7150/thno.31918>

© 2019 Ivyspring International Publisher. This is an open access article distributed under the terms of the Creative Commons Attribution (CC BY-NC) license (<https://creativecommons.org/licenses/by-nc/4.0/>). See <http://ivyspring.com/terms> for full terms and conditions.

Downloaded on 28 Aug 2022 08:02:25 SGT

Review

Bioengineering of Metal-organic Frameworks for Nanomedicine

Yuan Liu¹, Yanli Zhao²✉, Xiaoyuan Chen¹✉

1. Laboratory of Molecular Imaging and Nanomedicine, National Institute of Biomedical Imaging and Bioengineering, National Institutes of Health, Bethesda, Maryland 20892, United States
2. Division of Chemistry and Biological Chemistry, School of Physical and Mathematical Sciences, Nanyang Technological University, Singapore, 637371, Singapore

✉ Corresponding authors: zhaoyanli@ntu.edu.sg; shawn.chen@nih.gov

© Ivyspring International Publisher. This is an open access article distributed under the terms of the Creative Commons Attribution (CC BY-NC) license (<https://creativecommons.org/licenses/by-nc/4.0/>). See <http://ivyspring.com/terms> for full terms and conditions.

Received: 2018.12.01; Accepted: 2019.03.22; Published: 2019.05.18

Abstract

Controlled structure, tunable porosity, and readily chemical functionalizability make metal-organic frameworks (MOFs) a powerful biomedical tool. Nanoscale MOF particles have been increasingly studied as drug carriers, bioimaging agents, and therapeutic agents due to their excellent physicochemical properties. In this review, we start with MOF as a nanocarrier for drug delivery, covering therapeutic MOF agents followed by a comprehensive discussion of surface bioengineering of MOF for improved biostability, biocompatibility, and targeted delivery. Finally, we detail the challenges and prospects of the future of MOF research for biomedical applications.

Key words: Metal-organic frameworks, drug delivery, photodynamic therapy, surface engineering

Introduction

Metal-organic frameworks (MOFs), assembled from metal ions and organic linkers *via* coordination chemistry, have wide potentials in catalysis [1-5], energy [6-8], and biomedical applications [9, 10]. As a new type of porous crystalline material, MOF building blocks can themselves be functional, which is different from the other nanomaterials. In particular, the tunable porosity, controlled structure, and readily chemical functionalizability of MOFs make them good examples as nanocarriers in biomedical applications [11]. From bulk phase to nanoscale phase, the discovery of abundant applicable properties of MOFs has led to new applications in biomedicine, especially at nanoscale size. During the past few years, preparation of various uniform nanoscale MOFs has provided a significant platform to explore structure-orientated functions of MOFs [12]. From nanocarriers to nanocargoes, MOFs have been able to make themselves a functional entity by controlling their assembling units. As a consequence, multifunctional MOFs have been extensively studied *via* direct synthesis or post-synthesis modification for

biomedical applications. With a permanently porous structure, fluorescent dyes, small drug molecules, and even protein can be loaded into MOFs for targeted imaging and delivery by tuning the pore sizes [13]. Synergistic therapy is believed to be a promising way to enhance tumor therapy efficacy. On-demand drug delivery, such as immunotherapy by loading immune checkpoint inhibitors, photodynamic therapy by conjugating photosensitizer, and photothermal therapy by combining with photothermal agents, and radio therapy [14-18] has been demonstrated to significantly enhance the therapeutic outcomes.

Recently, efforts have been devoted to demonstrating that nanoscale MOFs have great potential in preclinical applications. The goal of this review is to provide an overview of surface functionalization of MOFs for nanomedicine and cancer therapy. Here, we shall highlight the recent progress of MOF as a theranostic platform, including drug delivery, bioimaging, and smart MOF-based nanomedicine for enhanced tumor therapy. In contrast to other interesting reviews which cover a

comprehensive survey of all MOF nanoparticles [9, 10, 19, 20], we highlight the surface modification-based biofunctionalization approaches of nanoscale MOFs. Factors that affect the drug delivery in terms of loading efficiency and stimulus-responsive release of the drugs will be discussed. In particular, the challenges and perspectives of MOFs to realize targeted delivery, enhanced therapeutics, and final clinical translation will also be discussed.

MOF loading with small molecules and proteins

Although various types of MOFs have been reported, MOFs that have nanoscale size showed significant potential in tumor therapy applications [16, 21-24]. The most popular MOF therapeutic agents are Zr-based MOF series, porphyrinic MOF series, zeolitic imidazolate frameworks (ZIF) series, and Fe-based MOF series which typically have excellent aqueous stability. Merits of MOF can be concluded as follows: (1) Permanent porous crystal structure. Compared with traditional inorganic colloidal nanoparticles which usually carry cargo *via* covalent or noncovalent surface conjugation, MOFs have a much higher cargo loading efficiency due to their porous structure. In addition, cargo loading can be realized directly either through a one-pot synthesis or post-synthesis diffusion. (2) Tunable size of the pores. The framework originates from the coordination of building units metal ions and organic linkers. The length of the organic linker and the way of coordination determine the size of the pore. Basically, the longer the linker, the larger the size of the pore. The loading cargo can range from small molecules to proteins. (3) High multifunctional efficiency. With a minimized functional units and short processing steps, MOFs can realize much higher functional efficiency than other traditional nanomaterials.

Due to their facile production at low cost, MOFs are attracting many researchers to explore their novel biochemical properties for nanomedical applications [25]. Typically, Zr-based MOF nanoparticles can be obtained by mixing a certain ratio of Zr source and organic linker in DMF and incubated for several hours at slightly elevated temperature [22]. Compared with the synthesis of traditional inorganic colloidal nanoparticles, which requires hydrophobic organic solvents and high temperature to achieve good quality [26-29], the preparation of nanoscale MOFs usually does not need ultrahigh temperature or tedious organic synthesis. With this benefit of preparation, one can easily make various MOF nanoparticles for further biochemical studies.

Early biomedical studies of MOF mainly focused on drug delivery using MOF as a carrier [13]. Drug

delivery efficiency is a key factor for improving therapeutic effects [30]. Most drug molecules are hydrophobic and cannot be delivered to the physiological environment directly. Conventionally, bioconjugation of the hydrophobic drugs to inorganic nanomaterials was studied as a major way for targeted delivery [31-34]. Nanocarriers such as polymer micelles [35-37] and liposomes [38-41], which have a higher delivery efficiency than inorganic bioconjugation techniques, were also developed for drug delivery. Both nanomaterial-based bioconjugation and liposome carriers rely on enhanced permeability and retention effects to deliver drug molecules to the target tissue [42-44]. For example, common organic linkers such as carboxylic acid, amine, and thiol have been applied to modify the surface of inorganic colloidal nanoparticles for further surface engineering through 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC)/ *N*-hydroxysuccinimide (NHS), NHS ester, and thiol-ene reactions. These crosslinkers provided very convenient platforms for conjugation of small molecules, polymers, peptides, enzymes, and proteins. Hollow liposome with membrane structure similar to cells with relative higher loading ability than inorganic nanoparticle conjugation is a very nice carrier. However, the stability of inorganic nanomaterials and liposomes is a very significant obstacle limiting the therapeutic efficacy. Poor colloidal stability of inorganic nanomaterials with a large size typically creates serious aggregation under physiological conditions and mostly accumulated in lung and liver thus lowering delivery efficiency. The phosphor lipid structure of liposomes with relatively high loading capability also have low physiological stability and can be easily diffused to another cell with similar membrane structure during circulation. So, MOF was believed to be a promising drug carrier when nanoscale MOF appeared.

Previously, iron carboxylate MOFs have been demonstrated as biocompatible, degradable, and flexible drug carriers that can deliver various drugs that are not easily loaded using existing nanocarriers [13, 45, 46]. As shown in Figure 1, the flexibility of iron carboxylate MOFs offered the opportunity to encapsulate not only the drugs from small molecules to relatively big molecules such as doxorubicin, but also the drugs from hydrophobic to hydrophilic. The biocompatibility and degradability of iron carboxylate MOFs solved the side effect issues that most other nanocarriers have. Besides single drug delivery, MOFs also provide a platform for co-delivery to enhance therapeutic efficacy through a synergistic effect [30, 47]. Due to its porous structure, cargos such as small drug molecules and fluorescent dyes can be

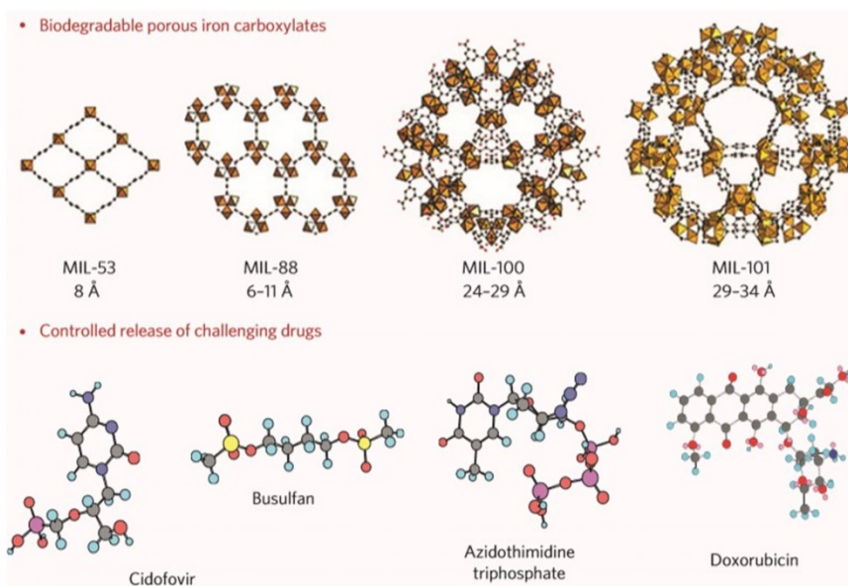


Figure 1. Porous iron carboxylate MOFs for dry delivery. Reprinted with permission from ref. [13]. Copyright (2010) Nature Publishing Group.

loaded into MOF structure on one hand. On the other hand, surface conjugation provides another opportunity to carry cargos of interest for a synergistic effect. For example, cisplatin prodrug and siRNA were co-loaded to an Universitetet i Oslo (UiO) MOF nanoparticle. A 12 wt % loading capability of cisplatin prodrug was achieved. SiRNA was bound to the surface of UiO MOF nanoparticles through multiple coordination between the phosphate backbone of SiRNA and Zr sites at the surface of MOF. This resulting co-delivery of cisplatin prodrug and siRNA significantly enhanced the *in vitro* chemotherapeutic efficacy.

Drug loading *via* physical adsorption by immersing the prepared MOF nanocarriers into cargo-containing solutions typically apply to the case when the size of cargo is smaller than the size of the pore of MOF nanoparticles. In other words, the size of pore determines whether the guest molecule can gain access to the pore of the MOF or not. Basically, the pore size of MOF and the size of the loading molecule have to be known. Physical adsorption on the surface of MOF may be obtained when the size of loading molecule is bigger than the pore size of MOF. To solve this size-dependent loading limitation, one-pot synthesis of cargo-loaded MOF nanoparticles has been developed [48-50]. For example, ZIFs have very small pores. Small molecules such as fluorescein and camptothecin cannot be diffused into the pore of ZIF nanoparticles and insert into the ZIF structure. With this facile one-pot synthesis, larger sized guest molecules that can be diffused into MOF can be encapsulated into the inner side of ZIF nanoparticles for efficient target delivery without premature release.

Compared with small molecule delivery, large molecule delivery such as peptides and proteins are encountering more challenges due to their size, surface charge, and component effects. First, direct conjugation of proteins to nanomaterials through covalent bonding typically yields a very low loading efficiency [34, 51-53]. Surface area determines the conjugation ability of inorganic colloidal nanoparticles. Second, the poor biological stability of protein-inorganic conjugates also significantly decreases the delivery efficiency. Third, the surface charge of the protein may hinder the cellular internalization. However, these three obstacles can be overcome by using MOF as a protein carrier [54, 55]. Physical adsorption by immersing the prepared MOF with large pore size into a guest protein-containing solution can significantly improve the protein loading efficiency. In addition, storing the protein inside the MOF structure can protect the protein from enzymatic degradation during transportation [56, 57]. In most cases, enzymes which have a large molecular weight and may decompose loading cargo in normal conditions will not be able to access loading cargo substrate when stored inside the porous MOF carrier. So, typically MOF can not only carry the loading cargos to target sites but also protect them from decomposing during transportation. Most importantly, the intracellular uptake of protein can be controlled by further surface modification of MOF, such as controlling the size and adjusting the surface charge. For example, insulin, the most important protein drug for the treatment of type I diabetes (Figure 2), cannot be directly applied by oral delivery because of extremely poor bioavailability and a low diffusion rate through the mucus layer. In the stomach acid environment, free insulin can be denatured by strong acid and digested by pepsin. However, when using MOF as a carrier for oral delivery of insulin, ultra-stable MOF in stomach acid environment can maintain the integrity of insulin while simultaneously excluding pepsin from getting access to the insulin, thus limiting its proteolysis [56]. This MOF carrier for insulin delivery provided insights to guide future protein and enzyme delivery. Further surface modification such as targeting molecule and aptamer may help realize a targeted delivery.

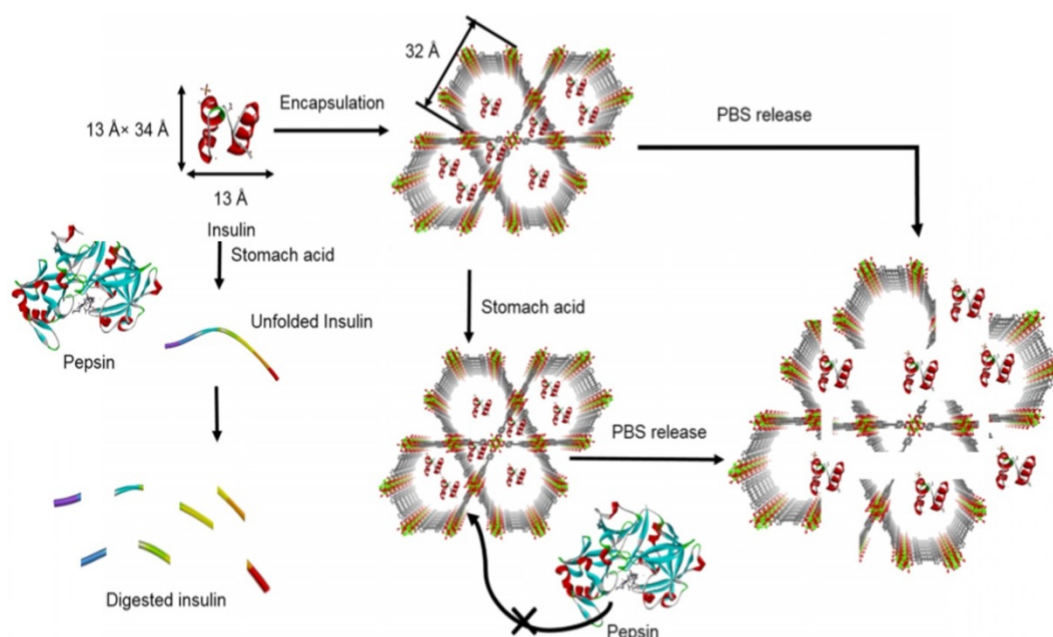


Figure 2. Free insulin loading to MOF NU-1000 and releasing in the presence of phosphate buffer saline. Pepsin which can digest insulin cannot access to the insulin that was stored in the porous MOF NU-1000 because of the large size of pepsin. Reprinted (adapted) with permission from ref. [56]. Copyright (2018) American Chemical Society.

Recently, MOFs with interconnected hierarchical mesoporous channels have been created as enzyme carriers for cell-free synthetic biology [58, 59]. Lactate dehydrogenase was encapsulated in the large pores of MOFs to access nicotinamide adenine dinucleotide coenzymes for an *in situ* coenzyme regeneration. Although enzymes or proteins can be absorbed into the porous MOF and the pore size of MOF can be controlled by adjusting the length of organic linkers, tedious synthesis of organic linkers for MOF has limited the preparation of MOF with large pore size to load proteins with high molecular weight. One-pot synthesis has been demonstrated to be able to encapsulate small molecules to ZIF and its small pore size. It can also be generalized for protein encapsulation [60].

Recently, the Willner group encapsulated both insulin and glucose oxidase into ZIF-8 nanoparticles to construct a smart sense-and-treat carrier [61]. In this smart sense-and-treat carrier, both insulin and glucose oxidase were encapsulated into ZIF-8 MOF particles. Insulin which can lower the blood glucose level has been applied to the treatment of type I diabetes. However, the usage of insulin may lead to hypoglycemia when using not properly. With glucose oxidase as a sensor, it can convert glucose to gluconic acid and lower the pH of the local environment. As a pH-sensitive carrier, lower pH can decompose the ZIF-8 carrier, thus releasing the loaded insulin to balance the blood glucose level. On the other hand, the lower blood glucose level balanced by insulin can also balance the pH of the local environment, thus balancing the release of insulin. So, this smart

glucose-responsive insulin release has the potential to decrease the risk of hypoglycemia.

Biocatalytic cascades driven by multienzyme-encapsulated MOFs *via* one-pot synthesis was also reported [62]. A model with three different enzymes β -galactosidase, glucose oxidase, and horseradish peroxidase was loaded into ZIF-8 nanoparticles. In the first step, β -galactosidase can convert lactose to glucose to provide a substrate for the second step. Subsequently, glucose oxidase can convert glucose and oxygen to gluconic acid and hydrogen peroxide which is a substrate for the third step. Finally, horseradish peroxidase can take advantage of hydrogen peroxide to convert amplex red to resorufin which is a fluorescent signal. Compared with the mixture of enzymatic catalysts in solution, a significant enhancement of catalytic cascades activity was obtained with this multienzyme-integrated MOF.

MOFs as smart drug carriers

Although the high loading capacity, low cytotoxicity, and effective cell and tissue permeation make MOFs excellent drug carriers, one of the significant issues of MOF as nanocarrier is the premature drug release. To solve this issue, stimuli-responsive drug release strategies have been designed. Typical stimuli such as pH, glutathione (GSH), ATP, and enzyme have been studied for controlled drug release [63-67]. An acidic environment in tumor tissue makes pH one of the most widely investigated stimulus for targeted and controlled drug release. ZIF-8 takes advantage of the pH sensitivity to realize a pH-responsive drug

delivery. In addition, other “smart” designs to lower the local pH and stimulate the drug release also have been reported [61]. For example, glucose oxidase and insulin integrated ZIF-8 can be triggered by glucose for insulin delivery. Nucleic acid with acidic pH sensitivity was modified on the MOF nanoparticles surface as a “lock” to control the drug release [68]. In a neutral pH environment, nucleic acids lock the drug inside the porous MOF nanoparticles. When the nucleic acid-modified MOF nanoparticles were transported to an acidic environment, such as pH=5.5, the nucleic acid would open the “lock” and slowly release the drug.

Compared with normal cells, the high intracellular concentration of GSH in cancer cells make GSH the second most important stimulus for controlled drug release [69-71]. Disulfide bond-containing molecules with GSH responsive properties have been widely studied not only in polymer-based drug delivery but also in inorganic nanoparticle-based drug release *via* surface functionalization [65]. In the case of MOF nanoparticles, disulfide bond-containing molecules or polymers were modified on the surface to block the premature drug release. Upon transporting to cancer cells with a high GSH level, the disulfide bond would be reduced and release the drug molecules. Using acidic pH and GSH as the stimuli-responsive drug delivery usually achieve targeted release and higher cancer therapy efficacy.

Adenosine triphosphate (ATP) is a very important complex organic chemical that living cells use it to provide energy to drive further many biological processes. Mitochondria, a double-layered membrane organelle in living cells is called an “energy factory” to generate energy. High concentration of ATP in the mitochondria of the cells also can contribute to stimuli-responsive drug delivery and intracellular imaging (Figure 3B) [64, 72, 73]. For example, encapsulation-leading fluorescence off of Rhodamine B in ZIF-90 provides a platform for intracellular ATP imaging based on the ATP triggered disassembly. The fluorescence of Rhodamine B was significantly suppressed after encapsulated into ZIF-90. However, the competitive coordination between the metal node of ZIF-90 and ATP can disassemble the structure of ZIF-90, thus releasing the Rhodamine B from the ZIF-90 nanoparticles. The dynamic images of mitochondria ATP in live cells have been observed through this stimuli-responsive system. Furthermore, an ATP-responsive ZIF-90 platform for cytosolic protein delivery and clustered regularly interspaced short palindromic repeats-associated protein 9 (CRISPR/Cas9) genome editing was developed using a similar system.

CRISPR/Cas9 has been demonstrated as a very promising genomic editing tool. With the one-pot synthesis method, the protein CRISPR/Cas9 was encapsulated into ZIF-90 without changing the function of CRISPR/Cas9. Upon delivering the CRISPR/Ca9 encapsulated ZIF-90, the high concentration of intracellular ATP will promote the disassembly of ZIF-90 to release CRISPR/Ca9. With this ATP-responsive delivery system, the genome editing protein CRISPR/Cas9 effectively knocked out the expression of the green fluorescent protein in HeLa cells. Furthermore, cytotoxic RNase A-encapsulated ZIF-90 significantly prohibited cancer cell growth.

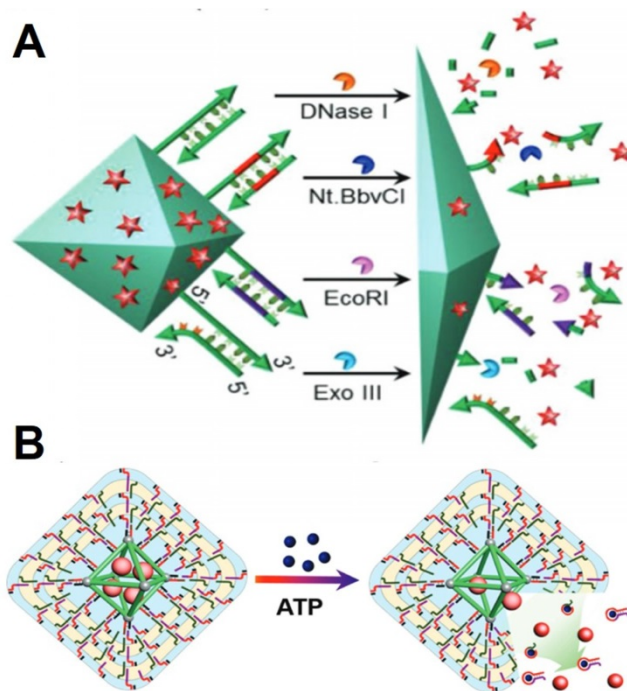


Figure 3. A) Cargo release from the duplex-capped MOF with different stimuli such as the DNase I, the nicking enzyme (Nt.BbvCI), the endonuclease (EcoRI), and the exonuclease (Exo III) as biocatalysts. Reprinted with permission from ref. [66] Copyright (2018) WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim. B) ATP triggered drug release. Reprinted with permission from ref. [63] Copyright (2017) WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.

The enzyme plays a very significant role in the balance of biological systems. Each enzyme has a specific function and can catalyze its substrate only under certain conditions. Oligonucleotides act as substrates for different enzymes, such as DNase I, endonuclease, and exonuclease III (Figure 3A) [66]. DNA functionalization on the surface of colloidal nanoparticles has been well studied. Both single stranded and double stranded DNA can be functionalized on the surface of MOFs for enzyme-responsive drug delivery. For example, the camptothecin-loaded and tailored hairpin DNA strands-capped MOF showed selective cytotoxicity toward MDA-MB-231 cancer cells that had a high

expression of exonuclease III. Low apoptosis to epithelial MCF-10A breast cells which has low expression of exonuclease III was also observed.

MOFs as photodynamic therapeutic agents

Photodynamic therapy, as a noninvasive treatment, has attracted tremendous interest owing to its fewer harmful side effects [74, 75]. However, the integration of photosensitizer to nanomaterials is often limited because of low loading efficiency, poor stability, and increased cytotoxicity. Recently, the preparation of photosensitizer-based MOF overcame the limitations of photosensitizers [76, 77], such as aggregation, self-quenching, and uncontrollable *in vivo* administration. With the uniform and well-defined porous crystalline structure, porphyrinic MOF allows $^3\text{O}_2$ and $^1\text{O}_2$ to diffuse freely in and out of the framework [77, 78]. In the past decade, various porphyrin and derivative linkers have been synthesized to prepare MOFs. The robust chemical structure and natural biological functions of porphyrins help preserve the functionality of porphyrins after coordinating with metal ions to form a MOF.

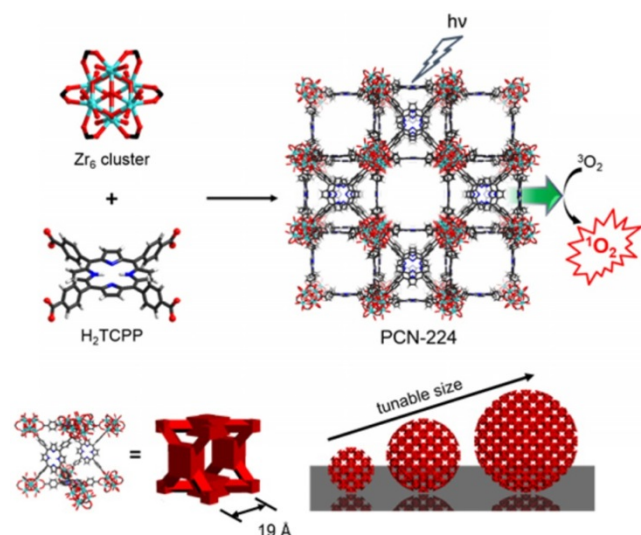


Figure 4. Size-controlled synthesis of porphyrinic Zr-MOF (PCN-224) for photodynamic therapy. Reprinted (adapted) with permission from ref. [76]. Copyright (2016) American Chemical Society.

In 2014, the first nanoscale MOF used for photodynamic therapy was reported by the Lin group [79]. A Hf-based MOF (Hf-MOF) nanoplate with 100 nm diameter and 10 nm thickness was prepared using the solvothermal method through coordination between Hf and 5,15-di(pbenzoato)-porphyrin. This Hf-MOF showed at least a two-fold increase in $^1\text{O}_2$ generation compared to free porphyrin. In 2016, the Zhou group developed a size-controlled synthesis of

Zr-based porphyrinic MOF (Zr-MOF) for targeted photodynamic therapy (Figure 4) [76]. A broad size range of Zr-MOFs with precise control was prepared for size-dependent cellular uptake and photodynamic therapy. Intracellular cytotoxicity studies indicated that the 90 nm Zr-MOF nanoparticles showed the best photodynamic therapy efficacy, suggesting a promising photodynamic therapy candidate. Later, various metal-based porphyrinic MOFs were prepared for synergistic tumor therapy, such as photodynamic-photothermal therapy, photodynamic-radio therapy, and photodynamic-immune therapy [70, 80-83].

Core-shell NP@MOF structure provides a multifunctional platform to extend the bioapplications of MOF in bioimaging, nanomedicine, and cancer therapy [83-87]. The Huo group overcame the challenge and successfully grew ZIF-8 on various colloidal nanoparticles. The structure of ZIF-8 did not change after encapsulating colloidal nanoparticles. However, collective properties such as photoluminescent, catalytic, and magnetic properties were obtained with the heterogenous MOF structures [87]. Photodynamic therapy using NIR light (980 nm) was achieved when upconversion nanoparticles @ MOF (UCNP@MOF) dimer was constructed through a fluorescence resonance energy transfer (FRET) strategy. The emission of UCNP with 980 nm light excitation at 650 nm can be adsorbed by porphyrinic MOF, thus generating toxic singlet oxygen for cancer cell therapy [83].

In the case of photodynamic-photothermal therapy, photothermal agent Au nanorods (AuNR) were used for photothermal therapy under the irradiation of NIR light [82]. Porphyrinic MOFs were used as singlet oxygen generator for photodynamic therapy. Nanoscale core-shell AuNR@MOF nanoparticles were prepared by growing a layer of porphyrinic MOF on the surface of Au nanorod. This core-shell AuNR@MOF provides a dual-therapy model for tumor inhibition. The synergistic function from NIR light 808 nm for Au nanorods to generate heat and 660 nm for porphyrinic MOF to generate singlet oxygen significantly enhanced the therapy efficacy both *in vitro* and *in vivo*.

Radiotherapy has been commonly applied to tumor therapy owing to its ability to control cancer cell growth. However, high dose of radiation typically causes a serious side effect. Heavy metals such as Au, Hf, and Ru are common radiosensitizers to enhance radiotherapy efficacy. For example, Au nanoparticles accumulated in the tumor site could enhance the radiotherapy [88]. Hf-based MOF has been demonstrated as an efficient agent for radiotherapy. The innovative combination of radiotherapy and

radiodynamic therapy also has been demonstrated to significantly suppress tumor growth with a low dose of radiation [89]. Recently, the Lin group developed a Hf-DBB-Ru [DBB-Ru = bis(2,2'-bipyridine) (5,5'-di(4-benzoato)-2,2'-bipyridine) ruthenium (II) chloride] nanoscale MOF for a combined radiotherapy and radiodynamic therapy (RT-RDT) [72]. With nanoscale Hf-DBB-Ru MOF as a carrier, both Hf and Ru were used as a radiosensitizer to enhance the radiotherapy efficacy. Upon irradiating with X-ray, hydroxyl radical and singlet oxygen can be generated by this Hf-DBB-Ru MOF nanoparticle. *In vitro* and *in vivo* study indicated that the mitochondria-targeted RT-RDT can depolarize the membrane of mitochondrial to initiate the apoptosis of cancer cells, thus significantly inhibit the tumor growth in mouse models.

Immunotherapy which activating or suppressing the immune system to treat cancers has attracted intensive interest in the past decades. Current immunotherapy methods such as non-specific immunotherapies, oncolytic virus therapy, monoclonal antibodies, and tumor-agnostic therapies, T-cell therapy, and cancer vaccines typically work by suppressing the cancer cells growth, stopping cancer cells from spreading, and helping the immune system to fight cancer cells. Recently, MOFs have been used to enhance checkpoint blockade immunotherapy [14, 17, 90, 91]. By incorporating radiosensitizers into MOF, enhanced radiotherapy was achieved to potentiate checkpoint blockade immunotherapy. In addition, combining anti-programmed death-ligand 1 antibody with MOF-mediated low-dose radiotherapy, the obvious abscopal effect was observed from a distant tumor. So, the local radiotherapy can trigger a local immune response by releasing immunostimulating signals to increase T cell infiltration to the tumor [92-94]. Later, combined low-dose X-ray radiotherapy and radiodynamic therapy using nanoscale MOF were also demonstrated to enhance the checkpoint blockade immunotherapy [14].

Surface engineering of MOFs

Surface functionalization of nanomaterials has always been very significant for biochemical applications, such as analytical detection, bioimaging, and cancer therapy [51, 52, 95]. The controlled manipulation of the external surface of MOFs to fit specific requirements and achieve the desired function is of paramount importance as it determines the overall performance of MOF nanoparticles [15, 96-98]. For example, PEGylation was typically used to improve the colloidal stability of inorganic nanoparticles [99-101]. Covalently anchoring a

fluorophore on the surface of the nanoparticles can be used for bioimaging. Surface functionalization of a targeting molecule, such as a peptide or aptamer, can realize target binding or targeted delivery [76, 81, 102]. Grafting functional polymers on the surface of the nanoparticles can achieve some stimuli-responsive properties. As a promising nanocarrier, surface functionalization of MOFs without changing their framework and porosity is also significant for the required biomedical applications. There are two popular post-synthesis incorporation ways to bioengineer the surface of MOFs [103-105]. Since MOFs are made of organic linkers and metal ions *via* coordination bonds, the first way is to modify an anchor on the organic linker before the synthesis of the MOF and then covalently conjugate the target molecule with the anchor on the surface of the as-prepared MOF [104, 106, 107]. The second method is to coordinate the target molecule on the surface of MOF directly in which the chelation between metal ions and target molecule acts as a bridge for the surface functionalization of the MOF [57, 105, 108].

The first example of anchor modification on an organic linker is the UiO-66 to UiO-66-N₃ nanoparticles. The organic linker of UiO-66, benzene-1,4-dicarboxylic acid does not have any anchor and the resulting UiO-66 cannot be functionalized through a covalent anchor [104]. In the case of UiO-66-N₃, the azide group in 2-azido-1,4-benzene dicarboxylic acid can react with the alkane group *via* click reactions, modifying the target molecule on the MOF surface. Any alkane terminal ligands can be functionalized on the surface of MOF with controlled loading through this click reaction. In addition, a dibenzylcyclooctyne terminal DNA sequence can be conjugated on the surface of UiO-66-N₃ nanoparticles.

Surface defects of nanoparticles are very common during synthesis. The unsaturated coordinative metal sites on the surface of MOFs provide opportunities for target molecules to bind to MOF nanoparticles through coordination. So far, different functional group-terminal ligands, such as carboxylate, phosphonate, histidine, and phenyl groups, have been reported to achieve incorporation [105, 106, 109]. As basic coordination, various metal-carboxylate bindings have been used to form different MOFs in an organic solvent. Naturally, carboxylate containing ligands can bind the unsaturated metal sites, thus functionalizing MOF. Binding affinity is a significant way to evaluate the binding strength between metal and ligand and it varies between different metals and ligands [105]. In the case of Zr-based MOFs, both carboxylate and

phosphonate can coordinate with Zr, thus capping the ligand on the surface of the Zr-based MOF. However, the binding affinity between Zr and phosphonate is stronger than that between Zr and carboxylate. The Farha group have demonstrated that both carboxylate- and phosphonate-terminal ligands can be incorporated on the surface of NU-1000 [94]. However, extra phosphonate-terminal ligands can decompose the structure of NU-1000, while carboxylate-terminal ligands cannot.

Histidine which can be readily integrated into proteins or peptides significantly extended the scope for targeting molecule-functionalized MOFs. The Lächelt group reported a coordinative incorporation of oligohistidine-tags with MOFs [109]. Despite different metal components, MIL-88A, HKUST-1, and Zr-*fum* exhibited considerable His-tag binding. Fluorescent models including His-carboxyfluorescein, His-green fluorescein protein, and His-ATTO 647N labeled human transferrin were selected to test the coordinative binding and cellular internalization using flow cytometry and confocal laser scanning microscopy. The His-tags binding to MOF demonstrated a general functionalization method of MOF with potential for protein and drug delivery. However, the limited histidine group in peptide and proteins may limit the application of this general surface functionalization strategy due to relatively weak binding affinity between histidine and the metal node of MOF nanoparticles.

Lipid coating to MOF is a facile method to functionalize MOF without changing its structural integrity and porosity [103, 110]. The lipid ligand 1,2-dioleoyl-sn-glycero-3-phosphate has been used to transfer MOF from aqueous phase to the organic phase by facile surface encapsulation. Similarly, bilayer lipid-coated MOF was developed to study the intracellular release of loading dye with 1,2-dioleoyl-sn-glycero-3-phosphocholine. This bilayer lipid not only effectively stores the dye molecules

inside the porous scaffold of MOF but also enables a high cellular uptake of MOF nanoparticles. Compared with artificial lipid layers, exosomes which have very similar membrane structures to cell membranes, are typically used for communication purposes by cells. They have the potential to form a protective coating on nanoparticles to bypass the immune system for longer circulation time for full biocompatibility. The Wuttke group overcame the challenge and successfully coated exosomes on the surface of MIL-88A using a fusion method [111]. A slow calcein release was observed with exosome-coated calcein-loaded MIL-88A nanoparticles in HeLa cells, indicating that exosome coating is a very promising drug delivery system. The combination of exosome and MOF solved the premature release issue and improved the biocompatibility of MOF nanoparticles.

Phenolic group-terminated ligand can also form a stable coordination to directly modify MOF nanoparticles [95]. Various metals such as Zr, Cr, Fe, Co, Cu, Zn, Al, In, and Eu have been demonstrated to chelate with phenolic group. The stable coordination can be attributed to a 5-member ring formed between metal ion and phenolic group. MOF nanoparticles including UiO-66, ZIF, HKUST, and MIL-101 were transferred to organic phase from aqueous phase with phenolic lipid. This phenolic group provides a versatile platform for MOF surface functionalization.

The three-dimensional oligonucleotides are of tremendous importance and have a wide application in biodetection, targeted binding, and genomic editing [112-116]. The study of chemical interface properties between MOFs and nucleic acids render a potential application of MOFs for analytical detection, bioimaging, drug delivery, and cancer therapy [96, 98]. A nucleic acid-MOF conjugate (Figure 5A) was constructed through a covalent click reaction with azido-anchored UiO-66 and dibenzylcyclooctyne functionalized DNA [104]. Because of the natural phosphate backbone structure of DNA,

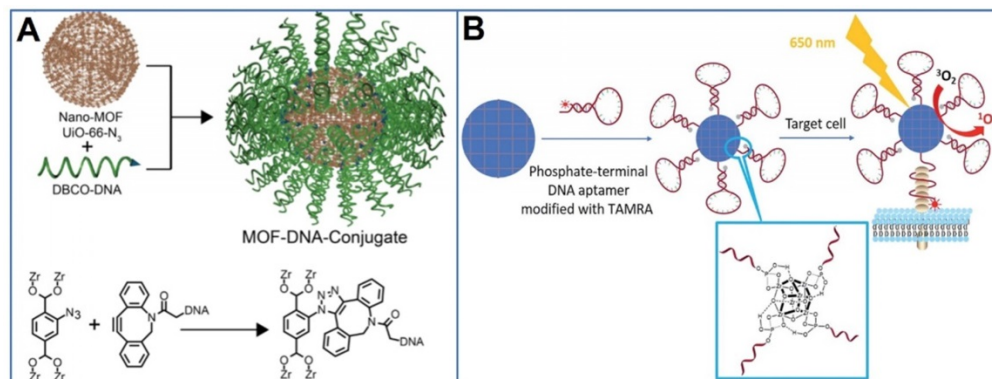


Figure 5. A) Nucleic acid-MOF conjugation through covalent click reaction. Reprinted (adapted) with permission from ref. [93]. Copyright (2014) American Chemical Society. B) Direct phosphate-terminal DNA conjugation to MOF. Reprinted (adapted) with permission from ref. [106]. Copyright (2018), Royal Society of Chemistry.

phosphate-terminal DNA was later found to be able to be directly integrated onto the surface of MOFs (Figure 5B) [108, 117]. With DNA capping on the surface, the colloidal stability and biocompatibility of MOF nanoparticles have been significantly improved. Aptamers are RNA and DNA oligonucleotides that bind their targets with high affinity and selectivity [118]. Targeted imaging and drug delivery were achieved by incorporating aptamers onto the surface of MOF nanoparticles.

Challenges and perspectives

The well-defined porous crystalline MOF has been demonstrated as a promising platform for drug delivery, bioimaging, and tumor therapy [16, 20, 24]. The preparation of various nanoscale MOF particles with facile cargo loading renders a wide range of biomedical applications. Surface engineering of MOFs for targeted stimuli-responsive drug delivery significantly enhances the tumor therapy efficacy. Despite the considerable progress, biomedical applications of MOFs still face many challenges.

First, the poor stability of MOFs in physiological conditions has significantly limited its biomedical applications. Zn-carboxylate MOFs are very unstable in aqueous solution because of low coordinative affinity. Zr-based MOF nanoparticles are very sensitive to phosphate containing buffers such as PBS and RPMI cell culture medium which have a high concentration of phosphate ion owing to a stronger binding affinity between phosphate ion and Zr ion [56-57]. Colloidal stability in aqueous solution due to large size (100 ~ 500 nm) of MOF also should be improved by surface functionalization. PEGylation or other hydrophilic ligand encapsulation is necessary to improve the colloidal stability of MOF nanoparticles for physiological studies [88-90]. Without solving the biostability of MOFs under physiological condition, any other biomedical applications of MOFs will be futile.

Second, therapeutic proteins can be exploited to produce potentially highly specific drugs, thus curing the disease without the conventional drugs [54]. The delivery of proteins without disrupting its bioavailability and activity depends on the delivery methods, and are affected by size, surface charge, and hydrophilicity [55]. Porous MOFs typically have pore/channel size about 1 to 3 nm. Small molecules and peptide/protein with a small molecular weight (< 7 kD) do not have a problem being loaded into the MOF particles [56-57]. However, the proteins with a large molecular weight (> 10 kD) typically need large pores/channels in order to be loaded into the MOF. Although the MOF particles with large pore/channel size need tedious work to synthesize their organic

linkers, it is worth it for developing MOF particle systems with large pore/channel size for therapeutic protein delivery.

The final biomedical goal of MOF nanoparticles is a clinical application. The side effects or toxicity of MOF nanoparticles is the most significant factor to determine whether MOF nanoparticles can be applied to clinical research or not. Toxicity of nanomaterials is concentration dependent. Coordinating unit metal ions or organic linkers with minimal toxicity should be considered when constructing MOF particles as drug carriers or therapeutic agents [13]. Controlled structure, tunable porosity, and readily chemical functionalizability make MOF a powerful biomedical tool for us to take advantage for biomedical applications. Bioengineering of MOF for nanomedicine is an interdisciplinary study. Future continued efforts need to focus on the biostability, biocompatibility, practicability, and efficacy to realize the full clinical applications of MOFs.

Acknowledgements

This work supported by the intramural research program of the National Institute of Biomedical Imaging and Bioengineering (NIBIB), National Institutes of Health (NIH).

Competing Interests

The authors have declared that no competing interest exists.

References

1. Guo Z, Xiao C, Maligal-Ganesh RV, Zhou L, Goh TW, Li X, et al. Pt nanoclusters confined within metal-organic framework cavities for chemoselective cinnamaldehyde hydrogenation. *ACS Catal.* 2014; 4: 1340-8.
2. Li Z, Peters AW, Platero-Prats AE, Liu J, Kung C-W, Noh H, et al. Fine-Tuning the Activity of Metal-Organic Framework-Supported Cobalt Catalysts for the Oxidative Dehydrogenation of Propane. *J Am Chem Soc.* 2017; 139: 15251-8.
3. Zhao M, Yuan K, Wang Y, Li G, Guo J, Gu L, et al. Metal-organic frameworks as selectivity regulators for hydrogenation reactions. *Nature.* 2016; 539: 76.
4. Li P, Moon S-Y, Guelta MA, Lin L, Gómez-Gualdrón DA, Snurr RQ, et al. Nanosizing a metal-organic framework enzyme carrier for accelerating nerve agent hydrolysis. *ACS Nano.* 2016; 10: 9174-82.
5. Shi W, Zhao X, Feng J, Liu J, Yang G, Wang G, et al. An Efficient, Visible-Light-Driven, Hydrogen Evolution Catalyst NiS/ZnxCd1-xS Nanocrystal Derived from a Metal-Organic Framework. *Angew Chem Int Ed Engl.* 2018; 57: 9790-4.
6. Furukawa H, Cordova KE, O'Keeffe M, Yaghi OM. The chemistry and applications of metal-organic frameworks. *Science.* 2013; 341: 1230444.
7. Schoedel A, Ji Z, Yaghi OM. The role of metal-organic frameworks in a carbon-neutral energy cycle. *Nat Energy.* 2016; 1: 16034.
8. Yuan D, Zhao D, Sun D, Zhou HC. An Isoreticular Series of Metal-Organic Frameworks with Dendritic Hexacarboxylate Ligands and Exceptionally High Gas-Uptake Capacity. *Angew Chem Int Ed Engl.* 2010; 122: 5485-9.
9. Horcajada P, Gref R, Baati T, Allan PK, Maurin G, Couvreur P, et al. Metal-organic frameworks in biomedicine. *Chem Rev.* 2011; 112: 1232-68.
10. Simon-Yarza T, Mielcarek A, Couvreur P, Serre C. Nanoparticles of Metal-Organic Frameworks: On the Road to In Vivo Efficacy in Biomedicine. *Adv Mater.* 2018; 30: 1707365.

11. Freund R, Lächelt U, Gruber T, Rühle B, Wuttke S. Multifunctional efficiency: extending the concept of atom economy to functional nanomaterials. *ACS Nano*. 2018; 12: 2094-105.
12. Cai W, Chu CC, Liu G, Wang YXJ. Metal-organic framework-based nanomedicine platforms for drug delivery and molecular imaging. *Small*. 2015; 11: 4806-22.
13. Horcajada P, Chalati T, Serre C, Gillet B, Sebrie C, Baati T, et al. Porous metal-organic-framework nanoscale carriers as a potential platform for drug delivery and imaging. *Nat Mater*. 2010; 9: 172.
14. Lu K, He C, Guo N, Chan C, Ni K, Lan G, et al. Low-dose X-ray radiotherapy-radiodynamic therapy via nanoscale metal-organic frameworks enhances checkpoint blockade immunotherapy. *Nat Biomed Engineering*. 2018; 2: 600-10.
15. Cai W, Gao H, Chu C, Wang X, Wang J, Zhang P, et al. Engineering Phototheranostic Nanoscale Metal-Organic Frameworks for Multimodal Imaging-Guided Cancer Therapy. *ACS Appl Mater Interfaces*. 2017; 9: 2040-51.
16. He Z, Dai Y, Li X, Guo D, Liu Y, Huang X, et al. Hybrid Nanomedicine Fabricated from Photosensitizer-Terminated Metal-Organic Framework Nanoparticles for Photodynamic Therapy and Hypoxia-Activated Cascade Chemotherapy. *Small*. 2019; 15: 1804131.
17. Lan G, Ni K, Xu Z, Veroneau SS, Song Y, Lin W. Nanoscale Metal-Organic Framework Overcomes Hypoxia for Photodynamic Therapy Primed Cancer Immunotherapy. *J Am Chem Soc*. 2018; 140: 5670-3.
18. Liu J, Yang Y, Zhu W, Yi X, Dong Z, Xu X, et al. Nanoscale metal-organic frameworks for combined photodynamic & radiation therapy in cancer treatment. *Biomaterials*. 2016; 97: 1-9.
19. Wuttke S, Lismont M, Escudero A, Rungtaweeworant B, Parak WJ. Positioning metal-organic framework nanoparticles within the context of drug delivery—a comparison with mesoporous silica nanoparticles and dendrimers. *Biomaterials*. 2017; 123: 172-83.
20. Wu MX, Yang YW. Metal - Organic Framework (MOF)-Based Drug/Cargo Delivery and Cancer Therapy. *Adv Mater*. 2017; 29: 1606134.
21. Wang S, McGuirk CM, d'Aquino A, Mason JA, Mirkin CA. Metal-Organic Framework Nanoparticles. *Adv Mater*. 2018: 1800202.
22. Wang XG, Cheng Q, Yu Y, Zhang XZ. Controlled Nucleation and Controlled Growth for Size Predictable Synthesis of Nanoscale Metal-Organic Frameworks (MOFs): A General and Scalable Approach. *Angew Chem Int Ed Engl*. 2018 ;57: 7836-40.
23. Li B, Wang X, Chen L, Zhou Y, Dang W, Chang J, et al. Ultrathin Cu-TCPP MOF nanosheets: a new theragnostic nanoplatform with magnetic resonance/near-infrared thermal imaging for synergistic phototherapy of cancers. *Theranostics*. 2018; 8: 4086-96.
24. Wang D, Zhou J, Shi R, Wu H, Chen R, Duan B, et al. Biodegradable Core-shell Dual-Metal-Organic-Frameworks Nanotheranostic Agent for Multiple Imaging Guided Combination Cancer Therapy. *Theranostics*. 2017; 7: 4605-17.
25. Sindoro M, Yanai N, Jee A-Y, Granick S. Colloidal-sized metal-organic frameworks: synthesis and applications. *Acc Chem Res*. 2013; 47: 459-69.
26. Liu Y, Hou W, Sun H, Cui C, Zhang L, Jiang Y, et al. Thiol-ene click chemistry: a biocompatible way for orthogonal bioconjugation of colloidal nanoparticles. *Chem Sci*. 2017; 8: 6182-7.
27. Liu Y, Purich DL, Wu C, Wu Y, Chen T, Cui C, et al. Ionic functionalization of hydrophobic colloidal nanoparticles to form ionic nanoparticles with enzymelike properties. *J Am Chem Soc*. 2015; 137: 14952-8.
28. Lynch J, Zhuang J, Wang T, LaMontagne D, Wu H, Cao YC. Gas-bubble effects on the formation of colloidal iron oxide nanocrystals. *J Am Chem Soc*. 2011; 133: 12664-74.
29. Park J, An K, Hwang Y, Park J-G, Noh H-J, Kim J-Y, et al. Ultra-large-scale syntheses of monodisperse nanocrystals. *Nat Mater*. 2004; 3: 891-5.
30. Chen Q, Xu M, Zheng W, Xu T, Deng H, Liu J. Se/Ru-Decorated Porous Metal-Organic Framework Nanoparticles for The Delivery of Pooled siRNAs to Reversing Multidrug Resistance in Taxol-Resistant Breast Cancer Cells. *ACS App Mater Interfaces* 2017; 9: 6712-24.
31. Biju V. Chemical modifications and bioconjugate reactions of nanomaterials for sensing, imaging, drug delivery and therapy. *Chem Soc Rev*. 2014; 43: 744-64.
32. Chen H, Zhang W, Zhu G, Xie J, Chen X. Rethinking cancer nanotheranostics. *Nat Rev Mater*. 2017; 2: 17024.
33. Howes PD, Chandrawati R, Stevens MM. Colloidal nanoparticles as advanced biological sensors. *Science*. 2014; 346: 1247390.
34. Liu Y, Chen T, Wu C, Qiu L, Hu R, Li J, et al. Facile surface functionalization of hydrophobic magnetic nanoparticles. *J Am Chem Soc*. 2014; 136: 12552-5.
35. Ahmad Z, Shah A, Siddiq M, Kraatz H-B. Polymeric micelles as drug delivery vehicles. *RSC Adv*. 2014; 4: 17028-38.
36. Zhang Y, Huang Y, Li S. Polymeric micelles: nanocarriers for cancer-targeted drug delivery. *AAAPS PharmSciTech*. 2014; 15: 862-71.
37. Movassaghian S, Merkel OM, Torchilin VP. Applications of polymer micelles for imaging and drug delivery. *Wiley Interdiscip Rev Nanomed Nanobiotechnol*. 2015; 7: 691-707.
38. Fouladi F, Steffen KJ, Mallik S. Enzyme-responsive liposomes for the delivery of anticancer drugs. *Bioconjug Chem*. 2017; 28: 857-68.
39. Nguyen TX, Huang L, Gauthier M, Yang G, Wang Q. Recent advances in liposome surface modification for oral drug delivery. *Nanomedicine*. 2016; 11: 1169-85.
40. Rengan AK, Bukhari AB, Pradhan A, Malhotra R, Banerjee R, Srivastava R, et al. In vivo analysis of biodegradable liposome gold nanoparticles as efficient agents for photothermal therapy of cancer. *Nano Lett*. 2015; 15: 842-8.
41. Sercombe L, Veerati T, Moheimani F, Wu SY, Sood AK, Hua S. Advances and challenges of liposome assisted drug delivery. *Front Pharmacol*. 2015; 6: 286.
42. Chertok B, Moffat BA, David AE, Yu F, Bergemann C, Ross BD, et al. Iron oxide nanoparticles as a drug delivery vehicle for MRI monitored magnetic targeting of brain tumors. *Biomaterials*. 2008; 29: 487-96.
43. Fang J, Nakamura H, Maeda H. The EPR effect: unique features of tumor blood vessels for drug delivery, factors involved, and limitations and augmentation of the effect. *Adv Drug Deliv Rev*. 2011; 63: 136-51.
44. Yang Z, Tian R, Wu J, Fan Q, Yung BC, Niu G, et al. Impact of semiconducting perylene diimide nanoparticle size on lymph node mapping and cancer imaging. *ACS Nano*. 2017; 11: 4247-55.
45. Baati T, Njim L, Neffati F, Kerkeni A, Bouttemi M, Gref R, et al. In depth analysis of the in vivo toxicity of nanoparticles of porous iron (III) metal-organic frameworks. *Chem Sci*. 2013; 4: 1597-607.
46. Simon-Yarza T, Baati T, Neffati F, Njim L, Couvreur P, Serre C, et al. In vivo behavior of MIL-100 nanoparticles at early times after intravenous administration. *Int J Pharm*. 2016; 511: 1042-7.
47. He C, Lu K, Liu D, Lin W. Nanoscale metal-organic frameworks for the co-delivery of cisplatin and pooled siRNAs to enhance therapeutic efficacy in drug-resistant ovarian cancer cells. *J Am Chem Soc*. 2014; 136: 5181-4.
48. Zheng H, Zhang Y, Liu L, Wan W, Guo P, Nyström AM, et al. One-pot synthesis of metal-organic frameworks with encapsulated target molecules and their applications for controlled drug delivery. *J Am Chem Soc*. 2016; 138: 962-8.
49. Zhuang J, Kuo C-H, Chou L-Y, Liu D-Y, Weerapana E, Tsung C-K. Optimized metal-organic-framework nanospheres for drug delivery: evaluation of small-molecule encapsulation. *ACS Nano*. 2014; 8: 2812-9.
50. Chen X, Tong R, Shi Z, Yang B, Liu H, Ding S, et al. MOF Nanoparticles with Encapsulated Autophagy Inhibitor in Controlled Drug Delivery System for Antitumor. *ACS Appl Mater Interfaces*. 2018; 10: 2328-37.
51. Sinha R, Kim GJ, Nie S, Shin DM. Nanotechnology in cancer therapeutics: bioconjugated nanoparticles for drug delivery. *Mol Cancer Ther*. 2006; 5: 1909-17.
52. Kango S, Kalia S, Celli A, Njuguna J, Habibi Y, Kumar R. Surface modification of inorganic nanoparticles for development of organic-inorganic nanocomposites—a review. *Prog in Polym Sci*. 2013; 38: 1232-61.
53. Xu ZP, Zeng QH, Lu GQ, Yu AB. Inorganic nanoparticles as carriers for efficient cellular delivery. *Chem Eng Sci*. 2006; 61: 1027-40.
54. Kato S, Otake K-i, Chen H, Akpınar I, Buru CT, Islamoglu T, et al. Zirconium-Based Metal-Organic Frameworks for the Removal of Protein-Bound Uremic Toxin from Human Serum Albumin. *J Am Chem Soc*. 2019; 141: 2568-76.
55. Lian X, Fang Y, Joseph E, Wang Q, Li J, Banerjee S, et al. Enzyme-MOF (metal-organic framework) composites. *Chem Soc Rev*. 2017; 46: 3386-401.
56. Chen Y, Li P, Modica JA, Drout RJ, Farha OK. Acid-Resistant Mesoporous Metal-Organic Framework toward Oral Insulin Delivery: Protein Encapsulation, Protection, and Release. *J Am Chem Soc*. 2018; 140: 5678-81.
57. Wang S, Chen Y, Wang S, Li P, Mirkin CA, Farha OK. DNA-Functionalized Metal-Organic Framework Nanoparticles for Intracellular Delivery of Proteins. *J Am Chem Soc*. 2019; 141: 2215-9.
58. Li P, Modica JA, Howarth AJ, Vargas E, Moghadam PZ, Snurr RQ, et al. Toward design rules for enzyme immobilization in hierarchical mesoporous metal-organic frameworks. *Chem*. 2016; 1: 154-69.
59. Li P, Chen Q, Wang TC, Vermeulen NA, Mehdi BL, Dohmalkova A, et al. Hierarchically Engineered Mesoporous Metal-Organic Frameworks toward Cell-free Immobilized Enzyme Systems. *Chem*. 2018; 4: 1022-34.

60. Duan Y, Ye F, Huang Y, Qin Y, He C, Zhao S. One-pot synthesis of a metal-organic framework-based drug carrier for intelligent glucose-responsive insulin delivery. *Chem Commun.* 2018; 54: 5377-80.
61. Chen W-H, Luo G-F, Vázquez-González M, Cazelles R, Sohn YS, Nechushtai R, et al. Glucose-Responsive Metal-Organic-Framework Nanoparticles Act as "Smart" Sense-and-Treat Carriers. *ACS Nano.* 2018; 12: 7538-45.
62. Chen W-H, Vazquez-Gonzalez M, Zoabi A, Abu-Reziq R, Willner I. Biocatalytic cascades driven by enzymes encapsulated in metal-organic framework nanoparticles. *Nat Catal.* 2018; 1: 689-95.
63. Chen W-H, Yu X, Ceconello A, Sohn YS, Nechushtai R, Willner I. Stimuli-responsive nucleic acid-functionalized metal-organic framework nanoparticles using pH-and metal-ion-dependent DNazymes as locks. *Chem Sci.* 2017; 8: 5769-80.
64. Chen WH, Yu X, Liao WC, Sohn YS, Ceconello A, Kozell A, et al. ATP-Responsive Aptamer-Based Metal - Organic Framework Nanoparticles (NMOFs) for the Controlled Release of Loads and Drugs. *Adv Funct Mater.* 2017; 27: 1702102.
65. Wang X-G, Dong Z-Y, Cheng H, Wan S-S, Chen W-H, Zou M-Z, et al. A multifunctional metal-organic framework based tumor targeting drug delivery system for cancer therapy. *Nanoscale.* 2015; 7: 16061-70.
66. Chen WH, Luo GF, Sohn YS, Nechushtai R, Willner I. Enzyme-Driven Release of Loads from Nucleic Acid-Capped Metal-Organic Framework Nanoparticles. *Adv Funct Mater.* 2019; 29: 1805341.
67. Ma Y, Li X, Li A, Yang P, Zhang C, Tang B. H₂S-Activable MOF Nanoparticle Photosensitizer for Effective Photodynamic Therapy against Cancer with Controllable Singlet-Oxygen Release. *Angew Chem Int Ed Engl.* 2017; 56: 13752-6.
68. Chen WH, Liao WC, Sohn YS, Fadeev M, Ceconello A, Nechushtai R, et al. Stimuli-Responsive Nucleic Acid-Based Polyacrylamide Hydrogel-Coated Metal - Organic Framework Nanoparticles for Controlled Drug Release. *Adv Funct Mater.* 2018; 28: 1705137.
69. Lin LS, Song J, Song L, Ke K, Liu Y, Zhou Z, et al. Simultaneous Fenton-like Ion Delivery and Glutathione Depletion by MnO₂-Based Nanoagent to Enhance Chemodynamic Therapy. *Angew Chem Int Ed Engl.* 2018; 130: 4996-5000.
70. Zhang W, Lu J, Gao X, Li P, Zhang W, Ma Y, et al. Enhanced Photodynamic Therapy by Reduced Levels of Intracellular Glutathione Obtained By Employing a Nano-MOF with CuII as the Active Center. *Angew Chem Int Ed Engl.* 2018; 130: 4985-90.
71. Fan H, Yan G, Zhao Z, Hu X, Zhang W, Liu H, et al. A smart photosensitizer-manganese dioxide nanosystem for enhanced photodynamic therapy by reducing glutathione levels in cancer cells. *Angew Chem Int Ed Engl.* 2016; 55: 5477-82.
72. Deng J, Wang K, Wang M, Yu P, Mao L. Mitochondria targeted nanoscale zeolitic imidazole framework-90 for ATP imaging in live cells. *J Am Chem Soc.* 2017; 139: 5877-82.
73. Yang X, Tang Q, Jiang Y, Zhang M, Wang M, Mao L. Nanoscale ATP-responsive Zeolitic Imidazole Framework-90 as a General Platform for Cytosolic Protein Delivery and Genome Editing. *J Am Chem Soc.* 2019; 141: 3782-6.
74. Fan W, Huang P, Chen X. Overcoming the Achilles' heel of photodynamic therapy. *Chem Soc Rev.* 2016; 45: 6488-519.
75. Lin J, Wang S, Huang P, Wang Z, Chen S, Niu G, et al. Photosensitizer-loaded gold vesicles with strong plasmonic coupling effect for imaging-guided photothermal/photodynamic therapy. *ACS Nano.* 2013; 7: 5320-9.
76. Park J, Jiang Q, Feng D, Mao L, Zhou H-C. Size-controlled synthesis of porphyrinic metal-organic framework and functionalization for targeted photodynamic therapy. *J Am Chem Soc.* 2016; 138: 3518-25.
77. Lismont M, Dreesen L, Wuttke S. Metal-Organic Framework Nanoparticles in Photodynamic Therapy: Current Status and Perspectives. *Adv Funct Mater.* 2017; 27: 1606314.
78. Liu Y, Sun H, Yang L, Zhu X, Wang X, Liang J, et al. Chelation-assisted assembly of multidentate colloidal nanoparticles into metal-organic nanoparticles. *Nanoscale.* 2018; 10: 21369-73.
79. Lu K, He C, Lin W. Nanoscale metal-organic framework for highly effective photodynamic therapy of resistant head and neck cancer. *J Am Chem Soc.* 2014; 136: 16712-5.
80. Lu K, He C, Guo N, Chan C, Ni K, Weichselbaum RR, et al. Chlorin-based nanoscale metal-organic framework systemically rejects colorectal cancers via synergistic photodynamic therapy and checkpoint blockade immunotherapy. *J Am Chem Soc.* 2016; 138: 12502-10.
81. Meng H-M, Hu X-X, Kong G-Z, Yang C, Fu T, Li Z-H, et al. Aptamer-functionalized nanoscale metal-organic frameworks for targeted photodynamic therapy. *Theranostics.* 2018; 8: 4332-44.
82. Zeng JY, Zhang MK, Peng MY, Gong D, Zhang XZ. Porphyrinic Metal-Organic Frameworks Coated Gold Nanorods as a Versatile Nanoplatform for Combined Photodynamic/Photothermal/Chemotherapy of Tumor. *Adv Funct Mater.* 2018; 28: 1705451.
83. Li Y, Di Z, Gao J, Cheng P, Di C, Zhang G, et al. Heterodimers Made of Upconversion Nanoparticles and Metal-Organic Frameworks. *J Am Chem Soc.* 2017; 139: 13804-10.
84. Li Y, Tang J, He L, Liu Y, Liu Y, Chen C, et al. Core-Shell Upconversion Nanoparticle@ Metal-Organic Framework Nanoprobes for Luminescent/Magnetic Dual-Mode Targeted Imaging. *Adv Mater.* 2015; 27: 4075-80.
85. Liu Y, He L, Pang K, Liu W, Tian Y, Chang L, et al. Core-Shell Noble-Metal@ Zeolitic-Imidazolate-Framework Nanocarriers with High Cancer Treatment Efficiency in Vitro. *J Mater Chem B.* 2019; 7: 1050-5.
86. Liu D, Wan J, Pang G, Tang Z. Hollow Metal -Organic-Framework Micro/Nanostructures and their Derivatives: Emerging Multifunctional Materials. *Adv Mater.* 2018: 1803291.
87. Lu G, Li S, Guo Z, Farha OK, Hauser BG, Qi X, et al. Imparting functionality to a metal-organic framework material by controlled nanoparticle encapsulation. *Nat Chem.* 2012; 4: 310-6.
88. Zhou Z, Chan A, Wang Z, Huang X, Yu G, Jacobson O, et al. Synchronous Chemoradiation Nanovesicles by X-Ray Triggered Cascade of Drug Release. *Angew Chem Int Ed Engl.* 2018; 130: 8599-603.
89. Xu J, Gao J, Wei Q. Combination of photodynamic therapy with radiotherapy for cancer treatment. *J Nanomater.* 2016: 8507924.
90. Duan X, Chan C, Lin W. Nanoparticle-Mediated Immunogenic Cell Death Enables and Potentiates Cancer Immunotherapy. *Angew Chem Int Ed Engl.* 2019; 58: 670-80.
91. Ni K, Lan G, Chan C, Quigley B, Lu K, Aung T, et al. Nanoscale metal-organic frameworks enhance radiotherapy to potentiate checkpoint blockade immunotherapy. *Nat Commun.* 2018; 9: 2351.
92. Lee Y, Auh SL, Wang Y, Burnette B, Wang Y, Meng Y, et al. Therapeutic effects of ablative radiation on local tumor require CD8+ T cells: changing strategies for cancer treatment. *Blood.* 2009; 114: 589-95.
93. Lugade AA, Moran JP, Gerber SA, Rose RC, Frelinger JG, Lord EM. Local radiation therapy of B16 melanoma tumors increases the generation of tumor antigen-specific effector cells that traffic to the tumor. *J Immunol.* 2005; 174: 7516-23.
94. Reynders K, Illidge T, Siva S, Chang JY, De Ruyscher D. The abscopal effect of local radiotherapy: using immunotherapy to make a rare event clinically relevant. *Cancer Treat Rev.* 2015; 41: 503-10.
95. Liu B, Ma M, Zacher D, Bétard A, Yuzenko K, Metzler-Nolte N, et al. Chemistry of SURMOFs: Layer-selective installation of functional groups and post-synthetic covalent modification probed by fluorescence microscopy. *J Am Chem Soc.* 2011; 133: 1734-7.
96. Huang X, He Z, Guo D, Liu Y, Song J, Yung BC, et al. "Three-in-one" Nanohybrids as Synergistic Nanoquencher to Enhance No-Wash Fluorescence Biosensors for Ratiometric Detection of Cancer Biomarkers. *Theranostics.* 2018; 8: 3461-73.
97. Liu D, Huxford RC, Lin W. Phosphorescent nanoscale coordination polymers as contrast agents for optical imaging. *Angew Chem Int Ed Engl.* 2011; 50: 3696-700.
98. Zhao M, Wang Y, Ma Q, Huang Y, Zhang X, Ping J, et al. Ultrathin 2D metal-organic framework nanosheets. *Adv Mater.* 2015; 27: 7372-8.
99. Cauda V, Argyo C, Bein T. Impact of different PEGylation patterns on the long-term bio-stability of colloidal mesoporous silica nanoparticles. *J Mater Chem.* 2010; 20: 8693-9.
100. Otsuka H, Nagasaki Y, Kataoka K. PEGylated nanoparticles for biological and pharmaceutical applications. *Adv Drug Deliv Rev.* 2012; 64: 246-55.
101. Shi Z, Chen X, Zhang L, Ding S, Wang X, Lei Q, et al. FA-PEG decorated MOF nanoparticles as a targeted drug delivery system for controlled release of an autophagy inhibitor. *Biomater Sci.* 2018; 6: 2582-90.
102. Wuttke S, Zimpel A, Bein T, Braig S, Stoiber K, Vollmar A, et al. Validating Metal-Organic Framework Nanoparticles for Their Nanosafety in Diverse Biomedical Applications. *Adv Health Mater.* 2017; 6: 1600818.
103. Wang S, Morris W, Liu Y, McQuirk CM, Zhou Y, Hupp JT, et al. Surface-Specific Functionalization of Nanoscale Metal - Organic Frameworks. *Angew Chem Int Ed Engl.* 2015; 54: 14738-42.
104. Morris W, Briley WE, Auyeung E, Cabezas MD, Mirkin CA. Nucleic Acid-Metal Organic Framework (MOF) Nanoparticle Conjugates. *J Am Chem Soc.* 2014; 136: 7261-4.
105. Deria P, Bury W, Hod I, Kung C-W, Karagiari O, Hupp JT, et al. MOF functionalization via solvent-assisted ligand incorporation: Phosphonates vs carboxylates. *Inorg Chem.* 2015; 54: 2185-92.
106. Zhu W, Xiang G, Shang J, Guo J, Motevalli B, Durfee P, et al. Versatile Surface Functionalization of Metal-Organic Frameworks through Direct Metal Coordination with a Phenolic Lipid Enables Diverse Applications. *Adv Funct Mater.* 2018; 28: 1705274.

107. Yi X-C, Xi F-G, Qi Y, Gao E-Q. Synthesis and click modification of an azido-functionalized Zr (iv) metal-organic framework and a catalytic study. *RSC Adv.* 2015; 5: 893-900.
108. Wang S, McGuirk CM, Ross MB, Wang S, Chen P, Xing H, et al. General and Direct Method for Preparing Oligonucleotide-Functionalized Metal-Organic Framework Nanoparticles. *J Am Chem Soc.* 2017; 139: 9827-30.
109. Röder R, Preiß T, Hirschle P, Steinborn B, Zimpel A, Hohn M, et al. Multifunctional nanoparticles by coordinative self-assembly of His-tagged units with metal-organic frameworks. *J Am Chem Soc.* 2017; 139: 2359-68.
110. Wuttke S, Braig S, Preiß T, Zimpel A, Sicklinger J, Bellomo C, et al. MOF nanoparticles coated by lipid bilayers and their uptake by cancer cells. *Chem Commun.* 2015; 51: 15752-5.
111. Illes B, Hirschle P, Barnert S, Cauda V, Wuttke S, Engelke H. Exosome-Coated Metal-Organic Framework Nanoparticles: An Efficient Drug Delivery Platform. *Chem Mater.* 2017; 29: 8042-6.
112. Lyu Y, Chen G, Shanguan D, Zhang L, Wan S, Wu Y, et al. Generating cell targeting aptamers for nanotheranostics using cell-SELEX. *Theranostics.* 2016; 6: 1440-52.
113. Tan W, Donovan MJ, Jiang J. Aptamers from cell-based selection for bioanalytical applications. *Chem Rev.* 2013; 113: 2842-62.
114. Zhu G, Zhang H, Jacobson O, Wang Z, Chen H, Yang X, et al. Combinatorial screening of DNA aptamers for molecular imaging of HER2 in cancer. *Bioconjug Chem.* 2017; 28: 1068-75.
115. Zhu G, Liu Y, Yang X, Kim Y-H, Zhang H, Jia R, et al. DNA-inorganic hybrid nanovaccine for cancer immunotherapy. *Nanoscale.* 2016; 8: 6684-92.
116. Zhu G, Mei L, Vishwasrao HD, Jacobson O, Wang Z, Liu Y, et al. Intertwining DNA-RNA nanocapsules loaded with tumor neoantigens as synergistic nanovaccines for cancer immunotherapy. *Nat Commun.* 2017; 8: 1482.
117. Liu Y, Hou W, Xia L, Cui C, Wan S, Jiang Y, et al. ZrMOF nanoparticles as quenchers to conjugate DNA aptamers for target-induced bioimaging and photodynamic therapy. *Chem Sci.* 2018; 9: 7505-9.
118. Zhu G, Chen X. Aptamer-based targeted therapy. *Adv Drug Deliv Rev.* 2018; 134: 65-78.