ISSN : 0082 - 6340 E-ISSN : 2337 - 876X Accredited : 30/E/KPT/2018



A JOURNAL ON ZOOLOGY OF THE INDO-AUSTRALIAN ARCHIPELAGO

Vol. 46, pp. 1–113 December 2019



Published by

RESEARCH CENTER FOR BIOLOGY INDONESIAN INSTITUTE OF SCIENCES BOGOR, INDONESIA

ISSN : 0082 - 6340 E-ISSN : 2337 - 876X Accredited : 30/E/KPT/2018



A JOURNAL ON ZOOLOGY OF THE INDO-AUSTRALIAN ARCHIPELAGO

Vol. 46, pp. 1–113 December 2019



Published by

RESEARCH CENTER FOR BIOLOGY INDONESIAN INSTITUTE OF SCIENCES BOGOR, INDONESIA

ISSN : 0082 - 6340 E-ISSN : 2337 - 876X Accredited : 30/E/KPT/2018

TREUBIA

A JOURNAL ON ZOOLOGY OF THE INDO-AUSTRALIAN ARCHIPELAGO Vol. 46, pp. 1–113, December 2019

Board of Editors:

Dr. Djunijanti Peggie, M.Sc. (Chief Editor) Dr. Dewi Malia Prawiradilaga, M.Rur.Sc.

Dr. Daisy Wowor, M.Sc.

Dr. Kartika Dewi

Dr. Dhian Dwibadra

Dr. Conni Margaretha Sidabalok, M.App.Sc.

International Editors:

Dr. Paul Bates, M.A. Harrison Institute Bowerwood House 15 Botolph's Road Sevenoaks,

Kent, TN13 3AQ, UK

Dr. Thomas von Rintelen Museum für Naturkunde Leibniz - Institut für Evolutions und

Biodiversitat sforschung an der Humboldt-University zu Berlin,

Invaliden straβe 43, 10115 Berlin, Germany

Dr. Alan T. Hitch University of California, Davis, CA 95616, USA

Referees:

Dr. Chris Reid Entomology, University New South Wales, Australian Museum,

1 William Street, Sydney, NSW 2010, Australia

Dr. Michael Balke Zoologische Staatssammlung München, Münchhausenstraße 21,

München, 81247 Germany

Dr. Chris J Müller Australian Museum, 6 College Street, Sydney, NSW 2010, Australia

Dr. Chooi Khim Phon Tropical Forest Biodiversity Centre, Forest Research Institute

Malaysia (FRIM), Kepong, Selangor; Zoological and Ecological Research Network, Institute of Biological Sciences, Faculty of

Science, University of Malaya, Kuala Lumpur, Malaysia

Prof. Dr. Masahiro Ôhara The Hokkaido University Museum, Japan

Dr. Peter Allsopp Queensland Museum, South Brisbane, Queensland, Australia

Dr. Aleš Bezděk Biology Centre of the Czech Academy of Sciences, Institute of

Entomology, Czech Republic

Dr. Mirza D. Kusrini Bogor Agricultural University, IPB-Department of Forest Resources

Conservation and Ecotourism, Bogor, Indonesia

Prof. Dr. Susumu Ohtsuka Graduate School of Integrated Sciences for Life, Setouchi Field

Science Center, Hiroshima University, Japan

Dr. J. C. von Vaupel Klein Rembrandtlaan 57, Bilthoven, Netherlands

Dr. R.I. Vane-Wright Durrell Institute of Conservation and Ecology, School of

Anthropology and Conservation, University of Kent, Canterbury, CT2 7NR, UK; Life Sciences, Natural History Museum, Cromwell Road,

London SW7 5BD, UK

Dr. Sih Kahono Zoology Division, Research Center for Biology, Indonesian Institute

of Sciences (LIPI), Indonesia

Eko Sulistyadi, M.Si. Zoology Division, Research Center for Biology, Indonesian Institute

of Sciences (LIPI), Indonesia

Maharadatunkamsi, M.Sc. Zoology Division, Research Center for Biology, Indonesian Institute

of Sciences (LIPI), Indonesia

Prof. Dr. Paul A. Racey Queensland Museum, South Brisbane, Queensland, Australia

Managing Assistant: Sri Wulan, S. Ikom.

Layout:

Liana Astuti

Distribution:

Rustan Nawawi

Subscription and Exchange

TREUBIA

RESEARCH CENTER FOR BIOLOGY - INDONESIAN INSTITUTE OF SCIENCES (LIPI)

Jl. Raya Jakarta-Bogor Km. 46, Cibinong-Bogor 16911, Indonesia
e-mail: treubia@gmail.com
http://e-journal.biologi.lipi.go.id/index.php/treubia

VOL. 46, DECEMBER 2019

CONTENT

Yaheita Yokoi, Hiroshi Makihara and Woro A. Noerdjito Callidiopini beetles (Coleoptera: Cerambycidae) in the collection of Museum Zoologicum Bogoriense, Indonesia	1–20
R.I. Vane-Wright The identity of <i>Euploea tulliolus goodenoughi</i> Carpenter, 1942, a crow butterfly (Lepidoptera: Nymphalidae, Danainae) from Papua New Guinea	21–34
Raden Pramesa Narakusumo and Michael Balke Four new species of <i>Epholcis</i> Waterhouse (Coleoptera: Scarabaeidae: Melolonthinae: Maechidiini) from the Moluccas, Indonesia	35–50
Mediyansyah, Amir Hamidy, Misbahul Munir and Masafumi Matsui A new tree frog of the genus <i>Kurixalus</i> Ye, Fei & Dubois, 1999 (Amphibia: Rhacophoridae) from West Kalimantan, Indonesia	51–72
Mulyadi New records and redescription of <i>Labidocera rotunda</i> Mori, 1929 (Copepoda, Calanoida, Pontellidae) from Sebatik Island, North Kalimantan, Indonesia, with notes on its species-group	73–84
Djunijanti Peggie Biological aspects of <i>Papilio peranthus</i> (Lepidoptera: Papilionidae) as observed at Butterfly Research Facility - LIPI, Cibinong, Indonesia	85–102
Susan M. Tsang and Sigit Wiantoro Review - Indonesian flying foxes: research and conservation status update	103–113

TREUBIA

(A JOURNAL ON ZOOLOGY OF THE INDO-AUSTRALIAN ARCHIPELAGO)

ISSN : 0082 - 6340 Date of issue: DECEMBER 2019 E-ISSN : 2337 - 876X

This abstract sheet may be reproduced without permission or charge

UDC: 595.76(594.53)

Yaheita Yokoi

Callidiopini beetles (Coleoptera: Cerambycidae) in the collection of Museum Zoologicum Bogoriense, Indonesia

TREUBIA, December 2019, Vol. 46, pp. 1–20.

Callidiopini species in the collection of Museum Zoologicum Bogoriense, Indonesian Institute of Sciences (LIPI) were examined. Three new species of the genus *Ceresium* Newman, 1842, are described, i.e. *C. clytinioides* sp. nov., *C. sugiartoi* sp. nov., both from Kalimantan, and *C. emarginatum* sp. nov. from Papua. One new species of the genus *Examnes* Pascoe, 1869, from Kalimantan, *E. subvermiculatus* sp. nov. is described.

(Yaheita Yokoi, Hiroshi Makihara and Woro A. Noerdjito)

Keywords: Asia, Kalimantan, longhorn beetle, New Guinea, taxonomy

UDC: 595.78.001.03(594.81)

R.I. Vane-Wright

The identity of *Euploea tulliolus* goodenoughi Carpenter, 1942, a crow butterfly (Lepidoptera: Nymphalidae, Danainae) from Papua New Guinea

TREUBIA, December 2019, Vol. 46, pp. 21–34.

The nominal taxon Euploea tulliolus goodenoughi Carpenter, 1942, based on a unique crow butterfly collected Goodenough Island in 1913, is shown to represent a small, aberrant female of the locally common Euploea leucostictos eustachius (Kirby, 1889). This new synonymy invalidates the only previous record of the Purple Crow, Euploea tulliolus (Fabricius, 1793), from the islands of Milne Bay Province, Papua New Guinea. However, two female Euploea tulliolus collected from islands in the Louisiade Archipelago during 2010 are reported here, constituting the first valid records of the Purple Crow from the Milne Bay islands.

(R.I. Vane-Wright)

Keywords: *tulliolus* species complex, new synonymy, new records, Milne Bay islands, *Euploea leucostictos*

UDC: 595.762(594.31)

Raden Pramesa Narakusumo

Four new species of *Epholcis* Waterhouse (Coleoptera: Scarabaeidae: Melolonthinae: Maechidiini) from the Moluccas, Indonesia

TREUBIA, December 2019, Vol. 46, pp. 35–50.

Here, we provide the first record of the chafer beetle genus *Epholcis* Waterhouse, 1875 from the Moluccas, Indonesia. We describe four new species: *E. acutus* sp. nov., *E. arcuatus* sp. nov., *E. cakalele* sp. nov., and *E. obiensis* sp. nov. A lectotype is designated for *Maechidius moluccanus* Moser, 1920, which is redescribed and transferred to the genus *Epholcis* as *E. moluccanus* (Moser) comb. nov.

(Raden Pramesa Narakusumo and Michael Balke)

Keywords: Coleoptera, *Epholcis*, Maechidiini, Melolonthinae, Moluccas

UDC: 597.82(594.17)

Mediyansyah

A new tree frog of the genus Kurixalus Ye, Fei & Dubois, 1999 (Amphibia: Rhacophoridae) from West Kalimantan, Indonesia

TREUBIA, December 2019, Vol. 46, pp. 51–72.

Kurixalus absconditus sp. nov., a new species of tree frog of the genus Kurixalus, described from West Kalimantan on the basis of molecular phylogenetic and morphological evidence. The new species can be distinguished from its congeners by a combination of following morphological characters: having smaller body size, more prominent of mandibular symphysis, skin smooth on throat, vomerine odontophores two oblique series touching anterior corner of choanae and widely separated, vomerine teeth thick, buccal cavity narrow and deep, choanae with teardrop shaped, single vocal slit, weakly crenulated dermal fringe on fore- and hindlimbs.

(Mediyansyah, Amir Hamidy, Misbahul Munir and Masafumi Matsui)

Keywords: *Kurixalus absconditus* sp. nov., new species, West Kalimantan

UDC: 594.34.001.03(594.11)

Mulyadi

New records and redescription of Labidocera rotunda Mori, 1929 (Copepoda, Calanoida, Pontellidae) from Sebatik Island, North Kalimantan, Indonesia, with notes on its species-group

TREUBIA, December 2019, Vol. 46, pp. 73–84.

During a plankton trip around Sebatik Island, North Kalimantan, a copepod Labidocera rotunda Mori, 1929 (Calanoida, Pontellidae) was collected for the first time in Indonesian waters. Both sexes redescribed and compared to previous descriptions. The geographical distribution of the species confirms that it is of Indo-Pacific origin. There has been a mix-up between L. rotunda described by Mori (1929) from Pusan, Korea and L. bipinnata from Sagami Bay, described by Tanaka (1936). Fleminger et al. (1982) have argued that the minor difference is based on the presence or absence of cephalic hooks and had synonymized *L. bipinnata* with *L. rotunda*.

(Mulyadi)

Keywords: copepods, Indonesia, *Labidocera rotunda*, new record, Pontellidae

UDC: 595.78:57.01(594.53)

Djunijanti Peggie

Biological aspects of *Papilio peranthus* (Lepidoptera: Papilionidae) as observed at Butterfly Research Facility - LIPI, Cibinong, Indonesia

TREUBIA, December 2019, Vol. 46, pp. 85–102.

Papilio peranthus is endemic to Indonesia, where it occurs on several islands and island groups. This beautiful butterfly is extensively traded, thus efforts to breed this species are very desirable. Captive breeding research was conducted on P. peranthus during September 2016 to December 2018. In total, 221 individuals were available for observation. Data on the life cycle of the species, together with observations on females being approached for mating, and female oviposition after presented. The result mating, are demonstrate that P. peranthus is not Observations on monogamous. other biological aspects are also reported.

(Djunijanti Peggie)

Keywords: egg-laying, mating, life cycle, *Papilio peranthus*, parent stocks

UDC: 599.41:001.891(594)

Susan M. Tsang

Review - Indonesian flying foxes: research and conservation status update

TREUBIA, December 2019, Vol. 46, pp. 103–113.

Flying foxes important are ecological keystone species on many archipelagoes, and Indonesia is home to over a third of all flying fox species globally. However, the amount of research on this clade belies their importance to natural systems, particularly as they are increasingly threatened by anthropogenic development and hunting. Here, we provide a review of the literature since the publication of the Old World Fruit Bat Action Plan and categorize research priorities as high, medium, or low based on the number of studies conducted. A majority of the research priorities for Indonesian endemics are categorized as medium or high priority. Low priority ratings were in multiple categories for widespread flying fox species found throughout Southeast Asia, though much of the data were from outside of the Indonesian extent of the species range. These research gaps tend to highlight broader patterns of research biases towards western Indonesia, whereas significant research effort is still needed in eastern Indonesia, particularly for vulnerable island taxa.

(Susan M. Tsang and Sigit Wiantoro)

Keywords: bats, conservation, Pteropodidae, *Pteropus*, threats

DOI:10.14203/treubia.v46i0.3795

BIOLOGICAL ASPECTS OF *PAPILIO PERANTHUS* (LEPIDOPTERA: PAPILIONIDAE) AS OBSERVED AT BUTTERFLY RESEARCH FACILITY - LIPI, CIBINONG, INDONESIA

Djunijanti Peggie*

Zoology Division (Museum Zoologicum Bogoriense), Research Center for Biology, Indonesian Institute of Sciences (LIPI), Jl. Raya Jakarta-Bogor Km. 46, Cibinong, Bogor 16911, Indonesia *e-mail: kupu2indonesia@gmail.com

Received: 14 November 2019; Accepted: 10 December 2019

ABSTRACT

Papilio peranthus is endemic to Indonesia, where it occurs on several islands and island groups. This beautiful butterfly is extensively traded, thus efforts to breed this species are very desirable. Captive breeding research was conducted on *P. peranthus* during September 2016 to December 2018. In total, 221 individuals were available for observation. Data on the life cycle of the species, together with observations on females being approached for mating, and female oviposition after mating, are presented. The result demonstrate that *P. peranthus* is not monogamous. Observations on other biological aspects are also reported.

Keywords: egg-laying, mating, life cycle, Papilio peranthus, parent stocks

ABSTRAK

Papilio peranthus merupakan spesies endemik Indonesia yang dijumpai di beberapa pulau di Indonesia. Spesies kupu-kupu yang indah ini diperdagangkan dalam jumlah besar sehingga diperlukan upaya penangkaran spesies ini. Penelitian penangkaran terhadap *P. peranthus* dilakukan dalam kurun waktu September 2016 hingga Desember 2018. Secara keseluruhan ada 221 individu yang diamati. Data mengenai siklus hidup spesies, dan pengamatan mengenai kupu-kupu betina didekati untuk kawin dan betina meletakkan telur setelah kawin dipaparkan di sini. Hasil studi ini menunjukkan bahwa *P. peranthus* bukan monogamous. Pengamatan terhadap aspek biologi lainnya juga dilaporkan.

Kata kunci: peletakan telur, kawin, siklus hidup, Papilio peranthus, induk

INTRODUCTION

Papilio peranthus Fabricius, 1787, which belongs to the peranthus-group of the subgenus Achillides, occurs in lowland areas of southern Sumatra (Hasanah et al., 2006; Nukmal et al., 2017), Java, Bawean, Bali, Nusa Tenggara, and Sulawesi (Tsukada & Nishiyama, 1982). Specimens deposited at Entomology Laboratory, Museum Zoologicum Bogoriense also confirm this geographical coverage for the species. This butterfly flies along streams and forest edges (Peggie & Amir, 2006). Many species of Papilio are highly sought after by collectors because of their beautiful wing colors, especially members of the subgenus Achillides. They have magnificent green or blue patches on forewings and hindwings, except Papilio paris and P. karna, which lack the patches on the forewings.

The high demands on trade of *P. peranthus* will put significant pressures on populations if they are taken directly from the habitat. To fulfil the demands, it is desirable to

have the butterflies harvested from captive breeding stocks, instead of capturing from wild populations. To achieve a better captive breeding operation for sustainable utilization of butterflies, various biological aspects need to be understood (Peggie, 2018). It is crucial to have the knowledge of the host plants (New et al., 1995) and how the species thrives in captivity. Previous detailed knowledge of the biology of *P. peranthus* is limited. The pre-adult stages were illustrated by Igarashi & Fukuda (2000), and observations of the mating behavior and male mate-locating strategy presented by Lukvitasari (2016) and Nukmal et al. (2017), respectively.

Each butterfly species has a close relationship with its hosts. The majority are oligotrophic, being limited to a few species of hosts. A female butterfly which has mated and is ready to lay eggs will search carefully for suitable plants and will not lay eggs if she cannot find the appropriate plants for her offspring (Vane-Wright, 2003; Ghosh et al., 2019). Eggs laid on inappropriate plants will usually hatch, but the larvae will either not feed, be poisoned, or develop very slowly, if at all. Igarashi & Fukuda (2000) recorded *Micromelum minutum*, which belongs to the family Rutaceae, as the host plant for *Papilio peranthus*. Nukmal et al. (2017) used another member of the Rutaceae, *Clausena excavata*, to rear *P. peranthus* at Lampung, southern Sumatra. *C. excavata* is known as the host plant for several other swallowtails, including *Papilio demoleus*, *P. polytes*, *P. palinurus*, and *P. daedalus* (Igarashi & Fukuda, 2000).

With the increasing need to know the life history and biological aspects of many Indonesian butterfly species, a butterfly research facility was built and established at Cibinong in August 2016, through the Biovillage program of the Indonesian Institute of Sciences (see below). To obtain life history data, observations on pre-adult stages were conducted in the rearing room. Observations on the adult behavior were made in the butterfly flight enclosure. Results from observations on flower visiting (nectaring), mating, oviposition, and other biological aspects of *Papilio peranthus*, are presented and discussed.

MATERIALS AND METHODS

Study area

The research was conducted at the Butterfly Research Facility of the Indonesian Institute of Sciences, located at Cibinong Science Center, Cibinong, Bogor, Indonesia. The butterfly facility includes a 10x20 sq. m. butterfly enclosure, a 4x6 sq. m. rearing room, and surrounding area for planting. Various plants have been grown inside and outside the butterfly enclosure to support butterflies, including larval host plants, nectar-producing plants, and shade plants. Rearing and research observations were conducted at ambient temperatures of 28–34°C.

Materials

Parent stocks (Fig. 1) for the research were obtained from several sources at different time, as follows:

- (I). The parent stocks on the first batch were pupae of *Papilio peranthus transiens* Fruhstorfer, 1897 from a butterfly supplier in Bali, a total of 29 pupae received on 21 September 2016.
- (II). Parent stocks on the second batch were pupae from a butterfly supplier in Bali, a total of 19 pupae received on 10 February 2017.
- (III). Parent stocks of the Javan subspecies, *Papilio peranthus peranthus*, were obtained from Citayam, Bogor, a total of 11 pupae received between 25 April 14 May 2017.
- (IV). Parent stocks from naturally arriving individuals of subspecies *peranthus* were obtained from outside the butterfly enclosure. To obtain more data on the biology of this species, they were caught and put into the butterfly enclosure (individual numbers 9188, 9315, 9347, and 9351).

Methods

Upon arrival of the parent stocks, each pupa was numbered by means of a label attached to the twig on which the pupa was hanging. The pupae were then put inside large plastic containers each with a mesh opening lid. When the adult butterfly emerged and its wings had fully expanded and dried, usually a few hours after emerging, the individual was marked using paint markers (Hagler & Jackson, 2001). The application of paint marker pens used in this research was developed to fulfil the need to recognize individuals. The dot marking was given on the underside of left forewing for easy handling (Fig. 2a). Paint marker pens with 10 different colors were used and applied consistently to indicate separate color for each number. White was used to indicate number 1, yellow = 2, purple = 3, brown = 4, red = 5, green = 6, blue = 7, orange = 8, silver = 9, gold = 0. This combination of 10 different colors proved effective. After being marked, the date of emergence and sex were noted in a data book. Males and females can readily be distinguished by the male androconial brand on the upperside of the forewings, and examination of the external genitalia at abdominal tip. Males have a pair of claspers or valves, while females have a rounded abdominal tip with a ventral opening between the 7th and 8th abdominal segments, the ostium bursae (Scoble, 1992, p. 103). Females of this species are slightly paler and, on average, larger than males (Tsukada & Nishiyama, 1982). Butterflies were released into the butterfly enclosure on the day of their eclosion. Observations were then commenced on the butterflies flying in the enclosure. Mating individuals were photographed whenever possible and the individual numbers recorded. Flower-visiting individuals were also noted or photographed. Other behavior such

as egg-laying was recorded whenever possible during the rearing period. To determine the life span of adults, regular searching for wings (Fig. 2b) was conducted as well as recording the activity of flying individuals.

After mating, or whenever ovipositing females were observed, the host plants were searched for eggs. Each day, the eggs were collected into a petri dish and brought to the rearing room for observation. After hatching, each first-instar caterpillar was placed in an individual plastic container. Fresh leaves of the host plants were added daily and excreta removed. Observation was conducted on the caterpillars as they grew and molted into subsequent instars, pupated, and the adults eclosed. All observations were recorded in the data book.



Figure 1. Pupae, as parent stocks of *Papilio peranthus*, obtained from several sources. The pupae were hung inside the wired pupal cabinet using cloth pins. (photograph: D. Peggie)



Figure 2. Marking of *Papilio peranthus* was done using paint marker pens (a) to indicate the individual number. The color combinations enabled individual butterflies to be recognized on inspection. Dead individuals were searched for daily to determine the life span of adults (b). (photograph: D. Peggie)

RESULTS

The rearing room observations gave data on the early stages of *Papilio peranthus*, whereas observations in the butterfly enclosure provided information on adult behavior. The observations of this butterfly species were conducted within the period of September 2016 through December 2018, encompassing 221 observed adults in total.

(I). The parental stock of the first batch of *P. peranthus transiens* from Bali all successfully eclosed between 25 September 2016 and 1 October 2016, to give 18 males and 11 females. Male number 238, which emerged on 26 September, was found at 10:00 mating with newly emerged female 239 on 27 September. Male 240, which emerged on 27 September, was found mating with newly emerged female 251 on 28 September. From this first batch, data on duration of adult life were obtained for five individuals, which ranged from 2-17 days. Ovipositing females were not observed for this first batch, but eggs were obtained from leaves of the host plant, Micromelum minutum. It was not possible to locate all eggs, determine which eggs were had been laid by particular females, or ascertain exactly when the eggs were laid. Most eggs were found on the underside of young leaves or young shoots (Fig. 3), glued with secretion from the colleterial glands (Chew & Robbins, 1984). Eggs were checked and taken every afternoon but some eggs might have been missed. The eggs were brought to the rearing room to rear the early stages. Thirty-seven eggs were obtained on 27 September, 104 eggs on 28 September, and some further eggs up to 9 October. At the same time, Papilio polytes individuals were in the enclosure, using the same host plants, so it was not easy to determine the eggs. The hatchlings were then moved to individual plastic jars and were given individual numbers. There were 81 individual F1 P. peranthus, 38 of which failed to develop into adulthood: 1 died at third instar, 4 died at fourth, 10 died at fifth, 10 failed at prepupal stage, 12 failed at pupal stage, and 1 failed to eclose. The 28 male and 15 female F1 individuals which emerged 29 October – 8 November did not manage to found a successful F2 generation. This was so despite F1 female 805 that emerged on 5 November mating almost immediately with F1 male 822.

- (II). The parental stock of the second batch of *P. peranthus transiens* comprised 19 pupae obtained on 10 February 2017 from Bali. Ten males and 1 female emerged between 13–19 February, 8 pupae failing at eclosion. No mating was observed, and no F1 generation resulted from this second batch.
- (III). A stock of the Javan subspecies, *P. peranthus peranthus*, from Citayam, Bogor, comprised 11 pupae obtained between 25 April and 14 May 2017. Six males and four females emerged, over the period 25 April to 27 May, with one pupa failing to eclose. No observations were obtained from this batch, and they did not produce an F1 generation.
- (IV). Parent stocks from naturally arriving individuals of subspecies *peranthus*. Local individuals of *P. peranthus* were observed outside the butterfly enclosure from time to time, visiting the host plants. On 3 June 2018 a male, numbered 9188, was put into the enclosure but there was no female present during the following days, so this male could not mate. On 6 July 2018 a female was seen outside and it was caught and numbered 9315. The activity of the butterfly seemed uninterrupted upon being put inside the enclosure, as it fed on available nectar plants and laid eggs on *Micromelum minutum*. This female, numbered 9315, laid eggs which resulted in 25 larvae. These produced an F1 generation of 12 male and 11 female adults which emerged 6–15 August 2018; 2 failed at pupal stage.

Table 1. Duration of each life stage of Papilio peranthus reared in captivity at the butterfly research

	egg	L1	L2	L3	L4	L5	prepupa	pupa	adult
Number individuals observed	156	155	155	153	148	138	128	107	20
Average (days)	4.45	3.04	3.35	3.12	2.78	2.90	1	10.58	6.8
Maximum (days)	7	4	4	4	4	4	1	12	21
Minimum (days)	1	2	2	2	2	2	1	8	2

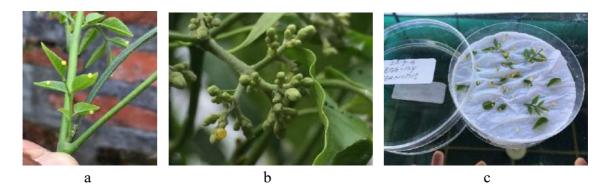


Figure 3. Examples of eggs of *Papilio peranthus* obtained during this research: (a) eggs on the underside of the host plant, (b) eggs laid on a young shoot of the host plant, (c) eggs taken to the rearing room for captive breeding. (photograph: D. Peggie)



Figure 4. Larvae of *Papilio peranthus*: (a) L1 – first instar, average length 3 mm, (b) L2 – second instar, average length 5–6 mm, (c) L3 – third instar, average length 12 mm, (d) L4 – fourth instar, average length 19 mm, and (e) L5 – fifth instar, average length 28 mm. (photograph: D. Peggie)

There were 32 F2 larvae resulting from the F1: 13 males and 10 females emerged 8–21 September 2018, and 9 failed (2 at L1, 3 at pupal stage, and 4 at eclosion). The F2 generation produced 13 F3 individuals, 7 males and 6 females emerging 20 October 2018 to 1 November 2018. The F3 resulted in 4 F4 individuals: 3 males and 1 female which emerged 28 November 2018 to 2 December 2018. Female 9315 had no further descendants.

Two more local *P. peranthus peranthus* females arrived on 28 August 2018, and were numbered 9347 and 9351. Eggs obtained on 31 August produced two females (9416 and 9417) which emerged on 30 September and 1 October 2018, respectively; another pupa failed at eclosion. These F1s did not produce offspring.

Based on daily observations of each individual and molting events to the next instar of larva, the duration period of each life stage is presented (Table 1), showing the average duration from eggs to adults is 31 days. However, it should be noted that the date of the eggs obtained might not necessarily mean the date of egg laying. The larval stage consists of 5 instars (Fig. 4), and each instar lasts about 3 days on average, giving a total of 15 days (see Table 1). For comparison, the larval duration of *Papilio polytes* reared at Nilgiri Hills, India was 14.41 days (Rajeswari & Jeyabalan, 2017). The process from prepupal to pupal stage (Fig. 5) takes one day. Pupal development lasts about 10 days. Observation in the butterfly enclosure showed that some adults could live over 2 weeks and one individual (number 9326) was recorded still alive for 21 days, but many of them lived only about a week. The

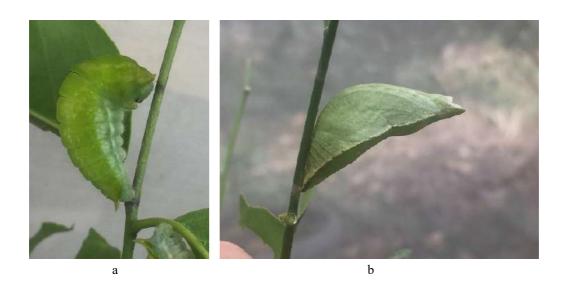


Figure 5. prepupa (a) is ready for pupal ecdysis (moult), which usually takes place within a day, to reveal the pupa (b). (photograph: D. Peggie)

determination of the life span of adults was obtained through finding broken wings (Fig. 2b) as well as recording the activity of flying individuals daily during the research period.

Adults eclosing from the pupal cases crept out and needed time to expand their wings. These processes would take 1–2 hours. When the butterflies were considered ready, with wings fully expanded and dried, they were marked with paint on the underside of left forewing, and then released into the enclosure. Some newly emerged butterflies were observed to perch on plants for a few minutes, others were ready to fly (Fig. 6).

Daily observations inside the enclosure revealed that newly emerged butterflies of this species do not visit flowers for nectar on the day of emergence. Subsequently, the butterflies visited Jatropha integerrima (Euphorbiaceae), Impatiens hawkeri (Balsaminaceae) (Fig. 8b), Ixora spp. (Rubiaceae), Caesalpinia pulcherrima (Fabaceae), Cuphea hyssopifolia (Lythraceae), Pseuderanthemum reticulatum (Acanthaceae), Zinnia sp. (Asteraceae), Cosmos caudatus (Asteraceae), Lantana camara (Verbenaceae), Jatropha podagrica (Euphorbiaceae), and Bougainvillea sp. (Nyctaginaceae). Later, after 3 days, the butterflies also visited Aloysia virgata (Verbenaceae). Clerodendrum paniculatum (Lamiaceae) is known also as a nectar plant for P. peranthus, as seen in some butterfly gardens, but during the period of research this plant species was not in bloom. Antigonon leptopus (Polygonaceae) was not visited.

Observations showed that males did not mate on the first day of emergence (Table 3, Fig. 7). Most females, on the other hand, were approached by males within minutes of their entrance to the enclosure and were then mated (Table 3). Some females were also observed mating on days 2 and 3 after emergence. One female (number 9400) was observed mating with two different males in the same morning. One male (number 9386) mated 3 times, with 3 different females on days 2, 3, and 6. Thus males and females of this species can be polygamous, mating multiple times with different individuals. This contradicts the conclusion drawn by Lukvitasari (2016) based on a very brief observation in a butterfly enclosure in Lampung. Of course, in nature some males and some females will not get the opportunity to mate more than once, if at all.

Mated females were seen laying eggs usually a day after mating (Table 2, Fig. 7b). These ovipositing females fluttered around the host plant, *Micromelum minutum*, several times before laying eggs on the underside of leaves or flower buds. Some females were observed visiting flowers between episodes of ovipositing activity.

'Mud-puddling' and urinating behavior was also seen (Table 3, Fig. 8b). Similar behavior was observed in *Papilio karna* at Cikaniki research station, Gunung Halimun-Salak National Park, April 2016.

Table 2. Adult activities of Papilio peranthus as recorded during the research in the butterfly enclosure

No.	Individual number	Date emergence	M/F	Observed mating	Other activities
1	239	27 September 2016	female	27 September 2016 at 10:00 with 238	
2	240	27 September 2016	male	28 September 2016 with 251	
3	805	5 November 2016	female	5 November 2016 with 822	
4	822	4 November 2016	male	5 November 2016 with 805	
5	9315	6 July 2018	female		laid eggs 6 July 2018
6	9319	8 August 2018	male	9 August 2018 at 8:50 with 9322	puddled 9 August 2018 at 12:00
7	9320	8 August 2018	male	9 August 2018 at 8:30 with 9323	puddled 9 August 2018 at 12:10
8	9322	9 August 2018	female	9 August 2018 at 8:50 (30 mins. upon release) with 9319 until 10:00	
9	9323	9 August 2018	female	9 August 2018 at 8:30 (10 mins. upon release) with 9320 until 10:50	laid eggs 10 August 2018
10	9327	10 August 2018	female	10 August 2018 at 10:20 with 9319, still in copula at 11:35	
11	9328	11 August 2018	male	13 August 2018 at 12:00 with 9335	
12	9326	10 August 2018	male		puddled 13 August 2018; still seen alive 31 August 2018
13	9335	13 August 2018	female	mated at 12:00 (10 mins. upon release) with 9328	
14	9339	14 August 2018	male	15 August 2018 with 9341	
15	9340	15 August 2018	female		laid eggs16 August 2018; laid 26 eggs in small cage 17 August 2018
16	9341	15 August 2018	female	15 August 2018 with 9339	
17	9385	12 August 2018	female	12 August 2018 with 9382	laid eggs 13 August 2018
18	9386	12 September 2018	male	13 September 2018 with 9389; 14 September 2018 with 9393; 17 September 2018 with 9400	
19	9387	13 September 2018	male	17 September 2018 with 9400	
20	9389	13 September 2018	female	4 minutes upon entering with 9386	
21	9393	14 September 2018	female	14 September 2018 at 9:50 with 9386	laid eggs 17 September 2018
22	9400	17 September 2018	female	17 September 2018 at 9:30 – 10:30 mated with 9387 and at 11:05 with 9386	
23	9401	17 September 2018	male	18 September 2018 with 9402	
24	9402	18 September 2018	female	18 September 2018 at 9:40 – 10:30 with 9401	laid eggs 20 September 2018



Figure 6. *Papilio peranthus* male (a) can be distinguished by the androconial brand on the upperside of the forewings; the female (b) is a little paler and lacks the androconia. (photograph: D. Peggie)



Figure 7. Mating of *Papilio peranthus* is shown here (a) with the female is usually above the male in position; when they needed to move, the female would take flight carrying the male trailing in the same position. An ovipositing female flutters around *Micromelum minutum* (b) to lay eggs singly on the underside of the leaves or young buds. (photograph: D. Peggie)

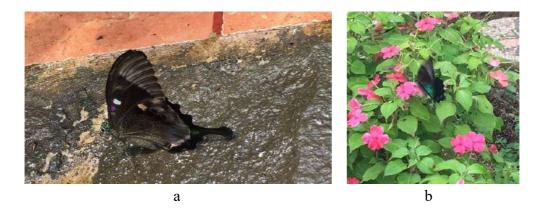


Figure 8. Puddling behavior (a) and flower visiting (b) of *Papilio peranthus* were observed in this research. (photograph: D. Peggie)

DISCUSSION

A total of 61 individuals (59 pupae and two adults) served as parental stocks in this investigation. Not all stocks were successful. From the first batch all pupae eclosed successfully and produced a first generation. From the second batch, eight pupae failed. From the Citayam parental stock, only one pupa failed to develop. However, the success rate for producing descendants was not only influenced by the number of potential parents, but also by the timing of emergence. The first batch of pupae eclosed within a few days (Fig. 9), so fresh males and females were available together to mate. The second batch only produced one female, which emerged very late compared to the males, which had emerged up to 6 days earlier (Fig. 10), so mating was unlikely. The stock from Citayam also failed to reproduce, even though there were 6 males and 4 females, most likely for the same reason, i.e. the time of emergence was too widely separated (Fig. 11).

The stock derived from naturally arriving individual 9315 was the most successful. Upon being captured and marked, this female resumed her activities – visiting flowers and laying eggs. Within that week, eggs were obtained from the leaves of the host plants. Ten days after the female was taken into the enclosure and laid eggs, first instar larvae started to hatch, and they grew well to adulthood. In exactly 31 days after the founder female was brought in, F1 individuals emerged. This cycle continued to a fourth generation.

Data on the life cycle of *Papilio peranthus* gave, at the ambient temperatures experienced, an average duration from egg to adult of 31 days. Apparent duration of egg development varied greatly. This was because the date of the eggs obtained might not necessarily represent the date at which the eggs were laid. Efforts had been taken to get all available eggs each day, but some eggs were almost certainly overlooked, and then gathered on subsequent days. Eggs were laid singly, as commonly observed for most species of Papilionidae (Stamp, 1980). The number of eggs that can be laid by a single female butterfly during her life time cannot be determined from the present data as there was only one enclosure available at the time. It was likewise not possible to know which eggs were laid by which female during the project. Therefore, no meaningful assessment of the fecundity of female *P. peranthus* can yet be made. It is hoped that further data will be presented at a later date as two small enclosures have been built recently. The success and survival rates of each larval instar also needs further research.

The host plant used by *P. peranthus* for this project was *Micromelum minutum*. Another member of the Rutaceae successfully used by *P. peranthus* is *Clausena excavata*: Nukmal et al. (2017) employed this species to rear *P. peranthus* at Lampung. *Clausena excavata* is also a known host for other species of *Papilio*, including *P. demoleus*, *P. polytes*, *P. palinurus* and *P. daedalus* (Igarashi & Fukuda, 2000). Studies on food plant preferences could be conducted in future, as additional enclosures are now available.

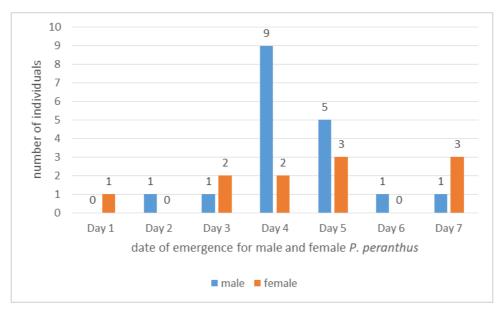


Figure 9. Adult emergence of parent stocks batch I of Papilio peranthus.

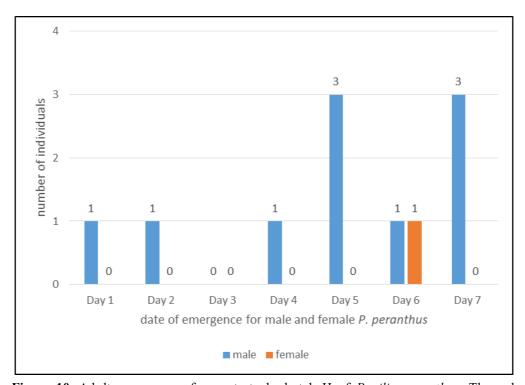


Figure 10. Adult emergence of parent stocks batch II of *Papilio peranthus*. The only female failed to mate.

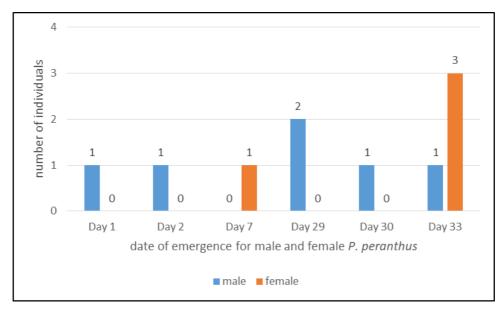


Figure 11. Adult emergence of parent stocks from Citayam, Bogor, of *Papilio peranthus*. No second generation was produced because the time of emergence of males and females was too widely separated.

The observations demonstrate that both males and females of this species are not monogamous (Table 2). They can mate multiple times with different individuals. Males approached newly emerged females relatively quickly, as little as four minutes after a female was released into the enclosure. Many female Lepidoptera release sex pheromones that can be recognized by the males conspecifics (Roelofs & Carde, 1974). Although there is evidence that this occurs in butterflies, details are sparse and the phenomenon is not well established for many butterflies (Boppré, 1984). The receptivity of female butterflies to males is influenced by various factors, including male pheromones (Boppré, 1984; Andersson et al., 2003; Wedell, 2005). Male pheromones can be dispersed from androconial scales, as suggested by an experiment with *Heliconius melpomene* in Panama, in which the specialized wing area with androconial scales was covered by a nail varnish, resulting in loss of receptivity by females (Darragh et al., 2017).

At the time of observation of *P. peranthus*, some other butterfly species were also inside the enclosure. At certain times the presence of *P. polytes* might have been a nuisance for *P. peranthus* as they were both making use of the same host plant. However, further study will need to be conducted to check any assumption or potential of competition. The presence of *Troides helena* (Papilionidae) and *Idea blanchardii* (Nymphalidae) did not seem to affect *P. peranthus*, neither of which feed on Rutaceae, as they continued to thrive. More quantitative assessments on the interactions among butterfly species co-existing in the enclosures could be undertaken. Daily activity patterns of the butterfly species co-occurring

within the enclosure could also be investigated, as Peixoto & Benson (2009) discussed for satyrine butterflies in a forest fragment in Brazil.

The associations of adult butterflies with plants for nectar are generally far less specific (Courtney, 1986; Shreeve, 1992). This research showed that adult butterflies would go to most flowers available for nectar. An exception was *Antigonon leptopus*, in which the small flower opening perhaps limits the ability of this butterfly species to obtain nectar. However, *Papilio polytes* and several other butterfly species have been recorded to take nectar from this plant (Raju et al. 2001, table 1) as observed also with other butterfly species in the enclosure. Flower preference by *P. peranthus* was not investigated in detail as the adults in general would visit almost any flowers with corolla depth that could be reached by the proboscis (Corbet, 2000; Tiple et al., 2009). Based on daily observations inside the enclosure, newly emerged butterflies of this species do not visit flowers for nectar on the day of emergence. This corresponds to observations on *Tirumala limniace* by Li et al. (2015). At times when there were many butterflies of different species in the enclosure, additional 10% sugar solution was placed on *Hibiscus* flowers, and *P. peranthus* would also feed on that.

Other behavior observed included "mud-puddling" (drinking) and urinating. Similar behavior was observed in *Papilio karna* at Cikaniki research station, Gunung Halimun-Salak National Park, April 2016. Many butterflies showed puddling behavior perhaps to obtain supplementary diet like sodium as suggested by Adler & Pearson (1982). The time of puddling observed here was between 12:00 – 12:20 and Patwardhan (2019) reported that puddling happened between 8:00 – 13:00 with peak activity at 11:30. This behavior was recorded only on three male individuals, two of which were observed to have mated that morning. This behavior may warrant further study.

Understanding the biology of this butterfly species and how it thrives in captivity may lead to a better captive breeding operation for sustainable utilization of the butterflies. Further study regarding the fecundity of female *P. peranthus*, the success and survival rates of each life stages will be useful. Observations on the effect of parasitoids and predators will complement the understanding toward better management of a butterfly facility.

CONCLUSION

The mean duration of life cycle of *Papilio peranthus* demonstrated in this study was 31 days. The success rates of each life stage needs to be further assessed. Adults can live over 21 days. The male and female of this species are not monogamous. Males exhibit water -puddling behavior. The adults used most available flowers as nectar sources except *Antigonon leptopus*.

ACKNOWLEDGMENTS

This research was supported by Biovillage program 2016-2017 coordinated by Research Center for Biotechnology-Indonesian Institute of Sciences (LIPI) and Biovillage program 2018 coordinated by Research Center for Biology-Indonesian Institute of Sciences (LIPI) through Indonesian government project DIPA 2016-2018. The author is very grateful for the immense help provided by Supadi and Guntoro in the butterfly research facility. Technicians of Entomology Laboratory: Sarino, Giyanto, Rina Rachmatiyah, and Fatimah have assisted with earlier rearing process. The author would like to extend sincere thanks to Dr. Sih Kahono for his help and suggestions regarding data presentation. I thank the reviewers who kindly read and improved the manuscript.

REFERENCES

- Adler, P.H. & Pearson, D.L. 1982. Why do male butterflies visit mud puddles? *Canadian Journal of Zoology*, 60: 322–325.
- Andersson, J., Borg-Karlson, A.-K. & Wiklund, C. 2003. Functional significance of citral as an aphrodisiac in the butterfly *Pieris napi*: Behavioural experiments and EAG tests. *J. Chem. Ecol.*, 29: 1489–1499.
- Boppré, M. 1984. Chemically mediated interactions between butterflies. In: R.I. Vane-Wright & P.R. Ackery, eds. *The Biology of Butterflies*. London: Academic Press: pp. 259–275.
- Chew, F.S. & Robbins, R.K. 1984. Egg-laying in butterflies. In: R.I. Vane-Wright & P.R. Ackery, eds. *The Biology of Butterflies*. London: Academic Press: pp. 65–79.
- Corbet, S.A. 2000. Butterfly nectaring flowers: butterfly morphology and flower form. *Entomol. Exp. Appl.*, 96: 289–298.
- Courtney, S.P. 1986. The ecology of pierid butterflies: dynamics and interactions. *Adv. Ecol. Res.*, 15: 51–116.
- Darragh, K., Vanjari, S., Mann, F., Gonzalez-Rojas, M.F., Morrison, C.R., Salazar, C., Pardo-Diaz, C., Merrill, R.M., McMillan, W.O., Schulz, S. & Jiggins, C.D. 2017. Male sex pheromone components in *Heliconius* butterflies released by the androconia affect female choice. *PeerJ.*, 5:e3953 https://doi.org/10.7717/peerj.3953
- Ghosh, A., Abedin, M.S. & Hossain, M.M. 2019. Captive rearing of *Papilio polymnestor* and *Chilasa clytia* butterflies in the campus of Jahangirnagar University, Bangladesh. *Journal of Entomology and Zoology Studies*, 7(3): 1046–1051.
- Hagler, J.R. & Jackson, C.G. 2001. Methods for marking insects: Current techniques and future prospects. *Annu. Rev. Entomol.*, 46: 511–543.
- Hasanah, N., Tabadepu, H., Sahari, B. & Buchori, D. 2006. Butterfly community structure in Bukit Barisan Selatan National Park, Sumatera. Final Report. Peduli Konservasi Alam (Peka) Indonesia Foundation (Center for Conservation and Insect Studies) and Wildlife Conservation Society-Indonesia Program (WCS IP), 20 pp.
- Igarashi, S. & Fukuda, H. 2000. *The Life Histories of Asian Butterflies. Vol. 2.* Tokyo (Japan): Tokai University Press: p. 335, pls. 43 & 401.

- Li, C., Wang, F., Chen, X., Zhou, C. & Yao, J. 2015. Adult behavior of *Tirumala limniace* (Lepidoptera: Danaidae). *J. Insect Sci.*, 15(1): 76. DOI: 10.1093/jisesa/iev061
- Lukvitasari, L. 2016. *Perilaku Kawin Kupu-kupu* Papilio peranthus (*Lepidoptera: Papilionidae*) di Kandang Penangkaran. Skripsi. Jurusan Biologi Fakultas Matematika dan Ilmu Pengetahuan Alam, Universitas Lampung, Bandar Lampung, 46 hal.
- New, T.R., Pyle, R.M., Thomas, J.A., Thomas, C.D. & Hammond, P.C. 1995. Butterfly conservation management. *Annual Review of Entomology*, 40: 57–83.
- Nukmal, N., Kanedi, M., Soekardi, H. & Lukvitasari, L. 2017. Ethogram of mating and male mate-locating strategy of *Papilio peranthus* (Lepidoptera: Papilionidae) at Gita Persada Butterfly Park, Lampung, Indonesia. *Journal of Entomology and Zoology Studies*, 5 (1): 869–874.
- Patwardhan, A. 2019. Some observations on the butterfly mud puddling in and around Mumbai. *Journal of Entomology and Zoology Studies*, 7(5): 296–304.
- Peggie, D. & Amir, M. 2006. Practical Guide to the Butterflies of Bogor Botanic Garden/ Panduan Praktis Kupu-kupu di Kebun Raya Bogor. Bogor (Indonesia): Bidang Zoologi, Pusat Penelitian Biologi, LIPI & Nagao Natural Environment Foundation, Japan: 126 pp.
- Peggie, D. 2018. Kajian diversitas kupu-kupu (Lepidoptera: Papilionoidea & Hesperioidea) dan potensi pemanfaatannya di Hutan Petungkriyono, Kabupaten Pekalongan, Jawa Tengah Petung Kriyono. *Kajen Jurnal Penelitian dan Pengembangan Pembangunan*, 2(2): 106–123.
- Peixoto, P. & Benson, W.W. 2009. Daily activity patterns of two co-occurring tropical satyrine butterflies. *Journal of Insect Science*, 9(54): 1–14. available online: insectscience.org/9.54
- Rajeswari, N.B. & Jeyabalan, D. 2017. Studies on biology and reproduction of butterflies (family: Papilionidae) in Nilgiris Hills, Southern Western Ghats, India. *Int. J. Adv. Res. Biol. Sci.*, 4(2): 1–11. DOI: http://dx.doi.org/10.22192/ijarbs.2017.04.02.001.
- Raju, A.J.S., Raju, V.K., Victor, P. & Naidu, S.A. 2001. Floral ecology, breeding system and pollination in *Antigonon leptopus* L. (Polygonaceae). *Plant Species Biology*, 16: 159–164.
- Roelofs, W.L. & Cardé, R. 1974. Sex pheromones in the reproductive isolation of lepidopterous species. In: M.C. Birch, ed. *Pheromones*. New York: Elsevier: pp. 96–114.
- Scoble, M.J. 1992. *The Lepidoptera. Form, Function and Diversity*. Natural History Museum Publications, Oxford: Oxford University Press: 404 pp.
- Shreeve, T.G. 1992. Adult behaviour. In: Dennis R.L.H., ed. *The Ecology of Butterflies in Britain*. Oxford (UK): Oxford University Press: pp. 22–45.
- Stamp, N.E. 1980. Egg deposition patterns in butterflies: why do some species cluster their eggs rather than deposit them singly? *Amer. Nat.*, 115: 367–380.
- Tiple, A.D., Khurad, A.M. & Dennis, R.L.H. 2009. Adult butterfly feeding-nectar flower associations: constraints of taxonomic affiliation, butterfly, and nectar flower morphology. *Journal of Natural History*, 43 (13): 855–884. DOI: 10.1080/00222930802610568.
- Tsukada, E. & Nishiyama, Y. 1982. Butterflies of the South East Asian Islands. Vol. I. Papilionidae. Tokyo (Japan): Plapac Co., Ltd.: 457 pp.

- Vane-Wright, R.I. 2003. *Butterflies*. Life Series. London: The Natural History Museum: 112 pp.
- Wedell, N. 2005. Female receptivity in butterflies and moths. *The Journal of Experimental Biology*, 208: 34333440. doi:10.1242/jeb.01774
- Westerman, E.L., Drucker, C.B. & Monteiro, A. 2014. Male and female mating behavior is dependent on social context in the butterfly *Bicyclus anynana*. *J. Insect Behav.*, 27(4): 478–495.

INSTRUCTIONS FOR AUTHORS

TREUBIA is a peer-reviewed, scientific zoological journal with focus on biosystematic aspects of terrestrial and aquatic fauna in the Indo-Australian region. TREUBIA is published yearly and accepts manuscripts within the scope of the journal. It is accessible online at http://e-journal.biologi.lipi.go.id/index.php/treubia.

The missions of TREUBIA are to: (1) promote sciences and disseminate information in animal systematics and on the biodiversity of the region; (2) participate in the effort of educating public through good quality of scientific media and available professional researchers; (3) establish linkages among zoologists particularly in the field of systematics.

TREUBIA accepts manuscripts based on original research, taxonomic review or short communication. The manuscript should not be offered for prior or simultaneous publication elsewhere. It must be written in English and should use the American English spelling. Manuscripts should be prepared double-spaced in Microsoft Word, using Times New Roman font 12, A4 paper size. Template is available through e-journal. An electronic file of the manuscript along with a formal cover letter – indicating the importance, stating its originality and its approval by all co-authors – should be submitted to the editors of TREUBIA through http://e-journal.biologi.lipi.go.id/index.php/treubia or through email address: treubia@gmail.com.

Concise writing is recommended. All numbers under 10 and any number forming the first word of a sentence must be spelled out, except in the Materials and Methods section of taxonomic papers. Year should be completely written. Names of genera and species should be in italic type. It is recommended to use metric measurements in abbreviation (for examples: kg, cm, ml). Please consult and refer to a recent issue of TREUBIA for an acceptable format. Please note that starting in 2018, we adopt Mendeley reference management application, with Harvard referencing style.

Manuscripts should be presented in the following order (with Conclusions and Appendices if necessary):

Title section. This includes the title of the paper (all capitalized), author's full name, author's institution and address (all with first letters capitalized), and e-mail address of the corresponding author. The title should be short, informative and without abbreviation.

Abstract. Except for short communications, articles should be accompanied by an abstract. The abstract consists of no more than 250 words in one paragraph which should clearly state the essence of the paper, with no references cited.

Keywords. Following the abstract, list up to 5 keywords, all typed in lowercase except a proper noun, separated by commas, presented in alphabetical order.

Introduction. The introduction must briefly justify the research and give the objectives. References related to the justification of the research should be cited in the introduction but extensive and elaborate discussion of relevant literature should be addressed in the Discussion section. References are to be cited in the text by the author's surname and year of publication. When citing multiple sources, place them in chronological order, for example: (Glaubrecht, 1999, 2006; Glaubrecht et al., 2009; Maaβ & Glaubrecht, 2012). For two authors, both names should be cited. For three authors or more, only the first author is given followed by et al.

Materials and Methods. Provide a clear explanation of materials and methods used in the research. The place of specimen depository should be mentioned here.

Results. The results can be presented in the form of tables and figures when appropriate. The text should explain and elaborate the data presented. Captions of tables, figures, and plates should be inserted where you want them to be inserted. All line drawings, photographs and other figures should be submitted separately in JPEG format and the image size should be at least 1024 by 768 pixels.

Discussion. The discussion should interpret the results clearly and concisely, and should discuss the findings in relation with previous publications.

Acknowledgments. Acknowledgments of grants, assistance and other matters can be written in one paragraph.

References. List of references should be in alphabetical order by the first or sole author's surname. Journal references should include author's surname and initials, year of publication, title of the paper, full title of the journal (typed in *italic*), volume number and inclusive page numbers. Book references should include author's surname and initials, year of publication, title of the book (typed in *italic*) or/and title of the chapter and editor (if part of a book), publisher, city of publication, and page numbers.

For example:

- Eaton, J.A., van Balen, B., Brickle, N.W. & Rheindt, F.E. 2016. *Birds of the Indonesian Archipelago: Greater Sundas and Wallacea*. 1st ed. Barcelona: Lynx Edicions.
- LaSalle, J. & Schauff, M.E. 1994. Systematics of the tribe Euderomphalini (Hymenoptera: Eulophidae): parasitoids of whiteflies (Homoptera: Aleyrodidae). *Systematic Entomology*, 19: 235–258.
- MacKinnon, J. & Phillips, K. 1993. *Field Guide to the Birds of Borneo, Sumatra, Java and Bali*. Oxford: Oxford University Press: 491 pp.
- Natural History Museum 2013. Wallace100 celebrating Alfred Russel Wallace's life and legacy. http://www.nhm.ac.uk/nature-online/science-of-natural-history/wallace/index.html 11 October 2013.
- Higgins, P., Christidis, L., Ford, H. & Bonan, A. 2017. Honeyeaters (Meliphagidae). In: J. del Hoyo,
 A. Elliott, J. Sargatal, D.A. Christie & E. de Juana, eds. *Handbook of the Birds of the World Alive*. Barcelona: Lynx Edicions. http://www.hbw.com.

Upon receiving a manuscript, a Treubia editor will check the compliance with these instructions and will send the manuscript to two reviewers. Based on comments from the reviewers and the suitability of the manuscript, Treubia editors will decide the acceptance or rejection of the manuscript. The author will be notified of the decision and will receive the manuscript with reviewers' comments.

Following the process of reviewing and revising, a final proof will be sent to the first or sole author for correction and approval. Five reprints are supplied free of charge but delivery cost will be charged. Joint authors will have to divide these copies among them at their discretion. Additional reprints can be provided at cost, the order should be placed before the final printing.

VOL. 46, DECEMBER 2019

CONTENT

Callidiopini beetles (Coleoptera: Cerambycidae) in the collection of Museum Zoologicum Bogoriense, Indonesia	1–20
R.I. Vane-Wright The identity of <i>Euploea tulliolus goodenoughi</i> Carpenter, 1942, a Crow Butterfly (Lepidoptera:	
Nymphalidae, Danainae) from Papua New Guinea	21–34
Raden Pramesa Narakusumo and Michael Balke Four new species of <i>Epholcis</i> Waterhouse (Coleoptera: Scarabaeidae: Melolonthinae: Maechidiini) from	
the Moluccas, Indonesia	35–50
Mediyansyah, Amir Hamidy, Misbahul Munir and Masafumi Matsui	
A new tree frog of the genus <i>Kurixalus</i> Ye, Fei & Dubois, 1999 (Amphibia: Rhacophoridae) from West Kalimantan, Indonesia	51-72
Mulyadi New records and redescription of <i>Labidocera rotunda</i> Mori, 1929 (Copepoda, Calanoida, Pontellidae)	
from Sebatik Island, North Kalimantan, Indonesia, with notes on its species-group	73–84
Djunijanti Peggie	
Biological aspects of <i>Papilio peranthus</i> (Lepidoptera: Papilionidae) as observed at Butterfly Research Facility - LIPI, Cibinong, Indonesia	85–102
Susan M. Tsang and Sigit Wiantoro Review - Indonesian flying foxes: research and conservation status undate	103 113
NEVIEW - INCOMESIAN ITYM9 TOXES, TESCAICH AND CONSETVALION STATUS UDGALE	1114_11