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Biomedical materials for wound dressing: recent advances and applications

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Wound healing is vital to maintain the physiological functions of the skin. The most common treatment is the use of a dressing to cover the wound and reduce infection risk and the rate of secondary injuries. Modern wound dressings have been the top priority choice for healing various types of wounds owing to their outstanding biocompatibility and biodegradability. In addition, they also maintain temperature and a moist environment, aid in pain relief, and improve hypoxic environments to stimulate wound healing. Due to the different types of wounds, as well as the variety of advanced wound dressing products, this review will provide information on the clinical characteristics of the wound, the properties of common modern dressings, and the *in vitro*, *in vivo* as well as the clinical trials on their effectiveness. The most popular types commonly used in producing modern dressings are hydrogels, hydrocolloids, alginates, foams, and films. In addition, the review also presents the polymer materials for dressing applications as well as the trend of developing these current modern dressings to maximize their function and create ideal dressings. The last is the discussion about dressing selection in wound treatment and an estimate of the current development tendency of new materials for wound healing dressings.

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1. Introduction

A wound is a type of injury that causes a loss of continuity in the skin, tissues, and mucous membranes. Based on the time and characteristics of the healing process, wounds are classified as acute wounds and chronic wounds. While acute wounds are skin wounds or surgical wounds, chronic wounds are pressure ulcers, leg ulcers, severe burns, and diabetic ulcers.¹ Acute wounds often heal completely and without complications within four weeks, following the expected or predictable rate of healing.² Compared to acute wounds, chronic wounds are more difficult to control the wound condition with slow healing time, persistence, and abnormal healing progress and can cause serious complications that require tissue removal.¹ Both acute and chronic wounds have a severely detrimental impact on the world's healthcare systems and economies.^{3,4} In the United States, injury treatment costs about 50 billion dollars annually.⁵ According to Medicare, a health insurance program, the cost of wound care for beneficiaries ranges from 28.1 to 96.8 billion dollars per year, with surgical wounds and diabetic ulcers the most expensive, accounting for 38.3 and 18.7 billion dollars, respectively.⁶ Chronic wounds are common in elders over 65

years old.⁷ Therefore, with the aging population, the number of patients with chronic wounds is expected to increase and continues to be a long-lasting problem in this population.⁸ Modern dressings are commonly used to cover wounds and generate a moist environment for wound healing. Traditional wound dressings are often used in clinical practice because they are economical, yet fibers stick to the granulation tissue, causing pain when removing the dressing. On the other hand, modern dressings maintain ideal temperature and humidity for the wound to stimulate wound healing and protect the wound from external bacteria and prevent cross-infection.⁹ Moreover, some types of dressing including alginate or hydrogel dressing have the property non-adhesive to tissues, causing less pain during dressing changes for patients, overcoming the limitation of traditional dressing.^{10,11} With their advantages, many studies on modern dressings such as foams, hydrogels, alginates, hydrocolloids, and films are carried out to solve clinical problems in treating wounds.

Here, we provide an overview of the modern dressings that have been developed for wound healing applications. We look over studies on the effectiveness of different dressings and consider the advantages and limits of wound treatment. In addition, the review also mentions new materials that are of interest to researchers for dressings applications in the future. Finally, we discuss the selection of modern dressings, and the trend of developing advanced materials to develop appropriate dressings in clinical treatment.

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2. Physiology of wound healing and treatment

2.1. Physiology of wound healing

Wound healing is a complicated biological process that restores tissue integrity. Acute wounds usually heal in a relatively short time frame from four to six weeks, depending on the size, depth, and extent of damage in the epidermis and dermis of the skin and the operation of growth factors, cytokines, and matrix proteins.¹² Physiologically, an acute wound healing is divided into five stages: hemostasis, inflammation, proliferation, re-epithelialization, and remodeling (Fig. 1).¹³

Hemostasis is the first stage of wound healing. Clotting factors are activated in hemostasis and form a platelet knot to reduce blood loss from the injuries.¹⁴ When bleeding is under control, growth factors such as vascular endothelial growth factor (VEGF), epithelial growth factor (EGF), and cytokines are released to recruit neutrophils, monocytes, and lymphocytes to reach the injured tissue and promote the inflammatory stage.¹⁵ The inflammatory stage involves a series of responses involving neutrophils and cytokines.¹⁶ The cells participate in clearing away cells debris, and pathogens as well as releasing cytokines such as tumor necrosis factor α (TNF- α), interleukin 6, 1 β (IL-6,

IL-1 β) to thrombolysis. The third stage is proliferation, in which platelets and leukocytes release cytokines, stimulate angiogenesis, fibroblast proliferation, collagen, and elastin synthesis to restore the dermis, leading to scar formation.^{15,17} The re-epithelialization stage is the re-establishment of intact epidermis over the newly formed tissue. Cells enhance collagen and elastin synthesis to increase skin elasticity and stability. Keratocytes migrate into the wound site while proteases released by macrophages remove excess extracellular matrix (ECM).¹⁸ The last stage of wound healing process is remodeling. During this stage, recently formed capillaries regress and most macrophages and fibroblasts undergo apoptosis.¹⁹

Chronic wounds last more than 12 weeks and heal at a much lower rate than acute wounds.²⁰ Chronic wounds have abnormal sequences of epithelial regeneration due to poor tissue blood supply, tissue necrosis, infection, persistent trauma, cancer, and a high amount of matrix metalloproteinase (MMP). In addition, chronic wounds have a low rate of cell division and high levels of pro-inflammatory cytokines and proteases.²¹ Other factors affecting wound healing include nutritional deficiency, vitamin-C, zinc deficiency, and hormone deficiency such as insulin in diabetics.^{22,23} These factors prolong the inflammatory stage and interfere with wound healing by continually

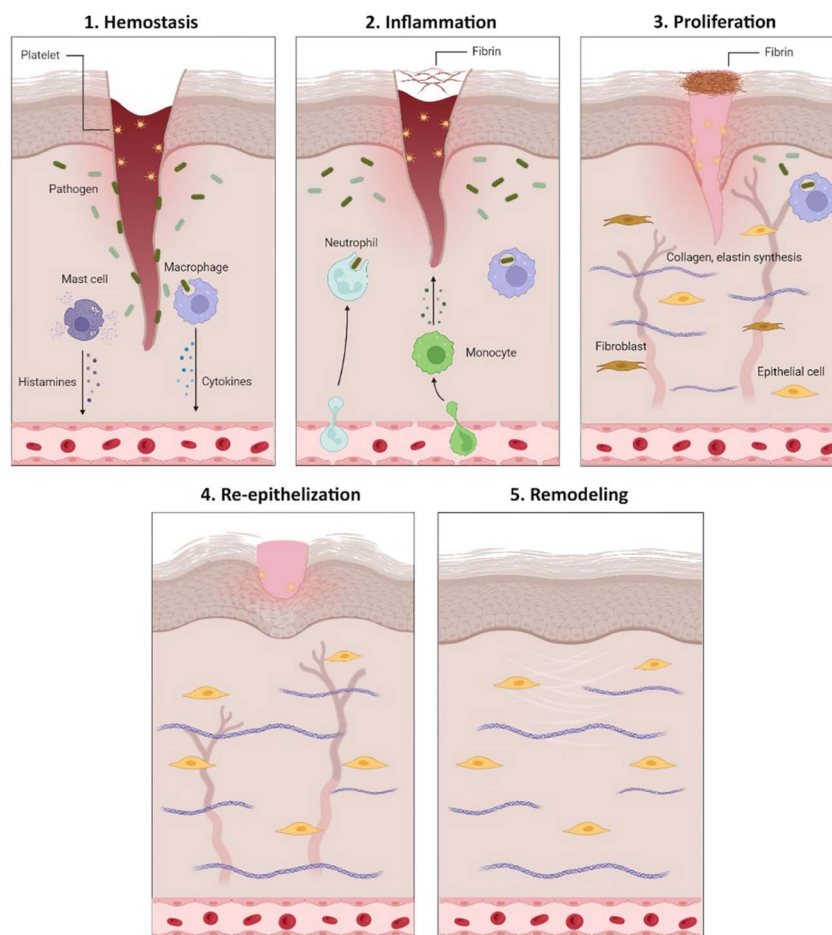


Fig. 1 Cell responses during five stages of acute wound healing.



attracting macrophages and neutrophils to the wound.^{24–26} Due to abnormalities in the successive stages of chronic wound healing, tissue and epithelial regeneration are disrupted, and the wound becomes a severe ulcer.

2.2. Wound treatment

In wound treatment, wound dressings cover damaged skin, maintain a moist environment and appropriate temperature required for healing, provide physical and microbiological protection, absorb excessive tissue fluid, and provide pain relief.⁹ In the 1960s, medical experts recommended that humidifying the wound environment dramatically affects the wound healing process.^{27,28} Dried wounds with eschar require extra moisture to optimize healing and soften the eschar.²⁹ On the other hand, excessive moisture leads to excessive hydration and damages the skin normal barrier function, causing ulceration.³⁰ Since then, various dressings have been designed to optimize moisture levels and create the ideal wound healing environment. Moreover, an ideal dressing not only protects a wound and maintains wound humidity but is also biocompatible, biodegradable, non-toxic, and non-allergenic while promoting gas exchange, granulation, and re-epithelialization.³¹ Modern wound dressings may also contain pharmacologically active substances such as antibiotics, non-steroidal anti-inflammatory, analgesic, and local anesthetics medicines or natural extracts with anti-inflammatory, epithelializing, antioxidant, and antimicrobial properties.^{32,33} Different wounds have different physiological conditions, depth, location, and extent of the wound, the amount of discharge, infection, and wound adhesion. Therefore, it is necessary to choose the proper dressing to promote the healing process or make it worse. Therefore, healthcare workers have to understand the condition of the wound and the characteristics of each dressing to choose appropriately and promote the healing rate and quality of healing.

3. Classification of modern wound dressings

For many years, traditional dressings such as cotton wool, lint, gauze have been widely used to ensure the wound clean and prevent getting bacteria infection. However, the dressings easily stick to the wound and do not create a suitable moist environment. Modern dressings have been developed with better-improved biocompatibility, degradability, pain relief, and moisture retention. Rather than just covering wound itself, modern wound dressings also act as facilitation for the function of the wound.³⁴ Several modern dressings currently used in clinical practice include hydrocolloid, alginate, hydrogel, foam, and film dressings (Fig. 2).

3.1. Hydrocolloid dressings

3.1.1 General characteristics. Hydrocolloid dressings consist of a hydrophilic and self-adhesive colloid granule coated with an external waterproof polyurethane (PU) film (Fig. 3).^{35,36} Colloid granules are commonly made of gelatin, pectin, and

carboxymethyl cellulose (CMC)^{35,37} and are available in various shapes, sizes, and thicknesses.³⁷ The outer layer protects the wound from bacteria, exotic agents, or other environmental impacts.³⁵

Hydrocolloid dressings can absorb a relatively large amount of wound fluids³⁸ as well as be virtually impermeable to water vapor, promoting the formation of a moist healing environment.³⁹ Furthermore, they are impermeable to oxygen, which accelerates epithelialization and collagen synthesis and decreases the wound exudates' pH, thus reducing the number of bacteria.⁴⁰ They also prevent contamination, promote autolysis to remove damaged or infected tissues, and do not require secondary dressings.^{35,37,41}

Hydrocolloid dressings are often incorporated with active ingredients in treating pressure ulcers or lower-extremity ulcers. They are indicated for low to moderate exuding wounds,⁴⁰ granular or necrotic wounds, and other acute wounds, including partial and full-thickness burns,³⁵ surgical or post-surgical wounds in children.⁴² Some hydrocolloid dressing products are shown in Table 1. However, this dressing is unsuitable for high exudate because there might lead to accumulation around the wound site.⁴³ To address the issues, the dressings need to be changed many times a week. Also, because of the adhesive nature that can damage the fragile surrounding skin, hydrocolloid dressings should not be used on infected wounds.³⁵

3.1.2 Trials and research. Since the first product was released to the market in 1982–1983, numerous studies have been conducted demonstrating the effectiveness of hydrocolloid dressings. Sung *et al.* evaluated the wound healing effect and antibacterial activity of hydrocolloid dressings containing benzalkonium chloride in *in vitro* and *in vivo* models. The results showed significant antibacterial activity against *Strep-tococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa*. The excision, infection, and abrasion wound sizes were reduced after using hydrocolloid dressings containing benzalkonium chloride on rat models.⁴⁴ In another study in mice with diabetes, there was a rapid decline in inflammatory M1 macrophages and the emergence of anti-inflammatory M2 macrophages. It also showed signs of wound healing, including re-epithelialization and angiogenesis.⁴⁵ In Sung and Lee's study, using DuoDERM Extra Thin hydrocolloid dressing on 12 neonatal extravasation injuries, all wounds healed with no deficiency in function and conspicuous scars.⁴⁶ Hydrocolloid dressings were proved to prevent nasotracheal tube-related pressure injury and improve significantly the endurance of the nasal skin in pediatric patients.⁴⁷ In the study by Shinohara *et al.*, ceramide-containing hydrocolloid dressings were investigated for hand-foot skin reaction (HFSR) on the soles of the feet in patients treated with sorafenib for metastatic renal cell carcinoma. The results indicated that 29% of the patients with grade 1 HFSR receiving hydrocolloid dressing containing ceramide developed to grade 2 and 3, lower than the control group with nearly 69%. Furthermore, the meantime for group A to have severe HFSR to grades 2 and 3 was longer than in group B, which meant that patients using hydrocolloid dressings containing ceramide might prolong time leading to the HFSR



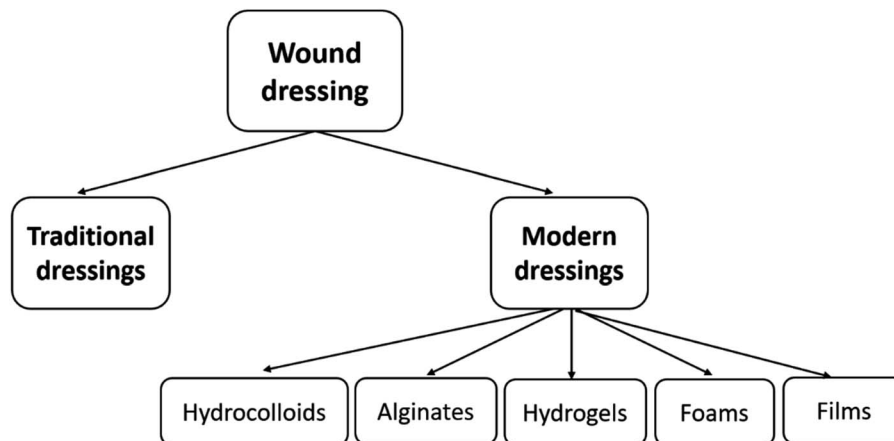


Fig. 2 Wound dressing classification.

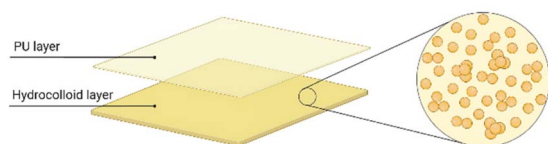


Fig. 3 Two layers structure of hydrocolloid dressings.

progression to grade 2 or 3. Therefore, ceramide-containing hydrocolloid dressing prevented the exacerbation of HFSR caused by sorafenib in patients with metastatic renal cell carcinoma.⁴⁸ Another clinical trial conducted by Sabando *et al.* showed that the novel hydrocolloid based on pectin, starch and plant extract decreased about 50% of topical edematous response. The pressure ulcer close completely without any adverse reactions.⁴⁹

3.2. Alginate dressings

3.2.1 General characteristics. Alginate is a natural polysaccharide extracted from brown marine algae, including *Laminaria* or *Ascophyllum*.⁷⁵ Alginate is also found in bacterial capsules of *Azotobacter* sp. and *Pseudomonas* sp.⁷⁶ It is a derivative of alginic acid and salts, such as calcium or sodium (Fig. 4).³⁵ The alginic acid structure consists of a linear copolymer of α -L-glucuronic acid and β -D-mannuronic acid and the glycoside linkage between these two saccharides influences the physical properties of the product.⁷⁷

Alginate is a popular biomaterial known for retaining a structure similar to ECM, exhibiting high biocompatibility. Alginate dressings absorb exudates from the wound, creating a moist environment for wound healing.⁷⁸ When applied to the wound surface, alginate forms a gel and easily sloughs when removing the dressing or rinsing with sterile saline.⁷⁹ It can be in the form of porous sheets or fibrous dressings when further processed.⁸⁰ Alginate performs high absorbent properties but also maintains structural integrity stability.^{81–83} Additionally, alginate dressings can reduce inflammation, wound odor, act as hemostatic agents, and have good permeability to oxygen, other

gases, or liquids.⁸⁴ Therefore, alginate is suitable for treating acute and chronic wounds such as diabetic foot ulcers, pressure ulcers, burns, and infected surgical wounds.⁷⁵

Since alginate cannot self-adhesive, a secondary dressing is required.⁸⁵ In addition, if the alginate does not absorb enough fluid to form the gel, it is possible to leave excess fibers in the wounds due to the fibrous nature of alginates.⁸⁶ This phenomenon may trigger inflammatory mechanisms against foreign agents. There had been reports of an allergy where there was insufficient moisture in the wound to form a gel⁸⁷ despite alginate's high biocompatibility.

3.2.2 Trials and research. Alginate dressings have been investigated for their effectiveness in many *in vitro*, *in vivo* studies, and clinical trials, showing that the dressing improves healing, hemostasis, and cell proliferation. Various alginate-based dressings in the market are presented in Table 1. In one study, alginate containing povidone-iodine and silver nanoparticles (AgNPs) had a marked affinity for microorganisms and required fewer dressings.⁸⁸

With high biocompatibility, to increase the effectiveness of treatment, the current bandage combines alginate with many antibacterial or anti-inflammatory compounds such as ZnO nanoparticles or Edaravone. The results of the *in vivo* model showed that the above combined materials are safe, capable of closing the wound through keratinocyte adhesion, cell proliferation⁸⁹ or reducing inflammation.⁹⁰ Shafei *et al.* study promotes tissue regeneration by exosome loaded alginate hydrogel.⁹¹ This bioactive dressing not only improved wound closure, collagen synthesis, and vessel formation but also was biodegradable and biocompatible.⁹¹ In addition, alginate is also combined with many naturally derived extractions such as *Malva sylvestris*⁹² or oregano essential oil,⁹³ which shows antimicrobial,⁹³ anti-inflammatory and high biocompatibility properties.⁹²

Clinically, alginate dressing had been shown to be effective in healing processes, causing less pain during dressing changes for patients.^{11,94} Another randomized study evaluated the clinical therapeutic effect of human granulocyte-macrophage colony-stimulating factor incorporated with alginate dressings





Table 1 Examples of commercially available modern dressings

Type	Product name	Components	Main features	Applications	References
Hydrocolloid	DuoDerm®	Pectin, gelatin, CMC and PU	The outer layer provides a waterproof barrier The matrix of hydrocolloid particles absorbs exudate They are keeping a moisture environment	Managing wounds with light to moderate exudate: Stage I and Stage II pressure ulcers, burn/scald, abrasions, lacerations, and reducing infection	50
	Comifeel® Plus	NaCMC and calcium alginate is added	Allows moisture to evaporate while maintaining a moist healing environment Water and bacteria proof to protect from external factors	Chronic wounds such as leg ulcers and pressure ulcers; acute wounds such as superficial burns, superficial partial-thickness burns, donor sites, postoperative wounds and skin abrasions	51
	Cutinova® Hydro	Absorbent colloid covered with semioclusive polyurethane film covering	Selective absorption: water is taken into the matrix and locked away Highly absorbent granules: absorb the fluid 10 times its own weight Does not form a gel substance	Venous leg ulceration, diabetic ulcers, slough, and necrotic tissues	52
	Replicare®	Absorbent colloid covered with polyurethane film	Cohesive properties keep the wound free of dressing residue	Partial to full thickness wound: ulcers (venous, arterial, diabetic) pressure sores, donor sites, surgical incisions and excision, burns (grade 1st and 2nd)	53
Alginate	Kaltostat®	Calcium alginate with guluronic acid	Creation and maintenance of a moist wound environment Supporting moist wound healing environment Promoting haemostatic, stop minor bleeding	Moderate to highly exuding wounds and for wounds with minor bleeding: leg ulcers, pressure ulcers, diabetic ulcers and fungating lesions, donor sites, abrasions, lacerations and post-surgical wounds	54
	Algicell® Ag	Calcium alginate with 1.4% silver	Calcium alginate with 1.4% silver Minimizing fibrous residue Maintaining a moist wound environment	Diabetic foot ulcer, leg ulcers, pressure ulcers, donor sites, and traumatic and surgical wounds	55
	Guardix-SG®	Containing sodium alginate and poloxamer	Antibacterial activity: <i>Staphylococcus aureus</i> , <i>Pseudomonas aeruginosa</i> , <i>Escherichia coli</i> , <i>Candida albicans</i>	Reduce the adhesion upon spine, thyroids, and abdominal surgery	56
	Algivon®	Alginate dressing impregnated with manuka honey	Prevents adhesion by forming a physical barrier on the surface of the wound tissue Anti-bacterial and anti-inflammatory Maintaining the ideal moist wound healing environment	Eliminates odors Apply for pressure ulcers, leg ulcers, diabetic ulcers, surgical wounds, burns, graft sites, infected wounds	57
	Fibracol™ Plus	90% collagen and 10% calcium alginate	Maintaining physiologically moist microenvironment to support granulation tissue formation, epithelialization and rapid wound healing	Full-thickness and partial-thickness wounds: venous ulcers, ulcers caused by mixed vascular etiologies, diabetic ulcers, second degree burns, donor sites and other bleeding surface wounds	58
	Tromboguard®	Alginate, chitosan, polyurethane	Haemostatic activity and control severe bleeding Antibacterial activity	Used to stop bleeding in traumatic and postoperative wounds, bleeding from accidents	59 and 60



Table 1 (Contd.)

Type	Product name	Components	Main features	Applications	References
Hydrogel	ActivHeal [®]	85% of water and collection of polymer chains	Rehydrating dry necrotic wounds Providing a moist wound environment Removing devitalized tissue Encouraging growth and migration of epithelial cells	Dry and sloughy wounds with nil to low exudate: pressure ulcers, cavity wounds, leg ulcers, graft and donor sites, diabetic ulcers, post op surgical wounds, lacerations and abrasions	61
	Restore [®]	Sodium polyacrylate, hyaluronic acid	Providing a moist wound environment Non-greasy, non-staining, and fragrance free	Stages II–IV pressure ulcers, diabetic skin ulcers, venous ulcers, 1st and 2nd degree burns, skin tears, cuts, abrasions, and conditions associated with peristomal care	62
	Suprasorb [®] G	CMC polymers, polyethylene and acrylic polymers	Gently removing necrotic tissue Supplying moisture Relieving wound pain Easy to mould Drying wounds and wounds with low amounts of exudate	Dry wounds and wounds with low amounts of exudate, acute or chronic superficial wounds (lower leg ulcers, pressure ulcers, first- and second-degree burns, scalds)	63
	Aquaderm [™]	2-Acrylamido-2 methyl-1 propanesulfonic acid sodium, poly(ethylene glycol) dimethacrylate, 2-hydroxy-2-methylpropiophenone with 38–55% water	Providing soothing coolness Non-traumatic, one-piece removal Supporting moist wound healing Bacteria and fluid barrier Absorbing minor exudate Transparent - easy to monitor wound Can be cut to fit wound site	Pressure ulcers, minor burn, radiation tissue damage	64
	Neoheal [®]	90% water and polyvinylpyrrolidone, PEG, agar	Maintaining of an optimal moist environment Cooling, soothing local pain and discomfort Protecting wounds from external contamination Absorbing wound secretions Permeable to water vapor and oxygen but impermeable to bacteria Allowing the dressing to be removed without trauma Conforms easily to any contour of the body Transparent to enable wound observation and assessment of healing process Biocompatible, non-cytotoxic, non-allergenic	First degree of burn wounds, second and third degree of burn wounds, ulcerations, bedsores	65

Table 1 (Contd.)

Type	Product name	Components	Main features	Applications	References
Foam	Allevyn life	Hydrophilic polyurethane foam	Providing coverage to the malleolus without the need for secondary retention Fitting the contours of the human body securely and allowing patients to shower Minimizing the visual impact of strikethrough, providing the patient with confidence that their dressing may not attract the negative attention of others Allowing the dressing to be repositioned Transporting rapidly wound exudate and ensuring a balanced moist wound environment Bounding safely germs and cell debris even under compression Soft and smooth with good padding properties Available in different sizes and shapes Preventing the dressing from sticking to the wound Absorbing wound exudate and releasing moisture Non-border dressing can be cut into the desired size and use a suitable secondary dressing Soft and conformable making it easy to use and for many situations Minimizing pain during dressing changes Minimizing the risk of leakage Inactivate microbial in wound (bacteria and fungi)	Pressure ulcers	66
	Permafoam®	Absorbent foam made of polyurethane		Medium to heavily exuding wounds, deeper wounds	67
	HydroTac®	Hydrated polyurethane polymer and propylene glycol		Wounds during the granulation and epithelialization phases with low to moderate exudation	68
	Mepilex Ag	Polyurethane foam containing Ag		Acute and chronic wounds	69
	Tegaderm™ Silicone foam dressing	Polyurethane foam with silicone adhesive	Absorbing and evaporating moisture to help reduce the potential for skin maceration Ability to access and assess skin Ability to manage microclimate Ease of application and removal Correcting dressing size for high-risk locations	Pressure injuries, venous leg ulcers, neuropathic ulcers, arterial ulcers, skin tears, and surgical wounds	70
Film	Bioclusive™ Plus	Transparent polyurethane film coated with acrylic adhesive	Providing a barrier to viruses 27 nm in diameter or larger and bacteria Allowing the transmission of oxygen and moisture vapor Preventing maceration by inhibiting the lateral movement of exudate	No or light levels of exudate, suitable for secondary cover dressing and securing of catheters; minor burns, donor sites, superficial pressure areas and leg ulcers Skin tears or abrasions, surgical excisions, second-degree burns, blistering conditions such as	71
	Mepitel	Two-sided wound contact layer coating with silicone			72



Table 1 (Contd.)

Type	Product name	Components	Main features	Applications	References
	Mepore®	Polyurethane coated with polyacrylic adhesive	Gentle adhesion helping remove with minimum pain and without damaging new tissue Bacterial and viral barrier for microbes larger than 25 nm Allowing moisture and skin transpiration to be evaporated	epidermolysis bullosa, partial and full thickness grafts, and skin damage following radiotherapy or steroid therapy Secondary dressing for fixation of medical devices such as wound dressings, tubes and cannulas	73
	Transeal®	Polyurethane coated with acrylic, pressure-sensitive adhesive	Maintaining a proven high moisture vapor transmission rate Impermeable to water and bacteria	Vascular access sites, first and second degree burns, superficial wounds, surgical incisions	74

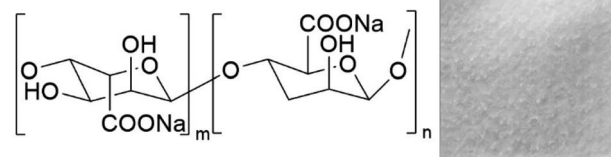


Fig. 4 Alginate dressing and chemical structure of sodium alginate.

in the treatment of refractory chronic skin ulcers. The combination showed many advantages, such as promoting granulation tissue growth, accelerating epithelial regeneration, and effectively alleviating wound pain, thus improving the patient's quality of life.⁹⁵

3.3. Hydrogel dressings

3.3.1 General characteristics. Hydrogels are hydrophilic three-dimensional polymer chains that can absorb a large volume of water due to the presence of hydrophilic moieties (Fig. 5).¹⁰ Based on the origin of component polymers, there are three types of hydrogels: natural, synthetic, and hybrid or semi-synthetic hydrogels.^{10,96} Hybrid hydrogels are mainly used as they possess both the high biocompatibility of natural polymers and elastic mechanical properties of synthetic polymers.^{10,96} Hydrogels can be applied either as an amorphous gel or as an elastic, solid sheet, film.^{10,31}

Hydrogel dressings can provide a moist environment in the wound site and make favorable conditions for tissue regeneration. In addition, this characteristic also gives a comfortable, soothing effect to the wound, especially severe wounds.^{97,98} However, this fluid accumulation can also lead to skin infections and bacterial growth, giving off foul odors in infected wounds.^{31,97} Moreover, the hydrogel wound dressing is non-adhesive to the wound or tissues, releasing pain during dressing changes yet not disrupting wound healing.^{83,96,99} The tuneable mechanical properties of hydrogels enhance their suitability for various wounds.⁹⁷

Hydrogel dressing products are usually suitable for pressure ulcers, diabetic foot ulcers, skin tears, and surgical wounds and burns, including minor burns, first and second-degree burns.^{10,99} Hydrogel dressings should be used either with a secondary dressing such as film or foam or without secondary dressing.⁹⁹

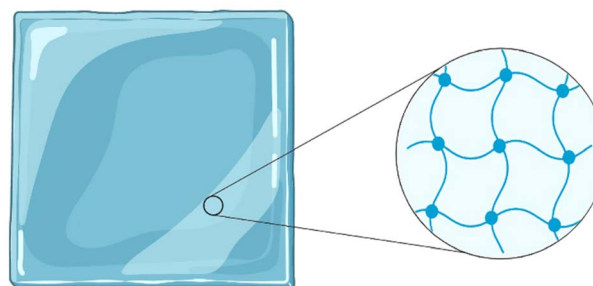


Fig. 5 Structure of hydrogel dressings.



3.3.2 Trials and research. Hydrogel wound dressings are a new type of high-end material with the necessary characteristics of ideal wound dressings due to the unique and flexible physicochemical properties.⁹⁷ For instance, a multifunctional hydrogel was prepared by coordinative cross-linking of multi-arm thiolated polyethylene glycol (SH-PEG) with silver nitrate followed by loading of an angiogenic drug, desferrioxamine.¹⁰⁰ This study by Chen *et al.* revealed that this versatile multifunctional hydrogel is a potential material in regeneration, particularly in diabetic skin wounds and open wounds, due to its flexible response with a high risk of infections and external mechanical stimuli.¹⁰⁰ Another hydrogel dressing was synthesized lately, named EHO-85.¹⁰¹ This new dressing is specially applied for moist wounds with antioxidant properties, capable of maintaining acidic environments, accelerating the healing of hard-to-heal chronic wounds and stimulating granulation tissue formation on other types of wounds.¹⁰¹ This outstanding characteristic can be explained based on the capability of adjusting pH and its antioxidant properties.¹⁰¹ Moreover, there are also many other trials and studies conducted to evaluate the effectiveness of hydrogel dressings in treating various types of wounds.^{102–104} A large range of hydrogel-related wound care products available in the market (Table 1).

Furthermore, there has been a growing interest in incorporating nanoparticles or nanostructures into hydrogels for improving the properties of hydrogel wound dressings.^{97,105} Silver is one of the most common nanomaterials combined with hydrogel dressings to enhance the treatment of infected wounds.^{105,106} Many randomized controlled trials were carried out to evaluate the effectiveness of hydrogel/AgNP dressings and showed the positive results in wound healing.^{105,106} However, the limitation of hydrogel/AgNP dressing is the high cost.¹⁰⁶ Besides AgNP, another study improved the mechanical of hydrogels by incorporating nano-clay with polydopamine and polyacrylamide.¹⁰⁷ This hydrogel displayed superior toughness owing to nanoreinforcement by clay and polydopamine-induced cooperative interactions with the hydrogel networks.¹⁰⁷ In addition, a new generation of smart hydrogel wound dressings that contain sensors has been developed rapidly for the ability to reveal wound conditions. Some remarkable studies can be mentioned such as flexible pH-sensing hydrogel fibers based on alginate in skin wounds,¹⁰⁸ polyvinyl alcohol/xyloglucan (PVA/XG) hydrogel membrane with the ability to absorb exudate and release biological factors.¹⁰⁹

3.4. Foam dressings

3.4.1 General characteristics. Foam is a porous structure that has the ability to absorb fluids into air-filled spaces based on capillary action.¹¹⁰ The most common foam dressings are made of polyurethane.¹¹⁰ Similar to the characteristics of hydrogel, foam dressing can maintain a moist environment around wounds, provides thermal insulation, and is highly absorbing, which is controlled by foam properties such as texture, thickness, and pore sizes.³¹ The porous structure with high absorbency makes foam dressing suitable for many exuding wounds.^{31,110}

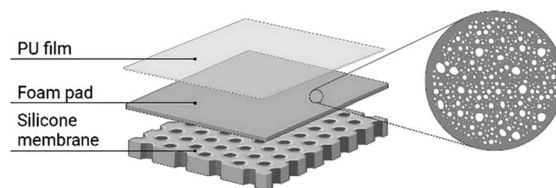


Fig. 6 Three layers structure of foam dressings.

To provide water and microbial resistant barrier to the environment, foam dressings are commonly supplied with a film-backing¹¹⁰ or a silicone membrane¹¹¹ for keeping the dressing in the right wound site and protecting the wound from trauma when changing the dressings (Fig. 6). Foam dressings can be kept for up to a week, depending on the level of exudate.¹¹² Nevertheless, this advantage is also the disadvantage of foam dressing because not frequent changes may affect the growth of new tissue and damage wounds when removing the dressing.¹¹³

Foam dressings are used to treat minimal to moderate wounds, proper for burns, chronic wounds, and deep ulcers.¹¹⁴ However, these dressings are not appreciated for epithelial dry wounds, necrotic wounds, and wounds requiring frequent care.

3.4.2 Trials and research. A study investigated the effectiveness of three commercial available foam dressings, namely Mepilex Border Flex (MxBF), Allevyn Life (AL), and Optifoam Gentle EX (OGEX) in treating chronic wounds, then confirmed that these foam dressings have particular effectiveness in treating wounds, especially chronic wounds.¹¹⁵ Moreover, many available foam products are integrated with silver component to enhance antibacterial activities. A review is conducted to analyze the results of *in vitro* trials to clinical data in the range of three foam dressings: Mepilex Ag, Mepilex Border Ag, and Mepilex Transfer Ag.¹¹⁶ The data showed that the silver foam dressings are effective in wound control, addressing local signs of infection in both acute and chronic wounds.¹¹⁶ These dressings are considered easy to use, provide a favorable environment to heal wounds, and do not damage when removed. Besides, treating wounds by using these dressings is cheaper than using antibiotics.¹¹⁶ In addition, the outstanding features and applications of some other foam products on the market are also provided in Table 1.

To meet the demand for treating various wounds, researchers have developed many new foam dressings. Namviriyachote *et al.* developed an innovative PU foam dressing comprising hydroxypropylmethylcellulose and alginate, containing silver and asiaticoside (AS).¹¹⁷ The findings showed that the foam dressing released AgNP at a 1 mg cm⁻² of silver dosage in the formula with 6% alginate and 5% AS for the most optimal antibacterial activity. This dressing improved wound healing both wound closure rate and histological parameters of skin wounds and had no dermatological reactions occurred.¹¹⁷ Another new foam dressing is mentioned to be the povidone-iodine foam dressing, known as Betafoam. To provide evidence about the effectiveness of Betafoam, a clinical trial was performed comparing Betafoam with Medifoam,



a commercially available foam dressing for treating diabetic foot ulcers.¹¹⁸ The trial suggests that Betafoam is safe, has a wound healing ability similar to Medifoam, and has no adverse effects.

3.5. Film dressing

3.5.1 General characteristics. Film dressings are thin, flexible, and transparent polyurethane sheets, designed to adhere to the wound-surrounding skin and maintain moisture in the healing environment.^{35,38} Film dressings are semi-permeable because of their permeability to water vapor, oxygen, and CO₂ but not water and microorganisms.³⁴ The solid adhesive property of film dressings allows them to be placed on moving surfaces such as joints but can cause damage to the wound-surrounding when removing the dressings.^{86,112} Initially, the film was made from a nylon derivative with an adhesive PU framework and was not used for exudative wounds due to its limited absorption capacity.³¹ Due to its highly elastic characteristics, and transparent nature which is appropriate for checking wound closure without removing,⁸⁶ these dressings are commonly used to cover newly healed and superficial wounds, including intravenous catheter sites and split skin graft sites.⁸⁵ However, one issue that needs to be addressed is preventing fluid accumulation beneath the films. The excess exudates can inactivate the adhesive,¹¹⁹ lead to maceration and break the seal to the external environment, thus facilitating the proliferation of bacteria.¹²⁰

3.5.2 Trials and research. In the study by Jafari *et al.*, the PU nanocomposite membrane consisting of high-molecular chitosan and titanium oxide reduced the amount of *Pseudomonas aeruginosa* bacteria by 63–69% independent of the concentration of nanochitosan.¹²¹ Li *et al.* had developed a multifunctional film dressing that combined segments of aniline trimer (AT), polyethylene glycol (PEG), and polycaprolactone (PCL). The combined PEG-PCL-AT dressing exhibited biocompatibility in both *in vitro* and *in vivo* models, antibacterial activity, and free radical scavenging ability, promoting wound healing. Furthermore, PEG-PCL-AT dressing with 12 wt% AT promotes collagen deposition and granulation tissue thickening.¹²² In addition, PU film dressings are also used to load drugs or antibiotics such as procaine.¹²³ Tang *et al.* developed a multifunctional elastomer film containing cetyltrimethylammonium bromide (CTAB). This dressing showed a superior antibacterial activity, with the bactericidal rate up to 90% within 12 h. In addition, the notably high collagen deposition proved the effectiveness in wound closure and healing process. The information about wound condition such as pH, temperature, and glucose level are provided real-time thanks to the fabricated sensor array within the dressing.¹²⁴ M. Kazanavičius *et al.* compared four types of dressing, including polyurethane (Mepilex), polyurethane with silicone membrane (Mepilex border), transparent breathable film (TBF; Mepitel film), and cotton gauze dressings. The wound healing time in the TBF group was the fastest, about ten days. Patients in the TBF group showed 66.7% of the donor sites healed by day 9, and painful feeling in this group was the mildest and shortest.¹²⁵ Another

study used hydrofilm on breast cancer patients and consequently, the severity of radiation dermatitis due to the whole-breast irradiation was reduced. Hydrofilm has also been shown to reduce erythema and hyperpigmentation, completely prevent scaling, and significantly reduce symptoms of itching, burning, pain, and little inconveniences of patient's daily activities at the same times.¹²⁶ The characteristics of some film dressings are presented in Table 1.

4. Polymer materials for wound dressings

Polymers are commonly used in pharmaceutical and biomedical areas due to their biocompatibility and their similarity with ECM. The generated substrate can mimic the biological environment that helps cells participate in proliferation, differentiation,¹²⁷ and repair damaged tissue.¹²⁸ In addition, some polymers also have biodegradable and bioresorbable properties to promote the reconstruction of new tissue without inducing the inflammation.¹²⁹ In this part, a brief introduction of the representative polymers and their recent application was presented.

4.1 Cellulose and bacterial cellulose

Cellulose is a major polysaccharide in the cell wall of a plant. Cellulose comprises many glucose units in a linear form which are linked together by β -1,4.¹³⁰ With the presence of hydrogen bonds that hold the oxygen atoms and hydroxyl groups together, the linear structure of cellulose is well maintained and exhibited the biomechanics property.^{130,131} As cellulose is formed from the glucose subunits, it is naturally biocompatible with human tissue and can be easily modified without affecting the structural and mechanical properties.¹³¹ Due to the semi-crystalline property in an aqueous state, cellulose can present biological effects when modified. However, pure cellulose has a major drawback due to its poor solubility in organic solvents.¹³² At below 300 °C, cellulose is completely insoluble and degraded when above this temperature.¹³³ To improve the dissolution ability, cellulose derivatives include cellulose esters (cellulose acetate), ethers (carboxymethyl cellulose, methyl cellulose, and ethyl cellulose), and cellulose sulfate represent alternatives to pure cellulose.¹³³

In contrast to plant cellulose, some bacteria, algae, and fungi produce cellulose through oxidative fermentation, which is called bacterial cellulose (BC) or microbial cellulose.¹³⁴ The BC is usually synthesized by Gram-negative bacteria such as *Ace-tobacter*, *Agrobacterium*, *Komagataeibacter* (formerly *Gluconacetobacter*), *Achromobacter*, *Azobacter*, *Rhizobium*, *Salmonella* or Gram-positive bacterium *Sarcina ventriculi*.¹³⁵ BC is considered a more biocompatible version of plant cellulose since it is free of lignin, hemicellulose, and pectin.¹³³ Therefore, BC exhibits specific characteristics, high water holding capacity due to being very hydrophilic, a large surface area, high crystallinity, and high mechanical strength.^{133,135,136} BC absorbs well fluid from the wound due to high water holding capacity, the water molecules bind to the hydroxyl group in the cellulose chain. The



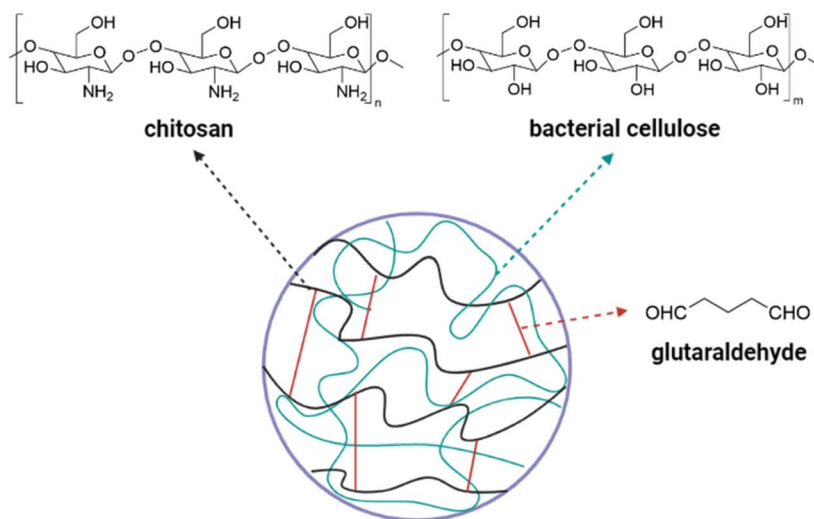


Fig. 7 Simple illustrated structure of semi-interpenetrating network.¹⁴⁰

high porosity combined with the large surface area suggests that BC can interact with antibacterial drugs or other active ingredients.¹³⁷ Despite the advanced characteristics, the high cost of BC production hinders industrial implementation. Therefore, the industrial wastes or by-products of fermentation media are utilized to improve the cost-effectiveness such as candied jujube waste water,¹³⁸ acetone-butanol-ethanol fermentation,¹³⁹ and pomegranate extract. Fig. 7 presented the semi-interpenetrating network of hydrogels based on fabricating BC and chitosan through glutaraldehyde linkage.¹⁴⁰

4.2 Collagen

Collagen is an abundant natural polymer found in the ECM, accounting for one-third of the protein in the body. Collagen exists mainly in epithelial and connective tissues such as bone, cartilage, ligaments, tendon, skin and is an essential component in cell interactions, regulation of cell anchoring, and cell migration.^{141,142} Cooperative with glycosaminoglycan, collagen has been an essential factor for cell attachment, proliferation, and differentiation.^{143,144} The high flexibility of collagen dressings is due to the three-dimensional structure, which can absorb liquids many times their weight thanks to the accessible surface and high capillarity.¹⁴⁵ Collagen dressings reduce the risk of secondary bacterial invasion by acting as chemo-attractant for neutrophils,¹⁴⁶ and affect other healing processes such as decreasing protease activity.¹⁴⁷ However, the disadvantages of collagen based-biomaterials are their rapid degradation rate and low stability, resulting in a significant loss of mechanical properties.¹⁴⁸

4.3 Chitosan

Chitosan (CS) is obtained by chitin deacetylation and extracted from the extracellular matrix of marine crustaceans, shrimp, crabs, shellfish, and some fungi.^{149,150} CS is a linear polysaccharide made up of D-glucosamine and N-acetyl-D-

glucosamine units. CS is a popular and renewable natural biomaterial for regenerative medicine with many valuable properties such as compatibility and biodegradability, inertness, allergenic, antibacterial, and hemostatic effects.^{151,152} It is also applied to treat wounds, especially chronic wounds, infections, or ulcers.^{151,153}

4.4 Hyaluronic acid

Hyaluronic acid (HA) is a natural polymer found in skin, lens, or synovial fluids, composed of D-glucuronic acid and N-acetyl-D-glucosamine disaccharide. HA is characterized by high viscosity and fast degradation rate.

HA is widely used in medical fields such as orthopedics and plastic surgery because its characteristics involve several structural properties of tissues, cell signaling, and critical elements of ECM.^{154,155} Additionally, HA plays a role in healing acute and chronic wounds by promoting early inflammation, increasing cellular infiltration, enhancing granulation tissue formation, and facilitating cell migration.^{156–158} HA also performs good swelling behavior due to the presence of the carboxyl group in the side chains and has a thick wall structure to support the strength of the material.¹⁵⁹ The swelling-reducing properties and stable structure may prove that HA can be a potential material for dressings. However, HA is sensitive to the molecular weight factor. According to Campo *et al.*, only medium molecular weight HA could enhance wound repair, while low molecular weights contributed to further inflammation, and high molecular weights may exert inflammatory pathologies such as rheumatoid arthritis.¹⁶⁰

4.5 Fibrinogen and fibrin

Fibrin and fibrinogen participate in various physiological functions such as fibrinolysis, cellular and matrix interactions, and the inflammatory response.¹⁶¹ In the wound healing process, fibrin and fibrinogen roles are expressed through



Table 2 Summary of recent studies on various types of polymer materials for wound dressings^a

Material	Name of dressing	Trial	Effectiveness	References
Cellulose	Cellulose nanocrystals and AgNPs	<i>In vivo, in vivo</i>	No toxic effects of the combination of cellulose and AgNPs Promoting rapid wound healing compared to control groups	174
	RPC/PB hydrogel	<i>In vitro, in vivo</i>	Exhibiting excellent antibacterial, skin tissue regeneration and wound closure capabilities	175
	Na CMC with merremia mammosa gel	<i>In vivo</i>	Not irritable, accelerating healing process through increasing collagen synthesis and angiogenesis	176
	Oxidized regenerated cellulose membrane	Clinical trials	Stop bleeding in patients with uncontrollable bleeding	177
Bacterial cellulose	BC reinforced chitosan-based hydrogel	<i>In vitro</i>	Showing good biocompatibility and excellent antibacterial activity against <i>E. coli</i> and <i>S. aureus</i>	178
	Dialdehyde carboxymethyl BC/CS composites	<i>In vivo</i>	Accelerating the wound healing rate and inhibit bacterial proliferation	179
	BC membrane	<i>In vivo</i>	Good biocompatibility and prevent fibrosis in trabeculectomy	180
	BC gel and associated film	Clinical trials	Decreasing significantly in the size of wound, lower dressing change frequency compared to group using Rayon®	181
	BC dressing	Clinical trials	Shorter healing time in managing second-degree burn wounds and skin graft donor sites compared to vaseline gauzes	182
Collagen	Modified collagen gel	<i>In vitro</i>	Enhancing macrophage attraction to the wound site, reducing proinflammatory virulence, promoting anti-inflammatory macrophage polarization, addressing wound inflammation, and improving angiogenesis	183
	Collagen-based composite dressing	Clinical trial	Forming granulation tissue, enhancing epithelialization, and having faster wound healing time	184 and 185
Chitosan	CS-based opticell dressing	<i>In vivo</i>	The total bleeding significantly decreased in excisional wounds mimicking debridement	186
	HemCon® dental dressing	Clinical trial	Pain values and post-extraction socket healing were lower after suture removal on treating anti-platelet patients	187
	Chitosan dressing	Clinical trial	Reducing wound size and wound depth on chronic, difficult-to-heal wounds such as diabetic ulcers, leg vein ulcers	188



Table 2 (Contd.)

Material	Name of dressing	Trial	Effectiveness	References
Hyaluronic acid	Incorporation of PVA/HA/cellulose nanocrystals as nanofiber	<i>In vitro</i>	Loading with L-arginine exhibited excellent proliferative and adhesive potential, high wound gap-closure, and showed antibacterial activity against <i>Klebsiella pneumoniae</i>	189
	0.2% HA	<i>In vivo</i>	Healing skin abrasions in rat's model	190
	PTE-NEs fabricated HA hydrogel	<i>In vitro, in vivo</i>	No toxicity, improve the wound healing through reducing inflammation, enhancing collagen synthesis, accelerating M2 macrophage polarization, and angiogenesis	191
	0.2% and 0.8% HA gel	Clinical trial	Complete epithelization. Pain and burning sensation scales were also lower. Color match scores were higher	192
	Healoderm	Clinical trial	The diabetic foot ulcer group had a higher complete healing rate, faster ulcer healing velocity, and shorter mean duration for achieving a 50% ulcer size reduction	193
Fibrinogen and fibrin	Fibrin combined with Na carboxymethylcellulose	<i>In vitro</i>	In the form of a mesh, supporting the fibroblast adhesion and proliferation, accelerating the wound healing	194
	Fibrin-based hydrogel load BNN6 mesoporous polydopamine nanoparticles	<i>In vitro, in vivo</i>	Clearing the infection of methicillin-resistant <i>S. aureus</i> through cell membrane and genetic metabolism damage under 808 nm laser irradiation. Accelerating wound healing through collagen deposition and the proliferation of hair follicles	195
	3D salmon fibrinogen and chitosan scaffold	<i>In vitro, in vivo</i>	The cell proliferate in the scaffold and the wound healing is more effective than the untreated group	196
	Alginate-fibrinogen-nisin hydrogel	<i>In vitro, in vivo</i>	Inhibiting the bacteria growth, accelerate the formation of blood clot, show the higher rates of wound healing, re-epithelialization, and collagen deposition	197
	Heterologous fibrin sealant	Clinical trial	Heterologous fibrin sealant is safe and non-immunogenic, showing good preliminary efficacy in chronic venous ulcers treatment	198
Polylysine	Gelatin nanofiber dressing contains ϵ PL	<i>In vitro, in vivo</i>	Eliciting bactericidal activity in burn wounds for fibroblasts migration and re-epithelialization. In partial thickness burns of porcine	199



Table 2 (Contd.)

Material	Name of dressing	Trial	Effectiveness	References
	Carbon dots and ϵ PL hydrogel	<i>In vitro, in vivo</i>	model, promoting wound closure and reduce hypertrophic scarring Having broad spectrum in antibacterial activity. Enhancing angiogenesis and epithelization that accelerate the wound healing rate	200
	Modified HA/ ϵ PL hydrogel	<i>In vitro, in vivo</i>	Killing bacteria in infected wound and improving the wound status in rat model	201
	ϵ PL modified natural silk fiber membrane	<i>In vivo</i>	Exhibiting thicker granulation tissue, higher collagen composition, help accelerate wound healing rate	202

^a AgNPs: silver nanoparticles, RPC: pH responsive cellulose, PB: poly(vinyl alcohol)/borax, CMC: carboxymethyl cellulose, BC: bacterial cellulose, CS: chitosan, PVA: poly(vinyl alcohol), PTE: *Poria cocos* triterpenes extract, NEs: nanoemulsions, HA: hyaluronic acid, BNN6: *N,N'*-disubutyl-*N,N'*-dinitroso-*p*-phenylenediamine, ϵ PL: ϵ -polylysine.

mediating both hemostasis and homeostasis.¹⁶² Fibrinogen molecules are made up of two sets of three different peptide chains including $A\alpha$, $B\beta$ and γ chains, and connected by disulfide bridges.¹⁶³ The formation of fibrin releases small peptides and is catalyzed by thrombin.^{161,164} Fibrin is highly extensible and has elasticity properties.¹⁶⁵ The scaffolds made of fibrin or fibrinogen can attain a high cell seeding efficiency and then proliferate, migrate, and differentiate into specific tissues/organs by secreting ECM.¹⁶⁶ Fibrinogen provides a surface for cellular attachment and proliferation and consists of a fibrous network for cell signaling and cell-matrix¹⁶⁷ which is a major advantage for healing. However, these proteins are easily degraded and have poor mechanical properties.¹⁶⁸

4.6 Polylysine

Polylysine is a cationic polymer synthesized through condensation polymerization or fermentation of amino acid lysine. Lysine is available in two chiral forms which are *L*-lysine and *D*-lysine resulting in α -polylysine or ϵ -polylysine.¹⁶⁹ ϵ -Poly-*L*-lysine is a naturally occurring polymer and presents various characteristics: water solubility,¹⁷⁰ biodegradability, non-toxic for drug delivery systems, biological adhesives,¹⁷¹ antibacterial ability.^{169,172} Because of its the biodegradability and biocompatibility characteristics, polylysine has raised the attention of medical applications.^{169,173} However, due to the limitation in mechanical properties, it should be modified or blended with other polymers to improve the mechanical strength.¹⁶⁹

All studies on dressings which are natural-based polymer for wound healing applications are summarized in Table 2.

5. Discussion

Using wound dressing for wound care and treatment is an effective method, proven by the positive results from numerous

experiments. Modern dressings are prioritized for research and development.^{31,203} Modern dressings ensure the essential factor of creating a moist environment for wound healing, pain reduction, and antibacterial. However, the effectiveness of these popular modern dressings is still limited to only a few clinical studies. Wound dressing efficacy trials are usually performed on flat wounds, with less complexity, creating an oversimplification of the actual clinical features.²⁰⁴

On the other hand, a wound is considered a complex clinical problem, and the effectiveness of treatment depends on several factors: diagnosis, patient comorbidities, anatomical location, physiological status, and wound size.^{203,204} Thus, the general recommendation is that dressing selection should be tailored to the wound and patient, under the guidance and consideration of physicians with expertise in wound treatment.^{112,205,206} That would help the patients receive the proper treatment with the right purpose, achieve optimal efficiency and limit unnecessary risks during treatment.

Table 3 shows the specific advantages, disadvantages, and application range of modern dressings for different wounds, which provide a clinical guideline for selecting suitable wound dressings for effective wound healing. In addition, it is still necessary to have more research and clinical trials to demonstrate the effectiveness and safety of various types of modern wound dressings in the future.

Developing new material platforms for modern dressings with positive results demonstrated through various tests has led to an innovation in improving the dressing products on the market. The current trend of creating ideal dressing products has been to combine materials with outstanding advantages needed for the wound healing process such as intrinsic antibacterial properties (AgNPs, ZnO), high biological compatibility, being environmentally friendly, and being easy to handle, particularly natural polymer materials. Novel materials have



Table 3 Summary advantages, disadvantages, and application of modern wound dressing

Dressing type	Advantages	Disadvantages	Application
Hydrocolloids	Self-adhesive, no need for extra tape	The gel formed can be thick, yellow, smelly, and easy to mistake for an infection	Low to moderate exudate wounds. Scratch, post-surgery wounds, pressure ulcers, shallow leg ulcers
	Creating a light layer of padding Moisturizing Painless during removing the dressing Easy to use	Not suitable for exudative wounds	
Alginates	High absorbency Hemostasis	Not suitable for dry wounds Sticking to granulation tissue easily if not changed frequently	Pressure ulcers, fluid lower-extremity ulcers, infected wounds
Hydrogels	Highly biocompatible Cools, soothe wounds, and relieve pain Changing the physicochemical properties, forming reactive materials, responding to changes in temperature, pH, and drug release	Not suitable for fluid oozing wounds Requiring tape for fixation	Burns, especially partial burns, foot ulcers
Foams	Absorbing and transmitting moisture Wound cushion Not necessary to change as often, depending on the amount of discharge	Effecting on new tissue growth ability and causing injury when removing the dressing if kept for too long	Burns, chronic wounds, deep ulcers, wounds in exudate cavities
Films	Thin and elastic, easy to pull, shape to the wound	Easy to fold and stick	Burns, wounds of joints, skin grafts with small thickness, superficial lacerations
	Transparent, easy to monitor wound condition Impervious to microorganisms	Poor absorbing, only suitable for wounds with little secretion No antibacterial properties	

been developed to respond to the different conditions and stages of wound healing. Moreover, wound dressings are loaded with bioactive components including antibiotics, growth factors, herbal extracts, essential oil, antioxidants, anti-inflammatory agents, and vitamins, to improve therapeutic outcomes or to overcome the limitation of the dressing. Simultaneously, wound dressings components play an essential role as drug delivery systems.²⁰⁷ So far, incorporating bioactive agents into the structure of the dressing may considerably enhance its biocompatibility and qualities, resulting in a significant healing process. However, this modification might have a detrimental impact on the dressing's ability to absorb exudate and its mechanical characteristics.²⁰⁸ Thus, there is still a necessity for research to minimize negative effects that may result from modifying materials with bioactive components, for instance loss of mechanical properties or decreased cell proliferation.

Nevertheless, a variety of new materials for modern wound dressings not only creates synergistic effects from the strengths of each material but also causes difficulties in complex synthesis processes, requiring many advanced techniques, limiting the popular, cost-effective, if scaled-up production. Therefore, these new materials platforms have been not yet widely applied in clinical practice. Thus, the future tasks are to focus on developing new materials and to find solutions that simplify the material synthesis process, aiming to develop potential materials into commercial products serving social

needs. In addition, future research on wound dressing production should not only stop enhancing healing treatment but also ensure aesthetic requirements after treatment and limit scar tissue formation.

6. Conclusions

Modern dressing has become a priority selection for wound care and treatment due to its advancements in promoting healing wound. With the rapid development of modern technology, more and more new potential biomaterials have been created and have been intensely exploited for wound healing applications. In this context, recent advances in the development of natural-based materials as well as the applications of modern wound dressings are presented. However, the limitations of new modern dressings are the complicated production process, the lack of quality assurance for biological materials, as well as the effectiveness of the component materials for widespread use. Thus, material scientists should have more trials and experiment to determine the actual effectiveness of new modern dressings in wound healing.

Conflicts of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.



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Notes and references

- 1 K. Jung, S. Covington, C. K. Sen, M. Januszzyk, R. S. Kirsner, G. C. Gurtner and N. H. Shah, *Wound Repair Regen.*, 2016, **24**, 181–188.
- 2 E. M. Tottoli, R. Dorati, I. Genta, E. Chiesa, S. Pisani and B. Conti, *Pharmaceutics*, 2020, **12**, 735.
- 3 J. F. Guest, G. W. Fuller and P. Vowden, *BMJ Open*, 2020, **10**, e045253.
- 4 Y. Hu, H. Li, X. Lv, Y. Xu, Y. Xie, L. Yuwen, Y. Song, S. Li, J. Shao and D. Yang, *Nanoscale*, 2022, **14**, 12967–12983.
- 5 C. E. Fife and M. J. Carter, *Wounds*, 2012, **24**, 10–17.
- 6 S. R. Nussbaum, M. J. Carter, C. E. Fife, J. DaVanzo, R. Haught, M. Nusgart and D. Cartwright, *Value Health*, 2018, **21**, 27–32.
- 7 L. Gould, P. Abadir, H. Brem, M. Carter, T. Conner-Kerr, J. Davidson, L. DiPietro, V. Falanga, C. Fife, S. Gardner, E. Grice, J. Harmon, W. R. Hazzard, K. P. High, P. Houghton, N. Jacobson, R. S. Kirsner, E. J. Kovacs, D. Margolis, F. McFarland Horne, M. J. Reed, D. H. Sullivan, S. Thom, M. Tomic-Canic, J. Walston, J. Whitney, J. Williams, S. Ziemann and K. Schmader, *J. Am. Geriatr. Soc.*, 2015, **23**, 1–13.
- 8 GlobeNewswire, *Advanced Wound Care Market worth over USD 13 billion by 2024*, Global Market Insights, Inc., <https://www.globenewswire.com/fr/news-release/2018/09/17/1571505/0/en/Advanced-Wound-Care-Market-worth-over-USD-13-billion-by-2024-Global-Market-Insights-Inc.html>, accessed 23 Nov 2022.
- 9 K. Heyer, M. Augustin, K. Protz, K. Herberger, C. Spehr and S. J. Rustenbach, *Dermatology*, 2013, **226**, 172–184.
- 10 S. H. Aswathy, U. Narendrakumar and I. Manjubala, *Heliyon*, 2020, **6**, e03719.
- 11 L. Sadati, R. Froozesh, A. Beyrami, Z. N. Khaneghah, S. A. Elahi, M. F. Asl and T. Salehi, *Adv. Skin Wound Care*, 2019, **32**, 1–5.
- 12 H.-R. Metelmann, T. v. Woedtke and K.-D. Weltmann, *Comprehensive Clinical Plasma Medicine: Cold Physical Plasma for Medical Application*, Springer Cham, 1st edn, 2018.
- 13 T. Velnar, T. Bailey and V. Smrkolj, *J. Int. Med. Res.*, 2009, **37**, 1528–1542.
- 14 M. Phillipson and P. Kubes, *Trends Immunol.*, 2019, **40**, 635–647.
- 15 M. Rodrigues, N. Kosaric, C. A. Bonham and G. C. Gurtner, *Physiol. Rev.*, 2018, **99**, 665–706.
- 16 G. Younan, F. Suber, W. Xing, T. Shi, Y. Kunori, M. Åbrink, G. Pejler, S. M. Schlenner, H.-R. Rodewald, F. D. Moore Jr, R. L. Stevens, R. Adachi, K. F. Austen and M. F. Gurish, *J. Immunol.*, 2010, **185**, 7681–7690.
- 17 S. Werner, T. Krieg and H. Smola, *J. Invest. Dermatol.*, 2007, **127**, 998–1008.
- 18 J. J. Tomasek, G. Gabbiani, B. Hinz, C. Chaponnier and R. A. Brown, *Nat. Rev. Mol. Cell Biol.*, 2002, **3**, 349–363.
- 19 J. M. Reinke and H. Sorg, *Eur. Surg. Res.*, 2012, **49**, 35–43.
- 20 K. R. Jones, K. Fennie and A. Lenihan, *Wounds*, 2007, **19**, 51–63.
- 21 D. T. Robles and D. Berg, *Clin. Dermatol.*, 2007, **25**, 26–32.
- 22 R. G. Frykberg and J. Banks, *Adv. Wound Care*, 2015, **4**, 560–582.
- 23 S. Guo and L. A. DiPietro, *J. Dent. Res.*, 2010, **89**, 219–229.
- 24 W. Y. J. Chen and A. A. Rogers, *Wound Repair Regen.*, 2007, **15**, 434–449.
- 25 S. Khanna, S. Biswas, Y. Shang, E. Collard, A. Azad, C. Kauh, V. Bhasker, G. M. Gordillo, C. K. Sen and S. Roy, *Plos One*, 2010, **5**, e9539.
- 26 C. Wetzler, H. Kämpfer, B. Stallmeyer, J. Pfeilschifter and S. Frank, *J. Invest. Dermatol.*, 2000, **115**, 245–253.
- 27 G. D. Winter, *Nature*, 1962, **193**, 293–294.
- 28 C. D. Hinman and H. Maibach, *Nature*, 1963, **200**, 377–378.
- 29 A. Flett, F. Russell, S. Stringfellow, P. J. Cooper, D. G. Gray and S. Lawton, *Nurs. Resid. Care*, 2002, **4**, 328–344.
- 30 L. Demarre, S. Verhaeghe, A. Van Hecke, E. Clays, M. Grypdonck and D. Beeckman, *J. Adv. Nurs.*, 2015, **71**, 391–403.
- 31 J. S. Boateng, K. H. Matthews, H. N. E. Stevens and G. M. Eccleston, *J. Pharm. Sci.*, 2008, **97**, 2892–2923.
- 32 L. J. Borda, F. E. Macquhae and R. S. Kirsner, *Curr. Dermatol. Rep.*, 2016, **5**, 287–297.
- 33 G. D. Mogoşanu and A. M. Grumezescu, *Int. J. Pharm.*, 2014, **463**, 127–136.
- 34 S. Dhivya, V. V. Padma and E. Santhini, *BioMedicine*, 2015, **5**, 22.
- 35 K. L. Andrews, K. M. Derby, T. M. Jacobson, B. A. Sievers and L. J. Kiemele, in *Braddom's Physical Medicine and Rehabilitation*, ed. D. X. Cifu, Elsevier, Philadelphia, 6th edn, 2021, ch. 24, pp. 469–484.
- 36 S. Rahmani and D. J. Mooney, in *The Diabetic Foot: Medical and Surgical Management*, ed. A. Veves, J. M. Giurini and R. J. Guzman, Humana Press, Totowa, New Jersey, 4th edn, 2018, ch. 15, pp. 247–256.
- 37 C. M. Wietlisbach, in *Cooper's Fundamentals of Hand Therapy*, ed. C. M. Wietlisbach, Mosby, St. Louis (MO), 3rd edn, 2020, ch. 17, pp. 154–166.
- 38 A. Agarwal, J. F. McAnulty, M. J. Schurr, C. J. Murphy and N. L. Abbott, in *Advanced Wound Repair Therapies*, ed. D. Farrar, Woodhead Publishing, 2011, pp. 186–208.
- 39 F. L. Bowling, S. T. Rashid and A. J. M. Boulton, *Nat. Rev. Endocrinol.*, 2015, **11**, 606–616.
- 40 N. M. Aruan, I. Sriyanti, D. Edikresnha, T. Suciati, M. M. Munir and K. Khairurrijal, *Procedia Eng.*, 2017, **170**, 31–35.
- 41 A. Janowska, M. Macchia and B. Paggi, in *Science and Practice of Pressure Ulcer Management*, ed. M. Romanelli, M. Clark, A. Gefen and G. Ciprandi, Spinger London, London, 2nd edn, 2018, ch. 12, pp. 159–173.
- 42 S. Thomas, *Int. Wound J.*, 2008, **5**, 602–613.
- 43 J. Z. M. Lim, N. S. L. Ng and C. Thomas, *J. R. Soc. Med.*, 2017, **110**, 104–109.



- 44 S. G. Jin, A. M. Yousaf, S. W. Jang, M.-W. Son, K. S. Kim, D.-W. Kim, D. X. Li, J. O. Kim, C. S. Yong and H.-G. Choi, *Drug Dev. Res.*, 2015, **76**, 157–165.
- 45 T. Takeuchi, M. Ito, S. Yamaguchi, S. Watanabe, M. Honda, T. Imahashi, T. Yamada and T. Kokubo, *Nagoya J. Med. Sci.*, 2020, **82**, 487–498.
- 46 K.-Y. Sung and S.-Y. Lee, *Wounds*, 2016, **28**, 145–151.
- 47 J. Chen, J. Chen, J. Yang, Y. Chen, Y. Liang and Y. Lin, *Pediatr. Crit. Care Med.*, 2020, **21**, e752–e758.
- 48 N. Shinohara, N. Nonomura, M. Eto, G. Kimura, H. Minami, S. Tokunaga and S. Naito, *Ann. Oncol.*, 2014, **25**, 472–476.
- 49 C. Sabando, W. Ide, M. Rodríguez-Díaz, G. Cabrera-Barjas, J. Castaño, R. Bouza, N. Müller, C. Gutiérrez, L. Barral, J. Rojas, F. Martínez and S. Rodríguez-Llamazares, *Curr. Top. Med. Chem.*, 2020, **20**, 280–292.
- 50 Convatec, *DuoDERM® Dressings*, <https://www.convatec.com/advanced-wound-care/duoderm-dressings/>, accessed Nov 23, 2022.
- 51 Coloplast, *Comfeel®Plus*, <https://www.coloplast.com/products/wound/comfeel-plus/>, accessed Nov 23, 2022.
- 52 Smith-Nephew, *Cutinova®Hydro*, <https://www.smith-nephew.com/professional/products/advanced-wound-management/other-wound-care-products/cutinova-hydro/>, accessed Nov 23, 2022.
- 53 Smith-Nephew, *Replicare®*, <https://www.smith-nephew.com/professional/products/advanced-wound-management/repicare/repicare/>, accessed Nov 23, 2022.
- 54 Convatec, *Kaltostat®*, <https://www.convatec.com/en-au/products/pc-wound-diabetic-foot-ulcers/kaltostat-alginate-calcium-sodium-dressing>, accessed Nov 23, 2022.
- 55 DermaSciences, *Algicell®Ag*, <https://www.integralife.com/algicell-ag-calcium-alginate-dressing-with-antimicrobial-silver/product/wound-reconstruction-care-outpatient-clinic-private-office-supportive-therapies-algicell-ag-calcium-alginate-dressing-with-antimicrobial-silver>, accessed Nov 23, 2022.
- 56 S. G. Kim, K. Y. Song, H. H. Lee, E. Y. Kim, J. H. Lee, H. M. Jeon, K. H. Jeon, H. M. Jin, D. J. Kim, W. Kim, H. M. Yoo, J. G. Kim and C. H. Park, *Medicine*, 2019, **98**, e15141.
- 57 A. Medical, *Activon® – Manuka Honey*, <https://uk.advancismedical.com/products/activon-manuka-honey/algivon>, accessed Nov 23, 2022.
- 58 3M™, *Fibracol™ Plus Collagen Wound Dressing with Alginate*, https://www.3m.com/3M/en_US/p/d/b5005265077/, accessed Nov 23, 2022.
- 59 M. K. Kucharska, M. H. Struszczyk, A. Niekraszewicz, D. Ciechańska, E. Witzczak, S. Tarkowska, K. Fortuniak, A. Gulbas-Diaz, A. Rogaczewska, I. Płoszaj, A. Pluta and T. Gąsiorowski, *Prog. Chem. Appl. Chitin Deriv.*, 2011, **16**, 121–130.
- 60 Tricomed, *Tromboguard®*, <https://tricomed.com/products/tromboguard/>, accessed Jun 26, 2022.
- 61 Activheal, *Activheal® hydrogel is an effective method for hydrating dry necrotic and sloughy wounds*, <https://activheal.com/wound-care-dressing-range/hydrogel-dressing/>, accessed Nov 23, 2022.
- 62 H. Woundcare, *Restore Hydrogel Dressings*, <https://www.hollister.com/-/media/files/pdfs-for-download/wound-care/restore-hydrogel-techsheet-911140-1110.ashx>, accessed Nov 23, 2022.
- 63 L. R. Globa, *Suprasorb® G Gel Wound Dressing*, <https://www.lohmann-rauscher.com/au-en/produkte-alt/wound-care/modern-wound-care/suprasorb-g-gel-dressing/>, accessed Nov 23, 2022.
- 64 DermaRite, *AquaDerm™ Hydrogel Sheet Dressing*, <https://dermarite.com/product/aquaderm/>, accessed Nov 23, 2022.
- 65 Kikgel, *Neoheal®Hydrogel dressing for wound management*, <https://kikgel.com.pl/en/products/neoheal/>, accessed Nov 23, 2022.
- 66 Smith-Nephew, *Allevyn Life*, <https://www.smith-nephew.com/professional/products/advanced-wound-management/allevyn/allevyn-life1/>, accessed Nov 23, 2022.
- 67 Hartmann, *PermaFoam® Classic*, <https://www.hartmann.info/en-dx/products/wound-management/hydroactive-wound-dressings/foam-wound-dressings/permafoam%C2%AE-classic>, accessed Nov 23, 2022.
- 68 Hartmann, *HydroTac®*, <https://www.hartmann.info/en-dx/products/wound-management/hydroactive-wound-dressings/foam-wound-dressings/hydrotac%C2%AE#products>, accessed Nov 23, 2022.
- 69 Mölnlycke, *Mepilex Ag*, <https://www.molnlycke.se/products-solutions/mepilex-ag/>, accessed Feb 2, 2023.
- 70 3M™, *Tegaderm™ Silicone Foam Dressings*, https://www.3m.com/3M/en_US/medical-us/tegaderm-silicone-foam-dressings/, accessed Nov 23, 2022.
- 71 3M™, *Bioclusive™ Plus Transparent Film Dressing*, <https://www.acerity.com/healthcare-professionals/global-product-catalog/catalog/bioclusive-plus-dressing>, accessed Nov 23, 2022.
- 72 Mölnlycke, *Mepitel*, <https://www.molnlycke.com/products-solutions/mepitel/>, accessed Nov 23, 2022.
- 73 Mölnlycke, *Mepore® Film*, <https://www.molnlycke.com/products-solutions/mepore-film/>, accessed Nov 23, 2022.
- 74 DeRoyal, *Transeal®*, <https://www.deroyal.com/products/search-catalog-item/catalog-item-preview/wc-burn-transeal>, accessed Nov 23, 2022.
- 75 E. A. Kamoun, E.-R. S. Kenawy and X. Chen, *J. Adv. Res.*, 2017, **8**, 217–233.
- 76 M. A. Taemeh, A. Shiravandi, M. A. Korayem and H. Daemi, *Carbohydr. Polym.*, 2020, **228**, 115419.
- 77 P. N. Sudha, S. Aisverya, R. Nithya and K. Vijayalakshmi, in *Advances in Food and Nutrition Research*, ed. S.-K. Kim, Academic Press, 2014, ch. 8, vol. 73, pp. 145–181.
- 78 R. Mahsood and M. MirafTAB, *J. Wound Care*, 2014, **23**, 153–159.
- 79 J. C. Dumville, S. J. Keogh, Z. Liu, N. Stubbs, R. M. Walker and M. Fortnam, *Cochrane Database Syst. Rev.*, 2015, CD011277.
- 80 B. A. Aderibigbe and B. Buyana, *Pharmaceutics*, 2018, **10**, 42.
- 81 Helmiyati and M. Aprilliza, *Presented in part at the IOP Conference Series: Materials Science and Engineering*, Bali, Indonesia, Jul 26–27, 2016.



- 82 M. Matyash, F. Despong, C. Ikonomidou and M. Gelinsky, *Tissue Eng., Part C*, 2013, **20**, 401–411.
- 83 S. K. Bajpai and N. Kirar, *Des. Monomers Polym.*, 2016, **19**, 89–98.
- 84 I. R. Sweeney, M. MirafTAB and G. Collyer, *Int. Wound J.*, 2012, **9**, 601–612.
- 85 A. Sood, M. S. Granick and N. L. Tomaselli, *Adv. Wound Care*, 2013, **3**, 511–529.
- 86 C. Weller, C. Weller and V. Team, in *Advanced Textiles for Wound Care* ed. S. Rajendran, Woodhead Publishing, 2nd edn, 2019, ch. 4, pp. 105–134.
- 87 S. McCarthy, V. Dvorakova, P. O'Sullivan and J. F. Bourke, *Contact Dermatitis*, 2018, **79**, 396–397.
- 88 M. Fischer, F. Gebhard, T. Hammer, C. Zurek, G. Meurer, C. Marquardt and D. Hofer, *J. Biomater. Appl.*, 2017, **31**, 1267–1276.
- 89 A. Doderò, S. Scarfi, M. Pozzolini, S. Vicini, M. Alloisio and M. Castellano, *ACS Appl. Mater. Interfaces*, 2020, **12**, 3371–3381.
- 90 Y. Fan, W. Wu, Y. Lei, C. Gaucher, S. Pei, J. Zhang and X. Xia, *Mar. Drugs*, 2019, **17**, 285.
- 91 S. Shafei, M. Khanmohammadi, R. Heidari, H. Ghanbari, V. Taghdiri Nooshabadi, S. Farzamfar, M. Akbariqomi, N. S. Sanikhani, M. Absalan and G. Tavoosidana, *J. Biomed. Mater. Res. A*, 2020, **108**, 545–556.
- 92 M. Contardi, A. M. d. M. d. Ayyoub, M. Summa, D. Kossyvak, M. Fadda, N. Liessi, A. Armirotti, D. Fragouli, R. Bertorelli and A. Athanassiou, *ACS Appl. Bio Mater.*, 2022, **5**, 2880–2893.
- 93 H. Lu, J. A. Butler, N. S. Britten, P. D. Venkatraman and S. S. Rahatekar, *Nanomaterials*, 2021, **11**, 2062.
- 94 S. O'Meara, M. Martyn-St James and U. J. Adderley, *Cochrane Database Syst. Rev.*, 2015, CD010182.
- 95 G. Huang, T. Sun, L. Zhang, Q. Wu, K. Zhang, Q. Tian and R. Huo, *Exp. Ther. Med.*, 2014, **7**, 1772–1776.
- 96 L. Devi and P. Gaba, *J. Crit. Rev.*, 2019, **6**, 1–10.
- 97 D. Zeng, S. Shen and D. Fan, *Chin. J. Chem. Eng.*, 2021, **30**, 308–320.
- 98 Y. Liang, J. He and B. Guo, *ACS Nano*, 2021, **15**, 12687–12722.
- 99 I. Firlar, M. Altunbek, C. McCarthy, M. Ramalingam and G. Camci-Unal, *Gels*, 2022, **8**, 127.
- 100 H. Chen, R. Cheng, X. Zhao, Y. Zhang, A. Tam, Y. Yan, H. Shen, Y. S. Zhang, J. Qi, Y. Feng, L. Liu, G. Pan, W. Cui and L. Deng, *NPG Asia Mater.*, 2019, **11**, 3.
- 101 L. L. M. Jose, R. M. Leocadio, L. Olga, B. P. Miriam, C. Dios-Guerra and L. M. Silvia, *J. Wound Care*, 2020, **29**, 202.
- 102 M. D. Holbert, B. R. Griffin, S. M. McPhail, R. S. Ware, K. Foster, D. C. Bertoni and R. M. Kimble, *Trials*, 2019, **20**, 13.
- 103 S. Niansheng, C. Binghuan, W. Cheng, G. Hua, B. Xu, Z. Ran and H. Ran, *Zhonghua Shao Shang Za Zhi*, 2021, **37**, 1085–1089.
- 104 L. Zhang, H. Yin, X. Lei, J. N. Y. Lau, M. Yuan, X. Wang, F. Zhang, F. Zhou, S. Qi, B. Shu and J. Wu, *Front. Bioeng. Biotechnol.*, 2019, **7**, 342.
- 105 K. A. Deo, G. Lokhande and A. K. Gaharwar, in *Encyclopedia of Tissue Engineering and Regenerative Medicine*, ed. R. L. Reis, Academic Press, Oxford, 2019, pp. 21–32.
- 106 E. Yahia, A. El-Sharkawey and M. Bayoumi, *Pakistan J. Medical Health Sci.*, 2021, **15**, 1571–1574.
- 107 L. Han, X. Lu, K. Liu, K. Wang, L. Fang, L.-T. Weng, H. Zhang, Y. Tang, F. Ren, C. Zhao, G. Sun, R. Liang and Z. Li, *ACS Nano*, 2017, **11**, 2561–2574.
- 108 A. Tamayol, M. Akbari, Y. Zilberman, M. Comotto, E. Lesha, L. Serex, S. Bagherifard, Y. Chen, G. Fu, S. K. Ameri, W. Ruan, E. L. Miller, M. R. Dokmeci, S. Sonkusale and A. Khademhosseini, *Adv. Healthcare Mater.*, 2016, **5**, 711–719.
- 109 C. Occhiuzzi, A. Ajovalasit, M. A. Sabatino, C. Dispenza and G. Marrocco, 2015.
- 110 J. Nielsen and K. Fogh, *Chronic Wound Care Manag. Res.*, 2015, **2**, 31–38.
- 111 A.-G. Niculescu and A. M. Grumezescu, *Polymers*, 2022, **14**, 421.
- 112 C. D. Weller, V. Team and G. Sussman, *Front. Pharmacol.*, 2020, **11**, 155.
- 113 S. M. Lee, I. K. Park, Y. S. Kim, H. J. Kim, H. Moon, S. Mueller and Y.-I. L. Jeong, *Biomater. Res.*, 2016, **20**, 15.
- 114 J.-A. Jung, K.-H. Yoo, S.-K. Han, E.-S. Dhong and W.-K. Kim, *Adv. Skin Wound Care*, 2016, **29**, 546–555.
- 115 O. M. Alvarez, M. S. Granick, A. Reyzelman and T. Serena, *J. Comp. Eff. Res.*, 2021, **10**, 481–493.
- 116 P. Davies, S. McCarty and K. Hamberg, *J. Wound Care*, 2017, **26**, S1–S32.
- 117 N. Namviriyachote, V. Lipipun, Y. Akkhawattanangkul, P. Charoonrut and G. C. Ritthidej, *Asian J. Pharm. Sci.*, 2019, **14**, 63–77.
- 118 H. C. Gwak, S. H. Han, J. Lee, S. Park, K.-S. Sung, H.-J. Kim, D. Chun, K. Lee, J.-H. Ahn, K. Kwak and H.-J. Chung, *Int. Wound J.*, 2020, **17**, 91–99.
- 119 K. Vowden and P. Vowden, *Surgery*, 2017, **35**, 489–494.
- 120 R. A. A. Dutra, G. M. Salomé, J. R. Alves, V. O. S. Pereira, F. D. Miranda, V. B. Vallim, M. J. A. de Brito and L. M. Ferreira, *J. Wound Care*, 2015, **24**, 268–275.
- 121 A. Jafari, S. Hassanajili, M. B. Karimi, A. Emami, F. Ghaffari and N. Azarpira, *J. Mech. Behav. Biomed. Mater.*, 2018, **88**, 395–405.
- 122 M. Li, J. Chen, M. Shi, H. Zhang, P. X. Ma and B. Guo, *Chem. Eng. J.*, 2019, **375**, 121999.
- 123 F. Gulmez, A. Yercan, B. Kocaaga and F. S. Guner, *J. Drug Deliv. Sci. Technol.*, 2021, **61**, 102160.
- 124 N. Tang, R. Zhang, Y. Zheng, J. Wang, M. Khatib, X. Jiang, C. Zhou, R. Omar, W. Saliba, W. Wu, M. Yuan, D. Cui and H. Haick, *Adv. Mater.*, 2022, **34**, 2106842.
- 125 M. Kazanavičius, A. Cepas, V. Kolaityte, R. Simoliuniene and R. Rimdeika, *J. Wound Care*, 2017, **26**, 281–291.
- 126 L. C. Schmeel, D. Koch, F. C. Schmeel, B. Bücheler, C. Leitzen, B. Mahlmann, D. Kunze, M. Heimann, D. Brüser, A.-V. Abramian, F. Schoroth, T. Müdder, F. Röhrner, S. Garbe, B. G. Baumert, H. H. Schild and T. M. Wilhelm-Buchstab, *Polymers*, 2019, **11**, 2112.



- 127 S. Hajebi, S. Mohammadi Nasr, N. Rabiee, M. Bagherzadeh, S. Ahmadi, M. Rabiee, M. Tahri, L. Tayebi and M. R. Hamblin, *Int. J. Polym. Mater. Polym. Biomater.*, 2021, **70**, 926–940.
- 128 M. Filippi, G. Born, M. Chaaban and A. Scherberich, *Front. Bioeng. Biotechnol.*, 2020, **8**, 474.
- 129 S. Huang and X. Fu, *J. Controlled Release*, 2010, **142**, 149–159.
- 130 G. Praveen Kumar, R. Shreeya Sai, P. Deepali Venkatesh, V. Priyadharsini, S. Vidhya, C. Chandrananthi, C. Shreya, S. Krithika and G. Keerthana, in *Cellulose*, ed. P. Alejandro Rodriguez and E. E. M. María, IntechOpen, Rijeka, 2019, ch. 4.
- 131 R. Naomi, R. Bt Hj Idrus and M. B. Fauzi, *Int. J. Environ. Res. Public Health*, 2020, **17**, 6803.
- 132 N. Mohd, S. F. S. Draman, M. S. N. Salleh and N. B. Yusof, *AIP Conf. Proc.*, 2017, **1809**, 020035.
- 133 M. Oprea and S. I. Voicu, *Carbohydr. Polym.*, 2020, **247**, 116683.
- 134 S. M. Choi, K. M. Rao, S. M. Zo, E. J. Shin and S. S. Han, *Polymers*, 2022, **14**, 1080.
- 135 J. D. de Amorim, C. J. da Silva Junior, A. D. de Medeiros, H. A. do Nascimento, M. Sarubbo, T. P. de Medeiros, A. F. Costa and L. A. Sarubbo, *Molecules*, 2022, **27**, 5580.
- 136 E. Tsouko, C. Kourmentza, D. Ladakis, N. Kopsahelis, I. Mandala, S. Papanikolaou, F. Paloukis, V. Alves and A. Koutinas, *Int. J. Mol. Sci.*, 2015, **16**, 14832–14849.
- 137 N. Shah, M. Ul-Islam, W. A. Khattak and J. K. Park, *Carbohydr. Polym.*, 2013, **98**, 1585–1598.
- 138 Z. Li, L. Wang, J. Hua, S. Jia, J. Zhang and H. Liu, *Carbohydr. Polym.*, 2015, **120**, 115–119.
- 139 C. Huang, X. Y. Yang, L. Xiong, H. J. Guo, J. Luo, B. Wang, H. R. Zhang, X. Q. Lin and X. D. Chen, *Lett. Appl. Microbiol.*, 2015, **60**, 491–496.
- 140 M. Ul-Islam, W. Alhajaim, A. Fatima, S. Yasir, T. Kamal, Y. Abbas, S. Khan, A. H. Khan, S. Manan, M. W. Ullah and G. Yang, *Int. J. Biol. Macromol.*, 2023, **231**, 123269.
- 141 F. J. O'Brien, *Mater. Today*, 2011, **14**, 88–95.
- 142 R. Parenteau-Bareil, R. Gauvin and F. Berthod, *Materials*, 2010, **3**, 1863–1887.
- 143 M. Abas, M. El Masry and H. Elgharably, in *Wound Healing, Tissue Repair, and Regeneration in Diabetes*, ed. D. Bagchi, A. Das and S. Roy, Academic Press, 2020, ch. 19, pp. 393–401.
- 144 I. N. Amirrah, M. F. Mohd Razip Wee, Y. Tabata, R. Bt Hj Idrus, A. Nordin and M. B. Fauzi, *Polymers*, 2020, **12**, 2168.
- 145 A. Gaspar-Pintilie, A.-M. Stanciuc and O. Craciunescu, *Int. J. Biol. Macromol.*, 2019, **138**, 854–865.
- 146 S. S. Mathew-Steiner, S. Roy and C. K. Sen, *Bioengineering*, 2021, **8**, 63.
- 147 C. Holmes, J. S. Wrobel, M. P. MacEachern and B. R. Boles, *Diabetes, Metab. Syndr. Obes.: Targets Ther.*, 2013, **6**, 17–29.
- 148 G. A. Rico-Llanos, S. Borrego-González, M. Moncayo-Donoso, J. Becerra and R. Visser, *Polymers*, 2021, **13**, 599.
- 149 N. Gull, S. M. Khan, M. T. Zahid Butt, S. Khalid, M. Shafiq, A. Islam, S. Asim, S. Hafeez and R. U. Khan, *RSC Adv.*, 2019, **9**, 31078–31091.
- 150 N. Gull, S. M. Khan, A. Islam and M. T. Z. Butt, in *Bio Monomers for Green Polymeric Composite Materials*, eds. P. M. Visakh, O. Bayraktar and G. Menon, Wiley, New York, United States, 2019, ch. 9, pp. 175–199.
- 151 A. M. Abdel-Mohsen, J. Frankova, R. M. Abdel-Rahman, A. A. Salem, N. M. Sahffie, I. Kubena and J. Jancar, *Int. J. Pharm.*, 2020, **582**, 119349.
- 152 R. A. Pérez, J.-E. Won, J. C. Knowles and H.-W. Kim, *Adv. Drug Delivery Rev.*, 2013, **65**, 471–496.
- 153 T. Phaechamud, K. Yodkhum, J. Charoenteeraboon and Y. Tabata, *Mater. Sci. Eng., C*, 2015, **50**, 210–225.
- 154 C. Chircov, A. M. Grumezescu and L. E. Bejenaru, *Rom. J. Morphol. Embryol.*, 2018, **59**, 71–76.
- 155 A. Fakhari and C. Berkland, *Acta Biomater.*, 2013, **9**, 7081–7092.
- 156 J. Voigt and V. R. Driver, *Wound Repair Regen.*, 2012, **20**, 317–331.
- 157 J. S. Frenkel, *Int. Wound J.*, 2014, **11**, 159–163.
- 158 M. Dovedytis, Z. J. Liu and S. Bartlett, *Eng. Regener.*, 2020, **1**, 102–113.
- 159 A. Ström, A. Larsson and O. Okay, *J. Appl. Polym. Sci.*, 2015, **132**, 42194.
- 160 G. M. Campo, A. Avenoso, A. D'Ascola, V. Prestipino, M. Scuruchi, G. Nastasi, A. Calatroni and S. Campo, *BioFactors*, 2012, **38**, 69–76.
- 161 P. A. Janmey, J. P. Winer and J. W. Weisel, *J. R. Soc., Interface*, 2008, **6**, 1–10.
- 162 P. Heher, S. Mühleder, R. Mittermayr, H. Redl and P. Slezak, *Adv. Drug Delivery Rev.*, 2018, **129**, 134–147.
- 163 N. Laurens, P. Koolwijk and M. P. M. De Maat, *J. Thromb. Haemostasis*, 2006, **4**, 932–939.
- 164 I. S. Bayer, *Molecules*, 2022, **27**, 4504.
- 165 N. Y. Becerra, L. M. Restrepo, Y. Galeano, A. C. Tobón, L. F. Turizo and M. Mesa, *Int. J. Biomater.*, 2021, **2021**, 9933331.
- 166 W. Bensaid, J. T. Triffitt, C. Blanchat, K. Oudina, L. Sedel and H. Petite, *Biomaterials*, 2003, **24**, 2497–2502.
- 167 S. A. Sell, P. S. Wolfe, K. Garg, J. M. McCool, I. A. Rodriguez and G. L. Bowlin, *Polymers*, 2010, **2**, 522–553.
- 168 C. Schneider-Barthold, S. Baganz, M. Wilhelmi, T. Scheper and I. Pepelanova, *BioNanoMaterials*, 2016, **17**, 3–12.
- 169 P. Zarrintaj, S. Ghorbani, M. Barani, N. P. Singh Chauhan, M. Khodadadi Yazdi, M. R. Saeb, J. D. Ramsey, M. R. Hamblin, M. Mozafari and E. Mostafavi, *Bioeng. Transl. Med.*, 2022, **7**, e10261.
- 170 S. C. Shukla, A. Singh, A. K. Pandey and A. Mishra, *Biochem. Eng. J.*, 2012, **65**, 70–81.
- 171 C. Shi, Y. He, X. Feng and D. Fu, *J. Biomater. Sci., Polym. Ed.*, 2015, **26**, 1343–1356.
- 172 T. Bo, P.-P. Han, Q.-Z. Su, P. Fu, F.-Z. Guo, Z.-X. Zheng, Z.-L. Tan, C. Zhong and S.-R. Jia, *Food Control*, 2016, **61**, 123–134.
- 173 N. A. Patil and B. Kandasubramanian, *Eur. Polym. J.*, 2021, **146**, 110248.
- 174 R. Singla, S. Soni, V. Patial, P. M. Kulurkar, A. Kumari, S. Mahesh, Y. S. Padwad and S. K. Yadav, *Sci. Rep.*, 2017, **7**, 10457.



- 175 G. Yang, Z. Zhang, K. Liu, X. Ji, P. Fatehi and J. Chen, *J. Nanobiotechnol.*, 2022, **20**, 312.
- 176 E.-E. Tudoroiu, C.-E. Dinu-Pîrvu, M. G. Albu Kaya, L. Popa, V. Anuta, R. M. Prisada and M. V. Ghica, *Pharmaceuticals*, 2021, **14**, 1215.
- 177 E. Masci, G. Faillace and M. Longoni, *BMC Res. Notes*, 2018, **11**, 239.
- 178 L. Deng, B. Wang, W. Li, Z. Han, S. Chen and H. Wang, *Int. J. Biol. Macromol.*, 2022, **217**, 77–87.
- 179 Y. Xie, K. Qiao, L. Yue, T. Tang, Y. Zheng, S. Zhu, H. Yang and Z. Fang, *Bioact. Mater.*, 2022, **17**, 248–260.
- 180 S. Yazgan, I. O. Tekin, N. Akpolat and O. Koc, *J. Glaucoma*, 2021, **30**, 1001–1010.
- 181 L. G. Silva, A. V. Albuquerque, F. C. M. Pinto, R. S. Ferraz-Carvalho, J. L. A. Aguiar and E. M. Lins, *J. Mater. Sci.: Mater. Med.*, 2021, **32**, 79.
- 182 X. Pan, C. Han, G. Chen and Y. Fan, *Evidence-Based Complementary Altern. Med.*, 2022, **2022**, 5217617.
- 183 A. Das, M. Abas, N. Biswas, P. Banerjee, N. Ghosh, A. Rawat, S. Khanna, S. Roy and C. K. Sen, *Sci. Rep.*, 2019, **9**, 14293.
- 184 B. Colak, S. Yormaz, I. Ece, A. Çalişir, K. Körez, M. Çınar and M. Sahin, *Int. J. Lower Extremity Wounds*, 2020, **21**, 279–289.
- 185 H.-C. Tsai, H.-C. Shu, L.-C. Huang and C.-M. Chen, *Formos. J. Surg.*, 2019, **52**, 52–56.
- 186 A.-H. Stricker-Krongrad, Z. Alikhassy, N. Matsangos, R. Sebastian, G. Marti, F. Lay and J. W. Harmon, *ePlasty*, 2018, **18**, e14.
- 187 R. Pippi, M. Santoro and A. Cafolla, *J. Oral Maxillofac. Surg.*, 2017, **75**, 1118–1123.
- 188 X. Mo, J. Cen, E. Gibson, R. Wang and S. L. Percival, *Wound Repair Regen.*, 2015, **23**, 518–524.
- 189 Y. Hussein, E. M. El-Fakharany, E. A. Kamoun, S. A. Loutfy, R. Amin, T. H. Taha, S. A. Salim and M. Amer, *Int. J. Biol. Macromol.*, 2020, **164**, 667–676.
- 190 M. N. Leite and M. A. C. Frade, *Heliyon*, 2021, **7**, e07572.
- 191 X. Ding, S. Li, M. Tian, P. Yang, Y. Ding, Y. Wang, G. Duan, D. Zhang, B. Chen and Q. Tan, *Int. J. Biol. Macromol.*, 2023, **226**, 1490–1499.
- 192 S. Yildirim, H. Ö. Özener, B. Doğan and B. Kuru, *J. Periodontol.*, 2018, **89**, 36–45.
- 193 M. Lee, S. H. Han, W. J. Choi, K. H. Chung and J. W. Lee, *Wound Repair Regen.*, 2016, **24**, 581–588.
- 194 M. Bacakova, J. Pajorova, T. Sopuch and L. Bacakova, *Materials*, 2018, **11**, 2314.
- 195 X. Lv, Y. Xu, X. Ruan, D. Yang, J. Shao, Y. Hu, W. Wang, Y. Cai, Y. Tu and X. Dong, *Acta Biomater.*, 2022, **146**, 107–118.
- 196 I. Laidmäe, K. Ērglis, A. Cēbers, P. A. Janmey and R. Uiibo, *J. Mater. Sci.: Mater. Med.*, 2018, **29**, 182.
- 197 M. Soleimanpour, S. S. Mirhaji, S. Jafari, H. Derakhshankhah, F. Mamashli, H. Nedaei, M. R. Karimi, H. Motasadizadeh, Y. Fatahi, A. Ghasemi, M. S. Nezamtaheri, M. Khajezade, M. Teimouri, B. Goliaei, C. Delattre and A. A. Saboury, *Sci. Rep.*, 2022, **12**, 7213.
- 198 L. P. F. Abbade, S. R. C. S. Barraviera, M. R. C. Silveiras, A. B. B. d. C. O. Lima, G. R. Haddad, M. A. N. Gatti, N. B. Medolago, M. T. Rigotto Carneiro, L. D. dos Santos, R. S. Ferreira and B. Barraviera, *Front. Immunol.*, 2021, **12**, 627541.
- 199 V. Mayandi, A. C. Wen Choong, C. Dhand, F. P. Lim, T. T. Aung, H. Sriram, N. Dwivedi, M. H. Periyah, S. Sridhar, M. Fazil, E. T. L. Goh, G. Orive, R. W. Beuerman, T. M. S. Barkham, X. J. Loh, Z. X. Liang, V. A. Barathi, S. Ramakrishna, S. J. Chong, N. K. Verma and R. Lakshminarayanan, *ACS Appl. Mater. Interfaces*, 2020, **12**, 15989–16005.
- 200 C. Mou, X. Wang, J. Teng, Z. Xie and M. Zheng, *J. Nanobiotechnol.*, 2022, **20**, 368.
- 201 S. Liu, X. Liu, Y. Ren, P. Wang, Y. Pu, R. Yang, X. Wang, X. Tan, Z. Ye, V. Maurizot and B. Chi, *ACS Appl. Mater. Interfaces*, 2020, **12**, 39936.
- 202 C. Li, Q. Zhang, D. Lan, M. Cai, Z. Liu, F. Dai and L. Cheng, *Int. J. Biol. Macromol.*, 2022, **220**, 1049–1059.
- 203 C. Shi, C. Wang, H. Liu, Q. Li, R. Li, Y. Zhang, Y. Liu, Y. Shao and J. Wang, *Front. Bioeng. Biotechnol.*, 2020, **8**, 182.
- 204 K. Harding, P. Chadwick, S. L. A. Jeffery, D. Gray, E. Lindsay, I. Younis, A. Sharpe, K. Cutting and M. Butcher, *J. Wound Care*, 2019, **28**, 497.
- 205 K. C. Broussard and J. G. Powers, *Am. J. Clin. Dermatol.*, 2013, **14**, 449–459.
- 206 M. Saco, N. Howe, R. Nathoo and B. Cherpelis, *J. Am. Acad. Dermatol.*, 2016, **74**, AB293.
- 207 S. Alven, S. Peter, Z. Mbese and B. A. Aderibigbe, *Polymers*, 2022, **14**, 724.
- 208 Y. P. Afsharian and M. Rahimnejad, *Polym. Test.*, 2021, **93**, 106952.

