Biomimetic Layer-by-Layer Assembly of Artificial Nacre

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Abstract

Nacre is a technologically remarkable organic-inorganic composite biomaterial. It consists of an ordered multilayer structure of crystalline calcium carbonate platelets separated by porous organic layers. This microstructure exhibits both optical iridescence and mechanical toughness which transcend those of its constituent components. Replication of nacre is essential to understanding this complex biomineral, and paves the way for tough coatings fabricated from cheap abundant materials. Fabricating a calcitic nacre imitation with biologically similar optical and mechanical properties will likely require following all steps taken in biogenic nacre synthesis. Here we present a route to artificial nacre which mimics the natural layer-by-layer approach to fabricate a hierarchical crystalline multilayer material. Its structure-function relationship was confirmed by nacrelike mechanical properties and striking optical iridescence. Our biomimetic route uses the interplay of polymer-mediated mineral growth, combined with layerby-layer deposition of porous organic films. This is the first successful attempt to replicate nacre, using CaCO₃.

Introduction

Nacre's biogenic synthesis process entails a symphony of mineral and organic species which finely control each others' deposition^{?, ?}. Nacre forms extracellularly in the isolated space between the shell and the epithelium mantle cells. These cells secrete porous hydrophobic chitin sheets that are covered by acidic silk proteins which form a boundary between mineral layers. Mineralisation then proceeds by the deposition of a chemically stabilised CaCO₃ amorphous precursor phase (ACC)^{?, ?} onto the chitin sheets, where it coalesces to form an ACC film. The repeated deposition of chitin and ACC followed by CaCO₃ crystallisation leads to a lamellar stacks of CaCO₃ tablets separated by organic layers. The porosity of the chitin layers is an important structural feature, allowing mineral

interconnects, which are responsible for crystal continuity between the platelets, improving the mechanical properties of the stack[?].

The characteristic nacre structure in Fig. ??a-c consists of 250-500 nm thick aragonite tablets, separated by 30-90 nm thick organic layers?, seen in Fig. ??b. Each tablet is part of a single crystal, and the crystallinity extends though tablet stacks, mediated by holes? (mineral interconnects) in the organic layers in Fig. ??c. The stack periodicity gives rise to the hallmark iridescence of nacre shown for a shell of *Haliotis tuberculata* in Fig. ??a.

While organic/inorganic composites are common in nature and technology, nacre, as well as sea-urchin spines and skeletons, has intrigued scientists because these organisms are able to mould large single-crystal shapes and create materials that fulfil specific functions for the organisms^{?,?,?}. Replicating nacre is interesting as it sheds light on the biological processes at work in growing this biomineral, and it equips us to fabricate novel tough coatings using low cost materials in ambient conditions.

While other studies have identified many complex biochemical pathways which lead to nacre's biomineral growth^{?,?,?}, here we propose a minimum of five essential stages which are necessary to successfully mimic nacre: (1) Stabilisation of ACC in solution, (2) Specific aggregation and continuous film formation on organic surfaces, (3) Deposition of a porous, suitably functionalised thin organic film on a previously formed mineral layer, (4) Crystallisation of the formed ACC layers to aragonite or calcite, and (5) Cyclical iteration of steps 1-4.

Previous attempts at mimicking nacre only aimed at some of these 5 steps. In particular, they lacked mineral continuity across the porous organics, and thus failed to achieve nacre's characteristic microstructure^{?,?,?}. Nacre has previously been retro-synthesised by recrystallising the decalcified insoluble organic matrix of an abalone shell[?]. The organic-inorganic hybrid principle of nacre has been followed in the creation of several non-calcareous systems^{?,?,?,?,?,?}.

Here, we report a route to a nacre-like $CaCO_3$ multilayer that includes all 5 steps outlined above, resulting in a stack of crystalline calcite layers, interconnected though porous organic films. Both the growth strategy and final material bear close resemblance to biogenic nacre.



Figure 1 Comparison of biogenic and artificial nacre. a, Photograph showing nacre's bright iridescence (scale bar 5 mm). **b**, Fractured surface SEM image of a stack of mineral tablets (scale bar $2 \mu m$). **c**, Organic inter-crystalline film which allows for vertical crystal continuity between tablets (scale bar 500 nm). **d**, Artificial nacre, exhibiting a similar coloration as in **a** (scale bar 5 mm). **e**, SEM image of fractured surface showing 7 aligned CaCO₃ tablets separated by organic films. The surface graininess is comparable to natural nacre (scale bar $1 \mu m$). **f**, SEM image of PVP film on calcite showing the a similar pore distribution as in **c** (scale bar 300 nm). **g**, AFM height image of the porous film (scale bar 300 nm).



Figure 2 | Scheme of artificial nacre synthesis. a, A continuous PAA/PVP film was formed by sequentially immersing a glass slide into PAA and PVP solutions for 1 min. each. b, Transformation of the layer into a porous film: (1) Immersion in a basic solution dissolves PAA and induces pores by dewetting. (2) Stabilisation by UV cross-linking. (3) Surface functionalisation by PAA immersion. c, Mineral film formation in close contact with gas permeable beaker wall. PILP droplets form by CO_2 diffusion, stabilised by included PAA and Mg to form a modified calcite (cal_{org}) film. d, Crystallisation of the most recently deposited cal_org layer. Each full cycle produces a single organic/mineral bilayer, which is repeated to create a stack.

Results

Fabrication

Our experimental procedure is illustrated in Fig. ??. Initially, layer-by-layer (LbL) deposition of two polyelectrolytes was used to form a continuous organic thin film^{?,?} by cyclicly immersing a glass substrate into poly (acrylic acid) (PAA) and poly (4-vinyl pyridine) (PVP) solutions until the desired film thickness was obtained (Fig. ??a). The film is then immersed into a pH 10 solution, dissolving the PAA and inducing a positive charge on the substrate and an in the PVP. The resulting repulsive interaction leads to nanopore formation by PVP dewetting, which is quenched and stabilised by UV cross-linking[?]. A final immersion into PAA solution functionalises the PVP surface with COO⁻ groups to promote mineral deposition (Fig. ??b).

Mineral growth in Fig ??c is based on the biological strategy using ACC^{?,?,?,?}. Using the the ammonium carbonate diffusion technique with a PAA-containing 1:5 Ca²⁺:Mg²⁺ solution (Mg²⁺ and PAA suppress calcite rhombohdra formation)[?] yields stable polymer-induced liquid precursor (PILP) droplets which wet the carboxyl terminated substrate and coalesce into a ACC film, rich in Mg and PAA (cal_{org})^{?,?}. A key advance in extending this mineral deposition technique to create large (10 cm²) high quality films is the confinement of the reaction volume by a gas permeable membrane, providing uniform distribution of CO₂ in the solution.

The ACC is then crystallised by exposure to high humidity, allowing for a dissolution-recrystallisation transformation? (Fig. ??d). The propagation of crystallisation



Figure 3 | Characterisation of artificial nacre crystallinity. **a**, SEM image showing early stage crystallisation through single PVP pores in a trilayer of cal_{org}-organic-cal_{org} (scale bar 500 nm) **b**, Optical micrograph showing 5 - 35 µm birefringent crystalline domains (right: though crossed polarisers) (scale bar 25 µm). **c**, TEM-SAED of a single tablet evidencing calcite single crystallinity. **d**, Cube corner indentation (imaged by AFM) caused crack-formation in a cal_{org} film (left) and plastic deformation in a 7-layer artificial nacre sample (right), similar to biogenic nacre (Supplementary Fig. S9) (scale bar 1 µm).

occurs through the organic pores, connecting the mineral layers. The fabrication of an organic/cal_{org} bilayer (an entire cycle of steps a - d) can be achieved in 5 hours, making it viable to grow many multilayers via repetition.

Microstructure

The resulting material is highly comparable to biogenic nacre (compare Fig. ??a-c to ??d-g) both in its morphology and growth route. In particular, a comparison of the scanning electron microscopy (SEM) fractured surfaces of biogenic (Fig. ??b) and artificial (Fig. ??e) nacre reveals very similar multilayers of 400 nm thick cal_{org} tablets with a nano-granular texture that is characteristic of nacre.

A key advantage of our route to artificial nacre is the fabrication of the porous organic inter-crystalline layers, which are comparable to those in natural nacre (Fig. ??c,f,g, Supplementary Table S1 and Supplementary Fig. S2). These pores allow bridges between the mineral films to form, providing vertical crystalline mineral continuity which enhances nacre's mechanical stability?,?. Without these interconnects the layered structure is unstable during growth and delaminates. This is readily demonstrated by modifying step b in Fig. ??, so that no pores are formed in the organic layers. This leads to highly unstable stacks of delaminating layers (Supplementary Fig. S3). Earlier approaches were also plagued by the delamination of non-connected layers caused by the stresses arising from the dehydration of ACC^{?, ?, ?}.

The pores allow the propagation of crystallisation across the organic layers, which promotes biomimetic crystal sizes that span a number of layers. Figure ??a confirms the role of the pores in crystal propagation. Quenching the crystallisation of the amorphous top layer after only 10 min. followed by dissolution of the remaining amorphous material reveals facetted seed crystallisation in the pores (also Supplementary Fig. S4,S5). Facets are suppressed while the mineral is in contact with a soft organic material, but crystallinity is nevertheless propagated through the pores, in agreement with previous studies^{?,?}.

While SEM and AFM images of the cal_{org} tablets do not resolve any crystal grain boundaries, optical microscopy between cross-polarisers reveals a polycrystalline structure with grain sizes in the 5-35 µm range (Fig. ??b). This is much larger than any other length scale (tablet thickness, pore-size and distance, etc.) in the material. The single-crystal nature of the large domains was confirmed by transmission electron microscopy selected area electron diffraction (TEM-SAED) of individual tablets (Fig. ??c & Fig. ??). The similarity of crystal domain size in Fig. ??b to natural nacre[?] is remarkable, given the close pore spacing of the organic interlayers.

The step-by step nature of our approach allows to elucidate the origin of the crystal domain size. The investigation of only on single layer shows a similar crystal morphology as the stack (Supplementary Fig. S6). The crystal domain size is therefore determined by the nucleation density of the first layer, which can be controlled by the relative Mg content?. Despite the high pore density and 10 nm pore size, the lateral crystal morphology is preserved in successive layers. To this end it is necessary that the crystal growth through the pores has the same spatial symmetry, leading to a nanogranular morphology of grains that have the same crystallographic orientation and therefore give rise to the clear diffractogram of Fig??c. Mineral continuity across the organic sheets is therefore important to maintain the co-orientation of all tablets in the columnar stack?, their crystal structure and the mechanical cohesion of the layers structure.

Mechanical Properties

The similar morphology of natural and artificial nacre is evidenced by comparable mechanical properties. Firstly, mean plain strain moduli of 69, 38, 176, 86, 10 GPa for natural and artificial nacre, aragonite, cal_{org} and PVP respectively were measured (Supplementary Fig. S7). The smaller elastic modulus of artificial nacre, compared with natural nacre, is a direct consequence of PAA and Mg inclusion within cal_{org}, seen in Supplementary Fig. S8. The modulus of both nacre types is well described by $1/E_{\text{nacre}} = \phi_1/E_1 + \phi_2/E_2$, where ϕ_i is the volume fraction of the (1) inorganic and (2) organic phases. The very similar morphologies therefore give rise to identical ratios of moduli of natural/artificial nacre to aragonite/cal_{org}, respectively.



Figure 4 | **Optical characterisation.** Reflectivity as a function of wavelength and reflection angle showing **a** biogenic and **b** artificial nacre containing 7 bilayers (bilayer = cal_{org}/organic). The wavelength-dependent angles of maximal intensity (red) extending diagonally across the images result from multilayer interference. **c**, Simulation of a 7-bilayer stack of artificial nacre. The mineral thickness was averaged over ± 15 nm to factor in disorder. **d**, The reflection profile from **b** at an angle of 35° is compared with a calculated spectrum of a perfect CaCO₃/PVP multilayer and a spectrum cut from the disordered simulation in **c**.

More important than the stiffness characteristics mimicking natural nacre, the fracture toughness was qualitatively probed by cube-corner indentation (Fig. ??d). This showed plastic deformation in artificial nacre, under loads which cracked solid cal_{org}, demonstrating the toughening of the alternating cal_{org}-PVP stack. This result is analogous to the comparison of toughening of aragonite in natural nacre (Supplementary Fig. S8).

Optical Properties

The similarities in iridescence between biological and artificial nacre (Fig. ??a,d) were spectroscopically investigated. Figures ??a,b show the specularly reflected signal as a function of incident angle and wavelength for the two types of nacre. Both spectra have the same salient features: the two prominent reflections bands that diagonally span the spectra correspond to second and third order interference which is characteristic of multiaver stacks. The similar location and width of these spectral features lies at the origin of the similar macroscopic optical appearance of Fig. ??a and c. The difference between the two stems from the variation of the intensity of the interference bands with angle, which arises from the increased scattering in biogenic nacre. The spectral response of the artificial nacre was compared to a simulated 7-bilayer system in Fig. ??c,d. A perfect stack has very narrow interference peaks and minor oscillations stemming from its finite size. Including irregularities by introducing layer thickness vari-



Figure 5 | TEM Cross section showing continuity of crystallinity across one organic layer. A trilayer of $cal_{org}/organics/cal_{org}$ was ion-beam milled to expose the cross-sectional surface. **a**, TEM imaging (scale bar 20 nm) and **b** electron diffraction reveals that pseudo-single crystallinity is maintained across this porous organic layer (bounded by the dashed lines in **a**).

ations of ± 15 nm gives rise to a simulated spectrum which qualitatively approaches the measured specular reflection of artificial nacre, seen in Fig. ??c,d. The broader interference bands in Fig. ??a,b also arise from scattering, which was not included in the simulation.

Single crystallinity across organic layers

Fig. ?? shows evidence of crystal continuity across one organic layer. The selected area diffraction in Fig. ??b was performed over a 1µm-wide area, including comparable areas on both sides of the organic layer (delimited by dashed lines in Fig. ??a). The bright indexed spots indicate a single crystalline material, although some weak background spots can be seen. This might be caused by a small minority of alternatively ordered crystals, although the signal could simply arise from the ion milling process.

It was not possible to find a cross-section that included a calcite bridge across an organic pore. We are, however, confident that the interconnecting mineral bridges are the source of this crystalline continuity because in their absence there is no mechanism for crystalline orientation to propagate across the 30 nm thick amorphous organic layers. The proliferation of crystallinity across a stack of tablets requires physical connectivity, and Fig. **??** is clear evidence that the crystallinity does indeed spread from one layer to the next.

These experiments were repeated along the organic interface, consistently showing that crystallinity was propagated from one cal_{org} tablet to the next across the organic film. The crystal orientation was seen to change occasionally during the scan, but the lateral boundaries between adjacent tables could not be resolved in the TEM crosssections.

Single-crystal diffraction of a small-particle assembly

The granular nature seen in Fig. ??a stems from the aggregation of PILP droplets which later crystallise into the tablets. The ED image in Fig. ??b reveals that this agglomeration of calcite nanoparticles diffracts as a single crystal, similarly to biogenic nacre. This seemingly paradoxical observation is common in biominerals, and is indicative of the oriented attachment of granular building blocks during calcium carbonate precipitation and crystallisation under biological (or biomimetic) conditions. This is a result of the amorphous precursor which forms nanoscale hydrated droplets that join to form an amorphous network, and later crystallise via a single nucleation event, or propagate the crystallinity of a crystal substrate. This is well documented for biological?,?,?,?,?,? and artificial^{?,?,?,?} systems and thus further emphasises the biomimetic nature of our artificial nacre.

Conclusions

We present a route to artificial nacre which resembles the biogenic material in terms of the detailed synthesis route, its structure as well as in it mechanical and optical properties. By carefully mimicking the steps in a cyclical deposition protocol, an organic-rich calcite mutlilayer stack that approximates biological nacre in all essential points was formed. These consist of (1) a 5-35 µm-sized polycrystalline cal_{org} structure organised in 400 nm-thick plates that are (2) interconnected via the holes in the PVP layers, which ensures crystal continuity. (3) The composite structure gives rise to nacre-like enhanced toughness. (4) The good control over the layer periodicity reproduces nacre's iridescence. The close replication of nacre allows to test each of the biological steps, elucidating for example the interplay of organic pore structure and crystal domain size. Technologically, this biomimetic process heralds the manufacture of tough surface coatings from cheap base materials made by low temperature sustainable methods.

Methods

Manufacture of artificial nacre. Poly (4-vinyl pyridine) (PVP, $M_{\rm n} = 60 \, \rm kg/mol$), poly (acrylic acid) (PAA, $M_{\rm n} = 1.8 \, \rm kg/mol$), poly (acrylic acid) sodium salt (PAA-Na, $M_{\rm n} = 5 \, \rm kg/mol$) and ammonium carbonate were bought from Sigma and used as received.

Glass slides were cleaned by immersion in freshly prepared piranha solution (concentrated $H_2SO_4: H_2O_2$ 3:1 by weight) for 1 h at 100 °C followed by rinsing with 18.2 M Ω cm water. The substrates were coated with a continuous PVP film by spin coating from a 1% solution in ethanol at 3000 rpm for 1 minute. This layer was cross-linked under 254 nm UV light for 5 minutes. Carboxyl functionalisation was achieved by dipping in a $10 \ {\rm wt/v\%}$ PAA solution for 1 minute followed by thorough rinsing in water. Samples were placed in a gaspermeable silicone beaker, secured against the side wall. The beaker was filled with 250 µl of PAA-Na stock solution $(20 \ {\rm wt/v\%})$ and 50 ml of a fresh aqueous solution of $16 \ {\rm mM} \ {\rm CaCl_2}$ and $80 \ {\rm mM} \ {\rm MgCl_2}$. The beaker was placed in a 11 sealed desiccator with 6g of freshly crushed ammonium carbonate and left for 2 h at room temperature. Amorphous calcium carbonate (ACC) was deposited via hydrated ${\rm Ca^{2+}/Mg^{2+}/PAA}$ ion clusters, which coalesce to form a smooth film. The samples were removed, rinsed in pure water and dried in a nitrogen stream.

Layer-by-layer (LbL) deposition was carried out by sequentially immersing samples in a 100 ml 1 mg/ml PAA methanol solution for 2 minutes, followed by rinsing in two pure methanol solutions for 1 minute each, then in a 100 ml 1 mg/ml PVP methanol solution followed by the same rinsing cycle. Using a Janome JR2400N robot, this two-step process was repeated 10–30 times to deposit a PAA/PVP layer with a well-defined film thickness. Samples were then placed in a pH 10 NaOH solution for 5 minutes. PAA dissolves out from the film during the first minute, followed by PVP dewetting into a porous membrane[?]. The films were rinsed in pure water for a further 5 minutes before the UV and PAA treatment and continued CaCO₃ growth as described above.

The ACC film was finally crystallised by placing dry multilayers in the same desiccator with 10 ml of warm water for 6 hours, causing a humidity of >95%. This gave rise to Mg and PAA-containing single crystal calcite. Note that in natural nacre aragonite forms from a similar $Ca^{2+}:Mg^{2+}$ ratio in water. ACC-mediated aragonite growth has however not been replicated in vitro.

This whole process forms a single bilayer and is repeated to form the artificial nacre.

Sample characterisation.

The biogenic and artificial nacre were imaged by optical microscopy with crossed polarisers. Reflection spectra were recorded using an angular resolved setup. The light from a incandescent source (Ocean Optics, with a wavelength-range of 230-2000 nm) was incident on the sample via an optical fibre illuminating and area of 3 mm^2 . The illumination angle ranged between $15-75^\circ$ with respect to the surface normal. The specularly reflected light was coupled into an optical fibre connected to an Ocean Optics QE65000 spectrometer.

Thermogravimetric Analysis (TGA) was performed on a TA Instruments Q500 system, running from 25 °C to 900 °C under air flow. The angular resolution of the setup was ~ 1°. X-ray analysis was performed using a Bruker D8 diffractometer with position-sensitive detector (Lynx-Eye). A single CaCO₃ layer was grown on a silicon wafer with (510) crystal orientation, which has no diffraction peaks in the 2θ region of interest. The diffraction pattern was compared to the spectra of calcite (PDF 830577) and dolomite (PDF 74-1687) crystals. For scanning electron microscopy, samples were fractured and sputtercoated with a 4 nm thick Au/Pd layer. Scanning electron microscopy (SEM) imaging was performed on a LEO 1530 operating at 5 kV at a sample angle of 45° . A Veeco Dimension 3100 atomic force microscope (AFM) was used in tapping mode to image the porous PVP layer. Transmission electron microscopy (TEM) selected area electron diffraction (SAED) to determine CaCO₃ crystallinity was carried out using a FEI Tecnai 20 on a CaCO₃ film on a holey carbon grid at 200 kV with a camera length of 890 mm. A 20 µm aperture was used for SAED of whole tablets, and 1 µm for cross-sections. Ion beam milling was performed on an FEI Philips Dualbeam Quanta 3D system.

Mechanical Measurements.

Nanoindentation tests were performed on a UBI 1 (now TI-700, Hysitron Inc., Minneapolis, MN) on a single crystal of aragonite, biogenic nacre sample, CaCO₃film, replica, and PVP. A Berkovich tip was employed for material property measurements in load control. A trapezoidal load profile was used, with 10 s loading, holding and unloading times. The peak load varied by sample to maintain indentation displacements in the range 30 - 150 nm (150, 250, 500 µN for each material respectively). Between 20 and 110 individual indentation tests were performed on each sample. The plain strain elastic modulus was calculated from the unloading stiffness via a calibrated tip-area function that was validated on fused silica and aluminium standard samples⁷.

A cube-corner tip was used to investigate the fracture behaviour via nanoindentation. A triangular load profile was used with 2 s loading and unloading times. The peak load was 5 mN for aragonite and nacre samples and 1.5 mN for CaCO₃ and Replica samples. Following cube-corner indentation, the residual indent was scanned in SPM mode at a contact force of 1.5 - 2 uN to ascertain whether cracking had occurred. Five individual indentation tests were scanned per sample.

References

- Cartwright, J. H. E. & Checa, A. G. The dynamics of nacre self-assembly. J. R. Soc. Interface 4, 491–504 (2007).
- Weiner, S. Biomineralization: A structural perspective. J Struct Biol. 163, 229–234 (2008).
- Weiss, I., Tuross, N. & Addadi, L. Mollusc larval shell formation: Amorphous calcium carbonate is a precursor phase for aragonite. J Exp Zool 293, 478–491 (2002).
- Addadi, L., Joester, D., Nudelman, F. & Weiner, S. Mollusk shell formation: A source of new concepts for understanding biomineralization processes. *Chem-Eur J* 12, 981–987 (2006).
- Song, F., Soh, A. K. & Bai, Y. L. Structural and mechanical properties of the organic matrix layers of nacre. *Biomaterials* 42, 3623–3631 (2003).

- Jackson, A., Vincent, J. & Turner, R. The mechanical design of nacre. P Roy Soc Lond B Bio 234, 415–440 (1988).
- Currey, J. D. The design of mineralised hard tissues for their mechanical functions. J Exp Biol 202, 3285–3294 (1999).
- Smith, B. *et al.* Molecular mechanistic origin of the toughness of natural adhesives, fibres and composites. *Nature* **399**, 761– 763 (1999).
- Schäffer, T. *et al.* Does abalone nacre form by heteroepitaxial nucleation or by growth through mineral bridges? *Chem Mater* 9, 1731–1740 (1997).
- Kato, T. Polymer/calcium carbonate layered thin-film composites. *Adv. Mater.* 12, 1543–1546 (2000).
- Wei, H., Ma, N., Shi, F., Wang, Z. & Zhang, X. Artificial nacre by alternating preparation of layer-by-layer polymer films and CaCO₃ strata. *Chem Mater* **19**, 1974–1978 (2007).
- Gong, H. *et al.* Multilayered CaCO₃/block-copolymer materials via amorphous precursor to crystal transformation. *Colloid Surface A* 354, 279–283 (2010).
- Gehrke, N. et al. Retrosynthesis of nacre via amorphous precursor particles. Chem Mater 17, 6514–6516 (2005).
- Tang, Z., Kotov, N., Magonov, S. & Ozturk, B. Nanostructured artificial nacre. Nat Mater 2, 413–418 (2003).
- Bonderer, L. J., Studart, A. R. & Gauckler, L. J. Bioinspired design and assembly of platelet reinforced polymer films. *Science* **319**, 1069–1073 (2008).
- Aksay, I. *et al.* Biomimetic pathways for assembling inorganic thin films. *Science* 273, 892–898 (1996).
- 17. Burghard, Z. *et al.* Toughening through nature-adapted nanoscale design. *Nano Lett* **9**, 4103–4108 (2009).
- Munch, E. *et al.* Tough, bio-inspired hybrid materials. *Science* 322, 1516–1520 (2008).
- Yao, H.-B., Tan, Z.-H., Fang, H.-Y. & Yu, S.-H. Artificial nacre-like bionanocomposite films from the self-assembly of chitosan-montmorillonite hybrid building blocks. *Angew Chem Int Edit* 49, 10127–10131 (2010).
- Decher, G., Hong, J. & Schmitt, J. Buildup of ultrathin multilayer films by a self-assembly process. *Thin Solid Films* 210, 831–835 (1992).
- Fu, Y. *et al.* Hydrogen-bonding-directed layer-by-layer multilayer assembly: Reconformation yielding microporous films. *Macromolecules* 35, 9451–9458 (2002).
- Burtovyy, R. & Luzinov, I. Reversibility of pH-induced dewetting of poly(vinyl pyridine) thin films on silicon oxide substrate. *Langmuir* 24, 5903–5910 (2008).
- Finnemore, A. S. *et al.* Nanostructured calcite single crystals with gyroid morphologies. *Adv. Mater.* 21, 3928–3932 (2009).
- Ludwigs, S., Steiner, U., Kulak, A. N., Lam, R. & Meldrum, F. C. Bioinspired polymer-inorganic hybrid materials. *Adv. Mater.* 18, 2270–2273 (2006).
- Wucher, B., Yue, W., Kulak, A. N. & Meldrum, F. C. Designer crystals: Single crystals with complex morphologies. *Chem Mater* **19**, 1111–1119 (2007).
- Aizenberg, J., Muller, D., Grazul, J. & Hamann, D. Direct fabrication of large micropatterned single crystals. *Science* 299, 1205–1208 (2003).
- Addadi, L., Moradian, J., Shay, E., Maroudas, N. & Weiner, S. A chemical-model for the cooperation of sulfates and carboxylates in calcite crystal nucleation - relevance to biomineralization. *P Natl Acad Sci Usa* 84, 2732–2736 (1987).
- Gower, L. & Odom, D. Deposition of calcium carbonate films by a polymer-induced liquid-precursor (PILP) process. *J Cryst Growth* **210**, 719–734 (2000).

- Cheng, X., Varona, P. L., Olszta, M. J. & Gower, L. B. Biomimetic synthesis of calcite films by a polymer-induced liquid-precursor (pilp) process 1. influence and incorporation of magnesium. *J Cryst Growth* **307**, 395–404 (2007).
- Xu, X., Han, J., Kim, D. & Cho, K. Two modes of transformation of amorphous calcium carbonate films in air. *J Phys Chem B* 110, 2764–2770 (2006).
- 31. Gilbert, P. *et al.* Gradual ordering in red abalone nacre. *J. Am. Chem. Soc.* **130**, 17519–17527 (2008).
- Politi, Y. *et al.* Transformation mechanism of amorphous calcium carbonate into calcite in the sea urchin larval spicule. *Proceedings of the National Academy of Sciences* **105**, 17362 (2008).
- Rousseau, M. et al. Multiscale structure of sheet nacre. Biomaterials 26, 6254–6262 (2005).
- Sethmann, I. & Wörheide, G. Structure and composition of calcareous sponge spicules: A review and comparison to structurally related biominerals. *Micron* 39, 209–228 (2008).
- Sethmann, I., Hinrichs, R., Wörheide, G. & Putnis, A. Nanocluster composite structure of calcitic sponge spicules—a case study of basic characteristics of biominerals. *J Inorg Biochem* 100, 88–96 (2006).
- 36. Wilt, F. Matrix and mineral in the sea urchin larval skeleton. Journal of Structural Biology **126**, 216–226 (1999).
- Zaremba, C. *et al.* Atomic-force microscopy of the nacreous layer in mollusk shells. *P Roy Soc Lond B Bio* 256, 17–23 (1994).
- Zhan, J., Lin, H. & Mou, C. Biomimetic formation of porous singlecrystalline caco3 via nanocrystal aggregation. Adv. Mater. 15, 621–623 (2003).

39. Oliver, W. & Pharr, G. An improved technique for determining hardness and elastic-modulus using load and displacement sensing indentation experiments. *Journal of Materials Research* **7**, 1564–1583 (1992).

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Author Contributions

All authors designed their experiments. A. F. performed fabrication experiments. T. S. Performed mechanical measurements. S. V. performed optical measurements. S. G. Performed x-ray diffraction experiments. A. F. and U. S. wrote the paper. All authors provided intellectual input, read and approved the manuscript.