

Brassinosteroids control root epidermal cell fate via direct regulation of a MYB-bHLH-WD40 complex by GSK3-like kinases

Yinwei Cheng¹, Wenjiao Zhu¹, Yuxiao Chen¹, Shinsaku Ito², Tadao Asami², Xuelu Wang^{1,3*}

¹State Key Laboratory of Genetic Engineering, and Institute of Plant Biology, School of Life Sciences, Fudan University, Shanghai, China; ²Graduate School of Agricultural and Life Sciences, The University of Tokyo, Tokyo, Japan; ³College of Life Science and Technology, Huazhong Agricultural University, Wuhan, China

Abstract In *Arabidopsis*, root hair and non-hair cell fates are determined by a MYB-bHLH-WD40 transcriptional complex and are regulated by many internal and environmental cues. Brassinosteroids play important roles in regulating root hair specification by unknown mechanisms. Here, we systematically examined root hair phenotypes in brassinosteroid-related mutants, and found that brassinosteroid signaling inhibits root hair formation through GSK3-like kinases or upstream components. We found that with enhanced brassinosteroid signaling, *GL2*, a cell fate marker for non-hair cells, is ectopically expressed in hair cells, while its expression in non-hair cells is suppressed when brassinosteroid signaling is reduced. Genetic analysis demonstrated that brassinosteroid-regulated root epidermal cell patterning is dependent on the WER-GL3/EGL3-TTG1 transcriptional complex. One of the GSK3-like kinases, BIN2, interacted with and phosphorylated EGL3, and EGL3s mutated at phosphorylation sites were retained in hair cell nuclei. BIN2 phosphorylated TTG1 to inhibit the activity of the WER-GL3/EGL3-TTG1 complex. Thus, our study provides insights into the mechanism of brassinosteroid regulation of root hair patterning.

DOI: 10.7554/eLife.02525.001

*For correspondence: xueluw@gmail.com

Competing interests: The authors declare that no competing interests exist.


Funding: See page 15

Received: 12 February 2014

Accepted: 01 April 2014

Published: 25 April 2014

Reviewing editor: Sheila McCormick, University of California, Berkeley and USDA Agricultural Research Service, United States

 Copyright Cheng et al. This article is distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use and redistribution provided that the original author and source are credited.

Introduction

The *Arabidopsis* root epidermal cell types are defined by position in a predictable manner (*Ishida et al., 2008*). Hair (H) cells, or trichoblasts, are specified early from root epidermal cells that lie over clefts between two underlying cortical cells, whereas the root epidermal cells that lie over a single cortical cell develop as non-hair (N) cells, or atrichoblasts (*Ishida et al., 2008*). Hair cell and non-hair cell files are patterned alternately in rows within the *Arabidopsis* root epidermis, with columns of hair cells interspersed with columns of non-hair cells (*Schiefelbein et al., 2009*). Prior to root hair outgrowth, root epidermal cells in the H position can be distinguished from those in the N position by many visible cellular features, including a greater rate of cell division (*Berger et al., 1998*), reduced cell length and vacuolation (*Dolan et al., 1994; Galway et al., 1994*), and enhanced cytoplasmic density (*Dolan et al., 1994*). It is proposed that positional signals and a putative receptor-like kinase SCRAMBLED (SCM) (*Kwak et al., 2005*) function through a MYB-bHLH-WD40 repeat transcriptional complex to determine root epidermal cell fate (*Schiefelbein et al., 2009*). Based on this model, in N cells, WEREWOLF (WER) (*Lee and Schiefelbein, 1999*), a R2R3 MYB-domain transcription factor, forms a complex with basic helix-loop-helix transcription factors, GLABRA3 (GL3)/ENHANCER OF GLABRA3 (EGL3) (*Bernhardt et al., 2003; Zhang et al., 2003*), and a WD40 repeat protein, TRANSPARENT

eLife digest Roots anchor a plant into the ground, and allow the plant to absorb water and mineral nutrients from the soil. As roots grow and branch, they increase the surface area of root exposed to the soil—and many plant cells in the root's outer layer have a hair-like projection to further increase this surface area. Thus, root hairs are where most water and mineral nutrients are absorbed. Many factors affect whether, or not, a plant cell will develop into a root hair. These factors include both external cues (such as the mineral content of the soil) and signals from the plant itself (such as hormones).

Brassinosteroids are plant hormones that regulate the development of shoots and roots, as well as the timing of when flowers begin to develop. These hormones are detected on the outside of plant cells, and activate a signaling pathway within the cell that causes changes in gene expression. Brassinosteroids also control if a root cell will become a hair cell or not, although the mechanism behind this activity is unclear.

Here, Cheng et al. have looked at the root hairs of mutant *Arabidopsis thaliana* plants that have had individual genes involved in brassinosteroid signaling knocked-out. Plant biologists commonly study this plant species because it is small and grows quickly—and *Arabidopsis* has regular stripes of root hair cells and 'non-hair cells' in the outer layer of its roots. Cheng et al. reveal that brassinosteroids prevent the formation of root hairs via signaling pathways that involve proteins called GSK3-like kinases. These hormones 'switch off' these kinases' activity, so knocking-out the genes that code for these kinases has the same effect as adding extra brassinosteroids to the plant roots: fewer root hair cells.

Cheng et al. show that one of the GSK3-like kinases binds and adds phosphate groups to protein complexes that control gene expression—and this causes these protein complexes to be less active. When GSK3-like kinase activity is switched off by brassinosteroids, these complexes instead become more active and trigger the expression of genes that direct a plant cell to become a non-hair cell.

The findings of Cheng et al. reveal the pathways that allow brassinosteroids to stop plant cells in roots from becoming hair cells, and that instead encourage these cells to become non-hair cells. However, further work is needed to uncover how the striped pattern of hair cells and non-hair cells on *Arabidopsis* roots is established, and how brassinosteroids work with other plant hormones to control this pattern.

DOI: [10.7554/eLife.02525.002](https://doi.org/10.7554/eLife.02525.002)

TESTA GLABRA1 (TTG1) (Galway et al., 1994), to promote expression of GLABRA2 (GL2) and CAPRICE (CPC) (Ryu et al., 2005; Song et al., 2011). GL2, a homeodomain/leucine zipper transcription factor, negatively regulates H cell fate and positively regulates N cell fate (Masucci et al., 1996). CPC (Wada et al., 1997), a MYB-type transcription factor, moves from N cells to H cells (Kurata et al., 2005a) to compete with WER for binding to GL3/EGL3 to form a CPC-GL3/EGL3-TTG1 complex, which is unable to induce GL2 expression (Song et al., 2011). In addition to CPC, the bHLH transcription factor GL3 is also a mobile protein (Bernhardt et al., 2005). GL3 and its homologue EGL3 are both expressed in H cells, but GL3 protein is only localized in the N cell nucleus, indicating that GL3 protein moves into the adjoining N cell nucleus to determine N cell fate (Bernhardt et al., 2003, 2005). Integration of existing genetic and biochemical data also supports an alternative mechanism centered on the movement of transcriptional factors between epidermal cells rather than a putative local activation of the WER gene function to determine root epidermis pattern formation (Savage et al., 2008).

In addition, root hair development is highly regulated by many external and internal cues, including phytohormones. For instance, abscisic acid (ABA) plays a role in the early stage of root epidermal cell specification (Van Hengel et al., 2004) and in inhibiting root hair tip growth in *Arabidopsis* (Schnall and Quatrano, 1992), while both ethylene and auxin may act downstream of TTG1 and GL2 to promote root hair formation and elongation (Masucci and Schiefelbein, 1994, 1996). Moreover, jasmonic acids (JAs) promote root hair formation through their interaction with ethylene (Zhu et al., 2006). However, the underlying cellular and molecular mechanisms of how these internal hormones integrate with environmental cues to regulate root hair cell fate determination are still poorly understood.

The plant steroid hormones, brassinosteroids (BRs), play essential roles in regulating many developmental processes, including shoot, root, and reproductive development (Savaldi-Goldstein et al., 2007;

Ye et al., 2010; Hacham et al., 2011; Yang et al., 2011). BRs are perceived by the receptor kinase BRASSINOSTEROID INSENSITIVE 1 (BRI1) (Li and Chory, 1997; Hothorn et al., 2011; She et al., 2011). The BR-activated BRI1 phosphorylates BRI1 KINASE INHIBITOR 1 (BK1) to release its inhibition (Wang and Chory, 2006), and then BK1 acts as a positive regulator by binding to a subset of 14-3-3 proteins (Wang et al., 2011). Another BRI1 substrate, BR-SIGNALING KINASE (BSK), transduces the BR signaling through *bri1* SUPPRESSORS 1 (BSU1) to inactivate a GSK3-like kinase BRASSINOSTEROID INSENSITIVE 2 (BIN2), which leads to accumulation of the dephosphorylated form of transcriptional factors BRI1 EMS SUPPRESSOR 1 (BES1)/BRASSINAZOLE RESISTANT 1 (BZR1) in the nucleus to regulate gene expression (Yang et al., 2011). A previous study suggests that BRs play an important role in determining root epidermal cell fate by regulating *WER* and *GL2* expression (Kuppusamy et al., 2009). However, the elaborate molecular mechanism by which BRs regulate root epidermal cell fate and development is still unknown.

Here, we first systematically examined root epidermal cell patterning and *PGL2::GUS* expression in a series of BR-deficient and signaling mutants. We found that BRs regulate root epidermal cell fate through promoting *GL2* expression in both H and N cells, which is mediated by GSK3-like kinases and the *WER-GL3/EGL3-TTG1* complex as indicated by genetic analysis and biochemical studies. Our study further demonstrated that BIN2, one of the GSK3-like kinases, interacted with and phosphorylated EGL3 on T399 and T209/T213, leading to its trafficking from nucleus to cytosol in H cells, which may facilitate its movement from H cells to N cells. BIN2 also phosphorylated TTG1 to inhibit the activity of the *WER-GL3/EGL3-TTG1* transcriptional complex. These results explain how BR signaling regulates both the formation and activity of the *WER-GL3/EGL3-TTG1* complex through GSK3-like kinases to coordinate root epidermal cell fate specification.

Results

BRs regulate root epidermal cell patterning through GSK3-like kinases

To broadly explore the role of BRs in root hair formation, we systematically examined the root hair phenotype of BR-biosynthetic mutants, *det2-1* and *cpd*, and BR-responsive mutants, including *bri1-116*, *BRI1-OX* (a *BRI1*-overexpression line), *bin2-3 bil1 bil2* (a triple knockout mutant of *BIN2* and its two closest homologues), and a *BES1-RNAi* line. We found that the relative hair number (=root hair density \times root hair cell length) was higher in *bri1-116* (4.67 ± 0.47), *det2-1* (4.75 ± 0.52), and *cpd* (4.65 ± 0.54), and significantly lower in *BRI1-OX* (3.40 ± 0.46) and *bin2-3 bil1 bil2* (2.86 ± 0.39) than in their corresponding wild types Col-0 (3.89 ± 0.43) and WS-2 (3.85 ± 0.41) (Figure 1—source data 1). However, there was no significant difference between *BES1-RNAi* and Col-0. Images at the highest magnification ($\times 100$; Figure 1) showed that in the wild type root (Figure 1A), the H cells and N cells were arranged in alternating files with the H cell columns regularly interspaced with the N cell columns; no adjacent H cell columns were found. However, in the BR signaling-inhibited mutants, including *bri1-116*, *det2-1*, and *cpd* (Figure 1B–D), many root hair columns were next to each other, leading to more root hairs, suggesting that some N cell fate might be changed into H cell fate. In contrast, the BR signaling-enhanced plants, including *BRI1-OX* and *bin2-3 bil1 bil2*, grew fewer root hairs than the wild type, due to the fact that they lacked root hairs in many H cell positions (Figure 1A,E–G). Interestingly, the *BES1-RNAi* line showed a similar root hair pattern as the wild type (Figure 1A,H).

To further test whether exogenously applied eBL (epibrassinolide), bikinin (a specific GSK3 kinase inhibitor) (De Rybel et al., 2009), or Brz (brassinazole, an inhibitor of BR biosynthesis) regulate root hair specification, we planted seeds on 1/2 MS (Murashige and Skoog) medium containing each of these chemicals or DMSO (as the mock treatment), and carefully observed their root hair phenotypes. We found that compared with plants grown on the mock medium (3.80 ± 0.45), the relative root hair number of plants grown on medium containing 100 nM eBL (3.22 ± 0.42) or 30 μ M bikinin (2.71 ± 0.50) was significantly reduced, while it was greatly increased in plants grown on medium containing 1 μ M Brz (4.17 ± 0.43) (Figure 1—source data 1). Images at higher magnification ($\times 100$) show that, compared to seedlings grown on the mock medium (Figure 1I), those grown on medium containing eBL or bikinin produced fewer root hairs in the H position of epidermal cells (Figure 1J,K), while those grown on medium containing Brz grew more root hairs in the N position (Figure 1L). We also found that the *bin2-3 bil1 bil2* seedlings grown on medium containing 30 μ M bikinin produced very few root hairs (Figure 1—figure supplement 1), suggesting that besides *BIN2*, *BIL1*, and *BIL2*, other GSK3-like kinases may also be involved in root hair specification in *Arabidopsis*. Taken together, these findings

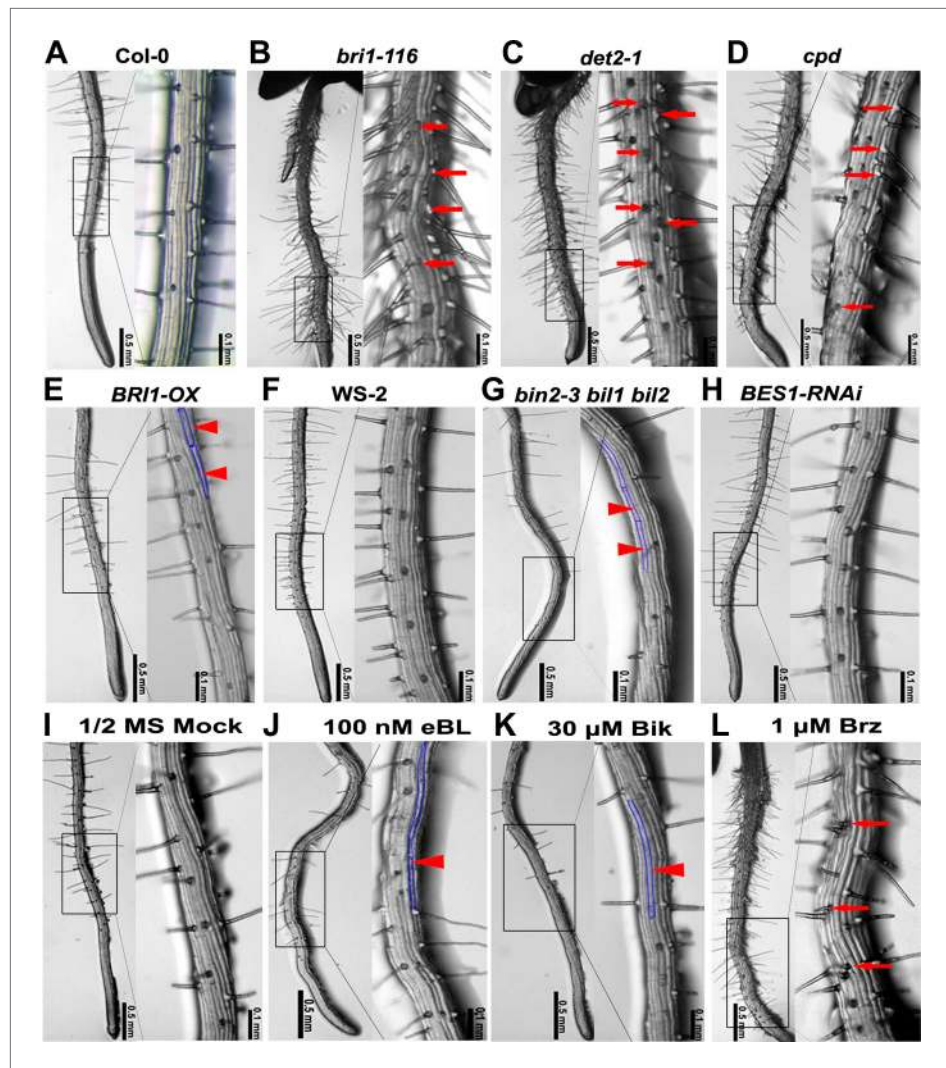


Figure 1. Root epidermal cell patterning is altered in the BR-related mutants. (A–H) Root hair patterning of the BR-related mutants and their wild type counterparts. *bin2-3 bil1 bil2* is in the WS-2 background, and all of the others are in the Col-0 background. (I–L) Root hair phenotype of the wild type plants grown on 1/2 MS (Murashige and Skoog) medium with DMSO (mock) (I), 100 nM epibrassinolide (eBL) (J), 30 μ M bikinin (Bik) (K), or 1 μ M brassinazole (Brz) (L). Right images are the outlined areas of left images with higher magnification. Arrows indicate ectopic root hair cells, and arrowheads indicate ectopic non-hair cells. Areas outlined with blue lines indicate the ectopic non-hair cells.

DOI: [10.7554/eLife.02525.003](https://doi.org/10.7554/eLife.02525.003)

The following source data and figure supplements are available for figure 1:

Source data 1. Root hair density, cell length, and relative hair number of the BR-related mutants and wild type plants treated with eBL, bikinin, Brz, or DMSO

DOI: [10.7554/eLife.02525.004](https://doi.org/10.7554/eLife.02525.004)

Figure supplement 1. Bikinin treatment inhibited H cell fate in *bin2-3 bil1 bil2* mutants.

DOI: [10.7554/eLife.02525.005](https://doi.org/10.7554/eLife.02525.005)

indicate that the BR-mediated root epidermal cell pattern formation largely relies on GSK3-like kinases and/or their upstream components.

BR signaling promotes N cell fate and inhibits H cell fate

GL2 has been widely used as a molecular maker of N cell fate determination (Masucci et al., 1996; Kuppusamy et al., 2009). In order to test whether the disordered root hair patterning in the BR-related mutants resulted from an altered root epidermal cell fate, we analyzed the *GL2* expression pattern in

these mutants using *PGL2::GUS* as a reporter. We found that in the wild type, 1.3% of epidermal cells in the N position lacked *GL2* expression in cross-sections (**Figure 2A,B**), and in the longitudinal view of root epidermis, the root epidermal cell files were arranged regularly, with *GL2*-expressing columns (N cell columns) interspaced with columns without *GL2* expression (H cell columns) (**Figure 2C**). However, about 15.3% of *bri1-116* and 18.8% of *det2-1* cells showed suppressed *PGL2::GUS* expression, and there were adjacent root epidermal cells without *GL2* expression in both *bri1-116* and *det2-1* (**Figure 2D–I**), which supports a previous finding with *bri1-116* and the Brz-treated wild type (**Kuppusamy et al., 2009**). These results indicated that adjacent root hairs in *bri1-116* and *det2-1* were caused by some N cell fate changing to H cell fate. In contrast, *GL2* was ectopically expressed in about 13.5% of H cells in *BRI1-OX* plants (**Figure 2J–L**) and in about 19.6% in *bin2-3 bil1 bil2* (**Figure 2M–O**), as compared with only 3.2% in the wild type (**Figure 2A–C**), indicating that lack of *GL2* expression in N cells may correspond to the ectopic root hairs observed in *bri1-116* and *det2-1*, and that the ectopically expressed *GL2* in H cells partially inhibits H cell fate in *BRI1-OX* and *bin2-3 bil1 bil2* plants. Taken together, the above results suggested that BR signaling has an important role in suppressing H cell fate and promoting N cell fate in both the N and the H positions, and BR signaling regulates root epidermal cell fate by controlling *GL2* expression through GSK3-like kinases, or their upstream components, but not through downstream transcription factors.

BR signaling acts upstream of WER and CPC to regulate root epidermal cell fate

It was known that *GL2* expression is directly regulated by the WER-GL3/EGL3-TTG1 but not the CPC-GL3/EGL3-TTG1 transcriptional complex (**Schieffelbein et al., 2009**). To explore whether the BR-regulated *GL2* expression and root epidermal cell fate determination are dependent on these complexes, we first created a set of double mutants of *cpc-1*, a mutant with fewer root hairs than its counterpart (**Figure 3A,B**), with *bri1-116* or *cpd* (**Figure 3C,D**). Similar to *cpc-1*, both double mutants *bri1-116 cpc-1* (**Figure 3E**) and *cpd cpc-1* (**Figure 3F**) produced few root hairs. We also generated double or multiple mutants of *wer-1*, a mutant with more root hairs than Col-0 (**Figure 3G,H**), with *BRI1-OX* or *bin2-3 bil1 bil2* (**Figure 3I,J**), and found that both *BRI1-OX wer-1* and *bin2-3 bil1 bil2 wer-1* (**Figure 3K,L**) were similar to *wer-1*, with many ectopic root hairs formed at the N cell position. These genetic analyses indicated that the WER-GL3/EGL3-TTG1 and CPC-GL3/EGL3-TTG1 transcriptional complexes act downstream of BR early signaling.

BIN2 phosphorylates EGL3 and TTG1

Therefore, we inferred that BR-mediated root epidermal cell fate may be dependent on GSK3-like kinases, the key negative components in the BR signaling pathway, acting upstream of the WER-EGL3/GL3-TTG1 or CPC-EGL3/GL3-TTG1 transcriptional complex. We then conducted yeast two-hybrid assays to test whether any components in the WER-EGL3/GL3-TTG1 or CPC-EGL3/GL3-TTG1 complex interact with BIN2, a well-studied GSK3-like kinase, and found that BIN2 can interact with EGL3 (**Figure 4A**) but not with CPC in yeast (**Figure 4—figure supplement 1**). However, due to strong auto-activation of WER and TTG1 fused with GAL4-DNA binding domain (DB) in yeast two-hybrid assays (**Figure 4—figure supplement 1**), we conducted GST pull-down and BiFC (biomolecular fluorescence complementation) assays to test their interactions, and found that BIN2 can interact with WER, EGL3, and TTG1 (**Figure 4B,C**). Furthermore, because BIN2 can regulate many transcription factors by phosphorylation (**Saidi et al., 2012**), and WER or CPC can interact with EGL3/GL3-TTG1 in vivo to form WER-EGL3/GL3-TTG1 or CPC-EGL3/GL3-TTG1 complexes, respectively (**Zhao et al., 2008; Song et al., 2011**), we then conducted in vitro kinase assays to test whether BIN2 can phosphorylate any of these components. We found that BIN2 did not phosphorylate WER and CPC, but was able to phosphorylate EGL3 and TTG1 (**Figure 5**).

GSK3-like kinase phosphorylates EGL3 to promote its movement to N cells

To investigate the biological relevance of EGL3 phosphorylation by a GSK3-like kinase BIN2, we used mass spectrometry and identified four potential phosphorylation sites (T209, T213, T399, and T403) of EGL3 by BIN2 (**Figure 5—figure supplement 1**), which are located in two regions that contain typical recognition sites of GSK3 kinases (**Cohen and Frame, 2001**). We then mutated threonine residues into alanine to make single- or double-site mutated forms of EGL3. In vitro phosphorylation assays showed that phosphorylation levels of EGL3^{T399A} and EGL3^{T209A/T213A} by BIN2 were significantly

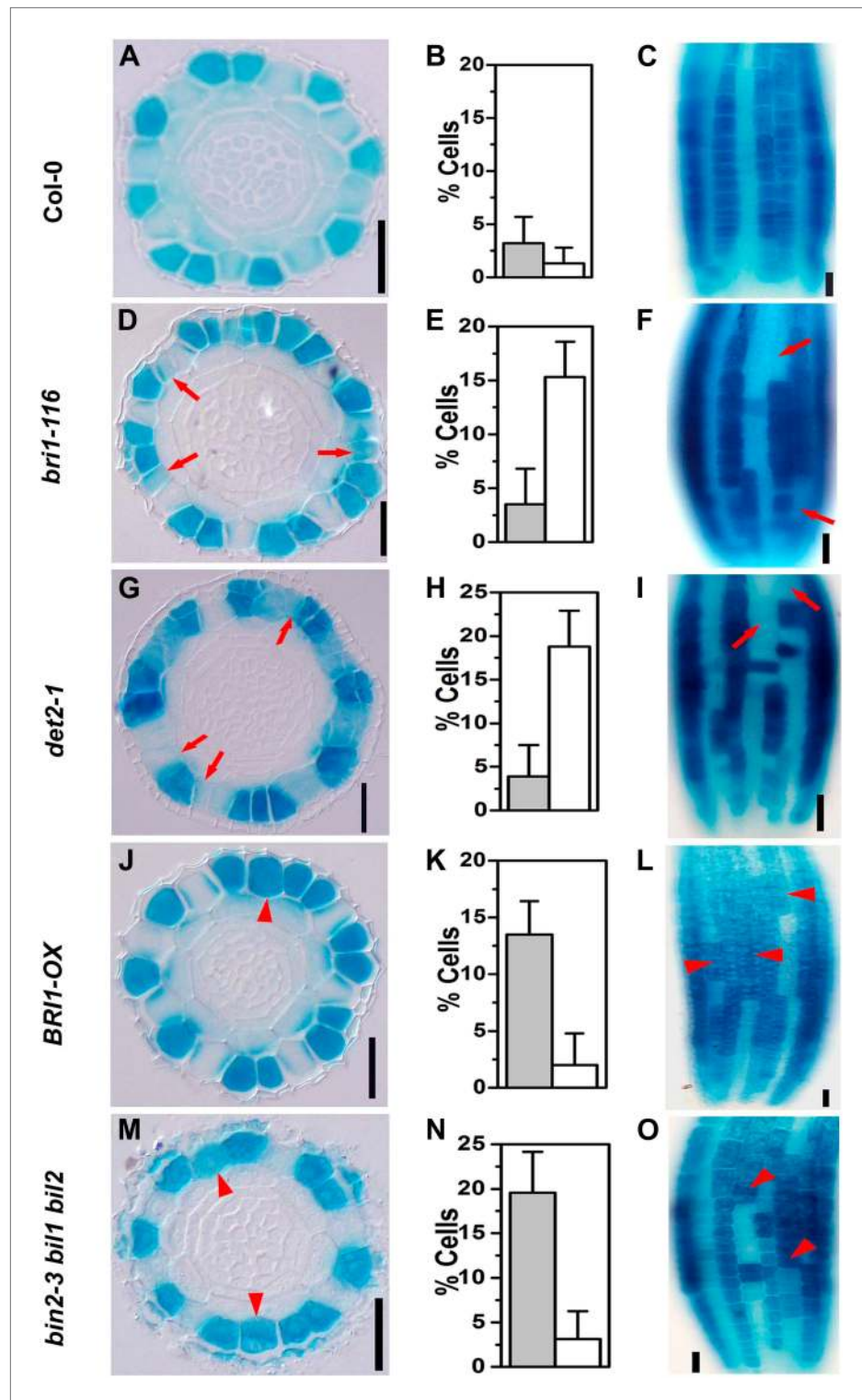


Figure 2. Expression pattern of *PGL2::GUS* is altered in the BR-related mutants. Transverse sections from root meristem of Col-0 (A), *bri1-116* (D), *det2-1* (G), *BRI1-OX* (J), and *bin2-3 bil1 bil2* (M). Frequency of cells without *PGL2::GUS* expression in the N cell position (open bars) and cells with ectopically expressed *PGL2::GUS* in the H cell position (solid bars) of Col-0 (B), *bri1-116* (E), *det2-1* (H), *BRI1-OX* (K), and *bin2-3 bil1 bil2* (N). Longitudinal images of the root epidermal cells in Col-0 (C), *bri1-116* (F), *det2-1* (I), *BRI1-OX* (L), and *bin2-3 bil1 bil2* (O). Scale bars, 25 μ m. Red arrows indicate N cells without *PGL2::GUS* expression, and red arrowheads indicate H cells ectopically expressing *PGL2::GUS*. For each genotype, n = 8. Error bars indicate standard deviation (SD).

DOI: [10.7554/eLife.02525.006](https://doi.org/10.7554/eLife.02525.006)

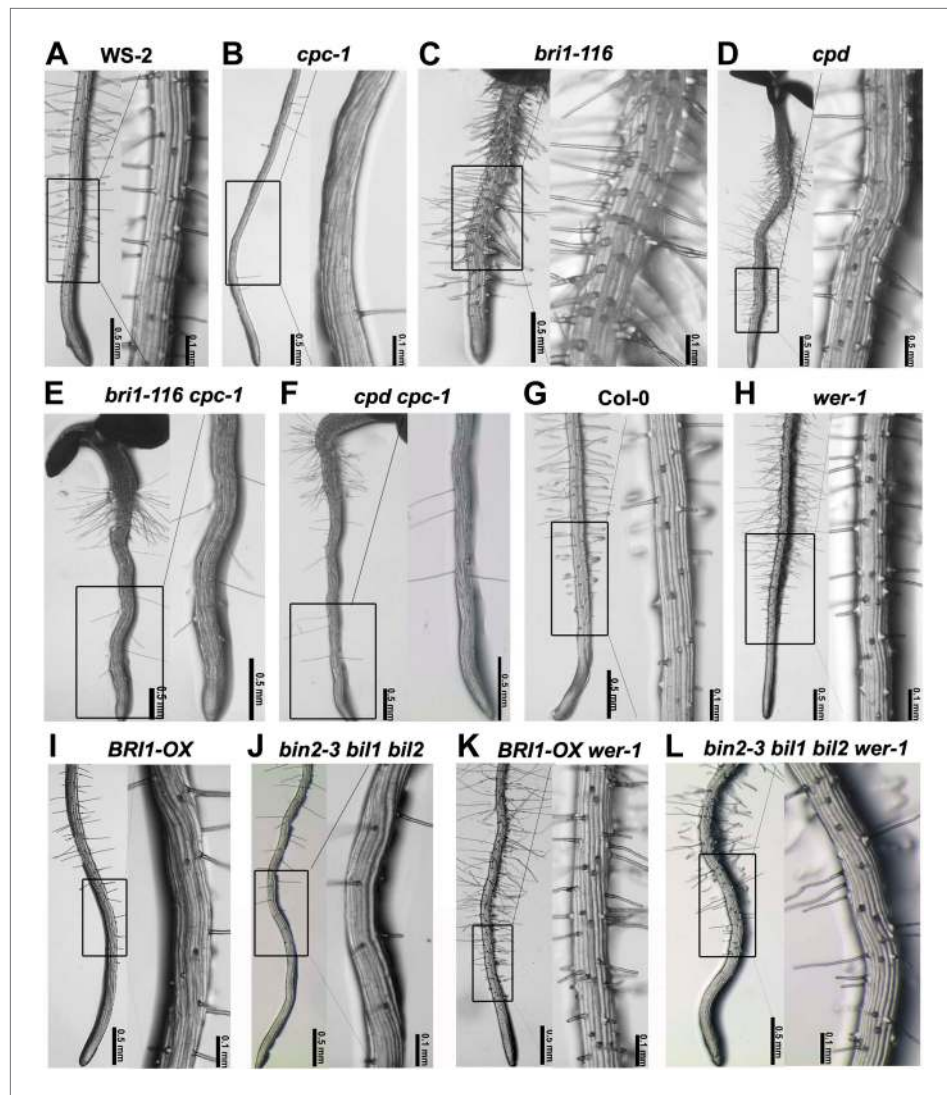


Figure 3. BR signaling acts upstream of CPC and WER to regulate root epidermal cell fate. Root hair phenotype of the wild type WS-2 (A) and double mutants of *cpc-1* (B) with *bri1-116* (C) or *cpd* (D), including *bri1-116 cpc-1* (E) and *cpd cpc-1* (F). Root hair phenotype of the wild type Col-0 (G) and the double/multiple mutants of *wer-1* (H) with *BRI1-OX* (I) or *bin2-3 bil1 bil2* (J), including *BRI1-OX wer-1* (K) and *bin2-3 bil1 bil2 wer-1* (L). *cpc-1* is in the WS-2 background. BR: brassinosteroid.

DOI: 10.7554/eLife.02525.007

reduced (Figure 6A), indicating that T399 and T209/T213 are the main phosphorylated residues. To investigate the biological function of EGL3 phosphorylation, we transformed EGL3 and its mutated forms driven by its own promoter into Col-0 to examine their subcellular localization. We found that the EGL3-GFP signal in N cells was apparently higher than that in H cells, and that it was mainly localized to cytosol in H cells, but to both cytosol and nucleus in N cells (Figure 6B). As EGL3 mRNA is only expressed in H cells (Bernhardt et al., 2005), this result indicated that, like its homologue GL3, EGL3 is also a mobile protein that moves from H cells to N cells. However, the EGL3^{T399A}-GFP and EGL3^{T209A/T213A}-GFP were solely localized to the nuclei of H cells (Figure 6C,D), indicating that EGL3 phosphorylation is required not only for its cytoplasmic accumulation, but also for its movement from H cells to N cells. In addition, we found that the root hair patterning of EGL3-GFP transgenic plants was not altered (Figure 6E; Table 1), indicating that correctly localized EGL3, mainly in N cell nuclei to promote N cell fate and less in H cell nuclei not to promote N cell fate, has no influence on root epidermal fate. Moreover, although root epidermal patterning in the EGL3^{T399A}-GFP transgenic plants was normal (Figure 6F; Table 1), the number of root hairs in EGL3^{T209A/T213A}-GFP plants was significantly

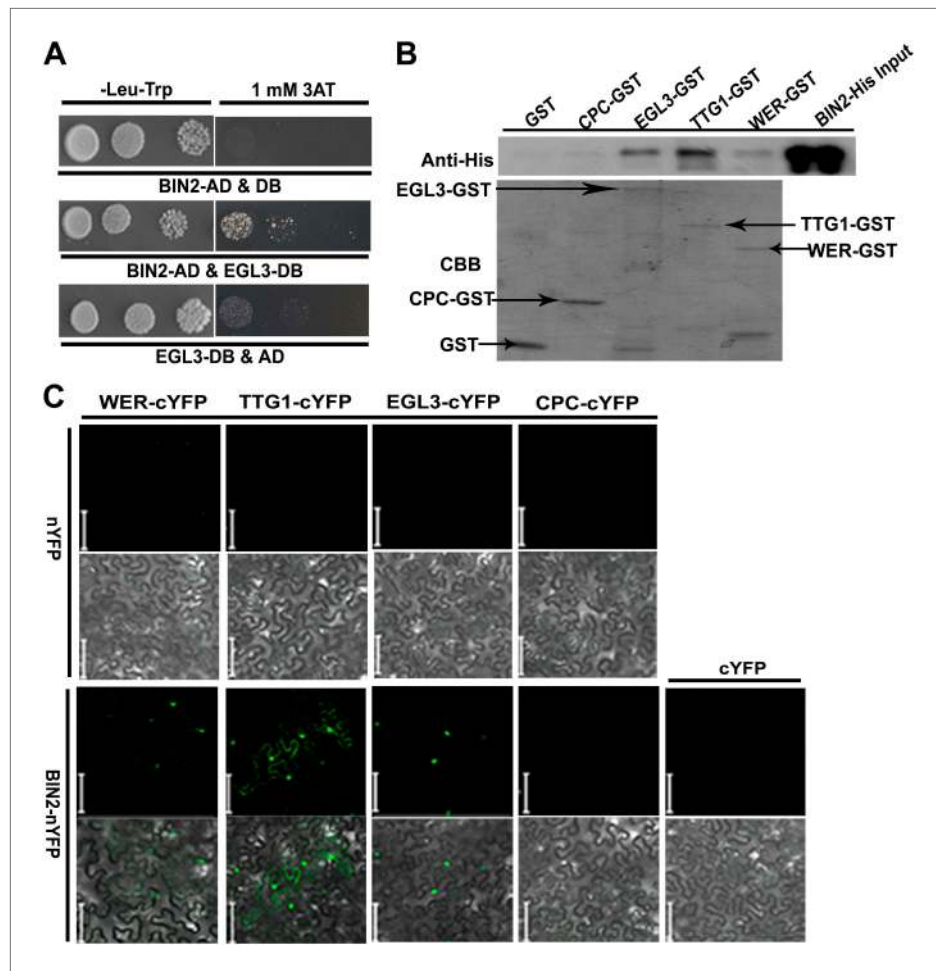


Figure 4. BIN2 interacts with EGL3, TTG1, and WER. **(A)** BIN2 interacts with EGL3 in yeast two-hybrid assays. **(B)** The interaction of BIN2-His with CPC-GST, EGL3-GST, TTG1-GST, and WER-GST in vitro. The BIN2-His pulled-down by CPC-GST, EGL3-GST, TTG1-GST, and WER-GST, or GST was detected by western blotting with anti-His antibody (top). The purified BIN2-His protein was used as inputs. An equal loading of recombinant proteins was indicated by Coomassie brilliant blue (CBB) staining (bottom). **(C)** BiFC assays of the interaction between BIN2 with EGL3, TTG1, and WER. Scale bars, 20 μm .

DOI: [10.7554/eLife.02525.008](https://doi.org/10.7554/eLife.02525.008)

The following figure supplements are available for figure 4:

Figure supplement 1. Yeast two-hybrid assays to test interactions of BIN2 with WER, TTG1, or CPC.

DOI: [10.7554/eLife.02525.009](https://doi.org/10.7554/eLife.02525.009)

reduced, likely due to a misspecification of non-hair cells in the H position (**Figure 6G**; **Table 1**), suggesting that the nucleus-localized EGL3 in H cells may determine N cell fate specification. Taken together, our results indicate that BIN2 phosphorylation on T399, T209, and/or T213 of EGL3 in H cells promotes EGL3 cytoplasmic localization, which likely helps its movement from H to N cells to regulate root epidermal cell fate.

GSK3-like kinases phosphorylate TTG1 to suppress the transcriptional activity of the WER-EGL3-TTG1 complex

TTG1 is required for a normal expression level of *GL2* (*Di Cristina et al., 1996*) but not for its expression pattern (*Hung et al., 1998*). Transgenic *TTG1-GFP* plants driven by its own promoter indicated that TTG1 was preferentially localized in the cytoplasm and slightly in the nucleus of both N and H cells (**Figure 7—figure supplement 1**), which was consistent with the subcellular localization of its petunia homologue AN11 (*De Vetten et al., 1997*). To understand the biological relevance of TTG1 phosphorylation by a GSK3-like kinase, we conducted transient transcription assays in *Nicotiana benthamiana*

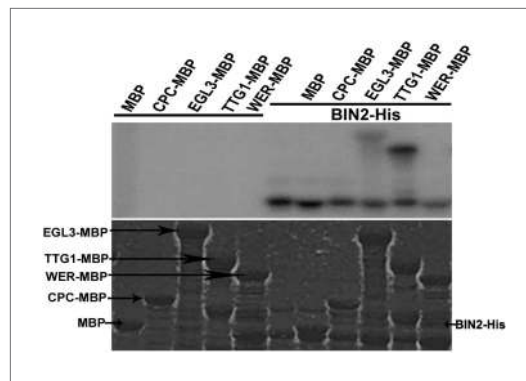


Figure 5. BIN2 phosphorylates EGL3 and TTG1, but not WER and CPC in vitro. An equal amount of recombinant BIN2 kinase indicated by Coomassie brilliant blue (CBB) staining (bottom) was incubated with recombinant MBP, WER-MBP, CPC-MBP, EGL3-MBP, or TTG1-MBP, separated by SDS-PAGE, and followed by autoradiography (top).

DOI: [10.7554/eLife.02525.010](https://doi.org/10.7554/eLife.02525.010)

The following figure supplements are available for figure 5:

Figure supplement 1. Mass spectrometry analysis of EGL3 phosphorylation sites.

DOI: [10.7554/eLife.02525.011](https://doi.org/10.7554/eLife.02525.011)

WER-EGL3-TTG1 complex (**Figure 7B**), indicating that TTG1 is mediating the negative effect of BIN2 on this transcriptional complex.

Discussion

BR signaling depends on GSK3-like kinases and the WER-EGL3-TTG1 complex to modulate root epidermal patterning

We provide several lines of evidence to strongly support an important role for BR signaling in directly regulating root hair cell fate. First, root hair patterning in the BR-biosynthetic and responsive mutants or in the wild type grown on eBL, bikinin, or BRZ, was dramatically altered, demonstrating that GSK3 kinases and/or their upstream components are mediating this regulation. Second, the expression pattern of the non-hair cell fate marker *PGL2::GUS* indicates that BR early signaling promotes N cell fate in the whole root epidermis, which is a reasonable explanation for the abnormal root hair patterning in the BR-related mutants: when BR signaling is enhanced, fewer root hairs are formed in the H position; when BR signaling is inhibited, more ectopic root hairs are produced in the N position. This finding supports the previous report that BRs positively regulate the expression of *WER* and *GL2* (**Kuppusamy et al., 2009**). Third, genetic analysis revealed that major components of the WER-EGL3-TTG1 or CPC-EGL3-TTG1 complex act downstream of BR signaling-mediated root epidermis patterning. Finally, BIN2 phosphorylation on EGL3 and TTG1 suggested that GSK3-like kinases directly regulate EGL3 movement and the transcription activity of the WER-EGL3-TTG1 complex to mediate root hair development.

Phosphorylation of EGL3 regulates its intercellular movement and controls root epidermal cell fate

This study revealed a key mechanism in the regulation of intercellular communication of transcription factors by a hormonal signal to determine epidermal cell specification. Non-cell autonomous movement of some transcriptional factors is an important mechanism to regulate some developmental processes (**Kurata et al., 2005b**), and cytoplasmic localization of these mobile proteins may be required for their intercellular movement. For example, in maize shoot apical meristem, a mutation in its potential nuclear localization signal (NLS) of *KNOTTED 1* abolished its intercellular movement

leaves to examine whether BIN2 regulated the complex's activity in a TTG1-dependent manner. We constructed a dual-luciferase reporter system using *PGL2::LUC* as a reporter gene and *35S::REN* as an internal control (**Figure 7A**). Because the protein of the gain-of-function mutation *bin2-1* (E263K) is more stable and has higher activity than the wild type BIN2 (**Peng et al., 2008**), and GSK3-like kinases are quite conserved among different species (**Saidi et al., 2012**), we used *bin2-1* to conduct this study. As shown in **Figure 7B**, transient expression of *WER* alone was able to slightly induce *PGL2::LUC* gene expression. In contrast, transient expression of *EGL3* alone was unable to induce reporter gene expression. Co-expression of *WER* and *EGL3* can dramatically promote *LUC* expression, which is consistent with a previous study (**Song et al., 2011**). Additional *bin2-1* did not alter the effect of *WER*, *EGL3*, or both *WER* and *EGL3* on *PGL2::LUC* expression. When *WER*, *EGL3*, and *TTG1* were used together, the expression of *PGL2::LUC* was further enhanced, indicating that *TTG1* can promote the activity of the *WER*-*EGL3* complex. Interestingly, additional *bin2-1* significantly inhibited reporter gene expression regulated by the

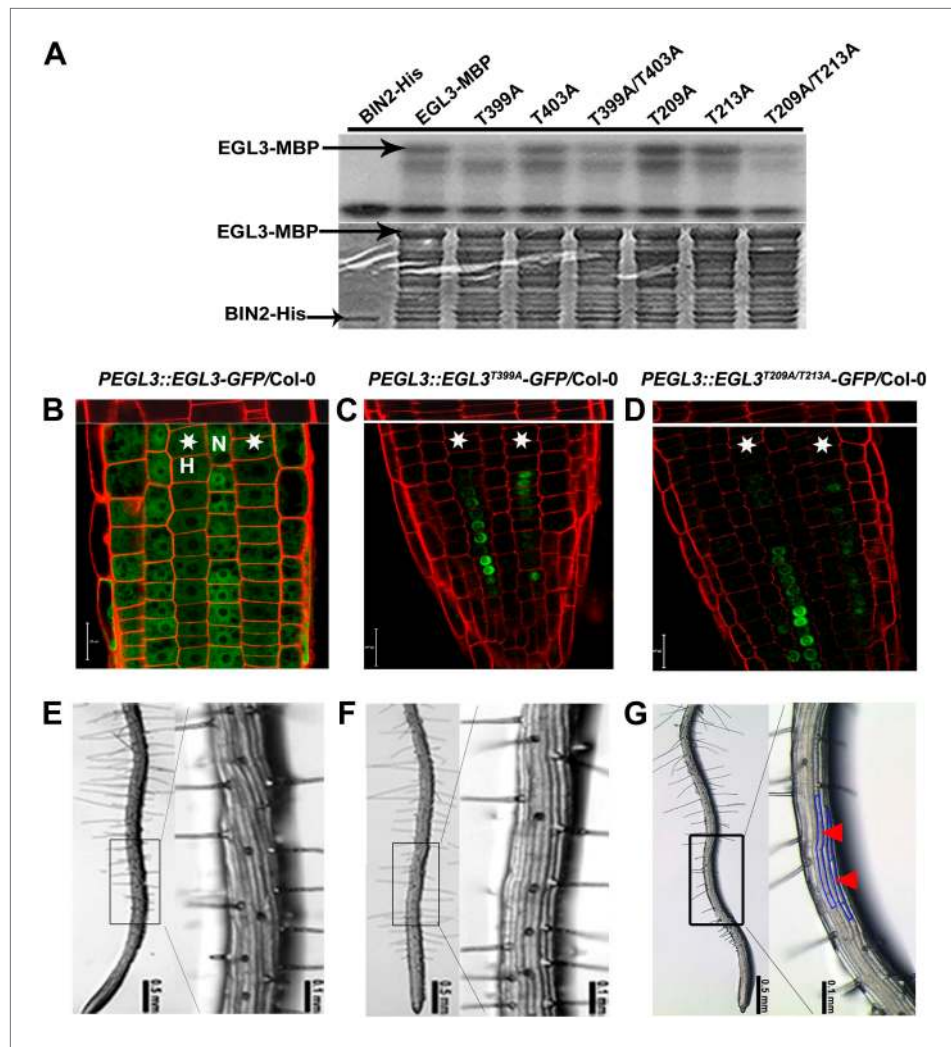


Figure 6. BIN2 phosphorylates EGL3 to regulate its subcellular localization and root epidermal cell fate. **(A)** BIN2 phosphorylates EGL3 on T399 and T209/T213. An equal amount of recombinant protein, as indicated by Coomassie brilliant blue (CBB) staining (bottom panel), was incubated in phosphorylation buffer, separated by SDS-PAGE, and followed by autoradiography (top panel). **(B)** EGL3-GFP is predominantly localized in N cell nuclei. Both EGL3^{T399A}-GFP **(C)** and EGL3^{T209A/T213A}-GFP **(D)** are solely localized in H cell nuclei. For **(B–D)**, the 5-day-old roots were stained with propidium iodide (red) for 10 s for visualizing the cell wall. The top panels show the underlying cortex. The stars indicate H cells. Scale bars, 20 μ m. **(E–G)** Root hair patterns of EGL3-GFP **(E)**, EGL3^{T399A}-GFP **(F)**, and EGL3^{T209A/T213A}-GFP **(G)** transgenic plants. Outlined areas in the left images are magnified in the right images. Red arrowheads and areas outlined with blue lines indicate ectopic non-root hair cells in the H position.

DOI: [10.7554/eLife.02525.012](https://doi.org/10.7554/eLife.02525.012)

The following source data and figure supplements are available for figure 6:

Source data 1. EGL3 amino acid sequence analysis

DOI: [10.7554/eLife.02525.013](https://doi.org/10.7554/eLife.02525.013)

Figure supplement 1. Alignment of EGL3 amino acid sequence with other bHLH homologues in *Arabidopsis*.

DOI: [10.7554/eLife.02525.014](https://doi.org/10.7554/eLife.02525.014)

(Vollbrecht et al., 1991; Lucas et al., 1995), and cytoplasmic localization of LEAFY (Schultz and Haughn, 1991), a transcriptional factor in floral identity, is also strongly correlated with its adjacent cell movement (Wu et al., 2003). Moreover, the movement of SHORT-ROOT, another mobile transcriptional factor in root radial patterning (Helariutta et al., 2000), was abolished when it was fused to a NLS, leading to its diminished cytoplasmic localization (Gallagher et al., 2004). However, it is largely unknown how the nuclear-cytoplasmic trafficking of transcription factors is regulated by internal cues to influence their intercellular movement. Although the intercellular movement of mobile factors in the

Table 1. The effect of EGL3 and its two mutated forms on root epidermal cell pattern formation

Genotype	Cells in the H position		Cells in the N position	
	Hair cells (%)	Non-hair cells (%)	Hair cells (%)	Non-hair cells (%)
Col-0	98.9 ± 3.3	1.1 ± 3.3	2.0 ± 4.5	98.0 ± 4.5
PEGL3::EGL3-GFP	95.8 ± 6.1	4.2 ± 6.1	1.9 ± 4.2	98.1 ± 4.2
PEGL3::EGL3 ^{T399A} -GFP	93.1 ± 6.4	6.9 ± 6.4	2.8 ± 6.0	97.2 ± 6.0
PEGL3::EGL3 ^{T209A/T213A} -GFP	85.5 ± 4.4*	14.5 ± 4.4*	1.0 ± 3.2	99.0 ± 3.2

At least 10 different 5-day-old roots were examined for each strain. Values represent means ± SD. For statistical analysis, the F test was used to determine the variance, and the two-tailed t test with equal variance or unequal variance was used to determine the significance level of the difference among the transgenic plants.

* $p < 0.05$.

DOI: 10.7554/eLife.02525.015

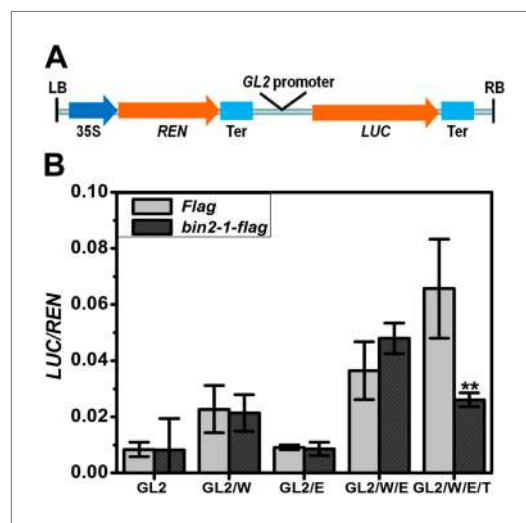


Figure 7. BIN2 inhibits the transcription activity of the WER-EGL3-TTG1 complex through TTG1. (A) Schematic diagram of the dual-luciferase reporter construct. The firefly luciferase (LUC) reporter gene was driven by GL2 promoter. The Renillia luciferase (REN) reporter gene was controlled by Cauliflower mosaic virus promoter (35S) and terminator (Ter). (B) bin2-1 inhibits PGL2::LUC expression only when TTG1 is co-expressed with WER and EGL3. Relative reporter activity in *Nicotiana benthamiana* leaf cells transiently transformed with the indicated effector, reporter, and regulatory constructs. G, W, E, and T indicate GL2, WER, EGL3, and TTG1, respectively. Error bars indicate SD. ** $p < 0.01$ determined by the two-tailed Student's t test.

DOI: 10.7554/eLife.02525.016

The following figure supplements are available for figure 7:

Figure supplement 1. Subcellular localization of TTG1-GFP in Col-0 root epidermal cells.

DOI: 10.7554/eLife.02525.017

WER/CPC-EGL3/GL3-TTG1 complex may determine root epidermis patterning (Savage et al., 2008), it is not clear how their movement is regulated by internal cues. Because EGL3 mRNA was expressed only in H cells, the major nuclear localization of EGL3 protein in N cells indicated that, like GL3 and CPC, EGL3 can also move from H to N cells. In addition, we found that EGL3^{T399A} and EGL3^{T209A/T213A} with abolished phosphorylation sites were solely localized in the nucleus of H cells, indicating that the unphosphorylated EGL3 may not move between cells. Although we do not know how T209/T213 phosphorylation regulated EGL3 subcellular localization, the T399 phosphorylation likely affected a NLS, because T399 is located in a predicted non-canonical NLS (Figure 6—source data 1), which is found in many other plant bHLHs (Galstyan et al., 2012). Although the EGL3^{T399A}-GFP plants showed normal root hair patterning, this can be explained by the close proximity of T399 to its bHLH domain, which may have affected its interaction with TTG1 and MYB (Zhang et al., 2003), leading to the nucleus-localized EGL3^{T399A} in H cells unable to induce GL2 expression. However, EGL3^{T209A/T213A}-GFP may still interact with TTG1 and have DNA-binding activity, because T209/T213 was located in the N-terminal region far away from the bHLH domain (Figure 6—figure supplement 1), which led to GL2 expression in H cells and EGL3^{T209A/T213A}-GFP plants growing fewer root hairs (Figure 6G; Table 1).

GSK3-like kinases inhibit GL2 expression through TTG1, which is required for the transcriptional activity of WER-EGL3-TTG1 complex

Besides the regulation of EGL3 nuclear-cytoplasmic trafficking, we also provided strong evidence to support GSK3-like kinases' inhibition on the transcriptional activity of the WER-EGL3-TTG1 complex through TTG1. In *Nicotiana benthamiana*

pavement cells, it was demonstrated that BIN2 has a negative role in WER-EGL3-TTG1 transcriptional activity, but has no effect on the activity of WER, EGL3, or both WER and EGL3. Although BIN2 phosphorylates EGL3, its failure to regulate WER-EGL3 transcriptional activity can be explained by a possible ubiquitous expression of WER, EGL3, and GSK3-like kinases in *Nicotiana benthamiana* leaves. Furthermore, it was reported that TTG1 interacts with EGL3 (Zhang et al., 2003), and TTG1 is necessary for the full functioning of other bHLH partners, such as GL3 and TRANSPARENT TESTA8 (Baudry et al., 2004; Zhao et al., 2008). Therefore, it is very likely that TTG1 phosphorylation by GSK3-like kinases may affect its regulation of EGL3 and the activity of the WER-EGL3-TTG1 complex.

BR signaling promotes the N cell fate of root epidermis

Our data also support the suggestion that the N cell is a default cell type in root epidermis, and that H cell fate is produced due to inhibition of N cell fate by internal or external cues. First, we observed that WER, a positive regulator for GL2 expression, is expressed in both N and H cells in the early root meristem (Figure 8—figure supplement 1), which is also supported by a previous report that WER exhibits uniform promoter activity in both N and H cells proximal to the initial cells (Savage et al., 2008). Second, in *Arabidopsis*, both EGL3 and GL3 are expressed in H cells, but their proteins move to adjacent cells to promote N cell fate (Bernhardt et al., 2005). If they stay in H cells with the ability to interact with WER and TTG1, the H cells may develop into N cells as shown in the *EGL3^{3T209A/T213A}-GFP* transgenic plants. Moreover, over-expression of GL3 and EGL3 promoted non-hair cell fate (Bernhardt et al., 2003). Third, TTG1 is localized in both N and H cells, and TTG1 and EGL3 may synergistically promote WER-EGL3 transcriptional activity and enhance N cell fate. Apparently, BR signaling can promote N cell fate in several ways. Besides the inhibition of BR signaling on EGL3 cell-cell movement and the promotion of TTG1 activity, BR signaling also promotes WER expression as WER up-regulation in *bin2-3 bil1 bil2* (Figure 8—figure supplement 1), which is consistent with the positive role of BRs in WER expression (Kuppusamy et al., 2009).

Thus, we proposed a model to illustrate how BR signaling regulates WER-EGL3-TTG1 complex formation and activity to control root epidermal cell fate. As shown in Figure 8, without BRs, WER-GL3/EGL3-TTG1 complex formation and activity is inhibited in both N and H cells, as WER expression is reduced in both H and N cells, and the activated GSK3-like kinases phosphorylate EGL3 in H cells to promote its cytoplasmic localization in both H cells and N cells, both of which lead to less WER-GL3/EGL3-TTG1 complex formation in nuclei and suppression of GL2 expression. The activity of some formed WER-bHLH-TTG1 complexes may be further inhibited by GSK3-like kinases phosphorylating TTG1. In contrast, enhanced BR early signaling inhibits GSK3-like kinases, leading to nuclear accumulation of the unphosphorylated EGL3 in H cells and normal function of unphosphorylated TTG1 in both cell types. Although CPC can move into H cells, due to enhanced WER expression and more efficient interaction of EGL3 with WER than with CPC (Song et al., 2011), more WER-EGL3-TTG1 complex is formed in H cells to promote GL2 expression and determine N cell fate. In N cells, the nucleus-localized GL3 can interact with WER and TTG1 to promote GL2 expression and maintain N cell fate. However, it remains to be investigated how TTG1 and WER expression is regulated by BR signaling. It is also not clear how BR signaling coordinates with positional signals and other phytohormones to regulate root hair patterning.

Materials and methods

Plant materials and growth conditions

The seeds of the *wer-1* and *PGL2::GUS* lines were obtained from Dr John Schiefelbein (University of Michigan), the *bin2-3 bil1 bil2* seeds were obtained from Jianming Li (University of Michigan), and the *cpc-1* seeds (CS6399) were obtained from the Arabidopsis Biological Resource Center (Ohio State University). Combinations of the BR-related mutants with the root hair mutants or the *PGL2::GUS* line were generated by crossing and selected by GUS staining based on the mutant phenotype, or antibiotic selection marker analysis. For root hair observation, seeds were grown on 1/2 MS medium (pH 5.8) with 1% sucrose, chilled for 3 d at 4°C, and grown for 5 d at 23°C under long-day conditions (16 hr light/8 hr dark). *Nicotiana benthamiana* plants were grown at 28°C under long-day conditions (16 hr light/8 hr dark).

Construction of double or multiple mutants

The double mutants or multiple mutants were derived from genetic crosses of the parental mutants (or transgenic lines). For generation of the *BRI1-OX wer-1* and *bil1 bil2 bil3 wer-1* double/multiple mutants, the *wer-1* was genotyped with its point mutation-derived cleaved amplified polymorphic

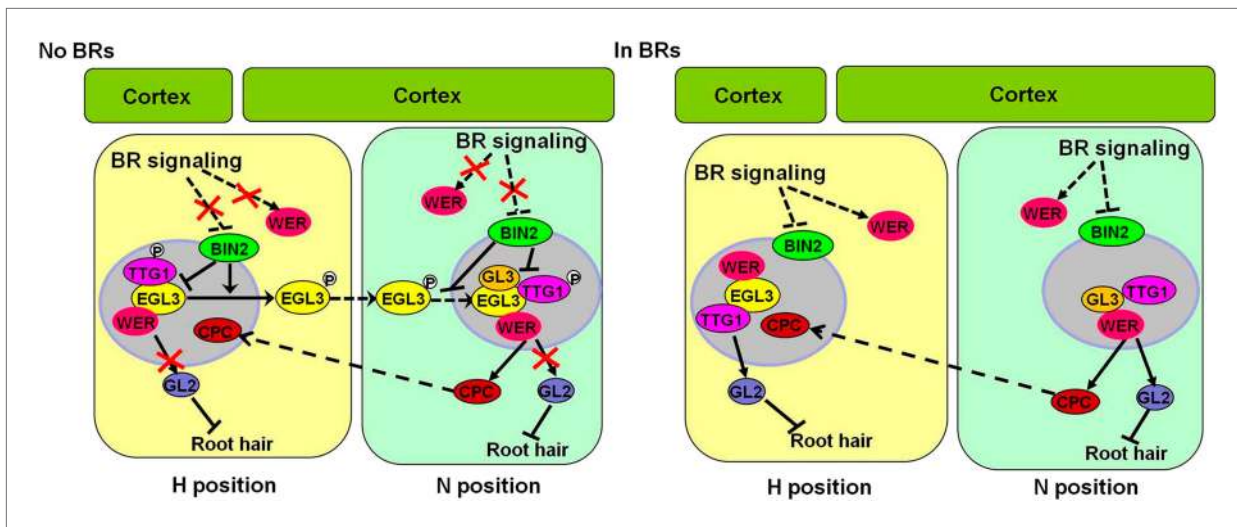


Figure 8. A proposed model to illustrate how BR signaling regulates root epidermal cell fate. Without BR early signaling, *WER* expression is reduced, and the activated GSK3-like kinases phosphorylate *EGL3* and *TTG1* in both H and N cells, leading to reduced formation and/or activity of the *WER-GL3/* *GL3-TTG1* complex, which inhibits *GL2* expression in some N cells. With enhanced BR early signaling, *WER* expression is enhanced in both H and N cells, and the GSK3-like kinases activity is inhibited, leading to reduced phosphorylation of *EGL3* and *TTG1* in both cell types. Thus, *WER-GL3-TTG1* and *WER-GL3-TTG1* complexes with transcriptional activity are formed in H and N cells, respectively, to promote *GL2* expression and non-root hair cell fate. BR: brassinosteroid.

DOI: 10.7554/eLife.02525.018

The following figure supplements are available for figure 8:

Figure supplement 1. *WER* expression pattern in the root early meristem and its expression level in the *bin2-3 bil1 bil2* and wild type Col-0.

DOI: 10.7554/eLife.02525.019

sequence (CAPS) marker (*Lee and Schiefelbein, 1999*) (**Supplementary file 1**), the *bin2-3 bil1 bil2* was genotyped as described (*Yan et al., 2009*), and the *BRI1-OX* was selected by the antibiotic selection markers. For generation of *bri1-116 cpc-1* and *cpd cpc-1* double mutants, the *cpc-1* was identified by PCR and phenotypic analysis, and the *bri1-116* and the *cpd* were isolated by phenotype.

Plasmid construction and recombinant protein purification

For GST pull-down assays, *BIN2* was cloned into the *PET28a* vector, and *EGL3* was cloned into the *pGEX4T-1* vector. For in vitro kinase assays, *WER*, *TTG1*, *CPC*, and *EGL3* were cloned into the *pMAL-C2X* vector. His-fused *BIN2* (*BIN2-His*), GST-fused *EGL3* (*EGL3-GST*), and MBP-fused *WER* (*WER-MBP*), *TTG1* (*TTG1-MBP*), *CPC* (*CPC-MBP*), and *EGL3* (*EGL3-MBP*) were expressed in BL21 (DE3) pLySs strain and purified with either Ni-NTA agarose (Clontech, Mountain View, CA), glutathione resin (Genescript, Piscataway, NJ), or amylose resin (NEB, Ipswich, MA), respectively.

Plant transformation and selection of transgenic plants

To generate plants expressing *GFP*-tagged *EGL3* or mutated *EGL3^{T399A}* and *EGL3^{T209A/T213A}*, the various *EGL3* cDNAs were cloned in-frame with *GFP* into the *pCAMBIA2302* vector and driven by the *EGL3* promoter (2 kb upstream of the start codon), and the plants expressing *GFP*-tagged *TTG1* were generated by cloning the *TTG1* cDNA in-frame with *GFP* into the *pCAMBIA2302* vector and driven by the *TTG1* promoter (2 kb upstream of the start codon). The constructs were transformed into *Agrobacterium tumefaciens* GV3101 strains. All transgenic plants were generated by floral dip transformation. T_0 seeds were harvested and screened by germinating on MS solid medium with antibiotic selection. For each transformation, at least five individual T_1 transgenic lines were selected. Transgenic lines of *PEGL3::EGL3-GFP*, *PEGL3::EGL3^{T399A}-GFP*, *PEGL3::EGL3^{T209A/T213A}-GFP*, and *PTTG1::TTG1-GFP* with T_2 or higher generations were used for further analysis.

Microscopy and histochemical analysis

The root hair pattern of the 5-day-old seedlings was observed, and images at $\times 100$ magnification were taken with a Leica MZ FLIII stereomicroscope (Leica Microsystems). The root hair density was counted as described (*Galway et al., 1994; Jones et al., 2002*) with some modifications. Any visible protrusion

from the epidermal cell was regarded as a root hair, regardless of length. The number of root hairs was counted from one side of a 1 mm segment from the imitated differentiation region of the 5-day-old roots, and at least eight roots were measured for each stain. The hair cell length was measured along the longitudinal plane at $\times 100$ magnification using the software Scion Image, and at least 10 root hair cells were measured for each root. The relative root number was calculated as root hair density \times root hair cell length for each root as described (Wada *et al.*, 1997).

The histochemical staining of 5-day-old roots harboring the *GUS* reporter was performed as described (Masucci *et al.*, 1996). Transverse sections of root meristem were prepared as described (Ye *et al.*, 2010) with modifications. The proportion of cells expressing or not expressing *PGL2::GUS* reporter in H cells or N cells was measured by examining sections at least from eight seedlings in each strain. For protein localization of EGL3-GFP and its mutated forms, the 5-day-old transgenic plants were examined by confocal microscope (Zeiss) after staining with 5 $\mu\text{g}/\text{ml}$ propidium iodide (PI) (Sigma, St. Louis, MO) for 10 s at room temperature, and images were captured at 489 nm and 538 nm laser excitation and at 509 nm and 617 nm emission for GFP and PI staining. The pattern of epidermal cell types was determined as described (Lee and Schiefelbein, 2002).

Yeast two-hybrid assay

For yeast two-hybrid assays, the full length cDNA of *BIN2* was cloned into vector *pEXP-AD502* (*BIN2-AD*) and used as a prey, and the full length cDNAs of *EGL3*, *WER*, *TTG1*, and *CPC* were cloned into the *pDBLeu* vector (*EGL3-DB*, *WER-DB*, *TTG1-DB*, *CPC-DB*), respectively, and used as a bait. The prey and bait plasmids were transformed into the yeast strains *AH109* and *Y187*, respectively. After yeast mating, the protein–protein interactions were tested on SD medium minus Leu, Trp, and His, and containing 2 mM 3-amino-1, 2, 4-triazole (3AT) (Sigma, St. Louis, MO).

Transient expression assays in *Nicotiana benthamiana* leaves

To generate the vector system for BiFC analysis, the full length cDNAs of *EGL3*, *WER*, *TTG1*, and *CPC* were cloned into the *pXY104* vector (*cYFP*), respectively, to generate *EGL3-cYFP*, *WER-cYFP*, *TTG1-cYFP*, and *CPC-cYFP* constructs, and *BIN2* cDNA was cloned into the *pXY106* (*nYFP*) vector to generate *BIN2-nYFP* construct. For transient expression, *Agrobacterium* strains (*GV3101*) carrying the constructs for testing the specific interaction were transformed into 4–5-week-old *Nicotiana benthamiana* leaves as described previously (Walter *et al.*, 2004). After infiltration for 4 d, the lower leaf epidermis cells were used for analyzing the fluorescence signal by confocal microscopy (Zeiss).

For dual-luciferase assays, cDNAs of the effectors *WER*, *EGL3*, and *TTG1*, and the regulator *bin2-1* were cloned with *Flag* tag into *pCAMBIA2302* driven by a 35S promoter. *GL2* promoter (2 kb upstream of the start codon) was cloned into the *pGreenII 0800-LUC* vector to be used as the reporter. The method of transient expression used was as previously described (Hellens *et al.*, 2005).

GST pull-down assays

The purified proteins, *EGL3-GST*, *WER-GST*, *TTG1-GST*, *CPC-GST*, and *GST*, were bound with 25 μl GST resin in binding buffer (10 mM phosphate buffer, pH 7.4, 140 mM NaCl, 3 mM KCl, 0.1% Triton X-100) for 2 hr at 4°C. After washing three times with the binding buffer, an equal amount of *BIN2-His* was added and rebound for 2 hr at 4°C. After boiling in SDS loading buffer for 5 min, the pull-down proteins were separated on 10% SDS–PAGE gels and detected by immunoblotting with anti-His antibody (Abmart, Shanghai, China).

In vitro kinase assays and phosphorylation site identification

In vitro kinase assays were performed in 24 μl reaction buffer (20 mM Tris, pH 7.5, 10 mM MgCl_2 , 5 mM DTT) containing 20 μM ATP and 1 μl of 10 μCi [^{32}P] γATP (PerkinElmer, Waltham, Massachusetts) and purified proteins. The reaction was carried out at 30°C for 1 hr and terminated by adding 6 μl of 5 \times SDS loading buffer. After boiling for 3 min, proteins were separated on 10% SDS–PAGE. Gels were stained with Coomassie brilliant blue, and then dried and autoradiographed. For phosphorylation site identification, in vitro kinase assays were performed, and protein bands were excised to be used for mass spectrometry analysis.

Site-directed mutagenesis of *EGL3*

The mutated forms of *EGL3* were generated by a PCR-based site-directed mutagenesis (Supplementary file 1).

Acknowledgements

We thank John Schiefelbein and Jianming Li (University of Michigan) for providing seeds, Ramin Yadegari (University of Arizona) and Hongquan Yang (Shanghai Jiao Tong University) for reviewing drafts of our manuscript, Hexige Saiyin and Donghong Chen (Fudan University) for providing advice on paraffin sections, Youheng Wei (Fudan University) for technical advice on confocal images, and Jianjun Jiang (Fudan University) for helpful discussions on the function of nuclear localization signals.

Additional information

Funding

Funder	Grant reference number	Author
National Basic Research Program of China	2012CB114300	Xuelu Wang
National Natural Science Foundation of China	91317302	Xuelu Wang
National Natural Science Foundation of China	30925020	Xuelu Wang
Shanghai Science and Technology Committee	10JC1400800	Xuelu Wang

The funder had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

Author contributions

YC, Conception and design, Acquisition of data, Analysis and interpretation of data, Drafting or revising the article; WZ, YC, Acquisition of data, Analysis and interpretation of data; SI, TA, Contributed unpublished essential data or reagents; XW, Conception and design, Analysis and interpretation of data, Drafting or revising the article

Additional files

Supplementary file

- Supplementary file 1. List of primers used in this study.

DOI: [10.7554/eLife.02525.020](https://doi.org/10.7554/eLife.02525.020)

References

- Baudry A**, Heim MA, Dubreucq B, Caboche M, Weissshaar B, Lepiniec L. 2004. TT2, TT8, and TTG1 synergistically specify the expression of *BANYULS* and proanthocyanidin biosynthesis in *Arabidopsis thaliana*. *The Plant Journal for Cell and Molecular Biology* **39**:366–380. doi: [10.1111/j.1365-313X.2004.02138.x](https://doi.org/10.1111/j.1365-313X.2004.02138.x).
- Berger F**, Hung CY, Dolan L, Schiefelbein J. 1998. Control of cell division in the root epidermis of *Arabidopsis thaliana*. *Developmental Biology* **194**:235–245. doi: [10.1006/dbio.1997.8813](https://doi.org/10.1006/dbio.1997.8813).
- Bernhardt C**, Lee MM, Gonzalez A, Zhang F, Lloyd A, Schiefelbein J. 2003. The bHLH genes *GLABRA3* (*GL3*) and *ENHANCER OF GLABRA3* (*EGL3*) specify epidermal cell fate in the *Arabidopsis* root. *Development (Cambridge, England)* **130**:6431–6439. doi: [10.1242/dev.00880](https://doi.org/10.1242/dev.00880).
- Bernhardt C**, Zhao M, Gonzalez A, Lloyd A, Schiefelbein J. 2005. The bHLH genes *GL3* and *EGL3* participate in an intercellular regulatory circuit that controls cell patterning in the *Arabidopsis* root epidermis. *Development (Cambridge, England)* **132**:291–298. doi: [10.1242/dev.01565](https://doi.org/10.1242/dev.01565).
- Cohen P**, Frame S. 2001. The renaissance of GSK3. *Nature Reviews Molecular Cell Biology* **2**:769–776. doi: [10.1038/35096075](https://doi.org/10.1038/35096075).
- De Rybel B**, Audenaert D, Vert G, Rozhon W, Mayerhofer J, Peelman F, Coutuer S, Denayer T, Jansen L, Nguyen L, Vanhoutte I, Beeckman T, Vleminckx K, Jonak C, Chory J, Inzé D, Russinova E, Beeckman T. 2009. Chemical inhibition of a subset of *Arabidopsis thaliana* GSK3-like kinases activates brassinosteroid signaling. *Chemistry & Biology* **16**:594–604. doi: [10.1016/j.chembiol.2009.04.008](https://doi.org/10.1016/j.chembiol.2009.04.008).
- De Vetten N**, Quattrocchio F, Mol J, Koes R. 1997. The an11 locus controlling flower pigmentation in petunia encodes a novel WD-repeat protein conserved in yeast, plants, and animals. *Genes & Development* **11**:1422–1434. doi: [10.1101/gad.11.11.1422](https://doi.org/10.1101/gad.11.11.1422).

- Di Cristina M**, Sessa G, Dolan L, Linstead P, Baima S, Ruberti I, Morelli G. 1996. The *Arabidopsis* Athb-10 (GLABRA2) is an HD-Zip protein required for regulation of root hair development. *The Plant Journal: for Cell and Molecular Biology* **10**:393–402. doi: [10.1046/j.1365-313X.1996.10030393.x](https://doi.org/10.1046/j.1365-313X.1996.10030393.x).
- Dolan L**, Duckett CM, Grierson C, Linstead P, Schneider K, Lawson E, Dean C, Poethig S, Roberts K. 1994. Clonal relationships and cell patterning in the root epidermis of *Arabidopsis*. *Development* **120**:2465–2474. doi: [10.1006/dbio.1997.8813](https://doi.org/10.1006/dbio.1997.8813).
- Gallagher KL**, Paquette AJ, Nakajima K, Benfey PN. 2004. Mechanisms regulating SHORT-ROOT intercellular movement. *Current Biology: CB* **14**:1847–1851. doi: [10.1016/j.cub.2004.09.081](https://doi.org/10.1016/j.cub.2004.09.081).
- Galstyan A**, Bou-Torrent J, Roig-Villanova I, Martínez-García JF. 2012. A dual mechanism controls nuclear localization in the atypical basic-helix-loop-helix protein PAR1 of *Arabidopsis thaliana*. *Molecular Plant* **5**:669–677. doi: [10.1093/mp/sss006](https://doi.org/10.1093/mp/sss006).
- Galway ME**, Masucci JD, Lloyd AM, Walbot V, Davis RW, Schiefelbein JW. 1994. The *TTG* gene is required to specify epidermal cell fate and cell patterning in the *Arabidopsis* root. *Developmental Biology* **166**:740–754. doi: [10.1006/dbio.1994.1352](https://doi.org/10.1006/dbio.1994.1352).
- Hacham Y**, Holland N, Butterfield C, Ubeda-Tomas S, Bennett MJ, Chory J, Savaldi-Goldstein S. 2011. Brassinosteroid perception in the epidermis controls root meristem size. *Development (Cambridge, England)* **138**:839–848. doi: [10.1242/dev.061804](https://doi.org/10.1242/dev.061804).
- Helariutta Y**, Fukaki H, Wysocka-Diller J, Nakajima K, Jung J, Sena G, Hauser MT, Benfey PN. 2000. The *SHORT-ROOT* gene controls radial patterning of the *Arabidopsis* root through radial signaling. *Cell* **101**:555–567. doi: [10.1016/S0092-8674\(00\)80865-X](https://doi.org/10.1016/S0092-8674(00)80865-X).
- Hellens RP**, Allan AC, Friel EN, Bolitho K, Grafton K, Templeton MD, Karunairetnam S, Gleave AP, Laing WA. 2005. Transient expression vectors for functional genomics, quantification of promoter activity and RNA silencing in plants. *Plant Methods* **1**:13. doi: [10.1186/1746-4811-1-13](https://doi.org/10.1186/1746-4811-1-13).
- Hothorn M**, Belkhadir Y, Dreux M, Dabi T, Noel JP, Wilson IA, Chory J. 2011. Structural basis of steroid hormone perception by the receptor kinase BRI1. *Nature* **474**:467–471. doi: [10.1038/nature10153](https://doi.org/10.1038/nature10153).
- Hung CY**, Lin Y, Zhang M, Pollock S, Marks MD, Schiefelbein J. 1998. A common position-dependent mechanism controls cell-type patterning and *GLABRA2* regulation in the root and hypocotyl epidermis of *Arabidopsis*. *Plant Physiology* **117**:73–84. doi: [10.1104/pp.117.1.73](https://doi.org/10.1104/pp.117.1.73).
- Ishida T**, Kurata T, Okada K, Wada T. 2008. A genetic regulatory network in the development of trichomes and root hairs. *Annual Review of Plant Biology* **59**:365–386. doi: [10.1146/annurev.arplant.59.032607.092949](https://doi.org/10.1146/annurev.arplant.59.032607.092949).
- Jones MA**, Shen JJ, Fu Y, Li H, Yang Z, Grierson CS. 2002. The *Arabidopsis* Rop2 GTPase is a positive regulator of both root hair initiation and tip growth. *The Plant Cell* **14**:763–776. doi: [10.1105/tpc.010359](https://doi.org/10.1105/tpc.010359).
- Kuppusamy KT**, Chen AY, Nemhauser JL. 2009. Steroids are required for epidermal cell fate establishment in *Arabidopsis* roots. *Proceedings of the National Academy of Sciences of the United States of America* **106**:8073–8076. doi: [10.1073/pnas.0811633106](https://doi.org/10.1073/pnas.0811633106).
- Kurata T**, Ishida T, Kawabata-Awai C, Noguchi M, Hattori S, Sano R, Nagasaka R, Tominaga R, Koshino-Kimura Y, Kato T, Sato S, Tabata S, Okada K, Wada T. 2005a. Cell-to-cell movement of the CAPRICE protein in *Arabidopsis* root epidermal cell differentiation. *Development (Cambridge, England)* **132**:5387–5398. doi: [10.1242/dev.02139](https://doi.org/10.1242/dev.02139).
- Kurata T**, Okada K, Wada T. 2005b. Intercellular movement of transcription factors. *Current Opinion in Plant Biology* **8**:600–605. doi: [10.1016/j.pbi.2005.09.005](https://doi.org/10.1016/j.pbi.2005.09.005).
- Kwak SH**, Shen R, Schiefelbein J. 2005. Positional signaling mediated by a receptor-like kinase in *Arabidopsis*. *Science* **307**:1111–1113. doi: [10.1126/science.1105373](https://doi.org/10.1126/science.1105373).
- Lee MM**, Schiefelbein J. 1999. WEREWOLF, a MYB-related protein in *Arabidopsis*, is a position-dependent regulator of epidermal cell patterning. *Cell* **99**:473–483. doi: [10.1016/S0092-8674\(00\)81536-6](https://doi.org/10.1016/S0092-8674(00)81536-6).
- Lee MM**, Schiefelbein J. 2002. Cell pattern in the *Arabidopsis* root epidermis determined by lateral inhibition with feedback. *The Plant Cell* **14**:611–618. doi: [10.1105/tpc.010434](https://doi.org/10.1105/tpc.010434).
- Li J**, Chory J. 1997. A putative leucine-rich repeat receptor kinase involved in brassinosteroid signal transduction. *Cell* **90**:929–938. doi: [10.1016/S0092-8674\(00\)80357-8](https://doi.org/10.1016/S0092-8674(00)80357-8).
- Lucas WJ**, Bouche-Pillon S, Jackson DP, Nguyen L, Baker L, Ding B, Hake S. 1995. Selective trafficking of KNOTTED1 homeodomain protein and its mRNA through plasmodesmata. *Science* **270**:1980–1983. doi: [10.1126/science.270.5244.1980](https://doi.org/10.1126/science.270.5244.1980).
- Masucci JD**, Rerie WG, Foreman DR, Zhang M, Galway ME, Marks MD, Schiefelbein JW. 1996. The homeobox gene *GLABRA2* is required for position-dependent cell differentiation in the root epidermis of *Arabidopsis thaliana*. *Development (Cambridge, England)* **122**:1253–1260.
- Masucci JD**, Schiefelbein JW. 1994. The *rhd6* mutation of *Arabidopsis thaliana* alters root-hair initiation through an auxin- and ethylene-associated process. *Plant Physiology* **106**:1335–1346. doi: [10.1104/pp.106.4.1335](https://doi.org/10.1104/pp.106.4.1335).
- Masucci JD**, Schiefelbein JW. 1996. Hormones act downstream of *TTG* and *GL2* to promote root hair outgrowth during epidermis development in the *Arabidopsis* root. *The Plant Cell* **8**:1505–1517. doi: [10.1105/tpc.8.9.1505](https://doi.org/10.1105/tpc.8.9.1505).
- Peng P**, Yan Z, Zhu Y, Li J. 2008. Regulation of the *Arabidopsis* GSK3-like kinase BRASSINOSTEROID-INSENSITIVE 2 through proteasome-mediated protein degradation. *Molecular Plant* **1**:338–346. doi: [10.1093/mp/ssn001](https://doi.org/10.1093/mp/ssn001).
- Ryu KH**, Kang YH, Park YH, Hwang I, Schiefelbein J, Lee MM. 2005. The WEREWOLF MYB protein directly regulates *CAPRICE* transcription during cell fate specification in the *Arabidopsis* root epidermis. *Development (Cambridge, England)* **132**:4765–4775. doi: [10.1242/dev.02055](https://doi.org/10.1242/dev.02055).
- Saidi Y**, Hearn TJ, Coates JC. 2012. Function and evolution of 'green' GSK3/Shaggy-like kinases. *Trends in Plant Science* **17**:39–46. doi: [10.1016/j.tplants.2011.10.002](https://doi.org/10.1016/j.tplants.2011.10.002).

- Savage NS**, Walker T, Wieckowski Y, Schiefelbein J, Dolan L, Monk NAM. 2008. A mutual support mechanism through intercellular movement of CAPRICE and GLABRA3 can pattern the *Arabidopsis* root epidermis. *PLOS Biology* **6**:e235. doi: [10.1371/journal.pbio.0060235](https://doi.org/10.1371/journal.pbio.0060235).
- Savaldi-Goldstein S**, Peto C, Chory J. 2007. The epidermis both drives and restricts plant shoot growth. *Nature* **446**:199–202. doi: [10.1038/nature05618](https://doi.org/10.1038/nature05618).
- Schiefelbein J**, Kwak SH, Wieckowski Y, Barron C, Bruex A. 2009. The gene regulatory network for root epidermal cell-type pattern formation in *Arabidopsis*. *Journal of Experimental Botany* **60**:1515–1521. doi: [10.1093/jxb/ern339](https://doi.org/10.1093/jxb/ern339).
- Schnall JA**, Quatrano RS. 1992. Abscisic acid elicits the water-stress response in root hairs of *Arabidopsis thaliana*. *Plant Physiology* **100**:216–218. doi: [10.1104/pp.100.1.216](https://doi.org/10.1104/pp.100.1.216).
- Schultz EA**, Haughn GW. 1991. *LEAFY*, a homeotic gene that regulates inflorescence development in *Arabidopsis*. *The Plant Cell* **3**:771–781. doi: [10.1105/tpc.3.8.771](https://doi.org/10.1105/tpc.3.8.771).
- She J**, Han Z, Kim TW, Wang J, Cheng W, Chang J, Shi S, Wang J, Yang M, Wang ZY, Chai J. 2011. Structural insight into brassinosteroid perception by BRI1. *Nature* **474**:472–476. doi: [10.1038/nature10178](https://doi.org/10.1038/nature10178).
- Song SK**, Ryu KH, Kang YH, Song JH, Cho YH, Yoo SD, Schiefelbein J, Lee MM. 2011. Cell fate in the *Arabidopsis* root epidermis is determined by competition between WEREWOLF and CAPRICE. *Plant Physiology* **157**:1196–1208. doi: [10.1104/pp.111.185785](https://doi.org/10.1104/pp.111.185785).
- Van Hengel AJ**, Barber C, Roberts K. 2004. The expression patterns of arabinogalactan-protein AtAGP30 and *GLABRA2* reveal a role for abscisic acid in the early stages of root epidermal patterning. *The Plant Journal: for Cell and Molecular Biology* **39**:70–83. doi: [10.1111/j.1365-313X.2004.02104.x](https://doi.org/10.1111/j.1365-313X.2004.02104.x).
- Vollbrecht E**, Veit B, Sinha N, Hake S. 1991. The developmental gene *Knotted-1* is a member of a maize homeobox gene family. *Nature* **350**:241–243. doi: [10.1038/350241a0](https://doi.org/10.1038/350241a0).
- Wada T**, Tachibana T, Shimura Y, Okada K. 1997. Epidermal cell differentiation in *Arabidopsis* determined by a *Myb* homolog, CPC. *Science* **277**:1113–1116. doi: [10.1126/science.277.5329.1113](https://doi.org/10.1126/science.277.5329.1113).
- Walter M**, Chaban C, Schütze K, Batistic O, Weckermann K, Näke C, Blazevic D, Grefen C, Schumacher K, Oecking C, Harter K, Kudla J. 2004. Visualization of protein interactions in living plant cells using bimolecular fluorescence complementation. *The Plant Journal: for Cell and Molecular Biology* **40**:428–438. doi: [10.1111/j.1365-313X.2004.02219.x](https://doi.org/10.1111/j.1365-313X.2004.02219.x).
- Wang H**, Yang C, Zhang C, Wang N, Lu D, Wang J, Zhang S, Wang ZX, Ma H, Wang X. 2011. Dual role of BK11 and 14-3-3 s in brassinosteroid signaling to link receptor with transcription factors. *Developmental Cell* **21**:825–834. doi: [10.1016/j.devcel.2011.08.018](https://doi.org/10.1016/j.devcel.2011.08.018).
- Wang X**, Chory J. 2006. Brassinosteroids regulate dissociation of BK11, a negative regulator of BRI1 signaling, from the plasma membrane. *Science* **313**:1118–1122. doi: [10.1126/science.1127593](https://doi.org/10.1126/science.1127593).
- Wu X**, Dinneny JR, Crawford KM, Rhee Y, Citovsky V, Zambryski PC, Weigel D. 2003. Modes of intercellular transcription factor movement in the *Arabidopsis* apex. *Development (Cambridge, England)* **130**:3735–3745. doi: [10.1242/dev.00577](https://doi.org/10.1242/dev.00577).
- Yan Z**, Zhao J, Peng P, Chihara RK, Li J. 2009. BIN2 functions redundantly with other *Arabidopsis* GSK3-like kinases to regulate brassinosteroid signaling. *Plant Physiology* **150**:710–721. doi: [10.1104/pp.109.138099](https://doi.org/10.1104/pp.109.138099).
- Yang CJ**, Zhang C, Lu YN, Jin JQ, Wang XL. 2011. The mechanisms of brassinosteroids' action: from signal transduction to plant development. *Molecular Plant* **4**:588–600. doi: [10.1093/mp/ssr020](https://doi.org/10.1093/mp/ssr020).
- Ye Q**, Zhu W, Li L, Zhang S, Yin Y, Ma H, Wang X. 2010. Brassinosteroids control male fertility by regulating the expression of key genes involved in *Arabidopsis* anther and pollen development. *Proceedings of the National Academy of Sciences of the United States of America* **107**:6100–6105. doi: [10.1073/pnas.0912333107](https://doi.org/10.1073/pnas.0912333107).
- Zhang F**, Gonzalez A, Zhao M, Payne CT, Lloyd A. 2003. A network of redundant bHLH proteins functions in all TTG1-dependent pathways of *Arabidopsis*. *Development* **130**:4859–4869. doi: [10.1242/dev.00681](https://doi.org/10.1242/dev.00681).
- Zhao M**, Morohashi K, Hatlestad G, Grotewold E, Lloyd A. 2008. The TTG1-bHLH-MYB complex controls trichome cell fate and patterning through direct targeting of regulatory loci. *Development (Cambridge, England)* **135**:1991–1999. doi: [10.1242/dev.016873](https://doi.org/10.1242/dev.016873).
- Zhu C**, Gan L, Shen Z, Xia K. 2006. Interactions between jasmonates and ethylene in the regulation of root hair development in *Arabidopsis*. *Journal of Experimental Botany* **57**:1299–1308. doi: [10.1093/jxb/erj103](https://doi.org/10.1093/jxb/erj103).