

Broad sarbecovirus neutralization by a human monoclonal antibody

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The recent emergence of SARS-CoV-2 variants of concern^{1–10} and the recurrent spillovers of coronaviruses^{11,12} into the human population highlight the need for broadly neutralizing antibodies that are not affected by the ongoing antigenic drift and that can prevent or treat future zoonotic infections. Here we describe a human monoclonal antibody designated S2X259, which recognizes a highly conserved cryptic epitope of the receptor-binding domain and cross-reacts with spikes from all clades of sarbecovirus. S2X259 broadly neutralizes spike-mediated cell entry of SARS-CoV-2, including variants of concern (B.1.1.7, B.1.351, P.1, and B.1.427/B.1.429), as well as a wide spectrum of human and potentially zoonotic sarbecoviruses through inhibition of angiotensin-converting enzyme 2 (ACE2) binding to the receptor-binding domain. Furthermore, deep-mutational scanning and in vitro escape selection experiments demonstrate that S2X259 possesses an escape profile that is limited to a single substitution, G504D. We show that prophylactic and therapeutic administration of S2X259 protects Syrian hamsters (*Mesocricetus auratus*) against challenge with the prototypic SARS-CoV-2 and the B.1.351 variant of concern, which suggests that this monoclonal antibody is a promising candidate for the prevention and treatment of emergent variants and zoonotic infections. Our data reveal a key antigenic site that is targeted by broadly neutralizing antibodies and will guide the design of vaccines that are effective against all sarbecoviruses.

SARS-CoV-2 has caused a global pandemic with over 182 million infections and more than 3 million fatalities as of July 2021. SARS-CoV-2 genetic drift has resulted in emerging variants of concern that are characterized by higher transmissibility, immune evasion and disease severity relative to the ancestral isolate^{1–10}. Countermeasures, such as vaccines and therapeutic agents, are needed to cope with SARS-CoV-2 evolution and to protect against future sarbecovirus spillovers.

The sarbecovirus spike (S) glycoprotein mediates viral entry into host cells, represents the main target of neutralizing antibodies and is the focus of vaccine design¹¹. S comprises an S1 subunit, which recognizes host cell receptors, and an S2 subunit that promotes viral–cell membrane fusion^{12,13}. The S1 subunit includes the receptor-binding domain (RBD), which in the case of SARS-CoV and SARS-CoV-2 interacts with ACE2 to allow virus entry^{12,14–16}.

A highly conserved region on sarbecovirus RBDs, designated antigenic site II¹⁷, has previously been shown to elicit SARS-CoV and SARS-CoV-2 cross-neutralizing antibodies^{18,19}. However, site II becomes accessible only when at least two RBDs in the S trimer adopt an open conformation and thus has limited immunogenicity^{17,19}. Here we describe a site-II-targeting monoclonal antibody (mAb) designated S2X259 that possesses exceptional neutralization breadth within the *Sarbecovirus* subgenus (including all SARS-CoV-2 variants of concern), and an escape profile that is limited to a sole and rare substitution. In addition, we show that prophylactic and therapeutic administration of S2X259 protects Syrian hamsters against SARS-CoV-2 challenge with the prototypic virus and the B.1.351 variant of concern. Our findings identify S2X259 as a promising countermeasure to protect against SARS-CoV-2 antigenic drift as well as potentially new zoonotic

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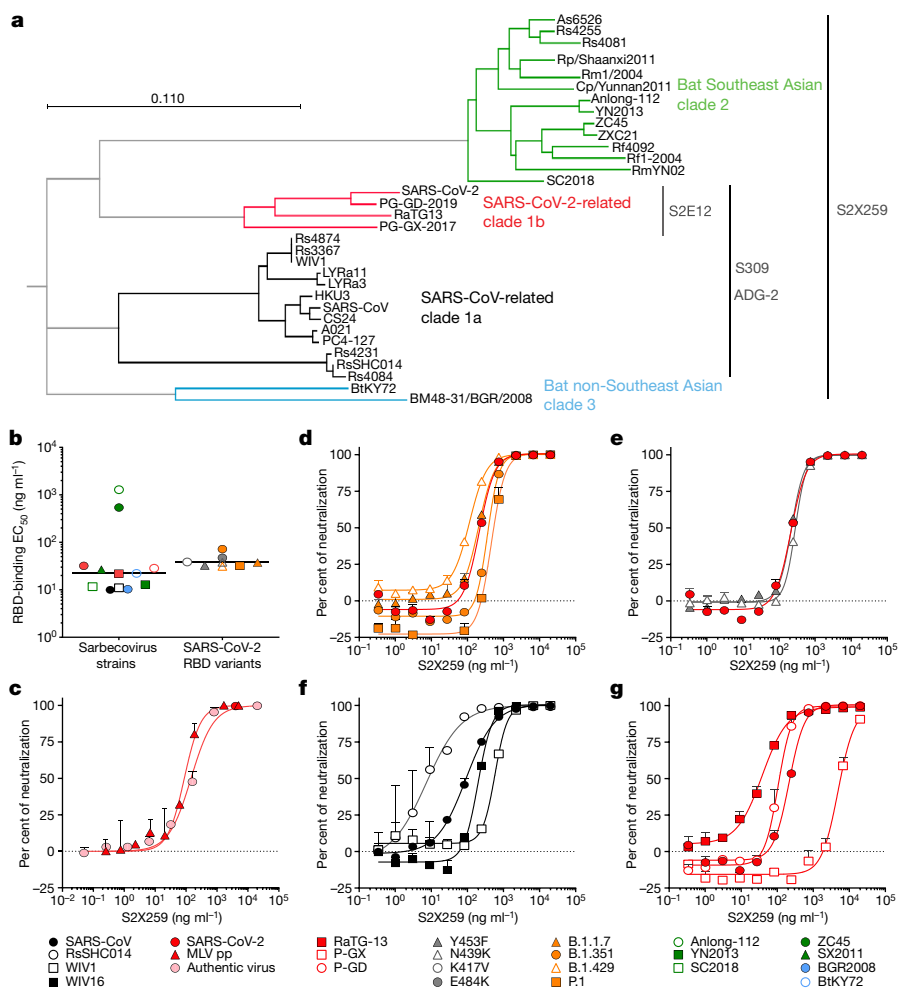


Fig. 1 | Identification of a broadly neutralizing sarbecovirus mAb.

a, Phylogenetic tree of sarbecovirus RBDs constructed using maximum likelihood analysis of amino acid sequences retrieved from GISAID and GenBank. Cross-reactivity within the *Sarbecovirus* subgenus for S2E12²¹, S309²² and ADG-2²⁰ is included for comparison. **b**, S2X259 binding to RBDs representative of the different sarbecovirus clades and SARS-CoV-2 variants as measured by ELISA. **c**, S2X259-mediated neutralization of SARS-CoV-2-Nluc

authentic virus and SARS-CoV-2 S MLV-pseudotyped virus (MLV pp). **d**, **e**, S2X259-mediated neutralization of VSV pseudotypes containing SARS-CoV-2 S from isolates representing the B.1.1.7, B.1.351, P.1 and B.1.429 variants of concern (**d**) as well as single RBD mutants (**e**). **f**, **g**, S2X259-mediated neutralization of VSV pseudotypes containing SARS-CoV-related (clade 1a) (**f**) or SARS-CoV-2-related (clade 1b) (**g**) S glycoproteins. $n = 2$ independent experiments. Error bars indicate s.d. of duplicates or triplicates.

infections, and highlight the importance of RBD site II for the design of vaccines eliciting antibodies against all sarbecoviruses.

A broadly neutralizing sarbecovirus mAb

We identified the mAb designated S2X259 from the memory B cells of an individual who had recovered from COVID-19 (Supplementary Table 1); this mAb cross-reacted with 25 out of 26 S glycoproteins representative of all clades of sarbecovirus^{18,20} (Fig. 1a, Extended Data Fig. 1a–c). However, no binding was observed to embecovirus S (HCoV-HKU1 and HCoV-OC43) or merbecovirus S (MERS-CoV) glycoproteins (Extended Data Fig. 1a, b).

The cross-reactivity of S2X259 within the *Sarbecovirus* subgenus was further confirmed using enzyme-linked immunosorbent assay (ELISA) with a panel of 12 RBDs (Fig. 1b). S2X259 bound tightly to all of the RBDs tested (half maximal effective concentration (EC_{50}) of less than 32 ng ml⁻¹) with the exception of two bat strains, ZC45 (EC_{50} of 539 ng ml⁻¹) and Anlong-112 (EC_{50} of 1,281 ng ml⁻¹) (Fig. 1b). The broad recognition of S from the divergent bat Southeast Asian and non-Southeast Asian clades (clades 2 and 3, respectively) highlights the exceptional breadth of S2X259 in comparison with previously reported mAbs^{17,21,22}

(Extended Data Fig. 1d). Surface plasmon resonance indicated that the S2X259 Fab fragment bound to clade 1a, 1b and 3 RBDs with nano- to picomolar affinities, and to clade 2 RBDs with micro- to nanomolar affinities. Of note, the affinity of S2X259 for the RBD of the Guanxi pangolin coronavirus was 10–100-fold lower compared to other clade 1b RBDs (Extended Data Fig. 2a). The S2X259 Fab recognized both the SARS-CoV-2 prefusion-stabilized S ectodomain trimer and the recombinant RBD with picomolar affinities, albeit with a slower on-rate for binding to S (presumably owing to limited epitope exposure)^{12,13} (Extended Data Fig. 2a). Finally, S2X259 binding was unaffected by several point mutants or constellation of mutations identified in circulating clinical isolates (K417V, N439K, Y453F and E484K) and in the RBDs of the B.1.1.7 (N501Y), B.1.351 (K417N/E484K/N501Y), B.1.427/B.1.429 (L452R) and P.1 (K417T/E484K/N501Y) lineages (Fig. 1b, Extended Data Fig. 2b).

We next evaluated the neutralization potency of S2X259 using authentic SARS-CoV-2-Nluc as well as murine leukaemia virus (MLV) and vesicular stomatitis virus (VSV) pseudotyping systems. S2X259 potently neutralized authentic SARS-CoV-2, SARS-CoV-2 S MLV and SARS-CoV-2 S VSV with half-maximal inhibitory concentrations (IC_{50}) of 144 ng ml⁻¹, 87 ng ml⁻¹ and 213 ng ml⁻¹, respectively (Fig. 1c, d). We subsequently evaluated S2X259-mediated neutralization against VSV

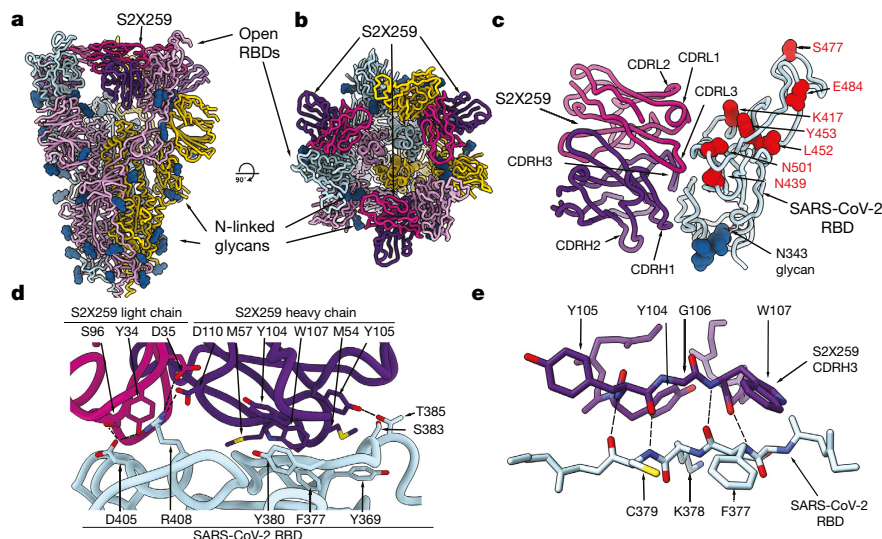


Fig. 2 | The S2X259 broadly neutralizing sarbecovirus mAb recognizes RBD antigenic site II. a, b, Cryo-electron microscopy structure of the prefusion SARS-CoV-2 S ectodomain trimer with three S2X259 Fab fragments bound to three open RBDs, viewed along two orthogonal orientations (Protein Data Bank (PDB) codes 7RA8 and 7RAL). **c,** The S2X259 binding mode involving contacts with multiple RBD regions. Residues corresponding to prevalent RBD

mutations are shown as red spheres (PDB code 7M7W). **d, e,** Close-up views showing selected interactions formed between S2X259 and the SARS-CoV-2 RBD. In **a–e**, each SARS-CoV-2 S protomer is coloured distinctly (cyan, pink and gold), and the S2X259 light and heavy chain variable domains are coloured magenta and purple, respectively. N-linked glycans are rendered as blue spheres in **a–c**.

pseudotypes containing SARS-CoV-2 S from the B.1.1.7, B.1.351, P.1, B.1.427/B.1.429 and B.1.617.1 and other lineages, as well as the N439K and Y453F single mutations^{5,23–25}. S2X259 neutralized all of these mutants with a similar potency, consistent with the identical binding affinities determined using RBD mutants (Fig. 1d, e, Extended Data Figs. 2b, 3a, b, Supplementary Table 2). Furthermore, S2X259 also neutralized a broad panel of ACE2-using sarbecoviruses with comparable potencies, including Guangdong pangolin coronavirus, RaTG13, SARS-CoV, WIV1, WIV16 and RsSHC014 (Fig. 1f, g, Supplementary Table 2). Consistent with the surface plasmon resonance data, S2X259 neutralized VSV pseudotypes containing S of Guanxi pangolin coronavirus but with reduced potency as compared to SARS-CoV-2 (Fig. 1g, Extended Data Fig. 2a, Supplementary Table 2).

Structural basis of sarbecovirus neutralization

Three-dimensional classification of the cryo-electron microscopy data showed the presence of SARS-CoV-2 S ectodomain trimers saturated with three S2X259 Fabs bound to open RBDs swung out to various extents, for which we determined a consensus structure at 3.2 Å resolution (Fig. 2a, b, Extended Data Fig. 4, Supplementary Table 3). We used local refinement to determine a 3.7 Å resolution map of the region corresponding to the S2X259 variable domains and RBD. In parallel, we crystallized the S2X259 Fab-bound SARS-CoV-2 RBD in the presence of the S2H97 Fab²⁶ and determined a 2.65 Å resolution X-ray structure (Fig. 2c–e, Supplementary Table 4).

S2X259 recognizes a glycan-free, cryptic epitope within antigenic site IIa, which was previously defined on the basis of the nonclonally related S2X35 mAb that was isolated from the same donor¹⁷. S2X259 binding requires opening of two RBDs to grant access to the Fab in the context of the S trimer (Fig. 2a, b). S2X259 contacts the RBD using both heavy and light chains, which respectively contribute two thirds and one third of the approximately 950 Å² paratope surface that is buried upon binding. Specifically, S2X259 uses complementary determining regions (CDRs) H1, H2, H3, L1 and L3 to contact RBD residues 369–386, which form two α-helices and an intervening β-strand belonging to the structurally conserved RBD β-sheet, as well as the residues 404–411

and 499–508, which form a continuous surface made up of an α-helix and a loop followed by an α-helix, respectively.

The S2X259 epitope is conserved in circulating SARS-CoV-2 isolates and does not comprise prevalent RBD mutations, such as S477N, N439K or L452R. S2X259 also circumvents residues 417 and 484, and contacts the backbone of residue N501 but not its side chain, explaining the high-affinity binding to the B.1.1.7, B.1.351 and P.1 RBDs and potent neutralization of pseudotyped variant of concern viruses (Fig. 1b, d Extended Data Fig. 2b). S2X259 broad neutralization results from the conservation of its epitope among sarbecoviruses and from the angle of approach of the Fab that circumvents the SARS-CoV N357 glycan present in all sarbecoviruses, except for SARS-CoV-2 (SARS-CoV-2 N370) (Extended Data Fig. 5). The SARS-CoV-2 RBD and that of Guanxi pangolin coronavirus differ at only two positions (G504N and Y508F), and the structural data indicate that the former substitution would probably dampen mAb binding through steric hindrance as indicated by in vitro neutralization data (Fig. 1g).

S2X259 binding to the RBD does not affect or prevent recognition by site-I or site-IV-targeting mAbs (Extended Data Fig. 6a, b), which represent the majority of mAbs currently approved or in the clinic^{17,22,27,28}. Competition binding assays suggest that serum antibodies directed to the RBD antigenic site II are subdominant in individuals who have recovered from SARS-CoV-2 infection and vaccinated individuals, as compared to antibodies directed towards site I (Extended Data Fig. 6c). Therefore, S2X259 can be used in combination with other site I or site IV mAbs to maximize breadth against currently circulating and emerging SARS-CoV-2 variants, as well as against future sarbecovirus infections from animal reservoirs.

S2X259 has an unusually narrow escape profile

Using deep-mutational scanning with a yeast-display RBD mutant library²⁹, we exhaustively mapped mutations that escape S2X259 binding. Notably, binding was strongly reduced by only a restricted number of amino acid substitutions, as compared to previously described neutralizing mAbs²⁶, with mutations at RBD site 504 yielding the most marked reduction in binding (Fig. 3a, Extended Data Fig. 7a–c). Indeed, substitution of G504 with most other amino acids reduced S2X259

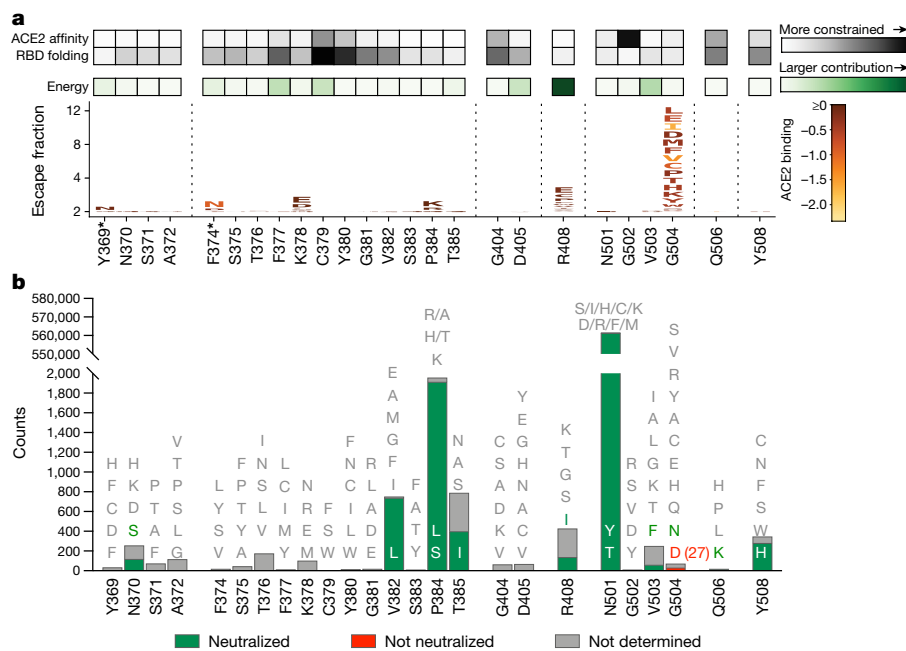


Fig. 3 | S2X259 is resilient to a broad spectrum of escape mutations.

a, Complete mapping of mutations reducing S2X259 binding using yeast-displayed RBD and deep-mutational scanning. Mean mutation effect on ACE2 affinity, RBD folding and contribution to S2X259 binding of substitutions at each position in the S2X259 epitope are reported. Asterisks denote

mutations introducing N-linked glycosylation sites that may not be tolerated in full S. **b**, Frequency of mutants within the S2X259 epitope on the basis of SARS-CoV-2 genome sequences available on GISAID as of May 2021. S2X259 neutralizing activity against selected mutations is reported.

binding compared to the wild-type (Wuhan-Hu-1) RBD, which highlights the importance of this residue for mAb recognition.

To further explore the functional importance of the deep-mutational scanning data, we passaged a replication-competent VSV-SARS-CoV-2 S chimera^{30,31} in Vero cells in the presence of S2X259 mAb. The selective pressure imposed by S2X259 led to the emergence of viral mutants containing the G504D mutation, which was the sole mutation present in each of the 18 neutralization-resistant plaques we sampled. This substitution did not impair viral fitness of the VSV-SARS-CoV-2 S chimera (Extended Data Fig. 7d, e, Supplementary Table 5). Under the same conditions, S2X35¹⁷ (which also targets antigenic site IIa) selected for seven escape mutants at five different positions (Y369S, K378E, D405N, V503D or V503G and G504D or G504S)²⁶ (Supplementary Table 5).

The G504D substitution is very rare: it has been detected in only 27 out of 1,345,708 (0.002%) SARS-CoV-2 genome sequences available on GISAID, as of May 2021 (Fig. 3b). Furthermore, in agreement with the deep-mutational scanning results, S2X259 efficiently neutralized VSV pseudoviruses containing S mutations located in S2X259-contact residues and found with high frequency in human isolates (Fig. 3b, Extended Data Fig. 7f). In the same analysis, we confirmed the loss of S2X259 neutralizing activity against the G504D mutant. S2X259 CDRL1 and CDRL3 contact G504 or surrounding residues, which explains why substituting a glycine with any other residue prevents mAb binding (via steric hindrance along with potential disruption of the interactions of the Fab with RBD residue D405, the latter being in close proximity to G504) (Extended Data Fig. 7g).

These data also support the hypothesis proposed above for the markedly reduced affinity and neutralization potency of S2X259 against Guanxi pangolin pseudovirus which contains the G504N mutation but maintains an aspartate at position 405 (Fig. 1g, Extended Data Figs. 2a, 7f, 8). However, S2X259 binding to clade 2S glycoproteins that bear G504D or G504E mutations (for example, those of Shanxi2011 or YN2013) was not abrogated (Fig. 1b), possibly owing to substitution at the SARS-CoV-2 equivalent position 405 with a serine and to substantial deletions in the receptor-binding motif (Extended Data Fig. 8).

The finding that the only escape mutation for S2X259 is the rare G504D substitution suggests this mAb might have a high barrier for the emergence of SARS-CoV-2 escape mutants, which might prove essential during the next stages of the pandemic.

S2X259 protects hamsters from SARS-CoV-2 challenge

Consistent with structural data that indicates a clash between S2X259 and ACE2 for binding to the RBD (Extended Data Fig. 9a), we found that S2X259 efficiently blocked binding of SARS-CoV-2 and SARS-CoV RBDs to human ACE2 (Extended Data Fig. 9b).

In addition, S2X259 binding efficiently promoted shedding of the S1 subunit from cell-surface-expressed full-length SARS-CoV-2 S, as has previously been shown with other RBD-specific mAbs^{17,32–35} but not with S2M11 (which locks S in the prefusion closed state)²¹ (Extended Data Fig. 9c). The efficient S1 shedding induced by S2X259 might be responsible for the lack of FcγRIIIa and FcγRIIIb activation, which we used as a proxy for antibody-dependent cellular phagocytosis and antibody-dependent cellular cytotoxicity, respectively (Extended Data Fig. 9d, e). Indeed, performing the same assays using target cells that express an uncleavable prefusion stabilized SARS-CoV-2 S protein (unable to release the S1 subunit), we confirmed the ability of S2X259 mAb to induce activation of FcγRIIIa and FcγRIIIb in these conditions (Extended Data Fig. 9f, g).

These data show that the primary mechanism of S2X259-mediated neutralization of SARS-CoV-2 and other sarbecoviruses relies on blocking viral attachment to host-cell receptors through competitive inhibition of ACE2 binding. Premature triggering of S1 subunit shedding could inactivate viruses before encountering target cells, albeit reducing activation of S2X259-mediated effector functions.

Next, we evaluated the prophylactic activity of S2X259 against challenge with the prototypic (Wuhan-Hu-1-related) SARS-CoV-2 in a Syrian hamster model³⁶. S2X259 was administered at 1, 5 and 25 mg per kg body weight via intraperitoneal injection before intranasal challenge, and the lungs of the hamsters were collected 4 days later for the quantification

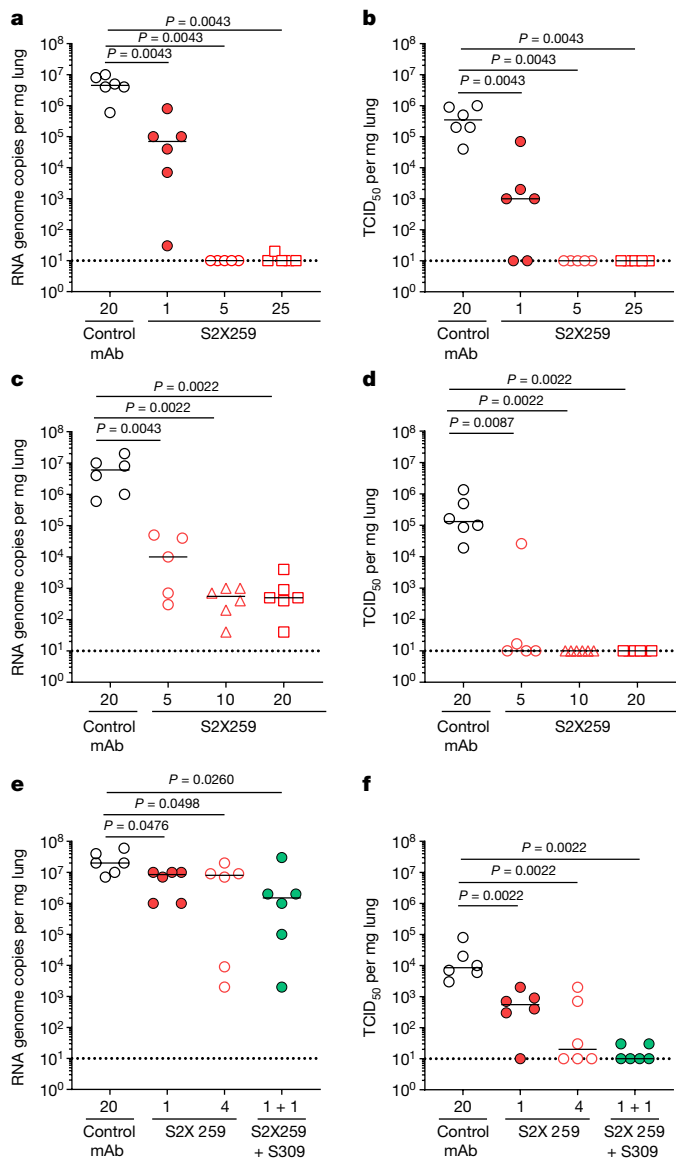


Fig. 4 | S2X259 protects hamsters challenged with prototypic and B.1.351 SARS-CoV-2. **a, b**, Viral RNA loads (**a**) and replicating virus titres (50% tissue culture infectious dose (TCID₅₀)) (**b**) in the lungs of Syrian hamsters at 4 days after intranasal infection with prototypic SARS-CoV-2 (Wuhan-Hu-1-related) following prophylactic administration of S2X259 mAb at 1 ($n = 6$ hamsters), 5 ($n = 5$ hamsters) and 25 ($n = 5$ hamsters) mg kg^{-1} . **c, d**, Viral RNA loads (**c**) and replicating virus titres (TCID₅₀) (**d**) in the lungs of Syrian hamsters after therapeutic administration of S2X259 at 5 ($n = 5$ hamsters), 10 ($n = 6$ hamsters) and 20 ($n = 6$ hamsters) mg kg^{-1} 24 h after infection with SARS-CoV-2 (Wuhan-Hu-1-related). **e, f**, Quantification of viral RNA loads (**e**) and replicating virus titres (TCID₅₀) (**f**) in the lungs of Syrian hamsters 4 days after intranasal challenge with B.1.351 SARS-CoV-2 variant of concern following prophylactic administration of S2X259 at 1 mg kg^{-1} ($n = 6$ hamsters), 4 mg kg^{-1} ($n = 6$ hamsters) and in combination with S309 (1 + 1 mg kg^{-1} , $n = 6$ hamsters). Data from one independent experiment are presented. Two-tailed Mann-Whitney test was used for statistical analysis of significance.

of viral RNA and replicating virus. Despite the lack of Fc-dependent effector functions *in vitro* and their proposed contribution to *in vivo* protection against SARS-CoV-2^{37,38}, administration of S2X259 at 1 mg kg^{-1} reduced the amount of viral RNA and infectious virus detected in the lungs by approximately 2 and 2.5 orders of magnitude, respectively, compared to hamsters that received a control mAb (Fig. 4a, b). Furthermore, complete abrogation of virus replication was observed

in the lungs of the hamsters that were treated with S2X259 at 5 and 25 mg kg^{-1} relative to the control group (Fig. 4b). Therapeutic administration (after exposure) of S2X259 at 5 mg kg^{-1} reduced the amounts of viral RNA detected by 3 log and abrogated viral replication in the lungs of all of the hamsters but one (Fig. 4c, d). Hamsters treated with S2X259 at 10 or 20 mg kg^{-1} displayed a further approximately 1 log reduction of viral RNA and complete abrogation of viral replication (Fig. 4c, d). Viral replication in the lungs detected at day 4 inversely correlated with serum mAb concentration measured at the time of infection or at day 4 after infection in prophylactic and therapeutic settings, respectively (Extended Data Fig. 10a, b).

We next assessed the prophylactic activity of S2X259 against challenge with B.1.351 SARS-CoV-2³⁹. S2X259 at 1 and 4 mg kg^{-1} , or in combination with S309 (each mAb at 1 mg kg^{-1}), significantly decreased the amount of viral RNA detected in the lungs relative to hamsters that received a control mAb (Fig. 4e). Moreover, S2X259 at 4 mg kg^{-1} or in combination with S309 (each mAb at 1 mg kg^{-1}) completely abrogated SARS-CoV-2 B.1.351 virus replication in most of the hamsters (Fig. 4f). As was the case for the prototypic SARS-CoV-2, the amount of infectious virus at day 4 detected in the group administered only with S2X259 also inversely correlated with serum mAb concentration measured at the time of infection (Extended Data Fig. 10c). Moreover, hamsters that received the mAb cocktail appeared to benefit from the additive contribution of the S309 mAb (Fig. 4f, Extended Data Fig. 10d, e).

Discussion

The large number of mutations in the S glycoprotein of SARS-CoV-2 variants of concern^{4,23,25} has been shown to abrogate or reduce the neutralization potency of several mAbs and vaccine-elicited antibodies^{5–10,40}. Moreover, the detection of a large pool of sarbecoviruses in bats and other mammals across several continents suggests that future cross-species transmission events are likely to occur^{41–43}. However, approved SARS-CoV-2 therapeutic agents and vaccines predominantly target or elicit immunity against immunodominant, but highly variable, epitopes and therefore have a limited efficacy against antigenic drift and genetically distinct strains with zoonotic potential^{40,44}.

Strategies for eliciting broadly neutralizing and protective sarbecovirus antibodies targeting the RBD have recently been proposed^{18,40}, and previous work has described mAbs that are now in the clinic with extended coverage within clade I sarbecoviruses^{20,22}. The recent results from a phase III study demonstrating efficacy (that is, 85% reduction of hospitalizations or deaths) of the cross-reactive derivative of the S309 mAb (sotrovimab) paves the way for the development of additional neutralizing mAbs targeting conserved RBD epitopes that may broaden the coverage across sarbecoviruses and increase the barrier to resistance.

Here we identify the S2X259 mAb that targets the RBD antigenic site II and uniquely cross-reacts with all of the SARS-CoV-2 variants of concern that we evaluated, as well as with 30 S trimers or RBDs from all four sarbecovirus clades. S2X259 broadly neutralizes pseudoviruses containing S proteins from the B.1.1.7, B.1.351, P.1, B.1.427/B.1.429, and B.1.617.1 variants, and from other representative members of SARS-CoV-2 and SARS-CoV clades (clades 1b and 1a, respectively). Of note, S2X259 neutralizes pseudotyped WIVIS and RsSHC014 S (clade 1a) viruses, which have previously been suggested to be potential zoonotic threats to human health^{41,42}. Moreover, S2X259 also recognizes S and RBDs from bat Southeast Asian and non-Southeast Asian sarbecovirus clades (clades 2 and 3, respectively). The coverage of clade 2 might be particularly important, as a previous study¹⁶ has shown that exogenous protease addition to pseudoviruses from this clade resulted in detectable S-mediated entry in human cells. Furthermore, although clade 2 and clade 3 sarbecoviruses have not been reported to bind human ACE2, mutation or recombination events could lead to human ACE2 binding and entry^{16,45}. Overall, these findings indicate that S2X259 is a

promising candidate for clinical development and could be stockpiled as part of a pandemic preparedness toolbox.

On the basis of the growing body of data demonstrating that antibodies targeting the SARS-CoV-2 RBD account for most neutralizing activity in sera from patients who have recovered from COVID-19^{17,46}, we propose that RBD-based vaccines could better promote elicitation of high titres of S2X259-like neutralizing antibodies, owing to the enhanced accessibility of this antigenic site compared to S-based vaccines^{40,47}. Our serological competition data confirm that the antibody responses against the conserved site targeted by S2X259 mAb is subdominant in donors who have recovered from COVID-19 as well as in individuals who have received S-based mRNA vaccines. We anticipate these data will guide future efforts to develop vaccines that overcome the emergence of SARS-CoV-2 variants and that are effective against all sarbecoviruses.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41586-021-03817-4>.

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Methods

Cell lines

Cell lines used in this study were obtained from ATCC (HEK293T, Vero and Vero E6), ThermoFisher Scientific (ExpiCHO-S cells, FreeStyle 293-F cells and Expi293F cells) or were generated via lentiviral transduction (ExpiCHO-S and HEK293T expressing ACE2 (HEK293T-ACE2)). None of the cell lines used was authenticated. Cell lines were routinely tested for mycoplasma contamination.

Sample donors

Samples from an individual who had recovered from COVID-19 (designated as donor X (a 52-year-old man)) was obtained 75 days after symptom onset under study protocols approved by the local Institutional Review Boards (Canton Ticino Ethics Committee, Switzerland). Samples from additional nine individuals who had recovered from COVID-19 and nine vaccinated individuals were obtained under study protocols approved by the Canton Ticino Ethics Committee, Switzerland and the Ethical committee of Luigi Sacco Hospital, Milan. Samples from individuals who had recovered from COVID-19 were collected between 14 and 75 days after symptom onset; samples from vaccinated individuals were collected between 13 and 20 days after the second dose of an mRNA-based SARS-CoV-2 vaccine. All donors provided written informed consent for the use of blood and blood components (such as peripheral blood mononuclear cells (PBMCs), sera or plasma).

Isolation of PBMCs, plasma and sera

PBMCs were isolated from blood draw performed using tubes pre-filled with heparin, followed by Ficoll density gradient centrifugation. PBMCs were either used freshly along SARS-CoV-2 S-protein-specific memory B cell sorting or stored in liquid nitrogen for later use. Sera were obtained from blood collected using tubes containing clot activator, followed by centrifugation and stored at -80°C .

B cell isolation and recombinant S2X259 mAb production

Starting from freshly isolated PBMCs or upon cells thawing, B cells were enriched by staining with CD19 PE-Cy7 and incubation with anti-PE beads, followed by positive selection using LS columns. Enriched B cells were stained with anti-IgM, anti-IgD, anti-CD14 and anti-IgA, all PE labelled, and perfusion SARS-CoV-2 S with a biotinylated avi tag conjugated to streptavidin Alexa-Fluor 647 (Life Technologies). SARS-CoV-2 S-specific IgG⁺ memory B cells were sorted by flow cytometry via gating for PE-negative and Alexa-Fluor 647-positive cells. Sorted IgG⁺ memory B cells were seeded in 384-well microtitre plates on a monolayer of mesenchymal stromal cells at 0.5 cells per well in the presence CpG2006, IL-2, IL-6 and IL-21, as previously described⁴⁸. After 7 days, the supernatants were screened for binding to SARS-CoV-2 RBD and perfusion S as well as for neutralization activity using a high-throughput VSV SARS-CoV-2 S-based microneutralization assay. Selection of the antibody of interest in secondary screening was based on reactivity to a panel of sarbecovirus RBDs as well as on the neutralizing activity. Antibody VH and VL sequences were obtained by RT-PCR and mAbs were expressed as recombinant human Fab fragment or as IgG1 (G1m3 allotype). ExpiCHO cells were transiently transfected with heavy and light chain expression vectors as previously described²².

Affinity purification was performed on ÄKTA Xpress FPLC (Cytiva) operated by UNICORN software version 5.11 (build 407) using HiTrap Protein A columns (Cytiva) for full length human and hamster mAbs and CaptureSelect CH1-XL MiniChrom columns (ThermoFisher Scientific) for Fab fragments, using PBS as mobile phase. Buffer exchange to the appropriate formulation buffer was performed with a HiTrap Fast desalting column (Cytiva). The final products were sterilized by filtration through 0.22- μm filters and stored at 4°C .

Sarbecovirus sequence analysis and SARS-CoV-2 mutant counts

Alignment and phylogenetic tree of the strains within the *Sarbecovirus* subgenus was generated using MEGA X and CLC Main work bench 21.0.3 (Qiagen). The following sequence was retrieved from GISAID and NCBI: A021 (AAV97986.1). The viral sequences were obtained from GISAID EpiCoV project (<https://www.gisaid.org/>). Analysis was performed on sequences submitted to GISAID up to 2 April 2021. The S protein sequences were either obtained directly from the protein dump provided by GISAID or, for the latest submitted sequences that were not yet incorporated in the protein dump on the day of data retrieval, from the genomic sequences with exonerate⁴⁹ 2.2.4.0-haf93ef1_3 (<https://quay.io/repository/biocontainers/exonerate?tab=tags>) using protein to DNA alignment with parameters `-m protein2dna --refine full --minintron 999999 --percent 20` and using accession YP_009724390.1 as a reference. Multiple sequence alignment of all human S proteins was performed with mafft⁵⁰ 7.475-h516909a_0 (<https://quay.io/repository/biocontainers/mafft?tab=tags>) with parameters `--auto --reorder --keeplength --addfragments` using the same reference as above. S sequences that contained more than 10% ambiguous amino acids or that were less than 80% of the canonical protein length were discarded. A total of 923,686 sequences were used for analysis. Variants were then extracted as compared to the reference with R 4.0.2 (<https://www.r-project.org/>) using Biostrings 2.56.0.

Binding to cell-surface-expressed sarbecovirus S proteins by flow cytometry

ExpiCHO-S cells were seeded at 6×10^6 cells per ml in a volume of 5 ml in a 50-ml bioreactor. S-encoding plasmids HKU3 (QND76020.1), Rs3367 (AGZ48818.1), YN2013 (AIA62330.1), Rs4874 (ATO98205.1), Rs4255 (ATO98193.1), Rs4231 (ATO98157.1), Rs4084 (ATO98132.1), ZXC21 (AVP78042.1), ZC45 (AVP78031.1), Rp/Shaanxi2011 (AGC74165.1), Rm1/2004 (ABD75332.1), Rf1-2004 (ABD75323.1), BM48-31/BGR/2008 (YP_003858584.1), RaTG13 (QHR63300.2), PC4-127 (AAU93318.1), SARS-CoV-2 (YP_009724390.1), LYRa3 (AHX37569.1), Cp/Yunnan2011 (AGC74176.1), SARS-CoV Urbani (AAP13441.1), As6526 (ATO98108.1), BtKY72 (APO40579.1), RmYN02 (EPI_ISL_412977), Pangolin_Guangdong-2019 (EPI_ISL_410721), Pangolin-Guanxi-2017 (EPI_ISL_410539) and HKU3-12 (ADE34812.1) were diluted in cold OptiPRO SFM (Life Technologies, 12309-050), mixed with ExpiFectamine CHO Reagent (Life Technologies, A29130) and added to the cells. Transfected cells were then incubated at 37°C with 8% CO_2 with an orbital shaking speed of 250 r.p.m. (orbital diameter of 25 mm) for 42 h. Transiently transfected ExpiCHO cells were collected and washed two times in wash buffer (PBS 2 mM FBS, 2 mM EDTA). Cells were counted and distributed into round-bottom 96-well plates (Corning, 3799) and incubated with S2X259 mAb serial dilutions from $50 \mu\text{g ml}^{-1}$. Alexa Fluor 647-labelled goat anti-human IgG secondary antibody (Jackson ImmunoResearch, 109-606-098) was prepared at $2 \mu\text{g ml}^{-1}$ and added onto cells after two washing steps. Cells were then washed twice and resuspended in wash buffer for data acquisition at ZES cytometer (Biorad).

Recombinant protein production

Wild-type SARS-CoV-2 RBD (with N-terminal signal peptide and 'ETGT', and C-terminal 8 \times His tag) was expressed in Expi293F cells at 37°C and 8% CO_2 in the presence of $10 \mu\text{M}$ kifunensine. Transfection was performed using the ExpiFectamine 293 Transfection Kit (Thermo Fisher Scientific). Cell culture supernatant was collected four days after transfection and supplemented with $10\times$ PBS to a final concentration of $2.5\times$ PBS (342.5 mM NaCl, 6.75 mM KCl and 29.75 mM phosphates). For crystallization, wild-type SARS-CoV-2 RBD was purified using a 5-ml HisTALON superflow cartridge (Takara Bio) followed by size-exclusion chromatography using a Superdex 200 Increase 10/300 GL column pre-equilibrated in 20 mM Tris-HCl pH 7.5, 150 mM NaCl. RBD was deglycosylated with EndoH and mixed with a 1.3-fold molar

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excess of S2X259 Fab and S2H97 Fab. The complex was purified on a Superdex 200 Increase 10/300 GL column pre-equilibrated with 20 mM Tris-HCl pH 7.5, 150 mM NaCl. SARS-CoV-2 S hexaprotein, used for cryo-electron microscopy (cryo-EM) single particle studies, was expressed and purified as previously described²¹.

ELISA

Ninety-six half area well-plates (Corning, 3690) were coated overnight at 4 °C with 25 µl of sarbecovirus RBD proteins WIV1 (AGZ48831.1), Anlong-112 (ARI44804.1), YN2013 (AIA62330.1), SC2018 (QDF43815.1), ZC45 (AVP78031.1), Rp/Shaanxi2011 (AGC74165.1), BM48-31/BGR/2008 (YP_003858584.1), RaTG13 (QHR63300.2), SARS-CoV-2 (YP_009724390.1), SARS-CoV Urbani (AAP13441.1), BtKY72 (APO40579.1), Pangolin_Guangdong-2019 (EPI_ISL_410721) and SARS-CoV-2 RBD mutants, prepared at 5 µg ml⁻¹ in PBS pH 7.2. Plates were then blocked with PBS 1% BSA (Sigma-Aldrich, A3059) and subsequently incubated with mAb serial dilutions for 1 h at room temperature. After 4 washing steps with PBS 0.05% Tween 20 (PBS-T) (Sigma-Aldrich, 93773), goat anti-human IgG secondary antibody (Southern Biotech, 2040-04) was added and incubated for 1 h at room temperature. Plates were then washed four times with PBS-T and 4-nitrophenyl phosphate (pNPP, Sigma-Aldrich, 71768) substrate was added. After 30 min incubation, absorbance at 405 nm was measured by a plate reader (Biotek) and data were plotted using Prism GraphPad 9.1.0.

S2X259 blockade-of-binding to RBD

S2X259 and S2H14 mAbs were biotinylated using EZ-Link NHS-PEG solid phase biotinylation kit (ThermoFisher Scientific) and binding to RBD was tested to set optimal concentration to be used in the assay after sample desalting using Zeba Spin Desalting Columns (ThermoFisher Scientific). Half-area 96-well-plates were coated overnight at 4 °C with SARS-CoV-2 RBD-mouse Fc Tag diluted at 1 µg ml⁻¹ in PBS. After a blocking step with Blocker Casein (ThermoFisher Scientific), serial plasma dilutions in Blocker Casein were incubated 30 min at room temperature. Biotinylated S2X259 or S2H14 were added at a concentration achieving 80% of maximal binding and the mixture was incubated for 30 min at room temperature. Alkaline-phosphatase conjugated streptavidin (Jackson ImmunoResearch) was diluted at 0.5 µg ml⁻¹ in Blocker Casein and added on plates previously washed 4 times with PBS 0.05% Tween 20. After 45 min incubation, plates were washed and 4-nitrophenyl phosphate substrate incubated for 1 h at room temperature. Absorbance at 405 nm was measured and percentage of inhibition was calculated as follows: $(1 - (\text{optical density (OD) sample} - \text{OD negative control}) / (\text{OD positive control} - \text{OD negative control})) \times 100$.

MLV-based pseudotyped virus production and neutralization

To generate SARS-CoV-2 S MLV pseudotyped virus, HEK293T cells were seeded in 10-cm dishes in DMEM supplemented with 10% FBS. The next day cells were transfected with a SARS-CoV-2 S glycoprotein-encoding plasmid containing the D19 C-terminal truncation⁵¹, an MLV Gag-Pol packaging construct and the reporter vector pTG-Luc, using the X-tremeGENE HP DNA transfection reagent (Roche) according to the manufacturer's instructions. Cells were then incubated at 37 °C with 5% CO₂ for 72 h. Supernatant was collected and cleared from cellular debris by centrifugation at 400g, and stored at -80 °C.

For neutralization assays, Vero E6 cells were seeded into white 96-well plates (PerkinElmer) at 20,000 cells per well and cultured overnight at 37 °C with 5% CO₂ in 100 µl DMEM supplemented with 10% FBS and 1% penicillin-streptomycin. The next day, MLV-SARS-CoV-2 pseudovirus was activated with 10 µg ml⁻¹ TPCK-treated trypsin (Worthington Biochem) for 1 h at 37 °C. Recombinant antibodies at various concentrations were incubated with activated pseudovirus for 1 h at 37 °C. The Vero E6 cells were then washed with DMEM, and 50 µl of pseudovirus-mAbs mixes were added and incubated for 2 h at 37 °C with 5% CO₂. After incubation, 50 µl of DMEM containing 20% FBS and 2%

penicillin-streptomycin were added and the cells were incubated 48 h at 37 °C with 5% CO₂. Following these 48 h of infection, culture medium was removed from the cells and 50 µl per well of Bio-Glo (Promega) diluted 1:2 with PBS with Ca²⁺Mg²⁺ (Thermo Fisher) was added to the cells and incubated in the dark for 15 min before reading on a Synergy HI Hybrid Multi-Mode plate reader (Biotek). Measurements were done in duplicate. Relative light unit (RLU) values were converted to percentage of neutralization and plotted with a nonlinear regression curve fit in GraphPad Prism 9.1.0.

VSV-based pseudotype virus production and neutralization assay

SARS-CoV-2 S (CAD0240757.1), RaTG13 S (QHR63300.2), Pangolin-Guangdong S (QLR06867.1), Pangolin-Guanxi S (QIA48623.1), SARS-CoV S (YP_009825051.1), WIV1 S (AGZ48831.1), WIV16 S (ALK02457.1), RsSHCO14 S (AGZ48806.1), the variant of concern B.1.429 S (QTC60823) and the variants of concern, N437K, Y453F, B.1.1.7 S, B.1.351 S and P.1 S with their corresponding mutations inserted in the SARS-CoV-2 S (CAD0240757.1) backbone were used to pseudotype VSV. Pseudotyped viruses were prepared using 293T cells seeded in 10-cm dishes. In brief, cells in DMEM supplemented with 10% FBS, 1% penicillin-streptomycin were transfected with the plasmid encoding for the corresponding S glycoprotein using lipofectamine 2000 (Life Technologies) following manufacturer's indications. One day after transfection, cells were infected with VSV (G*ΔG-luciferase) and after 2 h, infected cells were washed 4 times with DMEM before adding medium supplemented with anti-VSV-G antibody (II- mouse hybridoma supernatant diluted 1 to 50, from CRL-2700, ATCC). Particles were collected 18 h after inoculation, clarified from cellular debris by centrifugation at 2,000g for 5 min and concentrated 10 times using a 30-kDa cut-off membrane and used for neutralization experiments, aliquoted and frozen at -80 °C until use in neutralization experiments.

For neutralization, stable 293T cells expressing ACE2⁵² in DMEM supplemented with 10% FBS, 1% penicillin-streptomycin were seeded at 40,000 cells per well into clear-bottom white-walled 96-well plates and cultured overnight at 37 °C. Twelve-point threefold serial dilutions of the corresponding mAb were prepared in DMEM and pseudotyped VSV viruses were added 1:1 to each mAb dilution in the presence of anti-VSV-G antibody from II- mouse hybridoma supernatant diluted 50 times. After 45 min incubation at 37 °C, 40 µl of the mixture was added to the cells and 2 h after infection, 40 µl DMEM was added to the cells. After 17–20 h 50 µl per well of One-Glo-EX substrate (Promega) was added to the cells and incubated in the dark for 5–10 min before reading on a Varioskan LUX plate reader (ThermoFisher). Measurements were done in duplicate with two independent productions of pseudotyped viruses and RLU values were converted to percentage of neutralization and plotted with a nonlinear regression curve fit in GraphPad Prism 9.1.0.

Generation of VSV SARS-CoV-2 S pseudotypes from a selection of circulating variants

Replication-defective VSV pseudovirus expressing SARS-CoV-2 S protein were generated as previously described⁵³ with some modifications. Plasmids encoding the SARS-CoV-2 S glycoprotein corresponding to the variants of concern used to produce SARS-CoV-2-VSV were obtained using a multisep overlap extension PCR protocol⁵⁷. In brief, the mutations of the different variant of concern lineages were encoded on each primer pair used to amplify sequential, overlapping fragments of the SARS-CoV-2 S encoding DNA sequence from a pCDNA_SARS-CoV-2_S D19 plasmid encoding the C-terminally truncated S protein that has been shown to be better expressed at the cell³⁰. The amplified overlapping fragments were separated on agarose gel and purified using Illustra GFX PCR DNA and Gel Band Purification Kit (Merck KGaA). The purified overlapping fragments were joined by performing 10 cycles of PCR without the addition of primers, followed by 30 PCR cycles using

a single pair of external primers carrying 5' overhangs homologous to the vector backbone. The final PCR product was then cloned into the pCDNA3 vector using the Takara In-fusion HD cloning kit following manufacturer's instructions.

Lenti-X 293T cells (Takara, 632180) were seeded in 10-cm dishes at a density of 5×10^6 cells per cm^2 and the following day transfected with 10 μg of S expression plasmid with TransIT-Lenti (Mirus, 6600) according to the manufacturer's instructions. One day after transfection, cells were infected with VSV-luc (VSV-G) (Kerafast, EH1020-p.m.) for 1 h, rinsed three times with PBS, then incubated for an additional 24 h in complete medium at 37 °C. The cell supernatant was clarified by centrifugation, filtered (0.45 μm), aliquoted and frozen at -80 °C.

For VSV pseudovirus neutralization assays, Vero-E6 cells were grown in DMEM supplemented with 10% FBS and seeded into CulturePlate-96, White Opaque 96-well Microplate (PerkinElmer) at a density of 2×10^4 cells per well. The next day, mAbs were serially diluted in prewarmed complete medium, mixed at 1:1 ratio with pseudovirus and incubated for 1 h at 37 °C in round-bottom polypropylene plates. Medium from cells was aspirated and 50 μl of virus-mAb complexes were added to cells and then incubated for 1 h at 37 °C. An additional 100 μl of prewarmed complete medium was then added on top of complexes and cells incubated for an additional 16–24 h. Conditions were tested in duplicate wells.

Virus-mAb-containing medium was then aspirated from cells and 50 μl of a 1:2 dilution of SteadyLite Plus (Perkin Elmer) in PBS with $\text{Ca}^{2+}\text{Mg}^{2+}$ was added to cells. Plates were incubated for 10 min at room temperature and then analysed on the Synergy H1 Hybrid Multi-Mode reader (Biotek). Measurements were done in two independent experiments and RLU values were converted to percentage of neutralization and plotted with a nonlinear regression curve fit with an upper constraint equal to 100 and a lower constraint equal to 0 in Prism (version 9.1.0). Half-maximal inhibitory concentration (IC_{50}) values across biological replicates are presented as arithmetic mean \pm s.d.

Neutralization of SARS-CoV-2 B.1.351 pseudotyped VSV by S2X259 + S309 antibody cocktail

Vero E6 cells were plated into white 96-well plates (PerkinElmer) at 20,000 cells per well and cultured overnight at 37 °C 5% CO_2 in 100 μl culture medium (DMEM 10% FBS 1% penicillin-streptomycin). The next day, S2E12 and S309 antibodies were serially diluted in infection medium (DMEM 10% heat-inactivated FBS 1% penicillin-streptomycin) and mixed in a matrix format before incubation with SARS-CoV-2 B.1.351 pseudovirus (multiplicity of infection (MOI) 0.1) at 37 °C 5% CO_2 for 1 h. Cell culture medium was aspirated and 50 μl of mAbs-pseudovirus mix added onto cells. After incubation at 37 °C 5% CO_2 for 1 h, 100 μl of infection medium was added to cell and incubation continued for the following 20 h. For results read out, medium was removed and 50 μl per well of SteadyLite PLUS (PerkinElmer) 1:2 diluted in PBS + Ca^{2+} and Mg^{2+} was added and incubated 15 min at room temperature in the dark. Luminescence signals were measured using a Synergy H1 reader (Biotek) with 1-s integration time. Data were analysed using MacSynergy II and synergy plots obtained with 99.9% confidence were used for graphical elaboration with GraphPad Prism 9.1.0.

Neutralization of authentic SARS-CoV-2-Nluc virus

Neutralization of authentic SARS-CoV-2 by entry-inhibition assay was determined using SARS-CoV-2-Nluc, an infectious clone of SARS-CoV-2 (based on strain 2019-nCoV/USA_WA1/2020) that encodes nanoluciferase in place of the viral ORF7 and demonstrated comparable growth kinetics to wild-type virus⁵⁴. Vero E6 cells were seeded into black-walled, clear-bottom 96-well plates at 2×10^4 cells per well and cultured overnight at 37 °C. The next day, 9-point fourfold serial dilutions of mAbs were prepared in infection medium (DMEM + 10% FBS). SARS-CoV-2-Nluc was diluted in infection medium at a final MOI of 0.01 plaque-forming units per cell, added to the mAb dilutions and

incubated for 30 min at 37 °C. Medium was removed from the Vero E6 cells, mAb-virus complexes were added and incubated at 37 °C for 24 h. Medium was removed from the cells, Nano-Glo luciferase substrate (Promega) was added according to the manufacturer's recommendations, incubated for 10 min at room temperature and the luciferase signal was quantified on a VICTOR Nivo plate reader (Perkin Elmer).

Affinity determination by surface plasmon resonance

Surface plasmon resonance binding measurements were performed using a Biacore T200 instrument, running buffer was Cytiva HBS-EP+ (pH 7.4), and all measurements were performed at 25 °C. For Fab binding, anti-AviTag pAb (for capturing S proteins) or StrepTactin XT (for capturing RBDs) were covalently immobilized on CM5 chips. S2X259 Fab concentrations were 11, 33, 100, and 300 nM run as single-cycle kinetics. Double reference-subtracted data were fit to a 1:1 binding model using Biacore Evaluation software, which yields an 'apparent K_D ' ($K_{D,app}$) for the S-binding data because the kinetics also reflect S conformational dynamics. For SARS-CoV-2 S the dissociation rate was too slow to fit, so the $K_{D,app}$ is reported as an upper limit. The K_D above 1 μM is approximate and was determined from a fit in which R_{max} was set to a constant based on theoretical R_{max} .

Affinity determination using biolayer interferometry

Biotinylated RBD (wild type, N501Y, K417N-E484K-N501Y, or K417T-E484K-N501Y) were immobilized at 5 ng μl^{-1} in undiluted 10X Kinetics Buffer (Pall) to SA sensors until a load level of 1.1 nm. A 1:3 dilution series of Fab in undiluted kinetics buffer starting at 1.0 nM was used for 600 s association before 600 s dissociation to determine protein-protein affinity. The data were baseline-subtracted and the plots fitted using the Pall FortéBio/Sartorius analysis software (version 12.0). Data were plotted in Prism 9.1.0.

Competition assay by biolayer interferometry

Biolayer interferometry was used to assess S2X259 competition with S309 and S2E12 using an Octet Red96 (ForteBio). All reagents were prepared in kinetics buffer (PBS 0.01% BSA) at the indicated concentrations. His-tagged SARS-CoV-2 RBD was prepared at 8 $\mu\text{g ml}^{-1}$ and loaded on prehydrated anti-penta-HIS biosensors (Sartorius) for 2.5 min. Biosensors were then moved into a solution containing S2X259 mAb and association recorded for 7 min. A second association step was subsequently performed into S2X259 (as control), S309 and S2E12 mAbs solutions at 20 $\mu\text{g ml}^{-1}$ and recorded for 7 min. Response values were exported and plotted using GraphPad Prism 9.1.0.

Cell-surface mAb-mediated S1 shedding

CHO cells stably expressing wild-type SARS-CoV-2 S were resuspended in wash buffer (PBS 1% BSA, 2 mM EDTA) and treated with 10 $\mu\text{g ml}^{-1}$ TPCK-trypsin (Worthington Biochem) for 30 min at 37 °C. Cells were then washed and distributed into round bottom 96-well plates (90,000 cells per well). S2X259 was added to cells at 15 $\mu\text{g ml}^{-1}$ final concentration for 180 min at 37 °C. Cells were collected at different time points (5, 30, 60, 120 and 180 min), washed with wash buffer at 4 °C, and incubated with 1.5 $\mu\text{g ml}^{-1}$ secondary goat anti-human IgG, Fc fragment specific (Jackson ImmunoResearch) on ice for 20 min. Cells were washed and resuspended in wash buffer and analysed with ZE5 FACS (Biorad).

Measurement of Fc-effector functions

S2X259-dependent activation of human Fc γ RIIIa was performed with a bioluminescent reporter assay. ExpiCHO cells transiently expressing full-length wild-type SARS-CoV-2 S (target cells) or full-length prefusion stabilized SARS-CoV-2 S, which contains the 2P mutation and S1/S2 furin cleavage site mutation (RRARS to SGAG) as previously described¹², were incubated with different amounts of mAbs. After a 15-min incubation, Jurkat cells stably expressing Fc γ RIIIa receptor (V158 variant) or Fc γ RIIIa receptor (H131 variant) and NFAT-driven luciferase gene (effector cells)

were added at an effector to target ratio of 6:1 for FcγRIIIa and 5:1 for FcγRIIa. Signalling was quantified by the luciferase signal produced as a result of NFAT pathway activation. Luminescence was measured after 20 h of incubation at 37 °C with 5% CO₂ with a luminometer using the Bio-Glo Luciferase Assay Reagent according to the manufacturer's instructions (Promega).

In vivo mAb testing using a Syrian hamster model

KU LEUVEN R&D has developed and validated a SARS-CoV-2 Syrian hamster infection model^{36,39}. The SARS-CoV-2 (BetaCov/Belgium/GHB-03021/2020-EPI_ISL_109_407976|2020-02-03), which is most closely related to the prototypic Wuhan-Hu-1 2019-nCoV (GenBank accession number MN908947.3) strain, and B.1.351 (hCoV10519/Belgium/regal-1920/2021; EPI_ISL_896474, 2021-01-11) isolates used in this study were recovered from nasopharyngeal swabs taken from an asymptomatic returning traveller with RT-qPCR-confirmed SARS-CoV-2 infection in February 2020, and from a patient with respiratory symptoms in January 2021, respectively. A close relatedness with the prototypic Wuhan-Hu-1 2019 SARS-CoV-2 and with B.1.351 lineage was confirmed by sequencing and phylogenetic analysis. Infectious viruses were isolated by serial passaging on Vero E6 cells and passage 6 for SARS-CoV-2 Wuhan-Hu-1 and passage 2 for B.1.351 viruses were used for the study. The titre of the virus stock was determined by end-point dilution on Vero E6 cells by a previously published method⁵⁵. This work was conducted in the high-containment A3 and BSL3+ facilities of the KU Leuven Rega Institute (3CAPS) under licences AMV 30112018 SBB 219 2018 0892 and AMV 23102017 SBB 219 20170589 according to institutional guidelines.

Syrian hamsters were purchased from Janvier Laboratories and were housed per two in ventilated isolator cages (IsoCage N Bio-containment System, Tecniplast) with ad libitum access to food and water and cage enrichment (wood block). Housing conditions and experimental procedures were approved by the ethical committee of animal experimentation of KU Leuven (licence P065-2020). Hamsters were randomized and the sample size was chosen on the basis of previous experience with this animal model. To evaluate the prophylactic efficacy 6–10-week-old female hamsters were administered by intraperitoneal injection with S2X259 mAb at 1, 4, 5 and 25 mg kg⁻¹ or a combination of S2X259 and S309 mAbs (1 mg kg⁻¹ + 1 mg kg⁻¹) 48 h before intranasal infection with 1.89×10^6 TCID₅₀ in 50 µl inoculum. Blood samples were collected before infection and serum was obtained for pharmacokinetics analysis. To assess the therapeutic efficacy, 6–8-week-old Syrian hamsters were administered by intraperitoneally with isotype control antibody (20 mg kg⁻¹) or S2X259 mAb (at 20, 10 or 5 mg kg⁻¹) 24 h after infection with SARS-CoV-2. Blood samples were collected when hamsters were killed, and serum was obtained for pharmacokinetics analysis. Hamsters were monitored for appearance, behaviour and weight. At day 4 after infection, hamsters were euthanized by intraperitoneal injection of 500 µl Dolethal (200 mg ml⁻¹ sodium pentobarbital, Vétoquinol SA). Lungs were collected, homogenized using bead disruption (Precellys) in 350 µl RLT buffer (RNeasy Mini kit, Qiagen) and centrifuged (10,000 r.p.m., 5 min, 4 °C) to pellet the cell debris. RNA was extracted using a NucleoSpin kit (Macherey-Nagel) according to the manufacturer's instructions. RT-qPCR was performed on a LightCycler96 platform (Roche) using the iTaq Universal Probes One-Step RTqPCR kit (BioRad) with N2 primers and probes targeting the nucleocapsid³⁶. Standards of SARS-CoV-2 cDNA (IDT) were used to express viral genome copies per mg tissue or per ml serum. To quantify infectious SARS-CoV-2 particles, end-point titrations were performed on confluent Vero E6 cells in 96-well plates. The hamster-derived samples for RNA quantification and viral load titration were run by technicians who were blinded to treatment group of the analysed samples. Viral titres were calculated by a previously published method⁵⁵ and were expressed as TCID₅₀ per mg tissue.

Evaluation of escape mutants using deep-mutational scanning

A previously described deep-mutational scanning approach⁵⁶ was used to identify RBD mutations that escape S2X259 binding. In brief, duplicate libraries containing virtually all possible amino acid changes compatible with ACE2 binding and RBD folding within the Wuhan-Hu-1 SARS-CoV-2 RBD sequence were expressed on the surface of yeast^{29,56}. Libraries were labelled at 59 ng ml⁻¹ S2X259 antibody (the 90% effective concentration for binding to yeast-displayed SARS-CoV-2 RBD determined in isogenic pilot binding experiments), and fluorescence-activated cell sorting (FACS) was used to select RBD⁺ cells that exhibit reduced antibody binding, as previously described^{56,57}. Libraries were sequenced before and after selection to determine per-mutation escape fractions, as previously described⁵⁷. Experiments were performed in duplicate with independently generated mutant libraries, and we report the average mutant escape fraction across the duplicates. Raw escape fractions are available on GitHub: https://github.com/jbloombab/SARS-CoV-2-RBD_MAP_Vir_mAbs/blob/main/results/supp_data/s2X259_raw_data.csv. Detailed steps of analysis and code for the deep mutational scanning selections are available on GitHub: https://github.com/jbloombab/SARS-CoV-2-RBD_MAP_Vir_mAbs.

Selection of SARS-CoV-2 mAb escape mutants

VSV-SARS-CoV-2 chimera was used to select for SARS-CoV-2 S mAb-resistant mutants, as previously described^{30,31}. In brief, mAb-resistant mutants were recovered by plaque isolation on Vero cells with the indicated mAb in the overlay. The concentration of mAb in the overlay was determined by neutralization assays at a MOI of 100. Escape clones were plaque-purified on Vero cells in the presence of mAb, and plaques in agarose plugs were amplified on MA104 cells with the mAb present in the medium. Viral stocks were amplified on MA104 cells at an MOI of 0.01 in Medium 199 containing 2% FBS and 20 mM HEPES pH 7.7 (Millipore Sigma) at 34 °C. Viral supernatants were collected upon extensive cytopathic effect and clarified of cell debris by centrifugation at 1,000g for 5 min. Aliquots were maintained at -80 °C. Viral RNA was extracted from VSV-SARS-CoV-2 mutant viruses using RNeasy Mini kit (Qiagen), and S was amplified using OneStep RT-PCR Kit (Qiagen). The mutations were identified by Sanger sequencing (GENEWIZ). Their resistance was verified by subsequent virus infection in the presence or absence of antibody. In brief, Vero cells were seeded into 12-well plates for overnight. The virus was serially diluted using DMEM and cells were infected at 37 °C for 1 h. Cells were cultured with an agarose overlay in the presence or absence of mAb at 34 °C for 2 days. Plates were scanned on a biomolecular imager and expression of eGFP is shown at 48 h after infection.

Crystallization, data collection, structure determination and analysis

Crystals of the SARS-CoV-2-RBD-S2X259-S2H97 Fab complex were obtained at 20 °C by sitting drop vapour diffusion. A total of 200 nl complex at 5.7 mg ml⁻¹ was mixed with 200 nl mother liquor solution containing 0.12 M monosaccharides mix, 20% (v/v) ethylene glycol, 10% (w/v) PEG 8000, 0.1 M Tris (base)/bicine pH 8.5, 0.02 M sodium chloride, 0.01 M MES pH 6 and 3% (v/v) Jeffamine ED-2003. Crystals were flash-frozen in liquid nitrogen. Data were collected at Beamline 9-2 of the Stanford Synchrotron Radiation Lightsource facility. Data were processed with the XDS software package⁵⁸ for a final dataset of 2.65 Å in space group P21. The RBD-S2X259-S2H97 Fab complex structure was solved by molecular replacement using Phaser⁵⁹ from a starting model consisting of SARS-CoV-2 RBD (PDB code 7JX3) and homology models of the S2X259 and S2H97 Fabs built using the Molecular Operating Environment (MOE) software package from the Chemical Computing Group (<https://www.chemcomp.com>). Several subsequent rounds of model building and refinement were performed using Coot⁶⁰, ISOLDE⁶¹,

Refmac5⁶² and MOE (<https://www.chemcomp.com>), to arrive at a final model for the ternary complex.

Cryo-EM sample preparation, data collection and data processing

Recombinantly expressed and purified Fab S2X259 and SARS-CoV-2 S hexaprotein were incubated at 1 mg ml⁻¹ with a 1.2 molar excess of Fab at 4 °C during 1 h. Three microlitres of the complex mixture were loaded onto freshly glow discharged R 2/2 UltrAuFoil grids (200 mesh) or lacey grids covered with a thin layer of manually added carbon, before plunge-freezing using a vitrobot MarkIV (ThermoFisher Scientific) with a blot force of 0 and 7–7.5 s blot time (for the UltrAuFoil grids) or with a blot force of –1 and 2.5 s blot time (for the lacey thin carbon grids) at 100% humidity and 21 °C.

Data were acquired on a FEI Titan Krios transmission electron microscope operated at 300 kV and equipped with a Gatan K2 Summit direct detector and Gatan Quantum GIF energy filter, operated in zero-loss mode with a slit width of 20 eV. Automated data collection was carried out using Leginon⁶³ at a nominal magnification of 130,000× with a super-resolution pixel size of 0.525 Å. The dose rate was adjusted to 8 counts per pixel per s, and each movie was fractionated in 50 frames of 200 ms. Two datasets were collected from UltrAuFoil grids with the stage tilted 30° and 55° to circumvent particle preferential orientation. The third dataset was collected on lacey grids covered with a thin layer of carbon. The three datasets, with a total of 6,786 micrographs, were collected with a defocus range of between –0.8 and –2 µm. For each dataset, movie frame alignment, estimation of the microscope contrast-transfer function parameters, particle picking and extraction were carried out using Warp⁶⁴. Particle images were extracted with a box size of 800 pixels² and binned to 400 yielding a pixel size of 1.05 Å. The three datasets were merged and two rounds of reference-free 2D classification were performed using cryoSPARC⁶⁵. Subsequently, one round of 3D classification with 50 iterations, using PDB code 6VXX as initial model, was carried out using Relion^{66,67} without imposing symmetry. Three-dimensional refinements were carried out using nonuniform refinement⁶⁸. Particle images from each dataset were subjected to Bayesian polishing⁶⁹ using Relion before merging them to perform another round of nonuniform refinement in cryoSPARC followed by per-particle defocus refinement and again nonuniform refinement. To improve the density of the S–S2X259 interface, the particles were symmetry-expanded and subjected to a Relion focus 3D classification without refining angles and shifts using a soft mask encompassing the RBD and S2X259 variable domains. Particles belonging to classes with the best-resolved local density were selected and subjected to local refinement using cryoSPARC. Local resolution estimation, filtering and sharpening were carried out using CryoSPARC. Reported resolutions are based on the gold-standard Fourier shell correlation of 0.143 criterion and Fourier shell correlation curves were corrected for the effects of soft masking by high-resolution noise substitution⁷⁰.

Cryo-EM model building and analysis

UCSF Chimera⁷¹ and Coot⁶⁰ were used to fit atomic models (PDB code 6VXX or 6VYB) into the cryo-EM maps and the Fab variable domains were manually built. S2E12 was built in the locally refined map and subsequently validated using the Fab crystal structure. Models were refined and relaxed using Rosetta using both sharpened and unsharpened maps^{72,73}. Validation used Phenix⁷⁴, Molprobity⁷⁵ and Privateer⁷⁶. Figures were generated using UCSF ChimeraX⁷⁷.

Material availability

Materials generated in this study can be available on request and may require a material transfer agreement. The SARS-CoV-2 RBD mutant libraries (no. 1000000172) and unmutated parental plasmid (no. 166782) are available on Addgene

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this paper.

Data availability

All datasets generated and information presented in the study are available from the corresponding authors on reasonable request. The cryo-EM structure data and model are available from the PDB (and Electron Microscopy Data Bank) under accession codes 7RA8 (EMD-24347) for the global refinement of the SARS-CoV-2 S and S2X259 complex and 7RAL (EMD-24365) for the local refinement of the SARS-CoV-2 S RBD and S2X259 Fab variable domains. The X-ray structure data and model has been deposited at PDB code 7M7W for RBD–S2H97–S2X259. Interactive escape maps and structural visualizations can be found at: https://jbloombio.github.io/SARS-CoV-2-RBD_MAP_Vir_mAbs/. Raw Illumina sequencing data from deep mutational scanning experiments are available on NCBI SRA, BioSample SAMN18315604 (SARS-CoV-2 mutant selection data). A complete table of deep-mutational scanning antibody escape fractions is provided on GitHub: https://github.com/jbloombio/SARS-CoV-2-RBD_MAP_Vir_mAbs/blob/main/results/supp_data/all_antibodies_raw_data.csv.

Code availability

A repository containing all code, analysis and summary notebooks for the analysis of the SARS-CoV-2 deep-mutational scanning escape selections is available on GitHub at https://github.com/jbloombio/SARS-CoV-2-RBD_MAP_Vir_mAbs.

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Author contributions Experiment design was performed by M.A.T., N.C., T.N.S., R.M., A.C.W., S.P.J.W., J.D.B., D.C., D.V. and M.S.P.; E.C. and F.B. recruited donors and collected samples; R.M., F.Z., A.D.M., D.P. and M.B. processed PBMC samples; M.A.T., N.C., S.J., A.C.W., J.E.B., K.C., N.S., C.S., I.G.-S. and E.C. performed protein expression and purification; R.M., F.Z., A.D.M., D.P., M.B. and C.S.F. isolated mAbs; M.A.T., R.M., A.C.W., F.Z., D.P., M.B., M.P.H., F.A.L., E.D. Jr, H.K., M.M.-R., J.Z., S.K.Z. and C.H.-D. performed binding and neutralization assays; A.C.W., F.Z., A.D.M. and L.E.R. undertook bilayer interferometry and surface plasmon resonance assays; M.A.T., Z.W. and D.V. collected and processed cryo-EM data, and built the models; N.C. and G.S. performed crystallization experiments and determined the X-ray structure; D.P. and A.D.M. performed the ACE2 binding inhibition and S1 shedding; B.G. evaluated effector functions; T.N.S., A.A. and J.D.B. performed deep-mutational scanning; J.D.I., I.B. and A.T. undertook bioinformatic analysis of virus diversity and variants; Z.L. and S.P.J.W. selected and sequenced escape mutants; R.A., S.-Y.C.F., E.V., M.A.S., F.B., J.N., D.C. and M.S.P. performed hamster model experiments and data analysis; M.A.T., N.C., T.N.S., R.M., A.C.W., J.D.B., D.C., D.V. and M.S.P. analysed data; M.A.T., N.C., T.N.S., R.M., J.N., A.T., G.S., H.W.V., S.P.J.W., J.D.B., D.C., D.V. and M.S.P. wrote the manuscript; F.A.L., L.E.R., C.H.-D., M.A.S., F.B., E.C., J.N., A.T., G.S., H.W.V., S.P.J.W., J.D.B., D.C., D.V. and M.S.P. supervised the project; and J.D.B. and D.V. acquired funding.

Competing interests N.C., R.M., F.Z., S.J., J.D.I., A.D.M., D.P., M.B., I.B., M.H., F.A.L., L.E.R., E.D. Jr, H. K., M.M.-R., J.Z., K.C., N.S., C.S., E.V., I.G.-S., C.S.F., C.H.-D., M.A.S., F.B., E.C., A.T., G.S., H.W.V., D.C. and M.S.P. are employees of Vir Biotechnology Inc. and may hold shares in Vir Biotechnology Inc. D.C. is currently listed as an inventor on multiple patent applications, which disclose the subject matter described in this Article. The laboratory of J.N. has received sponsored research agreements from Vir Biotechnology Inc. After the submission of the initial version of this study, J.D.B. began consulting for Moderna on viral evolution and epidemiology. J.D.B. has the potential to receive a share of IP revenue as an inventor on a Fred Hutchinson Cancer Research Center-licensed technology and patent (application WO2020006494) related to deep-mutational scanning of viral proteins. H.W.V. is a founder of PierianDx and Casma Therapeutics. Neither company provided funding for this work or is performing related work. D.V. is a consultant for Vir Biotechnology Inc. The laboratory of D.V. has received a sponsored research agreement from Vir Biotechnology Inc. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

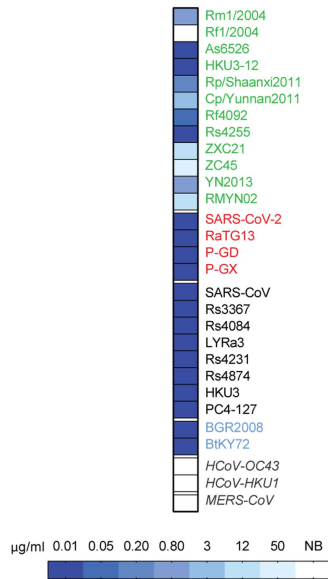
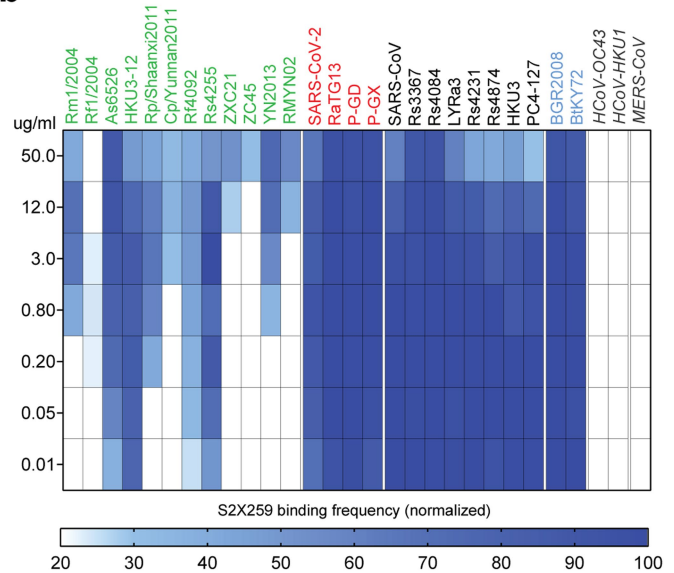
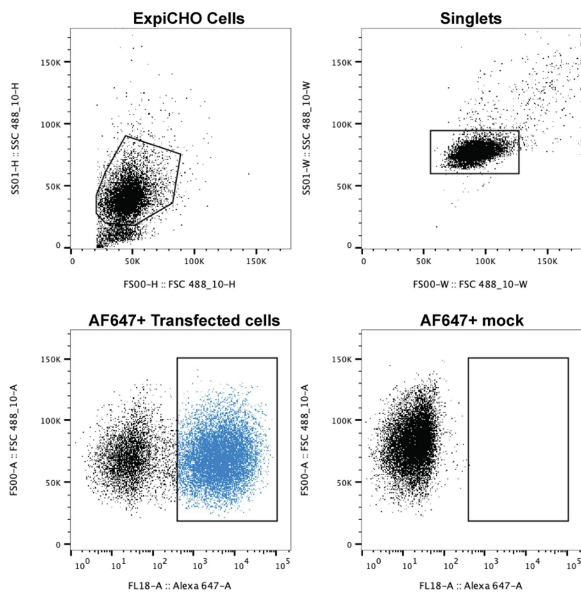
Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41586-021-03817-4>.

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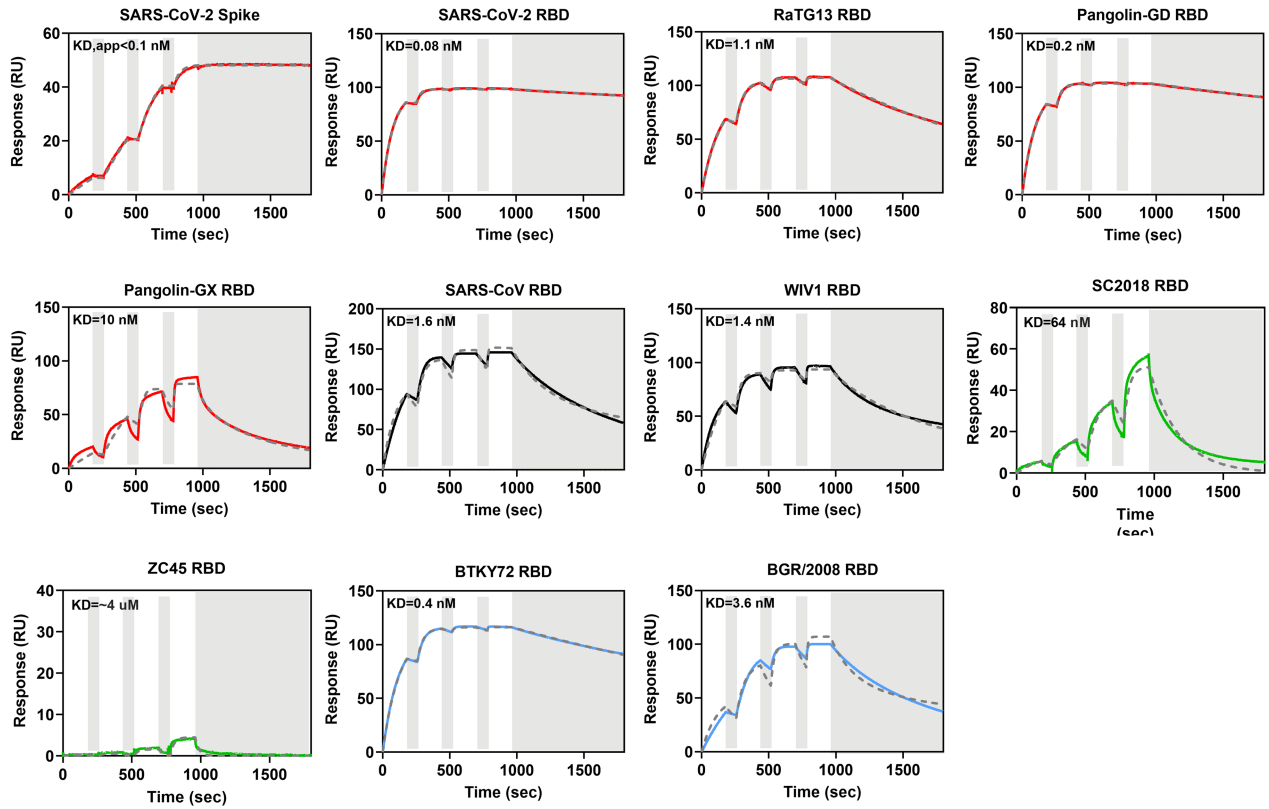
	S2X259	S2X35	S2E12	S309
SARS-CoV	9.97	25.58		15.52
WIV1	11.04			17.06
SARS-CoV-2	31.96	28.20	5.82	10.92
RatG13	21.88	36.81	7.50	17.93
PangGD	28.45	33.00	6.91	15.88
Anlong112	1281.00			
YN2013	12.86	348.20		
SC2018	11.63			
SX2011	26.50			239.80
ZC45	539.00			
BiKY72	22.18	27.24		
BGR2008	10.28	89.56		22.09

EC50 ng/ml 1-50 50-1000 1000-5000 No binding

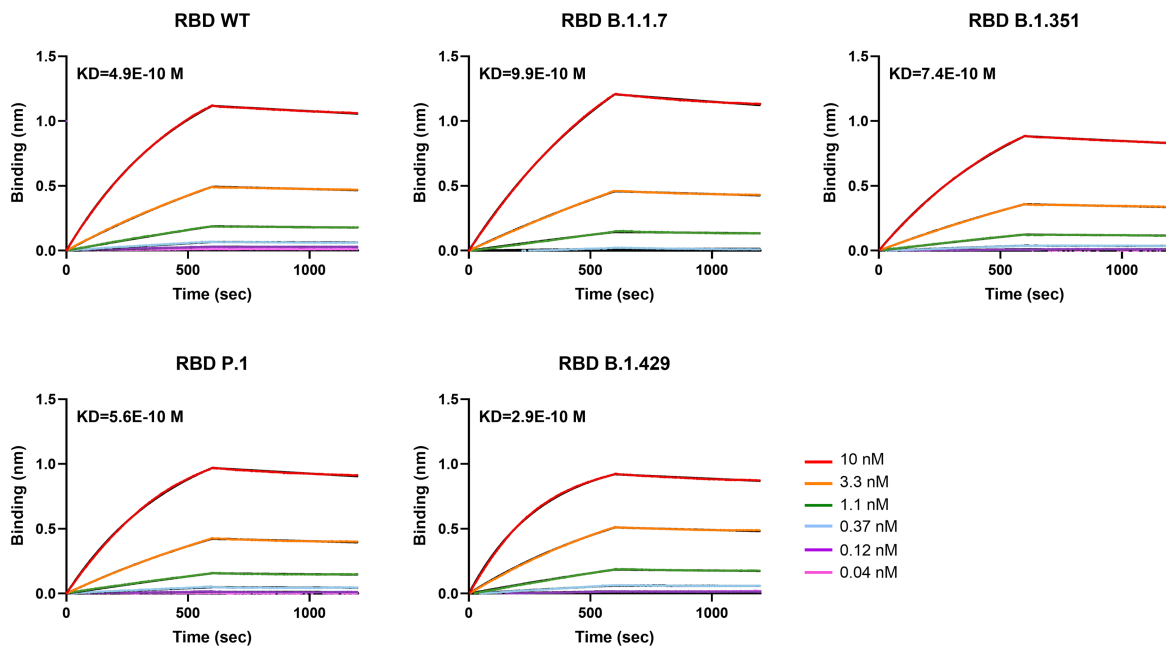
Extended Data Fig. 1 | Breadth of S2X259 mAb across the Sarbecovirus subgenus. a, Flow cytometry analysis of S2X259 cross-reactivity with a panel of 26 S glycoproteins representative of sarbecovirus clades 1a, 1b, 2 and 3. The colours represent the lowest concentration of mAb at which binding was observed. **b,** FACS binding of S2X259 to a panel of sarbecovirus S glycoproteins transiently expressed in ExpiCHO cells. Results represent the binding

frequency normalized using the ratio between mean fluorescence intensities for S2X259 and an expression control mAb⁷⁸. **c,** FACS gating strategy used to assess binding of S2X259 or of the expression control mAb. **d,** ELISA binding of site-I-targeting S2E12²¹, site-II-targeting S2X259 and S2X35¹⁷, and site-IV-targeting S309²² mAbs to RBDs spanning the different clades of the Sarbecovirus subgenus. EC₅₀s are reported. *n* = 2 independent experiments.

a

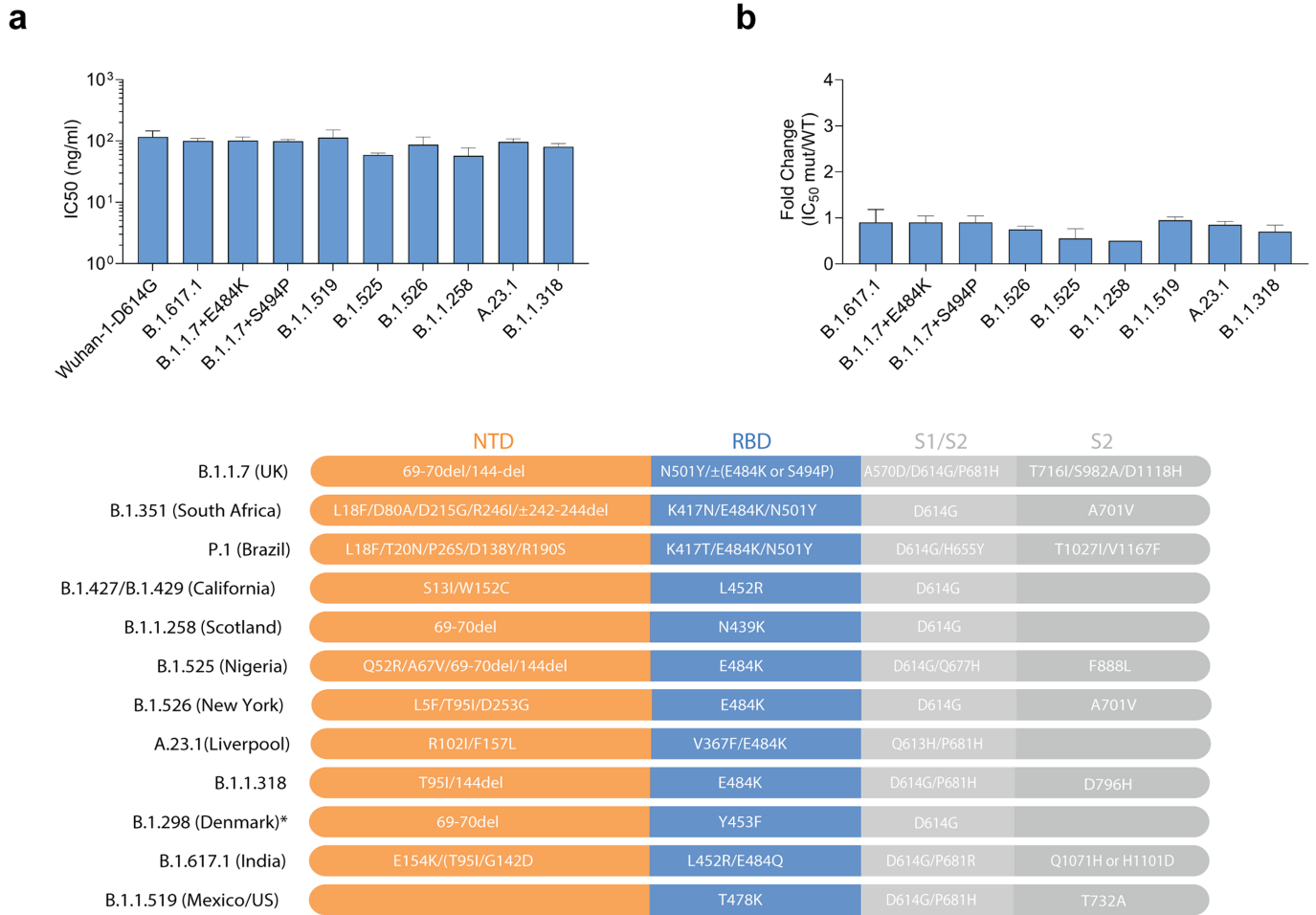


b



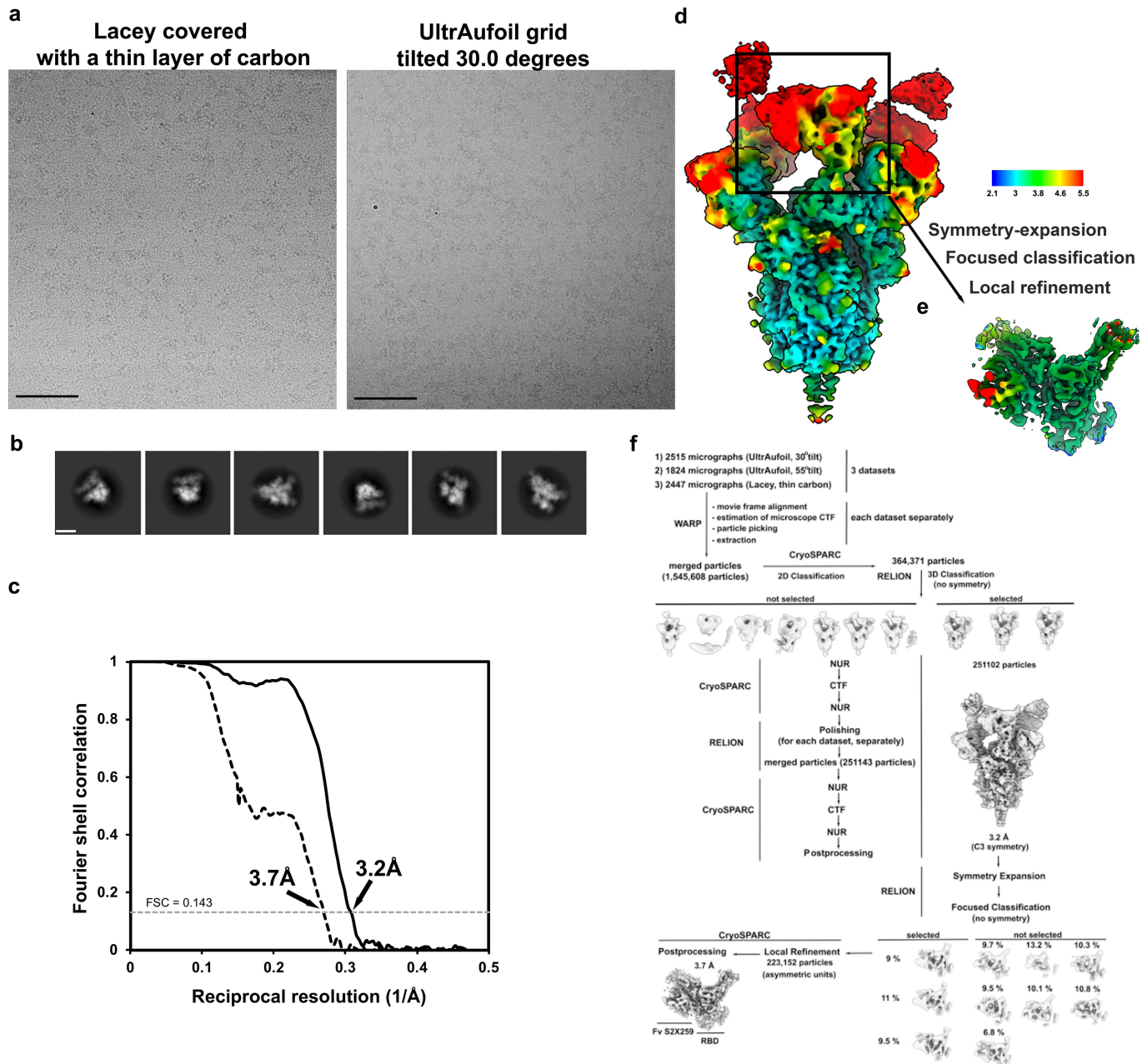
Extended Data Fig. 2 | S2X259 Fab binding to recombinant sarbecovirus RBDs, prefusion SARS-CoV-2 S ectodomain trimer and RBD variants. **a**, S or RBD antigens were captured on the sensor chip surface and binding to S2X259 Fab at 11, 33, 100 and 300 nM was monitored successively, in single-cycle kinetics format, by surface plasmon resonance. All data have been fit to a 1:1 binding model (black dashed line) and the equilibrium dissociation constant (K_D) is reported. For the S-binding data, we report an apparent K_D ($K_{D,app}$) as

kinetics are affected by conformational dynamics between open and closed RBD states. The colouring scheme matches the phylogenetic tree in Fig. 1a. **b**, Bi-layer interferometry binding analysis of the S2X259 Fab to wild-type or variant-of-concern SARS-CoV-2 biotinylated RBDs immobilized at the surface of SA biosensors. The data are coloured according to the key and fits to a 1:1 binding model are shown as black lines. Equilibrium dissociation constants (K_D) are reported above each plot.



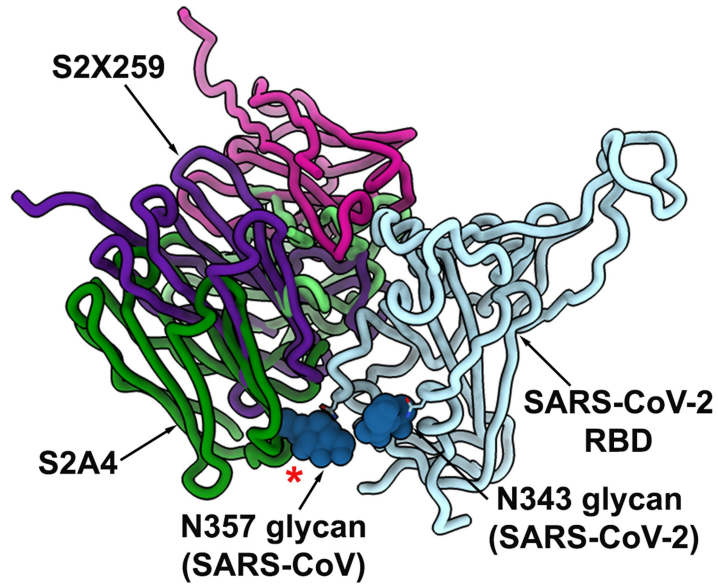
Extended Data Fig. 3 | S2X259 neutralizing activity is not affected by mutations present in circulating SARS-CoV-2 variant of concerns and recent lineages. a, b, Neutralization by S2X259 of SARS-CoV-2 VSV pseudotyped circulating variants shown as IC_{50} (a) and IC_{50} fold change as

compared to Wuhan-Hu-1 virus with D614G substitution (b). Schematic of the SARS-CoV-2 S and the mutation landscape in each variant is shown beneath the panels. Del, deletion. Asterisks denotes virus that was tested as single RBD mutant in Fig. 1e.



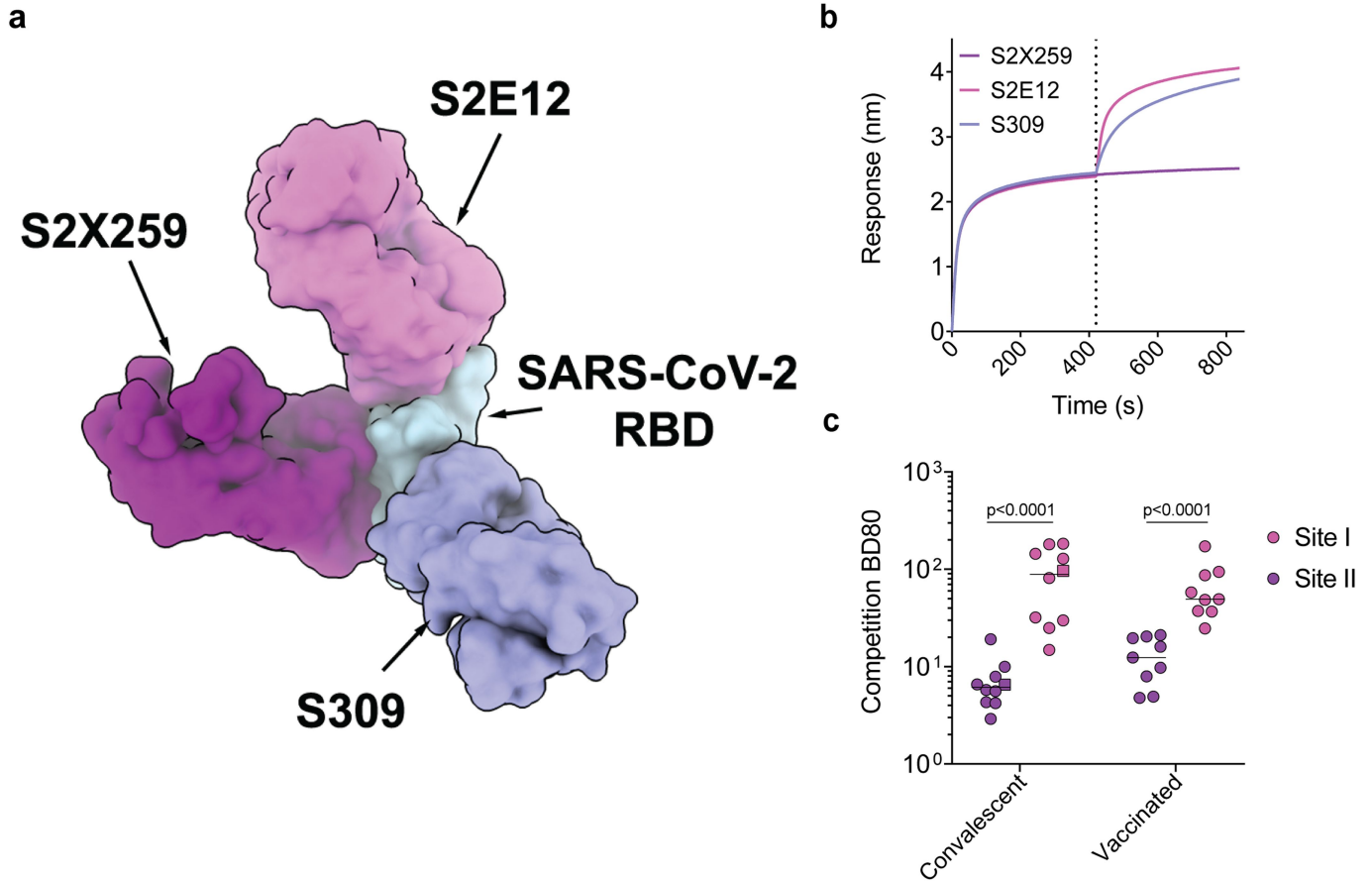
Extended Data Fig. 4 | Cryo-EM data processing and validation of S2X259-bound SARS-CoV-2 S. **a, b.** Representative electron micrographs (**a**) and class averages (**b**) of SARS-CoV-2 S in complex with the S2X259 Fab. Scale bars, 500 Å (**a**), 100 Å (**b**). **c.** Gold-standard Fourier shell correlation curves for the S trimer bound to three S2X259 Fabs (solid black line) and the locally

refined reconstruction of the RBD-S2X259 variable domains (dashed black line). The 0.143 cut-off is indicated by a horizontal dashed grey line. **d, e.** Local resolution map for the open S trimer bound to three S2X259 Fabs (**d**) and the locally refined reconstruction of the RBD-S2X259 variable domains (**e**). **f.** Cryo-EM data processing flowchart.



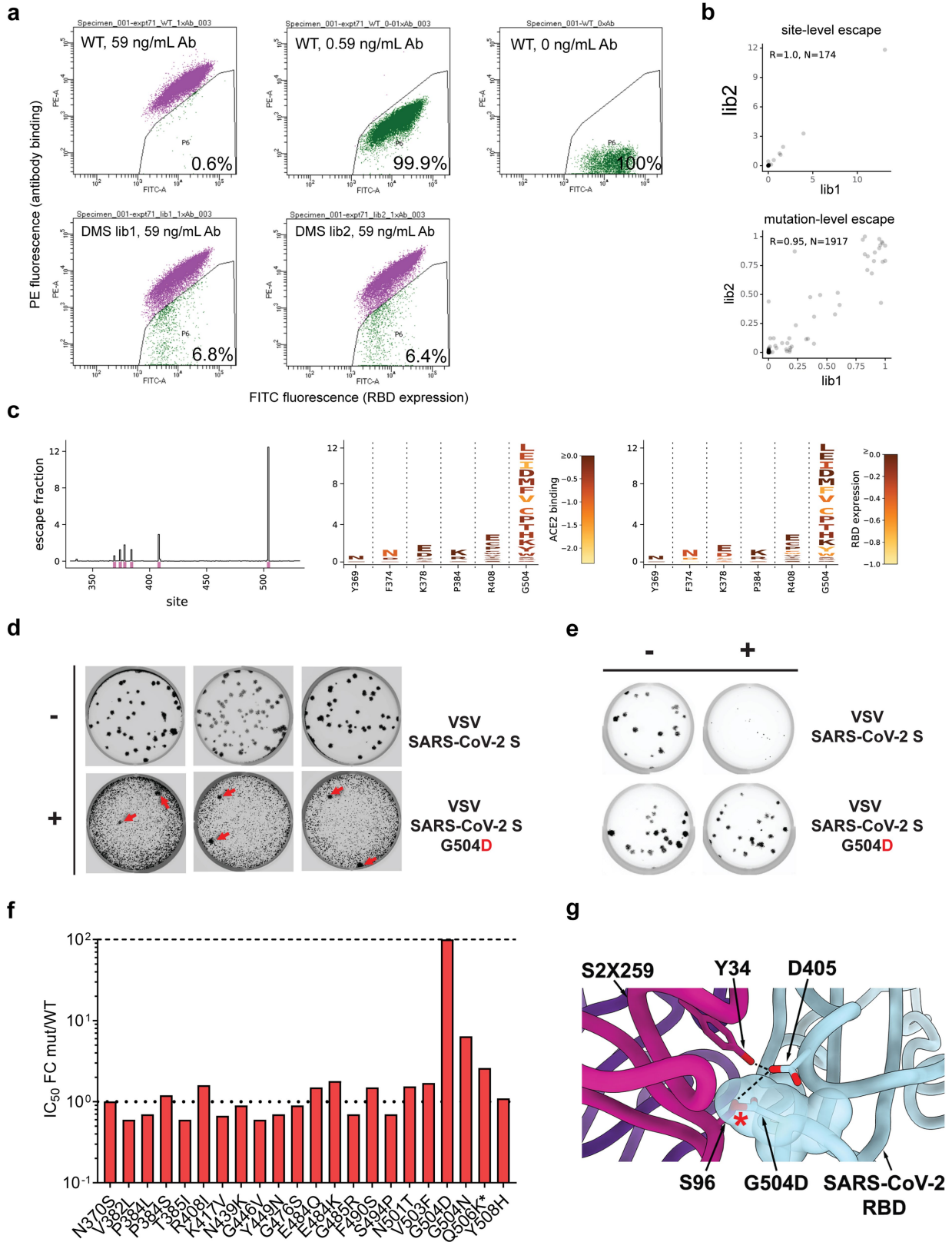
Extended Data Fig. 5 | The S2X259 angle of approach for binding to the SARS-CoV-2 RBD allows S2X259 to circumvent the SARS-CoV N357 glycan present in all sarbecovirus RBDs except SARS-CoV-2. Ribbon diagram showing a superimposition of the S2X259-bound and S2A4-bound (PDB code 7JVA) SARS-CoV-2 RBD¹⁷. The SARS-CoV glycan at position N357 was modelled

on the basis of the S230-bound SARS-CoV S structure (PDB code 6NB6³³) and is predicted to sterically hinder S2A4 binding (red star) but not S2X259 binding. The mAb light and heavy chains are coloured magenta and purple (S2X259) or light and dark green (S2A4), respectively. N-linked glycans are rendered as blue spheres.



Extended Data Fig. 6 | S2X259 can bind to the SARS-CoV-2 RBD in presence of site-I and -IV-targeting mAbs. **a**, View of site-I-targeting S2E12²¹ (pink), site-II-targeting S2X259 (magenta) and site-IV-targeting S309²² (purple) mAb bound to the SARS-CoV-2 RBD (light blue). **b**, Competition binding assays for S2X259 versus site-I-targeting S2E12 and site-IV-targeting S309 mAbs on SARS-CoV-2 RBD as measured by biolayer interferometry. One independent experiment out of two is shown. **c**, Competition ELISA (blockade-of-binding) between site-I-targeting S2H14¹⁷ or site-II-targeting S2X259 and sera or plasma

from individuals who had recovered from COVID-19 ($n=10$ biological samples) and vaccinated individuals ($n=9$ biological samples). Each plot shows the magnitude of inhibition of binding to immobilized RBD in the presence of each mAb, expressed as reciprocal sera or plasma dilution blocking 80% of the maximum binding response. The individual from whom S2X259 mAb was isolated is shown as a square. Statistical analysis was performed using two-tailed Mann-Whitney test.

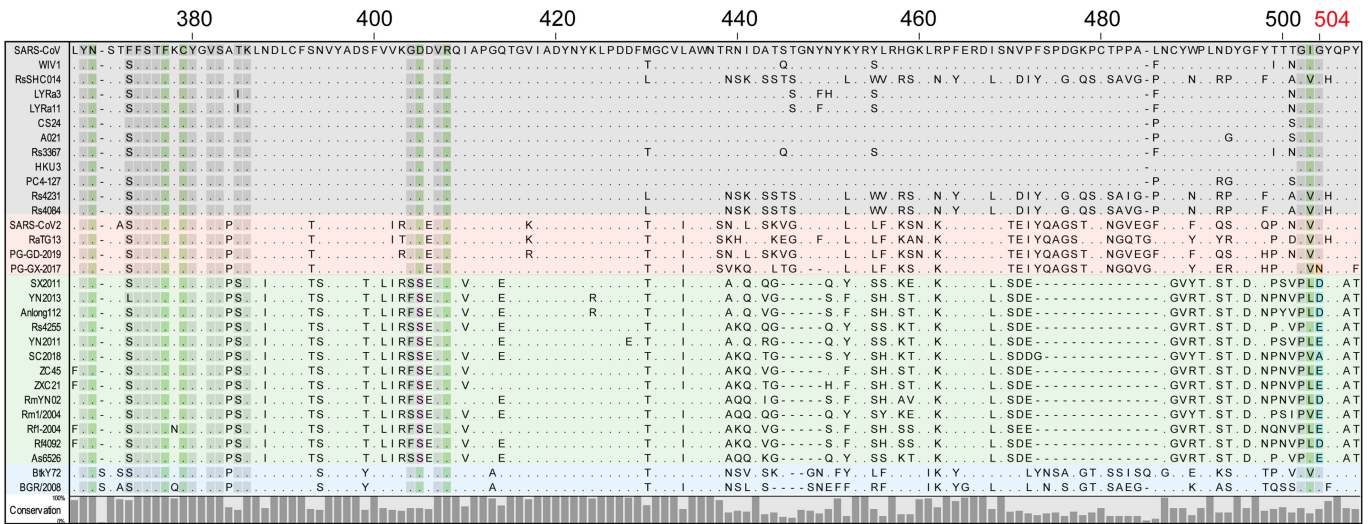


Extended Data Fig. 7 | See next page for caption.

Article

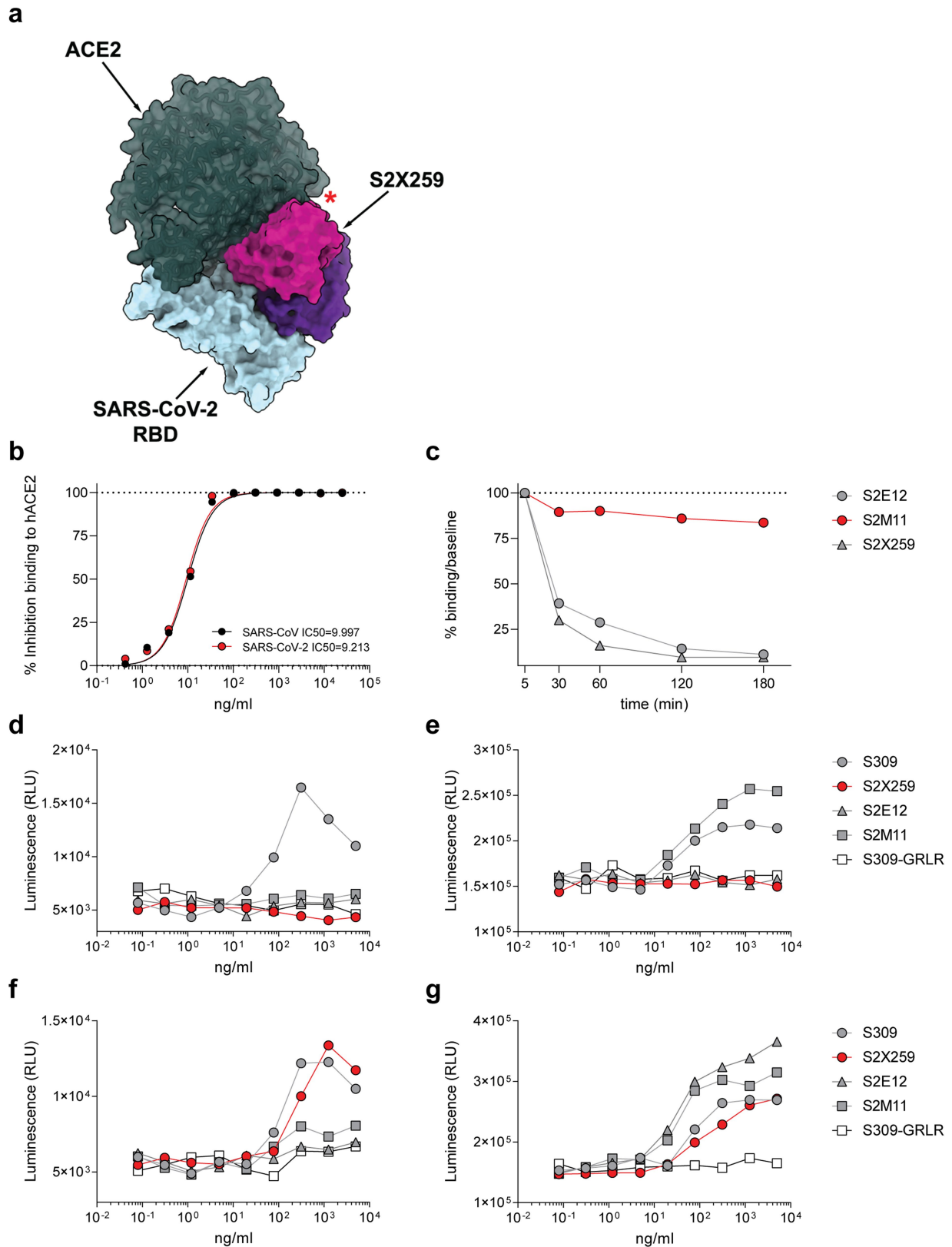
Extended Data Fig. 7 | S2X259 has a high barrier for the emergence of resistance mutants. **a**, FACS gates used in deep-mutational scanning to select escape variants. Top, yeast controls expressing unmutated SARS-CoV-2 RBD labelled at relevant S2X259 concentrations for setting of selection gates. Bottom, fraction of cells in SARS-CoV-2 mutant libraries falling into the antibody-escape bin. **b**, Correlation in site-level (top, sum of escape fractions for mutations at a site) and mutation-level (bottom) escape between independently generated and assayed RBD mutant libraries. **c**, Line plot of escape mutants along all positions in the SARS-CoV-2 RBD (left). Pink lines indicate sites that escape S2X259 binding illustrated at the mutation-level in logoplots (right). In logoplots, the height of a letter scales with its escape fraction. Letters are coloured according to their deleterious consequences for ACE2 binding (middle) or RBD expression (right) as determined in previous

deep-mutational scanning experiments²⁹. **d**, Plaque assay using VSV-SARS-CoV-2 chimeric virus on Vero cells with no mAb (-) or S2X259 (+) in the overlay to isolate escape mutants (red arrow). Data are representative of three independent experiments. **e**, Plaque assays performed to validate the VSV-SARS-CoV-2 G504D mutant in Vero cells in the presence (+) or absence (-) of S2X259 in the overlay. Representative image of two independent experiments is shown. **f**, S2X259 in vitro neutralizing activity against SARS-CoV-2 S VSV pseudotyped mutants. For each mutant the fold change of the IC_{50} geometric mean versus SARS-CoV-2 SD614G is reported. *Q506K displayed a tenfold reduction in viral entry in comparison to the other mutants. Results from two independent experiments are reported. **g**, Zoomed-in view of the S2X259-RBD interface showing that the G504D substitution would disrupt mAb binding owing to steric hindrance (indicated with a red star).



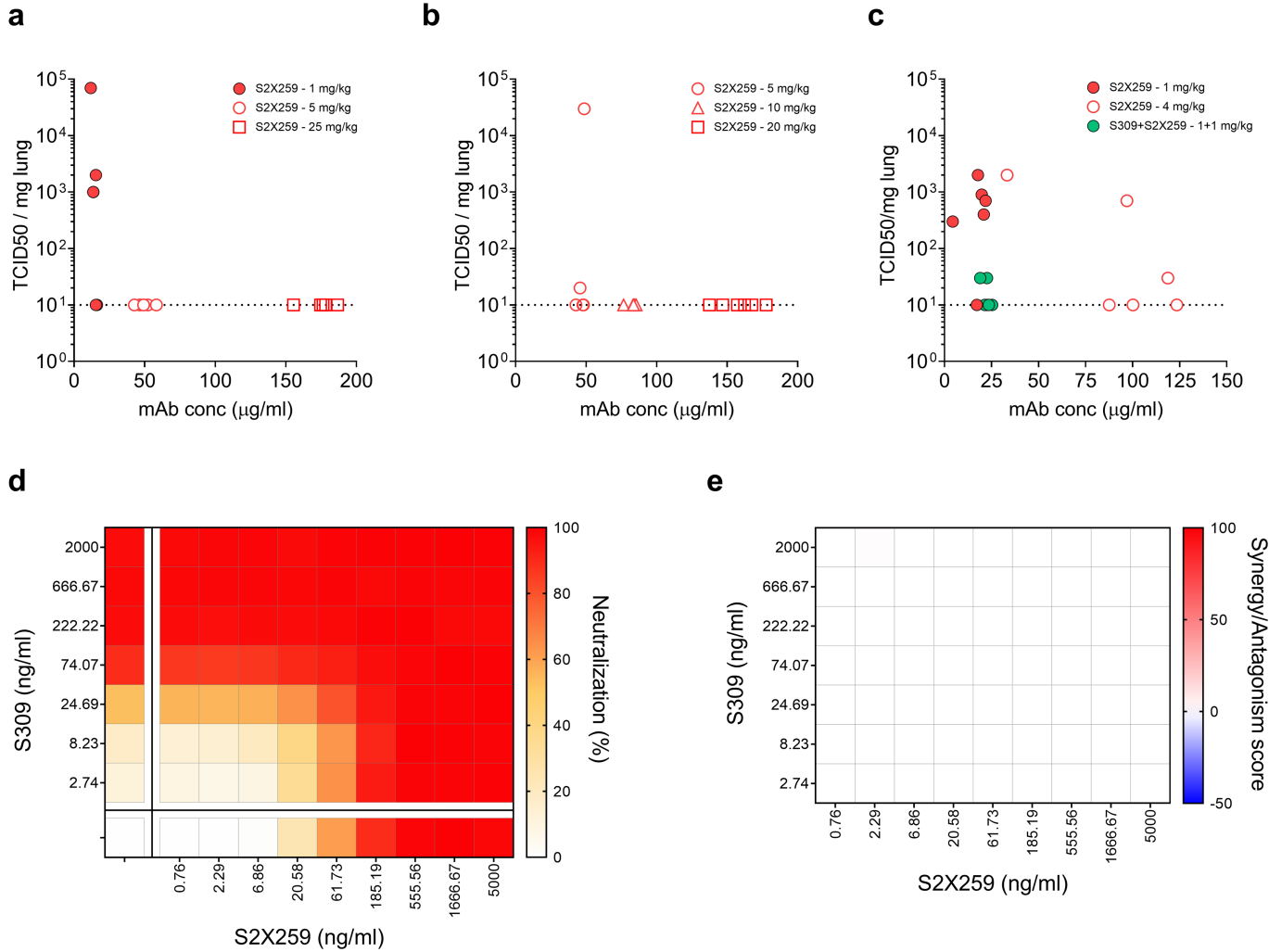
Extended Data Fig. 8 | S2X259 epitope conservation across sarbecovirus clades. Protein sequence alignment of representative sarbecovirus RBDs with strictly conserved residues shown as dots. Overall conservation is represented as a bar plot at the bottom of the figure. Residue positions are based on SARS-CoV-2. Residues determined to be most important for S2X259 binding

are denoted in dark green. Substitutions at positions D405 and G504 are indicated in pink and blue or orange, respectively. Additional residues representing the extended epitope are denoted in grey. Different clades within the *Sarbecovirus* subgenus are overlaid in grey (clade 1a), red (clade 1b), green (clade 2) and light blue (clade 3).



Extended Data Fig. 9 | Inhibition of ACE2 engagement, S1 subunit shedding and activation of Fc γ RIIa and Fc γ RIIIa in vitro. a, S2X259 (purple and pink) and ACE2 (dark green) bind partially overlapping binding sites on the SARS-CoV-2 RBD (blue). **b**, Pre-incubation of serial dilutions of S2X259 with SARS-CoV-2 (red) or the SARS-CoV (black) RBDs prevents binding to immobilized human ACE2 (hACE2) ectodomain in ELISA. **c**, mAb-mediated S1 subunit shedding from cell-surface-expressed SARS-CoV-2S as determined by flow cytometry. S2E12 was included as positive control and S2M11 was included

as negative control. **d, e**, NFAT-driven luciferase signal induced in Jurkat cells stably expressing Fc γ RIIa H131 (**d**) variant or Fc γ RIIIa V158 (**e**) variant by S2X259 binding to full-length wild-type SARS-CoV-2S on ExpiCHO target cells. **f, g**, NFAT-driven luciferase signal induced in Jurkat cells stably expressing Fc γ RIIa H131 (**f**) or Fc γ RIIIa V158 (**g**) variants by S2X259 binding to uncleavable full-length prefusion stabilized SARS-CoV-2S (unable to release the S1 subunit) transiently expressed in ExpiCHO cells. SE12, S2M11, S309 and S309-GRLR mAbs are included as controls.



Extended Data Fig. 10 | Correlation between mAb concentration and infectious virus in vivo and in vitro neutralizing activity of S2X259 + S309 antibody cocktail. a-c. Infectious virus titres in the lungs at 4 days after infection plotted as a function of serum mAb concentrations before infection (day 0) with prototypic SARS-CoV-2 (a) and SARS-CoV-2 B.1.351 (c) in prophylactic setting or at 4 days after infection with prototypic

SARS-CoV-2 (b) in therapeutic setting. **d, e.** SARS-CoV-2 B1.351 VSV-based pseudotype neutralization (d) and synergy score (e) measured combining S2X259 and S309 mAbs. Synergy was calculated using MacSynergyII (synergy score: 0.98, antagonism score: 0). Results represent four replicates of one independent experiment.

Reporting Summary

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Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

As described in Methods:

- * protein purification monitored by UNICORN software version 5.11
- * SPR binding assays collected and analyzed using Biacore Evaluation and Biacore Insight software
- * FACSDiva (version 6.1.3) and Everest 3.0 software for FACS data collection and cell sorting
- * Identification of escape mutants in virological assay was performed by Sanger sequencing (GENEWIZ)

Data analysis

Repository containing all code, analysis, and summary notebooks for the analysis of the SARS-CoV-2 deep mutational scanning escape selections available on GitHub: https://github.com/jbloomlab/SARS-CoV-2-RBD_MAP_Vir_mAbs. This code uses packages and versions as described in the Methods for various steps of computation.

Binding, Neutralization assays analyzed using PRISM (Version 9.1.0) as described in Methods.

Crystallography data analysis performed with XDS (VERSION Jan 31, 2020 BUILT=20200417), Coot (v. 0.9.5), ChimeraX (v. 1.1)/ISOLDE (v. 1.1), Refmac5 (v. 5.8.0267), and MOE (v. 2019.0102)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

- All datasets generated and information presented in the study are available from the corresponding author on reasonable request.
- CryoEM structure data and model are available with accession code PDB 7RA8 (EMD-24347) and PDB 7RAL (EMD-24365)
- The X-ray structure data and model has been deposited with accession code PDB 7M7W for RBD-S2H97-S2X259.
- Interactive escape maps and structural visualizations can be found at: https://jbloomlab.github.io/SARS-CoV-2-RBD_MAP_Vir_mAbs/.
- Raw Illumina sequencing data from deep mutational scanning experiments are available on NCBI SRA, BioSample SAMN18315604 (SARS-CoV-2 mutant selection data).
- Complete table of deep mutational scanning antibody escape fractions is provided on GitHub: https://github.com/jbloomlab/SARS-CoV-2-RBD_MAP_Vir_mAbs/blob/main/results/supp_data/all_antibodies_raw_data.csv.

Field-specific reporting

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- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	For in vivo studies the sample size was chosen based on prior experience with this animal model.
Data exclusions	Lack of detection of the human monoclonal antibody in the hamster sera at 48 post-administration (prophylactic setting) or at 4 days post-infection (therapeutic setting) was considered a criteria to identify animals that were not properly administered and to exclude them from the analysis
Replication	Experimental assays were performed in biological duplicate or triplicate (or more) according to or exceeding standards in the field. We conducted all neutralization and antibody functional assays in biological duplicate, triplicate, or more, as indicated in relevant figure legends. In all cases, representative figure displays were appropriately replicated.
Randomization	Hamsters were randomized to S2X259 and control treatment. For other experiments, randomization was not a relevant feature as we were applying a uniform set of techniques across a panel of candidate monoclonal antibodies
Blinding	The hamster derived samples for RNA quantification and viral load titration were run by technicians who were blinded to treatment group of the analyzed samples

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Antibodies |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Animals and other organisms |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern |

Methods

- | n/a | Involved in the study |
|-------------------------------------|--|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

Antibodies

Antibodies used	<p>For antibody isolation, B cells were stained with CD19 PE-Cy7 (BD Biosciences 557835), anti-IgM (BioLegend 314508), anti-IgD (BD Bioscience 555779), anti-CD14 (BD Bioscience 562691), and anti-IgA (Southern Biotech 2050-09), and spike labeled with streptavidin Alexa-Fluor 647 (Life Technologies S21374)</p> <p>* For FACS during deep mutational scanning, we used PE-conjugated goat anti-human-IgG (Jackson ImmunoResearch 109-115-098) and FITC-conjugated chicken anti-Myc (Immunology Consultants Lab, CYMC-45F).</p> <p>* For VSV pseudotyped neutralization assays, media was supplemented with anti-VSV-G antibody (I1-mouse hybridoma supertant, ATCC CRL-2700)</p> <p>* For SPR, immobilization was performed with anti-AviTag pAb (Genscript Cat # A00674-40) and goat anti-human IgG Fc pAb, (Southern Biotech Cat # 2014-01)</p> <p>* For RBD ELISA, antibody binding was detected via goat anti-human IgG (Southern Biotech, 2040-04)</p> <p>* For S1 shedding and binding FACS analysis, we used AF647-conjugated goat anti-human-IgG (Jackson ImmunoResearch 109-606-098)</p> <p>* For antibody blockade of ACE2 binding, RBD was detected via goat anti-mouse IgG (Southern Biotech 1030-04)</p> <p>* For detection of circulating mAb levels in hamster experiments, an anti-human LS mutation antibody was custom-made by BioRad AbD Serotec, and anti-human CH2 antibody from BioRad cat# MCA5748G clone R10Z8E9</p> <p>* Monoclonal antibodies (S309 and S2X259) were produced in-house using recombinant protein purification as described in the Methods. S2H97_GL and S2E12_GL were produced by ATUM.</p> <p>All the commercial antibodies used in the study have been indicted with supplier name, catalog number. The monoclonal antibodies used as controls or for combination experiments were previously described by our group.</p>
Validation	<p>We discovered and describe S2X259 monoclonal antibody in this study. Target validation of this antibody was performed with multiple binding assays, in addition to cryoEM and x-ray crystallography structures were determined. Reactivity of primary antibodies listed above is based on the information on manufacturer's homepages.</p>

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	Cell lines used in this study were obtained from ATCC (HEK293T, Vero and Vero-E6), ThermoFisher Scientific (Expi CHO cells, FreeStyle™ 293-F cells and Expi293F™ cells) or were generated via lentiviral trasduction (Expi CHO-S, HEK293T-ACE2).
Authentication	None of the cell lines used were authenticated
Mycoplasma contamination	Cell lines were tested for mycoplasma contamination
Commonly misidentified lines (See ICLAC register)	No commonly misidentified cell lines were used in the study

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals	6-10 week-old female Syrian hamsters (<i>Mesocricetus auratus</i>) were Syrian hamsters (<i>Mesocricetus auratus</i>) were purchased from Janvier Laboratories and used in the study
Wild animals	The study did not involve wild animals
Field-collected samples	No field collected samples were used in the study
Ethics oversight	This work was conducted in the high-containment A3 and BSL3+ facilities of the KU Leuven Rega Institute (3CAPS) under licenses AMV 30112018 SBB 219 2018 0892 and AMV 23102017 SBB 219 20170589 according to institutional guidelines. Housing conditions and experimental procedures were approved by the ethical committee of animal experimentation of KU Leuven (license P065-2020).

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Human research participants

Policy information about [studies involving human research participants](#)

Population characteristics	Participants were identified by prior SARS-CoV-2 infection for PBMC isolation. Blood drawn from donor S2X (male, 52 years-old) was obtained at day 75 after symptoms onset. For recovered individual samples were collected between 14 and 75 days after symptoms onset, for vaccinees samples were collected between 13 and 20 days after the second dose of mRNA-based SARS-CoV-2 vaccine.
Recruitment	Patients were recruited on the basis of prior SARS-CoV-2 infection or vaccination in the hospital or outpatient setting. Patients were healthy volunteers who donated blood after being informed about the study. The only exclusion criteria used were HIV or other debilitating disease, but other information about diagnosis and treatment was not collected.
Ethics oversight	Study protocols for antibody isolation were approved by the local Institutional Review Board (Canton Ticino Ethics

Ethics oversight

Committee, Switzerland), and all donors provided written informed consent for the use of blood and blood components. Study protocols for serological assays were approved by the local Institutional Review Boards relevant for each of three cohorts of samples (Canton Ticino Ethics Committee, Switzerland, the Ethical Committee of Luigi Sacco Hospital, Milan, Italy, and WCG North America, Princeton, NJ, USA). All donors provided written informed consent for the use of blood and blood components (such as PBMCs, sera or plasma) and were recruited at hospitals or as outpatients.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Flow Cytometry

Plots

Confirm that:

- The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).
- The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).
- All plots are contour plots with outliers or pseudocolor plots.
- A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation

Yeast were induced for cell-surface expression, incubated with the primary antibody from the panel, and secondary labeled with anti-Myc and anti-Fc fluorescent conjugates as described in the Methods.

Instrument

FACSAria II and ZE5 BioRad

Software

FACSDiva and Everest 3.0

Cell population abundance

We work with homogenous yeast libraries in our deep mutational scanning experiments, therefore cell population abundance is not a detail of our approach.

Gating strategy

Yeast expressing unmutated SARS-CoV-2 are labeled at 1x and 0.01x the concentration used for library labeling. Gates are drawn on the basis of these controls to standardize selection stringency across experiments. Gating schemes are shown in Extended Data Fig. 1c and 7a

- Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.