

## Bromine and Chlorine Determination in Cigarette Tobacco using Microwave-Induced Combustion and Inductively Coupled Plasma Optical Emission Spectrometry

Aline L. H. Müller,<sup>a,b</sup> Cezar A. Bizzi,<sup>a,b</sup> Juliana S. F. Pereira,<sup>a,b</sup> Marcia F. Mesko,<sup>c</sup>  
Diogo P. Moraes,<sup>d,b</sup> Erico M. M. Flores<sup>a,b</sup> and Edson I. Muller<sup>\*,a,b</sup>

<sup>a</sup>Departamento de Química, Universidade Federal de Santa Maria, 97105-900 Santa Maria-RS, Brazil

<sup>b</sup>Instituto Nacional de Ciência e Tecnologia de Bioanalítica, Campinas-SP, Brazil

<sup>c</sup>Instituto de Química e Geociências, Universidade Federal de Pelotas, 96010-610 Pelotas-RS, Brazil

<sup>d</sup>Instituto de Química, Universidade Federal do Rio Grande do Sul, 91501-970 Porto Alegre-RS, Brazil

A combustão iniciada com micro-ondas (MIC) foi aplicada para decomposição de amostras de tabaco de cigarro e subsequente determinação de bromo e cloro por espectrometria de emissão óptica com plasma indutivamente acoplado (ICP OES). Massas de amostra de até 500 mg foram decompostas em frascos fechados e pressurizados com 20 bar de oxigênio. A combustão foi completada em menos de 30 s e os analitos foram absorvidos em solução diluída de  $(\text{NH}_4)_2\text{CO}_3$ . A exatidão foi avaliada usando materiais de referência certificados e mediante a determinação utilizando ICP-MS. A concordância foi melhor do que 98% usando  $50 \text{ mmol L}^{-1}$  de  $(\text{NH}_4)_2\text{CO}_3$  como solução absorvedora e 5 min de refluxo. A temperatura durante a combustão foi superior a  $1400 \text{ }^\circ\text{C}$  e o conteúdo de carbono residual nos digeridos após MIC foi menor que 1%. Até oito amostras podem ser decompostas simultaneamente. Limites de quantificação utilizando MIC e determinação por ICP OES foram de 12 e  $6 \mu\text{g g}^{-1}$  para Br e Cl, respectivamente.

The microwave-induced combustion (MIC) was applied for cigarette tobacco samples digestion and further determination of bromine and chlorine by inductively coupled plasma optical emission spectrometry (ICP OES). Samples masses up to 500 mg were combusted in closed vessels using 20 bar of oxygen. Combustion was complete in less than 30 s and analytes were absorbed in diluted  $(\text{NH}_4)_2\text{CO}_3$  solution. Accuracy was evaluated using certified reference materials with similar matrix composition and comparison with results obtained using ICP-MS. The agreement was better than 98% using  $50 \text{ mmol L}^{-1}$   $(\text{NH}_4)_2\text{CO}_3$  as absorbing solution and 5 min of reflux. Temperature during combustion was higher than  $1400 \text{ }^\circ\text{C}$  and the residual carbon content in digest obtained after MIC was lower than 1%. Up to eight samples could be processed simultaneously and a single absorbing solution was suitable for both Br and Cl. Limit of quantification by MIC and further ICP OES determination was 12 and  $6 \mu\text{g g}^{-1}$  for Br and Cl, respectively.

**Keywords:** microwave-induced combustion, cigarette, bromine, chlorine, ICP OES

### Introduction

Tobacco leaves may be used in different forms such as smoking, chewing or sniffing. All of these forms result in a number of negative side effects on human health that are well-documented.<sup>1</sup> Chlorine and bromine are essential for growth of the plants and minor constituents of tobacco. The content of bromine and chlorine in tobacco could vary depending on tobacco type, soil and climatic

conditions. Typical concentration ranges for Br and Cl reported in tobacco are 100 to 200 and 700 to 30000  $\mu\text{g g}^{-1}$ , respectively. Over 85% of the all halogenated compounds found in tobacco and smoke are either halogenated agrochemicals, impurities found in the agrochemicals, or decomposition products from the agrochemicals. Up to 2.4% of the bromine and chlorine can be transferred to smoke. There are 242 identified halogenated compounds in tobacco and/or tobacco smoke and chlorine and bromine are present in 192 and 12 different compounds, respectively.<sup>2</sup>

\*e-mail: edson\_muller@yahoo.com.br

Several analytical methods have been used for Br and Cl determination in biological samples, such as neutron activation analysis,<sup>3,4</sup> total reflection X-ray fluorescence,<sup>5,6</sup> inductively coupled plasma optical emission spectrometry (ICP OES)<sup>7</sup> and inductively coupled plasma mass spectrometry (ICP-MS).<sup>8-10</sup> Instrumental neutron activation analysis (INAA) is a sensitive method for the determination of metals and non metals and has been accepted worldwide for tobacco analysis because it is easy to handle solid samples and produces little gas during irradiation. However, neutron activation requires a neutron flux source that limits its applicability for most of laboratories.<sup>11</sup>

Total reflection X-ray fluorescence has been used to the determination of halogens in tobacco. However, this technique is prone to interferences related to matrix composition that can affect the accuracy.<sup>12</sup> Inductively coupled plasma optical emission spectrometry and inductively coupled plasma mass spectrometry allow the determination of halogen providing suitable sensitivity and selectivity even at trace and ultra trace concentrations. In this sense, a complete digestion or extraction step is required before the determination. In general, extraction procedures with water have been used for halogens release from the matrix (especially for Cl) and further determination by potentiometric titration using silver nitrate solution. However, sometimes a complete extraction of halogens cannot be achieved and the titrimetric procedure with silver nitrate does not allow distinguishing for chlorine and bromine.<sup>13-15</sup>

The sample preparation step for tobacco digestion presents specific difficulties. Usually, tobacco digestion requires the use of mineral oxidizing acids (*e.g.* concentrated nitric acid) in order to obtain a more effective digestion.<sup>16</sup> This procedure can result in analyte losses due to volatility of chlorine and bromine generally as HX (X = Cl and Br) even using closed vessels.<sup>17-19</sup> In addition, the presence of high acid concentration in digests cannot be supported by some analytical techniques, such as ICP OES<sup>20</sup> and ICP-MS.<sup>21</sup> On the other hand, combustion procedures are particularly suitable to decompose organic materials with high carbon content because of the high temperature achieved.<sup>22</sup> In this case, diluted alkaline solutions can be used avoiding the normal requirement of concentrated reagents.

In recent years an important improvement in combustion methods was the development of microwave-induced combustion (MIC) system. This system combines some advantages of classical combustion procedures with those presented by conventional closed systems heated by microwave radiation.<sup>23</sup> This procedure is performed in the same system conventionally used for microwave-assisted wet digestion and the only instrumental change is a small quartz sample holder which is placed inside

the quartz vessel. A reflux step can be applied after combustion, assuring an effective washing of vessel walls and holder surface, which is a great advantage over the classic combustion systems, where the cleaning step of the vessel must be performed manually. This method has been successfully applied for digestion of biological samples,<sup>24,25</sup> elastomers,<sup>26</sup> coal<sup>27</sup> and crude oil products<sup>28-33</sup> for subsequent metal and halogens determinations.

In this work different cigarette tobacco samples were digested by MIC and further Br and Cl determination was performed by ICP OES. Determination of Br and Cl in MIC digests was also carried out using ICP MS. Sample mass, type and concentration of absorption solution were investigated and the efficiency of decomposition procedure was checked by the residual carbon content determination in digests. The residual carbon content was also determined using ICP OES by monitoring the carbon emission line at 193.091 nm.<sup>34</sup> The accuracy was evaluated using certified reference materials (CRM) with similar matrix composition. Recovery tests were also performed for both analytes. For comparison of results the determination of Br and Cl was also carried out using ICP-MS.

## Experimental

### *Samples and reagents*

Eight different cigarette samples from two different manufacturers were purchased in a local store. Firstly, cigarette filters and paper were removed and tobacco was ground in a cryogenic mill (Spex Certiprep, model 6750 Freezer Mill, Metuchen, NJ, EUA). Samples were frozen in liquid argon during 2 min and further ground during 2 min. The following certified reference materials (CRMs) were used for accuracy evaluation: Virginia tobacco leaves and Oriental tobacco leaves (CTA-VTL-2 and CTA-OTL-1, respectively, Polish certified reference material, Institute of Nuclear Chemistry and Technology, Poland), peach leaves (NIST 1547, National Institute of Standards & Technology, USA) and aquatic plant (BCR 60, Community Bureau of Reference, Brussels). All samples and CRMs were pressed as pellets (diameter of 13 mm) using a hydraulic press set at 3 ton by 1 min (Specac, Orpington, UK). A sample with lower Br and Cl content (sample G) was used for spike studies and for optimization of the absorbing solution.

Water was firstly distilled and subsequently deionized (18.2 M $\Omega$  cm) and analytical-grade reagents (Merck, Darmstadt, Germany) were used throughout. For the determination by ICP OES and ICP-MS standard solutions were prepared in an appropriated concentration of (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> solution. Ammonium nitrate solution (6 mol L<sup>-1</sup>)

was used as an ignition aid. A small disc of filter paper (15 mm in diameter, 12 mg) with low ash content (Black Ribbon Ashless, Schleicher & Schuell GmbH, Dassel, Germany) was used as aid for the combustion process. The filter paper was previously cleaned with absolute ethanol for 20 min in an ultrasonic bath and rinsed with water and dried in an oven for 2 h at 60 °C before use.

Water and diluted ammonium carbonate solutions were used as absorbing solutions for Br and Cl in MIC procedure. Ammonium carbonate was dissolved in water with final concentration ranging from 5 to 100 mmol L<sup>-1</sup>. Glass or quartz materials were soaked in 1.4 mol L<sup>-1</sup> HNO<sub>3</sub> for 24 h and thoroughly washed with water before use.

### Instrumentation

A Multiwave 3000 microwave sample preparation system (Anton Paar GmbH, Graz, Austria) equipped with high-pressure quartz vessels (internal volume of 80 mL, maximum temperature and pressure of 280 °C and 80 bar, respectively) was used in this study for proposed combustion procedure. The software version was v1.27-Synt and the microwave system was previously modified to run with a maximum pressure rate of 3 bar s<sup>-1</sup> (and not 0.8 bar s<sup>-1</sup> as in the original software). This change was necessary to prevent eventual interruption of the microwave irradiation before all the samples start the combustion. Pressure was monitored in each vessel for all the runs. A specially designed quartz device was used as sample holders for the MIC procedure. This device was developed in order to facilitate the oxygen diffusion by the pressed sample and to reduce the cool surfaces that might cause carbon deposits in the quartz surface.<sup>35</sup>

An inductively coupled plasma optical spectrometer model Spectro Ciros CCD simultaneous spectrometer with axial view configuration (Spectro Analytical Instruments,

Kleve, Germany) was used for Br, Cl and residual carbon determination. A crossflow nebulizer coupled to a Scott double pass type nebulization chamber was used throughout. Plasma operating conditions used in this work were a radio-frequency power generator of 1700 W, and argon flow rates of plasma, auxiliary and nebulizer were 14.0, 1.00 and 1.00 L min<sup>-1</sup>, respectively. The selected wavelengths used for Br, Cl and C determinations were 154.065, 134.724 and 193.091 nm, respectively. Argon of 99.996% purity (WhiteMartins, Praxair, São Paulo, Brazil) was used for plasma generation, nebulization and as auxiliary gas for the halogen determination (Table 1).

Halogen determinations by ICP OES were compared using an inductively coupled plasma mass spectrometer (PerkinElmer-SCIEX, Model Elan DRC II, Thornhill, Canada) equipped with a concentric nebulizer (Meinhard Associates, Golden, CA, USA), a baffled cyclonic spray chamber (Glass Expansion, Inc., West Melbourne, Australia) and a quartz torch with a quartz injector tube (2 mm i.d.) was used for the determination of Br and Cl after MIC. Instrumental performance optimization, including nebulizer gas flow rate, RF power and ion lens voltage, was performed and operational conditions are described in Table 1. Chlorine determination by ICP-MS was performed with DRC using ammonia (purity higher than 99.999%) as reaction gas. The DRC parameters were adjusted in order to minimize the interferences on <sup>35</sup>Cl<sup>+</sup> isotope according to previous work.<sup>36</sup>

The temperature during cigarette tobacco samples combustion was determined using an optical pyrometer (Ultimax Infrared Thermometer, Ircon, Niles, IL, USA) equipped with a close-up VX-CL1 lens.

### Proposed microwave-induced combustion procedure

The operation conditions for the proposed MIC procedure were evaluated using cigarette tobacco pellets

**Table 1.** Operational conditions of halogens determination by ICP OES and ICP-MS

Parameter	ICP OES	ICP-MS
RF power / W	1700	1400
Plasma gas flow rate / (L min <sup>-1</sup> )	14.0	15.0
Auxiliary gas flow rate / (L min <sup>-1</sup> )	1.00	1.2
Nebulizer gas flow rate / (L min <sup>-1</sup> )	1.00	1.15
Spray chamber	Scott type, double path	Cyclonic
Nebulizer	Crossflow	Concentric
Sampler and skimmer cones	--	Pt
Ion lens / V	--	Auto lens "on"
Dwell time / ms	--	50
Observation view	Axial	--
Isotope ( <i>m/z</i> )	--	<sup>35</sup> Cl and <sup>79</sup> Br
Emission lines / nm	Br (154.065), Cl (134.724) and C (193.091)	--

with sample masses up to 500 mg that were placed together the filter paper on the quartz holder. Quartz vessels were previously charged with 6 mL of absorbing solution (water or ammonium carbonate) and 50  $\mu\text{L}$  of 6 mol  $\text{L}^{-1}$   $\text{NH}_4\text{NO}_3$  solution were added to the filter paper.

The quartz holder was introduced into the quartz vessel and vessels were pressurized with 20 bar of oxygen using the valve originally designed for the pressure release when using conventional acid digestion system. Then, the rotor was placed inside the oven and the selected microwave heating program was started. The microwave irradiation program used was 1400 W for 5 min (optional reflux step) and 0 W for 20 min (cooling step). The time for the optional reflux step after the combustion and cooling time were set in accordance with previous studies.<sup>32,36</sup> After digestion, the pressure of each vessel was carefully released. In this work, each run was performed with a minimum of four vessels. The resultant solutions were diluted with water to 15 mL for further analysis by ICP OES. The concentration of ammonium nitrate solution, the minimum volume for the ignition process, as well as the mass of paper filter were based on previous work.<sup>23</sup> Cleaning of vessels and holders was carried out with 6 mL of concentrated  $\text{HNO}_3$  in the microwave oven at 1400 W for 5 min and 0 W for 20 min for cooling.

#### *Microwave-assisted extraction using water and alkaline solution*

A microwave-assisted water extraction procedure was also evaluated. In this sense, a mass of 500 mg of sample was placed in the quartz vessel and a volume of 6 mL of water was added. The same procedure was carried out using 100 mmol  $\text{L}^{-1}$   $(\text{NH}_4)_2\text{CO}_3$ . Heating program was 1400 W for 30 min (ramp of 10 min) and 0 W for 20 min (cooling step). The maximum temperature and pressure were 280 °C and 80 bar, respectively. After cooling, resultants solutions from extraction procedures were diluted with water to 20 mL in a polypropylene vessel and then centrifuged at 3000 rpm for 10 min prior to determination by ICP OES.

## Results and Discussion

#### *Determination of bromine and chlorine in cigarette tobacco samples by ICP OES after microwave-assisted extraction using water and alkaline solution*

A sample preparation procedure based on extraction with water in closed vessels was investigated for cigarette tobacco samples. Results obtained for CRM samples using water as extraction solution were with maximum agreement

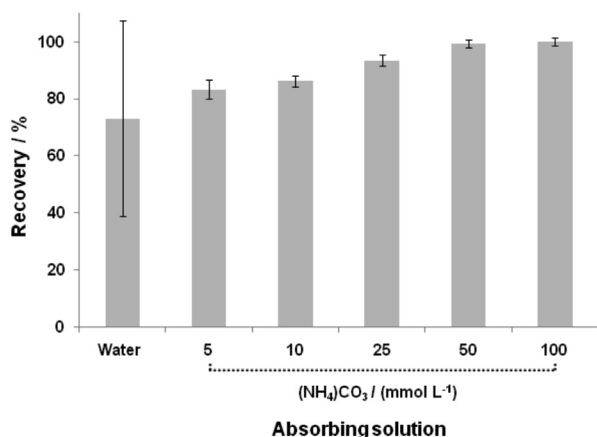
of 70% and 90% for Br and Cl, respectively. The use of alkaline extraction solution did not improve the recoveries for Br and Cl. Therefore, extraction using water or alkaline solution in closed vessels, even under high pressure and temperature, was not suitable for tobacco preparation for further Br and Cl determination.

#### *Optimization of microwave-induced combustion conditions*

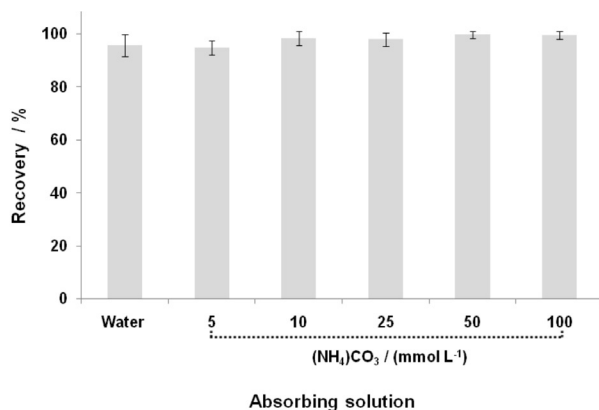
For the proposed procedure, cigarette tobacco samples were ground in a cryogenic mill and sample pellets of different masses were obtained (100, 250 and 500 mg). An initial study was performed in order to evaluate the characteristics of the MIC system such as maximum sample mass and maximum pressure during combustion. Using 100 mg of cigarette tobacco samples, the maximum pressure was about 29 bar and even using sample masses up to 500 mg the maximum pressure achieved during combustion was 39 bar that is about 49% of the maximum pressure recommended by the manufacturer for routine operation (80 bar). After combustion the sample was completely decomposed and negligible brown residues remained in quartz holder. The absorbing solution was completely clear and without visible particles. Therefore, as the procedure was considered safe for digestion up to 500 mg this mass was selected as the maximum mass of sample pellet for the proposed MIC procedure.

#### *Optimization of the absorbing solution for recovery of halogens after microwave-induced combustion*

The choice of the absorbing solution is important to achieve suitable recoveries. It is dependent on the analytes nature and should be compatible with the chosen determination procedures. In general, for halogens, acid solutions generally result in analyte losses by volatilization and also interferences in the determination step could be observed when chromatographic and even spectrometric techniques are used.<sup>19,32</sup> In addition, literature data recommend the use of water or diluted alkaline solutions for further halogen determination after digestion by MIC for different matrices.<sup>27,29,32</sup> In the present study, the suitability of ammonium carbonate as absorbing solution was evaluated for Br and Cl. Concentrations of  $\text{NH}_4\text{CO}_3$  solutions of 5, 10, 25, 50 and 100 mmol  $\text{L}^{-1}$  and also water were evaluated. Spike recoveries in cigarette tobacco samples were evaluated for each absorbing solution. Previous studies<sup>27,29,32</sup> also demonstrated that the use of a reflux step allows better recoveries for halogens determination when compared with combustion without reflux step. Results are shown in Figures 1 and 2 for Br and Cl, respectively.



**Figure 1.** Influence of absorbing solutions for Br determination in cigarette tobacco samples using H<sub>2</sub>O, (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> 10, 25, 50 or 100 mmol L<sup>-1</sup> and reflux step. Determination by ICP OES. (error bars are the standard deviation, n = 3).



**Figure 2.** Influence of absorbing solutions for Cl determination in cigarette tobacco samples using H<sub>2</sub>O, (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> 10, 25, 50 or 100 mmol L<sup>-1</sup> and reflux step. Determination by ICP OES. (error bars are the standard deviation, n = 3).

Recoveries below 75% were obtained for Br using water as absorbing solution (Figure 1). Moreover, the recovery of Br was dependent of (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> concentration. For 5, 10 and 25 mmol L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> the recoveries were not quantitative.

**Table 2.** Results for Br and Cl in cigarette samples digested by MIC using 50 mmol L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> as absorbing solution and determination by ICP OES and ICP-MS (mean ± standard deviation, µg g<sup>-1</sup>, n = 3)

Samples	Br		Cl	
	ICP OES	ICP-MS	ICP OES	ICP-MS
A	51.7 ± 1.5	52.2 ± 1.4	7086 ± 100	7075 ± 98
B	34.6 ± 1.8	34.0 ± 1.3	4589 ± 89	4621 ± 77
C	53.3 ± 2.0	54.1 ± 1.4	8249 ± 79	8201 ± 88
D	42.0 ± 1.7	41.5 ± 1.2	6743 ± 78	6798 ± 85
E	30.9 ± 2.0	31.3 ± 1.3	4253 ± 47	4268 ± 42
F	38.1 ± 2.0	37.1 ± 1.5	4876 ± 75	4866 ± 72
G	29.8 ± 2.1	30.0 ± 1.3	3871 ± 22	3865 ± 23
H	50.9 ± 2.2	50.1 ± 2.3	7965 ± 76	7951 ± 62

However, when 50 and 100 mmol L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> were used as absorbing solutions the recoveries were close to 100%. In Figure 2 it is possible to observe that the recoveries obtained for Cl using different absorbing solution were not significantly different (ANOVA, 95% confidence level) even using water. Therefore, 50 mmol L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> was chosen because it was a compromise condition and allows Br and Cl determination in cigarette tobacco samples.

#### *Determination of bromine and chlorine by ICP OES in cigarette tobacco samples after MIC digestion*

The proposed procedure was applied to digestion of eight cigarette tobacco samples using 50 mmol L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> as absorbing solution. The results for Br and Cl obtained using ICP OES are shown in Table 2.

Bromine and Cl concentrations ranged from 29.8 to 53.3 and 3871 to 8249 µg g<sup>-1</sup>, respectively. The relative standard deviations (RSDs) were lower than 7.0% and 1.9% for Br and Cl determination in commercial cigarette tobacco samples, respectively. Bromine and Cl were also determined by ICP-MS and no statistical difference (confidence level of 95%) was observed in comparison with results obtained by ICP OES. Based on this study, the use of concentrated acids or excessive amounts of reagents could be avoided in MIC, reducing blank values and minimizing problems concerning to differences between viscosity of standards and samples. In addition, MIC also reduces the residues generation according to recommendations of green chemistry. The limits of quantification (LOQ) for Br and Cl using ICP OES, calculated as ten times the standard deviation of the blank, were 12 and 6 µg g<sup>-1</sup>, respectively.

In order to check the accuracy, MIC was applied for four different CRMs. After digestion, Br and Cl were determined by ICP OES. Results are shown in Table 3.

When comparing MIC results and certified values for CTA-VTL-2, significant differences were not observed

**Table 3.** Results for Br and Cl in CRM samples digested by MIC using 50 mmol L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> as absorbing solution and determination by ICP OES (mean ± standard deviation, µg g<sup>-1</sup>, n = 5)

Certified reference material	Bromine		Chlorine	
	Certified value	MIC	Certified value	MIC
CTA-VTL-2	14.3 ± 1.4	14.0 ± 2.9	7430 ± 280	7350 ± 210
CTA-OTL-1	9.28 ± 1.06	< 12.0	2988 <sup>a</sup>	2995 ± 55
NIST 1547	11 <sup>a</sup>	< 12.0	360 ± 19	365 ± 20
BCR 60	20 <sup>a</sup>	19.5 ± 2.5	10 <sup>a</sup>	10.5 ± 1.5

<sup>a</sup>informed value.

for Br determination (*t*-test, 95% confidence level) and an agreement better than 98% was obtained. Statistical differences (*t* test, 95% confidence level) were also not observed for Cl concentration obtained by MIC and certified reference value for CTA-VTL-2 and NIST 1547 samples and an agreement better than 99% was obtained for Cl.

#### Evaluation of digestion efficiency of proposed method

In this work, the residual carbon content (RCC) was used in order to evaluate the efficiency of the proposed MIC procedure. Depending on the instrumental technique, the RCC value must be controlled. For MIC using the reflux step, RCC values below 1% were obtained after decomposition of cigarette tobacco samples. These values can be explained due to the high temperature reached (about 1500 °C) during the combustion. In this condition, practically all of the organic materials could be completely decomposed.

## Conclusions

Microwave-induced combustion was suitable for cigarette tobacco digestion for further Br and Cl determination by ICP OES. In spite of ICP OES has been considered suitable for analysis of cigarette tobacco samples, ICP-MS could provide lower LODs if it is necessary. Microwave-assisted water extraction presented poor recoveries for Br. The use of MIC showed good performance for cigarette tobacco digestion due to complete oxidation of the organic matrix using a relatively high sample mass (500 mg) that allowed obtaining low LODs. The operation was considered safe and a significant reduction of laboratory waste and analysis time was obtained that is important aspect for routine analysis. In addition, it was possible to obtain a single solution suitable for both analytes determination using different analytical techniques and with residual carbon content lower than 1%. Reproducibility was considered suitable for Cl and

Br with RSDs lower than 7%. Results obtained for CRMs samples under optimized conditions were in agreement to the certified values.

## Acknowledgments

The authors are grateful to INCT-Bioanalítica, CNPq, CAPES and FAPERGS for supporting this study.

## References

1. International Agency for Research on Cancer (IARC); *Tobacco Smoke and Involuntary Smoking, IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans*, vol. 83, International Agency for Research on Cancer, WHO: Lyon, France, 2004.
2. Rodgman, A.; Perfetti, T. A.; *The Chemical Components of Tobacco and Tobacco Smoke*, CRC Press: Boca Raton, FL, USA, 2009, pp. 857-893.
3. Serfor-Armah, Y.; Nyarko, B. J. B.; Akaho, E. H. K.; Kyere, A. W. K.; Osae, S.; Oppong-Boachie, K.; Osae, E. K.; *J. Radioanal. Nucl. Chem.* **2001**, *250*, 173.
4. Çam, N. F.; Yaprak, G.; Erduran, M. N.; *J. Radioanal. Nucl. Chem.* **1999**, *242*, 273.
5. Martinez, T.; Lartigue, J.; Zarazua, G.; Avila-Perez, P.; Navarrete, M.; Tejada, S.; *Spectrochim. Acta, Part B* **2008**, *63*, 1469.
6. Stephens, W. E.; Calder, A.; *Anal. Chim. Acta* **2004**, *527*, 89.
7. Naozuka J.; Veiga, M. A. M. S., Oliveira, P. V.; Oliveira, E.; *J. Anal. At. Spectrom.* **2003**, *18*, 917.
8. Hu Q.; Li L.; Shi J.; Chen Z.G.; Wang F.; Wang A.; *Spectrosc. Spect. Anal.* **2007**, *27*, 1210.
9. Swami, K.; Judd, C. D.; Orsini, J.; *Spectrosc. Lett.* **2009**, *42*, 479.
10. Torrence, K. M.; McDaniel, R. L.; Self, D. A.; Chang, M. J.; *Anal. Bioanal. Chem.* **2002**, *372*, 723.
11. Jervis, R. E.; *J. Radioanal. Nucl. Chem.* **1999**, *239*, 19.
12. Beckhoff, B.; Kanngießer, B.; Langhoff, N.; Wedell, R.; Wolff, H. eds.; *Handbook of Practical X-ray Fluorescence Analysis*, Springer: Germany, 2006, pp. 309-407.

13. Gilliam, J. W.; *Soil Sci. Soc. Am. J.* **1971**, *35*, 512.
14. Gaines, T. P.; Parker, M. B.; Gascho, G. J.; *Agron. J.* **1984**, *76*, 371.
15. Adriano, D. C.; Pratt, P. F.; Holtzclaw, K. M.; *Agron. J.* **1973**, *65*, 133.
16. Crispino, C. C.; Fernandes, K.G.; Kamogawa, M. Y.; Nóbrega, J. A.; Nogueira, A. R. A.; Ferreira, M. M. C.; *Anal. Sci.* **2007**, *23*, 435.
17. Di Narda, F.; Toniolo, R.; Bontempelli, G.; *Anal. Chim. Acta* **2001**, *436*, 245.
18. Fecher, P.A; Nagengast A.; *J. Anal. At. Spectrom.* **1994**, *9*, 1021.
19. Pereira, J. S. F.; Mello, P. A.; Moraes, D. P.; Duarte, F. A.; Dressler, V. L.; Knapp, G.; Flores, E. M. M.; *Spectrochim. Acta, Part B* **2009**, *64*, 554.
20. Trevizan, L. C.; Nóbrega, J. A.; *J. Braz. Chem. Soc.* **2007**, *18*, 678.
21. Tangen, A.; Lund, W.; *Spectrochim. Acta, Part B* **1999**, *54*, 1831.
22. Barin, J. S.; Flores, E. M. M.; Knapp, G. In *Trends in Sample Preparation*; Arruda, M. A. Z., ed.; Nova Science: New York, 2006, p. 288.
23. Flores, E. M. M.; Barin, J. S.; Paniz, J. N. G.; Medeiros, J. A.; Knapp, G.; *Anal. Chem.* **2004**, *76*, 3525.
24. Duarte F. A.; Pereira, J. S. F.; Barin, J. S.; Mesko, M. F.; Dressler, V. L.; Flores, E. M. M.; Knapp, G.; *J. Anal. At. Spectrom.* **2009**, *24*, 224.
25. Mesko, M. F.; Mello, P. A.; Bizzi, C. A.; Dressler, V. L.; Knapp, G.; Flores, E. M. M.; *Anal. Bioanal. Chem.* **2010**, *398*, 1125.
26. Moraes, D. P.; Mesko, M. F.; Mello, P. A.; Paniz, J. N. G.; Dressler, V. L.; Knapp, G.; Flores, E. M. M.; *Spectrochim Acta, Part B* **2007**, *62*, 1065.
27. Flores, E. M. M.; Mesko, M. F.; Moraes, D. P.; Pereira, J. S. F.; Mello, P. A.; Barin, J. S.; Knapp, G.; *Anal. Chem.* **2008**, *80*, 1865.
28. Mello, P. A.; Pereira, J. S. F.; Moraes, D. P.; Dressler, V. L.; Flores, E. M. M.; Knapp, G.; *J. Anal. At. Spectrom.* **2009**, *24*, 911.
29. Pereira, J. S. F.; Mello, P. A.; Moraes, D. P.; Duarte, F. A.; Dressler, V. L.; Knapp, G.; Flores, E. M. M.; *Spectrochim. Acta, Part B* **2009**, *64*, 554.
30. Pereira, J. S. F.; Mello, P. A.; Duarte, F. A.; Santos, M. F. P.; Guimarães, R. C. L.; Knapp, G.; Dressler, V. L.; Flores, E. M. M.; *Energy Fuels* **2009**, *23*, 6015.
31. Mello, P. A.; Giesbrecht, C. K.; Alencar, M. S.; Moreira, E. M.; Paniz, J. N. G.; Dressler, V. L.; Flores, E. M. M.; *Anal. Lett.* **2008**, *41*, 1623.
32. Pereira, J. S. F.; Diehl, L. O.; Duarte, F. A.; Santos, M. F. P.; Guimarães, R. C. L.; Dressler, V. L.; Flores, E. M. M.; *J. Chromatogr., A*, **2008**, *1213*, 249.
33. Pereira, J. S. F.; Moraes, D. P.; Antes, F. G.; Diehl, L. O.; Santos, M. F. P.; Guimarães, R. C. L.; Fonseca, T. C. O.; Dressler, V. L.; Flores, E. M. M.; *Microchem. J.* **2010**, *96*, 4.
34. Gouveia, S. T.; Silva, F. V.; Costa, L. M.; Nogueira, A. R. A.; Nóbrega, J. A.; *Anal. Chim. Acta* **2001**, *445*, 269.
35. Mesko, M. F.; Moraes, D. P.; Barin, J. S.; Dressler, V. L.; Knapp, G.; Flores, E. M. M.; *Microchem. J.* **2006**, *82*, 183.
36. Antes, F. G.; Duarte, F. A.; Paniz, J. N. G.; Santos, M. F. P.; Guimarães, R. C. L.; Flores, E. M. M.; Dressler, V. L.; *At. Spectrosc.* **2008**, *29*, 157.

Submitted: January 11, 2011

Published online: June 7, 2011

**FAPESP has sponsored the publication of this article.**