

Budget of Primary Production and Dinitrogen Fixation in a Highly Seasonal Red Sea Coral Reef

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ABSTRACT

Biological dinitrogen (N₂) fixation (diazotrophy, BNF) relieves marine primary producers of nitrogen (N) limitation in a large part of the world oceans. N concentrations are particularly low in tropical regions where coral reefs are located, and N is therefore a key limiting nutrient for these productive ecosystems. In this context, the importance of diazotrophy for reef productivity is still not resolved, with studies up to now lacking organismal and seasonal resolution. Here, we present a budget of gross primary production (GPP) and BNF for a highly seasonal Red Sea fringing reef, based on ecophysiological and benthic cover measurements combined with geospatial analyses. Benthic GPP varied from 215 to 262 mmol C m⁻² reef d⁻¹, with hard corals making the largest contribution (41–76%). Diazotrophy was

omnipresent in space and time, and benthic BNF varied from 0.16 to 0.92 mmol N m⁻² reef d⁻¹. Planktonic GPP and BNF rates were respectively approximately 60- and 20-fold lower than those of the benthos, emphasizing the importance of the benthic compartment in reef biogeochemical cycling. BNF showed higher sensitivity to seasonality than GPP, implying greater climatic control on reef BNF. Up to about 20% of net reef primary production could be supported by BNF during summer, suggesting a strong biogeochemical coupling between diazotrophy and the reef carbon cycle.

Key words: Diazotrophy; Photosynthesis; Productivity; Nutrient budget; Biogeochemical cycling; Gulf of Aqaba.

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INTRODUCTION

Nitrogen (N) is a fundamental component of all living organisms. In particular, N is needed by primary producers in capturing energy through photosynthesis and building biomass, leading to a tight coupling of the N and carbon (C) cycles (Gruber and Galloway 2008) and dictating constraints to the

flexibility of the ecosystem C:N stoichiometry (Geider and La Roche 2002). In unperturbed and oligotrophic marine systems, primary productivity is often limited by bioavailable forms of N, which are scarce due to low atmospheric inputs and N loss pathways (Vitousek and Howarth 1991). This limitation is of particular significance in coral reef ecosystems, as these are among the ecosystems displaying the highest rates of gross primary production (GPP), yet experiencing very low ambient concentrations of dissolved nutrients. Here, biological dinitrogen (N₂) fixation (diazotrophy, BNF) is thought to play an important role in replenishing the N pool, maintaining the ecosystem productivity and its biological storage of C (D'Elia and Wiebe 1990; Capone 1996; O'Neil and Capone 2008).

Since the early work by Wiebe and others (1975) on BNF in algal reef flats of the Marshall Islands, many studies have found high rates of BNF associated with several benthic substrates, ranging from sediments and cyanobacterial mats to macroalgae and scleractinian corals (for reviews, see O'Neil and Capone 2008; Cardini and others 2014). Moreover, after first evidence of a diazotroph–coral association by Shashar and others (1994a) and the subsequent discovery of diazotrophic cyanobacteria in the tissue of scleractinian corals of the genus *Montastraea* (Lesser and others 2004), there has been emergent recognition of the potential contribution of N₂-fixing symbioses in coral reefs (Fiore and others 2010; Cardini and others 2014). Although reef sediments and cyanobacterial mats show high rates of BNF (O'Neil and Capone 1989; Capone and others 1992; Shashar and others 1994b; Charpy-Roubaud and others 2001; Bednarz and others 2015b), N₂-fixing coral symbioses may also be responsible for significant inputs of N on the ecosystem level, particularly in reef habitats with high live coral coverage.

BNF in coral reef habitats is highly variable (O'Neil and Capone 2008) and potentially affected by global and local anthropogenic disturbances (Cardini and others 2014). Therefore, it is increasingly important to quantify BNF at the reef scale, and under changing environmental conditions, if we want to understand how diazotrophy is contributing to the functioning of reef ecosystems. Because of the uncertainty in (1) the distribution of BNF in coral reefs and in (2) the tightness of the coupling between BNF and primary production, the ecological significance of BNF in coral reef ecosystems is still not resolved, demonstrating the difficulty in determining BNF rates for whole communities (Cuet and others 2011). Thus, the objective of this paper is to provide an assessment

of the quantitative importance of the process of BNF for primary production in coral reef ecosystems. Although some studies have attempted this before (Capone and Carpenter 1982; Larkum and others 1988; Capone 1996; Casareto and others 2008), no systematic and comprehensive analyses considering the complex diversity of coral reef substrates have been performed so far that would allow for a synthesis. In particular, questions that we target are: (1) What is the relative contribution of key individual benthic categories to overall reef GPP and BNF? (2) How much are different benthic reef habitats contributing to these two processes, and is the water column contributing significantly? (3) To what extent is reef BNF important in sustaining net reef primary production (NPP)? (4) Are reef GPP and BNF affected by seasonality, and by seasonal shifts in benthic assemblages?

We address these questions using results from a year-long study at a highly seasonal fringing reef in the northern Red Sea, combining ecophysiological GPP and BNF measurements of all key benthic players with benthic surveys and environmental monitoring in high temporal resolution. Moreover, geographic information system (GIS) analyses allowed us to investigate the relative importance of all reef-associated habitats to total reef GPP and BNF. Finally, we indicate the most probable environmental factors driving BNF in coral reef habitats, and provide revised estimates of reef-wide BNF, with consequent implications for management and for predicting the potential effects of human alterations on reef biogeochemical cycling.

MATERIALS AND METHODS

Study Site

This study was carried out at a northern Red Sea fringing reef (Aqaba, Jordan) during two expeditions (January–April 2013 and August–December 2013). Four sampling campaigns, each encompassing 3 weeks, were undertaken in February, April, September, and November, respectively. The timing of these periods (hereinafter called winter, spring, summer, and autumn, respectively) was determined from literature to best cover the annual diversity of environmental conditions in this area of the Red Sea. In fact, the Gulf of Aqaba is characterized by an annual cycle of deep water mixing from December until May and stratification down to 200 m water depth from June until November (Silverman and others 2007; Carlson and others 2014). All measurements were conducted on site in the aquarium and laboratory facilities of the Mar-

ine Science Station (MSS, University of Jordan; location: 29°27'N, 34°58'E). Monitoring and sample collection took place on the adjacent fore reef slope at 10 m water depth. The fringing reef system extends approximately 1.1 km along its reef crest bordering the coastline. This reef has the typical morphology and zonation of Red Sea fringing reefs, and can be partitioned into a reef flat, a reef crest and slope (hereafter: reef crest), and a fore reef facing the open sea (Mergner and Schuhmacher 1974; Naumann and others 2012b). The fore reef consists of an upper, middle, and lower part characterized by distinctive morphological features and species composition (hereafter: sand belt, transition zone, and fore reef). The present study focuses on the reef area framed by the reef flat and the fore reef slope at the maximum investigated depth (depth range: 0.5–20.0 m).

Environmental Monitoring

Key environmental parameters were continuously monitored at the sampling site during all seasons. Light availability and water temperature were measured with data loggers (Onset HOBO Pendant UA-002-64; temperature accuracy: $\pm 0.53^\circ\text{C}$, spectral detection range 150–1200 nm) and a quantum sensor (Model LI-192SA; Li-Cor). Light measurements are presented as total mole of photons per square meter per day, while temperature values are daily (24 h) averages. Sampling for dissolved inorganic nitrogen ($\text{DIN} = \text{NH}_4^+ + \text{NO}_x$), dissolved inorganic phosphorous (DIP), chlorophyll *a* (Chl *a*), dissolved organic carbon (DOC), particulate organic carbon (POC), and particulate nitrogen (PN) was carried out once a week by SCUBA. For a detailed explanation of the analytical procedures, please refer to Bednarz and others (2015b) and Rix and others (2015).

Benthic Community Composition

Line-point intercept (LPI) surveys were conducted to quantify benthic community composition and to identify the dominant benthic categories. In each season, three LPI, each of 50 m length, were carried out at 0.5, 1.0, 5.0, 10.0, and 20.0 m water depth, at increasing distance from land along transects parallel to the shoreline, both in the north and south sections of the reef. This accounted for a total of 120 LPIs. The transect water depths were selected as in previous studies (Naumann and others 2012b) to include all major reef habitats and to be representative of the seafloor coverage by all dominant benthic categories. Intervals of 0.5 m between intercepts resulted in 101 data points per LPI. Results obtained from all LPI in each water

depth were used to calculate the percentage coverage for each reef habitat (that is, reef flat, reef crest, sand belt, transition zone, and fore reef) in each season (Table S1), and to select the benthic categories to be subsequently incubated that together accounted on average for ca. 90% of the slope 2D reef area (Figure S3, Table S2, S3).

Reef Bathymetry and Zonation

Bathymetric contours were obtained by merging data from the Aqaba Special Economic Zone Authority (ASEZA) GIS Unit and original bathymetric data obtained in the field, overlaid onto a high-resolution satellite image (© Digital Globe). Bathymetric contours were subsequently processed into a continuous raster using the ArcGis Topo to Raster tool (ESRI ArcGis®), with the final raster having a cell size of approximately 2.9 m. The habitat map was produced based on prior knowledge of the reef zonation (Mergner and Schuhmacher 1974), on the interpretation of the high-resolution satellite image, and on ground truthing during the LPI surveys. Finally, zonal statistics referring to the bathymetric raster were calculated for each habitat using ArcGis 3D Analyst tools. The slope 2D area (that is, considering the depth profile, see Figure S3) for each habitat was obtained from the planar 2D area using the bathymetric raster as base layer. The bathymetric and habitat maps are presented in Figure 1, and zonal statistics for each habitat are reported in Table S2 and were used in our subsequent ecological analyses.

Sample Collection and Maintenance

All benthic categories were sampled on the reef and transported back to the MSS facilities within 60 min of sampling. For an overview and description of the categories incubated and respective replication, see Table S3. Hard and soft corals, as well as turf algae covered branches, were fixed to ceramic tiles while coral rock fragments were stable without fixation. Macroalgae were fixed to Petri dishes with rubber bands, protecting the algal tissue from crushing. Sediment and cyanobacterial mat samples were collected with cylindrical sediment corers (internal diameter = 43 mm) and the top 1 cm of the core was extruded and placed in a Petri dish with the same internal diameter as the sediment corer. Coral rock was defined as pieces of biogenic reef framework with the carbonate structure clearly visible and open for settling organisms. Categories (such as coral rock and sediment) are hereafter defined 'bare' when they lack coverage by a dominant and visible epibenthic group and the

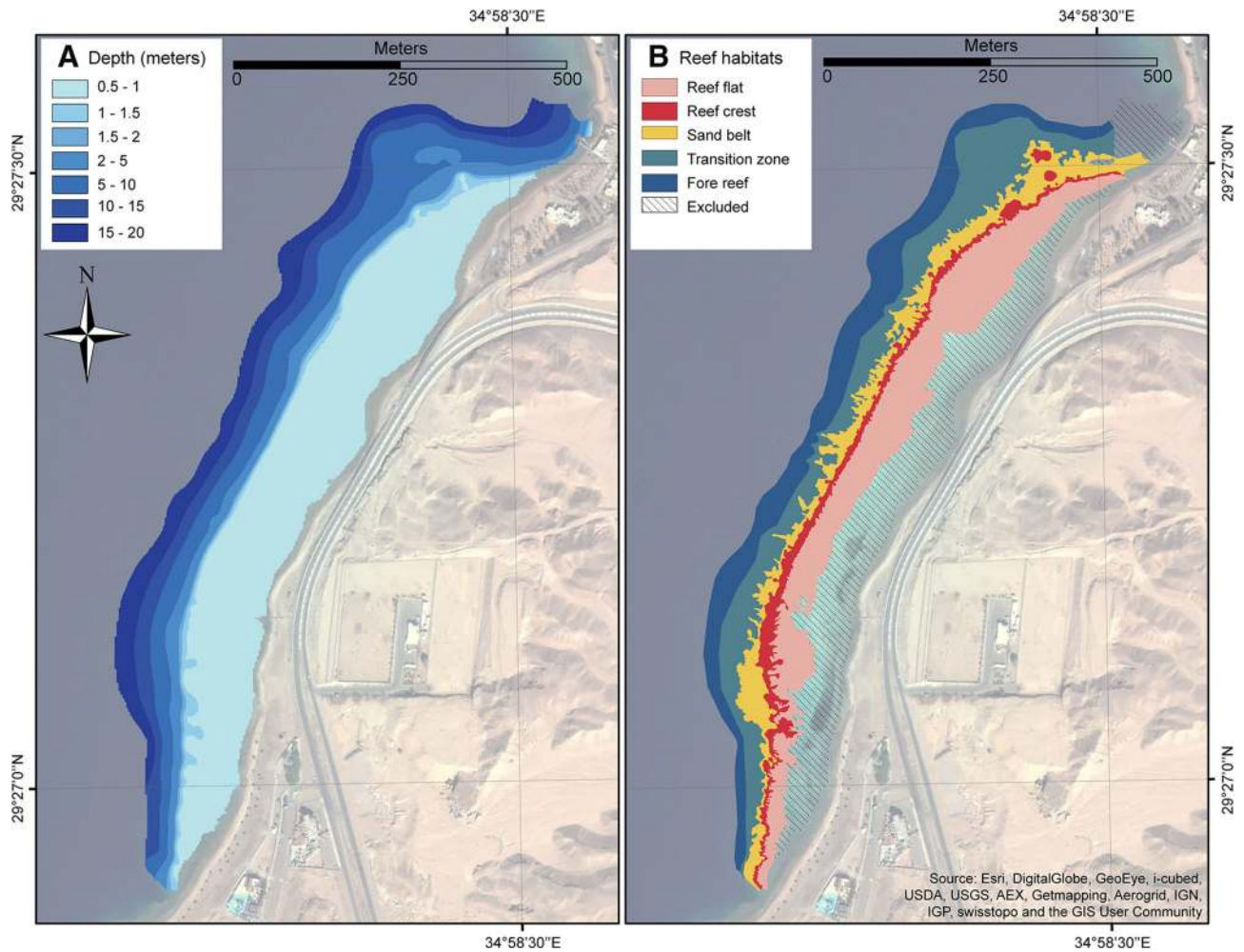


Figure 1. Bathymetry (A) and reef habitat (B) maps of the study site.

substrate is open for settling organisms although being associated with endobenthic algae, microbial biofilms, and sparse patches of crustose coralline algae (CCA), cyanobacteria, and filamentous algae (Rix and others 2015). All organisms and substrates were maintained in an outside flow-through aquarium of 800 L with an exchange rate of 4000 L h⁻¹. Water was pumped straight from the reef at 10 m water depth, thus resembling in situ conditions of temperature and inorganic nutrients. In situ light conditions at 10 m water depth were generated using layers of black mesh and monitored with Onset HOBO data loggers. Hard and soft corals were maintained for at least 7 days, and macroalgae for at least 24 h, to recover from the collection process. Sediment, turf algae, coral rock, and cyanobacterial mats were incubated on the day of collection.

Surface Area Quantification

The 3D surface areas of all benthic categories were quantified using various tools. 3D surface areas of hard and soft corals, turf algae, and coral rock were measured using Advanced Geometry (Naumann and others 2009; Bednarz and others 2015a). Macroalgae surface areas were assessed by digital image analysis of planar photographs using ImageJ v.1.48 (Schneider and others 2012), and doubling the one-sided surface area to obtain the total 3D specimen surface area as described in Haas and others (2010). Sediment and cyanobacterial mat surface areas were calculated from the circular surface area sampled by the sediment corer. Average 3D surface area for each benthic category (see Table S3 for information on replication for each category) calculated as described above, was related

to the planar projected 2D area of the same specimens quantified by digital image analysis to generate 2D to 3D area conversion factors (Naumann and others 2012b) as reported in Table S3. These conversion factors were subsequently used to calculate the benthic 3D area available to photosynthetic and/or N_2 -fixing organisms (Figure S3), and to extrapolate our physiological measurements to the reef ecosystem scale (Table S4). The normalization (surface area) and extrapolation (2D:3D conversion) methods used here, although being simple, yields good estimates of the target processes while allowing calculations of areal rates and best comparability of the present data to previous studies (for example, Larkum and others 1988; Eidens and others 2014).

Physiological Measurements

Specimens were incubated for measurements of NPP, dark respiration (R), and BNF. For a detailed explanation of the incubation and analytical procedures, please refer to Bednarz and others (2015b) and Rix and others (2015). Briefly, incubations were conducted under in situ conditions in a flow-through aquarium in closed glass chambers (0.5–1.0 L). Magnetic stirrer plates (CimarecTM i Telesystem Multipoint Stirrers, Thermo ScientificTM) and Teflon-coated stir bars in the chambers provided water movement during the incubations at 600 rpm. All material used to fix organisms was thoroughly cleaned before each incubation with a fine brush to remove potential epi-biota. NPP and R of each specimen were quantified by their respective oxygen (O_2) fluxes in the light and in the dark with a conductivity- and temperature-corrected O_2 optode probe (MultiLine[®] IDS 3430, WTW GmbH, Weilheim, Germany, accuracy: $\pm 0.5\%$ of measured value). Specimen BNF was quantified using an adapted acetylene (C_2H_2) reduction assay (Wilson and others 2012) during a full dark-light cycle incubation (24 h). Gas samples were collected from each chamber at time intervals and ethylene (C_2H_4) concentrations were measured using a customized reducing compound photometer (RCP) (Peak Laboratories, detection limit = 100 ppb). O_2 and C_2H_4 fluxes of the biological samples were corrected for unfiltered seawater control signals and normalized to incubation time and specimen 3D surface area. Unfiltered seawater signal was also recorded and normalized to per liter water volume.

Data Treatment

O_2 and C_2H_4 fluxes were converted into C and N fluxes assuming theoretical molar ratios of

$CO_2:O_2 = 1$ and $C_2H_4:N_2 = 4$. While we acknowledge that these ratios may vary depending on the biological system under study (Larkum 1988; Larkum and others 1988; Falkowski and Raven 2007), many systems display ratios not substantially different from these theoretical ratios (Capone 1988; Clavier and others 1994), and the use of the conservative theoretical ratios does allow here to compare GPP and BNF rates by different reef organisms. To estimate GPP, each specimen R measurement was added to its corresponding NPP measurement. NPP and GPP were extrapolated to per day estimates assuming 12 h of daylight, whereas daily R and BNF were calculated on a 24-h diel cycle. Daily C and N fluxes obtained for each benthic category were then normalized to the reef benthos 3D area considering the respective 2D:3D conversion factor and are reported in Table S4 expressed as $mmol (C \text{ or } N) m^{-2} \text{ benthos } 3D \text{ area } d^{-1}$. These represent fluxes of C and N from (or to) a square meter of reef benthos entirely composed by the respective category. Category-specific rates were then used to calculate the contribution of each benthic category to total benthic GPP, NPP, and BNF taking into account its relative 2D benthic cover in each reef habitat and season (Figure S1, Table S1, S2). Note that the physiological rates used here were measured under the average light intensity (10 m water depth) and may not capture the full depth-related profile of GPP and BNF on the reef. Consequently, our calculation only approximates the relative contribution of each reef habitat to reef-wide GPP and BNF. Nonetheless, it does provide a solid basis for assessing the overall significance and range of GPP and BNF that may occur at the reef ecosystem level. To allow comparison with other benthic systems, resulting contributions in $mmol (C \text{ or } N) d^{-1}$ were subsequently normalized to the reef planar 2D area and are expressed as $mmol (C \text{ or } N) m^{-2} \text{ planar } 2D \text{ area } d^{-1}$. To calculate the contribution of the water column to the reef ecosystem, GPP and BNF rates in $nmol (C \text{ or } N) m^{-3} d^{-1}$ (based on incubations of unfiltered seawater, Table S3) were extrapolated to the volume of the total reef area (Table S2) assuming relatively constant planktonic GPP and BNF in the surface mixed layer as it was previously observed in the Gulf of Aqaba (Foster and others 2009; Rahav and others 2015). We used the equations for the propagation of random errors (Miller and Miller 2005) to calculate uncertainties in the budget step-by-step calculations. The percentage of new production potentially achieved throughout N_2 fixation, as opposite to regenerated production that is supported by recycling of nutrients (Dugdale and

Goering 1967), was estimated using the percentage contribution of BNF to NPP, assuming a C:N ratio of 550:30 to estimate the demand of N for biomass generation of reef benthic autotrophs (Atkinson and Smith 1983; Atkinson and Falter 2003).

Statistical Analysis

Analyses were conducted using Primer-E v6 (Clarke and Gorley 2006) with the PERMANOVA extension (Anderson and others 2008). Differences in the benthic community cover among seasons (fixed, four levels) and reef habitats (fixed, five levels) were tested for significance using multivariate analysis of variance (PERMANOVA) on a Bray–Curtis similarity matrix of square-root transformed data. Type I (sequential) sum of squares was used with permutation of residuals under a reduced model (9999 permutations). Differences among habitats were visualized using principal coordinate analysis (PCO). Vector plots using Pearson ranking based on correlations greater than 0.4 were overlaid on the PCO plot to visualize relationships between variables and the ordination axes. Canonical analysis of principal coordinates (CAP, Anderson and others 2008) was used to identify specific groups in the benthic community responsible for differences among seasons, and benthic categories with correlations greater than 0.4 were overlaid on the plot as vectors. Differences in BNF, GPP, and R among seasons (four levels) were tested for significance using PERMANOVA on a Bray–Curtis similarity matrix of square-root transformed data. Type I (sequential) sum of

squares was used with unrestricted permutation of raw data (9999 permutations). If data did not conform to assumptions of homogeneity after transformation, tested with PERMDISP (Anderson and others 2008), a conservative α value of 0.01 was used (Underwood 1997). Differences among seasons were visualized using PCOs, with benthic categories correlation vectors overlaid on the plots.

RESULTS

Seasonal Environmental Conditions

Seawater temperature was stable around 22°C throughout winter and spring, but increased to 27°C in summer and decreased again in autumn (Table 1). Daily light availability increased from winter to spring and summer, and then decreased towards autumn. Inorganic nutrient and Chl *a* concentrations were significantly lower in summer and autumn than in winter and spring. Conversely, DOC showed higher concentrations in summer and autumn. POC concentrations did not differ between the seasons, while PN was lower in summer and autumn, with subsequent increase of the POC:PN ratio. Planktonic BNF was highly variable within seasons, with no significant seasonal differences, while planktonic GPP was significantly lower in summer and autumn compared to winter and spring (Table 1). Overall, the stratified season resembled warm and highly oligotrophic conditions, typical of tropical coral reefs worldwide, whereas the mixed season showed environmental conditions that are typical of marginal reef communities (Table 1, Kleypas and others 1999).

Table 1. Environmental Conditions during the Four Different Seasons at the Study Site (10 m water depth), and Comparison with Typical Values for Tropical Oligotrophic Waters as Reported by (a) Kleypas and others (1999); (b) Atkinson and Falter (2003); (c) Großkopf and others (2012); and (d) Torrèton and others (2010)

Parameter	Winter	Spring	Summer	Autumn	Typical conditions
Temperature (°C)	22.3 (0.1)	22.4 (0.1)	27.0 (0.2)	24.9 (0.1)	27.6 (1.1) ^a
Light (mol quanta m ⁻² d ⁻¹)	3.45 (0.26)	5.76 (0.13)	6.99 (0.32)	3.80 (0.34)	–
DIN (µM)	1.25 (0.13)	1.11 (0.07)	0.31 (0.06)	0.47 (0.09)	(0.1–1.1) ^b
DIP (µM)	0.11 (0.01)	0.10 (0.01)	0.04 (0.01)	0.04 (0.01)	(0.05–0.3) ^b
Chl <i>a</i> (µg L ⁻¹)	0.21 (0.01)	0.20 (0.02)	0.10 (0.00)	0.19 (0.02)	–
DOC (µM)	76.62 (3.82)	73.07 (3.13)	88.60 (1.93)	85.15 (1.46)	~100 ^b
POC (µM)	5.90 (0.56)	9.47 (1.39)	7.62 (1.01)	8.86 (0.38)	~10 ^b
PN (µM)	0.94 (0.05)	1.18 (0.14)	0.92 (0.11)	0.90 (0.04)	<1.0 ^b
POC:PN	6.81 (0.59)	7.95 (0.47)	8.25 (0.34)	9.86 (0.45)	–
BNF (nmol N L ⁻¹ d ⁻¹)	3.1 (4.8)	3.3 (5.6)	8.1 (9.2)	2.6 (4.0)	2.5 (0.0–12.4) ^c
GPP (µmol C L ⁻¹ d ⁻¹)	1.0 (0.4)	1.1 (0.7)	0.4 (0.8)	0.6 (0.2)	0.5 (0.1) ^d

Planktonic N₂ fixation rates are compared with BNF rates (¹⁵N dissolution method) obtained by Großkopf and others (2012) for oligotrophic ocean waters, while values from Torrèton and others (2010) are considered representative of GPP (¹⁴C-method) in coral reef waters isolated from terrestrial influence. Values are means with SD or range of values in parentheses.

Benthic Community Composition

Analysis of LPI data yielded the percentage coverage by all benthic categories as reported in Table S1. Multivariate analysis of the benthic community (Table S5) showed a significant effect of the factors 'Season' and 'Reef Habitat' (PERMANOVA: $P < 0.001$), but not of their interaction (PERMANOVA: $P > 0.1$). Distinctive communities (Figure 2) characterized the different reef habitats. The reef flat was correlated with coral rock, turf algae on rock, CCA, and zoanthids. The reef crest was dominated by coral rock, *Millepora*, turf algae on rock, and CCA. Conversely, the sand belt was correlated with 'bare' sediment, coral rubble, and turf algae on sediment, whereas hard and soft corals dominated the fore reef. The transition zone harbored an intermediate community between the sand belt and the fore reef, correlated with the presence of sponges, sediments, and microbial mats on sediment, but also hard and soft corals (Figure 2). Differences among seasons were not visible using the unconstrained ordination of the PCO technique. However, the CAP analysis revealed that seasonal changes were mainly attributable to macroalgae, microbial mats, and turf algae cover (Figure S1) that were higher in winter and spring (turf algae), only in spring (macroalgae and microbial mats on rock), or in spring and summer (microbial mats on sediment).

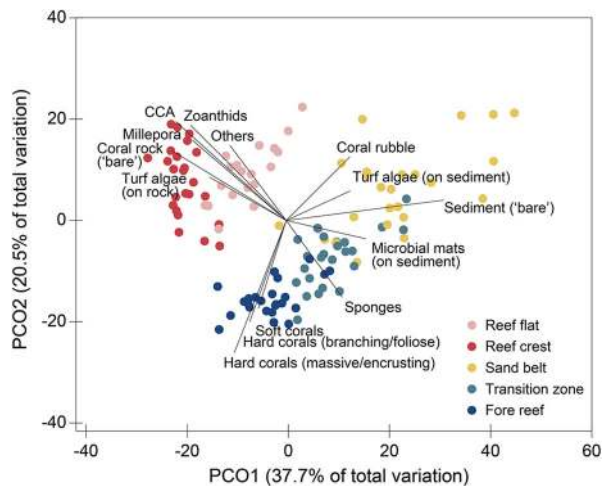


Figure 2. Principle coordinate analysis (PCO) of benthic community cover at the different reef habitats. Different colors represent different habitats as in the habitat map in Fig. 1. Vector overlay represents correlations greater than 0.4 based on Pearson ranking.

Metabolic Rates of Individual Benthic Categories

Benthic community metabolic rates showed significant differences between seasons (PERMANOVA: $P < 0.001$ and pair-wise comparisons, Table S6). GPP displayed a weak seasonal pattern, but generally increased in spring and summer compared to the other seasons for all benthic primary producers (Figure S2a). Hard corals, soft corals, microbial mats, and turf algae on rock displayed the highest rates of benthos 3D area-related GPP, whereas sediment showed the lowest rates (Table S4). R increased in summer for all benthic categories except for macroalgae and sediment that displayed higher R in spring compared to the other seasons (Figure S2b). The highest benthos 3D area-related R rates were associated with sponges and hard and soft corals, the lowest with sediment (Table S4). Compared to GPP and R, individual BNF rates showed a more pronounced seasonal pattern, with highest rates recorded in summer compared to all other seasons, except for sand-associated categories (macroalgae, sediment, and microbial mats) that showed the highest BNF in spring compared to the other seasons (Figure S2c). The highest benthos 3D area-related BNF rates were found for microbial mats on rock, the lowest for soft corals and sponges (Table S4).

Contribution of the Different Benthic Categories to Reef GPP and BNF

Hard corals contributed the largest share to benthic GPP (41–76%) over all seasons and in all reef habitats (Figure 3A), even in the sand belt where their benthic cover was less than 20% (Table S1). Soft corals contributed importantly to GPP in the fore reef (8–11%), in the transition zone (13–20%), and on the reef flat (7–16%), but their contribution was low in other reef habitats. The contribution of 'bare' sediment to GPP was only relevant in the sand belt (4–20%). 'Bare' hard substrates (that is, 'bare' coral rock and dead corals, see Table S3) contributed importantly to GPP in shallow and sandy reef habitats (that is, reef flat, reef crest, sand belt) in winter, summer, and autumn (11–26%), but their contribution was low in spring (5–8%) as a result of lower GPP rates (Figure S2, Table S4) and lower benthic coverage (Table S1). Turf algae provided a larger share of benthic GPP in shallow reef habitats (that is, reef flat, reef crest) in winter and spring (9–20%), but less in summer and autumn. Turf algae also contributed importantly in winter in the sand belt

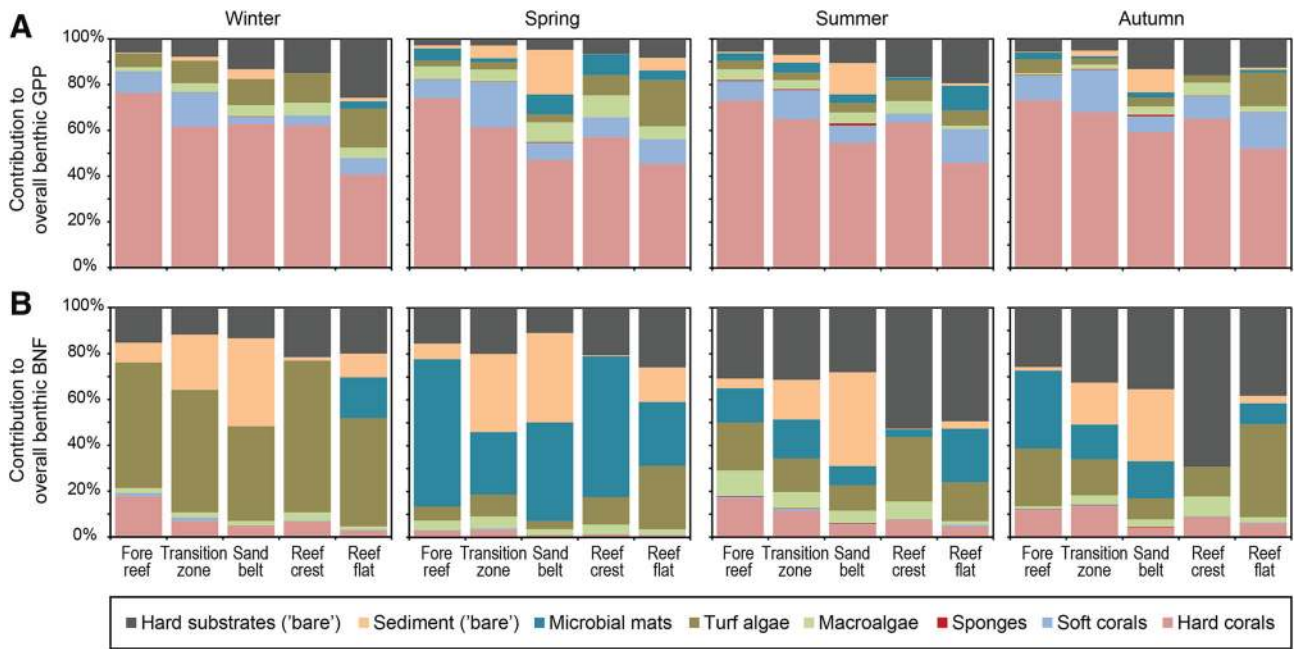


Figure 3. Contribution of the main benthic components to GPP (A) and BNF (B) for the four seasons and for each reef habitat. All hard coral morphologies are classified as one category; coral rock and dead corals are grouped into hard substrates ('bare'); macroalgae, microbial mats, and turf algae include categories 'on rock' and 'on sediment'.

Table 2. Community Dinitrogen Fixation (BNF), Gross Primary Production (GPP), Respiration (R), Net Primary Production (NPP) in $\text{mmol (C or N) m}^{-2}$ Planar 2D Area d^{-1} , and % Contribution of BNF to NPP in the Different Reef Habitats

Habitat	BNF (Larkum and others 1988)	BNF	GPP	R	NPP	% contribution
Water column	–	0.03 (0.02–0.06)	5 (3–7)	46 (30–55)	–	–
Reef flat	(0.67–1.33)	0.62 (0.22–1.18)	205 (178–240)	109 (86–145)	96 (88–112)	11.7 (4.4–22.9)
Reef crest	(0.05–0.11)	0.61 (0.15–1.07)	246 (222–266)	137 (121–165)	109 (88–139)	10.0 (2.6–19.2)
Sand belt	(0.02–0.04)	0.49 (0.14–0.83)	149 (117–173)	85 (75–97)	65 (42–95)	13.7 (4.0–23.8)
Transition zone	–	0.41 (0.14–0.80)	268 (234–303)	154 (134–186)	114 (95–156)	6.7 (2.6–14.2)
Fore reef	(0.18–0.40)	0.50 (0.10–0.78)	327 (284–359)	190 (167–223)	137 (116–187)	6.6 (1.6–12.0)

Values are means followed by the range in parentheses.

(11%), because of their high benthic coverage (Table S1). Macroalgae were unimportant to overall reef GPP, although their contribution was higher in shallow and sandy reef habitats in spring (6–10%) than in other seasons. Microbial mats contributed minor fractions to reef GPP, but their contribution was variable and sometimes important, depending on their benthic coverage and increasing up to 11% on the reef flat in summer (Figure 3A). Conversely, sponges showed negligible contributions to benthic GPP in all reef habitats and over all seasons.

Benthic categories contributing most to BNF were microbial mats, turf algae, 'bare' hard substrates, and 'bare' sediment (Figure 3B). Despite very low benthic coverage (0–1.7%) microbial mats provided a large fraction of BNF in spring in all reef habitats (27–64%), but their contribution to BNF was lower and variable in other seasons. Turf algae contributed the largest share of BNF in winter in all reef habitats (41–66%), but were also important in other seasons, particularly in the reef crest and reef flat (12–41%). 'Bare' hard substrates were important to BNF in all reef habitats and seasons (11–

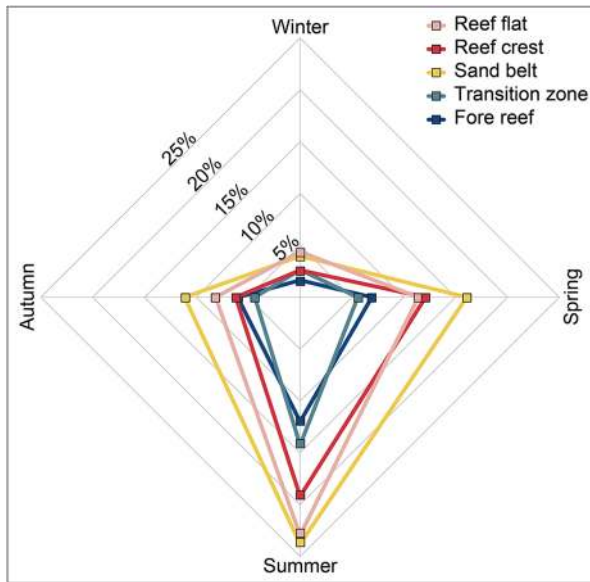


Figure 4. Contribution of BNF to NPP in all reef habitats and in all seasons. Different colors represent different habitats as in the habitat map in Fig. 1.

69%), particularly in summer and autumn in the reef crest and reef flat (38–69%). ‘Bare’ sediment contributed a large fraction of BNF in the sand belt in all seasons (31–41%), and was also important in the transition zone (17–34%). The contribution of hard corals was low but occasionally relevant (for example, in the fore reef, with up to 18% in winter and 17% in summer). Macroalgae showed significant individual BNF rates (Table S4), but very low benthic coverage (Table S1), consequently contributing a low fraction of reef BNF (Figure 3B). Sponges and soft corals showed negligible contributions to BNF in all habitats in all seasons (Figure 3B).

Areal GPP and BNF in the Different Reef Habitats

Areal physiological rates of BNF and GPP in the water column were respectively approximately 20- and 60-fold lower compared to areal rates of the benthos (Table 2). However, areal planktonic R rates were only about 3-fold lower than benthic R rates, and the water column was net heterotrophic throughout the year (Table 2). Areal BNF rates were highest on the reef flat and lowest on the fore reef, whereas GPP and R were lowest on the sand belt. In general, reef habitats with the highest live coral cover (that is, reef crest, transition zone, and fore reef, Table S1) showed higher areal GPP and NPP (Table 2). BNF contributed significantly to NPP in all reef habitats (Table 2). However, its contri-

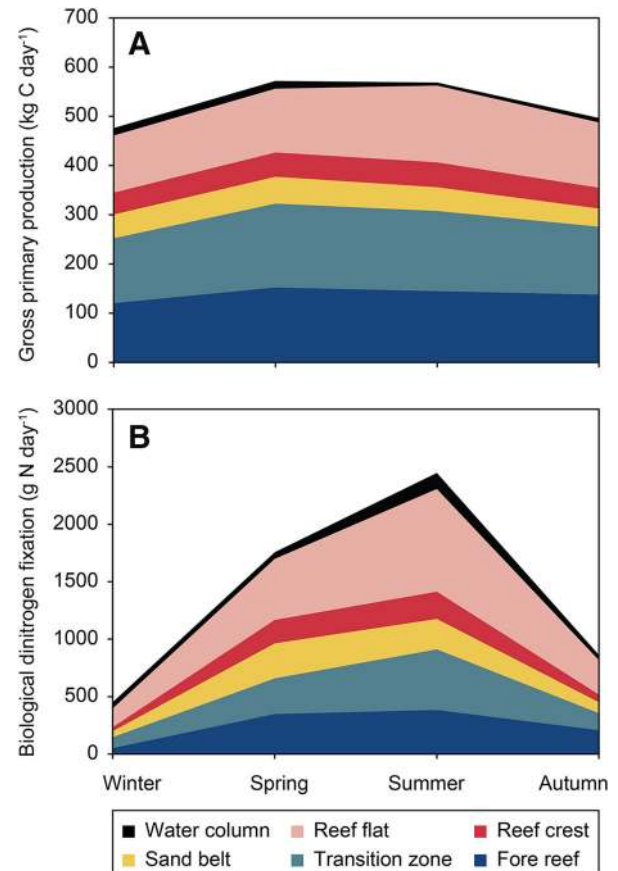


Figure 5. Contribution of each reef habitats to reef-wide GPP (A) and BNF (B) in the four seasons. Different colors represent different habitats as in the habitat map in Fig. 1, and black represents the contribution of the water column integrated for the entire reef water volume (see Table S2).

bution to NPP was highly seasonal, lowest in winter, and highest in summer in all reef habitats (Figure 4). The largest share of NPP was contributed by BNF in the sand belt, where NPP was lowest (Table 2; Figure 4), whereas contribution of BNF to NPP was generally low in the transition zone and fore reef (Figure 4). On average, BNF contributed about 10% of reef NPP on a yearly basis, while its contribution was 2-fold in summer (ca. 20%).

Reef-Wide GPP and BNF

The different reef habitats and the water column contributed differently to total reef-wide GPP and BNF (Figure 5A, B). The substantial areal extent and live coral cover of the fore reef, transition zone, and reef flat determined their important contribution to reef-wide GPP. Conversely, the sand belt and the reef crest contributed a lower share of reef-

wide GPP as these habitats show smaller areal extent (Figure 5A, Table S2). The contribution of the water column was always the lowest, and was insignificant for total reef-wide GPP (Figure 5A). To a lesser extent, this was also true for reef-wide BNF, where the contribution of the water column was comparable to that of benthic habitats only in winter (Figure 5B). Among benthic habitats, the reef flat contributed the largest share of reef-wide BNF, whereas the reef crest contributed the smallest (Figure 5B). BNF in the sand belt was highest in spring, whereas BNF in all other reef habitats and in the water column was highest in summer (Figure 5B). Overall, although total reef-wide GPP remained relatively stable year around (Figure 5B), total reef-wide BNF displayed a strong seasonal pattern with 4-fold higher rates in spring and summer compared to winter and autumn (Figure 5b).

DISCUSSION

Inputs of bioavailable N, particularly from BNF, may play a key role in the health and resilience of coral reef ecosystems (Wooldridge 2009; Holmes and Johnstone 2010; D'Angelo and Wiedenmann 2014). Thus, a detailed budget of nutrient fluxes in coral reefs is urgently needed for developing successful management actions (D'Angelo and Wiedenmann 2014). This study extends early fundamental work (Wiebe and others 1975; Larkum and others 1988) by generating a seasonal BNF budget for a coral reef community based on individual rates of all dominant benthic substrates and the reef-overlying water column. To our knowledge, this is the first attempt to combine substrate-specific GPP, R, and BNF rates extrapolated to their respective benthic 3D surface areas and to quantify their magnitude at the reef ecosystem level. Quantitatively expressing the uncertainty originating from direct measurements and natural variability in ecological studies involving step-by-step calculations is also important. In fact, this can improve the interpretation of data and simplify the estimation of risk associated with the management of an ecosystem (Lehrter and Cebrian 2010). Hence, in the present budget we used the equations for the propagation of random errors (Miller and Miller 2005) to constrain areal physiological rates with an assessment of the propagated uncertainty. GPP, R, and BNF rates presented here for the individual substrates fall well within those from previous studies that reported similar measurements from other reef environments (for example, see values in Nau-

mann and others 2013; Cardini and others 2014), and they appear robust against the associated propagated errors. However, there is considerable uncertainty in any such budget, and the increased complexity of the analyses introduced new sources of error in our budget. These include: (1) deviations from the theoretical $\text{CO}_2:\text{O}_2$ and $\text{C}_2\text{H}_4:\text{N}_2$ ratios in different benthic communities; (2) spatial variations in metabolic activity associated with the same substrates caused by the effect of water depth (for example, light availability) on the measured processes; (3) changes of ecosystem stoichiometry or deviations from the theoretical C:N ratio over space and time. Nonetheless, some results are solid and are thus discussed in the following sections, answering our key questions.

- (1) There was a clear decoupling between the substrates that contributed most to reef GPP and the ones that were important for reef BNF. This indicates that these two biogeochemical processes are uncoupled ecosystem functions where changes in the benthic community structure may differentially affect the availability of essential energy (C) or nutrients (N) at the ecosystem level. Hard corals were the key primary producers in all habitats and seasons, whereas categories such as turf algae, 'bare' hard substrates, and sediment were more important for reef BNF. This indicates that the latter substrates are also key constituents of a functioning coral reef as they provide important settlement space for diazotrophs (Wiebe and others 1975; Shashar and others 1994b; den Haan and others 2014; Rix and others 2015). Importantly, other reef organisms can provide either ecosystem functions, thanks to symbioses (for example, hard corals, Cardini and others 2015) or an inherent capability (for example, microbial mats, Charpy and others 2010). These benthic categories are fundamental to reef ecosystem GPP and BNF, with respective contributions strongly depending on their benthic coverage. For example, recently dead hard corals provide surfaces covered in organic detritus that can be hotspots of BNF. Fluxes of inorganic N following bleaching-induced mortality were estimated being sufficient to direct an affected ecosystem toward a phase shift from coral to algal dominance (Davey and others 2008; Holmes and Johnstone 2010).
- (2) In oligotrophic coral reefs such as the one investigated here, BNF and GPP are principally benthos-related processes (Atkinson 2011).

The entire water column above the reef contributed to BNF and GPP at rates per area that were approximately 20-fold and 60-fold lower, respectively, than those contributed by the benthos. The daily budget of GPP and R of the water column resulted in a negative metabolic balance in all seasons and in particular during the stratified summer period (van Hoytema and others 2016). This indicates net heterotrophy of the planktonic community. Similar conditions have been described for other reef planktonic communities, and it has been suggested that respiration of planktonic heterotrophs may be fueled by the supply of organic matter from the reef benthos (Naumann and others 2012a; Haas and others 2013). Additional evidence supporting net heterotrophy of the water column in the Gulf of Aqaba comes from a study that focused on the diazotrophic community and found a shift from mixed to predominantly heterotrophic diazotrophs in summer (Rahav and others 2015). In any case, the actual rates by which the reef biota can extract dissolved N from flowing seawater are physically constrained (Atkinson 2011). Conversely, BNF by benthos-associated prokaryotes has the potential to provide a source of N readily available to the reef benthos. This fixed N enters the food web through leakage of ammonium or dissolved organic N (DON), or by direct grazing, and can be assimilated *in situ* through processes that have limited or no interaction with the water column. On the benthos, reef habitats that are contiguous to the reef framework (that is, sand-dominated zones) contribute a large fraction of the newly fixed N to the reef ecosystem, as indicated by others (Capone and others 1992; Shashar and others 1994b; O'Neil and Capone 2008). However, the significance of the reef framework has previously been overlooked. In fact, these areas (for example, reef crest and fore reef) and the reef framework community in general, contribute newly fixed N at areal rates comparable if not higher than those of sand-dominated zones when accounting for their 3D surface area (Table 3). Moreover, our results confirm that high-coral cover habitats are unrivaled in terms of GPP, shaping coral reef ecosystem productivity, and growth.

(3) Despite the fact that the two processes of GPP and BNF appear to be provided by different benthic categories as uncoupled ecosystem functions, we did find a potentially strong biogeochemical link between the two at the ecosystem level. In fact, BNF could sustain

approximately 10% of NPP of the entire reef system on an annual basis, and up to about 20% of NPP in summer when inorganic nutrient concentrations were lowest. These results are similar to previous studies at other reef locations (Charpy-Roubaud and others 2001; Charpy and others 2007; Casareto and others 2008; Charpy and others 2010) and show that reef ecosystems, particularly those that experience oligotrophic conditions, rely significantly on BNF as an internal source of N to sustain their high ecosystem productivity. Our findings however confirm that BNF can only sustain a portion of reef NPP. Future studies should focus on the potentially important role of external sources (for example, of oceanic origin) of DON that may also sustain new production and growth in coral reefs. In fact, the DON pool is made up of poorly characterized compounds to which little attention has been given (see Suzuki and Casareto 2011), although mass balance calculations have highlighted its importance for benthic and pelagic production in tropical coral reef environments characterized by low inorganic N (Eyre and others 2008).

(4) Reef-wide GPP was increased in spring and summer (mostly driven by higher respiration, see also van Hoytema and others 2016), but relatively stable over the year despite strong seasonal shifts in environmental conditions. Conversely, reef-wide BNF was more susceptible to seasonality, being 5-fold higher in summer compared to winter. Several studies have shown that BNF is susceptible to changes in environmental factors such as temperature, nutrient concentrations, light, pH, and O₂, among others (see Carpenter and Capone (2008) and Cardini and others (2014) for syntheses). The present study further indicates that changes in BNF of single benthic categories have cascading effects at the reef ecosystem level. Moreover, although hard corals consistently showed the highest contribution to GPP in all habitats and seasons, substrates that mainly contributed to BNF changed from season to season, with turf algae contributing most in winter, microbial mats in spring, and 'bare' substrates in summer and autumn. Overall, these results suggest that climate exerts a strong control on BNF and that changes in benthic reef community composition drive strong feedbacks on the associated diazotrophic community and consequently on its provision of new N to the ecosystem.

Table 3. Estimate of the Total Contribution of BNF to the N Cycle in Benthic Coral Reef Environments, Compared with Previous Studies

Study location	Community	Time of year	Temperature (°C)	Areal rate ($\mu\text{mol N m}^{-2} \text{d}^{-1}$)	References
Aqaba, Jordan	Reef framework community	Annual average	(22.0–27.5)	536 (69)	This study
Aqaba, Jordan	Soft bottom community	Annual average	(22.0–27.5)	488 (62)	This study
Aqaba, Jordan	Reef framework community	Summer	27.0 (0.2)	956 (151)	This study
Aqaba, Jordan	Soft bottom community	Summer	27.0 (0.2)	723 (110)	This study
One Tree Reef	Various substrates	Annual range	(20–30)	(156–1330) ^a	Larkum and others (1988)
Eilat, Israel	Various substrates	Summer	n/a	(1152–1968) ^a	Shashar and others (1994b)

Values are means with propagated SE or range of values in parentheses.
^aCalculated values based on data from the original publications.

Responses to Environmental Forcing

Climatic controls on biogeochemical cycles are relevant in coral reefs because their biological activity is mainly driven by light intensity, water temperature, and nutrient availability. The fringing reef under study is exposed to seasonally changing environmental conditions. The stratified summer resembles typical conditions of tropical reef communities worldwide, which are characterized by warm and highly oligotrophic waters, whereas mixed season (winter) conditions are similar to marginal reef environments, where nutrient concentrations are often higher and temperature and light intensity are lower (Kleypas and others 1999). In the studied reef, BNF was highest in spring and summer, suggesting an important role of light availability, which showed a correlating seasonal pattern, as the main controlling environmental factor at the ecosystem level. The higher summer temperature (27°C) also likely exerted an important control on BNF, as summer individual BNF rates were highest of all seasons. These results are consistent with previous studies that found BNF on coral reef substrates to be primarily light and temperature dependent (Wilkinson and others 1985; Larkum and others 1988). Conversely, the relatively high spring DIN concentrations (>1 μM) did not cause a decrease of reef-wide BNF, suggesting that these concentrations were still low enough to prevent nitrogenase (that is, the enzyme responsible for BNF) inhibition (for example, Mulholland and others 2001). Moreover, BNF may still be a winning strategy in coral reef environments where energy sources are typically low in ammonium compared to carbon (that is, high C:N ratio, see

Atkinson 2011) and thus it is where organisms benefit from additional bioavailable N even at relatively high DIN concentrations. At this point however, it is difficult to predict which environmental factors dominate and how their complex interactive effects influence BNF in reef ecosystems, calling for experiments where separate environmental factors are manipulated to disentangle the single responses (for example, see Koop and others 2001).

Conclusions

GPP and BNF are closely linked in the marine environment for two reasons. First, BNF is a primary source of bioavailable N, which is a major component of chlorophyll, amino acids, energy-transfer compounds, and nucleic acids. Second, BNF requires energy that is supplied directly (photosynthetic diazotrophs) or indirectly (non-photosynthetic diazotrophs) by primary production. Thus, quantifying BNF is highly relevant for understanding the functioning of the marine environment. Here, we revised previous estimates of reef-wide BNF scaling physiological measurements for individual benthic categories to their 3D surface area and to the ecosystem level using a GIS analysis of the reef under study. No such detailed budget has been attempted to date for coral reef ecosystems. Whole-ecosystem measurements (for example, flow respirometry, eddy correlation) provide informative activity data on the entire community (Silverman and others 2007; Long and others 2013). However, there is not yet a whole-ecosystem method for reliably quantifying BNF. Moreover, our approach allowed us to differentiate

between the involved groups of organisms. At the same time, our whole-ecosystem estimates for GPP, R, and BNF fall in the range of those found for similar communities at other reef locations using both similar and different approaches (Atkinson and Grigg 1984; Hatcher 1988; Larkum and others 1988; Shashar and others 1994b; Silverman and others 2007; Long and others 2013; Eidens and others 2014), confirming the validity of our method. In Table 3, we present different estimates based on annual averages and on summer measurements only, as the latter ones may more closely approximate the scale and significance of BNF in other tropical reef systems (see section “Responses to Environmental Forcing”). These estimates, particularly those based on summer measurements, are similar to those presented by Larkum and others (1988) and by Shashar and others (1994b). However, importantly, the propagated errors associated with our estimates are relatively low despite taking into account the natural variability of a highly dynamic reef system, and should thus approximate the range of conditions found in other reef systems quite well. Consequently, the constrained estimates presented here of an important and so far poorly quantified reef N source can prove useful for environmental managers planning or evaluating the effects of nutrient management actions on the ecosystem biogeochemical cycling (for example, Boynton and others 2008). However, global and local anthropogenic environmental changes are predicted to profoundly alter coral reef communities and a variety of related ecosystem functions in the future (Hughes and others 2003; Hoegh-Guldberg 2011). As yet, large uncertainties remain in the potential responses of reef BNF and GPP to environmental change, requiring further assessments. It is clear, however, that the tight biogeochemical link between reef BNF and the C cycle will determine feedback loops, where perturbations in BNF will have repercussions in the C cycle, and vice versa. Ultimately, BNF dynamics need to be accounted for when projecting the future of coral reef ecosystem productivity in response to climate change.

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