

## Caloric restriction: powerful protection for the aging heart and vasculature

Edward P. Weiss<sup>1,2</sup> and Luigi Fontana<sup>2,3</sup>

<sup>1</sup>Department of Nutrition and Diabetics, Saint Louis University, Saint Louis, Missouri; <sup>2</sup>Division of Geriatrics and Nutritional Sciences, Department of Medicine, Washington University School of Medicine, Saint Louis, Missouri; and <sup>3</sup>Division of Nutrition and Aging, Istituto Superiore di Sanità, Rome, Italy

Submitted 8 July 2011; accepted in final form 8 August 2011

**Weiss EP, Fontana L.** Caloric restriction: powerful protection for the aging heart and vasculature. *Am J Physiol Heart Circ Physiol* 301: H1205–H1219, 2011. First published August 12, 2011; doi:10.1152/ajpheart.00685.2011.—Cardiovascular disease (CVD) is the leading cause of morbidity and mortality in the United States. Research has shown that the majority of the cardiometabolic alterations associated with an increased risk of CVD (e.g., insulin resistance/type 2 diabetes, abdominal obesity, dyslipidemia, hypertension, and inflammation) can be prevented, and even reversed, with the implementation of healthier diets and regular exercise. Data from animal and human studies indicate that more drastic interventions, i.e., caloric restriction with adequate nutrition (CR), may have additional beneficial effects on several metabolic and molecular factors that are modulating cardiovascular aging itself (e.g., cardiac and arterial stiffness and heart rate variability). The purpose of this article is to review the current knowledge on the effects of CR on the aging of the cardiovascular system and CVD risk in rodents, monkeys, and humans. Taken together, research shows that CR has numerous beneficial effects on the aging cardiovascular system, some of which are likely related to reductions in inflammation and oxidative stress. In the vasculature, CR appears to protect against endothelial dysfunction and arterial stiffness and attenuates atherogenesis by improving several cardiometabolic risk factors. In the heart, CR attenuates age-related changes in the myocardium (i.e., CR protects against fibrosis, reduces cardiomyocyte apoptosis, prevents myosin isoform shifts, etc.) and preserves or improves left ventricular diastolic function. These effects, in combination with other benefits of CR, such as protection against obesity, diabetes, hypertension, and cancer, suggest that CR may have a major beneficial effect on health span, life span, and quality of life in humans.

cardiovascular disease; inflammation; oxidative stress; arterial stiffness; cardiovascular aging

THIS ARTICLE is part of a collection on **Cardiovascular Response to Obesity and Diabetes**. Other articles appearing in this collection, as well as a full archive of all collections, can be found online at <http://ajpheart.physiology.org/>.

In the last century both life expectancy at birth and life expectancy beyond the age of 65 have markedly increased (146). The percentage of the total U.S. population over the age of 65 was 4% in 1900, has reached 13% in 2010, and is expected to increase to ~20% in 2030, amplifying concerns about health care spending. In fact, ~80% of older adults ( $\geq 65$  yr) have at least one chronic disease, and 50% have at least two chronic diseases, which often lead to disability (109).

Coronary heart disease, heart failure, stroke, and diabetes are the leading causes of morbidity and mortality, accounting for ~37% of total deaths in the United States (109). The incidence and prevalence of these diseases rise with advancing age because of the increased exposure time of the heart and arteries to harmful cardiometabolic risk factors (e.g., smoking, dyslip-

idemia, elevated blood pressure, insulin resistance, abdominal obesity, inflammation, and oxidative stress). Accordingly, participants of the Framingham Heart Study with low cardiometabolic risk (i.e., total cholesterol < 180 mg/dl, HDL-cholesterol > 40 mg/dl in men and > 50 mg/dl in women, blood pressure < 120/80 mmHg, fasting glycemia  $\leq 125$  mg/dl, body mass index < 25 kg/m<sup>2</sup>, and no smoking) at age 50 had substantially lower lifetime risk of developing cardiovascular disease (CVD) than participants with two or more major risk factors (5.2 vs. 68.9% in men; and 8.2 vs. 50.2% in women) (127). However, according to the most recent scientific literature, some of the criteria used in this study to define the low CVD risk profile are indeed nonoptimal. Optimal fasting glucose concentration should be below 86 mg/dl and LDL-cholesterol levels below 70 mg/dl, optimal blood pressure values should be below 115/75 mmHg, and optimal waist circumference should be  $\leq 94$  cm for men and  $\leq 88$  cm for women (3, 33, 41, 80, 145), suggesting that it is possible to achieve an even lower lifetime risk of developing coronary heart disease, heart failure, and stroke.

Nonetheless, aging has profound effects on the heart and arterial system, independently of the presence of clinical and subclinical CVD (114). There is a progressive deterioration in

Address for reprint requests and other correspondence: E. P. Weiss, Dept. of Nutrition and Diabetics, Doisy College of Health Sciences, St. Louis Univ., St. Louis, MO, 63104 (e-mail: eweiss4@slu.edu).

cardiovascular function and structure with advancing age, including a reduction in maximal heart rate (111), decreased heart rate variability (6, 60, 161), increased arterial stiffness (9, 10, 112, 134, 181, 192), diminished left ventricular systolic reverse capacity (112, 150), diastolic dysfunction (103, 112, 140), and impaired endothelial function (74). These anatomical and physiological changes that occur with normal aging and reduced physiological reserves of most body systems are not synonymous with disease but with an increased vulnerability to challenges which may decrease the ability of the organism to survive stressful conditions.

Several studies have now demonstrated that intrinsic cardiovascular aging can be affected by changes in food intake or mutations in single genes (61, 65, 80, 131, 200). Studies conducted on laboratory rodents have shown that calorie restriction without malnutrition (CR) promotes longevity and ameliorates the age-associated impairment of left ventricular diastolic function and arterial elasticity and improves heart rate variability (24, 130, 164, 179). The purpose of this article is to review the current knowledge on the effects of CR on the aging of the cardiovascular system and CVD risk in rodents, monkeys, and humans.

#### *Calorie Restriction Defined*

“Calorie restriction” refers to a state in which energy intake in animals or humans is minimized to low-normal levels while adequate intakes of protein and micronutrients are maintained at sufficient levels to avoid malnutrition. CR typically consists of an energy intake that is 30–50% below that which is required to maintain normal body weight and adiposity and thus results in a very lean phenotype. In the strict use of the term, CR is not an intervention in which excessive energy

intake is reduced. In animal studies, CR is typically introduced before physical maturation. Although it results in growth retardation, weight loss does not occur, which makes it clear that the health and longevity effects of CR in these studies are not attributable to weight loss. However, in human intervention studies, CR is introduced during adulthood and invariably results in weight loss, making it difficult to differentiate between the effects of weight loss and CR, per se. These and other aspects of animal and human CR study designs are presented in Table 1. The primary reason for studying CR in animals is to learn about human aging. In recent years, major advances in understanding the effects of CR on human aging and disease have been made by performing studies on human CR. However, despite this progress, none of the studies on humans can provide clear evidence of the effects of lifelong CR on life span or even major age-related diseases because of study design limitations.

The term “calorie restriction” has also been used less specifically to describe any reduction in energy intake, regardless of baseline energy intake, and is sometimes used to describe diet-induced weight loss in obesity. In this context, although calorie restriction results in weight loss, it is being used to alter the severity of a pathological/abnormal condition. This contrasts with the more strict use of the term, as described above, in which calorie restriction is intended to change a “normal” physiological state into “supernormal” state (i.e., a state in which physiological function is far better than that which would be necessary to be considered normal). For example, while a reduction in energy intake in overweight individuals has been shown to reduce serum C-reactive protein concentration from an abnormal level of 2.3 mg/l to a more clinically normal 1.6 mg/l (141), average serum C-reactive protein con-

Table 1. Comparison of animal versus human CR study design features

	Animal Studies	Human Studies
Energy intake	30-50% less than the intake of free-fed or modestly restricted animals (199)	Energy intake sufficiently low to achieve a lean phenotype or BMI of 18.5-21.0 kg/m <sup>2</sup> (64, 197) or energy intake that is sufficiently low to induce weight/fat loss in subjects with high-normal body weight or very mild overweight (i.e., BMI, 23.5-27.0 kg/m <sup>2</sup> ) (48, 86, 153)
Micronutrient and protein intake	Adequate to meet recommended intakes in some but not all studies (31)	Often stated that intakes are adequate to meet recommended intakes (64); however, micronutrient data have not been reported.
Initiation of CR	Typically after weaning but well before physical maturation (199)	Initiated in physically mature adults. Inevitably, this results in weight loss (48, 86, 153), making it difficult to differentiate between the effects of CR, per se, and weight loss.
Duration of CR	Generally lifelong after weaning (199)	6-12 mo in intervention trials (48, 86, 153); 2-30 yr in observational studies (64, 197)
Effects on growth	Growth retardation (131)	No growth retardation (initiated during adulthood)
Effects on body weight and fat mass	Due to growth retardation, body weight increases less than in control animals during early years; CR does not result in weight loss (199)	In response to the introduction of CR, body weight and fat mass decrease until energy balance is reestablished (48, 86, 153, 197). In cross-sectional research, body weight and fat mass are stable in CR practitioners (64).
Outcomes	Mean and maximal life span, disease prevention, and outcomes related to mechanism for life-span increases (65).	Risk factors for age-related diseases, biomarkers thought to reflect biological aging, and biomarkers shown in animal studies to change with CR (63, 67, 86, 89, 204).

CR, calorie restriction; BMI, body mass index.

centration in men and women practicing long-term CR is 0.2 mg/l (64), suggesting that CR results in a supernormal state of inflammatory control. This review article focuses specifically on the more strict use of the term “calorie restriction” as a means for optimizing the health and function of the cardiovascular function in the absence of overt preexisting disease. Studies in which dietary energy intake is reduced to correct obesity and related conditions will not be reviewed, as they are not within the scope of this paper.

Extensive research over the past seven decades has demonstrated that in animal species ranging from worms to rodents, 30–50% restriction of energy intake without introducing protein or micronutrient malnutrition (i.e., CR) increases life span by 30–50% (65). Part of this effect is mediated by preventing or postponing death because of chronic diseases such as cancer (up to 62% reduction in cancer incidence), obesity, type 2 diabetes, and autoimmune, cardiovascular, kidney, and neurodegenerative diseases (61, 131, 200). Furthermore, postmortem pathological studies have demonstrated that 30% of the rodents undergoing CR die in very old age without any evidence of lethal pathology (166), suggesting that it is possible to live a long life without overt disease. However, in addition to disease prevention, CR also slows the lifelong deterioration of structure and function in organs and tissues that occurs even in the absence of disease (61, 65, 131, 200). As a result, CR results in a more youthful biological phenotype and increases maximal life span (defined as the average age of the oldest 10% of animals in a cohort).

Ongoing studies are evaluating the effect of lifelong CR on aging and life span in nonhuman primates (i.e., Rhesus monkeys). Early evidence indicates that long-term CR results in many of the same adaptive responses that occur in rodents undergoing CR, such as increased insulin sensitivity, improved lipid profile, reduced blood pressure, decreased inflammatory and oxidative stress markers, lower serum triiodothyronine concentration and body temperature, and prevention of the age-associated decline in serum concentrations of dehydroepiandrosterone sulfate and melatonin (5, 61). The monkeys undergoing CR also appear to be partially protected against immune senescence, sarcopenia, and brain atrophy in subcortical regions that control motor and executive function (35, 36, 136). While it is premature to determine whether life span is increased by CR in these higher mammals, preliminary evidence based on small sample sizes suggests that CR may protect against cancer, CVD, and type 2 diabetes in monkeys (35). More definitive data on the effects of CR on life span in monkeys will likely be available in 10–15 years.

Data from studies on humans are far less definitive with respect to the effects of CR on life span than those from animal studies. The effect of CR on human longevity was evaluated in a retrospective analysis of data from men and women living in Okinawa Japan during the 1940s–1960s. The results indicate that these individuals were moderately calorie restricted (~1,785 kcal per day) with a lean phenotype (body mass index, ~21 kg/m<sup>2</sup>). When compared with residents of greater Japan and the United States, the Okinawans had markedly lower mortality from coronary heart disease and cancer and one of the highest numbers of centenarians in the world (~50 per 100,000 inhabitants) or about four to five times the average for most developed countries (93, 208). Studies have also shown that in individuals undergoing long-term self-imposed CR (i.e.,

~8 yr) and those undergoing 6–12 months of CR in randomized trials, CR results in adaptations that would be expected to reduce chronic disease risk, including greater insulin sensitivity, lower body fat levels, a better plasma lipid profile, lower blood pressure, less oxidative stress/damage, and lower levels of chronic systemic inflammation (61, 62, 64, 86, 89, 138). Furthermore, these studies also demonstrate that CR in humans results in lower leptin, sex hormones, and triiodothyronine hormone levels, lower core body temperature, and a lower metabolic rate (23, 63, 86, 169, 204), all of which are hallmark adaptations seen in response to CR in animals. However, a notable difference exists between human and animal studies of CR. In rodents, CR results in large reductions in insulin-like growth factor-1 (171); this adaptation may mediate the cancer-protective and life span-extending effects of CR (65). In contrast, in humans, CR does not lower insulin-like growth factor-1 levels unless protein intake is also restricted (67).

### *Cardiovascular System Aging and Calorie Restriction*

The cardiovascular system undergoes extensive structural and functional changes during the adult life span. Some of these changes are accelerated and exacerbated by the presence of harmful environmental and lifestyle factors and may eventually result in overt disease. For example, years of exposure to cardiometabolic risk factors cause atherosclerosis and may culminate in the rupture of an advanced plaque, ultimately resulting in ischemic heart disease and possibly heart failure. However, other age-related changes in the cardiovascular system may not necessarily be precursors to disease. For example, maximal heart rate decreases very predictably with increasing age (182, 203), yet this change is not a disease itself and is not known to be involved in disease pathogenesis. In this context, studies on the effects of CR on cardiovascular aging are especially interesting. Insights into the disease-protective effects of CR can be gained by evaluating the effects of CR on CVD risk factors and disease processes. However, because CR also appears to slow basic biological aging, independent of disease processes, studies on the effects of CR on disease-independent aspects of cardiovascular aging are important for learning about aging, per se.

### *Metabolic and Molecular Cardiovascular Targets of Calorie Restriction*

*Vascular oxidative stress.* Vascular aging is associated with increases in the production of reactive oxygen species (ROS) from vascular tissue mitochondria and decreases in endogenous antioxidant system activity (1, 44, 54, 92, 188, 193). In the healthy vasculature, oxidative stress activates the transcription factor NF-E2-related factor 2 (Nrf2), which moves into the nucleus, binds antioxidant-response elements, and induces proteins involved in protection against oxidative and free radical stress, including glutathione-S-transferases, NADPH:quinone oxidoreductase 1, and heme oxygenase 1 (91, 106, 144). With increasing age, the oxidative stress-induced activation of Nrf2 and the expression of its target genes is attenuated or becomes entirely absent (187, 190), thereby downregulating antioxidant system activity. This, together with increases in the production of ROS (92, 188, 193), results in increases in oxidative stress and damage that are common in old age (12, 187) (Fig. 1).



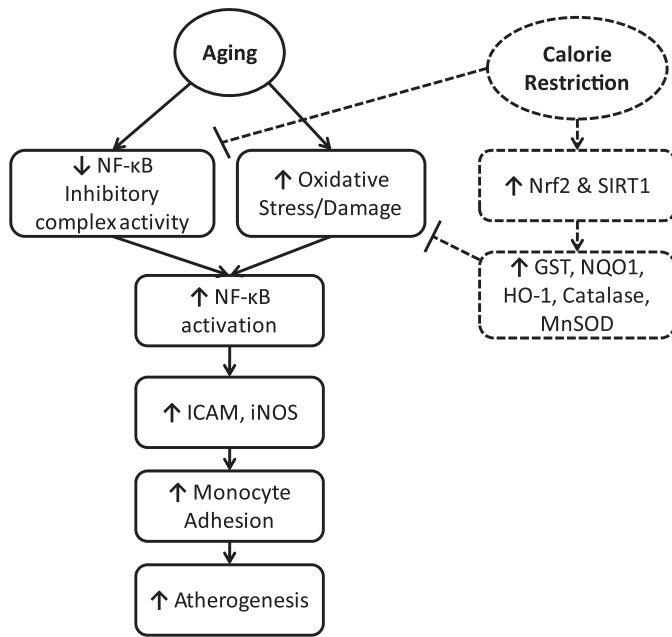


Fig. 1. The effects of aging and calorie restriction (CR) on oxidative stress and inflammation in the vasculature and their relationship to atherosclerosis. Increasing age is associated with increased production of reactive oxygen species, which results in oxidative stress and damage. Aging is also associated with a reduction in the inhibitory complex that prevents the transcription factor NF- $\kappa$ B from entering the nucleus where it can induce inflammatory genes. Together these factors result in increased expression of adhesion molecules (ICAM-1) and the proinflammatory enzyme inducible nitric oxide synthase (iNOS), thereby promoting monocytes adhesion to the endothelium and intimal infiltration and consequently initiating atherosclerosis. CR attenuates the age-related loss of NF- $\kappa$ B inhibition, thereby keeping NF- $\kappa$ B in the inactive state. Furthermore, by activating transcription factor NF-E2-related factor 2 (Nrf2) and sirtuin 1 (SIRT1), CR induces proteins involved in protection against oxidative and free radical stress including glutathione-S-transferases (GST), NADPH:quinone oxidoreductase 1 (NQO1), and heme oxygenase 1 (HO-1), manganese superoxide dismutase (MnSOD), and catalase. Together, these effects of CR prevent the NF- $\kappa$ B-mediated vascular inflammatory response and attenuate the age-related increase in atherosclerosis.

It has been theorized that CR slows the aging process, at least partly by attenuating the age-related increases in oxidative stress and the accumulation of oxidative damage (170). In support of this theory, at least in the context of the vasculature, CR has been shown to attenuate the production of ROS and oxidative damage (43, 80, 189) and increases levels of the endogenous antioxidants glutathione and ascorbate in the aorta (43). As has been found in other cells/tissues, the histone deacetylase and gene repressor sirtuin 1 (SIRT1) may be involved in the antioxidant effects of CR, as serum from CR animals induced SIRT1 in cultured endothelial cells and SIRT1 knockdown attenuated the CR-mediated reduction in ROS production (43). CR also induces endogenous antioxidant proteins by activating Nrf2 (149) (Fig. 1). Studies on humans also support the notion that CR attenuates oxidative stress. Twelve months of CR in humans have been shown to increase the activity of the antioxidant glutathione peroxidase in plasma (137), decrease plasma protein carbonyl levels as a marker of oxidative damage (137), and lower levels of oxidative damage to DNA and RNA in white blood cells and urine (86, 89).

**Vascular inflammation.** In addition to the direct effect of oxidative stress on the age-related deterioration of vascular

structure and function, oxidative stress also increases vascular inflammation including “endothelial activation” [i.e., increased expression of leukocyte recognition and adhesion molecules, a phenotypic change from antithrombotic to prothrombotic, and increased expression of cytokines and growth factors (17)]. As inflammation has been implicated as a major contributor to atherosclerosis (125), this may contribute to the development of atherosclerosis with increasing age. At least part of the mechanism by which oxidative stress promotes inflammation appears to involve the activation of the redox-sensitive transcription factor NF- $\kappa$ B (54, 177, 188), which has been implicated as an important contributor to atherosclerosis (83) (Fig. 1). Oxidative stress-induced activation of NF- $\kappa$ B has been shown to increase vascular expression of adhesion molecules and inducible nitric oxide (NO) synthase and to increase monocytes adhesion to endothelial cells (188), all of which promote atherosclerosis.

CR has several effects on vascular inflammation (Fig. 1). CR attenuates the age-related increase in expression of vascular adhesion molecules, an effect that coincided with reductions in ROS production (218). This effect of CR appears to be mediated by the prevention of the age-related reductions in NF- $\kappa$ B inhibitory complexes, thereby preventing NF- $\kappa$ B from entering the nucleus to promote inflammatory gene transcription. CR has also been shown to attenuate age-related increases in prostanoids in both serum (prostaglandin E<sub>2</sub>) and in the aorta (prostaglandin E<sub>2</sub> and thromboxanes A<sub>2</sub>); this effect appears to be mediated by an attenuation of the age-related increases in cyclooxygenase 2 and cytosolic phospholipase A<sub>2</sub>, both of which are involved in prostanoid synthesis. Although the mechanism for cyclooxygenase 2 and cytosolic phospholipase A<sub>2</sub> downregulation by CR is not clear, it is possible that NF- $\kappa$ B is involved because the genes for these proteins have NF- $\kappa$ B binding sites in their promoter regions (128, 211). Aging is also associated with increases in the proinflammatory cytokines in vascular tissue and serum from rodents (TNF- $\alpha$ , IL-1 $\beta$ , and IL-6) (45, 46) and in serum from humans (TNF- $\alpha$ , TNF receptor-1, and IL-6) (148, 175). CR has been shown to decrease levels of several inflammatory cytokines in rodents (80, 95, 172) and with seven to eight years of calorie restriction in humans (64) but not with shorter-term CR (i.e., 6–12 mo) in humans (118, 202).

#### *Effects of Calorie Restriction on Vascular Health and Function*

**Endothelial function.** Another means by which oxidative stress contributes to age-related vascular dysfunction is by altering the bioavailability of endothelium-derived free radical, NO. In the presence of oxidative stress, NO bonds with ROS, thereby eliminating its bioactivity. Furthermore, age-related reductions in endothelial production of NO also likely contribute to lower NO bioavailability, as a result of several age-related changes including reductions in the expression of endothelial NO synthase (eNOS) (44, 180), decreases in tetrahydrobiopterin (eNOS cofactor) levels (168), and decreases in intracellular L-arginine (eNOS substrate for NO production) availability (16). Among other compounds produced by the endothelium, NO serves an important role in the maintenance of vascular health and in preventing atherosclerosis, since it inhibits platelet and leukocyte adherence to the endothelium

(108), prevents platelet aggregation (174), and suppresses vascular smooth muscle cell proliferation (73). Endothelium-derived NO causes vascular smooth muscle relaxation and is the most potent endogenous vasodilator known (39). NO-mediated vasodilation is commonly measured as an index of NO bioavailability (152). In both rodents and humans, impaired NO-mediated vasodilation (i.e., endothelial dysfunction) develops with increasing age in both conduit and resistance vessels (15, 29, 97, 178). Not only does this blunt the minute-by-minute adjustments in blood vessel diameter that are important for regulating blood flow to target organs such as the heart (1, 191) and skeletal muscle (151, 185), but it is also associated with a greater likelihood of developing coronary artery disease (165, 213).

CR preserves endothelial function with advancing age through two known mechanisms. As reviewed earlier in this article, CR attenuates age-related increases in oxidative stress. This attenuates the degree to which NO is inactivated by ROS, improves NO bioavailability, and thus preserves endothelial function (43, 157, 214). It does not appear that these effects are mediated by alterations in the sensitivity of vascular smooth muscle to NO, as neither aging or CR alter the vasodilatory response to the endothelium-independent NO donor, *S*-nitroso-*N*-acetyl-penicillamine (43). CR also appears to prevent age-related losses in endothelial function by attenuating the age-related decrease in eNOS expression (157, 189) and activity (214). Although the means by which CR increases eNOS activity are not clear, SIRT1, which is induced by CR, has been shown to associate with eNOS in the cytosol of vascular endothelial cells, deacetylate eNOS, and increase eNOS activity (133). Furthermore, CR results in elevated levels of circulating adiponectin in both humans and rodents (62, 202, 217), which may activate eNOS in vascular tissue through a pathway involving the phosphorylation/activation of AMP-activated protein kinase (53). CR has also been shown to increase Akt phosphorylation in heart tissue (75), which may contribute to eNOS activation (71); however, it is not known whether CR alters Akt activity in the vasculature. Whether CR increases eNOS activity by altering levels of the eNOS cofactor tetrahydrobiopterin or the eNOS substrate *L*-arginine is not known.

**Arterial stiffness.** Aging is associated with increases in the stiffness of the large elastic arteries (102, 192). This hardening of the arteries occurs throughout adulthood (184) and in the absence of atherosclerosis (8, 192), thereby making it an attractive biomarker for biological aging. However, the process is also accelerated in the presence of diseases including hypertension, diabetes, and kidney dysfunction (162), and greater stiffness is associated with future development of coronary artery disease, and all cause mortality (194, 210). Age-related increases in arterial stiffness are primarily caused by structural alterations in the media including increases in collagen content (173), decreases in elastin (34), decreases in elastin cross linkage (which are responsible for elasticity) (198), accumulation of advanced glycation end products (163), smooth muscle atrophy (184), and calcification, especially in the elastin-rich regions (56). Age-related alterations also occur in the intima; however, these changes are more closely associated with atherogenesis, as covered later in this review.

CR has been shown to attenuate age-related increases in arterial stiffness, at least in animal models. In older rats, CR resulted in less arterial stiffness compared with free-fed rats, as

evidenced by greater fractional expansion of the aorta during the cardiac cycle (i.e., arterial distensibility) and slower pulse wave velocity (2). These effects of CR are accompanied by less fibrous matrix (i.e., collagen) accumulation, more elastin, and better preservation of vascular smooth muscle (2, 68). Furthermore, CR appears to prevent age-related vascular accumulation of the proteoglycan decorin (68), which has been shown to promote vascular fibrosis and mineralization through the activation of TGF- $\beta$  (158, 212). Less arterial stiffness has also been shown in arteries isolated from spontaneously hypertensive rats exposed to short-term CR (i.e., 5 wk) (53). To our knowledge, studies have not been published on the effects of CR on arterial stiffness in humans.

The mechanisms by which CR attenuates age-related increases in arterial stiffness may involve reductions in oxidative stress and systemic inflammation and alterations in NO bioavailability. Age-related increases in oxidative stress promote the production and accumulation of lipid peroxidation products including 4-hydroxynonenal (HNE) in the rodent aorta (24). HNE is known to promote fibrosis by inducing the fibrinogenic cytokine TGF- $\beta$  (122), at least partly through the activation of activator protein-1, which is a transcription factor for the TGF- $\beta$  gene (21). CR in rodents has been shown to attenuate the age-related increases in aortic HNE, TGF- $\beta$ , and fibrosis (24). The reductions in oxidative stress that result from CR, as well as increased eNOS levels, contribute to greater NO bioavailability in old age, which can also contribute to CR-mediated reductions in arterial stiffness (207). With respect to chronic inflammation, both cross-sectional (94, 196) and intervention studies (195) have demonstrated that systemic inflammation increases arterial stiffness. Furthermore, inflammation-reducing medications, including TNF- $\alpha$  antagonists and statins, have been shown to decrease arterial stiffness (72, 142). As outlined earlier in this article, CR has a potent anti-inflammatory effect in the vasculature, and this may be partly responsible for the protection against the development of arterial stiffness with increasing age.

**Atherosclerosis.** The development of atherosclerosis is a complex process and is not fully understood. Because atherogenesis can occur for many decades before atheromas develop and because many of the physiological conditions that promote atherogenesis (i.e., oxidative stress, chronic systemic inflammation, type 2 diabetes, endothelial dysfunction, high blood pressure, and dyslipidemia) develop in older age (18, 29, 70, 77, 82, 139, 186), advanced atherosclerotic lesions and their clinical sequelae (such as coronary artery disease and cerebrovascular disease) are far more common in older age than in young adulthood (114).

It is beyond the scope of this review to provide a detailed description of the mechanisms involved in atherogenesis; comprehensive reviews have been published elsewhere (84, 125). However, in brief, and as described in the review articles (84, 125), atherogenesis is a three-stage process. The earliest stage involves "injury" to the endothelium, resulting from oxidative stress and other factors. Blood constituents such as oxidized LDL leak through the injured endothelium and trigger an inflammatory response, in which macrophages phagocytose the invading agents to form foam cells. During the second stage, a fibrous cap consisting of smooth muscle, collagen, and calcium forms on the developing plaque to provide structural stability. Within the plaque, a core containing free cholesterol/lipid and

necrotic tissue forms and becomes enriched with procoagulants. At the most advanced stage, focal degradation of collagen and thinning of the fibrous cap make the plaque prone to rupture. Plaque rupture results in rapid activation of coagulation, arterial lumen obstruction, and ischemia to downstream tissues.

CR may protect against atherogenesis; however, data are limited. A major limitation to using rodents for studies on the effect of CR on atherogenesis is that rodents are generally not prone to atherosclerosis (primary cause of death is generally cancer). However, one study circumvented this problem by using a genetically engineered mouse model that is prone to atherosclerosis (i.e., the homozygous *apolipoprotein E* knockout) (80). In this study, mice undergoing 60% CR and those in a free-feeding control group all had early stage atherosclerosis, as evidenced by the presence of foam cells and free lipids in the wall of the aorta. However, the atherosclerotic lesions were 33% smaller in the CR group. Furthermore, CR appears to have blunted the development of more advanced atherosclerotic plaques, as evidenced by half as many mice with evidence of fibrous caps and plaque calcification and two-thirds fewer plaques with an "acellular" necrotic lipid core. These intriguing data were accompanied by lower levels of lipid hydroperoxide (oxidative damage marker) in the aorta and lower levels of the ROS production (i.e., superoxide and hydrogen peroxide) (80).

Preliminary data have been published on the effects of CR on atherosclerosis in nonhuman primates. In a preliminary report from a 20-year longitudinal study of Rhesus macaques (CR,  $n = 38$ ; and control,  $n = 38$ ) in which longevity will eventually be the primary outcome, 13% ( $n = 5$ ) of the CR-group monkeys have died from age-related causes, which is significantly fewer ( $P = 0.03$ ) than that in the control group (37%,  $n = 14$ ), indicating a significant longevity-enhancing effect of CR. Although only half as many monkeys in the CR group died of CVD (CR group,  $n = 2$ ; and control group,  $n = 4$ ), the number of cardiovascular deaths is far too small to make any conclusions. Furthermore, most of the CVD deaths were attributed to valvular endocardiosis, cardiomyopathy, and myocardial fibrosis, and thus these data do not provide insights regarding the effects of CR on atherogenesis. Another study of nonhuman primates used cynomolgus monkeys (26, 28), which are reported to be a good model for human atherosclerosis (201). The feeding intervention was four years in duration, was initiated in adulthood, and provided equal amounts of dietary cholesterol in the CR and control groups so that the effect of CR, per se, could be evaluated. Results showed no difference between the CR and control groups in terms of intimal thickness, as an index of atherosclerosis extent, in the coronary and abdominal arteries (26, 28). However, it has been argued that intimal thickening is not necessarily indicative of atherosclerosis, per se. Rather, a thicker intima simply reflects an aging artery that is more prone to atherogenesis (114). In this context, the data showing that CR in monkeys does not affect intimal thickness could be interpreted to indicate that four years of CR in adult monkeys is not sufficient for altering primary vascular aging, without drawing conclusions about the effects of CR on atherosclerosis.

The only data on the effect of CR on atherosclerosis in humans are based on the observational data from Japanese Okinawans. Men and women living in Okinawa during the

1940s–1960s were estimated to have been calorie restricted without evidence of malnutrition (208). Age-adjusted mortality from coronary heart disease in these individuals was 49% lower than that of other Japanese men and women and 87% lower than individuals residing in the United States (208). While this study provides evidence that moderate CR provides powerful protection against coronary artery disease, the information should be interpreted cautiously, as numerous other aspects of the Okinawans' diet (or lifestyle in general) could have contributed to protection against atherosclerosis. For example, they had high intakes of antioxidants from vegetables and fruits and their diet was rich in isoflavones from soy and other legumes; furthermore, they rarely consumed processed foods such as refined sugar (209).

*Risk factors for atherosclerosis.* In light of the difficulties in evaluating the effect of CR on atherogenesis, several studies have evaluated the effect of CR on risk factors for atherosclerosis. Not surprisingly, CR results in lower body weight and lower levels of both total and central/visceral fat mass in studies on numerous species including monkeys (27, 28, 37) and humans (49, 64, 153, 155). In humans, these changes corresponded with reductions in adipocyte size and with lower hepatic lipid content; however, intramyocellular lipid was not altered by CR (117).

One of the hallmark characteristics of CR in rodents and monkeys is lower plasma insulin levels and improved glucose regulation (14, 27, 79, 96, 101, 115, 132). Furthermore, 20 years of CR in monkeys has been shown to completely prevent glucoregulatory impairment, whereas in the control animals, 13% have developed diabetes and an additional 29% have developed prediabetes (35). While improvements in glycemic control have been hypothesized to contribute to the antiaging effect of CR (30), they would also be expected to protect against atherosclerosis and CVD, as hyperglycemia, insulin resistance, and diabetes are associated with atherosclerosis and CVD risk (13, 69, 76, 90, 215). Three randomized trials on men and women have demonstrated clear improvements in glucose regulation in response to CR. Two of these trials showed that CR decreases fasting insulin with no change in fasting glucose, indicating improved insulin action (48, 117); one of these trials also showed a strong tendency for CR-induced improvements in insulin action as measured by using an intravenous glucose tolerance test (117). Our trial showed that 20% CR for 12 months reduced fasting insulin (−37%) and oral glucose tolerance test insulin (area under curve, −24%); this, paired with a tendency for lower glycemia, resulted in a 26% improvement in insulin action (202). Additional insights about the effects of CR in humans comes from the Biosphere-2 experiment, in which eight men and women were sealed into a closed 3.14-acre dome for two years for the purposes of studying ecological factors that influence the earth's biosphere (4). During the two-year experiment, food production was inadequate to meet the needs of the crew, and as a result, they were calorie restricted but consumed a micronutrient-rich diet. Relative to preentry levels, fasting glucose decreased by 21% and fasting insulin decreased by 42% (197), suggesting a profound improvement in glucose regulation. While data from the randomized trials, the Biosphere-2 experiment, and the studies on nonhuman primates show clear and powerful effects of CR to improve glucose regulation, the results from our observational study on adults undergoing 3–20 years of strict self-imposed



CR are not as clear. When compared with nonobese control subjects consuming a typical western diet, the CR practitioners had 13% lower fasting glucose and 80% lower fasting insulin levels, suggesting that CR improved insulin action as shown in the other studies. However, 2 h glucose from an oral glucose tolerance test was 14% higher in the CR group, an effect that may have been attributable to somewhat lower postprandial insulin levels. Furthermore, 39% of the CR practitioners met the criterion for impaired glucose tolerance and another 21% had high-normal 2-h glucose values. An explanation for this unanticipated finding is not clear, and it is not known how this finding of low-fasting glycemia paired with high-postprandial glycemia would affect CVD risk. However, the relationship between aging/aging-related diseases and insulin resistance is confounded by associated factors such as excessive abdominal adiposity, decreased physical activity, hyperinsulinemia, dyslipidemia, inflammation, and other metabolic and hormonal components of the metabolic syndrome (14, 154). Interestingly, the CR individuals with high-postprandial glycemia values were extremely lean and had very low fasting plasma concentrations of glucose and insulin and an outstanding metabolic profile (very low triglyceride, high HDL-cholesterol, high adiponectin, and extremely low C-reactive protein concentrations) (62).

Studies on the effects of CR on blood lipids and lipoproteins in primates have yielded mixed results. Studies of adult cynomolgus monkeys exposed to four years of CR on a diet that was enriched with crystalline cholesterol (i.e., dietary cholesterol was equal in the CR and control groups) generally revealed negative findings. CR did not affect serum concentrations of total, HDL-, LDL-, or very low-density lipoprotein-cholesterol, triglycerides, or intimal thickness (28). Furthermore, despite strong evidence that CR reduces oxidative stress in the vasculature (reviewed above), LDL oxidation was not affected by CR (25). These findings suggest that CR, in the absence of a reduction in dietary cholesterol, does not alter lipid profile. In studies that did not enrich the diet with cholesterol and which also used a different species of monkey (i.e., rhesus), CR resulted in a very large 50% reduction in plasma triglycerides but did not alter total, HDL-, or LDL-cholesterol (55). Although LDL oxidation was not altered by CR (25), LDL particles from CR animals were found to have less proteoglycan binding (55), which would be expected to result in lower levels of LDL in the arterial wall, thereby making the LDL particles less atherogenic (22, 126). This

finding is supported by research on rodents showing that CR attenuated the age-related increase in elastin-associated LDL in the aortic wall (68). Plasma levels of lipoprotein (a) were lower in the CR monkeys, which is another adaptation that would be expected to reduce atherosclerotic disease risk (47, 216). Furthermore, although HDL-cholesterol was not altered by CR, the HDL<sub>2b</sub> subfraction was higher in CR monkeys. HDL<sub>2b</sub> is the HDL subfraction that is most closely associated protection against atherosclerosis (20, 98) and plasma concentrations were also found to be 38% higher in human centenarians compared with normolipidemic healthy weight controls (11).

In CR studies on humans, the effects of CR on lipoproteins are profound (Table 2). In the randomized intervention trials, total and LDL-cholesterol decreased by 5–15% (48, 66, 121), whereas in observational trials on more strict CR in humans, total, and LDL-cholesterol levels were ~30% lower than in control subjects with total cholesterol values of 130–160 mg/dl in the CR group (64, 197). Although one randomized trial and one observational trial reported increases in HDL-cholesterol (48, 64), others reported no change in HDL-cholesterol (66, 121) and one reported a large reduction (197). Large reductions in plasma triglyceride concentrations were seen in all studies with CR resulting in an average triglyceride concentration of ~85 mg/dl (48, 64, 66, 121, 197).

Blood pressure was not affected by four years of CR in adult cynomolgus monkeys (28). However, although there was no evidence of an age-related increase in blood pressure in rhesus monkeys, CR resulted in ~10 mmHg lower systolic and diastolic blood pressures (116). Randomized controlled trials on human CR have also produced equivocal results for blood pressure (Table 2). One reported no effect of 6 months of CR on blood pressure. While the other trial also reported no effect of CR on blood pressure (66), a subsequent analysis which excluded subjects who were not compliant with the CR intervention revealed a modest reduction in blood pressure (i.e., 10 mmHg systolic, and 6 mmHg diastolic) in response to 12 months of CR (156) (Table 2). Observational trials on longer-term strict CR showed substantial blood pressure lowering effects of 20–25% with CR, resulting in systolic pressures of ~95 mmHg and diastolic pressures of ~60 mmHg (64, 197).

#### *Effects of Calorie Restriction on Cardiac Function*

Increasing age is associated with a thickening of the myocardium and an increase in left ventricular mass/hypertrophy,

Table 2. Result from human trials on the effects of CR on plasma lipid concentrations, BP, and CRP

	TC	LDL-C	HDL-C	TG	Systolic BP	Diastolic BP	CRP
Randomized trials with 6-12-mo interventions							
Tufts University (48)	↓ 5	↓ 7	↑ 13	↓ 16	NR	NR	NR
PBRC (121)	NR	↓ 6	(↑ 9)	↓ 21	(↓ 2)	(↓ 2)	(↓ 29)
Washington University (66, 156)	↓ 10	↓ 14	(↑ 6)	↓ 23	↓ 8	↓ 9	(↓ 30)
Observational trials of long-term strict CR							
Biosphere 2 (197)	↓ 30	↓ 33	↓ 27	↓ 22	↓ 25	↓ 22	NR
CRON (64)	↓ 23	↓ 32	↑ 24	↓ 67	↓ 23	↓ 23	↓ 81
Mean of all trials	↓ 17	↓ 18	↑ 5	↓ 30	↓ 15	↓ 14	↓ 47

Values are percent changes from baseline. Values in parentheses were not significantly different from the control group in the Pennington Biomedical Research Center (PBRC), Washington University, and Washington University study on self-imposed calorie restriction with optimal nutrition (CRON) studies or were not significantly different from zero in the Tufts University and Biosphere-2 studies (these studies did not include a control group). BP, blood pressure; CRP, C-reactive protein; TC, total cholesterol; C, cholesterol; TG, tryglycerides; NR, not reported.

even after accounting for age-related increases in blood pressure, obesity, and vascular and valvular heart disease (124). Furthermore, aging is associated with myocardial fibrosis (40), increases in the number of collagen cross-linkages (19), and increases in the size and decreases in the number of cardiomyocytes (7, 147). These alterations result in an increase in stiffness, a reduction elastic recoil, a reduction in the passive suction-mediated early diastolic filling, and a greater reliance on the atrium for diastolic filling (32, 103, 140). There is also an age-related shift in myosin isoform in cardiac muscle from the fast VI ( $\alpha$ -myosin heavy chain) isoform to the slow V3 ( $\beta$ -myosin heavy chain) isoform (38, 129, 160), which may contribute to the slower contraction and relaxation of cardiac muscle seen in older hearts (110). Furthermore, age-related increases in arterial stiffness, as reviewed earlier in this article, result in a greater afterload on the heart (111). Although systolic function (based on ejection fraction) is generally preserved with increasing age in individuals without CVD (59), the greater myocardial work required to overcome the increased afterload contributes to further cardiac hypertrophy (32).

Studies on rodents have demonstrated that CR prevents part of the age-related deterioration in diastolic filling as evidenced by a greater ratio of Doppler-measured earlier to late diastolic filling velocity (i.e., E-wave-to-A-wave ratio) (179). Similar effects of CR have been demonstrated in nonobese spontaneously hypertensive rats (53) and in Dahl salt-sensitive rats (164), even in the presence of a high-salt diet (164). These effects may be partly mediated by reductions in blood pressure but were also accompanied by reductions in inflammatory cytokines (164) and improvements in vascular compliance and function (53). Histological studies have shown that CR also decreases cardiomyopathy, cardiac fibrosis, myocardial degeneration (100). Recent research has identified numerous other factors that are associated with CR-mediated preservation of diastolic function in rats, including reductions in cardiomyocyte size, less accumulation of  $\beta$ -galactosidase and lipofuscin (senescence markers), less myocyte apoptosis, less deterioration of intracellular calcium handling with faster diastolic relaxation, and increases in autophagy, which were associated with suppression of the mammalian target of rapamycin pathway (167) [inhibition of mammalian target of rapamycin with rapamycin has been shown to reverse cardiac hypertrophy (135) and may contribute to lifespan extension in mice (85)]. Insights into the mechanisms for the cardiac adaptive responses to CR also come from microarray gene expression studies on mouse hearts (52). Long-term CR, and to some extent, short-term CR alters the gene expression profile in a manner that is consistent with protection against age-related increases in myocardial remodeling and fibrosis and against age-related decreases in contractility and lipid metabolism. Additionally, and consistent with more youthful cardiac phenotype, immunohistochemical studies showed less perivascular collagen in the hearts of old CR mice and smaller myocyte size (52). Some of the beneficial effects of CR on the aged myocardium in rodents appear to be mediated by reductions in oxidative stress and damage, as evidenced by less dityrosine cross-linking of cardiac muscle proteins (120), better preservation of mitochondrial membrane fluidity (mitochondrial oxidative damage marker) and less lipid peroxidation (119), and less mitochondrial free radical produc-

tion and less oxidative damage to mitochondrial DNA in the myocardium (78).

Studies on humans generally corroborate the findings from rodent studies. In middle- to older-aged individuals, we showed that diastolic function was better in subjects who practiced strict CR for 3–15 years than that in healthy age- and sex-matched control subjects, as evidenced by Doppler studies showing lower late diastolic (A wave) velocities, greater early filling fraction, a higher E-wave-to-A-wave ratio, and greater early diastolic relaxation (E' wave from tissue Doppler imaging) (138) (Fig. 2). Insights into the physiological adaptations responsible for these improvements were obtained by using the parameterized diastolic filling (PDF) formalism (107), which indicated that CR subjects had less ventricular stiffness and less viscous loss of diastolic recoil (138), both of which would be consistent with less myocardial fibrosis. We also performed a randomized controlled trial to investigate the effects of one year of calorie restriction in middle-aged men and women with high-normal or slightly overweight body weight. In support of our cross-sectional study (138), results indicated that CR resulted in a lower isovolumic relaxation time and early diastolic filling with PDF-derived indexes, indicating less myocardial stiffness and less viscoelasticity (as an index of relaxation damping) (156) (Fig. 2). Furthermore, CR resulted in improvements in the PDF-based indexes of peak atrioventricular pres-

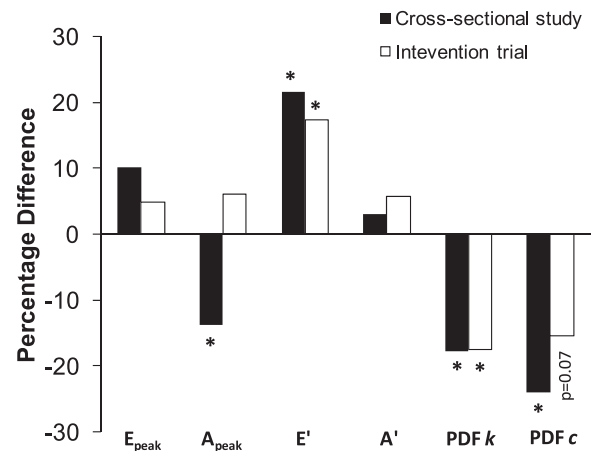


Fig. 2. Effects of CR on left ventricular diastolic function in humans. \* $P < 0.05$  vs. zero. Collectively, these data indicate that CR improves left ventricular diastolic function in healthy humans, partly by decreasing ventricular stiffness and partly by decreasing the damping of elastic recoil of the ventricle during diastole. For the cross-sectional data, values are percent difference between individuals undergoing long-term (3–15 yr) of self-imposed strict CR compared with age- and sex-matched healthy control subjects who were consuming a typical U.S. diet (138). For example, the 10% value for  $E_{peak}$  in the figure (black bar) indicates that the CR group had a 10% greater  $E_{peak}$  value than the control group. For the intervention trial, values are percent difference between baseline and follow-up measures in men and women undergoing a yearlong CR intervention (156). For example, the 5% value for  $E_{peak}$  in the figure (white bar) indicates that  $E_{peak}$  increased by 5% during the CR intervention.  $E_{peak}$ , early suction-mediated diastolic ventricular filling as measured with pulsed-Doppler transmitral flow assessments;  $A_{peak}$ , late/active atrium-driven diastolic ventricular filling as measured with pulsed-Doppler transmitral flow assessments;  $E'$ , early diastolic function as measured by tissue Doppler imaging of the septal aspect of the mitral valve annulus;  $A'$ , late-diastolic function as measured by tissue Doppler imaging of the septal aspect of the mitral valve annulus; PDF  $k$ , global ventricular stiffness parameter based on the parameterized diastolic filling (PDF) formalism (107); PDF  $c$ , index of viscoelastic loss of ventricular recoil during diastole as determined by using the PDF formalism.



sure gradient and indexes of stored strain energy, both of which would contribute to enhanced diastolic function (156).

Surprisingly, some research on rodents suggests that CR augments some age-related changes in the heart. Both long- and short-term CR was shown to augment the age-related increase in contraction and relaxation times in isolated rodent hearts (105). This alteration appears to coincide with a CR-induced increase in the age-related shift in cardiac myosin isoform from the fast V1 isoform to slow V3 (81, 104, 143). In our CR studies on humans, we saw no evidence for prolonged ventricular relaxation based on isovolumic relaxation time, E-wave duration, or early diastolic transmitral flow (E wave) or early diastolic mitral annulus velocity (E' wave) (138, 156).

Aging-related alterations in cardiac function are partly dependent on changes in autonomic function. These changes include decreases in the inotropic (50, 113) and chronotropic (176) responses to  $\beta$ -adrenergic stimulation, part of which are mediated by a reduction in myocardial  $\beta$ -adrenergic receptor density (206), reductions in maximal heart rate (182, 203), decreased heart rate variability (6, 60, 161), diminished left ventricular systolic reverse capacity (150), and a delayed cardiac response to carotid baroreceptor activation (58).

CR has been shown to alter several aspects of these age-related alterations in autonomic function. In rodents, CR has been shown to prevent the age-related decline in  $\beta$ -adrenergic receptor density, thereby preserving the inotropic and chronotropic responses to  $\beta$ -adrenergic (isoproterenol) stimulation (42, 105). Normal aging is associated with increases in circulating catecholamines (88), an effect that is associated with the induction of catecholamine biosynthetic enzymes in the adrenal medulla (99). CR has been shown to attenuate the expression of catecholamine biosynthetic enzymes (57) and reduce oxidative damage to the adrenal medulla (205). However, it is not clear whether long-term CR reduces circulating catecholamine levels (123). CR has also been shown to attenuate the age-related deterioration in the baroreceptor reflex in rodents (87, 183), suggesting an enhanced control of blood pressure through autonomic control of the heart and vasculature. Further evidence for a beneficial effect of CR on autonomic function comes from rodent studies showing a decrease in low-frequency diastolic blood pressure variability, suggesting lower sympathetic tone and greater high-frequency heart rate variability, implying greater parasympathetic tone (130). Studies using power spectral analysis of heart rate variability have also shown that six months of calorie restriction in nonobese humans results in decreases in sympathetic activity, increases in parasympathetic activity, and a decrease in the sympathetic/parasympathetic balance (51).

### Summary and Future Research Directions

Calorie restriction has numerous beneficial effects on the aging cardiovascular system, many of which appear to be mediated, at least partly, by reductions in oxidative stress and inflammation in the vasculature and heart. With respect to vascular benefits, animal studies have demonstrated that CR attenuates the age-related decline in the antiadhesion and vasodilatory functions of the endothelium and blunts the increase in arterial stiffness that occurs with advancing age. Future studies are needed to determine whether CR in humans also has beneficial effects on endothelial function and arterial stiffness. Evidence from animal studies also suggests that CR protects against atherosclerosis; however, these

findings are limited, largely because most animal species used in CR studies are not prone to atherosclerosis. While human CR studies involving direct measures of atherosclerosis are currently not practical or ethical for use in healthy human subjects, developing technologies for imaging of atherosclerosis [as reviewed elsewhere (159)] may make such studies feasible in the future. In the meantime, a large body of evidence, from both human and animal studies, indicates that CR has profound beneficial effects on risk factors for atherosclerosis. Furthermore, epidemiologic data from Japanese Okinawans suggest that CR protects against coronary heart disease.

CR also has beneficial effects on cardiac function. In animal models, CR has been shown to attenuate numerous age-related changes in the heart (including the development of hypertrophy and myocardial fibrosis), shifts in myosin isoform composition, histological changes, increases in cardiomyocyte apoptosis, and the deterioration of chronotropic and inotropic responses to adrenergic stimulation. Although such data are not available from human CR studies, echocardiographic studies have demonstrated that CR improves diastolic cardiac function in healthy nonobese humans and that these adaptations correspond with reductions in indexes of myocardial stiffness. Additional studies are needed, especially in humans, to elucidate other aspects of CR on the aging of the heart. For example, no studies have evaluated the effect of CR on the profound age-related loss of the ability of the heart to respond to acute physical stress/exercise (i.e., heart rate response and systolic reverse capacity) or on the shift in substrate metabolism that occurs in the aging heart. Additionally, more basic research is needed to elucidate the molecular adaptations that mediate the antiaging effects of CR on the cardiovascular system.

### Conclusion

Research on animals indicates that CR has a powerful ability to prevent many of the age-related changes in the structure and function of the cardiovascular system. Furthermore, research on humans has demonstrated that CR, when introduced in mature adults, may reverse some of the age-related changes in the cardiovascular system, although much more research on humans is needed. Taken together, these findings suggest that CR protects against the progressive decline in cardiovascular function that occurs with increasing age. They also show that CR has a powerful effect to prevent diseases of the cardiovascular system. These effects on the cardiovascular system, in combination with other benefits of CR, such as the protection against the development of excess adiposity and insulin resistance/type 2 diabetes, hypertension, and protection against cancer, suggest that CR may have a major beneficial effect on health span, life span, and quality of life in humans.

### GRANTS

This study was supported by National Institutes of Health Grants UL1-RR-024992, P30-DK-056341, and K01-DK-080886; by grants from Istituto Superiore di Sanità/National Institutes of Health Collaboration Program, Ministero della Salute, the Longer Life Foundation (an RGA/Washington University Partnership), and the Bakewell Foundation; and by a donation from the Scott and Annie Appleby Charitable Trust. The funding agencies provided financial support to the authors for their time in writing this manuscript and for their research on the effects of calorie restriction on the cardiovascular system but had no role in the analysis or interpretation of data or in the writing of this review article.

## DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

## REFERENCES

- Adler A, Messina E, Sherman B, Wang Z, Huang H, Linke A, Hintze TH. NAD(P)H oxidase-generated superoxide anion accounts for reduced control of myocardial O<sub>2</sub> consumption by NO in old Fischer 344 rats. *Am J Physiol Heart Circ Physiol* 285: H1015–H1022, 2003.
- Ahmet I, Tae HJ, de Cabo R, Lakatta EG, Talan MI. Effects of calorie restriction on cardioprotection and cardiovascular health. *J Mol Cell Cardiol* 51: 263–271, 2011.
- Alberti KG, Zimmet P, Shaw J. Metabolic syndrome—a new worldwide definition. A consensus statement from the International Diabetes Federation. *Diabet Med* 23: 469–480, 2006.
- Allen JP, Nelson M, Alling A. The legacy of Biosphere 2 for the study of biospherics and closed ecological systems. *Adv Space Res* 31: 1629–1639, 2003.
- Anderson RM, Shanmuganayagam D, Weindruch R. Caloric restriction and aging: studies in mice and monkeys. *Toxicol Pathol* 37: 47–51, 2009.
- Antelmi I, de Paula RS, Shinzato AR, Peres CA, Mansur AJ, Grupi CJ. Influence of age, gender, body mass index, and functional capacity on heart rate variability in a cohort of subjects without heart disease. *Am J Cardiol* 93: 381–385, 2004.
- Anversa P, Palackal T, Sonnenblick EH, Olivetti G, Meggs LG, Capasso JM. Myocyte cell loss and myocyte cellular hyperplasia in the hypertrophied aging rat heart. *Circ Res* 67: 871–885, 1990.
- Avolio A. Genetic and environmental factors in the function and structure of the arterial wall. *Hypertension* 26: 34–37, 1995.
- Avolio AP, Chen SG, Wang RP, Zhang CL, Li MF, O'Rourke MF. Effects of aging on changing arterial compliance and left ventricular load in a northern Chinese urban community. *Circulation* 68: 50–58, 1983.
- Avolio AP, Deng FQ, Li WQ, Luo YF, Huang ZD, Xing LF, O'Rourke MF. Effects of aging on arterial distensibility in populations with high and low prevalence of hypertension: comparison between urban and rural communities in China. *Circulation* 71: 202–210, 1985.
- Barbagallo CM, Averna MR, Frada G, Noto D, Cavera G, Notarbartolo A. Lipoprotein profile and high-density lipoproteins: subfractions distribution in centenarians. *Gerontology* 44: 106–110, 1998.
- Barja G. Free radicals and aging. *Trends Neurosci* 27: 595–600, 2004.
- Barrett-Connor E, Ferrara A. Isolated postchallenge hyperglycemia and the risk of fatal cardiovascular disease in older women and men. The Rancho Bernardo Study. *Diabetes Care* 21: 1236–1239, 1998.
- Barzilai N, Banerjee S, Hawkins M, Chen W, Rossetti L. Caloric restriction reverses hepatic insulin resistance in aging rats by decreasing visceral fat. *J Clin Invest* 101: 1353–1361, 1998.
- Behnke BJ, Delp MD. Aging blunts the dynamics of vasodilation in isolated skeletal muscle resistance vessels. *J Appl Physiol* 108: 14–20, 2010.
- Berkowitz DE, White R, Li D, Minhas KM, Cernetich A, Kim S, Burke S, Shoukas AA, Nyhan D, Champion HC, Hare JM. Arginase reciprocally regulates nitric oxide synthase activity and contributes to endothelial dysfunction in aging blood vessels. *Circulation* 108: 2000–2006, 2003.
- Blann AD. Endothelial cell activation, injury, damage and dysfunction: separate entities or mutual terms? *Blood Coagul Fibrinolysis* 11: 623–630, 2000.
- Bruunsgaard H, Skinhoj P, Pedersen AN, Schroll M, Pedersen BK. Ageing, tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ) and atherosclerosis. *Clin Exp Immunol* 121: 255–260, 2000.
- Bucala R, Cerami A. Advanced glycosylation: chemistry, biology, and implications for diabetes and aging. *Adv Pharmacol* 23: 1–34, 1992.
- Buring JE, O'Connor GT, Goldhaber SZ, Rosner B, Herbert PN, Blum CB, Breslow JL, Hennekens CH. Decreased HDL2 and HDL3 cholesterol, Apo A-I and Apo A-II, and increased risk of myocardial infarction. *Circulation* 85: 22–29, 1992.
- Camandola S, Scavazza A, Leonarduzzi G, Biasi F, Chiarpotto E, Azzì A, Poli G. Biogenic 4-hydroxy-2-nonenal activates transcription factor AP-1 but not NF- $\kappa$ B in cells of the macrophage lineage. *Biofactors* 6: 173–179, 1997.
- Camejo G, Waich S, Quintero G, Berrizbeitia ML, Lalaguna F. The affinity of low density lipoproteins for an arterial macromolecular complex. A study in ischemic heart disease and controls. *Atherosclerosis* 24: 341–354, 1976.
- Cangemi R, Friedmann AJ, Holloszy JO, Fontana L. Long-term effects of calorie restriction on serum sex-hormone concentrations in men. *Aging Cell* 9: 236–242, 2010.
- Castello L, Froio T, Cavallini G, Biasi F, Sapino A, Leonarduzzi G, Bergamini E, Poli G, Chiarpotto E. Calorie restriction protects against age-related rat aorta sclerosis. *FASEB J* 19: 1863–1865, 2005.
- Cefalu WT, Terry JG, Thomas MJ, Morgan TM, Edwards IJ, Rudel LL, Kemnitz JW, Weindruch R. In vitro oxidation of low-density lipoprotein in two species of nonhuman primates subjected to caloric restriction. *J Gerontol A Biol Sci Med Sci* 55: B355–B361, 2000.
- Cefalu WT, Wagner JD, Bell-Farrow AD, Edwards IJ, Terry JG, Weindruch R, Kemnitz JW. Influence of caloric restriction on the development of atherosclerosis in nonhuman primates: progress to date. *Toxicol Sci* 52: 49–55, 1999.
- Cefalu WT, Wagner JD, Wang ZQ, Bell-Farrow AD, Collins J, Haskell D, Bechtold R, Morgan T. A study of caloric restriction and cardiovascular aging in cynomolgus monkeys (*Macaca fascicularis*): a potential model for aging research. *J Gerontol A Biol Sci Med Sci* 52: B10–B19, 1997.
- Cefalu WT, Wang ZQ, Bell-Farrow AD, Collins J, Morgan T, Wagner JD. Caloric restriction and cardiovascular aging in cynomolgus monkeys (*Macaca fascicularis*): metabolic, physiologic, and atherosclerotic measures from a 4-year intervention trial. *J Gerontol A Biol Sci Med Sci* 59: 1007–1014, 2004.
- Celermajer DS, Sorensen KE, Spiegelhalter DJ, Georgakopoulos D, Robinson J, Deanfield JE. Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. *J Am Coll Cardiol* 24: 471–476, 1994.
- Cerami A. Hypothesis. Glucose as a mediator of aging. *J Am Geriatr Soc* 33: 626–634, 1985.
- Cerqueira FM, Kowaltowski AJ. Commonly adopted caloric restriction protocols often involve malnutrition. *Ageing Res Rev* 9: 424–430, 2010.
- Cheitlin MD. Cardiovascular physiology—changes with aging. *Am J Geriatr Cardiol* 12: 9–13, 2003.
- Chobanian AV, Bakris GL, Black HR, Cushman WC, Green LA, Izzo JL Jr, Jones DW, Materson BJ, Oparil S, Wright JT Jr, Roccella EJ. The seventh report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. *JAMA* 289: 2560–2572, 2003.
- Coggan AR, Spina RJ, King DS, Rogers MA, Brown M, Nemeth PM, Holloszy JO. Skeletal muscle adaptations to endurance training in 60- to 70-yr-old men and women. *J Appl Physiol* 72: 1780–1786, 1992.
- Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB, Cruzen C, Simmons HA, Kemnitz JW, Weindruch R. Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science* 325: 201–204, 2009.
- Colman RJ, Beasley TM, Allison DB, Weindruch R. Attenuation of sarcopenia by dietary restriction in rhesus monkeys. *J Gerontol A Biol Sci Med Sci* 63: 556–559, 2008.
- Colman RJ, Roecker EB, Ramsey JJ, Kemnitz JW. The effect of dietary restriction on body composition in adult male and female rhesus macaques. *Aging (Milano)* 10: 83–92, 1998.
- Compagno V, Di L, I, Cestelli A, Donatelli M. Effect of aging and hypertension on beta-myosin heavy chain in heart of spontaneously hypertensive rats. *Int J Mol Med* 7: 507–508, 2001.
- Cooke JP, Dzau VJ. Nitric oxide synthase: role in the genesis of vascular disease. *Annu Rev Med* 48: 489–509, 1997.
- Cornwell GG III, Thomas BP, Snyder DL. Myocardial fibrosis in aging germ-free and conventional Lobund-Wistar rats: the protective effect of diet restriction. *J Gerontol* 46: B167–B170, 1991.
- Coutinho M, Gerstein HC, Wang Y, Yusuf S. The relationship between glucose and incident cardiovascular events. A meta-regression analysis of published data from 20 studies of 95,783 individuals followed for 124 years. *Diabetes Care* 22: 233–240, 1999.
- Crandall DL, Lai FM, Huggins FJ, Tanikella TK, Cervoni P. Effect of caloric restriction on cardiac reactivity and beta-adrenoceptor concentration. *Am J Physiol Heart Circ Physiol* 244: H444–H448, 1983.
- Csiszar A, Labinskyy N, Jimenez R, Pinto JT, Ballabh P, Losonczy G, Pearson KJ, de Cabo R, Ungvari Z. Anti-oxidative and anti-inflammatory vasoprotective effects of caloric restriction in aging: role of circulating factors and SIRT1. *Mech Ageing Dev* 130: 518–527, 2009.



44. Csizsar A, Ungvari Z, Edwards JG, Kaminski P, Wolin MS, Koller A, Kaley G. Aging-induced phenotypic changes and oxidative stress impair coronary arteriolar function. *Circ Res* 90: 1159–1166, 2002.
45. Csizsar A, Ungvari Z, Koller A, Edwards JG, Kaley G. Aging-induced proinflammatory shift in cytokine expression profile in coronary arteries. *FASEB J* 17: 1183–1185, 2003.
46. Csizsar A, Ungvari Z, Koller A, Edwards JG, Kaley G. Proinflammatory phenotype of coronary arteries promotes endothelial apoptosis in aging. *Physiol Genomics* 17: 21–30, 2004.
47. Dahlen GH, Guyton JR, Attar M, Farmer JA, Kautz JA, Gotto AM Jr. Association of levels of lipoprotein Lp(a), plasma lipids, and other lipoproteins with coronary artery disease documented by angiography. *Circulation* 74: 758–765, 1986.
48. Das SK, Gilhooly CH, Golden JK, Pittas AG, Fuss PJ, Cheatham RA, Tyler S, Tsay M, McCrory MA, Lichtenstein AH, Dallal GE, Dutta C, Bhapkar MV, DeLany JP, Saltzman E, Roberts SB. Long-term effects of 2 energy-restricted diets differing in glycemic load on dietary adherence, body composition, and metabolism in CALERIE: a 1-y randomized controlled trial. *Am J Clin Nutr* 85: 1023–1030, 2007.
49. Das SK, Gilhooly CH, Golden JK, Pittas AG, Fuss PJ, Dallal GE, McCrory MA, Saltzman E, Roberts SB. Long term effects of energy-restricted diets differing in glycemic load on metabolic adaptation and body composition. *Open Nutr J* 85: 1023–1030, 2007.
50. Davies CH, Ferrara N, Harding SE. Beta-adrenoceptor function changes with age of subject in myocytes from non-failing human ventricle. *Cardiovasc Res* 31: 152–156, 1996.
51. de Jonge L, Moreira EA, Martin CK, Ravussin E. Impact of 6-month caloric restriction on autonomic nervous system activity in healthy, overweight, individuals. *Obesity (Silver Spring)* 18: 414–416, 2010.
52. Dhahbi JM, Tsuchiya T, Kim HJ, Mote PL, Spindler SR. Gene expression and physiologic responses of the heart to the initiation and withdrawal of caloric restriction. *J Gerontol A Biol Sci Med Sci* 61: 218–231, 2006.
53. Dolinsky VW, Morton JS, Oka T, Robillard-Frayne I, Bagdan M, Lopaschuk GD, Des RC, Walsh K, Davidge ST, Dyck JR. Calorie restriction prevents hypertension and cardiac hypertrophy in the spontaneously hypertensive rat. *Hypertension* 56: 412–421, 2010.
54. Donato AJ, Eskurza I, Silver AE, Levy AS, Pierce GL, Gates PE, Seals DR. Direct evidence of endothelial oxidative stress with aging in humans: relation to impaired endothelium-dependent dilation and up-regulation of nuclear factor-kappaB. *Circ Res* 100: 1659–1666, 2007.
55. Edwards IJ, Rudel LL, Terry JG, Kennitz JW, Weindruch R, Cefalu WT. Caloric restriction in rhesus monkeys reduces low density lipoprotein interaction with arterial proteoglycans. *J Gerontol A Biol Sci Med Sci* 53: B443–B448, 1998.
56. Elliott RJ, McGrath LT. Calcification of the human thoracic aorta during aging. *Calcif Tissue Int* 54: 268–273, 1994.
57. Erdos B, Broxson CS, Landa T, Scarpace PJ, Leeuwenburgh C, Zhang Y, Tumer N. Effects of life-long caloric restriction and voluntary exercise on age-related changes in levels of catecholamine biosynthetic enzymes and angiotensin II receptors in the rat adrenal medulla and hypothalamus. *Exp Gerontol* 42: 745–752, 2007.
58. Fisher JP, Kim A, Young CN, Ogoh S, Raven PB, Secher NH, Fadel PJ. Influence of ageing on carotid baroreflex peak response latency in humans. *J Physiol* 587: 5427–5439, 2009.
59. Fleg JL, O'Connor F, Gerstenblith G, Becker LC, Clulow J, Schulman SP, Lakatta EG. Impact of age on the cardiovascular response to dynamic upright exercise in healthy men and women. *J Appl Physiol* 78: 890–900, 1995.
60. Fluckiger L, Boivin JM, Quilliot D, Jeandel C, Zannad F. Differential effects of aging on heart rate variability and blood pressure variability. *J Gerontol A Biol Sci Med Sci* 54: B219–B224, 1999.
61. Fontana L, Klein S. Aging, adiposity and caloric restriction. *JAMA* 297: 986–994, 2007.
62. Fontana L, Klein S, Holloszy JO. Effects of long-term caloric restriction and endurance exercise on glucose tolerance, insulin action, and adipokine production. *Age (Dordr)* 32: 97–108, 2010.
63. Fontana L, Klein S, Holloszy JO, Premachandra BN. Effect of long-term caloric restriction with adequate protein and micronutrients on thyroid hormones. *J Clin Endocrinol Metab* 91: 3232–3235, 2006.
64. Fontana L, Meyer TE, Klein S, Holloszy JO. Long-term caloric restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc Natl Acad Sci USA* 101: 6659–6663, 2004.
65. Fontana L, Partridge L, Longo VD. Extending healthy life span—from yeast to humans. *Science* 328: 321–326, 2010.
66. Fontana L, Villareal DT, Weisz EP, Racette SB, Steger-May K, Klein S, Holloszy JO. Calorie restriction or exercise: effects on coronary heart disease risk factors. A randomized, controlled trial. *Am J Physiol Endocrinol Metab* 293: E197–E202, 2007.
67. Fontana L, Weiss EP, Villareal DT, Klein S, Holloszy JO. Long-term effects of calorie or protein restriction on serum IGF-1 and IGFBP-3 concentration in humans. *Aging Cell* 7: 681–687, 2008.
68. Fornieri C, Taparelli F, Quagliano D Jr, Contri MB, Davidson JM, Algeri S, Ronchetti IP. The effect of caloric restriction on the aortic tissue of aging rats. *Connect Tissue Res* 40: 131–143, 1999.
69. Fox CS, Pencina MJ, Wilson PW, Paynter NP, Vasan RS, D'Agostino RB Sr. Lifetime risk of cardiovascular disease among individuals with and without diabetes stratified by obesity status in the Framingham heart study. *Diabetes Care* 31: 1582–1584, 2008.
70. Franklin SS, Gustin W, Wong ND, Larson MG, Weber MA, Kannel WB, Levy D. Hemodynamic patterns of age-related changes in blood pressure. The Framingham Heart Study. *Circulation* 96: 308–315, 1997.
71. Fulton D, Gratton JP, McCabe TJ, Fontana J, Fujio Y, Walsh K, Franke TF, Papadopoulos A, Sessa WC. Regulation of endothelium-derived nitric oxide production by the protein kinase Akt. *Nature* 399: 597–601, 1999.
72. Galarraga B, Khan F, Kumar P, Pullar T, Belch JJ. Etanercept improves inflammation-associated arterial stiffness in rheumatoid arthritis. *Rheumatology (Oxford)* 48: 1418–1423, 2009.
73. Garg UC, Hassid A. Nitric oxide-generating vasodilators and 8-bromocyclic guanosine monophosphate inhibit mitogenesis and proliferation of cultured rat vascular smooth muscle cells. *J Clin Invest* 83: 1774–1777, 1989.
74. Gerhard M, Roddy MA, Creager SJ, Creager MA. Aging progressively impairs endothelium-dependent vasodilation in forearm resistance vessels of humans. *Hypertension* 27: 849–853, 1996.
75. Giani JF, Bonkowski MS, Munoz MC, Masternak MM, Turyn D, Bartke A, Dominici FP. Insulin signaling cascade in the hearts of long-lived growth hormone receptor knockout mice: effects of caloric restriction. *J Gerontol A Biol Sci Med Sci* 63: 788–797, 2008.
76. Goraya TY, Leibson CL, Palumbo PJ, Weston SA, Killian JM, Pfeifer EA, Jacobsen SJ, Frye RL, Roger VL. Coronary atherosclerosis in diabetes mellitus: a population-based autopsy study. *J Am Coll Cardiol* 40: 946–953, 2002.
77. Gostynski M, Gutzwiller F, Kuulasmaa K, Doring A, Ferrario M, Grafnetter D, Pajak A. Analysis of the relationship between total cholesterol, age, body mass index among males and females in the WHO MONICA Project. *Int J Obes Relat Metab Disord* 28: 1082–1090, 2004.
78. Gredilla R, Sanz A, Lopez-Torres M, Barja G. Caloric restriction decreases mitochondrial free radical generation at complex I and lowers oxidative damage to mitochondrial DNA in the rat heart. *FASEB J* 15: 1589–1591, 2001.
79. Gresl TA, Colman RJ, Roecker EB, Havighurst TC, Huang Z, Allison DB, Bergman RN, Kennitz JW. Dietary restriction and glucose regulation in aging rhesus monkeys: a follow-up report at 8.5 yr. *Am J Physiol Endocrinol Metab* 281: E757–E765, 2001.
80. Guo Z, Mitchell-Raymundo F, Yang H, Ikeno Y, Nelson J, Diaz V, Richardson A, Reddick R. Dietary restriction reduces atherosclerosis and oxidative stress in the aorta of apolipoprotein E-deficient mice. *Mech Ageing Dev* 123: 1121–1131, 2002.
81. Haddad F, Bodell PW, McCue SA, Herrick RE, Baldwin KM. Food restriction-induced transformations in cardiac functional and biochemical properties in rats. *J Appl Physiol* 74: 606–612, 1993.
82. Hajjar I, Kotchen TA. Trends in prevalence, awareness, treatment, and control of hypertension in the United States, 1988–2000. *JAMA* 290: 199–206, 2003.
83. Hajra L, Evans AI, Chen M, Hyduk SJ, Collins T, Cybulsky MI. The NF-kappa B signal transduction pathway in aortic endothelial cells is primed for activation in regions predisposed to atherosclerotic lesion formation. *Proc Natl Acad Sci USA* 97: 9052–9057, 2000.
84. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* 352: 1685–1695, 2005.
85. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* 460: 392–395, 2009.



86. Heilbronn LK, de Jonge L, Frisard MI, DeLany JP, Larson-Meyer DE, Rood J, Nguyen T, Martin CK, Volaufova J, Most MM, Greenway FL, Smith SR, Deutsch WA, Williamson DA, Ravussin E. Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. *JAMA* 295: 1539–1548, 2006.
87. Herlihy JT, Stacy C, Bertrand HA. Long-term calorie restriction enhances baroreflex responsiveness in Fischer 344 rats. *Am J Physiol Heart Circ Physiol* 263: H1021–H1025, 1992.
88. Hoeldtke RD, Cilmi KM. Effects of aging on catecholamine metabolism. *J Clin Endocrinol Metab* 60: 479–484, 1985.
89. Hofer T, Fontana L, Anton SD, Weiss EP, Villareal D, Malayappan B, Leeuwenburgh C. Long-term effects of caloric restriction or exercise on DNA and RNA oxidation levels in white blood cells and urine in humans. *Rejuvenation Res* 11: 793–799, 2008.
90. Howard G, O'Leary DH, Zaccaro D, Haffner S, Rewers M, Hamman R, Selby JV, Saad MF, Savage P, Bergman R. Insulin sensitivity and atherosclerosis. The Insulin Resistance Atherosclerosis Study (IRAS) Investigators. *Circulation* 93: 1809–1817, 1996.
91. Ishii T, Itoh K, Yamamoto M. Roles of Nrf2 in activation of antioxidant enzyme genes via antioxidant responsive elements. *Methods Enzymol* 348: 182–190, 2002.
92. Jacobson A, Yan C, Gao Q, Rincon-Skinner T, Rivera A, Edwards J, Huang A, Kaley G, Sun D. Aging enhances pressure-induced arterial superoxide formation. *Am J Physiol Heart Circ Physiol* 293: H1344–H1350, 2007.
93. Japan Ministry of Health, Labor, and Welfare. *Results from Ministry of Health, Labor and Welfare Nutrition Survey, 2000*. Tokyo, Japan, Dai-ichi, 2002.
94. Joly L, Djaballah W, Koehl G, Mandry D, Dolivet G, Marie PY, Benetos A. Aortic inflammation, as assessed by hybrid FDG-PET/CT imaging, is associated with enhanced aortic stiffness in addition to concurrent calcification. *Eur J Nucl Med Mol Imaging* 36: 979–985, 2009.
95. Kalani R, Judge S, Carter C, Pahor M, Leeuwenburgh C. Effects of caloric restriction and exercise on age-related, chronic inflammation assessed by C-reactive protein and interleukin-6. *J Gerontol A Biol Sci Med Sci* 61: 211–217, 2006.
96. Kalant N, Stewart J, Kaplan R. Effect of diet restriction on glucose metabolism and insulin responsiveness in aging rats. *Mech Ageing Dev* 46: 89–104, 1988.
97. Kang LS, Reyes RA, Muller-Delp JM. Aging impairs flow-induced dilation in coronary arterioles: role of NO and H<sub>2</sub>O<sub>2</sub>. *Am J Physiol Heart Circ Physiol* 297: H1087–H1095, 2009.
98. Katzel LI, Busby-Whitehead MJ, Rogus EM, Krauss RM, Goldberg AP. Reduced adipose tissue lipoprotein lipase responses, postprandial lipemia, and low high-density lipoprotein-2 subspecies levels in older athletes with silent myocardial ischemia. *Metabolism* 43: 190–198, 1994.
99. Kedzierski W, Porter JC. Quantitative study of tyrosine hydroxylase mRNA in catecholaminergic neurons and adrenals during development and aging. *Brain Res Mol Brain Res* 7: 45–51, 1990.
100. Kemi M, Keenan KP, McCoy C, Hoe CM, Soper KA, Ballam GC, van Zwieten MJ. The relative protective effects of moderate dietary restriction versus dietary modification on spontaneous cardiomyopathy in male Sprague-Dawley rats. *Toxicol Pathol* 28: 285–296, 2000.
101. Kemnitz JW, Roecker EB, Weindruch R, Elson DF, Baum ST, Bergman RN. Dietary restriction increases insulin sensitivity and lowers blood glucose in rhesus monkeys. *Am J Physiol Endocrinol Metab* 266: E540–E547, 1994.
102. Khoshdel AR, Thakkinstian A, Carney SL, Attia J. Estimation of an age-specific reference interval for pulse wave velocity: a meta-analysis. *J Hypertens* 24: 1231–1237, 2006.
103. Kitzman DW, Sheikh KH, Beere PA, Philips JL, Higginbotham MB. Age-related alterations of Doppler left ventricular filling indexes in normal subjects are independent of left ventricular mass, heart rate, contractility and loading conditions. *J Am Coll Cardiol* 18: 1243–1250, 1991.
104. Klebanov S, Herlihy JT. Effect of life-long food restriction on cardiac myosin composition. *J Gerontol A Biol Sci Med Sci* 52: B184–B189, 1997.
105. Klebanov S, Herlihy JT, Freeman GL. Effect of long-term food restriction on cardiac mechanics. *Am J Physiol Heart Circ Physiol* 273: H2333–H2342, 1997.
106. Kobayashi M, Yamamoto M. Nrf2-Keap1 regulation of cellular defense mechanisms against electrophiles and reactive oxygen species. *Adv Enzyme Regul* 46: 113–140, 2006.
107. Kovacs SJ Jr, Barzilai B, Perez JE. Evaluation of diastolic function with Doppler echocardiography: the PDF formalism. *Am J Physiol Heart Circ Physiol* 252: H178–H187, 1987.
108. Kubes P, Suzuki M, Granger DN. Nitric oxide: an endogenous modulator of leukocyte adhesion. *Proc Natl Acad Sci USA* 88: 4651–4655, 1991.
109. Kung H, Hoyert DL, Xu J, Murphy SL. Deaths: Final Data for 2005. *National Vital Statistics Reports* 56. Hyattsville, MD: National Center for Health Statistics, 2008.
110. Lakatta EG. Cardiac muscle changes in senescence. *Annu Rev Physiol* 49: 519–531, 1987.
111. Lakatta EG. Cardiovascular regulatory mechanisms in advanced age. *Physiol Rev* 73: 413–467, 1993.
112. Lakatta EG. Cardiovascular aging research: the next horizons. *J Am Geriatr Soc* 47: 613–625, 1999.
113. Lakatta EG, Gerstenblith G, Angell CS, Shock NW, Weisfeldt ML. Diminished inotropic response of aged myocardium to catecholamines. *Circ Res* 36: 262–269, 1975.
114. Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part I: aging arteries: a “set up” for vascular disease. *Circulation* 107: 139–146, 2003.
115. Lane MA, Ball SS, Ingram DK, Cutler RG, Engel J, Read V, Roth GS. Diet restriction in rhesus monkeys lowers fasting and glucose-stimulated glucoregulatory end points. *Am J Physiol Endocrinol Metab* 268: E941–E948, 1995.
116. Lane MA, Black A, Ingram DK, Roth GS. Calorie restriction in nonhuman primates: implications for age-related disease risk. *J Anti-Aging Med* 1: 315–326, 1998.
117. Larson-Meyer DE, Heilbronn LK, Redman LM, Newcomer BR, Frisard MI, Anton S, Smith SR, Alfonso A, Ravussin E. Effect of calorie restriction with or without exercise on insulin sensitivity, beta-cell function, fat cell size, and ectopic lipid in overweight subjects. *Diabetes Care* 29: 1337–1344, 2006.
118. Larson-Meyer DE, Newcomer BR, Heilbronn LK, Volaufova J, Smith SR, Alfonso AJ, Lefevre M, Rood JC, Williamson DA, Ravussin E. Effect of 6-month calorie restriction and exercise on serum and liver lipids and markers of liver function. *Obesity (Silver Spring)* 16: 1355–1362, 2008.
119. Lee J, Yu BP, Herlihy JT. Modulation of cardiac mitochondrial membrane fluidity by age and calorie intake. *Free Radic Biol Med* 26: 260–265, 1999.
120. Leeuwenburgh C, Wagner P, Holloszy JO, Sohal RS, Heinecke JW. Caloric restriction attenuates dityrosine cross-linking of cardiac and skeletal muscle proteins in aging mice. *Arch Biochem Biophys* 346: 74–80, 1997.
121. Lefevre M, Redman LM, Heilbronn LK, Smith JV, Martin CK, Rood JC, Greenway FL, Williamson DA, Smith SR, Ravussin E. Caloric restriction alone and with exercise improves CVD risk in healthy non-obese individuals. *Atherosclerosis* 203: 206–213, 2009.
122. Leonarduzzi G, Scavazza A, Biasi F, Chiarpotto E, Camandola S, Vogel S, Dargel R, Poli G. The lipid peroxidation end product 4-hydroxy-2,3-nonenal up-regulates transforming growth factor beta1 expression in the macrophage lineage: a link between oxidative injury and fibrosclerosis. *FASEB J* 11: 851–857, 1997.
123. Levay EA, Tammer AH, Penman J, Kent S, Paolini AG. Calorie restriction at increasing levels leads to augmented concentrations of corticosterone and decreasing concentrations of testosterone in rats. *Nutr Res* 30: 366–373, 2010.
124. Levy D, Anderson KM, Savage DD, Kannel WB, Christiansen JC, Castelli WP. Echocardiographically detected left ventricular hypertrophy: prevalence and risk factors. The Framingham Heart Study. *Ann Intern Med* 108: 7–13, 1988.
125. Libby P. Inflammation and cardiovascular disease mechanisms. *Am J Clin Nutr* 83: 456S–460S, 2006.
126. Linden T, Bondjers G, Camejo G, Bergstrand R, Wilhelmson L, Wiklund O. Affinity of LDL to a human arterial proteoglycan among male survivors of myocardial infarction. *Eur J Clin Invest* 19: 38–44, 1989.
127. Lloyd-Jones DM, Leip EP, Larson MG, D'Agostino RB, Beiser A, Wilson PW, Wolf PA, Levy D. Prediction of lifetime risk for cardio-

- vascular disease by risk factor burden at 50 years of age. *Circulation* 113: 791–798, 2006.
128. Luo SF, Lin CC, Chen HC, Lin WN, Lee IT, Lee CW, Hsiao LD, Yang CM. Involvement of MAPKs, NF-kappaB and p300 co-activator in IL-1beta-induced cytosolic phospholipase A2 expression in canine tracheal smooth muscle cells. *Toxicol Appl Pharmacol* 232: 396–407, 2008.
  129. Machida S, Tsujimoto H, Suzuki H, Kasuga N, Kobayashi K, Narusawa M. Age-related differences in the effect of running training on cardiac Myosin isozyme composition in rats. *J Gerontol A Biol Sci Med Sci* 57: B339–B343, 2002.
  130. Mager DE, Wan R, Brown M, Cheng A, Wareski P, Abernethy DR, Mattson MP. Caloric restriction and intermittent fasting alter spectral measures of heart rate and blood pressure variability in rats. *FASEB J* 20: 631–637, 2006.
  131. Masoro EJ. Overview of caloric restriction and ageing. *Mech Ageing Dev* 126: 913–922, 2005.
  132. Masoro EJ, McCarter RJ, Katz MS, McMahan CA. Dietary restriction alters characteristics of glucose fuel use. *J Gerontol* 47: B202–B208, 1992.
  133. Mattagajasingh I, Kim CS, Naqi A, Yamamori T, Hoffman TA, Jung SB, DeRicco J, Kasuno K, Irani K. SIRT1 promotes endothelium-dependent vascular relaxation by activating endothelial nitric oxide synthase. *Proc Natl Acad Sci USA* 104: 14855–14860, 2007.
  134. McGrath BP, Liang YL, Teede H, Shiel LM, Cameron JD, Dart A. Age-related deterioration in arterial structure and function in postmenopausal women: impact of hormone replacement therapy. *Arterioscler Thromb Vasc Biol* 18: 1149–1156, 1998.
  135. McMullen JR, Sherwood MC, Tarnavski O, Zhang L, Dorfman AL, Shioi T, Izumo S. Inhibition of mTOR signaling with rapamycin regresses established cardiac hypertrophy induced by pressure overload. *Circulation* 109: 3050–3055, 2004.
  136. Messaoudi I, Warner J, Fischer M, Park B, Hill B, Mattison J, Lane MA, Roth GS, Ingram DK, Picker LJ, Douek DC, Mori M, Nikolich-Zugich J. Delay of T cell senescence by caloric restriction in aged long-lived nonhuman primates. *Proc Natl Acad Sci USA* 103: 19448–19453, 2006.
  137. Meydani M, Das S, Band M, Epstein S, Roberts S. The effect of caloric restriction and glycemic load on measures of oxidative stress and antioxidants in humans: results from the CALERIE Trial of Human Caloric Restriction. *J Nutr Health Aging* 15: 456–460, 2011.
  138. Meyer TE, Kovacs SJ, Ehsani AA, Klein S, Holloszy JO, Fontana L. Long-term caloric restriction ameliorates the decline in diastolic function in humans. *J Am Coll Cardiol* 47: 398–402, 2006.
  139. Miles EA, Rees D, Banerjee T, Cazzola R, Lewis S, Wood R, Oates R, Tallant A, Cestaro B, Yaqoob P, Wahle KW, Calder PC. Age-related increases in circulating inflammatory markers in men are independent of BMI, blood pressure and blood lipid concentrations. *Atherosclerosis* 196: 298–305, 2008.
  140. Miller TR, Grossman SJ, Schechtman KB, Biello DR, Ludbrook PA, Ehsani AA. Left ventricular diastolic filling and its association with age. *Am J Cardiol* 58: 531–535, 1986.
  141. Mittendorfer B, Patterson BW, Klein S. Effect of weight loss on VLDL-triglyceride and apoB-100 kinetics in women with abdominal obesity. *Am J Physiol Endocrinol Metab* 284: E549–E556, 2003.
  142. Mizuguchi Y, Oishi Y, Miyoshi H, Iuchi A, Nagase N, Oki T. Impact of statin therapy on left ventricular function and carotid arterial stiffness in patients with hypercholesterolemia. *Circ J* 72: 538–544, 2008.
  143. Morris GS, Surdyka DG, Haddad F, Baldwin KM. Apparent influence of metabolism on cardiac isomyosin profile of food-restricted rats. *Am J Physiol Regul Integr Comp Physiol* 258: R346–R351, 1990.
  144. Nguyen T, Nioi P, Pickett CB. The Nrf2-antioxidant response element signaling pathway and its activation by oxidative stress. *J Biol Chem* 284: 13291–13295, 2009.
  145. O'Keefe JH Jr, Cordain L, Harris WH, Moe RM, Vogel R. Optimal low-density lipoprotein is 50 to 70 mg/dl: lower is better and physiologically normal. *J Am Coll Cardiol* 43: 2142–2146, 2004.
  146. Oeppen J, Vaupel Demography JW. Broken limits to life expectancy. *Science* 296: 1029–1031, 2002.
  147. Olivetti G, Melissari M, Capasso JM, Anversa P. Cardiomyopathy of the aging human heart. Myocyte loss and reactive cellular hypertrophy. *Circ Res* 68: 1560–1568, 1991.
  148. Paolisso G, Rizzo MR, Mazziotti G, Tagliamonte MR, Gambardella A, Rotondi M, Carella C, Giugliano D, Varricchio M, D'Onofrio F. Advancing age and insulin resistance: role of plasma tumor necrosis factor-alpha. *Am J Physiol Endocrinol Metab* 275: E294–E299, 1998.
  149. Pearson KJ, Lewis KN, Price NL, Chang JW, Perez E, Cascajo MV, Tamashiro KL, Poosala S, Csiszar A, Ungvari Z, Kensler TW, Yamamoto M, Egan JM, Longo DL, Ingram DK, Navas P, de Cabo R. Nrf2 mediates cancer protection but not longevity induced by caloric restriction. *Proc Natl Acad Sci USA* 105: 2325–2330, 2008.
  150. Port S, Cobb FR, Coleman RE, Jones RH. Effect of age on the response of the left ventricular ejection fraction to exercise. *N Engl J Med* 303: 1133–1137, 1980.
  151. Proctor DN, Parker BA. Vasodilation and vascular control in contracting muscle of the aging human. *Microcirculation* 13: 315–327, 2006.
  152. Pyke KE, Tschakovsky ME. The relationship between shear stress and flow-mediated dilatation: implications for the assessment of endothelial function. *J Physiol* 568: 357–369, 2005.
  153. Racette SB, Weiss EP, Villareal DT, Arif H, Steger-May K, Schechtman KB, Fontana L, Klein S, Holloszy JO; Washington University School of Medicine CALERIE Group. One year of caloric restriction in humans: feasibility and effects on body composition and abdominal adipose tissue. *J Gerontol A Biol Sci Med Sci* 61: 943–950, 2006.
  154. Reaven GM. Pathophysiology of insulin resistance in human disease. *Physiol Rev* 75: 473–486, 1995.
  155. Redman LM, Heilbronn LK, Martin CK, Alfonso A, Smith SR, Ravussin E. Effect of calorie restriction with or without exercise on body composition and fat distribution. *J Clin Endocrinol Metab* 92: 865–872, 2007.
  156. Riordan MM, Weiss EP, Meyer TE, Ehsani AA, Racette SB, Villareal DT, Fontana L, Holloszy JO, Kovacs SJ. The effects of caloric restriction- and exercise-induced weight loss on left ventricular diastolic function. *Am J Physiol Heart Circ Physiol* 294: H1174–H1182, 2008.
  157. Rippe C, Lesniewski L, Connell M, LaRocca T, Donato A, Seals D. Short-term calorie restriction reverses vascular endothelial dysfunction in old mice by increasing nitric oxide and reducing oxidative stress. *Aging Cell* 9: 304–312, 2010.
  158. Ruoslahti E, Yamaguchi Y. Proteoglycans as modulators of growth factor activities. *Cell* 64: 867–869, 1991.
  159. Sakalihan N, Michel JB. Functional imaging of atherosclerosis to advance vascular biology. *Eur J Vasc Endovasc Surg* 37: 728–734, 2009.
  160. Schuyler GT, Yarbrough LR. Effects of age on myosin and creatine kinase isoforms in left ventricles of Fischer 344 rats. *Mech Ageing Dev* 56: 23–38, 1990.
  161. Schwartz JB, Gibb WJ, Tran T. Aging effects on heart rate variation. *J Gerontol* 46: M99–M106, 1991.
  162. Selvin E, Najjar SS, Cornish TC, Halushka MK. A comprehensive histopathological evaluation of vascular medial fibrosis: insights into the pathophysiology of arterial stiffening. *Atherosclerosis* 208: 69–74, 2010.
  163. Semba RD, Najjar SS, Sun K, Lakatta EG, Ferrucci L. Serum carboxymethyl-lysine, an advanced glycation end product, is associated with increased aortic pulse wave velocity in adults. *Am J Hypertens* 22: 74–79, 2009.
  164. Seymour EM, Parikh RV, Singer AA, Bolling SF. Moderate calorie restriction improves cardiac remodeling and diastolic dysfunction in the Dahl-SS rat. *J Mol Cell Cardiol* 41: 661–668, 2006.
  165. Shechter M, Issachar A, Marai I, Koren-Morag N, Freinark D, Shahar Y, Shechter A, Feinberg MS. Long-term association of brachial artery flow-mediated vasodilation and cardiovascular events in middle-aged subjects with no apparent heart disease. *Int J Cardiol* 134: 52–58, 2009.
  166. Shimokawa I, Higami Y, Hubbard GB, McMahan CA, Masoro EJ, Yu BP. Diet and the suitability of the male Fischer 344 rat as a model for aging research. *J Gerontol* 48: B27–B32, 1993.
  167. Shinmura K, Tamaki K, Sano M, Murata M, Yamakawa H, Ishida H, Fukuda K. Impact of long-term caloric restriction on cardiac senescence: caloric restriction ameliorates cardiac diastolic dysfunction associated with aging. *J Mol Cell Cardiol* 50: 117–127, 2011.
  168. Sindler AL, Delp MD, Reyes R, Wu G, Muller-Delp JM. Effects of ageing and exercise training on eNOS uncoupling in skeletal muscle resistance arterioles. *J Physiol* 587: 3885–3897, 2009.
  169. Soare A, Cangemi R, Omodei D, Holloszy JO, Fontana L. Long-term calorie restriction, but not endurance exercise, lowers core body temperature in humans. *Aging (Albany NY)* 3: 374–379, 2011.
  170. Sohal RS, Weindruch R. Oxidative stress, caloric restriction, and aging. *Science* 273: 59–63, 1996.



171. Sonntag WE, Lynch CD, Cefalu WT, Ingram RL, Bennett SA, Thornton PL, Khan AS. Pleiotropic effects of growth hormone and insulin-like growth factor (IGF)-1 on biological aging: inferences from moderate caloric-restricted animals. *J Gerontol A Biol Sci Med Sci* 54: B521–B538, 1999.
172. Spaulding CC, Walford RL, Effros RB. Calorie restriction inhibits the age-related dysregulation of the cytokines TNF-alpha and IL-6 in C3B10RF1 mice. *Mech Ageing Dev* 93: 87–94, 1997.
173. Spina M, Garbisa S, Hinnie J, Hunter JC, Serafini-Fracassini A. Age-related changes in composition and mechanical properties of the tunica media of the upper thoracic human aorta. *Arteriosclerosis* 3: 64–76, 1983.
174. Stamler J, Mendelsohn ME, Amarante P, Smick D, Andon N, Davies PF, Cooke JP, Loscalzo J. N-acetylcysteine potentiates platelet inhibition by endothelium-derived relaxing factor. *Circ Res* 65: 789–795, 1989.
175. Stowe RP, Peek MK, Cutchin MP, Goodwin JS. Plasma cytokine levels in a population-based study: relation to age and ethnicity. *J Gerontol A Biol Sci Med Sci* 65: 429–433, 2010.
176. Stratton JR, Cerqueira MD, Schwartz RS, Levy WC, Veith RC, Kahn SE, Abrass IB. Differences in cardiovascular responses to isoproterenol in relation to age and exercise training in healthy men. *Circulation* 86: 504–512, 1992.
177. Sung B, Park S, Yu BP, Chung HY. Amelioration of age-related inflammation and oxidative stress by PPARgamma activator: suppression of NF-kappaB by 2,4-thiazolidinedione. *Exp Gerontol* 41: 590–599, 2006.
178. Taddei S, Virdis A, Mattei P, Ghiadoni L, Gennari A, Fasolo CB, Sudano I, Salvetti A. Aging and endothelial function in normotensive subjects and patients with essential hypertension. *Circulation* 91: 1981–1987, 1995.
179. Taffet GE, Pham TT, Hartley CJ. The age-associated alterations in late diastolic function in mice are improved by caloric restriction. *J Gerontol A Biol Sci Med Sci* 52: B285–B290, 1997.
180. Tanabe T, Maeda S, Miyauchi T, Iemitsu M, Takanashi M, Iruka-yama-Tomobe Y, Yokota T, Ohmori H, Matsuda M. Exercise training improves ageing-induced decrease in eNOS expression of the aorta. *Acta Physiol Scand* 178: 3–10, 2003.
181. Tanaka H, Dineno FA, Monahan KD, Clevenger CM, Desouza CA, Seals DR. Aging, habitual exercise, and dynamic arterial compliance. *Circulation* 102: 1270–1275, 2000.
182. Tanaka H, Monahan KD, Seals DR. Age-predicted maximal heart rate revisited. *J Am Coll Cardiol* 37: 153–156, 2001.
183. Thomas J, Bertrand H, Stacy C, Herlihy JT. Long-term caloric restriction improves baroreflex sensitivity in aging Fischer 344 rats. *J Gerontol* 48: B151–B155, 1993.
184. Toda T, Tsuda N, Nishimori I, Leszczynski DE, Kummerow FA. Morphometrical analysis of the aging process in human arteries and aorta. *Acta Anat (Basel)* 106: 35–44, 1980.
185. Trott DW, Seawright JW, Luttrell MJ, Woodman CR. NAD(P)H oxidase-derived reactive oxygen species contribute to age-related impairments of endothelium-dependent dilation in rat soleus feed arteries. *J Appl Physiol* 110: 1171–1180, 2011.
186. Tschudi MR, Barton M, Bersinger NA, Moreau P, Cosentino F, Noll G, Malinski T, Luscher TF. Effect of age on kinetics of nitric oxide release in rat aorta and pulmonary artery. *J Clin Invest* 98: 899–905, 1996.
187. Ungvari Z, Bailey-Downs L, Gautam T, Sosnowska D, Wang M, Monticone RE, Telljohann R, Pinto JT, de Cabo R, Sonntag WE, Lakatta EG, Csizsar A. Age-associated vascular oxidative stress, Nrf2 dysfunction, and NF-kB activation in the nonhuman primate macaca mulatta. *J Gerontol A Biol Sci Med Sci* 66: 866–875, 2011.
188. Ungvari Z, Orosz Z, Labinskyy N, Rivera A, Xiangmin Z, Smith K, Csizsar A. Increased mitochondrial H<sub>2</sub>O<sub>2</sub> production promotes endothelial NF-kB activation in aged rat arteries. *Am J Physiol Heart Circ Physiol* 293: H37–H47, 2007.
189. Ungvari Z, Parrado-Fernandez C, Csizsar A, de Cabo R. Mechanisms underlying caloric restriction and lifespan regulation: implications for vascular aging. *Circ Res* 102: 519–528, 2008.
190. Ungvari ZI, Bailey-Downs L, Sosnowska D, Gautam T, Koncz P, Losonczy G, Ballabh P, de Cabo R, Sonntag WE, Csizsar A. Vascular oxidative stress in aging: a homeostatic failure due to dysregulation of NRF2-mediated antioxidant response. *Am J Physiol Heart Circ Physiol* 301: H363–H372, 2011.
191. Uren NG, Camici PG, Melin JA, Bol A, de Bruyne B, Radvan J, Olivetto I, Rosen SD, Impallomeni M, Wijns W. Effect of aging on myocardial perfusion reserve. *J Nucl Med* 36: 2032–2036, 1995.
192. Vaitkevicius PV, Fleg JL, Engel JH, O'Connor FC, Wright JG, Lakatta LE, Yin FC, Lakatta EG. Effects of age and aerobic capacity on arterial stiffness in healthy adults. *Circulation* 88: 1456–1462, 1993.
193. van der LB, Labugger R, Skepper JN, Bachschmid M, Kilo J, Powell JM, Palacios-Callender M, Erusalimsky JD, Quaschnig T, Malinski T, Gygi D, Ullrich V, Luscher TF. Enhanced peroxynitrite formation is associated with vascular aging. *J Exp Med* 192: 1731–1744, 2000.
194. Vlachopoulos C, Aznaouridis K, Stefanadis C. Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. *J Am Coll Cardiol* 55: 1318–1327, 2010.
195. Vlachopoulos C, Dima I, Aznaouridis K, Vasiliadou C, Ioakeimidis N, Aggeli C, Toutouza M, Stefanadis C. Acute systemic inflammation increases arterial stiffness and decreases wave reflections in healthy individuals. *Circulation* 112: 2193–2200, 2005.
196. Vlachopoulos C, Ioakeimidis N, Aznaouridis K, Bratsas A, Baou K, Xaplanteris P, Lazaros G, Stefanadis C. Association of interleukin-18 levels with global arterial function and early structural changes in men without cardiovascular disease. *Am J Hypertens* 23: 351–357, 2010.
197. Walford RL, Mock D, Verdery R, MacCallum T. Calorie restriction in biosphere 2: alterations in physiologic, hematologic, hormonal, and biochemical parameters in humans restricted for a 2-year period. *J Gerontol A Biol Sci Med Sci* 57: B211–B224, 2002.
198. Watanabe M, Sawai T, Nagura H, Suyama K. Age-related alteration of cross-linking amino acids of elastin in human aorta. *Tohoku J Exp Med* 180: 115–130, 1996.
199. Weindruch R, Sohal RS. Seminars in medicine of the Beth Israel Deaconess Medical Center. Caloric intake and aging. *N Engl J Med* 337: 986–994, 1997.
200. Weindruch R, Walford RW. *The Retardation of Aging and Disease by Dietary Restriction*. Springfield, IL: Thomas, 1988.
201. Weingand KW. Atherosclerosis research in cynomolgus monkeys (*Macaca fascicularis*). *Exp Mol Pathol* 50: 1–15, 1989.
202. Weiss EP, Racette SB, Villareal DT, Fontana L, Steger-May K, Schechtman KB, Klein S, Holloszy JO. Improvements in glucose tolerance and insulin action induced by increasing energy expenditure or decreasing energy intake: a randomized controlled trial. *Am J Clin Nutr* 84: 1033–1042, 2006.
203. Weiss EP, Spina RJ, Holloszy JO, Ehsani AA. Gender differences in the decline in aerobic capacity and its physiological determinants during the later decades of life. *J Appl Physiol* 101: 938–944, 2006.
204. Weiss EP, Villareal DT, Racette SB, Steger-May K, Premachandra BN, Klein S, Fontana L. Caloric restriction but not exercise-induced reductions in fat mass decrease plasma triiodothyronine concentrations: a randomized controlled trial. *Rejuvenation Res* 11: 605–609, 2008.
205. Whidden MA, Kirichenko N, Halici Z, Erdos B, Foster TC, Tumer N. Lifelong caloric restriction prevents age-induced oxidative stress in the sympathoadrenal system of Fischer 344 × Brown Norway rats. *Biochem Biophys Res Commun* 408: 454–458, 2011.
206. White M, Roden R, Minobe W, Khan MF, Larrabee P, Wollmering M, Port JD, Anderson F, Campbell D, Feldman AM. Age-related changes in beta-adrenergic neuroeffector systems in the human heart. *Circulation* 90: 1225–1238, 1994.
207. Wilkinson IB, Qasem A, McEniery CM, Webb DJ, Avolio AP, Cockcroft JR. Nitric oxide regulates local arterial distensibility in vivo. *Circulation* 105: 213–217, 2002.
208. Willcox BJ, Willcox DC, Todoriki H, Fujiyoshi A, Yano K, He Q, Curb JD, Suzuki M. Caloric restriction, the traditional Okinawan diet, and healthy aging: the diet of the world's longest-lived people and its potential impact on morbidity and life span. *Ann NY Acad Sci* 1114: 434–455, 2007.
209. Willcox DC, Willcox BJ, Todoriki H, Suzuki M. The Okinawan diet: health implications of a low-calorie, nutrient-dense, antioxidant-rich dietary pattern low in glycemic load. *J Am Coll Nutr* 28, Suppl: 500S–516S, 2009.
210. Willum-Hansen T, Staessen JA, Torp-Pedersen C, Rasmussen S, Thijs L, Ibsen H, Jeppesen J. Prognostic value of aortic pulse wave velocity as index of arterial stiffness in the general population. *Circulation* 113: 664–670, 2006.
211. Wu D, Marko M, Claycombe K, Paulson KE, Meydani SN. Ceramide-induced and age-associated increase in macrophage COX-2 expres-



- sion is mediated through up-regulation of NF-kappa B activity. *J Biol Chem* 278: 10983–10992, 2003.
212. **Yan J, Stringer SE, Hamilton A, Charlton-Menys V, Gotting C, Muller B, Aeschlimann D, Alexander MY.** Decorin GAG synthesis and TGF-beta signaling mediate Ox-LDL-induced mineralization of human vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 31: 608–615, 2011.
213. **Yeboah J, Crouse JR, Hsu FC, Burke GL, Herrington DM.** Brachial flow-mediated dilation predicts incident cardiovascular events in older adults: the Cardiovascular Health Study. *Circulation* 115: 2390–2397, 2007.
214. **Zanetti M, Gortan CG, Burekovic I, Barazzoni R, Stebel M, Guarnieri G.** Caloric restriction improves endothelial dysfunction during vascular aging: Effects on nitric oxide synthase isoforms and oxidative stress in rat aorta. *Exp Gerontol* 45: 848–855, 2010.
215. **Zeina AR, Odeh M, Rosenschein U, Zaid G, Barmer E.** Coronary artery disease among asymptomatic diabetic and nondiabetic patients undergoing coronary computed tomography angiography. *Coron Artery Dis* 19: 37–41, 2008.
216. **Zenker G, Koltringer P, Bone G, Niederkorn K, Pfeiffer K, Jurgens G.** Lipoprotein(a) as a strong indicator for cerebrovascular disease. *Stroke* 17: 942–945, 1986.
217. **Zhu M, Miura J, Lu LX, Bernier M, DeCabo R, Lane MA, Roth GS, Ingram DK.** Circulating adiponectin levels increase in rats on caloric restriction: the potential for insulin sensitization. *Exp Gerontol* 39: 1049–1059, 2004.
218. **Zou Y, Jung KJ, Kim JW, Yu BP, Chung HY.** Alteration of soluble adhesion molecules during aging and their modulation by calorie restriction. *FASEB J* 18: 320–322, 2004.

