

PDF hosted at the Radboud Repository of the Radboud University Nijmegen

The following full text is a publisher's version.

For additional information about this publication click this link.

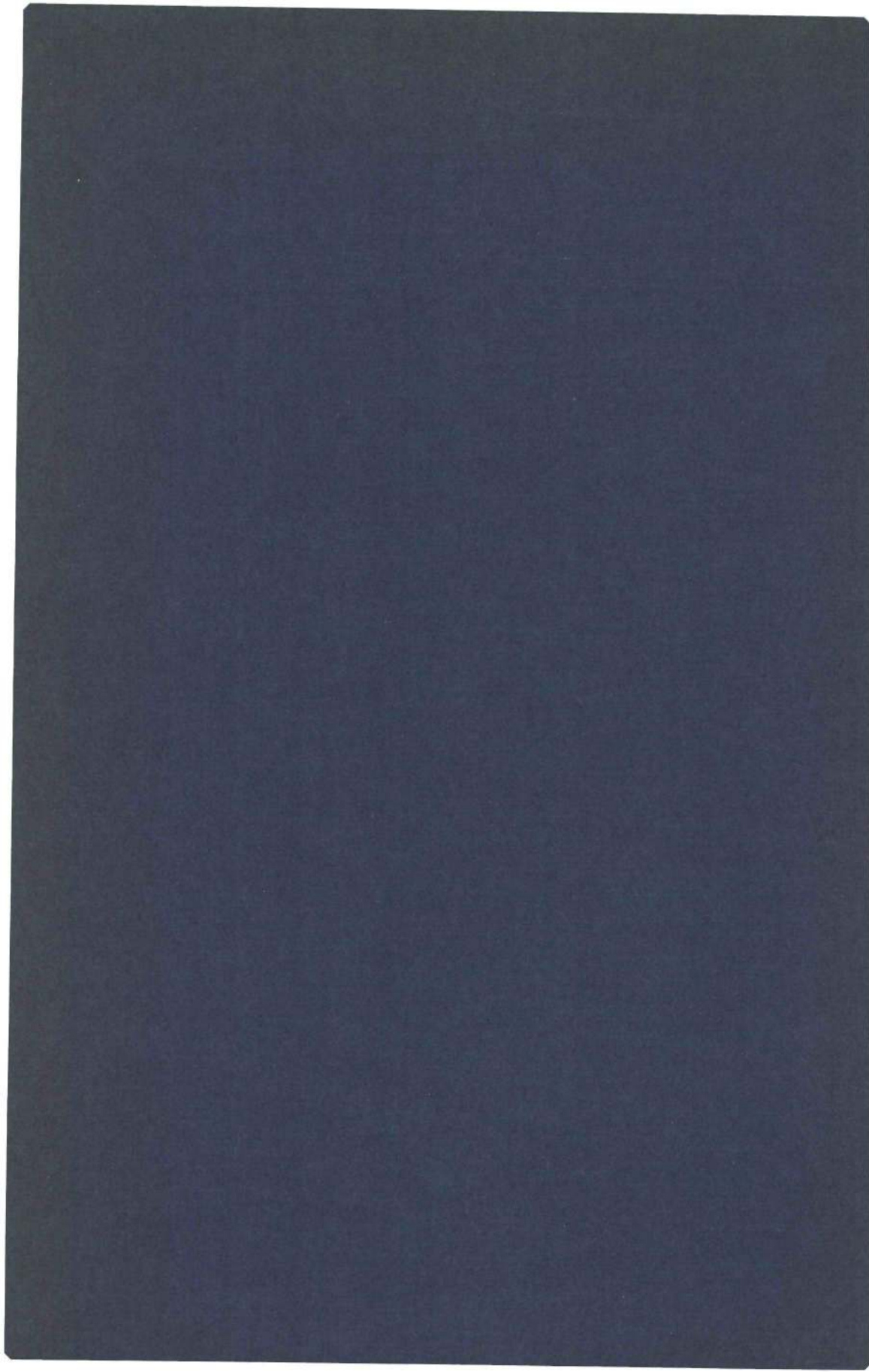
<http://hdl.handle.net/2066/147581>

Please be advised that this information was generated on 2022-08-22 and may be subject to change.

195

CATION TRANSPORT AND
COCHLEAR FUNCTION

W. KUIJPERS



CATION TRANSPORT AND
COCHLEAR FUNCTION

PROMOTORES:

Prof. Dr. S. L. BONTING

EN

Prof. Dr. W. F. B. BRINKMAN

CATION TRANSPORT AND COCHLEAR FUNCTION

PROEFSCHRIFT

TER VERKRIJGING VAN DE GRAAD VAN DOCTOR IN DE
WISKUNDE EN NATUURWETENSCHAPPEN
AAN DE KATHOLIEKE UNIVERSITEIT TE NIJMEGEN,
OP GEZAG VAN DE RECTOR MAGNIFICUS
DR. G. BRENNINKMEIJER,
HOGLERAAR IN DE FACULTEIT DER SOCIALE WETENSCHAPPEN,
VOLGENS BESLUIT VAN DE SENAAT
IN HET OPENBAAR TE VERDEDIGEN OP
VRIJDAG 19 DECEMBER 1969 DES NAMIDDAGS TE 2 UUR

DOOR

WILLIBRORDUS KUIJPERS

GEBOREN

TE KLOOSTERZANDE

1969

CENTRALE DRUKKERIJ NIJMEGEN

I am greatly indebted to Dr. J. F. G. Slegers for his interest and many valuable discussions throughout the course of this investigation. The technical assistance of Miss A. C. H. Janssen, Mr. A. C. van der Vleuten and Mr. P. Spaan and co-workers was greatly appreciated. I also wish to express my gratitude to Miss. A. E. Gonsalves and Miss. G. Kuijpers for typing and to Mrs. M. Duncan for correcting the manuscript.

The diagrams were prepared by Mr W. Maas and Mr. C. Reckers and the microphotographs by Mr. A. Reijnen of the department of medical illustration.

*Aan mijn Ouders.
Thea, Annemarie, Katrien
en Michiel.*

CONTENTS

GENERAL INTRODUCTION	9
CHAPTER I ANATOMY OF THE COCHLEA	
1. Introduction	11
2. Gross anatomy of the labyrinth	11
3. Anatomy of the mammalian cochlea	12
4. The lateral wall of the scala media	14
5. The vestibular wall of the scala media	16
6. The tympanal wall of the scala media	17
7. The ligamentum spirale and limbus spiralis	19
8. Vascularisation	19
9. Innervation	20
10. Sacculus endolymphaticus	20
11. Anatomy of the avian cochlea	21
CHAPTER II THE COCHLEAR FLUIDS	
1. Introduction	23
2. Composition	23
3. Origin and circulation	24
a. Perilymph	24
b. Endolymph	25
c. Cortilymph	27
CHAPTER III ELECTRICAL PHENOMENA IN THE MAMMALIAN COCHLEA	
1. Introduction	28
2. Endocochlear resting potential	28
3. Intracellular DC polarisations	30
4. Cochlear microphonic potential	31
5. Electrical phenomena in the avian cochlea	33
CHAPTER IV CATION TRANSPORT AND THE $\text{Na}^+\text{-K}^+\text{-ATPase}$ SYSTEM	
1. Introduction	34
2. Distribution of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system and correlation with cation transport	34
3. Properties of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system	35
4. Function of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system	37
5. Mechanism of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system	37
6. Purpose of this investigation	38
CHAPTER V MATERIALS AND METHODS	
1. Materials	39
2. Biochemical assays	39
a. Animals	39
b. Tissue preparation	39
c. ATPase determinations	42

3. Potential recording	44
a. Surgical procedure	44
b. Cochlear microphonic potential	44
c. Endolymphatic resting potential	45
d. Perfusion technique	46
CHAPTER VI- OCCURRENCE AND DISTRIBUTION OF THE Na⁺-K⁺-ATPase ACTIVITY IN THE CHICKEN COCHLEA	
1 Introduction	48
2 Distribution of the Na ⁺ -K ⁺ -ATPase activity in the chicken cochlea	48
3. Properties of ATPase activity	49
4. Discussion and conclusions	52
CHAPTER VII OCCURRENCE AND DISTRIBUTION OF Na⁺-K⁺-ATPase ACTIVITY IN THE COCHLEAR STRUCTURES OF THE GUINEA PIG	
1 Introduction	56
2. Presence and properties of the Na ⁺ -K ⁺ -ATPase system	56
3. Distribution of ATPase activities of the various cochlear structures	61
4. Discussion and conclusions	65
CHAPTER VIII THE EFFECT OF OUABAIN ON THE COCHLEAR POTENTIALS OF THE GUINEA PIG	
1. Introduction	69
2. Cochlear microphonic potential	70
3 Endolymphatic resting potential	73
4. Discussion and conclusions	76
CHAPTER IX THE NATURE OF THE ENDOLYMPHATIC RESTING POTENTIAL AND COMPOSITION OF THE PERILYMPH	
1. Introduction	79
2. Effect of electrolyte changes in the perilymph	81
3. Effect of pH on the potential	85
4. Experimental effects on the nature of the negative potential	86
5. Discussion and conclusions	88
SUMMARY	94
SAMENVATTING	96
REFERENCES	98

GENERAL INTRODUCTION

The cochlea is a very complicated structure composed of two compartments filled with fluid and lined with various kinds of epithelium. One of the most unusual phenomena of this organ is the intracellular-like ionic composition of one of these fluids, the endolymph. This situation is unique among extracellular body fluids. Moreover, several remarkable electrical phenomena, intimately connected with cochlear function, and dependent on the ionic composition of the fluids, can be measured in this organ.

In this investigation we have investigated the identity, location and role of the cation pump maintaining the characteristic cationic composition of the endolymph, and participating in the generation of the cochlear potentials. The first part of this thesis consists of a survey of the literature dealing with the histology and histochemistry of the cochlear structures and with the cochlear fluids and potentials. In addition the Na^+ - K^+ -activated ATPase system and its relation to cation transport is reviewed. The second part, dealing with our experiments, describes the experimental procedures and the results obtained and gives the discussion and conclusions of these results.

ANATOMY OF THE COCHLEA

1. Introduction

Our present knowledge of the development and structure of the cochlear tissues is derived from a series of light microscopic investigations (Held, 1926, Bast and Anson, 1949) and more specifically from electronmicroscopic studies performed during the last fifteen years (Smith, 1957, Rodriguez Echandia and Burgos, 1965; Engstrom et al., 1966; Iurato, 1967a, b).

In this chapter the fine structure of the cochlear tissues is described in addition to the general organisation of the whole labyrinth.

2. Gross anatomy of the labyrinth

The membranous labyrinth originates from the otic placode, an epithelial thickening in the region of the rhombencephalon in early embryonic life. The placode sinks inwards to form the closed otic vesicle. By intricate infolding processes this vesicle is transformed into the membranous labyrinth as we know it in the adult animal with its various compartments filled with endolymph and connected to each other by narrow passages (Fig. 1). These compartments can be divided in two functionally different groups: one which is involved in the function of equilibrium (sacculus, utriculus and semicircular canals), the other involved in the perception of sound (the ductus cochlearis or scala media).

In all these compartments the epithelial lining derived from the original epithelium of the otic vesicle, is differentiated into areas with specialized sensory cells, associated with branches of the vestibular or acoustic nerves, and into areas where the cells are thought to be more or less involved in the secretory or resorptive processes of the labyrinthine fluids. In the adult stage the whole membranous labyrinth is encased in a bony capsule and is separated from the latter by a space filled with fluid, the perilymph (Fig. 1)

This perilymphatic space originates from the disappearance of the mesenchymal tissue surrounding the membranous labyrinth during development. In the cochlea, this fluid has the important function of transferring the sound waves for stimulation of the sensory cells of the

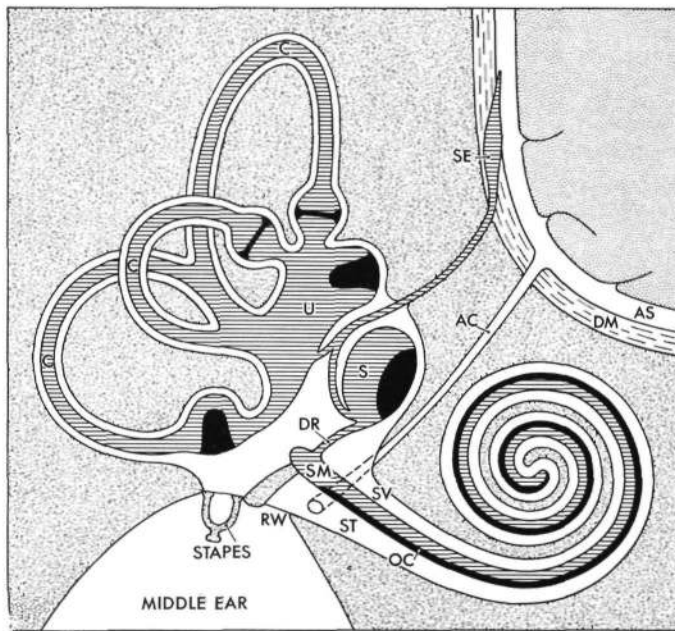


Fig. 1. Schematic drawing of the various compartments of the membranous labyrinth and its interconnections (redrawn and modified from von Ebner, 1903 and Bast and Anson, 1949). Black areas represent sensory cells. The spaces filled with endolymph are shaded and those filled with perilymph are white. AC: aqueductus cochleae, AS: arachnoid space, C: canales semicirculares, DM: dura mater, DR: ductus reuniens, OC: organ of Corti, RW: round window, S: sacculus, SE: saccus endolymphaticus, ST: scala tympani, SV: scala vestibuli, U: utriculus.

organ of Corti. The perilymphatic system is connected with the cerebrospinal fluid space by means of the aqueductus cochleae (Fig. 1). The endolymphatic space has a connection with the arachnoid space by means of the saccus endolymphaticus, a closed sac situated in the dura mater and connected to sacculus and utriculus by the ductus endolymphaticus (Fig. 1).

Since this thesis is mainly concerned with the cochlear physiology only the cochlear structures will be described in detail. After a more extensive description of the mammalian cochlea, a short description of the avian cochlear duct will be given, as some of our experiments were carried out on the cochlear structures of one day old chickens.

3. Anatomy of the mammalian cochlea

The mammalian cochlea consists of a three-fold tube, spirally wound about a central bony axis, the modiolus (Fig. 2), which contains the main blood vessels and the main trunk of the acoustic nerve. The num-

ber of turns varies from species to species. The cochlea of the guinea pig has $4\frac{1}{4}$ turns.

The middle tube is called the ductus cochlearis or scala media, and is filled with endolymph. This scala ends at the apical turn and at the basal turn is connected with the sacculus by means of the very small ductus reuniens. The scala media, triangular in transverse section, is surrounded by spaces filled with perilymph on two sides: the scala vestibuli running up from the oval window to the apex, where it passes into the scala tympani running down to the round window (Fig. 1).

Fig. 2

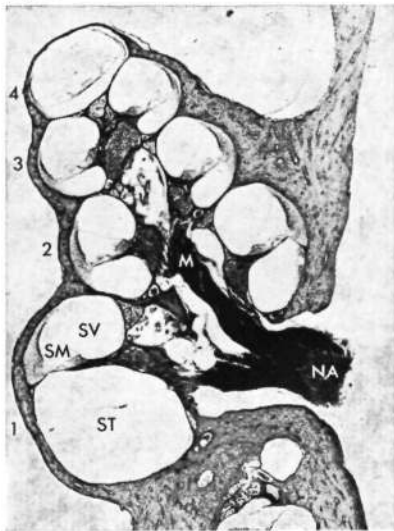


Fig. 3

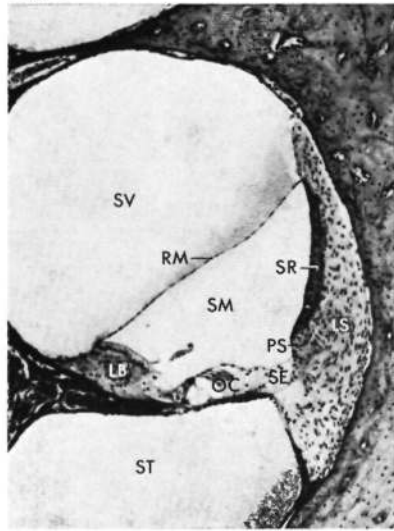


Fig. 2. Median section through the cochlea of the guinea pig (Hematoxylin-eosin, $\times 14$).

Fig. 3. Transverse section through the scala media (Hematoxylin-eosin, $\times 50$). LB: limbus spiralis, LS: ligamentum spirale, OC: organ of Corti, PS: prominentia spiralis, RM: Reissner's membrane, SE: sulcus externus, SM: scala media, ST: scala tympani, SV: scala vestibuli, M: modiolus, NA: nervus acusticus, SR: stria vascularis. Arabic figures refer to the turn numbers.

The scala vestibuli and tympani are separated from neighbouring turns by a bony shelf which contains the arteries and veins supplying the lateral wall of the scala media. These vessels are connected with the main vessels in the modiolus. The separation between scala media and scala vestibuli is formed by Reissner's membrane. The membrana basilaris separates the scala media from the scala tympani. On its lateral side, the epithelium of the scala media rests on a layer of connective tissue, the spiral ligament, which lies directly against the bone of the cochlear capsule.

4. The lateral wall of the scala media

The major part of the epithelial lining of the lateral wall of the scala media is formed by the stria vascularis. A minor part is formed by the epithelium of the prominentia spiralis and sulcus externus (Fig. 3). The fine structure of the stria vascularis has been described by several authors (Smith 1957, Rodriguez Echandia and Burgos, 1965, Rauch and Ruska, 1965, Hinojosa and Rodriguez Echandia, 1966; Spoendlin, 1967a).

It is a stratified epithelium consisting of three cell types with numerous capillaries embedded in it (Fig 4). The marginal cells bordering the endolymphatic space have a rather smooth surface with a few short microvilli and vesicular invaginations with a filamentous coating (Rodriguez Echandia and Burgos, 1965) Near their free surface adjacent cells are firmly connected by junctional complexes, consisting of zonulae occludentes and further down by zonulae adhaerentes and desmosomes as classified by Farquhar and Palade (1963). The basal cell wall shows extensive infoldings so that the linear extent of the free surface is only 3% of the basal surface. These processes interdigitate with each other or with projections of the other cell types, only very small extracellular spaces of about 30 μ width are seen between them (Rodriguez Echandia and Burgos, 1965). The cytoplasm of these infoldings is densely packed with mitochondria. Histochemical studies have revealed a very high concentration of enzymes participating in oxidative metabolism (Vosteen, 1961; Nakai and Hilding, 1968a) and a high incorporation of protein and RNA precursors (Koburg, 1961; Koburg and Plester, 1962) in the stria vascularis. Microrespirometric studies (Chou and Rodgers 1962) revealed a QO_2 for the stria vascularis even higher than that for kidney tissue. The cytoplasm in the apical part of the cell containing the nucleus shows only scattered mitochondria but is well furnished with free and membrane bound ribosomes, a Golgi complex and many vesicles, becoming more numerous and smaller towards the free surface. Many of these vesicles are similar in structure to the coated invaginations at the free surface of the cell. Such structures have been interpreted as a type of pinocytotic activity (Roth and Porter, 1964) but different from that in smooth muscle cells and endothelial cells.

The second layer is formed by the intermediate cells (Fig 4). These have large cytoplasmic processes running between those of the marginal cells or making contact with the underlying basal cells. The cytoplasm contains only a sparse endoplasmatic reticulum and a few mitochondria. The third type is formed by the basal cells, arranged in several firmly packed layers of flat cells connected to each other with desmosomes and zonulae occludentes (Spoendlin, 1967a). They gra-

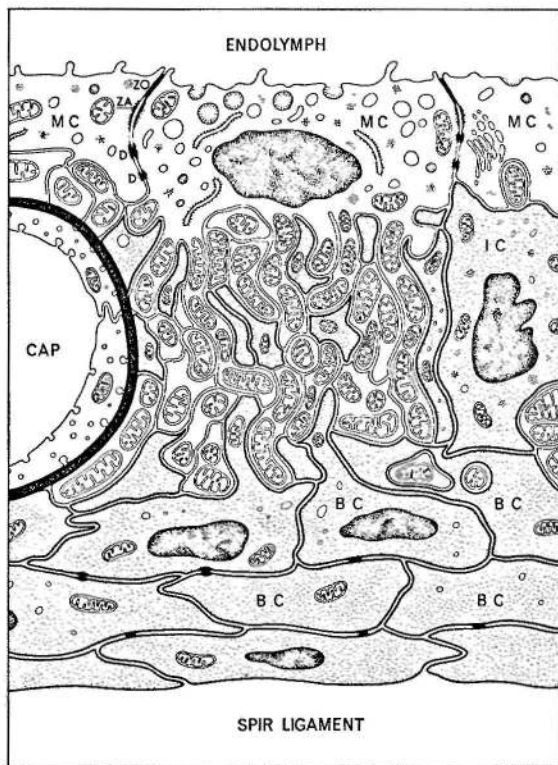


Fig. 4. Diagram of the stria vascularis (modified from Rodriguez Echandia and Burgos, 1965). The various cell types are indicated by MC (marginal cells), IC (intermediate cells) and BC (basal cells). The marginal cells are connected to each other with a junctional complex consisting of a zonula occludens (ZO), zonula adhaerens (ZA) and desmosomes (D). The apical surface shows pinocytotic activity and some microvilli, the basal cell membrane shows multiple infoldings, densely packed with mitochondria and interdigitating with those of the other cell types. Intermediate and basal cells contain only few cytoplasmic inclusions. The basal cells are connected to each other with desmosomes and zonulae occludentes. All types of cells make contact with the basal lamina of the capillaries (CAP) whose endothelial cells show a high pinocytotic activity.

dually change into the cells of the spiral ligament without the intervention of a basal lamina. Some of the basal cells have projections interdigitating with intermediate and marginal cells. As in the intermediate cells the cytoplasm contains only a few organelles.

The numerous capillaries embedded in the epithelium are surrounded by a basal lamina adjacent to the endothelial cells which show many pinocytotic vesicles (Fig. 4). All types of cells make contact with this basal lamina, although extensions of the marginal cells are most frequent.

The epithelium of the spiral prominence and sulcus externus forms a direct continuation of the stria vascularis (Fig. 3). It consists of only one cell layer separated from the spiral ligament by a distinct basal lamina. The epithelial cells of the prominence are cuboidal and possess a very irregular basal surface with numerous invaginations. The sulcus cells penetrate deeply into the spiral ligament. The free surface of the cells shows some microvilli. Adjacent cells are connected by zonulae occludentes near their free surface. The cytoplasm of the prominence cells contains a great amount of endoplasmatic reticulum, Golgi membranes and a few mitochondria. The sulcus cells, however, contain more mitochondria, vacuoles and only a very poorly developed endoplasmatic reticulum. In the spiral ligament behind this epithelium there are capillaries, irregular stroma cells and huge extracellular spaces penetrating into the epithelial invaginations. The stroma cells contain many mitochondria, vacuoles and a well developed endoplasmatic reticulum. Histochemical studies of this region revealed a high activity of several enzymes involved in the oxidative metabolism (Vosteen, 1961; Spoendlin and Balogh, 1963). Roots of the outer sulcus cells may penetrate deeply into this region, sometimes making contact with the stroma cells, and showing extensive membrane infolding associated with a great number of vacuoles (Spoendlin, 1967a; von Ilberg et al., 1968).

5. The vestibular wall of the scala media

The separation between endolymphatic space and scala vestibuli is formed by the avascular membrane of Reissner. Its fine structure has been described by Iurato (1967b); Duvall and Rhodes (1967a); von Ilberg (1968a). Reissner's membrane is composed of two cell layers: an outer layer of very flat cells, continuous with those lining the scala vestibuli (Duvall and Rhodes, 1967a) and an inner layer lining the endolymphatic space. The two cell layers are separated by a basal lamina, bordering the inner layer and a layer of intercellular substance (Fig. 5). Adjacent cells of the outer layer are not always attached to each other, so direct contact between the perilymph and the intercellular substance is present. The inner layer consists of cuboidal cells firmly connected to each other by a zonula occludens near the free surface, followed by a zonula adhaerens and desmosomes. The membrane facing the endolymph shows microvilli and also invaginations with a coated surface (Iurato, 1967b). These invaginations are similar to those found on the free surface of the stria vascularis suggested to be associated with a special form of pinocytosis (Rodriguez Echandia and Burgos, 1965; Hinojosa and Rodriguez Echandia, 1966). Pinocytotic vesicles like those normally found in endothelial cells have also been shown on the side of the basal lamina (Iurato, 1967b; von Ilberg, 1968a). The basal and also the lateral cell walls behind the junctional complex show

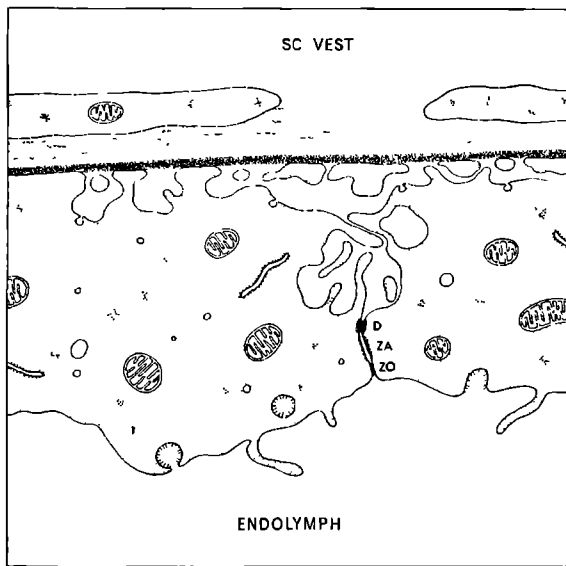


Fig. 5. Diagram of Reissner's membrane based on electron micrographs of Iurato (1957 b) and Duvall and Rhodes (1967 a). The epithelial cells on the endolymphatic side show microvilli and pinocytotic activity on their free surface and are connected to each other with a junctional complex consisting of a zonula occludens (ZO), a zonula adhaerens (ZA) and a desmosome (D). The lateral and basal cell membrane show many infoldings and pinocytotic activity. The epithelial cells on the perilymphatic side, separated from those on the endolymphatic side by a basal lamina and intercellular substance, do not form a continuous layer. Both types of cells contain only a few cytoplasmatic inclusions.

many irregular cell extensions making contact with the basal lamina or interdigitating with those of other cells. The cytoplasm of these cells contains a few mitochondria, some free and membrane bound ribosomes and vesicles of various sizes.

6. The tympanal wall of the scala media

The epithelial lining of the endolymphatic space on the side of the scala tympani consists of various types of cells (Engström et al., 1966; Iurato, 1967 a, b). The major part is formed by the cells of the organ of Corti (Fig. 6). These cells, except for the hair cells, rest directly on the membrana basilaris. This membrane consists of a ground substance with numerous filaments embedded in it. On the perilymphatic side this membrane is covered with a layer of connective tissue cells resembling the outer layer of Reissner's membrane and the rest of the epithelial lining of the perilymphatic space. A small vessel is present in this

membrane in the region of the hair cells, which is important for their nutrition (Kikuchi and Hilding, 1967).

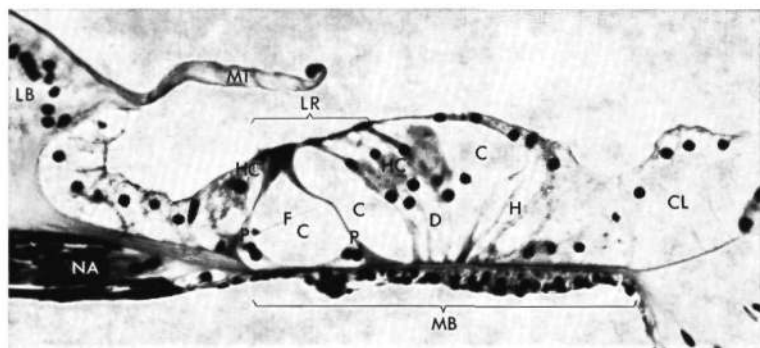


Fig. 6. The organ of Corti of the guinea pig (Hematoxylin-eosin, $\times 180$). C: cortilymph, CL: cells of Claudius, D: cells of Deiters, F: nerve fibres running to the hair cells, LB: limbus spiralis, LR: lamina reticularis, HC: hair cells, H: cells of Hensen, MT: membrana tectoria, MB: membrana basilaris, NA: nervus acusticus, P: pillar cells.

The cells of the organ of Corti can be distinguished in supporting cells with filaments (pillar cells and cells of Deiters) and those without filaments (cells of Hensen and Claudius) which pass into the cells of the sulcus externus. To the modiolar side the organ of Corti passes into the epithelium covering the limbus spiralis, a connective tissue structure resembling the ligamentum spirale. The membrana tectoria, composed of filaments embedded in an amorphous substance, is attached to the epithelium of the spiral limbus. The supporting cells have a rather clear cytoplasm with only a few organelles and vesicles. Moreover, the filamentous cells contain a great number of filaments running from the base to the apex, giving support to the hair cells. The adjacent cells are connected by zonulae occludentes alone in their apical part or followed by zonulae adhaerentes and desmosomes (Iurato, 1967b). The apical surface of the supporting cells shows many microvilli.

The highly differentiated sensory cells, divided into inner and outer hair cells, bear on their apical surface numerous hairs (stereocilia) which are in close contact with the membrana tectoria. The central axis of these hairs penetrates into the apical cytoplasm of the cells, consisting of a cuticular plate (lamina reticularis). Bending of the stereocilia caused by movements of the membrana basilaris, stimulates the hair cells and an action potential in the acoustic nerve is generated. Many mitochondria, vesicles and Golgi lamellae are found in the apical part

of the cell. A second concentration of mitochondria and vesicles is situated in the basal part, where synaptic contact is made with the nerve endings of the acoustic nerve. Along the lateral surface a distinct layer of membrane structures and mitochondria are found. Histochemical studies revealed a high content of enzymes of oxidative and glycolytic metabolism (Vosteen, 1961, Spoendlin and Balogh, 1963, Matschinsky and Thalmann, 1967) in the hair cells.

Around the hair cells several large extracellular spaces continuous with each other can be observed. These spaces which are traversed by the unmyelinated parts of the fibers of the acoustic nerve (Fig. 6), are filled with a fluid called the cortilymph, thought to have the same ionic composition as perilymph.

In view of its structural features, the scala media can be considered to be a closed sac filled with fluid. The tight junctions between the cells bordering the endolymphatic space rule out an intercellular fluid transport as proposed by Farquhar and Palade (1963). From a morphological point of view the stria vascularis would be expected to play the most important role in cochlear cation transport. There is a great resemblance between the marginal cells of the stria vascularis and those found in epithelia shown to be involved in the active transport of ions, such as the dogfish rectal gland (Bulger, 1963), the salt gland of sea birds (Doyle, 1960, Komnick and Komnick 1963) and the kidney tubules (Pease 1955). In the vestibular part of the labyrinth, areas with similar cells have been demonstrated (Dohlman, 1965, Nakai and Hilding, 1968b, Kimura 1969).

7. The ligamentum spirale and limbus spiralis

The ligamentum spirale and limbus spiralis (Figs. 2 and 3) are composed of a very loose fibrillar connective tissue with large extracellular spaces (Iurato, 1967a). These spaces are especially large in the part of the ligamentum spirale bordering the scala tympani, where they may freely communicate with the perilymph (Spoendlin and Balogh, 1963). The fibrous bundles situated in the extracellular spaces form an irregular network in which the connective tissue cells are embedded. In the adult stage most cells show a very poor cytoplasmatic content, except the layers bordering the scala vestibuli of both ligamentum spirale and limbus spiralis, and the cells behind the epithelium of the prominentia spiralis. They contain many mitochondria, which is in agreement with histo-enzymological studies (Vosteen, 1961, Spoendlin and Balogh, 1963).

8. Vascularisation

The vascular supply of the membranous inner ear structures is derived from the arteria cochlearis, located in the modiolus (Axelsson, 1968).

From this artery two main groups of vessels branch off the first group runs through the bony roof of the scala vestibuli and descends through the ligamentum spirale and the second group runs to the limbus spiralis and the organ of Corti. The first group gives off three groups of branches: one above Reissner's membrane which supplies this part of the ligamentum spirale, a second provides the capillary network of the stria vascularis and a third group supplies the prominentia spiralis. All these regions are drained by venules which descend to the collecting venules in the tympanal part of the ligamentum spirale, from whence they course through the floor of the scala tympani to the modiolar vein. The venous drainage of the organ of Corti and limbus spiralis is also connected with the modiolar vein.

9. Innervation

The main part of the cochlear innervation is formed by the afferent and efferent nerve fibres of the acoustic nerve, supplying the sensory cells of the organ of Corti and arising from the main nerve trunk in the modiolus (Figs. 2 and 6). These fibres lose their myelin sheath before they enter the cortilymph space. The innervation pattern of the hair cells has been described extensively by Smith and Sjostrand (1961) and Spoendlin (1967b, 1969). The adrenergic innervation consists of a dense perivascular network in the modiolus but these only accompany the vessels which course to the organ of Corti (Terrayama et al., 1966). Adrenergic fibres could not be demonstrated at all around the capillaries in the ligamentum spirale and stria vascularis. A second adrenergic system has been described, running along the cochlear nerve fibres to the organ of Corti but never reaching the hair cells. This system forms a terminal plexus near the organ of Corti, where the fibres of the acoustic nerve lose their myelin sheath (Spoendlin and Lichtensteiner, 1966).

10. Saccus endolymphaticus

Because of its possible role in endolymph circulation this structure will be briefly described. The saccus is located within two separated layers of the dura mater and is connected with sacculus and utriculus by the ductus endolymphaticus (Fig. 1). Because of its location it has also been suggested (Allen, 1964) that the endolymphatic sac, together with the aqueductus cochleae, represent a mechanism protecting the inner ear from variations in liquor pressure. The submicroscopical structure has been extensively described by Lundquist (1965). The epithelium, one cell layer thick, rests on a loose connective tissue layer permeated by capillaries. Different cell types can be discerned, some of them showing considerable pinocytotic and phagocytotic activity. The lumen of the sac is filled with many free floating macrophages and cell debris.

11. Anatomy of the avian cochlea

Although there are no essential differences in the gross anatomy of the avian and mammalian labyrinth, the avian cochlea shows some remarkable characteristics (Amerlinck, 1923; Held, 1926; Schwartzkopf and Winter, 1960). The avian cochlear duct (scala media), surrounded by perilymphatic spaces, is a sac filled with endolymph, which is only

Fig. 7

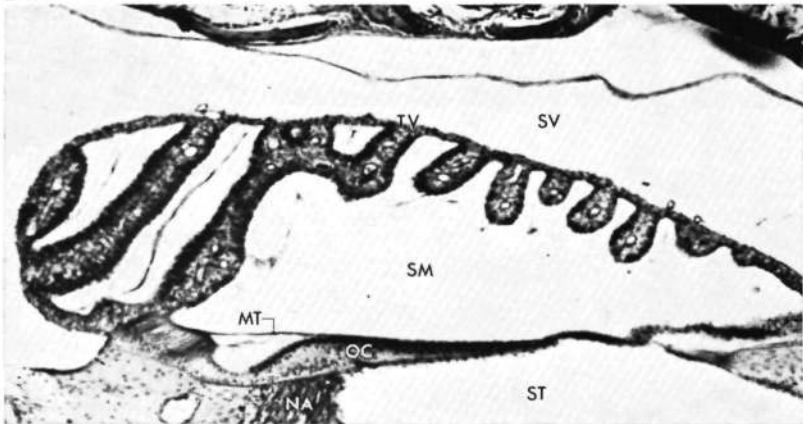


Fig. 8



Fig. 7. Longitudinal section through the cochlea of the one day chicken (Iron-Hematoxylin, $\times 80$).

Fig. 8. Detail of the tegmentum vasculosum showing the epithelial lining of the endolymphatic space, consisting of dark and light cells (Heidenhain's Iron-hematoxylin, $\times 350$). NA: nervus acusticus, MT: membrana tectoria, OC: organ of Corti, SM: scala media, ST: scala tympani, SV: scala vestibuli, TV: tegmentum vasculosum.

slightly curved. The epithelial lining of the scala media on the side of the scala tympani is formed by the sensory and supporting cells of the organ of Corti, resting on the membrana basilaris. The remaining part of the wall of the cochlear sac consists of the tegmentum vasculosum, a highly vascularised and richly folded epithelium with dark and light cells (Figs. 7 and 8). This structure may be considered phylogenetically to represent both Reissner's membrane and the stria vascularis of the mammalian cochlea.

Recently an electronmicroscopical study of the tegmentum was reported (Jahnke et al., 1969). The basal cell membrane of the dark cells showed a great number of invaginations interdigitating with those formed by the light cells. The cytoplasm of the dark cells was densely packed with mitochondria while that of the light cells was very clear.

THE COCHLEAR FLUIDS

1. Introduction

Besides a mechanical role in the transfer of sound vibrations from the middle ear to the organ of Corti, the inner ear fluids play an important role in providing the adequate environment for the function of the sensory cells. Alterations in the ionic composition of these fluids may result in drastic effects on the cochlear potentials (Chapter III) and even lead to degeneration of the epithelial lining of the endolymphatic space (Duvall and Rhodes, 1967b; Duvall, 1968).

This chapter deals with the chemical composition of the cochlear fluids and with the possible role of different cochlear structures in their production and absorption.

2. Composition

Many studies of the composition and biophysical characteristics of the inner ear fluids have been carried out. The first determinations of the electrolyte content of the mammalian inner ear fluids have been reported by Smith et al. (1954). They demonstrated a characteristic difference in the ionic composition between peri- and endolymph. The Na^+ and K^+ content of the perilymph agreed with that of extracellular fluids, but the endolymph proved to be high in K^+ and low in Na^+ thus resembling an intracellular fluid. After this study many authors have reported results of the chemical analysis of cochlear fluids in various animal species. These results have been reviewed by Maggio (1966) and Fernández (1967). Although there are variations in the reported data, it is consistently confirmed that both mammalian cochlear and vestibular endolymph contain a high K^+ and a low Na^+ concentration. Any variations, particularly in data on cochlear endolymph, may be the result of differences in techniques of collecting fluid samples. In view of the anatomical situation it is clear that endolymph samples may easily be contaminated with traces of blood or perilymph. Moreover, some data have been derived from dead animals, which may result in a decrease of K^+ and an increase in Na^+ in the endolymph as shown by Rodgers and Chou (1966); Boshier and Warren (1968); Mendel-

sohn and Konishi (1969). Therefore it seems likely that the most reliable data are those which are low in Na⁺. In Table 1 data on the electrolyte composition of the inner ear fluids reported by various authors, have been summarised.

Table 1
COMPOSITION OF COCHLEAR FLUIDS *

Species	Endolymph			Perilymph			Reference
	Na ⁺	K ⁺	Cl ⁻	Na ⁺	K ⁺	Cl ⁻	
Guinea pig (utricle)	26	142	110	148	5	120	Citron and Exley (1957)
Guinea pig (cochlea)	2	151		137	4		Johnstone et al (1963)
Guinea pig (cochlea)	1	154		138	7		Bosher and Warren (1968)
Guinea pig (cochlea)	5	144					Mendelsohn and Konishi (1969)
Man (cochlea)	13	144	114	139	4	118	Rauch (1964)
Cat (cochlea)	30	173		140	4		Silverstein (1966)

* All concentrations in mmole/l

3. Origin and circulation

Several microscopical and also some biochemical studies, dealing with the source and the circulation of cochlear fluids have been reported. In these studies various substances such as dyes, radioactive isotopes and particles of various origin were locally injected into the cochlear fluids, the cerebrospinal fluid or administered parenterally and were then traced with microscopical and chemical techniques. Results and conclusions have often been contradictory, which is presumably due to the techniques used, e.g. the use of dyes which may alter the membrane permeability or interfere with cellular metabolism. Moreover, because the inner ear structures are very delicate they may easily be injured or even disrupted by local injections, as may be concluded from some experiments.

a. Perilymph

Because the perilymphatic space is connected with the cerebrospinal space by the aqueductus cochleae and by perivascular and perineural spaces, the question whether perilymph is derived from cerebrospinal fluid, has drawn much attention.

Various substances injected into the liquor cerebrospinalis have been shown to be transported to the perilymph (Arnvig, 1951; Altmann and Waltner, 1950, Hughes and Chou, 1963; Schuknecht and El Seifi,

1963), indicating that the perilymph may be at least partially derived from the cerebrospinal fluid. However, occlusion of the aqueduct did not show histological or physiological alterations in the inner ear, suggesting that the cerebrospinal contribution is only a minor one (Schuknecht and El Seifi, 1963). This finding, together with differences in chemical composition, e.g. in protein content (Silverstein, 1966), and the swift appearance of parenterally administered radioisotopes in the perilymph (Rauch, 1964, Choo and Tabowitz, 1964), suggest that the perilymph is essentially a blood ultrafiltrate from the capillaries in the scala vestibuli. It has been demonstrated that perilymph can more or less freely circulate through ligamentum spirale and limbus spiralis from one scala to the other (Tonndorf et al., 1962; von Ilberg, 1968 b, c).

Absorption of perilymph has been shown to occur in the part of the ligamentum spirale bordering scala tympani by the perivascular tissue around the venules (Kırıkca et al., 1961) and by the stroma cells in the region of the prominentia spiralis (von Ilberg et al., 1968).

b. *Endolymph*

Most studies on endolymph circulation deal with the function of the saccus endolymphaticus (Fig. 1). Various substances injected into both cochlear (Guild, 1927; Lundquist, 1965; Ishii et al., 1966) and vestibular endolymph (Doi, 1939; Dohlman and Ormerod, 1960) have been found to accumulate in the saccus lumen, being phagocytised or removed by pinocytosis. Silverstein (1966) was able to show that the fluid flow from the cannulated saccus stopped after occlusion of the ductus endolymphaticus. Kimura (1967) and Beal (1968) showed that prolonged occlusion of the ductus resulted in a hydroph of the cochlear duct, so confirming the flow direction and the absorptive or filtering function of the saccus. Because of the long distance it seems very likely that this structure is only involved in a long term absorption process to clear away cell debris and excess of fluid from the endolymphatic system. Additional arguments in favour of this assumption are the low K^+ , high Na^+ and protein content of the saccus fluid (Silverstein, 1966) and the presence of acid phosphatase and proteolytic enzymes (Ishii et al., 1966).

However, several experiments have been reported indicating a local cochlear circulation which seems to be much more important in cochlear physiology. It has been demonstrated that a local intermixing of peri- and endolymph by rupturing Reissner's membrane only leads to a local disappearance of cochlear microphonic potential and a local degeneration of the epithelium (Lawrence et al., 1961; Schuknecht and El Seifi, 1963; Lawrence, 1966; Duvall and Rhodes, 1967 b; Duvall, 1968). A few mm from this tear the CMP was still present and the

epithelium intact, suggesting a segmental metabolic independence of the cochlear structures.

Von Ilberg (1968 a, b) demonstrated that thorium dioxide injected into the scala media was removed by pinocytotic activity of the stria vascularis and Reissner's membrane. Absorption of endolymph by the epithelium of spiral prominence and sulcus externus as shown by Yamamoto and Nakai (1964), using ferrodextran, could not be confirmed by von Ilberg et al. (1968) Thorium dioxide injected into perilymph could only reach the endolymph by pinocytosis of Reissner's membrane in the reverse direction (von Ilberg, 1968 a)

Naftalin and Harrison (1958) advanced the hypothesis that endolymph arises from perilymph passing through Reissner's membrane, while the stria vascularis extracts the excess Na^+ and pumps in K^+ to build up the required K^+ concentration against the leakage through Reissner's membrane. Support for this hypothesis can be derived from experiments with radioisotopes. Rauch (1964) showed that after intraperitoneal injection of K^{42} the isotope concentration reached a much higher value in endolymph than in the blood or perilymph This was confirmed by Choo and Tabowitz (1965) after intracardiac injection of K^{42} . These authors suggested a prominent role of the stria vascularis in K^+ transport from blood to endolymph. After intraperitoneal injection of Na^{22} , the isotope concentration was consistently lower in endolymph than in perilymph. (Choo and Tabowitz, 1964; Rauch, 1964). Within two minutes after injection of K^{42} into the perilymph of the scala vestibuli the isotope concentration in the endolymph became higher than in the perilymph, while the K^{42} concentration of the stria vascularis was also very high. This transport could be partly inhibited by NaCN , ouabain and iodoacetic acid. From these experiments Rauch (1964, 1966) concluded an active transport of K^{42} from perilymph to endolymph through Reissner's membrane and a resorption through the stria vascularis.

The entrance of Na^{22} from the vestibular perilymph into the endolymph was much slower, reaching half of the perilymph isotope concentration after approximately seven minutes only (Rauch, 1964). The same author failed to show any significant entrance of Na^+ and K^+ isotopes into the endolymph after injection into the perilymph of the scala tympani. Borghesan (1957) suggested that from an anatomical point of view the spiral prominence may also take part in endolymph formation, supposing that this structure produces the "plasmatic" fraction while the stria vascularis produces the "crystalloid" fraction.

Many other cochlear structures have been thought to take part in endolymph secretion or absorption, however, without any experimental evidence. Nevertheless, in spite of the lack of direct experimental evidence, the morphology and metabolic activity of the stria vascularis

suggest that this structure may play an important part in regulating the composition of cochlear endolymph. The role of the other structures may only be a more or less passive one.

c. Cortilymph

Although the exact composition of the fluid surrounding the cells of the organ of Corti (cortilymph) (Fig. 6) is not known, this fluid may be considered as extracellular in composition, resembling perilymph. As suggested by Tasaki (1957) it is highly unlikely that this fluid resembles endolymph, because unmyelinated fibres cannot function in a high K^+ medium. Support for this statement is given by the experimental work of von Ilberg (1968 d) and Schuknecht and El Seifi (1963), showing that perilymph can freely pass the basilar membrane. Rauch (1964) was able to show that cortilymph is rich in Na^+ and poor in K^+ . Therefore it may be concluded that the barrier between perilymph (i.e. cortilymph) and endolymph is formed by the lamina reticularis, the boundary of the cells of the organ of Corti to the endolymph.

ELECTRICAL PHENOMENA IN THE MAMMALIAN
COCHLEA

1. Introduction

Several electrical phenomena can be identified in the cochlea (Davis, 1957; Tasaki, 1957; Wever, 1966). They can be divided into resting DC polarisations and potentials that are generated after acoustic stimulation. The group of resting potentials consists of the endolymphatic or endocochlear potential (ERP), representing an extracellular polarisation of the endolymph which is positive with respect to blood and perilymph, and the negative intracellular potentials of the epithelial cells lining the endolymphatic space.

To the second group belong the action potentials (AP), representing the sum of many spikes generated in the nerve fibres of the acoustic nerve; the cochlear microphonic potentials (CMP), an alternating current response to sound stimuli; and the summing potential (SP), representing a positive or negative change in the resting potential. In this chapter the literature dealing with the nature and origin of the CMP and the ERP will be reviewed. Because the action potential (AP) and the summing potential (SP) have not been studied by us, they will not be discussed.

2. Endocochlear resting potential

The endocochlear resting potential (ERP), discovered by von Békésy in 1952, is an extracellular DC polarisation about 80 mV positive with respect to the perilymph, blood or any inactive tissue of the head. This potential can be measured in the entire endolymphatic space of the mammalian cochlea. This large, positive potential is remarkable in two ways. Most body cells with their high internal K^+ concentration have a negative resting potential of the same magnitude, which is closely related to the K^+ concentration gradient between the intra- and extracellular compartments (Hodgkin and Keynes, 1955; Adrian, 1956). Thus, on the basis of the ionic composition of the endolymph, one would have expected a negative, rather than a positive ERP. The other remarkable point is that the vestibular parts of the endolymphatic system, having the same ionic composition as the cochlear endolymph

(Table 1), have a resting potential of only a few millivolts (Smith et al., 1958, Schmidt, 1963 a). This potential has been shown to be independent of the cochlear resting potential (Schmidt, 1963 b).

Many experiments have been performed in order to demonstrate a dependence of the ERP on the ionic concentrations in peri- and endolymph. Introduction of a high K^+ solution in the endolymph depressed the ERP much less than injection of Na^+ -rich Ringer or perilymph (Tasaki et al., 1954; Davis et al., 1955; Konishi et al., 1966). Konishi and Kelsey (1968 a) reported only a slight effect on the ERP after perfusing the scala tympani with Na -free Ringer. Tetrodotoxin, which blocks the sodium channel in nerve membrane (Loewenstem et al., 1963, Ozaki and Sato, 1965), did not show any effect on the ERP when introduced into the scala tympani or scala media (Konishi and Kelsey, 1968 b). No effect could be demonstrated by injecting high K^+ solution into the scala tympani (Tasaki et al., 1954). Butler (1965), however, showed a decrease of about 50% in the ERP upon replacing the scala tympani perilymph with a solution containing 150 mM K^+ . Replacement of scala vestibuli perilymph with K^+ -rich Ringer had no effect on the ERP (Tasaki et al., 1954; Butler et al, 1962). Katsuki et al (1966) reported a 50% decrease of the ERP after iontophoretic introduction of tetraethylammonium chloride into the scala media, suggesting the blockage of the K^+ channel (Hagiwara and Saito, 1959). However, no effect was found after introduction of this substance into the hair cell region. From these findings it seems unlikely that the ERP represents merely a diffusion potential due to the ionic gradients between endo- and perilymph.

Several findings seem to support the assumption that the ERP is generated by the stria vascularis, a structure absent in the vestibular parts of the endolymphatic system. The ERP has been shown to be highly dependent on oxidative metabolism. A rapid and sharp decrease can be found after reduction of the blood oxygen tension, caused by clamping the trachea, by inhalation of pure N_2 , or after local interruption of the cochlear blood supply (Konishi et al., 1961). The same effect has been shown with local injection of $NaCN$ (Davis et al., 1955; Konishi and Kelsey, 1968 c). Since the stria is the only cochlear epithelial structure which is richly vascularised and has a high oxidative metabolism (Vosteen, 1961; Chou and Rodgers; 1962; Matchinsky and Thalmann, 1967) this structure seems very likely to be the generator of the ERP. Further support for this assumption has been supplied by Tasaki and Spyropoulos (1959). Moving an electrode along the epithelial wall of the scala media, after removal of Reissner's membrane, they found the highest positive potential near the surface of the stria vascularis. The organ of Corti is excluded as a source of the ERP, because this potential is present in animals which lack the organ of

Corti through hereditary or pathological causes (Davis et al., 1958; Tasaki and Spyropoulos, 1959).

A particularly intriguing phenomenon is that prolonged anoxemia lowers the ERP beyond zero to negative values of 40-50 mV within a few minutes, whereupon the potential gradually returns to zero in one or two hours (Konishi et al., 1961). Measurements of the endolymphatic K^+/Na^+ ratio during anoxemia revealed no great changes in this ratio in the first minutes when the potential becomes negative, while the ratio dropped concurrently with the return of the negative potential to zero, suggesting that the negative potential represents an ionic diffusion potential (Johnstone, 1965).

Johnstone (1967) proposed that the ERP could be a Na^+ diffusion potential of a similar nature to that developed during the peak of an action potential in a nerve. This would require a permeability ratio of $K:Na$ of 1:50. The negative potential arising after anoxemia was explained by assuming a change in this permeability ratio to 5:1, caused by lack of oxygen or accumulation of metabolites. According to Konishi et al. (1967) and Konishi and Kelsey (1968 c) this negative potential may represent a depolarisation potential of the cells of the organ of Corti. Alternatively Johnstone (1967) suggested that the positive potential could be generated directly by an electrogenic chloride and/or potassium pump

An active role of Reissner's membrane in the generation of the ERP has been proposed by von Ilberg and Imamura (1966). They reported that after perfusion of the scala vestibuli with oil the positive ERP was greatly and irreversibly decreased. Moreover, they demonstrated a potential difference generated by isolated pieces of Reissner's membrane, mounted in an Ussing chamber with normal Ringer solution on both sides. In two cases this potential could be affected by DNP, but not by ouabain.

Considering these findings no definitive conclusion can be drawn concerning the exact nature of the ERP. From the strong dependence of this potential on the oxidative metabolism the involvement of the stria vascularis in the maintenance of this potential seems highly likely.

3. Intracellular DC polarisations

The organ of Corti (sensory and supporting cells) has consistently been reported to have a negative polarisation of about 50-80 mV relative to the perilymph (von Békésy, 1952; Tasaki et al., 1954; Butler, 1965, Konishi and Kelsey, 1968 c). Only few data are available on the intracellular potentials of the epithelium lining the scala media. Von Békésy (1952) observed negative potentials of about 20 mV relative to perilymph in Reissner's membrane and of 40 mV in the supporting cells of the organ of Corti. This latter finding has been confirmed by

Lawrence (1967). The negative polarisation of the cells of the stria vascularis (von Békésy, 1952) has not been confirmed by other authors. Tasaki et al. (1954) only measured positive potentials in this area. To explain the difference between their observations and those of von Békésy, they suggested that their findings could be due to injury caused by the electrode, resulting in leakage of positive charge from the scala media. Butler (1965) demonstrated that the negative potential in the organ of Corti decreased linearly with the K^+ concentration in the scala tympani, and he suggested that this potential is of extracellular origin. However this seems highly unlikely in view of the high permeability of the membrana basilaris and the extracellular ionic contents of the cortilymph (Chapter II, 3 c). Konishi and Kelsey (1968 a, b) showed no change in this potential after the introduction of tetrodotoxin or Na^+ free solutions into the scala tympani. The effect of prolonged anoxemia was much slower than for the CMP and ERP, a decrease of only 40% being found after thirty minutes (Butler, 1965). A depressing effect of NaCN introduced into the scala tympani on this potential has also been described (Konishi and Kelsey, 1968 c).

In spite of the few data reported on the DC polarisations of the epithelium lining the endolymphatic space, the cell interior is most likely negatively charged with respect to the outer side. As for the cells of the organ of Corti it seems confirmed that this potential is mainly built up by the K^+ gradient across the cell membrane, as has been demonstrated in many other cells.

4. Cochlear microphonic potential

The cochlear microphonic potential (CMP) is an alternating electrical potential which at low and moderate sound intensities accurately and without measurable phase difference reflects the displacement of the basilar membrane or more exactly the displacement of the hair-bearing ends of the hair cells (lamina reticularis) relative to the membrana tectoria (von Békésy, 1960, Davis, 1957, 1961). There is no true threshold and no refractory period up to frequencies of 100 kc. The CMP can be measured on the outer side of the cochlea (e.g. at the round window) but also, and with greater amplitude, in the cochlear structures and fluids. The CMP for both low and high frequencies can be derived from the basal turns, in the apical turns only responses to low frequencies can be found (Tasaki et al., 1952). This agrees with the mechanical movement of the basilar membrane (von Bekesy, 1960). The CMP, like the positive ERP, has been shown to be highly dependent on oxidative metabolism (Misrahy et al., 1958 a; Konishi et al., 1961; Rice and Shinaberger, 1961; Chambers and Lucchina, 1966; Konishi and Kelsey, 1968 c), although an anoxic fraction of about 10% persists for hours after death (Davis, 1957).

Upon moving an electrode from the scala tympani through the basilar membrane to the scala vestibuli, the amplitude of the CMP was strongly enhanced in the region of the organ of Corti, sometimes reaching a maximum of a few mV (Tasaki, 1957). Upon entering the scala media, the phase of the CMP suddenly reversed, without change in the amplitude. During further penetration into the scala media no changes were observed. After penetration of Reissner's membrane the amplitude immediately decreased, without change in the phase however. These findings strongly suggest that the CMP is generated at or near the lamina reticularis. Davis, (1957, 1961) concluded that it represents a current flow across the hair bearing surface of the hair cells, as the membrane resistance changes with deflection of the hairs of the hair cells. This current was suggested to be driven by the ERP and the negative DC polarisation of the hair cells.

Additional support for this assumption can be derived from several experiments. A DC polarising current, with the positive pole in the scala media or vestibuli and the negative pole in the scala tympani, increased the amplitude of the CMP. A current flowing in the reverse direction diminished the amplitude (Tasaki and Fernández, 1952; Tasaki et al., 1954; Konishi and Yasuno, 1963). Application of hydrostatic pressure to the various cochlear compartments showed that moving the organ of Corti to the scala vestibuli reduced the ERP and moving it to the scala tympani increased that potential (Tasaki et al., 1954). These effects are presumably due to a decrease and increase of the leakage current from scala media to scala tympani (Johnstone and Johnstone, 1966). Moreover, Johnstone et al. (1966) showed a decrease of the effective resistance of the scala media from a loud sound resulting in a voltage drop of 6 mV.

In addition the CMP, like the positive ERP, was depressed, when endolymph was replaced by a K^+ -poor solution (Konishi et al., 1966). On replacement of scala vestibuli perilymph with K^+ -rich solutions no clear effect on the CMP could be found. However, when a K^+ -rich solution was introduced into the scala tympani CMP was strongly depressed (Tasaki and Fernández, 1952; Tasaki et al., 1954, Butler et al., 1962). From the lack of an effect on the CMP of replacing scala tympani perilymph with Na^+ -free solution or a solution containing tetrodotoxin, Konishi and Kelsey (1968 a, 1968 b) concluded that this potential is dependent on the high K^+ concentration in the endolymph. The lack of effect of tetrodotoxin has been confirmed by Katsuki et al. (1966), using an iontophoretical technique. Butler (1965) demonstrated that the decline of the CMP after anoxemia closely follows that of the algebraic sum of ERP and the negative potential of the organ of Corti, i.e. the DC gradient across the lamina reticularis, suggesting that the CMP represents a modulation of both resting potentials.

The involvement of an active cation pump in the generation of the CMP has recently been suggested by Matsuura et al (1968) He found that the CMP recorded from the hair cells of the saccular macula of the goldfish was inhibited by ouabain applied to the perilymphatic side but not to the endolymphatic side. Application of a solution containing 120 mM K^+ showed the same effect.

From the reported data it may be concluded that the existence of the CMP is highly dependent on the presence of the ERP and the negative DC polarisation of the hair cells. In addition many experiments are in favour of the hypothesis that the CMP would represent an alternating current, governed by changes in the resistance of the lamina reticularis.

5. Electrical phenomena in the avian cochlea

The same potentials demonstrated in the mammalian cochlea have been observed in the cochlea of birds, although they are smaller in magnitude (Schmidt and Fernández, 1962).

The ERP of about 20 mV in the avian cochlea appeared to be highly sensitive to anoxia and higher than the resting potentials in the labyrinthine parts of the endolymphatic system, as has also been demonstrated in mammals (Smith et al., 1958). The labyrinthine potentials have been shown to be independent of the cochlear resting potential (Schmidt, 1963 b).

In view of these findings it may be concluded that no essential differences seem to exist between the electrical phenomena in the mammalian and the avian cochlea.

CATION TRANSPORT AND THE $\text{Na}^+\text{-K}^+\text{-ATPase}$ SYSTEM

1. Introduction

It has now been widely accepted that the maintenance of the high K^+ and low Na^+ concentrations in most animal cells, against the reverse concentrations in the extracellular medium, depends on a cation pump or transport system. This system moves cations through cell membranes against their electrochemical gradients, which process is defined as active transport (Ussing, 1949). Since active transport is an energy-requiring process, it appeared likely that ATP provides the energy for this process. Definitive proof was supplied by experiments in which ATP was introduced into resealed erythrocyte ghosts (Gárdos, 1954) and into the squid giant axon (Caldwell et al., 1960). This explains the inhibition of active cation transport by inhibitors of energy metabolism.

In addition active cation transport can be specifically inhibited by cardiac glycosides, as first shown by Schatzmann (1953) in erythrocytes. In 1957 Skou reported that particles from crab nerve homogenates contained a Mg^{2+} activated ATPase activity, which could be increased by the simultaneous addition of Na^+ and K^+ ions. The increased activity could be inhibited by cardiac glycosides (Skou, 1960). These findings led him to suggest that this $\text{Na}^+\text{-K}^+$ activated ATPase ($\text{Na}^+\text{-K}^+\text{-ATPase}$) activity might be involved in active cation transport across the cell membrane. Subsequently, the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system has been demonstrated in a great variety of tissues and shown to be intimately connected with active cation transport in many biological systems.

2. Distribution of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system and correlation with cation transport

The discovery of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in crab nerve membranes has been followed by a great number of publications demonstrating the presence of this enzyme system in a great variety of tissues from various animal species, including invertebrates, and recently it has even been demonstrated in bacteria (Hafkenschied and Bonting, 1968, 1969).

Post et al. (1960) and Dunham and Glynn (1961) showed the presence of this enzyme system in erythrocyte membranes. Moreover, it appeared from experiments with resealed erythrocyte ghosts that the properties of the enzyme system were in good agreement with the properties of the cation transport system (Dunham and Glynn, 1961; Post et al., 1960; Whittam, 1962).

In a systematic study on the cat Bonting et al. (1961 a) showed the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity present in nearly all tissues. It was only absent in noncellular tissues or tissues with low cell density. The highest activities have been shown in tissues with a secretory function such as chorioid plexus (Vates et al., 1964), rectal gland of elasmobranchs (Bonting, 1966), nasal gland of marine-birds (Bonting et al., 1964 a), kidney (Bonting et al., 1961 a, 1962) or in tissues involved in excitatory processes such as nerve (Jarnefelt, 1961), brain (Bonting et al., 1961 a, 1962), retina (Bonting et al., 1964 c) and electroplax (Bonting et al., 1961 b). In another report Bonting and Caravaggio (1963) demonstrated a close quantitative correlation between the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity and the cation fluxes in a variety of tissues, calculating an average cation/ATP ratio of 3.

Tosteson et al. (1960) and Baker and Simmonds (1966) were able to show that the erythrocytes of certain varieties of sheep and opossum with a high content of potassium show a much higher $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity than those with a low concentration of potassium. Wiley (1969) demonstrated that erythrocytes from patients with hereditary spherocytosis, show a high sodium leak into the cell which was accompanied by an enhanced $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity. When ducks were transferred from a fresh water to a salt water diet, an increase was observed in the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity of the nasal gland, which serves as an extrarenal $\text{Na}^+\text{-secretory}$ system (Fletcher et al., 1967). In the developing brain a parallel increase has been found between $\text{Na}^+\text{-K}^+\text{-ATPase}$ and neural activity (Bignami et al., 1966; Abdel-Latif et al., 1967). These examples clearly show the role of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in active cation transport.

The localisation of the enzyme activity in the cell membrane, a prerequisite for its physiological role, has been confirmed by various authors: nervous tissue (Bonting et al., 1962; Cummins and Hyden, 1962), erythrocytes (Post et al., 1960; Dunham and Glynn, 1961), liver (Bonting et al., 1962; Emmelot and Bos, 1966), kidney (Landon and Norris, 1963), muscle (Portius and Repke, 1967), intestine (Berg and Chapman, 1965, Rosenberg and Rosenberg, 1968).

3. Properties of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system

The enzyme system requires for activation Na^+ and K^+ together, in addition to Mg^{2+} . K^+ can be replaced by NH_4^+ , Rb^+ and Cs^+ . Although

there are slight differences in the concentrations of Na^+ and K^+ giving half maximal activity in various tissues, activation by K^+ in the presence of Na^+ occurs at much lower concentrations than by Na^+ in the presence of K^+ (Skou, 1957; Post et al., 1960, Bonting et al., 1964 a; Bonting, 1966; Bakkeren and Bonting, 1968). From experiments with erythrocyte ghosts (Post et al., 1960; Dunham and Glynn, 1961; Whittam, 1962; Garrahan and Glynn, 1967 a, b) and nerve axons (Baker, 1963, Baker and Connelly, 1966, Baker et al., 1969) it has been shown that K^+ activates on the outside of the membrane and Na^+ on the inside of the membrane. A high concentration of K^+ on the inner side can compete with Na^+ and inhibit transport.

The only substrate that can be utilised is ATP, with UTP, ITP and GTP little or no activity could be found (Skou, 1960; Post et al., 1960). The pH optima for $\text{Na}^+\text{-K}^+\text{-ATPase}$ activities from various tissues may vary slightly (7.0-7.5) but they are very different from those of the glycoside-insensitive residual $\text{Mg}^{2+}\text{-ATPase}$, which range from 8.4-8.9 (Skou, 1960; Taylor, 1962, Bonting et al., 1964a; Bonting, 1966; Rosenberg and Rosenberg, 1968; Ridderstap and Bonting, 1969 a, b).

The $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity is specifically inhibited by cardiac glycosides. Although the sensitivity of the enzyme system in various tissues and animal species may vary, there is a striking resemblance between enzyme and transport system in relation to inhibition by cardiac glycosides (Post et al., 1960; Vates et al., 1964; Ridderstap and Bonting, 1969 a, b). The inhibitory effect of cardiac glycosides in both systems at partially inhibitory concentrations may be antagonised by raising the K^+ concentration (Dunham and Glynn, 1961; Bonting et al., 1963; Judah and Ahmed, 1964). A stimulatory effect of cardiac glycosides at very low concentrations has been demonstrated for the enzyme activity (Bonting et al., 1964 a; Bonting, 1966; Vates et al., 1964; Palmer et al., 1966), and this biphasic effect has also been shown in transport inhibition (Palmer and Nechay, 1964; Oppelt and Palmer, 1966; Ridderstap and Bonting, 1969 b). Although chemically different from the digitalis substances, erythropheum alkaloids are also strongly inhibitory to $\text{Na}^+\text{-K}^+\text{-ATPase}$ and cation transport (Bonting et al., 1964 b; Vates et al., 1964; Ridderstap and Bonting, 1969 b). Low concentrations of Ca^{2+} inhibit enzyme activity and cation transport (Skou, 1957; Hoffman, 1962) presumably by the formation of a $\text{Ca}^{2+}\text{-ATP}$ complex which may be competitive with the $\text{Mg}^{2+}\text{-ATP}$ substrate complex (Epstein and Whittam, 1966).

From these observations it may be concluded that de $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in a great variety of tissues represents the same enzyme system.

4. Function of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system

The primary function of the cation pump is to maintain the ionic concentration gradients between intra- and extracellular compartments necessary for cell metabolism and prevention of swelling of the cell. In addition the cation pump plays a central role in other processes. In excitatory tissues such as nerves and muscles, it maintains and restores the ionic gradients after depolarisation.

The enzyme system has also been shown to play an important role in secretory processes which depend on an active Na^+ transport, such as the formation of aqueous humor (Bonting and Becker, 1964), cerebrospinal fluid (Vates et al., 1964) pancreatic juice (Ridderstap and Bonting, 1969 a, b) and in sweat secretion (Slegers, 1968). In the nasal gland of marine birds (Bonting et al., 1964 a, Fletcher et al., 1967) and salt gland of elasmobranchs (Bonting, 1966) it is involved in removal of excess Na^+ from the blood. It also functions in the reabsorption of Na^+ in kidney (Palmer and Nechay, 1964; Katz and Epstein, 1967) and toad bladder (Bonting and Canady, 1964).

In addition, many instances have been found, where the transport of nonelectrolytes is coupled to the function of the cation pump (Crane, 1965); Rosenberg et al, 1965, Newey et al, 1968).

5. Mechanism of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system

A number of experiments during the past ten years have demonstrated that the hydrolysis of ATP by the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system does not occur in a single step. Skou (1960) showed that a membrane fraction, containing $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity, could also catalyze an ATP-ADP exchange reaction, suggesting the formation of a phosphorylated intermediate. The use of ^{32}P -labelled ATP as substrate for membrane fragments revealed that this ATP-ADP exchange was activated in the presence of Mg^{2+} and Na^+ and that the phosphate group was transferred to some intermediate in the membrane (Albers et al, 1963; Charnock and Post, 1963; Heinz and Hoffman, 1965). This suggests that the first step consists of a $\text{Na}^+\text{-Mg}^{2+}$ activated transfer of an energy-rich phosphate to the enzyme system. Upon contact with K^+ this phosphate group is released as inorganic phosphate into the cell interior (Bader et al, 1968; Fahn et al., 1968), representing the second step. This second step can be inhibited by very low concentrations of cardiac glycosides (Post et al, 1965, Ahmed and Judah, 1965), while the Na^+ dependent phosphorylation is only inhibited at very high concentrations of cardiac glycosides. The K^+ -activated dephosphorylation step is supposed to be intimately related to the glycoside sensitive K^+ -phosphatase activity observed in various tissues (Albers and Koval, 1966; Bader and Sen, 1966; Bakkeren, 1968).

Our knowledge of the structure of this enzyme system is still very limited, because it has not been solubilised and therefore the normal techniques of structural analysis of proteins cannot be applied. Some models for the system have been proposed. Opit and Charnock (1965) suggested that the enzyme system has a peptide back bone, which has many negatively charged sites capable of binding Na^+ and K^+ as counter ions. When the internal Na^+ concentration is high, most negative sites will be occupied by Na^+ , resulting in a phosphorylation of the enzyme system by ATP. This would cause the peptide chain to elongate slightly and to rotate in an outward direction between two adjacent phosphorylation sites. The Na^+ -carrying sites are now positioned outwardly and Na^+ is exchanged for K^+ followed by dephosphorylation and backward rotation of the membrane. A somewhat similar model has been proposed by Albers and coworkers (1968), assuming that the enzyme system may transform from a "cis" into a "trans" form. In the "cis" form the cation binding sites are oriented to the cell interior. Excess of Na^+ ions activates the transphosphorylation, and the enzyme is transformed into the "trans" form and the binding sites oriented to the outside, transporting Na^+ to the outside of the membrane. In the presence of K^+ , Na^+ is exchanged for K^+ and dephosphorylation takes place. The "trans" form then returns to the "cis" form, transporting K^+ ions into the cell.

6. Purpose of this investigation

The purpose of this investigation was to determine whether the cation gradients existing between peri- and endolymph, or blood and endolymph, are maintained by the Na^+ - K^+ -activated ATPase system, which has been shown to play an essential role in maintaining the cationic gradients in a great number of tissues.

The first object was to locate the site of the cation pump system. Since a reliable histochemical method for the demonstration of Na^+ - K^+ -ATPase activity is still lacking, we have obtained the quantitative distribution of the enzyme activity in the cochlea by using microdissection and ultra-micro enzyme assay techniques.

The second object was to demonstrate a relationship between the cochlear potentials and the functioning of the Na^+ - K^+ -ATPase system. For this purpose we made use of the specific inhibitory effect of ouabain on the Na^+ - K^+ -ATPase activity by perfusing this substance through the inner ear of the anaesthetised animal and simultaneously measuring its effect on the potentials.

Finally a number of experiments were carried out to elucidate the nature of the large, positive endolymphatic resting potential.

MATERIALS AND METHODS

1. Materials

The following chemicals and materials were used: adenosinetriphosphate, disodium salt (1); ethylenediamine-tetra-acetate, disodium salt (2); tris- (hydroxymethyl)-aminomethane (2); erythrophleine sulphate (2); ouabain (3); tris-adenosinetriphosphate (4); acetazolamide (5); lactic acid (6);

1. C. F. Boehringer und Soehne G.m.b.H., Mannheim, Germany.
2. E. Merck A.G., Darmstadt, Germany.
3. Nutritional Biochemicals Corporation, Cleveland, Ohio, U.S.A.
4. Sigma Chemical Company, St. Louis, Missouri, U.S.A.
5. American Cyanamid Company, Pearl River, New York, U.S.A.
6. The British Drug Houses Ltd., Poole, England.

2. Biochemical assays

a. *Animals*

Apart from the use of one day old chicken for ATPase assays, all further experiments have been carried out on guinea pigs.

For both biochemical and electrophysiological experiments young adult guinea pigs of a pigmented strain were used (250-350 g body weight). In the pigmented animals the stria vascularis is pigmented, facilitating its recognition and separation from the other inner ear structures for enzyme assay. Moreover, in most animals this band of pigmented cells, marking the lateral wall of the scala media, is visible through the bony cochlear wall, indicating the right place to pierce a microelectrode into the endolymphatic space.

b. *Tissue preparation*

For the enzyme assays the inner ear structures of one day old chickens and of guinea pigs were dissected within 45 minutes after death. Immediately after decapitation a piece of tissue containing the inner ear was separated from the skull and brought to 0° C. All further dissection procedures were carried out at 0° C. with the aid of a Zeiss stereo-microscope.

Chicken

In order to isolate the membranous inner ear of the one day old chicken a transverse section was made through the head in the otic region. The inner ear, extending from the middle ear to the middle of the base of the skull, could be recognised by its hyaline cartilaginous capsule. With a small piece of razor blade this capsule was carefully cut away until the membranous cochlear sac became exposed. After sectioning the acoustic nerve the whole sac could be lifted out of its capsule with fine forceps. For isolation of the tegmentum vasculosum the whole cochlear sac was placed on a glass slide and with a fine hair, mounted in glass, this lobulated structure could be separated from the rest. Because of the high viscosity of the avian endolymph, due to a high mucopolysaccharide content (Dohlman and Ormerod, 1960), a variable amount of this fluid could not be removed and adhered to the tegmentum vasculosum. This will result in a variable reduction of the true enzyme activity, based on tissue dry weight

Guinea pig

After decapitation the whole temporal bone of the guinea pig was dissected. The wall of the middle ear was removed to give wide exposure of the cochlea, protruding into the middle ear cavity (Fig 9). Afterwards the bony cochlear capsule was carefully chipped away and the membranous inner ear became visible. With the use of a small hook and fine forceps, a piece of tissue consisting of ligamentum spirale and stria vascularis was loosened and dissected turn by turn proceeding from apex to base. After stretching this tissue on a glass slide, the pigmented stria vascularis could be scraped from the underlying spiral ligament with the use of a hair mounted in a holder. In order to study the distribution of enzyme activity in the various parts of the ligamentum spirale, this structure was divided longitudinally into three parts with the aid of a piece of razor blade: part A behind the stria vascularis and bordering the scala vestibuli, part B in the region of the prominentia spiralis and sulcus externus and part C bordering the scala tympani (Fig 23). After dissection, all tissues were frozen on dry ice in small glass tubes, lyophilised for several hours in tubes with silicagel at -25°C and stored in vacuo at the same temperature until used. No significant loss of activity was noted over periods of storage up to four weeks. For the isolation of Reissner's membrane and the organ of Corti another method had to be used because these structures could only rarely be distinguished in the fresh inner ear.

Dissection of these structures was possible after lyophilising the whole cochlea. The cochlea was freed from adhering bone without damaging the bony cochlear capsule. Both oval and round windows

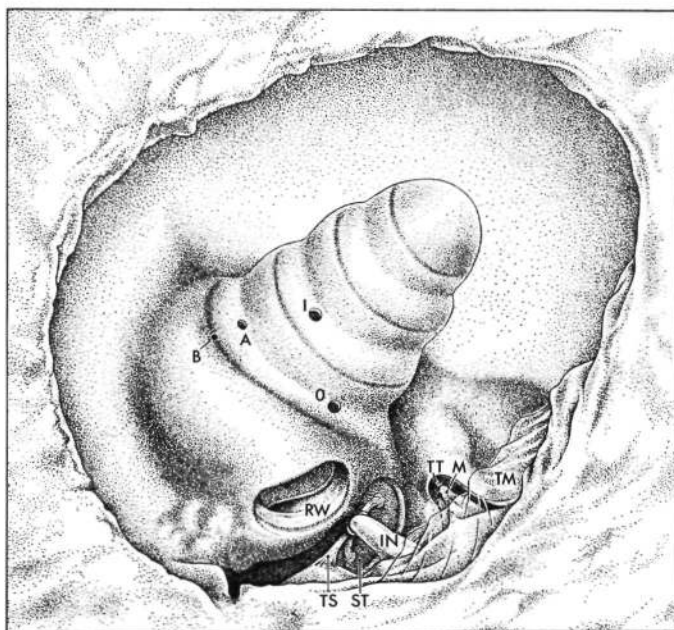


Fig. 9. Drawing of the position in the guinea pig middle ear of the cochlea. The cochlea was exposed by removal of part of the bony wall of the middle ear and of the main part of the tympanic membrane to obtain a better view of the windows. The dark area in the middle of the various turns indicates the stria vascularis. The holes made in the bony wall for placing the electrodes and for perfusing the scala vestibuli are also represented. A: position of CMP electrode, B: position of ERP electrode, I: inlet for perfusion, IN: incus, M: malleus, O: outlet for perfusion, RW: round window, ST: stapes, TM: tympanic membrane, TS: tendon of musculus stapedius, TT: tendon of musculus tensor tympani.

were opened and small pieces of filter paper were introduced to remove the perilymph. Thereafter the freezing and lyophilisation procedures previously mentioned were used. For dissection, the bony cochlear capsule was carefully removed. Reissner's membrane could then be easily discerned and isolated after loosening it from ligamentum spirale and limbus spiralis with the aid of a fine hair. Any dried lymph adhering to its surfaces was carefully removed. After removal of Reissner's membrane and the dried endolymph, the epithelial structures on the membrana basilaris became visible. The organ of Corti, forming a small ridge, was localised and lifted from the basilar membrane. Enzyme assays carried out on samples of the stria vascularis, isolated before and after lyophilisation, did not reveal any significant difference in ATPase activity.

c. ATPase determinations

(Na⁺-K⁺) activated and Mg²⁺-activated ATPase activities were determined with the method described by Bonting et al. (1963) by measuring inorganic phosphate liberated from the enzymatic breakdown of ATP in various substrate media (Table 2). This method is based on the

Table 2
COMPOSITION OF SUBSTRATE MEDIA USED FOR THE ASSAY
OF NA⁺-K⁺ ACTIVATED ATPase

	A	B	C	D	E
Mg ²⁺	1	1	1	1	1
Na ⁺	58	63	—	57	62
K ⁺	5	—	5	5	—
CN ⁻	10	10	5	10	10
EDTA	0.1	0.1	0.1	0.1	0.1
Tris-buffer pH 7.5	92	92	151	91	91
ATP	2	2	2	2	2
Ouabain	—	—	—	0.1	0.1

All concentrations in mmole/l final concentration, only other ionic species present was chloride

specific properties of the (Na⁺-K⁺)-ATPase system. it requires for activation both Na⁺ and K⁺ in the presence of Mg²⁺ and is specifically inhibited by cardiac glycosides such as ouabain, distinguishing it from the unspecific Mg²⁺-ATPase activity. Total ATPase activity was measured in medium A, containing Na⁺, K⁺, Mg²⁺ and no ouabain in addition to ATP and buffer. The media B, C, D and E are inhibitory to the Na⁺-K⁺-ATPase activity through the absence of K⁺ or Na⁺ or the presence of ouabain or the combination of absence of K⁺ and presence of ouabain. These media give only the Mg²⁺ activated ATPase activity. The difference between the activity in medium A and the activity in media B, C, D and E represents the Na⁺-K⁺-ATPase activity. Lyophilised pooled tissue samples of tegmentum vasculosum, stria vascularis and ligamentum spirale were weighed on a Cahn electrobalance and homogenised in twice distilled water in an all glass Potter-Elvehjem tissue grinder (0.04-0.15 mg dr wt/100 µl). Aliquots of 10 µl of the homogenates were added to 150 µl of each of the five media. From these mixtures six aliquots of 10 µl were transferred to microtest tubes, three of which were incubated for 1 hour at 37° C., the others remained in ice and served as enzyme blanks. Enzyme activity was stopped by adding 45 µl of 10% (w/v) trichloro-acetic acid to

each tube. The same quantity of TCA was added to the blanks immediately after placing the other tubes in the incubating bath. After centrifugation, to remove precipitated protein, 30 μ l of the supernatant was transferred to another microtube, containing 30 μ l of a colour reagent. The colour reagent was prepared by dissolving 400 mg FeSO_4 in 5 ml of a 1% NH_4 -molybdate solution in 1.15 N H_2SO_4 and was used within two hours after preparation. The resulting colour was measured between 2 and 120 minutes after mixing at 700 $\text{m}\mu$ in a Zeiss spectrophotometer (PMQ II) fitted with a microcuvette attachment. In each experiment inorganic phosphate standards were included to convert the extinction into moles of inorganic phosphate liberated/hour/kg dry wt. For this purpose 4 μ l of a solution of 50 mM KH_2PO_4 was added to 200 μ l and 400 μ l medium A. From this mixture three aliquots of 10 μ l were put into microtest tubes and 45 μ l TCA was added. To 30 μ l of this mixture 30 μ l of colour reagent was also added and the resulting colour was measured. Aliquots of 10 μ l medium A were treated in the same way and served as reagent blank. Because collection of quantities of Reissner's membrane and organ of Corti sufficient for homogenisation was not feasible, lyophilised samples of these structures (0.6-2.0 μ g) were weighed on a quartz-fiber balance (Lowry, 1953; Bonting and Mayron, 1961) and were directly transferred to 10 μ l of the incubation

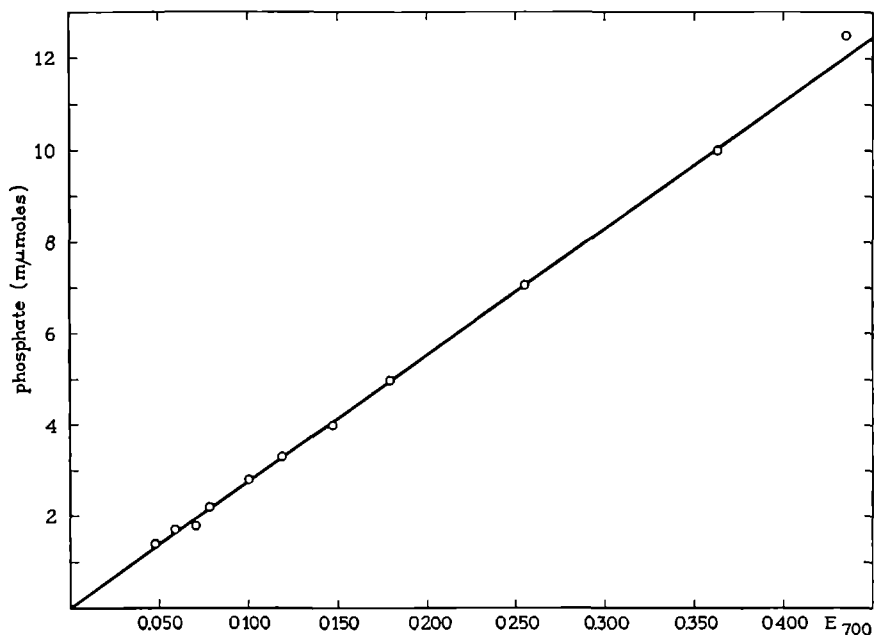


Fig 10 Calibration curve for the determination of inorganic phosphate.

media and treated as described before. In order to check the colorimetric method with our equipment we have carried out the whole procedure with medium A, to which were added various amounts of inorganic phosphate. The results shown in Fig. 10 demonstrate the required linearity.

3. Potential recording

a. *Surgical procedure*

The animals were anaesthetised with sodium pentobarbital (30 mg/kg body weight) administered intraperitoneally. During the experiment additional doses were given as required to maintain deep anaesthesia. The animals were tracheotomised and the trachea was cannulated to avoid airway obstruction. In addition this allowed us to make the animal respire gas mixtures of different composition. In order to reach the inner ear a skin incision was made in the posterior mandibular region.

The muscles and blood vessels were loosened from the mandibula and pushed away with a wound retractor. The mandibula was cut and the posterior portion removed. Thereafter the underlying muscles were retracted and the bulla tympanica became visible. From this point all further manipulations were carried out with the aid of a Zeiss operating microscope. With a dental drill part of the bony wall of the air filled bulla was carefully removed and the cochlea protruding into it was exposed (Fig. 9). In animals used for measuring the cochlear microphonic potentials, special care was taken to avoid damage of the tympanic membrane. Only animals with a middle ear cavity free of any sign of present or healed otitis were used in our experiments.

In order to perfuse part of the perilymphatic space two holes of approximately 100 μ were made in the bony wall of the scala vestibuli, one in the middle of the second turn and the other near the oval window (Fig. 9). The holes were drilled with a very fine motor driven drill, sharpened under a dissecting microscope and mounted on a micro-manipulator. In the animals used for measuring the cochlear microphonic potential a third hole was made between the previous two, for insertion of the wire electrode (Fig. 9). For measuring the endolymphatic potential a small window was made in the same region, just above the scala media, allowing us to place a glass capillary electrode into the endolymphatic space.

b. *Cochlear microphonic potential*

All potential recordings were carried out in a soundproof, electrically shielded room. The microphonic potential was recorded by means of a nichrome wire electrode (100 μ) placed in the hole made in the scala vestibuli in the middle of the first turn (Fig. 9) and an indifferent

electrode in the neck tissue. The recording electrode was insulated by dipping it into a resin solution, leaving small globules on the wire surface. The wire was cut just beyond one of those globules, thus preventing too deep penetration of the electrode into the scala vestibuli as well as outflow of the perfusion fluids.

The cochlear signals were amplified (1000 x) with a Tektronix differential amplifier and from there displayed on an oscilloscope screen and registered. The sound stimuli, consisting of pure tones in the frequency range of 500-8.000 cps, intensity 80 dB re 0.0002 dyn/cm², were produced by a condensor telephone driven by a beat frequency oscillator (Brüel and Kjoer, type 1014). This telephone was fixed in a small polyethylene container, which was fitted in an ear speculum. The speculum was securely fixed in the external auditory meatus. Sound pressure was measured with the aid of a probe microphone in the side wall of the container. The quality of the sound stimulus could be observed by visualizing the output of the probe microphone on the oscilloscope screen. The stimulating and recording circuit are schematically represented in fig. 11.

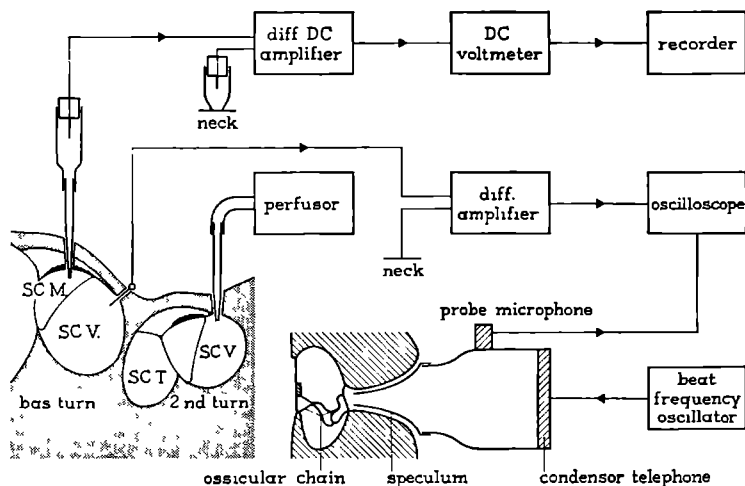


Fig. 11. Schematic representation of the equipment used for recording the cochlear potentials and for perfusing scala vestibuli. The upper part of this figure shows the recording circuit for the endolymphatic resting potential (ERP), the lower part the stimulating and recording circuit for the cochlear microphonic potential (CMP).

c. Endolymphatic resting potential

The endolymphatic resting potential was measured with glass pipette electrodes with tip diameters ranging from 5-15 μ and filled with isotonic KCl solution. The pipettes were fixed with wax into a wider glass

tube filled with the same solution and containing a chloride coated silver wire. The silver wire was connected to a DC voltage amplifier (S.M.I.: Rijswijk, The Netherlands, type MB-01) with an input impedance of $10^{12} \Omega$ and a grid current of 10^{-12} A . The output of the DC amplifier was delivered to a DC microvoltmeter (Philips, GM 6020) and read directly from the scale, or fed to a recorder for permanent recordings. A chloride coated silver wire, housed in a wide tube filled with Ringer and placed in the neck tissue served as the reference electrode.

Before penetrating the endolymphatic space the pipette electrode, mounted on a micromanipulator, was brought into contact with the ligamentum spirale, previously exposed by removing the bony capsule (Fig. 9). After compensating for the junction potential, the pipette was slowly advanced through the ligamentum spirale and stria vascularis until the positive endocochlear potential was observed, which remained constant upon further penetration. When measuring this potential, special care was taken to fix the head of the animal firmly to the operating table with a metal holder. Sudden movements would damage the cells of the stria vascularis in the neighbourhood of the electrode or even disrupt Reissner's membrane, resulting in a decrease of the potential.

At the end of the experiments the electrode was withdrawn and checked on the outer side of the ligamentum spirale for any change in the junction potential. The recording circuit is given in Fig. 11.

d. *Perfusion technique*

Perfusion of the perilymphatic space (scala vestibuli) was performed by inserting a small glass pipette (75-100 μ tip diameter) in the hole made in the second turn of the scala vestibuli (Figs. 9 and 11). Because this hole was tapered, the pipette could be inserted without any significant leakage of fluid. This glass pipette was connected by a small polyethylene tube to a mechanically driven all-glass syringe. The hole in the scala vestibuli near the oval window served as an outlet. The outflowing fluid was absorbed by small plugs of cotton wool in the bottom of the middle ear cavity. The excess fluid was removed by continuous suction, thus avoiding massive entry of the perfused substances into the circulation and at the same time keeping the fluid away from the windows. The cochlear microphonic potential appeared to be depressed when fluid accumulated at the windows, probably because sound transmission was decreased. Suction had to be done silently, since any noise close to the inner ear depressed the cochlear microphonic potential and also had some slight effect on the endolymphatic potential.

In most experiments the stability of the potentials was checked by perfusing the perilymphatic space for five minutes with a normal Ringer solution. In this way pressure effects, caused by obstruction of the outflow opening or injury of the epithelial wall of the endolymphatic space both resulting in changes in cochlear potentials, could be excluded.

The routinely used perfusion rate was 10 $\mu\text{l}/\text{min}$. The volume of the perfused space, as calculated from the cochlear dimensions reported by Fernández (1952), is approximately 3 μl , and the perilymph production varies between 1 and 3 $\mu\text{l}/\text{min}$. (Rauch, 1964). Therefore this perfusion rate seemed to be sufficient to replace the perilymph completely. All fluids used for perfusion were gassed with a mixture of 95% O_2 - 5% CO_2 and slightly warmed. The pH, measured immediately before the start of perfusion, was 7.3-7.4 and appeared to be raised to 7.8-7.9 after 60 min. perfusion. The composition of the standard perfusion fluid, which showed no effect on the cochlear potentials is given in Table 3. This fluid will be indicated as Ringer or normal Ringer's solution.

Table 3
COMPOSITION OF THE PERFUSING FLUID *

NaCl	124
KCl	3
CaCl ₂	1.3
MgCl ₂	0.8
NaHCO ₃	25
KH ₂ PO ₄	0.5
K ₂ HPO ₄	0.5

* All concentrations in mmole/l, pH adjusted to 7.4

OCCURRENCE AND DISTRIBUTION OF THE Na⁺-K⁺-ATPase ACTIVITY IN THE CHICKEN COCHLEA

1. Introduction

Although the available data are not as exact as those for mammalian cochlea, the endolymph of the avian cochlea appears to contain three times more potassium than sodium (Johnstone et al., 1963). This implies the existence of a cation gradient, which has to be maintained by an active cation transport mechanism. Moreover the cochlear potentials, which are present in mammals and have there been shown to be dependent on the cation composition of the cochlear fluids (Chapter III) have also been demonstrated in birds (Schmidt and Fernández, 1962, Schmidt, 1963). Therefore it seemed justified to look for the presence and localisation of the Na⁺-K⁺-ATPase activity, shown to be intimately involved in active cation transport. The dissection of the cochlear structures of one day old chickens is easier than in mammals, because the chicken skull consists mainly of cartilage and soft bone. Hence we started enzyme assays on these structures, rather than on the guinea pig cochlear structures.

2. Distribution of the Na⁺-K⁺-ATPase activity in the chicken cochlea

The cochlear structures of one day old chickens (Chapter I, 11) were dissected and lyophilised as reported in Chapter V, 2. The ATPase activities were determined in the various substrate media mentioned in Chapter V, 2. The Na⁺-K⁺-ATPase activity is represented by that part of the ATPase activity in the complete medium, that can be inhibited by the omission of Na⁺ or K⁺ or by the addition of ouabain. Enzyme assays on the whole cochlear sac revealed an absolute activity for Na⁺-K⁺-ATPase of 1.9 (SE: 0.4) and for Mg²⁺-ATPase of 4.2 (SE: 1.0) expressed in moles ATP hydrolysed per kg dry weight per hour at 37° C (Table 4).

In order to look for the distribution of the enzyme activity in the various structures of the cochlea, the tegmentum vasculosum was dissected and separately assayed for enzyme activity. The relative ATPase

Table 4

ATPase ACTIVITIES IN CHICKEN COCHLEAR TISSUES

RELATIVE ACTIVITIES IN TEGMENTUM VASCULOSUM HOMOGENATES		
<u>Medium</u>	<u>%</u>	
A (complete)	100	
B (no K ⁺)	66.6 ± 3.1	
C (no Na ⁺)	65.3 ± 4.2	
D (10 ⁻⁴ M ouabain)	66.3 ± 5.9	
E (no K ⁺ , 10 ⁻⁴ M ouabain)	59.3 ± 3.0	
Average for media B, C, D and E	64.4 ± 3.2	
ABSOLUTE ACTIVITIES		
	Na ⁺ -K ⁺ -ATPase	Mg ²⁺ -ATPase
Whole cochlear sac	1.9 ± 0.4	4.2 ± 1.0 (3)
Tegmentum vasculosum	5.0 ± 0.5	9.3 ± 1.0 (7)

Relative activities given with SE for 5 determinations. Absolute activities (means with SE and in parentheses number of determinations) are expressed in moles ATP hydrolysed per kg dry weight per hour at 37° C.

activities in the various substrate media and the absolute activities are given in Table 4. It appears from these data that the Na⁺-K⁺-ATPase activity is predominantly located in the tegmentum vasculosum, which has an activity of 5.0 moles/kg dry wt/hr.

The four inhibitory media caused approximately the same degree of inhibition, so the Na⁺-K⁺-ATPase activity represents 36% of the total ATPase activity.

3 Properties of ATPase activity

As demonstrated in various tissues there is an agreement in the effect of cations on the Na⁺-K⁺-ATPase activity and on the cation transport system (Chapter IV). Therefore the effects of Mg²⁺, Na⁺ and K⁺ on the Na⁺-K⁺-ATPase activity of the chicken tegmentum vasculosum were studied. In addition the inhibition by ouabain and the pH dependence were determined.

The effect of Mg²⁺ on the ATPase activities shown in Fig. 12, was investigated by varying the Mg²⁺ concentration from 0.6 mM in the media A and E containing 2 mM ATP. Medium E gave Mg²⁺-ATPase activity, while the difference between the activities in medium A and E gave Na⁺-K⁺-ATPase activity. A slight activity of both ATPase activities was present without added Mg²⁺. Na⁺-K⁺ activated ATPase was maximal at 1 mM Mg²⁺ and Mg²⁺ activated ATPase at 2 mM Mg²⁺.

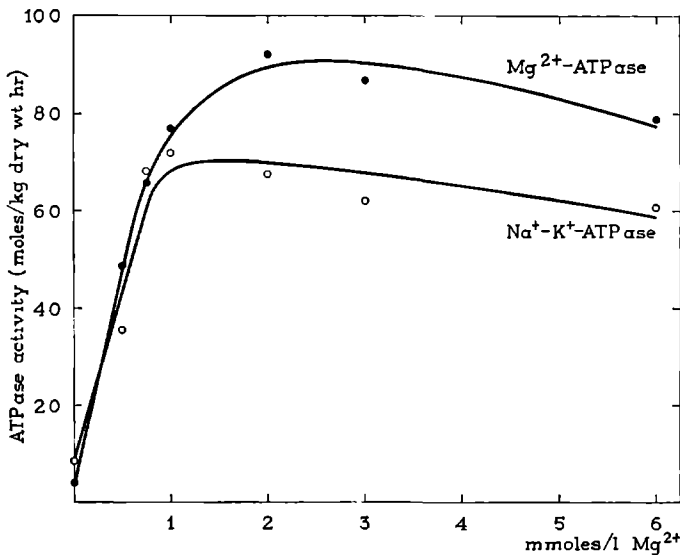


Fig 12 Effect of Mg^{2+} concentration on Na^+-K^+ ATPase activity (o—o) and Mg^{2+} -ATPase activity (●—●) in homogenates of chicken tegmentum vasculosum. Activities were measured in media A and E to which $MgCl_2$ (0.6 mM) was added. Medium E (no K^+ , 10^{-4} M ouabain) gave Mg^{2+} -ATPase activity, while the difference between activities in media A (complete) and E gave Na^+-K^+ -ATPase activity.

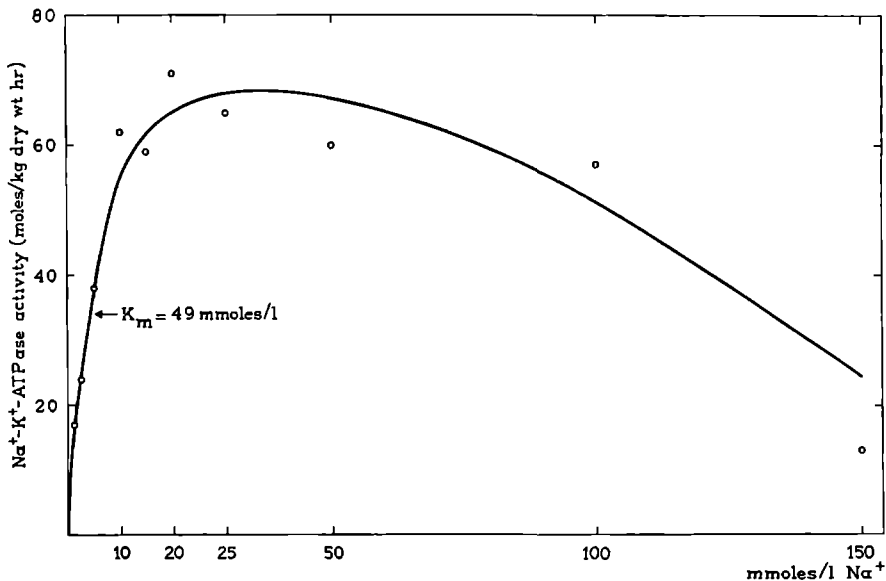


Fig 13 Effect of Na^+ concentration on Na^+-K^+ -ATPase activity in homogenates of chicken tegmentum vasculosum. Increasing amounts of $NaCl$ were added to medium C (no Na^+), while the K^+ concentration was kept at 5 mM.

Therefore the enzyme activities obtained from our routinely used medium, containing 1 mM Mg^{2+} represent the optimal Na^+K^+ -ATPase activity but only 84% of the optimal Mg^{2+} -ATPase activity.

The Na^+ -activation curve, in the presence of 5 mM K^+ for the Na^+K^+ -ATPase system is shown in Fig. 13. The activity was measured in medium C to which increasing amounts of NaCl were added (0-150 mM). Maximal activity was obtained at 25 mM Na^+ , while at higher Na^+ concentrations the enzyme activity decreased to about 30% of the maximal activity. Half maximal activation occurred at 4.9 mM Na^+ .

The activation of the Na^+K^+ -ATPase system by K^+ in the presence of 63 mM Na^+ is shown in Fig. 14. This curve was obtained by adding graded amounts of KCl to medium B (no K^+). Maximal activation of

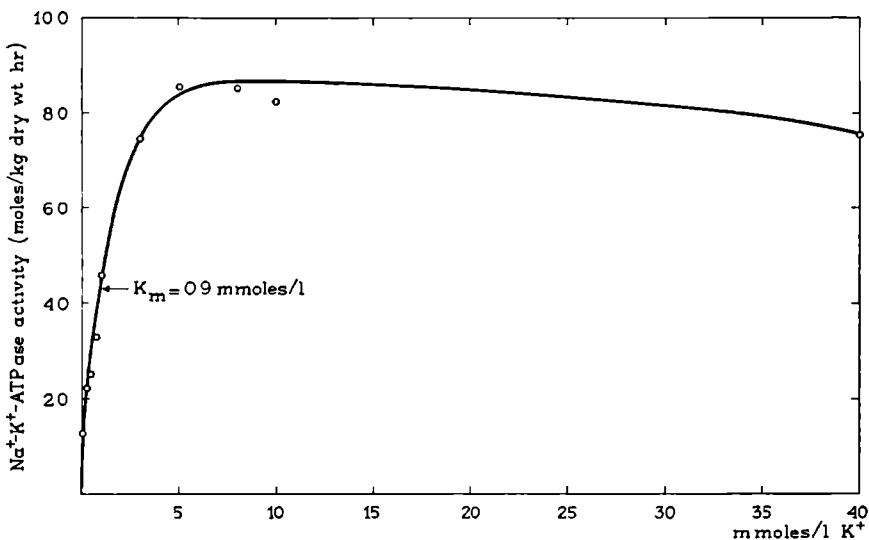


Fig 14 Effect of K^+ concentration on Na^+K^+ -ATPase activity in homogenates of chicken tegmentum vasculosum. Graded amounts of KCl were added to medium B (no K^+), keeping the Na^+ concentration at 63 mM. Medium E (no K^+ , 10^{-4} M ouabain) gave Mg^{2+} -ATPase activity.

the Na^+K^+ -ATPase was obtained with 5 mM K^+ , while higher concentrations were slightly inhibitory. Half maximal activation occurred at 0.9 mM K^+ . The slight activity without added KCl, is presumably due to partial activation of the enzyme by a small amount of tissue potassium, since the activity in medium B without K^+ appeared to be higher than in medium E (no K^+ , 10^{-4} M ouabain), which was considered to represent only Mg^{2+} -ATPase activity (Table 4).

The inhibitory effect of ouabain in various concentrations on the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity is given in Fig. 15. This inhibition was deter-

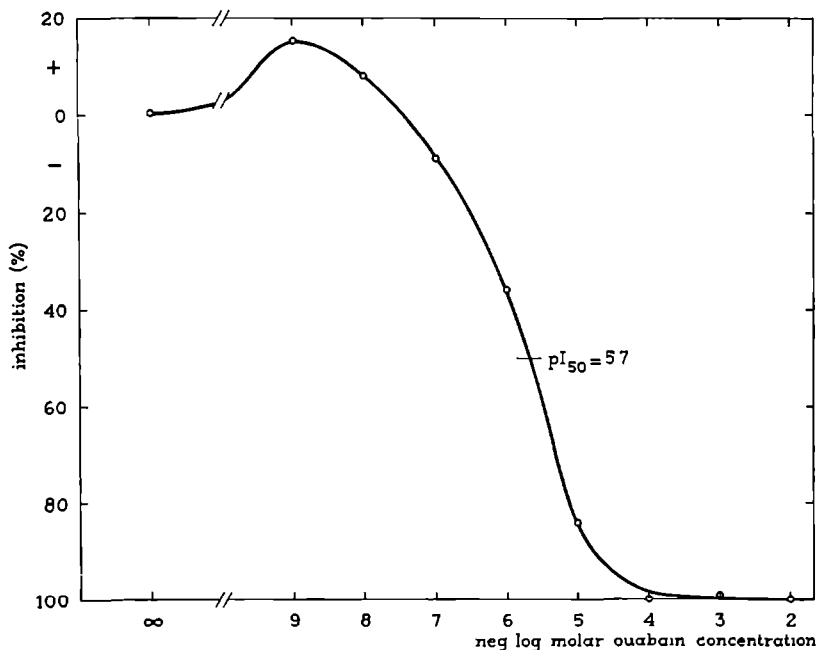


Fig. 15. Effect of ouabain on $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in homogenates of chicken tegmentum vasculosum. ATPase activity was determined in medium A containing various concentrations ouabain (10^{-9} - 10^{-2} M). The activity in medium E (no K^+ , 10^{-4} M ouabain) was taken to represent $\text{Mg}^{2+}\text{-ATPase}$ alone. The pI_{50} is the negative logarithm of the molar ouabain concentration causing 50% inhibition.

mined by measuring enzyme activity in medium A to which ouabain was added (10^{-2} - 10^{-9} M). The activity in medium E (no K^+ , 10^{-4} M ouabain) was taken to represent $\text{Mg}^{2+}\text{-ATPase}$ activity alone. The negative logarithm of the half maximal inhibition concentration was 5.7. Maximal inhibition was obtained at 10^{-4} M, the same concentration as routinely used in our substrate media. A slight stimulation of 15 (SE: 2.2)% occurred at very low ouabain concentrations (10^{-8} M).

The pH-activity curve was obtained by measuring ATPase activities in the media A and E, prepared with Tris-HCl buffers in the pH range from 6.5-9.5. The pH optimum for the $\text{Na}^+\text{-K}^+\text{-ATPase}$ was 7.3 and for $\text{Mg}^{2+}\text{-ATPase}$ 8.4 as shown in Fig. 16.

4. Discussion and conclusions

From the data presented in Table 4 and Figs. 12 to 16 it may be con-

Table 5

 PROPERTIES OF THE $\text{Na}^+\text{-K}^+\text{-ATPase}$ SYSTEM IN VARIOUS SPECIES
 AND TISSUES

Tissue	Species	$\text{Na}^+\text{-K}^+\text{-ATPase}$						$\text{Mg}^{2+}\text{-ATPase}$		Reference
		$\text{Mg}^{2+}\text{-ATP}$ opt	Na^+ K_m	K^+ K_m	pH opt	ouabain pI_{50}	activity MKH	activity MKH	pH opt	
Nerve	Crab	2	6-8	18	7.2	39	—	—	—	Skou (1957, 1960)
Intestine	Guinea pig	—	10	0.5	7.5-8.0	54	—	—	—	Taylor (1962)
Salt gland	Herring gull	15	12.5	15	7.2	63	2.7w	1.4w	8.7	Bonting et al (1964a)
Lens epithelium	Rabbit	—	—	0.4	7.3	59	0.15w	0.06w	8.4	Bonting (1965)
Rectal gland	Spiny dogfish	15	11.7	10	7.0	68	5.7d	2.9d	8.9	Bonting (1966)
E coli	E coli	1	—	4.3	7.7	± 4	0.24d	2.5d	8.7-8	Hafkenschied and Bonting (1968, 1969)
Liver	Rat	1	6	0.9	7.3	39	0.37	3.6d	8.7	Bakkeren and Bonting (1968)
Pancreas	Rabbit	15	10	0.8	7.2	54	0.23d	2.7d	8.8	Ridderstap and Bonting 1969a, b)
Kidney	Rat	0.5	8	0.7	7.4	39	6.8d	9.3d	8.7	v d Beek and Bonting, unpublished

Mg/ATP opt molar ratio of Mg^{2+} to ATP at which maximal activity occurs

Na^+K_m half-maximal activation concentration for Na^+ in meq/l at K^+ concentration of about 5 meq/l

K^+K_m half-maximal activation concentration for K^+ in meq/l at Na^+ concentration of about 60 meq/l

pH opt pH optimum

ouabain pI_{50} negative log molar concentration for 50% inhibition

activity MKH activity in moles/kg/hr, d=on dry wt basis, w=on wet wt basis

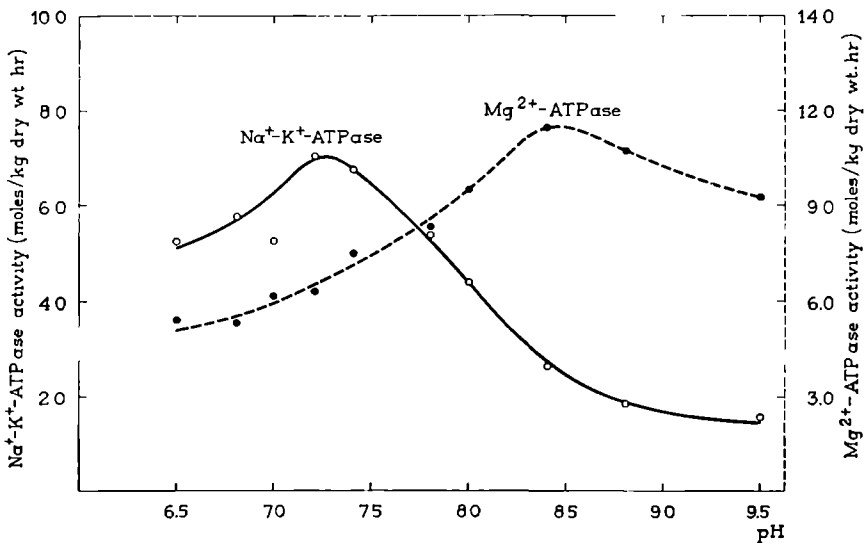


Fig. 16. Effect of pH on $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity $\circ\text{---}\circ$ and $\text{Mg}^{2+}\text{-ATPase}$ activity $\bullet\text{---}\bullet$ in homogenates of chicken tegmentum vasculosum. The pH dependence was measured in media A (complete) and E (no K^+ , 10^{-1} M ouabain), prepared with Tris-buffers in a pH range from 6.5-9.5. The $\text{Mg}^{2+}\text{-ATPase}$ activity was measured in medium E, while the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity was calculated from the difference between activities in media A and E.

cluded that the the inner ear structures of the one day chicken contain a $\text{Na}^+\text{-K}^+\text{-ATPase}$ system similar to that described by Skou (1957, 1960) in crab nerve, by Post et al. (1960) and by Dunham and Glynn (1961) in erythrocytes and by Bonting et al. (1962) and Bonting and Caravaggio (1963) in a great number of tissues in which cation transport against an electrochemical gradient occurs.

As demonstrated in Table 4 and Figs. 12 to 16, the enzyme system requires besides Mg^{2+} both Na^+ and K^+ for activation and is inhibited by ouabain. The concentrations of Na^+ ($K_m = 4.9$ mM) and K^+ ($K_m = 0.9$ mM) giving half maximal activation are in good agreement with those found in other tissues (Table 5). The same applies for the pH optimum of 7.3 of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity, while this optimum is very different from that found for the residual ouabain-insensitive $\text{Mg}^{2+}\text{-ATPase}$ activity (pH = 8.4). The half inhibition concentration of ouabain ($\text{pI}_{50} = 5.7$) is not very different from the value of 5.3, reported for the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in chicken kidney homogenates (Palmer and Nechay, 1964). The stimulatory effect of very low ouabain concentrations has also been demonstrated in the chicken kidney, both for the enzyme activity in homogenates and also for the tubular reabsorption of Na^+ and water in vivo (Palmer and Nechay,

1964). Similar biphasic effects have been reported for the production of cerebrospinal fluid (Oppelt and Palmer, 1966) and pancreatic juice (Ridderstap and Bonting, 1969 b).

As shown in Table 4, the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity appeared to be predominantly located in the tegmentum vasculosum. This structure representing about 30% of the dry weight of the whole cochlear sac, showed an activity of 5.0 moles/kg dry wt/hr, while the enzyme activity of the total structure is only 1.9 moles/kg dry wt/hr. This means that the rest of the cochlear structures has an activity of only 0.6 moles/kg dry wt/hr. The $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity of the tegmentum vasculosum is high in comparison with other tissues (Bonting et al., 1961 a). The actual value may be even higher, as may be concluded from the activities obtained in Figs. 12 to 16. In those experiments a more complete removal of the sticky endolymph from the tegmentum vasculosum could be obtained, resulting in an enhancement of the enzyme activity to about 7 moles/kg dry wt/hr. Such high $\text{Na}^+\text{-K}^+\text{-ATPase}$ activities have only been demonstrated in epithelia with a secretory function like the nasal gland of marine birds (Bonting et al., 1964 a), the rectal gland of elasmobranchs (Bonting, 1966), the choroid plexus (Vates et al., 1964), kidney (Bonting et al., 1961 a; Bonting et al., 1962) and in excitatory tissues like retina (Bonting et al., 1964 c), brain tissue (Bonting et al., 1962) and the electroplax of the electric eel (Bonting et al., 1961 b).

Bignami et al. (1966) showed that the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in developing chicken brain increased parallel with the electrical activity, reaching about 80% of the adult value at hatching. Since the inner ear of the one-day chicken has a nearly adult frequency response pattern (Vanzulli and Garcia Austt, 1963), it seems likely that the above enzyme activities closely approach those of the adult chicken cochlea.

It seems likely that the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system, present in high activity in the tegmentum vasculosum, functions in maintaining the high K^+ and low Na^+ concentration of the endolymph. An additional argument for this role of the tegmentum vasculosum can be derived from its fine structure. Jahnke et al. (1969) demonstrated a great structural similarity between the tegmentum vasculosum and epithelia involved in cation transport like kidney tubules (Pease, 1955), part of the labyrinthine epithelium (Dohman, 1965, Kimura, 1969) and the nasal gland of marine birds (Doyle, 1960). In addition, the assumption that phylogenetically the tegmentum vasculosum represents both stria vascularis and Reissner's membrane of the mammalian cochlea, is supported by the high $\text{Na}^+\text{-K}^+\text{-ATPase}$ activities in both tegmentum vasculosum and stria vascularis as will be shown in Chapter VII.

OCCURRENCE AND DISTRIBUTION OF $\text{Na}^+\text{-K}^+\text{-ATPase}$ ACTIVITY IN THE COCHLEAR STRUCTURES OF THE GUINEA PIG

1. Introduction

The presence and distribution of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the highly differentiated cochlear of the guinea pig was studied in order to investigate the involvement of this enzyme system in the maintenance of the high K^+ and low Na^+ concentrations in the endolymph. The anatomical position of the inner ear in this animal also permitted us to investigate the role of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the generation of the cochlear potentials by topical application of ouabain in the living animal. This appeared to be impossible in the chicken as will be discussed later on. The choice of the guinea pig was also guided by the considerations that the fine structure of the cochlear tissues, the cochlear fluid composition and the cochlear potentials have been studied in great detail (Chapters I, II and III), and that an ouabain-sensitive active transport of K^+ from peri- to endolymph has been demonstrated in this animal (Rauch, 1966).

2. Presence and properties of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system

Preliminary enzyme studies on lyophilised fragments of the total membranous structure of the guinea pig cochlear by incubation in the various substrate media, described in Chapter V, revealed a high ouabain-sensitive $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity of 0.8 (SE: 0.3) and a residual $\text{Mg}^{2+}\text{-ATPase}$ activity of 1.0 (SE: 0.4), both expressed in moles ATP hydrolysed per kg dry weight per hour. On further dissection it appeared that the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity was predominantly localised in the stria vascularis. The properties of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity have therefore been investigated in homogenates of the stria vascularis, collected from different turns. From an activity versus time curve it appeared that the hydrolysis of ATP at 37° C by $\text{Mg}^{2+}\text{-ATPase}$ and $\text{Na}^+\text{-K}^+\text{-ATPase}$ is linear up to 80 minutes (Fig. 17). Consequently, calculation of enzyme activity, based on an incubation time of 60 minu-

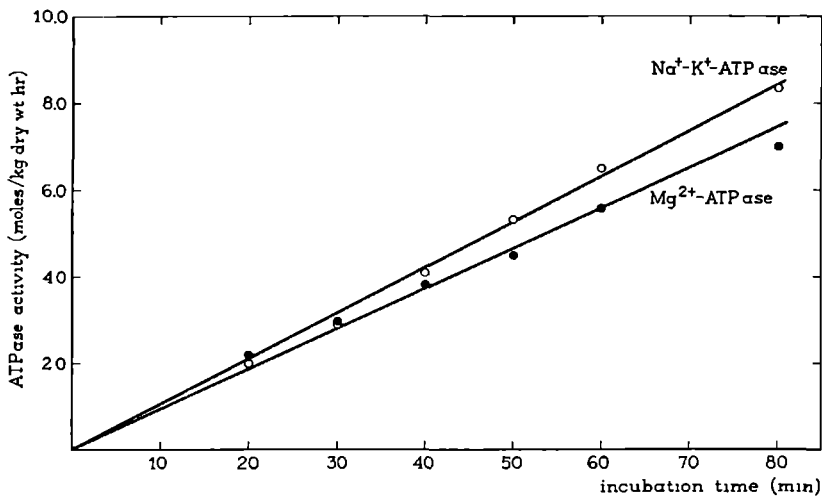


Fig. 17. ATPase activities in homogenates of guinea pig stria vascularis, determined in medium A (complete) and E (no K^+ , 10^{-1} M ouabain) after various incubation times. Na^+ - K^+ -ATPase activity \circ — \circ , Mg^{2+} -ATPase activity \bullet — \bullet

tes, will give a correct estimation of this activity.

The relative ATPase activities of the stria vascularis in the various substrate media are given in Table 6. The four inhibitory media caused

Table 6

RELATIVE ATPase ACTIVITIES OF GUINEA PIG STRIA VASCULARIS HOMOGENATES IN VARIOUS SUBSTRATE MEDIA

<i>Medium</i>	%
A (complete)	100
B (no K^+)	44.2 ± 1.9 (22)
C (no Na^+)	37.4 ± 2.2 (5)
D (10^{-1} M ouabain)	40.4 ± 1.9 (22)
E (no K^+ , 10^{-4} M ouabain)	40.5 ± 1.8 (22)
Average for media B, C, D and E	40.6 ± 1.4

Total ATPase activity in medium A is set at 100%.

Relative activities in media B, C, D, and E are given with standard errors and in parentheses number of determinations.

approximately the same degree of inhibition, indicating that the inhibited ATPase activity represents the Na^+ - K^+ -ATPase activity. Medium B gave slightly less inhibition than the other media. This is probably due to partial activation of the Na^+ - K^+ -ATPase by a small

amount of tissue K^+ present in the medium during incubation, since the K_m value for K^+ is only 0.9 (Fig. 20). Medium C showed slightly more inhibition than the other media. This effect may be due to a slight Na^+ dependence of the Mg^{2+} -ATPase, as has also been observed in other tissues (Bonting et al., 1964 a; Bakkeren and Bonting, 1968).

Fig. 18 shows the effect of increasing the Mg^{2+} concentration from 0-6 mM in the media A and E. Both Na^+ - K^+ -ATPase activity and

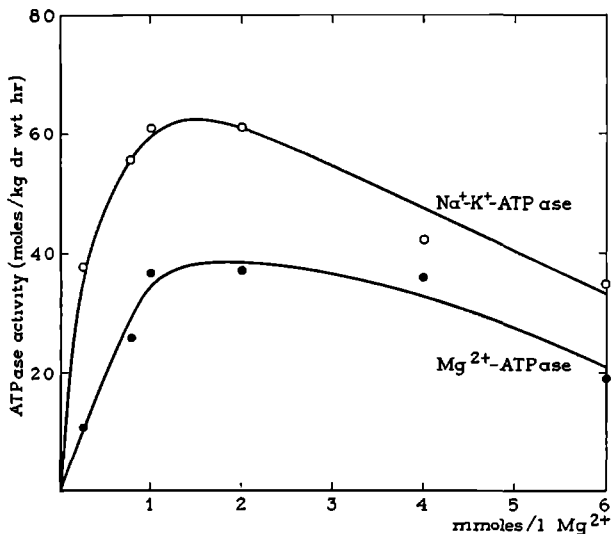


Fig. 18 Effect of Mg^{2+} concentration on Na^+ - K^+ -ATPase activity \circ — \circ and Mg^{2+} -ATPase activity \bullet — \bullet in homogenates of guinea pig stria vascularis. Activities were measured in media A and E to which was added $MgCl_2$ (0-6 mM). Medium E (no K^+ , 10^{-4} M ouabain) gave Mg^{2+} -ATPase activity, while the difference between the activities in media A (complete) and E gave Na^+ - K^+ -ATPase activity.

Mg^{2+} -ATPase activities were maximal at 1-2 mM Mg^{2+} present in the media at an ATP concentration of 2 mM and were depressed at higher Mg^{2+} concentrations. No ATPase activity could be demonstrated in the absence of Mg^{2+} .

The activation of the Na^+ - K^+ -ATPase by Na^+ in the presence of 5 mM K^+ is given in Fig. 19. The activity was measured in medium C to which increasing amounts of $NaCl$ were added (0-150 mM). Maximal activity was reached at 10 mM Na^+ , followed by a striking decrease between 10 and 20 mM; then the activity remained unchanged up to 150 mM. Half maximal activation occurred at 4.5 mM Na^+ /1. This value is somewhat lower than that for other tissues (Table 5).

Fig. 20 represents the K^+ activation curve of the Na^+ - K^+ -ATPase

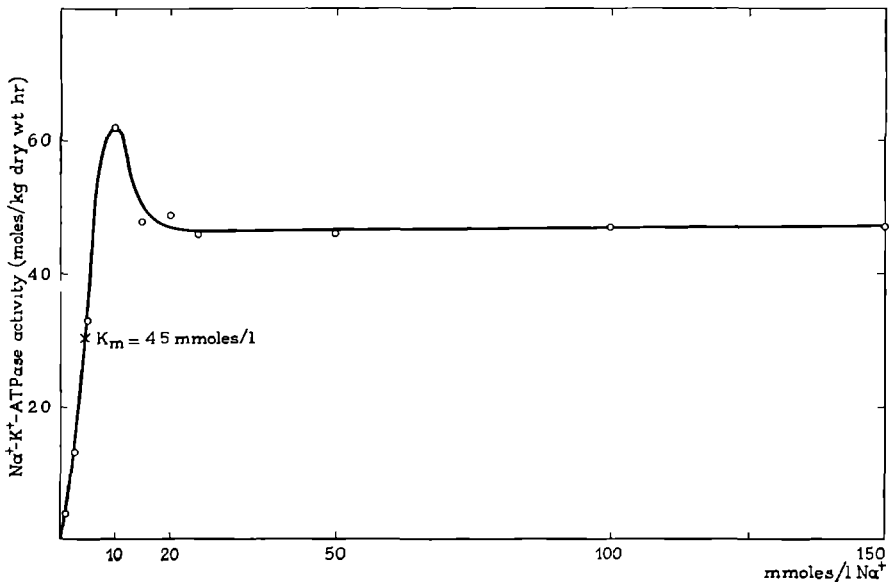


Fig 19 Effect of Na⁺ concentration on Na⁺-K⁺-ATPase activity in homogenates of guinea pig stria vascularis. Graded amounts of NaCl were added to medium C (no Na⁺), while the K⁺ concentration was kept at 5 mM. Activity in medium C without added NaCl gave Mg²⁺-ATPase activity.

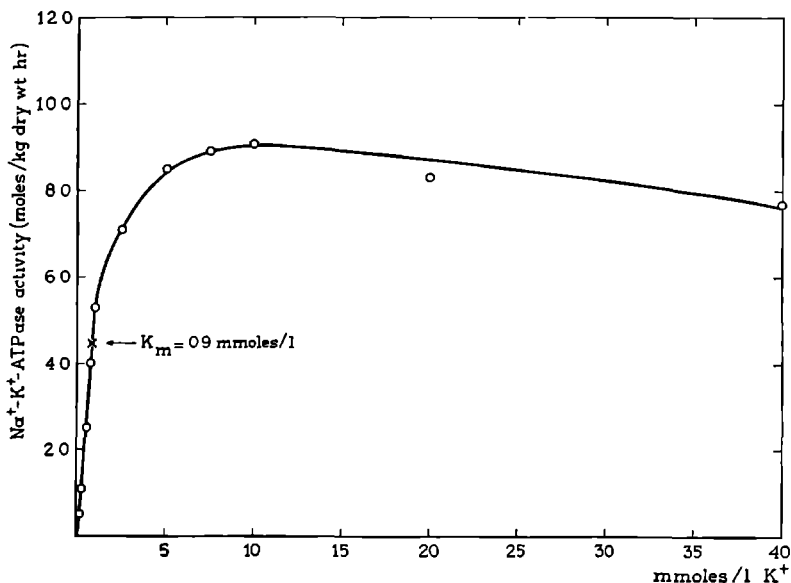


Fig 20 Effect of K⁺ concentration on Na⁺-K⁺-ATPase activity in homogenates of guinea pig stria vascularis. Graded amounts of KCl were added to medium B (no K⁺), while the Na⁺ concentration was kept at 63 mM. Activity in medium E (no K⁺, 10⁻⁴ M ouabain) gave Mg²⁺-ATPase activity.

activity in the presence of 63 mM Na⁺. This curve was obtained by measuring enzyme activity in medium B, containing various amounts of K⁺ (0-40 mM). Maximum activity was reached at 7.5 mM and half maximal activation at 0.9 mM K⁺/1. The activity at 5 mM K⁺, routinely used in the various substrate media, is only slightly different from the maximal activation obtained at 7.5 mM K⁺. The values for potassium activation are in good agreement with those reported for other tissues (Table 5).

The pH optimum for Na⁺-K⁺-ATPase is at 7.3 and for Mg²⁺-ATPase at 8.7 as shown in Fig. 21. These curves were obtained by

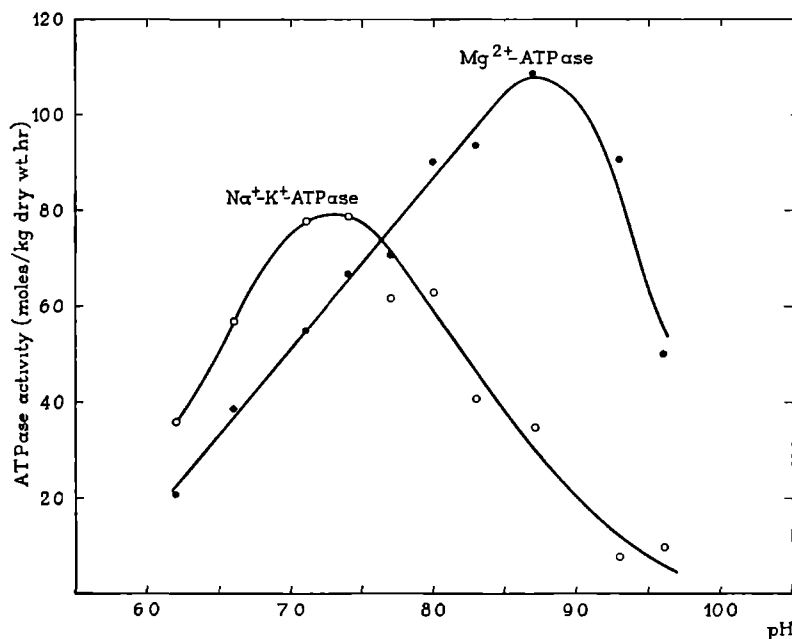


Fig 21 Effect of pH on Na⁺-K⁺-ATPase activity o—o and Mg²⁺-ATPase activity ●—● in homogenates of guinea pig stria vascularis. The pH dependence was measured in media A (complete) and E (no K⁺, 10⁻⁴ M ouabain), prepared with Tris buffers in a pH range from 6.2-9.6. The Mg²⁺-ATPase activity was measured in medium E, while the Na⁺-K⁺-ATPase activity was calculated from the difference between activities in media A and E.

measuring enzyme activities in the media A and E prepared with Tris-HCl buffers in a pH range from 7.4 to 9.6 and with Tris-histidine buffers in a pH range from 6.2-7.4. These optima are close to the values found in several other tissues (Table 5).

The inhibitory effect of ouabain on the Na⁺-K⁺-ATPase activity is presented in Fig. 22. This curve was determined by adding ouabain in

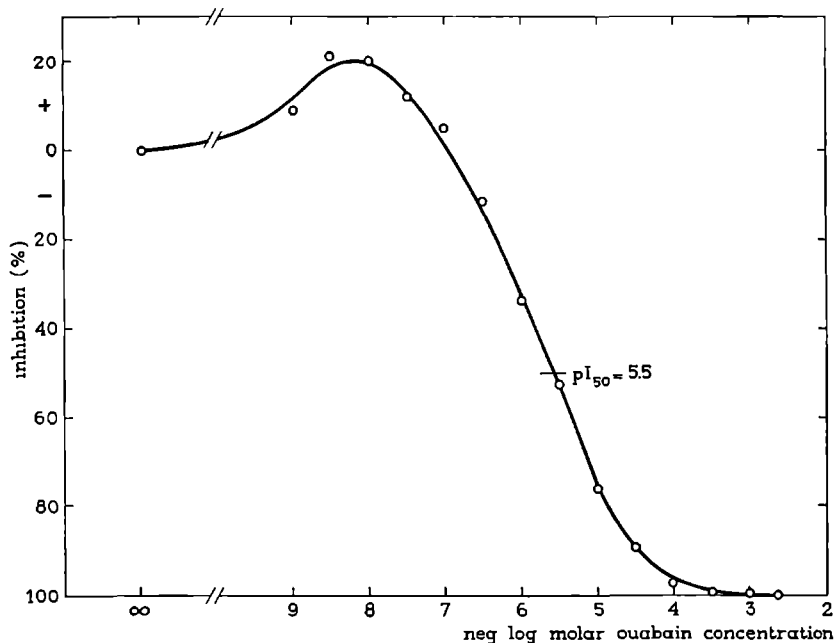


Fig. 22. Effect of ouabain on $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in homogenates of guinea pig stria vascularis. ATPase activity was determined in medium A containing various amounts of ouabain (10^{-9} - 10^{-2} M). The activity in medium E (no K^+ , 10^{-4} M ouabain) was taken to represent $\text{Mg}^{2+}\text{-ATPase}$ alone. The pI_{50} is the negative logarithm of the molar ouabain concentration causing 50% inhibition.

various concentrations (10^{-2} - 10^{-9} M) to medium A. The activity in medium E was considered to represent only $\text{Mg}^{2+}\text{-ATPase}$ activity. The negative logarithm of the half maximal inhibition concentration of ouabain was $\text{pI}_{50} = 5.5$, close to the value of 5.4 found for guinea pig intestine (Taylor, 1962). At very low ouabain concentrations (10^{-8} M) a stimulation of 20(SE: 8)% of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity was found, just as in the chicken tegmentum vasculosum (Chapter VI). This biphasic effect on $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity as well as on cation transport was also observed in the kidney (Palmer and Nechay, 1964), chorioid plexus (Oppelt and Palmer, 1966) and pancreas (Ridderstap and Bonting, 1969 b).

3. Distribution of ATPase activities of the various cochlear structures

Since preliminary studies demonstrated the presence of a very high $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in the stria vascularis, we made a systematic study of the distribution of this enzyme system in the cochlear structu-

res, especially in those which might be involved in cochlear cation transport. The dissection procedures used for isolating the various tissues have been described in Chapter V, 2.

The distribution of enzyme activities in the cochlear structures is presented in Table 7, and for the Na⁺-K⁺-ATPase activity alone in a

Table 7
ATPase ACTIVITIES IN HOMOGENATES OF THE VARIOUS COCHLEAR STRUCTURES OF THE GUINEA PIG

Structure	Na ⁺ -K ⁺ -ATPase		Mg ²⁺ -ATPase	Mean activity of turn No
	moles/kg dry wt/hr	% of total ATPase	moles/kg dry wt/hr	
Stria vascularis	8.0 ± 0.43 (16)	59 ± 1	5.6 ± 0.41	1, 2, 3
Lig spirale				
(A) behind stria vascularis	0.4 ± 0.09 (4)	50 ± 8	0.4 ± 0.09	2, 3
(B) comprising prominentia spiralis and sulcus externus	1.6 ± 0.39 (4)	55 ± 6	1.3 ± 0.43	2, 3
(C) bordering scala tympani	0.4 ± 0.07 (4)	40 ± 7	0.6 ± 0.07	2, 3
Reissner's membrane	0.4 ± 0.06 (12)	30 ± 4	0.9 ± 0.10	1, 2, 3, 4
Organ of Corti	0.5 ± 0.27 (3)	15 ± 6	2.8 ± 0.79	2, 3

Means with SE and in parentheses number of determinations

schematic cross section of the cochlea (Fig. 23). In the last column of Table 7 the numbers refer to the turns from which the various tissues were obtained. The Na⁺-K⁺-ATPase activity of the stria vascularis (8.0 moles/kg dry wt/hr) is extremely high and forms 59% of the total ATPase activity. The activity in this structure is about 12 times as high as the average activity in the ligamentum spirale, Reissner's membrane and organ of Corti, which range from 0.4-1.6 moles/kg dry wt/hr.

The ATPase activities of the ligamentum spirale appeared to be rather high, contrary to what could be expected from its connective tissue structure. In an attempt to localise this enzyme activity we divided the ligamentum spirale longitudinally into three parts and assayed them separately for ATPase activity. The results are represented in Table 7 and Fig. 23. The highest activity was found in part B of the ligamentum spirale, comprising the prominentia spiralis and sulcus externus. Histological sections of part B made after dissection showed that epithelial cells of the prominentia spiralis were absent.

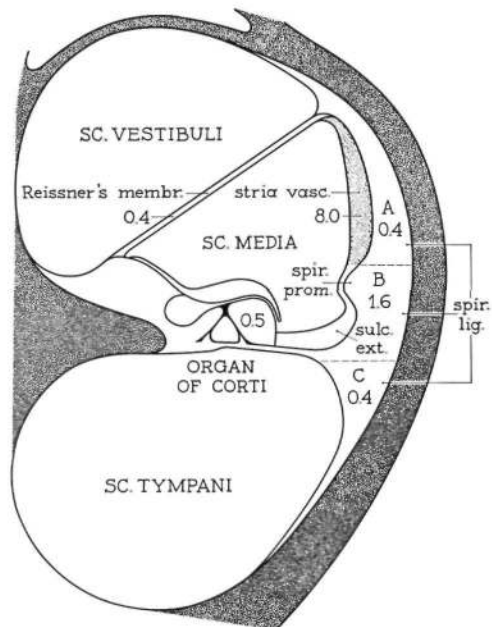


Fig. 23. Schematic cross section of the cochlea. $\text{Na}^+\text{-K}^+\text{-ATPase}$ activities in moles/kg dry wt/hr are indicated by the numbers in the various structures.

They were presumably removed together with the stria vascularis, which was dissected from the ligamentum spirale before dividing it into three parts. Thus, the enzyme activity of part B is probably mainly present in the sulcus externus epithelium and the stroma cells of the ligamentum spirale.

In order to ascertain possible differences in ATPase activities in the various cochlear turns, we have determined enzyme activities in the stria vascularis, ligamentum spirale and Reissner's membrane, obtained from individual turns. The bony connection between cochlea and middle ear wall was taken as the boundary line between the turns. The $\text{Na}^+\text{-K}^+\text{-ATPase}$ and $\text{Mg}^{2+}\text{-ATPase}$ activities of the stria vascularis in each turn are given in Table 8, and the total dry weight of this structure in each turn is included. There is a distinct decrease in both ATPase activities from base to apex. The ratio between $\text{Na}^+\text{-K}^+\text{-ATPase}$ and $\text{Mg}^{2+}\text{-ATPase}$ remained about the same. In these experiments the final tissue concentrations in the incubation media for third and fourth turns were considerably lower than for the first and second turns, ranging from 0.2 to 0.6 $\mu\text{g}/10 \mu\text{l}$. In order to exclude the possibility that this might cause the decrease in ATPase activities from first to fourth turn, we determined ATPase activities for tissue concen-

Table 8

ATPase ACTIVITIES IN HOMOGENATES OF GUINEA PIG STRIA VASCULARIS OF VARIOUS COCHLEAR TURNS

Turn no.	Na ⁺ -K ⁺ -ATPase		Mg ²⁺ -ATPase	Total dry weight of stria vasc. (μg)
	moles/kg dry wt/hr	% of total ATPase	moles/kg dry wt/hr	
1	6.4 ± 0.40 (5)	56 ± 3	5.0 ± 0.46	13
2	7.0 ± 0.88 (7)	62 ± 4	4.3 ± 0.35	12
3	4.6 ± 0.70 (5)	58 ± 6	3.4 ± 0.58	7
4	4.4 ± 0.55 (5)	59 ± 2	3.1 ± 0.37	3

Means with SE and in parentheses number of determinations. Basal turn is turn no. 1, apical turn is turn no. 4.

trations varying from 0.1 to 0.7 μg per 10 μl medium. The results are shown in Fig. 24. The linearity of these curves demonstrates that the decrease in enzyme activity from base to apex must be real. This effect

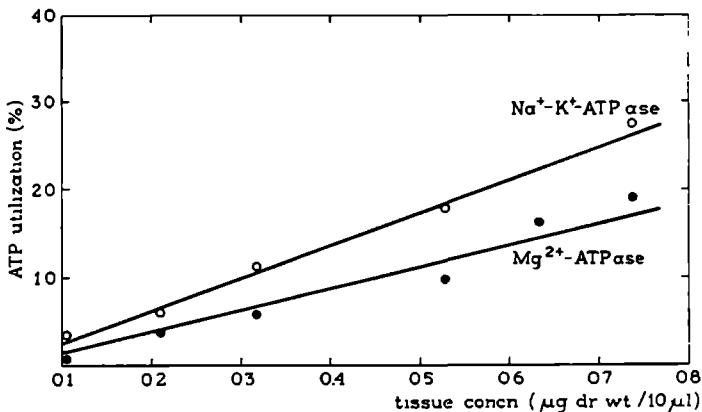


Fig 24. ATP utilisation in percent of total ATP present in the incubation medium by ATPase activities in homogenates of guinea pig stria vascularis at various tissue concentrations. Activities were determined in medium A (complete) and E (no K⁺, 10⁻¹ M ouabain). Na⁺-K⁺-ATPase activity o—o, Mg²⁺-ATPase activity ● ●.

is much more pronounced when the total dry weight of the stria vascularis of each turn, given in Table 8, is taken into account. This gives a Na⁺-K⁺-ATPase activity expressed in nanomoles/kg dry wt/hr of 83 for the first turn, of 84 for the second turn, of 32 for the third turn and of 13 for the fourth turn.

This decrease in enzyme activity could also be demonstrated for the ligamentum spirale of the various turns, as shown in Table 9. It was difficult to obtain ligamentum spirale of the first turn without adhering bone. Since bone fragments lead to high endogenous phosphate con-

Table 9

ATPase ACTIVITIES IN HOMOGENATES OF GUINEA PIG LIGAMENTUM SPIRALE OF VARIOUS COCHLEAR TURNS

Turn no.	Na ⁺ -K ⁺ -ATPase		Mg ²⁺ -ATPase
	moles/kg dry wt/hr	% of total ATPase	moles/kg dry wt/hr
2	15 ± 0.18 (4)	52 ± 2	14 ± 0.17
3	0.8 ± 0.09 (4)	38 ± 2	13 ± 0.13
4	0.7 ± 0.07 (3)	44 ± 1	0.9 ± 0.12

Means with SE and in parentheses number of determinations. No reliable data could be obtained for the basal turn (no 1)

centrations and incorrect weight determinations, no data on ATPase activities are given for this turn. In one experiment in which fragments of Reissner's membrane completely free of adhering dried lymph could be assayed separately for each turn, a similar decrease in enzyme activity was observed.

The absolute ATPase activity in Table 7 for the stria vascularis dissected in toto is 22% higher than the sum of the activities of the separate turns listed in Table 8. This is probably due to a slight loss of activity during the extra time needed for dissection of the turns separately, or to a slight loss of tissue. Since the fourth and the third turns were always isolated first, this loss should have primarily affected the results for the first and second turns. Hence this would tend to make the real difference between these turns and the third and fourth turns even greater. The enzyme activities for the organ of Corti and Reissner's membrane should not be influenced by the above-mentioned effect, because these structures were dissected from whole cochlea that were lyophilised immediately after decapitation as described in Chapter V, 2.

4. Discussion and conclusions

The data in Tables 6 and 7 and Figs. 18 to 23 demonstrate the presence of a Na⁺-K⁺-ATPase system in the guinea pig inner ear similar to that described by Skou (1957, 1960) in crab nerve, by Post et al.

(1960) in erythrocytes and by Bonting et al. (1961 a, 1962) in a great number of tissues of various animal species.

The properties of this $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in the guinea pig cochlea appeared to be quite similar to those found in the chicken inner ear (Chapter VI) and to those in many other tissues in which active cation transport occurs and in which a good agreement has been demonstrated between the properties of the enzyme system and the cation transport system (Post et al., 1960, Dunham and Glynn, 1961, Bonting and Caravaggio, 1963, Bonting and Canady, 1964). The stimulation by Na^+ , K^+ , and Mg^{2+} and the inhibition by ouabain of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity is much more pronounced than reported by Inuma (1967) for the stria vascularis and the ligamentum spirale together. These differences are probably due to less suitable methods used by the latter investigator e.g. the use of non-homogenised freshly dissected tissue samples without lyophilisation.

As demonstrated in Table 7 and Fig. 23 the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity of the stria vascularis, constituting 59% of the total ATPase activity is far higher than in any of the other cochlear structures. This was also true for enzymes involved in oxidative metabolism (Vosteen, 1961; Spoendlin and Balogh, 1963; Nakai and Hilding, 1968 a), and for the O_2 consumption of this structure, which is higher than that of kidney and brain tissue (Chou and Rodgers, 1962). The high oxidative metabolism of the stria vascularis presumably supplies the large amount of ATP required by the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system. Such a high $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity, as was also found in the tegmentum vasculosum of the chicken cochlea (Chapter VI), is typical for secretory tissues like kidney (Bonting et al., 1961 a; Bonting et al., 1962), rectal gland of elasmobranchs (Bonting, 1966), nasal gland of marine birds (Hokin, 1963; Bonting et al., 1964 a) and chorioid plexus (Vates et al., 1964) and for excitatory tissues like the electroplax of the electric eel (Bonting et al., 1961 a) and brain tissue (Bonting et al., 1962). An electron-microscopic study has shown the ATPase activity to be localised on the cell membranes of the marginal cells of the stria vascularis and in the basement membranes of the capillaries (Nakai and Hilding, 1966).

There is a clear decrease for both ATPase activities from first to fourth turn of the cochlea, not only in the stria vascularis but also in the ligamentum spirale and probably in Reissner's membrane, Tables 8 and 9. This phenomenon has also been demonstrated for lactic and malic dehydrogenase (Kurschner, 1968) and for the O_2 consumption of the stria vascularis (Meyer zum Gottesberge et al., 1965) and of the whole membranous structure (Mizukoshi and Daly, 1967). Furthermore the ERP has been shown to decrease from basal to apical turn (Mirashy et al., 1958 b). In contrast Inuma (1967) reported a more irregular distribution over the various turns for the $\text{Na}^+\text{-K}^+\text{-ATPase}$

activity of the stria vascularis-ligamentum spirale complex with a tendency to increase towards the apex, although for the phosphatase activities in the same complex an increase in activity from base to apex was found (Inuma and Daly, 1968). In our opinion Inuma's deviating results may be due to the use of non-homogenised and non-lyophilised fresh tissue causing difficulties in substrate penetration (Bonting and Rosenthal, 1960). Because the mass of this tissue complex is much larger for the basal than for the apical turns, the effect should be more pronounced in the basal turns.

The relatively high $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity in part B of the ligamentum spirale (Table 7, Fig 23) is noteworthy. In view of the absence of epithelial cells in the dissected part B of the prominentia spiralis the enzyme activity can only be localised in the sulcus externus and in the stroma cells behind the epithelium. This may be in accordance with the presence of a high activity of enzymes involved in oxidative metabolism in this region (Vosteen, 1961; Spöndlin and Balogh, 1963; Ishii and Nomura, 1968). Moreover these stroma cells show extensive infoldings of the cell membrane and many vacuoles (Spöndlin, 1967 a, von Ilberg et al., 1968) indicating that they are involved in fluid transport. A resorptive function of these cells has been demonstrated by von Ilberg et al. (1968). Therefore it may be concluded that this structure seems to be involved in cochlear fluid circulation.

The $\text{Na}^+\text{-K}^+\text{-ATPase}$ activities in the organ of Corti, Reissner's membrane and other parts of the ligamentum spirale are very low in comparison to the stria vascularis (Table 7, Fig. 23). The organ of Corti has a high $\text{Mg}^{2+}\text{-ATPase}$ activity. Nakai and Hilding (1967) demonstrated in this structure histochemically the presence of ATPase activity along the cell membranes. Moreover, an active metabolism has been demonstrated in this organ (Vosteen, 1961; Spöndlin and Balogh, 1963; Matschinsky and Thalmann, 1967) Since the $\text{Mg}^{2+}\text{-ATPase}$ system does not function in cation transport, the energy released by this enzyme in the organ of Corti must be used for other purposes than cation transport, probably in the perception and transduction of acoustic stimuli.

In view of its high $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity the stria vascularis would appear to be the site of the cation pump, which maintains the ionic concentration gradients between endolymph and perilymph or blood (Fig. 23). The assumption by Rauch (1964) that transport of K^+ from perilymph to endolymph would occur through Reissner's membrane must be considered highly unlikely in view of the low $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity of this structure. This activity is low on a weight basis, but even more so on an absolute basis. The ratio of the dry weights of stria vascularis and Reissner's membrane in the entire cochlea is approxi-

mately 3 l, hence the ratio of total $\text{Na}^+\text{-K}^+\text{-ATPase}$ activities in these structures is 68 l. This consideration leads us to the conclusion that the contribution of Reissner's membrane to the active cation transport can only be very minor. Additional arguments for this conclusion can be derived from the fact that the membrane is avascular and has a very high electrical resistance (Johnstone et al., 1966).

The transport of K^+ from the perilymph in the scala vestibuli to the endolymph, observed by Rauch (1964), could take place through the ligamentum spirale, through which perilymph can easily circulate (Tonndorf et al., 1962; von Ilberg, 1968 b). After arriving in the stria vascularis it would be actively secreted into the endolymph, while Na^+ would be actively removed from the endolymph by the same structure. An argument in favour of this supposition is the fact that in Rauch's experiments the ligamentum spirale and the stria vascularis showed a high isotope content, even when $^{42}\text{K}^+$ was injected immediately after interruption of the blood supply. This confirms that $^{42}\text{K}^+$ can easily diffuse from scala vestibuli to stria vascularis. In this connection it seems unlikely that Reissner's membrane should have nearly the same QO_2 as the stria vascularis (Chou, 1963), which is highly vascularised and has a far higher number of mitochondria in its cells than Reissner's membrane (Figs. 4 and 5). Recently Rauch (personal communication) has found a much lower O_2 consumption of this membrane than reported by Chou (1963). The $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the stria vascularis would therefore, appear to be the system which actively regulates the characteristic cation composition of the endolymph. The contribution of the other cochlear structures, judging from the $\text{Na}^+\text{-K}^+\text{-ATPase}$ distribution, can be considered to be negligible. In particular, such a role seems highly unlikely for Reissner's membrane. This conclusion would be in harmony with the hypothesis of Naftalin and Harrison (1958). These authors assume that there is only a passive leakage of Na^+ into and of K^+ out of the endolymph through Reissner's membrane. From the endolymph Na^+ would be removed actively, in exchange for K^+ , by a Na^+ pump in the stria vascularis, leading to the typical ionic composition of the endolymph.

In view of the strong evidence for a primary function of this enzyme system in cation transport in nerve (Baker, 1963, Baker and Connelly, 1966, Baker et al., 1969) and muscle (Corrie and Bonting, 1966) as well as in the formation of cerebrospinal fluid (Vates et al., 1964), aqueous humor (Bonting and Becker, 1964) and pancreatic juice (Ridderstap and Bonting, 1969 a, b), it would seem to be a reasonable assumption, that the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the stria vascularis may play the role of the cation pump in the endolymph formation.

THE EFFECT OF OUABAIN ON THE COCHLEAR POTENTIALS OF THE GUINEA PIG

1. Introduction

Both the cochlear endolymphatic resting potential (ERP) and the cochlear microphonic potential (CMP) have been shown to be dependent on the ionic composition of the cochlear fluids especially the high K^+ and low Na^+ concentration of the endolymph (Chapter III). These potentials are also dependent on oxidative metabolism, suggesting the involvement of oxidative metabolism in the maintenance of the typical intracellular-like ionic composition of the endolymph. In the previous chapters we reported the presence of a very high Na^+ - K^+ activated ATPase activity in the tissues of the inner ear. From these experiments it was concluded that this enzyme system is primarily localised in the stria vascularis of the guinea pig and in the analogous tegmentum vasculosum of the chicken, and that it represents most likely the cation pump responsible for the ionic composition of the endolymph.

The possible dependence of the cochlear potentials on the functioning of the Na^+ - K^+ -ATPase system was investigated by perfusing the perilymphatic space of the cochlea with Ringer solution containing ouabain, a specific inhibitor of the Na^+ - K^+ -ATPase system. The cochlear potentials were recorded simultaneously. The effect of ouabain was studied both on the ERP and on the CMP, which is elicited by acoustic stimulation.

An attempt to study these potentials in the chicken inner ear was unsuccessful. Although we succeeded after careful surgery to measure the CMP with a wire electrode placed on the round window, it was found to be impossible to perfuse the perilymphatic space without severely damaging the inner ear. This was concluded from the large decrease in the CMP immediately after opening the perilymphatic space. The guinea pig, which has frequently been used for electrophysiological studies on the inner ear, was more suitable for this purpose. In this animal the perilymphatic space could be opened and perfused with normal Ringer's solution without affecting the cochlear potentials, as previously reported by several authors (Tasaki et al., 1954; Honrubia et al., 1965; Konishi and Kelsey, 1968 a b)

It has been shown (Rauch, 1964) that $^{42}\text{K}^+$ introduced into the scala vestibuli is quickly transported to the endolymph, while $^{42}\text{K}^+$ injected into the scala tympani, reaches the endolymph much slower and to a much smaller extent. This indicates that the cation transport-system is more easily reached from the scala vestibuli than from the scala tympani. Therefore we chose the scala vestibuli for perfusion. Since the effect of ouabain on the CMP and the ERP had to be studied in separate experiments, we perfused in both cases the same region of the scala vestibuli, and placed the measuring electrodes in the same area (Fig. 11). Continuous removal of the outflowing perfusate was applied in order to avoid non-specific effects on heart frequency and respiration.

2. Cochlear Microphonic Potential

Because the CMP is generated in the basal turns, both by high and low sound frequencies, and in the apical turns only by low frequencies (Tasaki, and Fernández, 1952), we perfused the first and second turn in order to be able to record the effects of ouabain for a wide frequency range. Moreover, because of their smaller dimensions the structures of the apical turns are damaged more easily than those of the basal turns.

In preliminary experiments the CMP was recorded immediately before and at various times after perfusing the scala vestibuli for three minutes with about 150 μl Ringer's solution containing 10^{-2} M ouabain. Sound was administered by a loudspeaker situated in the experimental room, and driven by a beat frequency oscillator. Sound pressure was measured with a probe microphone placed near the head of the animal.

The results of a typical experiment are shown in Fig. 25. The CMP was strongly depressed, maximal decrease being obtained 20 minutes after ouabain application. Recovery to nearly normal values was found two hours after ouabain perfusion. This recovery, although variable in degree, was frequently found after short term perfusions. It is probably due to washing out of ouabain by the continuous outflow of perilymph through the holes in the bony capsule which was visually observed

Because of the limited frequency range, the considerable variation in intensity over the frequency range offered (ranging from 50 to 90 dB) and the poor quality of the sound produced by this system, all further experiments were carried out with the equipment described in Chapter V, 3. In this method sound was directly administered to the tympanic membrane. Thus a possible change in sound transmission by partial occlusion of the external auditory meatus during the experiment was avoided. Moreover, a constant sound pressure level of 80 dB for a frequency range from 500 to 16000 cps could be obtained. The sound stimulus consisted of continuous pure tones at 10 different frequencies in the frequency range of 500-8000 cps. In addition a continuous per-

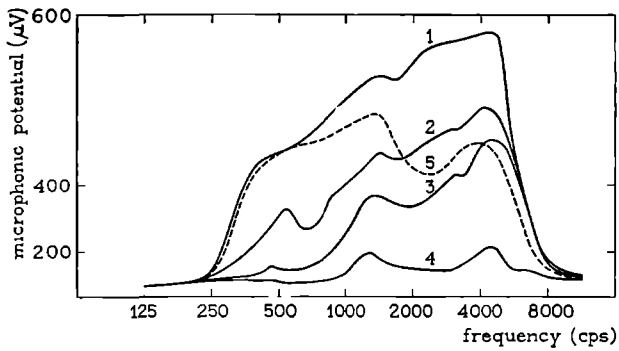


Fig. 25. Amplitude of the CMP at various frequencies before and after perfusing the scala vestibuli with about 150 μ l Ringer's solution, containing 10^{-2} M ouabain, for 3 minutes. 1. Before perfusion. 2. 3 minutes after perfusion. 3. 10 minutes after perfusion. 4. 20 minutes after perfusion. 5. 120 minutes after perfusion. The sound pressure varied from 50-90 dB in the frequency range of 500-6000cps.

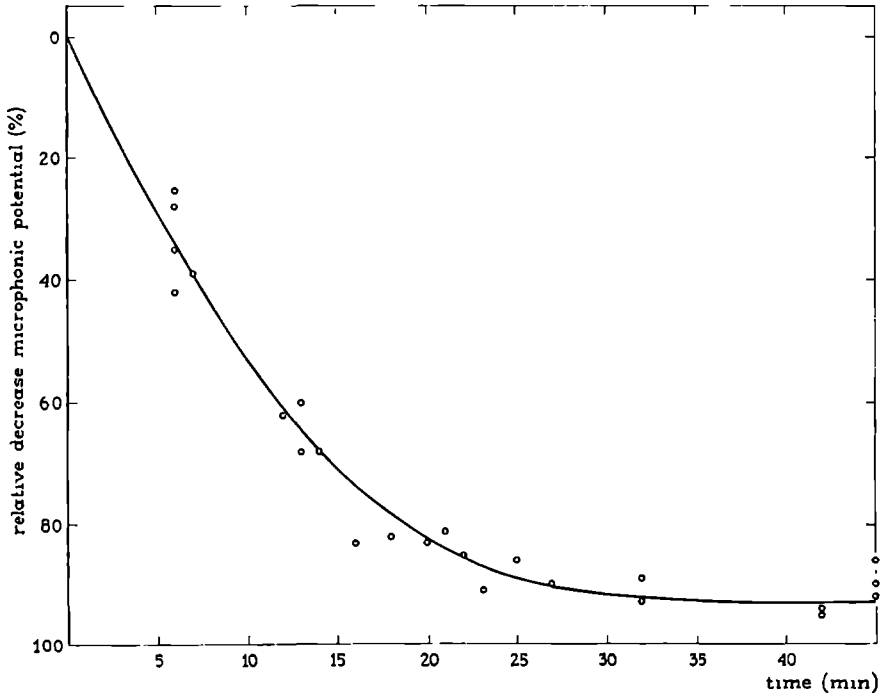


Fig. 26. Time course of relative decrease of the CMP, expressed in percentage of the original CMP value, upon continuous perfusion of the scala vestibuli with 10^{-3} M ouabain in Ringer's solution. Each point represents the mean value from five experiments, averaged for ten frequencies in the range of 500-8000 cps, intensity 80 dB.

fusing system (10 $\mu\text{l}/\text{min}$) was used, which permitted us to observe the effect of known and constant concentrations of ouabain.

The tendons of the middle ear muscles, musculus tensor tympani and musculus stapedius, attached to respectively malleus and stapes, were severed (Fig 9) in order to eliminate changes in sound transmission to the inner ear, which might occur from an effect of outflowing ouabain on muscular activity. Thus, a possible cause of unspecific changes in the CMP was avoided. Fig. 26 shows the time course for the decrease in the CMP, expressed in percentage of the original CMP value, upon perfusing the scala vestibuli for 45 min. with Ringer's solution containing 10^{-3} M ouabain. Each point represents the mean value from 5 experiments, averaged for ten frequencies in the range of 500-8000 cps. The amplitude of the CMP, measured directly after inserting

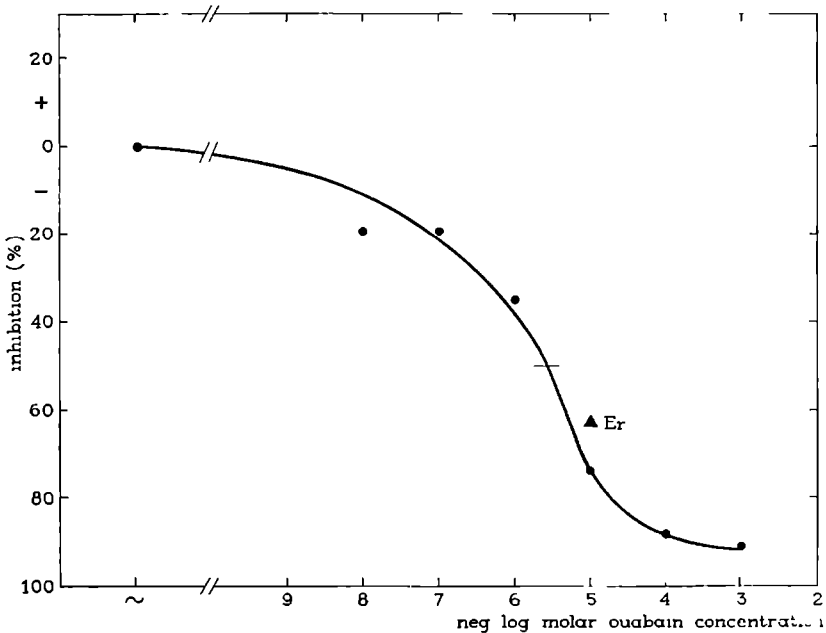


Fig 27 Effect on CMP of continuous perfusion of the scala vestibuli with Ringer's solution and Ringer containing various amounts of ouabain (10^{-8} - 10^{-3} M) ● — ● or 10^{-5} M erythropleine, ▲ Er, during 45 minutes. Each point represents the mean value from five experiments, averaged for ten frequencies in the range of 500-8000 cps, intensity 80 dB. The pI_{50} for ouabain is 5.5.

the pipet and before starting the perfusion, was set at 100%. Maximal inhibition (90%) was reached after 30 minutes, while no further decrease occurred upon continued perfusion. No significant difference was observed in the inhibition of the CMP at various frequencies.

After perfusion for about 45 minutes an inhibition was observed of 88(SE: 2.3)% at 500 cps. of 89(SE: 3.3)% at 4000 cps and of 88(SE: 2.7)% at 8000 cps. The persistence for several hours of about 10% of the original amplitude has also been shown after prolonged anoxemia (Davis, 1957).

Fig. 27 shows the effect on the CMP of perfusion for 45 minutes with normal Ringer's solution and Ringer's solution containing ouabain (10^{-3} M - 10^{-8} M) or erythropleine 10^{-5} M. Maximal inhibition was obtained at 10^{-3} M ouabain. The negative logarithm of the half maximal inhibition concentration (pI_{50}) was 5.5. The effect of 10^{-5} M erythropleine on the CMP was only slightly different from that of 10^{-5} M ouabain. This is in good agreement with the inhibitory effect of erythropleine on the $Na^+-K^+-ATPase$ activity (Bonting et al., 1964 b).

3. Endolymphatic Resting Potential

The endolymphatic resting potential was measured by introducing a glass capillary electrode, filled with 150 mM KCl, into the scala media as described in Chapter V, 3. The electrode resistance, measured in a solution approximating endolymph (147 mM KCl and 3 mM NaCl), was always less than 5 M Ω . In advancing the electrode slowly through ligamentum spirale and stria vascularis, often a variable positive potential was observed immediately before the high positive potential of the endolymphatic space appeared upon further penetration. Only when the electrode was advanced through the region of the prominentia spiralis a negative potential of about 50 mV was frequently recorded before the positive ERP was measured in the endolymphatic space. This negative potential represents most likely the depolarisation of the cells of the sulcus externus, which penetrate deeply into this region. This finding has previously been reported by Butler et al. (1962).

The values for the ERP in the various cochlear turns are summarised in Table 10. There is a clear decrease in potential going from basal to

Table 10
ENDOLYMPHATIC POTENTIAL (ERP) IN THE VARIOUS COCHLEAR
TURNS OF THE GUINEA PIG

Turn No.	ERP (mV)
1	75 \pm 2
2	70 \pm 2
3	55 \pm 3
4	49 \pm 3

Means with SE from four determinations.

apical turn, as has also been reported by Misrahy et al. (1958 b). We have demonstrated a similar decrease for the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity (Tables 8 and 9). Meyer zum Gottesberge et al. (1965) and Mizukoshi and Daly (1967) observed a similar distribution for the O_2 consumption and Kurschner (1968) for the distribution of lactic and malic dehydrogenase activity.

Fig. 28 shows the effect on the ERP of perfusing the scala vestibuli for 50 minutes with normal Ringer's solution and Ringer to which ouabain was added (10^{-3} - 10^{-8} M). No change was found upon perfusion with normal Ringer's solution. Ouabain caused a decrease of the

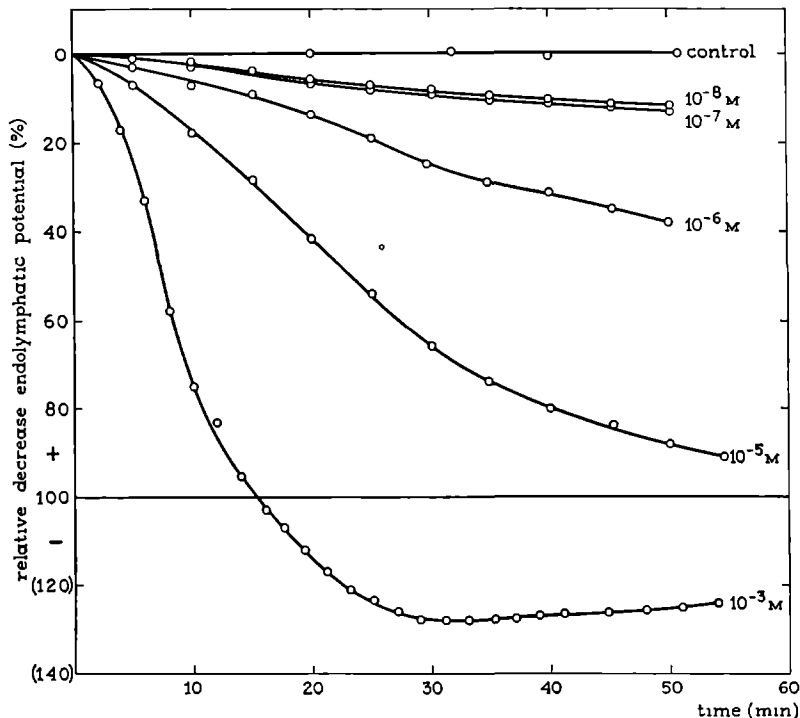


Fig 28 Effect on ERP of continuous perfusion of the scala vestibuli for 50 minutes with Ringer's solution containing various amounts of ouabain (10^{-8} - 10^{-3} M). The point at which ERP was zero was set at 100% inhibition. Inhibition larger than 100% indicates reversal of the potential. Each curve represents the mean value of four experiments.

ERP, which was dependent on its concentration. A remarkable effect was found with 10^{-3} M ouabain. The potential not only decreased to zero but became even negative, reaching a maximally negative value of about -25 mV after 30 minutes. Upon continued perfusion this negative

potential tended to increase slowly to zero. A similar behaviour of the ERP has been demonstrated during anoxemia, which will be discussed in Chapter IX. If the animal was made anoxic, when the potential upon perfusion with 10^{-3} M ouabain was decreased to a stable negative value, no further decrease could be observed. Therefore it may be concluded that the ERP had completely disappeared.

Fig. 29 shows the time course for the decrease of the ERP expressed in percentage of the original ERP value, upon perfusion of the scala vestibuli with 10^{-3} M ouabain in Ringer's solution. The corresponding

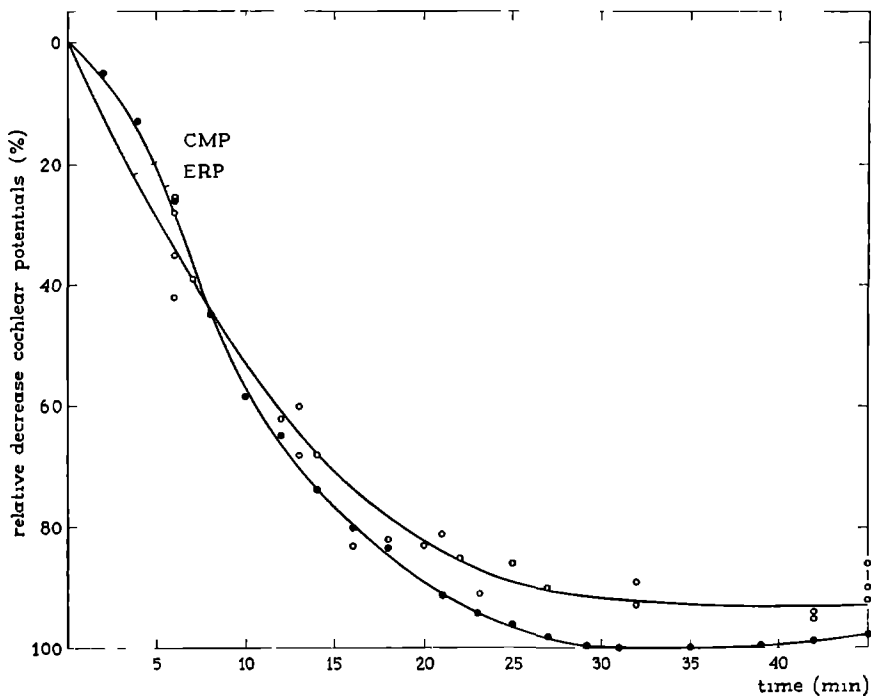


Fig. 29 Time course of relative decrease of ERP upon continuous perfusion of the scala vestibuli with 10^{-3} M ouabain in Ringer's solution. ● — ● The point at which the ERP was maximally negative (Fig. 28) was set at 100% inhibition. The corresponding curve for the CMP is also presented. ○ — ○

curve for the CMP is presented for purposes of comparison. The maximally negative value of the ERP after ouabain administration was taken as 100% inhibition. There is a striking resemblance in the time courses for both potentials, except that the CMP was only inhibited to 90% maximally.

Fig. 30 shows the inhibitory effect of various concentrations of

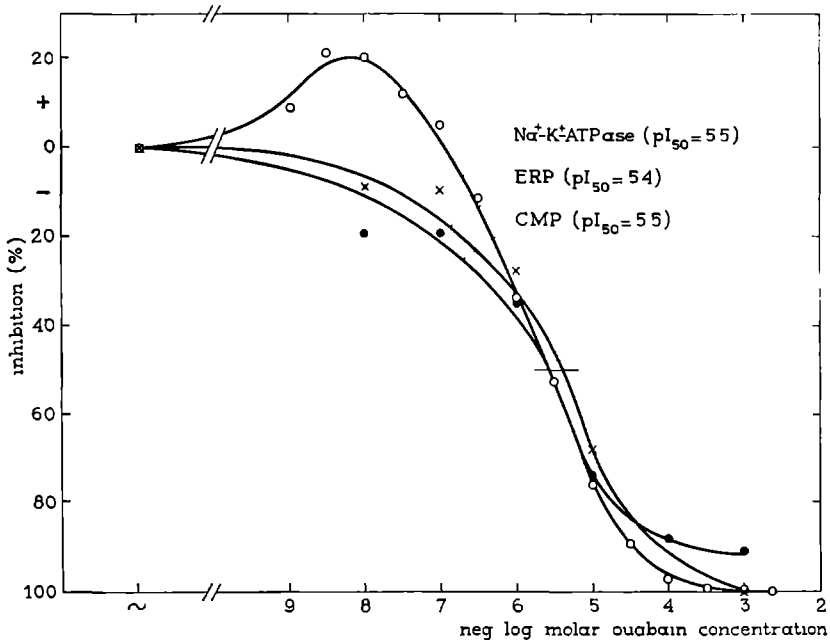


Fig 30 Effect on ERP of continuous perfusion of the scala vestibuli with Ringer's solution containing various amounts of ouabain (10^{-8} - 10^{-3} M) for 45 minutes x — x The inhibitory effects of ouabain on the CMP ● — ● and on the Na^+ K^+ -ATPase activity in homogenates of the stria vascularis o — o are presented for purposes of comparison

ouabain (10^{-8} - 10^{-3} M) on the ERP upon perfusing the scala vestibuli for 45 minutes. The curves for the inhibition of the CMP and the Na^+ - K^+ -ATPase activity in the stria vascularis homogenates are also presented in this figure. There is a striking similarity between these curves which have a nearly identical pI_{50} of 5.5 and 5.4. The stimulatory effect on the Na^+ - K^+ -ATPase activity at low concentrations of ouabain (10^{-8} M) was not observed for the cochlear potentials.

4. Discussion and conclusions

In view of the high Na^+ - K^+ -ATPase activity in the cochlear structures (Chapter VI and VII) and the effect of ouabain on both the ERP and the CMP (Figs 25 to 30), it may be concluded that the cochlear potentials depend on the functioning of the Na^+ - K^+ -ATPase system. Since this enzyme system is very predominantly located in the stria vascularis (Chapter VII), it seems very likely that this structure is responsible for the cochlear potentials.

Ouabain, perfused through the scala vestibuli, may easily reach the

stria vascularis by free diffusion through the extracellular spaces of the spiral ligament (Ilberg, 1968 b) or else it may enter the vessels running through the spiral ligament towards the stria vascularis. There it will block the $\text{Na}^+\text{-K}^+\text{-ATPase}$ cation pump system, which is located in the membrane of the marginal cells of the stria vascularis (Fig. 4) and which is responsible for maintaining the high K^+ and the low Na^+ concentration in the endolymph, necessary for cochlear function.

The striking similarity in the effects of ouabain on the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system of the stria vascularis and on the cochlear potentials with a pI_{50} of 5.5 for the inhibition of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system and the CMP and a pI_{50} of 5.4 for the ERP constitutes strong evidence for a direct relationship between the activity of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system and the occurrence of the cochlear potentials. The only discrepancy is the presence of a stimulatory effect of low concentrations of ouabain on the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity, which could not be demonstrated for the potentials. The fact that these low ouabain concentrations were slightly inhibitory, rather than stimulating, to the CMP and the ERP (Fig. 30) may be due to an adsorption of ouabain to the membranes of the cells of the stria vascularis during the 45-minute perfusion of the scala vestibuli. Such an adsorption has also been shown by Bonting and Becker (1964) for the cells of the ciliary epithelium, responsible for aqueous humor secretion.

An additional argument for a primary role of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the origin of at least the ERP may be derived from the decrease of the magnitude of the ERP from the basal to the apical turn (Table 10), since a similar gradient was found for the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity (Tables 8 and 9).

There is a remarkable similarity between the effects of ouabain, anoxemia (Konishi et al., 1961) and of cyanide (Konishi and Kelsey, 1968 c) on the cochlear potentials. This warrants the conclusion that the energy supply for the activity of the cation pump is supplied by the oxidative metabolism. This conclusion is supported by the findings of Matchinsky and Thalmann (1967), who demonstrated a very substantial decrease in the ATP content of the stria vascularis within one minute after anoxemia. The ERP has been shown to decrease at about the same rate, as will be discussed in Chapter IX.

On the basis of this evidence it seems reasonable to conclude that the ERP depends on the activity of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the stria vascularis. However this does not yet allow us to decide whether this potential arises directly from the activity of the cation pump, or secondarily from the ionic gradients maintained by the pump between endolymph and perilymph or plasma. This problem will be discussed in Chapter IX.

The similarity of the effects of ouabain on the CMP and the ERP

indicates a close relationship between the two potentials. Such a relationship is also indicated by the effects of anoxemia and of inhibitors of the oxidative metabolism. However the CMP, in contrast to the ERP, has also been shown to be dependent on the negative potential of the hair cells. Butler (1965) has demonstrated that the decrease of the CMP during anoxemia is similar to the decrease in the potential gradient across the lamina reticularis (Fig. 6), which is the sum of the ERP and the negative potential of the hair cells. This would be in agreement with the theory of Davis (1961), who proposes that the CMP originates from a change in the resistance of the lamina reticularis due to the bending of the cilia of the hair cells, which would regulate the amount and the direction of the current through this layer. This current is thought to be driven by both the ERP and the hair cell potential. The effects of ouabain perfusion on the ERP and the CMP, which we have observed, could be interpreted to support this hypothesis. Although we did not measure the potential of the hair cells, we cannot exclude the possibility that ouabain perfused through the scala vestibuli might through the ligamentum spirale reach the scala tympani and thence the organ of Corti. Ouabain could then inhibit the cation pump located in these cells and thus slowly decrease the potential of the hair cells, thus contributing to the decrease in the CMP. Although conclusive evidence for this theory has never been supplied, the persistence of about 10% of the CMP after continued anoxemia and also after ouabain perfusion might be the result of the presence of a potential gradient across the lamina reticularis, originating from the difference between the negative potential of the hair cells (Chapter III) and the negative potential in the scala media, arising upon anoxemia or ouabain perfusion (Chapter IX).

THE NATURE OF THE ENDOLYMPHATIC RESTING POTENTIAL AND COMPOSITION OF THE PERILYMPH

1. Introduction

In the previous chapter we showed the dependence of the endolymphatic resting potential (ERP) on the functioning of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the stria vascularis. This does, however, not yet explain the exact nature of this potential. To this problem we direct ourselves in this chapter. In most cells the potential difference across the cell membrane is a diffusion potential, which is due to the ionic concentration gradients between cytoplasm and extracellular fluid and which is determined by the relative permeabilities of the cell membrane for the ions. Such a potential is only indirectly dependent on the activity of the cation pump, namely in as far as the latter is responsible for maintaining the ion gradients responsible for the potential. The diffusion potential is given by the constant field equation formulated by Hodgkin and Katz (1949).

$$E = - \frac{RT}{F} \ln \frac{P_K (K^+_i) + P_{Na} (Na^+_i) + P_{Cl} (Cl^-_i)}{P_K (K^+_o) + P_{Na} (Na^+_o) + P_{Cl} (Cl^-_i)}$$

In this equation R represents the gassconstant, T the absolute temperature, F the Faraday constant and o and i refer to the concentrations of K^+ , Na^+ and Cl^- outside and inside the cell respectively. P_K , P_{Na} and P_{Cl} are the permeability constants of these ions. The validity of this relation has been shown for many types of biological membranes as nerve, muscle and lens. In the case of resting nerve and muscle and of the electroplax of the electric eel, the potential is mainly determined by the K^+ gradient, because the membrane is much more permeable to K^+ than to Na^+ and Cl^- . Because (K^+_i) is much larger than (K^+_o) , the inner side of the cell membrane is negative (about 80 mV) with respect to the outer side.

Since the ERP is positive with regard to plasma and perilymph, and (K^+) in the endolymph is high, this suggests that it cannot be a K^+ diffusion potential. This led Johnstone (1967) to assume that it could

be a Na^+ diffusion potential. Neglecting the Cl^- contribution, he could then arrive at a potential of +82 mV, if he set the permeability ratio $P_{\text{Na}}/P_{\text{K}}$ for the cochlear membranes at 50 as it is for the squid axon membrane during the rising phase of the action potential (Hodgkin and Katz, 1949, Nastuk and Hodgkin, 1950) and if he used the known cation concentrations in endolymph (K^+ , 150 mM; Na^+ , 3 mM) and plasma (Na^+ , 140 mM; K^+ , 5 mM). He further assumed that the fast decrease of the ERP in anoxemia, when no change in the Na^+/K^+ ratio has yet occurred (Johnstone, 1965), would be due to a change in the permeability ratio $P_{\text{Na}}/P_{\text{K}}$ to 0.2, which would result in the observed negative potential of -30 mV. If this hypothesis should be correct the Na^+/K^+ -ATPase system would only act to maintain the intracellular-like cation concentration of the endolymph, while the ERP would merely be the expression of the Na^+ gradient between endolymph and plasma or perilymph.

In an attempt to determine whether the ERP could be an ionic diffusion potential, we have observed the effects on the ERP of perfusing the perilymphatic space with solutions having various ionic contents (Table 11). In further experiments the effect on the ERP of rupturing Reissner's membrane was studied. In addition we have looked for a

Table 11
COMPOSITION OF PERFUSION FLUIDS *

	K^+ Ringer	Li^+ Ringer	SO_4^{2-} Ringer	Choline Ringer	Sucrose Ringer
Li^+	—	149	—	—	—
Na^+	—	—	149	—	—
K^+	153.5	4.5	4.5	4.5	4.5
Ca^{2+}	1.3	1.3	1.3	1.3	1.3
Mg^{2+}	0.8	0.8	0.8	0.8	0.8
Cl^-	131.2	153.2	2.6	166.2	4.2
HCO_3^-	25	3	3	3	3
HPO_4^{2-}	0.5	0.5	0.5	0.5	0.5
H_2PO_4^-	0.5	0.5	0.5	0.5	0.5
Choline	—	—	—	162	—
SO_4^{2-}	—	—	75.3	—	—
Sucrose	—	—	75	—	297

* All concentrations in mmole/l pH adjusted to 7.4

possible pH dependence of the ERP, because Rauch (1964) has suggested that a slight difference in pH between perilymph and endo-

lymph, as reported by Misrahy et al. (1958 c) could give an important contribution to the ERP. We have observed the ERP during perfusion of the perilymphatic space with Ringer's solutions of different pH and also while making the animal respire a gasmixture of 20% CO₂ and 80% O₂ so as to create a respiratory acidosis.

The nature of the negative potential occurring after anoxemia or upon applying ouabain to the perilymph was studied in a similar way, namely by changing the composition of the perfusing fluid.

The inner ear structures contain a very high activity of carboanhydrase (Erulkar and Maren, 1961) which is primarily localised in the stria vascularis (Eggemann and Bruchmuller, 1968) Intravenous administration of acetazolamide, a strong inhibitor of carboanhydrase, caused a large decrease in the K⁺ concentration of the endolymph (Erulkar and Maren, 1961) and a disappearance of the vacuoles in the marginal cells of the stria vascularis (Johnson and Spoendlin, 1966). Since these experiments suggested a role of the carboanhydrase system in cochlear cation transport, we also studied the effect on the ERP of perfusing acetazolamide through the scala vestibuli

2. Effect of electrolyte changes in the perilymph

The effect on the ERP of removing Na⁺ from the perilymph was studied by continuously perfusing the scala vestibuli with Ringer's solutions in which Na⁺ was replaced by Li⁺ or sucrose With sucrose Ringer, perfused at the rate of 10 µl/min, there was an increase of the ERP by about 5 mV within one minute after the start of the perfusion. The potential remained at this level for several minutes, and then it started to decrease. At a perfusion rate of 15 µl/min the increase appeared to be slightly higher, but it was nearly absent at 6 µl/min. Perfusion with normal Ringer at the same rates failed to show any change in the ERP Therefore, we ascribe the temporary increase in the ERP with the sucrose Ringer to an increase in hydrostatic pressure, caused by the high viscosity of this solution. A similar increase of the ERP after application of positive pressure to the scala vestibuli has been reported by Tasaki et al. (1954) In subsequent experiments we therefore used a perfusion rate of 6 µl/min for sucrose Ringer

The effects on the ERP during perfusion of the scala vestibuli with sucrose Ringer and Li⁺ Ringer is shown in Fig 31. Each curve represents mean values of six determinations. There is a remarkable similarity between the effects of Li⁺ and sucrose Ringer. The potential is decreased by 18% with Li⁺ Ringer and by 15% with sucrose Ringer after continuous perfusion for 22 minutes. The decrease of the ERP immediately ceased and could be reversed to a variable degree upon perfusing with normal Ringer. The recovery process varied to a certain extent in different experiments Sometimes the potential increased to

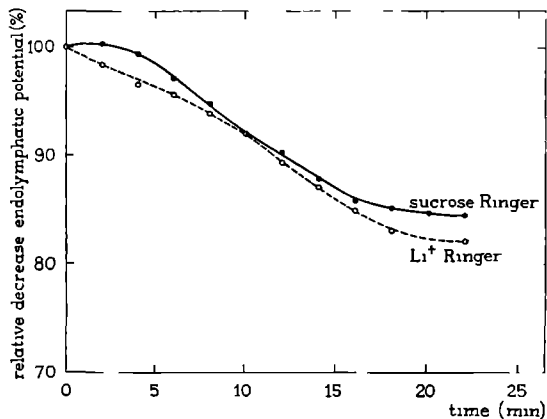


Fig 31 Effect on ERP of continuous perfusion of the scala vestibuli with sucrose Ringer ●—● and Li⁺ Ringer o—o Each curve represents the mean value of six experiments

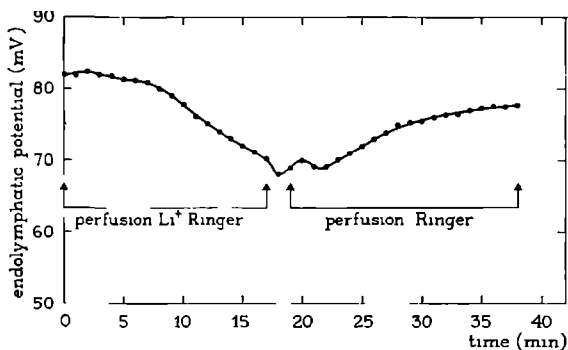


Fig 32 Effect on ERP of continuous perfusion of the scala vestibuli with Li⁺ Ringer and subsequent perfusion of normal Ringer's solution.

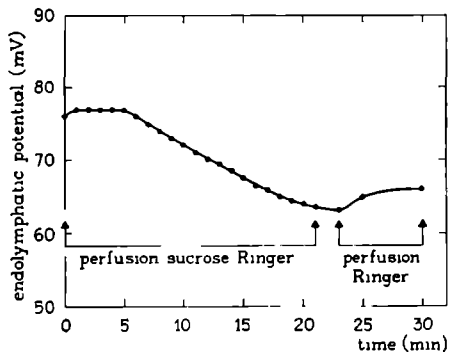


Fig 33 Effect on ERP of continuous perfusion of the scala vestibuli with sucrose Ringer and subsequent perfusion of normal Ringer's solution

nearly its original value (Fig. 32), but often only a slow and partial recovery was obtained (Figs. 33 and 34). The latter phenomenon was more frequently observed after longterm perfusion, suggesting that it might be due to damage of the cochlear membranes. However, this seems unlikely, because perfusion with normal Ringer for 45 minutes failed to show any effect on the ERP or CMP (Figs. 27 and 28). A more likely explanation is that perfusion with sucrose or Li^- would affect cell metabolism or membrane permeability somewhat. Additional evidence that the absence of Na^- in the perfusate is responsible for the decrease of the ERP is presented in Fig. 34. The decrease of the ERP, initiated by perfusion with sucrose Ringer, continued during

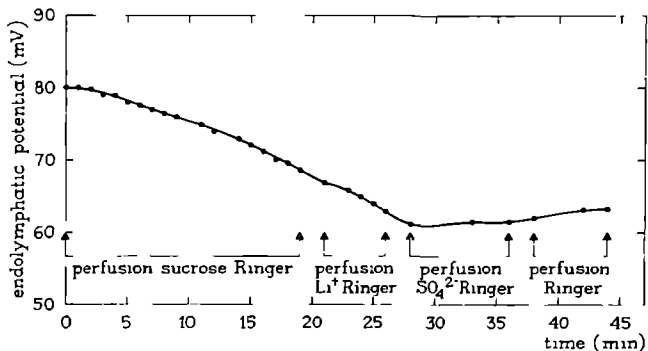


Fig. 34. Effect on ERP of continuous perfusion of the scala vestibuli with sucrose Ringer, followed by perfusion with Li^+ Ringer, SO_4^{2-} -Ringer and normal Ringer's solution.

subsequent perfusion with Li^+ Ringer, but it ceased when Na_2SO_4 Ringer was perfused and a slight recovery was obtained with normal Ringer.

In contrast to the slight effect of perfusing Na^+ free Ringer a dramatic effect on the ERP was obtained by perfusing the scala vestibuli with Ringer in which Na^+ was replaced by 150 mM K^+ . The results of three experiments are shown in Fig. 35. Within one minute after starting perfusion the potential rose sharply sometimes more than 20 mV and thereafter a fast decrease was observed to about 50% of the original value. When perfusion was stopped, the potential recovered slowly and only partially. This recovery, which is presumably due to exchange of the excess K^+ by Na^+ from the continuous flow of perilymph, could not be enhanced by perfusion with normal Ringer.

The contribution of Cl^- to the ERP was investigated by perfusing the scala vestibuli with Ringer solution in which Cl^- was replaced by SO_4^{2-} to which most membranes are much less permeable than to Cl^- .

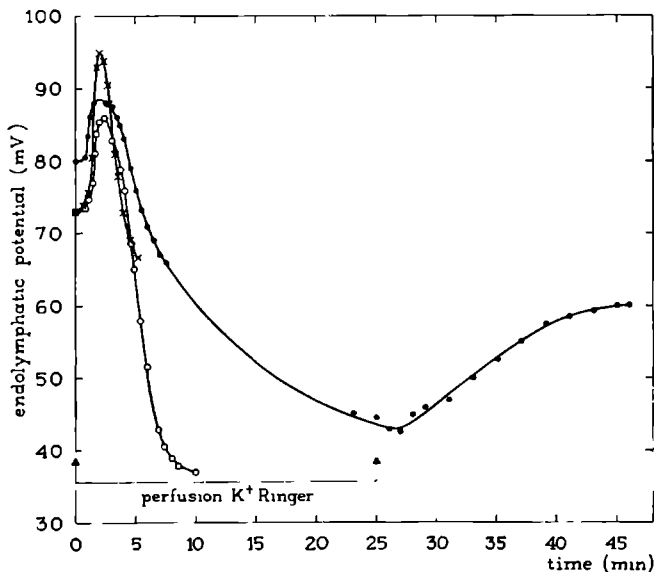


Fig. 35. Effect on ERP during and after continuous perfusion of the scala vestibuli with K^+ Ringer in three experiments.

The results, shown in Fig. 36, indicate that omission of Cl^- affected the potential to a very slight extent only, and the effect could be restored with normal Ringer. This is in agreement with the fact that sucrose Ringer, which has a very low Cl^- concentration, had the same effect on the ERP as Li^+ Ringer, which has a normal Cl^- content. The nearly equal Cl^- contents of endolymph and perilymph also indicate that Cl^- could not contribute significantly to the ERP (Table 1).

In an attempt to investigate the role of Reissner's membrane in the occurrence of the ERP, we ruptured this membrane. After careful removal of the bony wall of the scala vestibuli of the exposed part of the

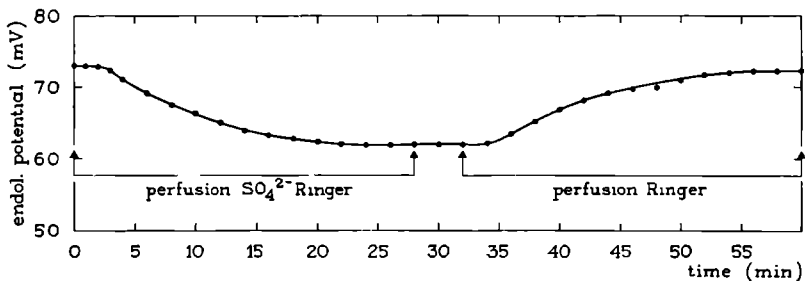


Fig. 36. Effect on ERP of continuous perfusion of the scala vestibuli with SO_4^{2-} Ringer and subsequent perfusion with normal Ringer.

basal turn, Reissner's membrane was ruptured thoroughly with a fine glass needle. The potential decreased rapidly to about + 15 mV and thereafter diminished slowly, becoming zero within about 15 minutes. However when immediately after rupturing Reissner's membrane the cochlear fluid was removed by suction, the potential increased by about 10 mV and then slowly decreased to about + 15 mV upon ceasing suction. The decrease to + 15 mV coincided with the rise of the fluid level in the cochlear space caused by perilymph production. This effect could be repeatedly found even when the potential was nearly zero, about fifteen minutes after the destruction of Reissner's membrane.

3. Effect of pH on the potential

The dependence of the ERP on a difference in pH between endolymph and perilymph was investigated by perfusing the scala vestibuli with Ringer's solutions in which the pH was brought to 6.0 and 8.0 by addition of lactic acid and of NaOH, respectively. Continuous perfusion with these solutions for 25 minutes failed to show any effect on the potential.

In a second group of experiments a general respiratory acidosis was established by making the animal respire a gas mixture consisting of 80% O₂ and 20% CO₂. The pH of the blood was decreased from 7.3 to 6.8 while the pCO₂ appeared to be more than 180 mm. Under these conditions Misrahy et al. (1958 b) measured a decrease in endolymph pH of the same magnitude. The ERP decreased to about 93% of its original value within ten minutes, and could not be returned to normal

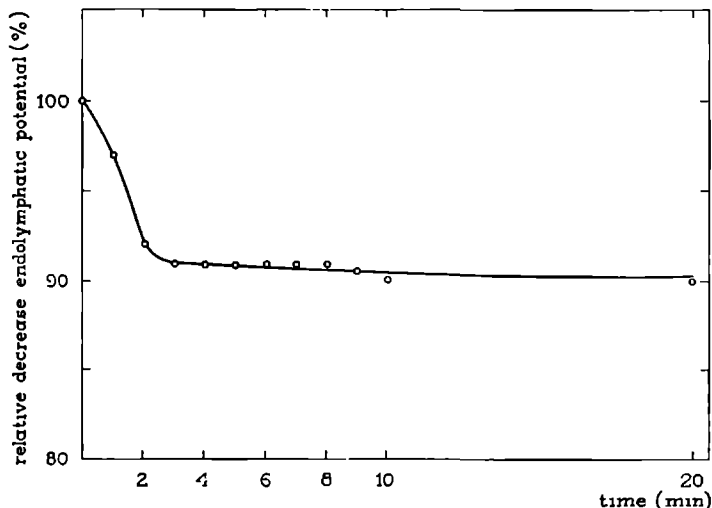


Fig 37 Effect on ERP of continuous perfusion of the scala vestibuli with 10^{-4} M acetazolamide in Ringer's solution (Mean value of four experiments)

by perfusing the scala vestibuli with normal Ringer of pH 7.4. When subsequently the animal was made to respire a gas mixture consisting of 95% O₂ and 5% CO₂, the potential returned within a few minutes to its original value.

The effect of acetazolamide, an inhibitor of carboanhydrase was studied by perfusing the scala vestibuli with Ringer's solution containing 10⁻³ M acetazolamide. The results of four experiments are shown in Fig. 37. The potential decreased rapidly to 91 (SE: 1.3) % of its original value within three minutes after starting perfusion. During the subsequent 17 minutes only a very slight decrease to 90% was observed. The resemblance between the effects of CO₂ inhalation and of acetazolamide perfusion may be due to an increase of intracellular CO₂ resulting in cellular acidosis in both cases (Slegers and Moons, 1968). The resulting decrease in intracellular pH could inhibit the cation pump, since the Na⁺-K⁺-ATPase system in the stria vascularis has a pH optimum of 7.3 and its activity falls considerably at lower pH (Fig. 21).

4. Experimental effects on the nature of the negative potential

The effect on the ERP and the venous O₂ pressure of anoxemia, induced by making the animal respire 20% CO₂ - 80% N₂, is demonstrated in Figs 38 and 39. The ERP decreased within four minutes to -12 mV, and thereafter slowly returned to zero in about two and a half hours. In our experiments the value of the negative potential varied between 10 and 35 mV. Assuming tentatively that the negative potential represents a K⁺ diffusion potential, we perfused the scala vestibuli of the anoxic animal with K⁺ free Ringer. Surprisingly this led to reversal of the potential. This has previously been reported by Honrubia et al. (1965), who suggested that it was due to removal of accumulated products of anaerobic metabolism. However, when we added 20 mM lactic acid to the perfusing fluid, the effect still occurred (Fig. 40), while upon stopping perfusion the potential returned again to its original negative value. This suggests that the potential reversal upon perfusion of the scala vestibuli is due to oxygen present in the perfusing fluid. To test this hypothesis 10 mM NaCN was added to the perfusing fluid. In this case the potential remained negative except for a slight transient increase (Fig. 41), which proves that the effect is indeed due to oxygen present in the perfusing fluid. In further experiments the negative potential was, therefore, obtained by addition of 10⁻³ M ouabain to the perfusing fluid. This has the added advantage that the general metabolism of the animal remains unaffected, because the very small amount of ouabain administered to the perfusate leaves almost entirely with the outflowing mixture of perilymph and perfusate.

The scala vestibuli was perfused with Ringer's solution containing

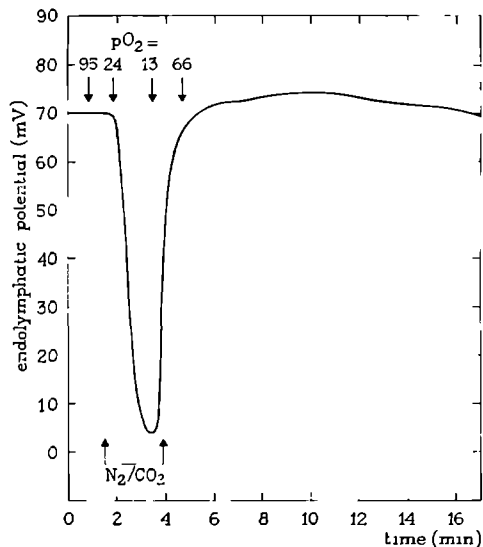


Fig. 38. The ERP and venous pO_2 before, during and after temporary anoxemia from inhalation of a gas mixture consisting of 20% CO_2 and 80% N_2 .

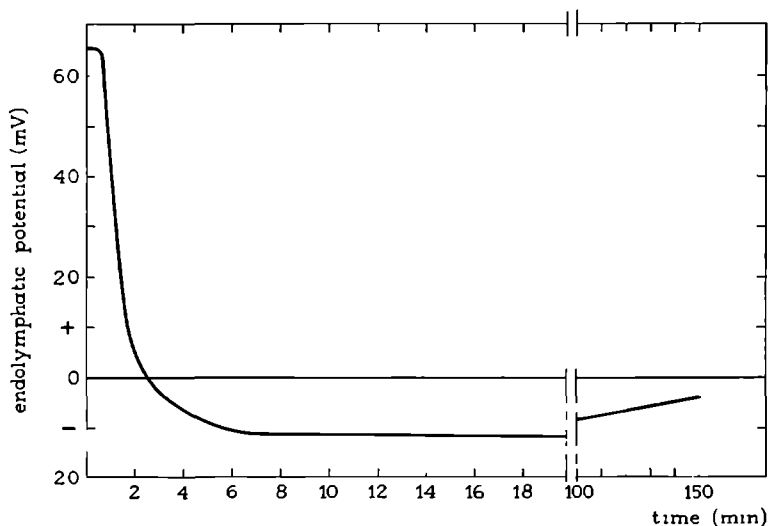


Fig. 39. Effect on ERP of prolonged anoxemia from inhalation of a gasmixture consisting of 20% CO_2 and 80% N_2 .

10^{-3} M ouabain until the potential had reached a stable negative value, usually, in about 30 minutes. Thereafter the scala vestibuli was perfused with solutions of various ionic compositions, but always con-

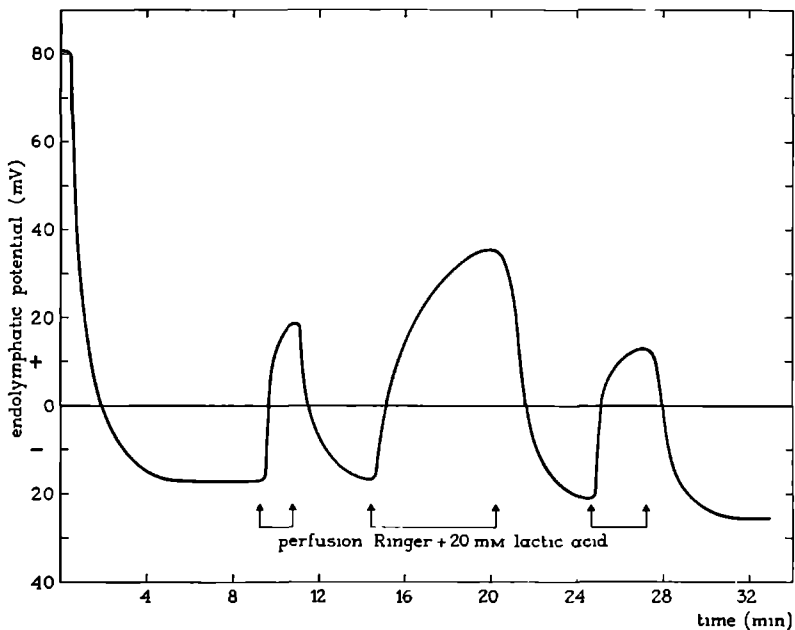


Fig 40 Effect on ERP of prolonged anoxemia and of perfusion of the scala vestibuli of the anoxic animal with 20 mM lactic acid in Ringer's solution. Anoxemia was induced at $t = 0$

taining 10^{-3} M ouabain. When Ringer's solution, in which Na^+ was replaced by choline was perfused, no effect on the negative potential was found (Fig. 42). The slight increase before choline Ringer perfusion was started may have been due to a movement of the electrode. When perfusion of K^+ Ringer (K^+ replacing Na^+) was started, the potential rose immediately to nearly zero within five minutes. A small fraction of the negative potential about 3 mV, was repeatedly found to persist. Subsequent perfusion with normal or choline Ringer's solution made the potential gradually decrease to negative values due to the removal of K^+ . If K^+ Ringer was again perfused, the potential immediately increased to nearly zero.

5 Discussion and conclusions

The large effect on the ERP of raising the K^+ concentration in the scala vestibuli and the small effect on this potential of lowering the Na^+ concentration in this compartment (Figs. 31 to 35) show clearly that the cochlear membranes are much more permeable for K^+ than for Na^+ . It is unlikely that the K^+ effect could be due to stimulation of the Na^+ - K^+ -ATPase system in the stria vascularis, because the enzyme is

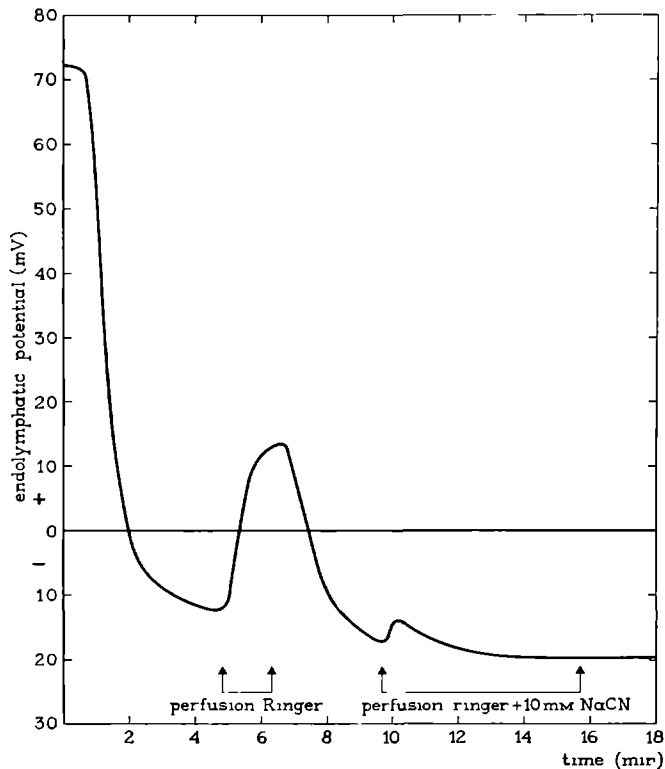


Fig. 41 Effect on ERP of prolonged anoxemia and of perfusion of the scala vestibuli of the anoxic animal with 10 mM NaCN in Ringer's solution. Anoxemia was induced at $t = 0$.

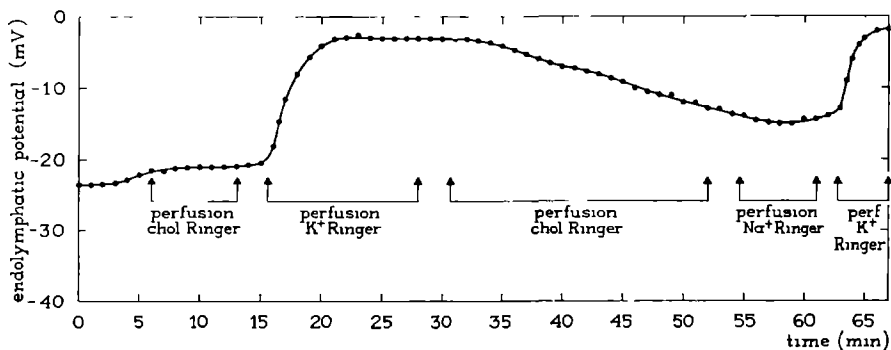


Fig 42. Effect on the negative endocochlear potential during perfusion of the scala vestibuli with solutions of various ionic composition. The negative potential was obtained by perfusing the scala vestibuli for 30 minutes with 10^{-3} M ouabain in Ringer's solution.

nearly maximally activated at 5 mM K^+ , which is the normal K^+ concentration in blood plasma. Therefore the 20 mV increase in the ERP must represent the disappearance of the K^+ diffusion potential, resulting from the unequal distribution of K^+ in perilymph and endolymph. This conclusion is confirmed by the disappearance of the negative potential caused by ouabain, upon perfusing the scala vestibuli with high- K^+ Ringer and the lack of an effect on the negative potential of perfusion with a Na^+ free Ringer.

The permeability characteristics of the tympanic wall of the scala media cannot be very different from those of the other cochlear membranes in view of the fact that Butler (1965) found an occasional increase of the ERP during perfusion of the scala tympani with 150 mM K^+ . No significant effect on the ERP could be found by perfusing the scala tympani with Na^+ -free solutions (Konishi and Kelsey, 1968 a). An additional argument against a high P_{Na}/P_K ratio for the cochlear membrane is the failure to affect the ERP with tetrodotoxin, which blocks Na^+ inflow during the rising phase of the nerve action potential (Katsuki et al., 1966, Konishi and Kelsey, 1968 b). The suggestion of Johnstone (1967) that the negative potential in anoxemia would be due to a change in the permeability ratio by the absence of oxygen is made unlikely by the occurrence of this negative potential after ouabain administration. These findings constitute convincing evidence against Johnstone's suggestion that the ERP would be a Na^+ diffusion potential. Johnstone (personal communication) has meanwhile obtained experimental evidence, which also argues against this suggestion.

The subsequent decrease of the ERP, which was observed by us after perfusing the scala vestibuli with high K^+ Ringer and also by Butler (1965) after perfusing the scala tympani with 150 mM KCl, might be due to an adverse effect of high extracellular K^+ concentration on cell metabolism (Schoffeniels, 1967). Significant increase in cell volume at high external K^+ concentration has been reported by Ussing et al. (1965). The incomplete recovery of the ERP upon perfusion with normal Ringer would also be in accordance with this explanation.

A significant contribution of Cl^- ions to the ERP can also be excluded on the basis of the experiment presented in Fig. 36. Likewise a contribution of H^+ ions is ruled out by the absence of a pH effect on the ERP.

The experiments with a high K^+ solution in the scala vestibuli, both on the ERP and on the negative potential due to ouabain, clearly show that the K^+ gradient between endolymph and perilymph does contribute to the ERP. This is, however, a negative contribution of about 20 mV. We must then conclude that the normal ERP of about +80 mV is the sum of two potentials, a positive potential of about +100 mV

(designated E_{+100}) and a negative potential of about -20 mV (designated E_{-20}). The E_{-20} is mainly a K^+ diffusion potential

We still have to explain the E_{+100} . We have shown that it cannot be an ion diffusion potential. This leaves the possibility that it is a secretion potential, generated by an electrogenic cation pump, which pumps a single ion without coupled transport of a counter ion. The cation pump must be the Na^+K^+ -ATPase system of the stria vascularis on the basis of our results reported in Chapter VIII. However, it cannot act as an electrogenic Na^+ pump in this case, since the endolymphatic resting potential is positive rather than negative.

There are two strong arguments in favour of an electrogenic pump being responsible for E_{+100} . First, there is the persistence of the ERP, though at a much lower value, after rupturing Reissner's membrane. All cation gradients across the stria vascularis and between scala media and scala vestibuli and tympani have been abolished in this case, and yet a potential remains. Secondly, in anoxemia the ERP disappears very rapidly, before the K^+/Na^+ ratio undergoes a significant change (Johnstone, 1965).

The occurrence of a positive secretion potential in connection with an electrogenic cation pump has been observed in the frog skin (Koefoed-Johnsen, 1957), the ciliary body of the eye (Cole, 1961), the nasal gland of marine birds (Thesleff and Schmidt-Nielsen, 1962), the chorioid plexus (Patlak, 1964) and the toad bladder (Herrera, 1968). With E_{+100} they have in common that they are highly sensitive to ouabain and that they are all due to the action of the Na^+K^+ -ATPase system (Bonting and Becker, 1964; Bonting et al., 1964a; Vates et al. 1964; Bonting and Canady, 1964). However there are two differences between these potentials and the E_{+100} . These potentials are small, ranging from 3-20 mV, and the positive potential is in the compartment to which Na^+ is being pumped. In the cochlea the potential is much larger and it is positive in the compartment from which Na^+ is extruded. Thus the E_{+100} cannot be due to an electrogenic Na^+ transport.

A logical conclusion would be that the E_{+100} is due to an electrogenic K^+ transport into the endolymph. Such a potential has so far only been found in the midgut of the silkworm (*Hyalophora cecropia*) (Haskell et al., 1965, Wood et al., 1969). In that case the potential was sensitive to anoxia, but not to 10^{-4} M ouabain. Although there is as yet no precedent for an ouabain-sensitive electrogenic K^+ pump, there is some experimental evidence in favour of this assumption. First, ^{42}K injected into the scala vestibuli rapidly appeared in the stria vascularis and in the endolymph, and this transport could be inhibited by ouabain (Rauch, 1966). Secondly after 30 minutes of anoxemia the decrease in the K^+ concentration of the endolymph is larger than the increase in

the Na^+ concentration (Bosher and Warren, 1968; Mendelsohn and Konishi, 1969), which suggest that the pump rate for K^- is higher than for Na^+ . Thirdly, the E_{+100} would be high enough to maintain a large part of the Na^+ gradient, because the Na^+ equilibrium potential, i.e. the potential that is just sufficient to maintain the observed Na^+ gradient (endolymph 1 mM, perilymph 136 mM), is about +135 mV. (Bosher and Warren, 1968). Thus the maintenance of the Na^+ gradient would require only a minor Na^+ pump activity.

Cl^- ions will tend to leak into the endolymph because of the high positive potential in this compartment. However, the actual Cl^- concentrations in the endolymph and perilymph are about equal. This suggests the existence of a Cl^- pump which extrudes Cl^- from the endolymph, as has previously been suggested by Johnstone (1967).

The increase of E_{+100} during perfusion of K^+ Ringer (Fig. 35) appeared to be much faster than that of E_{-20} under the same circumstances (Fig. 42). The slower increase of E_{-20} may be due to an effect of Cl^- . When the cochlear potential becomes negative, Cl^- ions will move out of the scala media and create a Cl^- concentration gradient between endolymph and blood and perilymph. This gradient could result in a small Cl^- diffusion potential, negative with respect to the perilymph. This small potential may also explain why the E_{-20} could never be completely abolished by perfusion with K^+ Ringer.

The decrease of the ERP after rupturing Reissner's membrane may be largely due to a deleterious effect of a low K^+ -high Na^+ medium on the stria vascularis. Duvall (1968), demonstrated that within a few hours after rupturing Reissner's membrane the stria vascularis showed degenerative changes. Moreover, Konishi et al. (1966) showed that when the scala media was perfused with high Na^+ solutions for about 30 minutes the ERP decreased slowly. This decrease could not be reversed by subsequent perfusion of high K^+ Ringer. The abolishing of the Na^+ gradient after rupturing Reissner's membrane could contribute another few mV to the decrease in potential.

The slight effect of CO_2 inhalation and acetazolamide on the potential suggests that carbonhydrase presumably is only involved in maintaining the pH of the cell. Prolonged inhibition of this enzyme may result in a change in the ionic composition of the endolymph, as found by Erulkar and Maren (1961), but only secondarily through an effect of the change in intracellular pH on cell metabolism.

Thus we feel that the ERP can be interpreted as the sum of a larger positive potential (E_{+100}) due to an ouabain-sensitive, electrogenic K^+ pump and a smaller negative K^- diffusion potential (E_{-20}). In addition we must assume an active Cl^- and Na^+ transport out of the endolymph, which may be coupled. Diffusion potentials for Na^+ , Cl^- and H^+ do not seem to contribute to the ERP to a significant extent

Certainty about these conclusions could be obtained by placing the stria vascularis with underlying ligamentum spirale in an Ussing chamber and measuring individual ion fluxes, potentials and short-circuit current. Our attempts to do this have failed probably due to the smallness or fragility of the structure.

SUMMARY

The cochlea contains two fluid compartments. One of these fluids, the endolymph, has an ionic composition as found inside cells, high potassium and low sodium content. The ionic composition of the perilymph is similar to that of other extracellular fluids. Some characteristic electrical potentials occur in the cochlea, which depend on the ionic composition of the cochlear fluids, and which are essential for the auditory process.

In most cells the high K^+ and the low Na^+ concentration is maintained by a cation pump, which is closely related or even identical with a Na^+-K^+ -activated ATPase system, which is specifically inhibited by ouabain and other cardiac glycosides.

In this thesis we have looked for a possible role of the Na^+-K^+ -ATPase system in the maintenance of the high K^+ and low Na^+ concentration of the endolymph. In addition, after establishing the presence of this enzyme system, the relationship between the Na^+-K^+ -ATPase system and the endolymphatic resting potential (ERP) and the cochlear microphonic potential (CMP) has been studied. This was investigated by perfusing the perilymphatic space with ouabain. The nature of the ERP was studied by changing the electrolyte composition of the perilymph, by rupturing Reissner's membrane and by application of anoxemia and ouabain.

In Chapter I our current knowledge of the anatomy of the cochlear structures is described. In Chapter II the composition and circulation of the cochlear fluids are discussed. The electrical potentials observed in the cochlea are reviewed in Chapter III. In Chapter IV the occurrence and properties of the Na^+-K^+ -ATPase system are described as well as its relation to cation transport. The methods used in our experiments are described in Chapter V.

The distribution and properties of the Na^+-K^+ -ATPase activity in the cochlear structures of the chicken and the guinea pig are reported in Chapters VI and VII, respectively. The enzyme system was shown to be present in very high activity in the stria vascularis of the guinea pig and in the comparable structure of the chicken, the tegmentum vasculosum, while the enzyme activity of the other cochlear structures was rather low. This points to stria vascularis and tegmentum vasculosum as the site of the cation pump involved in endolymph formation.

In Chapter VIII the effect of ouabain on the cochlear potentials in the guinea pig is described. Both the ERP and the CMP appeared to be affected strongly and to the same extent by ouabain. There was a remarkable similarity in the effect of ouabain on both potentials and on the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity. The pI_{50} for the CMP, the ERP and the $\text{Na}^+\text{-K}^+\text{-ATPase}$ activity were 5.5, 5.4 and 5.5 respectively. Thus, it was concluded that these potentials are dependent on the functioning of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the stria vascularis.

In Chapter IX the nature of the ERP is studied. The effects on the ERP of pH changes of anoxemia, cyanide and lactic acid, of rupturing Reissner's membrane and of changes in the electrolyte composition of the perilymph are reported. From these experiments it was concluded that the ERP is composed of a negative K^+ -diffusion potential and a positive potential due to an ouabain-sensitive electrogenic K^+ pump, represented by the $\text{Na}^+\text{-K}^+\text{-ATPase}$ system in the stria vascularis. In addition, an active Cl^- extrusion from the endolymph, possibly coupled with active Na^+ extrusion, was postulated in order to explain some of the observations.

SAMENVATTING

De cochlea bevat twee vloeistofcompartimenten. Eén van deze vloeistoffen, de endolymfhe, heeft een kationensamenstelling zoals die gevonden wordt in de cel. hoog kalium- en laag natriumgehalte. De perilymfhe heeft een kationensamenstelling die gelijk is aan die van andere extracellulaire vloeistoffen. In de cochlea zijn verder enige karakteristieke elektrische potentialen aanwezig, die afhankelijk zijn van de kationensamenstelling van de cochleaire vloeistoffen, en die van essentieel belang zijn voor het functioneren van het binnenoor.

In de meeste cellen wordt de hoge concentratie aan K^+ -ionen en de lage concentratie aan Na^+ -ionen in stand gehouden door een kationenpomp. Deze pomp is nauw verwant of zelfs identiek aan het Na^+K^- geactiveerde ATPase, dat specifiek geremd wordt door ouabaine en andere hartglycosiden.

In dit onderzoek werd nagegaan of het Na^+K^+ geactiveerde ATPase ook een rol speelt bij het in stand houden van de hoge K^+ en de lage Na^+ -concentratie van de endolymfhe. Daarnaast werd, nadat de aanwezigheid van het enzymstelsel was aangetoond, de afhankelijkheid van de cochleaire potentialen van het functioneren van het Na^+K^+ -ATPase systeem bestudeerd. Hiertoe werd het effect van ouabaine perfusie door de perilymphatische ruimte op de endolymphatische rustpotentiaal (ERP) en de cochleaire microphonische potentiaal (CMP) nagegaan. De aard van de ERP werd bestudeerd door wijziging van de electrolytsamenstelling van de perilymfhe, door verwijdering van het membraan van Reissner, en door toepassing van anoxaemie en ouabaine.

In hoofdstuk I wordt een overzicht gegeven van de anatomie van de verschillende cochleaire structuren. In hoofdstuk II worden samenstelling en circulatie van de cochleaire vloeistoffen besproken. De cochleaire potentialen worden beschreven in hoofdstuk III. Hoofdstuk IV geeft een overzicht van het voorkomen en de eigenschappen van het Na^+K^+ -ATPase en zijn relatie tot het transport van Na^+ en K^+ . De in dit onderzoek gebruikte methodieken worden besproken in hoofdstuk V.

De aanwezigheid, de eigenschappen en de verdeling van het Na^+K^+ geactiveerde ATPase over de verschillende cochleaire structuren worden beschreven in de hoofdstukken VI en VII respectievelijk voor de

cochlea van het kuiken en van de cavia. Het enzymstelsel blijkt in zeer hoge concentratie aanwezig te zijn in de stria vascularis van de cavia en in het hiermee vergelijkbare tegmentum vasculosum van het kuiken, terwijl de activiteit in de overige cochleaire structuren hiermee vergeleken bijzonder laag is. Dit duidt de stria vascularis en het tegmentum vasculosum aan als de plaats van de kationenpomp, betrokken bij de vorming van de endolymfe.

In hoofdstuk VIII wordt het effect van ouabaine op de cochleaire potentialen van de cavia beschreven. Zowel de ERP als de CMP bleken sterk en in dezelfde mate beïnvloed te worden door ouabaine. Er is een sterke overeenkomst in het remmend effect van ouabaine op de cochleaire potentialen en op het $\text{Na}^+\text{-K}^+$ geactiveerde ATPase systeem. De pI_{50} voor de CMP, de ERP en het $\text{Na}^+\text{-K}^+\text{-ATPase}$ zijn respectievelijk 5,5, 5,4 en 5,5. Hieruit werd geconcludeerd dat de potentialen afhankelijk zijn van het functioneren van het $\text{Na}^+\text{-K}^+\text{-ATPase}$ in de stria vascularis.

In hoofdstuk IX wordt de aard van de ERP bestudeerd. De effecten op de ERP van pH veranderingen, van anoxaemie, cyanide en melkzuur, van verwijdering van de membraan van Reissner en van veranderingen in de electrolytsamenstelling van de perilymphe worden beschreven. Uit deze resultaten werd geconcludeerd dat de ERP bestaat uit de som van een negatieve K^+ -diffusie potentiaal en een positieve component, veroorzaakt door een ouabaine-gevoelige electrogene K^+ -pomp, die vertegenwoordigd wordt door het $\text{Na}^+\text{-K}^+\text{-ATPase}$ systeem in de stria vascularis. Bovendien werd een actieve Cl^- extrusie vanuit de endolymfe, mogelijk gekoppeld met actieve Na^+ -extrusie, gepostuleerd ter verklaring van enkele waarnemingen.

REFERENCES

- Abdel-Latif, A. A., J. Brody and H. Ramahi* (1967) — Studies on sodium-potassium adenosine triphosphatase of the nerve endings and appearance of electrical activity in developing rat brain — *J. Neurochem.*, 14 1133.
- Adrian, R. H.* (1956) — The effect of the internal and external potassium concentration on the membrane potential of frog muscle — *J. Physiol. (London)*, 133 631.
- Ahmed, K. and J. D. Judah* (1965) — Identification of active phosphoprotein in a cation-activated ATPase — *Biochim. Biophys. Acta*, 104 112.
- Albers, R. W., S. Fahn and G. J. Koval* (1963) — Role of Na ions in the activation of *Electrophorus electric organ* ATPase — *Proc. Natl. Acad. Sci. U.S.A.*, 50 474.
- Albers, R. W. and G. J. Koval* (1966) — Na-K activated ATPase of *Electrophorus electric organ* III An associated K activated neutral phosphatase — *J. Biol. Chem.*, 241 1896.
- Albers, R. W., G. J. Koval and G. J. Siegel* (1968) — Interaction of ouabain and other cardioactive steroids with Na-K activated ATPase — *Molec. Pharmacol.*, 4 324.
- Allen, G. W.* (1964) — Endolymphatic sac and cochlear aqueduct. Their role in the regulation of labyrinthine pressure — *Arch. Otolaryng. (Chicago)*, 79: 322.
- Altmann, F. and J. G. Waltner* (1950) — Further investigations on the physiology of labyrinthine fluids — *Ann. Otol.*, 59 657.
- Amerlinck, A.* (1923) — Contribution à l'étude de la membrane de Reissner et de de l'épithélium de revêtement du canal cochleaire des oiseaux — *Arch. Biol. (Liège)*, 33 301.
- Arnvig, J.* (1951) — Relation of the ear to the subarachnoid space and absorption of the labyrinthine fluids — *Acta Otolaryng. (Stockholm)*, Suppl. 96.
- Axelsson, A.* (1968) — The vascular anatomy of the cochlea of the guinea pig and in man — *Acta Otolaryng. (Stockholm)*, Suppl. 243.
- Bader, H. and A. K. Sen* (1966) — (K^+) -dependent acylphosphatase as part of the $(Na^+ + K^+)$ -dependent ATPase of cell membranes — *Biochim. Biophys. Acta*, 118 116.
- Bader, H., R. L. Post and G. H. Bond* (1968) — Comparison of sources of a phosphorylated intermediate in transport ATPase — *Biochim. Biophys. Acta*, 150 41.
- Baker, P. F.* (1963) — Relation between phosphorus metabolism and the Na pump in intact crab nerve — *Biochim. Biophys. Acta*, 75 287.
- Baker, P. F. and C. M. Connelly* (1966) — Some properties of the external activation site of the Na pump in crab nerve — *J. Physiol. (London)*, 185 270.
- Baker, E. and W. J. Simmonds* (1966) — Membrane ATPase and electrolyte levels in marsupial erythrocytes — *Biochim. Biophys. Acta*, 126 492.
- Baker, P. F., M. P. Blaustein, R. D. Keynes, J. Manil, T. I. Shaw and R. A. Steinhart* (1969) — The ouabain sensitive fluxes of sodium and potassium in squid giant axons — *J. Physiol. (London)*, 200 459.
- Bakkeren, J. A. J. M.* (1968) — Active cation transport and $(Na^+ - K^+)$ -activated ATPase in rat liver — Thesis, Univ. of Nijmegen Nijmegen, The Netherlands.
- Bakkeren, J. A. J. M. and S. L. Bonting* (1968) — Studies on Na-K activated

- ATPase. XX. Properties of (Na⁺-K⁺)-activated ATPase in rat liver — *Biochim. Biophys. Acta*, 150 460.
- Bast, T. H. and B J. Anson* (1949) — The temporal bone and the ear — C. C. Thomas, Springfield
- Beal, D. D.* (1968) — Effect of endolymphatic sac ablation in the rabbit and cat — *Acta Otolaryng* (Stockholm), 66: 33.
- Bekesy, G. von* (1952) — DC resting potentials inside the cochlear partition — *J. Acoust. Soc. Amer.*, 24 72.
- Békésy, G. von* (1960) — Experiments in hearing — McGraw-Hill, New York.
- Berg, G. G. and B Chapman* (1965) — The sodium and potassium activated ATPase of intestinal epithelium. I Location of enzymatic activity in the cell. — *J. Cell. Comp. Physiol.*, 65 361.
- Bignam, A., G. Palladini and G Venturini* (1966) — Na-K ATPase in the developing chick brain — *Brain Res.* 3 207
- Bonting, S. L. and I. M Rosenthal* (1960) — Effect of the method of tissue preparation on the assay of tissue enzyme activities — *Nature*, 185 686.
- Bonting, S. L. and B R Mayron* (1961) — Construction, calibration and use of a modified quartz fiber fishpole" ultramicrobalance — *Microchem. J.*, 5 31.
- Bonting, S L, K. A Simon and N. M. Hawkins* (1961a) — Studies on sodium-potassium activated adenosinetriphosphatase no I Quantitative distribution in several tissues of the cat — *Arch Biochem*, 95 416.
- Bonting, S L., K A. Simon and L. L Caravaggio* (1961b) — Na-K activated ATPase and active cation transport — Abstracts 1st Ann. Mtg. Amer. Soc. Cell Biol, Chicago, p 19.
- Bonting, S L, L. L. Caravaggio and N. M. Hawkins* (1962) — Studies on sodium-potassium activated adenosinetriphosphatase no. IV. Correlation with cation transport sensitive to cardiac glycosides — *Arch. Biochem*, 98 413.
- Bonting, S. L. and L L. Caravaggio* (1963) — Studies on sodium-potassium activated adenosinetriphosphatase no. V. Correlation of enzyme activity with cation flux in six tissues — *Arch Biochem.*, 101 37.
- Bonting, S L, L L Caravaggio and N. M Hawkins* (1963) — Studies on sodium-potassium activated adenosinetriphosphatase no. VI. Its role in cation transport in the lens of cat, calf and rabbit — *Arch Biochem*, 101 47.
- Bonting, S. L. and B. Becker* (1964) — Studies on sodium-potassium activated adenosine-triphosphatase no XIV. Inhibition of enzyme activity and aqueous humor flow in the rabbit eye after intravitreal injection of ouabain — *Invest. Ophthal.*, 3 523.
- Bonting, S. L. and M. R. Canady* (1964) — Na-K activated ATPase and sodium transport in toad bladder (Studies on Na-K activated ATPase. XII) — *Amer. J. Physiol.*, 207 1005.
- Bonting, S L, I L Caravaggio, M R. Canady and N M. Hawkins* (1964a) — Studies on sodium-potassium activated adenosinetriphosphatase no. XI. The salt gland of the herring gull — *Arch. Biochem*, 106 49.
- Bonting, S. L., N M Hawkins and M R. Canady* (1964b) — Studies of sodium-potassium activated adenosinetriphosphatase. VII. Inhibition by erythrophleum alkaloids — *Biochem Pharmacol.*, 13 13.
- Bonting, S. L., L. L. Caravaggio and M. R. Canady* (1964c) — Studies on Na-K activated adenosinetriphosphatase no. X. Occurrence in retinal rods and relation to rhodopsin — *Exptl Eye Res.*, 3 47.
- Bonting, S L* (1966) — Studies on sodium-potassium activated adenosinetriphosphatase no XV. The rectal gland of the elasmobranchs — *Comp. Biochem. Physiol.*, 17 953
- Borghesan, E* (1957) — Modality of the cochlear humoral circulation — *Laryngoscope*, 67 1266.

- Bosher, S. K. and R. L. Warren* (1968) — Observations on the electrochemistry of the cochlear endolymph of the rat. a quantitative study of its electrical potential and ionic composition as determined by means of flame spectrophotometry — *Proc. Roy. Soc. (Biol.)*, 171: 227.
- Bulger, R. E.* (1963) — Fine structure of the rectal (salt secreting) gland of the spiny dogfish, *Squalus acanthias* — *Anat. Rec.*, 147: 95.
- Butler, R. A., V. Honrubia, B. M. Johnstone, and C. Fernández* (1962) — Cochlear function under metabolic impairment — *Ann. Otol.*, 71: 648.
- Butler, R. A.* (1965) — Some experimental observations on the resting potentials in the guinea pig cochlea — *J. Acoust. Soc. Amer.*, 37: 429.
- Caldwell, P. C., A. L. Hodgkin, R. D. Keynes and T. I. Shaw* (1960) — The effects of injecting "energy-rich" phosphate compounds on the active transport of ions in the giant axons of loloigo — *J. Physiol. (London)*, 152: 561.
- Chambers, A. H. and G. G. Lucchina* (1966) — Effects of dinitrophenol on cochlear potentials of the cat — *J. Audit. Res.*, 6: 13.
- Charnock, J. S. and R. L. Post* (1963) — Evidence of the mechanism of ouabain inhibition of cation-activated ATPase — *Nature*, 199: 910.
- Choo, Y. B. and D. Tabowitz* (1964) — The formation and flow of the cochlear fluids (Na^{22}) — *Ann. Otol.*, 73: 92.
- Choo, Y. B. and D. Tabowitz* (1965) — The formation and flow of the cochlear fluids (K^{42}) — *Ann. Otol.*, 74: 140.
- Chou, J. T. Y. and K. Rodgers* (1962) — Respiration of tissues lining the mammalian membranous labyrinth — *J. Laryng.*, 76: 341.
- Chou, J. T. Y.* (1963) — Respiration of Reissner's membrane of the guinea pig — *J. Laryng.*, 77: 374.
- Citron, L. and D. Exley* (1957) — Recent work on the biochemistry of the labyrinthine fluids — *Proc. Roy. Soc. Med.*, 50: 697.
- Cole, D. F.* (1961) — Electrochemical changes associated with the formation of the aqueous humor — *Brit. J. Ophthal.*, 45: 202.
- Corrie, W. S. and S. L. Bonting* (1966) — Studies on (Na^+ - K^+)-activated adenosine-triphosphatase no. XVII. Effect of ouabain and erythropleine on K concentration and membrane potential of frog sartorius muscle — *Biochim. Biophys. Acta*, 120: 91.
- Crane, R. V.* (1965) — Sodium dependent transport in the intestine and other animal tissues — *Fed. Proc.*, 24: 1000.
- Cummins, J. and H. Hyden* (1962) — ATP levels and ATPase in neurons, glia and neuronal membranes of the vestibular nucleus — *Biochim. Biophys. Acta*, 60: 271.
- Davis, H., I. Tasaki, C. A. Smith and G. H. Deatherage* (1955) — Cochlear potentials after intracochlear injections and anoxia — *Fed. Proc.*, 14: 112.
- Davis, H.* (1957) — Biophysics and physiology of the inner ear — *Physiol. Rev.*, 37: 1.
- Davis, H., B. H. Deatherage, B. Rosenblut, C. Fernández, R. Kimura and C. A. Smith* (1958) — Modification of cochlear potentials produced by streptomycin poisoning and by extensive venous obstruction — *Laryngoscope*, 68: 596.
- Davis, H.* (1961) — Some principles of sensory receptor action — *Physiol. Rev.*, 41: 391.
- Dohlman, G. and F. C. Ormerod* (1960) — The secretion and absorption of endolymph — *Acta otolaryng. (Stockholm)*, 51: 435.
- Dohlman, G. F.* (1965) — The mechanism of secretion and absorption of endolymph in the vestibular apparatus — *Acta Otolaryng. (Stockholm)*, 59: 275.
- Doi, S.* (1939) — Über die freien Zellen im Saccus endolymphaticus der Meer-schweinchen — *Zbl. Hals. Nas. Ohrenheilk.*, 31: 329.

- Doyle, W. L. (1960) — The principal cells of the salt-gland of marine birds — *Exp. Cell Res.*, 21: 386.
- Dunham, E. T. and I. M. Glynn (1961) — Adenosinetriphosphatase activity and the active movement of alkali metal ions — *J. Physiol. (London)*, 156: 274.
- Duvall, A. J. and V. T. Rhodes (1967a) — Reissner's membrane. An ultrastructural study — *Arch. Otolaryng. (Chicago)*, 86: 143.
- Duvall, A. J. and V. T. Rhodes (1967b). Ultrastructure of the organ of Corti following intermixing of cochlear fluids — *Ann. Otol.*, 76: 688.
- Duvall, A. J. (1968) — Ultrastructure of the lateral cochlear wall following intermixing of fluids — *Ann. Otol.*, 77: 317.
- Ebner, V. von (1903) — Schnecke und Schnecken gang. In: Köllikers Handbuch der Gewebelehre des Menschen — W. Engelmann, Leipzig, Bd. III, 2.
- Eggemann S. and W. Bruchmüller (1968) — Die kohlen säure anhydratase im Innenohr des Meerschweinchens und ihre Hemmung. — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 190: 450.
- Emmelot, P. and C. J. Bos (1966) — Plasma membranes. III. Mg^{2+} -ATPase, $(Na^+K^+Mg^{2+})$ -ATPase and 5'-nucleotidase activity of plasma membranes from rat liver — *Biochim. Biophys. Acta*, 120: 369.
- Engström, H., H. W. Ades and A. Andersson (1966) — Structural pattern of the organ of Corti — *Almqvist and Wiksell, Stockholm*.
- Epstein, F. H. and R. Whittam (1966) — Mode of inhibition by calcium of cell-membrane adenosinetriphosphatase activity — *Biochem. J.*, 99: 232.
- Erulkar, S. D. and T. H. Maren (1961) — Carbonic anhydrase and the inner ear — *Nature*, 189: 459.
- Fahn, S., G. J. Koval and R. W. Albers (1968) — Na-K activated ATPase of Electrophorus electric organ. V. Phosphorylation by ATP- 32 P. — *J. Biol. Chem.*, 243: 1993.
- Farquhar, M. G. and G. E. Palade (1963) — Junctional complexes in various epithelia — *J. Cell. Biol.*, 17: 375.
- Fernández, C. (1952) — Dimensions of the cochlea (guinea pig) — *J. Acoust. Soc. Amer.*, 24: 519.
- Fernández, C. (1967) — Biochemistry of labyrinthine fluids. Inorganic substances — *Arch. Otolaryng. (Chicago)*, 86: 222.
- Fletcher, G. L., I. M. Stainer and W. N. Holmes (1967) — Sequential changes in the adenosine triphosphatase activity and the electrolyte excretory capacity of the nasal glands of the duck (*Anas Platyrhynchos*) during the period of adaptation to hypertonic saline — *J. Exp. Biol.*, 47: 375.
- Gárdos, G. (1954) — Accumulation of K^+ ions in human blood cells — *Acta Physiol. Acad. Sci. Hung.*, 6: 191.
- Garrahan, P. J. and I. M. Glynn (1967a). The behaviour of the sodium pump in red cells in the absence of external potassium — *J. Physiol. (London)*, 192: 159.
- Garrahan, P. J. and I. M. Glynn (1967b) — Factors affecting the relative magnitudes of the sodium : potassium and sodium : sodium exchanges catalysed by the sodium pump — *J. Physiol. (London)*, 192: 189.
- Guild, S. R. (1927) — The circulation of the endolymph — *Amer. J. Anat.*, 39: 57.
- Hařkenschaid, J. C. M. and S. L. Bonting (1968) — Studies on (Na^+K^+) -activated ATPase, no. XIX. Occurrence and properties of a (Na^+K^+) -activated ATPase in *escherichia coli* — *Biochim. Biophys. Acta*, 151: 204.
- Hařkenschaid, J. C. M. and S. L. Bonting (1969) — Studies on Na-K activated adenosine triphosphatase no. XXIII. A Mg^{2+} ATPase in *E. Coli*, activated by monovalent cations — *Biochim. Biophys. Acta*, 178: 128.

- Hagiwara, S. and N. Saito* (1959) — Voltage-current relations in nerve cell membrane of onchidium veruculatum — *J. Physiol. (London)*, 148: 161.
- Haskell, J. A., R. D. Clemons and W. R. Harvey* (1965) — Active transport by the cecropia midgut. I. Inhibitors, stimulants, and potassium-transport — *J. Cell. Physiol.*, 65: 45.
- Heinz, E. and J. F. Hoffman* (1965) — Phosphate incorporation and Na-, K-ATPase activity in human red blood cell ghosts — *J. Cell Physiol.*, 65: 31.
- Held, H.* (1926) — Die Cochlea der Sauger und der Vogel, ihre Entwicklung und ihr Bau. In: *Handbuch der normale und pathologische Physiologie* — Springer, Berlin, Bd. 11: 467.
- Herrera, F. C.* (1968) — Bielectric properties and ionic content in toad bladder — *J. Gen. Physiol.*, 51: 261.
- Hinojosa, R. and E. L. Rodriguez-Echandia* (1966) — The fine structure of the stria vascularis of the cat inner ear — *Amer. J. Anat.*, 118: 631.
- Hodgkin, A. L. and B. Katz* (1949) — The effect of sodium ions on the electrical activity of the giant axon of the squid — *J. Physiol. (London)*, 108: 37.
- Hodgkin, A. L. and R. D. Keynes* (1955) — The potassium permeability of a giant nerve fibre — *J. Physiol. (London)*, 128: 61.
- Hoffman, J. F.* (1962) — Cation transport and structure of the red-cell plasma membrane — *Circulation*, 26: 1201.
- Hokin, M. R.* (1963) — Studies on a $\text{Na}^+\text{-K}^+$ dependent, ouabain-sensitive adenosinetriphosphatase in the avian salt gland — *Biochim. Biophys. Acta*, 77: 108.
- Honrubia, V., B. M. Johnstone and R. A. Butler* (1965) — Maintenance of cochlear potentials during asphyxia — *Acta Otolaryng. (Stockholm)*, 60: 105.
- Hughes, D. G. and J. T. Y. Chou* (1963) — The origin of the perilymph of the inner ear — *Life Sci.*, 2: 107.
- Iinuma, T.* (1967) — Evaluation of adenosinetriphosphatase activity in the stria vascularis and spiral ligament of normal guinea pigs — *Laryngoscope*, 77: 141.
- Iinuma, T. and J. F. Daly* (1968) — Evaluation of alkaline and acid phosphatases activity in the stria vascularis and spiral ligament of normal guinea pigs — *Laryngoscope*, 78: 1677.
- Ilberg, C. von, and A. Imamura* (1966) — Ein methodischer Beitrag zur Permeabilitätsmessung an der Reissner-Membran — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 187: 800
- Ilberg, C. von* (1968a) — Elektronenmikroskopische Untersuchung über Diffusion und Resorption von Thoriumdioxid an der Meerschweinchenschnecke. II. Reissnersche Membran — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 190: 426.
- Ilberg, C. von* (1968b) — Elektronenmikroskopische Untersuchungen über Diffusion und Resorption von Thoriumdioxid an der Meerschweinchenschnecke. I. Ligamentum spirale und Stria vascularis — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 190: 415.
- Ilberg, C. von* (1968c) — Elektronenmikroskopische Untersuchungen über Diffusion und Resorption von Thoriumdioxid an der Meerschweinchenschnecke. III. Limbus spiralis — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 192: 163.
- Ilberg, C. von* (1968d) — Elektronenmikroskopische Untersuchungen über Diffusion und Resorption von Thoriumdioxid an der Meerschweinchenschnecke. IV. Basilar Membran und Cortisches Organ — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 192: 384.
- Ilberg, C. von, H. Spoendlin and K. H. Vosteen* (1968) — Die Ultrastruktur und Funktion des Sulcus spiralis externus und der Prominentia spiralis der Meerschweinchenschnecke. Darstellung mittels Thorotrast — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 192: 124.

- Ishu, T H Silverstein and K Balogh jr* (1966) — Metabolic activities of the endolymphatic sac — *Acta Otolaryng* (Stockholm), 62 61
- Ishu, T and Y Nomura* (1968) — Histochemical study on adenosinetriphosphatase activity in the inner ear — *Pract Otorhinolaryng* (Basel), 30 237
- Iurato S* (1967a) — Submicroscopic structure of the inner ear — Pergamon, Oxford
- Iurato, S* (1967b) — Elektronenoptische Struktur der Innenohr-Membranen mit Ruckschlüssen auf ihre Eignung zum Stoffaustausch — *Arch Klin Exp Ohr Nas Kehlkopfheilk*, 189 113
- Jahnke V P G Lundquist and J Wersall* (1969) Some morphological aspects of sound perception in birds — *Acta Otolaryng* (Stockholm), 67 583
- Jarnefelt, J* (1961) — ATP dependent binding of Na by microsomes from brain — *Biochem Biophys Res Commun*, 6 285
- Johnson, R L and H H Spoendlin* (1966) — Structural evidence of secretion in the stria vascularis — *Ann Otol* 75 127
- Johnstone C G, R S Schmidt and B M Johnstone* (1963) — Sodium and potassium in vertebrate cochlear endolymph as determined by flame microspectrophotometry — *Comp Biochem Physiol* 9 335
- Johnstone B M* (1965) — The relation between endolymph and the endocochlear potential during anoxia — *Acta Otolaryng* (Stockholm) 60 113
- Johnstone, J R and B M Johnstone* (1966) — Origin of summating potential — *J Acoust Soc Amer* 40 1405
- Johnstone B M J R Johnstone and I D Pugsley* (1966) — Membrane resistance in endolymphatic walls of the first turn of the guinea pig cochlea — *J Acoust Soc Amer* 40 1398
- Johnstone, B M* (1967) — Genesis of the cochlear endolymphatic potential — *Curr Topics in Bioenerq* 2 335
- Judah, J D and K Ahmed* (1964) — Inhibitors of transport and cation activated ATP-ases — *J Cell Comp Physiol*, 64 355
- Katsuki Y K Yanagisawa and J Kanzaki* (1966) — Tetraethylammonium and tetrodotoxin effects on cochlear potentials — *Science* 151 1544
- Katz A I and F H Epstein* (1967) — Role of sodium-potassium activated adenosine triphosphatase in reabsorption of sodium by kidney — *J Clin Invest*, 46 1999
- Kikuchi, K and D A Hilding* (1967) — The spiral vessel and stria vascularis in shaker-1 mice — *Acta Otolaryng* (Stockholm), 63 395
- Kimura, R S* (1967) — Experimental blockage of the endolymphatic duct and sac and its effect on the inner ear of the guinea pig — *Ann Otol*, 76 664
- Kimura R S* (1969) — Distribution structure and function of dark cells in the vestibular labyrinth — *Ann Otol* 78 542
- Kiritake, I Y Nomura M Nagakura I Matsuba and S Sugiura* (1961) — A consideration on the circulation of the perilymph An experimental study — *Ann Otol*, 70 373
- Koburg E* (1961) — Autoradiographische Untersuchungen zum Nukleinsäurestoffwechsel der Gewebe der Cochlea — *Arch Klin Exp Ohr Nas Kehlkopfheilk* 178 150
- Koburg E and D Plester* (1962) — Zur Grosse des Eiweissstoffwechsels der Gewebe der Cochlea Autoradiographische Untersuchungen an Meerschweinchen mit H^3 Lysin — *Acta Otolaryng* (Stockholm), 54 319
- Koefoed-Johnsen, V* (1957) — Effect of g-strophanthin (ouabain) on the active transport of sodium through the isolated frog skin — *Acta Physiol Scand*, 42 Suppl 145 87
- Komnick, H and U Komnick* (1963) — Elektronenmikroskopische Untersuchungen

- zur funktionellen Morphologie des Ionentransportes in der Salzdrüse von *Larus argentatus* — *Z. Zellforsch.*, 60: 163.
- Konishi, T., R. A. Butler* and *C. Fernández* (1961) — Effect of anoxia on cochlear potentials — *J. Acoust. Soc. Amer.*, 33: 349.
- Konishi, T.* and *T. Yasuno* (1963) — Summating potential of the cochlea in the guinea pig — *J. Acoust. Soc. Amer.*, 35: 1448.
- Konishi, T., E. Kelsey* and *G. T. Singleton* (1966) — Effects of chemical alteration in the endolymph on the cochlear potentials — *Acta Otolaryng.* (Stockholm), 62: 393.
- Konishi, T., E. Kelsey* and *G. T. Singleton* (1967) — Negative potential in scala media during early stage of anoxia — *Acta Otolaryng.* (Stockholm), 64: 107.
- Konishi, T.* and *E. Kelsey* (1968a) — Effect of sodium deficiency on cochlear potentials — *J. Acoust. Soc. Amer.*, 43: 462.
- Konishi, T.* and *E. Kelsey* (1968b) — Effect of tetrodotoxin and procaine on cochlear potentials — *J. Acoust. Soc. Amer.*, 43: 471.
- Konishi, T.* and *E. Kelsey* (1968c) — Effects of cyanide on cochlear potentials — *Acta Otolaryng.* (Stockholm), 65: 381.
- Kürschner, H. D.* (1968) — Über die Lactat- und Malatdehydrogenase in den vier Windungen der Cochlea des Meerschweinchens — Thesis, Düsseldorf.
- Kuijpers, W., A. C. van der Vleuten* and *S. L. Bonting* (1967) — Cochlear function and sodium and potassium activated adenosine triphosphatase (Studies on Na-K activated adenosinetriphosphatase no. XVIII) — *Science*, 157: 949.
- Kuijpers, W.* and *S. L. Bonting* (1969) — Studies on (Na⁺-K⁺) activated ATPase no. XXIV. Localization and properties of ATPase in the inner ear of the guinea pig — *Biochim. Biophys. Acta*, 173: 477.
- Landon, E. J.* and *J. L. Norris* (1963) — Na- and K-dependent ATPase activity in a rat kidney endoplasmatic reticulum fraction — *Biochim. Biophys. Acta*, 71: 266.
- Lawrence, M., D. Wolsk* and *W. B. Litton* (1961) — Circulation of the inner ear fluids — *Ann. Otol.*, 70: 753.
- Lawrence, M.* (1966) — Histological evidence for localized radial flow of endolymph — *Arch. Otolaryng.* (Chicago), 83: 406.
- Lawrence, M.* (1967) — Electric polarization of the tectorial membrane — *Ann. Otol.*, 76: 287.
- Loewenstein, W. R., A. A. Terzuolo* and *Y. Washizu* (1963) — Separation of transducer and impulse-generating processes in sensory receptors — *Science*, 142: 1180.
- Lowry, O. H.* (1953) — The quantitative histochemistry of the brain histological sampling — *J. Histochem. Cytochem.*, 1: 420.
- Lundquist, P. G.* (1965) — The endolymphatic duct and sac in the guinea pig — *Acta Otolaryng.* (Stockholm), Suppl. 201.
- Maggio, E.* (1966) — The humoral system of the labyrinth — *Acta Otolaryng.* (Stockholm), Suppl. 218.
- Matschinsky, F. M.* and *R. Thalmann* (1967) — Quantitative histochemistry of microscopic structures of the cochlea — *Ann. Otol.*, 76: 638.
- Matsuura, S., K. Ikeda* and *T. Furukawa* (1968) — Ouabain and streptomycin: their different loci of action on saccular hair cells in goldfish — *Science*, 160: 1117.
- Mendelsohn, M.* and *T. Konishi* (1969) — The effect of local anoxia on the cation content of the endolymph — *Ann. Otol.*, 78: 65.
- Meyer zum Gottesberge, A., S. Rauch* and *E. Koburg* (1965) — Unterschiede in Metabolismus der einzelnen Schneckenwindungen — *Acta Otolaryng.* (Stockholm), 59: 116.
- Misrahy, G. A., E. W. Shinabarger* and *J. E. Arnold* (1958a) — Changes in cochlear endolymphatic oxygen availability, action potential and microphonics during

- and following asphyxia, hypoxia and exposure to loud sounds — *J. Acoust. Soc. Amer.*, 30: 701.
- Misrahy, G. A., K M Hildreth, E W. Shinabarger and W. J. Gannon (1958b)* — Electrical properties of wall of endolymphatic space of the cochlea (guinea pig) — *Amer J. Physiol*, 194 396.
- Misrahy, G. A., K. M. Hildreth, L. C. Clark and E. W. Shinabarger (1958c)* — Measurement of the pH of the endolymph in the cochlea of guinea pigs — *Amer. J. Physiol.*, 194 393.
- Mizukoshi, O. and J. F. Daly (1967)* — Oxygen Consumption in normal and kanamycin damaged cochleae — *Acta Otolaryng. (Stockholm)*, 64 45.
- Naftalin, L. and M. S. Harrison (1958)* — Circulation of labyrinthine fluids — *J. Laryng.*, 72 118.
- Nakai, Y. and D. A. Hilding (1966)* — Electronmicroscopic studies of adenosinetriphosphatase activity in the stria vascularis and spiral ligament — *Acta Otolaryng (Stockholm)*, 62 411.
- Nakai, Y. and D Hilding (1967)* — Adenosinetriphosphatase distribution in the organ of Corti Histochemical study by light and electron microscopy — *Acta Otolaryng. (Stockholm)*, 64 477.
- Nakai, Y. and D A. Hilding (1968a)* — Oxydative enzymes in the cochlea: An electron microscopic and histochemical study of succinic dehydrogenase and dihydronicotinamide adenine dinucleotide diaphorase — *Acta Otolaryng. (Stockholm)*, 65 459.
- Nakai, Y and D. Hilding (1968b)* — Vestibular endolymph producing epithelium. Electron microscopic study of the development and histochemistry of the dark cells of the crista ampullaris — *Acta Otolaryng. (Stockholm)* 66 120.
- Nastuk, W. L. and A L Hodgkin (1950)* — The electrical activity of single muscle fibers — *J. Cell. Physiol.*, 35. 39.
- Newey, H., P A Sanford and D H. Smyth (1968)* — Some effects of ouabain and K^+ on transport and metabolism in rat small intestine — *J. Physiol. (London)*, 194 237.
- Opit, L. J. and J. S. Charnock (1965)* — A molecular model for a Na pump — *Nature*, 208 471.
- Oppelt, W. W. and R. F. Palmer (1966)* — Stimulation of cerebrospinal fluid production by low doses of intraventricular ouabain — *J. Pharmacol. Exp. Ther.*, 154 581.
- Ozaki, M. and M. Sato (1965)* — Changes in the membrane potential and the membrane conductance associated with a sustained depolarization of the non-myelinated nerve terminal in Pacinian corpuscles — *J. Physiol. (London)*, 180. 186.
- Palmer, R. F. and B. R. Nechay (1964)* — Biphasic renal effects of ouabain in the chicken correlation with microsomal Na^+K^+ stimulated ATPase — *J. Pharmacol. Exp Ther.*, 146 92
- Palmer, R. F., K. C. Lassefer and S. L. Melvin (1966)* — Stimulation of Na^+K^+ dependent adenosinetriphosphatase by ouabain — *Arch. Biochem.*, 113. 629.
- Patlak, C. S. (1964)* — Potential difference of the ventricular fluid in vivo and in vitro in the dogfish — *Fed. Proc*, 23: 211.
- Pease, D. C. (1955)* — Electron microscopy of the tubular cells of the kidney cortex — *Anat. Rec*, 121. 723.
- Portius, H. I and K. Repke (1967)* — Preparation of the Na-K activated, Mg-dependent ATP phosphohydrolase system of heart muscle by isolation of the cell membrane — *Acta Biol. Med. German.*, 19 879.
- Post, R. L., C. R. Merritt, C. R. Kinsolving and C. D. Albright (1960)* — Membrane

- adenosine triphosphatase as a participant in the active transport of sodium and potassium in the human erythrocyte — *J Biol Chem*, 235 1796.
- Post, R L., A K. Sen and A S Rosenthal (1965) — A phosphorylated intermediate in adenosinetriphosphate-dependent sodium and potassium transport across kidney membranes — *J. Biol. Chem.*, 240 1437.
- Rauch, S. (1964) — *Biochemie des Hororgans* — Thieme Verlag, Stuttgart.
- Rauch, S and H Ruska (1965) — Zur elektronenoptischen Struktur und Funktion der Stria vascularis — *Arch. Klin Exp. Ohr. Nas. Kehlkopfheilk.*, 189 127.
- Rauch, S (1966) — Membrane problems of the inner ear and their significance — *J Laryng.*, 80 1144.
- Rice, E. A and E. W. Shinabarger (1961) — Studies on the endolymphatic DC potential of the guinea pig's cochlea — *J. Acoust Soc. Amer.*, 33 922.
- Ridderstap, A S and S L. Bonting (1969a) — $\text{Na}^+\text{-K}^+$ -activated ATPase and pancreatic secretion in the dog (Studies on Na-K activated adenosinetriphosphatase, no. XXII) — *Amer J. Physiol.*, 216 547.
- Ridderstap, A S. and S L Bonting (1969b) — $\text{Na}^+\text{-K}^+$ -activated ATPase and exocrine pancreatic secretion in vitro (Studies on Na-K activated adenosinetriphosphatase no. XXV) — *Amer. J Physiol.*, in press.
- Rodgers, K. and J. T. Y. Chou (1966) — Concentrations of inorganic ions in guinea pig inner ear fluids II. Post-mortem changes in the ionic composition of utricular endolymph and perilymph — *J Laryng*, 80 885.
- Rodriguez Echandia, E L. and M H Burgos (1965) — The fine structure of the stria vascularis of the guinea-pig inner ear — *Z. Zellforsch.*, 67 600
- Rosenberg, I. H., A L. Coleman and L E Rosenberg (1965) — The role of sodium ion in the transport of amino acids by the intestine — *Biochim. Biophys. Acta*, 102 161.
- Rosenberg, I. H. and L E. Rosenberg (1968) — Localization and characterization of adenosine triphosphatase in guinea pig intestinal epithelium — *Biochem. Physiol.*, 24 975
- Roth, T. F. and K. R. Porter (1964) — Yolk protein uptake in oocyte of the mosquito aedes aegypti — *J Cell. Biol.*, 20: 313.
- Schatzmann, H J. (1953) — Herzglykoside als Hemmstoffe für den aktiven Kalium- und Natrium Transport durch das Erythrocytenmembran — *Helv. Physiol. Pharmacol. Acta*, 11 346
- Schmidt, R S and C Fernandez (1962) — Labyrinthine DC potentials in representative vertebrates — *J. Cell Physiol.*, 59 311.
- Schmidt, R. S (1963a) — Types of endolymphatic potentials — *Comp. Biochem. Physiol.*, 10, 83.
- Schmidt, R S. (1963b) — Independence of the endovestibular potential in homeotherms — *J. Gen. Physiol.*, 47, 371.
- Schoffneels, E. (1967) — Cellular aspects of membrane permeability — Pergamon, Oxford, p 95.
- Schuknecht, H F. and A El Seifi (1963) — Experimental observations on the fluid physiology of the inner ear — *Ann Otol.*, 72 687.
- Schwartzkopf, J. and P Winter (1960) — Zur Anatomie der Vogel-Cochlea unter natürlichen Bedingungen — *Biol Zbl*, 79 607.
- Silverstein, H (1966) — Biochemical and physiologic studies of the endolymphatic sac in the cat — *Laryngoscope*, 76 498.
- Skou, J. C. (1957) — The influence of some cations on an adenosine triphosphatase from peripheral nerves — *Biochim. Biophys Acta*, 23 394.
- Skou, J. C. (1960) — Further investigations on a ($\text{Mg}^{++} + \text{Na}^+$)-activated adenosinetriphosphatase possibly related to the active linked transport of Na^+ and K^+ across the nerve membrane — *Biochim Biophys Acta*, 42: 6.

- Slegers, J. F. G.* (1968) — Ionic secretion by epithelial membranes. In: Control mechanisms in cystic fibrosis — Ciba foundation, London, p. 68.
- Slegers, J. F. G.* and *W. M. Moons* (1968) — Effect of acetazolamide on the chloride shift and the sodium pump in secretory cells — *Nature*, 220: 181.
- Smith, C. A., O. H. Lowry* and *M. L. Wu* (1954) — The electrolytes of the labyrinthine fluids — *Laryngoscope*, 64: 141.
- Smith, C. A.* (1957) — Structure of the stria vascularis and the spiral prominence — *Ann. Otol.*, 66: 521.
- Smith, C. A., H. Davis, B. H. Deatherage* and *C. F. Gessert* (1958) — DC potentials of the membranous labyrinth — *Amer. J. Physiol.*, 193: 203.
- Smith, C. A.* and *F. S. Sjöstrand* (1961) — Structure of the nerve endings on the external hair cells of the guinea pig cochlea as studied by serial sections — *J. Ultrastruct. Res.*, 5: 523.
- Spoendlin, H. H.* and *K. Balogh* (1963) — Histochemical localization of dehydrogenases in the cochlea of living animals — *Laryngoscope*, 73: 1061.
- Spoendlin, H.* and *W. Lichtensteiner* (1966) — The adrenergic innervation of the labyrinth — *Acta Otolaryng. (Stockholm)*, 61: 423.
- Spoendlin, H.* (1967a) — Vascular stria. In: *S. Iurato*, Submicroscopic structure of the inner ear — Pergamon, Oxford, p. 131.
- Spoendlin, H.* (1967b) — The innervation of the organ of Corti — *J. Laryng.*, 81: 717.
- Spoendlin, H.* (1969) — Innervation patterns in the organ of Corti of cat — *Acta Otolaryng. (Stockholm)*, 67: 239.
- Tasaki, I.* and *C. Fernández* (1952) — Modification of cochlear microphonics and action potentials by KCl solution and by direct currents — *J. Neurophysiol.*, 15: 497.
- Tasaki, I., H. Davis* and *J. P. Legoux* (1952) — The space-time pattern of the cochlear microphonics as recorded by differential electrodes — *J. Acoust. Soc. Amer.*, 24: 502.
- Tasaki, I., H. Davis* and *D. H. Eldredge* (1954) — Exploration of cochlear potentials in guinea pig with a microelectrode — *J. Acoust. Soc. Amer.*, 26: 765.
- Tasaki, I.* (1957) — Hearing — *Ann. Rev. Physiol.*, 19: 417.
- Tasaki, I.* and *C. S. Spyropoulos* (1959) — Stria vascularis as source of endocochlear potential — *J. Neurophysiol.*, 22: 149.
- Taylor, C. B.* (1962) — Cation stimulation of an ATPase system from the intestinal mucosa of the guinea pig — *Biochim. Biophys. Acta*, 60: 437.
- Terrayama, Y., E. Holz* and *C. Beck* (1966) — Adrenergic innervation of the cochlea — *Ann. Otol.*, 75: 69.
- Thesleff, S.* and *K. Schmidt-Nielsen* (1962) — An electrophysiological study of the salt gland of the herring gull — *Amer. J. Physiol.*, 202: 597.
- Tonndorf, J., A. J. Duvall* and *J. P. Reneau* (1962) — Permeability of intracochlear membranes to various vital stains — *Ann. Otol.*, 71: 801.
- Tosteson, D. C., R. H. Moulton* and *M. Blaustein* (1960) — Enzymatic basis for the difference in active cation transport in two genetic types of sheep red cells — *Fed. Proc.*, 19: 128.
- Ussing, H. H.* (1949) — Transport across cellular membranes — *Physiol. Rev.*, 29: 127.
- Ussing, H. H., T. V. Biber* and *N. S. Bricker* (1965) — Exposure of the isolated frog skin to high potassium concentration at the internal surface. II. Changes in epithelial cell volume, resistance and response to antidiuretic hormone — *J. Gen. Physiol.*, 48: 425.

- Vanzulli, A. and E Garcia-Austt* (1963) — Development of cochlear microphonic potentials in the chick embryo — *Acta Neurol. Lat. Amer*, 9 19.
- Vates, T S, S. L. Bonting and W. W. Oppelt* (1964) — Na-K activated adenosinetriphosphatase and formation of cerebrospinal fluid in the cat (Studies on Na-K activated adenosinetriphosphatase no IX) — *Amer J. Physiol*, 206 1165.
- Vosteen, K. H.* (1961) — Neue Aspekte zur Biologie und Pathologie des Innenohres — *Arch. Klin. Exp. Ohr. Nas. Kehlkopfheilk.*, 178 1.
- Wever, E. G.* (1966) — Electrical potentials of the cochlea — *Physiol. Rev.*, 46 102.
- Whittam, R.* (1962) — The asymmetrical stimulation of a membrane adenosine triphosphatase in relation to active cation transport — *Biochem. J.*, 84. 110.
- Wiley, J. S.* (1969) — Inheritance of an increased sodium pump in human red cells — *Nature*, 221 1222.
- Wood, J. L., P. S. Farrand and W. R. Harvey* (1969) — Active transport of potassium by the cecropia midgut. VI Microelectrode potential profile — *J. Exp. Biol.*, 50. 169.
- Yamamoto, K. and Y. Nakai* (1964) — Electronmicroscopic studies on the functions of the stria vascularis and the spiral ligament in the inner ear — *Ann Otol.*, 73. 332 .

Part of the results described in this thesis have been published

W. Kuijpers, A. C. van der Vleuten and S. L. Bonting (1967); W. Kuijpers and S. L. Bonting (1969).

STELLINGEN

I

De conclusie van Lawrence dat in de membrana tectoria geen rust-potentiaal aanwezig is, is zeer onwaarschijnlijk.

M Lawrence, *Ann Otol*, 76 287, 1967

II

De veronderstelling van Ling en Cope dat het merendeel van de intracellulaire K^+ -ionen van de kikkerspier geadsorbeerd is aan macromoleculen, is niet in overeenstemming te brengen met het iso-osmotisch evenwicht tussen intra- en extracellulair compartiment

G N Ling en F W Cope, *Science*, 163 1335, 1969

III

Wanneer bij ionentransport door biologische membranen de gemeten flux ratio overeenkomt met die berekend volgens de formule van Ussing, is er sprake van passief transport. Wanneer deze overeenkomst ontbreekt, mag echter niet automatisch tot het bestaan van actief transport geconcludeerd worden

IV

Op grond van de gebruikte isolatiemethode, mag de door Eaton en Moss gekarakteriseerde alkalische fosfatase activiteit niet als representatief beschouwd worden voor de alkalische fosfatase activiteit in het onderzochte bot.

R H Eaton en D W Moss, *Enzymologia*, 35 31, 1968

V

Gezien de gebruikte methodieken, mag uit de experimenten van Kasbekar en Durbin niet geconcludeerd worden dat er in het maagslijmvlies van *Rana catesbeiana* geen Na^+ - K^+ -geactiveerd ATPase voorkomt.

D K Kasbekar en R P Durbin, *Biochim Biophys Acta*, 105 472, 1965

VI

De conclusie dat *Streptomyces antibiotica* het glycolytische systeem van de zintuigcellen van het binnenoor aantasten, berust op een foutieve interpretatie van de experimentele gegevens.

H J Meuwissen en G C Robinson, *Clin Pediat (Phila)*, 6 262 1967

VII

De argumenten van Goodhill voor een inductieve werking van de annulus tympanicus op de vorming van het trommelvlies, zijn niet overtuigend

V Goodhill *Ann Otol* 75 866, 1966

VIII

De veronderstelling dat de bij het syndroom van van der Hoeve en de Kleijn beschreven geleidingshardhorendheid berust op een stapes-ankylose, identiek met die bij otosclerose, is zowel op klinische als op histopathologische gronden aanvechtbaar

H Wullstein R F Ogilvie en J S Hall, *J Laryng*, 74 67, 1960

P Clerc en L G Chevance, *Ann Otolaryng (Paris)*, 82 413, 1965

O Opheim, *Acta Otolaryng (Stockholm)*, 65 337 1968

IX

Het ontstaan van exogastrulae tijdens de embryogenese van *Limnaea stagnalis* na behandeling van eicellen rondom de tweede klievingsfase met LiCl hoeft niet uitsluitend het gevolg te zijn van deze behandeling. Dit verschijnsel kan mede veroorzaakt zijn door de samenstelling van het na de behandeling met LiCl gebruikte kweekmedium.

W L M Geilenkirchen, *J Embryol Exp Morph*, 17 367, 1967

L G Barth en L J Barth, *J Embryol Exp Morph*, 19 387, 1968

X

Bij gebruik van knaagdieren voor experimenteel onderzoek dient wegens het frequent voorkomen van otitis media bij ogenschijnlijk gezonde dieren een otoscopisch onderzoek te gebeuren

XI

Het is in het belang van het fundamenteel onderzoek in de kliniek, de hiervoor bestaande laboratoria in een instituut samen te brengen

W Kuipers, 1969.

