

University of Groningen

CCC- and WASH-mediated endosomal sorting of LDLR is required for normal clearance of circulating LDL

Bartuzi, Paulina; Billadeau, Daniel D.; Favier, Robert; Rong, Shunxing; Dekker, Daphne; Fedoseienko, Alina; Fieten, Hille; Wijers, Melinde; Levels, Johannes H.; Huijkman, Nicolette

Published in:
Nature Communications

DOI:
[10.1038/ncomms10961](https://doi.org/10.1038/ncomms10961)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2016

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Bartuzi, P., Billadeau, D. D., Favier, R., Rong, S., Dekker, D., Fedoseienko, A., Fieten, H., Wijers, M., Levels, J. H., Huijkman, N., Kloosterhuis, N., van der Molen, H., Brufau, G., Groen, A. K., Elliott, A. M., Kuivenhoven, J. A., Plecko, B., Grangl, G., McGaughran, J., ... van de Sluis, B. (2016). CCC- and WASH-mediated endosomal sorting of LDLR is required for normal clearance of circulating LDL. *Nature Communications*, 7, [10961]. <https://doi.org/10.1038/ncomms10961>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

ARTICLE

Received 18 Nov 2015 | Accepted 4 Feb 2016 | Published 11 Mar 2016

DOI: 10.1038/ncomms10961

OPEN

CCC- and WASH-mediated endosomal sorting of LDLR is required for normal clearance of circulating LDL

Paulina Bartuzi¹, Daniel D. Billadeau^{2,3}, Robert Favier⁴, Shunxing Rong⁵, Daphne Dekker¹, Alina Fedoseienko¹, Hille Fieten⁴, Melinde Wijers¹, Johannes H. Levels⁶, Nicolette Huijckman¹, Niels Kloosterhuis¹, Henk van der Molen¹, Gemma Brufau⁷, Albert K. Groen⁷, Alison M. Elliott⁸, Jan Albert Kuivenhoven¹, Barbara Plecko^{9,10}, Gernot Grangl⁹, Julie McGaughan^{11,12}, Jay D. Horton^{5,13}, Ezra Burstein^{13,14}, Marten H. Hofker¹ & Bart van de Sluis¹

The low-density lipoprotein receptor (LDLR) plays a pivotal role in clearing atherogenic circulating low-density lipoprotein (LDL) cholesterol. Here we show that the COMMD/CCDC22/CCDC93 (CCC) and the Wiskott-Aldrich syndrome protein and SCAR homologue (WASH) complexes are both crucial for endosomal sorting of LDLR and for its function. We find that patients with X-linked intellectual disability caused by mutations in *CCDC22* are hypercholesterolaemic, and that COMMD1-deficient dogs and liver-specific *Comm1* knockout mice have elevated plasma LDL cholesterol levels. Furthermore, *Comm1* depletion results in mislocalization of LDLR, accompanied by decreased LDL uptake. Increased total plasma cholesterol levels are also seen in hepatic COMMD9-deficient mice. Inactivation of the CCC-associated WASH complex causes LDLR mislocalization, increased lysosomal degradation of LDLR and impaired LDL uptake. Furthermore, a mutation in the WASH component *KIAA0196* (strumpellin) is associated with hypercholesterolaemia in humans. Altogether, this study provides valuable insights into the mechanisms regulating cholesterol homeostasis and LDLR trafficking.

¹Department of Pediatrics, Molecular Genetics Section, University of Groningen, University Medical Center Groningen, Antonius Deusinglaan 1, 9713 AV Groningen, The Netherlands. ²Department of Immunology, Mayo Clinic College of Medicine, Mayo Clinic, 200 First Street Southwest, Rochester, Minnesota 55905, USA. ³Department of Biochemistry and Molecular Biology, Mayo Clinic College of Medicine, Mayo Clinic, 200 First Street Southwest, Rochester, Minnesota 55905, USA. ⁴Department of Clinical Sciences of Companion Animals, Faculty of Veterinary Medicine, Utrecht University, Yalelaan 108, 3584 CM Utrecht, The Netherlands. ⁵Department of Molecular Genetics, University of Texas Southwestern Medical Center, 5323 Harry Hines Boulevard, Dallas, Texas 75390, USA. ⁶Department of Vascular and Experimental Vascular Medicine, Academic Medical Center, University of Amsterdam, Meibergdreef 9, 1105 AZ Amsterdam, The Netherlands. ⁷Department of Pediatrics, University of Groningen, University Medical Center Groningen, Antonius Deusinglaan 1, 9713 AV Groningen, The Netherlands. ⁸Department of Medical Genetics, University of British Columbia, 4500 Oak Street, Vancouver, British Columbia, Canada V6H 3N1. ⁹Department of Pediatrics and Adolescence Medicine, Division of Child Neurology, Medical University Graz, Auenbruggerplatz 2, A-8036 Graz, Austria. ¹⁰Division of Child Neurology, University Children's Hospital, Steinwiesstrasse 75, CH-8032 Zurich, Switzerland. ¹¹Genetic Health Queensland at the Royal Brisbane and Women's Hospital, Herston, University of Queensland, Herston, Brisbane, Queensland 4029, Australia. ¹²School of Medicine, University of Queensland, Herston, Brisbane, Queensland 4029, Australia. ¹³Department of Internal Medicine, University of Texas Southwestern Medical Center, 5323 Harry Hines Boulevard, Dallas, Texas 75390, USA. ¹⁴Department of Molecular Biology, University of Texas Southwestern Medical Center, 5323 Harry Hines Boulevard, Dallas, Texas 75390, USA. Correspondence and requests for materials should be addressed to B.v.d.S. (email: a.j.a.van.de.sluis@umcg.nl).

Various transmembrane proteins in the endosomal compartment depend on the Wiskott–Aldrich syndrome protein and SCAR homologue (WASH) complex to find their appropriate destination in the cell^{1–3}. This pentameric protein complex, which consists of WASH1, FAM21, strumpellin, KIAA1033 (also known as SWIP) and CCDC53, is recruited to endosomes by the retromer complex^{2,4–6}. Retromer is formed by the vacuolar protein-sorting (VPS) proteins VPS26, VPS29 and VPS35, and in concert with sorting nexins, it selectively mediates endosomal cargo sorting into recycling and retrieval pathways⁷. Recently, the COMMD/CCDC22/CCDC93 (CCC) complex has been identified to interact and colocalize with retromer and the WASH complex^{2,8,9}. The CCC complex consists of the copper metabolism MURR1 domain-containing (COMMD) proteins, coiled-coil domain-containing protein 22 (CCDC22), coiled-coil domain-containing protein 93 (CCDC93) and C16orf62 (ref. 8). Among the 10 COMMD proteins¹⁰, COMMD1, a gene product that is mutated in Bedlington terriers affected by a hepatic copper storage disorder resulting in copper toxicosis¹¹, was shown to regulate in concert with the WASH complex the recycling of the copper transporter ATP7A (ref. 8). In this same study, we reported that X-linked intellectual disability (XLID) patients carrying a mutation in *CCDC22* (c.49A>p.T17A) also have aberrant copper homeostasis, as serum copper and serum ceruloplasmin levels are increased in these patients⁸.

Given the pleiotropic function of COMMD1 (ref. 12) and CCDC22 (refs 2,8,9,13,14), and the large number of membrane proteins sorted by the WASH complex³, it is expected that the CCC complex is involved with numerous physiological processes. Here we identify that the CCC complex regulates the level of circulating low-density lipoprotein (LDL) cholesterol by mediating the endosomal trafficking of the low-density lipoprotein receptor (LDLR). Mutations affecting the formation of the CCC complex cause hypercholesterolaemia in humans, dogs and mice. We further show that LDLR is an endosomal cargo of the CCC-associated WASH complex, and inactivation of this complex results also in LDLR mislocalization and impaired LDL uptake. This study provides novel insights into the molecular mechanism causing hypercholesterolaemia, and highlights the importance of CCC and WASH complexes in cholesterol homeostasis.

Results

CCDC22 mutations are associated with hypercholesterolaemia. On further clinical analysis of a large XLID family affected with a *CCDC22* p.T17A mutation we discovered that these patients also have an increase in total plasma cholesterol and LDL cholesterol levels (Table 1 and Supplementary Fig. 1a), exceeding the 95th percentile corrected for age and gender^{15,16}. One of the patients (V-2, 4 years of age) is too young, and plasma cholesterol levels are not informative in this case¹⁷. In another XLID family with a *CCDC22* mutation (p.Y557C) (ref. 18), we found that the circulating total cholesterol (TC) and LDL cholesterol of the two patients carrying the mutation were also above the 95th percentile (Table 1 and Supplementary Fig. 1b). These observations suggest that mutations in the CCC component *CCDC22* are causally related to hypercholesterolaemia.

COMMD1 deficiency results in hypercholesterolaemia in dogs.

Since *CCDC22* mutation (p.T17A) and COMMD1 inactivation both impairs the formation of a stable CCC complex^{8,13}, we were prompted to investigate the plasma cholesterol levels in dogs with a *COMMD1* loss-of-function mutation^{11,19}. In addition to the expected copper accumulation in the liver of these animals (Supplementary Fig. 2a), the levels of the CCC components CCDC22 and CCDC93 were markedly reduced in liver homogenates from a COMMD1-deficient (*COMMD1*^{-/-}) dog (Fig. 1a). Importantly, similar to patients with *CCDC22* mutations, *COMMD1*^{-/-} dogs have elevated plasma TC levels, showing a ~50% increase in plasma cholesterol levels (Fig. 1b) without affecting plasma triglyceride (TG) concentrations (Fig. 1c). In unaffected littermates (*COMMD1*^{+/-} dogs), cholesterol is predominantly carried in high-density lipoprotein (HDL), due to the absence of cholesteryl ester transfer protein activity in dogs²⁰. In *COMMD1*^{-/-} dogs, the plasma cholesterol profile revealed that cholesterol is mainly present in the very low-density lipoprotein (VLDL) and LDL fractions (Fig. 1d).

High levels of copper in the liver have previously been described to be associated with reduced circulating cholesterol levels in mice²¹, we therefore evaluated a possible correlation between progressive hepatic copper accumulation and plasma cholesterol levels in dogs. As a model, we used Labrador retrievers

Table 1 | Plasma lipid levels of individuals with mutations in *CCDC22*.

Family	Patient	DOB	Age	Genotype <i>CCDC22</i>	Total cholesterol	HDL	LDL	TG
a	III-1	25/1/67	48	T17A	7.2 mmol l ⁻¹	1.1 mmol l ⁻¹	5.6 mmol l ⁻¹	1.1 mmol l ⁻¹
	IV-4	27/8/84	30	T17A	6.7 mmol l ⁻¹	ND	5.4 mmol l ⁻¹	ND
	IV-5	28/9/88	26	WT/T17A	5.3 mmol l ⁻¹	0.9 mmol l ⁻¹	3.5 mmol l ⁻¹	1.7 mmol l ⁻¹
	V-1	18/1/05	9	T17A	5.8 mmol l ⁻¹	2.8 mmol l ⁻¹	3.9 mmol l ⁻¹	0.3 mmol l ⁻¹
	V-2	9/7/10	4	T17A	4.4 mmol l ⁻¹	1.6 mmol l ⁻¹	2.5 mmol l ⁻¹	0.7 mmol l ⁻¹
	V-3	6/9/04	10	T17A	7.2 mmol l ⁻¹	2.1 mmol l ⁻¹	4.9 mmol l ⁻¹	0.4 mmol l ⁻¹
b	I-1	11/6/75	40	WT	5.2 mmol l ⁻¹	1.3 mmol l ⁻¹	3.5 mmol l ⁻¹	1.3 mmol l ⁻¹
	I-2	6/3/77	38	WT/Y557C	4.9 mmol l ⁻¹	1.1 mmol l ⁻¹	3.5 mmol l ⁻¹	1.1 mmol l ⁻¹
	II-1	14/7/96	19	Y557C	5.7 mmol l ⁻¹	1.6 mmol l ⁻¹	4.2 mmol l ⁻¹	0.7 mmol l ⁻¹
	II-2	26/8/03	11	Y557C	6.0 mmol l ⁻¹	2.0 mmol l ⁻¹	3.9 mmol l ⁻¹	0.7 mmol l ⁻¹
	III-3	25/9/98	16	WT	3.9 mmol l ⁻¹	1.4 mmol l ⁻¹	2.2 mmol l ⁻¹	1.0 mmol l ⁻¹

DOB, date of birth; ND, not determined.

In red levels >95th percentiles corrected for age and gender. The age is indicated at the moment of clinical evaluation.

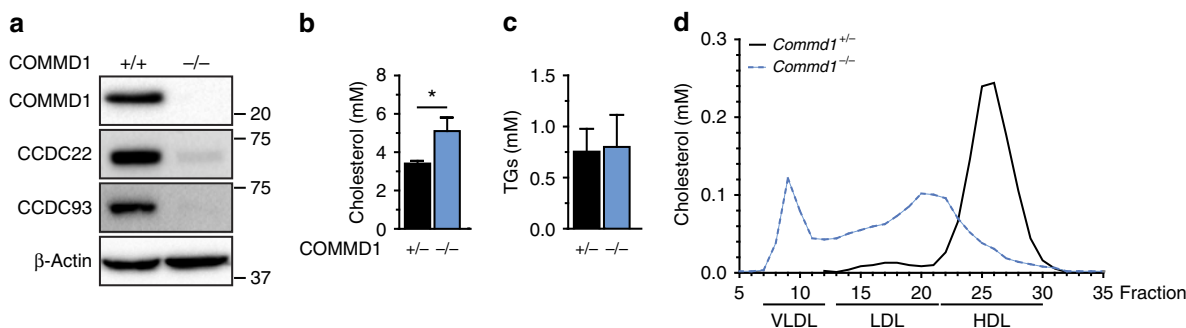


Figure 1 | COMMD1-deficient dogs are hypercholesterolaemic. (a) Western blot analysis of COMMD1, CCDC22 and CCDC93 in livers from an unaffected dog (*COMMD1*^{+/+}) and an affected dog homozygous for a loss-of-function mutation in *COMMD1* (*COMMD1*^{-/-}). (b) Plasma TC and (c) TG levels of dogs heterozygous (+/-) (*n* = 5) or homozygous (-/-) (*n* = 4) for a *COMMD1* mutation (d) FPLC lipoprotein profile of *COMMD1*^{+/+} (*n* = 5) and *COMMD1*^{-/-} dogs (*n* = 4). Pooled plasma samples of each experimental group were separated using FPLC gel filtration. Fifty fractions were collected from each separation. TC content was determined and lipoprotein profile was plotted. The results are presented as mean ± s.e.m., and significance was calculated relative to the control group by unpaired Student's *t*-test; **P* < 0.05.

affected by copper toxicosis^{22,23} (Supplementary Fig. 2b). The aetiology of this disorder is unknown, but *COMMD1* has been excluded to be the causal gene. Plasma cholesterol levels and lipoprotein profile were unchanged in affected Labrador retrievers (Supplementary Fig. 2c,d). These observations are in agreement with a recent study on Wilson disease, the main disorder of copper toxicosis in humans²⁴. Although Wilson disease patients accumulate copper to toxic levels in their livers, as *COMMD1*-deficient dogs, no unusual plasma cholesterol (total, HDL and LDL) levels were observed²⁴. Altogether, these data suggest that aberrant copper homeostasis and elevated plasma cholesterol are not causally linked in dogs or humans, and thus the CCC-deficient phenotype is due to pleiotropic effects.

Ablation of hepatic *Commd1* increases plasma LDL in mice.

LDL is the foremost cholesterol type that is elevated in patients and dogs carrying mutations affecting the CCC complex, and since the liver is the main organ involved in clearing circulating LDL cholesterol²⁵ we determined the cholesterol levels and the lipoprotein profiles of hepatocyte-specific *Commd1* knockout mice (*Commd1*^{ΔHep}) (ref. 26) fed either a chow diet or a high-fat high-cholesterol (HFC, cholesterol 0.2%) diet. Consistent with the increased plasma cholesterol concentrations found in *COMMD1*-deficient dogs, plasma cholesterol levels in *Commd1*^{ΔHep} mice were significantly higher than in *Commd1*^{loxP/loxP} wild-type (WT) littermates (Fig. 2a), with an increase of ~35% (chow-fed mice) and ~39% (HFC-fed mice). Plasma TG levels, hepatic cholesterol, hepatic TG concentrations and body weight were however unaffected by the loss of hepatic *Commd1* (Fig. 2b and Supplementary Tables 1 and 2). As shown previously²⁶, hepatic copper concentrations were not affected in *Commd1*^{ΔHep} mice when dietary copper was not increased (Supplementary Tables 1 and 2), which further supports that the increase in plasma cholesterol by CCC insufficiency is not caused by aberrant copper homeostasis. These data are in agreement with the fact that patients with *CCDC22* mutations have increased serum ceruloplasmin and copper levels, but do not exhibit clinical signs of copper overload or liver injury⁸. In addition, we observed no differences in cholesterol levels between WT mice and transgenic mice expressing Cre recombinase in hepatocytes (Alb-Cre), thus excluding the possibility that Cre recombinase expression in murine liver affects cholesterol metabolism (Supplementary Fig. 3a).

Next, we determined the lipoprotein profiles of pooled plasma samples from WT and *Commd1*^{ΔHep} mice fed either conventional chow (Fig. 2c) or HFC diet (Fig. 2d). Independent of the diet,

hepatic *Commd1* ablation resulted in significantly higher LDL cholesterol levels (Fig. 2c–f and Supplementary Fig. 3b,c,d,e). To assess whether the increase in plasma LDL in hepatocyte *COMMD1*-deficient mice is caused by aberrant VLDL homeostasis, VLDL production and secretion were measured. As shown in Supplementary Fig. 4a,b, no differences were found in VLDL production and secretion, indicating that *COMMD1* is not involved in VLDL homeostasis. Furthermore, we observed no alterations in hepatic expression of genes and proteins involved in cholesterol uptake, synthesis or efflux (Fig. 2g,h and Supplementary Fig. 4c). However, consistent with results from *COMMD1*-deficient dogs (Fig. 1a), *CCDC22* and *CCDC93* levels were both markedly decreased in *Commd1*-ablated hepatocytes (Fig. 2g,h). With the notion that all members of the *COMMD* family can associate with the CCC complex^{8,13,14}, we assessed if *COMMD9* is also involved in cholesterol homeostasis. We therefore deleted *Commd9* specifically in hepatocytes by intravenous injection of recombinant adenovirus carrying Cre recombinase (Ad-Cre) in conditional *Commd9* knockout mice (*Commd9*^{loxP/loxP}) (ref. 14). As a negative control mice were injected with adenovirus carrying the *LacZ* gene (Ad-LacZ). Cre-mediated ablation of *Commd9* resulted in a clear reduction in hepatic *Commd9* mRNA and *COMMD9* protein levels (Supplementary Fig. 5a,b). Similarly, as shown in hepatocytes lacking *COMMD1* (Fig. 2g,h), hepatic *COMMD9* inactivation resulted in a decrease in *CCDC22* and *CCDC93* protein levels (Supplementary Fig. 5b), which was accompanied by an increase in plasma cholesterol levels (Supplementary Fig. 5c). Together, these data indicate that mutations affecting the formation of the CCC complex cause hypercholesterolaemia.

COMMD1 binds to LDLR through its COMM domain. Plasma LDL cholesterol levels are largely dependent on functional LDL receptors in the liver, and since the CCC complex mediates the trafficking of various transmembrane proteins^{12,14}, we investigated whether *COMMD1* interacts with LDLR and is required for normal LDLR function. Pull-down and immunoprecipitation assays demonstrated a clear association between *COMMD1* and LDLR (Fig. 3a,b). Immunofluorescence staining in mouse embryonic fibroblasts (MEFs) and continuous sucrose gradient fractionation of fresh liver homogenates showed overlap in the cellular distribution of *COMMD1* and LDLR, which support a physical association between *COMMD1* and LDLR (Fig. 3c,d and Supplementary Fig. 6a). We found that this *COMMD1*–LDLR interaction depends on the NPxY-based trafficking motif within the cytoplasmic tail of LDLR and on

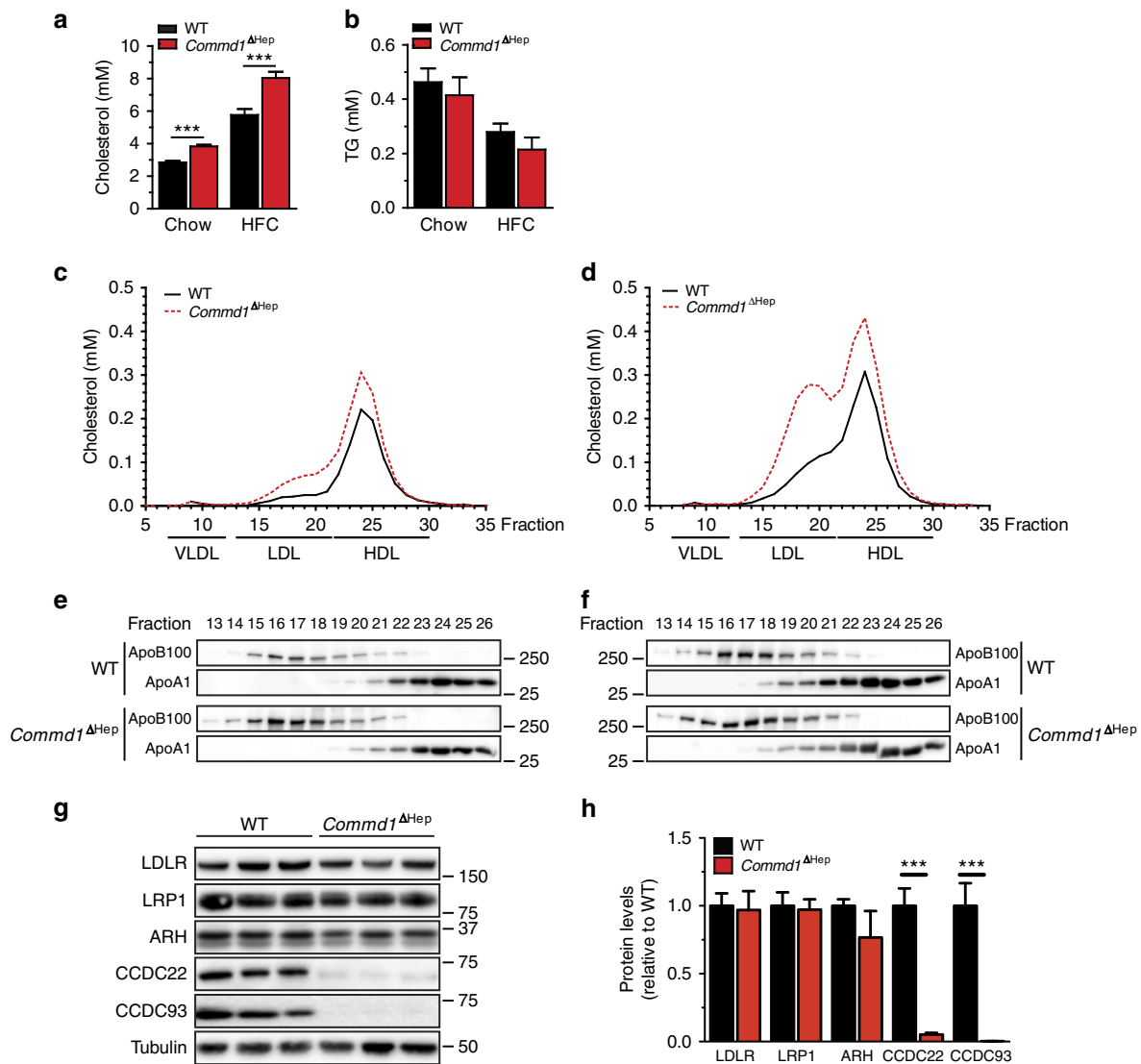


Figure 2 | Ablation of hepatic *CommD1* increases plasma total and LDL cholesterol levels. (a) Plasma TC and (b) TG levels of hepatic *CommD1* knockout mice (*CommD1*^{ΔHep}) and WT mice ($n = 6-8$) fed a chow diet or a HFC (0.2%) diet for 20 weeks. Pooled plasma samples of each experimental group of mice fed either the (c) chow or (d) the HFC diet were separated using FPLC gel filtration. Fifty fractions were collected from each separation. TC content was determined and lipoprotein profile was plotted. Fractions #13–26 containing cholesterol were collected and loaded on an SDS polyacrylamide gel and blotted using antibodies against apoA1 and apoB100 lipoproteins. The chow group (e) and HFC group (f) are shown. (g) Livers of chow-fed mice were homogenized, and 30 μ g of protein was subjected to immunoblot analysis. Levels of LDLR, LRP1, ARH, CCDC22, CCDC93 and tubulin were determined. Three representative samples from WT and *CommD1*^{ΔHep} mice are shown. (h) Relative levels of the proteins shown in g were determined by immunoblot analysis, and densitometry analysis was performed using tubulin as a loading control ($n = 6-8$ per genotype). The results are presented as mean \pm s.e.m., and significance was calculated relative to the control group by unpaired Student's *t*-test; *** $P < 0.001$. ARH, autosomal recessive hypercholesterolaemia protein.

the COMM domain of COMM1 (Fig. 3e and Supplementary Fig. 6b). Although additional studies are needed to determine whether the interaction between COMM1 and LDLR is direct, our results are consistent with previous studies, which demonstrated that the NPxY motif is essential for the binding of various LDLR adaptor proteins involved in the intracellular trafficking of LDLR²⁷.

WASH components colocalize and interact with LDLR.

Our prior work demonstrated that the CCC complex acts in conjunction with the multi-subunit WASH complex to mediate the trafficking of ATP7A (ref. 8), and this motivated us to assess the cellular distribution of the WASH complex and retromer

relative to LDLR. Isopycnic centrifugation of murine liver lysates showed a marked overlap in the cellular distribution of LDLR with WASH components (WASH1 and FAM21), as well as with CCC components (CCDC22 and COMM1), and to a lesser extent with the retromer component VPS35 (Fig. 3d). Furthermore, immunoprecipitation assays revealed that WASH1 and FAM21 can both bind to LDLR, but no physical association was detected between VPS35 and LDLR (Fig. 3f). In addition, the interaction between the PDZ-domain-containing sortin nexin 27 (SNX27) and LDLR was assessed as SNX27 is required for retrieval and recycling of numerous transmembrane proteins, including several members of the LDLR family (for example, VLDLR)²⁸. However, no interaction of LDLR and SNX27 was observed (Fig. 3f). Furthermore, the subcellular localization of

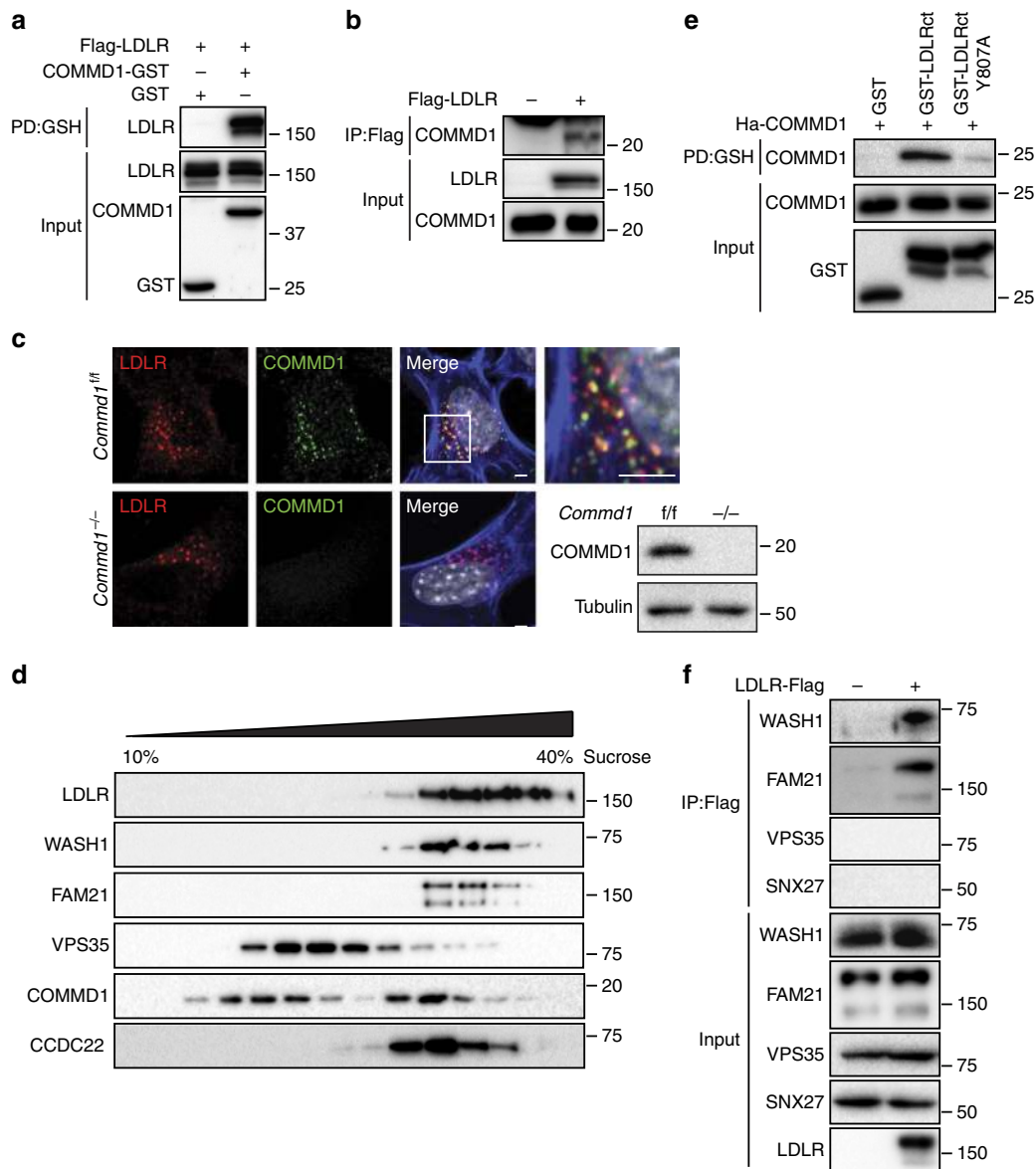


Figure 3 | LDLR associates with COMMD1 and the WASH complex. (a) Human embryonic kidney 293T (HEK293T) cells were transfected with constructs expressing Flag-LDLR with either COMMD1-GST or GST alone. Interaction with COMMD1 was detected via pull-down assay using glutathione sepharose beads. (b) HEK293T cells were transfected with Flag-LDLR vector, and interaction with endogenous COMMD1 was detected by immunoprecipitation with rabbit anti-Flag-antibody. (c) Colocalization of LDLR (red) and COMMD1 (green) in *Commd1^{f/f}* MEFs examined by immunofluorescence staining. Representative images are shown; scale bar, 5µm. LDLR (red) and COMMD1 (green) was stained in COMMD1-deficient MEFs (*Commd1^{-/-}*) and imaged by confocal fluorescence microscopy. COMMD1 levels in *Commd1^{f/f}* and in *Commd1^{-/-}* MEFs determined by immunoblot analysis. (d) Liver of a WT chow-fed mouse was homogenized and loaded on a continuous 10–40% sucrose gradient. Fractions were separated by ultracentrifugation and immunoblotted using antibodies against COMMD1, LDLR, WASH1, FAM21, VPS35 and CCDC22. The figure represents results of three independent experiments. (e) HEK293T cells were transfected with Ha-COMMD1 construct together with GST alone, GST-LDLRct (GST-tagged cytosolic domain of LDLR) or GST-LDLRct Y807A (GST-tagged mutated cytosolic domain of LDLR). Pull-down assay was performed to study the interaction between LDLRct and COMMD1. (f) Lysates of Flag-LDLR-transfected HEK293T cells were used for immunoprecipitation assays. Immunoprecipitates were washed, separated by SDS–polyacrylamide gel electrophoresis and immunoblotted as indicated.

SNX27 and its binding to FAM21 was not affected by COMMD1 deficiency (Supplementary Fig. 7). These observations suggest that the COMMD1-containing CCC complex acts in concert with the WASH complex to regulate LDLR endosomal trafficking.

COMMD1 and WASH facilitate LDLR trafficking. Next, we examined whether COMMD1 deficiency alters the subcellular localization of LDLR. As observed in HeLa cells⁸, immunofluorescence staining of COMMD1, WASH1 and

VPS35 demonstrated marked overlap in MEFs (Supplementary Fig. 8). In normal MEFs, LDLR is present in early endosomes, in EEA1⁺ and VPS35⁺ subdomains, as well as, to some extent, in LAMP1⁺ lysosomes. Loss of COMMD1 decreased the colocalization of LDLR with the early endosomal marker EEA1 and the lysosomal marker LAMP1, but increased LDLR localization to a VPS35-enriched endosomal compartment (Fig. 4a,b and Supplementary Fig. 9a), suggesting that LDLR might not be able to traffic normally from retromer-coated endosomes to other compartments. Next, we quantified surface

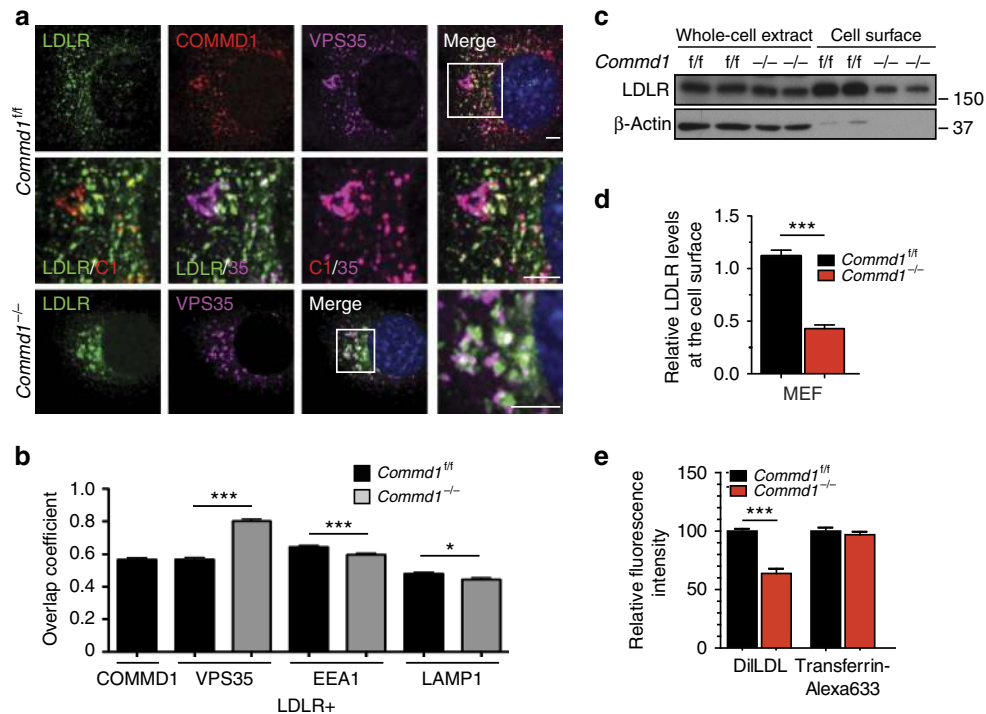


Figure 4 | COMMD1 deficiency impairs the function of LDLR. (a) LDLR (green), COMMD1 (red) and VPS35 (pink) were stained in *Commd1^{fl/fl}* and *Commd1^{-/-}* MEFs and imaged by confocal microscopy. Representative images are shown; scale bar, 5 μ m. (b) Quantification of the colocalization of LDLR with COMMD1, VPS35, EEA1 and LAMP1 was performed by the analysis of 30–40 cells. (c) Total and plasma membrane LDLR levels of *Commd1^{fl/fl}* and *Commd1^{-/-}* MEFs determined by biotinylation assay. Data represent three independent experiments, and (d) the relative levels of LDLR at the cell surface are quantified in all experiments. (e) *In vitro* LDL and transferrin uptake assay. Dil-labelled LDL (5 μ g ml⁻¹) or Alexa-633-labelled transferrin (5 μ g ml⁻¹) was added to serum-depleted medium and incubated with MEFs at 4 °C for 1 h and subsequently at 37 °C for 5 min. Dil-labelled LDL and Alexa-633-labelled transferrin uptake was measured by FACS analysis, and the relative uptake in triplicate is shown. The results are presented as mean \pm s.e.m.; significance was calculated relative to the control group by unpaired Student's *t*-test; **P* < 0.05, ****P* < 0.001.

levels of LDLR in WT and *Commd1^{-/-}* MEFs by using a surface biotinylation assay. Although total LDLR levels were not affected by the loss of COMMD1, cell surface levels of LDLR were markedly reduced (Fig. 4c,d). Decreased levels of LDLR was not only observed in *Commd1^{-/-}* MEFs but also in primary hepatocytes isolated from *Commd1^{ΔHep}* mice (Supplementary Fig. 9b,c). To assess the functional effect of LDLR mislocalization and decreased LDLR surface levels on LDL cellular uptake, we added fluorescently labelled LDL (DiI-LDL) to the medium and determined the level of endocytosed LDL. We observed a ~40% decrease in LDL uptake in *Commd1^{-/-}* MEFs compared with WT cells (Fig. 4e). To evaluate whether COMMD1 deficiency only affects LDL uptake, we also measured the uptake of labelled transferrin by the transferrin receptor (TfnR). Although the intracellular trafficking of TfnR shows many similarities to LDLR trafficking²⁹, no differences in transferrin uptake were observed between *Commd1^{-/-}* and WT cells (Fig. 4e). Altogether, these data indicate that COMMD1 is essential for maintaining proper levels of LDLR within the plasma membrane to clear LDL cholesterol.

Since the COMMD1-containing CCC complex is recruited to the endosomes by the WASH complex⁸, we assessed the contribution of the WASH complex on LDLR trafficking by determining the subcellular localization of LDLR in WASH1-deficient (*Wash1^{-/-}*) MEFs. *Wash1* ablation destabilizes the entire WASH complex³⁰, and in WASH-deficient MEFs the colocalization of LDLR and COMMD1 with VPS35, EEA1 and LAMP1 was increased (Fig. 5a–c and Supplementary Fig. 10). In addition, removal of WASH resulted in collapsed endosomal structures, as shown previously³⁰. Although COMMD1 levels

were not affected by WASH deficiency, LDLR protein levels were markedly reduced (Fig. 5d), but were restored by the treatment with the lysosome inhibitor bafilomycin A (Fig. 5e,f), as shown for epidermal growth factor receptor levels in WASH-deficient cells³⁰. In addition to a reduction in total LDLR levels (Fig. 5d), LDLR levels at the cell surface were decreased in *Wash1^{-/-}* MEFs, as demonstrated by a surface biotinylation assay (Fig. 5g,h). To assess whether mislocalization and reduced levels of LDLR in *Wash1^{-/-}* cells affect LDL uptake, DiI-LDL was added to the media. DiI-LDL uptake was visualized by microscopy and subsequently quantified. WASH1 depletion markedly diminished LDL cellular uptake (Fig. 5i,j). It has been demonstrated that WASH-mediated actin polymerization on the endosomal membrane is crucial for endosomal sorting of specific cargos, and loss of WASH1 results in endosomal collapse (Fig. 5a) (ref. 30). Indeed, restoring WT WASH1 in WASH1-deficient cells rescued the accumulation of LDLR in collapsed endosomes, which could not be prevented by expressing WASH1- Δ VCA, a deletion mutant that cannot activate the Arp2/3 complex (Supplementary Fig. 11). Altogether, these results indicate that WASH-mediated F-actin polymerization on endosomes is essential for the endosomal trafficking of LDLR.

A homozygous splice site mutation in *KIAA0196*, which encodes the WASH component strumpellin, causes Ritscher–Schinzel/3C syndrome (RSS) in a Canadian cohort³¹. This mutation results in a 60% reduction in strumpellin levels³¹, and RSS patients share many clinical features with patients with *CCDC22* mutations, described thus far^{18,31}. We assessed the plasma cholesterol levels of one molecularly confirmed RSS patient (age 25), and found that the *KIAA0196* mutation was also

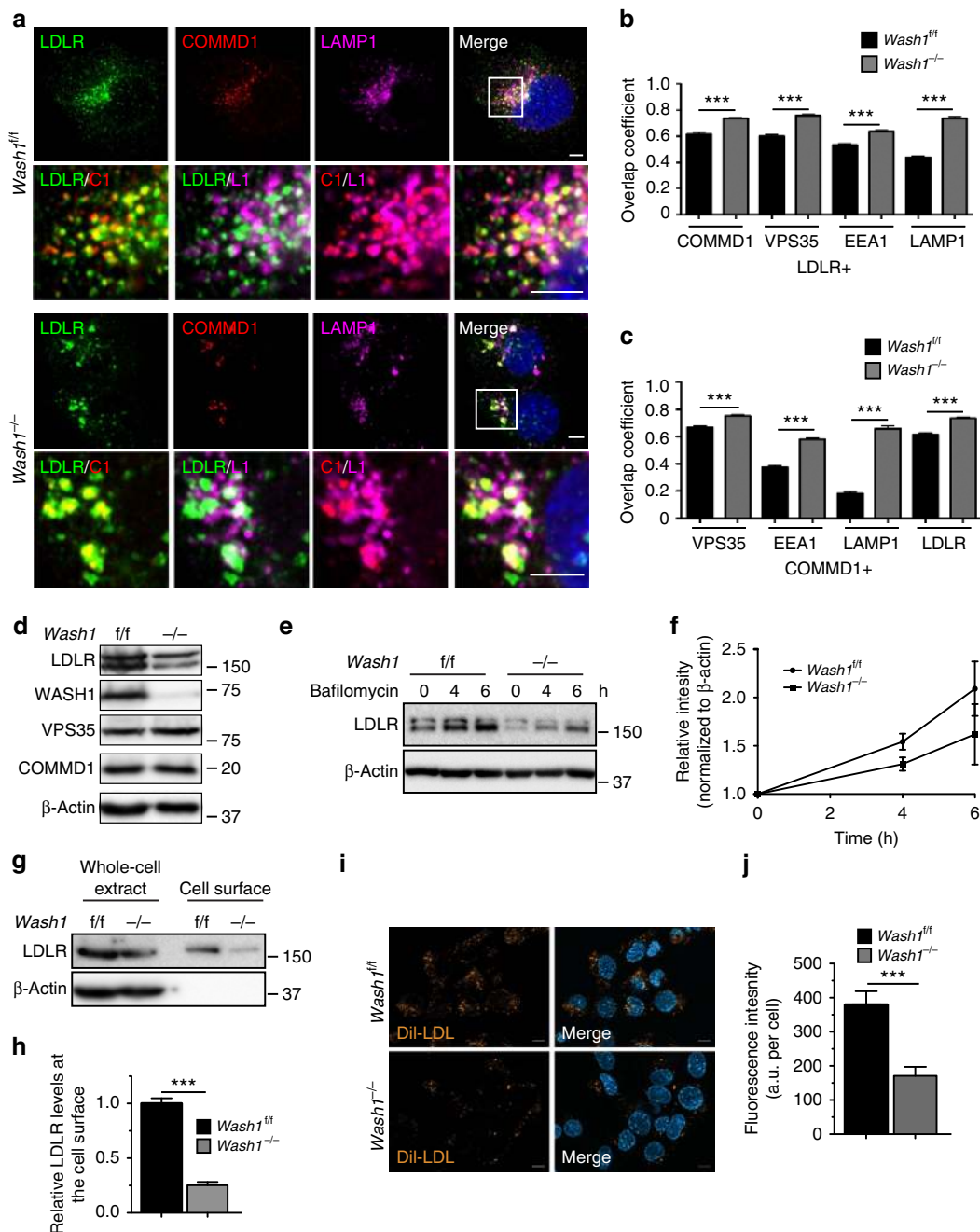


Figure 5 | The WASH complex is essential for endosomal sorting of LDLR. (a) Cellular localization of LDLR (green), COMMD1 (red) and LAMP1 (pink) in *Wash1^{f/f}* and *Wash1^{-/-}* MEFs was determined by immunofluorescence staining. Representative images are shown; scale bar, 5 μ m. Relative colocalization of (b) LDLR and (c) COMMD1 with VPS35, EEA1 and LAMP1 was quantified. (d) LDLR, WASH1, VPS35 and COMMD1 levels in *Wash1^{f/f}* and *Wash1^{-/-}* MEFs analysed by western blot. (e) Representative images ($n=3$) of immunoblot analysis of total LDLR levels in *Wash1^{f/f}* and *Wash1^{-/-}* MEFs treated with bafilomycin A (100 nM) for 0, 4 and 6 h. (f) Densitometry revealed the relative levels of LDLR in bafilomycin A-treated cells ($n=3$). (g) Representative images ($n=3$) of LDLR on the surface of *Wash1^{f/f}* and *Wash1^{-/-}* MEFs determined by surface biotinylation assay. (h) Densitometry revealed relative LDLR surface levels ($n=3$). (i) *Wash1^{f/f}* and *Wash1^{-/-}* MEFs were incubated with DiI-LDL for 30 min and imaged by fluorescence microscope. (j) Fluorescence intensity was quantified using ImageJ software and was normalized to the number of DAPI nuclei per image; >30 cells per condition were recorded. The results are presented as mean \pm s.e.m.; significance was calculated relative to the control group by unpaired Student's *t*-test; *** $P < 0.001$.

associated with elevated plasma TC and LDL cholesterol levels exceeding the 95th percentile corrected for age and gender (TC = 6.5 mmol l⁻¹, HDL = 1.5 mmol l⁻¹, LDL = 4.4 mmol l⁻¹ and TG = 1.2 mmol l⁻¹), comparable to the cholesterol increase seen in patients with *CCDC22* mutations (Table 1). These data indicate that both the CCC and WASH complexes regulate

homeostasis of circulating cholesterol through intracellular trafficking of LDLR.

Discussion

The LDL-LDLR axis is a potential therapeutic target for lowering atherogenic plasma cholesterol³², and the intracellular route of

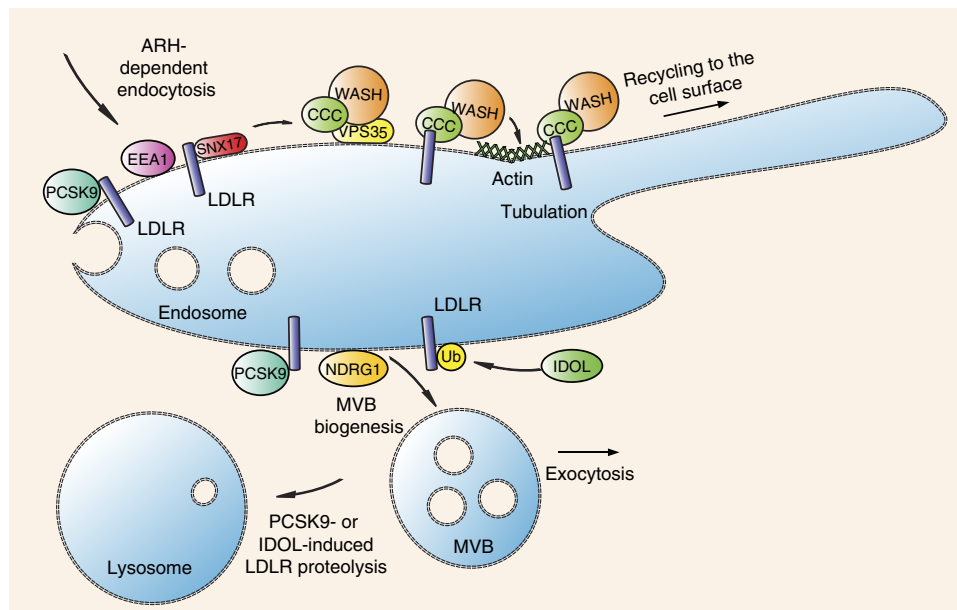


Figure 6 | Hypothetical model of CCC- and WASH-mediated endosomal LDLR sorting. After internalization mediated by ARH, LDLR is sorted in endosomes⁵⁸. LDLR is directed to the lysosome for proteolysis—which is induced by either PCSK9 (refs 59,60) or IDOL⁶¹—or retrieved and recycled back to the cell surface. SNX17 enhances LDL endocytosis likely by promoting LDLR retrieval⁴⁴, which might precede CCC and retromer/WASH-mediated endosomal trafficking. Through the interaction of the retromer component VPS35 with FAM21 the WASH complex and the CCC complex are recruited to the endosomes^{2,4,8}. Subsequently, CCC and WASH form a protein complex with LDLR; WASH facilitates the formation of branched actin patches to define restricted domains of the endosomes⁶² from where LDLR is sorted back to the cell surface. NDRG1 is required for the formation of MVB: loss of NDRG1 impairs LDL uptake and LDLR recycling, and causes the accumulation of IDOL-mediated ubiquitinated LDLR in MVB⁴⁸. Further research is needed to assess whether CCC and WASH are functionally related to SNX17, NDRG1 and to the lysosomal degradation pathways of LDLR. ARH, autosomal recessive hypercholesterolaemia protein; MVB, multivesicular body; PCSK9, proprotein convertase subtilisin/kexin type 9; Ub, ubiquitin; WASH, WASH1, FAM21, strumpellin, KIAA1033, CCDC53.

LDLR has been well described³³. However, the mechanisms regulating the trafficking of LDLR are not completely understood³⁴. Here we identified that the multi-subunit protein complexes CCC and WASH are required for LDLR endosomal sorting and for directing the receptor to the plasma membrane (Fig. 6). Mutations affecting the formation of these complexes are causally related to hypercholesterolaemia in humans, dogs and mice.

COMMD1 was initially identified as a regulator of copper homeostasis in mammals^{11,26}; however, here we uncover that COMMD1 is also critically important in cholesterol homeostasis. According to our previous work revealing that COMMD1 and the CCC complex regulate the endosomal sorting of the copper transporter ATP7A (ref. 8), we found that this same system similarly regulates the endosomal trafficking of LDLR. We demonstrated this in a variety of cellular models, and the physiologic importance of our observations has been substantiated in two independent mammalian models of COMMD1 deficiency, which display increased plasma LDL cholesterol. In agreement with these models, deleterious human mutations in *CCDC22* (refs 8,13,18) are causally linked to elevated plasma total and LDL cholesterol levels in affected patients. The atherogenic lipoprotein phenotype of these patients is similar to that of patients with familial hypercholesterolaemia caused by heterozygous *LDLR* mutations³⁵, but cholesterol levels do not reach the high levels seen in patients with homozygous mutations in either *LDLR*, *ARH* (autosomal recessive hypercholesterolaemia) or *APOB* (apolipoprotein B), or in patients with a gain-of-function mutation in *PCSK9* (proprotein convertase subtilisin/kexin type 9) (ref. 35). In addition, elevated TC and LDL cholesterol levels were found in a RSS patient, with a homozygous mutation in the *KIAA0196* gene³¹—encoding for a

component of the WASH complex—thus indicating that WASH dysfunction might also cause aberrant cholesterol homeostasis. Therefore, mutations in the *VPS35* gene associated with Parkinson's disease^{36,37}, and in components of the WASH complex, which are linked to autosomal recessive intellectual disability, and spastic paraplegia^{38–40}, warrant future attention with regards to their effects on cholesterol homeostasis.

In endosomal cargo sorting WASH mediates the F-actin formation on endosomes (Fig. 6) (ref. 3), whereas the CCC complex is likely involved in cargo recognition, in which the composition of the CCC complex defines which cargo is specifically sorted. This line of thought is reinforced by the observation that all 10 members of the COMMD family can engage in the CCC complex^{13,41}, and by our recent work showing that the trafficking of Notch, another cargo of this system, is particularly dependent on COMMD9 in various cell lines¹⁴. Although these observations advocate for the presence of CCC subcomplexes in cargo selectively trafficking, here we found that deletion of *Commd9* in hepatocytes also causes increased plasma cholesterol in mice, which indicates that the composition of these subcomplexes and their roles in cargo trafficking are likely cell type specific. Further research is therefore warranted to dissect the contribution of the COMMD proteins in cargo selectively trafficking.

The retromer–WASH-associated SNX27 (ref. 42) interacts with the NPxY-based trafficking motif⁴³, and here we assessed whether SNX27 is engaged in CCC/WASH-mediated LDLR trafficking. In agreement with prior work²⁸, we did not find SNX27 in the interactome of LDLR. Furthermore, ablation of COMMD1 did not affect SNX27–FAM21 interaction or subcellular localization of SNX27. Our results and the findings of Steinberg *et al.*²⁸ showing that the depletion of SNX27 in HeLa cells does not affect

the levels of LDLR on the cell surface—as shown for other members of the LDLR family—indicate that it is unlikely that SNX27 is involved in LDLR trafficking. Interestingly, the SNX27 family member SNX17 has been identified as an adaptor of LDLR. Like COMMD1, SNX17 binds to the NPxY motif of LDLR^{44–47} and promotes LDLR endocytosis⁴⁴. SNX17 and COMMD1 both colocalize to vesicles positive for the early endosome marker EEA1 (refs 12,44), but in contrast to COMMD1, localization studies did not demonstrate overlap between SNX17 and LDLR⁴⁴. Therefore, we hypothesize that the action of SNX17 on LDLR trafficking precedes CCC- and WASH-mediated endosomal sorting of LDLR (Fig. 6). In addition to SNX17, N-myc downstream regulated gene 1 (NDGR1) also regulates the trafficking of LDLR⁴⁸. Loss of NDRG1 reduces LDLR levels on the cell surface accompanied by impaired LDL uptake. In addition, cells deficient for NDRG1 display accumulation of inducible degrader of the LDLR (IDOL)-mediated ubiquitinated LDLR in multivesicular bodies, whose structure is deformed by NDRG1 deficiency (Fig. 6) (ref. 48). Whether CCC and WASH complexes are functionally connected to the aforementioned proteins or to other regulators of the LDLR recycling³⁴ warrants further investigation.

Altogether, this study uncovers CCC and WASH as novel multi-subunit protein complexes in the endosomal trafficking of LDLR, and provides valuable insights in the molecular mechanism of cholesterol homeostasis.

Methods

Human studies. All participating patients and their family members consented to the evaluations performed. Their participation in this study was approved by the appropriate institutional review boards (Women's and Children's Health Network, Adelaide, South Australia, Australia and Medical University Graz, Graz, Austria), as previously reported^{13,31}.

Animals. All animal studies were approved by the Institutional Animal Care and Use Committee, University of Groningen (Groningen, the Netherlands). Hepatocyte-specific *Commd1* knockout mice (*Commd1*^{ΔHep})²⁶ were backcrossed for more than eight generations in a C57BL/6J background. All mice were individually housed males, fed *ad libitum* with either a standard rodent chow diet (RMH-B, AB Diets, the Netherlands) or, starting at 8–9 weeks of age, a HFC diet (45% calories from butter fat) containing 0.2% cholesterol (SAFE Diets), *n* = 6–8. HFC feeding lasted for 20 weeks. *Commd1*^{loxP/loxP} mice¹⁴ were retro-orbitally injected with purified 1×10^{11} Ad-Cre or Ad-LacZ virus particles⁴⁹. Mice were killed following a 4-h morning fasting period. Tissues for mRNA and protein expression analysis were snap frozen in liquid nitrogen and stored at -80°C until further analysis. Blood was drawn by heart puncture, and plasma was isolated by centrifugation at 3,000 r.p.m. for 10 min at 4°C .

Dog samples were partly originated from earlier studies^{19,22}. Client owned Labrador retrievers were admitted to the Department of Clinical Sciences of Companion Animals, Faculty of Veterinary Medicine, Utrecht University (Utrecht, the Netherlands). Ultrasound-guided liver biopsies were obtained for diagnostic purposes from dogs under general anaesthesia. Copper concentrations in liver biopsies were measured using instrumental neutron activation analysis. Serum samples that were obtained for diagnostic purposes from Labrador retrievers were used for the current study, after informed consent was obtained from the owners. The procedures were approved by Utrecht University Ethical Committee, as required under Dutch legislation.

Cholesterol and TG analysis in plasma and liver homogenates. TC levels were determined using colorimetric assay (11489232, Roche Molecular Biochemicals) with cholesterol standard FS (DiaSys Diagnostic Systems) as a reference. TG levels were determined using Trig/GB kit (1187771, Roche Molecular Biochemicals) with Roche Precimat Glycerol standard (16658800) as a reference.

Hepatic lipid extraction. Liver homogenates prepared as 15% (w/v) solutions in PBS were subjected to lipid extraction according to the Bligh & Dyer method⁵⁰. In short, 600 μl of demi-water was mixed with 200 μl of liver homogenate, and 3 ml of chloroform/methanol was added and mixed. After 30 min of incubation 1.2 ml of H_2O and 1 ml of chloroform were added, mixed and subsequently centrifuged (10 min, 1,500 r.p.m. at room temperature). The chloroform layer was transferred into a new glass tube, and was evaporated using nitrogen at 50°C . Lipids were resolved in 1 ml of chloroform and used for further determination of cholesterol and TG content.

In vivo VLDL-TG production. The experiment was performed on 10- to 13-week-old chow-fed mice. After a 4-h morning fasting, animals were intraperitoneally injected with poloxamer 407 (BASF) solution in saline (1 g per kg body weight). Blood was drawn by retro-orbital puncture at the following time points: 0, 30, 60, 120 and 240 min. These samples were used for TG determination and calculation of VLDL-TG production rate.

Antibodies. In the experimental procedures described, the following antibodies were used: rabbit polyclonal antibody against COMMD1 (11938-1-AP, Proteintech Group, 1:1,000); rabbit polyclonal antibody against LDLR (PAB8804, Abnova, 1:1,000); rabbit polyclonal antibody against GST (Z-5) (sc-459, Santa Cruz Biotechnology, 1:5,000); goat anti-rabbit IgG (H + L)-horseradish peroxidase (HRP) conjugate (170-6515, Bio-Rad Laboratories, 1:10,000); goat anti-mouse IgG (H + L)-HRP conjugate (170-6516, Bio-Rad Laboratories, 1:10,000); mouse anti- β -actin (A5441, Sigma-Aldrich, 1:5,000); rabbit anti-tubulin (AB4047, Abcam, 1:2,000); rabbit anti-Rab11 (700184, Invitrogen, 1:1,000); rabbit anti-EEA-1 (AB2900, Abcam, 1:1,000); rabbit anti-CCDC22 (16636-1-AP, Proteintech, 1:2,000); and rabbit anti-CCDC93 (20861-1-AP, Proteintech, 1:5,000). Rabbit polyclonal antibodies against WASH1, FAM21 and VPS35 were previously described^{1,5}. Anti-apoB100 (1:1,000) and rabbit anti-apoA1 (1:1,000) antibodies were a gift from A.K. Groen. Rabbit polyclonal antibody against ARH (1:1,000) was a gift from H.H. Hobbs. Densitometry analysis of western blot bands was performed using Image Lab 3.0.1 software (Bio-Rad Laboratories). Images have been cropped for presentation. Full-size images are presented in Supplementary Fig. 12.

Expression constructs. The following vectors were used in the experiments described: pEBB-COMMD1-Flag; pEBB-GST; pEBB-COMMD1-GST; pEBB-1-118-GST; and pEBB-119-190-GST^{10,51}. Full-length Flag-tagged LDLR receptor, GST-LDL cytoplasmic tail (-cyt) and GST-LDL-cyt Y807A were obtained from N. Freedman⁵², and Flag-LDLR was subcloned into pDNA3.1. WT and mutant WASH1-GFP constructs have been described before⁵⁰.

Immunofluorescence staining. Staining was carried out as previously described¹. Cells were fixed in ice-cold fixative (4% paraformaldehyde and 0.5% glutaraldehyde in PBS) and incubated for 18 min at room temperature in the dark, followed by permeabilization with 0.2% Triton X-100 in PBS for 4 min. Cells were then cultured with primary antibodies in IF buffer (TBS plus human serum cocktail) overnight at 4°C in a humidifier chamber. Next day, cells were washed three times in PBS, and cells were incubated with secondary antibodies in blocking buffer for 1 h at room temperature. After four washes in PBS and addition of Hoechst 33342 nuclear stain, coverslips were rinsed in water and affixed to slides with Slowfade Antifade reagent (Invitrogen). Images were obtained with an LSM-710 laser scanning confocal microscope with a $\times 100/1.4$ Oil Plan-Apochromat objective lens using ZEN software (Carl Zeiss). Quantitative Pearson's correlation coefficients were obtained using Zen 2009 software (Carl Zeiss) by drawing a region of interest around the objects. The following antibodies were used: rabbit polyclonal antibody against LDLR (PAB8804, Abnova, 1:200); mouse monoclonal antibody against COMMD1 (MAB7526, R&D Systems, 1:400); rabbit polyclonal antibody against EEA1 (2411, Cell Signaling Technologies, 1:250); goat polyclonal antibody against VPS35 (ab10099-100, Abcam, 1:1,000); rat monoclonal antibody against LAMP1 (553792, BD Biosciences, 1:500); rabbit polyclonal antibody against WASH1 (1:1,000); and rabbit polyclonal antibody against FAM21 (1:1,000) (ref. 1).

Gene expression analysis. Pieces of murine liver of ~ 100 mg were homogenized in 1 ml QIAzol Lysis Reagent (Qiagen, Venlo, the Netherlands). Total RNA was isolated by chloroform extraction. Isopropanol-precipitated and ethanol-washed RNA pellets were dissolved in RNase-/DNase-free water. RNA (1 μg) was used to prepare cDNA with the Quantitect Reverse Transcription Kit (Qiagen), according to the manufacturer's protocol. cDNA (20 ng) was used for subsequent quantitative real-time PCR analysis using iTaq SYBR Green Supermix with ROX (Bio-Rad Laboratories BV) and 7900HT Fast Real-Time PCR Systems (Applied Biosystems). The following programme was used: $50^{\circ}\text{C}/2$ min; $95^{\circ}\text{C}/10$ min; 40 cycles with $95^{\circ}\text{C}/15$ s; and $60^{\circ}\text{C}/1$ min. Expression data were analysed using SDS 2.3 software (Applied Biosystems) and the standard curve method of calculation. Mouse cyclophilin A was used as an internal control gene. Primers used for the expression studies are listed in Supplementary Table 3.

Fast-performance liquid chromatography. Plasma samples within each murine or canine experimental group were pooled together and fractionated using the fast-performance liquid chromatography (FPLC) method. All 50 fractions were analysed to determine TC and TG content. Fractions containing LDL and HDL were further analysed by immunoblot using anti-apoA1 (1:1,000) and anti-apoB100 antibodies (1:1,000).

The TC distribution among the main lipoprotein classes of the individual plasma samples was measured using FPLC analysis as described previously but with some minor modifications⁵³. In brief, the system contained a PU-980 ternary pump with an LG-980-02 linear degasser and an UV-975 UV/VIS detector (Jasco).

An extra PU 2080i-plus pump (Jasco) was used for in-line cholesterol ready-to-use enzymatic reagent (Biomerieux) addition at a flow rate of 0.1 ml min⁻¹. EDTA plasma was diluted 1:1 with Tris-buffered saline, and 30 µl sample buffer mixture was loaded on a Superose 6 HR 10/30 column (GE Healthcare) for lipoprotein separation at a flow rate of 0.31 ml min⁻¹. Chromatographic profiles of commercially available plasma lipid standards (SKZL) served as reference.

Cell culture. Human embryonic kidney 293T cells (American Type Culture Collection) and MEFs^{30,34} were cultured using Dulbecco's Modified Eagles Medium (DMEM) with Glutamax (GIBCO), supplemented with 10% fetal bovine serum (Invitrogen) and antibiotics. Primary mouse hepatocytes were cultured in DMEM—low glucose (GIBCO), supplemented with 5% fetal calf serum (FCS) or normal calf lipoprotein-poor serum (NCLPPS) and antibiotics.

Sucrose gradients. Sucrose gradient separation of fractions obtained from fresh liver tissue was performed as described before⁵⁵ with few modifications. Chow-fed mice were fasted for 4 h before killing. A unit of 200 mg of liver was homogenized in 800 µl homogenization buffer (50 mM Tris-HCl, pH 7.4, 250 mM sucrose, 25 mM KCl, 5 mM MgCl₂, 3 mM imidazole and protease inhibitor mixture) using a Douncer homogenizer. Homogenates were centrifuged 1,000g, 10 min at 4 °C. A unit of 300 µg liver homogenate was loaded on a 3.7-ml continuous 10–40% sucrose gradient, and were centrifuged for 16 h using a Beckman Coulter ultracentrifuge equipped with a swinging bucket rotor SW55 Ti, at 40,000 r.p.m., and fractions of 285 µl were collected from the top of the tube; 1/10 of each fraction was mixed with SDS sample buffer and used for immunoblot analysis.

Biotinylation assay. Culture of primary hepatocytes and biotinylation assay were performed as previously described⁵⁶. Cells were washed (3 ×) with ice-cold PBS-CM buffer (PBS, 1 mM MgCl₂ and 0.1 mM CaCl₂), subsequently 0.5 mg ml⁻¹ biotin reagent solution (EZ-Link Sulfo-NHS-SS-Biotin, Thermo Scientific) in biotinylation buffer (10 mM triethanolamine, pH 8.0, 150 mM NaCl and 2 mM CaCl₂) was added to the cells for 30 min at 4 °C. Biotin reagent was removed and cells were washed 1 × with quenching buffer (PBS-CM, 25 mM Tris-HCl, pH 7.4 and 192 mM Glycine) for 30 min at 4 °C. Next, cells were washed 2 × with PBS-CM and 1 × with TBS-C (50 mM Tris-Cl, pH 7.4, 100 mM NaCl and 2 mM CaCl₂), and cells were collected by scraping in TBS-C. Cells were centrifuged (1,000g, 5 min, 4 °C), lysed in biotin lysis buffer (50 mM Tris-Cl, pH 7.4, 150 mM NaCl, 1% NP-40, 0.5% sodium deoxycholate, 5 mM EDTA and 5 mM EGTA), sonicated (10% output for 10 s) and incubated on ice for 15 min, subsequently centrifuged for 15 min at 12,000 g. Protein concentration was determined; 30 µg was used as input; 300 µg was used diluted in biotin lysis buffer (500 µl); 30 µl Neutravidin beads (Neutravidin plus ultralink beads, Thermo Scientific) were added and incubated for 4 h at 4 °C. Beads were collected by centrifugation (500g, 5 min), beads were washed 3 × with Biotin lysis buffer, 1 × with high-salt buffer (50 mM Tris-Cl, pH 7.4 and 500 mM NaCl) and 1 × with low-salt buffer (10 mM Tris-HCl). Finally the beads were resuspended in 30 µl 2 × loading buffer.

Immunoprecipitation analysis. The glutathione S-transferase (GST) pull-down assays and immunoprecipitation experiments were performed as described before⁵⁷. Cells were mainly lysed in NP-40 buffer (0.4 M NaCl, 0.1% NP-40, 10 mM Tris-HCl, pH 8.0 and 1 mM EDTA) supplemented with protease inhibitors. Immunoprecipitation of the WASH complex and retromer was performed in MRB lysis buffer (20 mM HEPES pH 7.2, 50 mM potassium acetate, 1 mM EDTA, 200 mM D-sorbitol and 0.1% Triton X-100) supplemented with protease inhibitors. Equal amount of protein was used for immunoprecipitation and pull-down assays. FLAG M2 agarose beads (Sigma-Aldrich) were used for Flag-immunoprecipitation experiments, Glutathione Sepharose 4B (GE Healthcare Life Sciences) for GST pull-down assays. Flag-, Ha-, GST-tagged proteins were detected by Flag-antibody (F1804, Sigma, 1:2,000), Ha-antibody (clone HA-7, H6533, Sigma-Aldrich, 1:5,000) and GST-antibody (sc-138, Santa Cruz, 1:5,000), respectively.

Dil-LDL uptake assay. Culture medium was replaced with DMEM culture medium supplemented with Dil-LDL (5 µg ml⁻¹; Molecular Probes, Invitrogen). Cells were kept for 1 h at 4 °C, then for 5 min at 37 °C. Then they were immediately placed on ice, washed with cold PBS and scraped. As a control for the specificity of the investigated uptake pathway, cells were incubated with Alexa633-transferrin (5 µg ml⁻¹). Cells were centrifuged at 300g for 5 min at 4 °C and resuspended in 50 µl of FACS buffer (PBS with 2% FCS and 5 mM EDTA) supplemented with additional 5% fetal bovine serum. Cell pellets were vortexed, and 2 ml of FACS buffer was added; samples were centrifuged as before and resuspended in 200 µl of FACS buffer. Cells were kept on ice at all times and subjected to immediate FACS analysis. The number of positive cells was counted and recorded as a percentage of the whole population. Data of four independent experiments were presented relative to the uptake results of the control WT population (set as 100%). Dil-LDL uptake in MEFs determined by fluorescence microscopy was performed with MEFs cultured for 16 h in DMEM medium supplemented with lipoprotein-deficient serum (10%). Medium was changed with Dil-LDL (5 µg ml⁻¹) containing 5% lipoprotein-deficient serum/DMEM medium, and after 30 min cells were fixed and

mounted with Vectashield mounting medium with DAPI (Vector Laboratories). Images were acquired with a Zeiss AxioObserver.Z1. Fluorescence intensity was quantified using ImageJ software (National Institutes of Health) and was normalized to the number of DAPI nuclei per image; >30 cells per condition were recorded.

Statistical analysis. All results are expressed as a mean ± s.e.m. Statistical analysis was performed using Prism 5.00 for Windows (GraphPad Software) and the unpaired Student's *t*-test. Results of *P* < 0.05 were considered statistically significant: **P* < 0.05, ***P* < 0.01 and ****P* < 0.001.

References

- Gomez, T. S. & Billadeau, D. D. A FAM21-containing WASH complex regulates retromer-dependent sorting. *Dev. Cell* **17**, 699–711 (2009).
- Harbour, M. E., Breusegem, S. Y. & Seaman, M. N. J. Recruitment of the endosomal WASH complex is mediated by the extended 'tail' of Fam21 binding to the retromer protein Vps35. *Biochem. J.* **442**, 209–220 (2012).
- Seaman, M. N. J., Gautreau, A. & Billadeau, D. D. Retromer-mediated endosomal protein sorting: all WASHed up! *Trends Cell Biol.* **23**, 522–528 (2013).
- Harbour, M. E. *et al.* The cargo-selective retromer complex is a recruiting hub for protein complexes that regulate endosomal tubule dynamics. *J. Cell. Sci.* **123**, 3703–3717 (2010).
- Jia, D., Gomez, T. S., Billadeau, D. D. & Rosen, M. K. Multiple repeat elements within the FAM21 tail link the WASH actin regulatory complex to the retromer. *Mol. Biol. Cell* **23**, 2352–2361 (2012).
- Helfer, E. *et al.* Endosomal recruitment of the WASH complex: active sequences and mutations impairing interaction with the retromer. *Biol. Cell* **105**, 191–207 (2013).
- Cullen, P. J. & Korswagen, H. C. Sorting nexins provide diversity for retromer-dependent trafficking events. *Nat. Cell Biol.* **14**, 29–37 (2012).
- Phillips-Krawczak, C. A. *et al.* COMMD1 is linked to the WASH complex and regulates endosomal trafficking of the copper transporter ATP7A. *Mol. Biol. Cell* **26**, 91–103 (2014).
- Freeman, C. L., Hesketh, G. & Seaman, M. N. J. RME-8 coordinates the activity of the WASH complex with the function of the retromer SNX dimer to control endosomal tubulation. *J. Cell Sci.* **127**, 2053–2070 (2014).
- Burstein, E. *et al.* COMMD proteins, a novel family of structural and functional homologs of MURR1. *J. Biol. Chem.* **280**, 22222–22232 (2005).
- van de Sluis, B., Rothuizen, J., Pearson, P. L., van Oost, B. A. & Wijnga, C. Identification of a new copper metabolism gene by positional cloning in a purebred dog population. *Hum. Mol. Genet.* **11**, 165–173 (2002).
- Fedoseenko, A., Bartuzi, P. & van de Sluis, B. Functional understanding of the versatile protein copper metabolism MURR1 domain 1 (COMMD1) in copper homeostasis. *Ann. N Y Acad. Sci.* **1314**, 6–14 (2014).
- Starokadomskyy, P. *et al.* CCDC22 deficiency in humans blunts activation of proinflammatory NF-κB signaling. *J. Clin. Invest.* **123**, 2244–2256 (2013).
- Li, H. *et al.* Endosomal sorting of Notch receptors through COMMD9-dependent pathways modulates Notch signaling. *J. Cell Biol.* **211**, 605–617 (2015).
- Porkka, K. V., Viikari, J. S., Rönnemaa, T., Marniemi, J. & Akerblom, H. K. Age and gender specific serum lipid and apolipoprotein fractions of Finnish children and young adults. The Cardiovascular Risk in Young Finns Study. *Acta Paediatr.* **83**, 838–848 (1994).
- Yip, P. M. *et al.* Pediatric reference intervals for lipids and apolipoproteins on the VITROS 5,1 FS Chemistry System. *Clin. Biochem.* **39**, 978–983 (2006).
- Wiegman, A. *et al.* Familial hypercholesterolaemia in children and adolescents: gaining decades of life by optimizing detection and treatment. *Eur. Heart J.* **36**, 2425–2437 (2015).
- Kolanczyk, M. *et al.* Missense variant in CCDC22 causes X-linked recessive intellectual disability with features of Ritscher-Schinzel/3C syndrome. *Eur. J. Hum. Genet.* **23**, 720 (2015).
- Favier, R. P. *et al.* COMMD1-deficient dogs accumulate copper in hepatocytes and provide a good model for chronic hepatitis and fibrosis. *PLoS ONE* **7**, e42158 (2012).
- Bailhache, E. *et al.* Metabolism of cholesterol ester of apolipoprotein B100-containing lipoproteins in dogs: evidence for disregarding cholesterol ester transfer. *Eur. J. Clin. Invest.* **34**, 527–534 (2004).
- Huster, D. *et al.* High copper selectively alters lipid metabolism and cell cycle machinery in the mouse model of Wilson disease. *J. Biol. Chem.* **282**, 8343–8355 (2007).
- Fieten, H. *et al.* Association of dietary copper and zinc levels with hepatic copper and zinc concentration in Labrador retrievers. *J. Vet. Intern. Med.* **26**, 1274–1280 (2012).
- Fieten, H., Leegwater, P. A. J., Watson, A. L. & Rothuizen, J. Canine models of copper toxicosis for understanding mammalian copper metabolism. *Mamm. Genome* **23**, 62–75 (2011).

24. Jessica Seessle *et al.* Alterations of lipid metabolism in Wilson disease. *Lipids Health Dis.* **10**, 83 (2011).
25. Osono, Y., Woollett, L. A., Herz, J. & Dietschy, J. M. Role of the low density lipoprotein receptor in the flux of cholesterol through the plasma and across the tissues of the mouse. *J. Clin. Invest.* **95**, 1124–1132 (1995).
26. Vonk, W. I. M. *et al.* Liver-specific *Comm1d1* knockout mice are susceptible to hepatic copper accumulation. *PLoS ONE* **6**, e29183 (2011).
27. Michaely, P. *et al.* Identification of a VLDL-induced, FDNVPVY-independent internalization mechanism for the LDLR. *EMBO J.* **26**, 3273–3282 (2007).
28. Steinberg, F. *et al.* A global analysis of SNX27-retromer assembly and cargo specificity reveals a function in glucose and metal ion transport. *Nat. Cell Biol.* **15**, 461–471 (2013).
29. Du, L. Macrophage colony-stimulating factor differentially regulates low density lipoprotein and transferrin receptors. *J. Lipid Res.* **45**, 1733–1740 (2004).
30. Gomez, T. S., Gorman, J. A., Artal-Martinez de Narvajias, A., Koenig, A. O. & Billadeau, D. D. Trafficking defects in WASH-knockout fibroblasts originate from collapsed endosomal and lysosomal networks. *Mol. Biol. Cell* **23**, 3215–3228 (2012).
31. Elliott, A. M. *et al.* A novel mutation in KIAA0196: identification of a gene involved in Ritscher-Schinzel/3C syndrome in a First Nations cohort. *J. Med. Genet.* **50**, 819–822 (2013).
32. Brautbar, A. & Ballantyne, C. M. Pharmacological strategies for lowering LDL cholesterol: statins and beyond. *Nat. Rev. Cardiol.* **8**, 253–265 (2011).
33. Brown, M. S. & Goldstein, J. L. A receptor-mediated pathway for cholesterol homeostasis. *Science* **232**, 34–47 (1986).
34. Wijers, M., Kuivenhoven, J. A. & van de Sluis, B. The life cycle of the low-density lipoprotein receptor: insights from cellular and in-vivo studies. *Curr. Opin. Lipidol.* **26**, 82–87 (2015).
35. Cuchel, M. *et al.* Homozygous familial hypercholesterolaemia: new insights and guidance for clinicians to improve detection and clinical management. A position paper from the Consensus Panel on Familial Hypercholesterolaemia of the European Atherosclerosis Society. *Eur. Heart J.* **35**, 2146–2157 (2014).
36. Zavodszky, E. *et al.* Mutation in VPS35 associated with Parkinson's disease impairs WASH complex association and inhibits autophagy. *Nat. Commun.* **5**, 3828 (2010).
37. Vilariño-Güell, C. *et al.* VPS35 mutations in Parkinson disease. *Am. J. Hum. Genet.* **89**, 162–167 (2011).
38. Ropers, F. *et al.* Identification of a novel candidate gene for non-syndromic autosomal recessive intellectual disability: the WASH complex member SWIP. *Hum. Mol. Genet.* **20**, 2585–2590 (2011).
39. de Bot, S. T. *et al.* Pure adult-onset spastic paraplegia caused by a novel mutation in the KIAA0196 (SPG8) gene. *J. Neurol.* **260**, 1765–1769 (2013).
40. Valdmanis, P. N. *et al.* Mutations in the KIAA0196 gene at the SPG8 locus cause hereditary spastic paraplegia. *Am. J. Hum. Genet.* **80**, 152–161 (2007).
41. Huttlin, E. L. *et al.* The BioPlex Network: a systematic exploration of the human interactome. *Cell* **162**, 425–440 (2015).
42. Temkin, P. *et al.* SNX27 mediates retromer tubule entry and endosome-to-plasma membrane trafficking of signalling receptors. *Nat. Cell Biol.* **13**, 717–723 (2011).
43. Ghai, R. *et al.* Phox homology band 4.1/ezrin/radixin/moesin-like proteins function as molecular scaffolds that interact with cargo receptors and Ras GTPases. *Proc. Natl Acad. Sci. USA* **108**, 7763–7768 (2011).
44. Stockinger, W. *et al.* The PX-domain protein SNX17 interacts with members of the LDL receptor family and modulates endocytosis of the LDL receptor. *EMBO J.* **21**, 4259–4267 (2002).
45. Burden, J. J., Sun, X.-M., García, A. B. G. & Soutar, A. K. Sorting motifs in the intracellular domain of the low density lipoprotein receptor interact with a novel domain of sorting nexin-17. *J. Biol. Chem.* **279**, 16237–16245 (2004).
46. Farfán, P. *et al.* A sorting Nexin 17-binding domain within the LRP1 cytoplasmic tail mediates receptor recycling through the basolateral sorting endosome. *Traffic* **14**, 823–838 (2013).
47. van Kerkhof, P. *et al.* Sorting nexin 17 facilitates LRP recycling in the early endosome. *EMBO J.* **24**, 2851–2861 (2005).
48. Pietiäinen, V. *et al.* NDRG1 functions in LDL receptor trafficking by regulating endosomal recycling and degradation. *J. Cell Sci.* **126**, 3961–3971 (2013).
49. Rohlmann, A., Gotthardt, M., Willnow, T. E., Hammer, R. E. & Herz, J. Sustained somatic gene inactivation by viral transfer of Cre recombinase. *Nat. Biotechnol.* **14**, 1562–1565 (1996).
50. Bartuzi, P. *et al.* A cell-type-specific role for murine *Comm1d1* in liver inflammation. *Biochim. Biophys. Acta* **1842**, 2257–2265 (2014).
51. Mao, X. *et al.* GCN5 is a required cofactor for a ubiquitin ligase that targets NF- κ B/RelA. *Genes Dev.* **23**, 849–861 (2009).
52. Wu, J.-H. *et al.* The adaptor protein beta-arrestin2 enhances endocytosis of the low density lipoprotein receptor. *J. Biol. Chem.* **278**, 44238–44245 (2003).
53. Levels, J. H. M., Lemaire, L. C. J. M., van den Ende, A. E., van Deventer, S. J. H. & van Lanschot, J. J. B. Lipid composition and lipopolysaccharide binding capacity of lipoproteins in plasma and lymph of patients with systemic inflammatory response syndrome and multiple organ failure. *Crit. Care Med.* **31**, 1647–1653 (2003).
54. Vonk, W. I. M. *et al.* The copper-transporting capacity of ATP7A mutants associated with Menkes disease is ameliorated by *COMM1D1* as a result of improved protein expression. *Cell. Mol. Life Sci.* **69**, 149–163 (2012).
55. Jones, C. *et al.* JCI—disruption of LDL but not VLDL clearance in autosomal recessive hypercholesterolemia. *J. Clin. Invest.* **117**, 165–174 (2007).
56. Lagace, T. A. *et al.* Secreted PCSK9 decreases the number of LDL receptors in hepatocytes and in livers of parabiotic mice. *J. Clin. Invest.* **116**, 2995–3005 (2006).
57. van de Sluis, B. *et al.* Increased activity of hypoxia-inducible factor 1 is associated with early embryonic lethality in *Comm1d1* null mice. *Mol. Cell. Biol.* **27**, 4142–4156 (2007).
58. Jones, C. *et al.* Normal sorting but defective endocytosis of the low density lipoprotein receptor in mice with autosomal recessive hypercholesterolemia. *J. Biol. Chem.* **278**, 29024–29030 (2003).
59. Maxwell, K. N. & Breslow, J. L. Adenoviral-mediated expression of *Pcsk9* in mice results in a low-density lipoprotein receptor knockout phenotype. *Proc. Natl Acad. Sci. USA* **101**, 7100–7105 (2004).
60. Maxwell, K. N., Fisher, E. A. & Breslow, J. L. Overexpression of PCSK9 accelerates the degradation of the LDLR in a post-endoplasmic reticulum compartment. *Proc. Natl Acad. Sci. USA* **102**, 2069–2074 (2005).
61. Zelcer, N., Hong, C., Boyadjian, R. & Tontonoz, P. LXR regulates cholesterol uptake through idol-dependent ubiquitination of the LDL receptor. *Science* **325**, 100–104 (2009).
62. Derivery, E. *et al.* The Arp2/3 activator WASH controls the fission of endosomes through a large multiprotein complex. *Dev. Cell* **17**, 712–723 (2009).

Acknowledgements

We are grateful to H.H. Hobbs (Department of Molecular Genetics, University of Texas Southwestern Medical Center, Dallas, TX, USA) for providing the rabbit polyclonal antibody against ARH and to N. Freedman (Department of Medicine, Duke University Medical Center, Durham, NC, USA) for providing the various LDLR expression constructs. We thank S. Ijzendoorn (Cell Biology-Membrane Cell Biology, University of Groningen, the Netherlands) for providing Alexa633-transferrin. We thank Sanne Wilbrink for technical assistance, and Laura Damiano for critically reading the manuscript. The work was partly funded by the Groningen University Institute for Drug Exploration (GUIDE; granted to B.v.d.S.), a NWO-ALW grant (817.02.022) to B.v.d.S., a Jan Kornelis de Cock foundation (University Medical Center Groningen) grant to P.B. and B.v.d.S., an EU TransCard (FP7-603091-2) grant to J.A.K./B.v.d.S., NIH grants R01 DK073639 to E.B. and R01 AI065474 to D.D.B., and a NIH grant (HL020948) to J.D.H. This work was supported by the Optical Microscopy Core of the Mayo Clinic Center for Cell Signaling in Gastroenterology (P30DK084567).

Author contributions

P.B., D.D.B., S.R., D.D., A.F., M.W., J.H.L., N.H., N.K., H.M., G.B. and B.v.d.S. carried out the experiments; D.D.B., J.D.H., E.B. and B.v.d.S. designed the experiments; R.F., H.F., A.M.E., B.P., G.G. and J.M. provided clinical data; J.D.H., E.B., M.H.H. and B.v.d.S. supervised the project; P.B. and B.v.d.S. wrote the manuscript, with input from D.D.B., A.K.G., A.M.E., J.A.K., B.P., G.G., J.D.H., E.B. and M.H.H.

Additional information

Supplementary Information accompanies this paper at <http://www.nature.com/naturecommunications>

Competing financial interests: The authors declare no competing financial interests.

Reprints and permission information is available online at <http://npg.nature.com/reprintsandpermissions/>

How to cite this article: Bartuzi, P. *et al.* CCC- and WASH-mediated endosomal sorting of LDLR is required for normal clearance of circulating LDL. *Nat. Commun.* **7**:10961 doi: 10.1038/ncomms10961 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>