



CCL5–Glutamate Cross-Talk in Astrocyte-Neuron Communication in Multiple Sclerosis

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The immune system (IS) and the central nervous system (CNS) are functionally coupled, and a large number of endogenous molecules (i.e., the chemokines for the IS and the classic neurotransmitters for the CNS) are shared in common between the two systems. These interactions are key elements for the elucidation of the pathogenesis of central inflammatory diseases. In recent years, evidence has been provided supporting the role of chemokines as modulators of central neurotransmission. It is the case of the chemokines CCL2 and CXCL12 that control pre- and/or post-synaptically the chemical transmission. This article aims to review the functional cross-talk linking another endogenous pro-inflammatory factor released by glial cells, i.e., the chemokine Regulated upon Activation Normal T-cell Expressed and Secreted (CCL5) and the principal neurotransmitter in CNS (i.e., glutamate) in physiological and pathological conditions. In particular, the review discusses preclinical data concerning the role of CCL5 as a modulator of central glutamatergic transmission in healthy and demyelinating disorders. The CCL5-mediated control of glutamate release at chemical synapses could be relevant either to the onset of psychiatric symptoms that often accompany the development of multiple sclerosis (MS), but also it might indirectly give a rationale for the progression of inflammation and demyelination. The impact of disease-modifying therapies for the cure of MS on the endogenous availability of CCL5 in CNS will be also summarized. We apologize in advance for omission in our coverage of the existing literature.

Keywords: CCL5, glutamate, nerve endings, multiple sclerosis, experimental autoimmune encephalomyelitis mice, release

INTRODUCTION

The immune system and the central nervous system (CNS) cross-talk, and this interaction are pivotal to the onset and the progression of central neurodegenerative diseases (i.e., Alzheimer's disease and amyotrophic lateral sclerosis), as well as in classic autoimmune-inflammatory disorders [i.e., multiple sclerosis (MS)]. Although inflammation probably does not represent an initiating factor, new evidence suggests that pro-inflammatory molecules contribute to the derangement of chemical synapses favoring disease progression (1). In fact, a number of endogenous molecules (i.e., cytokines and chemokines) released by the immunocompetent cells as well as by activated astrocytes control chemical transmission at active synapses, affecting the main functions of these cells including

transmitter release and second messenger production. These endogenous molecules are main transducers of the pathological “glial to neuron” cross-talk (1–5).

Chemokines: General Considerations

In mammals, the chemokine (chemotactic cytokines) universe comprised of approximately 50 endogenous small (8–14 kDa) peptides released by immune cells and 20 receptors (4, 5). Chemokines have a tertiary structure highly conserved and are subdivided into four groups (namely the CC, the CXC, the CX₂C, and the C subfamilies) based on the relative positions of two conserved cysteine residues near the N-terminus (6). Chemokines act at chemokine receptors that are seven transmembrane domain *Pertussis* toxin (PTx)-sensitive, G-protein-coupled receptors (GPCRs), which depending on the G protein involved, trigger enzymatic cascade of events controlling several intracellular pathways, mostly controlling Ca²⁺ ions mobilization, intracellular phosphorylation pathways, and small Rho GTPases signaling (7). First identified for their ability to mediate leukocyte chemo-attraction in inflammatory and autoimmune diseases (8), chemokines and their receptors are now recognized as a promiscuous and redundant system of signaling interactions and mutual binding relevant to inflammation, immunity and neuropathology. Most of the chemokine receptors bind more than one ligand, and several chemokines activate more than one receptor, accounting for the numerical mismatch among chemokines and relative receptors. In particular, CCR1, CCR3, and CCR5 are promiscuous receptors for different chemokines including CCL3 (macrophage inflammatory protein 1-alpha), CCL5 [Regulated upon Activation Normal T-cell Expressed and Secreted (RANTES)], and CCL7 (monocyte chemotactic protein-3). All these aspects have been largely documented in previous articles (3, 4, 9–19) and will not be further detailed.

CCL5

CCL5 plays a main role in inflammatory diseases and in cancer, because of its ability to control the movements of memory T lymphocytes, monocytes macrophages, and eosinophils (4–7). Evidence has been provided also showing a role of this chemokine in CNS diseases secondary to viral infections, such as the acquired immunodeficiency syndrome-related dementia, or involving neuro-inflammatory processes, such as MS, Alzheimer dementia, and Parkinson's disease (6, 12, 13, 20–23).

CCL5 is a 68-amino-acid protein that binds both pertussis toxin (PTx)-sensitive GPCRs (6, 8, 12) and PTx-insensitive GPCRs (24) in the CNS. As to the central role of CCL5, the chemokine controls positively the mobilization of cytosolic Ca²⁺ and second messenger production in cultured neurons (25–28), astrocytes (29), and microglia (30), but it also activates GPCRs negatively coupled to adenylyl cyclase (AC)-mediated signaling, leading to the reduction of the endogenous level of cytosolic cyclic adenosine monophosphate (cAMP) (Figure 1) (25, 29).

CCL5 PRODUCTION IN CNS

The endogenous level of CCL5 is very low, almost undetectable, in cerebral spinal fluid (CSF) of healthy individuals, but it increases

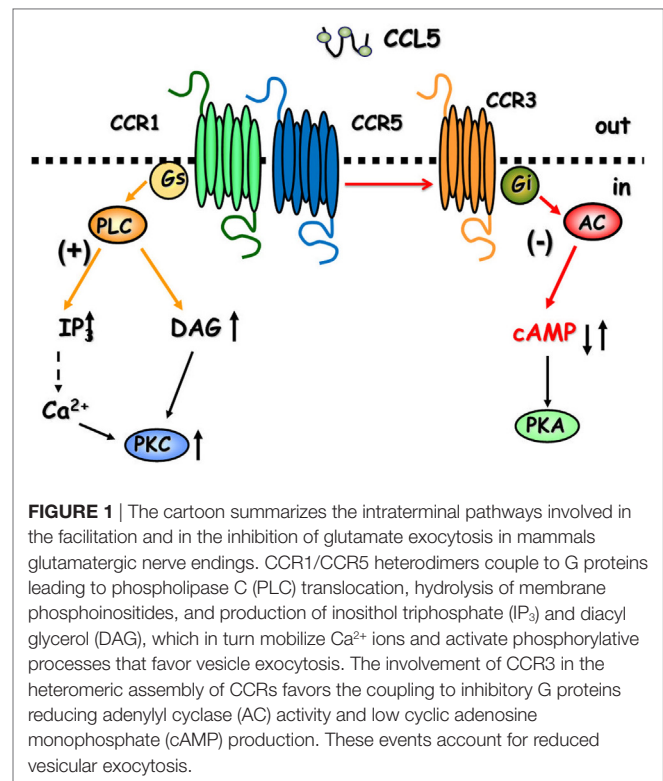


FIGURE 1 | The cartoon summarizes the intracellular pathways involved in the facilitation and in the inhibition of glutamate exocytosis in mammalian glutamatergic nerve endings. CCR1/CCR5 heterodimers couple to G proteins leading to phospholipase C (PLC) translocation, hydrolysis of membrane phosphoinositides, and production of inositol triphosphate (IP₃) and diacyl glycerol (DAG), which in turn mobilize Ca²⁺ ions and activate phosphorylation processes that favor vesicle exocytosis. The involvement of CCR3 in the heteromeric assembly of CCRs favors the coupling to inhibitory G proteins reducing adenylyl cyclase (AC) activity and low cyclic adenosine monophosphate (cAMP) production. These events account for reduced vesicular exocytosis.

dramatically when human immunodeficiency virus 1 (HIV-1) infection occurs (31–35), at the onset and during the progression of MS (9, 36–40). Increased central and peripheral CCL5 levels are also detected in mice suffering from the experimental autoimmune encephalomyelitis (EAE), an animal model reproducing most of the spinal cord pathological features of MS in humans [(41) and references therein]. Two are the mechanisms determining the dramatic increase of CCL5 bioavailability in CNS. First, the permeabilization of the blood–brain barrier that occurs in inflammation favors the entry of CCL5 from the periphery into the brain. Second, the concomitant massive local production of CCL5 from astrocytes and, to a lesser extent, from microglial cells triggered by pro-inflammatory cytokines.

Microglia cells coordinate brain innate immunity, rapidly expand their population, and then migrate chemotactically to sustain inflammation and cells death. Microglia exist in the “resting” and the “activated” forms. In the resting state, that is an active surveying state, microglia cells in close proximity to neurons and astrocytes participate to the central network by releasing regulatory agents and by controlling homeostasis. In the “active” state, microglia releases pro-inflammatory effectors including TNF-alpha and IL-1beta, which diffuse to neighboring astrocytes, influencing their functions (42). These factors are the principal inducers of chemokine overproductions from astrocytes. Microglia filopodia make dynamic contact with astrocytes and neurons and have a pivotal role in controlling synaptic plasticity (43–45). The intimate contact of microglia with chemical synapses also makes the neuronal control of microglial functions possible. Actually, microglia cells are endowed with receptors for transmitters (46) the activation of which controls their activation

state. Excitatory transmission favors the shift from the “resting” to the “activated” state of microglial cells, while inhibitory transmission reduces activation of microglial cells to the “resting” condition (47).

Astrocytes are innate immune sentinels that ensheath cerebral blood vessels controlling the entry of peripheral cells into the CNS parenchyma during infection. Astrocytoma cells as well as cultured astrocytes produce CCL5 upon incubation with the microglial effectors TNF-alpha or IL-1 beta, through an inflammatory-like response mediated by the p55 receptor-dependent signaling (42, 48). IFN-gamma from microglial cells, inactive on its own (42), seriously amplifies the effect of either TNF-alpha or IL-1 beta, worsening the pro-inflammatory signaling.

The expression and the release of CCL5 from astrocytes is tightly controlled by several receptor subtypes, including opioid receptors (49, 50), group III metabotropic glutamate receptors (2), alpha/beta noradrenergic receptors (51–53), and sphingosine-1-phosphate receptor (S1PR) subtype 1 (54), as well as by endogenous neurotoxin such as quinolinic acid acting at NMDA receptors (55). These receptors represent targets of therapeutics for inflammatory autoimmune disease typified by overexpression of CCL5.

Differently, data supporting the active production and release of the endogenous chemokine from neurons are lacking, suggesting that these cells do not release the chemokine.

EXPRESSION OF THE CHEMOKINE RECEPTORS TARGETED BY CCL5 IN GLIAL CELLS

Once released in the synaptic cleft, CCL5 regulates the function of glial cells (microglia and astrocytes) themselves through autocrine processes.

The autocrine control of microglia functions is permitted by the existence of CCR5 receptors, and, to a less extent, of CCR1 and CCR3 receptors in these cells, which also permit the chemotactic movements of microglial cells during inflammation (56). CCR5s are also the receptors functional to HIV-1-induced pathogenetic mechanisms (57–59). As to the astrocytes, the existence of CCR1, CCR3, and CCR5 proteins in these cells has been a matter of discussion, and, despite some initial discrepancies (60), it was definitively demonstrated that adult astrocytes possess CCR1, CCR3, and CCR5 (61–63). CCR1, CCR3, and CCR5 proteins are expressed in cultured fetal astrocytes and in adult astrocytes from mammalian brains [(29), but see for a concise review Ref. (60)], as well as in astrocytic processes [gliosomes (64, 65)] isolated from the cortex and the spinal cord of adult mice (**Table 1**) (66). In particular, in physiological conditions, the expression of CCR5 in astrocytes is low but rapidly increases to abnormal pathological levels upon stimulation with TNF-alpha and by IL-1beta released by neighboring microglial cells, which causes persistent adaptation leading to a significant increase in the expression of most of the CCR proteins (55). This cascade of events occurs during inflammation, so that the overexpression of the receptors targeted by CCL5 is an event intimately linked to pathological conditions.

CCL5 AND GLUTAMATE RELEASE FROM ASTROCYTES

In cultured astrocytes from both fetal simian and human brains, a significant increase in internal Ca^{2+} ions mobilization was observed upon exposure to CCL5 (29). Since intraterminal Ca^{2+} ions' mobilization is a prerequisite to transmitter exocytosis also in these cells [(77) and references therein] the CCL5-mediated control of cytosolic Ca^{2+} bioavailability might suggest that the chemokine could favor/modulate glutamate exocytosis from astrocytes. Despite the expectation, however, the spontaneous and the depolarization-evoked release of [^3H]D-aspartate ([^3H]D-Asp) from gliosomal particles isolated from human brain tissue was not modified by CCL5 (29), compatible with the idea that activation of these receptors cannot modify the glutamate outflow of the excitatory aminoacid. Similarly, exposure of human and mouse glial particles [we refer to as gliosomes (64)] to CCL5 in the absence or in the presence of a concomitant depolarizing stimulus (i.e., 20 mM KCl-enriched solution, KCl substituting for an equimolar amount of NaCl) did not significantly modify the outflow of glutamate. This is compatible with the idea that the amount of Ca^{2+} ions mobilized in gliosomal cytosol by CCL5 acting at its own receptors was insufficient to prime vesicle docking and fusion with astrocyte membranes (66, 71). So far, data concerning the impact of CCL5 on glutamate uptake in microglia and astrocytes are not available and further investigations are needed to address these aspects.

CCL5 IN NEURONS

Expression of Chemokine Receptors Targeted by CCL5 in Neurons

Once released in the biophase, CCL5 also reaches neurons through a mechanism of volume diffusion, to modulate neuronal functions via paracrine mechanisms, mediated by CCRs express in neurons. This regulation eventually occurs when the external concentration of the chemokine is high and assures a sufficient diffusion of the agent in the synaptic space.

The first report of the existence of chemokine receptors in neurons dates to 1997 [(67), **Table 1**]. In this article, by combining immune-histochemical staining with receptor binding studies, the authors demonstrated that cultured human neuronal cells express CXCR2, CXCR4, CCR1, and CCR5 receptors. The authors suggested that these entities represent the binding site for the viral envelope protein gp120 of the HIV-1 virus on neuronal plasmamembranes, allowing a CD4-independent interaction between the virus and the neurons. Soon after, in 1998, Meucci et al. (25) demonstrated that cultured hippocampal neurons are endowed with several chemokine receptors, including CCR1 and CCR5 subtypes. In line with these observations, in 1999, Klein et al. (29) provided clear evidence that a subpopulation of neurons in the cortex of human and macaque brains are endowed with CCR3, CCR5, and CXCR4 receptors. The existence of CCR1, CCR3, and CCR5 receptor proteins in neurons was then confirmed by other groups (20, 78–81), despite some discrepancies concerning their exact location (in the soma, on axonal processes and/or in

TABLE 1 | Distribution of CCR1, CCR3, and CCR5 in astrocytes and neurons in the central nervous system (CNS) of mammals.

	Cell types	CNS region	Species	mRNA	Protein	Reference
CCR1	Neurons	Cultured neurons	Human	+		(67)
		Fetal brain	Macacque			(68)
		Cerebellum	Rat			(27)
		Hippocampus	Rat	+		(25)
		Neonatal and adult cerebellum	Rat	+		(69)
		Neonatal and adult cerebellum	Rat	+		(70)
		Cortical nerve endings	Human			(71)
		Cortical and spinal cord nerve endings	Mouse			(66, 72)
	Astrocytes	Neonatal brain	Mouse	+		(73)
		Neonatal brain	Mouse	+	+	(74)
		Fetal brain	Human		+	(75)
		Neonatal and adult cerebellum	Rat	+	+	(69)
		Neonatal and adult cerebellum	Rat	+	+	(70)
		Spinal cord gliosomes	Mouse		+	(66)
CCR3	Neurons	Cerebellar neurons	Rat		+	(27)
		Cortical neurons	Fetal human		+	(29)
			Fetal macaque		+	(29)
		Cortical nerve endings	Human		+	(71)
		Cortical and spinal cord nerve endings	Mouse		+	(66, 72)
		Fetal brain neurons	Human		+	(20)
	Astrocytes	Primary astrocytes	Human	+	+	(62)
		Fetal and adult astrocytes	Human, macaque	+	+	(76)
		Spinal cord gliosomes	Mouse		+	(66)
		Fetal brain astrocytes	Human		+	(20)
CCR5	Neurons	Neonatal DRG	Rat	+		(68)
		Cerebellar neurons	Rat		+	(27)
		Cortical neurons	Fetal human		+	(29)
			Fetal macaque		+	(29)
		Hippocampal neurons	Rat	+		(25)
		Embryonic neurons	Human	+		(26)
		Cortical nerve endings	Human		+	(71)
		Cortical and spinal cord nerve endings	Mouse		+	(66, 72)
		Neonatal brain	Rat	+	+	(48)
		Brain	Human		+	(33)
	Astrocytes	Cortical astrocytes	Fetal human		+	(29)
			Fetal macaque		+	(29)
		Fetal brain neurons	Human		+	(20)
		Fetal and adult astrocytes	Human		+	(76)
Spinal cord gliosomes		Mouse		+	(66)	
Neonatal brain		Rat	+	+	(48)	
Brain	Human		+	(33)		

nerve terminals). As to this aspect, a relevant finding was that CCR1, CCR3, and CCR5 receptor proteins exist in both human and rodent cortical nerve endings, as well as in rodent spinal cord terminals, i.e., in those parts of neuron where transmitter exocytosis occurs (66, 71, 72).

CCL5 and Glutamate Release from Neurons

As observed in astrocytes, CCL5 controls the movement of Ca^{2+} ions in neurons (25, 29) but differently from astrocytes, this effect is sufficient to trigger changes in glutamate release efficiency. Based on these first observations, as well as on data published about a decade later (66, 71), the impact of CCL5 on glutamate release was found to represent a complex event, strictly dependent

on the activity of the neurons themselves and on the region of the CNS under study.

When studying the changes in Ca^{2+} cytosolic bioavailability in hippocampal cultured neurons, Meucci et al. (25) showed that nanomolar CCL5 favored Ca^{2+} ion mobilization in resting condition but significantly reduced the increase in cytosolic Ca^{2+} that follows exposure of neurons to a depolarizing stimulus. Similarly, low nanomolar concentrations of the human recombinant CCL5 (hCCL5) exert opposite control on glutamate release [measured as the release of the unmetabolizable marker of glutamate, the compound [3H]D-Asp (82–85)] from nerve endings (synaptosomes) isolated from cortical specimens that were removed from consenting patients undergoing neurosurgery to reach deeply located tumors (71). In particular, hCCL5 elicited a significant increase in the spontaneous release of [3H]D-Asp from these

terminals in basal conditions (i.e., in the absence of a depolarizing stimulus).

Facilitation of glutamate outflow relied on the activation of PTx-sensitive GPCRs positively coupled to phospholipase C (PLC)-mediated events, the activation of which led to the hydrolysis of membranes phosphoinositide and the mobilization of Ca²⁺ ions from Xestospingonin-C-sensitive, inositol triphosphate (IP₃)-dependent intraterminal stores located in the endoplasmic reticulum. Facilitation of glutamate release, however, turned to inhibition when hCCL5 was applied concomitantly to a mild depolarizing stimulus (i.e., 12 mM KCl). In this case, inhibition relied on the binding of hCCL5 to PTx-sensitive GPCRs negatively coupled to the AC/cAMP/protein kinase A (PKA) intraterminal enzymatic pathway (**Figure 1**). Comparable results were obtained when studying the impact of hCCL5 on human cortical slices. hCCL5 increased the basal release of [³H]D-Asp but significantly reduced the tritium overflow elicited by depolarizing stimuli (71).

Human recombinant CCL5-mediated facilitation of the spontaneous outflow of glutamate from both isolated nerve terminals and slices was prevented by MetRANTES, a broad-spectrum antagonist of the CCR1, CCR3, and CCR5 subtypes, confirming the involvement of these receptors (71). The impacts of the chemokine in the different experimental conditions (resting versus depolarized condition), however, seemed predictive of the existence of receptor subtype oligomers. Since antagonists able to discriminate among the different CCR subtypes were not available at that time, the pharmacological characterization of the receptor(s) accounting for the hCCL5-induced changes of glutamate outflow was carried out by pre-incubating human synaptosomes with antibodies raised against the N-terminal of the CCR1, the CCR3, and the CCR5 receptor proteins. By binding selectively to the outer side of the receptor protein, antibodies are expected to impede the interaction of the agonist with the orthosteric binding site, then mimicking receptor antagonists (86, 87). Pre-incubation of synaptosomes with antibodies raised against the extracellular NH₂ terminals of CCR1 or of CCR5 receptor proteins impeded the hCCL5-induced facilitation of glutamate outflow from cortical nerve endings, while pre-treatment with anti-CCR3 was ineffective. Differently, hCCL5-mediated inhibition of glutamate exocytosis was prevented by pre-incubating synaptosomes with anti-CCR1, anti-CCR3 or anti-CCR5 antibodies, consistent with the view that different CCR oligomers account for the opposite effects observed.

The main criticism to the results obtained with human nerve terminals concerned the potential confounding factors

originating from the origin of the human specimens, i.e., the brain of patients suffering from cerebral tumors. The receptor repertoire involved in the CCL5-mediated control of glutamate exocytosis in human specimens could have been altered because of the pathological overexpression of CCL5 in glioma cells. The effects of hCCL5 in human cortical synaptosomes, however, were soon after reproduced in glutamatergic nerve endings isolated from the cortex of mice, which represent healthy individuals, where the endogenous CCL5 level is expected to be low (88). Again, in mouse cortical terminals, the release of glutamate in basal condition (i.e., the absence of a depolarizing stimulus) was potentiated by CCL5, but the chemokine significantly inhibited the glutamate exocytosis evoked by a mild K⁺ depolarization (12 mM KCl) stimulus. The comparable effects observed in human and mice terminals allowed to conclude that the effects observed in human nerve endings were not influenced by the pathological origin of the tissue specimens.

Facilitation of the spontaneous release of glutamate as well as inhibition of 12 mM K⁺-evoked glutamate exocytosis from cortical synaptosomes was prevented by the selective CCR1 antagonist BX513 and by the selective CCR5 antagonist DAPTA, compatible with the involvement of CCR1/CCR5 heterodimers in the effect observed (66). Furthermore, the CCR3 antagonist, the compound SB 328437, failed to affect the CCL5-mediated facilitation of glutamate release in basal condition, but it strongly prevented the inhibitory effect exerted by the chemokine in depolarized nerve terminals. Comparable results could be drawn when using antibodies raised against the N-terminal of the CCR1, CCR3, and CCR5 receptor proteins, leading to conclude that (i) the receptor composition of the chemokine oligomers controlling glutamate release in mouse and human cortical nerve endings is largely conserved, (ii) the involvement of CCR3 in the oligomer expression dictates the coupling to inhibitory G proteins bridging negatively the chemokine receptor complex to the AC/cAMP/PKA transducing mechanism (**Figure 1**; **Table 2**), and (iii) the mouse brain tissue is appropriate to investigate the effects of CCL5 on central glutamatergic transmission (66).

CCL5-mediated control of glutamate release in nerve terminals was not restricted to the cortex. The chemokine also efficiently modulates the release of glutamate from spinal cord nerve endings. However, differently from what observed in the cortex of adult mice, the spontaneous release of glutamate from spinal cord glutamatergic nerve endings was unaffected by nanomolar CCL5, while the depolarization-evoked glutamate exocytosis was significantly increased. Again, facilitation of glutamate

TABLE 2 | Correlation between the composition of CCR oligomers and the CCL5-mediated changes to glutamate release.

	Human cortical synaptosomes		Mouse cortical synaptosomes		Mouse spinal cord synaptosomes	
	Basal glutamate release	12 mM KCl-evoked glutamate overflow	Basal glutamate release	12 mM KCl-evoked glutamate overflow	Basal glutamate release	15 mM KCl-evoked glutamate overflow
CCR1	↑	↓	↑	↓	↑	↑
CCR3	Not involved	↓	Not involved	↓	Not involved	Not involved
CCR5	↑	↓	↑	↓	↑	↑

The table resumes the modulatory action of CCL5 on the release of glutamate (measured as release of preloaded [³H]-aspartate) correlating these events to the CCR subunits involved in the expression of chemokine oligomers targeted by the chemokine.

↓, inhibition of glutamate release; ↑, facilitation of glutamate exocytosis.

exocytosis from these terminals involved CCR1/CCR5 heteromers positively coupled to PLC-induced IP₃-mediated enzymatic pathway, leading to increased mobilization of Ca²⁺ ions in the cytosol. Also in this region, the receptor protein composition was clarified by using selective CCR antagonists as well as anti CCR antibodies recognizing the N-terminal of the receptor protein (66). These studies unveiled that CCR1/CCR5 heterodimers mediates the CCL5-induced facilitation of glutamate exocytosis, further confirming the hypothesis that CCR1/CCR5 oligomers preferentially couples to stimulatory G protein positively coupled to PLC-mediated events (**Figure 1**; **Table 2**). Notably, CCR3 immunoreactivity was detected in spinal cord synaptosomal lysates, but this receptor subtype was not involved in the CCL5-mediated effect.

CCL5 IN DEMYELINATING DISORDERS

The serum level of CCL5 was found to be significantly increased in patients suffering from MS (89–91) as well as in EAE animals (15, 41, 92). The highest levels of the chemokine were detected in the peripheral blood mononuclear cells (PBMCs) of MS patients suffering from the secondary progressive form of the disease, while lower level was observed in the PBMCs from patients with the relapsing–remitting form of MS (93). CCL5 levels are also increased in the CNS of MS patients as well as of EAE mice (9, 13, 22, 88, 93–100). The huge elevation of the CCL5 levels in CNS mainly depends on the increased peripheral production of the chemokine, but it also reflects the local overexpression of the chemokine in astrocytes activated by IL-1beta, TNF-alpha, and IFN-gamma (55), from neighboring microglia cells. All these observations are predictive of the role of CCL5 in the onset and progression of disease in MS patients. Accordingly, a CCL5 polymorphism [the CCL5-403 G/A single nucleotide polymorphism (22, 99)] is associated to a higher risk of susceptibility to the onset of the disease, while modified CCL5 ligands are efficacious in controlling the symptoms and the neurodegenerative processes in EAE mice (15).

CCR1, CCR3, and CCR5 exist in different cell types, including T lymphocytes, monocytes/macrophages, and immature dendritic cells, but also exist in neurons and astrocytes (101–104). In particular, CCR1 and CCR3 are expressed by circulating T cells as well as in monocytes, which are occasionally found in perivascular infiltrates in the brain of MS patients. Differently, CCR5-positive T cells and macrophages are concentrated in the active demyelinating lesions in CNS of MS patients (13, 22). To note, the expression of CCRs in CNS correlates with disease severity (105–107) as proved by the observation that clinical symptoms are reduced in CCR1, CCR3, or CCR5 knockout (k.o.) EAE animals, which also suggest redundancy in the chemokine system (7, 8, 99, 108).

GLUTAMATE IN DEMYELINATING DISORDERS

Glutamate is the major excitatory neurotransmitter in CNS, where it mediates important physiological functions (i.e., synaptic

plasticity, learning, and memory), but also triggers excitotoxic degenerative processes. Glutamate concentration in the synaptic cleft is finely tuned by several cellular mechanisms including active re-uptake and release from nerve terminals as well as presynaptic mechanisms of control mediated by auto- and/or heteroreceptors (65, 82, 109–116). Glutamate bioavailability is also affected by neighboring astrocytes [i.e., the cells that take up and release the amino acid (77)] as well as by altered glutamate metabolism. L-Glutamate signaling, however, is not restricted to neuron/astrocyte compartments since glutamate receptors (GluR3-containing AMPA receptors and mGluR1/5 receptors) exist also in immune cells, including cells of the T lineage (117–119). Therefore, besides its role in controlling chemical transmission and excitotoxicity, glutamate may represent a chemo-attractant driving force for the recruitment and the migration of leukocytes and T cells into CNS site where glutamate release occurs (119).

Increased glutamate levels are found in the cerebrospinal fluid of MS patients (119–121) possibly because of the down-regulation of glutamate-metabolizing enzymes (glutamate dehydrogenase and glutamine synthase) and up-regulation of glutamate-producing enzyme glutaminase (122). In 2003, Sarchielli et al. (119) compared the levels of aspartate and glutamate in the CSF of patients suffering from different forms of MS and of controls healthy individuals. The authors find a significant increase of the glutamate levels in patients suffering from the relapsing–remitting form of MS. Interestingly, the glutamate levels were significantly higher in individuals suffering from the relapsing–remitting form of MS with active central lesions during the stable phase than in patients suffering from a similar form of disease, but without lesions. Inasmuch, high levels of glutamate were also detected in patients suffering from the secondary progressive form of MS.

Impaired glutamate bioavailability was also observed in EAE animals. However, depending on the animal model used and the brain region under study, opposite modifications of glutamate release efficiency were observed, consistent with the view that, in demyelinating disorders, impaired glutamate transmission at active synapses is a complex event. Increased glutamate release was detected in the spinal cord of EAE rats (123, 124) as well as in striatal and spinal cord nerve terminals of EAE mice (72, 88, 124–126), while reduced glutamate release was observed in cortical and hippocampal nerve endings of both mice and rats suffering from EAE disease (72, 88, 100, 126, 127). As to glutamate receptors, both metabotropic and ionotropic glutamate receptors (namely mGlu1/5 and mGlu4, mGlu2/3 receptors, and NMDA and AMPA receptors) control glutamate release (83, 109, 113, 115, 116, 128, 129). The expression and the function of these receptors were found to be altered in EAE mice when compared with controls (109, 128–136), suggesting that they represent suitable targets of drugs for the cure of MS symptoms. Besides receptors, also glutamate transporter expression is modified in EAE rats (124), determining increased glutamate bioavailability (137) and consequent neurotoxicity.

Generalized ongoing subclinical axonal degeneration in lesioned and non-lesioned white matter, as well as in gray matter, is detectable in CNS of MS patients and seems to occur independently from inflammation or demyelination, representing an

early cause of CNS damage in MS (138). Interestingly, besides the spinal cord, neurodegeneration also takes place in other brain regions, such as the cortex and the hippocampus, and could be responsible of cognitive and affective dysfunctions (121, 139–141) that represent common and early manifestations of MS (142, 143).

CCL5-MEDIATED CONTROL OF GLUTAMATE RELEASE IN EAE MICE

The thesis that central chemokines and classic transmitters functionally cross-talk in CNS has several implications and add new aspects of interest to the role of chemokines in the synaptic rearrangements that typify neuro-inflammatory central disease.

As already stated, an abnormal overproduction of CCL5 in the spinal cord, and to a lesser extent in the cortex, of EAE mice was evidenced in immunocytochemistry analysis and confirmed in tissue homogenate (88, 100, 126) and in blood (88). Concomitantly, changes in glutamate exocytosis from nerve endings isolated from the cortex and the spinal cord of EAE mice were observed. Quite interestingly, the modifications of glutamate exocytosis observed in cortical and spinal cord synaptosomes from EAE mice recall the presynaptic modulation elicited by CCL5 in nerve terminals in healthy mice. Actually, glutamate exocytosis was reduced in cortical nerve endings from EAE mice [i.e., in this CNS region, CCL5 inhibits glutamate exocytosis in control animals (66, 71)], but it was drastically increased in spinal cord terminals [where a positive role of CCL5 on glutamate release was observed in healthy animals (66)].

In both CNS regions, the altered glutamate exocytosis was paralleled by impaired second messenger production. Again, the alterations in cAMP and IP₃ productions observed in both cortical and spinal cord synaptosomal subpopulations from EAE mice were reminiscent of the modulatory presynaptic effects exerted by CCL5 on the corresponding enzymatic pathways in nerve terminals from control, non-immunized, mice. In fact, endogenous cAMP was drastically reduced in cortical synaptosomes but not in the spinal cord, where IP₃ production, the second messenger accounting for the CCL5-mediated presynaptic facilitation of glutamate exocytosis was significantly augmented (**Figure 1**) (66, 88).

The changes in second messenger productions and release efficiency could be explained by assuming that the prolonged *in vivo* CCR activation elicited by the high CCL5 could have triggered adaptive intraterminal changes in nerve terminals (144–146), which are retained in “*ex vivo*, *in vitro*” synaptosomal preparations and can emerge in “*in vitro*” functional studies as changes in glutamate release efficiency and second messenger production. As a matter of fact, the abnormal expression of the chemokine could have reverberated on the CCR repertoire presynaptically located on glutamatergic nerve terminals, leading to adaptation of the CCRs heteromers controlling glutamate exocytosis. These adaptations might lead to changes in the receptor expression and/or associated signaling that might account for the profound changes in glutamate exocytosis in

nerve terminals from EAE mice. The CCR composition of the presynaptic chemokinergic oligomers involved in the CCL5-mediated control of glutamate exocytosis in both cortical and spinal cord nerve endings of EAE mice, however, was conserved when compared to control mice, indirectly suggesting that adaptation in CCR subunits assembly were not involved in the EAE-induced changes to the CCL5-mediated control of glutamate exocytosis described above (72).

As to the second messengers, the strict correlation linking CCL5 levels, glutamate release efficiency and IP₃, and cAMP accumulation was confirmed by the observation that administration of drugs able to reduce the overexpression of CCL5 in CNS [i.e., the antidepressant desipramine (DMI)] (126) restored both presynaptic functions (i.e., transmitter exocytosis as well as second messenger production) at glutamatergic nerve endings in the cortex of mice suffering from EAE. The beneficial effects exerted by DMI were mediated by the change in noradrenaline bioavailability, due to blockade of the noradrenaline transporters. Actually, the transient increase in the endogenous amine in the synaptic cleft elicited by DMI activates the α and β receptors expressed in astrocytes in the near proximity of the noradrenergic nerve terminals, the activation of which hampers the central endogenous production and release of pro-inflammatory chemokines, including CCL5 (53, 126, 147) from these cells. Interestingly, the peripheral production of the chemokine (which at that stage of disease is already augmented) was unaffected (126).

To conclude, hampering the central overproduction of CCL5 leads to a marked amelioration of the presynaptic defects in terms of release of glutamate and second messenger production. Altogether, these observations clearly support a strict correlation between the increased production of CCL5 in the CNS and the onset of synaptic glutamatergic alteration in EAE mice, also strengthening the pathological role of the “glial to neuron” “CCL5–glutamate” interaction.

CCL5 IN MS: CLINICAL STUDIES

In 2016, Centonze et al. (40) demonstrated that the endogenous concentration of CCL5 in the cerebrospinal fluid of MS patients suffering from the active form of the disease was largely increased when compared to healthy individuals and to patients at the inactive stage of disease. Inasmuch, the authors showed a significant correlation between the endogenous level of CCL5 and the amount of IL-1 beta, used as a marker of gravity of the disease. CSF levels of RANTES were associated with enhanced cortical excitability in the cortex, as suggested by results from studies in which cortical excitability and plasticity was monitored with transcranial magnetic stimulation in MS patients. In these experiments, the authors highlighted a high correlation between the increased intracortical facilitation and the endogenous amount of CCL5. Differently, no correlation emerged when studying the relation linking CCL5 and a long-term potentiation-like synaptic plasticity measured through theta burst stimulation in the same patients. Despite the contrasting observation, the authors concluded that CCL5 couples inflammation and synaptic excitability in MS brains.

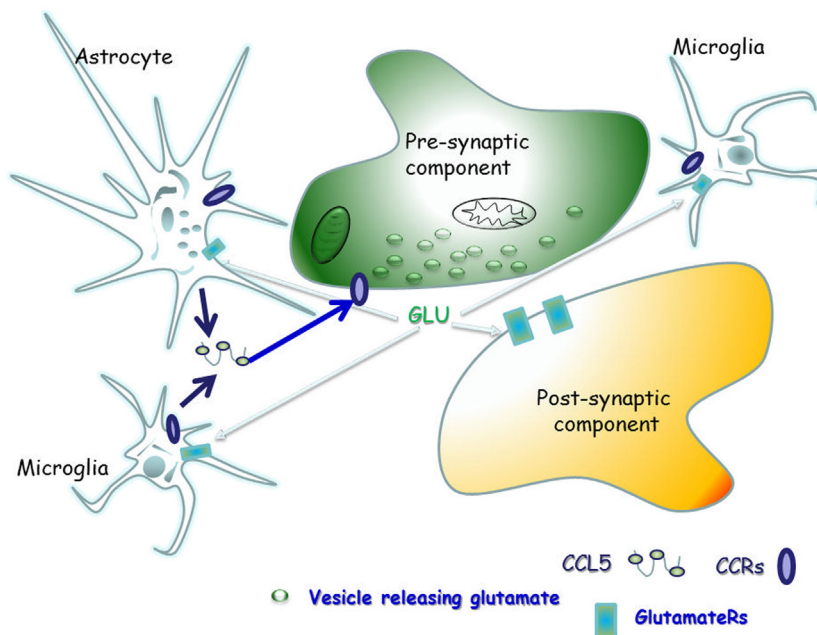


FIGURE 2 | The “quad partite” synapse is a functional structure consisting of neurons, astrocytes, and microglia cells. It represents the simplest central Unit, where adaptive and damaging processes occur in neuro-inflammatory disorders, including the demyelinating one. The concept originates from the tripartite synapse (152), but it is characterized by a highest level of complexity, since microglia is included as players of synaptic derangement. In a simplified model for demyelinating disorder, microglia cells migrating from periphery to central nervous system (CNS) as well as resident central microglia rapidly expand their populations and differentiate into the M1- and M2-cell subgroups, which exert various and mostly opposite functions in the brain. Microglia cells of the M1 group releases pro-inflammatory cytokines, including CCL5, which in turn activate astrocytes. In a whole these events sustain and worsen central inflammatory processes. Differently, M2 microglia secretes anti-inflammatory cytokine and its neuroprotective. Astrocytes are the most abundant glial cells in the human brain and represent the innate immune sentinels that sheath cerebral blood vessels controlling the entry of peripheral cells into the CNS during infection. Astrocytes are neuroprotective at the initial stage of disease, since they reduce local hyper-glutamatergicity by active glutamate uptake processes. Astrocyte activation, however, becomes pathological upon prolonged exposure to the pro-inflammatory compounds released from neighboring microglial cells. At this stage, reactive astrocytes become hypertrophic, do not uptake efficiently glutamate, but release much more cytokines (including CCL5), which accelerate neurodegenerative processes. CCL5 actively released by activated astrocytes and microglia by one side and the abnormal bioavailability of glutamate in the synaptic cleft, on the other side, reverberate onto neurons, eliciting structural and functional changes at chemical synapses.

IMPACT OF DISEASE-MODIFYING DRUGS ON THE ENDOGENOUS PRODUCTION OF CCL5 IN DEMYELINATING DISORDERS

As already stated in this review, CCL5 production is increased in MS patients suffering from both the remitting and the non-remitting form of the disease (15, 41, 89–92). Interestingly, most of the therapeutics currently in use for the cure of MS reduces significantly the overexpression of CCL5.

It is the case of interferon-beta-1b (IFN-beta-1b); administration of this drug prevents CCL5 overproduction in the sera, in the peripheral blood and in the adherent mononuclear cell supernatants during both the relapse and the remission stage of the pathology. These observations are compatible with the idea that CCL5 might be involved in determining the molecular events accounting for the action of IFN-beta-1b in MS patients (38).

Similarly, glatiramer acetate, an approved drug for the treatment of MS, was reported to reduce the TNF-alpha-induced CCL5 mRNA overexpression in human U-251 MG astrocytes. This effect was attributed to the inhibition of mRNA transcription and led to the conclusion that glatiramer acetate may exert

its therapeutic effect in MS also by inhibiting pro-inflammatory signaling (148).

Activation of cannabinoid receptors, which represents a therapeutic approach useful to control the progression of central neuroinflammation in EAE mice and MS patients, also reduces the endogenous availability of the chemokine CCL5 being concomitantly beneficial to the progression of the demyelinating disorder (41, 149).

Laquinimod is a novel orally administered drug for the treatment of relapsing–remitting MS. The molecular events accounting for its therapeutic effects are far from to be elucidated. Monocytes obtained from laquinimod-treated patients tended to secrete lower levels of the pro-inflammatory chemokines CCL2 or CCL5 (150).

Another orally active disease-modifying drug is Fingolimod. Fingolimod is a pro-drug, rapidly metabolized to its active form, the fingolimod-phosphate (fingolimod-P). By acting at the S1PRs in microglia cells, in circulating T cells, and in the spleen, fingolimod-P prevents the egress of lymphocytes and exerts central anti-inflammatory effects favoring remyelination (151). Recent data demonstrated that *in vivo* oral (the drug dissolved in the

drinking water) administration of this drug largely ameliorated the clinical symptoms in EAE mice. The treatment was beneficial to the inflammation and demyelination in the spinal cord of EAE mice, also significantly reducing the endogenous content of CCL5 in this CNS region (100).

Quite interestingly, the abovementioned therapeutics were also described to ameliorate glutamatergic synaptic transmission (152–155) further supporting the strict connection linking CCL5 overexpression and glutamatergic synaptic derangements.

CONCLUSION

The scope of this manuscript is to review the literature concerning the physio-pathological role of CCL5 in controlling glutamate transmission in the CNS of healthy mammals, as well as of individuals and animals suffering from demyelinating disease, in order to highlight the main role of CCL5 as a modulator of the neuroimmune cross-talk in the “quad partite” synapse in CNS (Figure 2).

These effects, together with the well-known chemo-attractant role of the chemokine toward glial cells, suggest that CCL5 exerts a dual role in the CNS of individuals suffering from MS. On one hand, the chemokine impairs the chemical transmission at asymmetric synapses in selected region of the CNS. On the other hand, it worsens the course of disease progression by favoring

the recruitment of pro-inflammatory glial cells in the site of the lesion.

When considering its role as modulator of glutamate transmission, the chemokine preferentially emerges as a key effector of the “astrocytes to neurons” signaling in CNS. Actually, the chemokine released from astrocyte and microglia is an efficient paracrine modulator of glutamate release at synaptic boutons of glutamatergic neurons in both healthy and demyelinating conditions, while its autocrine role of modulator of glutamate overflow from astrocytes is less evident (Table 1).

Therapeutic approaches aimed at containing the overexpression of the chemokine might represent therefore a useful approach to the cure of MS.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and approved it for publication.

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REFERENCES

- Hohlfeld R, Kerschensteiner M, Mein E. Dual role of inflammation in CNS disease. *Neurology* (2007) 68:558–63. doi:10.1212/01.wnl.0000275234.43506.9b
- Adler MW, Geller EB, Chen X, Rogers TJ. Viewing chemokines as a third major system of communication in the brain. *AAPS J* (2006) 7:E865–70. doi:10.1208/aapsj070484
- Rostene W, Kitabgi P, Parsadaniantz SM. Chemokines: a new class of neuro-modulator? *Nat Rev Neurosci* (2007) 8:895–903. doi:10.1038/nrn2255
- Rot A, von Andrian UH. Chemokines in innate and adaptive host defense: basic chemokines grammar for immune cells. *Annu Rev Immunol* (2004) 22:891–928. doi:10.1146/annurev.immunol.22.012703.104543
- Charo IF, Ransohoff RM. The many roles of chemokines and chemokine receptors in inflammation. *N Engl J Med* (2006) 354:610–21. doi:10.1056/NEJMra052723
- Bajetto A, Bonavia R, Barbero S, Schettini G. Characterization of chemokines and their receptors in the central nervous system: physiopathological implications. *J Neurochem* (2002) 82:1311–29. doi:10.1046/j.1471-4159.2002.01091.x
- Viola A, Luster AD. Chemokines and their receptors: drug targets in immunity and inflammation. *Annu Rev Pharmacol Toxicol* (2008) 48:171–97. doi:10.1146/annurev.pharmtox.48.121806.154841
- Ubogu EE, Cossoy MB, Ransohoff RM. The expression and function of chemokines involved in CNS inflammation. *Trends Pharmacol Sci* (2006) 27:48–55. doi:10.1016/j.tips.2005.11.002
- Sorensen TL, Tani M, Jensen J, Pierce V, Lucchinetti C, Folcik VA, et al. Expression of specific chemokines and chemokine receptors in the central nervous system of multiple sclerosis patients. *J Clin Invest* (1999) 103:807–15. doi:10.1172/JCI5150
- Cho C, Miller RJ. Chemokine receptors and neural function. *J Neurovirol* (2002) 8:573–84. doi:10.1080/13550280290101003
- Guillard SE, Lu M, Mastracci RM, Miller RJ. Expression of functional chemokine receptors by rat cerebellar neurons. *J Neuroimmunol* (2002) 124:16–28. doi:10.1016/S0165-5728(02)00005-X
- Cartier L, Hartley O, Dubois-Dauphin M, Krause KH. Chemokine receptors in the central nervous system: role in brain inflammation and neurodegenerative diseases. *Brain Res Brain Res Rev* (2005) 48:16–42. doi:10.1016/j.brainresrev.2004.07.021
- Szczuczinski A, Losy J. Chemokines and chemokine receptors in multiple sclerosis. Potential targets for new therapies. *Acta Neurol Scand* (2007) 115:137–46. doi:10.1111/j.1600-0404.2006.00749.x
- Hamann I, Zipp F, Infante-Duarte C. Therapeutic targeting of chemokine signaling in multiple sclerosis. *J Neurol Sci* (2008) 274:31–8. doi:10.1016/j.jns.2008.07.005
- Proudfoot A, Soares de Souza AL, Muzio V. The use of chemokine antagonists in EAE models. *J Neuroimmunol* (2008) 198:27–30. doi:10.1016/j.jneuroim.2008.04.007
- Sorte S, Myburgh R, Krause KH. The chemokine receptor CCR5 in the central nervous system. *Prog Neurobiol* (2011) 93:297–311. doi:10.1016/j.pneurobio.2010.12.003
- Louboutin JP, Strayer DS. Relationship between the chemokine receptor CCR5 and microglia in neurological disorders: consequences of targeting CCR5 on neuroinflammation, neuronal death and regeneration in a model of epilepsy. *CNS Neurol Disord Drug Targets* (2013) 12:815–29. doi:10.2174/18715273113126660173
- Williams JL, Holman DW, Klein RS. Chemokines in the balance: maintenance of homeostasis and protection at CNS barriers. *Front Cell Neurosci* (2014) 8:154. doi:10.3389/fncel.2014.00154
- Le Thuc O, Blondeau N, Nahon JL, Rovère C. The complex contribution of chemokines to neuroinflammation: switching from beneficial to detrimental effects. *Ann N Y Acad Sci* (2015) 1351:127–40. doi:10.1111/nyas.12855
- van der Meer P, Ulrich AM, Gonzalez-Scarano F, Lavi E. Immunohistochemical analysis of CCR2, CCR3, CCR5, and CXCR4 in the human brain: potential mechanisms for HIV dementia. *Exp Mol Pathol* (2000) 69:192–201. doi:10.1006/exmp.2000.2336
- Alvarez Losada S, Cantó-Nogués C, Muñoz-Fernández MA. A new possible mechanism of human immunodeficiency virus type 1 infection of neural cells. *Neurobiol Dis* (2002) 11:469–78. doi:10.1006/nbdi.2002.0566
- Tomioka R, Matsui M. Biomarkers for multiple sclerosis. *Intern Med* (2014) 53:361–5. doi:10.2169/internalmedicine.53.1246
- Kempuraj D, Thangavel R, Natteru PA, Selvakumar GP, Saeed D, Zahoor H, et al. Neuroinflammation induces neurodegeneration. *J Neurol Neurosurg Spine* (2016) 1:1003.

24. Ignatov A, Robert J, Gregory-Evans C, Schaller HC. RANTES stimulates Ca²⁺ mobilization and inositol trisphosphate (IP₃) formation in cells transfected with G protein-coupled receptor 75. *Br J Pharmacol* (2006) 149:490–7. doi:10.1038/sj.bjp.0706909
25. Meucci O, Fatatis A, Simen AA, Bushell TJ, Gray PW, Miller RJ. Chemokines regulate hippocampal neuronal signaling and gp120 neurotoxicity. *Proc Natl Acad Sci U S A* (1998) 95:14500–5. doi:10.1073/pnas.95.24.14500
26. Boutet A, Salim H, Leclerc P, Tardieu M. Cellular expression of functional chemokine receptor CCR5 and CXCR4 in human embryonic neurons. *Neurosci Lett* (2001) 311:105–8. doi:10.1016/S0304-3940(01)02149-8
27. Gillard SE, Lu M, Mastracci RM, Miller RJ. Expression of functional chemokine receptors by rat cerebellar neurons. *J Neuroimmunol* (2002) 124:16–28. doi:10.1016/S0165-5728(02)00005-X
28. Oh SB, Endoh T, Simen AA, Ren D, Miller RJ. Regulation of calcium currents by chemokines and their receptors. *J Neuroimmunol* (2002) 123:66–75. doi:10.1016/S0165-5728(01)00485-4
29. Klein RS, Williams KC, Alvarez-Hernandez X, Westmoreland S, Force T, Lackner AA, et al. Chemokine receptor expression and signaling in macaque and human fetal neurons and astrocytes: implications for the neuropathogenesis of AIDS. *J Immunol* (1999) 163:1636–46. doi:10.1002/glia.10181
30. Hegg CC, Hu S, Peterson PK, Thayer SA. Beta-chemokines and human immunodeficiency virus type-1 proteins evoke intracellular calcium increases in human microglia. *Neuroscience* (2000) 98:191–9. doi:10.1016/S0306-4522(00)00101-9
31. Kelder W, McArthur JC, Nance-Sproson T, McClernon D, Griffin DE. Beta-chemokines MCP-1 and RANTES are selectively increased in cerebrospinal fluid of patients with human immunodeficiency virus-associated dementia. *Ann Neurol* (1998) 44:831–5. doi:10.1002/ana.410440521
32. Kolb SA, Sporer B, Lahrtz F, Koedel U, Pfister HW, Fontana A. Identification of a T cell chemotactic factor in the cerebrospinal fluid of HIV-1-infected individuals as interferon-gamma inducible protein 10. *J Neuroimmunol* (1999) 93:172–81. doi:10.1016/S0165-5728(98)00223-9
33. McManus CM, Weidenheim K, Woodman SE, Nunez J, Hesselgesser J, Nath A, et al. Chemokine and chemokine-receptor expression in human glial elements: induction by the HIV protein, Tat, and chemokine autoregulation. *Am J Pathol* (2000) 156:1441–53. doi:10.1016/S0002-9440(10)65013-4
34. El-Hage N, Bruce-Keller AJ, Knapp PE, Hauser KF. CCL5/RANTES gene deletion attenuates opioid-induced increases in glial CCL2/MCP-1 immunoreactivity and activation in HIV-1 Tat-exposed mice. *J Neuroimmune Pharmacol* (2008) 3:275–85. doi:10.1007/s11481-008-9127-1
35. Hahn YK, Vo P, Fitting S, Block ML, Hauser KF, Knapp PE. beta-Chemokine production by neural and glial progenitor cells is enhanced by HIV-1 Tat: effects on microglial migration. *J Neurochem* (2010) 114:97–109. doi:10.1111/j.1471-4159.2010.06744.x
36. Ransohoff RM, Hamilton TA, Tani M, Stoler MH, Shick HE, Major JA, et al. Astrocyte expression of mRNA encoding cytokines IP-10 and JE/MCP-1 in experimental autoimmune encephalomyelitis. *FASEB J* (1993) 7:592–600. doi:10.4172/1948-5948.1000e131
37. Karpus WJ, Ransohoff RM. Chemokine regulation of experimental autoimmune encephalomyelitis: temporal and spatial expression patterns govern disease pathogenesis. *J Immunol* (1998) 161:2667–71.
38. Iarlori C, Reale M, Lugaesi A, De Luca G, Bonanni L, Di Iorio A, et al. RANTES production and expression is reduced in relapsing-remitting multiple sclerosis patients treated with interferon-beta-1b. *J Neuroimmunol* (2000) 107:100–7. doi:10.1016/S0165-5728(00)00261-7
39. Gentile A, Musella A, Bullitta S, Fresogna D, De Vito F, Fantozzi R, et al. Siponimod (BAF312) prevents synaptic neurodegeneration in experimental multiple sclerosis. *J Neuroinflammation* (2016) 13:207. doi:10.1186/s12974-016-0686-4
40. Mori F, Nisticò R, Nicoletti CG, Zagaglia S, Mandolesi G, Piccinin S, et al. RANTES correlates with inflammatory activity and synaptic excitability in multiple sclerosis. *Mult Scler* (2016) 22:1405–12. doi:10.1177/1352458515621796
41. Mecha M, Feliú A, Iñigo PM, Mestre L, Carrillo-Salinas FJ, Guaza C. Cannabidiol provides long-lasting protection against the deleterious effects of inflammation in a viral model of multiple sclerosis: a role for A2A receptors. *Neurobiol Dis* (2013) 59:141–50. doi:10.1016/j.nbd.2013.06.016
42. Barnes DA, Huston M, Holmes R, Benveniste EN, Yong VW, Scholz P, et al. Induction of RANTES expression by astrocytes and astrocytoma cell lines. *J Neuroimmunol* (1996) 71:207–14. doi:10.1016/S0165-5728(96)00154-3
43. Biber K, Neumann H, Inoue K, Boddeke HW. Neuronal 'on' and 'off' signals control microglia. *Trends Neurosci* (2007) 30:596–602. doi:10.1016/j.tins.2007.08.007
44. Kettenmann H, Kirchhoff F, Verkhratsky A. Microglia: new roles for the synaptic stripper. *Neuron* (2013) 77:10–8. doi:10.1016/j.neuron.2012.12.023
45. Wake H, Moorhouse AJ, Miyamoto A, Nabekura J. Microglia: actively surveying and shaping neuronal circuit structure and function. *Trends Neurosci* (2012) 36:209–17. doi:10.1016/j.tins.2012.11.007
46. Pocock JM, Kettenmann H. Neurotransmitter receptors on microglia. *Trends Neurosci* (2007) 30:527–35. doi:10.1016/j.tins.2007.07.007
47. Wong WT, Wang M, Li W. Regulation of microglia by ionotropic glutamatergic and GABAergic neurotransmission. *Neuron Glia Biol* (2011) 7:41–6. doi:10.1017/S1740925X11000123
48. Mennicken F, Maki R, de Souza EB, Quirion R. Chemokines and chemokine receptors in the CNS: a possible role in neuroinflammation and patterning. *Trends Pharmacol Sci* (1999) 20:73–8. doi:10.1016/S0165-6147(99)01308-5
49. Avdoshina V, Biggio F, Palchik G, Campbell LA, Mocchetti I. Morphine induces the release of CCL5 from astrocytes: potential neuroprotective mechanism against the HIV protein gp120. *Glia* (2010) 58:1630–9. doi:10.1002/glia.21035
50. Campbell LA, Avdoshina V, Rozzi S, Mocchetti I. CCL5 and cytokine expression in the rat brain: differential modulation by chronic morphine and morphine withdrawal. *Brain Behav Immun* (2013) 34:130–40. doi:10.1016/j.bbi.2013.08.006
51. Reynolds JL, Ignatowski TA, Sud R, Spengler RN. An antidepressant mechanism of desipramine is to decrease tumor necrosis factor-alpha production culminating in increases in noradrenergic neurotransmission. *Neuroscience* (2005) 133:519–31. doi:10.1016/j.neuroscience.2005.02.023
52. Szelényi J, Vizi ES. The catecholamine-cytokine balance: interaction between the brain and the immune system. *Ann N Y Acad Sci* (2007) 1113:311–24. doi:10.1196/annals.1391.026
53. O'Sullivan JB, Ryan KM, Harkin A, Connor TJ. Noradrenergic reuptake inhibitors inhibit expression of chemokines IP-10 and RANTES and cell adhesion molecules VCAM-1 and ICAM-1 in the CNS following a systemic inflammatory challenge. *J Neuroimmunol* (2010) 220:34–42. doi:10.1016/j.jneuroim.2009.12.007
54. Yester JW, Bryan L, Waters MR, Mierzanski B, Biswas DD, Gupta AS, et al. Sphingosine-1-phosphate inhibits IL-1-induced expression of C-C motif ligand 5 via c-Fos-dependent suppression of IFN-β amplification loop. *FASEB J* (2015) 29:4853–65. doi:10.1096/fj.15-275180
55. Croitoru-Lamoury J, Guillemin GJ, Dormont D, Brew BJ. Quinolinic acid up-regulates chemokine production and chemokine receptor expression in astrocytes. *Adv Exp Med Biol* (2003) 527:37–45. doi:10.1002/glia.10175
56. Barnes PJ. Cytokine-directed therapies for the treatment of chronic airway diseases. *Cytokine Growth Factor Rev* (2003) 14:511–22. doi:10.1016/S1359-6101(03)00058-3
57. Broder CC, Collman RG. Chemokine receptors and HIV. *J Leukoc Biol* (1997) 62:20–9.
58. Santoro F, Vassena L, Lusso P. Chemokine receptors as new molecular targets for antiviral therapy. *New Microbiol* (2004) 27(2 Suppl 1):17–29.
59. Yost R, Pasquale TR, Sahloff EG. Maraviroc: a coreceptor CCR5 antagonist for management of HIV infection. *Am J Health Syst Pharm* (2009) 66:715–26. doi:10.2146/ajhp080206
60. Dorf ME, Berman MA, Tanabe S, Heesen M, Luo Y. Astrocytes express functional chemokine receptors. *J Neuroimmunol* (2000) 111:109–21. doi:10.1016/S0165-5728(00)00371-4
61. Luo Y, Berman MA, Zhai Q, Fischer FR, Abromson-Leeman SR, Zhang Y, et al. RANTES stimulates inflammatory cascades and receptor modulation in murine astrocytes. *Glia* (2002) 39:19–30. doi:10.1002/glia.10079
62. Flynn G, Maru S, Loughlin J, Romero IA, Male D. Regulation of chemokine receptor expression in human microglia and astrocytes. *J Neuroimmunol* (2003) 136:84–93. doi:10.1016/S0165-5728(03)00009-2
63. McKimmie CS, Graham GJ. Astrocytes modulate the chemokine network in a pathogen-specific manner. *Biochem Biophys Res Commun* (2010) 394:1006–11. doi:10.1016/j.bbrc.2010.03.111
64. Stigliani S, Zappettini S, Raiteri L, Passalacqua M, Melloni E, Venturi C, et al. Glia re-sealed particles freshly prepared from adult rat brain are competent for exocytotic release of glutamate. *J Neurochem* (2006) 96:656–68. doi:10.1111/j.1471-4159.2005.03631.x

65. Pittaluga A. Presynaptic release-regulating mGlu1 receptors in central nervous system. *Front Pharmacol* (2016) 7:295. doi:10.3389/fphar.2016.00295
66. Di Prisco S, Summa M, Chellakudam V, Rossi PI, Pittaluga A. RANTES-mediated control of excitatory amino acid release in mouse spinal cord. *J Neurochem* (2012) 121:428–37. doi:10.1111/j.1471-4159.2012.07720.x
67. Hesselgesser J, Halks-Miller M, Del Vecchio V, Peiper SC, Hoxie J, Kolson DL, et al. CD4-independent association between HIV-1 gp120 and CXCR4: functional chemokine receptors are expressed in human neurons. *Curr Biol* (1997) 7:112–21. doi:10.1016/S0960-9822(06)00055-8
68. Oh SB, Tran PB, Gillard SE, Hurley RW, Hammond DL, Miller RJ. Chemokines and glycoprotein120 produce pain hypersensitivity by directly exciting primary nociceptive neurons. *J Neurosci* (2001) 21(14):5027–35.
69. Cowell RM, Silverstein FS. Developmental changes in the expression of chemokine receptor CCR1 in the rat cerebellum. *J Comp Neurol* (2003) 457(1):7–23. doi:10.1016/j.jneuroim.2006.01.005
70. Cowell RM, Xu H, Galasso JM, Silverstein FS. Hypoxic-ischemic injury induces macrophage inflammatory protein-1alpha expression in immature rat brain. *Stroke* (2002) 33(3):795–801. doi:10.1161/hs0302.103740
71. Musante V, Longordo F, Neri E, Pedrazzi M, Kalfas F, Severi P, et al. RANTES modulates the release of glutamate in human neocortex. *J Neurosci* (2008) 28:12231–40. doi:10.1523/JNEUROSCI.3212-08.2008
72. Di Prisco S, Meregá E, Pittaluga A. Functional adaptation of presynaptic chemokine receptors in EAE mouse central nervous system. *Synapse* (2014) 68:529–35. doi:10.1002/syn.21774
73. Tanabe S, Heesen M, Berman MA, Fischer MB, Yoshizawa I, Luo Y, et al. Murine astrocytes express a functional chemokine receptor. *J Neurosci* (1997) 17(17):6522–8. doi:10.1016/S0165-5728(00)00371-4
74. Han Y, Wang J, Zhou Z, Ransohoff RM. TGFbeta1 selectively up-regulates CCR1 expression in primary murine astrocytes. *Glia* (2000) 30:1–10. doi:10.1002/(SICI)1098-1136(200003)30:13.0
75. Andjelkovic AV, Kerkovich D, Shanley J, Pulliam L, Pachter JS. Expression of binding sites for beta chemokines on human astrocytes. *Glia* (1999) 28(3):225–35. doi:10.1002/(SICI)1098-1136(199912)28:3
76. Croitoru-Lamoury J, Guillemin GJ, Boussin FD, Mognetti B, Bigout LI, Chéret A, et al. Expression of chemokines and their receptors in human and simian astrocytes: evidence for a central role of TNF alpha and IFN gamma in CXCR4 and CCR5 modulation. *Glia* (2003) 41(4):354–70. doi:10.1002/glia.10181
77. Patti L, Raiteri L, Grilli M, Zappettini S, Bonanno G, Marchi M. Evidence that alpha7 nicotinic receptor modulates glutamate release from mouse neocortical gliosomes. *Neurochem Int* (2007) 51:1–7. doi:10.1016/j.neuint.2007.03.003
78. Coughlan CM, McManus CM, Sharron M, Gao Z, Murphy D, Jaffer S, et al. Expression of multiple functional chemokine receptors and monocyte chemoattractant protein-1 in human neurons. *Neuroscience* (2000) 97:591–600. doi:10.1016/S0306-4522(00)00024-5
79. Matsui M, Weaver J, Proudfoot AE, Wujek JR, Wei T, Richer E, et al. Treatment of experimental autoimmune encephalomyelitis with the chemokine receptor antagonist Met-RANTES. *J Neuroimmunol* (2002) 128:16–22. doi:10.1016/S0165-5728(02)00121-2
80. Tran PB, Miller RJ. Chemokine receptors in the brain: a developing story. *J Comp Neurol* (2003) 457:1–6. doi:10.1002/cne.10546
81. Zhang N, Rogers TJ, Caterina M, Oppenheim JJ. Proinflammatory chemokines, such as C-C chemokine ligand 3, desensitize mu-opioid receptors on dorsal root ganglia neurons. *J Immunol* (2004) 173:594–9. doi:10.4049/jimmunol.173.1.594
82. Grilli M, Raiteri L, Pittaluga A. Somatostatin inhibits glutamate release from mouse cerebrocortical nerve endings through presynaptic sst2 receptor linked to the adenylyl cyclase-protein kinase A pathway. *Neuropharmacology* (2004) 46:388–96. doi:10.1016/j.neuropharm.2003.09.012
83. Summa M, Di Prisco S, Grilli M, Usai C, Marchi M, Pittaluga A. Presynaptic mGlu7 receptors control GABA release in mouse hippocampus. *Neuropharmacology* (2013) 66:215–24. doi:10.1016/j.neuropharm.2012.04.020
84. Zucchini S, Pittaluga A, Summa M, Brocca-Cofano E, Fabris M, De Michele R, et al. Seizure susceptibility in Tat-transgenic mice: implications for the role of Tat in human immunodeficiency virus type 1 associated encephalopathy. *Neurobiol Dis* (2013) 62:354–9. doi:10.1016/j.nbd.2013.02.004
85. Zappettini S, Grilli M, Olivero G, Chen J, Padolecchia C, Pittaluga A, et al. Nicotinic alpha7 receptor activation selectively potentiates the function of NMDA receptors in glutamatergic terminals of the nucleus accumbens. *Front Cell Neurosci* (2014) 8:332. doi:10.3389/fncel.2014.00332
86. Wells TN, Power CA, Shaw JP, Proudfoot AE. Chemokine blockers-therapeutics in the making? *Trends Pharmacol Sci* (2006) 27:41–7. doi:10.1016/j.tips.2005.11.001
87. Heath H, Qin S, Rao P, Wu L, LaRosa G, Kassam N, et al. Chemokine receptor usage by human eosinophils. The importance of CCR3 demonstrated using an antagonistic monoclonal antibody. *J Clin Invest* (1997) 99:178–84. doi:10.1172/JCI119145
88. Di Prisco S, Meregá E, Milanese M, Summa M, Casazza S, Raffaghello L, et al. CCL5-Glutamate interaction in central nervous system: early and acute presynaptic defects in EAE mice. *Neuropharmacology* (2013) 75:337–46. doi:10.1016/j.neuropharm.2013.07.037
89. van Veen T, Nielsen J, Berkhof J, Barkhof F, Kamphorst W, Bö L, et al. CCL5 and CCR5 genotypes modify clinical, radiological and pathological features of multiple sclerosis. *J Neuroimmunol* (2007) 190:157–64. doi:10.1016/j.jneuroim.2007.08.005
90. Subileau EA, Rezaie P, Davies HA, Colyer FM, Greenwood J, Male DK, et al. Expression of chemokines and their receptors by human brain endothelium: implications for multiple sclerosis. *J Neuropathol Exp Neurol* (2009) 68:227–40. doi:10.1097/NEN.0b013e318197eca7
91. Comini-Frota ER, Teixeira AL, Angelo JP, Andrade MV, Brum DG, Kaimen-Maciel DR, et al. Evaluation of serum levels of chemokines during interferon-beta treatment in multiple sclerosis patients: a 1-year, observational cohort study. *CNS Drugs* (2011) 25:971–81. doi:10.2165/11595060-000000000-00000
92. Guo MF, Meng J, Li YH, Yu JZ, Liu CY, Feng L, et al. The inhibition of Rho kinase blocks cell migration and accumulation possibly by challenging inflammatory cytokines and chemokines on astrocytes. *J Neurol Sci* (2014) 343:69–75. doi:10.1016/j.jns.2014.05.034
93. Furlan R, Rovaris M, Martinelli Boneschi F, Khademi M, Bergami A, Gironi M, et al. Immunological patterns identifying disease course and evolution in multiple sclerosis patients. *J Neuroimmunol* (2005) 165:192–200. doi:10.1016/j.jneuroim.2005.04.012
94. Glabinski AR, Tuohy VK, Ransohoff RM. Expression of chemokines RANTES, MIP-1alpha and GRO-alpha correlates with inflammation in acute experimental autoimmune encephalomyelitis. *Neuroimmunomodulation* (1998) 5:166–71. doi:10.1159/000026333
95. Glabinski AR, Bielecki B, Ransohoff RM. Chemokine upregulation follows cytokine expression in chronic relapsing experimental autoimmune encephalomyelitis. *Scand J Immunol* (2003) 58:81–8. doi:10.1046/j.1365-3083.2003.01285.x
96. Jalonen TO, Pulkkinen K, Ukkonen M, Saarela M, Elovaara I. Differential intracellular expression of CCR5 and chemokines in multiple sclerosis subtypes. *J Neurol* (2002) 249:576–83. doi:10.1007/s004150200067
97. Ransohoff RM, Wei T, Pavelko KD, Lee JC, Murray PD, Rodriguez M. Chemokine expression in the central nervous system of mice with a viral disease resembling multiple sclerosis: roles of CD4+ and CD8+ T cells and viral persistence. *J Virol* (2002) 76:2217–24. doi:10.1128/jvi.76.5.2217-2224.2002
98. Bartosik-Psujek H, Stelmasiak Z. Correlations between IL-4, IL-12 levels and CCL2, CCL5 levels in serum and cerebrospinal fluid of multiple sclerosis patients. *J Neural Transm (Vienna)* (2005) 112:797–803. doi:10.1007/s00702-004-0225-9
99. Navratilova Z. Polymorphisms in CCL2 and CCL5 chemokines/chemokine receptors genes and their association with diseases. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* (2006) 150:191–204. doi:10.5507/bp.2006.028
100. Bonfiglio T, Olivero G, Meregá E, Di Prisco S, Padolecchia C, Grilli M, et al. Prophylactic versus therapeutic fingolimod: restoration of presynaptic defects in mice suffering from experimental autoimmune encephalomyelitis. *PLoS One* (2017) 12:e0170825. doi:10.1371/journal.pone.0170825
101. Calabresi PA, Martin R, Jacobson S. Chemokines in chronic progressive neurological diseases: HTLV-1 associated myelopathy and multiple sclerosis. *J Neurovirol* (1999) 5:102–8. doi:10.3109/13550289909029751
102. Haase CG, Schmidt S, Faustmann PM. Frequencies of the G-protein beta3 subunit C825T polymorphism and the delta 32 mutation of the chemokine receptor-5 in patients with multiple sclerosis. *Neurosci Lett* (2002) 330:293–5. doi:10.1016/S0304-3940(02)00807-8
103. Krakauer M, Sorensen PS, Khademi M, Olsson T, Sellebjerg F. Dynamic T-lymphocyte chemokine receptor expression induced by interferon-beta

- therapy in multiple sclerosis. *Scand J Immunol* (2006) 64:155–63. doi:10.1111/j.1365-3083.2006.01788.x
104. Uzawa A, Mori M, Hayakawa S, Masuda S, Nomura F, Kuwabara S. Expression of chemokine receptors on peripheral blood lymphocytes in multiple sclerosis and neuromyelitis optica. *BMC Neurol* (2010) 10:113. doi:10.1186/1471-2377-10-113
 105. Trebst C, Staugaitis SM, Tucky B, Wei T, Suzuki K, Aldape KD, et al. Chemokine receptors on infiltrating leucocytes in inflammatory pathologies of the central nervous system (CNS). *Neuropathol Appl Neurobiol* (2003) 29:584–95. doi:10.1046/j.0305-1846.2003.00507.x
 106. Savarin-Vuillaud C, Ransohoff RM. Chemokines and chemokine receptors in neurological disease: raise, retain, or reduce? *Neurotherapeutics* (2007) 4:590–601. doi:10.1016/j.nurt.2007.07.004
 107. Martin-Blondel G, Brassat D, Bauer J, Lassmann H, Liblau RS. CCR5 blockade for neuroinflammatory diseases – beyond control of HIV. *Nat Rev Neurol* (2016) 12:95–105. doi:10.1038/nrnneurol.2015.248
 108. Gu SM, Park MH, Yun HM, Han SB, Oh KW, Son DJ, et al. CCR5 knockout suppresses experimental autoimmune encephalomyelitis in C57BL/6 mice. *Oncotarget* (2016) 7:15382–93. doi:10.18632/oncotarget.8097
 109. Di Prisco S, Meregá E, Bonfiglio T, Olivero G, Cervetto C, Grilli M, et al. Presynaptic, release-regulating mGlu2-preferring and mGlu3-preferring autoreceptors in CNS: pharmacological profiles and functional roles in demyelinating disease. *Br J Pharmacol* (2016) 173:1465–77. doi:10.1111/bph.13442
 110. Di Prisco S, Olivero G, Meregá E, Bonfiglio T, Marchi M, Pittaluga A. CXCR4 and NMDA receptors are functionally coupled in rat hippocampal noradrenergic and glutamatergic nerve endings. *J Neuroimmune Pharmacol* (2016) 11:645–56. doi:10.1007/s11481-016-9677-6
 111. Pittaluga A, Asaro D, Pellegrini G, Raiteri M. Studies on [³H]GABA and endogenous GABA release in rat cerebral cortex suggest the presence of autoreceptors of the GABA_B type. *Eur J Pharmacol* (1987) 144:45–52. doi:10.1016/0014-2999(87)90007-0
 112. Pittaluga A, Thellung S, Maura G, Raiteri M. Characterization of two central AMPA-preferring receptors having distinct location, function and pharmacology Naunyn-Schmiedeb. *Arch Pharmacol* (1994) 349:555–8. doi:10.1007/BF01258458
 113. Pittaluga A, Feligioni M, Longordo F, Luccini E, Raiteri M. Trafficking of presynaptic AMPA receptors mediating neurotransmitter release: neuronal selectivity and relationships with sensitivity to cyclothiazide. *Neuropharmacology* (2006) 50:286–96. doi:10.1016/j.neuropharm.2005.09.004
 114. Musante V, Neri E, Feligioni M, Puliti A, Pedrazzi M, Conti V, et al. Presynaptic mGlu1 and mGlu5 autoreceptors facilitate glutamate exocytosis from mouse cortical nerve endings. *Neuropharmacology* (2008) 55:474–82. doi:10.1016/j.neuropharm.2008.06.056
 115. Longordo F, Feligioni M, Chiamonte G, Sbaffi PF, Raiteri M, Pittaluga A. The human immunodeficiency virus-1 protein transactivator of transcription up-regulates N-methyl-D-aspartate receptor function by acting at metabotropic glutamate receptor 1 receptors coexisting on human and rat brain noradrenergic neurons. *J Pharmacol Exp Ther* (2006) 317:1097–105. doi:10.1124/jpet.105.099630
 116. Luccini E, Musante V, Neri E, Brambilla Bas M, Severi P, Raiteri M, et al. Functional interactions between presynaptic NMDA receptors and metabotropic glutamate receptors co-expressed on rat and human noradrenergic terminals. *Br J Pharmacol* (2007) 151:1087–94. doi:10.1038/sj.bjp.0707280
 117. Ganor Y, Besser M, Ben-Zayak N, Unger T, Levite M. Human T-cell express a functional ionotropic glutamate receptor GLUR3, and glutamate by itself triggers integrin-mediated adhesion to laminin and fibronectin and chemotactic migration. *J Immunol* (2003) 170:4362–72. doi:10.4049/jimmunol.170.8.4362
 118. Chiocchetti A, Miglio G, Mesturini R, Varsaldi F, Mocellin M, Orilieri E, et al. Group I mGlu receptor stimulation inhibits activation-induced cell death of human T lymphocytes. *Br J Pharmacol* (2006) 148:760–8. doi:10.1038/sj.bjp.0706746
 119. Sarchielli P, Di Filippo M, Candelieri A, Chasserini D, Mattioni A, Tenaglia S, et al. Expression of ionotropic glutamate receptor GLUR3 and effects of glutamate on MBP and MOG-specific lymphocyte activation and chemotactic migration in multiple sclerosis patients. *J Neuroimmunol* (2007) 188:146–58. doi:10.1016/j.jneuroim.2007.05.021
 120. Schwartz M, Shaked I, Fisher J, Mizrahi T, Shori H. Protective autoimmunity against the enemy within: fighting glutamate toxicity. *Trends Neurosci* (2003) 26:297–302. doi:10.1016/S0166-2236(03)00126-7
 121. Lanz M, Hahn HK, Hildebrandt H. Brain atrophy and cognitive impairment in multiple sclerosis: a review. *J Neurol* (2007) 254:II43–8. doi:10.1007/s00415-007-2011-8
 122. Werner P, Pitt D, Raine CS. Multiple sclerosis: altered glutamate homeostasis in lesions correlates with oligodendrocyte and axonal damage. *Ann Neurol* (2001) 50:169–80. doi:10.1002/ana.1077
 123. Sulkowski G, Dabrowska-Bouta B, Kwiatkowska-Patzer B, Strużyńska L. Alterations in glutamate transport and group I metabotropic glutamate receptors in the rat brain during acute phase of experimental autoimmune encephalomyelitis. *Folia Neuropathol* (2009) 47:329–37.
 124. Marte A, Cavallero A, Morando S, Uccelli A, Raiteri M, Fedele E. Alterations of glutamate release in the spinal cord of mice with experimental autoimmune encephalomyelitis. *J Neurochem* (2010) 115:343–52. doi:10.1111/j.1471-4159.2010.06923.x
 125. Centonze D, Muzio L, Rossi S, Furlan R, Bernardi G, Martino G. The link between inflammation, synaptic transmission and neurodegeneration in multiple sclerosis. *Cell Death Differ* (2009) 17:1083–91. doi:10.1038/cdd.2009.179
 126. Di Prisco S, Meregá E, Lanfranco M, Casazza S, Uccelli A, Pittaluga A. Acute desipramine restores presynaptic cortical defects in murine experimental autoimmune encephalomyelitis by suppressing central CCL5 overproduction. *Br J Pharmacol* (2014) 171:2457–67. doi:10.1111/bph.12631
 127. Vilcaes AA, Furlan G, Roth GA. Inhibition of Ca²⁺-dependent glutamate release from cerebral cortex synaptosomes of rats with experimental autoimmune encephalomyelitis. *J Neurochem* (2009) 108:881–90. doi:10.1111/j.1471-4159.2008.05809.x
 128. Pittaluga A, Pattarini R, Feligioni M, Raiteri M. NMDA receptors mediating hippocampal noradrenaline and striatal dopamine release display differential sensitivity to quinolinic acid, the HIV-1 envelope protein gp120, external pH and PKC inhibition. *J Neurochem* (2001) 76:139–48. doi:10.1046/j.1471-4159.2001
 129. Musante V, Summa M, Cunha RA, Raiteri M, Pittaluga A. Presynaptic glycine GlyT1 transporter – NMDA receptor interaction: relevance to NMDA autoreceptor activation in the presence of Mg²⁺ ions. *J Neurochem* (2011) 117:516–27. doi:10.1111/j.1471-4159.2011.07223.x
 130. Fazio F, Notartomaso S, Aronica E, Storto M, Battaglia G, Vieira E, et al. Switch in the expression of mGlu1 and mGlu5 metabotropic glutamate receptors in the cerebellum of mice developing experimental autoimmune encephalomyelitis and in autaptic cerebellar samples from patients with multiple sclerosis. *Neuropharmacology* (2008) 55:491–9. doi:10.1016/j.neuropharm.2008.06.066
 131. Fallarino F, Volpi C, Fazio F, Notartomaso S, Vacca C, Busceti C, et al. Metabotropic glutamate receptor-4 modulates adaptive immunity and restrains neuroinflammation. *Nat Med* (2010) 16:897–902. doi:10.1038/nm.2183
 132. Geurts JJ, Wolswijk G, Bö L, van der Valk P, Polman CH, Troost D, et al. Altered expression patterns of group I and II metabotropic glutamate receptors in multiple sclerosis. *Brain* (2003) 126(Pt 8):1755–66. doi:10.1093/brain/awg179
 133. Taylor DL, Jones F, Kubota ES, Pocock JM. Stimulation of microglial metabotropic glutamate receptor mGlu2 triggers tumor necrosis factor alpha-induced. *J Neurosci* (2005) 25:2952–64. doi:10.1523/JNEUROSCI.4456-04.2005
 134. Pinteaux-Jones F, Sevastou IG, Fry VA, Heales S, Baker D, Pocock JM. Myelin-induced microglial neurotoxicity can be controlled by microglial metabotropic glutamate receptors. *J Neurochem* (2008) 106:442–54. doi:10.1111/j.1471-4159.2008.05426.x
 135. Grasselli G, Rossi S, Musella A, Gentile A, Loizzo S, Muzio L, et al. Abnormal NMDA receptor function exacerbates experimental autoimmune encephalomyelitis. *Br J Pharmacol* (2013) 168:502–17. doi:10.1111/j.1476-5381.2012.02178.x
 136. Nisticò R, Mango D, Mandolesi G, Piccinin S, Berretta N, Pignatelli M, et al. Inflammation subverts hippocampal synaptic plasticity in experimental multiple sclerosis. *PLoS One* (2013) 8:e54666. doi:10.1371/journal.pone.0054666

137. Ohgoh M, Hanada T, Smith T, Hashimoto T, Ueno M, Yamanishi Y, et al. Altered expression of glutamate transporters in experimental autoimmune encephalomyelitis. *J Neuroimmunol* (2002) 125:170–8. doi:10.1016/S0165-5728(02)00029-2
138. Trapp BD, Nave KA. Multiple sclerosis: an immune or neurodegenerative disorder? *Annu Rev Neurosci* (2008) 31:247–69. doi:10.1146/annurev.neuro.30.051606.094313
139. Centonze D, Muzio L, Rossi S, Cavasinni F, De Chiara V, Bergami A, et al. Inflammation triggers synaptic alteration and degeneration in experimental autoimmune encephalomyelitis. *J Neurosci* (2009) 29:3442–52. doi:10.1523/JNEUROSCI.5804-08.2009
140. Rao SM, Leo GJ, Bernardin L, Unverzagt F. Cognitive dysfunction in multiple sclerosis. I. Frequency, patterns, and prediction. *Neurology* (1991) 41:685–91. doi:10.1212/WNL.41.5.685
141. Siegert EJ, Abernethy DA. Depression in multiple sclerosis: a review. *J Neurol Neurosurg Psychiatry* (2005) 76:469–75. doi:10.1136/jnnp.2004.054635
142. Batista S, Zivadinov R, Hoogs M, Bergsland N, Heininen-Brown M, Dwyer MG, et al. Basal ganglia, thalamus and neocortical atrophy predicting slowed cognitive processing in multiple sclerosis. *J Neurol* (2011) 259:139–46. doi:10.1007/s00415-011-6147-1
143. Schubert DS, Foliart RH. Increased depression in multiple sclerosis patients: a meta-analysis. *Psychosomatics* (1993) 34:124–30. doi:10.1016/S0033-3182(93)71902-7
144. Pittaluga A, Raiteri L, Longordo F, Luccini E, Barbiero VS, Racagni G, et al. Antidepressant treatments and function of glutamate ionotropic receptors mediating amine release in hippocampus. *Neuropharmacology* (2007) 53:27–36. doi:10.1016/j.neuropharm.2007.04.006
145. Summa M, Di Prisco S, Grilli M, Marchi M, Pittaluga A. Hippocampal AMPA autoreceptors positively coupled to NMDA autoreceptors traffic in a constitutive manner and undergo adaptative changes following enriched environment training. *Neuropharmacology* (2011) 61:1282–90. doi:10.1016/j.neuropharm.2011.07.032
146. Grilli M, Summa M, Salmone A, Olivero G, Zappettini S, Di Prisco S, et al. In vitro exposure to nicotine induces endocytosis of presynaptic AMPA receptors modulating dopamine release in rat nucleus accumbens nerve terminals. *Neuropharmacology* (2012) 63:916–26. doi:10.1016/j.neuropharm.2012.06.049
147. O'Sullivan JB, Ryan KM, Curtin NM, Harkin A, Connor TJ. Noradrenaline reuptake inhibitors limit neuroinflammation in rat cortex following a systemic inflammatory challenge: implications for depression and neurodegeneration. *Int J Neuropsychopharmacol* (2009) 12:687–99. doi:10.1017/S146114570800967X
148. Li QQ, Burt DR, Bever CT. Glatiramer acetate inhibition of tumor necrosis factor- α -induced RANTES expression and release from U-251 MG human astrocytic cells. *J Neurochem* (2001) 77:1208–17. doi:10.1046/j.1471-4159.2001.00327.x
149. Palazuelos J, Davoust N, Julien B, Hatterer E, Aguado T, Mechoulam R, et al. The CB(2) cannabinoid receptor controls myeloid progenitor trafficking: involvement in the pathogenesis of an animal model of multiple sclerosis. *J Biol Chem* (2008) 283:13320–9. doi:10.1074/jbc.M70796020
150. Stasiolek M, Linker RA, Hayardeny L, Bar Ilan O, Gold R. Immune parameters of patients treated with laquinimod, a novel oral therapy for the treatment of multiple sclerosis: results from a double-blind placebo-controlled study. *Immun Inflamm Dis* (2015) 3:45–55. doi:10.1002/iid3.42
151. Schaier M, Vorwalder S, Sommerer C, Dikow R, Hug F, Gross ML, et al. Role of FTY720 on M1 and M2 macrophages, lymphocytes, and chemokines in 5/6 nephrectomized rats. *Am J Physiol Renal Physiol* (2009) 297:F769–80. doi:10.1152/ajprenal.90530.2008
152. Araque A, Parpura V, Sanzgiri RP, Haydon PG. Tripartite synapses: glia, the unacknowledged partner. *Trends Neurosci* (1999) 22:208–15. doi:10.1016/S0166-2236(98)01349
153. Shijie J, Takeuchi H, Yawata I, Harada Y, Sonobe Y, Doi Y, et al. Blockade of glutamate release from microglia attenuates experimental autoimmune encephalomyelitis in mice. *Tohoku J Exp Med* (2009) 217(2):87–92. doi:10.1620/tjem.217.87
154. Gentile A, Rossi S, Studer V, Motta C, De Chiara V, Musella A, et al. Glatiramer acetate protects against inflammatory synaptopathy in experimental autoimmune encephalomyelitis. *J Neuroimmune Pharmacol* (2013) 8:651–63. doi:10.1007/s11481-013-9436-x
155. de Lago E, Moreno-Martet M, Cabranes A, Ramos JA, Fernández-Ruiz J. Cannabinoids ameliorate disease progression in a model of multiple sclerosis in mice, acting preferentially through CB1 receptor-mediated anti-inflammatory effects. *Neuropharmacology* (2012) 62(7):2299–308. doi:10.1016/j.neuropharm.2012.01.030

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