SHORT COMMUNICATION

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cDNA cloning of a novel human gene *NAKAP95*, neighbor of A-kinase anchoring protein 95 (AKAP95) on chromosome 19p13.11–p13.12 region

Received: July 13, 1999 / Accepted: September 4, 1999

Abstract A-kinase anchoring protein 95 (AKAP95) is a nuclear protein which binds to the regulatory subunit (RII) of cyclic adenosine monophosphate (cAMP)-dependent protein kinase (PKA) and to DNA. A novel nuclear human gene which shares sequence homology with the human AKAP95 gene was identified by a nuclear transportation trap method. By polymerase chain reaction (PCR)-based analysis with both a human/rodent monochromosomal hybrid cell panel and a radiation hybrid panel, the gene was mapped to the chromosome 19p13.11-p13.12 region between markers WI-4669 and CHLC.GATA27C12. Furthermore, alignment with genomic sequences revealed that the gene and human AKAP95 resided tandemly only approximately 250bp apart from each other. We designated this gene as neighbor of AKAP95 (NAKAP95). The exonintron structure of NAKAP95 and AKAP95 was conserved, indicating that they may have evolved by gene duplication. The predicted protein product of the NAKAP95 gene consists of 646 amino acid residues, and NAKAP95 and AKAP95 had an overall 40% similarity, both having a potential nuclear localizing signal and two C2H2 type zinc finger motifs. The putative RII binding motif in AKAP95 was not conserved in NAKAP95. A reverse transcription coupled (RT)-PCR experiment revealed that the NAKAP95 gene was transcribed ubiquitously in various human tissues.

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N. Ueki · K. Yano · Y. Masuho · M. Muramatsu (\boxtimes) Helix Research Institute, 1532-3 Yana, Kisarazu, Chiba 292-0812, Japan Tel. +81-438-52-3966; Fax +81-438-52-3952 e-mail: mmasaaki@hri.co.jp Key words Cyclic AMP-dependent protein kinase (PKA) \cdot A-kinase anchoring proteins (AKAPs) \cdot AKAP95 \cdot Chromosome 19p13.11–p13.12 \cdot RH mapping \cdot Genomic structure \cdot Gene duplication

Introduction

A large number of hormones and neurotransmitters utilize cyclic adenosine monophosphate (cAMP) as an intracellular second messenger. Cyclic AMP regulates a number of key cellular processes such as metabolism, gene regulation, cell growth, cell differentiation, ion channel conductivity, and release of synaptic vesicles (Krebs and Beavo, 1979; Boynton and Whitfield, 1983; Edelman et al. 1987; Roesler et al. 1988; Taylor et al. 1990; McKnight 1991). The main intracellular target for cAMP in mammalian cells is cAMPdependent protein kinase (PKA or A-kinase). PKA type II is directed to different subcellular loci through interaction of the RII subunits with A-kinase anchoring proteins (AKAPs) (Scott and Macartney, 1994; Rubin 1994; Hausken et al. 1996; Hausken and Scotte, 1996; Faux and Scott 1996). A number of different AKAPs which direct different compartmentalizations have been found: AKAP79/75 direct the RII to postsynaptic densities and cortical actin (Carr et al. 1992; Li et al. 1996), AKAP250/ Gravin to filopodia (Nauert et al. 1997), AKAP350 to centrosomes (Schmidt et al. 1999), AKAP100 to sarcoplasmic reticulum (McCartney et al. 1995), AKAP220 to peroxisome (Lester et al. 1996), AKAP85 to Golgi apparatus (Kerver et al. 1993), and AKAP84/149 to mitochondria (Chen et al. 1997).

AKAP95 was originally isolated by an interaction cloning strategy with RII α as a probe from a rat pituitary (GH₄C₁) cDNA library (Coghlan et al. 1994). The rat AKAP95 contained both RII and DNA binding domains. The AKAP95 was detected in a nuclear matrix fraction, and immunofluorescence, using purified anti-AKAP95 antibodies, revealed distinct nuclear staining in a variety of cell types (Coghlan et al. 1994). It is proposed that AKAP95

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could play a role in targeting type II PKA for cAMPresponsive nuclear events. Recently, human AKAP95 was identified and sequenced, and was shown to be 89% homologous to rat AKAP95 (Eide et al. 1998).

We have recently developed a screening method, designated nuclear transportation trap (NTT), to systematically isolate nuclear proteins (Ueki et al. 1998). Using this method, several novel nuclear genes were isolated, such as *CLIM1/CLIM2*, *PIAS3*, and *HFB30* (Ueki et al. 1999a,b,c). We have also isolated a partial cDNA clone which had a sequence homologous to the human *AKAP95* gene. We describe here the complete cDNA sequence, expression profile, chromosomal assignment, and genomic structure of the gene, *NAKAP95*.

Cloning of human NAKAP95 gene

A partial cDNA clone (initially called HFB2018) was isolated from a human fetal library, using the NTT method (Ueki et al. 1998). BLAST search revealed that HFB2018 was most homologous to human AKAP95 (Eide et al. 1998). Specific primers were designed, according to the HFB2018 sequence, to obtain a full-length cDNA from a human fetal brain library, using GeneTrapper (GIBCO BRL, Gaithersburg, MD, USA). The resultant cDNA was 2057 bp in length and contained an open reading frame of 646 amino acids (Fig. 1a). The nucleotide sequence of the cDNA will appear in GenBank/EMBL/DDBJ databases under the accession number, AB025905. Since the gene was found to reside next to the human AKAP95 gene (see below and Fig. 2a) we designated this clone as neighbor of AKAP95 (NAKAP95). The predicted NAKAP protein initiated from the first putative initiation ATG codon was in accordance with Kozak's rule (Kozak 1987). A canonical polyadenylation signal, AATAAA, was located 14bp upstream of a poly(A) (Fig. 1a). The alignment of predicted amino acid sequences of human AKAP95, rat AKAP95, and human NAKAP95 proteins is shown in Fig. 1b. Human NAKAP95 and AKAP95 are 30% identical (40% similar) at the amino acid level. NAKAP95 and AKAP95 possess two C2H2 type zinc finger motifs at a similar position. The regions that included the zinc finger motifs showed the highest identity between the two proteins. A putative nuclear localization signal KKKKRK was found at residues 274–279 (Fig. 1a). From the alignment, the putative RII-binding region of AKAP95 (Coghlan et al. 1994; Hausken and Scott 1996) was not conserved in NAKAP95. Therefore, whether NAKAP95 can bind to RII remains elusive.

Expression profile of human NAKAP95 gene

We examined the distribution of the *NAKAP95* transcript in various human tissues by reverse transcription-coupled polymerase chain reaction (RT-PCR) as described previously (Seki et al. 1998; 1999). Primers used for the RT-PCR corresponded to the coding region of the gene (5'-TGG TGC CGC ATT TGG AGA CAG-3') and (5'-TGC CAA ACC CGA AAC CAA AGC-3'). The primer set gave a longer PCR product from genomic DNA, which was easily distinguished from the 459-bp product from the mRNA. The 459-bp PCR product was generated in all tissues examined, indicating that the transcript is ubiquitously expressed in a wide variety of human tissues (Fig. 3). Therefore, *NAKAP95* described in the present study seems to be involved in a basic house-keeping function of cells.

Chromosome mapping and genomic structure of human *NAKAP95* gene

Chromosomal assignment of the human NAKAP95 gene was done by PCR analysis of a human/rodent somatic cell hybrid panel (National Institute of General Medicine Service, Coriell Cell Repositories, Camden, NJ, USA) and a radiation hybrid panel (Genebridge 4; Research Genetics, Huntsville, AL, USA), as described previously (Saito et al. 1997; Seki et al. 1997). The human NAKAP95 specific PCR primers (5'-TCG GCT GCC CTC CCT CTT CTC-3', 5'-GGT CCG CCT CAT CTG CTT CAT-3') gave rise to an amplified product with a size of 139bp by genomic PCR. First, the specific amplified product for human was detected only from the hybrid containing human chromosome 19 (data not shown). Further mapping analysis, using a radiation hybrid panel with the same primer set, was done. Statistical analysis of the radiation hybrid data was performed using the RHMAPPER software package (http:// carbon.wi.mit.edu:8000/cgi-bin/contig/rhmapper.pl). The data vector for the human NAKAP95 gene was 000000101 0001000010 1000111110 0001010000 1100010101 1011010001 0001000110 1101011001 0001011100 001 and the consequent report indicated that the gene was placed to 3.89 cR distal from the marker CHLC.GATA27C12 (lod > 3.0). The region including the marker was cytogenetically mapped to the 19p13.11-p13.12 region (Fig. 2a). Since human NAKAP95 and AKAP95 genes have been mapped to the same region, we then searched for genomic sequences in the public database, and found a cosmid clone which contains both genes (accession number, AC005785). The human NAKAP95 and AKAP95 resided tandemly and the first exon of human AKAP95 began 258bp after the end of the NAKAP95 gene. We have no data about the orientation of NAKAP95 and AKAP95 in chromosome 19. To confirm that human AKAP95 and NAKAP95 are juxtaposed, primers were designed between the last exon of human NAKAP95 and the first exon of AKAP95 (5'-GAC AGC CCC GAG GAG GAG AAG-3' and 5'-CCC ACC AGC AGC CCC GTT TAC-3'; product size, 668bp). Genomic PCR using the primers generated a PCR product of the expected size (data not shown), proving that human AKAP95 and NAKAP95 are next to each other. Although the sequence between the genes would provide the promoter for the AKAP95 gene, it lacked a typical TATA box or a GC-rich region.

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gagcagcagaagccggcgtcgtcggatgttgtgttgcccgccaccATGAGCTACACAGGC 60 мзүт 5 G TTTGTCCAGGGATCTGAAACCACTTTGCAGTCGACATACTCGGATACCAGCGCTCAGCCC 120 F V Q G S E T T L Q S T Y S D T 25 SAO Ρ ACCTGTGATTATGGATATGGAACTTTGGAACTCTGGGACAAATAGAGGCTACGAGGGCTAT 180 T C D Y G Y G T W N S G TNRGY 45 EGY GGCTATGGCTATGGCTATGGCCAGGATAACACCACCAACTATGGGTATGGCTATGGCCACT 240 G Y G Y G YGQDN Т Т Ν Y GYGM Т 65 А TCACACTCTTGGGAAATGCCTAGCTCTGACACAAATGCAAACACTAGTGCCTCGGGTAGC 300 S 85 Н S W Ε М Р S S D Т Ν А Ν Т S А S G S GCCAGTGCCGATTCCGTTTTATCCAGAATTAACCAGCGCTTAGATATGGTGCCGCATTTG 360 S V L S R Ν Q R \mathbf{L} D V Ρ 105 Α S А D Ι М Н L GAGACAGACATGATGCAAGGAGGCGTGTACGGCTCAGGTGGAGAAAGGTATGACTCTTAT 420 E Т D М М Q G G V Υ G S G G Ε R Y D S Y 125 GAGTCCTGCGACTCGAGGGCCGTCCTGAGTGAGCGCGACCTGTACCGGTCAGGCTATGAC 480 E S С S R A V L S Е R D Y R S D 145 D \mathbf{L} G Y TACAGCGAGCTTGACCCTGAGATGGAAATGGCCTATGAGGGCCAATACGATGCCTACCGC 540 Y Ε L D Ρ Ε М E M А Y Ε G 0 165 S Y D Α Y GACCAGTTCCGCATGCGTGGCAACGACACCTTCGGTCCCAGGGCACAGGGCTGGGCCCGG 600 D 0 F М R G Ν D T F G Ρ R 185 R Α 0 G W A R GATGCCCGGAGCGGCCGGCCAATGGCCTCAGGCTATGGGCGCATGTGGGAAGACCCCATG 660 D А R S G R Р Μ А S G Y G R М W Е D Ρ М 205 GGGGCCCGGGGCCAGTGCATGTCTGGTGCCTCTCGGCTGCCCTCCTCTTCTCCCAGAAC 720 G А R 0 С S G А S R Ρ S F S 225 G М L L 0 Ň ATCATCCCCGAGTACGGCATGTTCCAGGGCATGCGAGGTGGGGGGCGCCTTCCCCGGGCGGC 780 245 Ι Ι Ρ Ē Υ G Μ F Q G М R G G G А F Ρ G G TCCCGCTTTGGTTTCGGGTTTGGCAATGGCATGAAGCAGATGAGGCGGACCTGGAAGACC 840 S R F G \mathbf{F} G F G Ν G М Κ Q М R R Τ W Κ Т 265 TGGACCACAGCCGACTTCCGAACCAAGAAGAAGAAGAAGAAGCAGGGCGGCAGTCCTGAT 900 W Τ Т А D \mathbf{F} RТ Κ Κ Κ Κ R K0 G G S Ρ D 285 GAGCCAGATAGCAAAGCCACCCGCACGGACTGCTCGGACAACAGCGACTCAGACAATGAT 960 305 E Р D S Κ А T RTDCSDN SDSD Ν D GAGGGCACCGAGGGGGAAGCCACAGAGGGCCTTGAAGGCACCGAGGCTGTGGAGAAGGGC 1020 ЕАТ EGLEGT ΕΑVΕ 325 E G Т Ε G Κ G 1080 S R V D G \mathbf{E} DΕ EGK EDGREE G Κ E 345 GATCCAGAGAAGGGGGGCCCTAACCACCCAGGATGAAAATGGCCAGACCAAGCGCAAGTTG 1140 L T T Ε 365 D Р E Κ G Α 0 D Ν G 0 T Κ R K T, 1200 CAGGCAGGCAAGAAGAGTCAGGACAAGCAGAAAAAGCGGCAGCGAGACCGCATGGTGGAA 385 0 А G Κ Κ S Q D Κ Q Κ Κ R 0 R D R М V Ε 1260 AGGATCCAGTTTGTGTGTTCTCTGTGCAAATACCGGACCTTCTATGAGGACGAGATGGCC 405 R Т 0 F V <u>C</u> S L C Κ Y R Т F Y Ε D Μ AGCCATCTTGACAGCAAGTTCCACAAGGAACACTTTAAGTACGTAGGCACCAAGCTCCCT 1320 <u>KFH</u>KEHFK Y V Т 425 S H L D S G Κ L Ρ AAGCAGACGGCTGACTTTCTGCAGGAGTACGTCACTAACAAGACCAAGAAGACAGAGGAG 1380 Κ 0 T ADFLQEY V Т Ν Κ Т Κ Κ Т Ε 445 CTCCGAAAAACCGTGGAGGACCTTGATGGCCTCATCCACCAAATCTACAGAGACCAGGAT 1440 V E DLDGL Η Q 465 L R Κ Т Т Т Y R D O D CTGACCCAGGAAATTGCCATGGAGCATTTTGTGAAGAAGGTGGAGGCAGCCCATTGTGCA 1500 І А М Е Н Ғ V К К V Е А А Н 485 \mathbf{L} Т 0 Ε С Α GCCTGCGACCTCTTCATTCCCATGCAGTTTGGGATCATCCAGAAGCATCTGAAGACCATG 1560 А С D \mathbf{L} F Т ΡΜΟΓ G Ι Ι <u>OKH</u> L Κ Т М 505 1620 Ν R R LMM Е Q S Κ Κ S S V 525 D Η Ν R T, М GCCCGCAGTATTCTCAACAACAAGCTCATCAGCAAGAAGCTGGAGCGCTACCTGAAGGGC 1680 А R S Ι \mathbf{L} Ν Ν Κ \mathbf{L} Ι S K K \mathbf{L} \mathbf{E} R Υ \mathbf{L} Κ G 545 GAGAACCCTTTCACCGACAGCCCCGAGGAGGAGGAGGAGGAGGAGGAGGCTGAGGGCGGT 1740 Т Ρ Ε 565 Е Ν Р F D S E E Κ E 0 E E А Е G G 1800 GCCCTGGACGAGGGGGGCGCAGGGCGAAGCGGCAGGGATCTCGGAGGGCGCAGAGGGCGTG

GΕΑ

ALDEGAO

AGIS

Ε

GAEGV

Fig. 1. a Nucleotide sequence and deduced amino acid sequence of human NAKAP95 gene. Asterisk denotes the stop codon. Two C2H2 zinc finger motifs are underlined and a putative nuclear localizing signal is shown in *italics*. The polyadenylation signal, aataaa, is double underlined. The nucleotide sequence of the NAKAP95 gene is deposited in GenBank/EMBL/DDBJ databases under the accession number, AB025905. The nucleotide sequences of both strands were determined by a primer walking method, using an ABI377 sequencer (Perkin Elmer, Norwalk, CT, USA). b Alignment of human AKAP95 (accession number, Y11997), rat AKAP95 (accession number, U01914), and human NAKAP95 (accession number, AB025905) proteins. Identities are indicated by black background and similar residues are shadowed. Asterisks denote the termination codon

CCGGCGCAGCCTCCCGTGCCCCCAGAGCCAGCCCCCGGGGCCGTGTCGCCGCCACCGCCG 1860 605 PAQPPVPPEPAPGAVSPPP 1920 P P P E E E E G A V P L L G G A L Q R 625 1980 Q I R G I P G L D V E D D E E G G G A 645 ${\tt CCGTGAcccgagctcggggcggggcggggcgcggggccgcgcgtggccgaacgtggaaaccaaacct} \underline{aa}$ 2040 P * 646 2057

a <u>taaa</u>gttttcccatccc

hAKAP95	1	MDOGYGGYGAWSAGPANTOGAYGUGVASWOGYENYNYYGAONTSVITGATYSYGPASWEA
rAKAP95	1	MEQSYGGYGAWSAGPANTOGTYGSGVASWQGYENYS
NAKAP95	1	~~~~MSYTGFVQGSETTLQSTYSDTSAQPTCDYGYGTWNSGTNRGYEGYGYGYGYGQDNT
hAKAP95	61	AKANDGGLAAGAPAMHMASYGPEPCTDNSDSLIAKINQRLDMMSKEGGRGGSGGGGG
rAKAP95	61	TKASDGGLAAGSSAMHVASFAPEPCTDNSDSLIAKINQRLDML
NAKAP95	57	TNYGYGMATSHSWEMPSSDTNANTSASGSASADSVLSRINQRLDMV.PHLETDMMQGGVY
hAKAP95 rAKAP95 NAKAP95	$\begin{array}{r} 118 \\ 118 \\ 116 \end{array}$	GIQDRESSFRFQPRESYDSRPCLPEHNPYRPSYSYDYEFDLGSDRNGSFGGQYSECRDPA GMQDRDSSFRFQPYESYDSRPCMPEHTPYRPSYSYDYDFDLGTDRNGSFGGTFNDCRDPT GSGGERYDSYESCDSRAVLSERDLYRSGYDYSELDPEMEMAYEGQYDAYRDQF
hAKAP95	178	RERGSLDGFMRGRGQGRFQDRSNPGTFMRSDPFVPPAASSEPLSTPWNELNYVGGRGLGG
rAKAP95	178	PERGALDGFLRGRGQGRFQDRSNSSTFIRSDPFMPPSASSEPLSTTWSELNYMGGRGLGG
NAKAP95	169	RMRGNDTFGPRAQGWARDARSGRPMASGYGRMWEDPMGARGQCMSG
hAKAP95	238	PSPSRPPPSLFSQSMAPDYGV.MGMQGAGGYDSTMPYGCGRSQPRMRDRDRPKRRGFDRF
rAKAP95	238	PSTNRPPPSLFSQSMAPDYSM.MGMQGVGGFGGTMPYGCGRSQTRIRDWPRRRGFERF
NAKAP95	215	ASRLPSLFSQNIIPFYGMFQGMRGGGAFPGGSRFGFGFGNGMKQMRRTWKTWTTADF
hAKAP95	297	GPDGTGRKRKQFQLYEEPDTKLARVD.SEGDFSENDDAA.GDFRSG
rAKAP95	295	GPDNMGRKRKPFPLYEEPDAKLARAD.SEGDLSENDDGA.GDLRSG
NAKAP95	272	RTKKKKRKQGGSPDEPDSKATRTDCSDNSDSDNDEGTEGEATEGLEGTEAVEKGSRVD
hAKAP95	341	DEEFKGEDELCDSGRORGEKEDEDEDVKKRREKORRRDRTRDRAADRIOF
rAKAP95	339	DEEFRGEDDLCDSRKORGEKEDEDEDVKKRREKORRRDRMRDRAADRIOF
NAKAP95	330	GEDEEGKEDGREEGKEDPEKGALTTODENGOTKRKLOAGKKSODKOKKRORDRMVERIOF
hAKAP95	391	ACSVCKFRSFDDEEIQKHLQSKFHKETLRFISTKLPDKTVEFLQEYIVNRNKKIEKRRQE
rAKAP95	389	ACSVCKFRSFEDEEIQKHLQSKFHKETLRFISTKLPDKTVEFLQEYIINRNKKIEKRRQE
NAKAP95	390	VCSECKYRFFYEDEMASHLDSKFHKE <mark>HFKYVG</mark> TKLP <mark>KQTAD</mark> FLQEYVTNKTKKTEELRKT
hAKAP95	451	LMEKETAKPKPDPFKGIGQEHFFKKIEAAHCLACDMLIPAQPQLLQRHLHSVDHNH
rAKAP95	449	LLEKESPKPKPDPFKGIGQEHFFKKIEAAHCLACDMLIPAQHQLLQRHLHSVDHNH
NAKAP95	450	VEDLDGLIHQIYRDQDLTQEIAMEHFVKKVEAAHCAACDIFIPMQFGITQKHLKTMDHNR
hAKAP95	507	NRRLAAEQFKKTSLHVAKSVLNN <mark>R</mark> HIVKMLEKYLKGEDPFTSETVDPEMEGDDNLG <mark>G</mark> EDK
rAKAP95	505	NRRLAAEQFKKTSLHVAKSVLNNKHIVKMLEKYLKGEDPF <mark>VNETADLETEGDE</mark> NLG.E
NAKAP95	510	NRRL <mark>MMEQSKKSSLMVARST</mark> LNNK <mark>LISKK</mark> LERYLKGENPFTDSPEFEKEQEFAEGGALDE
hakap95	567	KETPEEVAADVLAEVITAAVRAVDGEGAPAPESSGEPAEDEGPTDTAEAGSDPQAEQLLE
rakap95	563	KETPEEVAAEVLAEVITAAVKAVEGDGEPAAEHSDVLAEVEGPVDTAEAGSDSHTGKLLE
nakap95	570	GAQGEAAGISEGAEGVPAQ.PPVPPEPAPGAVSPPPPPPPEEEEEGAVPLLGGALQ
hakap95	6 2 7	EQVPCGTAHEKGVPKARSEAAEAGNGAETMAAEAESAQTRVAPAPAAADAEVEQTDAE
rakap95	6 2 3	EQT.CETASETRNMEDMARGEAAEARNEAAVPAAAAGSPVPVIA.IPGILEDELEQTDAE
nakap95	6 2 5	RQIRGIPGLDVEDDEEGGGGGAP*~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~
hAKAP95 rAKAP95 NAKAP95	685 681 647	SKDAVPTE* AKD.TPTE*

Fig. 1. Continued

b



Fig. 2. a Chromosomal placement of the *NAKAP95* gene at a relative distance to framework markers on the WICGR radiation hybrid map of the human genome (http://carbon.wi.mit.edu:8000/cgi-bin/contig/phys_map). The approximate corresponding cytogenetic location of

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the gene on chromosome 19p13.11–p13.12 region is indicated. Distances are in centirays (cR) from the top of the chromosome 19 linkage group. **b** Schematic exon-intron structure of human *NAKAP95* and *AKAP95* genes

Table 1. Intron-exon boundaries of the NAKAP95 gene

Exon No.	Exon size ^a	Splice acceptor ^b	Splice donor ^b
1	58		AGCTACACAG gt gggcctggc
2	75	ctttatgttt ag GCTTTGTCCA	TGTGATTATG gt aagtgtggac
3	33	tactttttgt ag GATATGGAAC	ACAAATAGAG gc aagtgtcatt
4	241	ccctcactgc ag GCTACGAGGG	GTGGAGAAAG gt gagtggacac
5	454	atgcctctgc ag GTATGACTCT	CGACTTCCGA gt gagtggaggc
6	97	cttatgaccc ag ACCAAGAAGA	TCAGACAATG gt gagcccacta
7	71	tttccctttc ag ATGAGGGCAC	CTCCAGAGTG gt aagaggetet
8	64	tgaaattgtt ag GACGGAGAGG	CCAGAGAAGG gt gagttttcct
9	109	ctgctcccac ag GGGCCCTAAC	TGGTGGAAAG gt aaccagcttc
10	142	tteetgtgge ag GATCCAGTTT	CTTTCTGCAG gt gageettgga
11	106	tatteettgeag GAGTACGTCA	CTGACCCAGG gt gaggagattt
12	131	teeccactac ag AAATTGCCAT	GAACCGCAGG gt gagtggccac
13	96	ccctgcccgc ag CTCATGATGG	CTACCTGAAG gt gaggcactgg
14	380	ccctccccac ag CCGCTCTTGG	AAGGGTAGGG

Intron-exon junctions were established by comparison of cDNA and genomic sequences ^aSize in basepairs

^bSequences at the splice junction. Exonic sequences are shown in capital letters, with intronic sequences shown in lowercase letters. Invariant nucleotides (ag/gt, gc) are in boldface type



Fig. 3. Tissue distribution analysis, using reverse transcription coupled-polymerase chain reaction (RT-PCR). The 12 tissues and genomic DNA examined are indicated *above each lane*. The templates of the human tissues of poly(A)+ RNAs were purchased from Clontech (Palo Alto, CA, USA). The cDNA templates for RT-PCR were synthesized from 2µg of poly(A)+, using excess amounts of Superscript II reverse transcriptase (GIBCO BRL, Gaithersburg, MD, USA) and random hexamer primers. PCR was carried out in a final volume of 10µl containing 1 × LA-PCR buffer (Takara, Kyoto, Japan), 2µM each primer, 200µM each dNTP, 1µl of template DNA, and 0.01 units of LA-Taq DNA polymerase (Takara). Temperatures and time schedule were: 30 cycles of 95°C for 20s and 62°C for 1 min. PCR products were separated on 2.5% Nusieve GTG agarose gel (FMC, Rockland, ME, USA) with a 1-kb ladder DNA marker (GIBCO BRL)

The exon-intron boundaries of the human NAKAP95 and AKAP95 genes were determined by aligning the cDNA sequence with the genomic sequence from the two cosmid clones (accession numbers, AC005785, AC006128) (Fig. 2b). As shown in Table 1, all but one of the splicing sites conformed to the canonical splicing acceptor and donor rule of AG-GT; one was AG-GC. The NAKAP95 gene was divided into 14 exons, which ranged in size from 33 bp (exon 3) to 454bp (exon 5). Exons 1 and 14 contained the ATG and TAG codons, respectively. The exon-intron boundary of the human AKAP95 gene was also determined in the same manner, revealing that human NAKAP95 and AKAP95 had a very similar gene structure (Fig. 2b). In both cases, the C2H2 type zinc finger motif resided in exons 10 and 12. These results strongly argue that the NAKAP95 and AKAP95 genes could have been established by tandem gene duplication.

References

- Boynton AL, Whitfield JF (1983) The role of cAMP in cell proliferration: a critical assessment of the evidence. Adv Cyclic Nucleotide Res 15:193–294
- Carr DW, Stofko-Hahn RE, Fraser ID, Cone RD, Scott JD (1992) Localization of the cAMP-dependent protein kinase to the postsynaptic densities by A-kinase anchoring proteins. Characterization

of AKAP 79. J Biol Chem 267:16816-16823

- Chen Q, Lin RY, Rubin CS (1997) Organelle-specific targeting of protein kinase AII (PKAII). Molecular and in situ characterization of murine A kinase anchor proteins that recruit regulatory subunits of PKAII to the cytoplasmic surface of mitochondria. J Biol Chem 272:15247–15257
- Coghlan VM, Langeberg LK, Fernandez A, Lamb NJ, Scott JD (1994) Cloning and characterization of AKAP 95, a nuclear protein that associates with the regulatory subunit of type II cAMP-dependent protein kinase. J Biol Chem 269:7658–7665
- Edelman AM, Blumenthal DK, Krebs EG (1987) Protein serine/threonine kinases. Annu Rev Biochem 56:567–613
- Eide T, Coghlan V, Orstavik S, Holsve C, Solberg R, Skalhegg BS, Lamb NJ, Langeberg L, Fernandez A, Scott JD, Jahnsen T, Tasken K (1998) Molecular cloning, chromosomal localization, and cell cycle-dependent subcellular distribution of the A-kinase anchoring protein, AKAP95 Exp Cell Res 238:305–316
- Faux MC, Scott JD (1996) Molecular glue: kinase anchoring and scaffold proteins. Cell 85:9–12
- Hausken ZE, Dell'Acqua ML, Coghlan VM, Scott JD (1996) Mutational analysis of the A-kinase anchoring protein (AKAP)-binding site on RII. Classification of side chain determinants for anchoring and isoform selective association with AKAPs. J Biol Chem 271:29016–29022
- Hausken ZE, Scott JD (1996) Properties of A-kinase anchoring proteins. Biochem Soc Trans 24:986–991
- Keryer G, Rios RM, Landmark BF, Skalhegg B, Lohmann SM, Bornens M (1993) A high-affinity binding protein for the regulatory subunit of cAMP-dependent protein kinase II in the centrosome of human cells. Exp Cell Res 204:230–240
- Kozak M (1987) At least six nucleotides preceding the AUG initiation codon enhance translation in mammalian cells. J Mol Biol 196:947– 950
- Krebs EG, Beavo JA (1979) Phosphorylation-dephosphorylation of enzymes. Annu Rev Biochem 48:923–959
- Lester LB, Coghlan VM, Nauert B, Scott JD (1996) Cloning and characterization of a novel A-kinase anchoring protein. AKAP 220, association with testicular peroxisomes. J Biol Chem 271:9460–9465
- Li Y, Ndubuka C, Rubin CS (1996) A kinase anchor protein 75 targets regulatory (RII) subunits of cAMP-dependent protein kinase II to the cortical actin cytoskeleton in non-neuronal cells. J Biol Chem 271:16862–16869
- McKnight GS (1991) Cyclic AMP second messenger systems. Curr Opin Cell Biol 3:213–217
- McCartney S, Little BM, Langeberg LK, Scott JD (1995) Cloning and characterization of A-kinase anchor protein 100 (AKAP100). A protein that targets A-kinase to the sarcoplasmic reticulum. J Biol Chem 270:9327–9333
- Nauert JB, Klauck TM, Langeberg LK, Scott JD (1997) Gravin, an autoantigen recognized by serum from myasthenia gravis patients, is a kinase scaffold protein. Curr Biol 7:52–62
- Roesler WJ, Vandenbark GR, Hanson RW (1988) Cyclic AMP and the induction of eukaryotic gene transcripton. J Biol Chem 263:9063– 9066
- Rubin CS (1994) A kinase anchor proteins and the intracellular targeting of signals carried by cyclic AMP. Biochim Biophys Acta 1224:467–479
- Saito T, Seki N, Ishii H, Ohira M, Hayashi A, Kozuma S, Hori T (1997) Complementary DNA cloning and chromosomal mapping of a novel phosphatidylinositol kinase gene. DNA Res 4:301–305
- Schmidt PH, Dransfield DT, Claudio JO, Hawley RG, Trotter KW, Milgram SL, Goldenring JR (1999) AKAP350, a multiply spliced protein kinase A-anchoring protein associated with centrosome. J Biol Chem 274:3055–3066
- Scott JD, Macartney S (1994) Localization of A-kinase through anchoring proteins. Mol Endocrinol 8:5–11
- Seki N, Nimura Y, Ohira M, Saito T, Ichimiya S, Nomura N, Nakagawara A (1997) Identification and chromosome assignment of a human gene encoding a novel phosphatidylinositol-3 kinase. DNA Res 4:355–358
- Seki N, Muramatsu M, Sugano S, Suzuki Y, Nakagawara A, Ohhira M, Hayashi A, Hori T, Saito T (1998) Cloning, expression analysis, and chromosomal localization of HIP1R, an isolog of huntingtin interacting protein (HIP1). J Hum Genet 43:268–271

- Seki N, Hattori A, Hayashi A, Kozuma S, Ohira M, Hori T, Saito T (1999) Structure, expression profile and chromosomal location of an isolog of DNA-PKcs interacting protein (KIP) gene. Biochim Biophys Acta 1444:143–147
- Taylor SS, Buechler JA, Yonemoto W (1990) cAMP-dependent protein kinase: framework for a diverse family of regulatory enzymes. Annu Rev Biochem 59:971–1005
- Ueki N, Oda T, Kondo M, Yano K, Noguchi T, Muramatsu M (1998) Selection system for genes encoding nuclear-targeted proteins. Nat Biotechnol 16:1338–1342
- Ueki N, Seki N, Yano K, Ohira M, Saito T, Masuho Y, Muramatsu M

(1999a) Isolation and chromosomal assignment of human genes encoding cofactor of LIM homeodomain proteins, CLIM1 and CLIM2. J Hum Genet 44:112–115

- Ueki N, Seki N, Yano K, Saito T, Masuho Y, Muramatsu M (1999b) Isolation and chromosomal assignment of a human gene encoding protein inhibitor of activated STAT3 (PIAS3). J Hum Genet 44:193– 196
- Ueki N, Seki N, Yano K, Saito T, Masuho Y, Muramatsu M (1999c) Isolation and characterization of a novel human gene (HFB30) which encodes a protein with a RING finger motif. Biochim Biophys Acta 1445:232–236