

Pierre Bouzat  
Nathalie Sala  
Tamarah Suys  
Jean-Baptiste Zerlauth  
Pedro Marques-Vidal  
François Feihl  
Jocelyne Bloch  
Mahmoud Messerer  
Marc Levivier  
Reto Meuli  
Pierre J. Magistretti  
Mauro Oddo

## Cerebral metabolic effects of exogenous lactate supplementation on the injured human brain

Received: 23 September 2013  
Accepted: 20 December 2013  
Published online: 30 January 2014  
© Springer-Verlag Berlin Heidelberg and ESICM 2014

**Take-home message:** Exogenous systemic supplemental lactate can be efficiently used by the injured human brain with beneficial sparing of cerebral glucose. These clinical interventional study further suggests resuscitation with hypertonic lactate may be a valid therapeutic strategy after TBI.

Clinicaltrials.gov identifier NCT01573507.

P. Bouzat · N. Sala · T. Suys ·  
M. Oddo (✉)  
Department of Intensive Care Medicine,  
Neuroscience Critical Care Research Group,  
Faculty of Biology and Medicine,  
University Hospital of Lausanne, Rue du  
Bugnon 46, 1011 Lausanne, Switzerland  
e-mail: mauro.oddo@chuv.ch  
Tel.: +41-79-5561246  
Fax: +41-21-3143045

P. Bouzat  
Joseph Fourier University, Grenoble, France  
e-mail: PBouzat@chu-grenoble.fr

J.-B. Zerlauth · R. Meuli  
Department of Radiology, Faculty of  
Biology and Medicine, University Hospital  
of Lausanne, 1011 Lausanne, Switzerland

P. Marques-Vidal  
Faculty of Biology and Medicine, Institute  
of Social and Preventive Medicine,  
University Hospital of Lausanne,  
1011 Lausanne, Switzerland

F. Feihl  
Division of Clinical Pathophysiology,  
Department of Internal Medicine, Faculty of  
Biology and Medicine, University Hospital  
of Lausanne, 1011 Lausanne, Switzerland

J. Bloch · M. Messerer · M. Levivier  
Division of Neurosurgery, Department of  
Clinical Neurosciences, Faculty of Biology  
and Medicine, University Hospital of  
Lausanne, 1011 Lausanne, Switzerland

P. J. Magistretti  
Laboratory of Neuroenergetics and Cellular  
Dynamics, Faculty of Biology and  
Medicine, Brain Mind Institute, Ecole  
Polytechnique Fédérale de Lausanne  
(EPFL), Centre de Neurosciences  
Psychiatriques, University Hospital of  
Lausanne, 1011 Lausanne, Switzerland

**Abstract Purpose:** Experimental evidence suggests that lactate is neuroprotective after acute brain injury; however, data in humans are lacking. We examined whether exogenous lactate supplementation improves cerebral energy metabolism in humans with traumatic brain injury (TBI). **Methods:** We prospectively studied 15 consecutive patients with severe TBI monitored with cerebral microdialysis (CMD), brain tissue  $PO_2$  ( $PbtO_2$ ), and intracranial pressure (ICP). Intervention consisted of a 3-h intravenous infusion of hypertonic sodium lactate (aiming to increase systemic lactate to ca. 5 mmol/L), administered in the

early phase following TBI. We examined the effect of sodium lactate on neurochemistry (CMD lactate, pyruvate, glucose, and glutamate),  $PbtO_2$ , and ICP. **Results:** Treatment was started on average  $33 \pm 16$  h after TBI. A mixed-effects multilevel regression model revealed that sodium lactate therapy was associated with a significant increase in CMD concentrations of lactate [coefficient 0.47 mmol/L, 95 % confidence interval (CI) 0.31–0.63 mmol/L], pyruvate [13.1 (8.78–17.4)  $\mu$ mol/L], and glucose [0.1 (0.04–0.16) mmol/L; all  $p < 0.01$ ]. A concomitant reduction of CMD glutamate [–0.95 (–1.94 to 0.06) mmol/L,  $p = 0.06$ ] and ICP [–0.86 (–1.47 to –0.24) mmHg,  $p < 0.01$ ] was also observed. **Conclusions:** Exogenous supplemental lactate can be utilized aerobically as a preferential energy substrate by the injured human brain, with sparing of cerebral glucose. Increased availability of cerebral extracellular pyruvate and glucose, coupled with a reduction of brain glutamate and ICP, suggests that hypertonic lactate therapy has beneficial cerebral metabolic and hemodynamic effects after TBI.

**Keywords** Brain metabolism · Lactate · Cerebral microdialysis · Traumatic brain injury · Hypertonic

## Introduction

Mechanisms of secondary cerebral damage after traumatic brain injury (TBI) rely on the complex interplay between cerebral blood flow, oxygen, and energy supply [1]. Clinical investigations using PET scans and the cerebral microdialysis (CMD) technique have shown that TBI is associated with increased cerebral glucose utilization that may eventually culminate into a state of energy dysfunction [2, 3]. In this setting, the supply to the injured brain of the main energy substrate—i.e., glucose [4]—may become limited, leading to reductions of cerebral extracellular glucose below critical thresholds, a scenario which is associated with worse outcome [5, 6].

Experimental evidence repeatedly demonstrated that the brain can use substrates other than glucose to sustain increased activity, including lactate [7–11]. Increased lactate can be the consequence of anaerobic metabolism, i.e., the formation of lactate in the presence of low oxygen. However, emerging evidence demonstrated that in patients with TBI increased lactate seems predominantly non-hypoxic/ischemic and rather the consequence of increased cerebral glycolysis [2, 12, 13]. Lactate formed under such glycolytic conditions is shuttled from one lactate-producing cell (astrocyte) to another lactate-consuming cell type (neuron) (*astrocyte–neuron lactate shuttle*) [14, 15]. Utilization of lactate as an alternative fuel may be an adaptive response to maximize energy function and limit substrate reduction. Uptake of endogenous cerebral lactate has indeed been demonstrated after TBI in animals [16] and in humans [3, 17].

Administration of supplemental lactate significantly contributes to brain metabolism: at supraphysiologic concentrations (ca. 5 mmol/L) “extra” lactate becomes the preferential energy substrate for neurons with sparing of glucose [18] and might be neuroprotective after experimental TBI [19]. The ability of the injured brain to use supplemental lactate as a glucose-sparing fuel may be important to overcome post-TBI increased energy demand. To test this hypothesis we designed a prospective interventional study to examine in patients with TBI monitored with CMD whether hypertonic lactate therapy—administered intravenously during the acute phase of injury—might have beneficial effects on cerebral energy metabolism.

## Methods

### Study type

This was a prospective phase II interventional study that was performed at the Department of Intensive Care Medicine, CHUV, Lausanne, Switzerland. The study was approved by the Ethical Research Committee of the

University of Lausanne. Informed consent was obtained from each patient’s next of kin.

### Patients

#### *Inclusion criteria*

Patients with severe TBI, defined by an admission Glasgow Coma Scale (GCS) < 9, aged 18–60 years, with abnormal admission brain CT scan (defined by a Marshall score  $\geq 2$ ), who underwent intracranial monitoring with CMD, brain tissue oxygen tension (PbtO<sub>2</sub>), and intracranial pressure (ICP).

#### *Exclusion criteria*

Penetrating TBI, age <18 or >60 years, history of neurological disease, brain death or expected death within 48 h, pregnancy, more than one major extracranial injury with sustained hemodynamic instability and persistent elevation of blood lactate >4 mmol/L for >6 h before intervention, time from TBI to study inclusion >72 h.

### Intracranial monitoring

Cerebral microdialysis consisted of a 70 MD bolt catheter with 20-kDa cutoff (M Dialysis AB<sup>®</sup>, Stockholm, Sweden). Microdialysis samples were collected hourly and analyzed for brain extracellular concentrations of glucose, lactate, pyruvate, and glutamate. PbtO<sub>2</sub> was measured using a Licox<sup>®</sup> catheter (Integra Neurosciences, Plainsboro, NJ, USA) calibrated to the patient’s temperature. Intracranial pressure was measured using a Codman<sup>®</sup> probe (Raynham, MA, USA). Probes were inserted through a triple-lumen bolt in the operating room by a neurosurgeon and placed into the brain parenchyma (subcortical white matter), in the right frontal lobe. Brain CT scan was performed at approximately 24 h to confirm the correct placement of intracranial monitors.

### General management

All patients were treated according to a standard protocol for the management of severe TBI following international guidelines [20, 21]. Sedation–analgesia consisted of propofol and sufentanil. All patients were mechanically ventilated to maintain PaO<sub>2</sub> at 90–100 mmHg and PaCO<sub>2</sub> at 35–40 mmHg. Brain physiological targets were set to maintain ICP < 20 mmHg and cerebral perfusion pressure (CPP = mean arterial pressure, MAP–ICP) >60 mmHg. Blood glucose was targeted at 6–8 mmol/L.

## Study intervention

The intervention consisted of a continuous infusion of hypertonic sodium lactate ( $\text{Na}^+$  1,000 mmol/L, lactate 1,000 mmol/L, prepared by the Division of Pharmacy, CHUV, Lausanne), administered over 3 h (at 40  $\mu\text{mol/kg/min}$  for 60 min, followed by 30  $\mu\text{mol/kg/min}$  for an additional 120 min) started within 72 h from TBI (clinicaltrials.gov identifier NCT01573507).

## Study endpoints

### Study efficacy

The treatment aimed to rapidly increase blood arterial lactate to supraphysiological concentrations (ca. 5 mmol/L) using a bolus dose over 30 min, followed by a maintenance dose up to 3 h. This infusion protocol was in line with previous clinical studies in healthy human subjects [22, 23] and performed according to a previous investigation at our institution in critically ill non-neurological patients [24]. The primary endpoint was to analyze the effect of sodium lactate infusion on CMD concentrations of lactate, pyruvate, and glucose. Secondary endpoints were to analyze the effect of lactate therapy on CMD glutamate,  $\text{PbtO}_2$ , and ICP.

### Study safety

Safety parameters included blood arterial pH, bicarbonate, sodium, and osmolarity. Safety thresholds were set at  $\text{pH} > 7.55$ , sodium  $> 155$  mmol/L, and osmolarity  $> 320$  mosm/L.

### Data collection and processing

Clinical and radiological variables included age, gender, admission GCS, Marshall CT scan score [25], type of injury, time from TBI to intracranial monitoring, and duration of monitoring. Neurological outcome at 6 months was assessed using the GOS. Cerebral microdialysis variables were collected hourly. Main brain physiological variables ( $\text{PbtO}_2$ , ICP, and CPP) were recorded every 60 s via a computerized medical chart system (Metavision<sup>®</sup>, IMD soft, Tel-Aviv, Israel) and were matched at the exact time of CMD analysis. For  $\text{PbtO}_2$ , data were corrected for temperature and  $\text{PaO}_2$ . Arterial blood was sampled hourly for concentration of pH,  $\text{PaO}_2$ ,  $\text{PaCO}_2$ , bicarbonate, lactate, glucose, sodium, chloride, and osmolarity.

### Statistical analysis

Each patient's individual response to intervention was graphically plotted for CMD lactate, pyruvate, and

glucose. Data distribution was tested for each variable and logarithmic transformation was applied for non-normal data. Results were expressed as mean  $\pm$  SD for the two main conditions, i.e., *baseline* (immediately before the start of sodium lactate infusion) and *sodium lactate* (maximum increase during intervention).

Data analyzed included CMD (lactate, pyruvate, glucose, glutamate) and main brain physiologic (ICP,  $\text{PbtO}_2$ , CPP) and systemic (lactate, glucose, pH, bicarbonate, sodium, chloride, osmolarity) variables. Longitudinal data analysis was used to account for repeated measures of physiological variables across different patients over time. Analysis was conducted using mixed-effects multilevel regression with time as an independent variable and allowing it to have a random intercept and a random slope according to the patient. The same model was also applied to analyze the data obtained during the 6 h preceding the intervention, to verify data stability prior to sodium lactate therapy. Correlations were tested using the Pearson's correlation coefficient. Statistical analyses were performed using STATA 12 (STATA<sup>®</sup> Corporation, College Station, TX, USA), and statistical significance was set at  $p < 0.05$ .

## Results

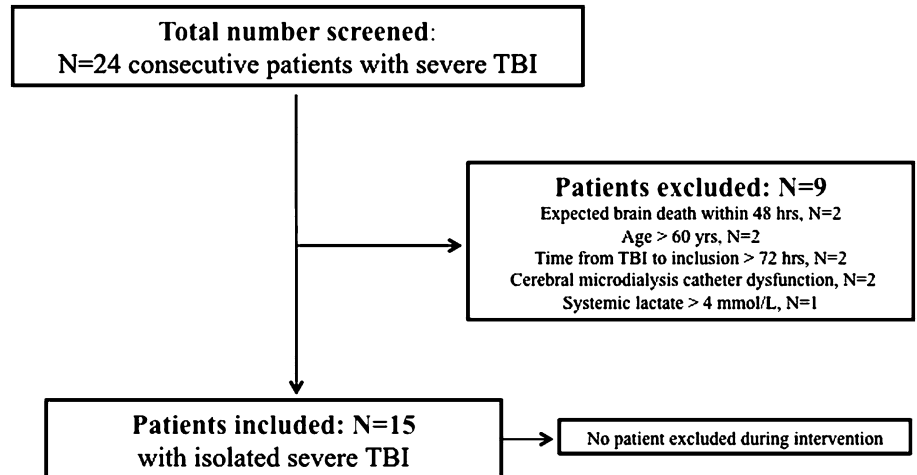
### Patient characteristics

From April 2012 to March 2013, a total of 24 consecutive patients were screened for the intervention, of which 9 were excluded, and 15 patients with isolated severe TBI were included (Fig. 1). Baseline characteristics of the 15 studied patients are summarized in Table 1. Mean age was  $40 \pm 15$  years and median admission GCS was 7 (range 3–8). Intracranial monitors were placed close to each other in the right frontal subcortical white matter in all patients, in an area that corresponded to visually normal brain parenchyma on control head CT scan (i.e., not around a contusion or hematoma, Fig. 2). Median time from TBI to intracranial monitoring was 6 h (interquartile range 3–15 h) and patients were monitored for a median of 4 (3.5–7.0) days. There was no catheter-related complication. At 6 months, 9 patients had a favorable neurological recovery [Glasgow Outcome Score (GOS) 4 or 5] and 6 patients had an unfavorable outcome [including 2 with severe disability (GOS 3), 1 with vegetative state (GOS 2), and 3 non-survivors (GOS 1)].

### Effect of sodium lactate therapy on cerebral variables

Sodium lactate therapy was started  $33 \pm 16$  h from TBI. Following sodium lactate infusion, blood lactate increased from a baseline pretreatment value of  $1.0 \pm 0.4$  mmol/L to a maximum of  $6.1 \pm 1.6$  mmol/L. Each patient's

**Fig. 1** Study flow chart. A total of 24 patients with isolated severe traumatic brain injury (TBI) were screened over the 1-year study period; 15 patients were included and concluded the study

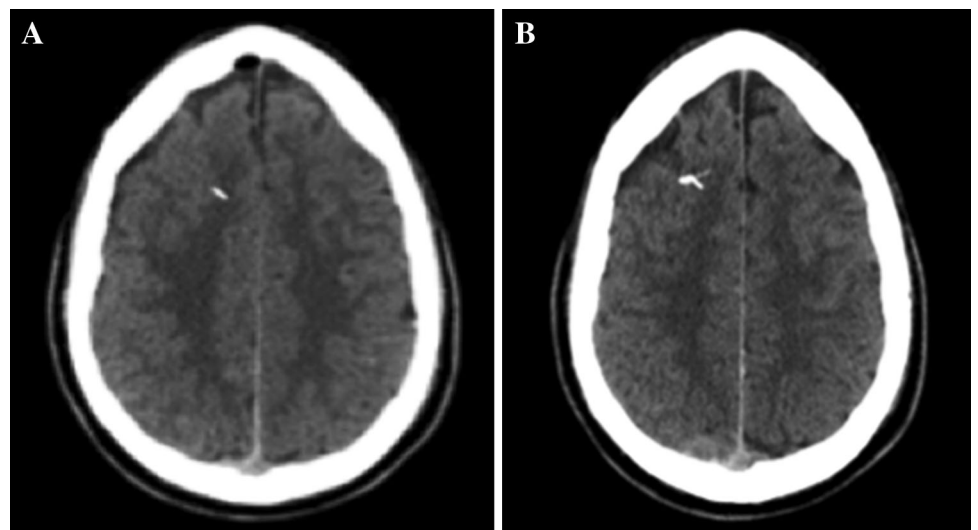


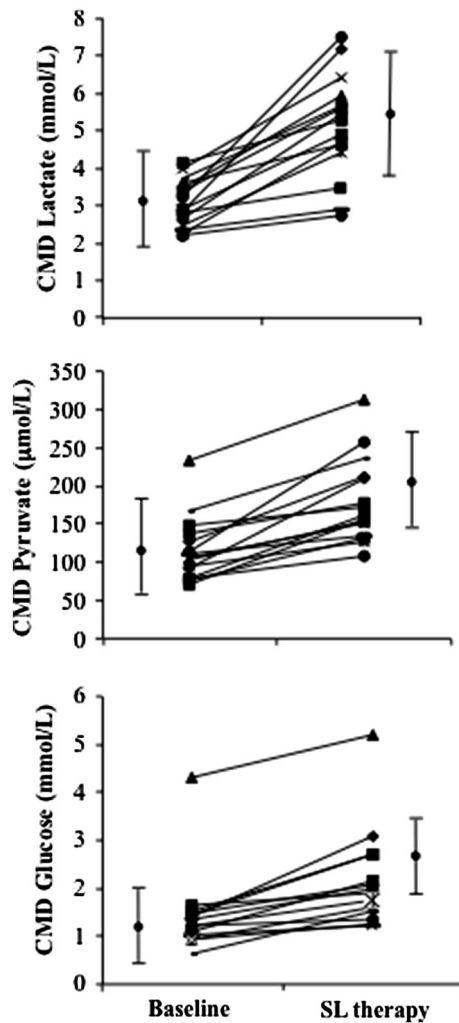
**Table 1** Patient characteristics

Patient no.	Age (years)	Gender	Admission GCS	Marshall score	Injury type	Time from TBI to intracranial monitoring (hours)	Time from TBI to sodium lactate therapy (hours)	Duration of intracranial monitoring (days)	GOS (at 6 months)
1	54	F	3	2	Diffuse	13	21	4	4
2	39	M	8	2	Focal	52	57	3	4
3	19	M	6	2	Diffuse	7	24	4	5
4	55	M	8	3	Diffuse	6	20	11	5
5	30	M	7	5	Diffuse	8	30	10	3
6	24	M	6	2	Diffuse	6	34	6	2
7	46	M	8	2	Diffuse	7	66	3	4
8	60	M	8	3	Focal	23	64	5	1
9	24	F	6	2	Diffuse	6	24	3	4
10	41	M	3	2	Diffuse	6	24	8	1
11	24	M	8	2	Diffuse	17	23	3	4
12	52	F	7	2	Diffuse	26	38	4	4
13	24	M	7	3	Diffuse	5	21	4	1
14	60	M	6	2	Diffuse	7	24	4	4
15	55	M	8	6	Diffuse	13	27	11	3

F female, GCS Glasgow Coma Scale, GOS Glasgow Outcome Score, M male, TBI traumatic brain injury

**Fig. 2** Location of cerebral microdialysis and brain tissue PO<sub>2</sub> catheters. **a** One illustrative example of probe location in the right frontal subcortical white matter, with close positioning of CMD and brain tissue PO<sub>2</sub> catheters (**b**)





**Fig. 3** Effect of sodium lactate therapy on cerebral energy metabolism across individual patients ( $n = 15$ ). Horizontal lines illustrate individual baseline and maximum values during sodium lactate (SL) therapy of cerebral microdialysis (CMD) concentrations of lactate, pyruvate, and glucose. Differences in means ( $\pm$ SD) of extracellular brain lactate concentration between baseline (left side of the graphic) versus SL therapy (right side of the graphic) are also shown (CMD lactate,  $3.1 \pm 0.6$  versus  $5.1 \pm 1.4$  mmol/L; CMD pyruvate,  $115.6 \pm 42.9$  versus  $180.2 \pm 55.7$   $\mu$ mol/L; CMD glucose,  $1.4 \pm 0.9$ – $2.2 \pm 1.0$  mmol/L)

individual response to intervention for CMD lactate, pyruvate, and glucose is shown in Fig. 3: sodium lactate was associated with an increase in CMD concentrations of lactate (from baseline  $3.1 \pm 0.6$  to a maximum of  $5.1 \pm 1.4$  mmol/L during sodium lactate therapy), CMD pyruvate ( $115.6 \pm 42.9$  vs.  $180.2 \pm 55.7$   $\mu$ mol/L), and CMD glucose ( $1.4 \pm 0.9$  vs.  $2.2 \pm 1.0$  mmol/L). All three patients with pretreatment low CMD glucose ( $<1$  mmol/L) had normalized brain glucose values at the end of sodium lactate therapy.

Results of mixed-effects analysis for changes of cerebral variables during intervention are shown in

Table 2. During sodium lactate therapy, we found a significant increase in CMD concentrations of lactate [coefficient 0.47 mmol/L, 95 % confidence interval (CI) 0.31–0.63 mmol/L], pyruvate [13.1 (8.78–17.4)  $\mu$ mol/L], and glucose [0.1 (0.04–0.16) mmol/L; all  $p < 0.01$ ]. A concomitant decrease of CMD glutamate [ $-0.95$  ( $-1.94$  to  $0.06$ ) mmol/L,  $p = 0.06$ ], PbtO<sub>2</sub> [ $-0.58$  ( $-1.14$  to  $-0.01$ ) mmHg,  $p = 0.04$ ], and ICP [ $-0.86$  ( $-1.47$  to  $-0.24$ ) mmHg,  $p < 0.01$ ] was also observed. The effect of sodium lactate on ICP reduction was greater in the two patients who had baseline ICP  $> 20$  mmHg. Cerebral perfusion pressure and lactate-to-pyruvate ratio (LPR) did not change significantly during the intervention. Altogether, our data indicate aerobic utilization of systemic lactate with sparing of cerebral glucose and overall metabolic benefit for brain tissue.

Of note, blood lactate as well as CMD lactate, pyruvate, and glucose did not change significantly over the 6 h prior to sodium lactate infusion (all  $p > 0.1$ , mixed-effects analysis, see example of an individual patient in Fig. 4), indicating data stability before treatment start. Concentrations of CMD lactate, pyruvate, and glucose returned to baseline values at 6 h post-treatment start.

A majority of patients (10/15, 67 %) displayed baseline LPR  $> 25$ , a marker of increased energy distress: interestingly, we observed a significant correlation between LPR and the percentage of CMD pyruvate increase (Pearson's  $r = 0.60$ ,  $p = 0.02$ ; Fig. 5), suggesting that metabolic conversion of lactate into pyruvate during sodium lactate therapy was more pronounced in patients who had a higher degree of cerebral energy demand.

#### Effect of sodium lactate therapy on systemic variables

Results of mixed-effects analysis for changes of systemic variables during sodium lactate therapy are shown in Table 2. Blood arterial concentrations of pH, bicarbonate, sodium, and osmolarity all increased during sodium lactate infusion (all  $p < 0.01$ ) and returned to pretreatment values within 6 h from the end of infusion. Systemic chloride did not change during treatment. Importantly, concentrations of systemic glucose did not change significantly during sodium lactate therapy ( $p = 0.39$ ). Means of other relevant systemic physiological variables (including MAP, heart rate, body temperature, PaO<sub>2</sub>, and PaCO<sub>2</sub>) were within normal ranges in all patients and a mixed-effects model did not show significant changes of these variables during the treatment.

## Discussion

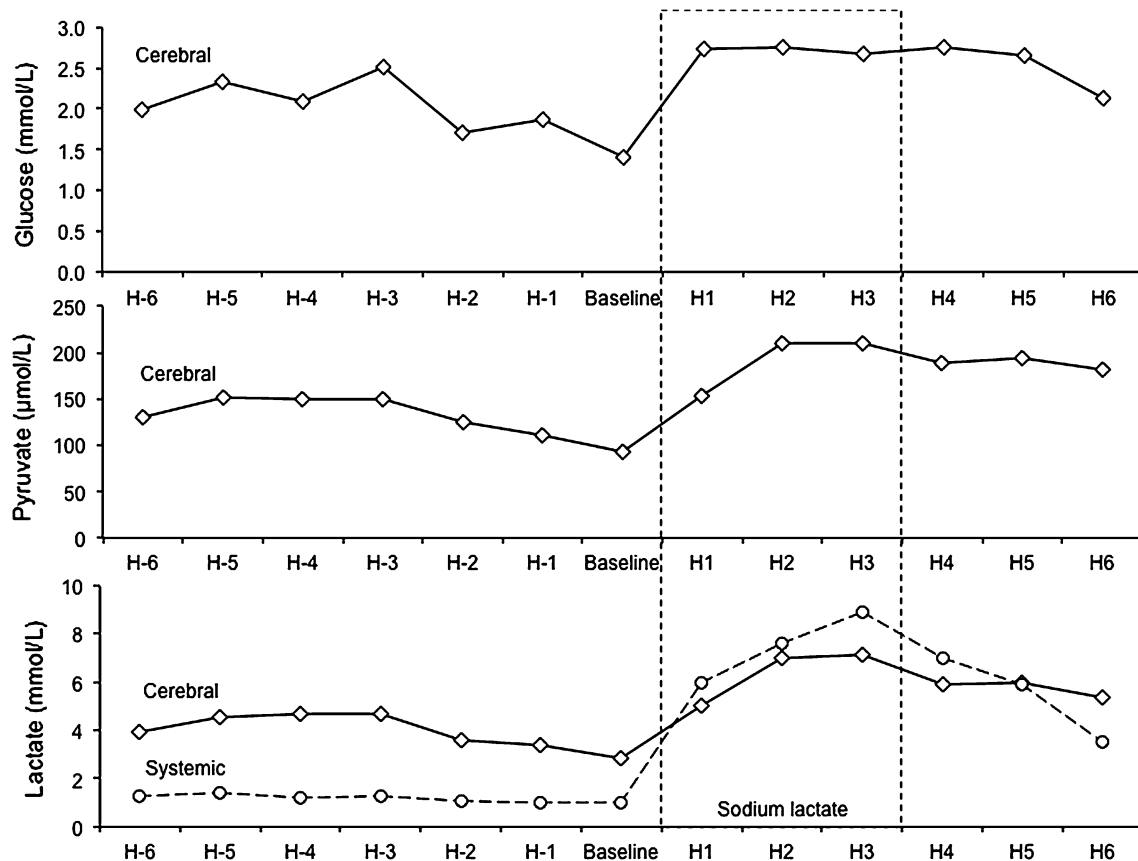
The findings of this study can be summarized as follows: (1) we showed for the first time in humans with acute

**Table 2** Changes of main cerebral and systemic variables during sodium lactate therapy

	Coefficient $\pm$ SE	95 % Confidence interval	<i>p</i> value
<b>Cerebral variables</b>			
CMD lactate (mmol/L)	0.47 $\pm$ 0.08	0.31 to 0.63	<0.01
CMD pyruvate ( $\mu$ mol/L)	13.10 $\pm$ 2.20	8.78 to 17.40	<0.01
CMD glucose (mmol/L)	0.10 $\pm$ 0.04	0.04 to 0.16	<0.01
CMD glutamate (mmol/L)	-0.95 $\pm$ 0.51	-1.94 to 0.06	0.06
PbtO <sub>2</sub> (mmHg)	-0.58 $\pm$ 0.29	-1.14 to -0.01	0.04
Intracranial pressure (mmHg)	-0.86 $\pm$ 0.32	-1.47 to -0.24	<0.01
<b>Systemic variables</b>			
Glucose (mmol/L)	0.08 $\pm$ 0.09	-0.11 to 0.28	0.39
Sodium (mmol/L)	1.96 $\pm$ 0.25	1.48 to 2.44	<0.01
Chloride (mmol/L)	-0.08 $\pm$ 0.25	-0.57 to 0.42	0.76
pH	0.04 $\pm$ 0.01	0.00 to 0.02	<0.01
Osmolarity (mosm/L)	3.57 $\pm$ 0.44	2.71 to 4.43	<0.01

Analysis was conducted using mixed-effects multilevel regression with time as an independent variable and allowing it to have a random intercept and a random slope according to patient

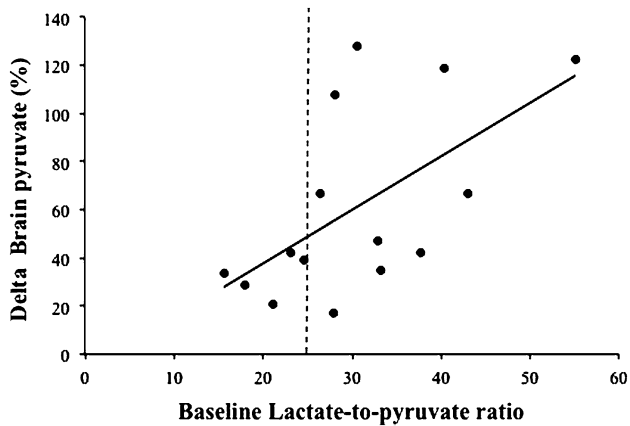
CMD cerebral microdialysis, PbtO<sub>2</sub> brain tissue oxygen tension



**Fig. 4** Dynamic changes over time of systemic lactate and brain energy metabolites upon sodium lactate therapy in one representative patient. The line graph shows the time course in one illustrative patient of cerebral microdialysis (CMD) glucose, CMD pyruvate, and systemic and CMD lactate during the 6 h prior to the intervention, during sodium lactate supplementation (box with dashed lines), and over the 3 h following the treatment

brain injury that exogenous systemic lactate may be utilized aerobically as a preferential energy substrate over glucose; (2) by increasing the availability of cerebral

extracellular pyruvate and glucose, exogenous lactate supplementation resulted in improved brain energetics during the early phase of TBI, (3) hypertonic lactate



**Fig. 5** Lactate conversion to pyruvate is proportional to baseline cerebral energy demand. The graph shows the positive linear correlation between baseline lactate-to-pyruvate ratio (marker of cerebral energy demand) and the extent of CMD pyruvate increase (delta brain pyruvate, expressed in % increase from baseline) observed during sodium lactate infusion ( $n = 15$  patients with severe TBI; Pearson's  $r = 0.60$ ,  $p = 0.02$ )

therapy was associated with a reduction of ICP and brain glutamate. Overall our findings suggest that hypertonic lactate therapy has both a positive effect on cerebral energy metabolism and on ICP reduction, suggesting that it may be a valid therapeutic option to test in the future in patients with TBI.

#### Evidence of lactate utilization by the injured human brain

Generally considered as a by-product of anaerobic metabolism, lactate was shown to support synaptic function [7] and to act as an alternative substrate to glucose [8–10]. Evidence of endogenous lactate utilization has been provided in humans with TBI [12] and subarachnoid hemorrhage (SAH) [26]. This may be important after TBI, where utilization of the predominant brain substrate (glucose) can be dramatically increased [2, 3, 27], leading to reduced glucose availability and worse outcome [5, 6].

We showed that supplemental lactate from the systemic circulation may be converted by the brain into pyruvate, and we found that cerebral pyruvate increase was proportionate to the extent of cerebral energy dysfunction. Importantly, pyruvate increase was coupled with a moderate but statistically significant decrease of PbtO<sub>2</sub>, whereas LPR remained stable during sodium lactate therapy. PbtO<sub>2</sub> is equal to the product of CBF with the difference between PaO<sub>2</sub> and PvO<sub>2</sub> [28]. During the study, CPP, PaCO<sub>2</sub>, MAP, and temperature (all major determinants of CBF) did not change. PaO<sub>2</sub> also was kept stable throughout the study. Actually, in a subset of patients ( $n = 6$ ) in whom transcranial Doppler was measured during sodium lactate infusion, a significant

increase in CBF velocities was found (data not shown), in line with experimental findings showing that lactate can act as local cerebral vasodilator [29]. Therefore, we can conclude that the observed reduction of PbtO<sub>2</sub> is secondary to a decrease in PvO<sub>2</sub>, indicating increased oxygen extraction by the tissue. These findings suggest that lactate was actively converted into pyruvate to be utilized aerobically by the injured human brain. Additional issues warrant further discussion. The first pertains to the relationship between lactate and oxygen. Lactate can increase as a consequence of anaerobic glycolysis, where increased lactate production is due to insufficient oxygen availability that will hamper the Krebs cycle metabolism of pyruvate. Under such conditions, exogenously administered lactate cannot be metabolized further. However, emerging clinical data suggest that lactate increase is mainly non-hypoxic in patients with TBI [13, 30, 31]. In the present study, TBI patients did not display low PbtO<sub>2</sub>, meaning that the administered lactate could indeed be converted to pyruvate, which is subsequently metabolized via the TCA cycle and the associated ATP-generating oxidative phosphorylation. This metabolic sequence occurs under normoxic conditions, and is also known as the Warburg effect [32]. In our study, lactate is provided exogenously under essentially normoxic conditions, which allows its utilization by lactate-consuming brain cells to produce ATP. The second issue relates to cerebral extracellular pyruvate increase. It should be kept in mind that the concentrations of lactate are about 10 times higher than those of pyruvate (2–3 mmol/L vs. 100 μmol/L). Thus, given the mass action effect, the normoxic conditions (which allow conversion of lactate to pyruvate) and the relative kinetic properties of lactate dehydrogenase (LDH) type A and pyruvate dehydrogenase, it is not surprising that more pyruvate is formed that can be metabolized through the TCA cycle, thereby resulting in a net increase of CMD pyruvate concentrations.

#### Lactate supplementation is associated with sparing of cerebral glucose

Sparing of cerebral glucose was demonstrated by the concomitant increase of CMD glucose concentration. Given that systemic glucose did not change during sodium lactate infusion, the observed increase of CMD glucose can only be attributable to preferential lactate utilization over glucose by the injured brain. Our study confirms previous experimental data and studies in healthy humans [18, 23]. Increasing the availability of cerebral glucose is important in the setting of human TBI where reductions of CMD glucose are frequent and are associated with worse prognosis [5, 6]. Restoration of cerebral glucose reserves by hypertonic lactate therapy indeed appears to be of benefit in the setting of TBI. Glucose is the essential substrate for brain metabolism,

participating in bioenergetics, reactive oxygen species regulation, neurotransmission, and biosynthesis [33]. The use of lactate as an alternative energy fuel could enable extra glucose to be available during the early phase of injury for these important rescue processes.

#### Additional effects of sodium lactate supplementation

Sodium lactate therapy was associated with a reduction of brain glutamate, which almost reached statistical significance ( $p = 0.06$ ). Given that glutamate was stable prior to sodium lactate therapy, we considered the observed glutamate decrease as physiologically and clinically relevant. Glutamate accounts for 80–90 % of the synapses in the brain and is the major excitatory neurotransmitter. The concentration of glutamate into the synaptic space rises dramatically after TBI, and failure of astrocytes to remove this excess of extracellular glutamate might lead to excitotoxic damage and neuronal death [34]. Increased glutamate release is a concomitant marker of energy failure; thus, providing energy in the form of lactate would allow neurons to control glutamate release and prevent glutamate “leak”. Lactate supplementation leads to increased glucose availability to sustain glutamate uptake in astrocytes.

Sodium lactate infusion also resulted in a significant reduction of ICP, which was most marked in patients with intracranial hypertension, further substantiating the view that hypertonic lactate may be a valid therapeutic option after TBI. Since the majority of patients did not have elevated ICP, additional studies are needed to confirm the effect of hypertonic lactate solutions in patients with intracranial hypertension. Lactate was administered as a hypertonic solution; therefore, we cannot completely rule out a potential anti-edematous effect: further discrimination between the metabolic and the anti-edematous effect of hypertonic lactate solutions requires future studies comparing hypertonic lactate to hypertonic saline.

#### Clinical implications of the study

Sodium lactate may be used for volume resuscitation, providing positive effects on brain energy metabolism, while avoiding adverse effects (hyperchloremic acidosis) of other standard resuscitative solutions, such as sodium chloride [35]. Hypertonic lactate could be an alternative for the treatment of intracranial hypertension [36, 37]. Altogether the findings of this prospective interventional study warrant further clinical investigation in a larger setting to confirm whether sodium lactate therapy is beneficial in patients with TBI and other forms of acute cerebral diseases, and to compare the effects of hypertonic lactate versus standard hypertonic saline solutions for the treatment of intracranial hypertension.

#### Study limitations

The main limitation of this study is the small sample size. However the cohort consisted of a homogenous group of patients with isolated severe TBI, with predominantly diffuse injury, who were all monitored in the same tissue region and were treated with a standardized algorithm. The time from TBI to sodium lactate therapy was variable among patients; however, this did not influence the results. Lactate therapy was given over a limited period of time (3 h). This was mainly to limit the potential side effects (hypernatremia, increased osmolarity, metabolic alkalosis), the relevance of which is unclear. Although the main relevant systemic physiological variables (MAP, heart rate, body temperature, PaO<sub>2</sub>, and PaCO<sub>2</sub>) did not change over time, we cannot entirely exclude potential confounders; therefore, our findings in this small cohort of patients must be considered as preliminary. Additional limitations include that brain metabolism was measured using CMD and PbtO<sub>2</sub>, i.e., with focal monitoring. This approach has several advantages, e.g., it allows the monitoring of dynamic changes of different brain metabolites upon therapy and it is accessible at the bedside. However, we did not use PET scans in this study and therefore we cannot provide global measurement of cerebral lactate uptake and CMRO<sub>2</sub>. Despite these technical limitations, we believe that combined monitoring of CMD, PbtO<sub>2</sub>, and ICP provided us with essential tools to test the dynamic changes of cerebral energy metabolism following sodium lactate therapy.

#### Conclusions

We showed that exogenous systemic lactate is utilized by the injured human brain as a preferential energy substrate with subsequent sparing of cerebral glucose. The increase of pyruvate and glucose in brain interstitial tissue— together with the decrease of cerebral glutamate and intracranial pressure— suggests that hypertonic lactate therapy has positive effects on cerebral energy metabolism and on ICP reduction in the early phase following TBI. Our findings support further investigation to test the value of this therapeutic strategy in patients with acute brain injury and to evaluate whether the observed cerebral biochemical and physiological effects of hypertonic lactate might translate into a better neurological outcome.

**Acknowledgments** The authors thank Béatrice Pellet, from the Division of Pharmacy, for the preparation of sodium lactate, as well as all neurosurgical fellows, ICU physicians, and ICU nurses for their help and support. This work was supported by grants from the Swiss National Science Foundation (Grant Nr. 320030\_138191, to M.O.), the European Critical Care Research Network, European Society of Intensive Care Medicine (to M.O.), and the Gueules Cassées Foundation (to P.B.). This work received the best abstract award at the European Society of Intensive Care Medicine congress in Paris, 2013.



**Conflicts of interest** The authors have no conflict of interest to declare.

## References

1. Maas AI, Stocchetti N, Bullock R (2008) Moderate and severe traumatic brain injury in adults. *Lancet Neurol* 7:728–741
2. Bergsneider M, Hovda DA, Shalmon E, Kelly DF, Vespa PM, Martin NA, Phelps ME, McArthur DL, Caron MJ, Kraus JF, Becker DP (1997) Cerebral hyperglycolysis following severe traumatic brain injury in humans: a positron emission tomography study. *J Neurosurg* 86:241–251
3. Glenn TC, Kelly DF, Boscardin WJ, McArthur DL, Vespa P, Oertel M, Hovda DA, Bergsneider M, Hillered L, Martin NA (2003) Energy dysfunction as a predictor of outcome after moderate or severe head injury: indices of oxygen, glucose, and lactate metabolism. *J Cereb Blood Flow Metab* 23:1239–1250
4. Simpson IA, Carruthers A, Vannucci SJ (2007) Supply and demand in cerebral energy metabolism: the role of nutrient transporters. *J Cereb Blood Flow Metab* 27:1766–1791
5. Oddo M, Schmidt JM, Carrera E, Badjatia N, Connolly ES, Presciutti M, Ostapkovich ND, Levine JM, Le Roux P, Mayer SA (2008) Impact of tight glycemic control on cerebral glucose metabolism after severe brain injury: a microdialysis study. *Crit Care Med* 36:3233–3238
6. Vespa PM, McArthur D, O'Phelan K, Glenn T, Etchepare M, Kelly D, Bergsneider M, Martin NA, Hovda DA (2003) Persistently low extracellular glucose correlates with poor outcome 6 months after human traumatic brain injury despite a lack of increased lactate: a microdialysis study. *J Cereb Blood Flow Metab* 23:865–877
7. Schurr A, West CA, Rigor BM (1988) Lactate-supported synaptic function in the rat hippocampal slice preparation. *Science* 240:1326–1328
8. Pellerin L, Pellegrini G, Bittar PG, Charnay Y, Bouras C, Martin JL, Stella N, Magistretti PJ (1998) Evidence supporting the existence of an activity-dependent astrocyte-neuron lactate shuttle. *Dev Neurosci* 20:291–299
9. Schurr A, Miller JJ, Payne RS, Rigor BM (1999) An increase in lactate output by brain tissue serves to meet the energy needs of glutamate-activated neurons. *J Neurosci* 19:34–39
10. Schurr A, Payne RS (2007) Lactate, not pyruvate, is neuronal aerobic glycolysis end product: an in vitro electrophysiological study. *Neuroscience* 147:613–619
11. Magistretti PJ (2008) Brain energy metabolism. In: Berg D, Bloom FE, du Lac S, Ghosh A, Spitzer NC, Squire LR (eds) *Fundamental neurosciences*. Academic, San Diego, pp 271–293
12. Gallagher CN, Carpenter KL, Grice P, Howe DJ, Mason A, Timofeev I, Menon DK, Kirkpatrick PJ, Pickard JD, Sutherland GR, Hutchinson PJ (2009) The human brain utilizes lactate via the tricarboxylic acid cycle: a <sup>13</sup>C-labelled microdialysis and high-resolution nuclear magnetic resonance study. *Brain* 132:2839–2849
13. Sala N, Suys T, Zerlauth JB, Bouzat P, Messerer M, Bloch J, Levivier M, Magistretti PJ, Meuli R, Oddo M (2013) Cerebral extracellular lactate increase is predominantly nonischemic in patients with severe traumatic brain injury. *J Cereb Blood Flow Metab* 33:1815–1822
14. Pellerin L, Magistretti PJ (1994) Glutamate uptake into astrocytes stimulates aerobic glycolysis: a mechanism coupling neuronal activity to glucose utilization. *Proc Natl Acad Sci U S A* 91:10625–10629
15. Pellerin L, Magistretti PJ (2012) Sweet sixteen for ANLS. *J Cereb Blood Flow Metab* 32:1152–1166
16. Chen T, Qian YZ, Rice A, Zhu JP, Di X, Bullock R (2000) Brain lactate uptake increases at the site of impact after traumatic brain injury. *Brain Res* 861:281–287
17. Jalloh I, Helmy A, Shannon RJ, Gallagher CN, Menon DK, Carpenter KL, Hutchinson PJ (2013) Lactate uptake by the injured human brain: evidence from an arteriovenous gradient and cerebral microdialysis study. *J Neurotrauma*. doi:10.1089/neu.2013.2947
18. Bouzier-Sore AK, Voisin P, Canioni P, Magistretti PJ, Pellerin L (2003) Lactate is a preferential oxidative energy substrate over glucose for neurons in culture. *J Cereb Blood Flow Metab* 23:1298–1306
19. Rice AC, Zsoldos R, Chen T, Wilson MS, Alessandri B, Hamm RJ, Bullock MR (2002) Lactate administration attenuates cognitive deficits following traumatic brain injury. *Brain Res* 928:156–159
20. Bratton SL, Chestnut RM, Ghajar J, McConnell Hammond FF, Harris OA, Hartl R, Manley GT, Nemecek A, Newell DW, Rosenthal G, Schouten J, Shutter L, Timmons SD, Ullman JS, Videtta W, Wilberger JE, Wright DW (2007) Guidelines for the management of severe traumatic brain injury. IX. Cerebral perfusion thresholds. *J Neurotrauma* 24(Suppl 1):S59–S64
21. Bratton SL, Chestnut RM, Ghajar J, McConnell Hammond FF, Harris OA, Hartl R, Manley GT, Nemecek A, Newell DW, Rosenthal G, Schouten J, Shutter L, Timmons SD, Ullman JS, Videtta W, Wilberger JE, Wright DW (2007) Guidelines for the management of severe traumatic brain injury. VIII. Intracranial pressure thresholds. *J Neurotrauma* 24(Suppl 1):S55–S58
22. Boumezebur F, Petersen KF, Cline GW, Mason GF, Behar KL, Shulman GI, Rothman DL (2010) The contribution of blood lactate to brain energy metabolism in humans measured by dynamic <sup>13</sup>C nuclear magnetic resonance spectroscopy. *J Neurosci* 30:13983–13991
23. Smith D, Pernet A, Hallett WA, Bingham E, Marsden PK, Amiel SA (2003) Lactate: a preferred fuel for human brain metabolism in vivo. *J Cereb Blood Flow Metab* 23:658–664
24. Revelly JP, Tappy L, Martinez A, Bollmann M, Cayeux MC, Berger MM, Chioloro RL (2005) Lactate and glucose metabolism in severe sepsis and cardiogenic shock. *Crit Care Med* 33:2235–2240
25. Marshall LF, Marshall SB, Klauber MR, Van Berkum Clark M, Eisenberg H, Jane JA, Luerksen TG, Marmarou A, Foulkes MA (1992) The diagnosis of head injury requires a classification based on computed axial tomography. *J Neurotrauma* 9(Suppl 1):S287–S292
26. Oddo M, Levine JM, Frangos S, Maloney-Wilensky E, Carrera E, Daniel RT, Levivier M, Magistretti PJ, LeRoux PD (2012) Brain lactate metabolism in humans with subarachnoid hemorrhage. *Stroke* 43:1418–1421
27. Bergsneider M, Hovda DA, McArthur DL, Etchepare M, Huang SC, Sehati N, Satz P, Phelps ME, Becker DP (2001) Metabolic recovery following human traumatic brain injury based on FDG-PET: time course and relationship to neurological disability. *J Head Trauma Rehabil* 16:135–148

- 
28. Rosenthal G, Hemphill JC 3rd, Sorani M, Martin C, Morabito D, Obrist WD, Manley GT (2008) Brain tissue oxygen tension is more indicative of oxygen diffusion than oxygen delivery and metabolism in patients with traumatic brain injury. *Crit Care Med* 36:1917–1924
  29. Gordon GR, Choi HB, Rungta RL, Ellis-Davies GC, MacVicar BA (2008) Brain metabolism dictates the polarity of astrocyte control over arterioles. *Nature* 456:745–749
  30. Hutchinson PJ, O’Connell MT, Seal A, Nortje J, Timofeev I, Al-Rawi PG, Coles JP, Fryer TD, Menon DK, Pickard JD, Carpenter KL (2009) A combined microdialysis and FDG-PET study of glucose metabolism in head injury. *Acta Neurochir (Wien)* 151:51–61; discussion 61
  31. Vespa P, Bergsneider M, Hattori N, Wu HM, Huang SC, Martin NA, Glenn TC, McArthur DL, Hovda DA (2005) Metabolic crisis without brain ischemia is common after traumatic brain injury: a combined microdialysis and positron emission tomography study. *J Cereb Blood Flow Metab* 25:763–774
  32. Vander Heiden MG, Cantley LC, Thompson CB (2009) Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 324:1029–1033
  33. Dienel GA (2012) Brain lactate metabolism: the discoveries and the controversies. *J Cereb Blood Flow Metab* 32:1107–1138
  34. McKenna MC (2007) The glutamate-glutamine cycle is not stoichiometric: fates of glutamate in brain. *J Neurosci Res* 85:3347–3358
  35. Roquilly A, Loutrel O, Cinotti R, Rosenczweig E, Flet L, Mahe PJ, Dumont R, Chupin AM, Peneaux C, Lejus C, Blanloeil Y, Volteau C, Asehnoune K (2013) Balanced versus chloride-rich solutions for fluid resuscitation in brain-injured patients: a randomized double-blind pilot study. *Crit Care* 17:R77
  36. Ichai C, Armando G, Orban JC, Berthier F, Rami L, Samat-Long C, Grimaud D, Leverve X (2009) Sodium lactate versus mannitol in the treatment of intracranial hypertensive episodes in severe traumatic brain-injured patients. *Intensive Care Med* 35:471–479
  37. Ichai C, Payen JF, Orban JC, Quintard H, Roth H, Legrand R, Francony G, Leverve XM (2013) Half-molar sodium lactate infusion to prevent intracranial hypertensive episodes in severe traumatic brain injured patients: a randomized controlled trial. *Intensive Care Med* 39:1413–1422