



Changes in the phenylalanine ammonia lyase activity, total phenolic compounds, and flavonoids in *Prosopis glandulosa* treated with cadmium and copper

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ABSTRACT

The aim of the present work is to evaluate the changes on the phenylalanine ammonia lyase (PAL) activity, phenolic compounds accumulation and photochemical efficiency in leaves of *P. glandulosa* treated with Cd²⁺ (0.001 M) and Cu²⁺ (0.52 M) concentrations for 96 h under hydroponic conditions. The results showed that only leaves treated with copper had a decrease in photochemical efficiency and leaf epidermal polyphenols in *P. glandulosa* leaves after 96 h of exposure. On the other hand the reverse-phase HPLC analysis revealed higher levels of phenolic compound (gallic, vanillic and caffeic acids) and flavonoids (rutin and kaempferol-3-O-glucosides) in plant leaves from Cu and Cd-treatments with respect to control plants. Finally, highest increments in PAL activity was observed in extracts of leaves treated with Cu and Cd (about 205 and 284%), respectively, with respect to control plants after 96 h treatment. These suggest that activation of phenylpropanoid pathway represent a source of nonenzymatic antioxidants that protect at *P. glandulosa* against oxidative stress when exposed to cadmium and copper. Hence future studies are necessary to elucidate the participation of phenylpropanoid pathway in the reduction of metal toxicity in *Prosopis* species.

Key words: heavy metals, *Prosopis glandulosa*, flavonoids, chlorophyll a fluorescence.

INTRODUCTION

As a result of a rapid industrialization and urbanization processes in the northwest of Mexico, the release of several contaminants like heavy metals in the ecosystem has increased and became

a serious problem in these ecoregion of Mexico (Páez-Osuna et al. 2015, Galindo et al. 2010). The heavy metals cause bioaccumulation affecting the entire ecosystem and pose harmful health consequences in all life forms. In the plants, these elements can interfere with different physiological and biochemical processes of the plant system (Sethy and Ghosh 2013). The presence of copper

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and cadmium in the environment generate negative effects in human health due to its high potential to enter and accumulate in the food chain (Pena et al. 2011, Tchounwou et al. 2012). The inadequate supply of both elements results in a variety of changes in plants like inactivation of photosynthesis, formation of free radical and reactive oxygen species, which result in an oxidative stress (Gonzalez-Mendoza et al. 2013). In recent years, some reports have shown that genus *Prosopis*, can be tolerant to essential and not essential metals even before any visible signs of toxicity are shown (Varun et al. 2011). Due to its ecological and commercial importance, *Prosopis* species has been extensively used in the last few years to study the effects of heavy metals in the environment (Beramendi-Orosco et al. 2013, Buendía-González et al. 2007). Currently different studies suggested that *Prosopis* species mainly *P. juliflora*, show the presence of a competent metal uptake and accumulation machinery in roots (Senthilkumar et al. 2005, Mokgalaka-Matlala et al. 2008, Usha et al. 2009).

However, even though the physiological and biochemical mechanisms of heavy metal tolerance or inactivation in *Prosopis* species have been previously studied, the influence of copper and cadmium in the phenylalanine ammonia lyase (PAL) activity and phenolic compounds accumulation in *Prosopis glandulosa* has been scarcely evaluated. Therefore, the aim of the present work is to evaluate the changes on the phenylalanine ammonia lyase (PAL) activity, phenolic compounds accumulation and photochemical efficiency in leaves of *P. glandulosa* exposed to cadmium and copper.

MATERIALS AND METHODS

DISINFESTION OF SEEDS

One hundred seeds of *Prosopis glandulosa* was disinfested with 1.5% NaOCl for 3 min, followed of a washed with abundant water. Later, the seedlings were cultivated with 12 h light:dark

photoperiods in a greenhouse, 60% relative air humidity temperature and day/night temperatures a range of 30/32 °C were used. The fertilization of seedlings was according to previous described by Gonzalez-Mendoza et al. (2013).

CADMIUM AND COPPER EXPOSURE

Twenty-three-days-old *P. glandulosa* seedlings were randomly selected and transferred to individual plastic containers with 250 ml Hoagland's solution prepared with any one of the following heavy metal exposure levels ($n=10$ per treatment level): (a) 0.001 M cadmium chloride (CdCl_2) and (b) 0.52 M copper sulfate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$). Control plants were transferred to plastic containers with 250 ml Hoagland's solution without CdCl_2 or $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$. Treated and control plants were exposed to their respective solutions for 96 h under hydroponics conditions.

DETERMINATION OF PHYSIOLOGICAL PARAMETERS

The determination of polyphenol contents (Phen) of the plant leaves were measured on the middle part of the third pair fully expanded leaves using an optical sensor (Dualox Scientific+™) according to (Cartelat et al. 2005). For pigment analysis, the content of polyphenols was expressed as Dualox units. Assessments were realized at 0, 24, 48, 72 and 96 h after treatment with copper and cadmium. The chlorophyll fluorescence was analyzed with an OS-30p Fluorometer (OPTI-SCIENCE, USA) on completely expanded leaves. Previously, the plant leaves were dark-adapted for 5 min. The potential photochemical yield (F_v/F_m) was recorded at a saturating actinic light (660 nm) intensity of 1100 $\mu\text{mol}/\text{m}^2/\text{s}$. For each treatment, the F_v/F_m of ten individual leaves were measured at 0, 24, 48, 72 and 96 after exposure to heavy metals according to proposed by Gonzalez-Mendoza et al. (2013).

IDENTIFICATION OF PHENOLIC ACIDS AND FLAVONOIDS BY HPLC

Qualitative and quantitative analyses of phenolic compounds and flavonoids in *P. glandulosa* samples were done by gradient elution high performance liquid chromatography (HPLC) analysis 96 h after of exposure to cadmium and copper prepared according to (Cervantes-Garcia et al. 2016). Separation of phenolic acids was carried out by The HPLC system consisted of quaternary pump model 9012, equipped with an UV detector model 9050, a Prodigy 5u ODS3 100A (Phenomenex, CA, USA) column (250 mm length, 4.6 mm of internal diameter and 5 µm particle size) with a C-18 guard column. The detector was set to 280 nm and flow rate to 1 ml/min (Varian Inc., Co. Palo Alto, CA, USA). The solutions of the phenolic acid mixture were prepared with 0.01 g of each phenolic acid and mixed with a solution of 1:1 ammonium acetate buffer 200 mM pH 5.4 and methanol.

The efficiency of the HPLC methodology for separation of phenolic acids and flavonoids was tested by preparing a solution containing the following commercially available standards: gallic acid, protocatechuic acid, hydroxybenzoic acid, vanillic acid, chlorogenic acid, caffeic acid, ferulic acid, *p*-coumaric acid, sinapic acid, *o*-coumaric acid, naringenin, kaempferol, quercetin, rutin, naringin and cinnamic acid (Sigma-Aldrich Chemical Co. St. Louis, MO, USA).

PHENYLALANINE AMMONIA-LYASE (PAL) ACTIVITY ASSAY

To assess PAL activity, 2 g of leaves tissues of *P. glandulosa* were homogenized in 5 mL of Tris-HCl buffer (0.1 M, pH 7.5) containing 1% polyvinylpyrrolidone and EDTA (1 mM) after 96 h of exposure to heavy metals. The homogenate was centrifuged at 12,000 rpm, at 4 °C for 10 min and was tested for PAL activity according to Cindi

et al. (2016) by measuring the absorbance of trans-cinnamic acid at 290 nm with some modifications. The reaction mixture contained the enzyme extract (100 µL), 200 µL of Tris-HCl (50 mM, pH 8.8) containing 20 mM L-phenylalanine and incubated for 30 min at 40 °C. After incubation time the reaction was stopped by the addition of 100 µL of 1 M HCl and the production of cinnamate was measured at 290 nm. One unit of the enzyme was defined as increase in absorbance of one unit per min. The activity of the enzyme was expressed as units per mg of soluble protein.

STATISTICAL ANALYSIS

Differences between the treatments were evaluated using one-way analyses of variance and the Tukey's test ($P \leq 0.05$), and SAS Version 9.0 (SAS Institute, 2002) was used.

RESULTS

EPIDERMAL POLYPHENOLS (Ephen) AND CHLOROPHYLL A FLUORESCENCE

The content of Ephen in control *P. glandulosa* leaf did not show any changes during the experiment (Fig. 1). However, when this species was exposed to a copper solution showed a decreased after 48 h of exposure to metal (Fig. 1). In contrast the Ephen in the *P. glandulosa* leaf exposed to cadmium did not exhibit significantly changes compared with the plants treated only with water (Fig. 1). The measurements of chlorophyll a fluorescence showed a significantly decrease ($P < 0.05$) on photochemical efficiency (F_v/F_m) value of *P. glandulosa* treated with copper dose from 48 hours followed by a dropped of 48% after 96 h of exposure with respect to control (Fig. 2). Finally, F_v/F_m values observed in leaf of *P. glandulosa* treated with cadmium did not show significantly changes with respect to control (Fig. 2).

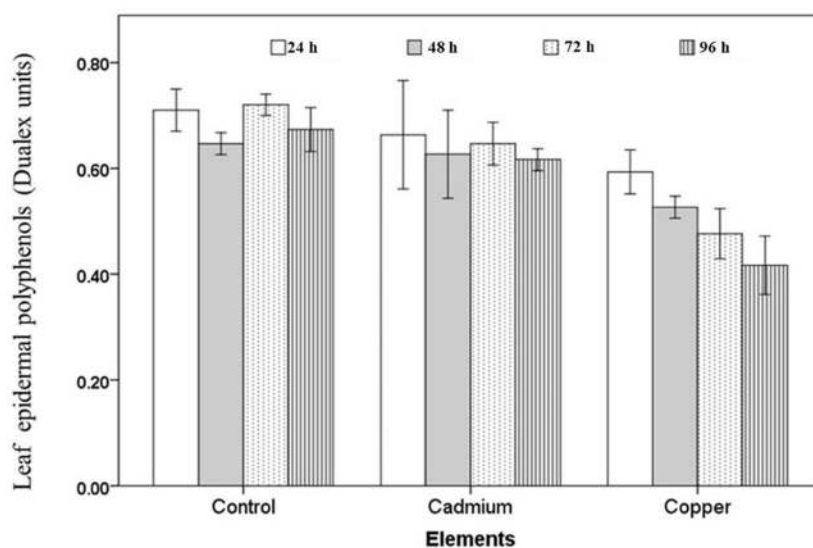


Figure 1 - Time course response of epidermal polyphenols (Dualex units) in leaves of *P. glandulosa* exposed to copper and cadmium after 96 h of treatment. Means \pm Standard error. $n=4$.

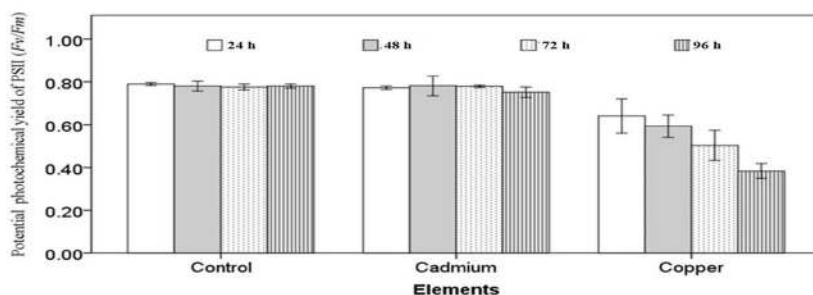


Figure 2 - Time course response of potential photochemical yield of PSII (F_v/F_m) measured in leaves of *P. glandulosa* exposed to copper and cadmium after 96 h of treatment. Means \pm Standard error. $n=4$.

IDENTIFICATION OF PHENOLIC ACIDS AND FLAVONOIDS BY HPLC

In the present study, seven individual phenolic compounds (gallic acid, hydroxybenzoic acid, chlorogenic acid, caffeic acid, vanillic acid, *p*-coumaric acid and ferulic acid) were analyzed and quantified using reverse-phase HPLC for all the plant treatments of *P. glandulosa* after 96 h of exposure (Fig. 3). Our result showed that Cd exposure increased gallic (10.5 ± 0.008), vanillic (3.14 ± 0.14) and caffeic acids (1.42 ± 0.2), in *P. glandulosa* leaves when compared to the control

treatment (Fig. 4). Also, there was a significant decrease in the concentrations of hydroxybenzoic acid (5.27 ± 0.5), *p*-coumaric acid (10.91 ± 0.29) and ferulic acid (1.35 ± 0.12) in leaves treated with Cd (Fig. 4). In contrast, our study revealed that plants treated with Cu, increased only caffeic acid (2.7 ± 0.02) in *P. glandulosa* leaves when compared to the control group after 96 h of treated (Fig. 4).

On the other hand, individual flavonoids compounds like naringenin, rutin, quercetin, kaempferol, kaempferol-3-O-rutinoside, kaempferol-3-O-glucoside and naringenin-chalcone were detected using reverse-phase HPLC

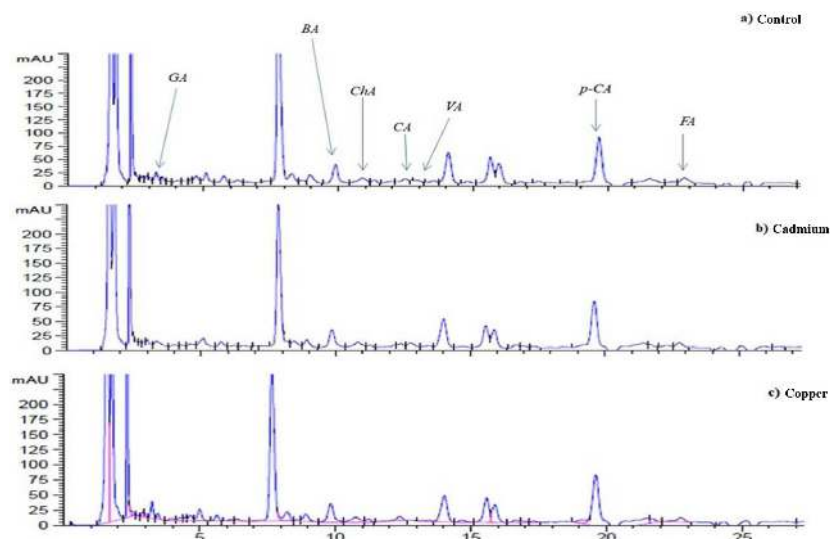


Figure 3 - Differences in phenolic compounds of hydrolysed extracts of *Prosopis glandulosa*: (a) control, (b) Cadmium (0.001 M) and (c) Copper (0.52 M) Peaks: GA, gallic acid; BA, hydroxybenzoic acid; ChA, chlorogenic acid; CA, caffeic acid; VA, vanillic acid; pCA, *p*-coumaric acid; FA, ferulic acid acid.

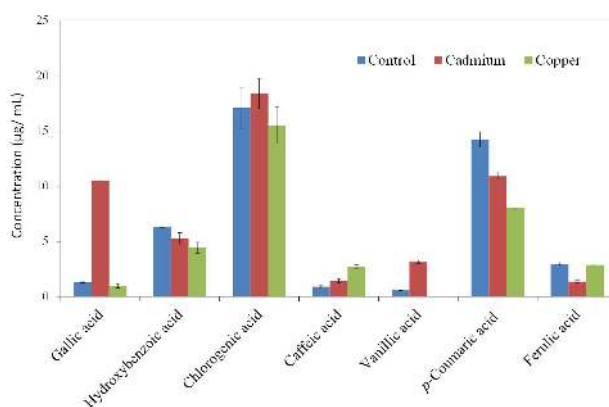


Figure 4 - Concentration of phenolic compounds of hydrolysed extracts of *Prosopis glandulosa* treated with Cadmium (0.001 M) and Copper (0.52 M). Means \pm Standard error. $n=4$.

for all the treatments of *P. glandulosa* after 96 h of exposure (Fig. 5). In the present study rutin and kaempferol-3-O-glucoside were present most abundantly in extracts of leaves from Cu and Cd-treatments with respect to control plants (Fig. 5). However, only Cd-treatment increased naringenin (2.65 ± 0.18) and kaempferol-3-O-rutinoside (3.14 ± 0.14) in *P. glandulosa* leaves when compared to the control treatment. In contrast, the highest amount of naringenin-chalcone was found in Cu-treatment

of leaf whereas minimum in Cd-treatment (Fig. 6). Finally, quercetin was significantly reduced in the leaves of all treatments with heavy metals but not in control plants (Fig. 6).

PHENYLALANINE AMMONIA-LYASE (PAL) ACTIVITY ASSAY

Our results showed changes in PAL activity of *P. glandulosa* treated with copper and cadmium after 96 h of exposure are shown in Figure 6. Highest

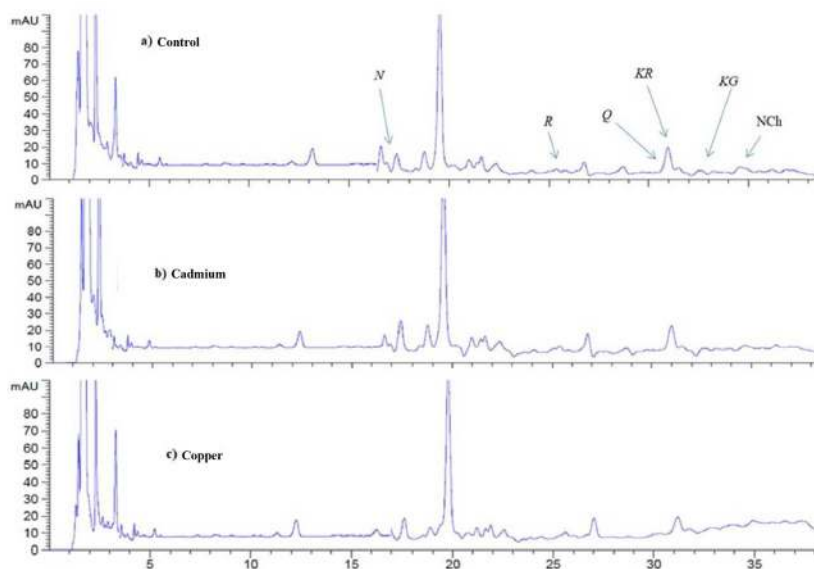


Figure 5 - Differences in flavonoid compounds of hydrolysed extracts of *Prosopis glandulosa*: (a) control, (b) Cadmium (0.001 M) and (c) Copper (0.52 M). Peaks: *N*, naringenin; *R*, rutin; *Q*, quercetin; *K*, kaempferol; *KR*, kaempferol-3-O-rutinoside; *KG*, kaempferol-3-O-glucoside; and *NCh*, naringenin-chalcone.

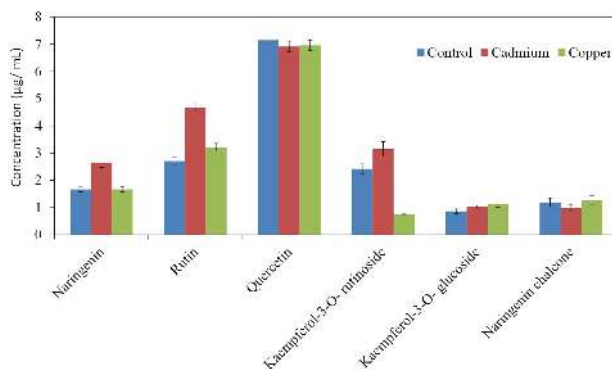


Figure 6 - Concentration of flavonoids of hydrolysed extracts of *Prosopis glandulosa* treated with Cadmium (0.001 M) and Copper (0.52 M). Means \pm Standard error. $n=4$.

increments in PAL activity was observed in extracts of leaves treated with Cu and Cd (about 205 and 284%), respectively, with respect to control plants (Table I).

DISCUSSION

Recent studies have suggested that the epidermal polyphenols (EPhen) content in plants, mainly flavonoids, may act as chelate agents of metal

ions reducing the presence of free radicals, and protect against oxidative stress (Mierziak et al. 2014). In the present study our results showed that the synthesis of EPhen was negatively affected when *P. glandulosa* was treated with copper. The low values of EPhen observed could be explained by impairment of antioxidative system due to the exposure to copper reducing the synthesis of new polyphenols.

TABLE I
Assessment of the activity of phenylalanine ammonia lyase (PAL) in *Prosopis glandulosa* after 96 h exposure to heavy metals.

Treatments	PAL (units/mg soluble protein)
<i>P. glandulosa</i> control	73.11 ± 2.79 ^a
<i>P. glandulosa</i> -Cadmium	207.73 ± 0.64 ^b
<i>P. glandulosa</i> -Copper	150.28 ± 8.71 ^c

Data are means ± S.D. ($n=4$). Those with different superscript letter (a, b, and c) in the same column are significantly different ($P < 0.05$, tukey multiple comparison).

Thus as a results of cooperation of EPhen with different antioxidant metabolites to scavenge the hydrogen peroxide which leaks out from mesophyll cells where it is produced in response to heavy metals such as copper (Sakihama et al. 2002, Mierziak et al. 2014). Surprisingly, no effects were observed when *P. glandulosa* was exposed to cadmium. This response to this metal may be the result of the presence of biochemistry strategies such as synthesis of metallothioneins which help in the reduction of stress caused by cadmium in different plants (Gonzalez-Mendoza et al. 2007, Gonzalez-Mendoza and Zapata-Perez 2008). On the other hand, the lack of increase in phenolic and flavonoid compounds in *P. glandulosa* exposed to 0.001 M Cd⁺² could be attributed to the impairment of antioxidative system responses due to the exposure to a high concentration of this metal, in such a way that this organism is not able to synthesize new phenolic and flavonoid compounds. Similar results has been reported in *Euglena gracilis*, which is not able to counteract the effects of exposure to high concentration of Cd⁺² (Cervantes-Garcia et al. 2016). Recent studies has reported that flavonoid and phenolic compounds could act as alternative antioxidants when antioxidative enzymes are affect by heavy metals (Márquez-García et al. 2009). In this sense our results showed that copper and cadmium stimulates the activity of PAL in *P. glandulosa*

leaves. This suggest that PAL activation and increased of phenolics and flavonoids compounds can act as source of nonenzymatic antioxidants and protect *P. glandulosa* from oxidative stress when exposed to heavy metals (Cd and Cu). These results are in agreement with previous studies reported by Pawlak-Sprada et al. (2011), that suggest antioxidant and metal chelating properties of phenolic and flavonoid compounds in plants exposed to heavy metals.

Additionally, our results showed that the reduction in *Fv/Fm* values observed in *P. glandulosa* could be explained by negative effects of copper on the structure and composition of reaction center complex (RC) of photosystem II reaction centers (Gonzalez-Mendoza et al. 2013). Nevertheless, the differences in the *Fv/Fm* values among Cd⁺² and Cu⁺² may be related to their capacity of acquisition and transport by *P. glandulosa* leaves. For example, previous studies showed the presence low values of cadmium-transportation index in *Prosopis juliflora* suggesting the participation of exclusion processes that regulate the metals uptake in leaves (Michel-López et al. 2016). Finally, according to our research, high production of phenolic and flavonoid compounds in plants of *P. glandulosa* under the cadmium and copper influence during the experiment are consistent with a significant increase in the PAL activity. This suggests that activation of phenylpropanoid pathway represent asource of nonenzymatic antioxidants that protect at *P. glandulosa* against oxidative stress when exposed to heavy metals. The decreased quercetin and hydroxybenzoic acid, *p*-coumaric acid and ferulic acid accumulation in *P. glandulosa* leaves in response to heavy metals, despite an increased PAL activity, suggests that the activated phenylpropanoid pathway was involved in the synthesis of secondary metabolites other than phenolics compounds. Hence future studies are necessary to elucidate the participation of phenylpropanoid pathway in the reduction of metal toxicity in *Prosopis* species.

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REFERENCES

- BERAMENDI-OROSCO LE, RODRIGUEZ-ESTRADA ML, MORTON-BERMEA O, ROMEO FM, GONZALEZ-HERNANDEZ G AND HERNANDEZ-ALVAREZ E. 2013. Correlations between metals in tree-rings of *Prosopis juliflora* as indicators of sources of heavy metal contamination. *Appl Geochem* 39: 78-84.
- BUENDÍA-GONZÁLEZ L, OROZCO-VILLAFUERTE J, CRUZ-SOSA F, CHÁVEZ-ÁVILA VM AND VERNON-CARTER EJ. 2007. Clonal propagation of mesquite tree (*Prosopis laevigata* Humb. & Bonpl. ex Willd. M.C. Johnston). I. Via cotyledonary nodes. *In Vitro Cell Dev Biol Plant* 43: 260-266.
- CARTELAT A ET AL. 2005. Optically assessed contents of leaf polyphenolics and chlorophyll as indicators of nitrogen deficiency in wheat (*Triticum aestivum* L.) *Field Crops Res* 91: 35-49.
- CERVANTES-GARCIA D, TRONCOSO-ROJAS R, SÁNCHEZ-ESTRADA A, GONZALEZ-MENDOZA D, GUTIERREZ-MICELI F, CECEÑA-DURAN C AND GRIMALDO-JUAREZ O. 2016. Effects of cadmium on total phenolic compounds and flavonoids in *Euglena gracilis*. *Gayana* 80: 1-5.
- CINDI MD, SOUNDY P, ROMANAZZI G AND SIVAKUMAR D. 2016. Different defense responses and brown rot control in two *Prunus persica* cultivars to essential oil vapours after storage. *Postharvest Biol Technol* 119: 9-17.
- GALINDO EAG, BARBOSA AM, VELASCO MRM, DAESLÉ LW AND OROZCO BORBÓN MV. 2010. Distribution and Enrichment of Silver and Cadmium in Coastal Sediments from Bahia Todos os Santos, Baja California, Mexico. *Bull Environ Contam Toxicol* 85: 391-396.
- GONZALEZ-MENDOZA D, FRANCISCO ESPADAS G, ESCOBOZA-GARCIA F, SANTAMARIA JM AND ZAPATA-PEREZ O. 2013. Copper Stress on Photosynthesis of Black Mangle (*Avicennia germinans*). *An Acad Bras Cienc* 85: 665-670.
- GONZALEZ-MENDOZA D, MORENO AQ AND ZAPATA-PEREZ O. 2007. Coordinated responses of phytochelatin synthase and metallothionein genes in black mangrove, *Avicennia germinans*, exposed to cadmium and copper. *Aquat Toxicol* 83: 306-314.
- GONZALEZ-MENDOZA D AND ZAPATA-PEREZ O. 2008. Mecanismos de tolerancia a elementos potencialmente tóxicos en plantas. *Bol Soc Bot México* 82: 53-61
- MÁRQUEZ-GARCÍA B, FERNÁNDEZ MA AND CÓRDOBA F. 2009. Phenolics composition in *Erica* sp. differentially exposed to metal pollution in the Iberian southwestern Pyritic Belt. *Bioresour Bioprocess* 100: 446-451.
- MICHEL-LÓPEZ C, ESPADA-GIL F, FUENTES-ORTIZ G, SANTAMARIA JM, GONZALEZ-MENDOZA D, CECEÑA-DURAN C AND GRIMALDO-JUAREZ O. 2016. Bioaccumulation and effect of cadmium in the photosynthetic apparatus of *Prosopis juliflora*. *Chemical Speciation and Bioavailability* 28: 1-4.
- MIERZIAK J, KOSTYN K AND KULMA A. 2014. Flavonoids as Important Molecules of Plant Interactions with the Environment. *Molecules* 19: 16240-16265
- MOKGALAKA-MATLALA NS, FLORES-TAVIZÓN E, CASTILLO-MICHEL H, PERALTA-VIDEA JR AND GARDEA-TORRESDEY JL. 2008. Toxicity of arsenic (III) and (V) on plant growth, element uptake, and total amyolytic activity of mesquite (*Prosopis juliflora* x *P. velutina*). *Int J Phytoremediation* 10: 47-60.
- PÁEZ-OSUNA F, BOJÓRQUEZ-LEYVA H, BERGÉS-TIZNADO M, RUBIO-HERNÁNDEZ OA, FIERRO-SAÑUDO JF, RAMÍREZ-ROCHÍN J AND LEÓN-CAÑEDO JA. 2015. Heavy metals in waters and suspended sediments affected by a mine tailing spill in the upper San Lorenzo River, Northwestern Mexico. *Bull Environ Contam Toxicol* 94: 583-588.
- PAWLAK-SPRADA S, ARASIMOWICZ-JELONEK M, PODGÓRSKA M AND DECKERT J. 2011. Activation of phenylpropanoid pathway in legume plants exposed to heavy metals. Part I. Effects of cadmium and lead on phenylalanine ammonia-lyase gene expression, enzyme activity and lignin content. *Acta Biochim Pol* 58: 211-216.
- PENA LB, AZPILICUETA CE AND GALLEGO SM. 2011. Sunflower cotyledons cope with copper stress by inducing catalase subunits less sensitive to oxidation. *J Trace Elem Med Biol* 25: 125-129.
- SAKIHAMA Y, COHEN MF, GRACE SC AND YAMASAKI H. 2002. Plant phenolic antioxidant and prooxidant activities: phenolics-induced oxidative damage mediated by metals in plants. *Toxicology* 177: 67-80.
- SENTHILKUMAR P, PRINCE W, SIVAKUMAR S AND SUBBHURAAM CV. 2005. *Prosopis juliflora* – A green solution to decontaminate heavy metal (Cu and Cd) contaminated soils. *Chemosphere* 60: 1493-1496.
- SETHY SK AND GHOSH S. 2013. Effect of heavy metals on germination of seeds. *J Nat Sc Biol Med* 4: 272-275.
- TCHOUNWOU PB, YEDJOU CG, PATLOLLA AK AND SUTTON DJ. 2012. Heavy Metals Toxicity and the Environment. *EXS* 101: 133-164.
- USHA B, VENKATARAMAN G AND PARIDA A. 2009. Heavy metal and abiotic stress inducible metallothionein isoforms from *Prosopis juliflora* (SW) DC show differences in binding to heavy metals *in vitro*. *Mol Genet Genomics* 281: 99-108.
- VARUN M, D'SOUZA R, PRATAS J AND PAUL MS. 2011. Phytoextraction potential of *Prosopis juliflora* (Sw.) DC. with specific reference to lead and cadmium. *Bull Environ Contam Toxicol* 87: 45-49.