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Characterization of recessive Parkinson's disease in a large multicenter study

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Abstract

Studies of the phenotype and population distribution of rare genetic forms of parkinsonism are required, now that gene-targeting approaches for Parkinson's disease have reached the clinical trial stage. We evaluated the frequencies of *PRKN*, *PINK1*, and *DJ-1* mutations in a cohort of 1587 cases. Mutations were found in 14.1% of patients: 27.6% were familial and 8% were isolated. *PRKN* was the gene most frequently mutated in Caucasians whereas *PINK1* mutations predominated in Arab-Berber individuals. Patients with *PRKN* mutations had an earlier age at onset, and less asymmetry, levodopa-induced motor complications, dysautonomia, and dementia than those without mutations.

Keywords: Parkinson's disease; *PRKN*; *PINK1*; autosomal recessive inheritance; genotype-phenotype correlations

Abbreviations: AAO, age-at-onset; AR, autosomal recessive; EO, early-onset; GLM, generalised linear model; PD, Parkinson's disease; LRRK2, leucine-rich repeat kinase 2; PINK1, PTEN-induced putative kinase 1

Our understanding of the genetic basis of Parkinson's disease (PD) has been improved with the identification of several disease-causing genes. Trials targeting these genes are underway and the development of cohorts ready for precision clinical trials that target genetic forms of PD are now required. 2,3

PRKN, *PINK1*, and *DJ-1* mutations are the most frequent cause of early-onset (EO) autosomal recessive (AR) typical PD. We investigated the frequency and nature of pathogenic variants of these three genes in a cohort of 1587 PD probands, comparing clinical characteristics between patients with *PRKN* mutations (*PRKN*-PD), and those without pathogenic variants of known PD genes (PD-NM).

Patients and Methods

Patient selection

Patients were enrolled between 1990 and 2018, through the French PDG network and North African and Turkish collaborations. PD was diagnosed according to the clinical diagnostic criteria of the UK Parkinson Disease Society Brain Bank (PDSBB).⁴ Cases with mutations responsible for recessive atypical parkinsonism or those carrying the common *LRRK2* Gly2019Ser variant were not included. We selected a cohort of 1587 probands from 497 AR PD families (at least two affected siblings and isolated cases born to consanguineous parents), and 1090 isolated cases, all screened for *PRKN*.

Standardized neurological examinations were performed by movement disorder experts, and 28 variables were used to obtain comparable data.

Most probands were Caucasian (n=1324, 83.4%; 927 French, 134 Turkish), Arab-Berber (n=213, 13.4%) or of other ethnicities (n=50, 3.2%). We included 1587 PD probands and 52 mutation-carrying relatives in the genotype/phenotype correlation analysis. Informed consent and approval from institutional review boards were obtained for sample collection.

Procedures

Probands were screened by denaturing high-performance liquid chromatography (dHPLC) and/or direct sequencing, ⁵⁻⁸ next-generation sequencing (NGS) with a targeted gene panel or whole-exome sequencing ^{9,10}.

PRKN was screened in 1587 probands, *PINK1* and *DJ-1* in 1223. Sanger sequencing was performed to confirm variants and cosegregation analyses, where possible. Exon rearrangements were detected by semi-quantitative PCR for $PRKN^5$ or with the MRC Holland Salsa MLPA P051/P052 Parkinson kits. Patients with an AAO \leq 40 years and lymphoblastoid cells available for RT-PCR analysis $(n=15)^5$ or unaffected relatives for cosegregation analysis (n=30) were investigated for possible PRKN rearrangements undetectable by MLPA.

Statistical analysis

The PD-NM and *PRKN*-PD groups were compared with Welch's *t*-tests for continuous variables and Fisher's exact tests for categorical variables.

We used generalized linear models (GLMs) to compare clinical features between *PRKN*-PD with bi-allelic or double heterozygous mutations and PD-NM, adjusting for sex, age-at-onset

(AAO), disease duration and dopaminergic medication. We used GLMs with identity links and normal distributions for continuous clinical features, and GLMs with logit links and Bernoulli distributions for binary clinical features. Interactions between AAO and disease duration were also included. Disease duration and dopaminergic medication were not included in models for clinical features at onset. Effects were assessed in Fisher type II tests, and effect size was estimated with Cohen's f2. Benjamini-Hochberg correction for multiple testing was performed. GLMs were generated for the 15/28 clinical features for at least 20% of the patients in each group with available data.

Results

Demographic and clinical data

In our cohort, men (n=951, 60%) and EO cases (mean AAO 40.2 [SD 12.1] years) were overrepresented, particularly among isolated cases (p<0.0001) (Table 1).

Distribution and nature of recessive PD-associated gene mutations

Bi-allelic or double heterozygous mutations of known AR PD-causing genes were present in 224 of the 1587 probands (14.1%): 27.6% (137/497) of familial and 8% (87/1090) of isolated cases. The most frequently mutated genes were PRKN (199/1587, 12.5%), then PINK1 (23/1223, 1.9%), and DJ-1 (2/1223, 0.16%). We identified 56 patients with single heterozygous variants in the three genes in whom AAO was significantly later than cases with bi-allelic or double heterozygous mutations (36.9 [SD 10.6] vs. 31.3 [SD 11.1], p=0.0009); they were removed from genotype/phenotype correlation analyses.

The 199 cases carried either homozygous (*n*=92), or compound heterozygous (*n*=59), confirmed by segregation analysis of all available unaffected relatives or double heterozygous (*n*=48) *PRKN* mutations for whom phasing was still unknown. These carriers displayed 77 different variants, including 19 absent from public databases (<u>www.mdsgene.org</u>)¹¹ (Fig 1A). RT-PCR identified a single French family with *PRKN* Ex3del/Ex3dup compound heterozygous rearrangements not detectable by MLPA but elucidated by co-segregation analysis.

Most of the 23 probands with pathogenic *PINK1* variants were Arab-Berbers (*n*=12, 52.2%); 8 carried the homozygous Gln456* mutation. Most variants were homozygous (*n*=18, 78.3%). We identified 21 different pathogenic variants, including three not in databases (Fig 1B).

Distribution of *PRKN* and *PINK1* mutation carriers by age-at-onset, pattern of presentation and ethnicity

The proportion of probands with PRKN mutations decreased with increasing AAO: 42.2% for an AAO \leq 20 years, 29% (21 to 30 years), 13% (31 to 40 years), and 4.4% (41 to 60 years) (Fig 2A). This decrease was more marked in isolated than familial cases (Fig 2B). PINKI mutations were less frequent than PRKN mutations (1.9% vs. 12.5%) but more evenly distributed among familial cases for AAOs up to 60 years. Neither PRKN nor PINKI variants were found in patients with onset after 60 (Fig 2A-D). PRKN mutations were more frequent in Caucasians (179/1324, 13.5%) than in Arab-Berbers (16/213, 7.5%) (Fig 2E). Conversely, PINKI mutations were more common in Arab-Berbers (12/188, 6.4%) than Caucasians (9/1005, 0.9%) (Fig 2E).

Comparison of *PRKN* mutation carriers (*PRKN*-PD) with non-mutation carriers (PD-NM): genotype-phenotype correlations

We compared the clinical features between the 228/241 *PRKN*-PD and 1181/1307 PD-NM subjects without missing data (Table 2; Supplementary Table 1). The proportion of men $(p_{adj}=0.016)$ and AAO $(p_{adj}<0.0001)$ were greater in the PD-NM than the *PRKN*-PD group. Dopaminergic treatment was similar between groups $(p_{adj}=0.69)$, but levodopa responsiveness was higher in *PRKN*-PD than PD-NM $(p_{adj}=0.045)$.

After adjustment for covariables, PRKN-PD had a higher initial frequency of tremor $(p_{adj}=0.0076)$, but lower frequencies of akinesia $(p_{adj}=0.0003)$, micrographia $(p_{adj}=0.010)$, and asymmetry $(p_{adj}=0.0005)$ than PD-NM patients (Table 2). Dystonia at onset and cardinal symptom (bradykinesia, rest tremor and rigidity) frequencies were similar in both groups. Motor severity after adjustment for disease duration was lower in PRKN-PD than in PD-NM (UPDRS score, $p_{adj}=0.014$), and PRKN-PD patients developed fewer levodopa-induced motor complications (dyskinesia: $p_{adj}=0.0005$; motor fluctuations: $p_{adj}<0.0001$). Non-motor symptoms, including dysautonomia $(p_{adj}<0.0001)$ and dementia $(p_{adj}=0.014)$, were less frequent in PRKN-PD patients.

Mutational and phenotypic characteristics of PINK1 and DJ-1 mutation carriers

The *PINK1*-associated phenotype of the 33 carriers resembled that of *PRKN*, but with a slightly later AAO (mean 34.6 [SD 12.2] years vs. 31.3 [SD 10.9] years; p=0.20), a lower

frequency of tremor (16.7% vs. 68.5%; p=0.0005) and a higher rate of dystonia (90% vs. 18.2%; p<0.0001) at onset (Supplementary Table 2). Non-motor symptoms, such as dysautonomia (72.2% vs. 19.6%; p<0.0001) and dementia (25% vs. 3.8%; p=0.0007) were more frequent in patients with PINKI variants, who were also more likely to display levodopa-induced dyskinesia (93.3% vs. 54.1%; p=0.0024) and motor fluctuations (66.7% vs. 46.2%; p=0.06).

The two pathogenic *DJ-1* variant carriers (one Caucasian, one North African) each carried a previously unknown homozygous Glu94* and a compound heterozygous variant affecting the same highly conserved amino acid (Thr154Ile/Thr154Ala). They developed PD, with the four cardinal signs, at the ages of 29 and 28. Dystonia at onset, dyskinesia and orthostatic hypotension were noted in one patient, without cognitive signs.

Discussion

We established the spectrum and relative frequencies of mutations in a large cohort of AR PD cases, elucidating the genotype-clinical phenotype relationship. Homozygous/compound or double heterozygous pathogenic variants of *PRKN*, *PINK1*, and *DJ-1* account for 14.1% of our PD patients, with *PRKN* the most frequently mutated. This study included a large number of genotyped and extensively phenotyped patients (276 mutation carriers) compared with a group of cases not carrying mutations of known AR PD-causing genes. However, the clinical data were cross-sectional, the numbers of patients with mutations of genes other than *PRKN* were small, and our populations were biased towards EO cases. In addition, 24% (48/199) of

our PD patients with two *PRKN* mutations lacked information on their mutational phasing. However, given that we found no mutations *in cis* in a co-segregation analysis of 59 index cases, we are confident that the vast majority of patients with two mutations carry them *in trans*. Nevertheless, our findings may have major implications for patient selection for genetic testing based on AAO, pattern of disease presentation, and ethnicity. Indeed, the frequency of pathogenic variants of AR PD-associated genes i) decreased with increasing AAO, to zero for an AAO beyond 60 years, ii) in cases with a positive family history or consanguinity was more than triple that in isolated cases, and iii) was much higher for *PRKN* than *PINK1* in Caucasians, but similar for these two genes in Arab-Berbers. However, pathogenic *PRKN* variants were more frequent in our EO PD cases (<50 years, 190/1273, 14.9%) than in four other cohorts (10.1% in a Taiwanese cohort¹² and 2.8% in a Norwegian cohort¹³, both with EO defined as <45 years; 5.9% in a UK series¹⁴ and 2.8% in a larger multicenter sample¹⁵, both with EO <50 years). A meta-analysis of more than 5800 PD patients, found *PRKN* variants in 8.6% of PD cases with an AAO <50.¹⁵ We provide more precise data for *PRKN* and *PINK1* genes, according to AAO.

GLM analyses revealed that AAO was lower, disease progression slower and the response to levodopa stronger in patients with *PRKN* mutations than in those without pathogenic variants, as previously reported.^{12,15,16,17} However, these mutations were not associated with higher rates of dystonia at onset, a trait more strongly associated with EO (see Supplementary Table 1) than with genetic status. Patients with *PRKN* mutations also had a distinctive non-motor symptom profile, with lower frequencies of dementia and dysautonomia, consistent with previous reports.¹⁸ After adjustment for disease duration and dopaminergic medication, these

patients had fewer treatment-induced complications, such as dyskinesia and motor fluctuations, than PD-NM patients. This very pure and slowly progressive phenotype makes patients with *PRKN* variants, very good candidates for deep-brain stimulation (DBS). ^{19,20}

The causal gene(s) remained unidentified for a number of families with AR PD (~72.5%), suggesting that pathogenic variants of known genes may have been missed or the involvement of unknown genes.

These findings will help to guide routine genetic testing and to establish cohorts of patients for clinical trials targeting the gene defects or their physio-pathological consequences.

Supplementary Materials

This file includes:

Supplementary Table 1

Supplementary Table 2

Supplementary Table 3

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Author Contributions

SL, J-CC, and AB contributed to the conception and design of the study. All authors contributed to the acquisition and analysis of data. SL, AL, MH, CT, ER, J-CC, AB contributed to the drafting of the text and figure preparation.

Potential Conflicts of Interest

References

- 1 Lunati A, Lesage S, Brice A. The genetic landscape of Parkinson's disease. Rev Neurol (Paris) 2018;174:628-643.
- 2 Strafella C, Caputo V, Galota MR, et al. Application of Precision Medicine in Neurodegenerative Diseases. Front Neurol 2018;9:701.
- Vollstedt EJ, Kasten M, Klein C; MJFF Global Genetic Parkinson's Disease Study Group. Using global team science to identify genetic parkinson's disease worldwide. Ann Neurol 2019;86:153-157.
- 4 Hughes AJ, Daniel SE, Kilford L, Lees AJ. Accuracy of clinical diagnosis of idiopathic Parkinson's disease: a clinico-pathological study of 100 cases. J Neurol Neurosurg Psychiatry 1992;55:181-184.
- 5 Lesage S, Magali P, Lohmann E, et al. Deletion of the parkin and PACRG gene promoter in early-onset parkinsonism. Hum Mutat 2007;28:27-32.
- 6 Ibáñez P, Lesage S, Lohmann E, et al. Mutational analysis of the PINK1 gene in early-onset parkinsonism in Europe and North Africa. Brain 2006;129:686-694.

- 7 Leutenegger AL, Salih MA, Ibáñez P, et al. Juvenile-onset Parkinsonism as a result of the first mutation in the adenosine triphosphate orientation domain of PINK1. Arch Neurol 2006;63:1257-1261.
- 8 Lohmann E, Dursun B, Lesage S, et al. Genetic bases and phenotypes of autosomal recessive Parkinson disease in a Turkish population. Eur J Neurol 2012;19:769-775.
- 9 Bouhouche A, Tesson C, Regragui W, et al. Mutation analysis of consanguineous Moroccan patients with Parkinson's disease combining microarray and gene panel. Front Neurol 2017;8:567.
- 10 Lesage S, Drouet V, Majounie E, et al. Loss of VPS13C function in autosomal-recessive Parkinsonism causes mitochondrial dysfunction and increases PINK1/Parkin-dependent mitophagy. Am J Hum Genet 2016;98:500-513.
- 11 Kasten M, Hartmann C, Hampf J, et al. Genotype-phenotype relations for the Parkinson's disease genes Parkin, PINK1, DJ1: MDSGene Systematic. Mov Disord 2018;33:730-741.
- 12 Lin CH, Chen PL, Tai CH, et al. A clinical and genetic study of early-onset and familial parkinsonism in Taiwan: An integrated approach combining gene dosage analysis and next-generation sequencing. Mov Disord 2019;34:506-515.
- 13 Gustavsson EK, Trinh J, McKenzie M, et al. Genetic identification in early onset Parkinsonism among Norwegian Patients. Mov Disord Clin Pract 2017;4:499-508.
- 14 Kilarski LL, Pearson JP, Newsway V, et al. Systematic review and UK-based study of PARK2 (parkin), PINK1, PARK7 (DJ-1) and LRRK2 in early-onset Parkinson's disease. Mov Disord 2012;27:1522-1529.

- 15 Marder KS, Tang MX, Mejia-Santana H, et al. Predictors of parkin pathogenic variants in early-onset Parkinson disease: the consortium on risk for early-onset Parkinson disease study. Arch Neurol 2010;67:731-738.
- 16 Lohmann E, Periquet M, Bonifati V, et al. How much phenotypic variation can be attributed to parkin genotype? Ann Neurol 2003;54:176-185.
- 17 Khan NL, Graham E, Critchley P, et al. Parkin disease: a phenotypic study of a large case series. Brain 2003;126:1279-1292.
- 18 Alcalay RN, Caccappolo E, Mejia-Santana H, et al. Cognitive and motor function in long-duration PARKIN-associated Parkinson disease. JAMA Neurol 2014;71:62-67.
- 19 Pal GD, Hall D, Ouyang B, et al. Genetic and clinical predictors of deep brain stimulation in young-onset Parkinson's disease. Mov Disord Clin Pract 2016;3:465-471.
- 20 Rizzone MG, Martone T, Balestrino R, Lopiano L. Genetic background and outcome of deep brain stimulation in Parkinson's disease. Parkinsonism Relat Disord 2018;64:8-19.

Figure Legends

FIGURE 1: Schematic representation of the (A) *PRKN* and (B) *PINK1* genes and respective proteins and associated disease-linked mutations.

Exonic deletions (in red), duplications (in green), or triplications (in blue) are shown in the upper panel and point mutations (missense, frameshift, nonsense, and splice) are shown in the lower panel. Newly identified mutations are shown in bold. Numbers in brackets indicate the number of mutation carriers. *PRKN* cDNA numbering: NM_004562.2; *PINK1* cDNA numbering: NM_032409.2.

Parkin protein: UBL, ubiquitin-like; RING, really interesting new gene; IBR, in-between RING; PINK1 protein: MTS, mitochondrial targeting sequence; TM, transmembrane helix.

FIGURE 2: Distribution of *PRKN* and *PINK1* mutation carriers by age-at-onset, pattern of disease presentation and ethnicity. (A) Proportion of probands, by age-at-onset, for *PRKN* (199 carriers among 1587 probands). (B) Proportion of *PRKN* mutation carriers from 497 cases with autosomal recessive PD (in blue) *vs.* 1090 isolated cases (in orange) by age-at-onset and pattern of presentation of PD. (C) Proportion of probands, by age-at-onset for *PINK1* (23 carriers among 1223 probands). (D) Proportion of *PINK1* mutation carriers from 386 cases with autosomal recessive PD (in blue) *vs.* 837 isolated cases (in orange) by age-at-onset and pattern of presentation of PD. (E) Proportion of *PRKN* and *PINK1* mutation carriers, according to their ethnicity: Caucasians (*n*=1324, in blue) or Arab-Berbers (*n*=213, in orange).

TABLE 1. Demographic data for our study population

	Whole PD group	n	AR PD, including isolated cases with consanguinity	n	Isolated cases	n	AR PD vs. isolated case p value
% men (n)	60.0 (951)	1587	57.1 (284)	497	61.2 (667)	1090	0.14
Eth nic background		1587		497		1090	<0.0001*
% Caucasian (n)	83.4 (1324)		75.5 (375)		87.1 (949)		
% Arab-Berbers (n)	13.4 (213)		22.1 (110)		9.4 (103)		
% others/mixed (n)	3.2 (50)		2.4 (12)		3.5 (38)		
% Consanguinity (n)	12.9 (202)	1564	40.9 (202)	494	0	1070	NA
Ag -at-onset (SD), y	40.2 (12.1)	1543	43.8 (15.0)	485	38.6 (10.1)	1058	<0.0001*
nge	2-81		3-81		2-74		
Age at examination (SD), y	49.7 (13.2)	1575	54.2 (14.5)	495	47.7 (12.0)	1080	<0.0001*
ge	9-87		16-87		9-79		
,							
ease duration (SD), y	9.8 (8.6)	1530	10.9 (9.6)	482	9.2 (8.0)	1048	0.0005*
Range	0-63		0-63		0-48		

Frequencies were compared in Fisher's exact tests for qualitative traits and means were compared in t-tests for continuous variables.

NA, Not appropriate.

^{*}p < 0.05

TABLE 2. Demographic and clinical characteristics of patients with Parkinson's disease with pathogenic *PRKN* variants (*PRKN*-PD) and of patients without pathogenic variants (PD-NM)

	PD-NM and PRKN-PD unadjusted comparisons				PD-NM and PRKN-PD adjusted comparisons			
Characteristics	PD-NM n=1181	<i>PRKN</i> -PD <i>n</i> =228	p value	p value adjusted ¥	Coefficient or odds ratio (OR) (SE)	Cohen's f2	p value	p value adjusted \{
Baseline								
Sex (% male)	727/1181	118/228	0.0063*	0.016*				
	(61.6%)	(51.8%)						
Age at	50.5 (13.2)	45.4 (12.9)	<0.0001*	<0.0001*				
examination								
(SD), y								
Disease duration	8.8 (7.8)	14.1 (10.4)	<0.0001*	<0.0001*				
(SD), y								
Age-at-onset	41.6 (12.0)	31.2 (10.7)	<0.0001*	<0.0001*				
(SD), y								
L-DOPA-treated	791/1181	148/228	0.54	0.69				
	(67%)	(64.9%)						
Levodopa	659/738	142/149	0.022*	0.045*	1.9 (0.82)	0.007	0.11	0.15
responsiveness#	(89.3%)	(95.3%)						
Motor symptoms and	l signs							
Dystonia at onset	164/990	37/201	0.54	0.69	0.84 (0.18)	0.001	0.42	0.46
J	(16.6%)	(18.4%)			` ,			
Akinesia at onset	590/946	99/206	0.0002*	0.0010*	0.51 (0.09)	0.015	0.0001*	0.0003*
	(61.3%)	(48.1%)			, , ,			
Tremor at onset	570/960	142/205	0.0091*	0.021*	1.7 (0.29)	0.007	0.0030*	0.0076*
	(59.4%)	(69.3%)						
Micrographia at	308/927	42/204	0.0003*	0.0013*	0.58 (0.11)	0.007	0.0047*	0.010*
onset	(33.2%)	(20.6%)						
Asymmetry	1001/1035	181/198	0.0016*	0.0049*	0.26 (0.09)	0.010	0.0002*	0.0005*
	(96.7%)	(91.4%)						
Bradykinesia	1033/1069	201/208	1.0000	1.0000	1.4 (0.63)	0.002	0.43	0.46
	(96.6%)	(96.6%)						
Rigidity	1008/1066	194/204	0.87	0.90	1.1 (0.42)	< 0.001	0.77	0.77
_	(94.6%)	(95.1%)	0.0044		4.7.40.00		0.0404	
Tremor	796/1056	168/205	0.031*	0.057	1.5 (0.32)	0.002	0.048*	0.072
LIDDDG D . III	(75.4%)	(82%)	0.0017*	0.0040*	2.2 (1.2)	0.000	0.0001*	0.01.4%
UPDRS-Part III	19.6 (13.8)	15.9 (11.9)	0.0017*	0.0049*	-3.3 (1.2)	0.008	0.0081*	0.014*
ON (SD)	2 (0.01)	2.00 (0.93)	0.61	0.74	0.14 (0.00)	0.003	0.13	0.16
Hoehn&Yahr ON (SD)	2 (0.91)	2.00 (0.93)	0.61	0.74	-0.14 (0.09)	0.003	0.13	0.10
Dyskinesia	457/667	97/178	0.0007*	0.0025*	0.44 (0.09)	0.020	0.0001*	0.0005*
Dyskillesia	(68.5%)	(54.5%)	0.0007	0.0023	0.44 (0.09)	0.020	0.0001	0.0003
Motor	485/663	82/177	<0.0001*	<0.0001*	0.32 (0.07)	0.041	<0.0001*	<0.0001*
fluctuations	(73.2%)	(46.3%)	10.0001	\0.0001	0.32 (0.07)	0.041	10.0001	10.0001
Non-motor symptom		(10.5 /0)						
	Ü	26402	.0.00011	.0.0001#	0.10 (0.07)	0.00-	. 0 00011	.0.0000
Dysautonomia	254/470	36/192	<0.0001*	<0.0001*	0.19 (0.05)	0.095	< 0.0001*	<0.0001*
	(54.0%)	(18.8%)	0.015	0.025	0.04 (0.16)	0.000	0.000=:	0.01.11
Dementia	67/667	6/153 (3.9%)	0.017*	0.037*	0.34 (0.16)	0.009	0.0087*	0.014*
	(10.0%)							

Data are expressed as mean (standard deviation) for continuous variables, and as counts (percentages) for categorical variables. We used *t*-tests to compare the two groups for continuous variables and Fisher's exact tests for binary variables.

Coefficients for continuous clinical features and odds ratios (ORs) for binary clinical features and standard error (SE), Cohen's f2 and p-values were calculated from GLMs with mutation status, sex, age-at-onset, disease duration, L-DOPA group and age-at-onset vs. disease duration for all 15 variables except for onset variables for which only mutation status, sex and age-at-onset were added. Linear models were used for continuous variables; GLMs with logit links and Bernouilli distributions were used for binary variables

#Levodopa responsiveness was defined as a >30% improvement in subjective perceived motor symptoms

Y p corrected for multiple testing by the Benjamini-Hochberg procedure

*p < 0.05



