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Characterization of the myometrial transcriptome and biological pathways of spontaneous human labor at term

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Abstract

Aims—To characterize the transcriptome of human myometrium during spontaneous labor at term.

Methods—Myometrium was obtained from women with (n=19) and without labor (n=20). Illumina[®] HumanHT-12 microarrays were utilized. Moderated t-tests and False Discovery Rate adjustment of p-values were applied. qRT-PCR was performed for a select set of differentially expressed genes in a separate set of samples. ELISA and Western Blot were utilized to confirm differential protein production in a third sample set.

Results—1) 471 genes were differentially expressed; 2) Gene Ontology analysis indicated enrichment of 103 biological processes and 18 molecular functions including: a) inflammatory response; b) cytokine activity; and c) chemokine activity; 3) systems biology pathway analysis using Signaling Pathway Impact Analysis indicated 6 significant pathways: a) cytokine-cytokine receptor interaction; b) Jak-Stat signaling; and c) complement and coagulation cascades; d) NOD-like receptor signaling pathway; e) Systemic Lupus Erythematosus; and f) Chemokine signaling pathway; 3) qRT-PCR confirmed over-expression of prostaglandin-endoperoxide synthase-2 (PTGS2/COX2), heparin binding EGF-like growth factor (HBEGF), chemokine C-C motif ligand 2 (CCL2/MCP1), leukocyte immunoglobulin-like receptor, subfamily A member 5 (LILRA5/

LIR9), IL-8, IL-6, chemokine C-X-C motif ligand 6 (CXCL6/GCP2), nuclear factor of kappa light chain gene enhancer in B-cells inhibitor zeta (NFKBIZ), suppressor of cytokine signaling 3 (SOCS3) and decreased expression of FK506 binding-protein 5 (FKBP5) and aldehyde dehydrogenase (ALDH2) in labor; 4) IL-6, CXCL6, CCL2 and SOCS3 protein expression was significantly higher in the term labor group compared to the term not in labor group.

Conclusions—Myometrium of women in spontaneous labor at term is characterized by a stereotypic gene expression pattern consistent with over-expression of the inflammatory response and leukocyte chemotaxis. Differential gene expression identified with microarray was confirmed with qRT-PCR using an independent set of samples. This study represents an unbiased description of the biological processes involved in spontaneous labor at term based on transcriptomics.

Keywords

inflammation; microarray; pregnancy; parturition; systems biology; aldehyde dehydrogenase; ALDH2; CCL2/MCP-1; CXCL6/GCP2; FK506 binding-protein 5; FKBP5; heparin binding EGF-like growth factor; HBEGF; IL-6; IL-8; leukocyte immunoglobulin-like receptor; subfamily A member 5; LILRA5/LIR9; nuclear factor of kappa light chain gene enhancer in B-cells inhibitor zeta; NFKBIZ; PTGS2/COX2; suppressor of cytokine signaling 3; SOCS3; progesterone; inflammasome

Introduction

Parturition is a complex process involving myometrial activation, cervical ripening, and membrane/decidual activation (the common pathway of labor) [2,19,30,34–37,41,114,149,169–171,173,210,212,217,220,236,243,247,298,313,327,333]. Evidence suggests that in preparation for labor, the myometrium attains an increasingly contractile phenotype, [27,29,53,55,59,62,66,68,100,103,106,108,110,112,113,122–124,146,174,177,179,181,183,188,190,196,216,225,280–282,296,310,315,321] while the cervix undergoes preparatory changes including cervical ripening and dilatation [61,68,112,113,132,134,143,147,151,176,184,185,218,219,225,272,275,276,282,296,297,311,312,321,330,337]. Labor disorders such as preterm labor and abnormal parturition at term represent abnormalities of one or more components of the common pathway of parturition and are associated with increased morbidity and mortality [101,102,117,118,121,148]. The application of high-dimensional biology techniques holds promise to assist in the understanding of parturition (term and preterm). While the process of labor and delivery is vital to the survival of most viviparous species, its physiology and pathology in humans remains to be elucidated.

High-dimensional biology techniques (genomics, transcriptomics, proteomics, etc.) provide the means by which comprehensive and unbiased insight into physiologic events, such as parturition, can be established [142,158,163,245,253,263,304]. Previous investigators have discovered differential gene and/or protein expression in the chorioamniotic membranes, [128] amniotic fluid, [23,24,43,50,125,197,222,224,248,270,322] umbilical cord blood, [182] uterine cervix, [132–135,143,205,234,324] and human myometrium [1,13,25,39,90,91,136,214] in preterm and term labor. However, the biological processes, molecular functions, and pathways associated with spontaneous term parturition have not been described and the regulatory mechanisms remain poorly understood. We undertook this study in order to characterize the transcriptome of human myometrium during normal labor at term to gain understanding of global changes in gene expression using an unbiased approach.

Materials and Methods

A prospective study was performed in which myometrium was obtained from women undergoing cesarean section at term (≥ 37 weeks) in the following groups: 1) not in labor ($n=20$); and 2) spontaneous labor ($n=19$). (please see supplementary material for details)

Eligible patients were enrolled at Hutzel Women's Hospital (Detroit, MI, USA). All women provided written informed consent prior to the collection of myometrial samples. The collection and utilization of the samples for research purposes was approved by the Institutional Review Board of the *Eunice Kennedy Shriver* National Institute of Child Health and Human Development (NICHD/NIH/DHHS, Bethesda, Maryland), and the Human Investigation Committee of Wayne State University (Detroit, MI, USA).

Sample collection

Myometrial tissue samples were collected from the hysterotomy site in the lower uterine segment during cesarean section following delivery of the placenta from the midpoint of the superior aspect of the uterine incision using Metzenbaum scissors and measured approximately 1.0cm^3 . Tissues were snap-frozen in liquid nitrogen, and were kept at -80°C until use.

RNA isolation

Total RNA was isolated from snap-frozen myometrium using TRI Reagent[®] combined with the Qiagen RNeasy Lipid Tissue kit protocol (Qiagen, Valencia, CA, USA) according to the manufacturers' recommendation. The RNA concentrations and the A260nm/A280nm ratio were assessed using a NanoDrop 1000 (Thermo Scientific, Wilmington, DE, USA). RNA integrity numbers were determined using the Bioanalyzer 2100 (Agilent Technologies, Wilmington, DE, USA).

Microarray analysis and Real-time quantitative reverse transcriptase-polymerase chain reaction (qRT-PCR)

The Illumina[®] HumanHT-12 v3 expression microarray platform (Illumina, San Diego, CA, USA) was used to assess the expression levels in each individual specimen following the manufacturer's instructions. qRT-PCR assays were performed for selected genes on an independent set of myometrium samples [term with ($n=10$) and without ($n=10$) labor] to determine whether the microarray results could be confirmed. The Biomark[™] System (Fluidigm, San Francisco, CA, USA) was utilized to perform high-throughput qRT-PCR confirmation. (please see supplementary material for details)

Enzyme-linked immunosorbent assay and immunoblot

A third independent set of myometrial samples were obtained from women at term with ($n=9$) and without labor ($n=11$). Myometrial concentrations of IL-6, CCL2, and IL-8 were determined with specific enzyme-linked immunoassays (R&D Systems, Inc, Minneapolis, MN) according to the manufacturer's instructions. Protein expression of SOCS3 and CXCL6 was analyzed using immunoblot (please see supplementary material for details).

Statistical analysis

Demographic and clinical characteristics of the study groups were compared using the Pearson's chi-square test and the Fisher's exact test for proportions and Kruskal-Wallis and the Mann-Whitney U test for continuous variables. SPSS v.12 (SPSS Inc, Chicago, IL) was used. A p -value < 0.05 (2-tailed) was considered significant.

Microarrays, qRT-PCR, and ELISA

A moderated t-test was applied to test differential expression, and a false discovery rate (FDR) adjustment of the p-value was performed to correct for multiple testing. Genes with an FDR <0.05 and a fold change >1.5 were considered significant. Gene ontology analysis was performed using an over-representation approach implemented in the Onto-Express [156,157] site and GOstats [95] software packages. Pathway analysis was performed on the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway database using an enrichment analysis as well as the Signaling Pathway Impact Analysis (SPIA) [69,303]. SPIA is based on a systems biology approach [69] and takes into account gene-gene signaling interactions as well as the magnitude and direction of gene expression changes to determine significantly impacted pathways. The MetaCore database (Gene Go, Inc, St. Joseph, MI) was also mined. (please see supplementary material for details)

Analysis for qRT-PCR was performed using an equal variance two-sample one-tailed t-test. Protein concentrations determined by ELISA were analyzed using one-tailed Mann-Whitney U tests. One-tailed tests were used because there was a prior hypothesis about the direction of change based upon microarray results.

Results

Table 1 displays the demographic and clinical characteristics of each study group. There were no differences among the groups.

Microarray Results

Discriminant analysis demonstrated significant changes in the transcriptome of human myometrium between women with and without labor. In total, 538 probes corresponding to 471 unique genes were differentially expressed. The top 50 differentially expressed probes overexpressed and underexpressed in spontaneous term labor are listed in Tables 2 and 3, respectively. The results of microarray profiling are depicted in Figure 1. The volcano plot (Figure 1A) shows the magnitude and significance of differential gene expression. Principal component analysis [302] (Figure 1B) was performed to reduce the dimensionality of the microarray data from that of all genes tested to three dimensions. The heat map in Figure 1C uses a color scale to show the consistency of the expression levels within each group of samples as well as the differences between the groups that led to positive test results.

In order to gain further insight into the biology of the differential gene expression, Gene Ontology enrichment analysis was employed. A total of 103 biological processes were associated with spontaneous term labor. The most significantly enriched biological processes included: 1) inflammatory response; 2) response to wounding; and 3) response to external stimulus. Eighteen molecular functions were enriched in spontaneous term labor including: 1) cytokine activity; 2) heparin binding; 3) receptor binding; 4) chemokine activity; and 5) chemokine receptor binding (Table 4).

Pathway Analysis

Pathway analysis on the KEGG database indicated significant enrichment of 4 pathways: 1) cytokine-cytokine receptor interaction; 2) Jak-STAT signaling pathway; 3) complement and coagulation cascade; and 4) ascorbate and aldarate metabolism. Based on over-representation analysis, 22 pathways in the MetaCore database were significant including interleukin-17 signaling. Of interest, 19 of 22 (86%) enriched pathways were involved in the biology of inflammation. The MetaCore map of differentially expressed genes in the interleukin-17 signaling pathway is depicted in Figure 2. Although gene regulation network models may be derived from experimental data spanning multiple systems and not be

completely applicable to a single system, an entire network module in the lower right quadrant of Figure 2, containing I- κ B, NF κ B, and several NF κ B targets, is upregulated in spontaneous term labor. This result, for the first time, presents a completely experimentally validated gene sub-network from myometrial transcriptome data.

While four significant pathways were identified by simple pathway enrichment methods, the systems biology-based pathway analysis implemented in SPIA yielded a total of six significant pathways: 1) cytokine-cytokine receptor interaction ($p < 0.001$); 2) complement and coagulation cascade ($p < 0.001$); 3) Jak-Stat signaling ($p < 0.001$); 4) NOD-like receptor signaling pathway ($p = 0.001$); 5) Systemic lupus erythematosus ($p = 0.01$); and 6) chemokine signaling pathway ($p = 0.01$). The inclusion of systemic lupus was attributed to differential expression of the complement C1q complex.

Given the significant enrichment of biological processes, molecular functions, and pathways centrally involved in the inflammatory response, we verified differential regulation of genes which are key components of the immune response as well as other genes of interest.

qRT-PCR Results

Thirty-one genes were tested for verification of microarray results. Fourteen genes were previously reported in the literature as being related to labor (Table 5A), while 17 genes were selected from among the microarray results (Table 5B).

qRT-PCR confirmed differential expression of 11 genes between the term labor and term not in labor groups. In addition to genes previously described as upregulated in spontaneous term labor [interleukin (IL)-8, IL-6, prostaglandin-endoperoxide synthase 2 (PTGS2/COX2), and CCL2], differential expression of a set of genes not previously described was confirmed. Novel genes with confirmed over-expression during spontaneous term labor included: heparin-binding EGF-like growth factor (HBEGF), leukocyte immunoglobulin-like receptor, subfamily A, member 5 (LILRA5), chemokine (C-X-C motif) ligand 6 (CXCL6), nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta (NFKBIZ), and suppressor of cytokine signaling 3 (SOCS3). Novel downregulated genes included FK506 binding protein 5 (FKBP5) and aldehyde dehydrogenase 2 (ALDH2) (see Figure 3). Of note, 8 of the 11 genes participate in the innate immune response.

Overall, the direction of change in gene expression by PCR was consistent with the microarray analysis in 29 out of 30 genes (96.7%). The comparison of microarray and PCR data for each gene is described in Table 6.

ELISA and Immunoblotting

Consistent with the microarray and qRT-PCR data, the median protein concentrations of CCL2 and IL-6 were higher in term labor compared to term not in labor myometrium (Figure 4). While the median concentration of IL-8 was higher in labor, this difference did not reach statistical significance (2.14 pg/mL interquartile range [IQR] 0–52.7 versus 21.97 pg/mL IQR 1.1–389.4; $p = 0.33$). Similar to the microarray and PCR results, the protein abundance of both CXCL6 and SOCS3 was higher in myometrium from women in labor compared to women not in labor as demonstrated by immunoblot (Figure 5).

Discussion

Principal findings of this study

1) The myometrial transcriptome of women in spontaneous labor at term was dramatically different from that of women not in labor; 471 genes were differentially expressed between

the groups; 2) Gene Ontology analysis indicated specific biological processes (e.g. inflammatory response, chemotaxis, and immune response) and molecular functions (e.g. cytokine activity, chemokine activity, chemokine receptor binding) associated with spontaneous term labor; 3) pathway analysis identified 6 pathways, all involved in inflammation, enriched in the myometrial transcriptome of labor; 4) a novel set of inflammation-related genes differentially expressed in human myometrium during labor was identified including LILRA5, CXCL6, NFKBIZ, and SOCS3 as well as additional genes not previously reported to be involved in term human parturition (FKBP5, HBEGF, and ALDH2); and 5) overall, the process of spontaneous term parturition is characterized by a molecular signature consistent with over-expression of genes involved in inflammation and leukocyte chemotaxis.

Inflammation: an integral component of labor at term in the myometrium

Inflammation has been implicated in key biological processes required for reproduction, including ovulation, implantation [63,97,203] and parturition [14,15,17,31,130,137,160,172,189,206,240–242,244,249,251,255–258,261,277,326]. Implantation of the blastocyst generates a pro-inflammatory response within the decidua (TH1-like polarization) [63,97,203]. After establishment of placentation, inflammation retreats, allowing a tolerogenic state required for the growth and development of the conceptus. This has been referred to as a TH2-like polarization present within the uterus and the maternal systemic circulation during most of pregnancy. Spontaneous parturition represents the re-emergence of this inflammatory state (TH1 polarization) [121,251]. Indeed, leukocyte infiltration of human myometrium has been described as a hallmark of the development of the uterus from a quiescent to a contractile organ [309]. Interestingly, gradients of leukocyte infiltration into the uterus and the uterine cervix [218] during labor have been reported. Thomson et al. [309] first described the region-specific leukocyte subpopulations in the fundal and lower uterine segments. The authors reported an increased density of neutrophils and macrophages in the myometrium of both regions during labor, while an increased density of T-cells was limited to the lower uterine segment. Overall, the inflammatory infiltrate was predominant in the lower uterine segment. However, the mechanisms responsible for leukocyte infiltration (chemokine signaling) and the biological processes induced by leukocytes in the uterus are still poorly elucidated.

Previous investigators have described the increased expression of chemotactic factors which may, in part, account for this accumulation of leukocytes in the uterus [259,264]. Interleukin-8 is a major chemokine that mediates neutrophil chemotaxis and activation. Increased expression of IL-8 has been reported in gestational tissues [218,241,305] and the cervix [132,219,272,275,276] in spontaneous labor. Our finding that IL-8 mRNA expression is dramatically overexpressed in human myometrium with spontaneous labor is consistent with these reports. These observations are consistent with our 1991 report demonstrating that the median amniotic fluid concentration of IL-8 was higher in women with spontaneous labor at term without infection/inflammation than in those not in labor at term [241]. It is noteworthy that IL-8 production by intrauterine tissues is regulated by progesterone [154]. Thus, a suspension of progesterone action as term approaches may be responsible for the increased production of IL-8, neutrophil infiltration and activation, and the subsequent inflammatory phenotype of the myometrium.

IL-6 mediates the acute phase response and functions as a myokine produced by contracting muscle [96]. Our results confirmed that there is a significant overexpression of IL-6 in myometrium during labor – this observation is consistent with the findings of Osman et al. [218] Furthermore, the current report provides evidence that IL-6 protein is elevated in the myometrium of women in labor at term, compared with women at term not in labor. These findings are also consistent with our report that IL-6 increases in the amniotic fluid of

women with spontaneous labor at term when compared to that of women at term not in labor [273]. Such observation (made in 1991) has subsequently been confirmed as the expression of IL-6 increases in gestational tissues [128,132,218,306,337]. This cytokine has become a valuable marker in the assessment of patients with preterm labor and preterm PROM, as well as in the definition of the fetal inflammatory response syndrome (which is associated with the onset of preterm labor) [118,121,239,250,251,260,265,266].

Some pro-inflammatory cytokines also promote uterine contractility by inducing the expression of PTGS2/COX2, the rate-limiting step of prostaglandin biosynthesis [5,10,67,71,94,98,126,131,153,231,232]. For example, we and others have demonstrated that IL-1 β , TNF α and IL-6 can increase prostaglandin production by amnion, decidua and myometrium [11,21,22,70,73,138–140,155,175,198,199,202,294,325]. PTGS2/COX2 is a rapidly inducible enzyme with reported increased expression in association with labor in myometrium [5,6,44,54,67,115,136,233,278,284,289,306] and the uterine cervix [26,67,132,186,230,297,313,330,331]. Our results are consistent with these findings.

We have previously reported that the concentration of CCL2 (also known as MCP-1) is increased in the amniotic fluid of women in spontaneous labor at term compared to those not in labor, including spontaneous preterm delivery [92,93]. Herein, we describe significant overexpression of CCL2, a monocyte chemoattractant also involved in macrophage activation, in spontaneous term parturition. Esplin et al.[91] described increased expression of CCL2 in human myometrium and the chorioamniotic membranes during labor using microarray analysis and Northern Blot analysis. Interestingly, a recent report demonstrated that progesterone attenuates the myometrial expression of CCL2 in pregnant rats, implicating CCL2 as a potential therapeutic target for the prevention of preterm labor [283]. In our analysis, we confirm increased CCL2 mRNA expression in the myometrium during labor and describe increased CCL2 protein concentrations in myometrium from women in spontaneous labor at term, providing further support for the role of this chemokine in the mechanisms of labor.

The use of systems biology to delineate the biological processes and pathways characterizing spontaneous term parturition

We report herein that 6 pathways are enriched in the myometrium during labor. Each of these pathways is central in the deployment of an inflammatory response: cytokine-cytokine receptor interaction, Jak-STAT signaling, the complement and coagulation pathway, NOD-like receptor signaling, systemic lupus erythematosus (specifically the complement C1q complex), and chemokine signaling. Interestingly, components of these pathways have been previously linked to both normal parturition [57,75,76,159,180,201,213,221,226–228,235,271,274,308] and the “Great Obstetrical Syndromes” [32,33,64,79–82,84,85,89,238,267,291,295,307,320], including preterm labor [28,58,74,77,83,86–88,119,120,164,165,211,223,246,252,254,290,292,293,316–319]. Indeed, our group has reported an association between activation of the NALP3 inflammasome and the common pathway of parturition [119]. Moreover, we have previously demonstrated that normal pregnancy is characterized by activation of the complement system [235]. The most significantly enriched biological process and molecular function in spontaneous term parturition were the “inflammatory response” and “cytokine activity”, respectively. Novel genes involved in these pathways with over-expression in myometrium during labor all have reported molecular functions in the initiation, maintenance, and regulation of physiologic inflammation including CXCL6, LILRA5, NFKBIZ, and SOCS3.

Chemokines play an integral role in both the innate and adaptive host response as well as immune homeostasis [299,300]. Induced by IL-1 β [332], CXCL6 shares high functional homology with IL-8. A potent neutrophil chemoattractant, over-expression of CXCL6 has

been reported in the chorioamniotic membranes during term labor [128]. However, unlike the closely related IL-8, amniotic fluid concentrations of CXCL6 do not significantly change with the onset of term labor, although CXCL6 concentrations in amniotic fluid are higher in women with preterm labor compared to those with preterm gestation who eventually deliver at term [200]. Herein, we report the novel findings that CXCL6 is expressed by human myometrium and CXCL6 mRNA and protein expression are significantly increased in human myometrium during spontaneous term labor in the absence of histologic chorioamnionitis. Indeed, the only gene with a more profound increase in gene expression was IL-8. Interestingly, CXCL6 has been described to interact synergistically with CCL2 to increase neutrophil infiltration 10-fold in gastrointestinal tumors [116]. The enhanced expression of CXCL6 might strongly contribute to the myometrial infiltration by leukocytes observed during labor.

Leukocyte immunoglobulin-like receptor subfamily A, member 5 (LILRA5; LIR9) is a member of the family of leukocyte immunoglobulin-like receptors. To date, the expression of LILRA5 in reproductive tissues has not been reported. First described in 2003 [18], this gene has been implicated in the early activation of the innate immune response. Cross-linking of LILRA5 molecules on the surface of monocytes induces a calcium flux resulting in secretion of the pro-inflammatory cytokines interleukin-1 β , IL-6 and TNF α . [18] Given that IL-1 β has been implicated in the initiation of myometrial contractions [7,60,218], the induction of IL-1 β secretion by LILRA5 suggests a role for this novel receptor in the onset of labor. While the precise role of LILRA5 in the process of human parturition remains to be elucidated, its increased expression in human myometrium during spontaneous labor is a new lead for the study of human parturition.

Another pivotal regulator of IL-6, NFKBIZ, was overexpressed in human myometrium during labor. NFKBIZ is a nuclear I κ B protein which maintains dual roles in the regulation of NF- κ B with both positive and negative effects [187,204]. The mechanisms determining the opposite actions are currently unknown, but are hypothesized to be cell-type specific [161,335]. Interestingly, recent studies in a knock-out mouse model have demonstrated that NFKBIZ expression is required for IL-6 production. Macrophages from NFKBIZ knock-out mice had a profoundly impaired ability to produce IL-6 in response to LPS, IL-1, and TLR ligands [334]. Moreover, suppression of NFKBIZ by small interfering RNA significantly decreased monocyte IL-6 production in response to LPS [279]. Given the role of NF- κ B in the regulation of immune responses, the involvement of this positive and negative regulator of inflammatory gene expression in human labor warrants further investigation.

Suppressor of cytokine signaling 3 (SOCS3) is a member of the family of cytokine signaling inhibitors which regulate cytokine signaling through the JAK/STAT pathway. In particular, SOCS3 has been described as the “central negative regulator” of macrophage IL-6 signaling [336]. In mice with a conditional SOCS3 gene deletion in macrophages, IL-6 hyper-responsiveness has been demonstrated [56]. We report SOCS3 over-expression in myometrium during labor. Contradictory data exist regarding the differential expression of SOCS3 in the chorioamniotic membranes as expression after term labor has been reported to be both under-[16] and over-expressed [128]. While inflammation characterizes term human parturition, the process would require close regulation to remain physiologic. Therefore, SOCS3 may play an integral role during normal labor to abrogate the potentially damaging effects of an uncontrolled inflammatory response.

Evidence for functional progesterone withdrawal in human parturition

Suspension of progesterone action is believed to be crucial for the initiation of parturition. In most animals, this is mediated by a decrease in circulating maternal progesterone concentrations [99,184,185]. Parturition in humans and other primates, however, occurs

without a systemic progesterone withdrawal in maternal serum [38,288,323]. Nonetheless, a role for progesterone withdrawal as a trigger for parturition is supported by the fact that administration of nuclear progesterone receptor (nPR) antagonists (e.g., RU486) augments myometrial contractility and excitability and initiates labor at all stages of pregnancy [8,46,47,72,104,109,111,129,301]. Interruption of progesterone/nPR signaling is sufficient to initiate the full cascade of parturition and therefore, it is generally considered that human parturition involves a functional rather than a systemic progesterone withdrawal whereby myometrial cells become desensitized to relaxatory nPR-mediated actions of progesterone [3,12,20,38,42,45,48,192–194,215,237,269,287,288,329]. To date, the precise mechanisms for a functional progesterone withdrawal in term labor remain to be elucidated. A role for changes in the nPR signaling pathway is supported by the following observations: 1) term parturition is associated with increased expression of inhibitory nPR isoforms in human myometrium [51,191,195]; 2) there is decreased expression of nPR co-activators in myometrium at term [52,152]; and 3) expression of nPR co-repressors increases during human labor [65]. Our microarray data indicate that nPR and nPR co-regulator expression is not different between the term labor and not in labor groups. However, this may be due to sensitivity issues as nPR and steroid co-regulator gene expression is relatively low in human myometrium and small changes in gene expression may not be detectable by microarray analysis. Nonetheless, several genes identified as differentially expressed with labor onset provide evidence for decreased nPR transcriptional activity, and therefore functional progesterone withdrawal, in laboring tissue: 1) IL-8, which is decreased by progesterone [154], and whose expression is increased in myometrium during labor; 2) PTGS2/COX2, which is also decreased by progesterone [78], whose expression in myometrium was also increased during spontaneous term labor; and 3) the immunophilin FK506 binding protein 5 (FKBP5; alias FKBP51), whose expression is normally increased by progesterone [144], but was decreased in the term labor group. Recently, the differential expression of FKBP5 has been described in cultured human chorion and decidua cells (obtained from women at term not in labor) treated with progesterone [207]. While FKBP5 expression increased in the chorion with exogenous progesterone treatment, its expression decreased in the decidua.

This is the first study reporting decreased expression of the immunophilin FKBP5 [285] in myometrium from women at term in labor compared to that obtained from women not in labor. FKBP5 is an immunophilin that participates with other proteins such as Hsp90 and p23 in forming the mature steroid hormone receptor complex capable of binding steroid with high efficacy and affinity [209,229,286]. Of note, FKBP52 (alias FKBP4; related immunophilin) null mice exhibit decreased uterine responsiveness to circulating progesterone leading to implantation failure [314]. However, the pregnancies of null mice with a specific genetic makeup (CD1 null mice) could be rescued with progesterone supplementation. Interestingly, the concentrations of progesterone treatment required to maintain pregnancy were specific to the stage of pregnancy-increasing concentrations were required as the pregnancy progressed [314]. Importantly, FKBP5 has been described to preferentially accumulate in the mature nPR complex [9,208], has a competitive advantage for progesterone receptor association over both the FKBP52 and Cyp-40 immunophilins [9], and its expression is upregulated by progesterone via the type-B nPR (PR-B) in breast cancer cell models [144,145]. In this context, decreased expression of this PR-B-responsive gene may reflect decreased PR-B-mediated progesterone activity in the myometrial cells. This is important because PR-B is thought to be the principal mediator of relaxatory actions of progesterone in human pregnant myometrium [51,52,191,195]. Therefore, a decrease in its transcriptional activity may reflect functional progesterone withdrawal. An alternative hypothesis is that decreased FKBP5 limits the capacity for progesterone to maintain a relaxed phenotype via PR-B since it participates as a molecular chaperone in the formation of functional PR-B complexes with high affinity for ligand. Therefore, the decreased myometrial expression of FKBP5 in association with term labor may not only reflect

functional progesterone withdrawal but may also be a novel mechanism for functional progesterone withdrawal by limiting progesterone actions via PR-B. Further studies are needed to test these hypotheses.

Additional genes associated with spontaneous labor at term

We report the novel finding of HBEGF over-expression in myometrium during labor compared to quiescent myometrium. This is consistent with evidence that exercise is associated with increased expression of HBEGF in skeletal muscle [105]. Of interest, in a rat model, the closely related epidermal growth factor (EGF) produced rhythmic uterine contractions that were abolished when tissue was treated with anti-EGF antibodies [107]. It is therefore plausible that HBEGF may also contribute to uterine contractility. HBEGF also acts as a mitogen for smooth muscle and fibroblasts while protecting against apoptosis during stress [141]. A transgenic mouse model demonstrated that increased expression of HBEGF is associated with increased glucose uptake and insulin sensitivity consistent with facilitation of glucose disposal [105]. During pregnancy, HBEGF expression has been associated with trophoblast survival [4,150,167,328] and we have previously reported decreased HBEGF placental expression in preeclampsia [168].

ALDH2, which was under-expressed during labor, is a mitochondrial enzyme essential for the oxidation and subsequent elimination of acetaldehyde [162]. The pathway of acetaldehyde metabolism interacts with that of retinol metabolism; interestingly, retinoic acid decreases transcription of the progesterone receptor gene [49] and increases expression of the oxytocin receptor gene in rat myometrium [166]. Indeed, gene deletion for retinoic acid receptor- α is associated with a decreased ethanol and acetaldehyde clearance with decreased activity of ALDH2 [127]. Myometrial ALDH2 expression has not been previously reported. The precise role of this gene in human parturition warrants further investigation.

Comparison with previous reports of functional genomics in the study of term human parturition

Aguan et al.[1] first reported the use of functional genomics to address the molecular mechanisms of labor in humans employing cDNA macroarrays to assess the expression of 588 genes from the myometrium of women with (n=3), and without term labor (n=3). Differential expression was observed in 21 genes of diverse functions. These results were not confirmed with a targeted approach (Northern blot or PCR). Using suppression subtractive hybridization (SSH), Chan et al.[39] compared myometrium from women at term not in labor and those with dysfunctional labor. The authors identified 400 clones. Thirty clones were differentially expressed and over-expression of cyclophilin, SOD2, and IL-8 was confirmed in patients with dysfunctional labor using qRT-PCR. We found overexpression of SOD2 and IL-8 in our microarray data. However, confirmatory qRT-PCR was significant only for IL-8. The molecular basis of dysfunctional term labor including arrest of dilatation and arrest of descent is an area for future study.

Havelock et al.[136] employed cDNA microarray in their analysis comparing spatial differential gene expression in myometrium from women with and without labor with pooled samples of 4 specimens per group. Differential expression of PTGS2/COX2 and calgranulin B (S100A9) was confirmed with qRT-PCR. In the current study, both PTGS2/COX2 and S100A9 were upregulated according to the microarray analysis, with confirmed over-expression of PTGS2/COX2. Using cDNA microarray, Esplin et al.[90] reported the differential expression of 56 unique genes involved in human parturition (term not in labor, n=5, term labor, n=5). The authors confirmed increased expression of 4 genes in labor: THBS1, SOD2, PBEF1, and NNMT. Our microarray findings were consistent with these

results. In contrast, verification of these findings using qRT-PCR in an independent set of samples did not yield significant results for THBS1, SOD2, or PBEF1. The authors also noted over-expression of CCL2 in myometrium during labor and confirmed this finding in a subsequent investigation.[91]

Bukowski et al [25] reported results describing the differences in transcriptomes among the uterine fundus, lower uterine segment, and cervix as they vary prior to (n=6) and during labor (n=7). However, after correction for multiple comparisons, the differences were no longer significant [262]. Validation studies with RT-PCR did confirm decreased expression of repressor of estrogen receptor alpha (REA) and retinoid X receptor alpha (RXR) in the uterine fundus during labor. Our findings in the present study could not confirm differential expression of REA or RXR in myometrium from the lower uterine segment of women in labor. Recently, the group of O'Brien et al.[214] utilized the Applied Biosystems Genome Survey Microarray version 2 to investigate the myometrial transcriptome of labor. A total of 698 genes were differentially expressed between the term labor (n=3) and term not in labor (n=3) groups. Over-expression of PSCDBP, EDNRB, TLR2, FLJ35383, TWIST1, and RGS12 was reported. Of interest, none of these genes were significantly overexpressed in the current study.

While differential gene expression in myometrium from women with and without labor has been described using global methods and targeted approaches, such as Northern blot analysis and qRT-PCR, differences in detection between methods are frequently noted, as demonstrated above. Similarly, overlap between different reports is also evident. The lack of uniform gene expression signatures is possibly due to differences in patient selection, experimental design, statistical analysis and the platforms used.

A noteworthy aspect of this study (for future in-depth functional analyses of the differentially expressed genes) is our finding that some differentially expressed genes in Tables 2 and 3 do not have a known function. Conserved among mammals, LOC100132684, synonymous with C14ORF132, (Table 3) encodes a novel 83-aa protein with no known domains. C5ORF4, encoding a 333-aa protein without a known function, is even more highly conserved throughout the animal kingdom. Our understanding of C5ORF4 may benefit from the annotation efforts of diverse organismal genome projects, as the mosquito (*Aedes aegypti*) homolog of this gene appears to be the first member of this family to be functionally annotated as a sterol desaturase. These results indicate that some platform annotations need updating or further interpretation aided by GenBank synonyms and homology searches to derive functional meanings.

Several differentially expressed loci in our study portray evidence of genomic complexity beyond protein-coding genes. In particular, STRBP (Table 3) hosts a small regulatory microRNA, miR-600, in its 3' untranslated region. Therefore, the functional regulatory output of this locus may not be limited to the STRBP protein, and may include downstream effects of the microRNA. We have previously reported differential expression of microRNAs in the uterine cervix during term labor [133]. Additionally, HIF1A and HOXA11 both have endogenous non-coding cis-antisense RNA transcripts [40,268]. Understanding the impact of functional RNA on the regulation of genes in term labor should aid our effort to place these genes into regulatory networks.

Strengths and limitations of the study

A major strength of this study is the large sample size included in the microarray analysis: the largest reported to date. In addition, the results of the microarray experiments for selected genes were confirmed with qRT-PCR in an independent set of samples (biological validation). Moreover, confirmation of differential protein expression between the groups

was performed using a third separate group of specimens. We have identified novel genes previously unrecognized to participate in human labor and verified the differential expression of genes previously implicated in the transformation of human myometrium from a quiescent to a contractile organ.

Our results also include the first description of the possible biological processes, molecular functions, and pathways associated with the transformation of human myometrium from a quiescent to a contractile organ derived from an unbiased and comprehensive analysis of the myometrial transcriptome. These findings provide a basis for future studies in which the differences and similarities in the myometrial transcriptome between women in term and preterm labor can be addressed. In those future studies, it may be helpful, for greater understanding of gene regulation in labor as well as for development of therapeutics, to define the high-level network regulators governing the entire functional module of genes emerging as different in term labor versus non-labor myometrium. Integration of genome-wide chromatin immunoprecipitation and sequencing (ChIP-seq) results from existing public studies of transcription factors involved in the processes we identified should help test the hypothesis that such transcription factors are master regulators. As a proof of principle, we asked whether any known human genes that are direct targets of NF κ B by ChIP-ditag sequencing [178] are differentially expressed in term labor. In fact, 10 differentially expressed genes from our study (ENC1, PDE4B, IL8, PIM1, RGS10, SOD2, IL8RB, IL1B, NFKBIZ, and IER3) are genomic direct targets of the RelA NF κ B subunit [178].

A potential shortcoming of our study is the racial polarity of our patient population, which is mainly African-American. The generalizability of our findings to other patient populations will require future investigation. Also, our results specifically describe the stereotypic transcriptome of myometrium from the lower uterine segment of the uterus and do not address the concurrent changes in the transcriptome of the uterine fundus during spontaneous labor at term. Furthermore, the relationship between differential expression of genes in the myometrium and the subsequent onset of labor cannot be studied using serial sampling in women. These studies would be important to establish causality, and would need animal experimentation.

In conclusion, spontaneous term labor is characterized by a stereotypic myometrial transcriptome including 471 differentially expressed genes. The application of high-dimensional biology techniques (transcriptomics) has enabled the identification of differentially expressed genes and processes involved in human parturition, and demonstrated the strong association between spontaneous labor and inflammation. These studies are essential for the understanding of parturition and will serve as the basis for understanding the differences between normal labor at term and dysfunctional labor.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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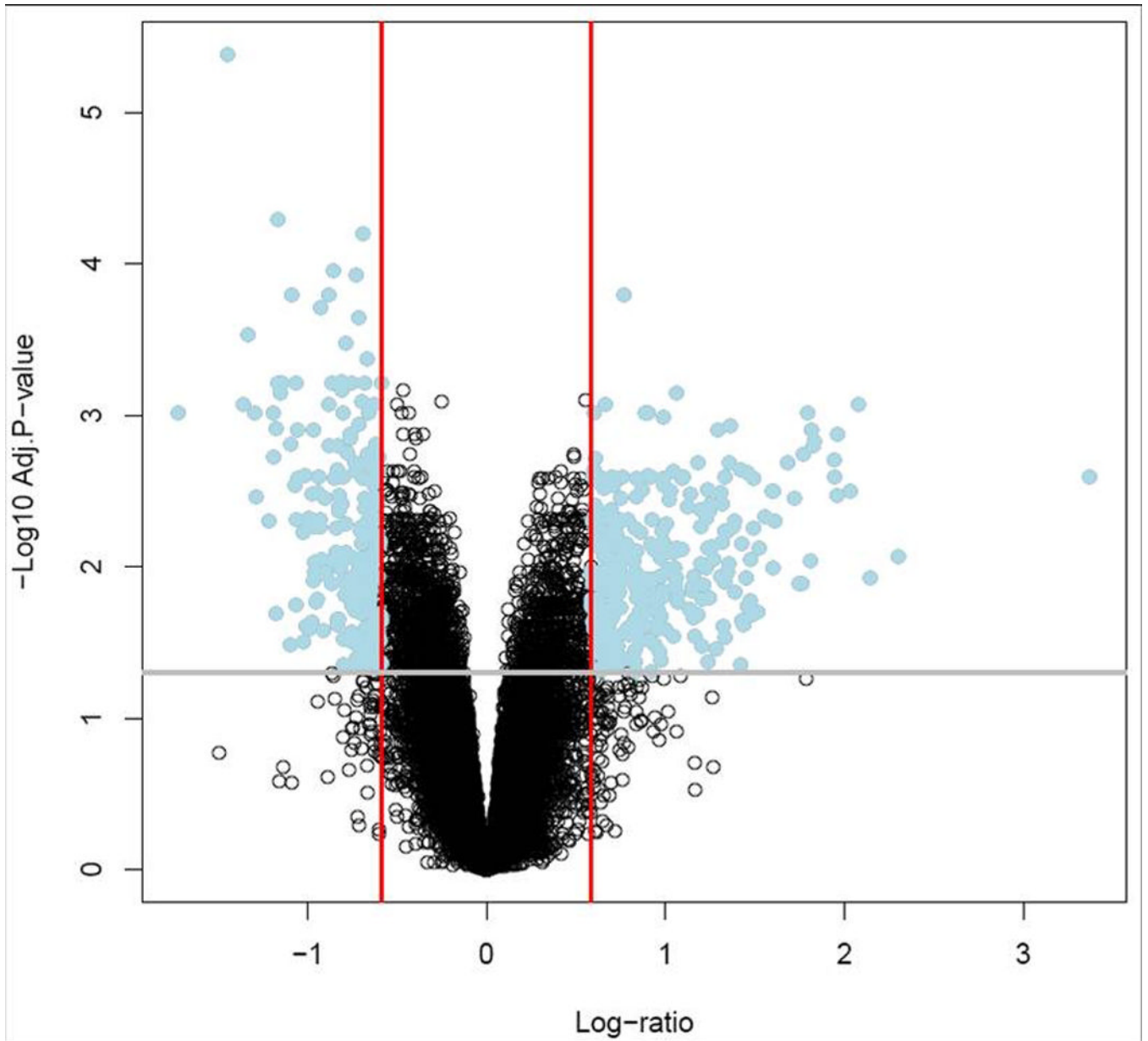
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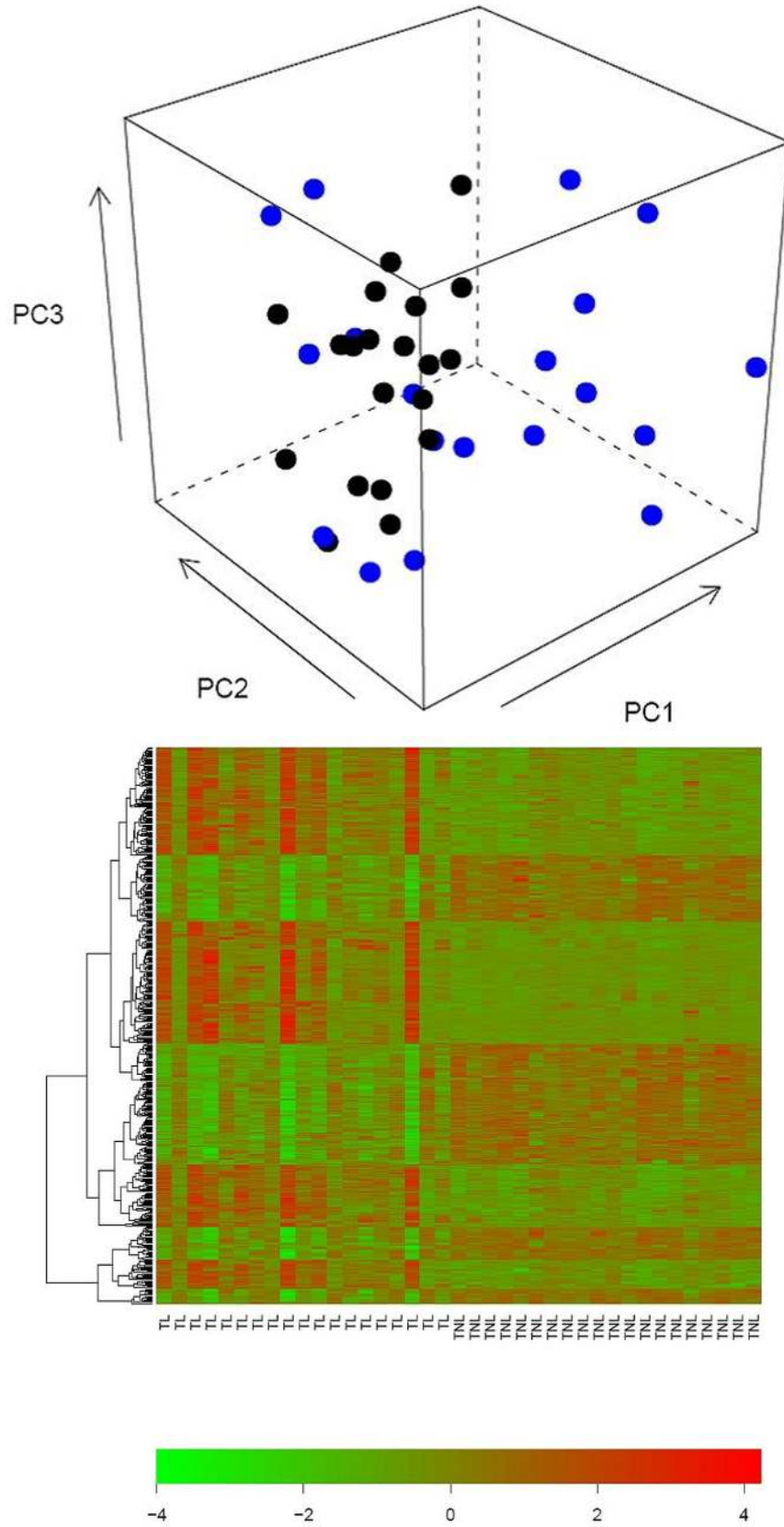


Figure 1.

Microarray analysis of the gene expression profiles of myometrium at term not in labor and spontaneous term labor.

Figure 1A. A Volcano plot showing the ratio between the average gene expression of the term not in labor (TNL) and term labor (TL) groups (x-axis) versus the significant p-values from the moderated t-test. Circles in the upper right and left quadrants represent genes with a fold change greater than 1.5 and a false discovery rate corrected p-value <0.05. With these criteria, 471 genes were differentially expressed between the myometrial transcriptomes of the two groups.

Figure 1B. Three-dimensional principal component analysis plot (PCA) demonstrating segregation of the TNL and TL groups based on gene expression levels. Black points indicate individual samples from the TNL groups while blue dots represent those from the group with TL.

Figure 1C. Heatmap of gene expression in TNL and TL clustered by genes. Rows correspond to genes while columns correspond to samples. High expression levels are shown in red, while low expression levels are in green. Data was log (base 2) transformed, and values were mean centered by rows. Color key equals \log_2 expression levels.

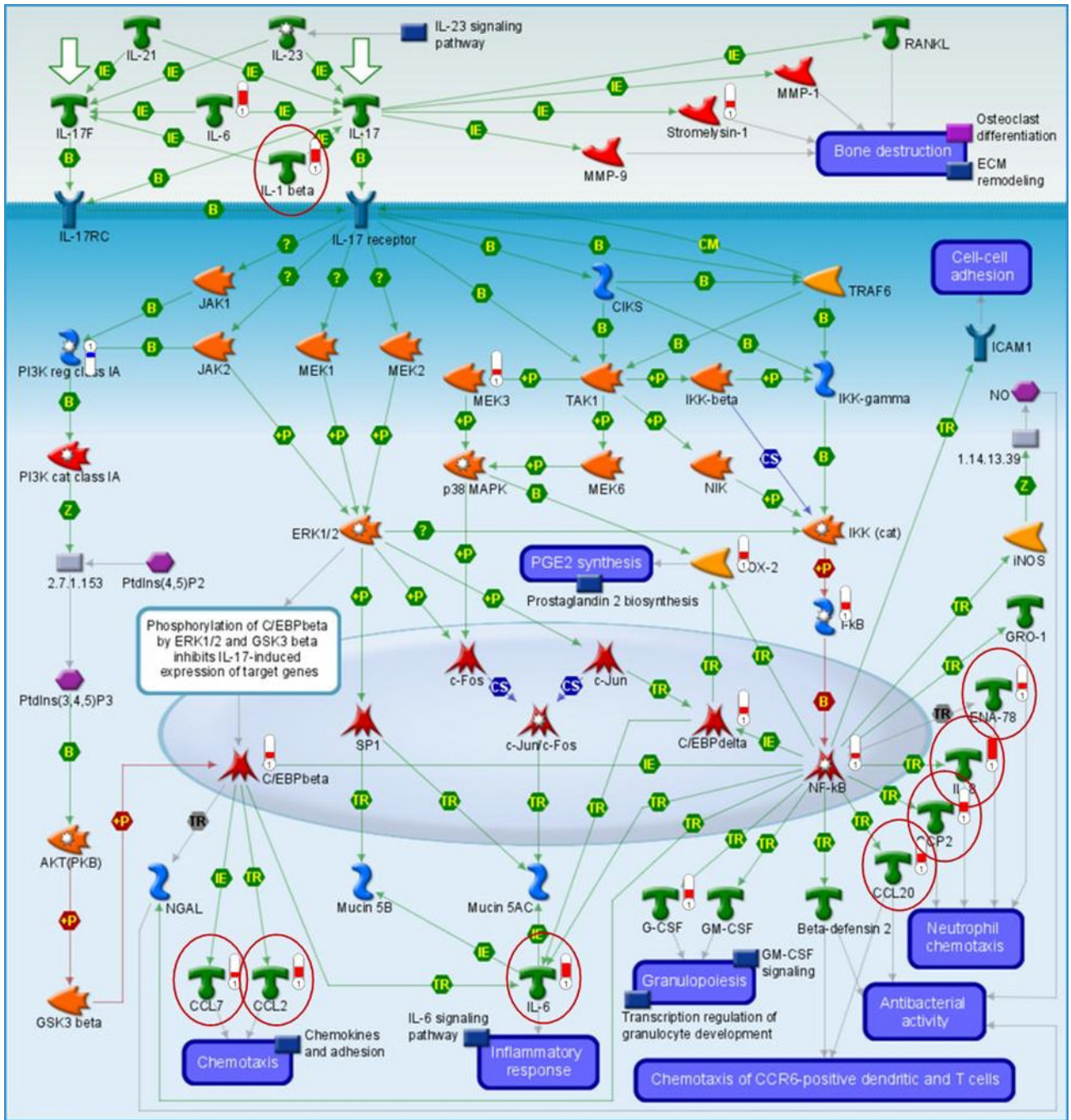


Figure 2. The Interleukin-17 signaling pathway (MetaCore)

Display of differentially expressed genes in human myometrium during term labor mapped on the MetaCore Interleukin-17 signaling pathway. Red thermometers indicate gene upregulation in labor while blue thermometers indicate downregulation in labor. Of note, the majority of differentially regulated genes in this pathway are those involved in inflammation and chemotaxis (circled genes).

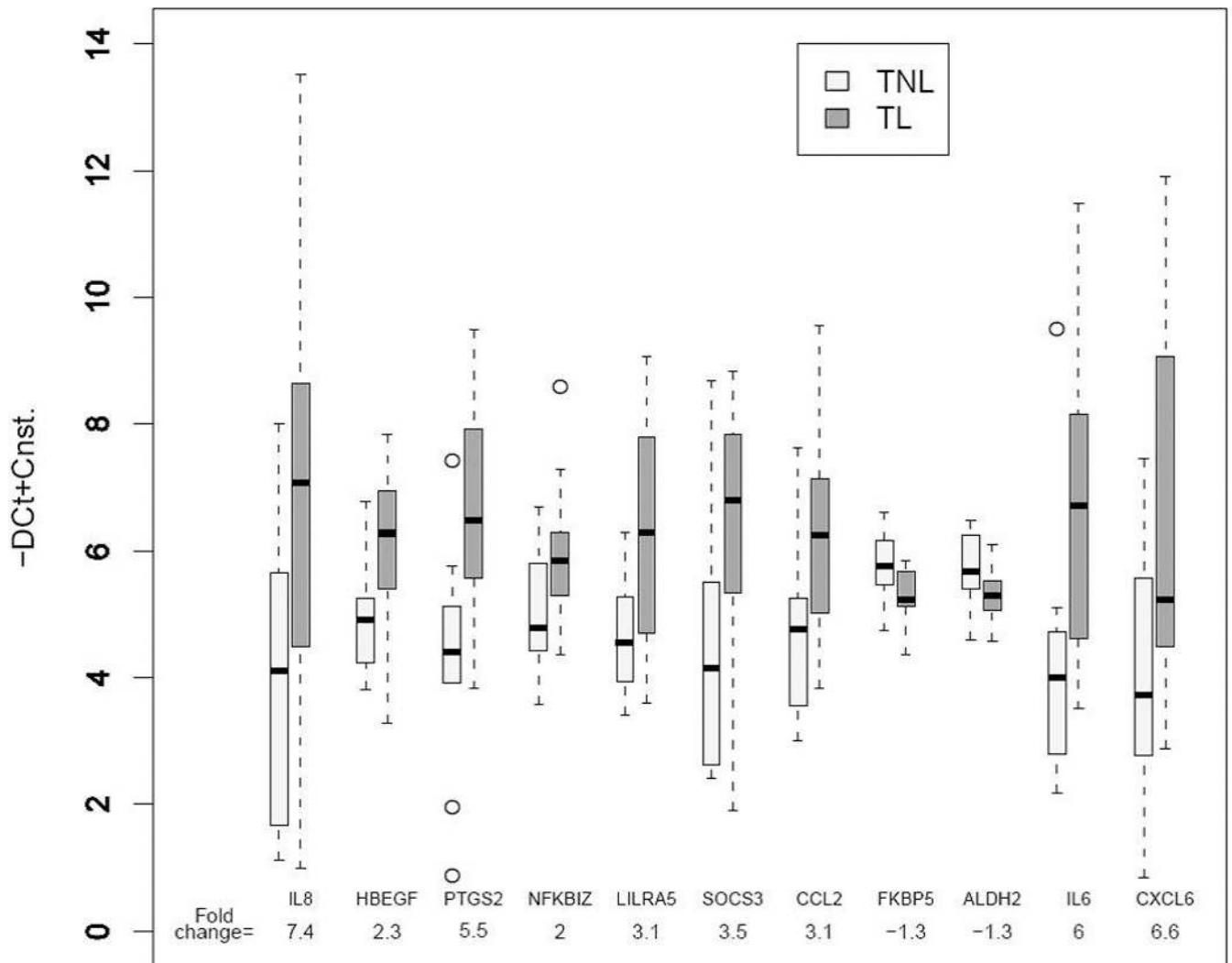
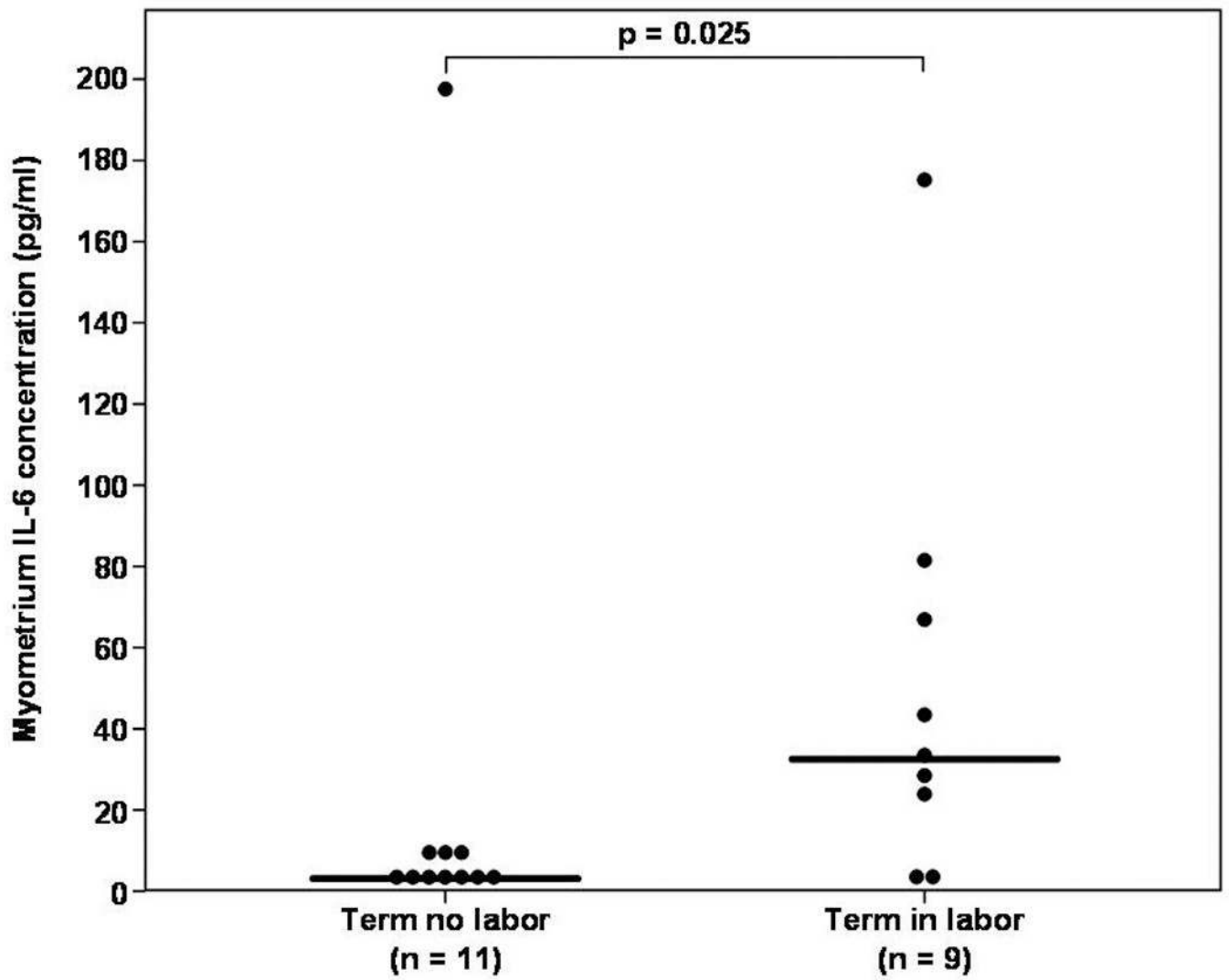


Figure 3. Box plots of significant qRT-PCR assays

The data is presented as $-DCt$ values (Ct reference gene- Ct target gene) which is a surrogate for gene expression (on a \log_2 scale). The boxes encompass 50% of the data from the 1st quartile to the 3rd quartile. The middle line represents the median value (50%) quantile. The whiskers extend to the most extreme data point, but do not exceed values 1.5 times the interquartile range from the box. The circles represent outliers. Significance was defined as a $P < 0.05$. TNLster not in labor, TLsspontaneous term labor, ILsinterleukin, HBEGFsheparin binding EGF-link growth factor, PTGS2sprostaglandin-endoperoxide synthase-2, NFKBIZsnuclear factor of kappa light chain gene enhancer in B-cells inhibitor zeta, LILRA5sleukocyte immunoglobulin-like receptor, subfamily A, member 5, SOCS3s suppressor of cytokine signaling 3, CCL2schemokine C-C motif ligand 2, FKBP5sFK506 binding-protein 5, ALDH2saldehyde dehydrogenase, CXCL6schemokine C-X-C motif ligand 6.



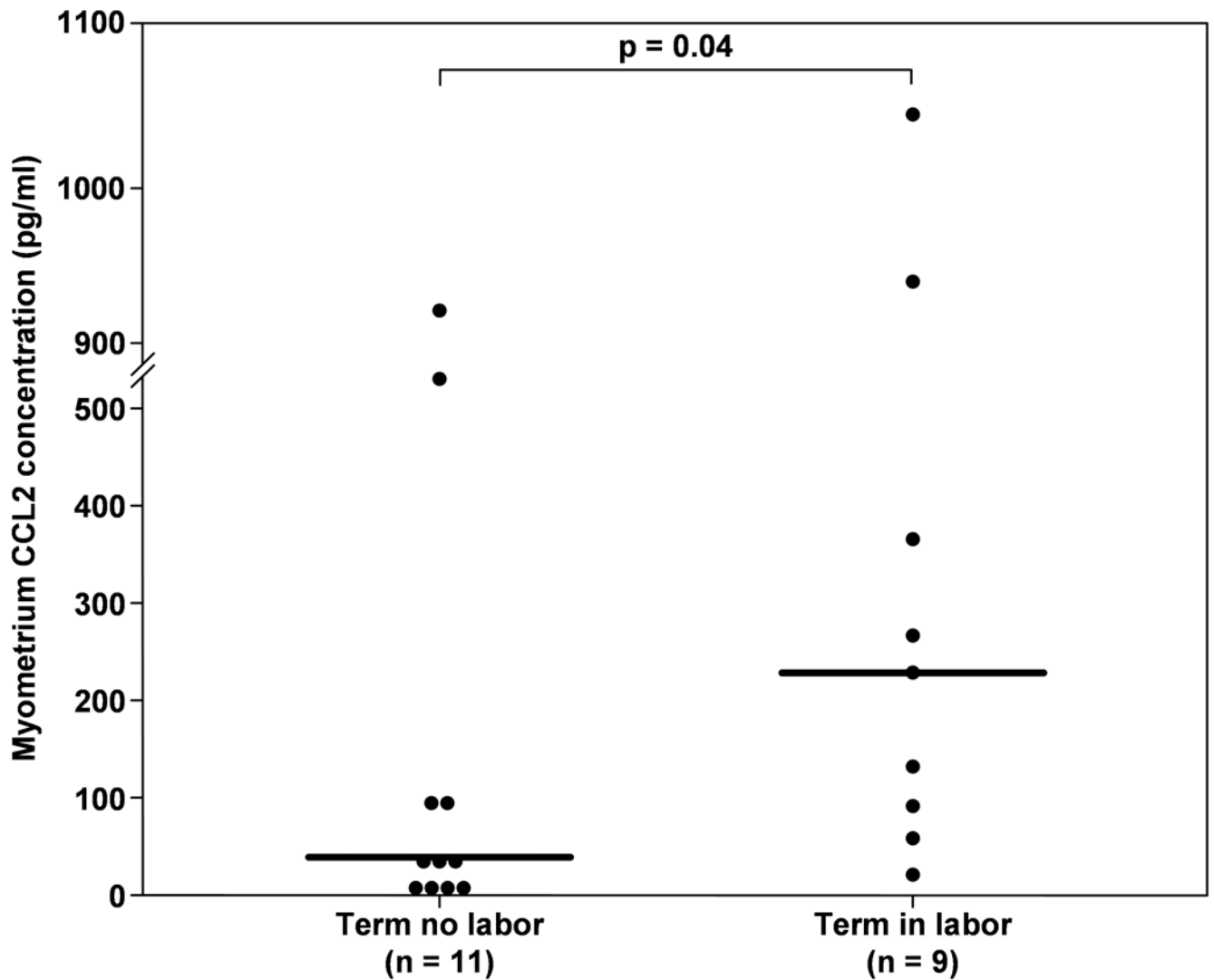


Figure 4.

Comparison of myometrial protein concentrations of interleukin-6 and CCL2 between term not in labor and spontaneous labor.

(A) The median interleukin-6 concentration was significantly higher in women at term in labor compared to those without labor (term not in labor 1.87 pg/mL interquartile range (IQR), IQR 0–9.0 vs. term labor 34.59 pg/mL, IQR 14–73; $P=0.25$).

(B) Median CCL2 protein concentration was also higher in myometrium from women in labor compared to those not in labor (term not in labor 35.8 pg/mL, IQR 6–107 vs. term labor 229.41 pg/mL, IQR 74–652; $P=0.04$). CCL2, chemokine C-C motif ligand 2, IL-8, interleukin.

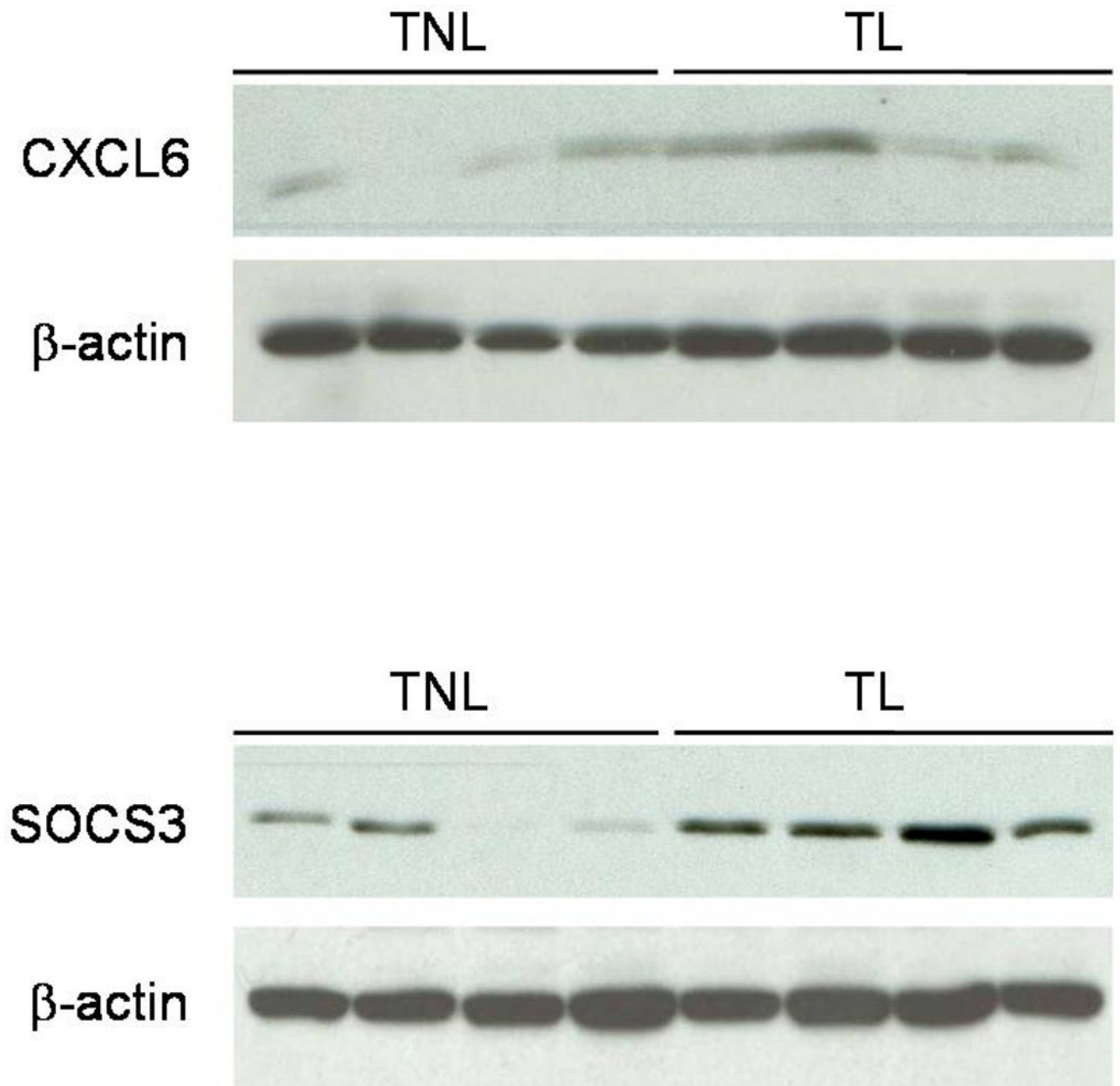


Figure 5. Immunoblotting analysis of CXCL6 and SOCS3 in term myometrium. Ten mg of total proteins were electrophoresed in 18% and 12% SDS-PAGE for CXCL6 and SOCS3, respectively. Protein concentrations of both CXCL6 and SOCS3 were higher in TL specimens compared to TNL. TNLsterm not in labor, TLsterm labor, CXCL6schemokine C-X-C motif ligand 6, SOCS3ssuppressor of cytokine signaling 3.

Table 1

Demographic and clinical characteristics of the study groups

| Category | Term not in labor Microarray (n=20) | Term labor Microarray (n=19) | Term not in labor qRT-PCR (n=10) | Term labor qRT-PCR (n=10) | Term not in labor Immunoblot/ ELISA (n=11) | Term labor Immunoblot/ ELISA (n=9) | p-value |
|-------------------------------------|---|------------------------------------|--|---------------------------------|---|---|---------|
| Maternal age (years) | 33 (21–39) | 27 (19–39) | 27 (19–33) | 26 (20–40) | 24 (19–30) | 27 (21–38) | NS |
| African-American ethnicity | 75 (15/20) | 70 (14/20) | 80 (8/10) | 90 (9/10) | 80 (8/10) | 80 (8/10) | NS |
| BMI (kg/m ²) | 28 (18.2–47.2) | 28.7 (22.1–54.6) | 31.8 (21.7–61) | 24.2 (21–31.3) | 29.6 (22.3–38.4) | 27.7 (21–33.4) | NS |
| Parity | 1 (0–4) | 0 (0–5) | 1 (0–6) | 1 (0–5) | 1 (0–4) | 1 (0–5) | NS |
| Gestational age at delivery (weeks) | 38.7 (37–41.9) | 39.3 (37–41.3) | 39.1 (38.9–41.3) | 38.9 (37–41) | 39.1 (38.7–41) | 39.2 (38.9–40.5) | NS |
| Birthweight (grams) | 3,070 (2,545–3,805) | 3,150 (2,570–3,740) | 3,330 (3,090–3,930) | 3,245 (2,645–3,740) | 3,490 (3,010–3,980) | 3,280 (2,870–3,840) | NS |

Values are expressed as percentage (number) or median (range) BMI: body mass index; NS: not significant.

Table 2

Microarray Results. Top 50 probes with overexpression in human myometrium during spontaneous term labor

| ENTREZ Gene ID | SYMBOL | Gene Name | Fold Change | FDR corrected P-value |
|----------------|-----------|--|-------------|-----------------------|
| 8507 | ENC1 | ectodermal-neural cortex | 1.70 | 0.0002 |
| 10630 | PDPN | podoplanin | 2.09 | 0.0007 |
| 8703 | B4GALT3 | UDP-Gal:betaGlcNAc beta 1,4-galactosyltransferase, polypeptide 3 | 1.59 | 0.0008 |
| 3872 | KRT17 | keratin 17 | 4.22 | 0.0008 |
| 10797 | MTHFD2 | methylenetetrahydrofolate dehydrogenase 2 | 1.87 | 0.0010 |
| 58477 | SRPRB | signal recognition particle receptor, B subunit | 1.52 | 0.0010 |
| 353514 | LILRA5 | leukocyte immunoglobulin-like receptor, subfamily A member 5 | 3.48 | 0.0010 |
| 22936 | ELL2 | elongation factor, RNA polymerase II, 2 | 1.85 | 0.0010 |
| 165 | AEBP1 | AE binding protein 1 | 1.98 | 0.0010 |
| 64651 | AXUD1 | AXIN1 up-regulated 1 | 2.57 | 0.0012 |
| 117247 | SLC16A10 | solute carrier family 16, member 10 | 2.45 | 0.0012 |
| 6280 | S100A9 | S100 calcium binding protein A9 | 3.52 | 0.0012 |
| 9941 | EXOG | endo/exonuclease (5'-3'), endonuclease G-like | 3.90 | 0.0013 |
| 4489 | MT1A | metallothionein 1A | 3.56 | 0.0015 |
| 366 | AQP9 | aquaporin 9 | 3.55 | 0.0016 |
| 4502 | MT2A | metallothionein 2A | 3.41 | 0.0018 |
| 732360 | LOC732360 | similar to G/T mismatch-specific thymine DNA glycosylase; pseudogene | 1.53 | 0.0019 |
| 6279 | S100A8 | S100 calcium binding protein A8 | 3.85 | 0.0020 |
| 10221 | TRIB1 | tribbles homolog 1 (Drosophila) | 2.56 | 0.0020 |
| 26585 | GREM1 | gremlin 1, cysteine knot superfamily, homolog (Xenopus laevis) | 3.22 | 0.0021 |
| 5142 | PDE4B | phosphodiesterase 4B, cAMP-specific | 2.27 | 0.0021 |
| 10135 | NAMPT | nicotinamide phosphoribosyltransferase | 2.68 | 0.0023 |
| 5744 | PTH1H | parathyroid hormone-like hormone | 1.52 | 0.0023 |
| 1647 | GADD45A | growth arrest and DNA-damage-inducible, alpha | 2.06 | 0.0023 |
| 8140 | SLC7A5 | solute carrier family 7 member 5 | 2.74 | 0.0024 |
| 4837 | NNMT | nicotinamide N-methyltransferase | 1.87 | 0.0025 |
| 3576 | IL8 | interleukin 8 | 10.36 | 0.0025 |
| 90007 | MIDN | midnolin | 1.70 | 0.0025 |
| 23560 | GTPBP4 | GTP binding protein 4 | 1.51 | 0.0025 |
| 1735 | DIO3 | deiodinase, iodothyronine, type III | 2.52 | 0.0025 |
| 85450 | ITPRIP | inositol 1,4,5-triphosphate receptor interacting protein | 2.15 | 0.0025 |
| 25819 | CCRN4L | CCR4 carbon catabolite repression 4-like (S. cerevisiae) | 1.90 | 0.0025 |
| 29950 | SERTAD1 | SERTA domain containing 1 | 1.62 | 0.0025 |
| 5743 | PTGS2 | prostaglandin-endoperoxide synthase 2 | 3.86 | 0.0025 |
| 4501 | MT1X | metallothionein 1X | 2.82 | 0.0026 |
| 22856 | CHSY1 | chondroitin sulfate synthase 1 | 1.79 | 0.0026 |
| 23764 | MAFF | v-maf musculoaponeurotic fibro sarcoma oncogene homolog F (avian) | 1.99 | 0.0026 |
| 1847 | DUSP5 | dual specificity phosphatase 5 | 2.13 | 0.0026 |

| ENTREZ Gene ID | SYMBOL | Gene Name | Fold Change | FDR corrected P-value |
|----------------|----------|--|-------------|-----------------------|
| 9446 | GSTO1 | glutathione S-transferase omega 1 | 1.57 | 0.0026 |
| 26585 | GREM1 | gremlin 1, cysteine knot superfamily, homolog (Xenopus laevis) | 1.63 | 0.0028 |
| 57761 | TRIB3 | tribbles homolog 3 (Drosophila) | 1.55 | 0.0028 |
| 5209 | PFKFB3 | 6-phosphofructo-2-kinase/fructose-2,6- biphosphatase 3 | 1.60 | 0.0028 |
| 3099 | HK2 | hexokinase 2 | 1.64 | 0.0029 |
| 4084 | MXD1 | MAX dimerization protein 1 | 1.99 | 0.0029 |
| 57647 | DHX37 | DEAH (Asp-Glu-Ala-His) box polypeptide 37 | 1.58 | 0.0031 |
| 5292 | PIM1 | pim-1 oncogene | 2.35 | 0.0031 |
| 3566 | IL4R | interleukin 4 receptor | 1.83 | 0.0031 |
| 51129 | ANGPTL4 | angiopoietin-like 4 | 3.04 | 0.0032 |
| 2357 | FPR1 | formyl peptide receptor 1 | 3.02 | 0.0032 |
| 199675 | C19orf59 | chromosome 19 open reading frame 59 (mast cell-expressed membrane protein 1) | 4.10 | 0.0032 |

Table 3

Microarray Results. Top 50 probes with underexpression in human myometrium during spontaneous term labor

| ENTREZ Gene ID | SYMBOL | Gene Name | Fold Change | FDR corrected P-value |
|----------------|--------------|--|-------------|-----------------------|
| 1580 | CYP4B1 | cytochrome P450, family 4, subfamily B, polypeptide 1 | -2.73 | 4.12 E-06 |
| 1580 | CYP4B1 | cytochrome P450, family 4, subfamily B, polypeptide 1 | -2.24 | 0.00005 |
| 55342 | STRBP | spermatid perinuclear RNA binding protein | -1.62 | 0.00006 |
| 3207 | HOXA11 | homeobox A11 | -1.81 | 0.00011 |
| 126823 | KLHDC9 | kelch domain containing 9 | -1.66 | 0.00012 |
| 1728 | NQO1 | NAD(P)H dehydrogenase, quinone 1 | -2.13 | 0.00016 |
| 7743 | ZNF189 | zinc finger protein 189 | -1.85 | 0.00016 |
| 126823 | KLHDC9 | kelch domain containing 9 | -1.90 | 0.00019 |
| 5641 | LGMN | legumain | -1.64 | 0.00023 |
| 85004 | RERG | RAS-like, estrogen-regulated, growth inhibitor | -2.52 | 0.00030 |
| 145781 | GCOM1 | GRINL1A complex locus | -1.72 | 0.00033 |
| 2289 | FKBP5 | FK506 binding protein 5 | -1.59 | 0.00042 |
| 65009 | NDRG4 | NDRG family member 4 | -1.75 | 0.00060 |
| 100132684 | LOC100132684 | hypothetical protein LOC100132684 (C14ORF132) | -2.09 | 0.00062 |
| 6001 | RGS10 | regulator of G-protein signaling 10 | -1.50 | 0.00062 |
| 2861 | GPR37 | G protein-coupled receptor 37 | -2.20 | 0.00062 |
| 25959 | KANK2 | KN motif and ankyrin repeat domains 2 | -1.82 | 0.00062 |
| 51284 | TLR7 | toll-like receptor 7 | -1.69 | 0.00062 |
| 5733 | PTGER3 | prostaglandin E receptor 3 | -2.24 | 0.00062 |
| 284 | ANGPT1 | angiopoietin 1 | -1.75 | 0.00069 |
| 284 | ANGPT1 | angiopoietin 1 | -2.22 | 0.00071 |
| 3489 | IGFBP6 | insulin-like growth factor binding protein 6 | -1.62 | 0.00080 |
| 8718 | TNFRSF25 | tumor necrosis factor receptor superfamily, member 25 | -1.54 | 0.00084 |
| 5334 | PLCL1 | phospholipase C-like 1 | -2.57 | 0.00084 |
| 2857 | GPR34 | G protein-coupled receptor 34 | -1.84 | 0.00084 |
| 6414 | SEPP1 | selenoprotein P, plasma, 1 | -2.46 | 0.00096 |
| 256691 | MAMDC2 | MAM domain containing 2 | -3.30 | 0.00096 |
| 3757 | KCNH2 | potassium voltage-gated channel, subfamily H, member 2 | -2.29 | 0.00096 |
| 145781 | GCOM1 | GRINL1A complex locus | -1.58 | 0.00097 |
| 27175 | TUBG2 | tubulin, gamma 2 | -1.74 | 0.00097 |
| 51435 | SCARA3 | scavenger receptor class A, member 3 | -1.64 | 0.00115 |
| 6035 | RNASE1 | ribonuclease, RNase A family, 1 | -2.26 | 0.00123 |
| 6035 | RNASE1 | ribonuclease, RNase A family, 1 | -2.08 | 0.00125 |
| 6543 | SLC8A2 | solute carrier family 8, member 2 | -1.95 | 0.00125 |
| 130814 | PQLC3 | PQ loop repeat containing 3 | -1.70 | 0.00133 |
| 10608 | MXD4 | MAX dimerization protein 4 | -1.70 | 0.00143 |
| 4286 | MITF | microphthalmia-associated transcription factor | -1.54 | 0.00154 |

| ENTREZ Gene ID | SYMBOL | Gene Name | Fold Change | FDR corrected P-value |
|----------------|---------|--|-------------|-----------------------|
| 3033 | HADH | hydroxyacyl-Coenzyme A dehydrogenase | -2.13 | 0.00154 |
| 5733 | PTGER3 | prostaglandin E receptor 3 | -1.84 | 0.00160 |
| 4056 | LTC4S | leukotriene C4 synthase | -1.78 | 0.00160 |
| 79921 | TCEAL4 | transcription elongation factor A (SII)-like 4 | -1.71 | 0.00177 |
| 2018 | EMX2 | empty spiracles homeobox 2 | -1.58 | 0.00181 |
| 463 | ZFH3 | zinc finger homeobox 3 | -1.52 | 0.00188 |
| 10826 | C5orf4 | chromosome 5 open reading frame 4 | -2.28 | 0.00190 |
| 9459 | ARHGEF6 | Rac/Cdc42 guanine nucleotide exchange factor 6 | -1.60 | 0.00201 |
| 9068 | ANGPTL1 | angiopoietin-like 1 | -1.59 | 0.00204 |
| 79148 | MMP28 | matrix metalloproteinase 28 | -1.67 | 0.00207 |
| 10742 | RAI2 | retinoic acid induced 2 | -1.55 | 0.00222 |
| 54510 | PCDH18 | protocadherin 18 | -1.78 | 0.00230 |
| 7106 | TSPAN4 | tetraspanin 4 | -1.52 | 0.002339 |

Table 4

A. Gene Ontology analysis: Biological Processes enriched in the differentially expressed genes between spontaneous term labor and term not in labor Partial list (total of 103 significant processes)

| Biological Process Category | Differentially expressed genes/ number of total genes in GO term | Adjusted P-value |
|----------------------------------|---|------------------|
| Inflammatory response | 36/257 | 1.42E-09 |
| Response to wounding | 44/371 | 1.42E-09 |
| Response to external stimulus | 55/576 | 8.95E-09 |
| Defense response | 47/458 | 2.10E-08 |
| Behavior | 32/266 | 5.40E-07 |
| Chemotaxis | 21/123 | 5.66E-07 |
| Taxis | 21/123 | 5.66E-07 |
| Locomotion | 35/334 | 2.24E-06 |
| Locomotory behavior | 24/175 | 2.87E-06 |
| Immune system process | 56/723 | 4.52E-06 |
| Response to stimulus | 116/2042 | 7.27E-06 |
| Response to stress | 76/1146 | 7.55E-06 |
| Regulation of cell proliferation | 42/488 | 1.39E-05 |
| Immune response | 42/500 | 2.54E-05 |
| Multicellular organismal process | 134/2592 | 9.32E-05 |
| Response to chemical stimulus | 46/604 | 9.53E-05 |

B. Gene Ontology analysis: Molecular Functions enriched in the differentially expressed genes between spontaneous term labor and term not in labor

| Molecular Function Category | Differentially expressed genes/ number of total genes GO term | Adjusted P-value |
|---|--|------------------|
| Cytokine activity | 22/135 | 6.02E-07 |
| Heparin binding | 11/63 | 0.0025 |
| Receptor binding | 42/619 | 0.0029 |
| Chemokine activity | 8/35 | 0.0029 |
| Chemokine receptor binding | 8/36 | 0.0029 |
| Glycosaminoglycan binding | 12/85 | 0.0030 |
| G-protein-coupled receptor binding | 10/61 | 0.0030 |
| Polysaccharide binding | 12/87 | 0.0030 |
| Carbohydrate binding | 20/228 | 0.0075 |
| Pattern binding | 12/101 | 0.0104 |
| Growth factor activity | 13/123 | 0.0167 |
| Cytokine binding | 10/81 | 0.0220 |
| Oxidoreductase activity | 5/21 | 0.0287 |
| Cadmium ion binding | 3/6 | 0.0332 |
| NADPH:quinone reductase activity | 2/2 | 0.0434 |
| Thyroxine 5'-deiodinase activity | 2/2 | 0.0434 |
| Interleukin-8 receptor activity | 2/2 | 0.0434 |
| Oxidoreductase activity NAD or NADP as acceptor | 4/15 | 0.0479 |

Table 5

A. Description of genes selected for qRT-PCR based upon previous reports and results of each gene in current microarray study.

| Symbol | Gene Name | Function | Microarray results in labor |
|---------|---------------------------------------|--|-----------------------------|
| OXTR | Human Oxytocin Receptor | The receptor for the hormone and neurotransmitter oxytocin | N/A |
| THBS1 | Thrombospondin I | Secreted protein associated with the extracellular matrix | Over-expression |
| SOD2 | Superoxide Dismutase 2 | Mitochondrial enzyme, antioxidant | Over-expression |
| GJA1 | Gap Junction Protein, Alpha-1 | Connexin 43, intercellular communication | NS |
| IL8 | Interleukin 8 | Mediates chemotaxis and activation of neutrophils | Over-expression |
| PTGS2 | Prostaglandin-endoperoxide synthase 2 | COX 2, regulation of prostaglandin synthesis | Over-expression |
| CCL2 | chemokine (C-C motif) ligand 2 | Monocyte chemoattractant | Over-expression |
| IL6 | Interleukin 6 | Cytokine mediator of acute phase response | Over-expression |
| IL1B | Interleukin 1-β | Stimulation of T cells, enhanced proliferation of B cells, proinflammatory | Over-expression |
| NAMPT | nicotinamidephosphoribosyltransferase | Visfatin: adipocytokine with anti-apoptotic functions | Over-expression |
| PTGES | prostaglandin E synthase | Enzyme catalyzing production of prostaglandin E from prostaglandin H2 | Over-expression |
| ESR1 | Estrogen receptor-alpha | Nuclear receptor | NS |
| HSP90B1 | heat shock protein 90kDa beta | Molecular chaperone protein required for the proper functioning of steroid receptors | NS |
| S100A8 | S100 calcium binding protein A8 | Innate immune response, expressed by macrophages | Over-expression |

| B. Description of genes selected for qRT-PCR based upon microarray results | | | |
|---|--|---|--|
| Symbol | Gene Name | Function | |
| HBEGF | heparin-binding EGF-like growth factor | Mitogen for fibroblasts and smooth muscle growth; anti-apoptotic | |
| LILRA5 | leukocyte immunoglobulin-like receptor, subfamily A, member 5 | Leukocyte IG-like receptor on monocyte surface, induces secretion of pro-inflammatory cytokines | |
| CXCL6 | chemokine (C-X-C motif) ligand 6 | Neutrophil chemoattractant | |
| NFKBIZ | nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta | Regulation of NF-κB, induction of IL6 secretion | |
| FKBP5 | FK506 binding protein 5 | Progesterone receptor-associated immunophilin required for functionally mature steroid receptor | |
| SOCS3 | suppressor of cytokine signaling 3 | Inhibition of IL6, IL10, and interferon-gamma | |
| ALDH2 | aldehyde dehydrogenase 2 family | Mitochondrial enzyme required for acetaldehyde metabolism | |
| IER3 | immediate early response 3 | Cellular resistance to apoptosis | |
| ALDH7A1 | aldehyde dehydrogenase 7 family, member A1 | Detoxification of aldehydes generated by lipid peroxidation | |
| HIF1A | hypoxia inducible factor 1, alpha subunit | Transcription factor with roles in systemic and cellular responses to hypoxia | |
| HOXA11 | homeobox A11 | Extracellular matrix metabolism inducing collagen III expression | |
| IL24 | Interleukin 24 | Pro-apoptotic cytokine, member of IL10 superfamily | |
| PSAT1 | phosphoserine aminotransferase 1 | Progesterone-induced protein | |
| MMP10 | matrix metalloproteinase 10 | Degrades proteoglycans and fibronectin | |

| B. Description of genes selected for qRT-PCR based upon microarray results | | |
|---|---|--|
| Symbol | Gene Name | Function |
| PROK2 | prokineticin 2 | Output molecule from the suprachiasmatic nucleus circadian clock |
| ALDH1A3 | aldehyde dehydrogenase 1 family, member A3 | Retinoic acid synthesizing enzyme |
| EXOG | endo/exonuclease (5'-3'), endonuclease G-like | Mitochondrial enzyme involved in programmed cell death |

NS: not significant; NA: not available; NS: not significant

Table 6

Comparison of qRT-PCR and microarray analysis of select genes Direction of change denotes change in spontaneous term labor.

| Gene symbol | P-value qRT-PCR | Fold change qRT-PCR | Direction of change in labor qRT-PCR | Corrected P-value microarray | Fold change microarray | Direction of change in labor microarray |
|-------------|-----------------|---------------------|--------------------------------------|------------------------------|------------------------|---|
| PTGS2* | 0.004 | 5.47 | ↑ | 0.003 | 3.86 | ↑ |
| HBEGF* | 0.011 | 2.32 | ↑ | 0.010 | 2.56 | ↑ |
| CCL2* | 0.015 | 3.08 | ↑ | 0.013 | 3.40 | ↑ |
| LILRA5* | 0.016 | 3.09 | ↑ | 0.001 | 3.48 | ↑ |
| IL8* | 0.022 | 7.38 | ↑ | 0.003 | 10.36 | ↑ |
| IL6* | 0.011 | 6.01 | ↑ | 0.009 | 4.92 | ↑ |
| CXCL6* | 0.028 | 6.60 | ↑ | 0.005 | 2.95 | ↑ |
| NFKB1Z* | 0.027 | 2.04 | ↑ | 0.007 | 2.69 | ↑ |
| FKBP5* | 0.038 | -1.34 | ↓ | 0.000 | -1.59 | ↓ |
| SOC3* | 0.040 | 3.46 | ↑ | 0.006 | 2.52 | ↑ |
| ALDH2* | 0.046 | -1.31 | ↓ | 0.007 | -1.52 | ↓ |
| IER3 | 0.066 | 1.91 | ↑ | 0.006 | 2.55 | ↑ |
| SOD2 | 0.075 | 1.79 | ↑ | 0.012 | 2.74 | ↑ |
| ALDH7A1 | 0.086 | -1.27 | ↓ | 0.008 | -1.54 | ↓ |
| IL1B | 0.093 | 3.15 | ↑ | 0.012 | 4.43 | ↑ |
| NAMPT | 0.096 | 2.00 | ↑ | 0.002 | 2.68 | ↑ |
| HSP90B1 | 0.135 | 1.25 | ↑ | 0.143 | 1.17 | ↑ |
| HIF1A | 0.170 | 1.34 | ↑ | 0.014 | 1.62 | ↑ |
| S100A8 | 0.187 | 1.88 | ↑ | 0.002 | 3.85 | ↑ |
| HOXA11 | 0.206 | -1.23 | ↓ | <0.001 | -1.81 | ↓ |
| IL24 | 0.206 | 2.73 | ↑ | 0.018 | 2.75 | ↑ |
| PSAT1 | 0.219 | 1.92 | ↑ | 0.010 | 1.67 | ↑ |
| PTGES | 0.237 | 1.77 | ↑ | 0.013 | 3.36 | ↑ |
| MMP10 | 0.331 | 1.71 | ↑ | 0.020 | 2.87 | ↑ |

| Gene symbol | P-value qRT-PCR | Fold change qRT-PCR | Direction of change in labor qRT-PCR | Corrected P-value microarray | Fold change microarray | Direction of change in labor microarray |
|-------------|-----------------|---------------------|--------------------------------------|------------------------------|------------------------|---|
| GJA1 | 0.394 | 1.09 | ↑ | 0.405 | 1.17 | ↑ |
| PROK2 | 0.406 | 1.19 | ↑ | 0.003 | 3.89 | ↑ |
| ESR1 | 0.419 | 1.04 | ↑ | 0.964 | 1.01 | ↑ |
| ALDH1A3 | 0.497 | 1.00 | ↑ | 0.004 | 2.29 | ↑ |
| EXOG | 0.830 | -1.20 | ↓ | 0.001 | 3.90 | ↑ |
| OXTR | 0.856 | -1.15 | ↓ | NA | NA | NA |
| THBS1 | 0.266 | 1.42 | ↑ | 0.016 | 2.38 | ↑ |

* Genes with significant results by microarray analysis and confirmed differential expression by qRT-PCR

NA: Not applicable. A probe set for this gene is not included on the Illumina® HumanHT-12 v3 expression microarray platform

↑ means increased expression in the Term Labor group compared to the Term Not in Labor group

↓ means decreased expression in the Term Labor group compared to the Term Not in Labor group