ORIGINAL PAPER



Chemical composition of leguminous seeds: part I—content of basic nutrients, amino acids, phytochemical compounds, and antioxidant activity

Eugeniusz R. Grela¹ · Bożena Kiczorowska¹ · Wioleta Samolińska¹ · Jan Matras¹ · Piotr Kiczorowski² · Wojciech Rybiński³ · Ewa Hanczakowska⁴

Received: 9 August 2016 / Revised: 16 November 2016 / Accepted: 19 January 2017 / Published online: 1 March 2017 © The Author(s) 2017. This article is published with open access at Springerlink.com

Abstract The aim of the study was to determine and analyse the content of essential nutrients, the amino acid profile, phytochemical compounds, and antioxidant activity in the seeds of selected species and varieties of Fabaceae plants. The highest total protein content was detected in the seeds of the yellow and Andean lupines. The lupine seeds were characterised by a high (P < 0.05) level of crude fibre, NDF, and CEL. The highest proportion of EAAI (77%) was noted in the protein of chickpea, broad bean, grasspea, and pea. Among the Fabaceae plants analysed, chickpea seeds exhibited the highest (P < 0.05) levels of total phenols and polyphenols. The greatest (P < 0.05) antioxidant activity (DPPH) was noted for the lentil and yellow lupine seeds. Compared to the other legume representatives, the importance of lupine is increasing, as the chemical composition of its seeds makes the plant an important nutraceutical component.

Keywords Leguminous seeds · Basic nutrients · Amino acids · Anti-nutritional factors · Antioxidant activity

Bożena Kiczorowska bozena.kiczorowska@up.lublin.pl

- ¹ Department of Bromatology and Food Physiology, Institute of Animal Nutrition and Bromatology, Akademicka Street 13, University of Life Science, 20-950 Lublin, Poland
- ² Department of Biological Bases of Food and Feed Technologies, University of Life Sciences in Lublin, Głęboka 28, 20-612 Lublin, Poland
- ³ Institute of Plant Genetics, Polish Academy of Sciences, Poznań, Poland
- ⁴ National Research Institute of Animal Production, Kraków, Poland

Introduction

Currently, the proportion of legume seeds in the daily diet has declined in favour of meat and highly processed products. An imbalance in these proportions in the diet has a negative impact on human health. Therefore, nutritionists recommend a return to the consumption of legume seeds. The year 2016 has been declared as the International Year of Pulses (IYP) by the UNO to promote the use of legumes in human nutrition [1]. Due to their high nutritional values, these plants are widely used as the main source of protein, in particular in the vegan diet. Besides protein, these seeds provide many other important components, i.e. minerals and B-complex vitamins, and have other vital health protective compounds (phenolics, inositol phosphates, and oligosaccharides). Their attribute is the low glycaemic index, therefore they may be recommended for people suffering from diabetes. Leguminous plants comprise many species, whose seeds differ considerably in their chemical composition and nutritive value [2]. Besides nutrients, leguminous seeds contain undesirable biologically active substances, including anti-nutritional factors (ANFs)-tannins, alkaloids, glucosides, protease inhibitors, lectins, and others. They can impair food digestion and worsen the food flavour; additionally, those with toxic properties may aggravate the mineral health status. Besides the ANFs, legume seeds contain phytoestrogens, which possess antioxidant properties and diminishes the risk of cancer [3].

The aim of the study was to determine and analyse the content of essential nutrients, the amino acid profile, phytochemical compounds, and antioxidant activity in the seeds of selected species and varieties of leguminous plants.

Materials and methods

Basic composition

The studied general-utility and consumption seeds of legumes were obtained in amounts of ~5 kg from cultivation areas of South and East Poland in 2014. A random sample was taken after harvesting for chemical analysis. The content of dry matter and basic nutrients in ground seed samples (250 g of each accession) was determined according to standard AOAC [4] procedures. Fibre fractions, i.e. neutral-detergent (NDF) and acidic-detergent fibre (ADF), were assayed according to the method by Goering and Van Soest [5]. The measurements were performed using an Ankom220 Fiber Analyzer (ANKOM Technology Corp., Fairport, New York, USA). The content of nitrogen-free extracts (NFE) was calculated as dry matter–crude protein–crude fat–crude fibre–crude ash.

Amino acid (AA) determination

The contents of amino acids were determined on a Sykam Amino Acid Analyzer (Laserchrom HPLC Laboratories Ltd. Inc., Rochester, UK). Prior to analysis, the samples were hydrolysed with 6 N HCl at 110 °C for 24 h. Methionine and Cys were analysed as methionine sulfone and cysteic acid after cold performic acid oxidation overnight before the hydrolysis. Tryptophan was determined after NaOH hydrolysis at 110 °C for 22 h. The EAAI was calculated as the geometric mean according to the following formula [6]:

EAAI =
$$\sqrt[n]{\left(\frac{a_1}{a_{1s}}\right) \times 100 \times \dots \times \left(\frac{a_n}{a_{ns}}\right) \times 100} = 10^{\log \text{EAAI}}$$

where a_n is the AA content in the protein tested and a_{ns} the AA content in the reference protein.

The CS values were calculated for all amino acids according to the following formula [7]:

$$\mathrm{CS} = \frac{a_n}{a_{ns}}.$$

To emphasise the value of the seed protein in terms of the content of limiting amino acids, only the CS_{Lys} and $CS_{Met+Cys}$ values are presented in the paper. The recommended levels of exogenous amino acids were as follows: Lys—7.0 g·16 g⁻¹·N, Met+Cys—5.7 g·16 g⁻¹ N, Thr— 4.7 g·16·g⁻¹ N, Ile—5.4 g·16 g⁻¹ N, Trp—1.7 g·16 g⁻¹ N, Val—6.6 g·16 g⁻¹ N, Leu—8.6 g·16 g⁻¹ N, His—2.2 g·16 g⁻¹ N, Phe+Tyr—9.3 g·16 g⁻¹ N, for the whole egg protein standard [8].

Analysis of anti-nutritional factors

The tannin content in legumes seeds was determined with the Folin–Ciocalteu reagent and pyrogallol as a standard [9]. The analyses were carried out at 760 nm in a UV–VIS spectrophotometer (Nicolet Evolution 300 LC, Thermo Fisher Scientific, Waltham, USA).

The trypsin inhibitor activity (TIA) was analysed according to PN EN ISO 14902:2005 [10], expressed as trypsin inhibitor units (TIU) per milligram of protein, and calculated from absorbance read at 410 nm (UV–VIS spectrophotometer, Nicolet Evolution 300 LC, Thermo Fisher Scientific, Waltham, USA) against a reagent blank. One TIU was defined as a value that yields an increase in absorbance of 0.01 relative to trypsin control reactions using a 10-mL assay volume.

Total phenolic content

The total phenolic content was determined using the spectrophotometric method [11]. A methanolic solution of the extract at a concentration of 1 mg/ml was used in the analysis. The reaction mixture was prepared by mixing 0.5 ml of the methanolic solution of the extract, 2.5 ml of the 10% Folin–Ciocalteu reagent dissolved in water, and 2.5 ml of 7.5% NaHCO₃. A blank was concomitantly prepared and contained 0.5 ml of methanol, 2.5 ml of the 10% Folin–Ciocalteu reagent dissolved in water, and 2.5 ml of 7.5% NaHCO₃. The samples were thereafter incubated in a thermostat at 45 °C for 45 min. The results were expressed as GAE (gallic acid equivalents).

Determination of individual polyphenols

Homogenised freeze-dried legume samples (0.5 g) were extracted with 5×5 ml methanol, in an UltraTurrax T25 (IKA.Werke, Janke & Kunkel) homogeniser. Aliquots of the methanolic legume extracts (0.1 ml) were transferred to GC vials and 50 µl of an internal standard was added (3-[4-hydroxyphenyl]-1-propanol solution, 19.2 μ g/ml); next, the sample was evaporated to dryness under nitrogen and derivatised by addition of 250 µl N, O-bis(trimethylsilyl)trifluoroacetamide (BSTFA) at 70 °C for 20 min. Analysis of the samples was performed by an Agilent (Wallborn, Germany) HP series GC 6890N coupled with a HP 5973 MS detector (EI, 70 eV), a split-splitless injector, and an HP 7683 autosampler in an HP-5 MS capillary column (5% phenyl-95% methyl siloxane, 30 m \times 0.25 mm \times 250 µm). As standard the 3-(4-hydroxyphenyl)-1-propanol was used.

DPPH radical scavenging method for measuring antioxidant activity

The method consisted in spectrophotometric measurement of the intensity of the colour change in the solution depending on the amount of the DPPH·(2,2-diphenyl-1picrylhydrazyl radical). The reaction was initiated by mixing 1 mL of the methanolic extract with 3 mL of methanol and then adding 1 mL of the DPPH·(0.012 g/100 mL). Absorbance at λ max of 517 nm (UV mini-1240 Shimadzu spectrophotometer, Fischer Bioblock, France) was checked at 0, 0.5, and at every 0.5 min until the reaction reached a stable state. This plateau was reached within 60 min. The activity of the extract in DPPH·scavenging was calculated as follows:

%DPPH scavenging = $\left[\frac{\text{absorbance of control} - \text{absorbance of sample}}{\text{absorbance of control}}\right] \times 100$

Total antioxidant capacity was also expressed as a Trolox equivalent on the basis of the standard curve ($R^2 = 0.9974$).

Statistical analysis

Analyses were performed in triplicate and all data were expressed as means. All the data of the chemical composition of the leguminous seeds were analysed with the Statistica software version 10.0 [12]. The normality of data and homogeneity of variances were tested using the Shapiro-Wilk and Brown-Forsythe tests, respectively. The data obtained were analysed statistically using species as an independent variable in the general linear model (GLM) of one-way ANOVA analysis of variance or a non-parametric Kruskal-Wallis test (a non-parametric equivalent of oneway analysis of variance). Detailed comparisons between the means of the species legume were conducted using the post hoc Duncan or Dunn test. Pearson's or Spearman's correlation tests were conducted to determine the linear correlations among some variables. All statements of significance were based on the 0.05 and 0.01 probability levels.

Results and discussion

The analysed seeds were characterised by high dry matter (DM) content, i.e. over 900 g in 1 kg (Table 1). The analysed seeds exhibited a significant (P < 0.05) variation of the crude ash content ranging from ca. 44.3 g kg⁻¹ DM in the white and yellow lupine and grasspea to even 25.3 g kg⁻¹ DM in the pea (Pomorski cv.). The seeds of the legume plants were characterised by high protein content, in particular the yellow and Andean lupine seeds (427.4 g kg⁻¹ DM). The high content of this valuable nutrient, especially in the yellow lupine, was reported by Sujak et al. [13]. The authors report that some cultivars of the species may contain as much as $482 \text{ g kg}^{-1} \text{ DM}$ (Kroton cv.) or 471 g kg⁻¹ DM (Markiz cv.) of protein. The lowest (P < 0.05) total protein content in the analysed seeds was found for the common bean, pea, chickpea, and broad bean seeds (219 g kg^{-1} DM on average). Similar total protein levels in these leguminous species were reported by other researchers [2, 14]. The nutritional role of these plants in the human organism is determined not only by the content of protein, but also its structure and functions. Two protein fractions, albumins and globulins, are distinguished. Albumins account for ca. 10-25% of total protein and serve a structural and enzymatic function in the human organism. Higher albumin levels contribute to a greater nutritional value of seeds. In turn, globulins are mainly regarded as immune proteins [15]. Globulins can account for even 60–75% of proteins in pea, soya, and lupine seeds and 80–90% in common and broad bean seeds [16].

The fat content ranged (P < 0.05) from 7.91 to 23.81 g kg⁻¹ DM in the seeds of a majority of the species analysed; higher levels of fat, i.e. 53 g kg⁻¹ DM on average, were detected in the seeds of the blue and yellow lupines and chickpea. In turn, the white lupine (100.3 g kg⁻¹ DM) and the Andean lupine (155 g kg⁻¹ DM) proved to be oilbearing species. Similar ranges of the content of this nutrient in Fabaceae plant seeds were reported in literature [2, 13, 14, 17].

Nutritionists emphasise the need for consumption of leguminous plant seeds, due to their content of various dietary fibre fractions that are essential for the digestion process and actively counteract lifestyle and cancer diseases. Investigations conducted by Farvid et al. [18] support the hypothesis that a higher fibre intake reduces the risk of breast cancer and suggest that consumption of large amounts of insoluble dietary fibre during the adolescence and early adulthood periods may have a preventive character. Especially high nutritional-therapeutic importance in this aspect is ascribed to lupine seeds, which are characterised by a high (P < 0.05) proportion of crude fibre (on average 117 g kg⁻¹ DM), in contrast to the chickpea and lentil seeds, which contained by ca. 78% lower levels of this nutrient (Table 1). The lupines and beans analysed exhibited a large level of the NDF fraction (on average 228 g kg^{-1} DM) and CEL (on average 153 g kg⁻¹ DM). Literature data show that the fibre content in seeds may differ significantly in different species and their varieties [19]. A special health-enhancing role is played by hardly soluble dietary fibre fractions, as they, e.g. improve the parameters of large intestine physiological status. They exhibit an ability to retain water, which directly increases the stool volume, thereby eliminating secondary bile acids and other toxic substances from the

Table 1 Content of basic nutrients, detergent fibre, and anti-nutritional factors in some legume species ($g kg^{-1} DM$)

Species	DM g kg ⁻¹	Crude ash	Crude protein	Ether extract	Crude fibre	NDF	ADF	ADL	HCEL	CEL
Blue lupine										
cv. Regent	933.2	35.47	319.2	67.31	116.4	209.3	172.1	12.21	37.20	159.89
cv. Zeus	918.5	29.15	398.4	46.78	94.45	245.4	209.1	15.13	36.30	193.97
Mean value	925.8	32.31 ^b	358.8 ^b	57.04 ^c	105.4 ^b	227.3 ^a	190.6 ^a	13.67 ^c	36.70 ^g	176.93 ^a
White lupine										
cv. Butan	939.7	39.06	355.2	101.9	117.1	214.4	138.2	19.14	76.20	119.06
cv. Bardo	915.4	46.52	336.7	98.74	135.4	237.8	164.4	16.42	73.40	147.98
Mean value	927.5	42.79 ^a	345.9 ^b	100.3 ^b	126.2 ^a	226.1 ^a	151.3 ^b	17.78 ^b	74.80 ^c	133.52 ^b
Yellow lupine										
cv. Parys	938.9	45.80	428.4	55.28	118.4	232.2	179.1	13.43	53.10	165.67
cv. Iryd	908.1	42.38	454.2	40.18	121.5	256.7	213.3	15.31	43.40	197.99
Mean value	923.5	44.09 ^a	441.3 ^a	47.73 ^c	119.9 ^a	244.4 ^a	196.2 ^a	14.37 ^c	48.20^{f}	181.83 ^a
Andean lupine										
Mean value	922.6	37.45 ^{ab}	413.5 ^a	155.1 ^a	73.84 ^c	178.8 ^b	127.1 ^c	10.52 ^d	51.70 ^e	116.58 ^c
Pea										
cv. Medal	926.4	29.58	237.1	12.52	48.25	176.1	85.01	7.53	91.09	77.48
cv. Mentor	901.2	25.34	201.5	9.78	43.18	101.4	73.44	4.84	27.96	68.60
Mean value	913.8	27.46 ^c	219.3 ^{cd}	11.15 ^e	45.71 ^d	138.7 ^{bc}	79.23 ^d	6.18 ^e	59.48 ^d	73.04 ^d
Chickpea										
Mean value	929.8	31.30 ^b	226.2 ^c	54.85 ^c	22.16 ^f	168.3 ^b	75.72 ^d	11.43 ^d	92.58 ^b	64.29 ^e
Lentil										
Mean value	924.3	28.13 ^c	268.9 ^c	7.91 ^e	31.05 ^e	89.52 ^c	53.65 ^e	4.81^{f}	35.87 ^g	48.84^{f}
Grasspea										
cv. Derek	925.5	31.66	276.9	9.72	45.49	148.5	68.21	9.64	80.29	58.57
cv. Krab	903.2	55.69	312.4	8.24	44.19	163.7	71.63	6.62	92.07	65.01
Mean value	914.35	43.67 ^a	294.65 ^{bc}	8.98 ^e	44.84 ^d	156.1 ^b	69.92 ^d	8.13 ^e	86.18 ^b	61.79 ^e
Common bean										
cv. Mela	901.2	41.82	194.5	23.81	44.34	216.8	163.4	0.13	53.40	163.3
Mean value	901.2	41.82 ^a	194.5 ^d	23.81 ^d	44.34 ^d	216.8 ^{ab}	163.4 ^b	0.13 ^g	53.40 ^d	163.3 ^a
Broad bean										
cv. Bonus	900.1	31.81	236.4	9.39	62.81	236.9	112.3	96.31	124.60	15.99
Mean value	900.1	31.81 ^b	236.4 ^c	9.39 ^e	62.81 ^d	236.9 ^a	112.3 ^c	96.31 ^a	124.60 ^a	15.99 ^f
Pool SEM	8.79	0.39	2.84	0.67	0.93	1.46	2.67	0.12	0.45	2.03
P value	0.152	0.0.37	0.024	0.016	0.037	0.019	0.048	0.025	0.033	0.026

CEL cellulose (calculated values: ADF-ADL), ADF acid-detergent fiber, NDF neutral-detergent fiber, ADL lignin, HCEL hemicellulose (calculated values: NDF-ADF)

^{a, b, c, d, e, f}Statistical differences between mean values (P < 0.05)

organism. Fibre fractions in lupine seeds not only can regulate the intestinal pH and moisture levels, but also have a beneficial effect on the morphometric parameters of the colon, e.g. the length of plica mucosae and the thickness of the muscular layer. Soluble fibre fractions and α -galactosides exhibit considerable proliferative activity due to the rapid fermentation processes [20]. These processes occurring within reasonable limits are beneficial to health, as they contribute to stool loosening and regular defecation. The high fibre content in legume seeds ensures their basificating properties, which balance the acidifying effects of meat, eggs, and cereal products. The combination of legume seeds with meat, eggs, or cheese increases the digestibility of proteins contained therein [21].

The amino acid profiles of crude protein and the values of their indexes are compiled in Table 2. Noteworthy, there are differences in the contents of both endo- and exogenous amino acids between the species as well as within the varieties. The greatest amount of essential amino

Table 2Endogenous aminoacid profile of legume species

 $(g \cdot 16 g^{-1} N)$

Species	Ala	Arg	Asp	Glu	Gly	Ser	Pro	ΣΝΕΑΑ
Blue lupine								
cv. Regent	3.59	8.73	14.74	16.85	3.79	4.97	4.40	57.07
cv. Zeus	3.08	9.28	9.15	18.34	3.51	3.58	3.86	50.80
Mean value	3.34	9.01	11.95	17.60 ^b	3.65	4.28	4.13	53.94
White lupine								
cv. Butan	3.71	8.96	12.20	18.54	3.83	5.60	3.74	56.60
cv. Bardo	3.25	9.54	9.67	19.96	3.66	4.93	3.37	54.38
Mean value	3.48	9.25	10.94	19.25 ^{ab}	3.75	5.27	3.56	55.49
Yellow lupine								
cv. Parys	3.83	9.87	11.26	21.78	4.03	5.40	4.18	60.34
cv. Iryd	4.83	9.21	8.83	20.68	5.31	4.63	1.59	55.08
Mean value	4.33	9.54	10.05	21.23 ^a	4.67	5.02	2.89	57.71
Andean lupine								
Mean value	3.98	9.47	10.88	21.68 ^a	3.95	5.36	3.93	59.24
Pea								
cv. Medal	4.46	8.29	12.66	17.30	4.17	5.05	4.18	56.12
cv. Mentor	3.56	7.89	8.79	16.54	3.86	4.53	3.45	48.62
Mean value	4.01	8.09	10.73	16.92 ^b	4.02	4.79	3.82	52.37
Chickpea								
Mean value	4.90	8.30	12.60	16.50 ^b	4.11	5.56	4.50	56.48
Lentil								
Mean value	3.77	9.60	12.91	18.30 ^b	3.80	5.19	4.10	57.67
Grasspea								
cv. Derek	4.99	9.33	11.55	17.32	4.37	4.92	4.30	56.78
cv. Krab	5.78	6.75	10.27	17.89	6.07	5.10	2.15	54.01
Mean value	5.39	8.04	10.91	17.61 ^b	5.22	5.01	3.23	55.40
Common bean								
cv. Mela	3.45	7.59	9.25	16.15	4.59	4.13	3.39	48.55
Mean value	3.45	7.59	9.25	16.15 ^b	4.59	4.13	3.39	48.55
Broad bean								
cv. Bonus	3.84	8.57	10.73	16.69	4.56	4.96	3.86	53.21
Mean value	3.84	8.57	10.73	16.69 ^b	4.56	4.96	3.86	53.21
Pooled SEM	0.196	0.224	0.454	0.487	0.175	0.139	0.211	0.904
P value	0.160	0.568	0.968	0.016	0.549	0.622	0.950	0.610

NEAA non-essential amino acids

^{a, b, c, d, e, f}Statistical differences between mean values (P < 0.05)

acids was noted in grass pea cv. Krab, in chickpea, broad bean, and pea (over 40 g·16 g⁻¹ N) (Table 3). However, the differences in the mean levels of essential amino acids between the legume species were not confirmed statistically (P=0.198), except for the glutamine content with the highest proportion in the protein of the Andean and yellow lupine seeds, in comparison with the lentil, grasspea, blue lupine, pea, broad bean, chickpea, and common bean seeds (P < 0.05). The highest CS_{met+cys} index was noted for crude protein of the yellow lupine (54%) and grasspea (52%) and the lowest for the seeds of common bean (30%), blue lupine (37%), and white lupine (39%). The highest EAAI (77%) was detected in the protein of chickpea as well as the protein of the broad bean (76%), grasspea (75%), and pea (74%), whereas the lowest values of this index were noted for the common bean and lupine protein (62–70%). Similarly, in the case of the indicators of the biological value of the protein, the statistical analysis did not confirm the differences between the Fabaceae species. In the analysed seeds of the legume plants (lupines, chickpea, pea, lentil, grasspea, broad and common bean), the content of glutamine and asparagine was the highest of all the non-essential amino acids (NEAA). Similar results were obtained by Iqbal et al. [22] in their investigations of the chemical composition and nutritional value of chickpea, cowpea, lentil,

and green pea seeds as well as Khattab et al. [23] in their analyses of cowpea, pea, and kidney bean seeds.

The protein of Fabaceae seeds is characterised by a lower value than that of animal protein and a higher value than that of cereal protein [23]. This was confirmed in the present study by the high content of exogenous amino acids and the high biological value of the legume proteins. In this study, the content of essential amino acids (EAA) ranged from 41 g 16 g^{-1} N in chickpea to 34 $g \cdot 16 g^{-1}$ N in common bean seeds, compared with the value of 51 g \cdot 16 g $^{-1}$ N in hen's egg white used in the assessment as an amino acid standard [8]. The Pearson correlation analysis revealed a high positive (P < 0.05) correlation between EAAI and the protein content in the analysed Fabaceae seeds (Table 5). Similar results were obtained in the investigations conducted by Khattab et al. [23], where the total essential amino acid content in different seeds ranged from 31.29 g-16 g⁻¹ N in the Egyptian pea to 36.12 g-16 g⁻¹ N in the Canadian kidney bean. In the present study, the EAAI value ranged from 77% for the chickpea to 62% to the bean, which was reflected in the CS_{Met+Cvs} value. The lowest value of this parameter, i.e. 30%, was estimated for the bean, while the yellow lupine exhibited the highest value (54%). The content of cystine and methionine in the lupine and grasspea seeds was strongly positively correlated with the protein content (P < 0.05) (Table 5). The available literature shows a high proportion of lysine and a relatively low level of sulphur amino acids and tryptophan in legume plant seeds [22, 23]. This was largely confirmed by the present results, as the $\mbox{CS}_{\mbox{Lys}}$ value for all the analysed Fabaceae seeds was high (on average ca. 87%), and sulphur amino acids were the first limiting amino acids, with the exception of the Andean lupine, lentil, and yellow lupine protein, where tryptophan was the first limiting amino acid (20, 29, and 38% CS_{trp}, respectively).

Anti-nutritional substances comprise compounds that significantly diminish the nutritional value of leguminous plants, e.g. chymotrypsin and trypsin inhibitors (Table 4). Their negative impact is reflected not only in deterioration of protein digestibility through complexation thereof, but also in blocking the activity of proteolytic enzymes. This may result in pancreas hypertrophy and, due to intensified enzyme secretion, an increased demand of the organism for amino acids, in particular for methionine and cysteine [24]. The strong relationships between protein and trypsin inhibitors are confirmed by the high values of correlation coefficients noted for nearly all legume seeds (Table 5). Yet, their effect on the organism may be positive in some circumstances. Trypsin inhibitors are applied in treatment of obesity, type 2 diabetes, and cancer disease as well as prevention of infarction and cardiovascular diseases [25]. The greatest content of these compounds was determined in the grasspea and pea seeds (on average 129.3 and 47.06 TUI⁻¹ mg), and the lowest levels were noted for the yellow lupine and common bean seeds (on average 1.64 and 2.62 TUI⁻¹ mg) (Table 4). A similar variability in the content of these substances was reported by other researchers [26]. However, despite the relatively small content of protease inhibitors in the lupine seeds, recent studies have suggested their unusual properties [27]. They contain anticancer MMP-9 inhibitors, which are the most effective of those present in leguminous plants, inhibiting both gelatinases and HT29 migration and growth. Some authors suggest possible application of legume protein MMPIs as novel metalloproteinase inhibitors in anti-cancer pharmacological and nutritional strategies.

Tannins present in legume seeds are classified as antinutritional compounds. Among the analysed cultivars, the highest tannin content was detected in the old variety of Andean lupine (16.34 g kg⁻¹ DM), whereas the new blue and yellow lupine varieties had by ca. 70% lower content of these compounds (Table 4). The lowest levels of hydrolysable tannins, i.e. 2.58 g kg⁻¹ DM, were detected in the white lupine seeds. Among the other species analysed, only the lentil seeds exhibited a relatively high level of these compounds, i.e. 8.72 g kg⁻¹ DM. A strong correlation (P < 0.05) was observed in this study between the tannin content and protein level in the blue lupine, yellow lupine, Andean lupine, lentil, grasspea, and broad bean (+) and in the white lupine and pea (-) (Table 5). Similar amounts of these anti-nutritional compounds in legume seeds were reported by Księżak and Bojarszczuk [28] in their investigations covering a very large area of Poland. They did not observe a significant effect of the site of cultivation on the tannin level, but correlated it with the species and cultivar. The present study shows a high correlation between tannins and protein. Tannins are polyphenolic compounds capable of formation of stable complexes with proteins, minerals, and vitamins A and B₁₂, which results in deterioration of the digestibility and availability of nutrients. This harmful effect of tannins is alleviated by the thermal treatment of seeds [29].

Fabaceae plant seeds are increasingly referred to as a source of antioxidants. Besides tannins, they contain phenolic compounds with important antioxidant activity, e.g. quercetin glycosides, kaempferol, and myricetins as well as caffeic, ferulic, and *p*-coumaric acids. They are mainly present in seed coats, and seeds of varieties with colourful flowers contain higher amounts of these compounds. These substances are believed to protect against cancer and cardiovascular diseases [30]. The beneficial effect of polyphenols involves removal of existing reactive oxygen species from blood as well as prevention of ROS formation. Polyphenols can also inhibit oxidation of endogenous antioxidants in the oxidation process, e.g. querce-tin may prevent oxidation of vitamin C [31]. Among the

\mathbf{z}
ad
$\cdot 16$
<u>છ</u>
lues
va
nal
iti
nutı
their
s and
e
peci
le s
ung
f leg
e of
offle
pre
acid
10
imi
al
enti
Ess
ິ
Table
Tal

Species	Essential AA	I AA											INULTION	Nutritional values		
	His	Val	Ile	Leu	Lys	Met	Cys	Met+Cys	Phe	Tyr	Thr	Trp	ΣEAA	$\mathrm{CS}_{\mathrm{Lys}}$	CS _{Met+Cys}	EAAI
Blue lupine															-	
cv. Regent	2.74	4.40	3.42	7.05	6.81	1.37	1.10	2.48	4.03	2.79	4.03	0.74	38.49	76	43	72
cv. Zeus	2.92	3.81	3.75	6.53	5.09	0.74	1.02	1.76	3.56	3.54	3.22	0.68	34.86	73	31	64
Mean value	2.83^{ab}	4.11 ^b	3.59^{ab}	6.79^{b}	5.95 ^{ab}	1.06^{b}	1.06^{d}	2.12	3.80	3.17^{ab}	3.63^{b}	$0.71^{\rm bc}$	36.68	85	37	68
White lupine																
cv. Butan	2.71	3.57	3.06	7.20	5.80	0.68	1.12	1.80	3.33	2.79	3.83	0.65	34.72	83	32	64
cv. Bardo	2.26	4.09	2.83	7.38	5.09	0.87	1.69	2.56	3.98	3.68	3.69	0.78	36.34	73	45	67
Mean value	2.49^{b}	3.83^{b}	2.95^{b}	7.29^{ab}	5.45 ^b	0.78°	1.41 ^{bc}	2.18 ^c	3.66^{b}	3.24^{a}	3.76^{ab}	0.72^{bc}	35.53	78	38.5	65.5
Yellow lupine																
cv. Parys	2.93	3.70	3.36	7.51	5.57	0.92	1.75	2.67	3.78	2.56	4.00	0.71	36.79	80	47	69
cv. Iryd	2.91	4.38	3.84	8.15	4.76	1.34	2.16	3.50	3.84	2.71	3.19	0.59	37.87	68	61	70
Mean value	2.92^{a}	4.04 ^b	3.60^{ab}	7.83 ^b	$5.17^{\rm b}$	1.13^{b}	1.96^{a}	3.09^{a}	3.81^{ab}	$2.64^{\rm bc}$	3.60^{b}	0.65°	37.33	74	54	69.5
Andean lupine																
Mean value	3.03^{a}	3.67^{b}	3.90^{a}	$6.77^{\rm bc}$	6.1^{ab}	0.85°	1.64^{b}	2.50^{b}	3.57^{b}	3.57^{a}	4.10^{a}	0.38^{d}	37.61	88	44	99
Pea																
cv. Medal	2.65	4.41	3.65	6.97	7.10	1.40	1.31	2.71	4.52	2.69	4.37	1.06	40.13	101	48	LL
cv. Pomorski	2.23	4.23	4.31	6.29	6.23	0.95	1.09	2.04	4.36	2.39	3.98	0.98	37.04	89	36	70
Mean value	2.44^{b}	4.32^{ab}	3.98^{a}	$6.63^{\rm bc}$	6.67^{a}	1.18^{ab}	1.20°	2.38^{b}	4.44^{a}	2.54°	4.18^{a}	1.02^{a}	38.59	95	42	73.5
Chickpea																
Mean value	2.91^{a}	4.61 ^a	3.90^{a}	7.47^{ab}	6.60^{a}	1.10^{b}	1.55 ^b	2.65 ^b	4.60^{a}	2.49 ^c	4.48^{a}	0.84^{b}	40.55	94	46	LL
Lentil																
Mean value	2.88^{ab}	4.63 ^a	3.49^{ab}	7.08^{ab}	6.92^{a}	1.08^{b}	1.50^{b}	2.58 ^b	4.71 ^a	2.40°	4.42^{a}	0.49^{d}	39.59	66	45	71
Grasspea																
cv. Derek	3.07	4.84	3.74	6.98	6.20	1.07	1.40	2.47	4.10	2.82	4.06	0.98	39.27	89	43	76
cv. Krab	3.06	3.89	4.39	7.69	60.9	1.56	1.94	3.50	4.02	4.21	4.19	0.45	41.49	87	61	74
Mean value	3.07^{a}	4.37^{a}	4.07^{a}	7.34^{b}	6.15^{ab}	1.32^{a}	$1.67^{\rm b}$	2.99^{a}	4.06^{ab}	3.52 ^a	4.13^{a}	0.72^{bc}	40.38	88	52	75
Common bean																
cv. Mela	2.42	4.21	3.72	6.44	5.46	0.69	1.03	1.72	2.61	2.56	3.89	0.61	33.64	78	30	62
Mean value	2.42	4.21 ^{ab}	3.72^{a}	6.44°	5.46^{b}	0.69°	1.03^{d}	1.72 ^d	2.61°	2.56^{bc}	3.89^{ab}	0.61°	33.64	78	30	62
Broad bean																
cv. Albus	3.05	4.79	3.54	8.53	6.33	0.74	1.98	2.72	3.94	2.79	3.89	0.84	40.42	90	48	76
Mean value	3.05^{a}	4.79 ^a	3.54^{ab}	8.53^{a}	6.33^{ab}	0.74 ^c	1.98^{a}	2.72^{ab}	3.94^{ab}	2.79^{b}	3.89^{ab}	0.84^{b}	40.42	90	48	76
Pooled SEM	0.134	0.175	0.121	0.297	0.173	0.132	0.119	0.084	0.164	0.127	0.203	0.347	0.611	0.978	0.734	0.695
P value	0.027	0.038	0.017	0.016	0.049	0.022	0.036	0.025	0.048	0.027	0.016	0.034	0.198	0.341	0.127	0.279

Table 4 Content of anti-nutritional factors and antioxidant activity in some legume species (g kg⁻¹ DM)

Species	Inhibitors trypsin	Tannins	Total phenols ¹	Polypheno	ls µg/100 g DM	A		DPPH ²
	TIU mg ⁻¹ protein	g kg ⁻¹ DM	mg/100 g DM	Quercetin	Caffeic acid	Ferulic acid	<i>p</i> -Coumaric acid	(mmol trolox equivalents/g DM)
Blue lupine								
cv. Regent	3.28	4.93	313.7	47.62	78.64	88.52	41.84	7.91
cv. Zeus	3.45	5.12	294.2	45.69	63.43	84.75	32.56	7.42
Mean value	3.36 ^f	5.02 ^d	304.0 ^c	46.65 ^b	71.03 ^b	86.63 ^b	37.2 ^d	7.66 ^b
White lupine								
cv. Butan	4.17	2.54	289.3	43.53	63.54	86.34	18.73	3.73
cv. Bardo	4.28	2.62	278.4	39.41	45.83	87.16	15.38	6.34
Mean value	4.23 ^f	2.58 ^g	283.9 ^c	41.47 ^b	54.69 ^c	86.75 ^b	17.06	5.04 ^c
Yellow lupine								
cv. Parys	1.52	4.63	328.9	45.63	123.4	83.43	53.84	8.75
cv. Iryd	1.75	4.96	273.5	41.27	116.8	81.64	47.96	7.64
Mean value	1.64 ^h	4.80 ^e	301.2 ^c	43.45 ^b	120.1 ^a	82.54 ^b	50.90 ^c	8.20 ^a
Andean lupine								
Mean value	3.94^{f}	16.34 ^b	297.8 ^c	48.64 ^b	59.81 ^c	76.41 ^c	44.91 ^{cd}	5.31 ^c
Pea								
cv. Medal	42.35	4.16	225.6	nd	32.64	56.83	45.67	2.89
cv. Mentor	51.76	5.21	189.7	nd	30.86	52.74	46.93	2.56
Mean value	47.06 ^b	4.69 ^e	207.6 ^d	nd	31.75 ^d	54.79 ^d	46.30 ^c	2.73 ^e
Chickpea								
Mean value	11.23 ^d	3.78^{f}	459.7 ^a	101.5 ^a	126.4 ^a	134.7 ^a	90.72 ^a	2.97 ^e
Lentil								
Mean value	31.47 ^c	8.72 ^c	398.3 ^b	24.53 ^c	36.86 ^d	29.75 ^f	72.93 ^b	8.13 ^a
Grasspea								
cv. Derek	123.5	3.54	249.7	nd	27.61	_	67.94	4.69
cv. Krab	135.2	3.86	294.1	nd	26.43	_	66.38	3.09
Mean value	129.3 ^a	3.70 ^f	271.9 ^c	nd	27.02 ^e	_	67.16 ^b	3.89 ^d
Common bean								
cv. Mela	2.62	1.53	418.4	1.52	19.83	26.28	21.97 ^e	5.77
Mean value	2.62 ^g	1.53 ^h	418.4 ^a	1.52 ^d	19.83 ^f	26.28^{f}	21.97	5.77 ^c
Broad bean								
cv. Bonus	5.67	2.16	366.7	nd	27.34	41.96	32.34	7.33
Mean value	5.67 ^e	2.16 ^g	366.7 ^b	nd	27.34 ^e	41.96 ^e	32.34 ^d	7.33 ^b
Pool SEM	0.16	0.38	4.56	0.27	0.97	0.54	0.19	0.06
P value	0.037	0.028	0.025	0.019	0.043	0.037	0.014	0.018

DPPH free radical scavenging capacity

^{a, b, c, d, e, f}Statistical differences between mean values (P < 0.05)

¹mg gallic acid equivalents/g

Fabaceae seeds analysed, the chickpea and common bean seeds proved to have the highest levels of total phenols (on average 439 mg 100 g⁻¹DM), and slightly lower contents of these compounds were determined in the broad bean and lentil seeds (on average 400 mg 100 g⁻¹DM) (Table 4). The lowest amounts, i.e. lower by 50%, were noted in the pea seeds. The chickpea seeds were characterised by the highest levels of polyphenols (P < 0.05). Approximately twofold lower levels of polyphenols were

determined in the seeds of the lupine varieties, with the yellow lupine characterised by the highest content of caffeic acid and *p*-coumaric acid. Similar results were obtained by Siger et al. [32], who reported the highest level of these compounds in yellow lupine seeds (in particular in the Paris cv.). Furthermore, the authors emphasised that the antioxidant apigenin-6,8-di-Cp-glucopyranoside and apigenin 7-O- β -Apium furanosyl-6,8-di-Cp-glucopyranoside discovered for the first time in

Table 5Correlation coefficients between the measured characteristics in legume seeds	coefficients betwe	en the measured chi	aracteristics in legum	ne seeds						
Variables	Blue lupine	White lupine	Yellow lupine	Andean lupine	Pea	Chickpea	Lentil	Grasspea	Common bean	Broad bean
Protein										
Met+Cys	0.567*	0.598*	0.618*	0.536^{*}	0.283	0.378	0.435	0.503*	0.369	0.416
EAAI	0.697*	0.674^{*}	0.524*	0.534^{*}	0.667*	0.571^{*}	0.628*	0.689*	0.653*	0.623^{*}
Inhibitors trypsin	0.736^{*}	-0.545*	0.616*	0.584^{*}	-0.597*	0.593*	0.495*	0.423*	0.387	0.678*
Tannins	0.587*	-0.451*	0.598*	0.578*	-0.535*	0.462	0.567*	0.597*	0.212	0.808*
DPPH										
Tannins	-0.574*	-0.364	-0.525*	-0.578*	-0.407*	-0.412	-0.374	-0.437*	-0.143	-0.509
Quercetin	0.485	-0.576^{*}	0.578*	0.678*	Ι	0.577*	0.589*	I	0.548^{*}	0.415
Caffeic acid	0.433	-0.489	0.467	0.378	0.385	0.451	0.247	0.389	0.267	0.378
Ferulic acid	0.587*	0.556^{*}	0.637*	0.645*	0.674^{*}	0.614^{*}	0.648*	I	0.649*	0.576^{*}
<i>p</i> - Coumaric acid	0.145	-0.278	0.156	0.234	-0.203	0.271	0.154	0.374	0.245	0.421
Total phenols	0.193	0.578*	0.345	0.457	0.646^{*}	0.695*	0.378	0.278	0.348	0.247
*Statistical differences ($P < 0.05$)	s (P < 0.05)									

Deringer

Eur Food Res Technol (2017) 243:1385–1395

lupine seeds further enhanced the nutritional value of this plant not only as a high-protein raw material, but also as a rich source of antioxidants. A considerable variation in the content of antioxidant compounds in Fabaceae seeds has been reported by many authors [21, 30, 33, 34].

The antioxidant properties of the legume seeds were determined by the ability to scavenge the DPPH radical. The highest (P < 0.05) antioxidant activity (DPPH) was determined for the lentil and yellow lupine seeds (on average 8.16 mmol trolox equivalents g DM^{-1}). In the broad bean, blue, white, and Andean lupines as well as common bean, DPPH exhibited a level of ca. 7-5 mmol Trolox equivalents g DM-1. The lowest DPPH value was determined in the grasspea, chickpea, and pea seeds (respectively, 3.89, 2.97, 2.73 mmol Trolox equivalents g DM^{-1}) (Table 4). In the present study, there were correlations between DPPH and each polyphenol (Table 5). In the case of quercetin, a high correlation (P < 0.05) was noted for the white lupine (-), yellow lupine, Andean lupine, pea, chickpea, and common bean. In turn, a high correlation (P < 0.05) was found between total phenols and DPPH for the seeds of the white lupine, pea, and chickpea. There are literature reports of a moderate or even low correlation between phenols and DPPH measured with the DPPH and ORAC methods in lentil seeds [34]. However, some authors report that the activity measured with the FRAP assay exhibited a strong correlation (over 0.8) with phenolic compounds and condensed tannins. Li at al. [35] explain that this phenomenon can also be observed in other food products, which is related to the fact that phenolic compounds may have not only antioxidant properties. Potent antioxidant activity, particularly in relation to linoleic acid, was exhibited by aqueous and ethanol extracts of lentil, pea, lupine, or bean husks. Simultaneously, these preparations have been observed to have anti-inflammatory activity and a capability of selective 15-LOX inhibition, moderate COX-1 and COX 2 inhibition, as well as cyclooxygenase pathway inhibition [36]. Given these properties, Fabaceae seeds can be regarded as a source of high-value nutrients, especially of protein, and can be useful in enhancement of human health and reduction of the risk of incidence of lifestyle diseases, i.e. cancer, degenerative diseases, obesity, and diabetes.

Conclusions

Given their unique and very rich chemical composition and high nutritional value, legume plant seeds can be an important component of the daily diet. However, the content of biologically active substances that have so far been regarded as inessential or even anti-nutritional can promote the suitability of the seeds to be used as functional, health-enhancing foodstuffs, particularly for prevention of diabetes and obesity, which determine the quality of life for millions of people worldwide. In this aspect, lupine seeds, which have so far been used mainly as animal feed and are currently gaining importance as a nutraceutical component, should be given special attention.

Compliance with ethical standards

Conflict of interest None.

Compliance with ethics requirements This article does not contain any studies with human or animal subjects.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

- United Nations (2014) Resolution adopted by the General Assembly on 20 December 2013, 68/231. International Year of Pulses 2016. A/RES/68/231.http://www.fao.org/pulses-2016/en/
- Grela E R, Günter KD (1995) Fatty acid composition and tocopherol content of some legume seeds. Anim Feed Sci Technol 52(3):325–331
- Gemede HF, Ratta N (2014) Antinutritional factors in plant foods: potential health benefits and adverse effects. Int J Nutr Food Sci 3(4):284–289
- AOAC (2011) Official methods of analysis of AOAC International. 18th Edition, Revision 4, In: Horwitz W, Latimer G.W. Jr. (eds) AOAC International. Gaithersburg, Maryland, USA
- Goering HK, Van Soest PJ (1970) Forage fiber analysis. USDA Agriculture Handbook 379–381
- Oser BL (1959) An integrated essential amino acid index for predicting the biological value of proteins. Protein Amino Acid Nutr 281–289
- Block RJ, Mitchell HH (1946) The correlation of the amino acid composition of proteins with their nutritive value. Nutr Abstracts Rev 16(2):249–278
- Hidvegi M, Bekes F (1984) Mathematical modeling of protein Nutritional Quality from Amino Acid Composition. In: Lazity R, Hidvegi M (eds) Proceedings of the International Association for Cereal Chemistry Symposium, Academic Kiado. Budapest, pp 205–286
- 9. European Pharmacopoeia (2014) 8th edition, 2.8.14. Tannins in herbal drugs
- PN EN ISO 14902:2005. Animal Feeding Stuffs Determination of trypsin inhibitor activity of soya products. Polish Committee for Standardization
- Singleton VL, Rossi JA (1965) Colorimetry of total phenolics with phosphotungstic-phosphomolybdic acid reagents. Am J Enol Viticult 16:144–158

- 12. StatSoft, Inc. Tulsa, OK, USA. STATISTICA (data analysis software system), version 10. http://www.statsoft.com. 2011
- Sujak A, Kotlarz A, Strobel W (2006) Compositional and nutritional evaluation of several lupin seeds. Food Chem 98:711–719
- Hanczakowska E, Świątkiewicz M (2013) Legume seeds and rapeseed press cake as substitutes for soybean meal in sow and piglet feed. Agric Food Sci 22(4):435–444
- Rubio LA, Pérez A, Ruiz R, Guzman M, Aranda-Olmedo I, Clemente A (2014) Characterization of pea (*Pisum sativum*) seed protein fractions. J Sci Food Agric 94(2):280–287
- Dziuba J, Szerszunowicz I, Nałęcz D, Dziuba M (2014) Proteomic analysis of albumin and globulin fractions of pea (*Pisum sativum* L.) seeds. Acta Scient Pol Technol Aliment 13:181–190
- Kiczorowska B, Samolińska W, Andrejko D (2016) Effect of micronized pea seeds (*Pisum sativum L.*) as a substitute of soybean meal on blood lipid parameters, tissue fatty acids composition and meat quality of broiler chickens. Anim Sci J. doi:10.1111/asj.12592
- Farvid MS, Eliassen AH, Cho E, Liao X, Chen WY, Willett WC (2016) Dietary fiber intake in young adults and breast cancer risk. Pediatrics 137(3):1–11
- Wang N, Daun JK (2006) Effects of variety and crude protein content on nutrients and anti-nutrients in lentils (*Lens culinaris*). Food Chem 95(3):493–502
- Kapravelou G, Martínez R, Andrade AM, Sánchez C, Chaves CL, López-Jurado M, Aranda P, Cantarero S, Arrebola F, Fernández-Segura E, Galisteo M (2013) Health promoting effects of Lupin (*Lupinus albus* var. multolupa) protein hydrolyzate and insoluble fiber in a diet-induced animal experimental model of hypercholesterolemia. Food Res Int 54(2):1471–1481
- Hayat I, Ahmad A, Masud T, Ahmed A, Bashir S (2014) Nutritional and health perspectives of beans (*Phaseolus vulgaris* L.): an overview. Crit Rev Food Sci 54(5): 580–592
- Iqbal A, Khalil IA, Ateeq N, Khan MS (2006) Nutritional quality of important food legumes. Food Chem 97(2):331–335
- Khattab RY, Arntfild SD, Nyachoti CM (2009) Nutritional quality of legume seeds as affected by some physical treatments, Part 1: protein quality evaluation. Food Sci Technol 42(6):1107–1112
- Wink M (2013) Evolution of secondary metabolites in legumes (Fabaceae). South Afr J Bot 89:164–175
- Kobayashi H (2013) Prevention of cancer and inflammation by soybean protease inhibitors. Front Biosci (Elite Ed) 5:966–973
- Olanca B, Ozay DS (2015) Effects of natural protease inhibitors on high protease activity flours. J Cereal Sci 65:290–297
- Lima AIG, Mota J, Monteiro SAVS, Ferreira RMSB (2016) Legume seeds and colorectal cancer revisited: Protease inhibitors reduce MMP-9 activity and colon cancer cell migration. Food Chem 197:30–38
- Księżak J, Bojarszczuk J (2014) Evaluation of the variation of the contents of anti-nutrients and nutrients in the seeds of legumes. Biotechnol Anim Husb 30(1):153–166
- Sashikala VB, Sreerama YN, Pratape VM, Narasimha H (2015) Effect of thermal processing on protein solubility of green gram (*Phaseolus aureus*) legume cultivars. J Food Sci Technol 52(3):1552–1560
- Gupta RK, Patel AK, Shah N, Chaudhary AK, Jha UK, Yadav UC, Gupta PK, Pakuwal U (2014) Oxidative Stress and Antioxidants in Disease and Cancer: A. Asian Pac J Cancer Prev 15(11):4405–4409
- 31. Gheribi E (2013) The role of fruits and vegetables polyphenols in the dietotherapy of atherosclerosis. Med Rodz 4:149–153
- Siger A, Czubinski J, Kachlicki P, Dwiecki K, Lampart-Szczapa E, Nogala-Kalucka M (2012) Antioxidant activity and phenolic content in three lupin species. J Food Compos Anal 25(2):190–197

- Sánchez-Chino X, Jiménez-Martínez C, Dávila-Ortiz G, Álvarez-González I, Madrigal-Bujaidar E (2015) Nutrient and nonnutrient components of legumes, and its chemopreventive activity: a review. Nutr Cancer 67(3):401–410
- 34. Zhao Y, Du SK, Wang H, Cai M (2014) In vitro antioxidant activity of extracts from common legumes. Food Chem 152:462–466
- Li H, Deng Z, Wu T, Liu R, Loewen S, Tsao R (2012) Microwave-assisted extraction of phenolics with maximal antioxidant activities in tomatoes. Food Chem 130(4):928–936
- Boudjou S, Oomah BD, Zaidi F, Hosseinian F (2013) Phenolics content and antioxidant and anti-inflammatory activities of legume fractions. Food Chem 138(2):1543–1550