

### Short Communication

## Chemical fingerprinting of Saudi Arabian snake venoms using gel filtration chromatography.

Abdulrahman Al Asmari<sup>1</sup>, Haseeb Ahmad Khan<sup>2</sup>, Rajamohammed Abbas Manthiri<sup>1</sup>

<sup>1</sup>Research Center, Prince Sultan Military Medical City, Riyadh, Saudi Arabia

<sup>2</sup>Department of Biochemistry, College of Science, King Saud University, Riyadh, Saudi Arabia

### Abstract

Snake bite is crucial medical emergency while the success of immediate care and anti-venom therapy mainly depend on the correct identification of culprit snake. We present a simple method for identification of six snake species including *Echiscoloratus*, *Echispyramidum*, *Cerastescerastesgaspreti*, *Bitisarietans*, *Najahaje Arabica*, and *Walterinnesiaegyptia*, using the gel filtration chromatographic profiles of their venoms. The chromatograms of venoms from different snake species showed peculiar patterns based on the number and location of peaks. Our findings suggest that chromatographic profiles of snake venoms provide a simple and reproducible chemical fingerprinting method for quick identification of snake species.

**Keywords:** Snake venom, Chromatographic profiles, Elapidae, Viperidae, Identification

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### Introduction

The venoms from Elapidae and Viperidae snakes are complex mixtures containing different components such as metalloproteinases, proteolytic enzymes, phospholipase, serine proteinase, presynaptic and postsynaptic neurotoxins, potassium channel-binding neurotoxins, cytotoxins, cardiotoxins and platelet aggregation inhibitors [1-3]. Newton et al [4] have suggested that analysis of venom components can produce a unique fingerprint to be used as a valuable reference tool in taxonomic analysis as a complementary method to morphology and behavioral characterization for species identification and classification. Compositional differences between snake venoms can be employed as a taxonomy signature for unambiguous species identification independently of geographic origin and morphological characteristics [5]. Recently, Calvete [6] have pointed out that use of proteomics approaches in identification of evolutionary and immunoreactivity trends among homologous and heterologous venoms may aid in the replacement of the traditional geographic- and phylogenetic-driven hypotheses for antivenom production strategies.

In continuation to our previous work on chemical fingerprinting of scorpion venoms [7], we performed the gel filtration chromatography of snake venoms from four

species from the family Viperidae (*Echiscoloratus*, *Echispyramidum*, *Cerastescerastesgaspreti* and *Bitisarietans*) and two species from the family Elapidae (*Najahaje Arabica* and *Walterinnesiaegyptia*) and compared the chromatographic profiles for their application in species identification.

### Materials and Methods

We collected six species of snakes (Fig. 1) including *Echiscoloratus*, *Echispyramidum*, *Cerastescerastesgaspreti*, *Bitisarietans*, *Najahaje Arabica* and *Walterinnesiaegyptia* from the Riyadh region of Saudi Arabia. The snakes were kept in plastic boxes and fed on mice and water *ad libitum*. The crude venom was diluted with distilled water, properly mixed, and centrifuged at 10,000 rpm at 4°C for 20 min to separate the mucus. The clear supernatant was filtered through 0.20 µm filter before chromatography.

Gelfiltration chromatography was used for venom fractionation on Superdex200 PC 3.2/30 column. Venom solution was diluted in 0.05 M sodium phosphate buffer containing 0.15 M NaCl (pH 7.0). An aliquot (25 µl) of venom solution was loaded in a previously equilibrated column with the same buffer. The sample was injected using an Auto injector. The flow rate was adjusted at 0.4

mL/min and the UV range was 0.02 AUFS (absorbance unit full scale). Column operational pressure was 1.5 MPa. The elution profile was monitored at 280 nm by a UV spectrophotometer (AKTA Micro System). All the samples were run in triplicate to confirm the reproducibility of their chromatographic patterns.

## Results and Discussion

All the snake species showed peculiar chromatographic profiles of their venoms depending on the location and height of the peaks (Fig. 2). The minimum numbers of peaks were observed with the venom of *Cerastes cerastes gaspretti* (3 peaks) and the maximum number of peaks with the venom of *Walterinnesia aegyptia* (7 peaks). Both the members of the genus *Echis* (*Echis coloratus* and *Echis pyramidum*) showed 5 peaks each (3 common peaks) whereas the venoms of *Naja haje Arabica* and *Bitis ariatans* resulted in 4 and 6 peaks respectively (Fig. 2).

The venom profile-based chemical fingerprinting clearly differentiated the two species of the family Elapidae (*Naja Arabica* and *Walterinnesia Aegyptia*) from the members of the family Viperidae. The venom profile of *Echis coloratus* was more closely related to the venom

profile of *Cerastes gaspretti* instead of *Echis pyramidum* (Fig. 2). Aird [8] compared the gel filtration profiles of crude snake venoms from 38 *Crotalus viridis*, representing the subspecies *concolor*, *viridis* and *lutosus* and readily distinguished these taxa. John and Kaiser [9] conducted a comparative study of the venoms from *Notechis cutatus*, *Notechisater serventyi*, *Notechisater humphreysi* and *Notechisater using* gel filtration resulting in slightly different elution profiles on a Superose-12 gel filtration column. The protein profile of venoms of Elapidae was identified using electrofocusing technique; the two species could easily be differentiated whereas the differences between the two sub-species were more difficult to evidence [10]. The elution profiles of the venoms of seven *Bothrops* species fractionated on a Mono-Q FPLC column resulted in reproducible chromatograms however there was a considerable overlap of active proteins in different species venoms [11].

In conclusion, each snake's venom has a unique chromatographic profile that can be used as a fingerprint to differentiate one species from the other. Our findings suggest that gel filtration chromatography of snake venom is a simple and reproducible method for identification of snake species.



Figure 1. Snake species used in this study.

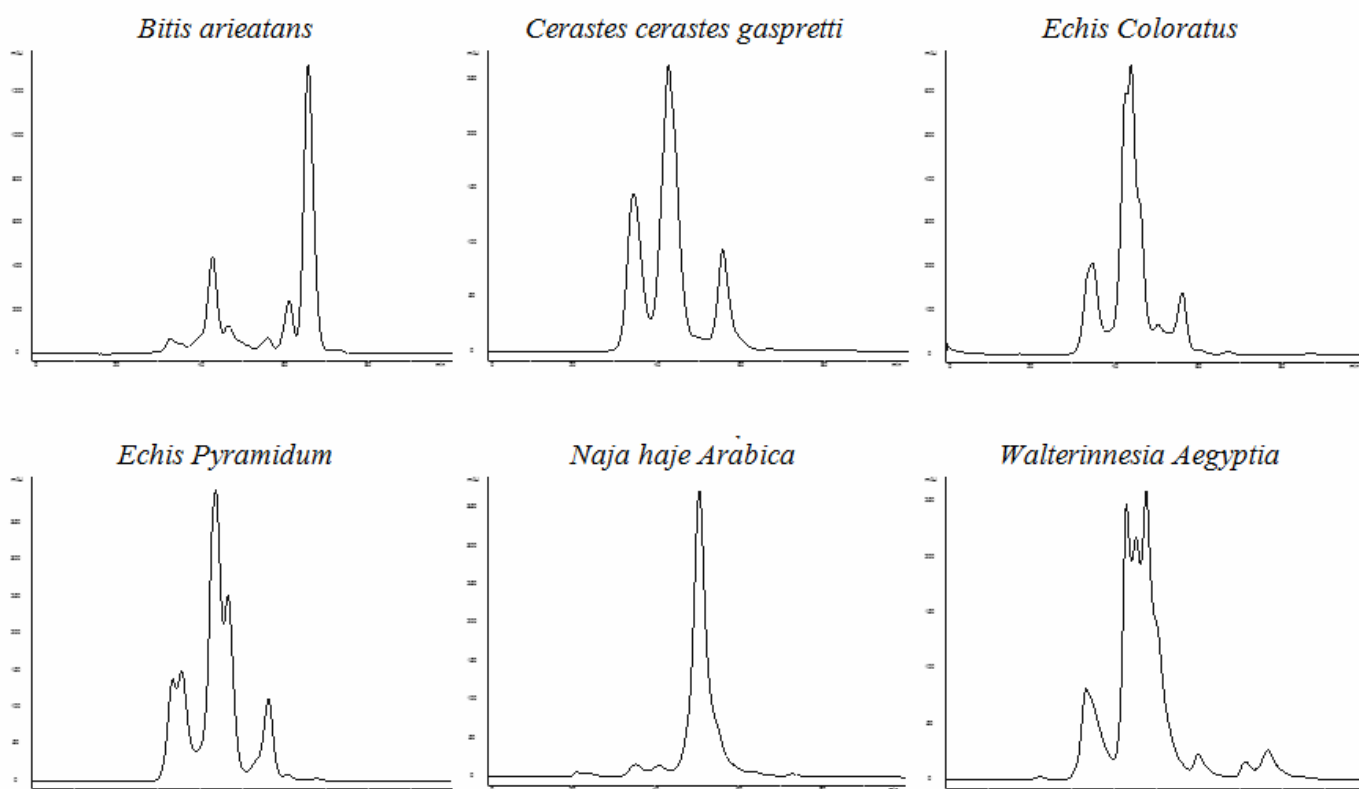


Figure 2. Gel filtration chromatographic profiles of venoms from different snake species.

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## Correspondence to:

Abdulrahman Al-Asmari  
 Director of Research Center  
 Prince Sultan Military Medical City  
 P.O. Box 7897, Riyadh 11159  
 Saudi Arabia