

# 1 **Title: Chromosome 10q24.32 Variants Associate with Brain Arterial** 2 **Diameters in Diverse Populations: A Genome-Wide Association Study**

3 **Authors:** Minghua Liu<sup>1</sup>, Farid Khasiyev<sup>2</sup>, Sanjeev Sariya<sup>1,3,4</sup>, Antonio Spagnolo-Allende<sup>1</sup>,  
4 Danurys L Sanchez<sup>1,3,4</sup>, Howard Andrews<sup>5</sup>, Qiong Yang<sup>6</sup>, Alexa Beiser<sup>6</sup>, Ye Qiao<sup>7</sup>, Emy A  
5 Thomas<sup>8</sup>, Jose Rafael Romero<sup>9</sup>, Tatjana Rundek<sup>10,11,12</sup>, Adam M Brickman<sup>1,3,4</sup>, Jennifer J  
6 Manly<sup>1,3,4</sup>, Mitchell SV Elkind<sup>1,13</sup>, Sudha Seshadri<sup>9,14</sup>, Christopher Chen<sup>15</sup>, Ralph L Sacco<sup>10,11,12</sup>,  
7 Saima Hilal<sup>15</sup>, Bruce A Wasserman<sup>7,16</sup>, Giuseppe Tosto<sup>1,3,4</sup>, Myriam Fornage<sup>8,17</sup>, Jose Gutierrez<sup>1</sup>.

## 8 9 **Affiliations:**

10 1 Department of Neurology, Vagelos College of Physicians and Surgeons, Columbia University,  
11 New York, NY, USA

12 2 Department of Neurology, Saint Louis University School of Medicine, St. Louis, MO, USA

13 3 Taub Institute for Research on Alzheimer's Disease and the Aging Brain, Vagelos College of  
14 Physicians and Surgeons, Columbia University, New York, NY, USA

15 4 The Gertrude H. Sergievsky Center, Vagelos College of Physicians and Surgeons, Columbia  
16 University, New York, NY, USA

17 5 Biostatistics Department, Mailman School of Public Health, Columbia University, New York,  
18 NY, USA

19 6 Department of Biostatistics, School of Public Health, Boston University, Boston, MA, USA

20 7 Johns Hopkins University School of Medicine, Baltimore, MD, USA

21 8 Brown Foundation Institute of Molecular Medicine, Mc Govern Medical School, The  
22 University of Texas Health Science Center at Houston, Houston, TX, USA

23 9 Department of Neurology, Boston University School of Medicine, Boston, MA, USA

24 10 Department of Neurology, University of Miami Miller School of Medicine, Miami, FL, USA

25 11 Department of Public Health Sciences, University of Miami Miller School of Medicine,  
26 Miami, FL, USA

27 12 Evelyn F. McKnight Brain Institute, University of Miami Miller School of Medicine, Miami,  
28 FL, USA

29 13 Department of Epidemiology, Mailman School of Public Health, Columbia University, New  
30 York, NY, USA

31 14 The Glenn Biggs Institute for Alzheimer's and Neurodegenerative Diseases, University of  
32 Texas Health Sciences Center, San Antonio, TX, USA

33 15 Memory Aging and Cognition Center, Department of Pharmacology, Yong Loo Lin School of  
34 Medicine, National University of Singapore, Singapore

35 16 University of Maryland School of Medicine, Baltimore, MD, USA

36 17 Human Genetics Center, School of Public Health, The University of Texas Health Science  
37 Center at Houston, Houston, TX, USA

## 38 39 **Contact author:**

40 Jose Gutierrez, MD, MPH.

41 Address: 710 W 168th Street, 6th floor, Suite 639

42 New York, NY, 10032.

43 Email: [jg3233@cumc.columbia.edu](mailto:jg3233@cumc.columbia.edu)

44 Phone: (212) 305-1710; Fax: (212) 305-165

45

46

---

<b>Author Name</b>	<b>Email Address</b>
Minghua Liu	ml3883@cumc.columbia.edu
Farid Khasiyev	farid.khasiyev@health.slu.edu
Sanjeev Sariya	ss5505@cumc.columbia.edu
Antonio Spagnolo-Allende	ajs2363@cumc.columbia.edu
Danurys L Sanchez	dls167@cumc.columbia.edu
Howard Andrews	Howard.Andrews@nyspi.columbia.edu
Qiong Yang	qyang@bu.edu
Alexa Beiser	alexab@bu.edu
Ye Qiao	yqiao4@jhmi.edu
Emy A Thomas	Emy.Anu.Thomas@uth.tmc.edu
Jose Rafael Romero	joromero@bu.edu
Tatjana Rundek	TRundek@med.miami.edu
Adam M Brickman	amb2139@cumc.columbia.edu
Jennifer J Manly	jjm71@cumc.columbia.edu
Mitchell SV Elkind	mse13@cumc.columbia.edu
Sudha Seshadri	seshadri@uthscsa.edu
Christopher Chen	phccclh@nus.edu.sg
Ralph L Sacco	rsacco@med.miami.edu
Saima Hilal	saimahilal@nus.edu.sg
Bruce A Wasserman	BWasser@som.umaryland.edu
Giuseppe Tosto	gt2260@cumc.columbia.edu
Myriam Fornage	Myriam.Fornage@uth.tmc.edu
Jose Gutierrez	jg3233@cumc.columbia.edu

---

47

48

49 **Abstract**

50

51 **Background:** Brain arterial diameters are novel imaging biomarkers of cerebrovascular disease,  
52 cognitive decline and dementia. Traditional vascular risk factors have been associated with brain  
53 arterial diameters but whether there may be genetic determinants of brain arterial diameters is  
54 unknown.

55

56 **Results:** We studied 4150 participants from six geographically diverse population-based cohorts  
57 (40% European, 14% African, 22% Hispanic, 24% Asian ancestries). We measured brain arterial  
58 diameters for 13 segments and averaged them to obtain a global measure of brain arterial  
59 diameters as well as the posterior and anterior circulations. A genome-wide association study  
60 (GWAS) revealed 14 variants at one locus associated with global brain arterial diameter at  
61 genome-wide significance ( $P < 5 \times 10^{-8}$ ) (top SNP, rs7921574;  $\beta = 0.06$ ,  $P = 1.54 \times 10^{-8}$ ). This locus  
62 mapped to an intron of *CNNM2*. A trans-ancestry GWAS meta-analysis identified two more loci  
63 at *NT5C2* (rs10748839;  $P = 2.54 \times 10^{-8}$ ) and at *AS3MT* (rs10786721;  $P = 4.97 \times 10^{-8}$ ), associated with  
64 global brain arterial diameter. In addition, two SNPs co-localized with expression of *CNNM2*  
65 (rs7897654,  $\beta = 0.12$ ,  $P = 6.17 \times 10^{-7}$ ) and *AL356608.1* (rs10786719,  $\beta = -0.17$ ,  $P = 6.60 \times 10^{-6}$ ) in brain  
66 tissue. For the posterior brain arterial diameter, two variants at one locus mapped to an intron of  
67 *TCF25* were identified (top SNP, rs35994878;  $\beta = 0.11$ ,  $P = 2.94 \times 10^{-8}$ ). For the anterior brain  
68 arterial diameter, one locus at *ADAP1* was identified in trans-ancestry genome-wide association  
69 analysis (rs34217249;  $P = 3.11 \times 10^{-8}$ ).

70

71 **Conclusion:** Our study reveals three novel risk loci (*CNNM2*, *NT5C2* and *AS3MT*) associated  
72 with brain arterial diameters. Our finding may elucidate the mechanisms by which brain arterial  
73 diameters influence the risk of stroke and dementia.

74

75 **Key words:** Larger brain arterial diameters, Genome-wide association studies, Chromosome  
76 10q24.32, Cyclin and CBS Domain Divalent Metal Cation Transport Mediator 2 (*CNNM2*)

77

## 78 **Background**

79 Dolichoectasia has been defined by elongated and tortuous arteries<sup>1</sup> and it is usually associated  
80 with smoking, male sex and aging.<sup>2</sup> The diagnosis of dolichoectasia has been historically  
81 ascertained by visual inspection of neuroimaging or more recently using a fixed arterial diameter  
82 cutoffs for the basilar artery.<sup>3</sup> Although these methods are easy to use, they simplify the  
83 biological meaning of the continuum of intracranial arterial diameters in brain health and neglect  
84 arterial-size expectations based on age, sex and head size.<sup>4</sup> Therefore, we proposed and validated  
85 the principle that arterial diameters measured continuously and adjusted for head size relate to  
86 health outcomes in a non-linear fashion and that people with very small or very large arterial  
87 diameters are at a higher risk of vascular events.<sup>5</sup> Furthermore, dilated brain arterial diameters  
88 are associated with a higher risk of dementia<sup>6</sup> and steeper cognitive decline.<sup>7</sup> Smaller arterial  
89 diameters causing stenosis, usually related to atherosclerosis, are intuitively related to adverse  
90 health outcomes<sup>8,9</sup> but less is known about the underlying nature of dilated brain arteries.

91  
92 Larger arterial diameters have been described in people with connective tissue disorders such as  
93 Marfan syndrome,<sup>10</sup> Ehlers-Danlos,<sup>11</sup> and Arterial Tortuosity Syndrome,<sup>12</sup> among others. These  
94 monogenic diseases are usually rare and detected in younger patients. Furthermore, there is clear  
95 association between larger arterial diameters and vascular risk factors, especially  
96 hypertension.<sup>13,14</sup> Although hypertension is highly prevalent in elderly populations, the  
97 heterogeneity of brain arterial phenotypes in people with vascular risk factors suggests that a  
98 specific genetic profile might partially be responsible for higher risk of brain arterial dilatation.  
99 Consequently, we hypothesize that in the general population less pathogenic but more frequent  
100 genetic variants may relate to brain arterial diameters. Identifying such a genetic profile may  
101 shed light into possible mechanistic links between large brain arterial diameters and the observed  
102 brain outcomes. To test our hypothesis, we leveraged diverse population cohorts within and  
103 outside the United States to investigate associations between brain arterial diameters and  
104 Alzheimer's disease, stroke, and white matter hyperintensities volume.

## 105 **Results**

### 107 **Multi-ancestry GWAS Identifies a Novel Locus Associated with Brain Arterial Diameter**

108 We conducted a multi-ancestry GWAS for brain arterial diameter levels in 4150 participants,  
109 including 1650 from European, 583 from African, 920 from Hispanic, and 997 from Asian  
110 ancestries. Mean age of the participants across studies ranged from 70 to 76 years, with  
111 proportions of women ranging from 52 to 64%. Detailed demographic information is presented  
112 in Table 1.

113  
114 We identified 14 variants at one locus associated with global brain arterial diameter at genome-  
115 wide significance ( $P < 5 \times 10^{-8}$ ; Figure 1A). This locus mapped to an intron of *CNNM2* (Cyclin and  
116 CBS Domain Divalent Metal Cation Transport Mediator 2). One copy of the C allele (minor  
117 allele frequency [MAF], 0.42) for the lead single-nucleotide polymorphism (SNP) rs7921574  
118 was associated with 6% increased global brain arterial diameter ( $P = 1.54 \times 10^{-8}$ ) (Table 2; Figure  
119 S1A). We also identified 2 intronic variants in *TCF25* (Transcription Factor 25) associated with  
120 posterior brain arterial diameter at genome-wide significance (Figure 1B). One copy of the C  
121 allele (MAF= 0.29) for the lead SNP rs35994878 was associated with 11% increased posterior  
122 brain arterial diameter ( $P = 2.94 \times 10^{-8}$ ) (Table 2; Figure S1B). We did not observe any genome-  
123 wide significant association for anterior brain arterial diameter (Figure 1C). No genomic

124 inflation was observed for any of the brain arterial diameter analyses (Figure S2). We also  
125 performed ancestry-specific regional Manhattan plots for top SNPs in global (rs7921574),  
126 anterior (rs7921574), and posterior (rs35994878) diameter meta-analysis, respectively (Figure  
127 S3, S4, and S5), but did not observe ancestry-specific genome-wide significant associations.  
128

129 In the trans-ancestry genome-wide association analysis for global brain arterial diameter, in  
130 addition to SNPs in *CNNM2*, we identified one genome-wide significant SNP near *NT5C2*  
131 (rs10748839;  $P=2.54 \times 10^{-8}$ ) and one in *AS3MT* (rs10786721;  $P=4.97 \times 10^{-8}$ ). All were located  
132 within 10q24.32. For anterior brain arterial diameter, we identified one locus at *ADAPI*  
133 (rs34217249;  $P=3.11 \times 10^{-8}$ ) (Table 3). In the Hispanic-specific analysis, we identified a genome-  
134 wide significant locus at *LOC107986223* for global brain arterial diameter; three loci at  
135 *TGFBR2*, *LOC105374506*, and *LOC105376292* for posterior brain arterial diameter (Table S5).  
136 We did not observe genome-wide significant association in European, African, or Asian  
137 ancestries (Table S3, S4 and S6) for global, anterior, or posterior brain arterial diameter. No  
138 genomic inflation was observed in trans-ancestry analysis (Figure S6, S7, and S8).  
139

### 140 **Variant Effects Predictions on Protein Coding Sequence**

141 We investigated the predicted deleterious effects of brain arterial diameter-associated loci using  
142 the Combined Annotation Dependent Depletion (CADD) scores. The SNPs and their proxies  
143 with CADD scores are shown in Table 2 and Table 3. We did not observe significant CADD  
144 score among the genome-wide significant loci associated with brain arterial diameter. A SNP  
145 associated with posterior brain arterial diameter in the *RAD52* region (rs140934041) showed  
146 significant CADD score (13.45) in Hispanic-specific analysis (Table S5). Additionally, SNPs  
147 associated with anterior brain arterial diameter in the *RAPGEF4* and posterior brain arterial  
148 diameter in *PODXL* region showed significant CADD scores (rs2290378, 16.31; rs888608,  
149 16.63) in Asian-specific analysis (Table S6).  
150

### 151 **Gene-Based Association Test and Gene-Set Enrichment**

152 The Multi-Marker Analysis of GenoMic Annotation gene-based association analysis identified  
153 one locus associated with global brain arterial diameter ( $P < 1.50 \times 10^{-5}$ ) (Table S7). The  
154 significant associations for global brain arterial diameter included the GWAS located at *AS3MT*,  
155 *CNNM2*, *NT5C2*, *ARL3*, *TMEM180*, *C10orf32* and *C10orf32-ASMT*. Genes mapped to GWAS  
156 associations with  $P < 1 \times 10^{-5}$  were further investigated for gene-set enrichment (Table S8). Three  
157 genome-wide significant loci for global brain arterial diameter, *AS3MT*, *CNNM2* and *NT5C2*,  
158 were enriched in the white matter lesion progression gene set from GWAS catalog database  
159 (adjusted  $P=7.60 \times 10^{-7}$ ).  
160

### 161 **Tissue-Specific Colocalization Analyses**

162 We performed colocalization analysis for the locus identified in the GWAS and MTAG analysis  
163 with gene expression using Genotype-Tissue Expression v8 eQTL data (Table S9). We identified  
164 SNPs associated with *AS3MT* and *C10orf32* expression and global brain arterial diameter in all  
165 13 brain tissues. We also identified SNPs at *TMEM180* in caudate basal ganglia, cerebellar  
166 hemisphere, nucleus accumbent basal ganglia, putamen basal ganglia, and spinal cord cervical c-  
167 1; SNPs at *CNNM2* in caudate basal ganglia tissues and SNPs at *NT5C2* and *ARL3* in cerebellum  
168 tissue, which colocalized with global brain arterial diameter. We also performed a transcriptome-  
169 wide association analysis for the loci identified in the GWAS with gene expression using



170 BrainMeta project data in global, anterior, and posterior brain arterial diameter (Table 4). At the  
171 transcriptome-wide significance level ( $P < 8.4 \times 10^{-6}$ ), we identified SNPs associated with *CNNM2*  
172 ( $P = 6.17 \times 10^{-7}$ ) and *AL356608.1* ( $P = 6.6 \times 10^{-6}$ ) expression in global brain arterial diameter (Figure  
173 2A). We did not observe transcriptome-wide significant association in anterior or posterior brain  
174 arterial diameter (Figure 2B, 2C).

175

### 176 **Causal Pathway from Brain Arterial Diameter to Alzheimer's disease, stroke, and white** 177 **matter hyperintensities volume**

178 To establish a causal pathway from brain arterial diameter to Alzheimer's disease, stroke, and  
179 white matter hyperintensities volume, we performed a Mendelian Randomization (MR) analysis  
180 (Table S10, S11). We did not observe any association of brain arterial diameter with Alzheimer's  
181 disease, stroke, or white matter hyperintensities volume.

182

### 183 **Pleiotropic Locus for Anterior and Posterior Brain Arterial Diameter**

184 MTAG analysis used the fixed-effect meta-analysis estimates for anterior and posterior brain  
185 arterial diameter. Since global brain arterial diameter is the average of anterior and posterior  
186 brain arterial diameter, the global estimate was excluded from multivariate analysis. No genomic  
187 inflation was observed in trans-ancestry analysis (Figure S9). MTAG results of joint analysis  
188 brain arterial diameter did not show any genome-wide significant SNPs (Table S12, S13).

189

## 190 **Discussion**

191 This is the first study to examine the genetic determinants of brain arterial diameter in an  
192 ancestrally diverse population, where we identified associations of novel genetic loci with brain  
193 arterial diameter genetic architecture. Beyond mapping to the nearest genes, we also showed the  
194 biological impact of our findings using in silico functional analyses. Our results demonstrated  
195 that multiple genetic loci were coupled with gene expression information, which imply  
196 biologically relevant pathways.

197

198 We identified a novel brain arterial diameter locus at 10q24.32 mapped to *CNNM2*. *CNNM2*  
199 encodes Cyclin M2, which is a member of magnesium ( $Mg^{2+}$ ) transporters. As an abundant  
200 intracellular divalent cation in the human body, magnesium ( $Mg^{2+}$ ) plays an important role in  
201 numerous biological processes such as the synthesis of RNA, DNA and protein, and the  
202 production and storage of cellular energy<sup>42</sup>. *CNNM2* is involved in brain development,  
203 neurological functioning and  $Mg^{2+}$  homeostasis<sup>43</sup>. Heterozygous variants in the *CNNM2* gene  
204 can cause renal hypomagnesemia (HOMG6 [MIM 613882]), seizures, and intellectual disability  
205 (HOMGSMR1 [MIM 616418])<sup>44</sup>. In our study, variant rs7897654 was associated with decreased  
206 brain arterial diameter ( $\beta = -0.06$ ) and an increased *CNNM2* expression ( $\beta = 0.02$ ) (Table 4). The  
207 variant rs7897654 colocalized with eQTLs of *CNNM2*, confirming its functional relationship to  
208 this gene. Our study also identified a variant in *TCF25* associated at genome-wide significance  
209 with posterior brain arterial diameter. *TCF25* is a member of the basic helix-loop-helix (bHLH)  
210 family of transcription factors that are important in embryonic development<sup>45</sup>. These two results  
211 suggest that the effects of these genetic variants on arterial size might be present early in life, but  
212 how aging interacts with these variants remains unknown.

213

214 The *NT5C2* encodes a phosphatase involved in cellular purine metabolism, which is associated  
215 with disorders characterized by psychiatric and psychomotor disturbances<sup>46,47</sup>. *NT5C2* has a

216 high affinity for adenosine monophosphate and is involved in the extensive transcriptional  
217 programming which regulates cell maintenance, proliferation, migration, and differentiation  
218 during neurodevelopment<sup>48-51</sup>. *NT5C2* has also been shown to negatively regulate  
219 phosphorylation of the alpha subunit of 5'-adenosine monophosphate-activated protein kinase  
220 (AMPK alpha) and protein translation<sup>52</sup>. Studies in the Chinese Han population report that  
221 *NT5C2* rs2148198 is associated with coronary heart disease susceptibility, and *NT5C2*  
222 rs11191580 is associated with schizophrenia and symptom severity<sup>53,54</sup>. In addition, a zebrafish  
223 study provides evidence that *NT5C2* and *CNNM2* are most likely the causal genes within a blood  
224 pressure locus at the 10q24.32<sup>55</sup>. Our trans-ancestry GWAS analysis identified a significant  
225 variant rs10748839, mapped on the 2KB upstream of *NT5C2*, which is promoter variant that  
226 control expression of *NT5C2*<sup>68</sup>.

227  
228 The *AS3MT* gene, located in 10q24.32, encodes a cytosolic protein which is a cysteine rich  
229 enzyme that transfers a methyl group from S-adenosyl-L-methionine to trivalent arsenical<sup>56,57</sup>.  
230 *AS3MT* plays an important role in catalysis of biomethylation of arsenic in vivo and in vitro.  
231 *AS3MT* is mainly expressed in human adrenal glands, liver, heart, kidney, and brain<sup>58</sup>.  
232 Additionally, *AS3MT* expression is highly expressed in adult human neurons and astrocytes  
233 during human stem cell differentiation toward neuronal fates and in brains of patients with  
234 schizophrenia compared with controls<sup>59</sup> and with attention deficit or hyperactivity disorder<sup>60</sup>.  
235 Notably, *AS3MT* rs7085104 as a schizophrenia-associated risk SNP altered striatal dopamine  
236 synthesis capacity. Moreover, the *AS3MT-CNNM2-NT5C2* gene cluster region is involved in  
237 etiology and pathogenesis of schizophrenia and the three genes have been confirmed as  
238 schizophrenia susceptibility gene cluster<sup>61,62</sup>. Our study identified *AS3MT* rs10786721 variants  
239 with genome-wide significance in global brain arterial diameter. In addition, *AS3MT* rs72841270  
240 is a lead variant associated with global brain arterial diameter in Hispanic-specific population.  
241 Whether neuronal connectivity or network formation indirectly or directly impacts brain arterial  
242 diameters is unclear but should be further studied.

243  
244 Lysosomes play a critical role in maintenance of the integrity of neuronal function, and  
245 mutations in genes that contribute to lysosome formation, transport, and activity are associated  
246 with neurodegenerative disorders<sup>63,64</sup>. Recently, the multi-subunit complex, BLOC-one-related  
247 complex (BORC), has been shown to be involved in positioning lysosomes within the cytoplasm,  
248 although the consequences of altered BORC function in adult animals have not been established  
249<sup>65,66</sup>. A study in mice identifies *BORCS7* (*C10orf32*) as a central factor in axonal transport of  
250 lysosomes and a possible target for improving disease-related disturbances in this important  
251 function; additionally, the Q87X mutation in the *BORCS7* subunit results in motor deficits and  
252 dystrophic axonopathy in mice<sup>67</sup>. In our gene-based MAGMA analysis, the significant  
253 associations for global brain arterial diameter included *AS3MT*, *C10orf32*, *CNNM2* and *NT5C2*;  
254 we suspect that this four-gene cluster region may be involved with the etiology and pathogenesis  
255 of brain arterial diameter, but the underlying mechanism is not clear.

256  
257 Our study is the first to explore the risk variants of brain arterial diameter in a large multi-  
258 ancestry GWAS. We detected novel SNPs located in genomic region 10q24.32 which are  
259 associated with brain arterial diameter. Due to the modest sample sizes of African, Asian, and  
260 Hispanic participants, the statistical power to detect ancestry-specific associations or functional  
261 associations in these ancestries were limited. Similarly, disentangling the effects of these variants

262 on overall brain health versus AD specific pathways is difficult without functional analyses of  
263 genes related to arterial diameters, but exploring such pathways may reveal novel vascular  
264 contribution to Alzheimer's disease and related dementias. Based on our results, we anticipate  
265 that the association between these genetic variants and brain arterial diameters will be consistent  
266 across populations, although the effect size might vary given the presence of common  
267 confounders such as vascular risk factors and environmental exposures.

268

## 269 **Conclusions**

270 In summary, we identified a novel genome-wide significant locus for brain arterial diameter,  
271 *CNNM2*, *NT5C2* and *AS3MT*, in a large multi-ancestry population. Our study provides a  
272 potential biological mechanism for the association between 10q24.32 variation and brain arterial  
273 diameter. Identifying genes associated with these loci and their function may help us to elucidate  
274 the mechanism by which brain arterial diameters may influence cerebrovascular health.

275

## 276 **Methods**

### 277 **Sampled populations**

#### 278 Atherosclerosis Risk in Communities (ARIC) study

279 The ARIC study is a population-based prospective cohort study of vascular risks and includes  
280 15,792 persons aged 45-64 years at baseline (1987-89), randomly chosen from four US  
281 communities.<sup>15</sup> Cohort members completed seven clinic examinations, conducted between 1987  
282 and 2019. Written informed consent was provided by all study participants, and the study design  
283 and methods were approved by institutional review boards at the collaborating medical  
284 institutions (The Johns Hopkins University, Wake Forest University, University of Mississippi  
285 Medical Center, and University of Minnesota). Dementia and dementia subtypes were  
286 adjudicated beginning in 2011 using in-person interviews and cognitive testing, chart reviews  
287 and telephone surveys.<sup>16</sup>

288

#### 289 The Northern Manhattan Study (NOMAS)

290 The NOMAS is an ongoing prospective cohort initially focused on determining the incidence of  
291 stroke and vascular events in a diverse urban population. Participants were recruited using  
292 random digit dialing between 1993 and 2001 with the following eligibility criteria: (1) age 40 or  
293 older, (2) clinically stroke free, and (3) resident of Northern Manhattan for at least 3 months. In  
294 person cognitive testing has been done three times since 2011 in surviving participants,  
295 Dementia was adjudicated by consensus between a neurologist and a neuropsychologist.  
296 The institutional review boards at Columbia University Medical Center and the University of  
297 Miami approved the study. All participants provided written informed consent.

298

#### 299 Washington Heights–Inwood Columbia Aging Project (WHICAP study)

300 WHICAP is a prospective, population-based study of aging and dementia. Established through  
301 several recruitment waves, participants were first recruited in 1992 from a random sample of  
302 Medicare-eligible adults (age  $\geq 65$ ) residing in the neighborhoods of Washington Heights and  
303 Inwood in northern Manhattan. Participants are evaluated longitudinally every 18–24 months,  
304 with a comprehensive neuropsychological battery, medical and neurologic examination, and  
305 survey about health-related outcomes.<sup>17</sup> Dementia and dementia subtypes are adjudicated in a  
306 consensus conference that includes neurologists and neuropsychologists.

307



308 Epidemiology of Dementia In Singapore (EDIS study)

309 The EDIS study is a population-based cohort study conducted in southwestern Singapore  
310 between 2004 and 2011. It recruited participants who participated in the baseline visit of the  
311 Singapore Epidemiology of Eye Diseases (SEED) which comprised 10,033 adults of Chinese,  
312 Malay, and Indian ancestry, 40-80 years old<sup>18-21</sup>. Briefly, the EDIS study consisted of three  
313 independent population cohorts with a common protocol. In all studies, individuals 40-80 years  
314 old were selected by an age-stratified random sampling method from a computer-generated  
315 random list of names provided by the Ministry of Home Affairs. The study was approved by the  
316 SERI Institutional Review Board. Written informed consent was obtained, in the preferred  
317 language of participants, by bilingual study coordinators prior to recruitment into the study.

318

319 Memory Clinic in Singapore (MCS study)

320 The MCS study included patients attending the National University Hospital (NUH) and St  
321 Luke's Hospital memory clinics between 2009 and 2015. Patients were referred by primary care  
322 as well as secondary and tertiary care facilities because of consistent memory complaints and  
323 were assessed by a team of clinicians, psychologists, and nurses in the Memory Aging and  
324 Cognition Center, National University of Singapore.

325

326 Framingham Heart Study (FHS)

327 FHS started enrolling community-based participants in 1949. In 1971, all descendants of the  
328 original cohort (i.e., offspring cohort, requiring at least one parent from the original cohort) and  
329 their spouses were invited to participate in a follow-up study, and since then, they have been  
330 followed prospectively. The initial cohort consisted of 5124 men and women; 88% of survivors  
331 (3539/4031) participated in examination 7 in 1998-2001. Participants who survived to the 7th  
332 examination were invited to undergo a brain MRI (1999-2005), with a final sample of 2144  
333 stroke-free, community-based participants. For these analyses, we used a FHS subsample with  
334 available MRA as part of the stroke case study.

335

336 **Measurement of brain arterial diameter**

337 Brain magnetic resonance angiogram acquisition parameters by cohort are reported in  
338 supplemental Table 1. Brain arterial diameters and lengths were obtained from all available  
339 MRA images using commercial software (LAVA, Leiden University Medical Center, The  
340 Netherlands, build date Oct 19, 2018). Briefly, this software uses a flexible 3D tubular Non-  
341 Uniform Rational B-Splines model to automatically identify the margins of the arterial lumen  
342 based on voxel intensity<sup>22</sup> with excellent reliability.<sup>23</sup> The 13 arterial segments measured  
343 included the bilateral intracranial internal carotid (ICA); middle cerebral (MCA), anterior  
344 cerebral (ACA), posterior cerebral (PCA), vertebral (VA), and posterior communicating  
345 (Pcomm) arteries, plus the basilar artery (BA). The location of measurement was aimed at the  
346 largest portion of a given segment free of focal stenosis, with good to excellent reliability.<sup>24</sup> For  
347 arteries visualized in the axial source MRA images but not large enough to be reconstructed, we  
348 systematically assigned the smallest measured diameters for the artery in the sample minus 10%.  
349 We counted arteries not visualized in axial source MRA images to create a score of absent  
350 arteries. We transformed each artery diameter distribution into normal scores and obtained the  
351 global (all 13 arteries), anterior (ICA, MCA, ACA and Pcomm if available) and posterior (VA,  
352 BA and PCA if available) arterial diameter scores by cohort as the principal dependent variable.

353

## 354 **Genotyping and Imputation**

355 Detailed description of genotyping, quality control and imputation in each study is provided in  
356 supplemental Table 2. All analyses were conducted on autosomal chromosomes. Genotypes with  
357 missing rate greater than 10%, significant Hardy-Weinberg Equilibrium p-value (HWE p-value  
358  $<5 \times 10^{-8}$ ), or poor imputation quality ( $r^2 < 0.3$ ) were excluded from the analyses.

## 359 **Genome-wide Association Analysis**

361 In each study and self-reported racial/ancestry strata, linear regression models were used to test  
362 the association between genetic variants and brain arterial diameter (global, anterior and  
363 posterior scores) using an additive genetic model, adjusted for sex, age, head size, number of  
364 absent arteries and population-specific principal components of ancestry (PCs). Genome-wide  
365 association studies (GWAS) results were subjected to quality control analyses using EasyQC  
366 (Winkler et al., 2013) and combined by meta-analysis using a fixed-effect inverse-variance-  
367 based method implemented in METAL<sup>25</sup> Variants with minor allele frequencies (MAF)  $< 1\%$   
368 and those do not present in at least two studies were excluded after the meta-analyses. Cross-  
369 study heterogeneity was assessed using Cochran's Q-test, and variants with heterogeneity p-  
370 value  $< 0.05$  were excluded. A trans-ancestry meta-analysis of GWAS was conducted to account  
371 for heterogeneity in allelic effect that is correlated with ancestry by Meta-Regression of Multi-  
372 Ethnic Genetic Association (MR-MEGA).<sup>26</sup> Ancestry-specific meta-analyses were also  
373 performed to identify ancestry-specific variants. Multi-Trait Analysis of GWAS (MTAG) tool<sup>27</sup>  
374 was used for multivariate analysis of anterior and posterior brain arterial diameter to boost the  
375 statistical power to detect genetic associations. An association with a p-value  $< 5 \times 10^{-8}$  was  
376 considered genome-wide significant, whereas p-value  $< 1.0 \times 10^{-5}$  were used as suggestive  
377 evidence for marker associations.

## 378 **Gene-based Association Analysis and Gene-set Enrichment**

380 We performed a gene-based association analysis based on summary statistics using Multi-marker  
381 Analysis of GenoMic Annotation (MAGMA v.1.07),<sup>28</sup> implemented by FUnctional Mapping  
382 and Annotation (FUMA).<sup>29</sup> Variants with p-value  $< 1 \times 10^{-5}$  were mapped to the nearest gene  
383 within 50kb or an expression quantitative trait locus (eQTL) genes in Genotype-Tissue  
384 Expression (GTEx) project data version 8 (v8)<sup>30</sup> from brain tissues: amygdala, anterior cingulate  
385 cortex (BA24), caudate (basal ganglia), cerebellar hemisphere, cerebellum, cortex, frontal Cortex  
386 (BA9), hippocampus, hypothalamus, nucleus accumbens (basal ganglia), putamen (basal  
387 ganglia), spinal cord (cervical c-1), substantia nigra. Mapped genes were then tested for tissue  
388 specificity in 30 general GTEx tissues using the pre-calculated differentially expressed gene  
389 (DEG) sets integrated in the GENE2FUNC of FUMA.<sup>29</sup> Hypergeometric enrichment tests were  
390 performed via GENE2FUNC against pre-defined gene sets obtained from Molecular signatures  
391 database (MsigDB)<sup>31</sup>, WikiPathways<sup>32</sup> and GWAS catalog.<sup>33</sup> We used Bonferroni corrected p-  
392 value  $< 0.05$  to define statistical significance in gene-based analyses.

## 393 **Identification of Genomic Risk Loci and Deleteriousness of Lead SNPs**

395 Variants that had at least suggestive evidence (p-value  $< 10^{-5}$ ) were filtered and LD-clumped at  
396  $r^2 < 0.1$  to identify independent loci using FUMA's SNP2GENE function<sup>29</sup> based on the relevant  
397 1000G reference. To investigate the protein coding consequences of lead independent variants  
398 associated with brain arterial diameter, the Combined Annotation Dependent Depletion (CADD)  
399 score was estimated. We used the threshold of 12.37 to determine whether a lead variant was

400 deleterious.<sup>34</sup> When the CADD score of a lead variant was smaller than 12.37, we assessed  
401 whether its proxy variants ( $r^2 > 0.8$ ) were deleterious instead.

402

### 403 **Pleiotropic Association Analysis with Gene Expression**

404 We used the SMR software<sup>35</sup> to test for pleiotropic association between brain arterial diameter  
405 traits and gene expression. We used summary-level data from our GWAS analyses and data on  
406 expression quantitative trait loci (eQTL) from the BrainMeta project version 2.<sup>36</sup> There are 5967  
407 cis-eQTLs with eQTL  $p$ -value  $< 5 \times 10^{-8}$ . We used a Bonferroni corrected  $p$ -value  $< 8.4 \times 10^{-6}$   
408 (0.05/5967) to define statistical significance in pleiotropic association analyses.

409

### 410 **Two Sample Mendelian Randomization Analysis**

411 We conducted a two-sample Mendelian randomization (MR) analysis using genetic instruments  
412 from the present analyses to assess whether brain arterial diameter is a causal factor for  
413 Alzheimer's disease, stroke and white matter hyperintensities volume. The summary statistics for  
414 Alzheimer's disease, stroke and white matter hyperintensities volume were used in this analysis.

415 <sup>37-39</sup> To avoid bias driven by correlated instruments, variants with brain arterial diameter  
416 association  $p$ -value  $< 1.0 \times 10^{-5}$  were LD-clumped at  $r^2 < 0.01$ <sup>40</sup> against the 1000 Genome LD  
417 reference calculated for African, European, Asian and Hispanic populations. Variants with  
418 MAFs  $< 0.01$  in the reference population were excluded from MR analysis. Causal association  
419 was primarily evaluated using the inverse-variance weighted method. To assess the presence of  
420 horizontal pleiotropy (i.e., that variants influence the outcome trait via independent pathways  
421 other than the exposure trait), we used the simple mode method, weighted mode method,  
422 inverse-variance weighted method (IVW), median-based method, and MR-Egger method. All  
423 MR analyses were performed using the "TwoSampleMR" R package.<sup>41</sup>

424

425

426 **Declarations**

427 **Ethics approval and consent to participate**

428 The included studies have been approved by local ethics committees: Atherosclerosis Risk in  
429 Communities (ARIC) study: The institutional review board at The Johns Hopkins University,  
430 Wake Forest University, University of Mississippi Medical Center, and University of  
431 Minnesota; The Northern Manhattan Study (NOMAS): The institutional review boards at  
432 Columbia University Medical Center and the University of Miami; Washington Heights-  
433 Inwood Columbia Aging Project (WHICAP): The institutional review boards at Columbia  
434 University Medical Center; Epidemiology of Dementia In Singapore (EDIS): The  
435 institutional review boards at Singapore Eye Research Institute; Memory Clinic in Singapore  
436 (MCS): The institutional review boards at National University Hospital; Framingham Heart  
437 Study (FHS): The institutional review boards at Boston Medical Center.

438

439 **Consent for publication**

440 Informed consent has been obtained from all participants included in the analyzed studies.

441

442 **Availability of data and materials**

443 Studies participating in this meta-analysis have separate and specific data request and  
444 approval policies, depending on local, national, and international laws and regulations.  
445 Because of restrictions based on such privacy laws and regulations and informed consent of  
446 the participants, data cannot be made freely available in a public repository for any of the  
447 participating studies. Requests for information on procedures and formal data requests can be  
448 submitted to investigators from the corresponding author (Jose Gutierrez).

449

450 **Competing interests**

451 The authors declare that they have no competing interests.

452

453 **Funding**

454 This investigation was supported by National Institutes of Health grant R01 AG057709.  
455 The Atherosclerosis Risk in Communities (ARIC) Study is carried out as a collaborative  
456 study supported by National Heart, Lung, and Blood Institute contracts (75N92022D00001,  
457 75N92022D00002, 75N92022D00003, 75N92022D00004, 75N92022D00005). The ARIC  
458 Neurocognitive Study is supported by U01HL096812, U01HL096814, U01HL096899,  
459 U01HL096902, and U01HL096917 from the NIH (NHLBI, NINDS, NIA and NIDCD).  
460 Funding was also supported by R01AG054491, R01HL087641 and R01HL086694; National  
461 Human Genome Research Institute contract U01HG004402; and National Institutes of Health  
462 contract HHSN268200625226C. Infrastructure was partly supported by Grant Number  
463 UL1RR025005, a component of the National Institutes of Health and NIH Roadmap for  
464 Medical Research. The Northern Manhattan Study (NOMAS) was supported by National  
465 Institutes of Health (R01 AG066162, R01 NS36286, R01 NS29993). The Washington  
466 Heights–Inwood Columbia Aging Project (WHICAP) was supported by National Institutes of  
467 Health (R01 AG072474, R01 AG037212, RF1 AG054023). The Epidemiology of Dementia  
468 In Singapore (EIDS) study was supported by the National Medical Research Council,  
469 Singapore (NMRC/CG/NUHS/2010 [Grant no: R-184-006-184-511]). The Memory Clinic in  
470 Singapore (MCS) was supported by U01 AG052409. The Framingham Heart Study (FHS)  
471 was supported by National Heart, Lung and Blood Institute contracts (N01 HC25195,

472 HHSN268201500001I, 75N92019D00031) with additional support from National Institutes  
473 of Health grants (R01 AG047645, R01 HL131029) and an American Heart Association  
474 Award (15GPGGC24800006).

475

#### 476 **Authors' contributions**

477 ML performed statistical analyses, and drafted and revised the manuscript. FK, SS, AS, YQ,  
478 ET, SS, JR participated in data acquisition and revised the manuscript. DS, HA, QY, AB,  
479 SH, TR, JJ, GT participated in data analysis and interpretation and revised the manuscript.  
480 AB, ME, RS, CC, BW, MF were responsible for obtaining funding and revising the  
481 manuscript. JG was responsible for the study concept and design, obtaining funding, and  
482 drafting and revising the manuscript. All authors read and approved the final manuscript.

483

#### 484 **Acknowledgements**

485 The authors thank the staff and participants of the ARIC, NOMAS, WHICAP, EDIS, MCS  
486 and FHS studies for their important contributions.

487

#### 488 **References**

489

- 490 1. Morgagni G. *De sedibus et causis morborum per anatomen indigatis libri quinque. Venice: ex*  
491 *typographica Remondiana.* 1761.
- 492 2. Gutierrez J, Sacco RL, Wright CB. Dolichoectasia-an evolving arterial disease. *Nat Rev Neurol.*  
493 2011;7(1):41-50.
- 494 3. Smoker WR, Corbett JJ, Gentry LR, Keyes WD, Price MJ, McKusker S. High-resolution  
495 computed tomography of the basilar artery: 2. Vertebrobasilar dolichoectasia: clinical-pathologic  
496 correlation and review. *AJNR Am J Neuroradiol.* 1986;7(1):61-72.
- 497 4. Gutierrez J, Bagci A, Gardener H, et al. Dolichoectasia diagnostic methods in a multi-ethnic,  
498 stroke-free cohort: results from the northern Manhattan study. *J Neuroimaging.* 2014;24(3):226-  
499 231.
- 500 5. Gutierrez J, Cheung K, Bagci A, et al. Brain Arterial Diameters as a Risk Factor for Vascular  
501 Events. *Journal of the American Heart Association.* 2015;4(8):e002289.
- 502 6. Gutierrez J, Guzman V, Khasiyev F, et al. Brain arterial dilatation and the risk of Alzheimer's  
503 disease. *Alzheimers Dement.* 2019;15(5):666-674.
- 504 7. Gutierrez J, Kulick E, Park Moon Y, et al. Brain Arterial Diameters and Cognitive Performance:  
505 The Northern Manhattan Study. *J Int Neuropsychol Soc.* 2017:1-12.
- 506 8. Pasterkamp G, Schoneveld AH, van Wolferen W, et al. The impact of atherosclerotic arterial  
507 remodeling on percentage of luminal stenosis varies widely within the arterial system. A  
508 postmortem study. *Arterioscler Thromb Vasc Biol.* 1997;17(11):3057-3063.
- 509 9. Xu WH, Li ML, Gao S, et al. In vivo high-resolution MR imaging of symptomatic and  
510 asymptomatic middle cerebral artery atherosclerotic stenosis. *Atherosclerosis.* 2010;212(2):507-  
511 511.
- 512 10. Groenink M, den Hartog AW, Franken R, et al. Losartan reduces aortic dilatation rate in adults  
513 with Marfan syndrome: a randomized controlled trial. *Eur Heart J.* 2013;34(45):3491-3500.
- 514 11. Morris SA, Orbach DB, Geva T, Singh MN, Gauvreau K, Lacro RV. Increased vertebral artery  
515 tortuosity index is associated with adverse outcomes in children and young adults with connective  
516 tissue disorders. *Circulation.* 2011;124(4):388-396.
- 517 12. Naunheim MR, Walcott BP, Nahed BV, MacRae CA, Levinson JR, Ogilvy CS. Arterial  
518 tortuosity syndrome with multiple intracranial aneurysms: a case report. *Arch Neurol.*  
519 2011;68(3):369-371.



- 520 13. Gutierrez J, Bagci A, Del Brutto V, et al. Correlates of Dolichoectasia in an Urban, Stroke-free  
521 Cohort: Results From the Northern Manhattan Study. *Stroke*. 2014;45(Suppl 1):ATP152.
- 522 14. Gutierrez J, DiTullio M, K Cheung YK, et al. Brain arterial dilatation modifies the association  
523 between extracranial pulsatile hemodynamics and brain perivascular spaces: the Northern  
524 Manhattan Study. *Hypertension Research*. 2019.
- 525 15. Wright JD, Folsom AR, Coresh J, et al. The ARIC (Atherosclerosis Risk In Communities) Study.  
526 *Journal of the American College of Cardiology*. 2021;77(23):2939-2959.
- 527 16. Knopman DS, Gottesman RF, Sharrett AR, et al. Mild Cognitive Impairment and Dementia  
528 Prevalence: The Atherosclerosis Risk in Communities Neurocognitive Study (ARIC-NCS).  
529 *Alzheimer's & dementia (Amsterdam, Netherlands)*. 2016;2:1-11.
- 530 17. Manly JJ, Bell-McGinty S, Tang MX, Schupf N, Stern Y, Mayeux R. Implementing diagnostic  
531 criteria and estimating frequency of mild cognitive impairment in an urban community. *Archives*  
532 *of neurology*. 2005;62(11):1739-1746.
- 533 18. Foong AW, Saw SM, Loo JL, et al. Rationale and methodology for a population-based study of  
534 eye diseases in Malay people: The Singapore Malay eye study (SiMES). *Ophthalmic*  
535 *epidemiology*. 2007;14(1):25-35.
- 536 19. Lavanya R, Jeganathan VS, Zheng Y, et al. Methodology of the Singapore Indian Chinese Cohort  
537 (SICC) eye study: quantifying ethnic variations in the epidemiology of eye diseases in Asians.  
538 *Ophthalmic epidemiology*. 2009;16(6):325-336.
- 539 20. Wong TY, Chong EW, Wong WL, et al. Prevalence and causes of low vision and blindness in an  
540 urban malay population: the Singapore Malay Eye Study. *Archives of ophthalmology (Chicago,*  
541 *Ill : 1960)*. 2008;126(8):1091-1099.
- 542 21. Zheng Y, Lavanya R, Wu R, et al. Prevalence and causes of visual impairment and blindness in  
543 an urban Indian population: the Singapore Indian Eye Study. *Ophthalmology*. 2011;118(9):1798-  
544 1804.
- 545 22. Suinesiaputra A, de Koning PJ, Zudilova-Seinstra E, Reiber JH, van der Geest RJ. Automated  
546 quantification of carotid artery stenosis on contrast-enhanced MRA data using a deformable  
547 vascular tube model. *Int J Cardiovasc Imaging*. 2012;28(6):1513-1524.
- 548 23. Qiao Y, Guallar E, Suri FK, et al. MR Imaging Measures of Intracranial Atherosclerosis in a  
549 Population-based Study. *Radiology*. 2016;280(3):860-868.
- 550 24. Gutierrez J, Khasiyev F, Liu M, et al. Determinants and Outcomes of Asymptomatic Intracranial  
551 Atherosclerotic Stenosis. *J Am Coll Cardiol*. 2021;78(6):562-571.
- 552 25. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide  
553 association scans. *Bioinformatics*. 2010;26(17):2190-2191.
- 554 26. Magi R, Horikoshi M, Sofer T, et al. Trans-ethnic meta-regression of genome-wide association  
555 studies accounting for ancestry increases power for discovery and improves fine-mapping  
556 resolution. *Human molecular genetics*. 2017;26(18):3639-3650.
- 557 27. Turley P, Walters RK, Maghzian O, et al. Multi-trait analysis of genome-wide association  
558 summary statistics using MTAG. *Nature genetics*. 2018;50(2):229-237.
- 559 28. de Leeuw CA, Mooij JM, Heskes T, Posthuma D. MAGMA: generalized gene-set analysis of  
560 GWAS data. *PLoS Comput Biol*. 2015;11(4):e1004219.
- 561 29. Watanabe K, Taskesen E, van Bochoven A, Posthuma D. Functional mapping and annotation of  
562 genetic associations with FUMA. *Nat Commun*. 2017;8(1):1826.
- 563 30. Consortium GT. Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis:  
564 multitissue gene regulation in humans. *Science*. 2015;348(6235):648-660.
- 565 31. Liberzon A, Birger C, Thorvaldsdottir H, Ghandi M, Mesirov JP, Tamayo P. The Molecular  
566 Signatures Database (MSigDB) hallmark gene set collection. *Cell Syst*. 2015;1(6):417-425.
- 567 32. Slenter DN, Kutmon M, Hanspers K, et al. WikiPathways: a multifaceted pathway database  
568 bridging metabolomics to other omics research. *Nucleic Acids Res*. 2018;46(D1):D661-D667.

- 569 33. Buniello A, MacArthur JAL, Cerezo M, et al. The NHGRI-EBI GWAS Catalog of published  
570 genome-wide association studies, targeted arrays and summary statistics 2019. *Nucleic Acids Res.*  
571 2019;47(D1):D1005-D1012.
- 572 34. Kircher M, Witten DM, Jain P, O'Roak BJ, Cooper GM, Shendure J. A general framework for  
573 estimating the relative pathogenicity of human genetic variants. *Nature genetics.* 2014;46(3):310-  
574 315.
- 575 35. Zhu Z, Zhang F, Hu H, et al. Integration of summary data from GWAS and eQTL studies predicts  
576 complex trait gene targets. *Nature genetics.* 2016;48(5):481-487.
- 577 36. Qi T, Wu Y, Zeng J, et al. Identifying gene targets for brain-related traits using transcriptomic  
578 and methylomic data from blood. *Nat Commun.* 2018;9(1):2282.
- 579 37. Bellenguez C, Küçükali F, Jansen IE, et al. New insights into the genetic etiology of Alzheimer's  
580 disease and related dementias. *Nature genetics.* 2022;54(4):412-436.
- 581 38. Malik R, Rannikmäe K, Traylor M, et al. Genome-wide meta-analysis identifies 3 novel loci  
582 associated with stroke. *Ann Neurol.* 2018;84(6):934-939.
- 583 39. Elliott LT, Sharp K, Alfaro-Almagro F, et al. Genome-wide association studies of brain imaging  
584 phenotypes in UK Biobank. *Nature.* 2018;562(7726):210-216.
- 585 40. Davies NM, Hill WD, Anderson EL, Sanderson E, Deary IJ, Davey Smith G. Multivariable two-  
586 sample Mendelian randomization estimates of the effects of intelligence and education on health.  
587 *eLife.* 2019;8.
- 588 41. Hemani G, Zheng J, Elsworth B, et al. The MR-Base platform supports systematic causal  
589 inference across the human phenome. *eLife.* 2018;7.
- 590 42. Volpe SL. Magnesium in disease prevention and overall health. *Advances in nutrition (Bethesda,*  
591 *Md).* 2013;4(3):378s-383s.
- 592 43. Accogli A, Scala M, Calcagno A, et al. CNNM2 homozygous mutations cause severe refractory  
593 hypomagnesemia, epileptic encephalopathy and brain malformations. *Eur J Med Genet.*  
594 2019;62(3):198-203.
- 595 44. Stuiver M, Lainez S, Will C, et al. CNNM2, encoding a basolateral protein required for renal  
596 Mg<sup>2+</sup> handling, is mutated in dominant hypomagnesemia. *American journal of human genetics.*  
597 2011;88(3):333-343.
- 598 45. Steen H, Lindholm D. Nuclear localized protein-1 (Nulp1) increases cell death of human  
599 osteosarcoma cells and binds the X-linked inhibitor of apoptosis protein. *Biochemical and*  
600 *biophysical research communications.* 2008;366(2):432-437.
- 601 46. Duarte RRR, Bachtel ND, Cotel MC, et al. The Psychiatric Risk Gene NT5C2 Regulates  
602 Adenosine Monophosphate-Activated Protein Kinase Signaling and Protein Translation in  
603 Human Neural Progenitor Cells. *Biol Psychiatry.* 2019;86(2):120-130.
- 604 47. Singgih EL, van der Voet M, Schimmel-Naber M, Brinkmann EL, Schenck A, Franke B.  
605 Investigating cytosolic 5'-nucleotidase II family genes as candidates for neuropsychiatric  
606 disorders in Drosophila (114/150 chr). *Translational psychiatry.* 2021;11(1):55.
- 607 48. Itoh R. Enzymatic properties and physiological roles of cytosolic 5'-nucleotidase II. *Curr Med*  
608 *Chem.* 2013;20(34):4260-4284.
- 609 49. Hoffman GE, Schrode N, Flaherty E, Brennand KJ. New considerations for hiPSC-based models  
610 of neuropsychiatric disorders. *Mol Psychiatry.* 2019;24(1):49-66.
- 611 50. Tang Y, Illes P. Regulation of adult neural progenitor cell functions by purinergic signaling. *Glia.*  
612 2017;65(2):213-230.
- 613 51. Tornroth-Horsefield S, Neutze R. Opening and closing the metabolite gate. *Proceedings of the*  
614 *National Academy of Sciences of the United States of America.* 2008;105(50):19565-19566.
- 615 52. Kulkarni SS, Karlsson HK, Szekeres F, Chibalin AV, Krook A, Zierath JR. Suppression of 5'-  
616 nucleotidase enzymes promotes AMP-activated protein kinase (AMPK) phosphorylation and  
617 metabolism in human and mouse skeletal muscle. *J Biol Chem.* 2011;286(40):34567-34574.
- 618 53. Chen X, Zhang Z, Wang X, Chen Y, Wang C. NT5C2 Gene Polymorphisms and the Risk of  
619 Coronary Heart Disease. *Public health genomics.* 2020;23(3-4):90-99.

- 620 54. Li Z, Jiang J, Long J, et al. The rs11191580 variant of the NT5C2 gene is associated with  
621 schizophrenia and symptom severity in a South Chinese Han population: evidence from GWAS.  
622 *Revista brasileira de psiquiatria (Sao Paulo, Brazil : 1999)*. 2017;39(2):104-109.
- 623 55. Vishnolia KK, Hoene C, Tarhbalouti K, Revenstorff J, Aherrahrou Z, Erdmann J. Studies in  
624 Zebrafish Demonstrate That CNNM2 and NT5C2 Are Most Likely the Causal Genes at the Blood  
625 Pressure-Associated Locus on Human Chromosome 10q24.32. *Front Cardiovasc Med*.  
626 2020;7:135.
- 627 56. Lin S, Shi Q, Nix FB, et al. A novel S-adenosyl-L-methionine:arsenic(III) methyltransferase from  
628 rat liver cytosol. *J Biol Chem*. 2002;277(13):10795-10803.
- 629 57. Liu WS, Wang XY, Lu J, et al. Polymorphisms in arsenic (+3 oxidation state)  
630 methyltransferase (AS3MT) predict the occurrence of hyperleukocytosis and arsenic metabolism  
631 in APL patients treated with As(2)O(3). *Archives of toxicology*. 2020;94(4):1203-1213.
- 632 58. Su AI, Cooke MP, Ching KA, et al. Large-scale analysis of the human and mouse transcriptomes.  
633 *Proceedings of the National Academy of Sciences of the United States of America*.  
634 2002;99(7):4465-4470.
- 635 59. Li M, Jaffe AE, Straub RE, et al. A human-specific AS3MT isoform and BORCS7 are molecular  
636 risk factors in the 10q24.32 schizophrenia-associated locus. *Nat Med*. 2016;22(6):649-656.
- 637 60. Zhao W, Zhang Q, Chen X, et al. The VNTR of the AS3MT gene is associated with brain  
638 activations during a memory span task and their training-induced plasticity. *Psychological  
639 medicine*. 2021;51(11):1927-1932.
- 640 61. Duarte RRR, Troakes C, Nolan M, Srivastava DP, Murray RM, Bray NJ. Genome-wide  
641 significant schizophrenia risk variation on chromosome 10q24 is associated with altered cis-  
642 regulation of BORCS7, AS3MT, and NT5C2 in the human brain. *American journal of medical  
643 genetics Part B, Neuropsychiatric genetics: the official publication of the International Society of  
644 Psychiatric Genetics*. 2016;171(6):806-814.
- 645 62. Guan F, Zhang T, Li L, et al. Two-stage replication of previous genome-wide association studies  
646 of AS3MT-CNNM2-NT5C2 gene cluster region in a large schizophrenia case-control sample  
647 from Han Chinese population. *Schizophrenia research*. 2016;176(2-3):125-130.
- 648 63. Navone F, Genevini P, Borgese N. Autophagy and Neurodegeneration: Insights from a Cultured  
649 Cell Model of ALS. *Cells*. 2015;4(3):354-386.
- 650 64. Menzies FM, Fleming A, Caricasole A, et al. Autophagy and Neurodegeneration: Pathogenic  
651 Mechanisms and Therapeutic Opportunities. *Neuron*. 2017;93(5):1015-1034.
- 652 65. Pu J, Schindler C, Jia R, Jarnik M, Backlund P, Bonifacino JS. BORC, a multisubunit complex  
653 that regulates lysosome positioning. *Developmental cell*. 2015;33(2):176-188.
- 654 66. Jia R, Guardia CM, Pu J, Chen Y, Bonifacino JS. BORC coordinates encounter and fusion of  
655 lysosomes with autophagosomes. *Autophagy*. 2017;13(10):1648-1663.
- 656 67. Snouwaert JN, Church RJ, Jania L, et al. A Mutation in the Borcs7 Subunit of the Lysosome  
657 Regulatory BORC Complex Results in Motor Deficits and Dystrophic Axonopathy in Mice. *Cell  
658 reports*. 2018;24(5):1254-1265.
- 659 68. Mitra AK, Crews KR, Pounds S, et al. Genetic variants in cytosolic 5'-nucleotidase II are  
660 associated with its expression and cytarabine sensitivity in HapMap cell lines and in patients with  
661 acute myeloid leukemia. *J Pharmacol Exp Ther*. 2011;339(1), 9-23.
- 662  
663  
664

## Tables

Table 1 Demographic information of studies

Cohort	ARIC	NOMAS	WHICAP	EDIS	MCS	FHS
N	1565	1092	290	647	350	206
Age, years, mean (sd)	75.8 (5.3)	70.1 (8.4)	77.2 (6.5)	70.2 (6.6)	71.0 (8.2)	72.7 (11.7)
Female (%)	58.8	60.5	64.1	51.7	55.7	54.3
Self-reported racial/ethnic identity or geographic ancestry (%)						
European/white	74.3	13.8	44.8	-	-	100.0
African/black/African American	25.7	16.6	55.2	-	-	-
Hispanic	-	69.6	-	-	-	-
Asian	-	-	-	100.0	100.0	-
Hypertension (%)	74.9	68.1	63.1	80.8	67.8	68.4
Diabetes (%)	33.9	19.0	20.2	37.3	31.8	23.9
Dyslipidemia (%)	56.0	45.0	33.9	76.0	71.4	51.6
Smoking (%)	6.0	52.7	5.2	27.9	7.1	9.3

ARIC, Atherosclerosis Risk in Communities study; NOMAS, The Northern Manhattan Study; WHICAP, Washington Heights-Inwood Community Aging Project; EDIS, Epidemiology of Dementia In Singapore; MCS, Memory Clinic in Singapore; FHS, Framingham Heart Study.

Table 2 Variants ( $P < 5 \times 10^{-8}$ ) associated with brain arterial diameter

SNP	Chr	Position (hg 19)	Nearest gene	Relation to gene	Allele1	Allele2	AF	beta	se	P value	CADD
<b>Global</b>											
rs7921574	10	104840970	CNNM2	ncRNA_intronic	C	T	0.42	0.06	0.01	1.54E-08	0.33
rs943035	10	104839152	CNNM2	ncRNA_intronic	C	T	0.42	0.06	0.01	1.57E-08	0.71
rs10883826*	10	104830819	CNNM2	ncRNA_intronic	G	A	0.42	0.06	0.01	1.58E-08	2.25
rs943036	10	104836047	CNNM2	ncRNA_intronic	C	T	0.42	0.06	0.01	1.66E-08	5.85
rs10883824	10	104812897	CNNM2	ncRNA_intronic	G	A	0.42	0.06	0.01	2.12E-08	2.98
rs8139	10	104848123	CNNM2	ncRNA_intronic	A	G	0.42	0.06	0.01	2.31E-08	7.82
rs3740387	10	104849468	CNNM2	ncRNA_intronic	A	G	0.42	0.06	0.01	2.33E-08	8.71
rs10883817	10	104755431	CNNM2	ncRNA_intronic	A	G	0.42	0.06	0.01	2.82E-08	0.37
rs3902934	10	104746649	CNNM2	ncRNA_intronic	G	A	0.42	0.06	0.01	2.84E-08	1.24
rs10883823	10	104812331	CNNM2	ncRNA_intronic	C	T	0.42	0.06	0.01	3.35E-08	0.25
rs7911789	10	104756374	CNNM2	ncRNA_intronic	C	T	0.42	0.06	0.01	3.84E-08	0.58
rs67908413	10	104764989	CNNM2	ncRNA_intronic	C	T	0.42	0.06	0.01	3.84E-08	6.84
rs1890184	10	104748459	CNNM2	ncRNA_intronic	C	A	0.42	0.06	0.01	4.26E-08	0.62
rs10786733	10	104794947	CNNM2	ncRNA_intronic	A	G	0.42	0.06	0.01	4.84E-08	4.21
<b>Posterior</b>											
rs35994878	16	89949033	TCF25	Intronic	C	T	0.29	0.11	0.02	2.94E-08	1.06
rs8061025	16	89948397	TCF25	Intronic	A	G	0.29	0.11	0.02	4.49E-08	0.42

\* Independent variant associated with brain arterial diameter. Nearest gene with a functional protein or RNA product that either overlaps with the variant or for intergenic variants, the nearest genes up- and downstream, respectively. The statistics are based on Allele1. Allele1 indicates effect allele, allele2 is another allele. AF: allele 1 frequency. CADD, combined annotation dependent depletion score. Chr, chromosome.



Table 3 Trans-ancestry Genome-Wide Significant Associations

SNP	Chr	Position (hg 19)	Nearest gene	Relation to gene	Allele1	Allele2	AF	P value	P_Het ANCS	P_Res Het	CADD
<b>Global</b>											
rs10883805	10	104708251	CNNM2	ncRNA_intronic	C	T	0.43	1.88E-08	0.003	0.333	4.39
rs10748839	10	104953547	NT5C2	upstream	C	T	0.42	2.54E-08	0.001	0.526	3.34
rs3902934	10	104746649	CNNM2	ncRNA_intronic	G	A	0.42	4.55E-08	0.026	0.329	1.24
rs10883814	10	104737404	CNNM2	ncRNA_intronic	C	T	0.43	4.79E-08	0.004	0.324	3.01
rs12569617	10	104729996	CNNM2	ncRNA_intronic	C	T	0.43	4.85E-08	0.004	0.323	2.87
rs10786721	10	104654383	AS3MT	ncRNA_intronic	A	C	0.43	4.97E-08	0.004	0.384	4.52
<b>Anterior</b>											
rs34217249	7	960642	ADAP1	Intronic	A	G	0.20	3.11E-08	2.03E-08	0.36	2.26

Nearest gene with a functional protein or RNA product that either overlaps with the variant or for intergenic variants, the nearest genes up- and downstream, respectively. The statistics are based on Allele1. Allele1 indicates effect allele, allele2 is another allele. AF: allele 1 frequency. CADD, combined annotation dependent depletion score. Chr, chromosome. P\_Het ANCS P-value for heterogeneity correlated with ancestry. P\_Res Het is the residual heterogeneity.

Table 4 Top 10 Co-localization of brain arterial diameter GWAS and eQTL associations

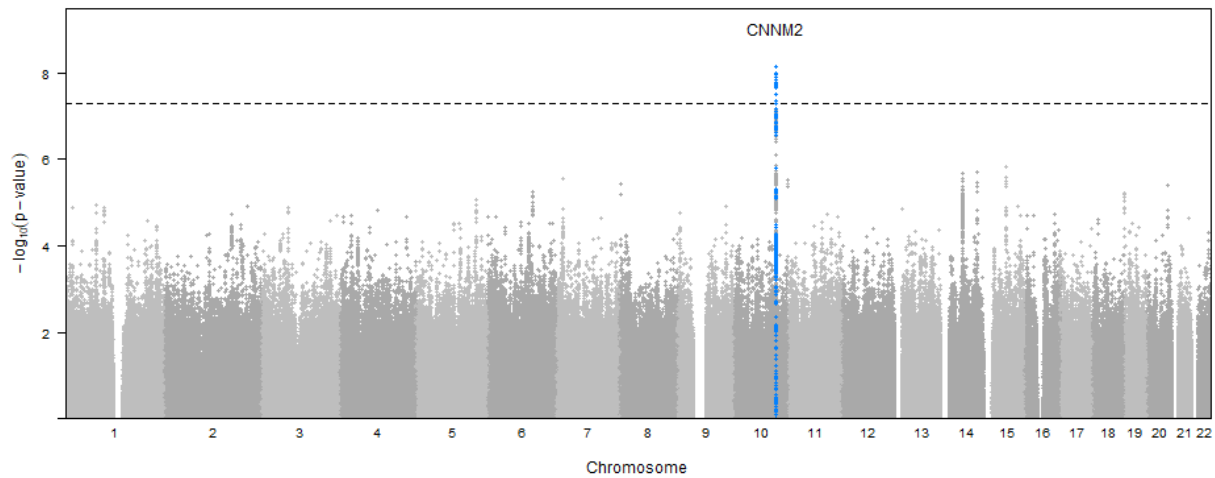
Probe ID	Probe Chr	Gene	Probe_bp	SNP	SNP chr	SNP bp	Allele1	Allele2	AF	beta	se	p
<b>Global</b>												
ENSG00000148842.18	10	CNNM2	104764015	rs7897654	10	104662458	C	T	0.28	0.12	0.02	6.17E-07*
ENSG00000272912.1	10	AL356608.1	104674752	rs10786719	10	104637992	G	A	0.41	-0.17	0.04	6.60E-06*
ENSG00000137760.15	11	ALKBH8	107404960	rs2037827	11	107408592	C	T	0.21	-0.11	0.03	3.22E-04
ENSG00000186715.11	1	MST1L	17089068	rs7513616	1	17298496	G	A	0.43	-0.06	0.02	6.88E-04
ENSG00000140265.12	15	ZSCAN29	43656796	rs523156	15	43811843	G	C	0.53	0.08	0.03	7.75E-04
ENSG00000014123.10	6	UFL1	96986312	rs11153023	6	96968525	T	C	0.15	-0.12	0.03	8.19E-04
ENSG00000237624.1	1	OXCT2P1	39981395	rs12028034	1	40039707	A	G	0.24	0.08	0.02	8.29E-04
ENSG00000159363.19	1	ATP13A2	17325438	rs7513616	1	17298496	G	A	0.43	0.09	0.03	8.53E-04
ENSG00000065717.15	19	TLE2	3022635	rs11150	19	2997897	A	G	0.17	-0.17	0.05	9.46E-04
ENSG00000268869.6	1	ESPNP	17030243	rs7513616	1	17298496	G	A	0.43	-0.12	0.04	1.36E-03
<b>Anterior</b>												
ENSG00000272912.1	10	AL356608.1	104674752	rs10786719	10	104637992	G	A	0.41	-0.17	0.04	4.98E-05
ENSG00000148842.18	10	CNNM2	104764015	rs7897654	10	104662458	C	T	0.28	0.10	0.02	6.62E-05
ENSG00000162669.16	1	HFM1	91798368	rs17131417	1	91848784	T	C	0.11	0.06	0.02	7.78E-05
ENSG00000162461.8	1	SLC25A34	16065320	rs41393951	1	16053493	A	G	0.31	-0.05	0.01	3.75E-04
ENSG00000137760.15	11	ALKBH8	107404960	rs2037827	11	107408592	C	T	0.21	-0.11	0.03	3.79E-04
ENSG00000065060.17	6	UHRF1BP1	34805354	rs6906129	6	34801160	C	T	0.51	-0.05	0.02	4.60E-04
ENSG00000156052.11	9	GNAQ	80488870	rs4582625	9	80520544	C	T	0.26	-0.08	0.02	6.44E-04
ENSG00000257354.2	12	AC048341.1	63006262	rs17731893	12	63013773	A	G	0.18	0.27	0.08	6.70E-04
ENSG00000231305.4	3	AC112484.1	128584923	rs789217	3	128593201	A	G	0.26	0.04	0.01	7.64E-04
ENSG00000016402.13	6	IL2ORA	137343712	rs9494644	6	137403294	G	C	0.31	0.21	0.06	7.80E-04
<b>Posterior</b>												
ENSG00000138111.14	10	MFSD13A	104228977	rs11593583	10	104228149	G	A	0.56	-0.16	0.04	1.23E-04
ENSG00000168386.18	3	FILIP1L	99691171	rs6809988	3	99656615	A	G	0.22	0.13	0.03	2.75E-04
ENSG00000213903.9	14	LTB4R	24783949	rs11158632	14	24769663	G	T	0.22	-0.23	0.06	4.01E-04
ENSG00000148842.18	10	CNNM2	104764015	rs7897654	10	104662458	C	T	0.28	0.11	0.03	4.11E-04

ENSG00000223959.8	16	AFG3L1P	90053782	rs2270459	16	89979851	A	C	0.11	-0.21	0.06	4.48E-04
ENSG00000111615.14	12	KRR1	75895030	rs2070162	12	75900588	G	A	0.25	-0.06	0.02	4.60E-04
ENSG00000214043.8	12	LINC02347	126940360	rs17577161	12	126842742	G	A	0.34	0.06	0.02	5.97E-04
ENSG00000255595.5	12	AC007368.1	126797235	rs17577161	12	126842742	G	A	0.34	-0.07	0.02	6.18E-04
ENSG00000258839.4	16	MC1R	89982954	rs2270459	16	89979851	A	C	0.11	0.22	0.06	6.36E-04
ENSG00000256310.1	12	NDUFA5P6	127008983	rs17577161	12	126842742	G	A	0.34	0.09	0.02	6.78E-04

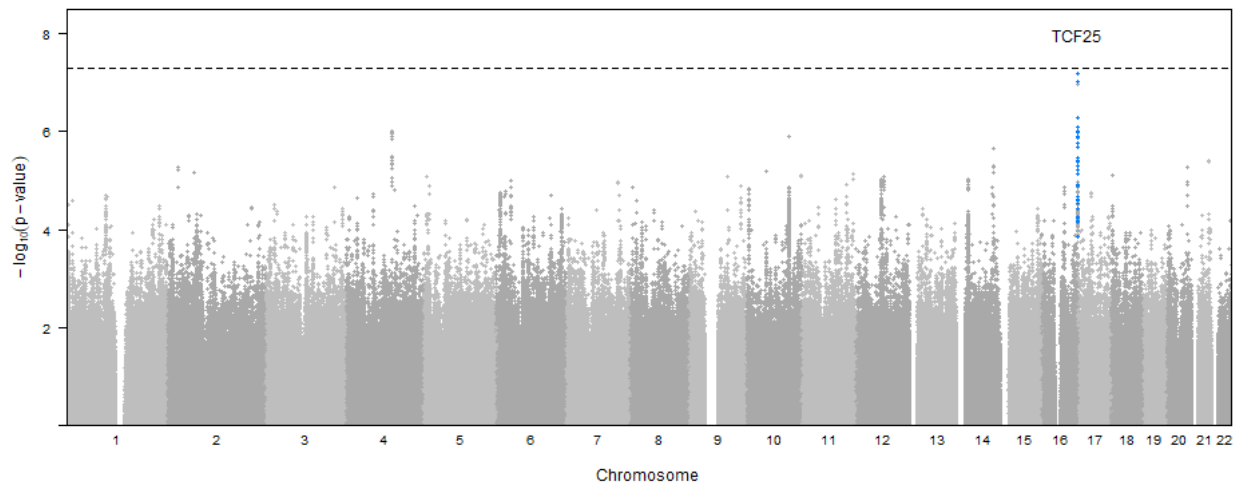
Allele1 indicates effect allele, allele2 is another allele. AF: allele 1 frequency. \*Significant functional genes at P value < 8.4E-06

## Figures

A



B



C

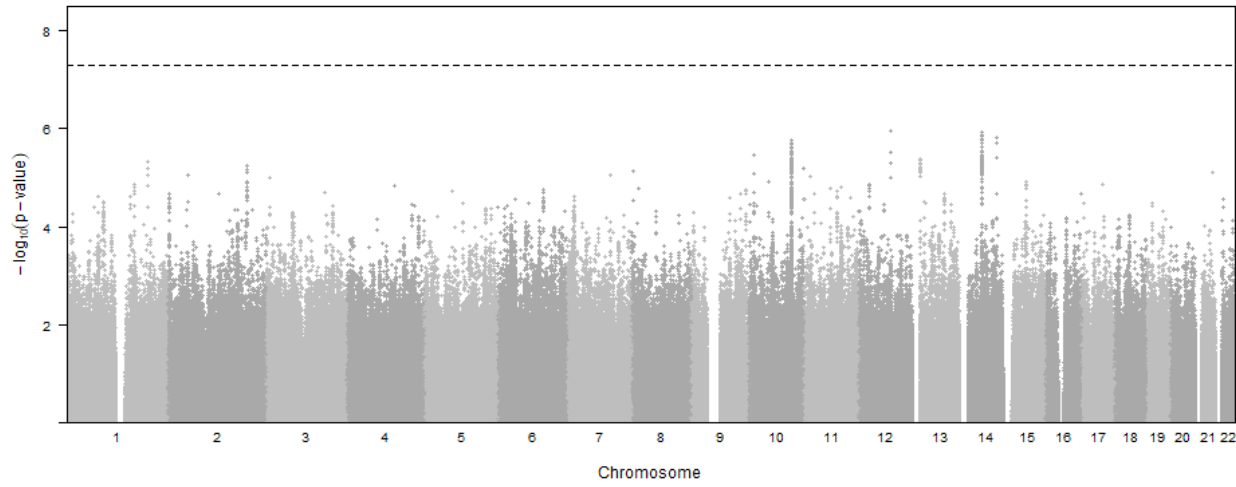
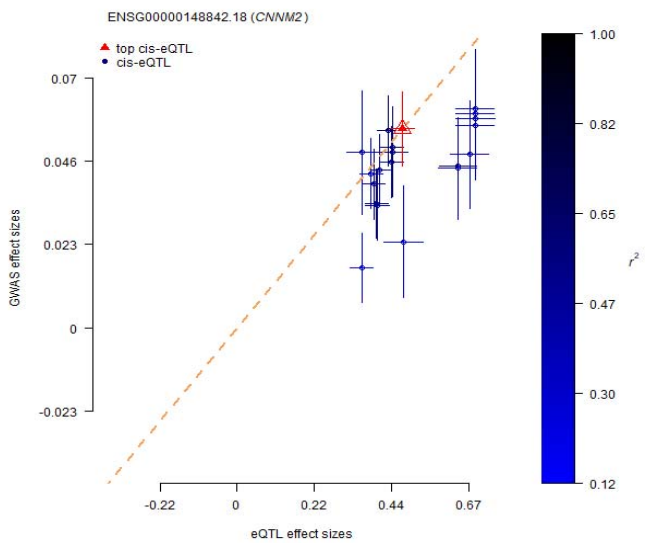
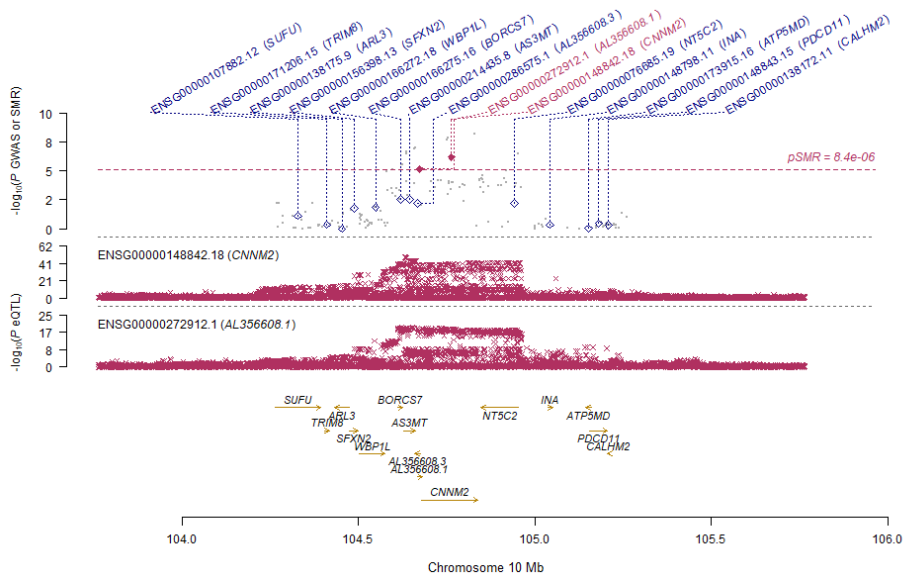


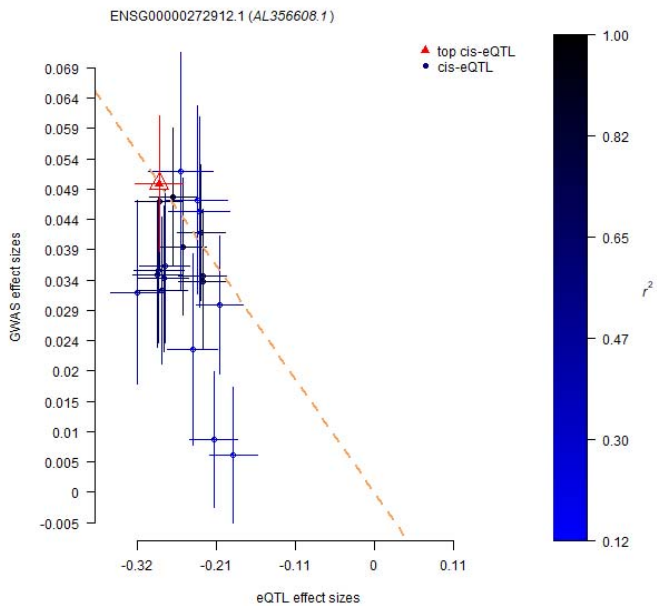
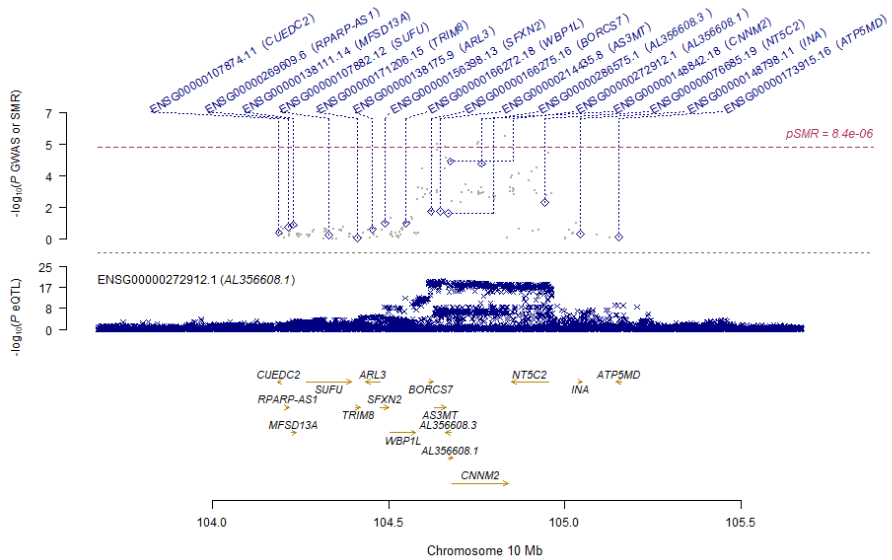
Figure 1 Genome-wide associations in brain arterial diameter. Manhattan plots for brain arterial diameter show combined genome-wide associations from 6 population-based studies.



A



B



C

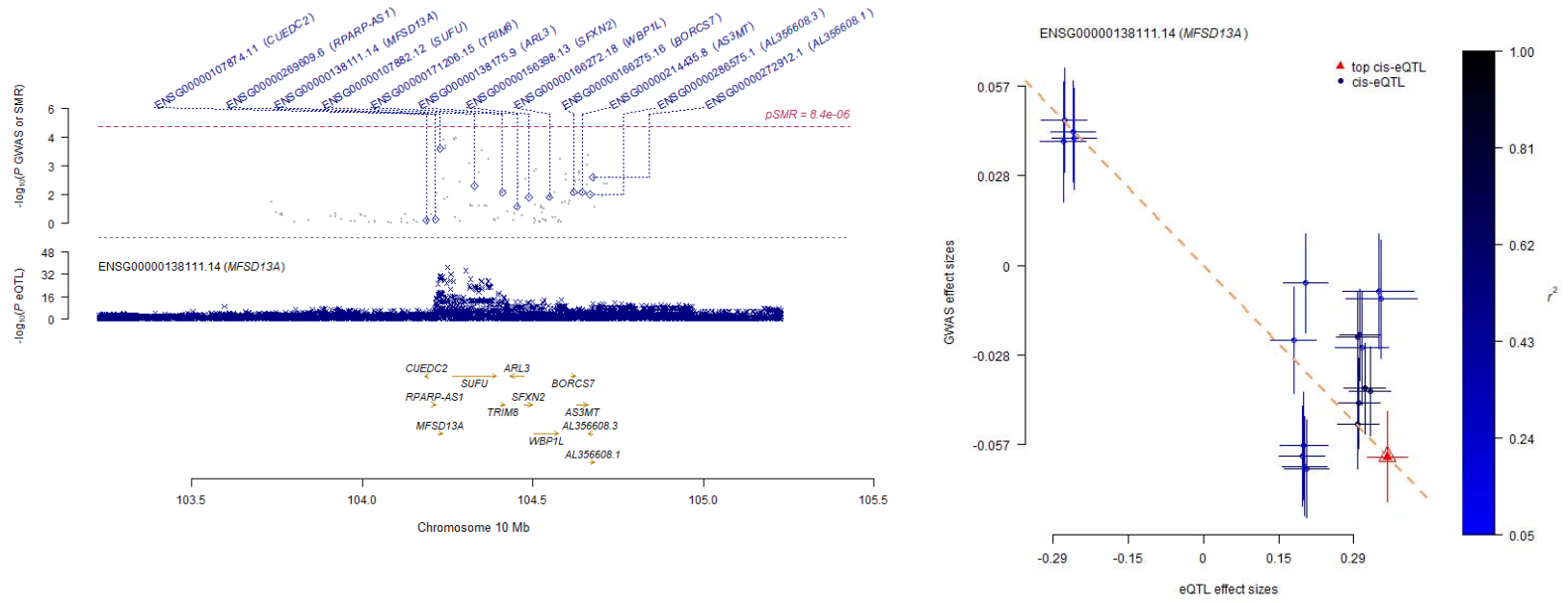


Figure 2 Locus plot and effect sizes plot of genome-wide association studies (GWAS) and expression quantitative trait locus (eQTL) associations.