

# Circulating semicarbazide-sensitive amine oxidase is raised both in Type I (insulin-dependent), in Type II (non-insulin-dependent) diabetes mellitus and even in childhood Type I diabetes at first clinical diagnosis

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**Summary** Plasma semicarbazide-sensitive amine oxidase is raised in patients with Type I (insulin-dependent) diabetes mellitus. It has been suggested that this enzyme is involved in the development of microvascular damage through its ability to convert amines (e.g. methylamine and aminoacetone) into aldehydes, hydrogen peroxide and ammonia. Plasma semicarbazide-sensitive amine oxidase was found to be equally raised both in patients with Type I diabetes (n = 73) and Type II (non-insulin-dependent) diabetes mellitus (n = 88) compared with control subjects  $(621 \pm 209 \text{ and } 619 \pm 202 \text{ vs } 352 \pm 102 \text{ mU/l},$ p < 0.0001) and to correlate in multiple regression analysis with HbA<sub>1c</sub>. Since the enzyme could protect the islets from the inhibitory effects of methylamine on insulin secretion, we also tested sera of 100 children, collected consecutively at first diagnosis of Type I diabetes, for semicarbazide-sensitive amine oxidase. The activity was greatly increased compared with serum values of 76 control (siblings) children  $(757 \pm 300 \text{ vs } 455 \pm 138 \text{ mU/l}, p < 0.0001)$ , but not associated with HbA<sub>1c</sub>. Our study confirms the increase of plasma semicarbazide-sensitive amine oxidase in Type I diabetes and extends this finding to Type II diabetes as well as to childhood Type I at first clinical diagnosis. In the last case increased enzyme activities could serve to protect the islets from inhibitory effects of methylamine but cause damage by generation of hydrogen peroxide, aldehydes and ammonia. In the long run the increased enzyme activities could also contribute to vascular damage by direct cytotoxic action on endothelial cells, including increased oxidative stress and glycosylation of proteins. [Diabetologia (1999) 42: 233–237]

**Keywords** Type I diabetes, Type II diabetes, semicarbazide-sensitive amine oxidase, methylamine.

In patients with either Type I (insulin-dependent) diabetes mellitus or Type II (non-insulin-dependent) diabetes mellitus, progression of the disease is often accompanied by serious microvascular complications leading to nephropathy, retinopathy and neuropathy. In this context it has been suggested that the semicarbazide-sensitive amine oxidase (SSAO) enzyme

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Corresponding author: Dr. F. Boomsma, Department of Internal Medicine I, University Hospital Dijkzigt, Dr. Molewaterplein 40, 3015 GD Rotterdam, The Netherlands Abbreviations: SSAO, Semicarbazide-sensitive amine oxidase; GAD, glutamic acid decarboxylase; CI, confidence interval.

plays a part in the development of microvascular damage [1].

Semicarbazide-sensitive amine oxidases are a group of heterogeneous enzymes and are present in various mammalian tissues (especially vascular smooth muscle cells of arteries) and also in plasma [2–9]. They readily convert endogenous methylamine and aminoacetone to the corresponding cytotoxic aldehydes formaldehyde and methylglyoxal, while generating ammonia and hydrogen peroxide [1, 7, 10–13]. The formation of the reaction compounds can cause damage to endothelial cells in three different ways: 1) directly, presumably mostly by direct action of hydrogen peroxide; 2) by reaction of aldehydes with proteins leading to AGE's; and 3) by the formation of oxidative radicals. In Type I diabetes

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plasma SSAO was increased compared with control subjects; the increase was greater in patients with, than in those without, microvascular complications [14]. No data on plasma SSAO activity in Type II diabetes are as yet available.

The SSAO substrate, methylamine, is an inhibitor of the glucose-stimulated insulin release [15,16]. Plasma SSAO becomes increased gradually in rats made diabetic with streptozotocin or alloxan, but not in those resistant to the diabetogenic action of alloxan [17]. We therefore examined SSAO activity at the onset of diabetic symptoms. For this, blood samples were taken at first diagnosis in children with Type I diabetes where hyperglycaemia presents more abruptly than in Type II diabetic patients.

The aim of our study was to establish whether plasma SSAO is also raised in Type II diabetes and at first clinical manifestation of Type I diabetes in children.

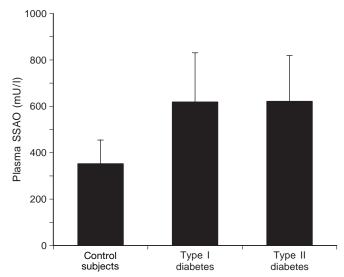
# **Subjects and methods**

Patients and controls. The investigations conformed with the principles outlined in the Declaration of Helsinki. The adults gave informed oral consent, the children over 12 years of age provided signed written consent as did the parents/guardians for those under 12 years of age.

Plasma was obtained from 161 consecutive adult diabetic patients who visited the out-patient clinic of our hospital. Of these patients 73 were classified as having Type I and 88 Type II diabetes (Table 1). The absence or presence of retinopathy (investigated by an experienced ophtalmologist), nephropathy (urinary albumin excretion > 30 mg/24 h), and neuropathy (by biothesiometry) were assessed in each patient. The Quetelet Index is calculated as bodyweight (in kg) divided by the squared height (in m). Patients with Type I diabetes mellitus were younger and had had diabetes for longer than patients with Type II diabetes. Retinopathy occurred more often in Type I and nephropathy and neuropathy more often in Type II diabetes.

Serum was obtained from 100 children (48 males) who had been consecutively diagnosed as having Type I diabetes during a regional prospective Type I diabetes family study. The patients met the World Health Organisation/American Diabetes Association criteria and had islet cell cytoplasmatic antibodies or glutamic acid decarboxylase (GAD) antibodies or both in 82 %. Their age was  $9.3 \pm 4.4$  years (range 0.4–19.5 year). In view of reports of higher SSAO activities in normal children than in adults [18, 19] and the difference between plasma and serum [20], we used 76 serum samples from siblings of the diabetic children (40 males, age  $10.9 \pm 4.8$  years, range 2.3–20.4 years) as control subjects.

Methods. The activity of SSAO was determined as described previously [20]. All plasma and serum was immediately separated and stored at –80 °C until assay. Briefly, plasma or serum, after preincubation with clorgyline (0.9 mmol/l) to inactivate possibly present monoamine oxidase, was incubated for 1 h with the SSAO-substrate benzylamine at 37 °C. The amount of benzaldehyde generated was measured by high-performance liquid chromatography with fluorimetric detection after derivatization with dimedone (5,5-dimethyl-1,3-cyclohexanedione). The SSAO activity is expressed as pmol benzaldehyde



**Fig. 1.** Plasma SSAO activities (means + SD) in control, patients with Type I, and patients with Type II diabetes

formed per ml per min, or mU/l. Normal values of SSAO in adults are  $352 \pm 102$  mU/l in plasma and values in serum are 15% lower [20].

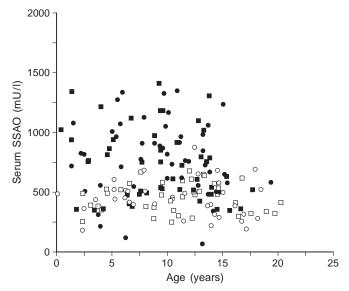
Glycosylated haemoglobin (HbA<sub>1c</sub>%) was measured by HPLC within 1 week of first clinical diagnosis in 45 of the 100 diabetic children (normal range 4.0%-6.2%). Previously we found a weak association between HbA1c at clinical onset and GAD antibodies [21]. We measured the GAD antibodies within a week of diagnosis in 43 of the 100 Type I diabetic children with a minor modification of a method described previously [22]. Briefly, 2 µl of serum was incubated overnight in a microplate with [35 S]methionine-labelled in vitro translated human recombinant GAD65. Immune complexes were isolated using protein A Sepharose (CL4B Pharmacia, Uppsala, Sweden) and washed using the multiscreen system (Millipore, Bedford, Mass., USA). Precipitated activity was counted in a microbeta-plate-reader (EG&E Wallac, Turku, Finland) and expressed relative to internal reference sera as the GAD index or titre.

Data are reported as means  $\pm$  SD. Comparison of groups was by Student's unpaired t-test, Mann Whitney U-test, and chi-squared analysis. A p-value of 0.05 or smaller was considered significant.

# **Results**

Adult Type I and Type II diabetes mellitus. Plasma SSAO was equally raised in both groups of diabetic patients compared with control subjects (p < 0.0001; Fig. 1). Plasma SSAO correlated with age in Type I diabetes (r = 0.38, p = 0.0009) and with duration of diabetes (r = 0.27, p = 0.0124) and plasma glucose (r = 0.36, p = 0.0007) in Type II diabetes. When the two diabetes types were subdivided according to the absence or presence of complications (retinopathy, nephropathy and neuropathy), no significant differences in plasma SSAO were found.

In a multiple regression analysis with (ln) SSAO as a dependent variable and type of diabetes, sex, age,



**Fig. 2.** Serum SSAO activities at first clinical diagnosis compared with age in children (n = 100) with Type I diabetes (filled symbols) and control subjects (n = 76; open symbols). Squares, males; Circles, females

**Table 1.** Characteristics of the adult patients with diabetes mellitus

	Type I diabetes	Type II diabetes	<i>p</i> -value
Number	73	87	
Sex (male/female)	36/37	43/44	NS
Age (years)	$43 \pm 15 (38)$	$63 \pm 11 (62)$	< 0.0001
QI (kg/m2)	$26 \pm 6 (25)$	$30 \pm 5 (29)$	< 0.0001
Diabetes duration (years)	$20 \pm 13 (19)$	$11 \pm 7 \ (10)$	< 0.001
Retinopathy (%)	49	39	0.0276
Background	15	21	
Proliferative	34	18	
Nephropathy (%)	22	58	< 0.0001
Neuropathy (%)	32	58	0.0027
Plasma			
Creatinine (µmol/l)	$78 \pm 22 (77)$	$81 \pm 24 (75)$	NS
Glucose (mmol/l)	$8.6 \pm 4.6 (7.4)$	$8.4 \pm 3.8 (7.2)$	NS
$HbA_{1c}(\%)$	$8.2 \pm 1.2 \ (8.2)$	$8.4 \pm 1.4 \ (8.3)$	NS

Data are means  $\pm$  SD (median)

duration of diabetes, Quetelet Index,  $HbA_{1c}$  and presence or absence of the complications retinopathy (background or proliferative), nephropathy and neuropathy as independent variables, only  $HbA_{1c}$  (t = 2.21, p = 0.029, B coefficient 0.042) and Quetelet Index (t = -2.74, p = 0.007, B coefficient 0.013) were found to be significant, whereas age approached significance (t = 1.81, p = 0.073, B coefficient 0.0044) in the adult (Type I and Type II) diabetic patients.

Children at first diagnosis of Type I diabetes mellitus and two siblings at risk. In the 100 diabetic children serum SSAO at first diagnosis was much higher than in the control subjects ( $757 \pm 300$ , confidence interval

(CI) 697–817 mU/l vs  $455 \pm 138$ , CI 424–487 mU/l, p < 0.0001). Regression analysis showed no correlation between serum SSAO and age (Fig.2). In 45 of the newly diagnosed Type I diabetic children the average HbA<sub>1c</sub> was 10.6%, median 10.3%. In these children SSAO was not associated with HbA<sub>1c</sub> (p > 0.1) nor with GAD indices (p > 0.1). Serum SSAO was also not different between boys and girls.

Two of the control children (76 siblings of the childhood patients) had raised GAD indices as well as islet cell cytoplasmatic antibodies of more than 20 Juvenile Diabetes Foundation units. Their SSAO activities were 450 and 650 mU/l, i.e. in the middle, respectively the high part of the normal SSAO range.

## **Discussion**

In the adult diabetic patients SSAO was raised both in Type I and in Type II diabetes; in fact, both groups had equally high plasma SSAO activities. The Type II diabetic group was older than the Type I diabetic group and plasma SSAO could increase a little with age [23]; in this study, however, age did not reach statistical significance in multiple regression analysis. Even compared to an older control group, however, the activity in the Type II diabetic group was clearly raised (619 vs 455 mU/l, p < 0.0001) [23]. The mean activity of 621 mU/l in Type I was a little higher than the mean activity we found previously in Type I diabetic patients without complications (486 mU/l) but similar to the activities in those with Type I with complications (581–641 mU/l) [14]. This fits in well with the higher percentage of patients with complications in this than in the earlier study. The correlation between plasma SSAO and HbA<sub>1c</sub>, found previously in prevalent Type I diabetes is confirmed in the present study. The negative association with the Quetelet Index needs to be explored otherwise [24].

The study of the Type I diabetic children shows that serum SSAO is substantially raised even at first diagnosis: 757 vs 455 mU/l. Half of the patients had serum activities higher than the means + 2SD of the control subjects. When corrected for the mean difference between plasma and serum, plasma SSAO values of the control children would be 535 mU/l, which agrees well with the value of  $554 \pm 206$  (median 534, range 47–1117) mU/l we have found in 105 plasma samples of children (58 boys, median age 6.6, range 0.1–15.1 years) collected with the specific purpose of establishing normal ranges for various plasma variables. The SSAO activities in children are thus higher than in adults, in agreement with earlier reports [18, 19] about plasma monoamine oxidase (now believed to have been SSAO) which described no differences in SSAO with age during adulthood but higher levels in children. Similarly, plasma SSAO activity in Type I diabetic children (891 mU/l, after correction for the plasma-serum difference) is higher than in diabetic adults. Within the age-range of both the control and diabetic children of our study, no correlation between age and SSAO was found.

The raised blood SSAO activities in patients with diabetes could constitute a mechanism for damage to the vascular endothelium, through conversion of the endogenous substrates methylamine and aminoacetone to cytotoxic formaldehyde, methylglyoxal, hydrogen peroxide and ammonia. Both methylamine, formed by metabolism of, e.g. creatinine and adrenaline, and aminoacetone, formed from catabolism of threonine and glycine, have been suggested to be increased in diabetes [1, 25]. The possibility that plasma SSAO increases, as a result of such damage, by leakage of SSAO from the membranes of the vascular smooth muscle cells of arteries into the plasma, has been discussed before [14]. It is notable that blood concentrations of methylglyoxal, as well as of the enzymes of the glyoxalase system breaking down methylglyoxal, are reported to be increased in diabetes and correlate with diabetic microvascular complications [26]. Besides being cytotoxic, methylglyoxal is also involved in the modification of proteins and in the formation of advanced glycosylation products. Further, the methylglyoxal scavenger aminoguanidine has been proposed as a prophylactic agent for preventive therapy of diabetic complications [27, 28]. That pancreatic islet homogenates catalyse the incorporation of radioactivity from [14 C]methylamine in N,N-dimethylcasein [15] could also possibly be attributed to transformation of methylamine by SSAO into formaldehyde. Methylglyoxal is produced by enzymatic and non-enzymatic elimination of phosphate from dihydroxyacetone phosphate and glyceraldehyde 3-phosphate [29]; whether under (patho)physiological conditions it also stems from oxidation by SSAO from aminoacetone remains to be established. In the latter case, or if formation of formaldehyde from methylamine is substantial or both, treatment with an SSAO inhibitor could prove useful in preventing vascular damage during chronic diabetes mellitus.

The finding that SSAO is already raised at first diagnosis of Type I diabetes in children suggests involvement of pancreatic endothelial cells in the course of insulitis, as the islets receive  $10\,\%$  of the total pancreatic blood flow [30]. It is conceivable that the enhanced SSAO activity serves to protect the islets from increases in methylamine emerging with the insulitis. The report that methylamine inhibits the glucose-mediated release of insulin [16], raises the possibility that increased concentrations of methylamine might be such a triggering factor for increases in SSAO causing pancreatic islet damage, in particular through the formation of hydrogen peroxide [31]. Duration and degree of hyperglycaemia (HbA<sub>1c</sub>) in the weeks prior to clinical diagnosis was

apparently no such trigger nor were concentrations of GAD antibodies associated with SSAO.

In summary, we have found that plasma SSAO activity is raised both in patients with Type I and patients with Type II diabetes and relates to  $HbA_{1c}$ . In children with Type I diabetes, plasma SSAO is considerably raised even at first clinical diagnosis, but without being associated with  $HbA_{1c}$  or GAD antibody indices.

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### References

- 1. Yu PH, Zuo D-M (1993) Oxidative deamination of methylamine by semicarbazide-sensitive amine oxidase leads to cytotoxic damage in endothelial cells. Diabetes 42: 594–603
- McEwen C, Cohen JD (1963) An amine oxidase in normal human serum. J Lab Clin Med 62: 766
- Lewinsohn R (1981) Amine oxidase in human blood vessels and non-vascular smooth muscle. J Pharm Pharmacol 33: 569–575
- Norqvist A, Oreland L, Fowler CJ (1982) Some properties of monoamine oxidase and a semicarbazide sensitive amine oxidase capable of the deamination of 5-hydroxytryptamine from porcine dental pulp. Biochem Pharmacol 31: 2739–2744
- Barrand MA, Callingham BA (1982) Monoamine oxidase activities in brown adipose tissue of the rat: some properties and subcellular distribution. Biochem Pharmacol 31: 2177–2184
- Falk MC, Staton AJ, Williams TJ (1983) Heterogeneity of pig plasma amine oxidase: molecular and catalytic properties of chromatographically isolated forms. Biochem 22: 3746–3751
- 7. Precious E, Gunn CE, Lyles GA (1988) Deamination of methylamine by semicarbazide-sensitive amine oxidase in human umbilical artery and rat aorta. Biochem Pharmacol 37: 707–713
- 8. Elliott J, Callingham BA, Sharman DF (1989) Semicarbazide-sensitive amine oxidase (SSAO) of the rat aorta. Biochem Pharmacol 38: 1507–1515
- Boor PJ, Hysmith RM, Sanduja R (1990) A role for a new vascular enzyme in the metabolism of xenobiotic amines. Circ Res 66: 249–252
- Lyles GA, Holt A, Marshall CMS (1990) Further studies on the metabolism of methylamine by semicarbazide-sensitive amine oxidase activities in human plasma, umbilical artery and rat aorta. J Pharm Pharmacol 42: 322–338
- 11. Elliott WH (1960) Methylglyoxal formation from aminoacetone by ox plasma. Nature 184: 467–468
- Ray S, Ray M (1983) Formation of methylglyoxal from aminoacetone by amine oxidase from goat plasma. J Biol Chem 258: 3461–3462
- Lyles GA, Chalmers J (1992) The metabolism of aminoacetone to methylglyoxal by semicarbazide-sensitive amine oxidase in human umbilical artery. Biochem Pharmacol 43: 1409–1414

- 14. Boomsma F, Derkx FHM, van den Meiracker AH, Man in 't Veld AJ, Schalekamp MADH (1995) Plasma semicarbazide-sensitive amine oxidase activity is elevated in diabetes mellitus and correlates with glycosylated hemoglobin. Clin Sci 88: 675–679
- Gomis R, Sener A, Malaisse-Lagae F, Malaisse WJ (1983) Transglutaminase activity in pancreatic islets. Biochim Biophys Acta 760: 384–388
- Lebrun P, Atwater I, Rosario LM, Herchuelz A, Malaisse WJ (1985) Dissociation by methylamine of insulin release from glucose-induced electrical activity in isolated mouse islets of Langerhans. Metabolism 34: 1122–1127
- 17. Hayes BE, Clarke DE (1990) Semicarbazide-sensitive amine oxidase activity in streptozotocin diabetic rats. Res Commun Chem Pathol Pharmacol 69: 71–83
- Tryding N, Nilsson SE, Tufvesson G et al. (1969) Physiological and pathological influences on serum monoamine oxidase level. Scand J Clin Lab Invest 23: 79–84
- Murphy DL, Wright C, Buchsbaum M, Nichols A, Costa JL, Wyatt RJ (1976) Platelet and plasma amine oxidase activity in 680 normals: sex and age differences and stability over time. Biochem Med 16: 254–265
- 20. Dijk J van, Boomsma F, Alberts G, Man in 't Veld AJ, Schalekamp MADH (1995) Determination of semicarbazide-sensitive amine oxidase in human plasma by high-performance liquid chromatography with fluorimetric detection. J Chromatogr B Biomed Appl 663: 43–50
- 21. Batstra M, Pina M, Quan J, Mulder P et al. (1997) Fluctuations in GAD65 antibodies after clinical diagnosis of IDDM in young children. Diabetes Care 20: 642–644
- 22. Petersen JS, Hejnaes KR, Moody A, Karlsen AE, Marshall MO, Hoier-Madsen M (1994) Detection of GAD65 antibodies in diabetes and other autoimmune diseases using a simple radioligand assay. Diabetes 43: 459–467

- 23. Boomsma F, van Veldhuisen DJ, de Kam PJ et al. (1997) Plasma semicarbazide-sensitive amine oxidase is elevated in patients with congestive heart failure. Cardiovasc Res 33: 387–391
- 24. Enrique-Taracon G, Marti L, Morin N et al. (1998) Role of semicarbazide-sensitive amine oxidase on glucose transport and GLUT4 recruitment to the cell surface in adipose cells. J Biol Chem 273: 8025–8032
- 25. Callingham BA, Crosbie AE, Rous BA (1995) Some aspects of the pathophysiology of semicarbazide-sensitive amine oxidase enzymes. In: Yu PM, Tipton KF, Boulton AA (eds) Progress in brain research, vol. 106. Elsevier, Amsterdam, pp 305–321
- 26. McLellan AC, Thornalley PJ, Benn J, Sonksen PH (1994) Glyoxalase system in clinical diabetes mellitus and correlation with diabetic complications. Clin Sci 87: 21–29
- 27. Lo TWC, Selwood T, Thornalley PJ (1994) The reaction of methylglyoxal with aminoguanidine under physiological conditions and prevention of methylglyoxal binding to plasma proteins. Biochem Pharmacol 48: 1865–1870
- Yu PH, Zuo DM (1997) Aminoguanidine inhibits semicarbazide-sensitive amine oxidase activity: implications for advanced glycation and diabetic complications. Diabetologia 40: 1243–1250
- Thornalley PJ (1990)The glyoxalase system: new developments towards functional characterization of a metabolic pathway fundamental to biological life. Biochem J 269: 1–11
- 30. Jansson L (1994) The regulation of pancreatic islet blood flow. Diabetes Metab Rev 10: 407-4-16
- 31. Nomikos IN, Wang Y, Lafferty KJ (1989) Involvement of O<sub>2</sub> radicals in 'autoimmune' diabetes. Immunol Cell Biol 67: 85–87