



# CLEC10A Is a Specific Marker for Human CD1c<sup>+</sup> Dendritic Cells and Enhances Their Toll-Like Receptor 7/8-Induced Cytokine Secretion

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## OPEN ACCESS

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### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 06 January 2018

**Accepted:** 26 March 2018

**Published:** 27 April 2018

### Citation:

Heger L, Balk S, Lühr JJ, Heidkamp GF, Lehmann CHK, Hatscher L, Purbojo A, Hartmann A, Garcia-Martin F, Nishimura S-I, Cesnjevar R, Nimmerjahn F and Dudziak D (2018) CLEC10A Is a Specific Marker for Human CD1c<sup>+</sup> Dendritic Cells and Enhances Their Toll-Like Receptor 7/8-Induced Cytokine Secretion. *Front. Immunol.* 9:744. doi: 10.3389/fimmu.2018.00744

Dendritic cells (DCs) are major players for the induction of immune responses. Apart from plasmacytoid DCs (pDCs), human DCs can be categorized into two types of conventional DCs: CD141<sup>+</sup> DCs (cDC1) and CD1c<sup>+</sup> DCs (cDC2). Defining uniquely expressed surface markers on human immune cells is not only important for the identification of DC subpopulations but also a prerequisite for harnessing the DC subset-specific potential in immunomodulatory approaches, such as antibody-mediated antigen targeting. Although others identified CLEC9A as a specific endocytic receptor for CD141<sup>+</sup> DCs, such a receptor for CD1c<sup>+</sup> DCs has not been discovered, yet. By performing transcriptomic and flow cytometric analyses on human DC subpopulations from different lymphohematopoietic tissues, we identified CLEC10A (CD301, macrophage galactose-type C-type lectin) as a specific marker for human CD1c<sup>+</sup> DCs. We further demonstrate that CLEC10A rapidly internalizes into human CD1c<sup>+</sup> DCs upon binding of a monoclonal antibody directed against CLEC10A. The binding of a CLEC10A-specific bivalent ligand (the MUC-1 peptide glycosylated with N-acetylgalactosamine) is limited to CD1c<sup>+</sup> DCs and enhances the cytokine secretion (namely TNF $\alpha$ , IL-8, and IL-10) induced by TLR 7/8 stimulation. Thus, CLEC10A represents not only a candidate to better define CD1c<sup>+</sup> DCs—due to its high endocytic potential—CLEC10A also exhibits an interesting candidate receptor for future antigen-targeting approaches.

**Keywords:** dendritic cell subpopulations, CLEC10A, lineage marker, antigen targeting, cDC2, CD1c<sup>+</sup> dendritic cells

## INTRODUCTION

Dendritic cells (DCs) are potent antigen-presenting cells capable of inducing and regulating adaptive immune responses (1). They continuously sample pathogenic and harmless antigens from their surroundings. This is followed by the migration into lymphoid organs, accompanied by antigen processing and the presentation of the uptaken antigens as peptide MHC complexes to T cells. Tolerogenic T cell reactions are induced if danger signals are missing. In contrast, additional danger signals (such

as TLR-mediated signals) allow for the full maturation of DCs and thereby the induction of life-long immunity against invading pathogens (1, 2).

Dendritic cells can be distinguished into plasmacytoid DCs (pDCs) and conventional DCs. pDCs are known for their ability to secrete high amounts of type 1 interferon in response to viral infections and a lower T cell stimulatory capacity (3–5). In mice and man, two major conventional DC subpopulations have been described. Murine conventional DCs can be separated into CD8<sup>+</sup>CD11b<sup>-</sup>/CD103<sup>+</sup>CD11b<sup>-</sup> DCs (also named cDC1) and CD8<sup>-</sup>CD11b<sup>+</sup> DCs (also named cDC2) (6–8). The first excel in cross-presentation of antigens to cytotoxic CD8<sup>+</sup> T cells as well as T<sub>H</sub>1 CD4<sup>+</sup> T cell responses, whereas the latter induce preferentially T<sub>H</sub>2 and T<sub>H</sub>17 responses (9–14). According to their expression profiles, human conventional DCs can be distinguished into CD141<sup>+</sup> and CD1c<sup>+</sup> DCs. CD141<sup>+</sup> DCs share the expression of BATF3, IRF8, and XCR1 with murine CD8<sup>+</sup>CD11b<sup>-</sup>/CD103<sup>+</sup>CD11b<sup>-</sup> DCs, whereas CD1c<sup>+</sup> DCs are homologous to murine CD8<sup>-</sup>CD11b<sup>+</sup> DCs in regard to their expression of SIRP $\alpha$  and IRF4 (15–18).

Due to their unique ability to induce naïve T cell responses, DCs are suitable targets for immunotherapeutic approaches for the therapy of cancer as well as infectious diseases. One promising strategy is the *in vivo* delivery of antigens to DCs using antibodies directed against endocytic surface receptors (19). Hereby, it is possible to induce protective as well as therapeutic immune responses (19–27). In order to harness DCs for antigen-targeting approaches, it is necessary to identify endocytic receptors specifically expressed on DCs. One suitable subclass of such endocytic receptors are C-type lectin receptors (CLRs). In mice, the specific expression of the CLRs DEC205 and DCIR2 allowed for the distinct targeting of the conventional DC subsets, leading to CD8<sup>+</sup> or CD4<sup>+</sup> T cell responses, respectively (9, 20, 28). In humans, DEC205 and DCIR (a homolog of murine DCIR2) are not only expressed by one specific DC subset, thereby hindering the direct translation into the human system (15, 29–31). Recently, CLEC9A was identified as a uniquely expressed CLR on murine CD8<sup>+</sup>CD11b<sup>-</sup>/CD103<sup>+</sup>CD11b<sup>-</sup> DCs and human CD141<sup>+</sup> DCs (21, 22, 32–35). However, a potential targeting receptor specifically expressed on human CD1c<sup>+</sup> DCs is still missing.

Transcriptional data of human primary DC subpopulations suggest that the type 1 CLR CLEC10A [CD301, macrophage galactose-type C-type lectin (MGL), and CLECSF14] might be an interesting candidate expressed on human CD1c<sup>+</sup> DCs (15, 17, 36) and human CD103<sup>+</sup>SIRP $\alpha$ <sup>+</sup> DCs, the equivalent of CD1c<sup>+</sup> DCs in the human gut (16). Although transcriptomic analyses of human primary monocytes revealed human CLEC10A mRNA expression in intermediate monocytes (CD14<sup>++</sup>CD16<sup>+</sup>), only very low protein expression could be detected in these cells (37). Originally, human CLEC10A was identified as a CLR expressed on immature monocyte-derived DCs (moDCs), but not or to a lower extent on mature moDCs (38). It was further demonstrated that the carbohydrate recognition domain of CLEC10A recognizes galactose/*N*-acetylgalactosamine (Tn antigen) (38). In mice, two homologs of CLEC10A can be distinguished (CD301a/MGL1 and CD301b/MGL2), which show different expression patterns and binding specificities (39, 40). MGL1 was specifically

present on macrophages, whereas MGL2 is expressed on DCs (41). Taking together, these data imply CLEC10A as a useful marker to distinguish DC subsets (16).

The unique transcriptional expression pattern of CLEC10A in human lymphohematopoietic tissues prompted us to dissect CLEC10A protein expression in blood, thymus, and spleen. We here show a CD1c<sup>+</sup> DC specific, but organ-irrespective expression profile of CLEC10A. Our data further indicate rapid receptor internalization upon binding of an anti-CLEC10A antibody or a natural ligand (with Tn glycosylated MUC-1 peptide) of the receptor. Furthermore, this ligand in combination with the TLR7/8 ligand R848 led to an enhanced secretion of the cytokines IL-8, IL-10, and TNF $\alpha$  by CD1c<sup>+</sup> DCs. Due to its unique expression profile and its endocytic activity, we are here proposing that CLEC10A might be not only a very good marker for the distinction of human CD1c<sup>+</sup> DCs from monocytic lineages in lymphoid tissues but also a suitable receptor for the *in vivo* delivery of antigens to human CD1c<sup>+</sup> DCs.

## MATERIALS AND METHODS

### Human Tissue Preparation

Leukocyte reduction cones were retrieved from anonymous healthy adult donors. Thymus samples were retrieved from cardiac surgeries of otherwise healthy children. The sources of spleen samples were patients requiring therapeutic splenectomy. All samples were received under local ethical committee approvals (Ethikkommission der Friedrich-Alexander-Universität Erlangen-Nürnberg), and informed written consents were obtained in accordance with the Declaration of Helsinki.

All tissues were freshly processed as described earlier (15). In brief, thymic and splenic tissues were chopped into small pieces using forceps and scalpel. Then, the tissue was transferred into C-tubes (Miltenyi Biotec), filled with 5 ml RPMI1640, further mechanically disrupted using a Gentle MACS tissue dissociator (Miltenyi Biotec), and enzymatically digested with 400 U/ml collagenase D (Serva) and 100  $\mu$ g (spleen) or 300  $\mu$ g (thymus) deoxyribonuclease I (Sigma). After filtering the cell suspension twice, cell suspension of splenic and thymic tissue as well as the leukocyte enriched fraction of human blood was diluted with RPMI1640 and a density gradient centrifugation using Human Pancoll ( $\rho = 1.077$  g/ml; Pan Biotech) was performed as described earlier. After the centrifugation, the interphase containing the mononuclear cells was collected, washed twice with RPMI1640, and used for experiments.

### Microarray Analysis

Published microarray data were analyzed for relative expression of CLEC10A (15). Microarray data are available in the Gene Expression Omnibus database ([www.ncbi.nlm.nih.gov/gds](http://www.ncbi.nlm.nih.gov/gds)) under the accession number GSE77671. Transcriptome data of whole Human Genome Oligo microarray (Agilent) of human CD1c<sup>+</sup> DCs, CD141<sup>+</sup> DCs, and pDCs from three blood, spleen, and thymus donors as well as blood monocytes, B cells, and CD4<sup>+</sup> and CD8<sup>+</sup> T cells were used. Raw values generated by automated feature extraction have been RMA background corrected and quantile normalized using R (Windows, x64, 3.3.1) (42). Relative

expression values were plotted taking advantage of the gplots package of R (43).

## Flow Cytometry

Flow cytometric analyses of single cell suspensions of blood, spleen, and thymus were performed on a BD LSRFortessa and analyzed using FlowJo software (Treestar).  $5 \times 10^6$  (blood) to  $8 \times 10^6$  (thymus, spleen) cells were stained with the following antibody cocktail to discriminate different immune cell populations for 15 min at 4°C: CD1c-APC/Cy7 (L161; BioLegend), CD3-BUV395 (UCHT1; BD Biosciences), CD4-BV510 (OKT4; BioLegend), CD8-APC (RPA-T8; eBioscience) or CD8-BUV737 (SK1; BD Biosciences), CD11b-PE/Cy5 (M1/70; BioLegend), CD11c-PE/Cy7 (3.9; BioLegend), CD14-A700 (HCD14; BioLegend), CD19-BV605 (SJ25C1; BioLegend), CD20-BV605 (2H7; BioLegend), CD56-BV421 (5.1H11; BioLegend), CD123-BV650 (6H6; BioLegend), CD141-BV711 (1A4; BD Bioscience), CD303-PerCP/Cy5.5 (201A; BioLegend), CLEC10A-PE or -APC (H037G3; BioLegend), HLA-DR-PE-CF594 (G46-6; BD Bioscience), and NKp46-BV421 (9E2; BioLegend). As appropriate isotype control for CLEC10A-PE and -APC, the cells were stained with a PE- or APC-labeled mouse IgG2a isotype control (MOPC-173; BioLegend). For some experiments, CLEC10A was stained using 10 µg/ml of an FITC-coupled MUC-1 peptide ( $\beta$ Ala-GVTSAPDTRPAPGSTAPPAHGVT-NH<sub>2</sub>) that was glycosylated with N-acetylgalactosamine (Tn antigen) at Serine 4 and Threonine 15 [referred to as MUC-1-(Tn)<sub>2</sub>] or a non-glycosylated FITC- $\beta$ Ala-MUC-1 peptide as control (referred to as MUC-1). Synthesis and characterization of the compounds have been described earlier (44). After staining for 30 min on ice, the cells were washed and stained for different immune cell populations as before. After washing the cells, they were resuspended in PBS + 2% human sera + 0.1 mg/ml 4',6-diamidino-2-phenylindole and acquired using a BD LSRFortessa.

## Internalization Assay

For the internalization assay,  $5 \times 10^6$  cells were stained with anti-CLEC10A-PE (H037G3; BioLegend) or an appropriate isotype control (mouse IgG2a, MOPC-173; BioLegend) for 15 min on ice. Then, the cells were incubated for different time points at 37°C (5, 15, 30, and 60 min) or kept on ice for 60 min. In a next step, surface CLEC10A was stained with an anti-PE antibody (polyclonal goat IgG; Novus Biologicals) and after washing with an anti-goat-A647 (polyclonal donkey IgG; Southern Biotech). In a last step, the cells were stained for the discrimination of different immune cell populations as described under flow cytometry and samples acquired using a BD LSRFortessa.

## Generation of moDCs

PBMCs were isolated by gradient-density centrifugation.  $5 \times 10^7$  PBMCs in 10 ml DMEM were adhered to a human IgG-coated tissue culture dish (BD Falcon) and incubated for 1 h at 37°C. Then, the medium was replenished with 10 ml moDC medium [RPMI-1640/containing 1% penicillin/streptomycin, 1% L-glutamine, 1% HEPES, and 2% human sera type AB (Lonza)]. On the next day, the medium was replenished with fresh moDC medium containing 800 U/ml GM-CSF and 50 U/ml IL-4. On day 3 and day

5, 4 ml of fresh moDC medium was added containing 800 U/ml GM-CSF/50 U/ml IL-4 and 400 U/ml GM-CSF/25 U/ml IL-4, respectively. On day 6, half of the cells were matured using a maturation cocktail consisting of 13.2 ng/ml IL-1 $\beta$ , 10,000 U/ml IL-6, 10 ng/ml TNF $\alpha$ , and 1 µg/ml prostaglandin E<sub>2</sub> for 24 h. IL-1 $\beta$ , IL-6, and TNF $\alpha$  were purchased from PeproTech and prostaglandin E<sub>2</sub> from Sigma Aldrich.

## Enrichment of DCs

In order to perform cell sorts and for internalization experiments using thymic DCs, DCs were enriched prior to the experiments. Therefore, blood DCs were enriched with the EasySep Pan-DC Pre-Enrichment Kit (Stemcell Technologies) as described in the manual of the manufacturer. Briefly, PBMCs were diluted in PBS + 2% human sera + 1 mM EDTA and stained with anti-human CD32 (Fc gamma RII) Blocker (Stemcell Technologies) and the EasySep Pan-DC Pre-Enrichment Cocktail for 45 min at room temperature (RT). Then, magnetic Dextran Rapidspheres were added, and the cells were incubated for 10 min at RT followed by incubation for 5 min inside "The Big Easy" EasySep Magnet (Stemcell Technologies). The unlabeled DCs were poured off into a new tube and stained for cell sorts.

Thymic DCs were enriched using biotinylated antibodies against thymocytes (CD7, clone eBio124-1D1; eBioscience), T cells (CD3, SK7, BioLegend; CD8, RPA-T8, BioLegend), B cells (CD19, HIB19; BD Bioscience), and NK cells (CD56, MEM-188; BioLegend), and MojoSort Streptavidin Nanobeads (BioLegend). The cell suspension was stained with the cocktail of biotinylated antibodies in PBS + 2% human sera + 1 mM EDTA for 30 min on ice. Then, the cells were washed, resuspended in PBS + 2% human sera + 1 mM EDTA, and MojoSort Streptavidin Nanobeads added. After incubation for 15 min on ice, the tube was transferred into "The Big Easy" EasySep Magnet and incubated for 5 min at RT. Next, the unlabeled DCs were poured off into a new tube and used for further experiments.

## Cell Sorting of Human DC Subpopulations

Dendritic cell-enriched cell suspensions were stained with an antibody cocktail consisting of the following antibodies in PBS + 2% human sera for 30 min on ice: CD1c-APC/Cy7 (L161; BioLegend), CD3-BUV395 (UCHT1; BD Bioscience), CD11b-A700 (M1/70; BioLegend), CD11c-PE/Cy7 (3.9; BioLegend), CD14-A700 (HCD14; BioLegend), CD19-V450 (HIB19; BD Bioscience), CD20-eFluor 450 (2H7; eBioscience), CD56-BV421 (5.1H11; BioLegend), CD123-BV605 (6H6; BioLegend), CD141-BV711 (1A4; BD Bioscience), CD303-PerCP/Cy5.5 (201A; BioLegend), HLA-DR-PE-CF594 (G46-6; BD Bioscience), and NKp46-BV421 (9E2; BioLegend). After washing the cells, they were resuspended in PBS + 2% human sera + 0.1 mg/ml 4',6-diamidino-2-phenylindole and cell sorted using a BD FACSAria II cell sorter into CD1c<sup>+</sup> DCs (CD3<sup>-</sup>CD11b<sup>-</sup>CD14<sup>-</sup>CD19<sup>-</sup>CD20<sup>-</sup>CD56<sup>-</sup>CD123<sup>-</sup>NKp46<sup>-</sup>HLA-DR<sup>+</sup>CD11c<sup>+</sup>CD1c<sup>+</sup>), CD141<sup>+</sup> DCs (CD1c<sup>-</sup>CD3<sup>-</sup>CD11b<sup>-</sup>CD14<sup>-</sup>CD19<sup>-</sup>CD20<sup>-</sup>CD56<sup>-</sup>CD123<sup>-</sup>NKp46<sup>-</sup>HLA-DR<sup>+</sup>CD11c<sup>+</sup>CD141<sup>+</sup>), pDCs (CD1c<sup>-</sup>CD3<sup>-</sup>CD11b<sup>-</sup>CD14<sup>-</sup>CD19<sup>-</sup>CD20<sup>-</sup>CD56<sup>-</sup>NKp46<sup>-</sup>HLA-DR<sup>+</sup>CD123<sup>+</sup>CD303<sup>+</sup>), and monocytes (CD3<sup>-</sup>CD19<sup>-</sup>CD20<sup>-</sup>CD56<sup>-</sup>CD123<sup>-</sup>NKp46<sup>-</sup>HLA-DR<sup>+</sup>CD11b<sup>+</sup>CD14<sup>+</sup>). The purity of sorted cell populations was reanalyzed and routinely above 95%.



## Stimulation of Sorted DCs

Cell-sorted DC subpopulations and monocytes were resuspended in DC medium (RPMI1640 + 5% human sera + 5% Pannexin NTA + 5% Pannexin NTS + 1% glutamine + 1% sodium pyruvate + 1% penicillin/streptomycin) to a concentration of  $2 \times 10^5$  cells/ml. Cells were cultured in sterile 96-well plates (v-bottom) and stimulated either with one of the TLR ligands pIC (5  $\mu$ g/ml) or R848 (5  $\mu$ g/ml), the CLEC10A ligand MUC-1-(Tn)<sub>2</sub> (10  $\mu$ g/ml) or a non-glycosylated MUC-1 control peptide (10  $\mu$ g/ml), or a combination of both. TLR ligands were purchased from Invivogen and reconstituted in water. After 12 h of stimulation, the cells were analyzed by flow cytometry for the expression of CLEC10A (CLEC10A-APC, clone: H037G3, BioLegend) and activation markers (CD40-PE, clone: 5C3, BioLegend; CD86-PE-CF594, 2331(FUN-1), BD Biosciences) together with sorted DCs and monocytes of the very same donor, which were kept on ice until analysis. Supernatants of stimulated cells were harvested and stored at  $-80^\circ\text{C}$  until analysis with the LEGENDplex Human Anti-Virus Response Panel (BioLegend). Acquisition of the samples was performed using a BD LSRFortessa and data analyzed using FlowJo (CLEC10A, CD40, CD86) or LEGENDplex software (VigeneTech; cytokine secretion).

## Intracellular Signaling in Sorted CD1c<sup>+</sup> DCs

CD1c<sup>+</sup> DCs were sorted from PBMCs of healthy donors, resuspended in DC medium to a final concentration of  $2 \times 10^5$  cells/ml and stimulated either with 10  $\mu$ g/ml MUC-1-(Tn)<sub>2</sub> or non-glycosylated MUC-1 in combination with 5  $\mu$ g/ml R848 and incubated at  $37^\circ\text{C}$  for different time points (0, 5, 15, 30, and 60 min). Afterward, the cells were washed and fixed with Cytofix/Cytoperm Buffer (BD Bioscience) for 20 min on ice. Then, the cells were washed and permeabilized with Perm Buffer III (BD Bioscience) for 30 min on ice. After washing, the cells were stained with BV421 Mouse Anti-Human NF- $\kappa$ B p65 (pS529), Alexa Fluor<sup>®</sup> 647 Mouse anti-JNK (pT183/pY185), Alexa Fluor<sup>®</sup> 488 Mouse Anti-CREB (pS133)/ATF-1 (pS63), PE-CF594 Mouse Anti-p38 MAPK (pT180/pY182), PE-C7 Mouse anti-ERK1/2 (pT202/pY204), and PE Mouse anti-Human IKK $\gamma$  (pS376) for 30 min at RT. All antibodies were purchased from BD Bioscience. The cells were washed and then acquired using a BD LSRFortessa.

## RESULTS

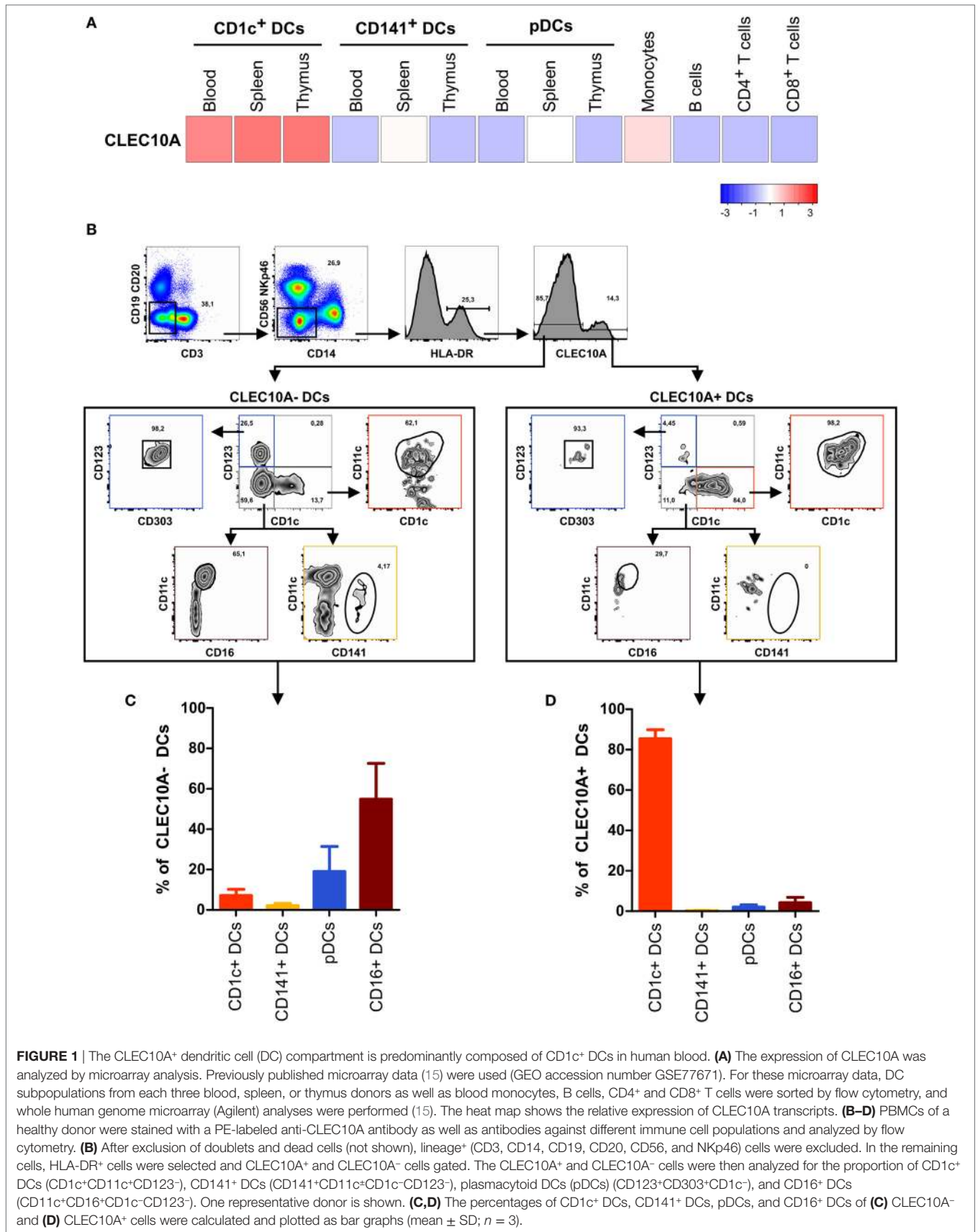
### CLEC10A Is Specifically Expressed on Human CD1c<sup>+</sup> DCs in Various Lymphoid Tissues

In our recently performed transcriptional analysis of human lymphohematopoietic organ DC subpopulations (blood, thymus, and spleen), we demonstrated a strong expression of the CLEC10A mRNA in CD1c<sup>+</sup> DCs in all tested tissues (Figure 1A) (15). In line with an earlier report about intermediate monocytes (CD14<sup>++</sup>CD16<sup>+</sup>), we could confirm a low CLEC10A mRNA expression in monocytes (Figure 1A) (37). To prove that CLEC10A is also expressed on protein level, we performed flow cytometric

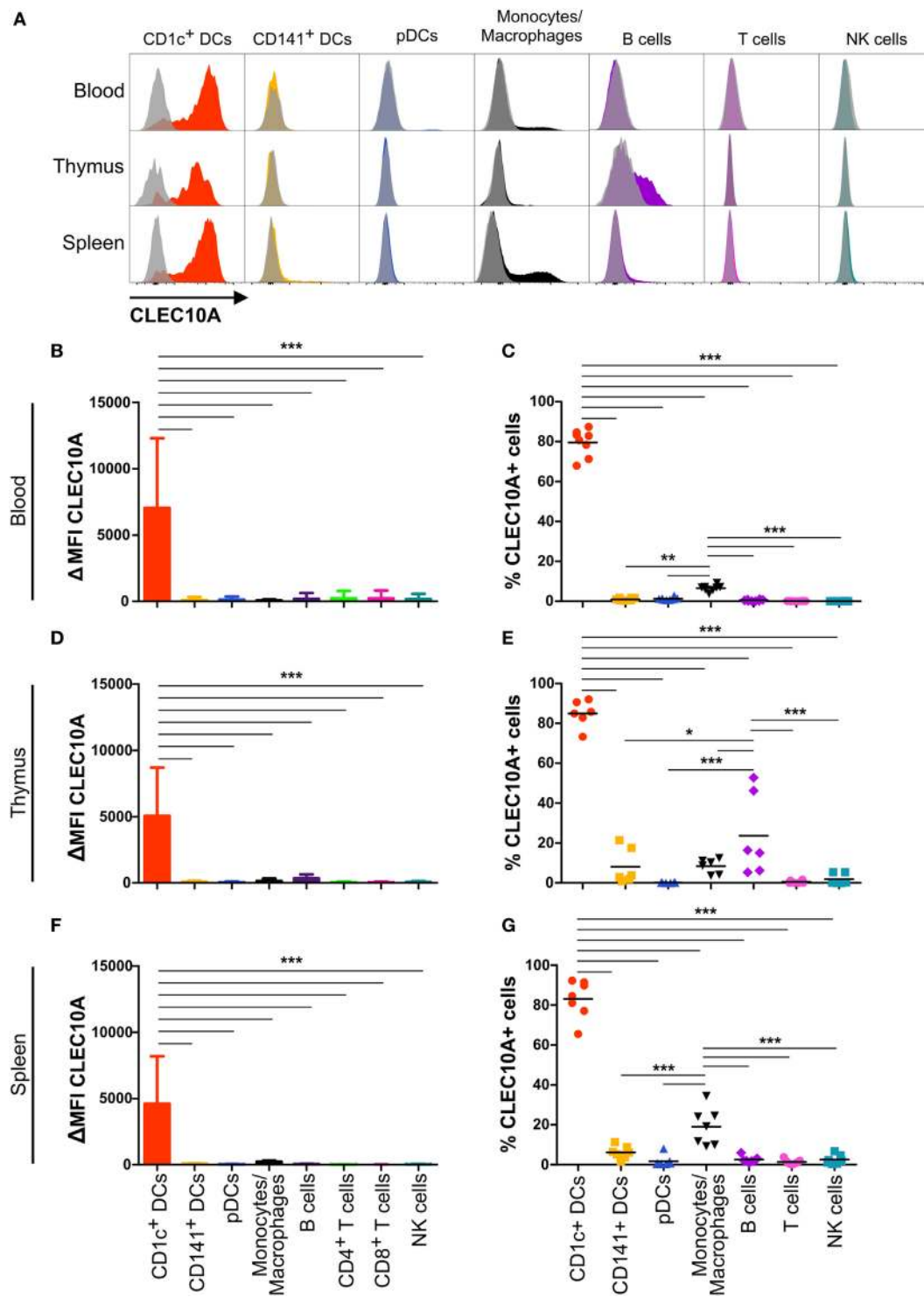
analyses on single cell suspensions of human blood. We found that the CLEC10A<sup>+</sup> lineage (CD3, CD14, CD19, CD20, CD56, and NKp46) negative cells consisted almost exclusively of CD1c<sup>+</sup> DCs (Figures 1B,D). In contrast, the CLEC10A<sup>-</sup> lineage negative cells consisted mainly of CD16<sup>+</sup> DCs, CD141<sup>+</sup> DCs, and pDCs (Figures 1B,C). In addition, a low amount of CD1c<sup>+</sup> DCs were represented in the CLEC10A<sup>-</sup> DC compartment (Figure 1C). We further extended our analyses to thymus and splenic tissue and analyzed the expression of CLEC10A on other cells of the immune system (Figure 2). Remarkably, also in these lymphoid tissues CLEC10A demonstrated a selective expression on human CD1c<sup>+</sup> DCs comparable to the blood (Figures 2A,B,D,F). Only to a low extent we found CLEC10A protein expression on thymic B cells and blood as well as splenic monocytes/macrophages (Figure 2). Notably, no expression could be detected on CD141<sup>+</sup> DCs, pDCs, T cells, or NK cells (Figure 2). Taken together, we found that CLEC10A was expressed on about 80% of CD1c<sup>+</sup> DCs in all tested tissues, while it was only expressed on about 5% of blood monocytes, 20% of thymic B cells, and 20% of splenic monocytes/macrophages (Figures 2C,E,G). In summary, our data suggest a prevalent expression profile of CLEC10A protein on human CD1c<sup>+</sup> DCs.

### CLEC10A Is Rapidly Internalized Into Human CD1c<sup>+</sup> DCs

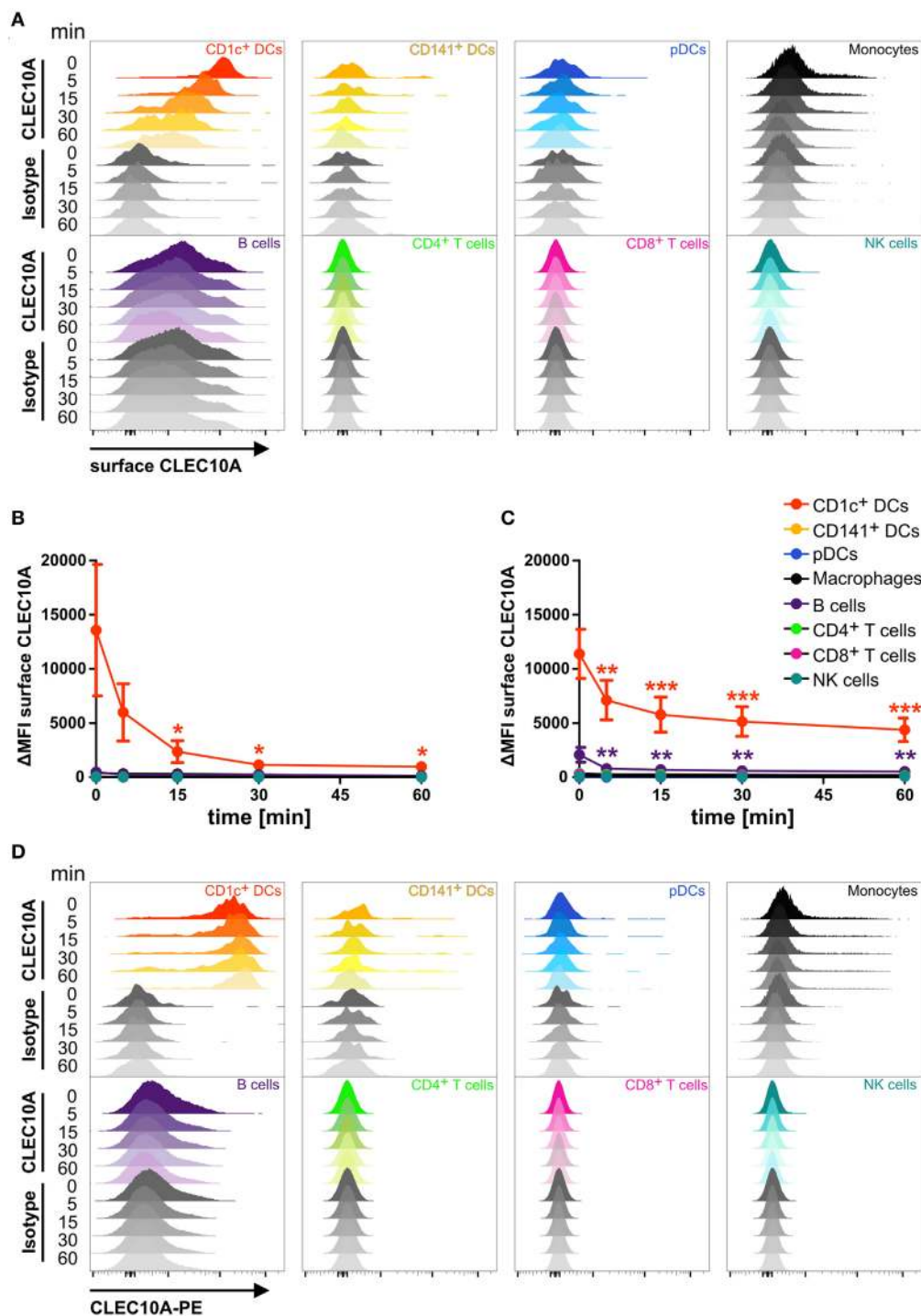
As a potential receptor for antigen-targeting approaches, it is necessary that antibodies bound to its specific receptor can internalize into the cell. To investigate the endocytic activity of the CLEC10A receptor, we performed an internalization assay. We therefore stained single cell suspensions of blood and thymic tissue with a PE-coupled anti-CLEC10A antibody. After binding of the antibody, the cells were washed and incubated at  $37^\circ\text{C}$  for different time points to allow the internalization of the antibody bound to CLEC10A. To distinguish between antibodies that were internalized from remaining antibodies at the surface of the cells, we stained the cells with a secondary antibody recognizing the PE fluorochrome coupled to the anti-CLEC10A antibody. For controlling the internalization efficacy, the secondary surface-bound antibody was visualized by a tertiary A647-labeled antibody directed against the Fc part of the secondary antibody. In cells that were kept on ice, the PE-labeled primary anti-CLEC10A antibody remained on the surface and thereby the cells displayed a strong staining for the A647-labeled tertiary antibody (Figure 3). With increasing incubation time, the A647 signal was reduced (Figures 3A–C), whereas the PE signal of the primary PE-labeled anti-CLEC10A antibody was stable over the whole incubation time (Figure 3D). Our data indicate that already after 5 min at  $37^\circ\text{C}$  large amounts of the PE-labeled anti-CLEC10A antibody were internalized into CD1c<sup>+</sup> DCs. We did not observe other cells, which expressed or internalized the antibody except for a low internalization into thymic B cells (Figure 3). Interestingly, whereas blood CD1c<sup>+</sup> DCs completely internalized the anti-CLEC10A antibody within 30 min (Figure 4B), about 50% of the signal remained on the surface of thymic CD1c<sup>+</sup> DCs (Figure 3C). Thus, our data suggest a strong endocytic activity of CLEC10A upon antibody binding.



**FIGURE 1** | The CLEC10A+ dendritic cell (DC) compartment is predominantly composed of CD1c+ DCs in human blood. **(A)** The expression of CLEC10A was analyzed by microarray analysis. Previously published microarray data (15) were used (GEO accession number GSE77671). For these microarray data, DC subpopulations from each three blood, spleen, or thymus donors as well as blood monocytes, B cells, CD4+ and CD8+ T cells were sorted by flow cytometry, and whole human genome microarray (Agilent) analyses were performed (15). The heat map shows the relative expression of CLEC10A transcripts. **(B–D)** PBMCs of a healthy donor were stained with a PE-labeled anti-CLEC10A antibody as well as antibodies against different immune cell populations and analyzed by flow cytometry. **(B)** After exclusion of doublets and dead cells (not shown), lineage+ (CD3, CD14, CD19, CD20, CD56, and NKp46) cells were excluded. In the remaining cells, HLA-DR+ cells were selected and CLEC10A+ and CLEC10A- cells gated. The CLEC10A+ and CLEC10A- cells were then analyzed for the proportion of CD1c+ DCs (CD1c+CD11c+CD123-), CD141+ DCs (CD141+CD11c+CD1c-CD123-), plasmacytoid DCs (pDCs) (CD123+CD303+CD1c-), and CD16+ DCs (CD11c+CD16+CD1c-CD123-). One representative donor is shown. **(C,D)** The percentages of CD1c+ DCs, CD141+ DCs, pDCs, and CD16+ DCs of **(C)** CLEC10A- and **(D)** CLEC10A+ cells were calculated and plotted as bar graphs (mean ± SD; n = 3).

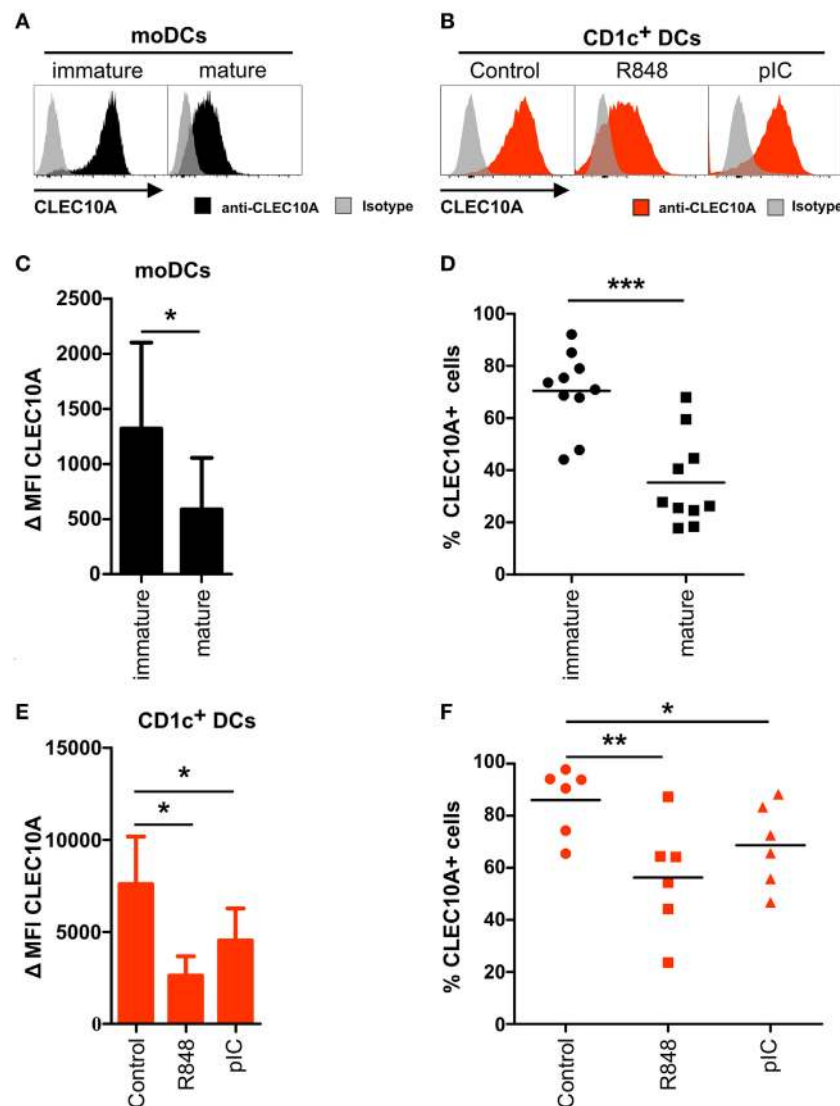


**FIGURE 2 |** CLEC10A is mainly expressed on human CD1c+ dendritic cells (DCs) in different tissues in the steady state. The expression of CLEC10A was analyzed by flow cytometric analysis on human DC subpopulations in blood, thymus, and spleen. **(A)** Depicted are overlay histograms of single cell suspensions of one representative donor each of blood, thymus, and spleen analyzed by flow cytometry for an anti-CLEC10A antibody (colored) and appropriate isotype control (gray, transparent). Bar graphs in **(B,D,F)** show the  $\Delta$ MFI for CLEC10A (median  $\pm$  SD, blood  $n = 8$ ; thymus  $n = 6$ ; spleen  $n = 7$ ). **(C,E,G)** In each leukocyte population, the CLEC10A+ cells were determined using an appropriate isotype control and plotted as a scatter plot with mean as horizontal line. Significant differences in the **(B,D,F)** expression of CLEC10A or **(C,E,G)** percentages of CLEC10A+ cells were calculated by one-way ANOVA with Bonferroni posttest using GraphPad Prism ( $*p < 0.05$ ,  $**p < 0.01$ , and  $***p < 0.001$ ).



**FIGURE 3** | CLEC10A is rapidly internalized into blood and thymic CD1c<sup>+</sup> dendritic cells (DCs). Single cell suspensions of blood (**A,B,D**) and thymus (**C**) were incubated with a PE-coupled anti-CLEC10A antibody or an appropriate isotype control at 4°C for 15 min, washed, and resuspended in PBS containing 2% human sera. Subsequently, the cells were incubated at 37°C for different time points (0, 5, 15, 30, and 60 min) to allow for the internalization of the primary PE-coupled antibody. Afterward, the cells were stained for surface CLEC10A by an anti-PE secondary antibody (polyclonal goat IgG) and an A647-labeled tertiary anti-goat antibody. Then, the cells were stained for different immune cell populations.  $2.5 \times 10^6$  cells were acquired using a BD LSRFortessa and analyzed using FlowJo. **(A)** Overlay histograms show surface expression of CLEC10A for CD1c<sup>+</sup> DCs (red), CD141<sup>+</sup> DCs (yellow-orange), plasmacytoid DCs (pDCs) (blue), monocytes (black), B cells (purple), CD4<sup>+</sup> T cells (green), CD8<sup>+</sup> T cells (pink), and NK cells (cyan). **(B,C)** PBMCs (**B**) and thymic (**C**) cells were analyzed as in **(A)**. Plot shows mean  $\pm$  SD of surface CLEC10A ( $MFI_{CLEC10A} - MFI_{isotype\ control}$ ) of three (**C**) or five (**B**) donors. Statistical analysis was performed using one-way ANOVA with Dunnett posttest for each cell population ( $\Delta MFI$  0 min as control; \* $p < 0.05$ , \*\* $p < 0.01$ , and \*\*\* $p < 0.001$ ). **(D)** Overlay histograms show the PE-signal of the primary PE-coupled anti-CLEC10A antibody or the appropriate isotype control for the same donor as in **(A)**.





**FIGURE 4** | CLEC10A is downregulated on activated human CD1c<sup>+</sup> dendritic cells (DCs). **(A,C,E)** Monocyte-derived DCs were generated from blood monocytes of a healthy donor. After 6 days, half of the cells were matured using a maturation cocktail consisting of IL-1 $\beta$ , IL-6, TNF $\alpha$ , and PGE<sub>2</sub> for 24 h. **(B,D,F)** Blood CD1c<sup>+</sup> DCs (red) were sorted from PBMCs of a healthy donor and subsequently stimulated either with R848 (5  $\mu$ g/ml), pIC (5  $\mu$ g/ml), or cultured in DC medium for 12 h at 37°C. Afterward, the cells were stained with an APC-labeled anti-CLEC10A antibody or an appropriate isotype control and analyzed using a BD LSRFortessa and FlowJo software. **(A,B)** Overlay histograms demonstrate a representative donor for **(A)** moDCs (black: CLEC10A; gray-transparent: isotype control) and **(B)** CD1c<sup>+</sup> DCs (red: CLEC10A; gray-transparent: isotype control). **(C,E)** Bar graphs show the expression of CLEC10A [mean  $\pm$  SD of  $\Delta$ MFI (MFI<sub>CLEC10A</sub> – MFI<sub>isotype control</sub>)] on **(C)** moDCs ( $n = 10$ ) and **(E)** CD1c<sup>+</sup> DCs ( $n = 6$ ). **(D,F)** CLEC10A<sup>+</sup> **(D)** moDCs (black symbols) and **(F)** CD1c<sup>+</sup> DCs (red symbols) were determined using an appropriate isotype control and blotted as scatter plots. Significant differences in the **(C,E)** expression of CLEC10A or **(D,F)** percentages of CLEC10A<sup>+</sup> cells were calculated using  $t$  test (\* $p < 0.05$ , \*\* $p < 0.01$ , and \*\*\* $p < 0.001$ ).

## CLEC10A Is Downregulated on Activated CD1c<sup>+</sup> DCs

Human CLEC10A was identified as a CLR expressed on immature moDCs. To place our data into the context of previous findings, we performed flow cytometric analyses on immature and mature moDCs. Accordingly, we could detect a low to intermediate surface expression of CLEC10A on immature moDCs (**Figures 4A,C**). In line with earlier published data, displaying an absent expression of CLEC10A-mRNA in mature moDCs (38),

we found that after stimulation of moDCs with a maturation cocktail consisting of IL-1 $\beta$ , IL-6, TNF $\alpha$ , and prostaglandin E<sub>2</sub>, the expression of CLEC10A was significantly reduced, but not absent, on mature moDCs (**Figures 4A,C,D**).

To investigate potential expression changes of CLEC10A on activated human CD1c<sup>+</sup> DCs, we stimulated sorted human blood CD1c<sup>+</sup> DCs with TLR ligands R848 (TLR7/8) or pIC (TLR3) and analyzed the cells by flow cytometry. In line with the results obtained with moDCs and independent on the used stimulus,



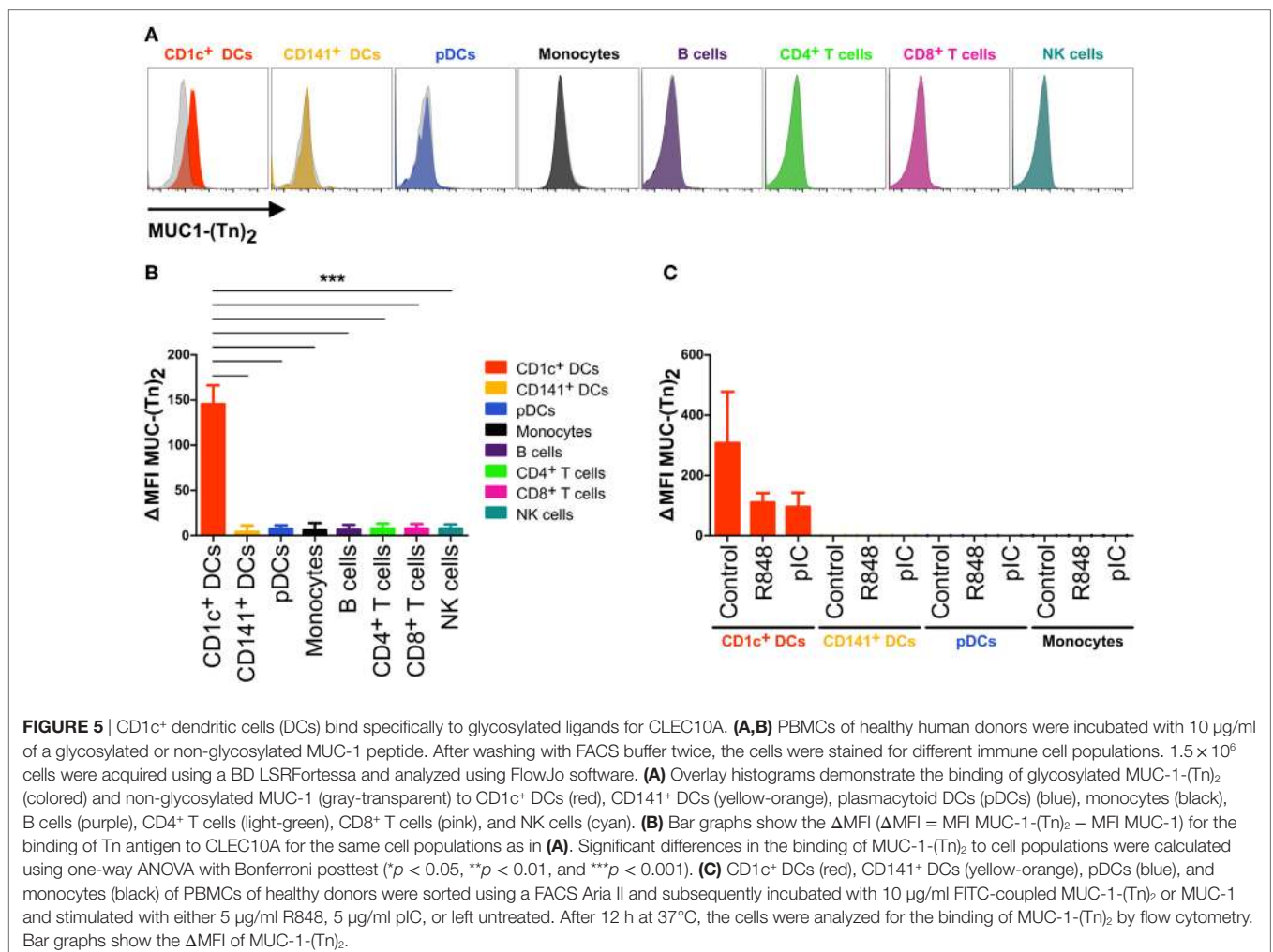
12 h upon TLR ligand stimulation we found a reduced surface expression of CLEC10A on blood CD1c<sup>+</sup> DCs (Figures 4B,E), in which TLR7/8 stimulation was superior to TLR3 stimulation, as the first demonstrated a stronger reduction of the CLEC10A surface expression. Comparable to data generated from moDCs, TLR stimulated CD1c<sup>+</sup> DCs remained partially positive for CLEC10A (Figure 4F). Our data suggest a typical behavior of CLEC10A compared to other type I CLR receptors such as DCIR, DC-Sign, or CLEC9A upon cell stimulation.

## CD1c<sup>+</sup> DCs Specifically Bind Glycosylated Ligands for CLEC10A

In order to better define the CLEC10A-specific expression profile, we performed FACS staining experiments applying a previously described bivalent ligand of CLEC10A (44). The use of such natural ligands is not only of importance for the identification of interaction partners and cell activation, natural ligands can also be used for the delivery of antigens to endocytic receptors as it was demonstrated for the chemokine ligand receptor pair XCL1 and XCR1 (45, 46). Thus, PBMCs of healthy donors were incubated either with 10 µg/ml of an FITC-coupled MUC-1 peptide

(sequence: βAla-GVTSAPDTRPAPGSTAPPAHGVT-NH<sub>2</sub>) that was glycosylated with N-acetylgalactosamine (Tn antigen) at Serine 4 and Threonine 15 (referred to as CLEC10A ligand MUC-1-(Tn)<sub>2</sub>) or with 10 µg/ml of a non-glycosylated FITC-coupled MUC-1 peptide (referred to as MUC-1) (44). After ligand staining, the cells were incubated with antibodies for the detection of different immune cell populations and analyzed by flow cytometry. Analogously to the expression profile we have seen upon application of an anti-CLEC10A antibody, we detected a specific binding of the glycosylated form of MUC-1 [MUC-1-(Tn)<sub>2</sub>] to CD1c<sup>+</sup> DCs (Figure 5). No other immune cell populations, such as T cells, B cells, NK cells, monocytes, nor CD141<sup>+</sup> DCs, and pDCs demonstrated an interaction with the glycosylated MUC-1-(Tn)<sub>2</sub> (Figures 5A,B).

As we found that upon TLR ligand activation of CD1c<sup>+</sup> DCs the expression of CLEC10A was downregulated, we next investigated CLEC10A ligand binding on activated CD1c<sup>+</sup> DCs. Therefore, we sorted CD1c<sup>+</sup> DCs together with CD141<sup>+</sup> DCs, pDCs, and monocytes from PBMCs of healthy donors and stimulated them simultaneously with the glycosylated or the non-glycosylated MUC-1 peptide [MUC-1-(Tn)<sub>2</sub> or MUC-1] in combination with one of the TLR ligands R848 or pIC. We found that CD1c<sup>+</sup> DCs



incubated without a TLR ligand bound high amounts of MUC-1-(Tn)<sub>2</sub>, whereas CD1c<sup>+</sup> DCs stimulated with either R848 or pIC showed a lower binding of MUC-1-(Tn)<sub>2</sub> (Figure 5C). This is in accordance with the induced downregulation of CLEC10A upon stimulation with the TLR ligands R848 and pIC (Figure 4).

## Stimulation of CD1c<sup>+</sup> DCs With CLEC10A Ligands Enhances TLR-Induced IL-8, IL-10, and TNF $\alpha$ Secretion

As some CLR receptors have been described to induce signaling upon binding of their ligands, we were interested to understand whether stimulation of CD1c<sup>+</sup> DCs with the CLEC10A ligand MUC-1-(Tn)<sub>2</sub> would induce signaling and thereby activation and cytokine secretion by the DCs. Therefore, CD1c<sup>+</sup> DCs were sorted from PBMCs of healthy donors and incubated with glycosylated MUC-1-(Tn)<sub>2</sub> or the non-glycosylated MUC-1 in the presence or absence of the TLR ligands R848 or pIC. In the absence of further TLR ligand stimulation, we found no activation of CD1c<sup>+</sup> DCs by the glycosylated MUC-1-(Tn)<sub>2</sub> or the non-glycosylated MUC-1 (Figure 6A). As shown earlier (47, 48), stimulation with R848 or pIC induced high expression of the co-stimulatory molecules CD40 and CD86, which was not enhanced by binding of MUC-1-(Tn)<sub>2</sub> to CLEC10A (Figure 6A). We further investigated, if this glycosylated ligand might influence the cytokine secretion of CD1c<sup>+</sup> DCs, as it was shown that signaling *via* CLEC10A can enhance TLR-induced secretion of cytokines by moDCs (49). We therefore analyzed the supernatants of CD1c<sup>+</sup> DCs for IL-6, IL-8, IL-10, IL-12p70, IL-23, and TNF $\alpha$ . Neither the incubation with the glycosylated MUC-1-(Tn)<sub>2</sub> nor the non-glycosylated MUC-1 alone induced strong cytokine secretion by CD1c<sup>+</sup> DCs (Figure 6B). However, we found a slightly, but statistically significant increased secretion of the cytokines IL-8, IL-10, and TNF $\alpha$  after stimulation with glycosylated MUC-1-(Tn)<sub>2</sub> in combination with R848, but not pIC (Figure 6B). In contrast, glycosylated MUC-1-(Tn)<sub>2</sub> did not enhance IL-6, IL-12p70, or IL-23 secretion induced by TLR7/8 or TLR3 stimulation (Figure 6B).

Since the combination of MUC-1-(Tn)<sub>2</sub> and R848 enhanced the secretion of cytokines by CD1c<sup>+</sup> DCs, we wondered, if key signaling pathways were influenced in CD1c<sup>+</sup> DCs. We therefore incubated the cells with the glycosylated MUC-1-(Tn)<sub>2</sub> or the non-glycosylated MUC-1 in the presence or absence of the TLR ligand R848 as described earlier. We performed intracellular FACS staining for the detection of phosphorylation in signaling cascade proteins and of transcription factors, such as NF $\kappa$ B p65, p38 MAPK, IKK $\gamma$ , JNK, ERK-1/2, or CREB/ATF-1. Although all of the tested proteins are important components in intracellular signal transduction, we could not detect any differences in their phosphorylation states after costimulation of CD1c<sup>+</sup> with MUC-1-(Tn)<sub>2</sub> and R848 (Figure 7).

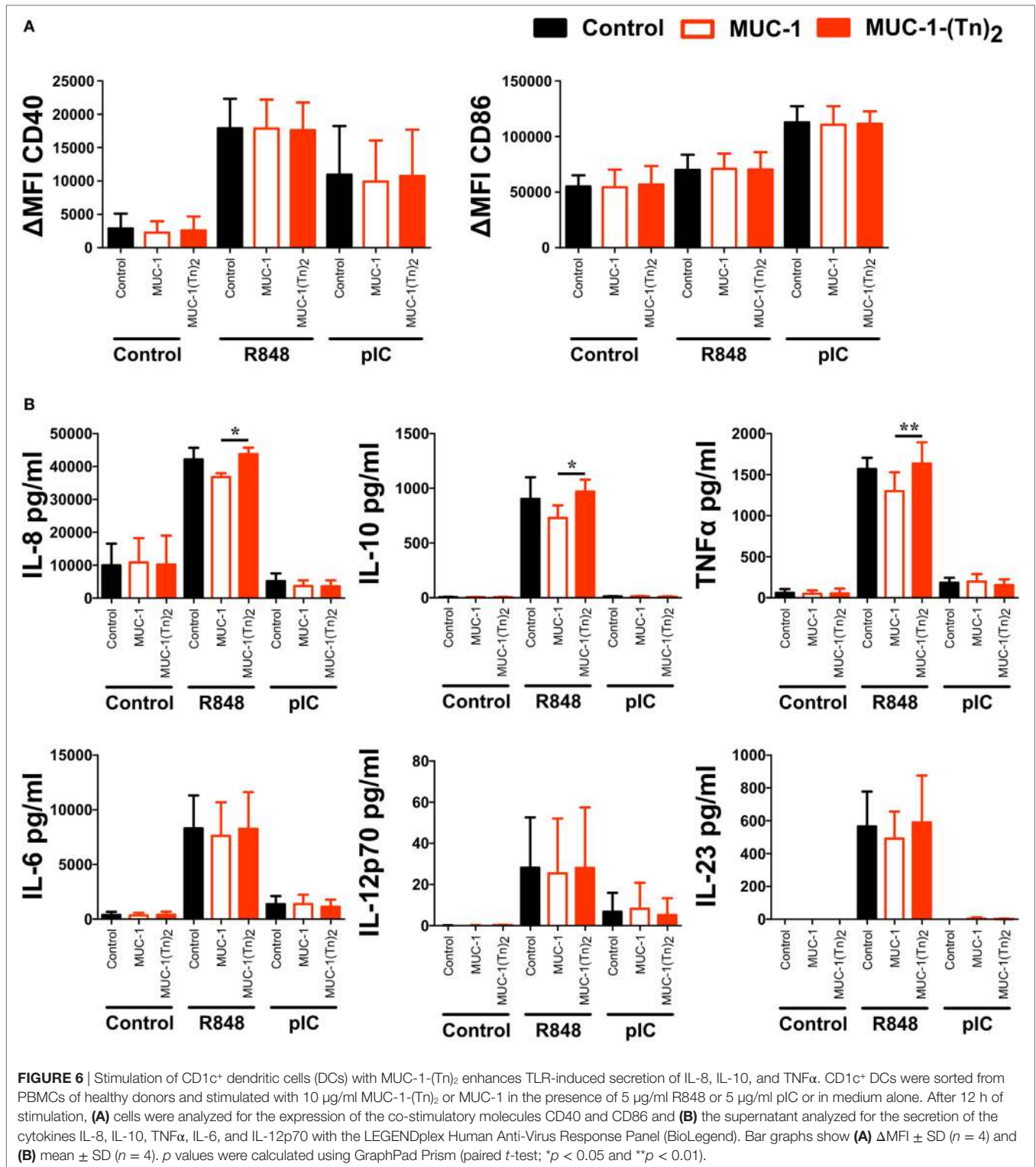
## DISCUSSION

Here, we investigated the expression of CLEC10A in various immune cell populations and found a selective presence of this CLR on CD1c<sup>+</sup> DCs. Our data indicate that CLEC10A is rapidly internalized upon antibody binding. Although MUC-1-(Tn)<sub>2</sub>, a

natural ligand for CLEC10A, specifically binds to this DC subset, signaling events were not directly induced. However, we detected a slightly enhanced secretion of selective cytokines induced by the TLR7/8 ligand R848 upon coadministration of MUC-1-(Tn)<sub>2</sub>.

CLEC10A belongs to the family of CLRs. CLRs are responsible for the recognition of sugar structures on bacteria and cancer cells (40, 50–52). For CLEC10A, the specific ligand N-acetylgalactosamine was recently described (41, 44). Although CLEC10A was originally identified on human moDCs (38), we previously found a high mRNA expression in human CD1c<sup>+</sup> DCs isolated from blood, spleen, and thymus (Figure 1) (15), which is in accordance to other published transcriptomic data (16, 17, 36). This led us to the investigation of CLEC10A protein levels on immune cell populations. Thereby, we found a selective CLEC10A protein expression on human CD1c<sup>+</sup> DCs. While protein expression of CLEC10A on a human subpopulation of blood CD1c<sup>+</sup> DCs was already reported by Lundberg et al. (53, 54), we here show that CLEC10A is expressed on the majority of blood as well as lymphoid tissue CD1c<sup>+</sup> DCs. In contrast to Lundberg et al., we demonstrate that CLEC10A is specifically expressed on CD1c<sup>+</sup> DCs, but not on CD141<sup>+</sup> DCs. Our data are further supported by the transcriptional profiling of human lymphohematopoietic DC subpopulations [Figure 1; (15)] as well as our ligand binding assays using the CLEC10A ligand MUC-1-(Tn)<sub>2</sub> on various immune cell populations (Figure 5). Although we and others found a CLEC10A mRNA expression in monocytes in lymphohematopoietic tissues, we could detect no or a very moderate protein expression (Figure 2) (37). The same was true for CD141<sup>+</sup> DCs and pDCs (Figures 1 and 2). We here suggest that the lectin receptor CLEC10A might be a useful marker for a more delicate gating of CD1c<sup>+</sup> DCs, resulting in a better separation from monocyte-related cells.

In dependency on the antibody clone used, recent studies showed different results on the expression of CLEC10A on immune cells. Wong et al. compared the transcriptome of classical, non-classical, and intermediate monocytes and found a specific expression of the mRNA in intermediate monocytes compared to the other two populations of monocytes (37). However, the protein expression of CLEC10A on intermediate monocytes was only marginal and neither the mRNA nor the protein expression was compared to other cell populations of the immune system (37). In our experiments, we identified a small population of CLEC10A<sup>+</sup> monocytes in the blood (Figure 2), which might represent the described intermediate monocytes of Wong et al. (37). In another study, Li et al. generated an antibody against CLEC10A (here named DC-ASGPR) (55). In contrast to our obtained data, the anti-DC-ASGPR antibody (clone 49C11) bound to all myeloid DCs (HLA-DR<sup>+</sup> CD11c<sup>+</sup> CD123<sup>-</sup>), CD14<sup>+</sup> monocytes, and CD19<sup>+</sup> B cells (55), while we could not observe binding to blood B cells and the majority of monocytes (Figure 2). Moreover, our data reveal that only CD1c<sup>+</sup> DCs were positive for CLEC10A, while other CD11c<sup>+</sup> DCs such as CD16<sup>+</sup> DCs and CD141<sup>+</sup> DCs were represented among the CLEC10A<sup>-</sup> DCs (Figure 1). As Valladeau et al. reported that short and long isoforms of CLEC10A (DC-ASGPR) exist (56), we here speculate that the antibody used in our study (clone: H037G3) and the antibody used by Li et al. (49C11) might recognize different isoforms

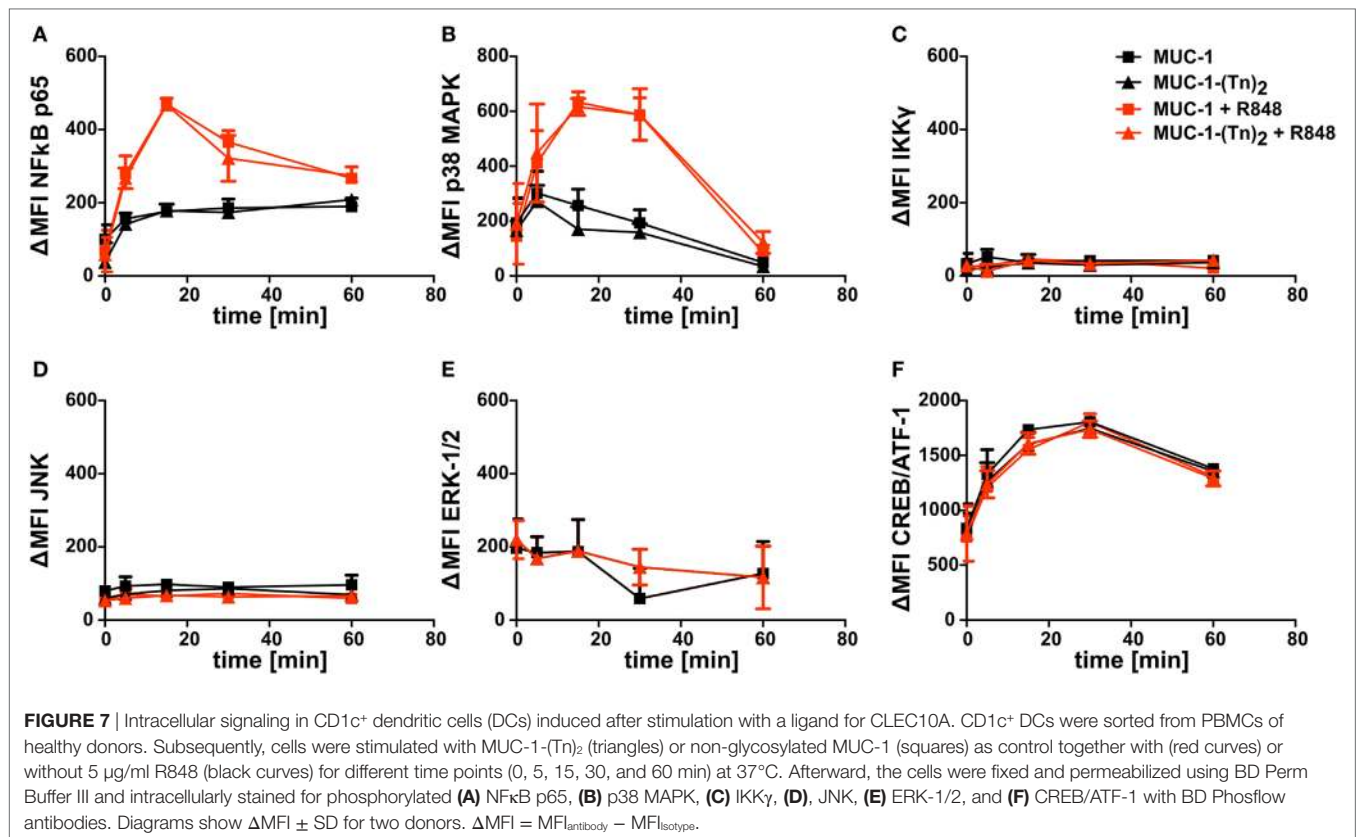


**FIGURE 6 |** Stimulation of CD1c+ dendritic cells (DCs) with MUC-1-(Tn)<sub>2</sub> enhances TLR-induced secretion of IL-8, IL-10, and TNFα. CD1c+ DCs were sorted from PBMCs of healthy donors and stimulated with 10 μg/ml MUC-1-(Tn)<sub>2</sub> or MUC-1 in the presence of 5 μg/ml R848 or 5 μg/ml pIC or in medium alone. After 12 h of stimulation, (A) cells were analyzed for the expression of the co-stimulatory molecules CD40 and CD86 and (B) the supernatant analyzed for the secretion of the cytokines IL-8, IL-10, TNFα, IL-6, and IL-12p70 with the LEGENDplex Human Anti-Virus Response Panel (BioLegend). Bar graphs show (A) ΔMFI ± SD (n = 4) and (B) mean ± SD (n = 4). p values were calculated using GraphPad Prism (paired t-test; \*p < 0.05 and \*\*p < 0.01).

of CLEC10A, which could explain the observed expression differences. The system of different CLEC10A isoforms expressed on either DCs or monocytes/macrophages would also reflect a high similarity to the murine system with CD301a and CD301b expressed on macrophages and DCs, respectively (41, 57). In

summary, as the natural ligand of CLEC10A (44) demonstrated a similar binding behavior as the CLEC10A antibody (Figure 5) and as our and other's transcriptomic analyses (Figure 1) revealed a strong specificity of CLEC10A for CD1c+ DCs, the receptor CLEC10A is approved to be a good marker for CD1c+ DCs.





Interestingly, data of Breton et al. indicate that, in addition to fully differentiated CD1c<sup>+</sup> DCs, CD1c<sup>+</sup> DC-committed precDCs also selectively expressed the mRNA of CLEC10A (17). In contrast, non-committed as well as CD141<sup>+</sup> DC-committed precDCs lacked the mRNA expression for CLEC10A (17). Although the protein expression has to be confirmed, these data suggest the usage of CLEC10A as a selective lineage marker in the development of CD1c<sup>+</sup> DCs (Figure 1).

As most *in vivo* studies on immunological questions are performed in the murine system, it is also of major importance to identify the most similar antigen-presenting cells in mice in man. For the CD8<sup>+</sup>/CD103<sup>+</sup> subset in mice and the CD141<sup>+</sup> DCs as their potential human counterpart, XCR1 and CLEC9A have been suggested as universal subset-defining markers for DCs, which potentially excel in cross-presentation of engulfed antigens (33, 34, 58–61). However, there is no such marker described for murine CD11b<sup>+</sup> DCs and human CD1c<sup>+</sup> DCs, which is not also expressed by monocytic lineages (such as CD172a, also known as SIRPα). In mice, two homologs of CLEC10A exist, named MGL1 (CD301a) and MGL2 (CD301b), respectively. These two CLRs exhibit different expression patterns as MGL1 was found on macrophages in the skin, whereas MGL2 was expressed on dermal CD11b<sup>+</sup> DCs (57). Human CLEC10A is more closely related to murine MGL2 as both can recognize similar ligands (40, 44). Although there are controversial reports on the protein expression of MGL2 on murine splenic CD11b<sup>+</sup> DCs (16, 41, 51),

we here propose that CLEC10A might not only be a suitable lineage but also a cross-species marker.

In addition to the selective expression of CLEC10A on CD1c<sup>+</sup> DCs, we further demonstrated that CLEC10A internalizes rapidly into CD1c<sup>+</sup> DCs upon binding of a specific antibody (Figure 3). A similar behavior has been reported for DEC205 (9, 62, 63). The rapid uptake is a prerequisite for the usage of CLEC10A as a potential antigen-targeting receptor. Antigen targeting takes advantage of fusing a receptor-specific antibody with an antigen of choice, such as viral, bacterial, or cancer antigens. This technique has successfully been used for the induction of immune responses against various antigens (9, 19–22, 24, 28, 64). However, till date there was no suitable CD1c<sup>+</sup> DC-specific targeting receptor described. We here propose CLEC10A as a potential antigen-targeting receptor for human CD1c<sup>+</sup> DCs.

In future studies, it will be important to analyze the compartmentalization of CLEC10A by the bound antigen-targeting antibody or the interacting natural ligand. Other lectin receptors such as the murine and human DEC205 receptors localize to late endosomes after uptake into the cell (63, 65). As the efficiency of crosspresentation in human DCs is partially dependent on the antigen routing to late or early endosomes, it will be important to follow the internalization of the CLEC10A receptor, its ligands, or its respective antigen-targeting antibodies (65–67). Importantly, we found no intracellular activation upon binding of the anti-CLEC10A antibody to its receptor (data not shown), supporting our previous findings that internalization and signaling might be



separate events (28). As CLEC10A was recently demonstrated to leap into HLA I and II rich compartments in human immature moDCs (68), we propose that targeting of CLEC10A using antigen-targeting antibodies will lead to a presentation of antigens, both on HLA I and II.

Moreover, we investigated the influence of a natural ligand of CLEC10A, MUC-1-(Tn)<sub>2</sub>, on CD1c<sup>+</sup> DCs. As expected, MUC-1-(Tn)<sub>2</sub> bound solely to CD1c<sup>+</sup> DCs, but not to other immune cells (**Figure 5**). In our analyses, we have not observed a stimulatory capacity of the ligand alone on CD1c<sup>+</sup> DCs. However, an additional costimulation of sorted CD1c<sup>+</sup> DCs with a ligand for CLEC10A and the TLR7/8 ligand R848 induced a slightly increased secretion of the cytokines IL-8, IL-10, and TNF $\alpha$  in comparison to R848 alone (**Figure 6**). This is in line with a study by van Vliet et al., which also showed an enhanced secretion of IL-10 and TNF $\alpha$  after stimulation of moDCs with TLR2 ligands in combination with either anti-CLEC10A antibodies or CLEC10A ligands (49). Furthermore, Li et al. showed that stimulation of IFN-DCs, which were generated from blood monocytes using GM-CSF and IFN $\alpha$ , with an anti-CLEC10A antibody induced ERK and p38 phosphorylation and secretion of IL-10 (55). In contrast to the study by van Vliet et al., an additional TLR stimulation was not necessary. This might be due to the culture with IFN $\alpha$ , which led to a sufficient activation of moDCs (49, 55). In contrast to these studies, we could not observe any additional phosphorylation of ERK in CD1c<sup>+</sup> DCs upon stimulation with ligands for CLEC10A by Phosflow analyses with or without TLR stimulation (**Figure 7**) (49, 55).

As the TLR7/8 ligand R848 alone induces high secretion of IL-10 by CD1c<sup>+</sup> DCs (**Figure 6**), the additional stimulation of CLEC10A could lead to only subtle changes in ERK phosphorylation, which we could not detect with Phosflow analysis. Furthermore, the induced signaling by a TLR is influenced by the localization of the TLR (69, 70). Therefore, the different localization of TLR7/8 and TLR2 could lead to the activation of different signaling molecules. Even TLR ligands targeting the same receptor induce different signaling events dependent on the targeted ectodomain of the TLR. Although both R848 and RNA40 activated TLR8, only R848 induced phosphorylation of ERK1 (71). Additionally, the used cell type influences the induced signaling after TLR stimulation, too (72). Since moDCs are more similar to human inflammatory DCs than to CD1c<sup>+</sup> DCs (73), this might additionally explain the observed difference in TLR signaling.

Stimulation of CD1c<sup>+</sup> DCs with the TLR3 ligand pIC together with glycosylated MUC1-(Tn)<sub>2</sub> did not induce enhanced cytokine secretion, although the CD1c<sup>+</sup> DCs showed a high expression of costimulatory molecules (**Figure 6**). Overall, the detected cytokine levels were notably lower compared to stimulation with the TLR7/8 ligand R848. This is in accordance with published data by other groups, who also showed a differential cytokine pattern induced by TLR3 and TLR7/8 stimulation (47, 48, 74). While TLR7/8 stimulation induces inflammatory cytokines, such as IL-6, IL-12, and TNF $\alpha$ , stimulation with pIC results mainly in secretion of IFN $\lambda$  as well as chemokines, such as RANTES and IP-10 (47, 48, 74). As TLR3, in contrast to TLR7/8, signals independent of MyD88 *via* TRIF and IRF3, we

assume that this difference in cytokine secretion is attributed to the differential signaling pathways that are induced by TLR3 and TLR7/8 (75).

Overall, CLEC10A showed a specific expression on CD1c<sup>+</sup> DCs in different lymphohematopoietic tissues rendering it as a unique marker for this subset to clearly separate it from monocytic lineages. CLEC10A did not only bind its natural ligand but also internalized rapidly upon binding of a specific antibody. Furthermore, ligands for CLEC10A alone did not induce activation or cytokine secretion by CD1c<sup>+</sup> DCs. This is a favorable feature for future immunotherapeutic approaches, since thereby antigen targeting of CLEC10A could not only be used to induce immunity but also tolerance under non-stimulatory conditions (22, 76–78).

## ETHICS STATEMENT

This study was carried out in accordance with the recommendations of the local ethical committee (Ethikkommission der Friedrich-Alexander-Universität Erlangen-Nürnberg) with written informed consent from all subjects. All subjects gave written informed consent in accordance with the Declaration of Helsinki. The protocol was approved by the local ethical committee (Ethikkommission der Friedrich-Alexander-Universität Erlangen-Nürnberg).

## AUTHOR CONTRIBUTIONS

LH performed the experiments with participation by GFH, SB, and LH; and JJJ, FG-M, and S-IN provided MGL ligands. GH provided microarray data. CL analyzed microarray data. AH, AP, and RC ensured human tissue sample supply. GH and CL contributed to the review of the manuscript. LH, FN, and DD contributed to the data analysis and interpretation as well as discussions. LH and DD designed the study; and LH, CL, and DD wrote and revised the manuscript.

## ACKNOWLEDGMENTS

We thank S. Beck and N. Eissing for technical support. We are grateful for cell-sorting support especially by M. Mroz and D. Schönhöfer (Core Unit Cell Sorting and Immunomonitoring) as well as the support by the Medical Immunology Campus Erlangen (MICE). We thank the Department of Transfusion Medicine of the University Hospital Erlangen. We are grateful for forwarding CLEC10A (generated by FG-M) ligands by Bernd Lepenies (University of Veterinary Medicine Hannover). We thank the members of the Dudziak laboratory for their critical comments.

## FUNDING

This work was partly supported by grants from the German Research Foundation [Deutsche Forschungsgemeinschaft (DFG)] to DD (CRC1181-TPA7, RTG1962, and RTG1660), FN (CRC1181-TPA7). DD and FN are funded by the Emerging Fields Initiative BIG-THERA of the Friedrich-Alexander University Erlangen-Nürnberg. LH and GH were supported by Erlanger Leistungsbezogene Anschubfinanzierung und

Nachwuchsförderung (ELAN) (DE-17-09-15-1-Heger, DE-14-10-17-1-Heidkamp). CL, FN, and DD received support from Interdisziplinäres Zentrum für Klinische Forschung (IZKF) (IZKF-J54, IZKF-A65, and IZKF-A68). DD and FN were both

supported by BayGene. DD was a fellow of the “Young Colleague” of the Bavarian Academy of Sciences. FG-M was supported by JSPS Wakate B KAKENHI Grant 24710242 and The Naito Foundation. S-IN was funded by JSPS Kakenhi Grant 25220206.

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