

Current reviews of allergy and clinical immunology

(Supported by an unrestricted educational grant from Genentech, Inc. and Novartis Pharmaceuticals Corporation)

Series editor: Harold S. Nelson, MD

Clinical and pathologic perspectives on aspirin sensitivity and asthma

Donald D. Stevenson, MD,^a and Andrew Szczeklik, MD^b La Jolla, Calif, and Krakow, Poland

This activity is available for CME credit. See page 38A for important information.

Aspirin and other nonsteroidal anti-inflammatory drugs that inhibit COX-1 induce unique nonallergic reactions, consisting of attacks of rhinitis and asthma. These hypersensitivity reactions occur in a subset of asthmatic subjects, thus identifying them as having this exclusive clinical presentation. We refer to these patients as having aspirin-exacerbated respiratory disease, a disease process that produces devastating eosinophilic inflammation of both the upper and lower respiratory tracts. This review focuses on a description of patients with aspirin-exacerbated respiratory disease, methods available to diagnose their condition, the unique ability of all nonsteroidal anti-inflammatory drugs that inhibit COX-1 to cross-react with aspirin, an update on pathogenesis, and current thoughts about treatment. (J Allergy Clin Immunol 2006;118:773-86.)

Key words: Aspirin, nonsteroidal anti-inflammatory drugs, asthma, nasal polyps, chronic hyperplastic eosinophilic sinusitis, aspirin-exacerbated respiratory disease, aspirin desensitization

In 1922, Widal et al¹ published the first article describing the association of aspirin sensitivity, asthma, and nasal polyposis. They also conducted the first aspirin challenges and desensitization. This syndrome was not widely recognized, however, until Samter published 2 articles in the late 1960s and called the condition *Samter's*

Abbreviations used

AERD:	Aspirin-exacerbated respiratory disease
CHES:	Chronic hyperplastic eosinophilic sinusitis
cysLT ₁ RA:	Cysteinyl leukotriene receptor antagonist 1
EP:	E-prostanoid
FLAP:	5-Lipoxygenase-activating protein
HETE:	Hydroxy-eicosatetraenoic acid
5-LO:	5-Lipoxygenase
5-LOINH:	5-Lipoxygenase inhibitor
LT:	Leukotriene
LTC ₄ S:	Leukotriene C ₄ synthase
NSAID:	Nonsteroidal anti-inflammatory drug
PG:	Prostaglandin
SNP:	Single nucleotide polymorphism
uLTE ₄ :	Urinary leukotriene E ₄

triad (asthma, nasal polyps, and aspirin reactions).^{2,3} Most clinical investigators now include chronic hyperplastic eosinophilic sinusitis (CHES)⁴ as a fourth hallmark of aspirin-exacerbated respiratory disease (AERD).⁵

Many other terms have been used to describe this respiratory disease: *aspirin-induced asthma*, *aspirin-sensitive asthma*, *aspirin hypersensitivity*, *aspirin idiosyncrasy*, and *aspirin intolerance*. All terms refer to the same patients who are afflicted with intractable inflammation in both the upper and lower respiratory tracts (nasal polyps, CHES, and asthma). Exposure to aspirin does not initiate or even perpetuate the underlying respiratory inflammatory disease. However, once the disease is ongoing, aspirin and nonsteroidal anti-inflammatory drugs (NSAIDs) induce release or synthesis of critical mediators, which then cause all of the clinical manifestations of the characteristic respiratory reactions. The recent consensus nomenclature to describe aspirin-induced respiratory reactions is *nonallergic hypersensitivity reactions*.⁶

PREVALENCE

Identifying the exact prevalence of AERD is difficult. Many patients have the disease but do not know it because

From ^athe Division of Allergy, Asthma and Immunology and the Department of Medicine, Scripps Clinic and the Scripps Research Institute, La Jolla, and ^bthe Department of Medicine, Jagellonian University School of Medicine, Krakow.

Disclosure of potential conflict of interest: D. D. Stevenson has received the Skaggs Scholarship Institutional Grant and study grants from Merck and Novartis, is employed by the Scripps Clinic and The Scripps Research Institute, and is on the speakers' bureau for Merck and Critical Therapeutics. A. Szczeklik declares that he has no conflict of interest.

Received for publication June 16, 2006; revised July 6, 2006; accepted for publication July 7, 2006.

Available online September 4, 2006.

Reprint requests: Donald D. Stevenson, MD, Scripps Clinic, 10666 N Torrey Pines Rd, La Jolla, CA 92037. E-mail: dstevemd@AOL.com or Stevenson.donald@scrippshealth.org.

0091-6749/\$32.00

© 2006 American Academy of Allergy, Asthma and Immunology

doi:10.1016/j.jaci.2006.07.024

they have not ingested aspirin and other NSAIDs (usually acetaminophen users). Alternatively, they experienced mild asthma attacks after ingesting NSAIDs but did not correlate these events. In a large survey of pharmacy-reported reactions to analgesics in asthmatic subjects, the prevalence of AERD was listed as 3% in adults and 2% in children.⁷ In large population surveys, using questionnaires to ask about asthma and aspirin inducing shortness of breath or asthma attacks, the prevalence was 1.2% (sample size, 4300),⁸ but the incidence of aspirin sensitivity was much higher in patients whose physician made a diagnosis of asthma (8.8%). In a large population survey in Poland (12,971 adults), 4.3% of asthmatic subjects identified themselves as reacting to aspirin with asthmatic attacks.⁹ In Perth, Australia, in a survey of 516 asthmatic patients and 1298 randomly selected individuals, the prevalence was reported to be 11% among asthmatic subjects and 2.5% among the general population.¹⁰

Investigators have performed prospective oral aspirin challenges on various populations of asthmatic patients to circumvent the problem of prior nonexposure to aspirin/NSAIDs in asthmatic patients. During such aspirin challenge studies in adult asthmatic subjects, the incidence of AERD has ranged from 10% to 20% in the pre-1990 literature.¹¹⁻¹⁴ In a meta-analysis of 15 studies performed after 1990 using oral aspirin challenges to detect aspirin hypersensitivity in asthmatic populations, the combined prevalence was 21% (CI, 14% to 29%), whereas in 5 studies in children (0-18 years) the combined prevalence was only 5% (CI, 0% to 14%).⁷

When target populations of asthmatic subjects are further stratified and include only those who also had nasal polyps and chronic sinusitis, the prevalence of aspirin hypersensitivity, discovered by means of prospective oral aspirin challenges, was found to be even higher, in the range of 30% to 40%.^{11,15} In fact, the one study in children that stands out as having a high incidence of positive oral aspirin challenge results (28%) was actually in a population of teenagers who had nasal polyps, chronic sinusitis, severe asthma, and a high prevalence of steroid dependency.¹⁶ Teenagers are probably more like adults than prepubertal children with respect to being vulnerable to acquiring AERD. The general experience is that aspirin/NSAID hypersensitivity in preschool asthmatic subjects is incredibly rare.

Asthmatic patients who reported a history of aspirin- or NSAID-induced asthma attacks (ie, believed they were "aspirin sensitive") experienced positive oral aspirin challenges of 66%,¹³ 84%,¹⁷ and 97%¹¹ of the time in these 3 studies. These observations point out the problem of overdiagnosing AERD when relying on a history of asthma after ingestion of aspirin or NSAIDs. In some patients a coincidence occurred in that aspirin or an NSAID was ingested within 3 hours of the asthma attack, but the 2 events were unrelated to each other. The best histories include ingestion of a therapeutic dose of any NSAID that preferentially inhibits COX-1, followed by a severe asthma attack requiring emergency intervention, an NSAID-associated asthma attack at another time, or both.

NATURAL HISTORY AND CLINICAL PRESENTATION OF AERD

Consistent with a low prevalence of AERD in preteens, AERD is an acquired disorder with an onset of symptoms beginning somewhere between the teenage years and age 40 years. The average ages of onset were 34 and 29 years in 2 large studies involving 300 and 500 patients with AERD, respectively.^{18,19} There are more female than male subjects who acquire this disease: in a study of 300 patients from the United States,¹⁸ a 3:2 ratio of female/male sex was found, whereas in Europe¹⁹ a 2.3:1 ratio was found. There is no racial or ethnic predilection for acquiring AERD. Family histories of AERD are rare and reported in 6% of patients in the European survey¹⁹ but in only 1% of patients in the US series.¹⁸

Aspirin hypersensitivity can appear in patients who already have allergic rhinitis and allergic asthma or any other provoking factor for their asthma (eg, gastroesophageal reflux disease, viral respiratory tract infections, irritant inhalation, or exercise) or it can appear in patients *de novo* who have never had any prior respiratory disease. In a review of 103 German and Polish patients with AERD, 34% had positive skin prick test responses to at least one aeroallergen.¹⁹ In the United States series of 300 patients with AERD, the prevalence of positive wheal-and-flare skin test responses was 64% but included both prick and intradermal testing.¹⁸

The first clinical manifestation of AERD is usually nasal congestion, but it might be superimposed on patients who already have allergic rhinitis. Many patients remember an upper respiratory tract viral infection as the inciting event ("My cold never went away"). In 1988, Szczeklik²⁰ presented a theory that a viral respiratory tract infection might be an inciting event that starts the inflammatory cascade, leading to AERD in genetically susceptible individuals. Another idea is that diesel exhaust and cigarette smoke exposure, both of which contain polyaromatic hydrocarbons, such as benzopyrene and phenanthrene, stimulate respiratory epithelial cells to synthesize cytokines (IL-1, IL-6, IL-16, and GM-CSF), which drives a T_H2 response.²¹⁻²⁵ In an epidemiologic study a significantly higher prevalence of exposure to passive cigarette smoke during childhood and young adult life was found in patients with AERD when compared with their asymptomatic spouses, acting as control subjects (unpublished data, J. Martin and D. D. Stevenson). Hyposmia or anosmia occurs in most patients with AERD. In fact, normal olfaction correlates with not having AERD.²⁶

In AERD the original chronic rhinitis progresses to CHES with nasal polyposis. Computed tomography or plain radiography of the sinuses revealed opacifications in 99% of patients in one study.¹⁸ In this same study 94% of patients with AERD had undergone at least 1 and averaged 3 prior sinus and polyp operations.¹⁸

Asthma was either previously present in childhood and young adult life (usually IgE mediated) or begins *de novo* between 3 months and up to 5 years (average, 2 years) after onset of nasal congestion and polyposis.¹⁹ Aspirin/

NSAID-induced hypersensitivity respiratory reactions can appear at any time in the course of the disease, but until such an event occurs, the diagnosis of AERD cannot be considered. Despite avoidance of aspirin and NSAIDs, mucosal inflammation of the upper and lower respiratory tracts persists and progresses. This strongly supports the fact that ingestion of aspirin and the older NSAIDs exacerbate an inflammatory condition that is already active, rather than being responsible for inducing the disease in the first place.

RESPIRATORY REACTIONS TO ASPIRIN AND NSAIDS

Cross-reactions among the NSAIDs that inhibit COX-1

Patients with AERD react to aspirin, as well as all older NSAIDs that preferentially inhibit COX-1 (Table I), inducing a spectrum of respiratory reactions, including rhinitis and conjunctivitis, laryngeal spasm, and asthma attacks.²⁷⁻³¹ The reactions usually occur within 30 to 60 minutes after ingesting full therapeutic doses of aspirin or NSAIDs but can occur up to 3 hours later, particularly when patients are challenged with smaller aspirin doses in the range of 30 to 100 mg. Assuming that the doses of NSAIDs are in the therapeutic range, cross-reactivity among NSAIDs that inhibit COX-1 is 100% (Table I).

In a study of 300 patients in the United States with documented AERD, aspirin was the most commonly reported NSAID (80%) to elicit prior respiratory reactions, followed by ibuprofen (41%).¹⁸ Of the 300 patients, 36% experienced 3 or more previous respiratory reactions to aspirin and NSAIDs, indicating that a third of patients had a significant delay in diagnosis. This suggested either observational confusion on the part of the patients and their physicians or insufficient education by health care providers regarding avoidance strategies.¹⁸

Partial cross-reactivity with poor inhibitors of COX-1

Salsalate and acetaminophen are poor inhibitors of COX-1 (Table II). However, at high doses, both can induce mild respiratory reactions in patients with AERD. Most patients with AERD can safely tolerate up to 500 mg of acetaminophen, but 28% experienced mild respiratory reactions to 1000 mg of acetaminophen, and another 6% reacted when doses were increased to 1500 mg.³² Patients with AERD can take salsalate in doses of less than 2000 mg.³³ However, when 2000 mg was given to patients with AERD, mild respiratory reactions occurred in 10% of patients undergoing oral challenges with salsalate. When reactions are elicited at higher doses of acetaminophen and salsalate, they tend to be milder than those observed with the older NSAIDs.

Partial cross-reactivity with partially selective COX-2 inhibitors

Meloxicam and nimesulide (Table II) are 2 anti-inflammatory drugs that preferentially inhibit COX-2 at lower

TABLE I. Universal cross-reactions between aspirin and non-NSAIDs occur

All of the following preferentially inhibit COX-1:*

Piroxicam (Feldene)
Indomethacin (Indocin)
Sulindac (Clinoril)
Tolmetin (Tolectin)
Ibuprofen (Motrin, Rufen, Advil)
Naproxen (Naprosyn)
Naproxen sodium (Anaprox, Aleve)
Fenoprofen (Nalfon)
Meclofenamate (Meclomen)
Mefenamic acid (Ponstel)
Flurbiprofen (Ansaid)
Diflunisal (Dolbid)
Ketoprofen (Orudis, Oruval)
Diclofenac (Voltaren, Cataflam)
Ketorolac (Toradol)
Etodolac (Lodine)
Nabumetone (Relafen)
Oxaprozin (Daypro)

*Generic name (brand names).

concentrations but inhibit COX-1 at higher therapeutic concentrations (ie, 15 mg of meloxicam).³⁴⁻³⁶ Similar to salsalate and acetaminophen, respiratory reactions can occur with higher doses of these 2 drugs and tend to be relatively mild.^{37,38} Only meloxicam is available in the United States, but both are available worldwide.

Unusual cross-reactivity with selective COX-2 inhibitors

Selective COX-2 inhibitors, such as rofecoxib, celecoxib, valdecoxib, etoricoxib, parecoxib, and lumiracoxib, are the most recent category of NSAIDs to enter and, for some, to then exit the market (Table III). Many clinicians are apprehensive about prescribing selective COX-2 inhibitors to patients with AERD because warning labels on all coxibs list “aspirin triad” as a contraindication for prescribing these drugs. Since 2001, well-designed studies, however, demonstrated that selective COX-2 inhibitors, given in therapeutic dosages, have not cross-reacted with aspirin or NSAIDs in any patients with AERD participating in these studies.

In a double-blind placebo-controlled trial, Stevenson and Simon³⁹ challenged 60 patients with AERD with rofecoxib. None of the patients reacted after ingesting 12.5 mg and 25 mg of rofecoxib. Martin-Garcia et al⁴⁰ and Szczeklik et al⁴¹ conducted single-blind challenges in a total of 52 more patients with AERD with up to 25 mg of rofecoxib, and none reacted. Woessner³¹ challenged 60 patients with AERD with even higher doses of rofecoxib (50 mg or 2 times the therapeutic dose), and again, none of the patients had any adverse reactions. Rofecoxib has been withdrawn from the market.

Celecoxib has also been studied in patients with AERD. Yoshida et al⁴² challenged 17 patients with AERD with 200 mg of celecoxib, and none reacted. Woessner et al⁴³

TABLE II. Cross-reactions with aspirin occur with higher doses of these drugs

A. NSAIDs that are poor inhibitors of COX-1
Acetaminophen (paracetamol) (Tylenol)
Salsalate (Disalcid)
B. NSAIDs that preferentially inhibit COX-2 at lower doses but also inhibit COX-1 when higher doses are given
Nimesulide (Aulin, Nimesil)
Meloxicam (Mobic)

conducted double-blind placebo-controlled challenges in which 60 patients with AERD were given 200 mg of celecoxib, and none of the patients reacted. Gyllfors et al⁴⁴ challenged 32 patients with AERD with 400 mg of celecoxib, and again, none of the patients reacted. Furthermore, there was no increase in urinary leukotriene E₄ (uLTE₄) levels during celecoxib challenges, but the usual increase in uLTE₄ levels occurred when the same patients were challenged with aspirin.

Valdecoxib is also a COX-2 inhibitor that entered and then exited the US market. Woessner³¹ completed a double-blind placebo-controlled study of 70 patients with proved AERD. None of the patients had adverse reactions to 20 mg of valdecoxib. Etoricoxib is another highly selective COX-2 inhibitor and is available by prescription outside the United States. By experience, it has been well tolerated by patients with AERD. In a recent report from Italy, 27 subjects with mostly urticarial and angioedema reactions to NSAIDs, only 3 of whom had AERD, underwent challenges with a new selective COX-2 inhibitor, parecoxib.⁴⁵ None reacted to parecoxib, which is the first injectable coxib. Lumiracoxib is another coxib available outside the United States.⁴⁶ We did not find case reports of respiratory reactions to lumiracoxib.

However, a word of caution; despite all of the safety data and lack of cross-reactivity for use of selective COX-2 inhibitors in large numbers of patients with AERD, rare case reports of respiratory reactions to selective COX-2 inhibitors have appeared in the literature. A most instructive case was reported by Baldassarre et al.⁴⁷ They described a 45-year-old woman with AERD who experienced asthma attacks by history after paracetamol (500 mg) and by oral challenge after aspirin (10 mg) and celecoxib (15 mg). In addition, during her respiratory reaction to celecoxib, her uLTE₄ level increased from a baseline value of 368 to 1318 pg/mg creatinine. Her unusual sensitivity to aspirin (10 mg) is in contrast to the average aspirin provoking dose of 60 mg (range, 30-100 mg), which is characteristic of large numbers of patients with AERD undergoing oral aspirin challenges.⁴⁸ One of the authors of this review (AS) has recently worked with a 30-year-old woman with AERD who had very high baseline uLTE₄ levels (40 times the upper limit of normal). During oral challenges, she reacted with bronchospasm after celecoxib and went into anaphylactoid shock after aspirin. Similar case reports for celecoxib,⁴⁹ rofecoxib,^{38,50} and etoricoxib⁵¹ make it impossible to take the position that coxibs never induce respiratory reactions in patients with AERD.

TABLE III. Selective COX-2 inhibitors preferentially inhibit COX-2

Celecoxib (Celebrex)*
Rofecoxib (Vioxx)†
Valdecoxib (Bextra)†
Etoricoxib (Arcoxia)‡
Parecoxib (Dynastat)‡
Lumiracoxib (Prexige)‡

In vitro studies show that in supertherapeutic concentrations of these drugs, weak inhibition of COX-1 occurs.

*Available worldwide.

†Removed from the world market in 2004 and 2005.

‡Available outside the United States.

It is difficult to understand the mechanisms that might account for the case reports described above. COX-2 inhibitors are designed with a side arm so that their entrance into the smaller COX-1 channel is prevented by the reduced size of the entrance and lack of a side pocket within the enzyme.⁵² Because of this feature, COX-2 inhibitors are between 5- and 50-fold more selective for COX-2 over COX-1.⁵³ Therefore with very high doses of selective COX-2 inhibitors, one could envision such a high drug concentration around the mouth of the COX-1 enzymes that arachidonic acid could not enter the channel. However, low to usual therapeutic doses of COX-2 inhibitors have participated in what on the surface appear to be cross-reactions.⁴⁷ We are not able to explain why these rare reactions occur, but genetic differences in the structure of the COX-1 channel would be interesting to investigate.

Some patients with AERD rarely experience respiratory, urticarial, and anaphylactic reactions through immune recognition of a specific COX-2 inhibitor.⁵⁴ It is well known that COX-2 inhibitors, like any of the NSAIDs, can induce IgE-mediated reactions.⁵⁴⁻⁵⁶ In other words, patients with AERD are not protected from experiencing other types of specific immune reactions that have nothing to do with inhibition of COX-1. Whatever the mechanisms, as prescribing physicians, we can never guarantee that patients will not react to a selective COX-2 inhibitor. It seems logical to us to give the first full dose of a COX-2 inhibitor in the physician's office to patients with AERD or asthmatic subjects with unknown sensitivities. Unfortunately, because of an increase in cardiovascular adverse events, rofecoxib and valdecoxib were withdrawn from the world market. Currently, the only remaining selective COX-2 inhibitor in the United States is celecoxib. Outside the United States, etoricoxib, parecoxib, and lumiracoxib are available (Table III).

DIAGNOSIS OF AERD

The diagnosis of AERD can be definitively established only through provocative aspirin challenges.²⁶ There is no reliable *in vitro* test, but the search for one continues.^{57,58} There are 4 types of provocation challenges, depending on

the route of administration and challenge drug: oral,^{26,59} inhalational,⁶⁰⁻⁶² nasal,⁶³⁻⁶⁵ and intravenous.⁶⁶

In the United States oral aspirin challenges are available.²⁶ Details for conducting these challenges can be found in a prior reference.²⁶ Instead of starting with 30 mg of ASA during oral challenges, one can cut ASA 81 mg in half and then a quarter and start with 20.25 mg of ASA.

Patients are instructed to continue oral and topical corticosteroids, long-acting bronchodilators, cysteinyl leukotriene receptor antagonist 1 (cysLT₁RA), 5-lipoxygenase inhibitor (5-LOINH), and systemic corticosteroids because discontinuing these medications can lead to an increase of hyperirritable airways. Some medications should be discontinued 24 hours before challenge, including antihistamines and short-acting inhaled β -agonists or anticholinergics. Antihistamines can block upper respiratory tract reactions to aspirin, which can interfere with accurate identification of patients with AERD.⁶⁷ Use of short-acting β -agonists or anticholinergics can lead to false-positive reactions because once the short-acting bronchodilator effect has disappeared, a rapid decrease in lung function can occur. If a decrease in FEV₁ is greater than 15%, diagnostic misinterpretation might develop.⁶⁸ CysLT₁RA and 5-LOINH do not block upper airway reactions but do prevent or modify bronchospastic reactions during oral aspirin challenges.^{69,70} CysLT₁RA shifts target organ responses from lower respiratory tract reactions to mostly upper respiratory tract reactions.⁶⁹

Although not available in the United States, several European centers use bronchial challenges with aspirin-lysine.^{62,71} Nizankowska et al⁶² studied the diagnostic value of bronchial inhalation challenge with L-lysine-aspirin in 35 patients with AERD who were suspected of having AERD on the basis of a prior history of NSAID reactions. Thirty-one (89%) of 35 suspected asthmatic subjects experienced bronchospastic reactions during oral aspirin challenges. In the same patients, 27 (77%) of 35 or, more accurately, 27 (87%) of 31 known reactors had bronchospastic reactions during inhaled aspirin-lysine challenges.

Aspirin-lysine nasal challenges are also conducted in Europe and have a satisfactory diagnostic capability.^{63-65,72} In a recent study by Micheletto et al,⁶⁴ both asthmatic patients with AERD and aspirin-tolerant asthmatic patients underwent nasal challenges with up to 25 mg of aspirin-lysine. Positive nasal responses and increased uLTE₄ levels occurred only in the patients with AERD. Despite increased synthesis of leukotrienes (LTs), as measured in uLTE₄, none of the patients with AERD experienced asthma attacks during nasal challenges, despite significant nasal responses. The major advantage of intranasal aspirin-lysine challenges is avoidance of aspirin-induced bronchospasm. The disadvantages are that pure lower respiratory tract reactors might not be identified, and in some patients with AERD, occluding nasal polyps interfere with nasal flow rates and make it impossible to perform the test.⁶⁵ In the United States a new diagnostic test for nasal challenge using a dilute solution of ketorolac has recently completed diagnostic trials.⁷³ Ketorolac solutions (8 mg/mL),

delivered as a nasal spray in increasing doses every 30 minutes, offer an alternative to aspirin-lysine nasal challenge in the United States because aspirin-lysine has not been approved for use in human subjects by the US Food and Drug Administration.

PATHOGENESIS

Underlying respiratory disease (Fig 1)

The pathophysiology of AERD has been partially elucidated. Nasal tissue biopsy specimens from patients with AERD reveal extensive infiltration of eosinophils and degranulated mast cells.⁷⁴ Bronchial biopsy specimens also contain increased numbers of eosinophils and mast cells when compared with biopsy specimens from patients with aspirin-tolerant asthma.⁷⁵ Why the eosinophils and activated mast cells infiltrate the respiratory mucosa in the first place is not clear and difficult to study because any prior inciting events, such as viral infections or exposure to air pollution and cigarette smoke, might have occurred years earlier. Nevertheless, once AERD appears, levels of proinflammatory cytokines synthesized by epithelial cells and activated T_H2 lymphocytes are found to be increased. These include IL-2, IL-3, IL-4, IL-5, IL-13, GM-CSF, and eotaxin.^{4,76-78} The cytokines IL-3, IL-4, IL-5, IL-13, and GM-CSF all skew T-lymphocyte responses toward T_H2, stimulate bone marrow precursor cells, recruit eosinophils, and dramatically increase the lifespan of eosinophils *in vitro* by inhibiting apoptosis.⁷⁷ IL-5 and GM-CSF are overexpressed in inflammatory cells from mucosal biopsy specimens in patients with AERD.⁷⁹ Eotaxin is also an important chemokine, the primary function of which is recruitment and activation of eosinophils, as well as contribution to tissue damage through induction of reactive oxygen radicals.⁸⁰ LTC₄, LTD₄, and LTE₄ are also chemotactic for eosinophils.⁸¹ Mast cells contain LTA₄ hydrolase and therefore can convert LTA₄ to LTB₄, which stimulates bone marrow to form mast cell progenitors and eosinophils.⁸² LTC₄ synthase (LTC₄S) converts LTA₄ to LTC₄,⁸² thus activating this potent pathway.^{83,84} The end result is a striking increase in numbers of eosinophils and mast cells in the respiratory mucosa of patients with AERD. Activated eosinophils can also release cytotoxic molecules (eg, eosinophilic cationic protein, major basic protein, eosinophil-derived neurotoxin, and eosinophil peroxidase), leading to respiratory mucosal inflammation and damage.⁸⁵ Despite identification of the above mechanisms, none are restricted to patients with AERD. In fact, these same inflammatory pathways and patterns can be found in non-aspirin-sensitive asthmatic subjects with nasal polyps and CHES or in allergic asthmatic subjects.

There is clear evidence that most, but not all, patients with AERD synthesize excessive amounts of LTs, even before any exposure to aspirin or NSAIDs.⁸⁶ Higher concentrations of LTC₄ and thromboxane B₂ were found in bronchoalveolar lavage fluid taken from patients with AERD compared with those seen in samples from control asthmatic subjects and healthy patients.^{75,87} Christie et al⁸⁸

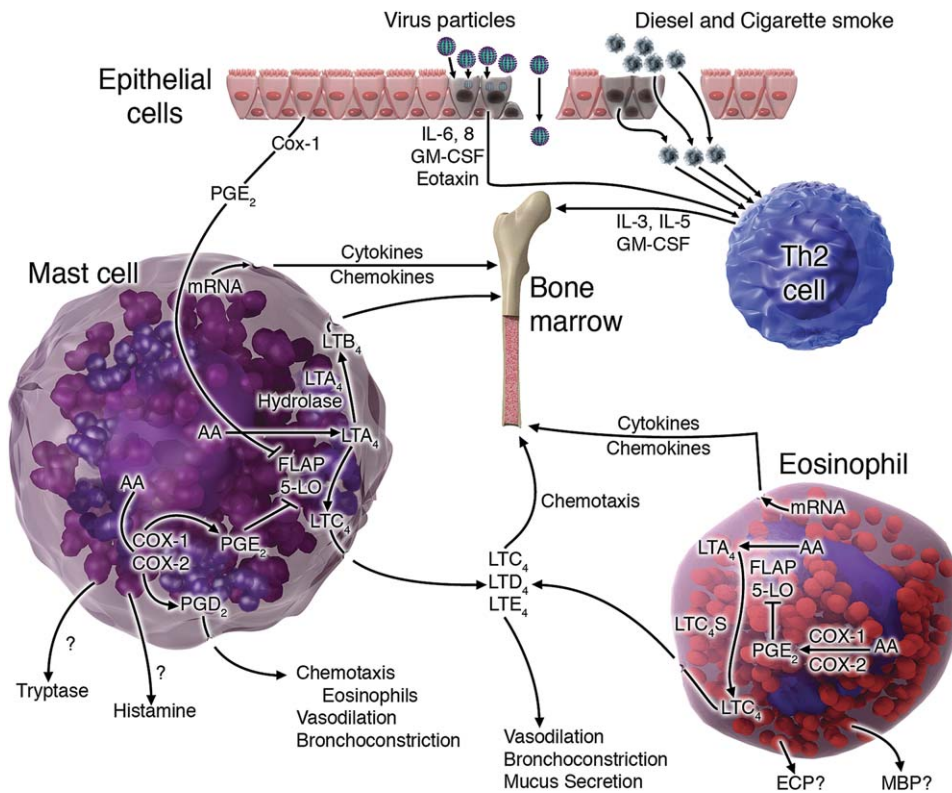


FIG 1. Pathogenesis of AERD. Presumably, a damaging stimulus of respiratory epithelial cells initiates bone marrow formation of mast cell progenitor cells and eosinophils. In AERD these cells are the major contributors to local inflammation. Density and polymorphism of EP2 receptors, as well as cell-activating receptors, are not shown. Chemotaxis of new mast cell progenitor cells and eosinophils from bone marrow perpetuates the disease. AA, Arachidonic acid; ECP, eosinophil cationic protein; MBP, major basic protein.

and Smith et al⁸⁹ were the first to measure increased levels of LTE₄ in the urine of patients with AERD before aspirin challenges. Many, but not all, patients with AERD demonstrate overexpression of LTC₄S in eosinophils and mast cells in their bronchial biopsy specimens,⁸³ and their circulating eosinophils carry more mRNA for LTC₄S.⁹⁰ Sanak et al⁹¹ discovered genetic polymorphisms of the LTC₄S promoter region and identified an increased prevalence of its variant type in Polish patients with aspirin-induced asthma, although the same finding also occurred in samples from some healthy individuals. Attempts to find similar patterns in more heterogeneous asthmatic subjects with AERD did not identify an increase in polymorphisms for the flanking region of the gene encoding for LTC₄S.^{92,93} Not only is there overproduction of LTs in AERD, but Sousa et al⁹⁴ demonstrated that in asthmatic subjects with AERD, but not in aspirin-tolerant asthmatic subjects, nasal inflammatory cells expressed more cysLT₁ receptors. In patients with AERD, not only is there overproduction of LTs, but more receptors are available to receive these mediators.

Arachidonic acid is synthesized to 5-hydroxyperoxy-eicosotetraenoic acid by 5-lipoxygenase (5-LO) and 5-lipoxygenase-activating protein (FLAP), with synthesis then proceeding down the LTA₄ pathway or into the

alternative 5-hydroxy-eicosatetraenoic acid (HETE) pathway. Leukocytes and platelets synthesize a microsomal enzyme called 5-hydroxyeicosanoid dehydrogenase, which selectively converts 5-HETE to 5-oxo-6,8,11,14-eicosatetraenoic acid (5-OXO-ETE).⁹⁵ Respiratory stress and oxidative stress of polymorphonuclear cells activate synthesis of 5-OXO-ETE, which is a potent chemoattractant for PMNs and eosinophils. Although not specifically investigated in AERD, this proinflammatory mechanism might also be active in AERD, the extent of which is currently unknown.

Just as overproduction of cysLTs is the hallmark of AERD, underproduction of lipoxins correlates with having AERD.⁹⁶ Lipoxins are anti-inflammatory derivatives of arachidonic acid and products of lipoxygenation. They require 2 or more lipoxygenase enzymes for biosynthesis (5-LO and 15-LO). They are generated by transcellular cooperation and are functional antagonists of LTs.⁹⁷⁻⁹⁹ Therefore diminished capacity to generate lipoxin and 15-epimer lipoxin might contribute to uncontrolled and protracted inflammation in patients with AERD.^{96,99} The possible relationship between this phenomenon and the accumulation of 15-HETE after stimulation with aspirin of peripheral leukocytes from patients with AERD¹⁰⁰ is interesting but needs further study.

Prostaglandin (PG) D₂, a mast cell–derived prostanoid that is synthesized through the COX-1 and COX-2 pathways, is oversynthesized and secreted in asthmatic subjects with AERD.¹⁰¹ PGD₂ causes vasodilatation and bronchoconstriction. It is also a potent chemoattractant for eosinophils, operating through prostaglandin D₂ receptors [DP(2)] receptors on eosinophils.^{102,103} Therefore in patients with AERD, not only is there an increase in 5-LO products, but certain prostanoids are also proinflammatory and tend to be oversynthesized.

A special role for PGE₂ in the pathogenesis of AERD has been suggested.¹⁰⁴ Peripheral blood macrophage cells (PBMCs) of some patients with AERD undersynthesize PGE₂ at baseline.¹⁰⁵ This places patients with AERD at the disadvantage of lacking sufficient concentrations of cellular or transcellular PGE₂ to stabilize mast cells and slow synthesis of LTs. There are 4 receptors for PGE₂, E-prostanoid (EP) receptors 1 through 4. In one study single nucleotide polymorphisms (SNPs) in the promoter region of the gene encoding EP2 were significantly associated with AERD.¹⁰⁶ Thus decreased transcription of the EP2 receptor for PGE₂ might render that subset of patients with AERD unable to efficiently inhibit 5-LO and FLAP, even if they synthesized normal amounts of PGE₂. Nasal mucosal biopsy specimens from patients with AERD, when immunostained for all 4 EP receptors, showed a significant reduction in numbers of neutrophils, mast cells, eosinophils, and T cells expressing EP2 receptors but not EP1, EP3, and EP4 receptors.¹⁰⁷ Another study demonstrated a special role for EP3 receptors in suppressing allergic inflammation and suggested that patients with AERD might depend more on the PGE₂-EP3 receptor pathway than do aspirin-tolerant asthmatic subjects and healthy subjects.¹⁰⁸ Any of these discovered defects, under synthesis of PGE₂ or its EP2 or EP3 receptors, diminishes the blocking capabilities of PGE₂ on 5-LO and FLAP or an inhibitory effect on mast cells.¹⁰⁹

Recently, several authors addressed the problem of genetic polymorphism.¹¹⁰⁻¹¹⁴ The most comprehensive study of genetic associations with AERD was published by Jinnai et al.¹⁰⁶ Using 370 SNPs from 63 candidate genes, almost 200 patients with AERD were compared with aspirin-tolerant asthmatic subjects and control subjects. A gene coding for EP receptor 2 was the only one significantly associated with AERD. The investigators, using 24 SNPs' haplotypes, demonstrated that a region located 11 kb upstream from the translation start of the gene had the best correlation with the AERD phenotype. A particular allele, –12 813A instead of G, *in vitro* had diminished transcriptional activity when stimulated by IL-4 through the IL-4 receptor–signal transducer and activator of transcription 6 pathway. This finding recaptured attention when Ying et al.¹⁰⁷ demonstrated that inflammatory cells infiltrating the nasal mucosa of patients with AERD were deficient in EP2.

Nearly a decade after the initial description of an association between the HLA DPB1 locus and AERD,¹¹² these data were replicated by Choi et al.¹¹¹ Despite a quite different ethnicity (Polish vs Korean), the same allele,

DPB1*0301, was overrepresented in aspirin-hypersensitive asthmatic subjects. Thus an adaptive immune response to MHC class II antigens has probably occurred to mediate at least a step in the development of AERD.

Attempting to put this whole picture into perspective, many of the inflammatory pathways described above are also found in patients who have no reactions to aspirin and other NSAIDs. However, one theme that runs through most studies is that in patients with AERD, measurements of cytokines, eicosanoid mediators (uLTE₄ and PGD₂), and cysLT₁ receptors tend to group in the high end of most study results. Yet clearly separating individual asthmatic subjects with AERD from aspirin-tolerant asthmatic subjects cannot be accomplished on the basis of their inflammatory profiles, including their uLTE₄ levels, because of considerable overlap between patients with AERD and aspirin-tolerant asthmatic subjects. To make matters more difficult, patients with AERD can be mild, moderate, or severe in their clinical disease presentation and their inflammatory profiles.⁸⁶ Despite these comments, a pattern is emerging in which many defects in either overstimulation of inflammation or underproduction of countermeasures, especially in the eicosanoid family, is found in patients with AERD. Furthermore, a single genetic defect or promoter gene that accounts for all patients with AERD has not been discovered. Rather, some patients with AERD are upregulating LTC₄S, others are upregulating cysLT₁ receptors, and still others are oversynthesizing specific cytokines. At the opposite end of the equation, some patients with AERD are undersynthesizing lipoxins, PGE₂, or EP2 or EP3 receptors. These facts would fit a theory in which multiple divergences in inflammatory pathways could occur in different patients with AERD, yet rendering all patients vulnerable to aspirin.

Aspirin- and NSAID-induced hypersensitivity reactions (Fig 2)

In patients with AERD, acute respiratory reactions induced by aspirin or NSAIDs encompass all the features of immediate IgE-mediated hypersensitivity reactions, yet such a mechanism has never been demonstrated.⁵⁴ This fact is logical when one considers that all structurally distinct NSAIDs that inhibit COX-1 can cause respiratory tract reactions in all patients with AERD on first exposure to the new NSAID.^{27,28,115} Thus drug hapten-antibody recognition cannot be responsible for these reactions.³

When patients with AERD undergo aspirin challenges, very substantial increases in uLTE₄ levels and decreases in COX-1 products are recorded during bronchospastic reactions^{86,88,116} or nasal responses.⁶⁴ Increases in LTC₄ and histamine levels in both nasal¹¹⁷⁻¹¹⁹ and bronchial lavage⁷⁵ fluid after oral aspirin challenges in patients with AERD, but not aspirin-tolerant control subjects, have also been measured. As shown in Fig 2, during aspirin-induced respiratory reactions, mast cells release histamine and tryptase and synthesize prostanoids (PGD₂) and LTs, and eosinophils secrete toxic molecules and synthesize LTs.^{109,120}

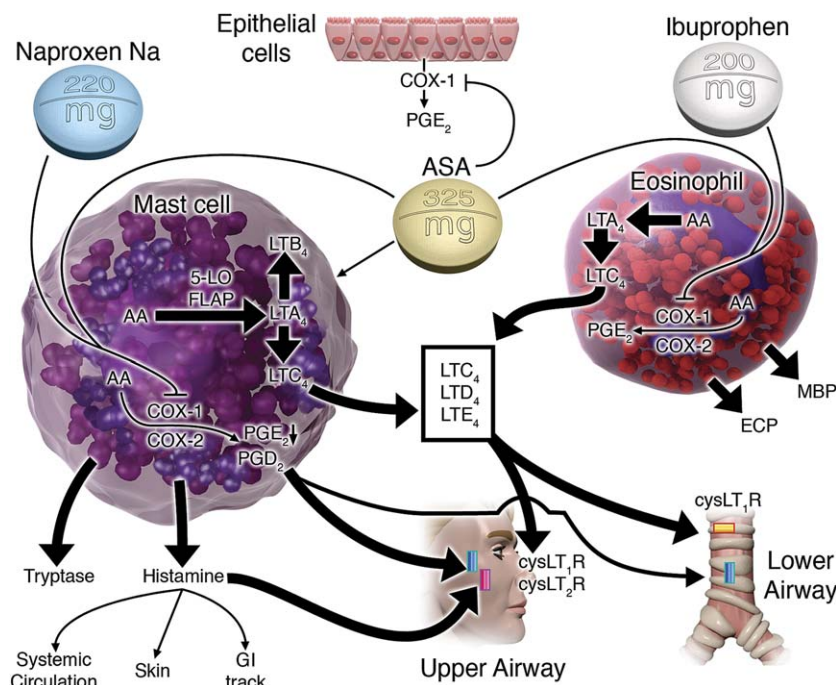


FIG 2. Aspirin or NSAIDs induced respiratory reactions. Aspirin and NSAIDs inhibit COX-1, thus depriving mast cells and eosinophils of PGE₂ to block the ongoing synthesis of LTs. Alteration in function of PGE₂ receptors (particularly EP2) might contribute to the inability of PGE₂ to sustain the blockade. From mast cells, PGD₂ is oversynthesized, and histamine and tryptase are released during reactions. Eosinophil cationic protein (ECP) and major basic protein (MBP) are released from eosinophils. AA, Arachidonic acid.

Over the years, it has become clear that excessive synthesis of LTs in patients with AERD undergoing respiratory reactions to NSAIDs or aspirin is secondary to competitive inhibition or disabling of COX-1 enzymes (Fig 2).¹²⁰ COX-1 is a constitutively expressed enzyme that is present in most mammalian cells, including respiratory and gastrointestinal epithelial cells, as well as inflammatory cells. COX-2, by contrast, is only expressed in inflammatory cells and is an inducible enzyme that is highly upregulated by proinflammatory mediators, such as cytokines, growth factors, and molecules coming from tissue injury.¹²⁰ When COX-1 is inhibited by aspirin or cross-reacting NSAIDs, there is a rapid decrease in the synthesis of COX-1 products, including PGE₂. In healthy individuals and, to a lesser extent, patients with AERD, PGE₂ inhibits 5-LO and FLAP enzymes that otherwise would generate potent proinflammatory mediators, such as LTC, LTD, and LTE₄ and LTB₄.

When COX-1 enzymes are inhibited by aspirin and NSAIDs, the braking effects of PGE₂ are further reduced or disappear, and 5-LO is unopposed, allowing large increases in synthesis of LTs.¹⁰⁴ Reduced PGE₂ synthesis also results in decreased mast cell stability and increased release of histamine and tryptase.¹⁰⁹ During oral aspirin challenges, when patients with AERD were pretreated with inhaled PGE₂, they did not experience respiratory reactions, and uLTE₄ levels did not increase.^{121,122} The failure of COX-2 inhibitors to cross-react in patients with AERD is further evidence that COX-2 enzymes do

not synthesize enough PGE₂ to make a difference in the loss of PGE₂. This should not be surprising because the small numbers of inflammatory cells synthesizing PGE₂, when compared with the millions of cells expressing constitutive COX-1, all epithelial and endothelial cells of the respiratory tract, makes inhibition of COX-2 a minor event from the standpoint of available PGE₂. Furthermore, expression of COX-2 is diminished and its activity is reduced in patients with AERD.^{123,124} The fact that meloxicam does not cross-react with aspirin in AERD with low doses (at which COX-2 is inhibited) but does cross-react at high doses (at which COX-1 is inhibited) provides further evidence that COX-1 inhibition is the key event in the induction of aspirin/NSAID-induced respiratory reactions. Finally, the defect of undersynthesizing PGE₂ or decreased transcription of EP2 receptors for PGE₂ might well render patients with AERD preferentially unable to inhibit 5-LO and FLAP when challenged with aspirin or NSAIDs. This area of research is critical because explaining why inhibition of COX-1 and COX-2 in healthy subjects or aspirin-tolerant asthmatic subjects does not lead to the same respiratory reactions hinges on as-yet-undiscovered fundamental defects unique to patients with AERD.

In addition to increased synthesis of LTs, most patients with AERD also have increased expression of cysLT₁ receptors on their inflammatory cells.⁹⁴ Thus not only is there an absence of braking effects by PGE₂ and a surge of new LT molecules, but the number of cysLT₁ receptors on target cells

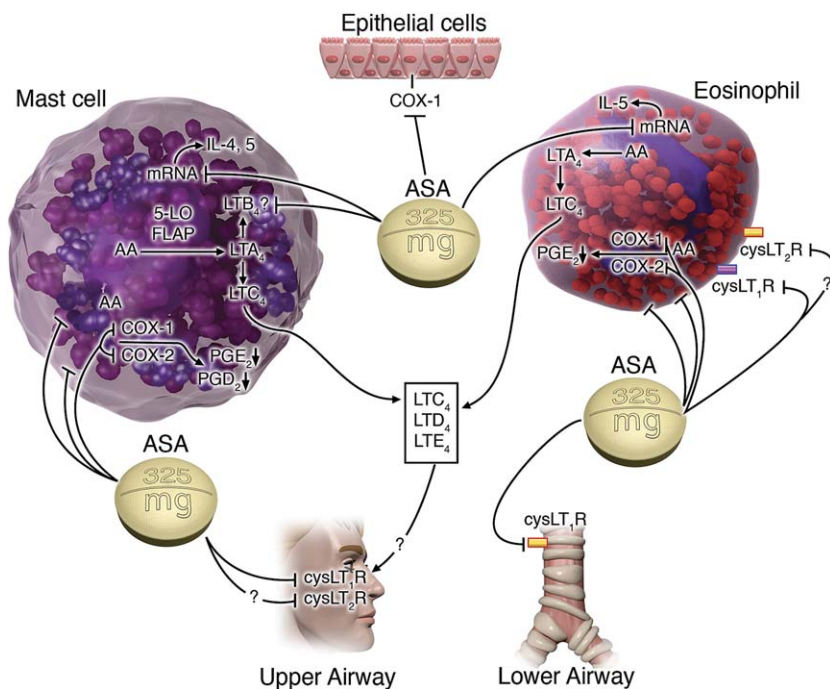


FIG 3. Aspirin desensitization. After aspirin disables COX-1 and COX-2, inflammatory cells are unable to synthesize prostanoids, including the proinflammatory PGD₂ and the anti-inflammatory PGE₂. Histamine and tryptase are no longer released from mast cells. Eosinophils stop releasing toxic molecules. CysLT receptors are underexpressed after aspirin treatment, and it is likely that intracellular transcription factors are also inhibited by aspirin.

are also increased. This tips the equation toward a pronounced increase in end-organ responses to LTs.¹²⁵

LTs are not the only mediators released or synthesized during aspirin-induced respiratory reactions. Nasal secretions obtained during aspirin-induced reactions contained increased concentrations of histamine, tryptase, LTB₄, LTC₄, and PGD₂ and decreased concentrations of PGE₂.^{117-119,126} The same is true for bronchial secretions during aspirin-lysine-induced lower respiratory tract reactions.⁷⁵ PGD₂ metabolites increase in serum samples during aspirin-induced respiratory reactions.¹⁰¹ During oral aspirin challenges, a minority of patients with AERD have extrapulmonary reactions.⁵⁴ In such patients histamine and tryptase levels were found to be increased in the systemic circulation.¹²⁷

Aspirin desensitization (Fig 3)

Aspirin desensitization is one of the most consistent and least understood features of aspirin and asthma. Almost all patients with AERD can be desensitized to aspirin and then take aspirin indefinitely to maintain their desensitized state. During aspirin desensitization, uLTE₄ levels returned to baseline,⁸⁶ and cysLT₁ receptor levels decreased significantly.⁹⁴ At 2 weeks after daily treatment with aspirin, 650 mg twice daily, LTB₄ synthesis by peripheral monocytes also decreased significantly in patients with AERD.¹²⁸ A reduction in synthesis of LTB₄ would be therapeutically useful in reducing chemotaxis of eosinophils and polymorphonuclear leukocytes, as well as mast

cell progenitors. LTC₄ and histamine disappeared in nasal secretions at the onset of aspirin desensitization.¹¹⁷ Biochemically, we can measure specific changes that downregulate inflammation and accompany aspirin desensitization. However, why aspirin desensitization occurs in the first place continues to be a challenging question.

TREATMENT

Avoidance of aspirin and NSAIDs and treatment of reactions

Education of patients regarding complete avoidance of COX-1 inhibitors is important. In particular, asthmatic patients need to be vigilant when ingesting over-the-counter remedies that might include aspirin or NSAIDs (eg, Alka Seltzer Plus Flu, which contains 500 mg of aspirin). In addition, appropriate flagging of charts and communication among health care professionals, including physician and pharmacy computers, is vital in preventing future inappropriate administration of cross-reacting NSAIDs in those patients with AERD already given a diagnosis. We are not advocating universal avoidance of NSAIDs for all asthmatic patients. Such a position denies 80% to 90% of asthmatic subjects access to these important medications. However, use of COX-2 inhibitors, extra care, and thoughtful use of first-dose ingestion of COX-1 inhibitors in physicians' offices might prevent some inadvertent bronchospastic catastrophes. The major

danger of death from asthma in AERD is the first NSAID-induced reaction, particularly if this occurs far from a medical facility.

Depending on the severity, acute respiratory reactions caused by accidental aspirin or NSAID ingestion are treated with inhaled β -agonists by using multiple dosing (5 inhalations, wait 5 minutes, and then keep repeating inhalation treatments), antihistamines, systemic corticosteroids, and, if systemic histamine release is present, intramuscular epinephrine. Patients who experience laryngospasm respond rapidly to racemic epinephrine administered by means of nebulization. Some patients will need more intense medical attention, including admission to the intensive care unit, intubation, and mechanical ventilation.

Treatment of the underlying respiratory tract disease

Long-term control of both upper and lower airway inflammation is the goal of treatment. High doses of intranasal steroids are helpful in reducing inflammation and retarding nasal polyp formation in some patients.¹²⁹ During acute bacterial sinus infections, extended courses of broad-spectrum antibiotics are frequently required.¹³⁰ Often patients will also respond to a 2- to 3-week burst of systemic corticosteroids to aid in shrinking nasal polyps and reestablishing temporary sinus drainage. A subset of patients will gradually require continuous systemic corticosteroids. In a review of 300 patients with AERD, systemic corticosteroids were used as short courses in 134 (45%), on a daily basis in 95 (32%), and not at all in 71 (23%).¹⁸ Unfortunately, as daily or frequent bursts of systemic doses of corticosteroids escalate, significant adverse side effects also begin to accumulate.

Zileuton (a 5-LOINH) and montelukast (a cysLT₁RA), are commonly used in patients with AERD, with variable success. Dahlen et al^{131,132} studied the effect of zileuton on the clinical course of 40 patients with AERD in a double-blind placebo-controlled treatment trial and demonstrated efficacy. Montelukast has also been studied in a double-blind placebo-controlled treatment trial in 80 patients with AERD, in which efficacy was also demonstrated.¹³³ CysLT₁RA treatment success is distinctly better in the carriers of the variant C allele of LTC₄S¹³⁴⁻¹³⁷ and in individuals with the HLA-DPB1*0301 marker.¹³⁸ In our experience LT-modifier drugs are generally helpful as adjunctive therapy. Particularly in view of the fact that AERD is largely the result of overproduction of cysLTs, the addition of a 5-LOINH or a cysLT₁RA to a baseline of topical corticosteroids is now fairly routine treatment for AERD. The use of both zileuton and a cysLT₁RA has never been formally studied in patients with AERD but is used by clinicians with anecdotal success.

In patients with AERD who are also atopic, treatment of underlying allergic inflammation should also be maximized. Allergen avoidance, antihistamines, immunotherapy, and anti-IgE treatment should be strongly considered as adjunctive treatment in patients with AERD with this concomitant disorder. It does not make any sense to ignore

a concomitant disease, such as allergic rhinitis and asthma, and conclude that AERD by itself is the only important mechanism that is driving eosinophilic inflammation in patients with both AERD and allergic respiratory disease. In a study of 300 patients with AERD, two thirds had positive wheal-and-flare skin test responses to relevant allergens.¹⁸

When maximal medical management of nasal polyposis has failed, which is common, referral to a competent otolaryngologist should be initiated. In fact, many patients begin their medical journey with an otolaryngologist because nasal polyps and anosmia are an early and devastating manifestation of the disease. Nasal polypectomies, resection of eosinophilic inflammatory tissue, and widening of sinus ostia can be performed to help reestablish proper drainage. In addition, at the time of surgery, specimens can be sent for cultures and pathologic investigation, which can be helpful in choosing appropriate antibiotic or antifungal therapy. McFadden et al¹³⁹ followed 22 patients with AERD who underwent endoscopic sinus surgery for 1 year. Patients showed improvement in pulmonary function testing, need for topical and systemic corticosteroids, and quality-of-life measures. Kennedy¹⁴⁰ reported that patients with AERD have the same long-term, postsurgical outcomes as aspirin-tolerant patients, if they have the same degree of mucosal disease. However, patients with AERD as a group tend to have a larger burden of polypoid tissue, and postsurgical regrowth of polypoid tissue remains a significant problem.¹⁴¹ On average, reoperation for nasal polyps is required every 3 years in patients with AERD.^{18,142}

Aspirin desensitization

Aspirin desensitization is an effective yet underused means of treating patients with AERD. Almost all patients with AERD can be desensitized to aspirin.¹⁴³ Once desensitized and maintained with daily ingestion of aspirin, patients not only enjoy significant improvement in both upper and lower respiratory symptoms but can also ingest any of the cross-reacting NSAIDs without acute respiratory reactions.^{144,145} Four long-term studies of patients who underwent aspirin desensitization followed by daily aspirin ingestion have demonstrated efficacy in reducing upper airway congestion and nasal polyp formation and improving lower airway asthma control.^{142,145-147} One study involved 172 patients desensitized between 1995 and 2000 and treated with aspirin, 650 mg twice daily, for 1 to 5 years.¹⁴⁵ Significant reductions in sinus infections and number of rescue oral steroid courses were observed, as well as improvements in anosmia, rhinitis, and asthma symptom scores. Of the 126 patients who completed a year or more of aspirin treatment, 87% experienced good or excellent improvement in their clinical courses. In another study patients began to enjoy improvement in upper airway congestion as early as 4 weeks after starting aspirin treatment.¹⁴⁴

Aspirin desensitization should be considered as add-on treatment in patients with uncontrolled upper and lower respiratory symptoms, patients requiring multiple polypectomies and/or sinus operations, patients requiring unacceptably high intermittent or chronic systemic

corticosteroids, and patients requiring aspirin/NSAIDs for treatment of other diseases, such as prophylaxis for coronary artery disease or postcoronary stent protection against thrombosis and acute myocardial infarction.

Performing aspirin challenges followed by desensitization is relatively safe.¹⁴⁷ We are not aware of any published or unpublished reports of deaths during controlled oral aspirin challenges. The risk of aspirin-induced asthma can be significantly reduced by using pretreatment with cysLT₁RAAs.^{69,70} Because the reaction severity is ASA dose dependent, the degree of respiratory reactions is almost always smaller than the original reported aspirin/NSAID-induced reaction with full therapeutic doses of drug.

Chronic aspirin therapy can result in well-known adverse side effects in a minority of patients. In the most recent Scripps Clinic study, 24 (14%) of 172 patients had to discontinue aspirin therapy because of side effects: 14 had epigastric pain, 2 had gastrointestinal bleeding, 2 had bleeding from the nose and ear, and 6 had aspirin-induced urticaria.¹⁴⁵ Patients with a prior history of gastritis, gastroduodenal ulcers, and gastroesophageal reflux might be at higher risk for development of aspirin-induced gastritis, but no prospective study of such patients has been conducted.

CONCLUSION

AERD is a distinct clinical entity that is characterized by aspirin-induced respiratory reactions, asthma, nasal polyposis, and CHES. If not recognized and treated appropriately, AERD has the potential to cause significant morbidity and even mortality, particularly when full doses of aspirin or NSAIDs are ingested away from an acute-care medical facility. Patients must be educated regarding avoidance of aspirin and cross-reacting COX-1 inhibitors to prevent potential life-threatening asthma exacerbations. Treatment of nasal polyp and sinus disease is also essential to effectively control asthma, as well as to significantly prevent secondary respiratory tract infections and improve patients' quality of life. Aspirin desensitization should be considered as add-on treatment for AERD in many patients. Hopefully, etoricoxib, parecoxib, and lumiracoxib, new selective COX-2 inhibitors available in Europe and elsewhere, will find their way into the US market over the next few years. Further pharmacologic advances, such as a new 5-LO or FLAP inhibitor with improved pharmacokinetics might also be on the horizon.

REFERENCES

1. Widal MF, Abrami P, Lermeyez J. Anaphylaxie et idiosyncrasie. *Presse Med* 1922;30:189-92.
2. Samter M, Beers RF. Concerning the nature of the intolerance to aspirin. *J Allergy* 1967;40:281-93.
3. Samter M, Beers R Jr. Intolerance to aspirin: clinical studies and consideration of its pathogenesis. *Ann Intern Med* 1968;68:975-83.
4. Borish L. Sinusitis and asthma: entering the realm of evidence-based medicine. *J Allergy Clin Immunol* 2002;109:606-7.
5. Stevenson D, Sanchez-Borges M, Szczeklik A. Classification of allergic and pseudoallergic reactions to drugs that inhibit cyclooxygenase enzymes. *Ann Allergy Asthma Immunol* 2001;87:177-80.
6. Johannessen SGO, Bieber T, Dahl R, Friedmann PS, Lanier BO, Lockey RF, et al. Revised nomenclature for allergy for global use: report of the Nomenclature Review Committee of the World Allergy Organization, Oct 2003. *J Allergy Clin Immunol* 2004;113:832-6.
7. Jenkins C, Costello J, Hodge L. Systematic review of prevalence of aspirin-induced asthma and its implications for clinical practice. *BMJ* 2004;328:434-7.
8. Hedman J, Kaprio J, Poussa T, Nieminen MM. Prevalence of asthma, aspirin intolerance, nasal polyps and chronic obstructive pulmonary disease in a population-based study. *Int J Epidemiol* 1999;28:717-22.
9. Kasper L, Sladek K, Duplaga M, Bochenek G, Liebhart J, Gladysz U, et al. Prevalence of asthma with aspirin hypersensitivity in the adult population of Poland. *Allergy* 2003;58:1064-6.
10. Vally H, Taylor M, Thompson PJ. The prevalence of aspirin intolerant asthma in Australian asthmatic patients. *Thorax* 2002;57:569-74.
11. Delaney JC. The diagnosis of aspirin idiosyncrasy by analgesic challenge. *Clin Allergy* 1976;6:177-81.
12. McDonald J, Mathison DA, Stevenson DD. Aspirin intolerance in asthma—detection by challenge. *J Allergy Clin Immunol* 1972;50:198-207.
13. Spector SL, Wangaard CH, Farr RS. Aspirin and concomitant idiosyncrasies in adult asthmatic patients. *J Allergy Clin Immunol* 1979;64:500-6.
14. Stevenson DD, Mathison D, Tan E, Vaughn J. A study of provoking factors in bronchial asthma. *Arch Intern Med* 1975;135:777-83.
15. Weber RW, Hoffman M, Raine DA, Nelson HS. Incidence of bronchoconstriction due to aspirin, azo dyes, non-azo dyes, and preservatives in a population of perennial asthmatics. *J Allergy Clin Immunol* 1979;64:32-7.
16. Rachelefsky GS, Coulson A, Siegel SC, Stiehm ER. Aspirin intolerance in chronic childhood asthma: detection by oral challenge. *Pediatrics* 1975;56:443-8.
17. Pleskow WW, Stevenson DD, Mathison DA, Simon RA, Schatz M, Zieger RS. Aspirin-sensitive rhinosinusitis/asthma: spectrum of adverse reactions to aspirin. *J Allergy Clin Immunol* 1983;71:574-9.
18. Berges-Gimeno M, Simon RA, Stevenson DD. The natural history and clinical characteristics of aspirin exacerbated respiratory disease. *Ann Allergy Asthma Immunol* 2002;89:474-8.
19. Szczeklik A, Nizankowska E, Duplaga M. Natural history of aspirin-induced asthma. *AIANE Investigators. European Network on Aspirin-Induced Asthma. Eur Respir J* 2000;16:432-6.
20. Szczeklik A. Aspirin-induced asthma as a viral disease. *Clin Allergy* 1988;18:15-20.
21. Diaz-Sanchez D, Garcia MP, Wang M, Jyrala M, Saxson A. Nasal challenge with diesel exhaust particles can induce sensitization to a neoallergen in mice. *J Allergy Clin Immunol* 1999;104:1183-8.
22. Finkelman FD, Yang M, Orekhova T, Clyne E, Bernstein L, Whitekus M, et al. Diesel exhaust particles suppress in vivo IFN-gamma production by inhibiting cytokine effects on NK and NKT cells. *J Immunol* 2004;172:3808-13.
23. Heo Y, Saxson A, Hankinson O. Effect of diesel exhaust particles and their components on the allergen-specific IgE and IgG1 response in mice. *Toxicology* 2001;159:143-58.
24. Johnstone RAW, Plimmer J. The chemical constituents of tobacco and tobacco smoke. *Chem Rev* 1959;59:885-936.
25. Wjst M, Heinrich J, Liu P, Dold S, Wassmer G, Merkel G, et al. Indoor factors and serum IgE levels in children. *Allergy* 1994;49:766-71.
26. Stevenson D, Simon RR, Zuraw BL. Sensitivity to aspirin and NSAIDs. In: Adkinson NJ, Yunginger JW, Busse WW, Bochner BS, Holgate ST, Simon FE, editors. *Allergy: principles and practice*. 6th ed. Philadelphia: CV Mosby and Co; 2003. p. 1695-710.
27. Szczeklik A, Gryglewski RJ, Czerniawska-Mysik G. Clinical patterns of hypersensitivity to nonsteroidal anti-inflammatory drugs and their pathogenesis. *J Allergy Clin Immunol* 1977;60:276-84.
28. Vanselow NA, Smith JR. Bronchial asthma induced by indomethacin. *Ann Intern Med* 1967;66:568-73.
29. Szczeklik A, Gryglewski RJ, Czerniawska-Mysik G. Relationship of inhibition of prostaglandin biosynthesis by analgesics to asthma attacks in aspirin-sensitive patients. *BMJ* 1975;1:67-9.
30. Mathison DA, Stevenson DD. Hypersensitivity to non-steroidal anti-inflammatory drugs: indications and methods for oral challenge. *J Allergy Clin Immunol* 1979;64:669-74.

31. Woessner K. Cross-reacting drugs and chemicals. *Clin Rev Allergy Immunol* 2003;24:149-58.
32. Settipane RA, Stevenson DD. Cross sensitivity with acetaminophen in aspirin sensitive asthmatics. *J Allergy Clin Immunol* 1989;84:26-33.
33. Stevenson DD, Hougham A, Schrank P, Goldlust B, Wilson R. Disalicyd cross-sensitivity in aspirin sensitive asthmatics. *J Allergy Clin Immunol* 1990;86:749-58.
34. Asero R. Multiple sensitivities to NSAIDs. *Allergy* 2000;55:893-4.
35. Quaratino D, Romano A, Di Fonso M, Papa G, Perrone MR, D'Anbro-sia FP, et al. Tolerability of meloxicam in patients with histories of adverse reactions to nonsteroidal anti-inflammatory drugs. *Ann Allergy Asthma Immunol* 2000;84:613-7.
36. Bavbek S, Celik G, Ediger D, Mungan D, Demirel YS, Misirligil Z. The use of nimesulide in patients with acetylsalicylic acid and nonsteroidal anti-inflammatory drug intolerance. *J Asthma* 1999;36:657-63.
37. Vaghi A. Tolerance of meloxicam in aspirin-sensitive asthmatics. *Am J Respir Crit Care Med* 1998;157:715.
38. Bavbek S, Celik G, Ozer F, Mungan D, Misirligil Z. Safety of selective COX-2 inhibitors in aspirin/NSAID intolerant patients: comparison of nimesulide, meloxicam and rofecoxib. *J Asthma* 2004;41:67-75.
39. Stevenson DD, Simon RA. Lack of cross-reactivity between rofecoxib and aspirin in aspirin sensitive asthmatic patients. *J Allergy Clin Immunol* 2001;108:47-51.
40. Martin-Garcia C, Hinojosa M, Berges P. Safety of a cyclooxygenase-2 inhibitor in patients with aspirin-sensitive asthma(*). *Chest* 2002;121:1812-7.
41. Szczeklik A, Nizankowska E, Bochenek G, Nagraba K, Mejza F, Swierczyńska M. Safety of a specific COX-2 inhibitor in aspirin-induced asthma. *Clin Exp Allergy* 2001;31:219-25.
42. Yoshida S, Ishizaki Y, Onuma K, Shoji T, Nakagawa H, Amayasu H. Selective cyclo-oxygenase 2 inhibitor in patients with aspirin-induced asthma. *J Allergy Clin Immunol* 2000;106:1201-2.
43. Woessner KM, Simon RA, Stevenson DD. The safety of celecoxib in aspirin exacerbated respiratory disease. *Arthritis Rheum* 2002;46:2201-6.
44. Gyllfors BG, Overholt J, Drupka D, Kumlin M, Sheller J, Nizankowska E, et al. Biochemical and clinical evidence that aspirin-intolerant asthmatic subjects tolerate the cyclooxygenase 2-selective analgetic drug celecoxib. *J Allergy Clin Immunol* 2003;111:1116-21.
45. Vola M, Quaratino D, Volpetti S, Gaeta F, Romano A. Parecoxib tolerability in patients with hypersensitivity to nonsteroidal anti-inflammatory drugs. *J Allergy Clin Immunol* 2006;117:1189-90.
46. Rordorf CM, Choi I, Marshall P, Mangold JP. Clinical and pharmacology of lumiracoxib: a selective cyclo-oxygenase-2 inhibitor. *Clin Pharmacokinet* 2005;44:1247-66.
47. Baldassarre S, Schandene L, Choufani G, Michils A. Asthma attacks induced by low doses of celecoxib, aspirin and acetaminophen. *J Allergy Clin Immunol* 2006;117:215-7.
48. Woessner KW, Simon RA, Stevenson DD. Safety of high-dose rofecoxib in patients with aspirin-exacerbated respiratory disease. *Ann Allergy Asthma Immunol* 2004;93:1-6.
49. Mastalerz L, Sanak M, Gawlewicz A, Gielicz A, Faber J, Szczeklik A. Different eicosanoid profile of the hypersensitivity reactions triggered by aspirin and celecoxib in a patient with sinusitis and asthma. *J Allergy Clin Immunol* 2006;118:957-8.
50. Passero M. Cyclo-oxygenase-2 inhibitors in aspirin sensitive asthma. *Chest* 2003;123:2155-6.
51. Morias-Almeida Marinho S, Rosa S, Rosado-Pinto JE. Multiple drug intolerance, including etoricoxib. *Allergy* 2006;61:144-5.
52. Hawley C. COX-2 inhibitors. *Lancet* 1999;353:307-14.
53. Warner TD, Guiliano F, Vojnovic I, Bukasa A, Mitchell JA, Vane JR. Nonsteroidal drug selectivity for cyclo-oxygenase-1 rather than cyclo-oxygenase-2 are associated with human gastrointestinal toxicity: a full in vitro analysis. *Proc Natl Acad Sci U S A* 1999;96:9666.
54. Stevenson DD. Anaphylactic and anaphylactoid reactions to aspirin and nonsteroidal anti-inflammatory drugs. *Immunol Allergy Clin North Am* 2001;21:745-68.
55. Levy MB, Fink JN. Anaphylaxis to celecoxib. *Ann Allergy Asthma Immunol* 2001;87:72-3.
56. Kaur C, Sakar R, Kanwar AJ. Fixed drug eruption to rofecoxib with cross-reactivity to sulfonamides. *Dermatology* 2001;203:351.
57. Brune K, Hines B. The discovery and development of anti-inflammatory drugs. *Arthritis Rheum* 2004;50:2391-9.
58. Gamboa P, Sanz M, Caballero MR, Urrutia I, Antepará I, Esparza R, et al. The flow-cytometric determination of basophil activation by aspirin and other non-steroidal anti-inflammatory drugs (NSAIDs) is useful for in vitro diagnosis of the NSAID hypersensitivity syndrome. *Clin Exp Allergy* 2004;34:1448-57.
59. Stevenson DD. Oral challenges to detect aspirin and sulfite sensitivity in asthma. *New England Regional Allergy Proceedings* 1988;9:135-42.
60. Dahlen B, Zetterstrom O. Comparison of bronchial and per oral provocation with aspirin in aspirin-sensitive asthmatics. *Eur Respir J* 1990;3:527-34.
61. Melillo G, Podovano A, Cocco G, Masi C. Dosimeter inhalation test with lysine acetylsalicylate for the detection of aspirin-induced asthma. *Ann Allergy* 1993;71:61-5.
62. Nizankowska E, Bestynska-Krypel A, Cmiel A, Szczeklik A. Oral and bronchial provocation tests with aspirin for diagnosis of aspirin-induced asthma. *Eur Respir J* 2000;15:863-9.
63. Alonso-Llamazares A, Martinez-Cocera C, Dominguez-Ortega J, Robledo-Echarren T, Cimarra-Alvarez M, Mesa del Castillo M. Nasal provocation test (NPT) with aspirin: a sensitive and safe method to diagnose aspirin-induced asthma. *Allergy* 2002;57:632-5.
64. Micheletto C, Tognella S, Visconti M, Trevisan F, Dal Negro RW. Changes in urinary LTE4 and nasal function following nasal provocation tests with ASA-tolerant and -intolerant asthmatics. *Respir Med* 2006; May 4 [Epub ahead of print].
65. Milewski M, Mastalerz L, Nizankowska E, Szczeklik A. Nasal provocation test with lysine-aspirin for diagnosis of aspirin-sensitive asthma. *J Allergy Clin Immunol* 1998;101:581-6.
66. Mita H, Endoh S, Kudoh M, Akiyama K. Possible involvement of mast-cell activation in aspirin provocation of aspirin-induced asthma. *Allergy* 2001;56:1061-7.
67. Szczeklik A, Serwonska M. Inhibition of idiosyncratic reactions to aspirin in asthmatic patients by clemastine. *Thorax* 1979;34:654-8.
68. Stevenson DD. Oral challenge, aspirin, NSAID, tartrazine, and sulfites. *N Engl J Regional Allergy Proc* 1984;5:111-20.
69. Berges-Gimeno M, Simon RA, Stevenson DD. The effect of leukotriene modifier drugs on asa-induced asthma and rhinitis reactions. *Clin Exp Allergy* 2002;32:1491-6.
70. White AA, Simon RA. The blocking effect of essential controller medications during aspirin challenges in patients with aspirin exacerbated respiratory disease. *Ann Allergy Asthma Immunol* 2005;95:330-5.
71. Melillo G, Balzano G, Bianco S, Dahlen B, Godard P, Kowalski ML. Report of the INTERASMA Working Group on Standardization of Inhalation Provocation Tests in Aspirin-induced Asthma. Oral and inhalation provocation tests for the diagnosis of aspirin-induced asthma. *Allergy* 2001;56:899-911.
72. Pawlowicz A, Williams WR, Davies BH. Inhalation and nasal challenge in the diagnosis of aspirin induced asthma. *Allergy* 1991;46:405-9.
73. White AA, Stevenson DD. Intranasal ketorolac challenge for the diagnosis of aspirin exacerbated respiratory disease. *Ann Allergy Asthma Immunol*. 2006 In press.
74. Yamashita T, Tsuyi H, Maeda N, Tomoda K, Kumazawa T. Etiology of nasal polyps associated with aspirin-sensitive asthma. *Rhinology* 1989;8:15-24.
75. Sladek K, Dworski R, Soja J, Sheller JR, Nizankowska E, Oates JA, et al. Eicosanoids in bronchoalveolar lavage fluid of aspirin-intolerant patients with asthma after aspirin challenge. *Am J Respir Crit Care Med* 1994;149:940-6.
76. Bachert C, Wagenmann M, Hauser U, Rudack C. IL-5 synthesis is upregulated in human nasal polyp tissue. *J Allergy Clin Immunol* 1997;99:837-42.
77. Bachert C, Wagenmann M, Rudack C, Hopken K, Hillebrandt M, Wang D, et al. The role of cytokines in infectious sinusitis and nasal polyposis. *Allergy* 1998;53:2-13.
78. Hamilos D, Leung DYM, Wood R, Cunningham L, Bean DK, Yasruel Z. Evidence for distinct cytokine expression in allergic versus non-allergic chronic sinusitis. *J Allergy Clin Immunol* 1995;96:537-44.
79. Sousa AR, Lams BE, Pfister R, Christie PE, Schmitz M, Lee TH. Expression of interleukin-5 and granulocyte-macrophage colony-stimulating factor in aspirin-sensitive and non-aspirin-sensitive asthmatic airways. *Am J Respir Crit Care Med* 1997;156:1384-9.

80. Elsner J, Hochstetter R, Kimming D, Kapp A. Human eotaxin represents a potent activator of the respiratory burst in human eosinophils. *Eur J Immunol* 1996;26:1919-25.
81. Kanaoka Y, Boyce J. Cysteinyl leukotrienes and their receptors: cellular distribution and function in immune and inflammatory responses. *J Immunol* 2004;173:1503-10.
82. Weller CL, Collington S, Brown KW, Miller HR, Al-Kashi A, Clark P, et al. Leukotriene B₄, activation product of mast cells, is a chemoattractant for their progenitors. *J Exp Med* 2005;201:1961-71.
83. Cowburn AS, Sladek K, Soja J, Adamek L, Nizankowska E, Szczeklik A, et al. Over expression of leukotriene C₄ synthase in bronchial biopsies from patients with aspirin-intolerant asthma. *J Clin Invest* 1998;101:834-46.
84. Zirolfi NE, Na H, Chow JM, Stankiewicz JA, Samter M, Young MR. Aspirin-sensitive versus non-aspirin-sensitive nasal polyp patients: analysis of leukotrienes/Fas and Fas-ligand expression. *Otolaryngol Head Neck Surg* 2002;126:141-6.
85. Bachert C, Gevaert P, Van Cauwenberge P. Nasal polyposis—a new concept on the formation of polyps. *ACI Int* 1999;11:130-5.
86. Daffern P, Muilenburg D, Hugli TE, Stevenson DD. Association of urinary leukotriene E₄ excretion during aspirin challenges with severity of respiratory responses. *J Allergy Clin Immunol* 1999;104:559-64.
87. Szczeklik A, Sladek K, Dworski R, Nizankowska E, Soja J, Oates J. Bronchial aspirin challenge causes specific eicosanoid response in aspirin-sensitive asthmatics. *Am J Respir Crit Care Med* 1996;154:1608-14.
88. Christie PE, Tagari P, Ford-Hutchinson AW, Charlesson S, Chee P, Arm JP. Urinary leukotriene E₄ concentrations increase after aspirin challenge in aspirin-sensitive asthmatic subjects. *Am Rev Respir Dis* 1991;143:1025-9.
89. Smith CM, Hawksworth RJ, Thien FC, Christie PE, Lee TH. Urinary leukotriene E₄ in bronchial asthma. *Eur Respir J* 1992;5:693-9.
90. Sanak M, Pierzchalska M, Bazan-Socha S, Szczeklik A. Enhanced expression of the leukotriene C₄ synthase due to overactive transcription of an allelic variant associated with aspirin-intolerant asthma. *Am J Respir Cell Mol Biol* 2000;23:290-6.
91. Sanak M, Simon HU, Szczeklik A. Leukotriene C₄ synthase promoter polymorphism and risk of aspirin-induced asthma. *Lancet* 1997;350:1599-600.
92. Van Sambeek R, Stevenson DD, Baldasaro M, Lam BK, Zhao J, Yoshida S, et al. 5' flanking region polymorphism of the gene encoding leukotriene C₄ synthase does not correlate with the aspirin-intolerant asthma phenotype in the United States. *J Allergy Clin Immunol* 2000;106:72-6.
93. Sanak M, Szczeklik A. Leukotriene C₄ synthase polymorphism and aspirin-induced asthma. *J Allergy Clin Immunol* 2001;107:561-2.
94. Sousa A, Parikh A, Scadding G, Corrigan CJ, Lee TH. Leukotriene-receptor expression on nasal mucosal inflammatory cells in aspirin-sensitive rhinosinusitis. *N Engl J Med* 2002;347:1524-6.
95. Powell WS, Rokach J. Biochemistry, biology, chemistry of the 5-lipoxygenase product 5-oxo-E₄. *Prog Lipid Res* 2005;44:154-83.
96. Sanak M, Levy BD, Clish CB, Chiang N, Gronert K, Mastalerz L, et al. Aspirin-tolerant asthmatics generate more lipoxins than aspirin-intolerant asthmatics. *Eur Respir J* 2000;16:44-9.
97. Nasser MS. Products of 15-LO: Are they important in asthma? *Clin Exp Allergy* 2002;32:1540-2.
98. Levy BD, DeSanctis GT, Devhand PR, Kim E, Ackerman K, Schmidt BA, et al. Multi-pronged inhibition of airway hyper-responsiveness and inflammation by lipoxin A₄. *Nat Med* 2002;8:1018-23.
99. Perez-Novo CA, Watelet JB, Claeys C, Van Cauwenberge P, Bachert C. Prostaglandin, leukotriene and lipoxin balance in chronic rhinosinusitis with and without nasal polyps. *J Allergy Clin Immunol* 2005;115:1189-96.
100. Kowalski ML, Bienkiewicz B, Pawliczak R, DuBuske L. Differential effects of aspirin and misoprostol on 15-hydroxyeicosatetraenoic acid generated by leukocytes from aspirin sensitive asthmatic patients. *J Allergy Clin Immunol* 2003;112:505-12.
101. Bochenek G, Nagraba K, Nizankowska E, Szczeklik A. A controlled study of 9αphal1βeta-PGF₂ (a PGD₂ metabolite) in plasma and urine of patients with bronchial asthma and healthy controls after aspirin challenges. *J Allergy Clin Immunol* 2003;111:743-9.
102. Monneret G, Cossett C, Gravel S, Rokach J, Powell WS. 15R-methyl-prostaglandin D₂ is a potent and selective CRTH/DP₂ receptor agonist in human eosinophils. *J Pharmacol Exp Ther* 2003;304:349-55.
103. Powell WS. A novel PGD₂ receptor expressed in eosinophils. *Prostaglandins Leukot Essent Fatty Acids* 2003;69:179-85.
104. Szczeklik A. Prostaglandin E₂ and aspirin-induced asthma [letter]. *Lancet* 1995;345:1056.
105. Schaefer D, Gode UC, Baenkler HW. Dynamics of eicosanoids in peripheral blood cells during bronchial provocation in aspirin-intolerant asthmatics. *Eur Respir J* 1999;13:638-46.
106. Jinnai N, Sakagami T, Sekigawa T, Kakiyama M, Nakjima T, Yoshia K, et al. Polymorphisms in the prostaglandin E₂ receptor subtype 2 gene confer susceptibility to aspirin-intolerant asthma: a candidate gene approach. *Hum Mol Genet* 2004;15:3203-17.
107. Ying S, Meng Q, Scadding G, Parikh A, Corrigan CJ, Lee TH. Aspirin-sensitive rhinosinusitis is associated with reduced E-prostanoid 2 receptor expression. *J Allergy Clin Immunol* 2006;117:312-8.
108. Kunikata T, Yamane H, Segi E, Matsuoka T, Sugimoto Y, Tanaka S, et al. Suppression of allergic inflammation by the prostaglandin E receptor subtype EP₃. *Nat Immunol* 2005;6:524-31.
109. Szczeklik A, Sanak M. The broken balance in aspirin hypersensitivity. *Eur J Pharmacol* 2006;533:145-55.
110. Choi JH, Park H, Oh HB, Lee JH, Suh YJ, Park CS, et al. Polymorphism of tandem repeat in promoter of 5-lipoxygenase in ASA-intolerant asthma: a positive association with airway hyper responsiveness. *Hum Genet* 2004;114:337-44.
111. Choi JH, Lee KW, Oh HB, Lee KJ, Suh YJ, Park CS, et al. HLA association in aspirin intolerant asthma: DPB1*0301 as a strong marker in a Korean population. *J Allergy Clin Immunol* 2004;113:562-4.
112. Dekker JW, Nizankowska E, Schmitz-Schumann M, Pile K, Bochenek G, Dyczek A, et al. Aspirin-induced asthma and HLA-DRB1 and HLA-DPB1 genotypes. *Clin Exp Allergy* 1997;27:574-7.
113. Kedda MA, Worsley P, Shi J, Phelps S, Duffy D, Thompson PJ. Polymorphisms in the 5-lipoxygenase activating protein (ALOX5AP) gene are not associated with asthma in an Australian population. *Clin Exp Allergy* 2005;35:332-8.
114. Kim SH, Bae J, Suh CH, Nahm DH, Holloway JW, Park HS. Polymorphism of tandem repeat in promoter of 5-lipoxygenase in ASA-intolerant asthma: a positive association with airway hyperresponsiveness. *Allergy* 2005;60:760-5.
115. Szczeklik A. Adverse reactions to aspirin and nonsteroidal anti-inflammatory drugs. *Ann Allergy* 1987;59:113-8.
116. Kumlin M, Dahlen B, Bjorck T, Zetterstrom O, Granstrom E, Dahlen SE. Urinary excretion of leukotriene E₄ and 11-dehydro-thromboxane B₂ in response to bronchial provocations with allergen, aspirin, leukotriene D₄, and histamine in asthmatics. *Am Rev Respir Dis* 1992;146:96-103.
117. Ferreri NR, Howland WC, Stevenson DD, Spiegelberg HL. Release of leukotrienes, prostaglandins, and histamine into nasal secretions of aspirin-sensitive asthmatics during reaction to aspirin. *Am Rev Respir Dis* 1988;137:847-54.
118. Fischer AR, Rosenberg MA, Lilly CM, Callery JL, Rubin P, Cohn P. Direct evidence for a role of the mast cell in the nasal response to aspirin in aspirin-sensitive asthma. *J Allergy Clin Immunol* 1994;94:1046-56.
119. Kowalski ML, Sliwiska-Kowalska M, Igarashi Y, White MV, Wojciechowska B, Brayron P, et al. Nasal secretions in response to acetylsalicylic acid. *J Allergy Clin Immunol* 1993;91:580-98.
120. Szczeklik A, Stevenson DD. Aspirin-induced asthma: advances in pathogenesis, diagnosis and management. *J Allergy Clin Immunol* 2003;111:913-21.
121. Sestini P, Armetti L, Gambaro G, Peroni MG, Refini RM, Sala A, et al. Inhaled PGE₂ prevents aspirin-induced bronchoconstriction and urinary LTE₄ excretion in aspirin-sensitive asthma. *Am J Respir Crit Care Med* 1996;153:572-5.
122. Szczeklik A, Mastalerz L, Nizankowska E, Cmiel A. Protective and bronchodilator effects of prostaglandin E and salbutamol in aspirin-induced asthma. *Am J Respir Crit Care Med* 1996;153:567-71.
123. Picado C, Fernandez-Morata JC, Juan M, Roca-Ferrer J, Fuentes M, Xaubert A, et al. Cyclooxygenase-2 mRNA is down expressed in nasal polyps from aspirin-sensitive asthmatics. *Am J Respir Crit Care Med* 1999;160:291-6.
124. Pujols L, Mullol J, Alobid I, Roca-Ferrer J, Xaubert A, Picado C. Dynamics of COX-2 in nasal mucosa and nasal polyps from aspirin-tolerant and aspirin-intolerant patients with asthma. *J Allergy Clin Immunol* 2004;114:814-9.

125. Arm JP, O'Hickey SP, Spur BW, Lee TH. Airway responsiveness to histamine and leukotriene E4 in subjects with aspirin-induced asthma. *Am Rev Respir Dis* 1989;140:148-53.
126. Kowalski M, Grzegorzcy J, Wojciechowska B, Sponiatowska M. Intranasal challenge with aspirin induces cell influx and activation of eosinophils and mast cells in nasal secretions of ASA-sensitive patients. *Clin Exp Allergy* 1996;26:807-14.
127. Bosso JV, Schwartz LB, Stevenson DD. Tryptase and histamine release during aspirin-induced respiratory reactions. *J Allergy Clin Immunol* 1991;88:830-7.
128. Juergens UR, Christiansen SC, Stevenson DD, Zuraw BL. Inhibition of monocyte leukotriene B4 production following aspirin desensitization. *J Allergy Clin Immunol* 1995;96:148-56.
129. Mastalerz L, Milewski M, Duplaga M, Nizankowska E, Szczeklik A. Intranasal fluticasone propionate for chronic eosinophilic rhinitis in patients with aspirin-induced asthma. *Allergy* 1997;52:895-900.
130. Mathison DA, Stevenson DD. Aspirin sensitivity in rhinosinusitis and asthma. *Immunol Allergy Pract* 1983;5:340-50.
131. Dahlen SE, Nizankowska E, Dahlen B, Szczeklik A. The Swedish-Polish treatment study with the 5-lipoxygenase inhibitor Zileuton in aspirin-intolerant asthmatics. *Am J Respir Crit Care Med* 1995;151:376.
132. Dahlen B, Nizankowska E, Szczeklik A, Zetterstrom O, Bochenek G, Kumlin M, et al. Benefits from adding the 5-lipoxygenase inhibitor zileuton to conventional therapy in aspirin-intolerant asthmatics. *Am J Respir Crit Care Med* 1998;157:1187-94.
133. Dahlen S, Malstrom K, Nizankowska E, Dahlen B, Kuna P, Kowalski M, et al. Improvement of aspirin-intolerant asthma by montelukast, a leukotriene antagonist: A randomized, double blind, placebo controlled trial. *Am J Respir Crit Care Med* 2002;165:9-14.
134. Mastalerz L, Nizankowska E, Sanak M, Mejza F, Pierzchalska M, Bazan-Socha S, et al. Clinical and genetic features underlying the response of patients with bronchial asthma to treatment with a leukotriene receptor antagonist. *Eur J Clin Invest* 2002;32:949-55.
135. Mastalerz L, Nizankowska E, Ćmiel A, Szczeklik A. Protection against exercise-induced bronchoconstriction by montelukast in aspirin-sensitive and aspirin-tolerant patients with asthma. *Clin Exp Allergy* 2002;32:1360-5.
136. Sampson AP, Siddiqui S, Buchanan D, Howarth PH, Holgate ST, Holloway JW, et al. Variant LTC(4) synthase allele modifies cysteinyl leukotriene synthesis in eosinophils and predicts clinical response to zafirlukast. *Thorax* 2000;55(suppl):S28-31.
137. Asano K, Shiomi T, Hasegawa N, Nakamura H, Kudo H, Matsuzaki T, et al. Leukotriene C4 synthase gene A(-444)C polymorphism and clinical response to an LT(1) antagonist, pranlukast, in Japanese patients with moderate asthma. *Pharmacogenetics* 2002;12:565-70.
138. Park HE, Kim SH, Sampson A, Lee KW, Park CS. The HLA-DPB1*0301 marker might predict the requirement for leukotriene receptor antagonist in patients with aspirin-intolerant asthma. *J Allergy Clin Immunol* 2004;114:688-9.
139. McFadden EA, Woodson BT, Fink JN, Toohill RJ. Surgical treatment of aspirin triad sinusitis. *Am J Rhinol* 1997;11:263-70.
140. Kennedy DW. Prognostic factors, outcomes and staging in ethmoid sinus surgery. *Laryngoscope* 1992;102:1-18.
141. Amar YG, Frenkiel S, Sobol SE. Outcome analysis of endoscopic sinus surgery for chronic sinusitis in patients having Samter's triad. *J Otolaryngol* 2000;29:7-12.
142. Sweet JA, Stevenson DD, Simon RA, Mathison DA. Long term effects of aspirin desensitization treatment for aspirin sensitive rhinosinusitis asthma. *J Allergy Clin Immunol* 1990;86:59-65.
143. Pleskow WW, Stevenson DD, Mathison DA, Simon RA, Schatz M, Zieger RS. Aspirin desensitization in aspirin sensitive asthmatic patients: Clinical manifestations and characterization of the refractory period. *J Allergy Clin Immunol* 1982;69:11-9.
144. Berges-Gimeno M, Simon RA, Stevenson DD. Early effects of aspirin desensitization treatment in asthmatics with aspirin exacerbated respiratory disease. *Ann Allergy Asthma Immunol* 2003;90:1-4.
145. Berges-Gimeno M, Simon RA, Stevenson DD. Treatment with aspirin desensitization in patients with aspirin exacerbated respiratory disease. *J Allergy Clin Immunol* 2003;111:180-6.
146. Stevenson DD, Hankammer MA, Mathison DA, Christensen SC, Simon RA. Long term ASA desensitization-treatment of aspirin sensitive asthmatic patients: clinical outcome studies. *J Allergy Clin Immunol* 1996;98:751-8.
147. Stevenson DD. Aspirin desensitization in patients with AERD. *Clin Rev Allergy Immunol* 2003;24:159-67.