

MDPI

Review

Clinical Epigenomic Explanation of the Epidemiology of Cannabinoid Genotoxicity Manifesting as Transgenerational Teratogenesis, Cancerogenesis and Aging Acceleration

Albert Stuart Reece 1,2,* and Gary Kenneth Hulse 1,2

- Division of Psychiatry, University of Western Australia, Crawley, WA 6009, Australia
- ² School of Medical and Health Sciences, Edith Cowan University, Joondalup, WA 6027, Australia
- * Correspondence: stuart.reece@uwa.edu.au

Abstract: As global interest in the therapeutic potential of cannabis and its' derivatives for the management of selected diseases increases, it is increasingly imperative that the toxic profile of cannabinoids be thoroughly understood in order to correctly assess the balance between the therapeutic risks and benefits. Modern studies across a number of jurisdictions, including Canada, Australia, the US and Europe have confirmed that some of the most worrying and severe historical reports of both congenital anomalies and cancer induction following cannabis exposure actually underestimate the multisystem thousand megabase-scale transgenerational genetic damage. These findings from teratogenic and carcinogenic literature are supported by recent data showing the accelerated patterns of chronic disease and the advanced DNA methylation epigenomic clock age in cannabis exposed patients. Together, the increased multisystem carcinogenesis, teratogenesis and accelerated aging point strongly to cannabinoid-related genotoxicity being much more clinically significant than it is widely supposed and, thus, of very considerable public health and multigenerational impact. Recently reported longitudinal epigenome-wide association studies elegantly explain many of these observed effects with considerable methodological sophistication, including multiple pathways for the inhibition of the normal chromosomal segregation and DNA repair, the inhibition of the basic epigenetic machinery for DNA methylation and the demethylation and telomerase acceleration of the epigenomic promoter hypermethylation characterizing aging. For cancer, 810 hits were also noted. The types of malignancy which were observed have all been documented epidemiologically. Detailed epigenomic explications of the brain, heart, face, uronephrological, gastrointestinal and limb development were provided, which amply explained the observed teratological patterns, including the inhibition of the key morphogenic gradients. Hence, these major epigenomic insights constituted a powerful new series of arguments which advanced both our understanding of the downstream sequalae of multisystem multigenerational cannabinoid genotoxicity and also, since mechanisms are key to the causal argument, inveighed strongly in favor of the causal nature of the relationship. In this introductory conceptual overview, we present the various aspects of this novel synthetic paradigmatic framework. Such concepts suggest and, indeed, indicate numerous fields for further investigation and basic science research to advance the exploration of many important issues in biology, clinical medicine and population health. Given this, it is imperative we correctly appraise the risk-benefit ratio for each potential cannabis application, considering the potency, severity of disease, stage of human development and duration of use.

Keywords: tobacco; alcohol; cannabis; cannabinoid; cancer; cancerogenesis; mutagenesis; oncogenesis; genotoxicity; epigenotoxicity; transgenerational inheritance



Citation: Reece, A.S.; Hulse, G.K. Clinical Epigenomic Explanation of the Epidemiology of Cannabinoid Genotoxicity Manifesting as Transgenerational Teratogenesis, Cancerogenesis and Aging Acceleration. *Int. J. Environ. Res. Public Health* 2023, 20, 3360. https://doi.org/10.3390/ijerph20043360

Academic Editor: Sarabjit Mastana

Received: 10 December 2022 Revised: 7 February 2023 Accepted: 13 February 2023 Published: 14 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Cannabinoid genotoxicity is not controversial and is widely acknowledged by both government drug regulators and the cannabis industry. The official prescribing information for both Sativex ($\Delta 9$ -tetrahydrocannabinol (THC)—cannabidiol) and Epidiolex

(cannabidiol) registered with the US Food and Drug Administration (FDA), the European Medicines Agency (EMA) and the Medicines and Health Care Products Regulatory Agency (MHCPRA) of the UK [1,2] carry warnings against the use of these products in pregnancy and lactation. Further, some products released directly by the cannabis industry, and thus not formally regulated, contain similar reproductive warnings.

However, while cannabinoid genotoxicity has been well established experimentally in a variety of different in vitro systems [3–19], this information is strikingly absent from most public health discussions on the place of cannabinoids and the management of morbidity. The public health implications of the documented cannabinoid genotoxicity have more recently begun to be appreciated with large, newly published epidemiological investigations of cannabinoid-related cancerogenesis, teratogenesis, heritable teratogenic cancerogenesis and the acceleration of aging.

With rising interest in the pharmaceutical use of cannabis for the management of selected diseases and a strong commercial and popular interest in the therapeutic potential of cannabis derivatives, it becomes increasingly important that the toxic profile of cannabinoids is thoroughly understood in order to parse the classical balance between the therapeutic risks and benefits correctly and astutely. Inherently, this risk-benefit ratio must be correctly appraised for each potential cannabis application, considering the potency, severity of disease, stage of human development and duration of use.

The correct and astute appraisal of cannabinoid toxicity becomes even more important in the case of cannabinoids where, at the time of writing, reports from both the Institute of Medicine and the National Academy of Sciences described the objective evidentiary basis underpinning cannabis prescription as often "weak" to "moderate at best", and most of its hypothetical indications as being of anecdotal level only, and so of weak evidentiary power [20,21].

Contrariwise, strong epidemiological evidence from many jurisdictions showed the likely involvement of cannabinoids in many cancers, dozens of serious birth defects and cellular and organismal aging [22–54]. Moreover, there is a remarkably close uniformity between the findings from different jurisdictions [38,39], which not only confirms the findings of single studies but also, according to the established Hill criteria of causality, lends formal credence to the likely causal relationship between epidemiologically assessed cannabis exposure and the observed morbidity.

While it is acknowledged that some historical literature assessing the cannabis morbidity associations produced data showing no effect, the field is plagued with numerous methodological difficulties, including the conduct of many widely quoted studies in earlier eras when cannabis was of a much lower THC potency and the increase in the community exposure since that time, in terms of not only the prevalence of cannabis consumption but the intensity of cannabis consumption. In some widely quoted studies [55], the systematic deletion of individuals who had experienced a high dose exposure was practiced. Clearly, it is in patients with this higher exposure level where more significant effects might reasonably be expected. The issue of small sample sizes is also a major and common shortcoming of many studies. It should also be noted that relatively sudden increases in cannabis use prevalence, THC potency and cannabis daily use intensity all occurring at once will launch the community relatively abruptly into the higher exposure levels where, due to the exponential dose-response relationship for many genotoxic [3,5–8,10,11,14,16,18,19] and mitochondrial metabolic [4,9,12,13,15,17] consequences, the negative outcomes become both more common and more severe.

Importantly, in attaining a cannabis dose-response threshold, there has been an increase in the cannabis exposure for some communities, arising not only from an increased prevalence in cannabis consumption but from an increased potency and frequency of use [56,57], where the well-documented exponential genotoxic dose-response effect curves shown in many laboratory [3,5–8,10,11,14,16,18,19] and preclinical animal model assays [58–60] take effect. Since the cannabis use prevalence, intensity of daily use and THC potency are all rising *simultaneously*, these features can be expected to operate syn-

ergistically as a relatively abrupt switch where severe adverse genotoxic and neurotoxic outcomes relatively suddenly become commonplace due to the underlying exponential dose-response effects [56,57].

It is well established that the trend of cannabinoid exposure in both the US and Europe has been upwards and positive (increasing) in recent decades. This rising trend carries two major implications. Firstly, in the US, it has been shown that the prevalence of cannabis use has increased, the rate of daily cannabis use has more than doubled [61,62] and the THC content of cannabis has increased from 3.4% in 1993 to 17.1% in 2017 [63–66]. Similar changes are reported in Europe [56,57]. Moreover, it has also been shown that the combination of these three metrics (of the prevalence, intensity and THC concentration) is the most powerful predictor of the teratogenic genotoxic outcomes [40]. Were these three changes occurring in isolation, they would be cause for concern in themselves. However, their occurrence in many places *simultaneously* implies a major paradigmatic shift in the community exposure from a relatively low level to a much higher region where genotoxic outcomes become more common.

Secondly, it would be expected that the rising trend in cannabinoid exposure shows a simple bivariate statistical correlation with any rising trend in the population with ill health. Accordingly, recent research has established a robust relationship with multisystem tumorigenesis and teratogenesis, which is robust to multivariable adjustment, is verified in combined formal spatiotemporal analyses and satisfies the modern quantitative criteria of causality with very similar results reflected in both the US and European datasets [22–31,36,37,40,41,44–46,67].

It must be underscored that the triple convergence of cannabinoid carcinogenesis, cannabinoid teratogenesis and the cannabinoid acceleration of aging together forms strong and theoretically robust evidence for a clinically and highly significant genotoxicity [68] severe enough to impact numerous metrics of the population health adversely.

Furthermore, both in vitro and clinical studies implicate many different cannabinoid moieties, suggesting that genotoxicity is a class effect shared by many cannabinoids [69,70]—a feature now well confirmed by many epidemiological studies. This includes such allegedly benign cannabinoid species as $\Delta 9 THC$, $\Delta 8 THC$ and cannabidiol, among several others [22–24,28,71,72].

For the reader who is unfamiliar with this epidemiological literature, it should be pointed out that most of the modern epidemiological studies referred to are not just observational ecological studies of convenience which happen to show a particular association. Many of the best studies used a formal space—time analysis and the quantitative tools of causal inference to introduce a pseudo-randomized quasi-experimental paradigm from which it is entirely appropriate to invoke causal associations [28,36,37,40,45,51,71,73–82].

It is the purpose of the present paper to explore the manner in which the diverse recent laboratory and clinical results, which are the outcomes of the three primary expressions of cannabinoid genotoxicity in cancerogenesis, teratogenesis and aging, are explained using cannabinoid epigenotoxicity and the mechanistic insights which follow an improved understanding and appreciation of the magnitude and breadth of the scope of the epigenotoxic profile of cannabinoids. Since this is only beginning to be explored, the present review is necessarily limited to an introductory oversight, but it does lead to the formulation of a broad ranging experimental investigation into the ways in which these insights can be developed and explored further with far reaching implications across the spectrum of clinical medicine and the basic biological sciences.

2. Cannabinoid Genotoxic Phenomenology

2.1. Cannabinoid Genotoxic Carcinogenesis

There is impressive overlap between the US and European data for cannabis exposure and tumors. A recent review of 28 US cancer tumors that were significantly associated with $\Delta 9$ THC included acute myeloid leukemia, breast, oropharynx, thyroid, liver, pancreas, chronic myeloid leukemia, testis and kidney [22]. The cancers which were significantly as-

sociated with cannabidiol were prostate, bladder, ovary, all cancers, colorectum, Hodgkin's, brain, non-Hodgkin's lymphoma, esophagus, breast and stomach [22–24]. Eight cancers significantly associated with $\Delta 8 THC$ on bivariate testing included corpus uteri, liver, gastric cardia, breast and post-menopausal breast, anorectum, pancreas and thyroid [72]. An additional 18 tumors demonstrated positive marginal effects after the multivariable adjustment, including stomach, Hodgkin's and non-Hodgkin's lymphomas, ovary, cervix uteri, gall bladder, oropharynx, bladder, lung, esophagus, colorectal cancer and all cancers (excluding non-melanoma skin cancer) [72].

Similarly, in a review of 40 European cancers, 27 tumors were related to various metrics of cannabis exposure, including daily use. The tumor overlap included all cancers (excluding non-melanoma skin cancer), oropharynx, the four major leukemias and Hodgkin's and non-Hodgkin's lymphoma, liver, pancreas, brain medulloblastoma, anus, kidney, thyroid, testis (seminoma and non-seminoma), ovary and ovarian germ cell tumors. They also identified hepatocellular, skin melanoma, mesothelioma, Kaposi sarcoma, penis, prostate, vulva and vaginal cancers [83].

Hence, there is impressive overlap between the US and European data for those tumors which are listed as common.

It is mechanistically noteworthy that, for several of the tumors aforementioned, chromosomal translocation is an important and well-established pathway in their oncogenesis. The presumed mode of action here is to constitutively activate the proto-oncogenes or suppress the tumor suppressor genes. These comments apply particularly to acute myeloid and lymphoid leukemias (AML and ALL) and to testicular cancer [25,84–87]. Indeed, if one adds up all the chromosomes commonly implicated in ALL and testicular cancer, they result in 1254 megabases and 645 megabases, which represent 41.8% and 21.5% of the 3000 megabases of the human genome, respectively.

2.2. Cannabinoid Genotoxic Teratogenesis

When 62 congenital anomalies were tracked longitudinally across the US, 45 were shown to be related to the metrics of cannabis exposure, particularly those from the cardiovascular, chromosomal, gastrointestinal, limb, urinary, body wall and face [28].

When a series of 95 European congenital anomalies were studied, 89 were shown to be relatable to the various metrics of cannabis exposure and were related particularly to the anomalies affecting the cardiovascular, gastrointestinal, uronephrological and central nervous systems, as well as the chromosomal, limb, body wall, face and general (unallocated) [40].

Again, a significant overlap was shown where the anomalies listed were in common [36]. It is of interest that if one adds the lengths of the chromosomes directly impacted by these chromosomal anomalies together, it results in 388 of the 3000 megabases of the human genome, or 12.9%.

2.3. Cannabinoid Genotoxic Aging Acceleration

The data from the twelve separate empirical streams both independently and more strongly collectively provide a convincing empirical case for cannabis-induced accelerated aging. These include hepatotoxic [88], immunological [89–99], genotoxic [3,5–8,10,11,14,16,18,19], epigenotoxic [42,100–108], disruption of chromosomal physiology [109–115], endocrine [116–120], congenital anomalies [28,36,40,45,46,51] and cancers [22–24,26,30,34,41,51], including inheritable tumorigenesis [25,26], telomerase inhibition [42,121], mitochondriopathic [122–126], cardiovascular [33] and elevated mortality [127–138]. Cannabis dependence not only recapitulates many of the key features of aging, but is characterized by both age-defining and age-generating illnesses, including hepatoinflammatory disorders, immunomodulation, many psychiatric syndromes with a neuroinflammatory basis [139–161], genotoxicity [3,5–8,10,11,14,16,18,19] and epigenotoxicity [162–166].

A recent detailed report from a large electronic health record database in Hawaii showed that cannabis users were subject to elevated rates of an impressive array of acute

illnesses, including myocardial infarction, stroke, acute bronchitis, cyclic vomiting, injuries, poisonings, car wrecks, falls and several chronic diseases, including coronary artery disease, hypertension, chronic obstructive pulmonary disease, chronic pain, behavioral health disorders, addictions and poverty [53]. Overall, this pattern of chronic ill health includes many age-defining illnesses and provides solid clinical evidence of accelerated aging [163–166].

This important study was followed by another pivotal study, which demonstrated a dramatic 30% increase in cellular aging at a median chronological age of 30 years using a late-generation epigenomic clock based on DNA methylation, demonstrating the acceleration of the aging process from cannabinoid exposure in somatic (non-germ cell) tissues utilizing 'state-of-the-art' technologies [54].

However, arguably of greater concern, is that fact that the nuclear blebs and bridges and chromosomal breaks and translocations well-described for sperm and oocytes in the experimental cannabis literature [114,167] are signs of advanced cellular aging [162]. This leads to the conclusion that the fertilized zygote must also be prematurely aged from the time of conception since both component parts are also aged. Given that the explosion of evidence from epigenomics has now placed the Barker hypothesis of the prenatal origins of adult disease [168–171] on a firm evidentiary basis, such a finding of cannabis-induced advanced aging carries far reaching and grave public health and transgenerational implications.

3. Mechanisms of Cannabinoid Genotoxicity

Central to assigning a potentially causal relationship between the exposure and observed epidemiological effects was the presence of the plausible biological mechanisms of action [172]. The past published research is awash with a flotilla of biological mechanisms by which the cannabinoids effects in the reproductive tracts [173,174], including at the level of both the male and female gametes [173–175], chromosomal breaks and translocations [11,109,110,115], the nucleotide bases of DNA [11], single and double stranded DNA breaks [11], the mitochondrial metabolic machinery which forms the (small molecule co-substrates, energetic and intracellular intraorganellar signaling) basis of epigenetic regulation [122–126,176–180] and the epigenomic machinery itself [42], have all been implicated in prior studies.

While these many different modalities were explored in a number of previous publications and reviews [22–36,42,43,49,101,102], the extraordinary detail, predictive power and keen mechanistic insights provided by recent fine-level studies of the epigenetic changes to human sperm DNA methylation following cannabis exposure and withdrawal [42] provided a truly extraordinary insight not only into the various tumors, congenital anomalies and aging-related changes reported following cannabis exposure but also their impressive breadth, diversity and variety. Importantly, these methylation changes to the human sperm epigenome showed an intricate and intimate concordance with the morbidity identified in the large-scale community cannabis exposure epidemiological studies. Paradoxically, this important feature was not readily cited or understood. For these reasons, it was of considerable importance to explore these epigenomic findings in light of the most modern and penetrating epidemiological studies.

3.1. Fundamental Primacy of the Epigenomic Effects

3.1.1. Layers of Epigenomic Regulation

Many layers of epigenomic regulation are described and the list appears to be rapidly increasing. While they may be listed individually, they are not independent and are coordinated across the various layers. The key parameters include DNA methylation, the histone post-translational modifications, various short and long non-protein coding RNAs, over 100 post-transcriptional modifications to the RNAs, including m6-adenosine RNA methylation (also referred to as epitranscriptomics), the position with respect to the nuclear lamina (which is suppressive of the gene transcription), the chromatin state (euchromatin or suppressive heterochromatin), the presence within the transcriptional factories of the

topologically defined domains (and their controlling boundary elements) and the presence of the tethering elements (especially important during the development) [181].

3.1.2. Epigenomic Functions

It is now well understood that the cell lineage specification (that is, whether a cell develops as a muscle cell or a neuron, etc.) is controlled epigenomically. This issue was first formalized in the epigenetic valley hypothesis of Conrad Waddington [182]. It is also well established that the state of the cell differentiation from a pluripotent embryonic cell to a mature fully differentiated cell is also controlled epigenomically.

For a long time, the mechanisms of aging were not understood and many competing and often complimentary and overlapping theories were advanced [162]. However recent studies have confirmed that, while there are many different pathways to induce age-related damage, the major controller of cellular age is actually the epigenomic state of the cell on which other pathways likely converge [183–185]. Thus, robust evidence of the reversal of epigenomic clock aging, biological age and the youthful/neonatal functional capacity has now been convincingly demonstrated in many systems, including optic nerve crush injury, congenital glaucoma and ocular aging, progeroid mouse models, cardiac and skeletal muscle and fibroblasts [184,186–188]. This view is concordant with the well-established control of the state of the cell differentiation by the epigenomic machinery.

This implies that the epigenomic state is central and pivotal to the control of cancer, cell development and aging, which are the three principal themes of the present discussion.

3.2. Epigenomic Impacts of Cannabis Exposure and Withdrawal

A recent detailed epigenome-wide association study (EWAS) by Schrott and colleges investigating the DNA methylation changes of human and mouse sperm both in cannabis dependence and withdrawal provides a 359-page Supplementary Appendix listing the detailed methylation changes [42]. These researchers looked at the differential DNA methylation of the cannabis-dependent humans and mice compared to the cannabis free controls and again after an 11-week period of washout following a documented period of abstinence and detoxification from the cannabis. This longitudinal design is a very powerful way to design an epigenomic study. Close study of this dataset revealed the following remarkable findings.

3.2.1. Disruption of the Epigenetic Machinery

There was widespread disruption between the main readers, writers and erasers of the epigenetic code. Hence, there were five hits for the DNA methyltransferases which added the methylation mark to the CpG islands and one hit for TET1 (ten-eleven translocase) which began the process of removing it. There was one hit for telomerase which controlled the end length of the chromosomes, and thus protected them against aging, three hits for polycomb repressors, five hits for the chromatin remodelers (SMARCA's, SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin, subfamily A) and three hits for the UHRF (Ubiquitin-like with PHD and ring finger domains) family which controlled both DNA methylation and histone methylation and integrated the signaling in the two classes of pathways.

There were 161 hits for the histone methyltransferases with methylate histones and 199 hits for the histone demethylases that remove this mark. There were eleven hits for both the histone acetyltransferases, which acetylate histone tailed and thereby made the genome accessible to the transcription machinery, and eleven hits for the deacetylases which removed this mark.

3.2.2. Stem Cell Renewal Factors

Considering the key stem cell induction factors identified by Yamanaka, Oct3/4, Sox2, Klf4 and Myc [186], all four were positively identified in this EWAS screen. When the EWAS screen was widened somewhat to include the other stem cell factors identified by

the Yamanaka group and others [188], again many were positively identified, including Ras, catenins, Kit and the Lin28 microRNA.

3.2.3. Chromosomal Disorders

As noted above, chromosomal disorders hold a prominent place in the patterns of cannabinoid-related carcinogenic and teratogenic disorders. Indeed, when the length of all the chromosomes involved was summed (omitting duplications), it was found that an impressive 1765 megabases of the 3000 megabases of the human genome were directly implicated in the cannabinoid-related genotoxic disorders, which is 58.8% of the human genome. For this reason, the epigenomic findings of the Schrott EWAS dataset were of immense importance.

Additionally, the rays of the mitotic spindle were composed of microtubules of polymerized tubulin. There were 106 hits in the Schrott EWAS database for tubulins. Importantly, tubulin undergoes numerous post-translational modifications which are thought to govern its intracellular trafficking and organellar addressing [189–191]. The cannabis withdrawal disrupted the alpha tubulin acetyl transferase which was tasked with acetylating tubulin and thereby made the microtubules flexible and increased its tensile and torsional strength. This was important as the microtubules are normally bent during the spindle formation and tensioning. Failure of completing this action leads to microtubular breaks and fractures and, thus, the chromosomal derailment during the anaphase.

The centromeres are the critical central portions of chromosomes where binding occurs to the mitotic spindle. Centrosomal protein A (CENPA) was a modified version of histone 3 (H3) and CENPA replaced H3 in the centromere, which marked the location of the centromere. Upon this CENPA basis, a complicated scaffold of 17 proteins assembled which then was bound to the microtubules of the spindle via other kinetochore scaffolding proteins [192–196]. Fifteen different CENPs were identified in this EWAS screen, including 86 hits for CENPN, which was the second protein to bind to the centromere complex.

The proteins which cohere the ends of the human oocyte meiotic spindle so that two (and only two) spindle poles are formed, guiding the formation of two (and only two) daughter cells, are called centrosomal organizers. There were three hits for these proteins, including the nuclear mitotic apparatus protein (NUMA).

The motor proteins which actually move the chromosomes along the microtubules towards minus end of the microtubules and the spindle poles after the anaphase checkpoint is released are called dynein motors which are controlled by a binding partner known as dynactin. There were seven EWAS hits for dynein–dynactin. Interestingly the intracellular kinesin motor moved protein and other cargo in the opposite direction towards the positive end of the microtubule and 218 hits were recorded for kinesin motors.

Sumoylation was shown to be a key post-translational modification of the key proteins, which organizes the chromosomes and are known as the remodelers of the structure of the chromatin (RSC) complexes in yeast [197]. Sumoylation involves the addition of small ubiquitin-like molecules, often in chains, to the key signaling residues of the proteins. Sumoylation of the RSC forms the founder post-translational modification upon which a string of subsequent post-translational modifications may be established [198]. These are believed to control the RSC complex activity. This RSC was shown to be centrally involved in the key chromosomal functions, such as the DNA break repair, chromosomal segregation and chromosomal duplication [198]. $\Delta 9 THC$ inhibited this sumoylation process directly [100], disrupting the downstream signaling through the epigenetic histone code to H3 mono-, diand tri-methylation, H3/H4 acetylation and H2B lysine 123 ubiquitylation [198].

It was also noted that the EWAS screen showed nine hits against RAD51, which was the key member of the high-fidelity homologous recombination (HR) pathway, but only one hit against RAD52, which was part of the low-fidelity non-homologous end-joining pathway [42]. It was previously shown that the inhibition of the high-fidelity HR leads to the activation of the low-fidelity default microhomology end-joining repair pathway [68].

Thus, these many results clearly impacted and disrupted all the major functions of the chromosomes and likely provided a powerful epigenomic underpinning for the epidemiologically observed carcinogenic and teratogenic pathophysiology. Moreover, the DNA breakage was shown to be a prominent feature of the cannabis exposure of oocytes, sperm, lymphocytes and many other cells, and these finding imply that these lesions were preferentially repaired low-fidelity rather than high-fidelity pathways due to the epigenomic dysregulatory mechanisms.

3.3. Brain Development and Brain Aging

The Schrott EWAS study [42] revealed a widespread disruption to the receptor-based signaling, including 132 of the ionotropic AMPA receptors (GRIA), the main workhorse excitatory receptor of the CNS, 165 hits on the kainate glutamate receptor (GRIK), 26 hits on the NMDA glutamate receptor (GRIN) that mediates neuroplasticity and long-term potentiation, 11 hits on the delta glutamate receptor (GRID), 122 hits on the glutamate metabotropic receptor (GRM), 125 hits on the inhibitory GABA A receptor (GABRA), 22 hits on the GABA B receptor (GABRB), 85 hits on the "feel good" serotonin receptor (HTR), 17 hits on the dopamine receptors, five hits on each of the μ - and δ -opioid receptors and seven hits on the oxytocin "feel great" receptor.

There were ten hits each on neurexin and neuroligin, which are a ligand–receptor pair that mediate the receptor formation and scaffolding. There were eight hits on discs large homolog-associated protein 2 (DLGAP2), which is a protein known to be involved in synaptic scaffolding and the previously implicated ion autism development [102]. Similarly, there were 14 hits on the Down syndrome cell adhesion molecule (DSCAM), which is involved in axonal and dendritic pathfinding, self-avoidance and olfaction.

Recent studies have shown that the massive overgrowth of the human cerebral cortex relative to other species is controlled by signaling between the Slit-Robo ligand–receptor pair [199–201]. This was shown to be inhibited by cannabis [202–204]. There were 351 hits for the Slit signaling in the Schrott dataset and 40 hits for Robo. Moreover, there were eight hits for a key activating enzyme in this pathway—the Slit-Robo Rho GTPase activating protein (SRGAP2). These findings imply the impeded brain and neocortical outgrowth.

Another key study found that the very high gradient of retinoic acid at the frontal pole of the forebrain was responsible for driving the frontal lobe outgrowth [205]. The gradient was maintained using a retinoic acid synthesizing enzyme—alcohol dehydrogenase 1 (ALDH1)—at the frontal pole, transduced by the retinoic acid receptors RXRG and RARB, and was dissipated using the metabolic enzymes of the CYP26B1 group which had a high concentration at the posterior of the frontal lobe and the premotor cortex. There were 13 hits in the Schrott EWAS dataset for ALDH1, ten hits for RXRG and RARB and ten hits for the CYP2 series cytochrome metabolizing enzymes.

These data showed that the cannabinoid stimulated epigenomic pathways disrupted the synaptic processing across a broad range of receptor subtypes, synaptic scaffolding using several routes, and neural progenitor and forebrain outgrowth by inhibiting several of the main pathways responsible for this key proliferation action. Such findings indicated that mental illness and congenital neurological conditions, including autistic spectrum disorders and developmental disorders such as microcephaly and anencephaly, were more likely, as observed in an increasing number of large epidemiological studies on community cannabis exposure [28,29,40,45]. Since these disorders were also characterized by impaired brain development, they may be seen as broadly degenerative in nature and, thus, consistent with an advanced broadly defined aging profile.

This was, in turn, systemically important as brain aging has been well demonstrated to drive whole organism aging [164–166,206]. Indeed, accelerated systemic aging accompanies many syndromes where brain aging features prominently, including progeria and Down syndrome [207–211].

3.4. Vascular Aging

The issue of vascular aging has broader implications than simply the cardiovascular system since it has been aphoristically said that "you are as old as your arteries" [164,165,212,213]. This is true not only because most people succumb to macrovascular cardiovascular disorders [214] but because most stem cell niches contain a microvascular compartment which is key to stem cell function generally.

Cannabis exposure has been shown to advance human cardiovascular age in an ecological longitudinal study [33].

The key genes in arterial development are sonic hedgehog (shh), the vascular endothelial growth factor (VEGF) and notch and ephrinB2 signaling [215].

Importantly, when investigating the EWAS-identified epigenetic methylation changes to human sperm, sonic hedgehog signaling was shown to be disrupted by nine hits on both the patched co-receptor and elsewhere, in addition to 185 hits on the key Gli3 transcription factor which signaled to the nuclear genome [42]. Notch, VEGF and ephrinB2 were disrupted at 18, five and one hits, respectively [42].

The point of these findings was not only to identify that cardiovascular development can be disrupted in these ways but that the induced arterial aging can also induce the system-wide whole organism aging processes through an impairment of the stem cell quiescence/multiplication balance both directly and indirectly.

3.5. Epigenomic Disruptions by Organ System

The Schrott EWAS contained 73 hits for central nervous system dysfunctions, including the brain, neurological, synaptic, cerebral, neuronal and eye derangements.

At total of 29 hits were noted for cardiovascular disorders, including the heart, atria, ventricles, atrioventricular valves and vessels.

Additionally, 22 hits were noted for orofacial genetic lesions, including the head, sensory organs, palate, nose, anterior eye and ear derangements.

Six hits were identified for limb development directly. Further exploration of a more exhaustive list of the limb morphogens revealed 130 hits for most of the key morphogens involved in limb and digit development, including the fibroblast growth factors (FGFs), retinoid signaling, Wnt signaling, bone morphogenetic pathway signaling and five genes from the sonic hedgehog (shh) pathway, namely MEGF8 (multiple EGF-like domains 8), TMEM107 (transmembrane protein 107), Gli3 (GLI gamily zinc finger 3), CHD7 (chromodomain helicase DNA-binding protein 7) and the patched receptor cofactor. Indeed, 185 hits for the key shh transcription factor Gli3 were found in the Schrott EWAS.

There were 37 hits for development of the gastrointestinal tract, including references to the esophagus, large intestine, liver and pancreas. This epigenomic pattern was noted to be consistent with the pattern of the anomalies observed in the population cannabis exposure data from both the US and Europe [28,36,40,51].

There were 23 hits observed for the urinary system, including the kidneys. When a more detailed exploration of gene regulation guided by a recent developmental renal cell map was used as a guide for data mining [216], a total of 51 hits were identified for renal development. In addition 18, 27 and 18 hits were identified for the key renal tract morphogens—notch, sonic hedgehog and transforming growth factor β , respectively.

A total of 15 hits were identified for the body wall and embryo.

Additionally, 60 hits were noted for the general otherwise unclassified disorders, including embryonic growth, DNA, mitochondria, microtubules, body trunk, body axis, ovarian reserve, breast disorders, granulocytes, myogenesis, vertebral growth and bone development.

Hence, an abundance of epigenomic evidence exists to explain the broad spectrum and high severity of the teratological patterns observed in the many jurisdictions described. These findings are described in further quantitative detail elsewhere [217].

3.6. Cancer Hits in the Schrott EWAS

The Schrott supplementary file lists 487 hits for "cancer", 112 hits for "tumor", 126 hits for "carcinoma", 36 hits for "neoplasm", 32 hits for "leukemia" and 17 hits for "lymphoma". This totals 810 hits for cancer and its synonyms, making this one of the standout and major findings of this EWAS report.

The report specifically mentioned many leukemias, lymphomas, myeloma and tumors of the breast, ovary colorectum, thyroid, liver, brain, pancreas, melanoma, stomach, esophagus and upper aerodigestive tract.

As noted, all of these cancers have been described in association with cannabis exposure in historical [22–26,30,41,51,218–228] and recent reports [22–24,26,30,34,41,51,83,229]. Further quantitative details in the description of this material is provided elsewhere [229].

By listing over 30 cancers by name and the genes whose epigenomic modulation was linked with them, these results provided a powerful pan-cancer mechanistic contributory explanation for the patterns of cancer epidemiologically observed in human populations.

It is also worth noting that many of the more recent epidemiological reports proceeded beyond the methodologies commonly adopted in observation cohort studies [22–24,26,30,34,41,51,83,229,230]. By applying the formal techniques of causal inference including inverse probability weighting and E-values to quantitatively exclude extraneous unmeasured confounding these investigators have constructed a pseudo-randomized analytical framework and, therefore, reported the causal relationships in preference to the more commonly noted ecological associations [22–24,26,30,34,41,51,83,229,230].

3.7. Aging Implications of the Schrott EWAS Dataset

A concise overview and introduction to aging was provided in the preceding sections. Since DNA methylation was shown to be a key determinant of the progressive decline of the function which characterizes the aging process, and since cannabis dependence and withdrawal was shown to widely disrupt both DNA methylation and demethylation and the histone code with which it is coordinated, the disruption of the aging process itself is not unexpected. As noted above, this was confirmed in somatic tissues experimentally and found to be of high magnitude at 30% at 30 years of age [54].

It was also shown that cannabinoids can reduce the telomerase activity in a rat hepatocarcinogenesis model [121]. The Schrott EWAS dataset showed that the telomerase activity was epigenetically reduced with a significance of $p = 2.82 \times 10^{-6}$ and a multiplicity-corrected p-value of 0.01258 [42]. Indeed, since cannabis dependence inhibits TET1 ($p = 1.18 \times 10^{-5}$, multiplicity-corrected p-value = 0.02278) and this is the main counterbalancing force to the promoter hypermethylation of aging, it is easy to understand how the accelerated aging process is not only established initially, but how it might become a positive feed-forward process with time as age-related epigenomic changes are predisposed to further pro-aging epigenomic processes.

Two of the key tissues with which we were concerned in the present context were the male and female gametes. It was understood that none of the presently available epigenetic clocks were suitable for the application to measure the relatively very hypomethylated ages of the gametes. However it does stand to reason that it may be possible to develop such an algorithmic clock mathematically. What the negative ages might mean, as they may relate to ages prior to birth, has yet to be determined biologically. Hence, it was not possible to measure gametal age directly or epigenomically at the time of writing.

However, it was emphasized that the well-described presence of the characteristic aging nuclear changes on the sperm of the DNA chromosomal breaks and translocations [115] and oocyte nuclear blebs and bridges [114] provided strong genetic evidence of the changes of accelerated aging. The likelihood then that both gametes and the fertilized zygote are "old prior to conception" clearly has far reaching public health and multigenerational implications in terms of the prenatal origin of many disorders in later life [171,231].

3.8. Strengths and Limitations

There are various strengths and limitations to the present conceptualization. The strengths include the remarkable consistency across the many epidemiological studies, which clearly demonstrates the genotoxic harms of cannabis exposure in several different international jurisdictions, in relation to both the congenital anomalies [28,36,37,40,45,46,49,51,71,73–82] and cancer [22–26,30,41,232], and, indeed, now also in aging [53,54]. Similar results in many different studies are clearly mutually supportive and strengthen the overall quality of the body of evidence. Similarly, there is a striking concordance between the many epigenomic studies of gestational cannabis exposure in relation to global DNA hypomethylation and the disruption of DNA methylation levels at key promoter and enhancer sites, which control the regulation of many critical genes [42,101–103,105–108,233,234].

The major limitation of the present work is its preliminary nature in that we present an introductory conceptual framework which needs to be filled out and completed by numerous further laboratory studies. The purpose of the present paper is merely to draw attention to this remarkable concordance of cross-disciplinary results and indicate to researchers in the basic sciences that the field is ripe for detailed exploration in many studies with far-ranging consequences.

3.9. Future Directions

In the same way that Harvard researchers were recently able to advance cellular and organismal age forwards and backwards by the induction of just a few DNA breaks and then demonstrate their phenotypical rescue with the OSK Yamanaka stem cell factors [235] so too models of cannabis exposure lend themselves to similar exploration by experimental induction of aging in cells and model organisms and then their rescue either with a subset of the Yamanaka factors [184–188,235] or chemical cocktails [236], which are similarly directed. Thus, cannabinoid research could intersect powerfully with aging research in general at a time when the whole field is making important advances.

Moreover, the 810 differentially methylated genes implicated with cancer by the work of the Murphy lab [42] have not yet been explored. In much the same way that potentially important breakthroughs may occur by exploring and developing cannabis-based models of aging, they could also be developed in cancer. While the present paper serves to sketch a general outline based on the now considerable body of evidence from DNA methylation work, there is much of the story which urgently needs to be investigated. Due to its obvious public health importance from the widespread and now intergenerational cannabinoid exposure, which is occurring in many places, it is appropriate that cannabinoid oncogenesis serves as an important model for cancer research.

The important questions that need to be considered include the following.

- How is the landscape of the post-translational histone modifications changed by cannabis exposure, particularly in relation to the key modifications, such as histone acetylation, which reliably mark active genes?
- How are the critically important issues of the modulation of super-enhancers and super-anchors affected by cannabinoid exposure?
- Can tissue de-differentiation be demonstrated experimentally from cannabinoids and, if so, which tissues are the most susceptible? What are their time course? What are their dose-response effect? In particular, in the ovaries, testes, brain, liver, heart, respiratory tract and immunocytes. Is this de-differentiation premalignant? Do cannabinoids induce premalignant field changes and cancerization from adverse genomic (DNA breaks), epigenomic or metabolomic effects or some interaction between all of these and more?
- Oocytes are particularly genomically fragile and non-renewable cells and have a very long life of many decades. Their genomic, metabolic and epigenomic vulnerability to cannabinoids needs to be characterized in detail. What are the effects of cannabinoids on oogenic stem cells both prenatally and postnatally?

- Since cannabinoids affect the mitochondria and cell metabolism adversely and this is closely related to maintenance of both the genome and epigenome, how does this compare to the Warburg effect observed in stem cells and cancer cells [237]? How is it similar? How is it different?
- Such studies would provide an invaluable window of insight into the way the metabolome and epigenome are coordinated and bidirectionally co-regulate each other.
- In particular, the effect of lactylation as a key post-translational modification of the key metabolic enzymes and histones needs to be quantified, as this has also been shown to be a critical post-translational modification for stem cells, cancer cells and cancer stem cells with potentially far-reaching and cross-disciplinary applications in aging medicine and cancer biology [238,239].
- As described, cannabis widely disrupts many of the key enzymes of the epigenetic machinery itself. What are the implications of this?
- As discussed, cannabinoids disrupt key elements of the mitotic spindle, microtubule physiology, spindle pole formation and kinetochore and centrosome formation. These are very far-reaching findings as chromosomal mis-segregation has implications in many fields, including male and female fertility optimization, sperm and egg storage, the preservation and fertility of medicine, the biology of aging, cancerogenesis, congenital anomalies in early pediatrics and neurodevelopmental alterations in later pediatrics. This field has been largely ignored but is very much in need of detailed exploration and explication at the cellular and molecular levels from the point of view of the impacts of cannabinoids.
- As described by epigenomically disrupting both CTCF and the major components
 of the cohesion complex, it seems inevitable that cannabis must disrupt the basic
 machinery of chromatin looping and gene expression itself. What are the implications
 of this potentially very far-reaching derangement?
- What is it about testicular germ cells that makes them particularly susceptible to cannabinoid oncogenesis? Is it the DNA hypomethylation of germ cells to start?
- Techniques are emerging to allow the characterization of the non-protein coding, regulatory and repeat sequences in the genome. In what way is this normal physiology perturbed by cannabinoid exposure?
- DNA hypomethylation of the gene bodies and gene deserts is likely to result in the mobilization of transposable elements in the genome, which precipitates genomic instability, cytoplasmic and extracellular immune activation (via cGAS—STING and downstream IL1β, IL6 and interferons) and, thus, aging and oncogenesis in both the immune and genomic pathways. This field needs to be explored and further developed.
- The advancement of human epigenomic age from cannabis exposure at a single age has been mentioned [54]. What is the time course of this across the lifespan? How does it progress? Dose it rise as a linear function of age or as a polynomial or exponential function of age as suggested by the biophysical clinical studies [33]? What are its dose-response characteristics?
- Many different cannabinoids need to be profiled in vitro to characterize their multichannel epigenomic effects (DNA methylation, many histone methylation and acetylation modifications, super-enhancers) in the modern era to define the breadth of their epigenotoxicity as a possible or likely class effect.
- The well-documented exponentiation of the cannabinoid dose-response effects in many cellular and metabolic assays needs to be formally explored in the modern epigenomic context and its public health significance needs to be carefully considered.

These and a host of similar questions are now present for careful review and exploration by modern laboratory techniques with almost certainly far-reaching implications for both the basic sciences and clinical medicine.

4. Conclusions

The above overview of several large datasets is notable for many reasons. There is a surprising uniformity in the findings from the three domains of cannabinoid-related teratogenesis, carcinogenesis and the acceleration of aging processes, which together imply a clinically significant if widely underrated cannabinoid genotoxicity. Moreover, the severity of the findings in all three domains is considerably greater than is commonly supposed.

The other very striking feature is that the predictive power of the epigenomic profiles provided by the recent epigenome wide studies, particularly that of Schrott and colleagues [42], is the predictive power of the morbidity data arising from both the European and US datasets following increased cannabis exposure. The identified DNA methylation changes and associated biological mechanistic pathways explain these observed patterns of disorders.

The wide spectrum of cancers is specifically explained and the organ-specific nature of many congenital anomalies is explained. The pathophysiology of the wide spectrum of chromosomal disorders which occur as both malignant disorders and congenital anomalies is also elegantly explained by several mechanisms. The pattern of limb anomalies, including syndactyly and polydactyly, is explained by the widespread epigenomic disruption of most of the key limb morphogen gradients. Most particularly, the patterns of accelerated aging are explained by the epigenomic inhibition of DNA methylation, demethylation and telomerase, among others, and confirmed by the recent pattern of chronic illness studies and epigenomic clock investigations.

Most concerning, the very real prospects that both sperm and oocytes are epigenomically aged prior to conception carry severe and dire prospects for public health and multigenerational epigenomic transmission and beg a formal assessment using gamete-appropriate epigenomic clocks whose development must be considered a major research priority.

With the increasing interest in the pharmaceutical use of cannabis and cannabis derivatives for medical management, it is increasingly important that the balance between the therapeutic risks and benefits for each potential cannabis/cannabinoid application is thoroughly understood. The mandate of regulatory global authorities is to not succumb to strong commercial and popular interest in the promotion of cannabis/cannabinoids until the risk-benefit appraisal can be accurately undertaken for each potential cannabis application, considering, among other items, the potency, severity of disease, stage of human development and duration of use.

Author Contributions: A.S.R. assembled the data, designed and conducted the analyses and wrote the first manuscript draft. G.K.H. provided technical and logistic support, co-wrote the paper, assisted with gaining the ethical approval, provided advice on the manuscript preparation and general guidance for the study conduct. A.S.R. had the idea for the article, performed the literature search, wrote the first draft and is the guarantor for the article. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding. No funding organization played any role in the design and conduct of the study; collection, management, analysis or interpretation of the data; preparation, review or approval of the manuscript; or the decision to submit the manuscript for publication.

Institutional Review Board Statement: Ethics approval and consent to participate. The Human Research Ethics Committee of the University of Western Australia provided ethical approval for the study to be undertaken on 24 September 2021 (No. RA/4/20/4724).

Informed Consent Statement: Patient consent was not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: All authors had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

Conflicts of Interest: The authors declare that they have no competing interests.

Abbreviations

Abbreviation Meaning

ALDH1 Alcohol dehydrogenase 1
ALL Acute lymphoid leukemia
AML Acute myeloid leukemia

AMPA α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid

CENPA Centrosomal protein A
CENPN Centrosomal protein N

CHD7 Chromodomain helicase DNA-binding protein 7

CYP Cytochrome

DLGAP2 Discs large homolog-associated protein 2 DSCAM Down syndrome cell adhesion molecule

EMA European Medicines Agency
EWAS Epigenome-wide association study
FDA Food and Drug Administration

FGF Fibroblast growth factor
GABA γ-aminobutyric acid
GABRA GABA A receptor
GABRB GABA B receptor
Gli3 GLI family zinc finger 3

GRIA Glutamate ionotropic receptor AMPA-type subunit
GRID Glutamate ionotropic receptor Delta-type subunit
GRIK Glutamate ionotropic receptor Kainate-type subunit
GRIN Glutamate ionotropic receptor NMDA-type subunit

GRM Glutamate metabotropic receptor

HTR Serotonin receptor
Klf4 Kruppel-like factor 4
MEGF8 Multiple EGF-like domains 8

MHCPRA Medicines and Health Care Products Regulatory Agency

Myc Myc proto-oncogene, bHLH transcription factor

NMDA N-methyl-D-aspartate

NUMA Nuclear-mitotic apparatus protein Oct3/4 POU5F1, POU class 5 homeobox 1

RAD51 RAD51 recombinase

RAD52 RAD52 homologue, DNA repair protein

RARB Retinoic acid receptor beta

RSC Remodelers of the structure of Chromatin

RXRG Retinoid X receptor gamma

shh Sonic hedgehog

SMARCA SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin,

subfamily A

Sox2 SRY-box transcription factor 2

SRGAP2 Slit-Robo Rho GTPase-activating protein 2

TET1 Ten-eleven translocase
 THC Δ9-tetrahydrocannabinol
 TMEM107 Transmembrane protein 107

UHRF Ubiquitin-like with PHD and ring finger domains

VEGF Vascular endothelial growth factor

Wnt Wnt family member
Δ8THC Δ8-tetrahydrocannabinol
Δ9THC Δ9-tetrahydrocannabinol

References

- Package Leaflet: Information for the Patient: Sativex[®] Oromucosal Spray. Available online: https://www.medicines.org.uk/emc/files/pil.602.pdf (accessed on 17 March 2021).
- 2. Greenwich Biosciences. *Epidiolex: Highlights of Prescribing Information*; Food and Drug Administration: Silver Springs, MD, USA, 2018; Volume 1, p. 1.

- 3. Busch, F.W.; Seid, D.A.; Wei, E.T. Mutagenic activity of marihuana smoke condensates. Cancer Lett. 1979, 6, 319–324. [CrossRef]
- 4. Fisar, Z.; Singh, N.; Hroudova, J. Cannabinoid-induced changes in respiration of brain mitochondria. *Toxicol. Lett.* **2014**, 231, 62–71. [CrossRef]
- 5. Fish, E.W.; Murdaugh, L.B.; Zhang, C.; Boschen, K.E.; Boa-Amponsem, O.; Mendoza-Romero, H.N.; Tarpley, M.; Chdid, L.; Mukhopadhyay, S.; Cole, G.J.; et al. Cannabinoids Exacerbate Alcohol Teratogenesis by a CB1-Hedgehog Interaction. *Sci. Rep.* **2019**, *9*, 16057–16075. [CrossRef] [PubMed]
- 6. Hölzel, B.N.; Pfannkuche, K.; Allner, B.; Allner, H.T.; Hescheler, J.; Derichsweiler, D.; Hollert, H.; Schiwy, A.; Brendt, J.; Schaffeld, M.; et al. Following the adverse outcome pathway from micronucleus to cancer using H2B-eGFP transgenic healthy stem cells. *Arch. Toxicol.* **2020**, *94*, 3265–3280. [CrossRef]
- 7. Koller, V.J.; Auwarter, V.; Grummt, T.; Moosmann, B.; Misik, M.; Knasmuller, S. Investigation of the in vitro toxicological properties of the synthetic cannabimimetic drug CP-47,497-C8. *Toxicol. Appl. Pharmacol.* **2014**, 277, 164–171. [CrossRef]
- 8. Koller, V.J.; Ferk, F.; Al-Serori, H.; Misik, M.; Nersesyan, A.; Auwarter, V.; Grummt, T.; Knasmuller, S. Genotoxic properties of representatives of alkylindazoles and aminoalkyl-indoles which are consumed as synthetic cannabinoids. *Food Chem. Toxicol.* **2015**, *80*, 130–136. [CrossRef]
- 9. Morimoto, S.; Tanaka, Y.; Sasaki, K.; Tanaka, H.; Fukamizu, T.; Shoyama, Y.; Shoyama, Y.; Taura, F. Identification and characterization of cannabinoids that induce cell death through mitochondrial permeability transition in Cannabis leaf cells. *J. Biol. Chem.* **2007**, 282, 20739–20751. [CrossRef]
- 10. Price, P.J.; Suk, W.A.; Spahn, G.J.; Freeman, A.E. Transformation of Fischer rat embryo cells by the combined action of murine leukemia virus and (-)-trans-9-tetrahydrocannabinol. *Proc. Soc. Exp. Biol. Med.* **1972**, *140*, 454–456. [CrossRef]
- 11. Russo, C.; Ferk, F.; Mišík, M.; Ropek, N.; Nersesyan, A.; Mejri, D.; Holzmann, K.; Lavorgna, M.; Isidori, M.; Knasmüller, S. Low doses of widely consumed cannabinoids (cannabidiol and cannabidivarin) cause DNA damage and chromosomal aberrations in human-derived cells. *Arch. Toxicol.* **2019**, *93*, 179–188. [CrossRef]
- 12. Sarafian, T.A.; Habib, N.; Oldham, M.; Seeram, N.; Lee, R.P.; Lin, L.; Tashkin, D.P.; Roth, M.D. Inhaled marijuana smoke disrupts mitochondrial energetics in pulmonary epithelial cells in vivo. *Am. J. Physiol.* **2006**, 290, L1202–L1209. [CrossRef]
- 13. Sarafian, T.A.; Kouyoumjian, S.; Khoshaghideh, F.; Tashkin, D.P.; Roth, M.D. Delta 9-tetrahydrocannabinol disrupts mitochondrial function and cell energetics. *Am. J. Physiol.* **2003**, 284, L298–L306. [CrossRef]
- 14. Shoyama, Y.; Sugawa, C.; Tanaka, H.; Morimoto, S. Cannabinoids act as necrosis-inducing factors in Cannabis sativa. *Plant Signal. Behav.* **2008**, *3*, 1111–1112. [CrossRef]
- 15. Singh, N.; Hroudova, J.; Fisar, Z. Cannabinoid-Induced Changes in the Activity of Electron Transport Chain Complexes of Brain Mitochondria. *J. Mol. Neurosci.* **2015**, *56*, 926–931. [CrossRef]
- 16. Tahir, S.K.; Trogadis, J.E.; Stevens, J.K.; Zimmerman, A.M. Cytoskeletal organization following cannabinoid treatment in undifferentiated and differentiated PC12 cells. *Biochem. Cell Biol.* 1992, 70, 1159–1173. [CrossRef] [PubMed]
- 17. Tahir, S.K.; Zimmerman, A.M. Influence of marihuana on cellular structures and biochemical activities. *Pharmacol. Biochem. Behav.* **1991**, 40, 617–623. [CrossRef] [PubMed]
- 18. Vela, G.; Martin, S.; Garcia-Gil, L.; Crespo, J.A.; Ruiz-Gayo, M.; Fernandez-Ruiz, J.J.; Garcia-Lecumberri, C.; Pelaprat, D.; Fuentes, J.A.; Ramos, J.A.; et al. Maternal exposure to delta9-tetrahydrocannabinol facilitates morphine self-administration behavior and changes regional binding to central mu opioid receptors in adult offspring female rats. *Brain Res.* 1998, 807, 101–109. [CrossRef] [PubMed]
- 19. Zimmerman, A.M.; Raj, A.Y. Influence of cannabinoids on somatic cells in vivo. *Pharmacology* **1980**, 21, 277–287. [CrossRef] [PubMed]
- 20. Watson, S.J.; Benson, J.A., Jr.; Joy, J.E. Marijuana and medicine: Assessing the science base: A summary of the 1999 Institute of Medicine report. *Arch Gen. Psychiatry* **2000**, *57*, 547–552. [CrossRef] [PubMed]
- 21. National Academies of Sciences, Engineering, and Medicine. The Health Effects of Cannabis and Cannabinoids: The Current State of Evidence and Recommendations for Research. In *The Health Effects of Cannabis and Cannabinoids: The Current State of Evidence and Recommendations for Research;* The National Academies Collection: Reports funded by National Institutes of Health, Ed.; National Academies Press: Washington, DC, USA, 2017; Volume 1.
- 22. Reece, A.S.; Hulse, G.K. Geotemporospatial and Causal Inferential Epidemiological Overview and Survey of USA Cannabis, Cannabidiol and Cannabinoid Genotoxicity Expressed in Cancer Incidence 2003–2017: Part 1—Continuous Bivariate Analysis. *Arch. Public Health* 2022, 80, 99–133. [CrossRef] [PubMed]
- 23. Reece, A.S.; Hulse, G.K. Geotemporospatial and Causal Inferential Epidemiological Overview and Survey of USA Cannabis, Cannabidiol and Cannabinoid Genotoxicity Expressed in Cancer Incidence 2003–2017: Part 2—Categorical Bivariate Analysis and Attributable Fractions. *Arch. Public Health* 2022, 80, 100–135. [CrossRef] [PubMed]
- 24. Reece, A.S.; Hulse, G.K. Geotemporospatial and Causal Inferential Epidemiological Overview and Survey of USA Cannabis, Cannabidiol and Cannabinoid Genotoxicity Expressed in Cancer Incidence 2003–2017: Part 3—Spatiotemporal, Multivariable and Causal Inferential Pathfinding and Exploratory Analyses of Prostate and Ovarian Cancers. Arch. Public Health 2022, 80, 100–136. [CrossRef] [PubMed]
- 25. Reece, A.S.; Hulse, G.K. Cannabinoid exposure as a major driver of pediatric acute lymphoid Leukaemia rates across the USA: Combined geospatial, multiple imputation and causal inference study. *BMC Cancer* **2021**, 21, 984. [CrossRef] [PubMed]

- 26. Reece, A.S.; Hulse, G.K. A geospatiotemporal and causal inference epidemiological exploration of substance and cannabinoid exposure as drivers of rising US pediatric cancer rates. *BMC Cancer* **2021**, *21*, 197. [CrossRef]
- 27. Reece, A.S.; Hulse, G.K. Contemporary epidemiology of rising atrial septal defect trends across USA 1991-2016: A combined ecological geospatiotemporal and causal inferential study. *BMC Pediatr.* **2020**, *20*, 539. [CrossRef]
- 28. Reece, A.S.; Hulse, G.K. Geotemporospatial and causal inference epidemiological analysis of US survey and overview of cannabis, cannabidiol and cannabinoid genotoxicity in relation to congenital anomalies 2001–2015. *BMC Pediatr.* 2022, 22, 47. [CrossRef] [PubMed]
- 29. Reece, A.S.; Hulse, G.K. Broad Spectrum epidemiological contribution of cannabis and other substances to the teratological profile of northern New South Wales: Geospatial and causal inference analysis. *BMC Pharm. Toxicol.* **2020**, *21*, 75. [CrossRef]
- 30. Reece, A.S.; Hulse, G.K. Causal inference multiple imputation investigation of the impact of cannabinoids and other substances on ethnic differentials in US testicular cancer incidence. *BMC Pharm. Toxicol.* **2021**, 22, 40. [CrossRef] [PubMed]
- 31. Reece, A.S.; Hulse, G.K. Co-occurrence across time and space of drug- and cannabinoid- exposure and adverse mental health outcomes in the National Survey of Drug Use and Health: Combined geotemporospatial and causal inference analysis. *BMC Public Health* 2020, 20, 1655. [CrossRef]
- 32. Reece, A.S. Rapid Response: Known Cannabis Teratogenicity Needs to be Carefully Considered. BMJ 2018, 362, k3357. [CrossRef]
- 33. Reece, A.S.; Norman, A.; Hulse, G.K. Cannabis exposure as an interactive cardiovascular risk factor and accelerant of organismal ageing: A longitudinal study. *BMJ Open* **2016**, *6*, e011891–e011901. [CrossRef]
- Reece, A.S. Cannabinoid Genotoxic Trifecta—Cancerogenesis, Clinical Teratogenesis and Cellular Ageing. Br. Med. J. 2022, 376, n3114, Rapid Response. [CrossRef]
- 35. Reece, A.S. Limblessness: Cannabinoids Inhibit Key Embryonic Morphogens both Directly and Epigenomically. *Br. Med. J.* **2022**. Available online: https://www.bmj.com/content/376/bmj.n3114/rr-2 (accessed on 12 February 2023).
- 36. Reece, A.S.; Hulse, G.K. Cannabinoid Genotoxicity and Congenital Anomalies: A Convergent Synthesis of European and USA Datasets. In *Cannabis, Cannabinoids and Endocannabinoids*; Preedy, V., Patel, V., Eds.; Elsevier: London, UK, 2022; Volume 1, *in press*.
- 37. Reece, A.S.; Hulse, G.K. Cannabis Teratology Explains Current Patterns of Coloradan Congenital Defects: The Contribution of Increased Cannabinoid Exposure to Rising Teratological Trends. *Clin. Pediatr.* **2019**, *58*, 1085–1123. [CrossRef]
- 38. Reece, A.S.; Hulse, G.K. Epidemiological Associations of Various Substances and Multiple Cannabinoids with Autism in USA. *Clin. Pediatr.* **2019**, *4*, 155. [CrossRef]
- Reece, A.S.; Hulse, G.K. Effect of Cannabis Legalization on US Autism Incidence and Medium Term Projections. Clin. Pediatr. 2019, 4. [CrossRef]
- 40. Reece, A.S.; Hulse, G.K. Cannabinoid- and Substance- Relationships of European Congenital Anomaly Patterns: A Space-Time Panel Regression and Causal Inferential Study. *Environ. Epigenet.* **2022**, *8*, dvab015. [CrossRef] [PubMed]
- 41. Reece, A.S.; Hulse, G.K. Geospatiotemporal and Causal Inference Study of Cannabis and Other Drugs as Risk Factors for Female Breast Cancer USA 2003–2017. *Environ. Epigenet.* **2022**, 2022, dvac006. [CrossRef] [PubMed]
- 42. Schrott, R.; Murphy, S.K.; Modliszewski, J.L.; King, D.E.; Hill, B.; Itchon-Ramos, N.; Raburn, D.; Price, T.; Levin, E.D.; Vandrey, R.; et al. Refraining from use diminishes cannabis-associated epigenetic changes in human sperm. *Environ. Epigenet.* **2021**, 7, dvab009. [CrossRef] [PubMed]
- 43. Reece, A.S.; Hulse, G.K. Impacts of cannabinoid epigenetics on human development: Reflections on Murphy et. al. 'cannabinoid exposure and altered DNA methylation in rat and human sperm' epigenetics 2018; 13: 1208-1221. *Epigenetics* 2019, 14, 1041–1056. [CrossRef]
- 44. Reece, A.S.; Hulse, G.K. Cannabis Consumption Patterns Explain the East-West Gradient in Canadian Neural Tube Defect Incidence: An Ecological Study. *Glob. Pediatr. Health* **2019**, *6*, 2333794X19894798. [CrossRef] [PubMed]
- 45. Reece, A.S.; Hulse, G.K. Canadian Cannabis Consumption and Patterns of Congenital Anomalies: An Ecological Geospatial Analysis. *J. Addict. Med.* **2020**, *14*, e195–e210. [CrossRef] [PubMed]
- 46. Forrester, M.B.; Merz, R.D. Risk of selected birth defects with prenatal illicit drug use, Hawaii, 1986-2002. *J. Toxicol. Environ. Health* **2007**, 70, 7–18. [CrossRef]
- 47. Reece, A.S.; Hulse, G.K. Gastroschisis and Autism-Dual Canaries in the Californian Coalmine. *JAMA Surg.* **2019**, *154*, 366–367. [CrossRef]
- 48. Reece, A.S.; Hulse, G.K. Cannabis and Pregnancy Don't Mix. Mo. Med. 2020, 117, 530-531.
- 49. Reece, A.S.; Hulse, G.K. Chromothripsis and epigenomics complete causality criteria for cannabis- and addiction-connected carcinogenicity, congenital toxicity and heritable genotoxicity. *Mutat Res.* **2016**, 789, 15–25. [CrossRef]
- 50. Reece, A.S.; Hulse, G.K. Cannabis in Pregnancy—Rejoinder, Exposition and Cautionary Tales. *Psychiatr. Times* **2020**, 37. Available online: https://www.psychiatrictimes.com/view/cannabis-pregnancy-rejoinder-exposition-cautionary-tales (accessed on 10 September 2020).
- 51. Reece, A.S.; Hulse, G.K. Epidemiological Overview of Multidimensional Chromosomal and Genome Toxicity of Cannabis Exposure in Congenital Anomalies and Cancer Development. *Sci. Rep.* **2021**, *11*, 13892. [CrossRef]
- 52. Reece, A.S.; Hulse, G.K. Epidemiological association of cannabinoid- and drug- exposures and sociodemographic factors with limb reduction defects across USA 1989–2016: A geotemporospatial study. *Spat. Spatio-Temporal Epidemiol.* **2022**, *41*, 100480–100490. [CrossRef]

- 53. Phillips, K.T.; Pedula, K.L.; Choi, N.G.; Tawara, K.K.; Simiola, V.; Satre, D.D.; Owen-Smith, A.; Lynch, F.F.; Dickerson, J. Chronic health conditions, acute health events, and healthcare utilization among adults over age 50 in Hawai'i who use cannabis: A matched cohort study. *Drug Alcohol. Depend.* **2022**, 234, 109387. [CrossRef]
- 54. Allen, J.P.; Danoff, J.S.; Costello, M.A.; Hunt, G.L.; Hellwig, A.F.; Krol, K.M.; Gregory, S.G.; Giamberardino, S.N.; Sugden, K.; Connelly, J.J. Lifetime marijuana use and epigenetic age acceleration: A 17-year prospective examination. *Drug Alcohol. Depend.* **2022**, 233, 109363. [CrossRef]
- 55. Hashibe, M.; Morgenstern, H.; Cui, Y.; Tashkin, D.P.; Zhang, Z.F.; Cozen, W.; Mack, T.M.; Greenland, S. Marijuana use and the risk of lung and upper aerodigestive tract cancers: Results of a population-based case-control study. *Cancer Epidemiol. Biomark. Prev.* **2006**, *15*, 1829–1834. [CrossRef] [PubMed]
- 56. Reece, A.S.; Hulse, G.K. Quadruple convergence—Rising cannabis prevalence, intensity, concentration and use disorder treatment. *Lancet Reg. Health* **2021**, *10*, 100245–100246. [CrossRef] [PubMed]
- 57. Manthey, J.; Freeman, T.P.; Kilian, C.; Lopez-Pelayo, H.; Rehm, J. Public health monitoring of cannabis use in Europe: Prevalence of use, cannabis potency, and treatment rates. *Lancet Reg. Health* **2021**, *10*, 100227–200237. [CrossRef] [PubMed]
- 58. Geber, W.F.; Schramm, L.C. Teratogenicity of marihuana extract as influenced by plant origin and seasonal variation. *Arch Int. Pharm. Ther.* **1969**, 177, 224–230.
- 59. Geber, W.F.; Schramm, L.C. Effect of marihuana extract on fetal hamsters and rabbits. *Toxicol. Appl. Pharmacol.* **1969**, 14, 276–282. [CrossRef]
- 60. Graham, J.D.P. Cannabis and Health. In *Cannabis and Health*, 1st ed.; Graham, J.D.P., Ed.; Academic Press: London, UK; New York, NY, USA; San Francisco, CA, USA, 1976; Volume 1, pp. 271–320.
- 61. Substance Abuse and Mental Health Services Administration. National Survey of Drug Use and Health (NSDUH 2018). Available online: https://www.datafiles.samhsa.gov/study/national-survey-drug-use-and-health-nsduh-2018-nid18757# (accessed on 26 April 2020).
- 62. United National Office of Drugs and Crime. World Drug Report 2021; World Health Organization Office of Drugs and Crime, Ed.; United National World Health Organization: Geneva, Switzerland, 2021; Volume 1–5, Available online: https://wdr.unodc.org/wdr2019/index.html (accessed on 10 June 2022).
- 63. Freeman, T.P.; Craft, S.; Wilson, J.; Stylianou, S.; ElSohly, M.; Di Forti, M.; Lynskey, M.T. Changes in delta-9-tetrahydrocannabinol (THC) and cannabidiol (CBD) concentrations in cannabis over time: Systematic review and meta-analysis. *Addiction* **2021**, *116*, 1000–1010. [CrossRef]
- 64. ElSohly, M.A.; Mehmedic, Z.; Foster, S.; Gon, C.; Chandra, S.; Church, J.C. Changes in Cannabis Potency Over the Last 2 Decades (1995-2014): Analysis of Current Data in the United States. *Biol. Psychiatry* **2016**, *79*, 613–619. [CrossRef]
- 65. Chandra, S.; Radwan, M.M.; Majumdar, C.G.; Church, J.C.; Freeman, T.P.; ElSohly, M.A. New trends in cannabis potency in USA and Europe during the last decade (2008-2017). *Eur. Arch. Psychiatry Clin. Neurosci.* **2019**, 269, 5–15. [CrossRef]
- 66. ElSohly, M.A.; Ross, S.A.; Mehmedic, Z.; Arafat, R.; Yi, B.; Banahan, B.F., 3rd. Potency trends of delta9-THC and other cannabinoids in confiscated marijuana from 1980-1997. *J. Forensic Sci.* **2000**, 45, 24–30. [CrossRef]
- 67. Reece, A.S.; Hulse, G.K. Clinical Epigenomics Explains Epidemiology of Cannabinoid Genotoxicity Manifesting as Transgenerational Teratogenesis, Cancerogenesis and Aging Acceleration. *Int. J. Environ. Res. Public Health*, 2023; submitted.
- 68. Yilmaz, D.; Furst, A.; Meaburn, K.; Lezaja, A.; Wen, Y.; Altmeyer, M.; Reina-San-Martin, B.; Soutoglou, E. Activation of homologous recombination in G1 preserves centromeric integrity. *Nature* **2021**, *600*, 748–753. [CrossRef] [PubMed]
- 69. Nahas, G.G. Cannabis Physiopathology Epidemiology Detection; CRC Press: Boca Raton, FL, USA, 1990; Volume 1.
- 70. Nahas, G.G. Keep Off the Grass; Eriksson, P.S., Ed.; Elsevier: Middlebury, VT, USA, 1990; Volume 1, p. 300.
- 71. Reece, A.S.; Hulse, G.K. Congenital Anomaly Epidemiological Correlates of Δ8THC Across USA 2003-2016: Panel Regression and Causal Inferential Study. *Environ. Epigenet.* **2022**, *8*, dvac012. [CrossRef] [PubMed]
- 72. Reece, A.S.; Hulse, G.K. Epidemiology of Δ8THC–Related Carcinogenesis in USA: A Panel Regression and Causal Inferential Study. *Int. J. Environ. Res. Public Health* **2022**, 19, 7726. [CrossRef] [PubMed]
- 73. Reece, A.S.; Hulse, G.K. European Epidemiological Patterns of Cannabis- and Substance- Related Congenital Body Wall Anomalies: Geospatiotemporal and Causal Inferential Study. *Int. J. Environ. Res. Public Health* **2022**, 19, 9027. [CrossRef] [PubMed]
- 74. Reece, A.S.; Hulse, G.K. European Epidemiological Patterns of Cannabis- and Substance-Related Congenital Chromosomal Anomalies: Geospatiotemporal and Causal Inferential Study. *Int. J. Environ. Res. Public Health*, 2022; submitted.
- 75. Reece, A.S.; Hulse, G.K. European Epidemiological Patterns of Cannabis- and Substance- Related Congenital Cardiovascular Anomalies: Geospatiotemporal and Causal Inferential Study. *Environ. Epigenet.* **2022**, *8*, dvac015. [CrossRef]
- 76. Reece, A.S.; Hulse, G.K. European Epidemiological Patterns of Cannabis- and Substance- Related Congenital Neurological Anomalies: Geospatiotemporal and Causal Inferential Study. *Int. J. Environ. Res. Public Health* **2022**, *20*, 441. [CrossRef]
- 77. Reece, A.S.; Hulse, G.K. Effects of Cannabis on Congenital Limb Anomalies in 14 European Nations: A Geospatiotemporal and Causal Inferential Study. *Environ. Epigenet.* **2022**, *8*, dvac016. [CrossRef]
- 78. Reece, A.S.; Hulse, G.K. European Epidemiological Patterns of Cannabis- and Substance- Related Congenital Uronephrological Anomalies: Geospatiotemporal and Causal Inferential Study. *Int. J. Environ. Res. Public Health* **2022**, *19*, 13769. [CrossRef]
- 79. Reece, A.S.; Hulse, G.K. Cannabis- and Substance-Related Epidemiological Patterns of Chromosomal Congenital Anomalies in Europe: Geospatiotemporal and Causal Inferential Study. *Int. J. Environ. Res. Public Health* **2022**, *19*, 11208. [CrossRef]

- 80. Reece, A.S.; Hulse, G.K. Geospatiotemporal and Causal Inferential Study of European Epidemiological Patterns of Cannabis- and Substance- Related Congenital Orofacial Anomalies. *J. Xenobiotics* **2023**, *13*, 42–74. [CrossRef]
- 81. Reece, A.S.; Hulse, G.K. Patterns of Cannabis- and Substance- Related Congenital General Anomalies in Europe: A Geospatiotemporal and Causal Inferential Study. *Pediatr. Rep.* **2023**, *15*, 69–118. [CrossRef]
- 82. Reece, A.S.; Hulse, G.K. Chapter 3: Geospatiotemporal and Causal Inferential Analysis of United States Congenital Anomalies as a Function of Multiple Cannabinoid- and Substance- Exposures: Phenocopying Thalidomide and Hundred Megabase-Scale Genotoxicity. In *Epidemiology of Cannabis: Genotoxicity and Neurotoxicity, Epigenomics and Aging*; Elsevier: New York, NY, USA, 2023; Volume 1, p. 2500, *in press*.
- 83. Reece, A.S.; Hulse, G.K. Cannabis- and Substance-Related Carcinogenesis in Europe: A Lagged Causal Inferential Panel Regression Modelling Study. *Int. J. Environ. Res. Public Health*, 2023; *in press*.
- 84. Shen, H.; Shih, J.; Hollern, D.P.; Wang, L.; Bowlby, R.; Tickoo, S.K.; Thorsson, V.; Mungall, A.J.; Newton, Y.; Hegde, A.M.; et al. Integrated Molecular Characterization of Testicular Germ Cell Tumors. *Cell Rep.* **2018**, 23, 3392–3406. [CrossRef] [PubMed]
- 85. Khwaja, A.; Bjorkholm, M.; Gale, R.E.; Levine, R.L.; Jordan, C.T.; Ehninger, G.; Bloomfield, C.D.; Estey, E.; Burnett, A.; Cornelissen, J.J.; et al. Acute myeloid leukaemia. *Nat. Rev. Dis. Prim.* **2016**, *2*, 16010. [CrossRef] [PubMed]
- 86. Miles, L.A.; Bowman, R.L.; Merlinsky, T.R.; Csete, I.S.; Ooi, A.T.; Durruthy-Durruthy, R.; Bowman, M.; Famulare, C.; Patel, M.A.; Mendez, P.; et al. Single-cell mutation analysis of clonal evolution in myeloid malignancies. *Nature* **2020**, *587*, 477–482. [CrossRef]
- 87. Malouf, C.; Ottersbach, K. Molecular processes involved in B cell acute lymphoblastic leukaemia. *Cell. Mol. Life Sci.* **2018**, 75, 417–446. [CrossRef]
- 88. Mukhopadhyay, B.; Cinar, R.; Yin, S.; Liu, J.; Tam, J.; Godlewski, G.; Harvey-White, J.; Mordi, I.; Cravatt, B.F.; Lotersztajn, S.; et al. Hyperactivation of anandamide synthesis and regulation of cell-cycle progression via cannabinoid type 1 (CB1) receptors in the regenerating liver. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 6323–6328. [CrossRef]
- 89. Heller, J.E.; Baty, D.E.; Zhang, M.; Li, H.; Adler, M.; Ganea, D.; Gaughan, J.; Loftus, C.M.; Jallo, J.I.; Tuma, R.F. The combination of selective inhibition of the cannabinoid CB1 receptor and activation of the cannabinoid CB2 receptor yields improved attenuation of motor and autonomic deficits in a mouse model of spinal cord injury. *Clin. Neurosurg.* **2009**, *56*, 84–92. [CrossRef]
- 90. Greineisen, W.E.; Turner, H. Immunoactive effects of cannabinoids: Considerations for the therapeutic use of cannabinoid receptor agonists and antagonists. *Int. Immunopharmacol.* **2010**, *10*, 547–555. [CrossRef]
- 91. Rieder, S.A.; Chauhan, A.; Singh, U.; Nagarkatti, M.; Nagarkatti, P. Cannabinoid-induced apoptosis in immune cells as a pathway to immunosuppression. *Immunobiology* **2010**, *215*, 598–605. [CrossRef] [PubMed]
- 92. Robinson, R.H.; Meissler, J.J.; Breslow-Deckman, J.M.; Gaughan, J.; Adler, M.W.; Eisenstein, T.K. Cannabinoids inhibit T-cells via cannabinoid receptor 2 in an in vitro assay for graft rejection, the mixed lymphocyte reaction. *J. Neuroimmune. Pharm.* **2013**, *8*, 1239–1250. [CrossRef] [PubMed]
- 93. Alshaarawy, O.; Anthony, J.C. Cannabis smoking and serum C-reactive protein: A quantile regressions approach based on NHANES 2005-2010. *Drug Alcohol. Depend.* **2015**, 147, 203–207. [CrossRef]
- 94. Chandra, L.C.; Kumar, V.; Torben, W.; Vande Stouwe, C.; Winsauer, P.; Amedee, A.; Molina, P.E.; Mohan, M. Chronic administration of Delta9-tetrahydrocannabinol induces intestinal anti-inflammatory microRNA expression during acute simian immunodeficiency virus infection of rhesus macaques. *J. Virol.* 2015, 89, 1168–1181. [CrossRef] [PubMed]
- 95. Eisenstein, T.K.; Meissler, J.J. Effects of Cannabinoids on T-cell Function and Resistance to Infection. *J. Neuroimmune. Pharm.* **2015**, 10, 204–216. [CrossRef] [PubMed]
- 96. Zumbrun, E.E.; Sido, J.M.; Nagarkatti, P.S.; Nagarkatti, M. Epigenetic Regulation of Immunological Alterations Following Prenatal Exposure to Marijuana Cannabinoids and its Long Term Consequences in Offspring. *J. Neuroimmune. Pharm.* **2015**, *10*, 245–254. [CrossRef] [PubMed]
- 97. Chiurchiu, V. Endocannabinoids and Immunity. Cannabis. Cannabinoid Res. 2016, 1, 59–66. [CrossRef]
- 98. Gallily, R.; Yekhtin, Z. Avidekel Cannabis extracts and cannabidiol are as efficient as Copaxone in suppressing EAE in SJL/J mice. *Inflammopharmacology* **2018**, 27, 167–173. [CrossRef]
- 99. Kaplan, B.L.F. Evaluation of Marijuana Compounds on Neuroimmune Endpoints in Experimental Autoimmune Encephalomyelitis. *Curr. Protoc. Toxicol.* **2018**, 75, 11–25. [CrossRef]
- 100. Gowran, A.; Murphy, C.E.; Campbell, V.A. Delta(9)-tetrahydrocannabinol regulates the p53 post-translational modifiers Murine double minute 2 and the Small Ubiquitin MOdifier protein in the rat brain. *FEBS Lett.* **2009**, *583*, 3412–3418. [CrossRef]
- 101. Murphy, S.K.; Itchon-Ramos, N.; Visco, Z.; Huang, Z.; Grenier, C.; Schrott, R.; Acharya, K.; Boudreau, M.H.; Price, T.M.; Raburn, D.J.; et al. Cannabinoid exposure and altered DNA methylation in rat and human sperm. *Epigenetics* **2018**, *13*, 1208–1221. [CrossRef] [PubMed]
- 102. Schrott, R.; Acharya, K.; Itchon-Ramos, N.; Hawkey, A.B.; Pippen, E.; Mitchell, J.T.; Kollins, S.H.; Levin, E.D.; Murphy, S.K. Cannabis use is associated with potentially heritable widespread changes in autism candidate gene DLGAP2 DNA methylation in sperm. *Epigenetics* **2019**, *15*, 161–173. [CrossRef] [PubMed]
- 103. DiNieri, J.A.; Wang, X.; Szutorisz, H.; Spano, S.M.; Kaur, J.; Casaccia, P.; Dow-Edwards, D.; Hurd, Y.L. Maternal cannabis use alters ventral striatal dopamine D2 gene regulation in the offspring. *Biol. Psychiatry* 2011, 70, 763–769. [CrossRef] [PubMed]
- 104. Szutorisz, H.; DiNieri, J.A.; Sweet, E.; Egervari, G.; Michaelides, M.; Carter, J.M.; Ren, Y.; Miller, M.L.; Blitzer, R.D.; Hurd, Y.L. Parental THC exposure leads to compulsive heroin-seeking and altered striatal synaptic plasticity in the subsequent generation. *Neuropsychopharmacology* **2014**, *39*, 1315–1323. [CrossRef] [PubMed]

- 105. Watson, C.T.; Szutorisz, H.; Garg, P.; Martin, Q.; Landry, J.A.; Sharp, A.J.; Hurd, Y.L. Genome-Wide DNA Methylation Profiling Reveals Epigenetic Changes in the Rat Nucleus Accumbens Associated With Cross-Generational Effects of Adolescent THC Exposure. Neuropsychopharmacology 2015, 40, 2993–3005. [CrossRef] [PubMed]
- 106. Szutorisz, H.; Hurd, Y.L. Epigenetic Effects of Cannabis Exposure. Biol. Psychiatry 2016, 79, 586–594. [CrossRef] [PubMed]
- 107. Szutorisz, H.; Hurd, Y.L. High times for cannabis: Epigenetic imprint and its legacy on brain and behavior. *Neurosci. Biobehav. Rev.* **2018**, *85*, 93–101. [CrossRef]
- 108. Ellis, R.J.; Bara, A.; Vargas, C.A.; Frick, A.L.; Loh, E.; Landry, J.; Uzamere, T.O.; Callens, J.E.; Martin, Q.; Rajarajan, P.; et al. Prenatal Δ(9)-Tetrahydrocannabinol Exposure in Males Leads to Motivational Disturbances Related to Striatal Epigenetic Dysregulation. *Biol. Psychiatry* **2021**, *92*, 127–138. [CrossRef]
- 109. Stenchever, M.A.; Kunysz, T.J.; Allen, M.A. Chromosome breakage in users of marihuana. *Am. J. Obs. Gynecol.* **1974**, *118*, 106–113. [CrossRef]
- 110. Leuchtenberger, C.; Leuchtenberger, R. Morphological and cytochemical effects of marijuana cigarette smoke on epithelioid cells of lung explants from mice. *Nature* **1971**, 234, 227–229. [CrossRef]
- 111. Nahas, G.G.; Morishima, A.; Desoize, B. Effects of cannabinoids on macromolecular synthesis and replication of cultured lymphocytes. *Fed. Proc.* **1977**, *36*, 1748–1752. [PubMed]
- 112. Morishima, A.; Henrich, R.T.; Jayaraman, J.; Nahas, G.G. Hypoploid metaphases in cultured lymphocytes of marihuana smokers. *Adv. Biosci.* **1978**, 22–23, 371–376.
- 113. Henrich, R.T.; Nogawa, T.; Morishima, A. In vitro induction of segregational errors of chromosomes by natural cannabinoids in normal human lymphocytes. *Environ. Mutagen* **1980**, *2*, 139–147. [CrossRef]
- 114. Morishima, A. Effects of cannabis and natural cannabinoids on chromosomes and ova. *NIDA Res. Monogr.* **1984**, 44, 25–45. [PubMed]
- 115. Huang, H.F.S.; Nahas, G.G.; Hembree, W.C. Effects of Marijuana Inhalation on Spermatogenesis of the Rat. In *Marijuana in Medicine*; Nahas, G.G., Sutin, K.M., Harvey, D.J., Agurell, S., Eds.; Human Press: Totowa, NJ, USA, 1999; Volume 1, pp. 359–366.
- 116. Mendelson, J.H.; Mello, N.K. Effects of marijuana on neuroendocrine hormones in human males and females. *NIDA Res. Monogr.* **1984**, *44*, 97–114. [PubMed]
- 117. Smith, C.G.; Asch, R.H. Acute, short-term, and chronic effects of marijuana on the female primate reproductive function. *NIDA Res. Monogr.* **1984**, 44, 82–96. [PubMed]
- 118. Hillard, C.J. Endocannabinoids and the Endocrine System in Health and Disease. In *Endocannabinoids*; Pertwee, R.G., Ed.; Springer International Publishing: Cham, Switzerland, 2015; pp. 317–339. [CrossRef]
- 119. Meah, F.; Lundholm, M.; Emanuele, N.; Amjed, H.; Poku, C.; Agrawal, L.; Emanuele, M.A. The effects of cannabis and cannabinoids on the endocrine system. *Rev. Endocr. Metab. Disord.* 2021, 23, 401–420. [CrossRef] [PubMed]
- 120. Borowska, M.; Czarnywojtek, A.; Sawicka-Gutaj, N.; Woliński, K.; Płazińska, M.T.; Mikołajczak, P.; Ruchała, M. The effects of cannabinoids on the endocrine system. *Endokrynol. Pol.* **2018**, *69*, 705–719. [CrossRef]
- 121. Hussein, N.A.E.M.; El-Toukhy, M.A.E.-F.; Kazem, A.H.; Ali, M.E.-S.; Ahmad, M.A.E.-R.; Ghazy, H.M.R.; El-Din, A.M.G. Protective and therapeutic effects of cannabis plant extract on liver cancer induced by dimethylnitrosamine in mice. *Alex. J. Med.* **2014**, *50*, 241–251. [CrossRef]
- 122. Chan, J.Z.; Duncan, R.E. Regulatory Effects of Cannabidiol on Mitochondrial Functions: A Review. Cells 2021, 10, 1251. [CrossRef]
- 123. Olivas-Aguirre, M.; Torres-López, L.; Pottosin, I.; Dobrovinskaya, O. Phenolic Compounds Cannabidiol, Curcumin and Quercetin Cause Mitochondrial Dysfunction and Suppress Acute Lymphoblastic Leukemia Cells. *Int. J. Mol. Sci.* 2020, 22, 204. [CrossRef] [PubMed]
- 124. Ryan, D.; Drysdale, A.J.; Lafourcade, C.; Pertwee, R.G.; Platt, B. Cannabidiol targets mitochondria to regulate intracellular Ca²⁺ levels. *J. Neurosci.* **2009**, *29*, 2053–2063. [CrossRef] [PubMed]
- 125. Winklmayr, M.; Gaisberger, M.; Kittl, M.; Fuchs, J.; Ritter, M.; Jakab, M. Dose-Dependent Cannabidiol-Induced Elevation of Intracellular Calcium and Apoptosis in Human Articular Chondrocytes. *J. Orthop. Res.* **2019**, *37*, 2540–2549. [CrossRef] [PubMed]
- 126. Wu, H.Y.; Huang, C.H.; Lin, Y.H.; Wang, C.C.; Jan, T.R. Cannabidiol induced apoptosis in human monocytes through mitochondrial permeability transition pore-mediated ROS production. *Free Radic. Biol. Med.* **2018**, 124, 311–318. [CrossRef]
- 127. von Greiff, N.; Skogens, L.; Berlin, M.; Bergmark, A. Mortality and Cause of Death-A 30-Year Follow-Up of Substance Misusers in Sweden. Subst. Use Misuse 2018, 53, 2043–2051. [CrossRef]
- 128. Arendt, M.; Munk-Jorgensen, P.; Sher, L.; Jensen, S.O. Mortality among individuals with cannabis, cocaine, amphetamine, MDMA, and opioid use disorders: A nationwide follow-up study of Danish substance users in treatment. *Drug Alcohol. Depend.* **2011**, 114, 134–139. [CrossRef]
- 129. Calabria, B.; Degenhardt, L.; Hall, W.; Lynskey, M. Does cannabis use increase the risk of death? Systematic review of epidemiological evidence on adverse effects of cannabis use. *Drug Alcohol. Rev.* **2010**, 29, 318–330. [CrossRef]
- 130. Callaghan, R.C.; Cunningham, J.K.; Verdichevski, M.; Sykes, J.; Jaffer, S.R.; Kish, S.J. All-cause mortality among individuals with disorders related to the use of methamphetamine: A comparative cohort study. *Drug Alcohol. Depend.* **2012**, 125, 290–294. [CrossRef]
- 131. Davstad, I.; Allebeck, P.; Leifman, A.; Stenbacka, M.; Romelsjo, A. Self-reported drug use and mortality among a nationwide sample of Swedish conscripts—A 35-year follow-up. *Drug Alcohol. Depend.* **2011**, *118*, 383–390. [CrossRef]

- 132. DeFilippis, E.M.; Singh, A.; Divakaran, S.; Gupta, A.; Collins, B.L.; Biery, D.; Qamar, A.; Fatima, A.; Ramsis, M.; Piplas, D.; et al. Cocaine and Marijuana Use among Young Adults Presenting with Myocardial Infarction: The Partners YOUNG-MI Registry. *J. Am. Coll. Cardiol.* 2018, *in press.* [CrossRef]
- 133. Desai, R.; Patel, U.; Sharma, S.; Amin, P.; Bhuva, R.; Patel, M.S.; Sharma, N.; Shah, M.; Patel, S.; Savani, S.; et al. Recreational Marijuana Use and Acute Myocardial Infarction: Insights from Nationwide Inpatient Sample in the United States. *Cureus* 2017, 9, e1816. [CrossRef]
- 134. Fridell, M.; Bäckström, M.; Hesse, M.; Krantz, P.; Perrin, S.; Nyhlén, A. Prediction of psychiatric comorbidity on premature death in a cohort of patients with substance use disorders: A 42-year follow-up. *BMC Psychiatry* **2019**, *19*, 150. [CrossRef]
- 135. Frost, L.; Mostofsky, E.; Rosenbloom, J.I.; Mukamal, K.J.; Mittleman, M.A. Marijuana use and long-term mortality among survivors of acute myocardial infarction. *Am. Heart J.* **2013**, *165*, 170–175. [CrossRef] [PubMed]
- 136. Hser, Y.I.; Kagihara, J.; Huang, D.; Evans, E.; Messina, N. Mortality among substance-using mothers in California: A 10-year prospective study. *Addiction* **2012**, 107, 215–222. [CrossRef] [PubMed]
- 137. Muhuri, P.K.; Gfroerer, J.C. Mortality associated with illegal drug use among adults in the United States. *Am. J. Drug. Alcohol. Abus.* **2011**, *37*, 155–164. [CrossRef]
- 138. Pavarin, R.M.; Berardi, D. Mortality risk in a cohort of subjects reported by authorities for cannabis possession for personal use. Results of a longitudinal study. *Epidemiol. Prev.* **2011**, *35*, 89–93. [PubMed]
- 139. Luzi, S.; Morrison, P.D.; Powell, J.; di Forti, M.; Murray, R.M. What is the mechanism whereby cannabis use increases risk of psychosis? *Neurotox Res.* **2008**, *14*, 105–112. [CrossRef]
- 140. Veling, W.; Mackenbach, J.P.; van Os, J.; Hoek, H.W. Cannabis use and genetic predisposition for schizophrenia: A case-control study. *Psychol. Med.* **2008**, *38*, 1251–1256. [CrossRef]
- 141. Hall, W. The adverse health effects of cannabis use: What are they, and what are their implications for policy? *Int. J. Drug Policy* **2009**, 20, 458–466. [CrossRef]
- 142. Hall, W.; Lynskey, M. The challenges in developing a rational cannabis policy. Curr. Opin. Psychiatry 2009, 22, 258–262. [CrossRef]
- 143. Hall, W.D. Challenges in reducing cannabis-related harm in Australia. Drug. Alcohol. Rev. 2009, 28, 110–116. [CrossRef] [PubMed]
- 144. Henquet, C.; Rosa, A.; Delespaul, P.; Papiol, S.; Fananas, L.; van Os, J.; Myin-Germeys, I. COMT ValMet moderation of cannabis-induced psychosis: A momentary assessment study of 'switching on' hallucinations in the flow of daily life. *Acta Psychiatr. Scand.* **2009**, *119*, 156–160. [CrossRef] [PubMed]
- 145. Borgan, F.; Laurikainen, H.; Veronese, M.; Marques, T.R.; Haaparanta-Solin, M.; Solin, O.; Dahoun, T.; Rogdaki, M.; Salokangas, R.K.; Karukivi, M.; et al. In Vivo Availability of Cannabinoid 1 Receptor Levels in Patients With First-Episode PsychosisImaging Study of Cannabinoid 1 Receptor Levels in First-Episode PsychosisImaging Study of Cannabinoid 1 Receptor Levels in First-Episode Psychosis. *JAMA Psychiatry* 2019, 76, 1074–1084. [CrossRef] [PubMed]
- 146. Fine, J.D.; Moreau, A.L.; Karcher, N.R.; Agrawal, A.; Rogers, C.E.; Barch, D.M.; Bogdan, R. Association of Prenatal Cannabis Exposure With Psychosis Proneness Among Children in the Adolescent Brain Cognitive Development (ABCD) Study. *JAMA Psychiatry* 2019, 76, 762–764. [CrossRef]
- 147. Gobbi, G.; Atkin, T.; Zytynski, T.; Wang, S.; Askari, S.; Boruff, J.; Ware, M.; Marmorstein, N.; Cipriani, A.; Dendukuri, N.; et al. Association of Cannabis Use in Adolescence and Risk of Depression, Anxiety, and Suicidality in Young Adulthood: A Systematic Review and Meta-analysisCannabis Use in Adolescence and Risk of Depression, Anxiety, and Suicidality in Young Adulthood. *JAMA Psychiatry* 2019, 76, 426–434. [CrossRef]
- 148. Ecker, A.H.; Buckner, J.D. Cannabis-Related Problems and Social Anxiety: The Mediational Role of Post-Event Processing. *Subst. Use Misuse* **2018**, *53*, 36–41. [CrossRef]
- 149. Duperrouzel, J.; Hawes, S.W.; Lopez-Quintero, C.; Pacheco-Colon, I.; Comer, J.; Gonzalez, R. The association between adolescent cannabis use and anxiety: A parallel process analysis. *Addict. Behav.* **2018**, *78*, 107–113. [CrossRef]
- 150. Otten, R.; Huizink, A.C.; Monshouwer, K.; Creemers, H.E.; Onrust, S. Cannabis use and symptoms of anxiety in adolescence and the moderating effect of the serotonin transporter gene. *Addict Biol.* **2017**, 22, 1081–1089. [CrossRef]
- 151. Lisboa, S.F.; Gomes, F.V.; Terzian, A.L.; Aguiar, D.C.; Moreira, F.A.; Resstel, L.B.; Guimaraes, F.S. The Endocannabinoid System and Anxiety. *Vitam. Horm.* **2017**, 103, 193–279. [CrossRef]
- 152. Huckins, L.M. Linking cannabis use to depression and suicidal thoughts and behaviours. *Lancet. Psychiatry* **2017**, *4*, 654–656. [CrossRef]
- 153. Smolkina, M.; Morley, K.I.; Rijsdijk, F.; Agrawal, A.; Bergin, J.E.; Nelson, E.C.; Statham, D.; Martin, N.G.; Lynskey, M.T. Cannabis and Depression: A Twin Model Approach to Co-morbidity. *Behav. Genet.* **2017**, *47*, 394–404. [CrossRef] [PubMed]
- 154. Troup, L.J.; Torrence, R.D.; Andrzejewski, J.A.; Braunwalder, J.T. Effects of cannabis use and subclinical depression on the P3 event-related potential in an emotion processing task. *Medicine* **2017**, *96*, e6385. [CrossRef] [PubMed]
- 155. Dierker, L.; Selya, A.; Lanza, S.; Li, R.; Rose, J. Depression and marijuana use disorder symptoms among current marijuana users. *Addict. Behav.* **2018**, *76*, 161–168. [CrossRef] [PubMed]
- 156. Filbey, F.M.; Aslan, S.; Lu, H.; Peng, S.L. Residual Effects of THC via Novel Measures of Brain Perfusion and Metabolism in a Large Group of Chronic Cannabis Users. *Neuropsychopharmacology* **2018**, *43*, 700–707. [CrossRef]
- 157. Bartoli, F.; Crocamo, C.; Carra, G. Cannabis use disorder and suicide attempts in bipolar disorder: A meta-analysis. *Neurosci. Biobehav. Rev.* **2019**, 103, 14–20. [CrossRef]

- 158. Waterreus, A.; Di Prinzio, P.; Badcock, J.C.; Martin-Iverson, M.; Jablensky, A.; Morgan, V.A. Is cannabis a risk factor for suicide attempts in men and women with psychotic illness? *Psychopharmacology* **2018**, 235, 2275–2285. [CrossRef]
- 159. Kimbrel, N.A.; Newins, A.R.; Dedert, E.A.; Van Voorhees, E.E.; Elbogen, E.B.; Naylor, J.C.; Ryan Wagner, H.; Brancu, M.; Workgroup, V.A.M.-A.M.; Beckham, J.C.; et al. Cannabis use disorder and suicide attempts in Iraq/Afghanistan-era veterans. *J. Psychiatr. Res.* 2017, 89, 1–5. [CrossRef]
- 160. Feingold, D.; Rehm, J.; Lev-Ran, S. Cannabis use and the course and outcome of major depressive disorder: A population based longitudinal study. *Psychiatry Res.* **2017**, 251, 225–234. [CrossRef]
- 161. Borges, G.; Benjet, C.; Orozco, R.; Medina-Mora, M.E.; Menendez, D. Alcohol, cannabis and other drugs and subsequent suicide ideation and attempt among young Mexicans. *J. Psychiatr. Res.* **2017**, *91*, 74–82. [CrossRef]
- 162. Lopez-Otin, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. Cell 2013, 153, 1194–1217. [CrossRef]
- 163. Lombard, D.B.; Chua, K.F.; Mostoslavsky, R.; Franco, S.; Gostissa, M.; Alt, F.W. DNA repair, genome stability, and aging. *Cell* **2005**, 120, 497–512. [CrossRef] [PubMed]
- 164. Hadley, E.C.; Lakatta, E.G.; Morrison-Bogorad, M.; Warner, H.R.; Hodes, R.J. The future of aging therapies. *Cell* **2005**, 120, 557–567. [CrossRef] [PubMed]
- 165. Chien, K.R.; Karsenty, G. Longevity and lineages: Toward the integrative biology of degenerative diseases in heart, muscle, and bone. *Cell* **2005**, *120*, 533–544. [CrossRef] [PubMed]
- 166. Kirkwood, T.B. Understanding the odd science of aging. Cell 2005, 120, 437-447. [CrossRef]
- 167. Zimmerman, A.M.; Zimmerman, S.; Raj, A.Y. Effects of Cannabinoids on Spermatogensis in Mice. In *Marijuana and Medicine*, 1st ed.; Nahas, G.G., Sutin, K.M., Harvey, D.J., Agurell, S., Eds.; Humana Press: Totowa, NJ, USA, 1999; Volume 1, pp. 347–358.
- 168. Barker, D.J.B. Fetal and Infant Origins of Adult Disease; BMJ Publishing Group: London, UK, 1992.
- 169. Barker, D.J. Low intelligence and month of birth. Acta Genet. Et Stat. Med. 1966, 16, 383-393. [CrossRef]
- 170. Barker, D.J.; Fall, C.H. Fetal and infant origins of cardiovascular disease. Arch. Dis. Child 1993, 68, 797–799. [CrossRef]
- 171. Barker, D.J. The fetal and infant origins of adult disease. BMJ 1990, 301, 1111. [CrossRef]
- 172. Hill, A.B. The Environment and Disease: Association or Causation? Proc. R. Soc. Med. 1965, 58, 295–300. [CrossRef]
- 173. Rossato, M.; Pagano, C.; Vettor, R. The cannabinoid system and male reproductive functions. *J. Neuroendocrinol.* **2008**, 20 (Suppl. 1), 90–93. [CrossRef]
- 174. Chioccarelli, T.; Cacciola, G.; Altucci, L.; Lewis, S.E.; Simon, L.; Ricci, G.; Ledent, C.; Meccariello, R.; Fasano, S.; Pierantoni, R.; et al. Cannabinoid receptor 1 influences chromatin remodeling in mouse spermatids by affecting content of transition protein 2 mRNA and histone displacement. *Endocrinology* **2010**, *151*, 5017–5029. [CrossRef]
- 175. Rossato, M.; Ion Popa, F.; Ferigo, M.; Clari, G.; Foresta, C. Human sperm express cannabinoid receptor Cb1, the activation of which inhibits motility, acrosome reaction, and mitochondrial function. *J. Clin. Endocrinol. Metab.* **2005**, *90*, 984–991. [CrossRef] [PubMed]
- 176. Avitabile, D.; Magenta, A.; Lauri, A.; Gambini, E.; Spaltro, G.; Vinci, M.C. Metaboloepigenetics: The Emerging Network in Stem Cell Homeostasis Regulation. *Curr. Stem. Cell Res. Ther.* **2016**, *11*, 352–369. [CrossRef] [PubMed]
- 177. Harkany, T.; Horvath, T.L. (S)Pot on Mitochondria: Cannabinoids Disrupt Cellular Respiration to Limit Neuronal Activity. *Cell Metab.* 2017, 25, 8–10. [CrossRef] [PubMed]
- 178. Hebert-Chatelain, E.; Desprez, T.; Serrat, R.; Bellocchio, L.; Soria-Gomez, E.; Busquets-Garcia, A.; Pagano Zottola, A.C.; Delamarre, A.; Cannich, A.; Vincent, P.; et al. A cannabinoid link between mitochondria and memory. *Nature* **2016**, *539*, *555*–*559*. [CrossRef] [PubMed]
- 179. Koch, M.; Varela, L.; Kim, J.G.; Kim, J.D.; Hernandez-Nuno, F.; Simonds, S.E.; Castorena, C.M.; Vianna, C.R.; Elmquist, J.K.; Morozov, Y.M.; et al. Hypothalamic POMC neurons promote cannabinoid-induced feeding. *Nature* **2015**, *519*, 45–50. [CrossRef] [PubMed]
- 180. Chiu, P.; Karler, R.; Craven, C.; Olsen, D.M.; Turkanis, S.A. The influence of delta9-tetrahydrocannabinol, cannabinol and cannabidiol on tissue oxygen consumption. *Res. Commun. Chem. Pathol. Pharmacol.* **1975**, 12, 267–286.
- 181. Batut, P.J.; Bing, X.Y.; Sisco, Z.; Raimundo, J.; Levo, M.; Levine, M.S. Genome organization controls transcriptional dynamics during development. *Science* **2022**, *375*, 566–570. [CrossRef]
- 182. Waddington, C.H. Organizers and Genes; Cambridge University Press: Cambridge, UK, 1940; Volume 1.
- 183. Mkrtchyan, G.V.; Abdelmohsen, K.; Andreux, P.; Bagdonaite, I.; Barzilai, N.; Brunak, S.; Cabreiro, F.; de Cabo, R.; Campisi, J.; Cuervo, A.M.; et al. ARDD 2020: From aging mechanisms to interventions. *Aging* **2020**, *12*, 24484–24503. [CrossRef]
- 184. Lu, Y.; Brommer, B.; Tian, X.; Krishnan, A.; Meer, M.; Wang, C.; Vera, D.L.; Zeng, Q.; Yu, D.; Bonkowski, M.S.; et al. Reprogramming to recover youthful epigenetic information and restore vision. *Nature* **2020**, *588*, 124–129. [CrossRef]
- 185. Schultz, M.B.; Sinclair, D.A. When stem cells grow old: Phenotypes and mechanisms of stem cell aging. *Development* **2016**, 143, 3–14. [CrossRef]
- 186. Takahashi, K.; Yamanaka, S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* **2006**, *126*, 663–676. [CrossRef] [PubMed]
- 187. Ocampo, A.; Reddy, P.; Martinez-Redondo, P.; Platero-Luengo, A.; Hatanaka, F.; Hishida, T.; Li, M.; Lam, D.; Kurita, M.; Beyret, E.; et al. In Vivo Amelioration of Age-Associated Hallmarks by Partial Reprogramming. *Cell* **2016**, *167*, 1719–1733.e12. [CrossRef] [PubMed]

- 188. Yu, J.; Vodyanik, M.A.; Smuga-Otto, K.; Antosiewicz-Bourget, J.; Frane, J.L.; Tian, S.; Nie, J.; Jonsdottir, G.A.; Ruotti, V.; Stewart, R.; et al. Induced pluripotent stem cell lines derived from human somatic cells. *Science* **2007**, *318*, 1917–1920. [CrossRef] [PubMed]
- 189. Gadadhar, S.; Alvarez Viar, G.; Hansen, J.N.; Gong, A.; Kostarev, A.; Ialy-Radio, C.; Leboucher, S.; Whitfield, M.; Ziyyat, A.; Touré, A.; et al. Tubulin glycylation controls axonemal dynein activity, flagellar beat, and male fertility. *Science* **2021**, *371*, eabd4914. [CrossRef] [PubMed]
- 190. Janke, C.; Magiera, M.M. The tubulin code and its role in controlling microtubule properties and functions. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 307–326. [CrossRef] [PubMed]
- 191. Moutin, M.J.; Bosc, C.; Peris, L.; Andrieux, A. Tubulin post-translational modifications control neuronal development and functions. *Dev. Neurobiol.* **2021**, *81*, 253–272. [CrossRef] [PubMed]
- 192. Beh, T.T.; Kalitsis, P. Centromeres and Kinetochores. In Centromeres and Kinetochores; Springer: Berlin/Heidelberg, Germany, 2017.
- 193. Black, B.E. Preface to: Centromeres and Kinetochores. In *Centromeres and Kinetochores*; Springer: Cham, Switzerland, 2017; Volume 1, pp. 5–8.
- 194. Corbett, K.D. Molecular Mechanisms of Spindle Assembly Checkpoint Activation and Silencing. In *Centromeres and Kinetochores*; Black, B.E., Ed.; Springer: Philadelphia, PA, USA, 2017; Volume 1, pp. 1–554.
- 195. French, B.T.; Straight, A.F. The Power of Xenopus Egg Extract for Reconstitution of Centromere and Kinetochore Function. In *Centromeres and Kinetochores*; Black, B.E., Ed.; Springer: Philadelphia, PA, USA, 2017; Volume 1, pp. 1–554.
- 196. Hara, M.; Fukagawa, T. Critical Foundation of the Kinetochore: The Constitutive Centromere—Associated Network (CCAN). In *Centromeres and Kinetochores*; Black, B.E., Ed.; Springer: Philadelphia, PA, USA, 2017; Volume 1, pp. 1–554.
- 197. Hsu, J.M.; Huang, J.; Meluh, P.B.; Laurent, B.C. The yeast RSC chromatin-remodeling complex is required for kinetochore function in chromosome segregation. *Mol. Cell. Biol.* **2003**, 23, 3202–3215. [CrossRef]
- 198. Ryu, H.Y.; Hochstrasser, M. Histone sumoylation and chromatin dynamics. Nucleic. Acids Res. 2021, 49, 6043–6052. [CrossRef]
- 199. Borrell, V.; Cardenas, A.; Ciceri, G.; Galceran, J.; Flames, N.; Pla, R.; Nobrega-Pereira, S.; Garcia-Frigola, C.; Peregrin, S.; Zhao, Z.; et al. Slit/Robo signaling modulates the proliferation of central nervous system progenitors. *Neuron* **2012**, *76*, 338–352. [CrossRef]
- 200. Cardenas, A.; Villalba, A.; de Juan Romero, C.; Pico, E.; Kyrousi, C.; Tzika, A.C.; Tessier-Lavigne, M.; Ma, L.; Drukker, M.; Cappello, S.; et al. Evolution of Cortical Neurogenesis in Amniotes Controlled by Robo Signaling Levels. *Cell* 2018, 174, 590–606.e521. [CrossRef]
- 201. Yeh, M.L.; Gonda, Y.; Mommersteeg, M.T.; Barber, M.; Ypsilanti, A.R.; Hanashima, C.; Parnavelas, J.G.; Andrews, W.D. Robol modulates proliferation and neurogenesis in the developing neocortex. *J. Neurosci.* **2014**, *34*, *5717–5731*. [CrossRef]
- 202. Alpar, A.; Tortoriello, G.; Calvigioni, D.; Niphakis, M.J.; Milenkovic, I.; Bakker, J.; Cameron, G.A.; Hanics, J.; Morris, C.V.; Fuzik, J.; et al. Endocannabinoids modulate cortical development by configuring Slit2/Robo1 signalling. *Nat. Commun.* **2014**, *5*, 4421. [CrossRef] [PubMed]
- 203. Lu, T.; Newton, C.; Perkins, I.; Friedman, H.; Klein, T.W. Cannabinoid treatment suppresses the T-helper cell-polarizing function of mouse dendritic cells stimulated with Legionella pneumophila infection. *J. Pharm. Exp. Ther.* **2006**, 319, 269–276. [CrossRef]
- 204. Newton, C.A.; Chou, P.J.; Perkins, I.; Klein, T.W. CB(1) and CB(2) cannabinoid receptors mediate different aspects of delta-9-tetrahydrocannabinol (THC)-induced T helper cell shift following immune activation by Legionella pneumophila infection. *J. Neuroimmune Pharm.* 2009, 4, 92–102. [CrossRef]
- 205. Shibata, M.; Pattabiraman, K.; Lorente-Galdos, B.; Andrijevic, D.; Kim, S.K.; Kaur, N.; Muchnik, S.K.; Xing, X.; Santpere, G.; Sousa, A.M.M.; et al. Regulation of prefrontal patterning and connectivity by retinoic acid. *Nature* **2021**, *598*, 483–488. [CrossRef] [PubMed]
- 206. Tanzi, R.E.; Bertram, L. Twenty years of the Alzheimer's disease amyloid hypothesis: A genetic perspective. *Cell* **2005**, 120, 545–555. [CrossRef] [PubMed]
- 207. Reece, A.S.; Hulse, G.K. Socioeconomic, Ethnocultural, Substance- and Cannabinoid- Related Epidemiology of Down Syndrome USA 1986-2016: Combined Geotemporospatial and Causal Inference Investigation. *Int. J. Environ. Res. Public Health* 2022, 19, 13340. [CrossRef]
- 208. Horvath, S.; Garagnani, P.; Bacalini, M.G.; Pirazzini, C.; Salvioli, S.; Gentilini, D.; Di Blasio, A.M.; Giuliani, C.; Tung, S.; Vinters, H.V.; et al. Accelerated epigenetic aging in Down syndrome. *Aging Cell* **2015**, *14*, 491–495. [CrossRef]
- 209. Labuhn, M.; Perkins, K.; Matzk, S.; Varghese, L.; Garnett, C.; Papaemmanuil, E.; Metzner, M.; Kennedy, A.; Amstislavskiy, V.; Risch, T.; et al. Mechanisms of Progression of Myeloid Preleukemia to Transformed Myeloid Leukemia in Children with Down Syndrome. *Cancer Cell* 2019, 36, 123–138.e10. [CrossRef]
- 210. Kudlow, B.A.; Kennedy, B.K.; Monnat, R.J., Jr. Werner and Hutchinson-Gilford progeria syndromes: Mechanistic basis of human progeroid diseases. *Nat. Rev. Mol. Cell Biol.* **2007**, *8*, 394–404. [CrossRef]
- 211. Bejaoui, Y.; Razzaq, A.; Yousri, N.A.; Oshima, J.; Megarbane, A.; Qannan, A.; Potabattula, R.; Alam, T.; Martin, G.M.; Horn, H.F.; et al. DNA methylation signatures in Blood DNA of Hutchinson-Gilford Progeria syndrome. *Aging Cell* **2022**, *21*, e13555. [CrossRef] [PubMed]
- 212. Lakatta, E.G. Arterial aging is risky. J. Appl. Physiol. 2008, 105, 1321-1322. [CrossRef] [PubMed]
- 213. Wang, M.; Khazan, B.; Lakatta, E.G. Central Arterial Aging and Angiotensin II Signaling. *Curr. Hypertens. Rev.* **2010**, *6*, 266–281. [CrossRef] [PubMed]

- 214. Jameson, J.L.; Fauci, A.S.; Hauser, S.L.; Longo, D.L.; Jameson, J.L.; Loscalzo, J. (Eds.) *Harrison's Principles of Internal Medicine*, 20th ed.; McGraw Hill: New York, NY, USA, 2018; Volume 1–2, p. 2754.
- 215. Carlson, B.M. Human Embryology and Developmental Biology, 6th ed.; Elsevier: Philadelphia, PA, USA, 2019; Volume 1, p. 506.
- 216. Ungricht, R.; Guibbal, L.; Lasbennes, M.C.; Orsini, V.; Beibel, M.; Waldt, A.; Cuttat, R.; Carbone, W.; Basler, A.; Roma, G.; et al. Genome-wide screening in human kidney organoids identifies developmental and disease-related aspects of nephrogenesis. *Cell Stem Cell* 2022, 29, 160–175.e7. [CrossRef] [PubMed]
- 217. Reece, A.S.; Hulse, G.K. Epigenomic and Other Evidence for Cannabis-Induced Aging Contextualized in a Synthetic Epidemiologic Overview of Cannabinoid-Related Teratogenesis and Cannabinoid-Related Carcinogenesis. *Int. J. Environ. Res. Public Health* **2022**, *19*, 16721. [CrossRef]
- 218. Aldington, S.; Harwood, M.; Cox, B.; Weatherall, M.; Beckert, L.; Hansell, A.; Pritchard, A.; Robinson, G.; Beasley, R. Cannabis use and risk of lung cancer: A case-control study. *Eur. Respir. J.* **2008**, *31*, 280–286. [CrossRef]
- 219. Voirin, N.; Berthiller, J.; Benhaim-Luzon, V.; Boniol, M.; Straif, K.; Ayoub, W.B.; Ayed, F.B.; Sasco, A.J. Risk of lung cancer and past use of cannabis in Tunisia. *J. Thorac. Oncol.* **2006**, *1*, 577–579. [CrossRef]
- 220. Berthiller, J.; Straif, K.; Boniol, M.; Voirin, N.; Benhaim-Luzon, V.; Ayoub, W.B.; Dari, I.; Laouamri, S.; Hamdi-Cherif, M.; Bartal, M.; et al. Cannabis smoking and risk of lung cancer in men: A pooled analysis of three studies in Maghreb. *J. Thorac. Oncol.* 2008, 3, 1398–1403. [CrossRef]
- 221. Zhang, Z.F.; Morgenstern, H.; Spitz, M.R.; Tashkin, D.P.; Yu, G.P.; Marshall, J.R.; Hsu, T.C.; Schantz, S.P. Marijuana use and increased risk of squamous cell carcinoma of the head and neck. *Cancer Epidemiol. Biomark. Prev.* 1999, *8*, 1071–1078.
- 222. Hashibe, M.; Ford, D.E.; Zhang, Z.F. Marijuana smoking and head and neck cancer. *J. Clin. Pharmacol.* **2002**, 42, 103S–107S. [CrossRef]
- 223. Sidney, S.; Quesenberry, C.P., Jr.; Friedman, G.D.; Tekawa, I.S. Marijuana use and cancer incidence (California, United States). *Cancer Causes Control* **1997**, *8*, 722–728. [CrossRef] [PubMed]
- 224. Daling, J.R.; Doody, D.R.; Sun, X.; Trabert, B.L.; Weiss, N.S.; Chen, C.; Biggs, M.L.; Starr, J.R.; Dey, S.K.; Schwartz, S.M. Association of marijuana use and the incidence of testicular germ cell tumors. *Cancer* 2009, 115, 1215–1223. [CrossRef] [PubMed]
- 225. Efird, J.T.; Friedman, G.D.; Sidney, S.; Klatsky, A.; Habel, L.A.; Udaltsova, N.V.; Van den Eeden, S.; Nelson, L.M. The risk for malignant primary adult-onset glioma in a large, multiethnic, managed-care cohort: Cigarette smoking and other lifestyle behaviors. *J. Neuro-Oncol.* 2004, 68, 57–69. [CrossRef]
- 226. Moiche Bokobo, P.; Atxa de la Presa, M.A.; Cuesta Angulo, J. Transitional cell carcinoma in a young heavy marihuana smoker. *Arch. Esp. De Urol.* **2001**, *54*, 165–167.
- 227. Chacko, J.A.; Heiner, J.G.; Siu, W.; Macy, M.; Terris, M.K. Association between marijuana use and transitional cell carcinoma. *Urology* **2006**, *67*, 100–104. [CrossRef]
- 228. Nieder, A.M.; Lipke, M.C.; Madjar, S. Transitional cell carcinoma associated with marijuana: Case report and review of the literature. *Urology* **2006**, *67*, 200. [CrossRef]
- 229. Reece, A.S.; Hulse, G.K. Novel Insights into Potential Cannabis-Related Cancerogenesis from Recent Key Whole Epigenome Screen of Cannabis Dependence and Withdrawal: Epidemiological Comment and Explication of Schrott et al. *Genes* 2022, 14, 32. [CrossRef]
- 230. Reece, A.S.; Hulse, G.K. Response to Chen et.al. Arch. Public Health 2022, 80, 235–236. [CrossRef]
- 231. Barker, D.J. Low intelligence. Its relation to length of gestation and rate of foetal growth. *Br. J. Prev. Soc. Med.* **1966**, 20, 58–66. [CrossRef]
- 232. Reece, A.S.; Hulse, G.K. State Trends of Cannabis Liberalization as a Causal Driver of Increasing Testicular Cancer Rates across the USA. *Int. J. Environ. Res. Public Health* **2022**, *19*, 12759. [CrossRef]
- 233. Schrott, R.; Greeson, K.W.; King, D.; Symosko Crow, K.M.; Easley, C.A.t.; Murphy, S.K. Cannabis alters DNA methylation at maternally imprinted and autism candidate genes in spermatogenic cells. *Syst. Biol. Reprod. Med.* **2022**, *68*, 357–369. [CrossRef] [PubMed]
- 234. Schrott, R.; Modliszewski, J.L.; Hawkey, A.B.; Grenier, C.; Holloway, Z.; Evans, J.; Pippen, E.; Corcoran, D.L.; Levin, E.D.; Murphy, S.K. Sperm DNA methylation alterations from cannabis extract exposure are evident in offspring. *Epigenet. Chromatin* **2022**, *15*, 33. [CrossRef] [PubMed]
- 235. Yang, J.H.; Hayano, M.; Griffin, P.T.; Amorim, J.A.; Bonkowski, M.S.; Apostolides, J.K.; Salfati, E.L.; Blanchette, M.; Munding, E.M.; Bhakta, M.; et al. Loss of epigenetic information as a cause of mammalian aging. *Cell* **2023**, *186*, 305–326.e27. [CrossRef] [PubMed]
- 236. Du, J.; Zheng, L.; Gao, P.; Yang, H.; Yang, W.J.; Guo, F.; Liang, R.; Feng, M.; Wang, Z.; Zhang, Z.; et al. A small-molecule cocktail promotes mammalian cardiomyocyte proliferation and heart regeneration. *Cell Stem Cell* **2022**, 29, 545–558.e13. [CrossRef]
- 237. Warburg, O. On respiratory impairment in cancer cells. Science 1956, 124, 269–270. [CrossRef]

- 238. Li, X.; Yang, Y.; Zhang, B.; Lin, X.; Fu, X.; An, Y.; Zou, Y.; Wang, J.-X.; Wang, Z.; Yu, T. Lactate metabolism in human health and disease. *Signal Transduct. Target. Ther.* **2022**, *7*, 305. [CrossRef]
- 239. Yang, Z.; Yan, C.; Ma, J.; Peng, P.; Ren, X.; Cai, S.; Shen, X.; Wu, Y.; Zhang, S.; Wang, X.; et al. Lactylome analysis suggests lactylation-dependent mechanisms of metabolic adaptation in hepatocellular carcinoma. *Nat. Metab.* 2023, 5, 61–79. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.