



## Cloning and Sequence Analysis of the Spike Gene of Porcine Epidemic Diarrhea Virus Chinju99\*

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**Abstract.** The spike (S) gene of the porcine epidemic diarrhea virus (PEDV) Chinju99 which was previously isolated in Chinju, Korea was cloned and sequenced to aid in the development of genetically engineered vaccines and diagnostic reagents against PEDV. The nucleotide sequence encoding the entire S gene open reading frame (ORF) of Chinju99 was 4152 bases long encoding 1383 amino acids. It consisted of 1001 adenine (24.1%), 849 cytosine (20.4%), 877 guanine (21.1%) and 1425 thymine (34.3%) residues. The Chinju99 S ORF nucleotide sequence was 94.5% homologous with that of the Br1/87 and CV777 strains, respectively. The Chinju99 S protein had 92.8% amino acid identity with that of Br1/87 and CV777, respectively. The amino acid sequence contained 27 potential sites for asparagine (N)-linked glycosylation and there was a stretch of highly hydrophobic residues at position 1325–1350.

**Key words:** nucleotide sequence, PEDV, S gene

### Introduction

Since porcine epidemic diarrhea (PED) was first reported in Belgium [1] and the UK [2] in 1978, it has occurred in many countries and causes economic losses in pig farming [3–6]. Pigs in all age groups are susceptible to the disease and show acute diarrhea and dehydration, leading to death with high mortality, especially up to 90% in 1–2 weeks old piglets [7–9]. PED is clinically and pathologically similar to transmissible gastroenteritis (TGE), which obscures the differentiation of the two diseases. Immunity is,

therefore, important for the prevention and control of the disease. Moreover, maternal antibodies derived from immunized sows are the sole source for immunity to the disease in highly susceptible neonates during the first few weeks after birth [7,9]. However, no effective vaccines have been developed for the mucosal immunity in the gut of pigs where the etiologic agent causes severe enteritis [9].

The etiologic agent, porcine epidemic diarrhea virus (PEDV), belongs to the family *Coronaviridae* of the order *Nidovirales* and has a genome of positive sense, ss-RNA [7,10]. The subgenomic mRNAs, which are transcribed from the genome, produce viral proteins such as the spike (S, 180–220 kDa), membrane (M, 27–32 kDa) and nucleocapsid (N, 55–58 kDa) proteins [10–12]. Recently, Pensaert [9] suggested a molecular mass of 85–135 kDa for the S protein. Among the proteins, S, a glycoprotein peplomer on the viral surface, plays an important role

\*Nucleotide sequence data reported is available in the GenBank database under the accession no. AY167585.

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in the attachment of viral particles to the receptors of host cells with subsequent penetration into the cells by membrane fusion. The S glycoprotein also stimulates induction of neutralizing antibodies in the host [13]. The S gene is, therefore, an important target for cloning and expression in the development of genetically engineered vaccines. Duarte et al. [14] reported a sequence of 1741 nucleotides in the region between the S and N genes of the British, Br1/87 and Belgian, CV777 strains of PEDV. Duarte and Laude [13] also reported a sequence of 4162 nucleotides including the coding region of the Br1/87 S gene. More recently, the complete S gene sequence has been reported within the complete genome sequence of PEDV CV777 [15].

PED occurs frequently in Korea since the virus was first isolated there [6], and developmental efforts should target accurate diagnosis and control of the disease. Studies on the nucleotide sequences of the S gene of Korean PEDV isolates have not yet been reported. In the present study, DNA clones of the full-length S gene open reading frame (ORF) of PEDV isolated in Chinju, Korea (PEDV Chinju99) were constructed. The nucleotide and deduced amino acid sequences of the S gene were determined, and further analyzed and aligned with those of other PEDVs to aid in the production of genetically engineered vaccines and diagnostic reagents.

## Materials and Methods

### *Virus*

PEDV Chinju99, first isolated by the Virology Laboratory of Gyeongsang National University College of Veterinary Medicine, Chinju, Korea from

the intestinal tissues of piglets suffering from severe diarrhea (data not shown), was used for cloning of the S gene. The virus was propagated in Vero cells grown in minimal essential medium (MEM) containing streptomycin (100 µg/ml), penicillin (100 U/ml) and trypsin (10 µg/ml) in a 5% CO<sub>2</sub> incubator at 37°C following the methods of Hofmann and Wyler [16].

### *Extraction of Viral RNA*

The medium was removed at 15 h post-infection at the early stage of cytopathic effects such as rounding, degeneration and syncytia formation. Cells were lysed by Trizol<sup>®</sup> reagent (Invitrogen, USA) at 2 ml per tissue culture flask (25 cm<sup>2</sup>) and homogenized by passing the cell lysate several times through a pipette. Viral RNA was extracted from the homogenate following the manufacturer's suggestions and dissolved in diethyl pyrocarbonate-treated distilled water.

### *Primers used for cDNA Synthesis*

Pairs of sense and antisense primers were designed and aligned based on the nucleotide sequence of the Br1/87 S gene [13] from the GenBank data base (National Center for Biotechnology Information, USA). These primers were used to generate cDNA for the S gene. The nucleotide sequences and relative position of the primers are shown in Table 1 and Fig. 1, respectively.

### *cDNA Synthesis for the S Gene*

Synthesis of the first-strand cDNA for the S gene was carried out by reverse transcription (RT) using

Table 1. Primers used in cDNA synthesis for PEDV S gene

Primers	Nucleotide Sequences	Mers	% GC	Strand
SF1	5'ACGTAAACAAATGAGGTC 3'	18	39.0	+
SF2	5'TACCTTCTACTGTCAGGG 3'	18	50.0	+
SF3	5'ACTCTGGTGCATGGTAC 3'	18	48.6	+
SR1	5'CTGTTTCGTGACTCAGAAG 3'	18	50.0	-
SR2	5'CTCGCTTGAACAGCATAG 3'	18	50.0	-
SR3	5'ACATCACTGCACGTGGAC 3'	18	61.0	-

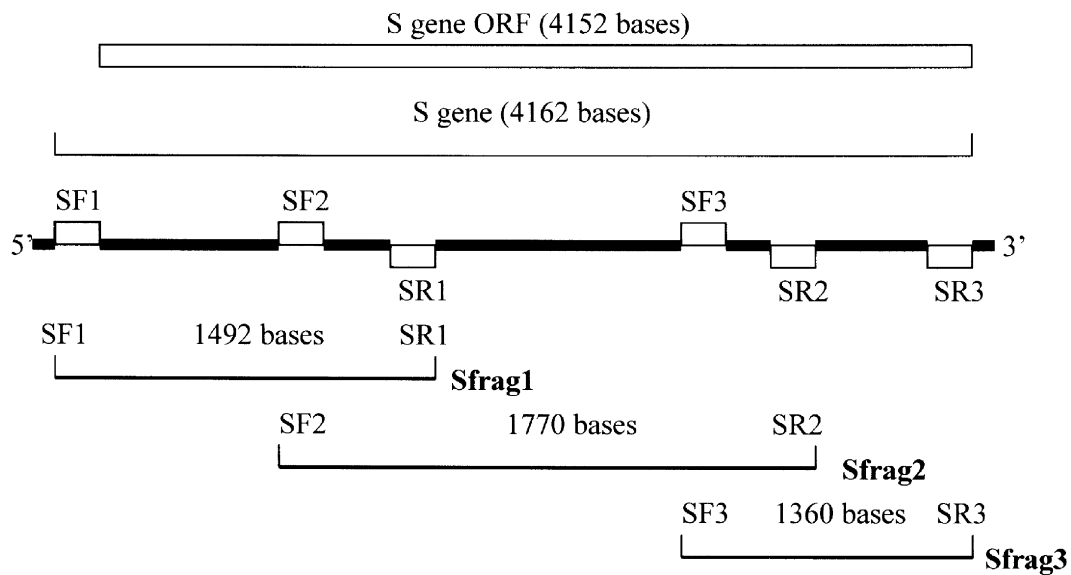


Fig. 1. Construction of cDNA clones for the full-length S gene of PEDV Chinju99 strain by RT-PCR using sense (SF) and antisense (SR) primers: diagrammatic representation of the S gene of viral RNA (solid bar) and S gene ORF (long open rectangle) show primer-binding sites (small open rectangles); three DNA fragments amplified by PCR and cloned into pUC19 vector are denoted as recombinant DNA clones Sfrag1, Sfrag2 and Sfrag3.

Superscript II<sup>®</sup> reverse transcriptase reagent kit (Invitrogen) following the manufacturer's suggestions. The viral RNA was mixed with 1  $\mu$ l of 100 pM of the appropriate antisense primer, 4  $\mu$ l of 5X first-strand buffer, 1  $\mu$ l of 10 mM dNTP mixture, 2  $\mu$ l of 0.1 M DTT, 1  $\mu$ l of RNase inhibitor (40 U/ $\mu$ l), 1  $\mu$ l of reverse transcriptase (200 U/ $\mu$ l) and brought to 20  $\mu$ l with distilled water. The reaction mixture was incubated for 50 min at 42°C, and the reaction was stopped by heat at 70°C for 15 min. RNase H (1 U) treatment was done for 20 min at 37°C to degrade the RNA template.

The ds-cDNA for the S gene was synthesized by polymerase chain reaction (PCR) using a reagent kit (Perkin-Elmer, USA). A 10  $\mu$ l portion of the first-strand cDNA template was added to 5  $\mu$ l of 10X PCR buffer, 4  $\mu$ l of 25 mM MgCl<sub>2</sub>, 1  $\mu$ l of 10 mM dNTP mixture, 1  $\mu$ l of each 100 pM sense and antisense primers, 1  $\mu$ l of *Taq* DNA polymerase (5 U/ $\mu$ l) and brought to 50  $\mu$ l with distilled water. The PCR was carried out in a thermocycler (Perkin-Elmer) following the program of 2 min at 94°C and 30 cycles of 1 min at 94°C, 1 min at 45–50°C depending on the primers and 1 min at 72°C, and a final extension at 72°C for 5 min. The PCR

products were resolved by electrophoresis in 1% agarose gels.

#### Cloning of cDNA

The PCR-generated S gene ds-cDNAs were blunt-ended with Klenow enzyme (2 U) and 1  $\mu$ l of 0.5 mM dNTPs (Invitrogen) in 20- $\mu$ l reaction volume and cloned into the *Sma*I site of pUC19 plasmid DNA by ligation using T4 DNA ligase (1 U) (Invitrogen) [17]. The recombinant plasmid DNAs were transformed into competent *Escherichia coli* DH5 $\alpha$  cells by heat shock for 45 s at 42°C. After adding SOC medium (0.5% yeast extract, 2% tryptone, 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl<sub>2</sub>, 20 mM MgSO<sub>4</sub>, 20 mM glucose), the tube was shaken for 1 h at 220 rpm, 37°C. The transformed cells were plated onto Luria Bertani (LB) agar (Invitrogen) containing ampicillin (50  $\mu$ g/ml), X-gal (40  $\mu$ g/ml) and isopropylthio- $\beta$ -galactoside (20  $\mu$ g/ml) (Invitrogen) and incubated at 37°C, overnight. Transformed colonies were cultured in LB broth with ampicillin (50  $\mu$ g/ml) by shaking at 220 rpm, at 37°C, overnight, and were subjected to DNA extraction by alkaline-lysis, restriction enzyme



Chinju99	CTCAGTATGG CCAAGTCAAG ATTGCACCCA CGGTTACTGG GAATATTAGT ATTCCCACCA ACITTAGTAT GAGTATTAGA ACAGAATATT TACAGCTTTA	2400
Br1/87		2400
Chinju99	CAACACGCCT GTTAGTGTG ATTGTGCTAC ATATGTTTGT AATGGTAACT CTCGGTTGTAA ACAATTACTT ACCCAGTACA CTGCAGCATG TAAGACCATA	2500
Br1/87		2500
Chinju99	GAGTCAGCAT TAGAACTCAG CGCTAGGCTT GAGTCTGTTG AAGTTAACTC TATGCTTACT ATTTCTGAAG AGGCCCTACA GTTAGCTACC ATCAGTTCGT	2600
Br1/87		2600
Chinju99	TTAATGGTGA TGGGATAAC TTTACTAATG TGTTGGGTGT TTCTGTGTAC GATTCTCAA GTGGCAGGGT GATACACGAA AGGTCTTTTA TTGAAGACCT	2700
Br1/87		2700
Chinju99	GCTTTTTAAT AAGTGGTGA CTAATGGCCT TGGTACTGTT GATGAAGACT ATAAGCGCTG TTCTAATGGT CGCTCTGTGG CTGATAAAGT CTGTGCGCAG	2800
Br1/87		2800
Chinju99	TATTACTCTG <u>GTCTCATGGT</u> ACTACCTGGG GTTGTGACG CTGAGAAGCT TCACATGTAT AGTGCCTCTC TCATCGGTGG TATGGCGCTA GGAGGTTTAA	2900
Br1/87		2900
Chinju99	CTGCCGCAGT GGCATTGCCT TTTAGCTATG <u>CTGTTCAAGC</u> GAGACTCAAT TATCTTGCTT TACAGACGGA TGTTTTACAG CGGAACCAGC AACTGCTTGC	3000
Br1/87		3000
Chinju99	TGAGTCTTTT AACTCTGCTA TTGTAATAT AACCTCAGCC TTTGCGAGTG TTAAGAGGC TATCAGTCAA ACTTCTAAGG GTTTGAATAC TGTGGCTCAT	3100
Br1/87		3100
Chinju99	GCGCTACTA AAGTTCAAGA GGTGTTAAT TCGCAGGGTT CAGCTTTGAC TCAACTTACC ATACAGCTGC AACATAACTT CCAAGCCATT TCTAGTTCTA	3200
Br1/87		3200
Chinju99	CTGATGACAT TTAATCCCGA CTGGACATTC TTTACGCCGA TGTTCAGGTT GATGGTCTCA TCACCCGGCAG ATTATCAGCA CTTAATGCTT TTGTTGCTCA	3300
Br1/87		3300
Chinju99	AACCCTCACT AAGTATACTG AGGTTCAGGC TAGCAGGAAG CTAGCACAGC AAAAGGTTAA TGAGTGCCTC AAATCGCAAT CTCAGCGTTA TGGTTTTTGT	3400
Br1/87		3400
Chinju99	GGTGGTGATG GCGAGCACAT TTTCTCTCTT GTACAGGCCG CACCCAGGGC CCTGCTGTTT TTACACACAG TACTTGTACC GGGTGATTTT GTAAATGTTA	3500
Br1/87		3500
Chinju99	TTGCCATCGC AGGCTTATGC GTTAATGGTG ATATTGCCTT GACTCTACGT GAGCCTGGCT TAGTCTTGGT TACGCATGAA CTTCAAACCT ATACTGCGAC	3600
Br1/87		3600
Chinju99	GGAATATTTT GTTTCATGCG GACGTATGTT TGAACCTAGA AACCTACCG TTAGTGATTT TGTTCAAATT CAGAGTTGTG TGGTACCTA TGTCAATCTG	3700
Br1/87		3700
Chinju99	ACTAGCGACC AACTACCAGA TGTAATCCCA GATTACATCG ATGTTAACAA AACACTTGAT GAGATTTTAG CTTCTCTGCC CAATAGAACT GGTCCAAATC	3800
Br1/87		3800
Chinju99	TTCCCCTAGA TGTTTTTAAAT GCCACTTATC TTAATCTCAC TGGTGAAATT GCAGATCTAG AGCAGCGTTC AGAGTCTCTC CGTAATACTA CAGAAGAGCT	3900
Br1/87		3900
Chinju99	CCGAAGTCTT ATATAACAATA TCAACAACAC ACTTGTGAC CTGAGTGGC TCAACCGAGT TGAGACTTAT ATCAAGTGGC CGTGGTGGGT TTGTTGATT	4000
Br1/87		4000
Chinju99	ATTTTTATTG GTCTCATCTT TGTTGTGCA TTATTAGTGT TCTGCTGCAT TTCACGGGT TGTTGTGGAT GCTGCGGTTG CTGCGGTGCT TGTTTTTCAG	4100
Br1/87		4100
Chinju99	GTTGTTGGAG GGGTCCTAGA CTTCAACCTT <u>ACGAAGGTTT</u> TGAAAAGGTC <u>CACGTGCAGT</u> GA	4162
Br1/87		4162

Fig. 2. (Continued)

digestion and electrophoresis through 1% agarose gels for identification of recombinant DNA clones.

#### Nucleotide Sequencing

Nucleotide sequencing was done for the S gene-recombinant DNA clones using Dye Terminator Cycle Sequencing kit (Perkin-Elmer) by the automatic

sequencer (ABI prism 377, Advanced Biotechnologies, USA).

#### Analysis of Sequences of Nucleotides and Amino Acids

The sequences of nucleotides and deduced amino acids were analyzed by ClustalW, version 1.82 using

Chinju99	MRSLIYFLLF	VPVLP <del>TL</del> SLP	QDVYRCSANT	NFR <del>RR</del> FFSKFN	VQAPAVVVLG	GYLPSVKHAG	WYCAADIQTA	SGVHGIFLSH	IRGGHGFEIG	ISQEPFDP	PSG	100																			
Br1/87		W L L		T QST			MNSSS	GTG E		Y DS Q		100																			
CV777		W L L		T QST			MNSSS	GTG E		Y DS Q		100																			
Chinju99	YQLYLHKATN	<u>GNT</u> NATARLR	ICQFPSSkTL	GPTANDVTTG	RNCLFNKAIP	AHMSDGKHSV	VGITWDNDRV	TVFADKIYHF	YLNKDW	SRVA	TNCYNSGGCA	200																			
Br1/87			DN	V		Y R DI					R RRS	200																			
CV777		I	DN	V		Y R DI					R RRS	200																			
Chinju99	MQVYVEPIYY	<u>MLNV</u> TSAGED	GISYQPCTAN	CIGYAANVFA	TESNGHIPEG	FSFNWFLLS	<u>NDSTL</u> FHGKV	VSNQPLL <del>VNC</del>	LWAI	PKIYGL	GHFFSF <del>NQ</del> TM	300																			
Br1/87		T	Y E	T	D		L		L		Q H	300																			
CV777		T	Y E	T	D		L		L		Q H	300																			
Chinju99	DGVCNGATAY	RAPQALRFNI	<u>NDISV</u> ILAEG	SIVLHTALGT	<u>NLSFVCS</u> NSS	DPHKAI	IFSIP	LGATQV	PPYYC	FLKVD	TYNST	VYKFP	PAVLP	TVREIVITKY	400																
Br1/87		AVD E				L A	E				L S			400																	
CV777		AVD E				L A	E				L P			400																	
Chinju99	GDVYVNGFGY	LHLGLLDAVT	<u>INF</u> TGHGTND	DVSGFWTIAS	TNFDALIEV	QATAIQ	RILY	CDDPVS	QLKC	SQV	SFDLDDG	FYPI	SSRNLL	SHEQPI	SFVT	500															
Br1/87			Y	D	V	G S				A						500															
CV777				D	V	G S				A						500															
Chinju99	LPSFNDHSFV	<u>NI</u> TVSAAF <del>GD</del>	SGGANLVASD	TTINGFSSFC	VDRQYITRL	<u>FYN</u> VNSYGY	VSKSQDSNCP	FTLQSV	VNDYL	SFSKFC	VSTS	LLAGACT	IDL			600															
Br1/87			G LSS		F T											600															
CV777			G LSS		F T											600															
Chinju99	FGYPAFGSGV	NYRSLYQFT	KGEWITWAPK	PLNGLLKG <del>GF</del>	MTLDVCTKYT	IYGFKGEGII	<u>TLT</u> NS <del>SL</del> LAG	VYYTSD	SGQL	LAFK	<u>NV</u> ISGA	VYSVTP	CSFS			700															
Br1/87		KLT	L GT	E ITDVS												700															
CV777		KLT	L GT	E ITDVS												700															
Chinju99	EQAAYVND <del>DI</del>	VGVI	<u>SSLS</u> NS	<u>TFN</u> NTRELPG	FFYHSNDGSN	<u>C</u> TEPVLVYSN	IGVCKSGSIG	YVPSQY	GOVK	IAPT	V	TGNIS	<u>I</u> PTNF	SMSIR	TEYLQ	LYNTP	800														
Br1/87																	800														
CV777																	800														
Chinju99	VSDCATYVC	NGNSRCKQLL	TQYTAACKTI	ESALELSARL	ESVEVNSMLT	I	SEALQ	LAT	ISSFNG	DGYN	<u>FTN</u> VLGVS	VY	DSQSGR	VIHE	RSFIED	LLFN	900														
Br1/87				Q							A	PA	VQK	V			900														
CV777				Q							A	PA	VQK	V			900														
Chinju99	KVVTNGLGTV	DEDYKRCNSG	RSVADK	VCAQ	YYSGLM	VLPG	VVDAE	KLHM	Y	SASL	IGGMAL	GGFTA	AAVALP	FSYAVQ	ARLN	YLALQ	TDVLQ	RNQQLL	AESF	1000											
Br1/87			L	V								I	A							1000											
CV777			L	V								I	A							1000											
Chinju99	NSAIGN	<u>IT</u> SA	FASVKEAISQ	TSKGLN	TVAH	ALTKVQ	EVVN	SGQSAL	TQLT	IQLQ	HN	FQAI	SSSTDD	IYSR	LDIL	SADVQV	DRLIT	GRLSA	LNAFVA	QTLT	1100										
Br1/87		E						N	V			I		L							1100										
CV777		E						N	V			I									1100										
Chinju99	KYTEVQASRK	LAQQK	VNECV	KSQSQR	YGFC	GGDGEH	I	FSL	VQAA	PQGLLF	LHTVL	VP	GDF	VNVIA	IAGLC	VNGD	I	AL	TLR	EPGLV	LV	THE	LQTY	TATEYF	1200						
Br1/87														L	E		F								1200						
CV777														L	E		F								1200						
Chinju99	VSSRRMFEP	R	KPTV	SDFVQI	QSCV	VTVN	<u>TS</u> DQLPDVIP	DYID	VN	KTLD	EILAS	LP	NRT	GP	NL	PL	DV	FN	ATY	L	N	L	T	GEI	ADLE	QR	SESL	RNTTEEL	RSL	1300	
Br1/87				E										S																1300	
CV777				E										S																1300	
Chinju99	IYNIN	<u>N</u> ILVD	LEWLN	RVETY	IKW	WWW	WLI	IFIG	L	IFVVS	LLV	FCC	ISTG	CCG	CCG	CCGA	C	FSG	C	WR	GPR	LQ	P	Y	E	G	F	E	KV	HVQ*	1383
Br1/87		N						V	V								C														1383
CV777		N						V	V								C														1383

Fig. 3. Comparison of the amino acid sequence deduced from Chinju99 S gene with that of Br1/87 and CV777 strains: only amino acids of Br1/87 and CV777 which mismatched those of Chinju99 are included; \*, translation termination; 27 potential sites for asparagine (N)-linked glycosylation are underlined.

data available from GenBank and the European Molecular Biology Laboratory (EMBL). S gene nucleotide and amino acid sequences of Chinju99 were compared with Br1/87 [14] (EMBL accession No. Z25483) and CV777 (GenBank accession No. AF353511).

## Results

### Cloning of the S Gene

In the production of ds-cDNA of the Chinju99 S gene, three overlapping DNA fragments were amplified by

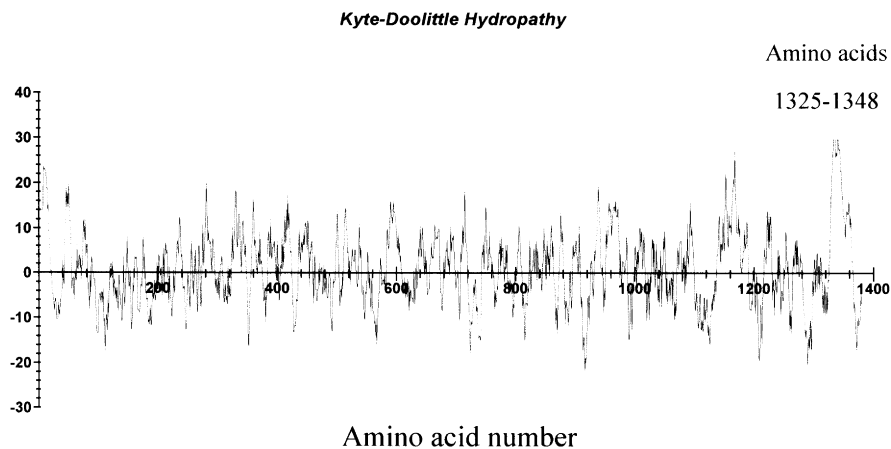


Fig. 4. Hydropathicity of Chinju99 amino acid sequence: a stretch of highly hydrophobic residues was found at position 1325–1348 (>1.6 on the Kyte–Doolittle scale).

RT-PCR using appropriate pairs of sense (SF) and antisense (SR) primers. The DNAs, designated as Sfrag1 (1.5 kb), Sfrag2 (1.8 kb) and Sfrag3 (1.4 kb), were each cloned into pUC19 vector DNAs (Fig. 1), and sequenced.

#### *Analysis of S Gene Nucleotide and Amino Acid Sequences*

The nucleotide sequence encoding the entire Chinju99 S gene was 4162 bases in length and contained a single 4152 bases long ORF starting with an initiator ATG at nucleotide 11 and ending with a terminator TGA at 4160. The sequence GUAAAC, found at 8 nucleotides upstream of the ATG, was considered part of the leader sequence preceding the ORF. The coding region of the gene had 229 nucleotide mismatches compared to Br1/87 and CV777, respectively (Fig. 2). It consisted of 1001 adenine (24.1%), 849 cytosine (20.4%), 877 guanine (21.1%) and 1425 thymine (34.3%) nucleotides, and a GC content of 41.5%. The gene showed 94.5% nucleotide sequence homology to that of Br1/87 and CV777, respectively.

The putative translation product of the PEDV Chinju99 S gene consisted of 1383 amino acids starting at the ATG at nucleotide 11 and ending at the terminator TGA at nucleotide 4160. There were 27 potential asparagine (N)-linked glycosylation sites recognized in the protein. The Chinju99 S protein, had 100 and 99 amino acid mismatches compared to those of Br1/87 and CV777, respectively (Fig. 3).

There was also a stretch of highly hydrophobic residues at position 1325–1348 (>1.6 on the Kyte–Doolittle scale) (Fig. 4). The Chinju99 S protein showed 92.8% amino acid sequence identity with that of Br1/87 and CV777, respectively.

#### **Discussion**

The S gene of the Korean Chinju99 isolate of PEDV was cloned as a series of three overlapping cDNA clones, and the resulting sequence data revealed a single large ORF of 4152 nucleotides which encodes 1383 amino acids to the terminator TGA. Previous studies showed the same lengths for the nucleotide and deduced amino acid sequences of the S gene in Br1/87 [13] and CV777. A single ORF of 4149 nucleotides was identified with the potential to produce a coronavirus S protein [18]. The Chinju99 S gene also had a sequence GUAAAC at 8 nucleotides upstream of the initiator ATG as previously recognized in Br1/87 [13]. This is a common hexameric motif in coronaviruses and similar to the motifs XUA(A/G)AC found adjacent to other PEDV ORFs [12]. The hexameric motifs were suggested as the site for transcription of the subgenomic mRNAs [18].

The Chinju99 S protein contained 27 potential N-linked glycosylation sites and a region of hydrophobic residues at position 1325–1348 was assumed to function as a membrane anchor. Similarly, the

Br1/87 S protein [13] had 29 potential N-linked glycosylation sites and a hydrophobic stretch at 1322–1337. Consequently, the conformational features of the Chinju99 S protein are probably highly conserved with those of Br1/87. Nevertheless, the Chinju99 S gene showed 94.5% nucleotide sequence homology and 92.8% amino acid sequence identity with those of both Br1/87 and CV777, respectively, although the ratios of adenine (24.0–24.1%), cytosine (20.4%), guanine (21.1%) and thymine (34.3–34.4%) were identical or very similar among the three strains. Accordingly, the Chinju99 S protein had only minimal differences in overall structure compared to that of Br1/87 and CV777. Nevertheless the minor differences in the structural features of the S protein could help in elucidating aspects related to both molecular pathogenesis and antigenic structures of PEDV isolates in relation to prevention and control of the disease.

This is the first published report on the nucleotide sequences of the S gene of any Korean PEDV isolate. In the present study, the complete nucleotide sequence of the Chinju99 S gene was determined, and its amino acid composition was analyzed in comparison to other PEDVs. The nucleotide sequence of the Chinju99 S gene can form the basis for further studies on the development of genetically engineered vaccines and diagnostic reagents for PEDV isolates in Korea.

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