

1 **Cold tolerance in two large maize inbred panels adapted to European climates**

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1 **Abstract**

2 Maize (*Zea mays* L.) for northern growing areas requires cold tolerance for extending the  
3 vegetative period. Our objectives were to evaluate two large panels of maize inbred lines  
4 adapted to Europe for cold tolerance and to estimate the effects of cold-related traits on  
5 biomass production. Two inbred panels were evaluated for cold tolerance *per se* and in  
6 testcrosses under cold and control conditions in a growth chamber and under field  
7 conditions. Comparisons of inbreds and groups of inbreds were made taking into account  
8 the SNP-based genetic structure of the panels, and the factors affecting biomass  
9 production were studied. Eight flint and one dent inbreds with diverse origins were the  
10 most cold tolerant. The most cold tolerant dent and flint groups were the Iodent Ph207  
11 and the Northern Flint D171 groups, respectively. The relationships between inbred *per se*  
12 and testcross performance and between controlled and field conditions were low.  
13 Regressions with dry matter yield in the field as dependent variable identified plant height  
14 ( $R^2=0.285$ ) as the main independent variable, followed by quantum efficiency of  
15 photosystem II ( $R^2=0.034$ ) and other traits with minor contributions. Cold tolerance-  
16 related traits had low and negative effects on dry matter yield. Models intending the  
17 prediction of final performance from traits scored in early developmental stages are not  
18 expected to be precise enough for breeding. For improving cold tolerance, inbreds released  
19 from crosses among the No Iodent group and the Northern Flint group may show high  
20 combining ability, as well as between both groups and the Northern Flint D171 group.  
21 **Key words:** maize, cold tolerance, abiotic stress, germplasm.

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1 Maize (*Zea mays* L.) originated in the tropical highlands of America and is currently grown  
2 in northern growing areas all over the world, where temperatures are well below the  
3 optimum for this crop. Such wide adaptation is the result of either selection for a short  
4 vegetation period in order to escape cold stress or improvement of cold tolerance for  
5 surviving under cold conditions. The first strategy is associated with low yields, while the  
6 second should allow long vegetation periods that potentially increase yield (Revilla et al.,  
7 2005; Strigens et al., 2012). Indeed, early sowing of maize allows for a longer vegetation  
8 period that can potentially increase yield and stability, and the probability of escaping  
9 summer drought stress (Kucharik, 2006). This is particularly true in some temperate areas,  
10 where springs are cold and rainy and summers are hot and dry. But early sowing in  
11 temperate areas requires cold tolerance and, consequently, the interest of breeders in cold  
12 tolerance is increasing (Darkó et al., 2011; Frascaroli and Landi, 2013; Revilla et al., 2005;  
13 Strigens et al., 2012, 2013). In the northwest of Spain, a breeding goal would be to advance  
14 maize sowing two weeks, i.e. from May to mid April, because there are no late frosts.  
15 However, in northern areas the gain could be just a few days.

16         The main handicap for breeding programs intending to improve cold tolerance in  
17 maize has been the narrow genetic base for this trait (Greaves, 1996; Revilla et al., 2005).  
18 Rodríguez et al. (2010) evaluated for cold tolerance the largest collection of germplasm so  
19 far published, the European Union Maize Landrace Core Collection (EUMLCC). Their  
20 results were not encouraging because most cold tolerant populations from the EUMLCC  
21 were not more cold tolerant than the checks. Actually, they were similar to the improved  
22 populations and checks already known, including commercial checks and the best cold  
23 tolerant hybrids. Furthermore, their agronomic performance was not at the level of  
24 commercial standards.

25         Apparently classical maize breeding for cold tolerance has reached a ceiling (Revilla  
26 et al., 2005). The incorporation of new techniques, such as molecular markers, has not

1 solved the problem so far (Leipner et al., 2008). Although several studies have identified  
2 QTLs for cold tolerance, most of them were not reliable enough for marker-assisted  
3 selection and were associated with secondary traits such as chlorophyll content or function  
4 (Jompuk et al., 2005; Rodríguez et al., 2008). Actually, the main effect of cold temperatures  
5 is the reduction of chlorophyll synthesis (Rodríguez et al., 2013). However, some QTLs  
6 were consistent when clearly distinct parents of the segregating population under study  
7 were used (Presterl et al., 2007). Genome selection has recently been suggested by Strigens  
8 et al. (2013), who carried out genome-wide association mapping for cold tolerance in a  
9 collection of maize inbred lines. They obtained 19 highly significant association signals,  
10 explaining between 5.7 and 52.5% of the phenotypic variance for early growth and  
11 chlorophyll fluorescence. They propose the use of whole genome prediction approaches  
12 rather than classical marker assisted selection to improve the chilling tolerance of maize.

13         Other major obstacles for breeders are that cold tolerance has large experimental  
14 errors, a strong genotype by environment interaction, and a complex genetic regulation  
15 (Revilla et al., 2000, Strigens et al., 2013). Moreover, evaluations for cold tolerance are not  
16 accurate enough for a precise discrimination of cold tolerance. This is due to two facts.  
17 First, field trials are not reliable because the occurrence of cold temperatures in a concrete  
18 year is not guaranteed, and second, controlled conditions in growth chambers are not  
19 clearly associated to real conditions in the field. Frascaroli and Landi (2013) pointed out  
20 that breeding programs for cold tolerance are efficient for this trait, but may also yield  
21 some undesirable associated response in other agronomic traits. As an example, high cold  
22 tolerance is generally associated with early flowering, short plants or fewer leaves. Also,  
23 several authors have reported a poor relationship between cold tolerance and agronomic  
24 traits, e.g. early vigor is neither positively associated with grain yield (Revilla et al., 2000)  
25 nor with dry matter accumulation (Leipner et al., 2008). Strigens et al. (2012) found weak  
26 associations between early growth and dry matter accumulation, although the association

1 was larger with biomass accumulation before flowering. The associations between early  
2 traits and final yield depend both on the circumstances of the experiments and the  
3 germplasm involved. Therefore, large sets of genotypes thoroughly evaluated should  
4 provide more reliable estimates of the relationships between cold tolerance traits under  
5 controlled and field conditions.

6 Cold tolerance is an important challenge that must be faced with the powerful tools  
7 available nowadays. First of all, the breeding base should be enlarged as much as possible  
8 by screening larger collections of germplasm; besides, the evaluation methods should be  
9 more precise, involving cold and control conditions as well as field trials, and the traits for  
10 which to select should be carefully chosen in order to accurately discriminate tolerant from  
11 susceptible genotypes. Breeders generally assume that European Flints are more cold  
12 tolerant than Corn Belt Dents, a belief that has been experimentally demonstrated to some  
13 extent (Frascaroli and Landi, 2013; Strigens et al., 2013). Several efforts for searching  
14 sources of cold tolerance have been carried out in limited collections of European  
15 germplasm (Lee et al., 2002; Mosely et al., 1984; Revilla et al., 2000; Rodríguez et al., 2010;  
16 Semuguruka et al., 1981; Verheul et al., 1996). Given that most previous reports have faced  
17 the problem by using limited resources, we believe that a global evaluation of large  
18 collections of genotypes for cold tolerance is still lacking. Therefore, the objectives of this  
19 research were to thoroughly evaluate cold tolerance in two large panels of maize inbred  
20 lines adapted to European conditions *per se* and in testcrosses, and to estimate the effects of  
21 cold tolerance-related traits on biomass production.

22

## 1 **Material and Methods**

### 2 *Plant material: inbred lines of the flint and dent panels and their genetic structure*

3 Two panels of maize inbred lines adapted to European conditions, consisting of  
4 306 dent and 292 flint inbred lines, were evaluated *per se* (Appendix 1) and in testcrosses  
5 (Appendices 2 and 3). The panels were built from the collections of Spanish, French, and  
6 German breeders involved in this research. They come from Western Europe and the  
7 USA. The inbreds are public and have been released throughout the history of maize  
8 breeding. The inbreds were chosen based on diversity and adaptation to the range of  
9 European conditions. Although the range of environmental conditions is wide, the  
10 European area represented here has some common characteristics; namely it is temperate  
11 with short growing cycles and cold springs. Seed of the inbreds *per se* was multiplied by  
12 each station and sent to Pontevedra (Spain) for evaluations. The dent inbreds were crossed  
13 to the flint tester UH007 and the flint inbreds to the dent tester F353 in a winter nursery in  
14 2010.

15 We used the genotyping, diversity, and relationship matrices of Rincent et al.  
16 (2012). These authors genotyped the same diversity panels with the Illumina MaizeSNP50  
17 BeadChip described by Ganai et al. (2011) that includes 49,585 SNPs. These authors did  
18 not use the data from individuals or markers with a missing rate above 0.1 and 0.2  
19 respectively, or with a heterozygosity above 0.05 and 0.15, respectively. In total, 261 flint  
20 lines and 261 dent lines passed the genotyping and phenotyping filter criteria after  
21 removing possible contaminations.

### 22 *Growth chamber trials*

23 The cold chamber was built inside a laboratory with modulated panels, isolated with  
24 injected polyurethane. The 598 inbreds *per se* from the flint and dent panels, six checks  
25 (C105, CO109, D152, EA1027, F816, FP1) repeated in both panels, and their testcrosses  
26 were evaluated for cold and for control conditions in consecutive runs of the cold

1 chamber. In each trial entries were grown in a single 20 m<sup>3</sup> growth chamber following a  
2 randomized complete block design with six replications. Each panel (*per se* or in testcrosses)  
3 was evaluated in the chamber for each treatment, but the confounding effects for blocks  
4 were limited because the six repetitions were together. Confounding effects are always  
5 present in this kind of trials because it is not possible to evaluate under cold and warm  
6 conditions in the same space and time. Confounding effects increase the error term and  
7 reduce the power to identify significant effects.

8 Maize seeds were planted in seedbeds filled with sterilized peat (Gramoflor GmbH  
9 & Co. KG, Vechta, Germany) with one kernel per plot (altogether six plants per inbred or  
10 testcross were used in each growth chamber trial). Each seed was sown in a cell with a  
11 surface of 3 cm x 2.5 cm and 5 cm depth; therefore, average distances were 3 cm between  
12 seedlings within each column and 2.5 cm between seedlings within each row. The  
13 experiments were watered after planting; afterwards the trials were watered as needed.  
14 Temperature conditions were set up at 14 °C/14 h light and 8 °C/10 h dark for the cold  
15 experiments and 25 °C/14 h light and 20 °C/10 h dark for the control experiments. The  
16 cold conditions were chosen for screening for cold tolerance alone, removing any other  
17 stress that the seed can find in the field. Cool light was provided by seven VHO (very high  
18 output) fluorescent lamps per shelf with a photosynthetic photon flux (PPF) of 228 μmol  
19 m<sup>-2</sup> s<sup>-1</sup>. Distance between shelf and fluorescent lamps was 0.5 m.

20 In both the inbred *per se* and the testcross trials, data were recorded at the three-leaf  
21 (V3) stage to assure that plants were at the same developmental stage. Four cold-tolerance  
22 related traits were recorded: number of days from sowing to emergence, relative leaf  
23 chlorophyll content (SPAD) using a hand-held CCM-200 Chlorophyll Content Meter  
24 (Opti-Sciences, Tyngsboro, Massachusetts, USA), quantum efficiency of photosystem II  
25 (ΦPSII) recorded using an OS-30p Chlorophyll Fluorometer (Opti-Sciences, , Tyngsboro,  
26 Massachusetts, USA), and dry weight of the testcrosses. For inbreds *per se*, instead of dry

1 weight we scored early vigor using a visual scale from 1=weak plants to 9=vigorous plants.  
2 For the traits recorded at the three-leaf stage, each trait was taken simultaneously for all  
3 plants. Indeed, simultaneous measurements of a trait in several plants produce distortions  
4 as does the measurements of the same trait in different days. We choose the day for  
5 measurements based on the average development of the trial as a whole, as most breeders  
6 do because it is more precise to carry out simultaneous harvests than picking plants from  
7 the plots individually as they reach the appropriate stage.

8

### 9 *Field trials*

10 The main trials were carried out in the growth chamber, with field trials as  
11 references for testing the performance of the flint and dent panel testcrosses in the field.  
12 These field trials were used also for comparing the evaluation under control conditions  
13 with a real control in the field under normal conditions.

14 The 595 testcrosses (306 dents and 289 flints) were evaluated in field trials during  
15 two years (2010 and 2011) in Pontevedra, Spain (42° 24' N, 8° 38' W, 20 m above sea level).  
16 This location has a humid climate with an annual rainfall of about 1600 mm. The soil is a  
17 humic cambisol with a sandy loam texture (53% sand, 28% silt, 18% clay). A previous  
18 analysis showed that the soil had 12.3% moisture, 9% organic matter, and pH=6.6. The  
19 weather in this location during these years was favorable for maize growth; therefore, field  
20 trials were carried out under optimum conditions. The field trials were planted on 19 May  
21 2010 and 12 May 2011. The flint and dent panels were evaluated in adjacent trials following  
22 a modified augmented design. The experimental design for evaluations of the 289 flint  
23 testcrosses involved 17 blocks (eight included the early testcrosses and nine included the  
24 late testcrosses) with 20 entries per block and a total of 340 experimental plots; in each  
25 block 17 entries were unrepeated and three were repeated once elsewhere throughout the  
26 other blocks; the 51 replicated testcrosses were used for estimating the experimental error.



1 Likewise, the 306 dent testcrosses were evaluated in 18 blocks (nine included the early  
2 testcrosses and nine included the late testcrosses) with 20 entries per block and a total of  
3 360 experimental plots; in each block 17 entries were unrepeated and three were repeated  
4 once elsewhere throughout the other blocks; the 54 replicated testcrosses were used for  
5 estimating the experimental error.

6 Each experimental plot consisted of one row with 27 hills per row and two grains  
7 per hill. Rows were spaced 0.80 m apart and hills were spaced 0.14 m apart. Hills were  
8 thinned to one plant, achieving a final plant density of approximately 90,000 plants ha<sup>-1</sup>.  
9 Currently accepted management and cultural practices were used in both trials and trials  
10 were harvested at physiological maturity. We measured percentage of emergence, early  
11 vigor, dry weight of 5-week old plants, percentage of dry matter in 5-week old plants,  
12 relative leaf chlorophyll content and  $\Phi$ PSII, plus the vegetative traits days to silking, days  
13 to pollen shedding, plant height, dry matter yield, and dry matter content. Data were  
14 recorded on 3 plants per plot for traits measured on individual plants. Leaf chlorophyll  
15 content and  $\Phi$ PSII were taken in the central hours of the morning in sunny days.

#### 16 *Statistical analysis*

17 To analyze growth chamber trials, analyses of variance over control and cold  
18 conditions were performed for dent inbreds, flint inbreds, dent testcrosses, and flint  
19 testcrosses separately. The combined analysis of dent and flint inbreds *per se* was made  
20 considering repetitions and inbreds as random effects, while the population effect (flint or  
21 dent) was fixed. The analysis of each panel under a given condition considered repetitions  
22 as random effects while inbreds or testcrosses were considered fixed effects in order to  
23 compare the performance of inbred lines for identifying cold tolerant inbreds. Least  
24 squares means for inbreds and testcrosses were calculated for each trait. The analyses of  
25 variance were made using the Mixed procedure of SAS (SAS Institute, 2008). Mean  
26 comparisons were made for each trait individually with independent cutting levels for

1 global comparisons of cold tolerance; in other words, the inbreds that did not differ  
2 significantly from the best inbred for all traits were considered the most cold tolerant and  
3 conversely, the lines that were not significantly different from the worst inbred for all traits  
4 were considered the most cold susceptible.

5         Analyses of variance for the field trials over years were performed for each trait.  
6 The testcrosses of the flint and dent panels were analyzed separately; the source of  
7 variation ‘years’ was considered random and ‘genotype’ fixed. Least square means were  
8 estimated for each trait. The analyses were made using the GLM procedure of SAS (SAS  
9 Institute, 2008).

10         Correlation analyses between traits were calculated by using the CORR procedure  
11 of SAS. Besides, regression analyses were made by using dry matter yield at harvest or dry  
12 matter yield five weeks after planting (both in the field) as the dependent variables; all other  
13 previously recorded traits were included as the independent variables. For these regression  
14 analyses, we used the REG procedure of SAS with the stepwise method.

15         In order to find a comprehensive method for classifying genotypes as tolerant or  
16 susceptible to cold conditions, principal component analyses were performed for inbreds  
17 *per se* in cold conditions using the least squares means of days to emergence,  $\Phi$ PSII,  
18 chlorophyll content, and early vigor. These analyses were made for both flint and dent  
19 panels after standardizing the traits. In order to use the principal components for  
20 classifying the inbreds as cold tolerant or susceptible, we calculated an index of  
21 susceptibility by modifying PC2, i.e. combining only days to emergence and early vigor.  
22 Therefore, inbreds with scores on  $PC1 \geq 0$  and  $PC2 \leq 0$  are cold tolerant.

23         Rincent et al (2012) used the markers developed from sequences of the founder  
24 lines of the US nested association mapping population (PANZEA SNPs; Gore et al., 2009)  
25 to estimate Nei's index of diversity (Nei, 1978) and relationship coefficients. Nei's index of  
26 diversity of each SNP was calculated and averaged over the genome for estimating

1 genotype diversity in the two panels (Appendix 4). Rincent et al (2012) characterized the  
2 panels using molecular data with the structure analysis with ‘admixture’ (Alexander et al.,  
3 2009) from  $K=2$  to  $K=8$ . One inbred was classified in a group if  $Q_K > 0.60$  for any cluster  
4 for  $K = 2$  and if  $Q_K > 0.50$  for  $K=3, 4, 5, 6, 7,$  and  $8$ ; the other inbreds were classified as  
5 “mixed” (Appendix 4) ( $Q$ =estimated membership coefficients for each inbred in each  
6 cluster,  $K$ =number of clusters). Both dent and flint inbreds were classified in  $K=2, 3, 4, 5,$   
7  $6, 7,$  and  $8$  groups based on the genetic structure and pedigree knowledge (Rincent et al.,  
8 2012), with several possible alternative classifications of inbreds. Differences between  
9 group means were calculated for all classifications. This analysis was conducted only for  
10 inbreds *per se* and cold and control conditions were analyzed separately.

11

## 1 **Results and discussion**

### 2 *Evaluation of inbreds per se: flints vs. dents*

3           In the present study the analyses of variance over cold and control environmental  
4 conditions in the growth chamber confirmed that environments were significantly different  
5 and that the genotype  $\times$  environment interactions were significant ( $P < 0.05$ ) for the four  
6 traits in both flint and dent panels (results not shown). In the combined analyses,  
7 differences among inbreds were significant ( $P < 0.05$ ) for chlorophyll content and early  
8 vigor, and not significant for days to emergence or  $\Phi$ PSII both in the flint and dent panels.  
9 The analyses of variance by panel and growth condition showed that the differences among  
10 dent inbreds were highly significant ( $P < 0.001$ ) for all traits under both cold and control  
11 conditions. The differences among flint inbreds were also significantly different ( $P < 0.001$ )  
12 for all traits except days to emergence with  $P < 0.01$  under cold conditions and  $P < 0.05$   
13 under control conditions.

14           The dent inbreds evaluated *per se* germinated earlier and had a higher early vigor  
15 than the flints, but the flints had more chlorophyll and a higher  $\Phi$ PSII than the dents  
16 (Table 1, Appendix 1). When performances under control and cold conditions were  
17 compared, flints had a larger increase in days to emergence and smaller reductions in  
18 chlorophyll content and  $\Phi$ PSII than dents, while the decrease in early vigor was similar  
19 between dents and flints (Table 1). Flints and dents did not behave consistently as  
20 differentiated groups for cold tolerance. However, according to the literature, European  
21 flints and dents have diverse origins and history, and they are two clearly distinct genetic  
22 groups. Both panels evaluated contain inbreds that are adapted to European conditions;  
23 most inbreds from the dent panel or their parents originated from the US Corn Belt and  
24 were introduced in Europe during the second half of the 20<sup>th</sup> century, while the ancestors  
25 of the flint inbreds probably were introduced along the four preceding centuries. Previous  
26 studies have shown that within the flints, there are at least two main origins of European

1 genotypes (Rebourg et al., 2003; Revilla et al., 2003). First, maize from Central America was  
2 introduced through the south of Spain and was the origin of the Mediterranean maize;  
3 second, several North American flint populations were introduced through the European  
4 Atlantic coast and were the origin of the European flints. While the Mediterranean maize is  
5 not expected to be cold tolerant, the European flints are believed to be more tolerant to  
6 cold conditions because they are more adapted to northern latitudes than the Corn Belt  
7 Dents. Rodríguez et al. (2010) concluded that the European flints had some potential value  
8 for improving cold tolerance of maize. Other authors have found results supporting this  
9 conclusion because the flint kernel phenotype was associated to cold tolerance (Frascaroli  
10 and Landi, 2013). Strigens et al (2013) concluded that flint and dent inbreds adapted to  
11 European conditions have diverse mechanisms underlying that adaptation. Revilla et al.  
12 (1998) pointed out that the origin of a variety in a cold region does not warrant cold  
13 tolerance, because genotypes with short growing cycle escape cold temperatures when  
14 planted late.

#### 15 *Evaluation of inbreds per se: variability within panels*

16       The genetic diversity and the genomic relationship matrix showed that the diversity  
17 was higher in the dent than in the flint panel (Rincent et al. 2012). Most of the coefficients  
18 of similarity between inbreds were low, but there were some pairs of closely related  
19 inbreds. In the present study we made sets of inbreds with close genetic relationships  
20 within each group (see below). There was no consistency for cold tolerance within each set  
21 except for those sets with few inbreds that were all cold susceptible. However, most  
22 inbreds were cold susceptible and, therefore, most sets were also cold susceptible, except  
23 one mixed set that had four cold susceptible inbreds (UHF084, UHF105, UHF070, and  
24 UHF082) and six cold tolerant inbreds (UHF093, UHF098, UHL058, UHF023, UHF091,  
25 and UH006).

1           Based on the genetic structure and pedigree knowledge, several alternative  
2   classifications of inbreds are possible (Appendix 4). Assignments of flint and dent inbreds  
3   to the different groups for scenarios with K=2 to K=8 are shown in Appendix 4. In the  
4   dent panel, the best discriminating ability was obtained for K=6 groups which were  
5   designated as Iodent Ph207, Iodent UH4068, Lancaster Oh43, No Iodent, other No  
6   Iodent F252, and Stiff Stalk. Among these, the Iodent Ph207 group was the most cold  
7   tolerant and the Stiff Stalk group the most cold sensitive (Table 2). Cold tolerance was  
8   similar for the Iodent UH4068 and the Lancaster Oh43 groups.

9           For flint inbreds the results are more clear for K=6 groups which were designated  
10   as FV7, Northern Flint (NF), NF D171, No NF, Southern EC18, and Southern Flint from  
11   open pollinated varieties (Table 2). The most cold tolerant group was NF D171 (except for  
12   chlorophyll content), and the most cold susceptible was No NF; the Southern EC18 group  
13   had the highest chlorophyll content and  $\Phi$ PSII under cold conditions.

14          When looking at individual inbreds, the nine most cold tolerant inbreds (those that  
15   were simultaneously not significantly different from the best inbred for the four traits) were  
16   EV18, UHL058, CH34, UHP024, EC51, F364, FV71, F471, and UHF043 (Table 3). Eight  
17   of them were flint and one was dent, supporting the conclusion that European Flints are  
18   more cold tolerant than Corn Belt Dents. In this outstanding group three inbreds were  
19   from Germany, three from France, two from Northern Spain, and one from Switzerland.  
20   Therefore, European flint material is at large promising for finding sources of tolerance to  
21   cold conditions. Among the 92 inbreds that were not significantly different from the best  
22   inbred for three traits, flints and dents were similar as there were 47 flints and 45 dents.  
23   The inbreds with less than nine days from sowing to germination under cold conditions  
24   were D09, EV23, EZ53, F922, A310, and UHF018, although many others were not  
25   significantly different (Appendix 1). Inbreds with a chlorophyll content above 10 were  
26   EV18, EP1, FV75, EC237, and EP66.  $\Phi$ PSII was over 0.650 for EC237, UN2065, EC248,

1 UHP042, EV18, EC242C, PB57, and F471. Finally, the early vigor score was higher than 5  
2 for EV18, FV353, UH2551, FV335, PHT77, F816, D06, UH6145, CH16.1-295, UHP033,  
3 CH113-379, EC326A, B111, FC1571, ML606, F922, EP74, EP27, F362, C105, and EC237.  
4 Even though the inbreds come from a wide range of latitudes from Spain to Germany,  
5 there are no clear patterns of geographical variability. However, if we classify the inbreds of  
6 Appendix 1 in five groups (those with high performance for 4, 3, 2, 1, and 0 traits) most of  
7 the Spanish or French inbreds are in the group with 1 outstanding trait while most of the  
8 German inbreds are in the group with 2 outstanding traits, which suggests natural selection  
9 for adaptation to cold environments in Germany during inbred development. On the other  
10 hand, most inbreds with high chlorophyll content or high  $\Phi$ PSII were from Spain, perhaps  
11 as a consequence of the traditional focus on selection for early vigor and dark green color  
12 in northern Spain.

### 13 *Evaluation of Inbreds per se: principal component analysis*

14 The principal component analyses for both panels evaluated under cold conditions  
15 identified two principal components (PC) explaining 50% and 29% of the variability,  
16 respectively. PC1 had a negative contribution for days to emergence (eigenvector = -0.33)  
17 and a positive contribution for chlorophyll (0.55),  $\Phi$ PSII (0.56), and early vigor (0.53). PC2  
18 had a positive contribution for days to emergence (eigenvector = 0.73), chlorophyll (0.42),  
19 and  $\Phi$ PSII (0.38), and a negative effect for early vigor (-0.38). Therefore, PC1 is an index  
20 of cold tolerance while PC2 represents plants that grow less and slower in cold conditions  
21 than in normal conditions, but with more chlorophyll and a higher photosynthetic  
22 efficiency.

23 In the principal components, inbreds with scores on  $PC1 \geq 0$  and  $PC2 \leq 0$  are cold  
24 tolerant (Figure 1, Appendix 5). Considering inbreds with  $PC1 \geq 2$  and  $PC2 \leq -1$ , the most  
25 cold tolerant flint inbreds were EV18, UHL058, F471, UHL048, EC51, F364, CO255,  
26 CH34, UH006, H113-379, UHF093, and UHF091, and the most cold tolerant dent inbreds

1 were UHP024, LH85, EP74, FV335, PHT77, UH2551, EC140, UHP033, EZ19, and  
2 Pa374. These selected groups based on the principal component analysis agreed reasonably  
3 with the previous selection based on mean comparisons among groups ( $K=6$ ), although  
4 the agreement between both criteria was better for the flints than for the dents.  
5 Furthermore, the dent inbreds had on average lower scores in PC1 than the flint inbreds;  
6 therefore, we expect that dents would be more cold susceptible than flints.

7         Among the inbreds of these panels, some were included in large flint and dent half  
8 sib panels (Bauer et al., 2013). Some of the parents of the dent half sib panel were cold  
9 tolerant based on our present data (Table 3), namely the dent inbreds D06, Mo17, and  
10 UH304, and the flint inbreds UH007, D152, and UH006. Considering that the common  
11 parent of the flint half sib panel (UH007) was cold tolerant, the half sib panels provide  
12 valuable material for studying the genetics of cold tolerance in segregating populations.

13         Combining the information from the groups and the individual inbreds, we found  
14 that among the nine best inbreds (Table 3) the most cold tolerant dent inbred (UHP024)  
15 belongs to the most cold tolerant group (Iodent Ph207) (Appendix 4). On the other hand,  
16 among the eight most cold tolerant flint inbreds, only two (UHL058 and UHF043) belong  
17 to the most cold tolerant group (Northern Flint D171), while CH34 and F364 are  
18 Northern Flint, EV18, FV71, and F471 come from southern open pollinated varieties, and  
19 EC51 belongs to the Southern EC18 group. However, differences among groups for cold  
20 tolerance are not very clear and both cold tolerant and non-tolerant lines exist in most  
21 groups. Considering the most cold tolerant flint and dent inbreds based on the principal  
22 component analyses, five out of the 12 flint inbreds belong to the Northern Flint D171  
23 group (UHL058, UHL048, UH006, UHF093, and UHF091), three to Northern Flint  
24 (F364, CH34, and CH113-379), two to the Southern open pollinated varieties (EV18 and  
25 F471), one to the Southern EC18 group (EC51) and one to the group FV7. Among the  
26 dent inbreds, only two (UHP024 and UHP033) of the most cold tolerant 10 inbreds belong



1 to the most cold tolerant group (Iodent Ph207) while the other eight cold tolerant inbreds  
2 (LH85, EP74, FV335, PHT77, UH2551, EC140, EZ19, and Pa374) belong to the cold  
3 susceptible group No Iodent. The concordance between the different analyses is better for  
4 the flint than for the dent inbreds. These lacks of agreement suggest that the most efficient  
5 way for identifying cold tolerant genotypes is the comparison among genotypes *per se*  
6 itself.

#### 7 *Evaluation of testcrosses*

8         The analysis of variance for testcrosses combined over panels and environments in  
9 the growth chamber revealed significant differences among testcrosses for chlorophyll and  
10 biomass in the V3-stage. The genotype  $\times$  environment interaction was not significant for  
11 days to emergence and for  $\Phi$ PSII in both dent and flint panels (results not shown). For  
12 chlorophyll content, both the differences between testcrosses and the genotype  $\times$   
13 environment interaction were significant in both panels. Finally, for biomass in the V3-  
14 stage, differences among testcrosses were significant but the genotype  $\times$  environment  
15 interaction was not significant in both panels.

16         Separate analyses of variance for each panel and environmental condition showed  
17 significant differences for chlorophyll content and for biomass in the V3-stage in all cases  
18 (results not shown). Differences were significant for  $\Phi$ PSII in all cases except for the flint  
19 panel in control conditions. For days to emergence, differences were significant only under  
20 cold conditions for both panels. Differences between inbreds *per se* were more often  
21 significant than between testcrosses, probably because of reduced genetic variance in  
22 testcrosses compared to inbreds. Besides, the flint tester UH007 was also evaluated as  
23 inbred *per se* showing a good performance under cold conditions except for chlorophyll  
24 content. Therefore, it was difficult to find differences between testcrosses in the dent  
25 panel. The dent tester F353 was not evaluated *per se* so we do not know its performance

1 under cold conditions. Furthermore, as both panels were crossed to different testers,  
2 comparisons between flints and dents are not possible.

3 In the cold chamber, the testcross with highest cold tolerance was EC35G × F353  
4 and did not significantly differ ( $P < 0.05$ ) from the best testcrosses for any of the four traits  
5 (Table 4). EC35G was also among the inbreds with high cold tolerance *per se* (Table 3).  
6 There were other inbreds that were cold tolerant both *per se* and in testcrosses, namely  
7 CH34, CH16.1-295, UHP017, and F670 (Tables 2 and 3). However, most inbreds with  
8 high cold tolerance *per se* did not produce a cold tolerant testcross and vice versa. There has  
9 been some controversy on the issue of predicting hybrid cold tolerance from inbred  
10 performance. Maryam and Jones (1983) stated that hybrid performance could be predicted  
11 from their parents, Hodges et al. (1997) found that it is not possible to reliably predict  
12 hybrid cold tolerance from the parents' performance, and Revilla et al. (2000) stated that  
13 their results partially support the notion that hybrid cold tolerance can be predicted from  
14 the performance of the inbred parents. Presterl et al. (2007) found consistent QTLs for  
15 cold tolerance in inbreds and their testcrosses, suggesting that cold tolerance of inbreds and  
16 hybrids was genetically associated. This strongly depends on the genotypes evaluated, the  
17 testers, and the methods being used. Previous reports have shown that it is not always  
18 possible to reliably predict hybrid cold tolerance from inbred performance (Revilla et al.,  
19 2005). The inheritance of cold tolerance is complex and variable. For instance, McConnell  
20 and Gardner (1979) found epistatic, additive, and dominance gene effects for germination  
21 under cool conditions, and mainly additive and dominance effects for seedling vigor in  
22 crosses among three warm-season and three cool-season inbreds. Eagles (1982) found  
23 additive and dominance effects for rate of seedling growth, and Revilla et al. (2000)  
24 concluded that the genetic regulation of cold-tolerance traits conformed to an additive-  
25 dominance model in a diallel among European flints.

1           We evaluated testcrosses from the flint and the dent panels in separate but adjacent  
2 field trials for two years in the field. In the flint panel, differences were not significant  
3 among testcrosses and the year  $\times$  testcross interaction was not significant for emergence,  
4 early vigor, dry weight of 5-week old plants, dry matter content in 5-week old plants,  
5  $\Phi$ PSII, days to silking, and days to pollen shedding (data not shown). Differences between  
6 flint testcrosses were significant for leaf chlorophyll content, plant height, dry matter yield,  
7 and dry matter content. The year  $\times$  testcross interaction was only significant for plant  
8 height in the flint panel. In the dent panel, differences between testcrosses were not  
9 significant for percentage of emergence, dry weight of 5-week old plants dry matter content  
10 in 5-week old plants,  $\Phi$ PSII, days to silking, and days to pollen shedding. Differences  
11 between dent testcrosses were significant for early vigor, leaf chlorophyll content, plant  
12 height, dry matter yield, and dry matter content. The year  $\times$  testcross interaction was not  
13 significant for dent testcrosses. All testcrosses were evaluated under favorable conditions in  
14 the field. The weather conditions were fine for growth at early stages in both years and as a  
15 consequence, testcrosses did not show significant differences for early traits such as  
16 percentage of emergence or dry weight of 5-week old plants. Although field conditions are  
17 unpredictable, evaluations for cold tolerance could be more informative when sown earlier  
18 or in cooler environments.

#### 19 *Relationships among traits*

20           Simple correlations between traits were calculated using means of inbreds and  
21 testcrosses separately for both panels and all environments. Most correlations were low and  
22 only the significant correlations with values above 0.5 are shown and discussed here.  
23 Highly significant ( $P < 0.01$ ) correlations above 0.5 were detected for chlorophyll content  
24 and  $\Phi$ PSII of inbreds in cold conditions (0.61), early vigor and dry matter in 5-week old  
25 plants of testcrosses in the field (0.52), days to pollen shedding and silking of testcrosses in  
26 the field (0.95), and plant height and dry matter yield of testcrosses in the field (0.53).

1 Analyzing separately flint and dent panels, correlations were above 0.5 for chlorophyll  
2 content and  $\Phi$ PSII of inbreds in cold conditions (flint: 0.56, dent: 0.64), early vigor and dry  
3 matter in 5-week old plants of testcrosses in the field (flint: 0.55, dent: 0.50) and also for  
4 testcrosses under control conditions but only in the flint panel (0.58), days to pollen and  
5 silking of testcrosses in the field (flint: 0.94, dent: 0.92), chlorophyll content and dry matter  
6 content in 5-week old plants of testcrosses in control conditions (0.58 in flint) and in cold  
7 conditions (0.50 in flint), and dry matter content in 5-week old plants of testcrosses in cold  
8 and in control conditions (0.51 in flint). Within the dent panel, the only noteworthy  
9 correlation was between early vigor and percentage of emergence of inbreds in cold  
10 conditions (-0.52). None of the correlations for any testcross trait measured in the field and  
11 under cold conditions was above 0.5, showing that the evaluations in the growth chamber  
12 were not clearly associated with performance in the field. However, evaluations in the field  
13 were closer to optimum than to cold conditions and the evaluation of testcrosses provides  
14 limited opportunities for differentiating genotypes in the cold chamber due to reduced  
15 genetic variance. Several authors have shown that correlations between performance under  
16 cold conditions and in the field were positive when growing conditions in the field were  
17 colder than in our experiments (Hodges et al., 1995; Bhosale et al., 2007).

18 In order to check the effect of the early traits on dry matter yield, regression  
19 analyses were carried out considering dry matter yield of testcrosses in the field as the  
20 dependent variable and the traits recorded in the growth chamber as the independent  
21 variables. Most of the latter had significant effects on dry matter yield and were consistent  
22 over panels in the field, and also in the control and under cold conditions, although only  
23 chlorophyll content explained more than 5% of the variability (data not shown). When the  
24 dependent variable was early dry weight, the only trait with a relevant significant effect was  
25 early vigor that explained around 27% of the variation (data not shown).

1 Regression analyses were carried out considering dry matter yield of testcrosses in  
2 the field as the dependent variable and the rest of traits as the independent variables.  
3 Multiple regression with a stepwise selection method for both panels and considering  
4 inbreds and testcrosses as independent variables revealed that the main factor affecting dry  
5 matter yield was plant height ( $R^2=0.285$ ) followed by  $\Phi$ PSII ( $R^2=0.034$ ) and six other traits  
6 with minor contributions (Table 5). All significant variables had a positive coefficient of  
7 regression on dry matter yield, except dry weight at 5 weeks and days to emergence under  
8 cold conditions. When the same analysis was made for the dent panel, plant height  
9 ( $R^2=0.200$ ) was again the main factor affecting dry matter yield, the number of significant  
10 variables was smaller and the signs of the coefficients were the same for the common  
11 variables. The analysis of the flint panel showed a similar result concerning the  
12 predominance of plant height ( $R^2=0.220$ ) and some other variables with smaller effects;  
13 one of which was early vigor under cold conditions, that had a negative coefficient of  
14 regression on dry matter yield. The effect of plant height on dry matter yield is well known  
15 and the other significant traits were quite consistent over the two panels. Among the cold  
16 tolerance-related traits, only days to emergence and early vigor under cold conditions were  
17 included in the final model. Both, as well as dry weight at 5 weeks in the field, had a  
18 negative coefficient of regression. The other cold tolerance traits ( $\Phi$ PSII and chlorophyll  
19 content) had significant positive effects on dry matter yield, but the proportion of variance  
20 explained was very low (Table 5). According to other authors, the effects of cold  
21 temperatures on final yield is due to leaf size rather than to leaf function (Louarn et al.,  
22 2008), which is not in agreement with our results because early vigor in cold conditions had  
23 a negative effect on dry matter yield, and high  $\Phi$ PSII values had a positive effect. Negative  
24 effects of cold tolerance-related traits on dry matter yield are not surprising because our  
25 experience shows that when we improve either early growth or early vigor, we obtain  
26 smaller plants with less dry matter yield. Based on this, both size and function should be

1 taken into consideration as cold-related factors affecting plant growth, as the same authors  
2 concluded later (Louarn et al., 2010). Our results indicate, however, that predictive models  
3 based on plant performance under cold conditions cannot explain large proportions of the  
4 variance. This might be due to the low differentiation among testcrosses in the field under  
5 optimal conditions. When more clearly distinct genotypes are used and biomass is  
6 measured under cold conditions, the results can differ (Presterl et al., 2007). Contrarily,  
7 Frascaroli and Landi (2013) concluded that inbred performance could be used to predict  
8 testcrosses germination measured as the difference between cold and control conditions,  
9 although most previous studies have stated that the ability to predict hybrid performance  
10 from inbred value was limited. Certainly, the relationship between inbreds and testcrosses  
11 for cold tolerance depends on the genotypes, the testers used, and the environments  
12 involved.

### 13 *Conclusions*

14         There is large variability for cold tolerance among the inbred lines adapted to  
15 European environments. Some of the inbreds investigated in our study can be used as  
16 sources of cold tolerance in breeding populations for improving cold tolerance and for  
17 further genetic studies. On the other hand, some of the traits related to the performance of  
18 young plants had significant negative, though small, effects on dry matter yield of adult  
19 plants.

20         For breeding purposes, two groups of cold tolerant inbreds can be suggested as  
21 base germplasm, namely the groups Northern Flint and No Iodent, particularly the  
22 Northern Flint D171 group with UHF043, UHL058, UHL048, UH006, UHF093, and  
23 UHF091, and the No Iodent group with LH85, EP74, FV335, PHT77, UH2551, EC140,  
24 EZ19, and Pa374. These two groups could yield second cycle inbreds with high combining  
25 ability that could also combine favorably with the cold tolerant inbreds UHP024, UHP033,  
26 and D171.

1

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9

## 1 **References**

- 2 Alexander, D.H., J. Novembre, and K. Lange. 2009. Fast model-based estimation of  
3 ancestry in unrelated individuals. *Gen. Res.* 19:1655–1664.
- 4 Bhosale, S.U., B. Rymen, G.T.S. Beemster, A.E. Melchinger, and J.C. Reif. 2007. Chilling  
5 tolerance of central European maize lines and their factorial crosses. *Ann. Bot.*  
6 100:1315–1321.
- 7 Bauer, E., M. Falque, H. Walter, C. Bauland, C. Camisan, L. Campo, N. Meyer, N. Ranc,  
8 R. Rincent, W. Schipprack, V. Wimmer, T. Altmann, P. Flament, A.E. Melchinger,  
9 M. Menz, J. Moreno-González, M. Ouzunova, P. Revilla, A. Charcosset, O.C.  
10 Martin, C.C. Schön. 2013. Intraspecific variation of recombination rate in maize.  
11 *Genome Biology* 14:R103.
- 12 Darkó, É., J. Fodor, S. Dulai, H. Ambrus, A. Szenzenstein, Z. Kira, and B. Barnaba. 2011.  
13 Improved cold and drought tolerance of doubled haploid maize plants selected for  
14 resistance to prooxidant tert-butyl hydroperoxide. *J. Agron. Crop. Sci.* 197: 454–465.
- 15 Eagles, H.A. 1982. Inheritance of emergence time and seedling growth at low temperatures  
16 in four lines of maize. *Theor. Appl. Genet.* 62:81–87.
- 17 Frascaroli, E., and P. Landi. 2013. Divergent selection in a maize population for  
18 germination at low temperature in controlled environment: study of the direct  
19 response, of the trait inheritance and of correlated responses in the field. *Theor.*  
20 *Appl. Genet.* 126:733–746.
- 21 Ganal, M.W., G. Durstewitz, A. Polley, A. Bérard, E.S. Buckler, A. Charcosset, J.D. Clarke,  
22 E.-M. Graner, M. Hansen, J. Joets, M.-C. Le Paslier, M.D. McMullen, P. Montalent,  
23 M. Rose, C.-C. Schön, Q. Sun, H. Walter, O.C. Martin, and M. Falque. 2011. A large  
24 maize (*Zea mays* L.) SNP genotyping array: development and germplasm genotyping,  
25 and genetic mapping to compare with the B73 reference genome. *PLoS ONE*  
26 6:e28334.



- 1 Gore, M.A., J.-M. Chia, R.J. Elshire, Q. Sun, E.S. Ersoz, B.L. Hurwitz, J.A. Peiffer, M.D.  
2 McMullen, G.S. Grills, J. Ross-Ibarra, D.H. Ware, and E.S. Buckler. 2009. A first-  
3 generation haplotype map of maize. *Science* 326:1115–1117.
- 4 Greaves, J.A. 1996. Improving suboptimal temperature tolerance in maize - the search for  
5 variation. *J. Exp. Bot.* 47:307–323.
- 6 Hodges, D.M., C.J. Andrews, D.A. Johnson, R.I. Hamilton. 1997. Sensitivity of maize  
7 hybrids to chilling and their combining abilities at two developmental stages. *Crop*  
8 *Sci.* 37:850–856.
- 9 Hodges, D.M., R.I. Hamilton, and C. Charest. 1995. A chilling response test for early  
10 growth phase maize. *Agron. J.* 87:970–974.
- 11 Jompuk, C., Y. Fracheboud, P. Stamp, and J. Leipner. 2005. Mapping of quantitative trait  
12 loci associated with chilling tolerance in maize (*Zea mays* L.) seedlings grown under  
13 field conditions. *J. Exp. Bot.* 56:1153–63.
- 14 Kucharik, C.J. 2006. A multidecadal trend of earlier corn planting in the central USA.  
15 *Agron. J.* 98:1544–1550.
- 16 Lee, E.A., M.A. Staebler, and M. Tollenaar. 2002. Genetic variation in physiological  
17 discriminators for cold tolerant-early autotrophic phase of maize development. *Crop*  
18 *Sci.* 42:1919–1929.
- 19 Leipner, J., C. Jompuk, K. Camp, P. Stamp, Y. Fracheboud. 2008. QTL studies reveal little  
20 relevance of chilling-related seedling traits for yield in maize. *Theor. Appl. Genet.*  
21 116:555–562.
- 22 Louarn, G., K. Chenu, C. Fournier, B. Andrieu, and C. Giauffret. 2008. Relative  
23 contributions of light interception and radiation use efficiency to the reduction of  
24 maize productivity under cold temperatures. *Funct. Plant. Biol.* 35:885–899.

- 1 Louarn, G., B. Andrieu, and C. Giauffret. 2010. A size-mediated effect can compensate for  
2 transient chilling stress affecting maize (*Zea mays*) leaf extension. *New Phytol.*  
3 187:106–118.
- 4 Maryam, B., and D.A. Jones. 1983. The genetics of maize (*Zea mays* L.) growing at low  
5 temperatures. I. Germination of inbred lines and their F<sub>1</sub>s. *Euphytica* 32:535–542
- 6 McConnell, R.L., and C.O. Gardner. 1979. Inheritance of several cold tolerance traits in  
7 corn. *Crop Sci.* 19:847–852.
- 8 Mosely, P.R., T.M. Crosbie, and J.J. Mock. 1984. Mass selection for improved cold and  
9 density tolerance of two maize populations. *Euphytica* 33:263–269.
- 10 Nei, M. 1978. Estimation of average heterozygosity and genetic distance from a small  
11 number of individuals. *Genetics* 89: 583.
- 12 Presterl, T., M. Ouzunova, W. Schmidt, E.M. Moeller, F.K. Roeber, C. Knaak, K. Ernst, P.  
13 Westhoff, H.H. Geiger. 2007. Quantitative trait loci for early plant vigour of maize  
14 grown in chilly environments. *Theor. Appl. Genet.* 114:1059–1070.
- 15 Rebouq, C., M. Chastanet, B. Gouesnard, C. Welcker, P. Dubreuil, and A. Charcosset.  
16 2003. Maize introduction into Europe: The history reviewed in the light of molecular  
17 data. *Theor. Appl. Genet.* 106:895–903.
- 18 Revilla, P., A. Butrón, M.E. Cartea, R.A. Malvar, and A. Ordás. 2005. Breeding for cold  
19 tolerance. In: M. Ashraf y PJC Harris (eds). *Abiotic Stresses. Plant resistance through*  
20 *breeding and molecular approaches.* The Haworth Press, Inc., New York.
- 21 Revilla, P., R.A. Malvar, M.E. Cartea, A. Butrón, and A. Ordás. 2000. Inheritance of cold  
22 tolerance at emergence and during early season growth in maize. *Crop Sci.* 40:1579–  
23 1585.
- 24 Revilla, P., R.A. Malvar, M.E. Cartea, and A. Ordás. 1998. Identifying open-pollinated  
25 populations of field corn as source of cold tolerance for improving sweet corn.  
26 *Euphytica* 101:239–247.

- 1    Revilla, P., P. Soengas, M.E. Cartea, R.A. Malvar, and A. Ordás. 2003. Isozyme variability  
2           among European maize populations and the introduction of maize in Europe.  
3           *Maydica* 48:141–152.
- 4    Rincent, R., D. Laloë, S. Nicolas, T. Altmann, D. Brunel, P. Revilla, V.M. Rodriguez, J.  
5           Moreno-Gonzales, A.E. Melchinger, E. Bauer, C.-C. Schön, N. Meyer, C. Giauffret,  
6           C. Bauland, P. Jamin, J. Laborde, H. Monod, P. Flament, A. Charcosset, and L.  
7           Moreau. 2012. Maximizing the reliability of genomic selection by optimizing the  
8           calibration set of reference individuals: comparison of methods in two diverse groups  
9           of maize inbreds (*Zea mays* L.). *Genetics* 192:715–728.
- 10   Rodríguez, V.M., A. Butrón, R.A. Malvar., A. Ordás, and P. Revilla. 2008. QTLs for cold  
11           tolerance in the maize IBM population. *Int. J. Plant Sci.* 169:551–556.
- 12   Rodríguez V.M., A. Butrón, M.O.A. Rady, P. Soengas, and P. Revilla. 2013. Identification  
13           of QTLs involved in the response to cold stress in maize (*Zea mays* L.). *Mol Breed.*  
14           33:363–371.
- 15   Rodríguez, V.M., M.C. Romay, A. Ordás, and P. Revilla. 2010. Evaluation of the European  
16           maize (*Zea mays* L.) germplasm under cold conditions. *Gen. Res. Crop Evol.* 57:329–  
17           335.
- 18   Rodríguez V.M., P. Velasco, J.L. Garrido, P. Revilla, A. Ordás, and A Butrón. 2013.  
19           Genetic regulation of cold-induced albinism in the maize inbred line A661. *J. Exp.*  
20           *Bot.* 64:3657–3667.
- 21   SAS Institute Inc., 2008. Cary, North Carolina.
- 22   Semuguruka, G.H., W.A. Compton, C.Y. Sullivan, and M.A. Thomas. 1981. Some  
23           measures of temperature response in corn (*Zea mays* L.). *Maydica* 26:209–218.
- 24   Strigens, A., C. Grieder, B.I.G. Haussmann, and A.E. Melchinger. 2012. Genetic variation  
25           among inbred lines and testcrosses of maize for early growth parameters and their  
26           relationship to final dry matter yield. *Crop Sci.* 52:1084–1092.

- 1 Strigens, A., N.M. Freitag, X. Gilbert, C. Grieder, C. Riedelsheimer, T.A. Schrag, R.  
2 Messmer, and A.E. Melchinger. 2013. Association mapping for chilling tolerance in  
3 elite flint and dent maize inbred lines evaluated in growth chamber and field  
4 experiments. *Plant Cell Env.* 36:1871–1887.
- 5 Verheul, M.J., C. Picatto, and P. Stamp. 1996. Growth and development of maize (*Zea mays*  
6 L.) seedlings under chilling conditions in the field. *Eur. J. Agron.* 5:31–43.
- 7

1 Figure 1. Principal component analyses for maize inbreds *per se* in cold conditions using the  
2 least squares means of days to emergence,  $\Phi$ PSII, chlorophyll content, and early vigor after  
3 standardizing the traits. A) Cold tolerant dent inbreds with  $PRIN1 > 2$  and  $PRIN2 < -1$ . B)  
4 Cold tolerant flint inbreds with  $PRIN1 > 2$  and  $PRIN2 < -1$ . C) Dent panel. D) Flint panel.  
5

| Table 1. Mean comparisons between flint and dent maize inbred panels <i>per se</i> for four traits recorded in cold and control conditions in a growth chamber according to an F test ( $p$ = probability of significant differences)  |                   |         |   |         |                          |         |                          |         |
|--|-------------------|---------|---|---------|--------------------------|---------|--------------------------|---------|
| Type   | Days to emergence |         | Relative chlorophyll content <sup>†</sup> |         | $\Phi$ PSII <sup>‡</sup> |         | Early vigor <sup>§</sup> |         |
|  | cold              | control | cold                                      | control | Cold                     | control | cold                     | control |
| Dent   | 11.42             | 3.39    | 4.61                                      | 11.82   | 0.39                     | 0.72    | 4.00                     | 5.07    |
| Flint  | 11.79             | 3.39    | 5.54                                      | 12.63   | 0.43                     | 0.71    | 3.86                     | 4.93    |
| $p$  | 0.001             | 0.856   | <0.0001                                   | 0.010   | 0.0023                   | <0.0001 | 0.007                    | 0.003   |
| <sup>†</sup> Relative chlorophyll content (SPAD) recorded using a hand-held Chlorophyll Content Meter, CCM-200 (Opti-Sciences, Tyngsboro, Massachusetts, USA)<br><sup>‡</sup> Recorded using an OS-30p Chlorophyll Fluorometer (Opti-Sciences, Inc., USA)<br><sup>§</sup> Subjective score from 1=weak plants to 9=vigorous plants |                   |         |   |         |                          |         |                          |         |

Table 2. Comparisons among groups of maize germplasm in the dent and flint panels for cold tolerance-related traits under cold and control conditions evaluated in a growth chamber

| Germplasm group at K=6<br>(Number of inbreds in the group)  | Days to emergence |         | Chlorophyll content <sup>†</sup> |          | ΦPSII <sup>‡</sup> |         | Early vigor <sup>§</sup> |          |
|---|-------------------|---------|----------------------------------|----------|--------------------|---------|--------------------------|----------|
|   | Cold              | Control | Cold                             | Control  | Cold               | Control | Cold                     | Control  |
| <b>Dent panel</b>   |                   |         |                                  |          |                    |         |                          |          |
| Iodent Ph207 (42 inbreds)   | 11.31 b           | 3.34 b  | 5.72 a                           | 13.59 a  | 0.47 a             | 0.71 c  | 4.07 a                   | 5.26 a   |
| Iodent UH4068 (16)  | 10.81 b           | 3.39 ab | 4.01 bc                          | 13.88 a  | 0.42 a             | 0.73 a  | 4.19 a                   | 5.07 abc |
| Lancaster Oh43 (12)   | 12.18 a           | 3.42 a  | 4.82 b                           | 10.15 c  | 0.44 a             | 0.72 ab | 4.25 a                   | 5.01 bc  |
| No Iodent (80)  | 11.80 a           | 3.40 a  | 4.35 b                           | 12.01 b  | 0.34 b             | 0.72 b  | 3.87 b                   | 5.05 b   |
| Other No Iodent F252 (21)   | 10.74 b           | 3.34 ab | 4.63 b                           | 12.61 a  | 0.47 a             | 0.73 ab | 4.25 a                   | 4.86 c   |
| Stiff Stalk (36)  | 11.01 b           | 3.38 ab | 3.65 c                           | 12.41 b  | 0.24 c             | 0.73 ab | 3.92 b                   | 5.12 ab  |
| <b>Flint panel</b>  |                   |         |                                  |          |                    |         |                          |          |
| FV7 (24)  | 12.11 b           | 3.4 ab  | 5.09 ab                          | 11.73 bc | 0.44 b             | 0.70 b  | 3.85 b                   | 4.78 b   |
| Northern Flint (44)   | 11.85 b           | 3.41 b  | 5.85 a                           | 14.18 a  | 0.42 b             | 0.72 a  | 3.91 b                   | 5.01 a   |
| Northern Flint D171 (43)  | 11.30 a           | 3.35 a  | 5.79 a                           | 13.91 a  | 0.50 a             | 0.70 b  | 4.14 a                   | 5.12 a   |
| No Northern Flint (36)  | 12.27 b           | 3.39 a  | 4.74 b                           | 12.4 b   | 0.37 c             | 0.71 ab | 3.58 c                   | 4.83 b   |
| Southern EC18 (16)  | 12.59 b           | 3.46 b  | 6.49 a                           | 11.89 bc | 0.51 a             | 0.73 a  | 3.85 bc                  | 5.01 ab  |
| S. Open Pollinated Varieties (19)   | 11.48 ab          | 3.35 a  | 4.45 b                           | 9.9 c    | 0.34 c             | 0.72 ab | 3.66 bc                  | 5.14 a   |
| <sup>†</sup> Relative chlorophyll content (SPAD) recorded using a hand-held Chlorophyll Content Meter, CCM-200 (Opti-Sciences, Tyngsboro, Massachusetts, USA)<br><sup>‡</sup> Recorded using an OS-30p Chlorophyll Fluorometer (Opti-Sciences, Inc., USA)<br><sup>§</sup> Subjective scale from 1=weak plants to 9=vigorous plants<br>Means followed by the same letter, within the same column and panel, were not significantly different |                   |         |                                  |          |                    |         |                          |          |

Table 3. List of 101 inbreds of maize with the highest cold tolerance, i.e. those that were not significantly different from the best inbred for four (in bold) or three traits when evaluated *per se* in cold conditions in a growth chamber. Ranking goes from top to bottom and from left to right, with EV18 being the first and UHP017 the last in the ranking.

| Type         | Inbred        | Type  | Inbred     | Type  | Inbred | Type  | Inbred    | Type  | Inbred |
|--------------|---------------|-------|------------|-------|--------|-------|-----------|-------|--------|
| <b>Flint</b> | <b>EV18</b>   | Flint | PB57       | Dent  | PH207  | Flint | CH31A     | Dent  | UHS002 |
| <b>Flint</b> | <b>F364</b>   | Flint | EP71       | Flint | FV70   | Flint | PLS41     | Flint | UHF098 |
| <b>Flint</b> | <b>F471</b>   | Dent  | F7025      | Flint | UH5250 | Dent  | PHV78     | Flint | UHF023 |
| <b>Flint</b> | <b>EC51</b>   | Dent  | EV30       | Flint | F02803 | Flint | UHF091    | Flint | UHL038 |
| <b>Flint</b> | <b>UHL058</b> | Dent  | UHP074     | Dent  | EP72   | Dent  | UH6132    | Dent  | UH6102 |
| <b>Flint</b> | <b>CH34</b>   | Flint | F591       | Dent  | C105   | Dent  | UHP033    | Dent  | B99    |
| <b>Dent</b>  | <b>UHP024</b> | Flint | CH16.1-295 | Dent  | EP74   | Dent  | Pa31      | Dent  | UH304  |
| <b>Flint</b> | <b>FV71</b>   | Flint | CH4.2      | Dent  | LH85   | Flint | F362      | Flint | PP87   |
| <b>Flint</b> | <b>UHF043</b> | Flint | EA1349     | Dent  | UHP042 | Dent  | UH2551    | Flint | UH007  |
| Flint        | EP1           | Flint | UH1494     | Dent  | FV335  | Dent  | UH8513    | Dent  | F816   |
| Flint        | FV75          | Flint | UHF035     | Flint | UH006  | Dent  | EZ19      | Dent  | D06    |
| Flint        | EC237         | Flint | EZ16A      | Flint | UHL048 | Flint | FV65      | Dent  | Pa35   |
| Flint        | PLS6          | Flint | FC1571     | Dent  | Pa374  | Dent  | H99       | Dent  | Mo17   |
| Flint        | EC35G         | Flint | UH5231     | Flint | EP45   | Dent  | EC151     | Flint | FV344  |
| Flint        | UH5113        | Dent  | UH6148     | Dent  | FC1890 | Flint | CH113-379 | Flint | PB6R   |
| Dent         | NQ508         | Dent  | EC242C     | Flint | EZ53   | Dent  | FV277     | Dent  | F908   |
| Flint        | D152          | Flint | FV79       | Flint | CO255  | Flint | RT9       | Dent  | EC326A |
| Flint        | PB268         | Flint | FV131      | Flint | UHF093 | Dent  | W602S     | Dent  | F838   |
| Flint        | UHF106        | Dent  | F7028      | Flint | FV355b | Flint | EZ21      | Dent  | FV113  |
| Dent         | LP325         | Dent  | W604S      | Dent  | FC1852 | Dent  | LH82      | Dent  | UHP017 |
| Dent         | F670          |       |            |       |        |       |           |       |        |



Table 4. List of 23 testcrosses<sup>†</sup> from maize inbreds with the highest cold tolerance, i.e. those that were not significantly different for four (in bold) or three traits from the best when evaluated in cold conditions in a growth chamber. Ranking goes from top to bottom and from left to right, with EC35G being the first and EZ11A the last in the ranking.

| Type  | Inbred       | Type  | Inbred |
|-------|--------------|-------|--------|
| Flint | <b>EC35G</b> | Flint | FV66   |
| Flint | CH34         | Dent  | F7059  |
| Flint | CH8.7        | Dent  | NS701  |
| Flint | CH27-12      | Flint | FV72   |
| Flint | PP85         | Dent  | UHP017 |
| Dent  | B37          | Dent  | ML606  |
| Flint | F47          | Dent  | B14A   |
| Flint | IL101        | Flint | UH1291 |
| Flint | CH16.1-295   | Flint | EC50   |
| Flint | FV345        | Dent  | F670   |
| Dent  | FC1819       | Dent  | EZ11A  |
| Flint | CH28-2       |       |        |

<sup>†</sup> The dent inbreds were crossed to the flint tester UH007 and the flint inbreds to the dent tester F353

Table 5. Multiple stepwise regressions for biomass yield of testcrosses from two panels of flint and dent maize inbred lines evaluated in the field, and in a growth chamber under control and cold conditions (only significant variables are shown)

| Significant independent variables  | Cumulated R <sup>2</sup> | Coefficient  |
|--|--------------------------|--------------|
| <b>Dry matter yield of testcrosses from both panels evaluated in the field</b>     |                          |              |
| Plant height of testcrosses (field)  | 0.285                    | 0.056±0.004  |
| ΦPSII of testcrosses (field)   | 0.319                    | 0.011±0.002  |
| Days to pollen of testcrosses (field)  | 0.334                    | 0.113±0.032  |
| Dry early weight of testcrosses (control conditions)                               | 0.348                    | 6.830±2.080  |
| Chlorophyll content of testcrosses (field)   | 0.356                    | 0.037±0.015  |
| Dry weight at 5 weeks of testcrosses (field)                                       | 0.363                    | -0.060±0.017 |
| Early vigor of testcrosses (field)   | 0.372                    | 0.307±0.115  |
| Days to emergence (cold conditions)  | 0.376                    | -0.101±0.050 |
| <b>Dry matter yield of testcrosses from the dent panel evaluated in the field</b>  |                          |              |
| Plant height of testcrosses (field)  | 0.200                    | 0.055±0.006  |
| Days to emergence (cold conditions)  | 0.217                    | -0.180±0.064 |
| Dry early weight of testcrosses (control conditions)                               | 0.230                    | 6.330±3.013  |
| ΦPSII of testcrosses (field)   | 0.242                    | 0.007±0.003  |
| Dry weight at 5 weeks of testcrosses (field)                                       | 0.255                    | -0.040±0.019 |
| <b>Dry matter yield of testcrosses from the flint panel evaluated in the field</b> |                          |              |
| Plant height of testcrosses (field)  | 0.220                    | 0.053±0.006  |
| ΦPSII of testcrosses (field)   | 0.258                    | 0.013±0.004  |
| Dry weight of testcrosses (control conditions)                                     | 0.276                    | 9.048±2.789  |
| ΦPSII of inbreds (cold conditions)   | 0.293                    | 0.003±0.001  |
| Early vigor of inbreds (cold conditions)   | 0.316                    | -0.453±0.180 |
| Dry weight at 5 weeks of testcrosses (field)                                       | 0.330                    | -0.093±0.027 |
| Early vigor of testcrosses (field)   | 0.349                    | 0.453±0.168  |