



Combined Vaccination with B Cell Peptides Targeting Her-2/neu and Immune Checkpoints as Emerging Treatment Option in Cancer

Joshua Tobias ^{1,*}, Mirjana Drinić ¹, Anna Schmid ¹, Anastasiya Hladik ², Martin L. Watzenböck ², Claire Battin ³, Erika Garner-Spitzer ¹, Peter Steinberger ³, Michael Kundi ⁴, Sylvia Knapp ², Christoph C. Zielinski ⁵ and Ursula Wiedermann ^{1,*}

- ¹ Center for Pathophysiology, Infectiology and Immunology, Institute of Specific Prophylaxis and Tropical Medicine, Medical University of Vienna, 1090 Vienna, Austria
- ² Department of Medicine I, Research Division of Infection Biology, Medical University of Vienna, 1090 Vienna, Austria
- ³ Center for Pathophysiology, Infectiology and Immunology, Division of Immune Receptors and T Cell Activation, Institute of Immunology, Medical University of Vienna, 1090 Vienna, Austria
- ⁴ Department of Environmental Health, Center for Public Health, Medical University of Vienna, 1090 Vienna, Austria
- ⁵ Central European Cancer Center, Wiener Privatklinik and Central European Cooperative Oncology Group (CECOG), 1090 Vienna, Austria
- * Correspondence: joshua.tobias@meduniwien.ac.at (J.T.); ursula.wiedermann@meduniwien.ac.at (U.W.)

Simple Summary: Therapies with monoclonal antibodies (mAbs) targeting tumor-associated antigens (TAAs) or immune checkpoint inhibitors (ICIs) have revolutionized cancer treatment. Nevertheless, the inevitable development of resistance and the failure to respond are among this approach's disadvantages, limiting the duration of disease- or progression-free and overall survival. As an alternative to therapeutically efficacious monoclonal antibodies, the concept of active immunization with vaccines has been repeatedly discussed. In particular, mimotopes, representing the B cell epitope of therapeutic mAbs, have been shown to induce immunological memory and effectively produce antibodies with similar functionality to the respective mAbs/ICIs. This review focuses on a new frontier of vaccinations directed against two cancer-relevant targets, addresses concerns about the safety of active immunization targeting PD-1 and discusses limitations and outlooks.

Abstract: The application of monoclonal antibodies (mAbs), targeting tumor-associated (TAAs) or tumor-specific antigens or immune checkpoints (ICs), has shown tremendous success in cancer therapy. However, the application of mAbs suffers from a series of limitations, including the necessity of frequent administration, the limited duration of clinical response and the emergence of frequently pronounced immune-related adverse events. However, the introduction of mAbs has also resulted in a multitude of novel developments for the treatment of cancers, including vaccinations against various tumor cell-associated epitopes. Here, we reviewed recent clinical trials involving combination therapies with mAbs targeting the PD-1/PD-L1 axis and Her-2/neu, which was chosen as a paradigm for a clinically highly relevant TAA. Our recent findings from murine immunizations against the PD-1 pathway and Her-2/neu with peptides representing the mimotopes/B cell peptides of therapeutic antibodies targeting these molecules are an important focus of the present review. Moreover, concerns regarding the safety of vaccination approaches targeting PD-1, in the context of the continuing immune response, as a result of induced immunological memory, are also addressed. Hence, we describe a new frontier of cancer treatment by active immunization using combined mimotopes/B cell peptides aimed at various targets relevant to cancer biology.

Keywords: Her-2/neu; immune checkpoints; combination therapy; vaccination; active immunization; mimotopes/B cell peptides; anti-tumor effect; safety



Citation: Tobias, J.; Drinić, M.; Schmid, A.; Hladik, A.; Watzenböck, M.L.; Battin, C.; Garner-Spitzer, E.; Steinberger, P.; Kundi, M.; Knapp, S.; et al. Combined Vaccination with B Cell Peptides Targeting Her-2/neu and Immune Checkpoints as Emerging Treatment Option in Cancer. *Cancers* **2022**, *14*, 5678. https://doi.org/10.3390/ cancers14225678

Academic Editor: Armand Bensussan

Received: 12 October 2022 Accepted: 12 November 2022 Published: 18 November 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

1. Introduction

A range of cancer immunotherapy approaches enabling and reactivating the patient's anti-tumor immune response have shown remarkable advances in recent years [1,2]. The application of therapeutic and tumor-targeting monoclonal antibodies (mAbs), possessing an intrinsic antineoplastic activity, represents the passive form of immunotherapy [3–6]. Contrarily, anticancer vaccines enhance the activation of the host's immune system [7,8], referred to as active immunotherapy. Cancer immunotherapy based on the application of mAbs has been successfully established in recent years [9–13]; however, tumor heterogeneity, intra-tumoral factors, the interaction between cancer cells and the immune system, as well as the tumor microenvironment, are amongst factors limiting the efficacy of therapeutically applied monoclonal antibodies [14].

The interaction between T cell receptors and antigens presented in the context of MHC molecules and costimulatory receptors (such as CD28 or CD86) results in the activation of T cells [15]. However, co-inhibitory receptors on T cells result in counterbalanced stimulatory signals. Such co-inhibitory molecules, i.e., immune checkpoints (ICs) include programmed cell death 1 (PD-1), which binds to PD-L1 (B7-H1 or CD274) and PD-L2 (B7-DC or CD273), and cytotoxic T lymphocyte antigen 4 (CTLA-4), which interacts with CD80 or CD86 [16–19]. Preclinical and clinical evidence has repeatedly demonstrated the ability of tumors to escape immunosurveillance via the expression of surface ligands that engage inhibitory receptors on tumor-specific T cells, thus, resulting in immune tolerance and failure to induce tumor cell death due to T cell anergy/exhaustion [7,20]. Consequently, the blockade of the PD-1/PD-L1 interaction by various mAbs, i.e., immune checkpoint inhibitors (ICIs) [6,21–23], which target PD-1 (e.g., nivolumab, pembrolizumab and cemiplimab), PD-L1 (e.g., atezolizumab, avelumab and durvalumab) or CTLA-4 (ipilimumab and tremelimumab) [23–25], is considered a milestone in cancer treatment. This has been registered in manifold indications [26–28], for example, in the treatment of gastroesophageal cancer [29], hepatocellular carcinoma [30], cervical cancer [31], head and neck cancer [32], urothelial carcinoma [33] and lymphoma [34]. Treatment with ICIs still holds tremendous promise, be it as monotherapy or in combination with either chemotherapy, targeted therapies or other immunomodulatory compounds [35].

Her-2/neu, a 185 kDa transmembrane protein, is a member of the human epidermal growth factor receptors (EGFR) family, and its overexpression has been demonstrated in approximately 15–30% of breast and gastric cancers [36–42]. The receptor is an attractive tumor-associated antigen (TAA) for cancer therapy due to the association of Her-2/neu overexpression with an aggressive biological cancer phenotype and reduced survival in patients with Her-2/neu-positive tumors, as well as improvable response to traditional chemotherapy and, consequently, poor prognosis [42–46]. The extracellular domain of Her-2/neu is divided into four subdomains (I, II, III, IV), with its intracellular domain exhibiting tyrosine kinase activity similar to the other receptors in this family, i.e., Her-1 or EGFR, Her-3 and Her-4 [47,48]. Because no ligand has been identified for Her-2/neu and it is in a constitutively active conformation, the receptor is a preferred partner for dimerization with other members via the dimerization loop located on its extracellular subdomain II. Once paired, the tyrosine residues on the receptor's intracellular domains are mutually phosphorylated, leading to the initiation of signaling pathways, including the phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt) pathway, and, consequently, tumor cell proliferation, angiogenesis, invasion and metastasis [49–52]. By way of different mechanisms, such as antibody-dependent cellular cytotoxicity (ADCC), dimerization inhibition in a ligand-independent manner, the receptor's degradation and/or internalization and PI3K-AKT signaling pathway inhibition, the first FDA-approved anti-Her-2/neu humanized mAb, trastuzumab, interferes with Her-2/neu signaling [53,54] (Figure 1). The initial therapeutic effect evaluation of trastuzumab showed that treatment with mAb after adjuvant chemotherapy improved overall survival (OS) in women with Her-2/neu-positive metastatic breast cancer [55] and resulted in significantly improved disease-free survival one year after the treatment [56]. As a standard first line of care, trastuzumab is used for the

treatment of both early-stage and metastatic Her-2/neu-positive breast cancer. Differing from trastuzumab in its mechanisms of action, pertuzumab, the second FDA-approved anti-Her-2/neu mAb, which is used for the treatment of Her-2/neu-positive metastatic breast cancer, targets the dimerization domain/loop of Her-2/neu and, consequently, results in the inhibition of the receptor's ligand-induced/dependent dimerization [57,58] (Figure 1). Therefore, by complementary mechanisms of action and in a synergistic manner, which results in a maximal blockade of the Her-2/neu oncogenic pathway [58], the inhibition of tumor growth by the combination of trastuzumab and pertuzumab in both in vitro and in vivo preclinical models has been shown [59,60]. The addition of pertuzumab to trastuzumab and docetaxel was shown in the phase III CLEOPATRA trial to significantly prolong median OS by 15.7 months compared to placebo plus trastuzumab and docetaxel [61].



Figure 1. Schematic description of passive immunotherapy with Her-2/neu-targeting therapeutic mAbs pertuzumab and trastuzumab. The mAbs bind to their respective B cell epitopes on extracellular domain II (pertuzumab) and IV (Trastuzumab) of Her-2/neu. The mAbs' modes of action upon their binding to the receptor are shown in the respective boxes. Adapted with permission from Ref. [62]. Elsevier, 2022.

ICIs have been proposed to strengthen an emerging immune response following vaccination against TAAs. Here, we summarize the latest reports on the application of Her-2/neu-targeting mAbs and ICIs and vaccination strategies for targeting Her-2/neu and immune checkpoints.

2. Combination of Her-2/neu-Targeted Therapy with Immune Checkpoint Blockade *2.1. Basic Considerations*

Trastuzumab's capacity in upregulating PD-L1 expression, by the recruitment of IFNgamma-secreting immune effector cells, has been shown as a mechanism of resistance to trastuzumab [63]. Furthermore, the mAbs that are bound to tumor cells engage innate

immune effector cells via their Fc receptor, resulting in ADCC [64,65], which in turn also results in the upregulation of PD-L1 [66]. It has been shown that the increased ability of tumor cells to evade the immune system is attributed to their PD-L1 expression capacity, allowing their interaction with PD-1-expressing immune cells [67]. Because the application of ICIs aims to inhibit the interaction of PD-1 with PD-L1 and increase T cell survival and proliferation, consequently leading to an enhanced tumor-directed immune response, it has been suggested that combining ICIs with anti-Her-2/neu mAbs, such as Trastuzumab, could act in a synergistic manner to delay or even prevent resistance to the mAb [68]. Thus, the progression of Her-2/neu-positive metastatic breast cancer, as a result of primary or secondary resistance to Her-2/neu-targeted therapies with trastuzumab [69], might be hindered by combining with an immune checkpoint blockade [70,71].

2.2. Clinical Trials for Combination Therapies of Her-2/neu-Targeting Compounds plus ICIs

A phase II trial (ClinicalTrials.gov Identifier: NCT02129556) enrolled 52 patients with advanced Her-2/neu-positive breast cancer who had progressed on trastuzumab treatment [72] (Table 1). Patients received anti-PD-1 mAb pembrolizumab and standard trastuzumab. Patients with PD-L1-positive tumors demonstrated a disease control rate of 25% (90% CI, 14–39), with a 12-month progression-free survival (PFS) and OS of 12% and 65%, respectively, whereas patients with PD-L1-negative tumors did not experience a benefit from such treatment. The results from this trial were promising and indicated that Her-2/neu-positive PD-L1 overexpressing breast cancer patients benefited from anti-PD-1 therapy [72]. In a recently completed randomized phase II trial (ClinicalTrials.gov Identifier: NCT02924883), the addition of anti-PD-L1 mAb atezolizumab to T-DM1 (trastuzumab covalently linked to the cytotoxic agent DM1) [73] was evaluated in patients with unresectable or metastatic Her-2/neu-positive breast cancer who had received prior trastuzumab- and taxane-based therapy [74] (Table 1). Patients with PD-L1-positive breast cancers from the atezolizumab group had longer PFS than those with PD-L1-negative tumors. Furthermore, patients in the atezolizumab group whose tumors had \geq 5% infiltrating-tumor lymphocytes (TILs) had longer PFS than patients with a TILs of <5% [74]. On the basis of the results of this trial, a randomized phase III trial (ClinicalTrials.gov Identifier: NCT04873362; Table 1) is now being conducted to evaluate T-DM1 \pm atezolizumab in PD-L1 overexpressing breast cancers with residual invasive Her-2/neu-positive breast cancer following neoadjuvant taxane-based and Her-2/neu-targeted therapy.

As also shown in the case of other types of cancer [75], patients with PD-L1-positive tumors may potentially benefit more from PD-1 or PD-L1 inhibition than those with PD-L1-negative tumors. Ongoing trials aim to evaluate PD-1-, PD-L1- or CTLA4-targeting ICIs in combination with standard anti-Her-2/neu therapy for Her-2/neu-positive breast cancer (Table 1).

Evaluated Drugs						NCT/		
Anti-Her-2 mAb	ICI	Biological/ Other Drug	Condition/Disease	Phase	Status	Trial's Identifier	Setting	Reference
-	Avelumab	Taxane and anthracycline	Metastatic or locally advanced solid tumors	Ι	Completed	NCT01772004	Dose escalation trial	[76]
Trastuzumab, pertuzumab and trastuzumab emtansine	Atezolizumab	Carboplatin, docetaxel, doxorubicin and cyclophosphamide	Her-2/neu-positive and negative metastatic breast cancer and locally advanced early breast cancer	Ι	Completed	NCT02605915	Two cohorts with several arms, which evaluated the different combinations of the examined drugs	[77]
Trastuzumab	Pembrolizumab		Advanced, trastuzumab-resistant Her-2/neu-positive metastatic breast cancer	Ib, II	Completed	NCT02129556 (PANACEA)	trastuzumab with pembrolizumab	[72]
Trastuzumab emtansine	Atezolizumab		Her-2/neu-positive locally advanced or metastatic breast cancer	П	Completed	NCT02924883	Arm 1: trastuzumab emtansine and placebo Arm 2: trastuzumab emtansine and atezolizumab	[74]
Trastuzumab and pertuzumab	Atezolizumab		Her-2/neu-positive metastatic breast cancer that has spread to the brain	П	Active	NCT03417544	Arm: trastuzumab, pertuzumab and atezolizumab	Clinicaltrials.gov
Trastuzumab and pertuzumab	Atezolizumab	Doxorubicin, cyclophosphamid and paclitaxel	Early Her-2/neu-positive breast cancer	ш	Active	NCT03726879	Arm 1: placebo, doxorubicin, cyclophosphamide, paclitaxel, trastuzumab, pertuzumab and trastuzumab emtansine Arm 2: atezolizumab, doxorubicin, cyclophosphamide, paclitaxel, trastuzumab, pertuzumab and trastuzumab emtansine	Clinicaltrials.gov

 Table 1. List of main completed/active/recruiting clinical trials with ICIs and anti-Her-2/neu-targeted therapies, with or without combination with biological drugs.

Evaluated Drugs						NCT/		
Anti-Her-2 mAb	ICI	Biological/ Other Drug	Condition/Disease	Phase	Status	Trial's Identifier	Setting	Reference
Trastuzumab and pertuzumab	Atezolizumab	Taxane, paclitaxel and docetaxel	Neoadjuvant treatment of Her-2/neu-positive early high-risk and locally advanced breast cancer	Ш	Active	NCT03595592	Arm 1: trastuzumab, pertuzumab, paclitaxel and carboplatin Arm 2: trastuzumab, pertuzumab, paclitaxel, carboplatin, doxorubicin, cyclophosphamide and atezolizumab Arm 3: trastuzumab, pertuzumab, paclitaxel, carboplatin and atezolizumab	Clinicaltrials.gov
Trastuzumab and pertuzumab	Atezolizumab	Taxane, paclitaxel and docetaxel	Her-2/neu-positive metastatic breast cancer	Ш	Active	NCT03199885	Arm 1: trastuzumab, pertuzumab, taxane therapy and placebo Arm 2: trastuzumab, pertuzumab, taxane therapy and atezolizumab	Clinicaltrials.gov
Trastuzumab deruxtecan (T-DXd), trastuzumab and pertuzumab	Durvalumab	Deruxtecan, paclitaxel and tucatinib	Her-2/neu-positive metastatic breast cancer, without (part 1) or with brain metastases (part 2)	I, II	Recruiting	NCT04538742	Part 1: Arm 1: T-DXd Arm 2: T-DXd and durvalumab Arm 3: T-DXd and pertuzumab Arm 4: T-DXd and paclitaxel Arm 5: T-DXd and durvalumab and paclitaxel Arm 6: T-DXd and tucatinib Part 2: Arm 7: T-DXd and tucatinib Part 2: Arm 7: T-DXd and tucatinib	Clinicaltrials.gov

Table 1. Cont.

Evaluated Drugs NCT/ Condition/Disease Phase Status Trial's Setting Reference **Biological**/ Anti-Her-2 mAb ICI Identifier Other Drug Arm 1: VRP-Her-2/neu vaccine Arm 2: VRP-Her-2/neu and Patients with Her-2/neu breast Π NCT03632941 pembrolizumab Recruiting [78] pembrolizumab cancer Arm 3: VRP-Her-2/neu vaccine and pembrolizumab Arm: Her-2/neu-positive advanced/ Trastuzumab Atezolizumab Vinorelbine Π Recruiting NCT04759248 trastuzumab, atezolizumab Clinicaltrials.gov metastatic breast cancer and vinorelbine Arm: Tremelimumab and Her-2/neu-positive metastatic breastcancer-II BCT 1703 durvalumab, tremelimumab Trastuzumab Recruiting durvalumab breast cancer trials.org.au and trastuzumab Arm 1: trastuzumab and vinorelbine Arm 2: trastuzumab, vinorelbine Avelumab and Advanced Her-2/neu-positive and avelumab Trastuzumab Vinorelbine Π Recruiting NCT03414658 Clinicaltrials.gov utomilumab [79] breast cancer Arm 3: trastuzumab, vinorelbine, avelumab and utomilumab Arm 4: trastuzumab, avelumab and utomilumab Arm 1: trastuzumab, pertuzumab and paclitaxel Chemo naive patients with Arm 2: trastuzumab, Trastuzumab and invasive Her-2/neu-positive Π Clinicaltrials.gov Pembrolizumab Paclitaxel Recruiting NCT03747120 pertuzumab, pertuzumab pembrolizumab and breast cancer paclitaxel Arm 3: trastuzumab, pembrolizumab and paclitaxel

Table 1. Cont.

Evaluated Drugs						NCT/		
Anti-Her-2 mAb	ICI	Biological/ Other Drug	Condition/Disease	Phase	Status	Trial's Identifier	Setting	Reference
Trastuzumab emtansine	Atezolizumab		Her-2/neu-positive breast cancer at high risk of recurrence following preoperative therapy	III	Recruiting	NCT04873362	Arm 1: placebo and trastuzumab emtansine Arm 2: atezolizumab and trastuzumab emtansine	Clinicaltrials.gov
Trastuzumab emtansine	Atezolizumab		Her-2/neu-positive and PD-L1-positive locally advanced or metastatic breast cancer	III	Recruiting	NCT04740918	Arm 1: T-DM1 and placebo Arm 2: T-DM1 and atezolizumab	Clinicaltrials.gov
Trastuzumab emtansine	Atezolizumab		Her-2/neu-positive primary breast cancer	Ш	Recruiting	NCT04873362	Arm 1: trastuzumab emtansine and placebo Arm 2: trastuzumab emtansine and atezolizumab	Clinicaltrials.gov

ICI: Immune Checkpoint Inhibitor; and VRP-Her-2/neu: virus-like replicon particles (VRP) packaged with an alphaviral vector encoding the extracellular domain and transmembrane regions of Her-2/neu.

Table 1. Cont.

2.3. B Cell Peptides/Mimotopes-Based Vaccination Targeting Her-2/neu and ICs

Despite tremendous therapeutic success with the use of mAbs, their continuous application over a long period, their half-life [80] and lack of capacity to induce immunological memory [81] may limit the duration of therapy, resulting in only temporary disease control, in particular, once the tumor has metastasized [82–89]. Further drawbacks of treatments with mAbs include the high frequencies of non-responsiveness [90], the development of resistance [91], and immune-related adverse events and hypersensitivity to treatments with mAbs [92,93], possibly due to the high doses of mAbs that ensure their immediate therapeutic effect [94,95]. In contrast, vaccines based on TAAs or mimotopes representing therapeutic mAb binding epitopes can induce prolonged activation of the immune system and mount immunological memory, which, in association with booster vaccinations, potentially results in tumor involution [96].

Mimotopes are peptides that mimic and represent the immunodominant epitopes on a target protein (TAA or tumor-specific antigens) or the binding epitopes of therapeutic mAbs [97]. Mimotopes identified by antibodies or therapeutic mAbs are solely a representation of the antibodies' B cell epitopes and, thus, are considered as B cell peptides. Such mimotopes, consequently, not only inhibit the binding of the antibodies to the respective antigen or protein but also, upon conjugation to an immunogenic carrier protein, induce an epitope-specific antibody response. As depicted in Figure 1, the application of mAbs represents the passive immunotherapy approach, while vaccination with B cell peptides/mimotopes is the active immunotherapy approach (Figure 2). Endogenously generated antibodies after vaccination can induce anti-tumor responses for prolonged periods of time by the induction of immunological memory [98].

The identification of mimotopes can be based on the use of phase display strategies [99,100] or, alternatively, as applied in our group, by computer algorithms or the use of synthesized overlapping bio-peptides from the sequence of the respective protein [62,101–103]. The peptides are screened and the mimotope candidates are selected based on their capacity to inhibit the binding of the examined mAb to the respective protein [62,101,102,104]. For the evaluation of immunogenicity by vaccination strategy and the anti-tumor effect, the mimotopes (B cell peptides) are conjugated to a carrier protein and administered together with an adjuvant [62,101,102,104] (Figure 2). Mimotopes have demonstrated a promising approach in the field of allergy [105,106], infectious diseases [107–109] and for cancer therapy, by inducing an anti-tumor effect targeting PD-1 [101], and Her-2/neu-expressing solid tumors [62,100] and lung metastasis [62,102]. The use of overlapping peptides as mimotopes with T cell immunodominant epitopes has also been evaluated in a phase I immunotherapeutic trial against human papillomavirus 16, and shown tolerability and the induction of the T cell response, even in the patients with end-stage disease [110].

2.3.1. Her-2/neu B Cell Peptide/Mimotope

In line with active immunization/vaccination with B cell peptides [8,62,101–103], we have formulated the anti-Her-2/neu vaccine HER-Vaxx [111] consisting of a hybrid peptide (P467) [111,112]. The cross-reacting material CRM197, a mutated and nontoxic form of diphtheria toxin [113] with the capacity to activate B cells and CD4+ T cells with a heterogeneous Th1 and Th2 cytokine profile [114], is used as a carrier protein conjugated to the peptide, and the conjugate is administered with the Th1/Th2-driving adjuvant Montanide (ISA-51-VG) [62,102,103,111,115,116]. The Her-2/neu-directed B cell peptide-based vaccine HER-Vaxx has been evaluated in an array of preclinical experiments [102,111,112,117]. In phase 1b and Phase II trials involving patients suffering from Her-2/neu overexpressing metastatic or advanced adenocarcinoma of the stomach the vaccine was shown to be safe, immunogenic, and to prolong progression-free survival [118–120].



Figure 2. Schematic description of active immunotherapy with B cell peptides/mimotopes representing the B cell epitopes of therapeutic mAbs on TAAs, for example, a B cell peptide/mimotope of pertuzumab. The binding epitope of pertuzumab indicates the mAb B cell epitope on Her-2/neu. B cell peptides/mimotopes representing the mAb binding epitopes are identified, as described in the text and previously [62,101–103]. The selected B cell peptide/mimotope of the mAbs is conjugated to an immunogenic carrier protein, and an admixture-based vaccine comprising the conjugate and an adjuvant is then formulated. The formulated vaccine directly activates B cells for the production of antibodies in the host. The production of the antibodies is also induced further by the presence of T cell epitopes in the carrier protein, resulting in stimulation of T cells and further activation of B cells. The endogenously produced antibodies have the same functionality and modes of action as the respective therapeutic mAbs, such as pertuzumab. Adapted with permission from Ref. [62]. Elsevier, 2022.

Given that, in Her-2/neu-positive metastatic breast cancer patients, the combined treatment of trastuzumab with pertuzumab has established an incredible achievement, and to broaden the binding spectrum of the induced antibodies by our vaccine, we recently constructed a multi-peptide B cell vaccine comprising HER-Vaxx and pertuzumab's mimotope (HSGICELHCPALVTYNTDTFESMPNPEGRYTFGASCVTACPY; amino acids (AAs) 260–301, Uniprot P04626) and tested its capacity to prevent the formation of metastases in a mouse model with Her-2/neu lung metastases [62,102]. Active immunization with the multi-peptide vaccine combining HER-Vaxx and the mimotope of pertuzumab resulted in a significant reduction of lung metastasis formation, reflected by the reduction of lung weight and the size of the metastases, which was shown to be prominently associated with the formation of Her-2/neu-negative tumors with increased PD-L1 expression [102]. Because ICIs have been proposed to strengthen an emerging immune response following vaccination against TAAs, this observation suggests that a combination of a multi-peptide B cell Her-2/neu vaccine together with anti-PD-L1 ICI or active immunization with a mimotope from PD-L1 might serve as a suitable intervention to prevent metastasis formation.

2.3.2. PD-L1 B Cell Peptide/Mimotope

We recently used avelumab as one of the therapeutic mAbs targeting PD-L1 to identify its mimotope. On the basis of the crystal structure of human PD-L1 (hPD-L1) complexed with the single-chain Fv fragment of avelumab, it has been shown that the mAb binds to the IgV domain of the immune checkpoint [121]. By applying our platform for the identification of avelumab mimotope [101], 20-mer overlapping peptides spanning the IgV domain of hPD-L1 were tested and two B cell epitopes (EKQLDLAALIVYWEMEDKNIIQFVH, AAs 45-69; VYRCMISYGGADYKR, AAs 111–125; Uniprot Q9NZQ7) were selected. The peptide 'hybrid hPDL1-mimotope' consisting of the two mimotopes linked with a flexible glycine linker (GGGG) was generated. A cellular binding assay [122] using Jurkat reporter cells expressing hPD-L1 was employed to examine the inhibitory capacity of the hybrid peptide. As shown in Figure 3, avelumab alone could effectively bind to hPD-L1-expressing Jurkat cells. However, pre-incubation of the mAb with the peptide dose-dependently inhibited the binding of the mAb. On the basis of these data, in vivo investigations are ongoing to examine the effect of vaccination/active immunization with the hybrid peptide on inducing anti-tumor effects in a mouse model with syngeneic tumors expressing Her-2/neu [101].



Figure 3. Examination of the 'hybrid hPDL1-mimotope' in cellular assay. Jurkat T cells expressing hPD-L1 were used in a cellular assay to assess the binding of avelumab (30 ng/mL) alone or after pre-incubation with different concentrations of the examined peptide. The values represent the mean and standard deviation of the geometric mean of the fluorescence intensity (gMFI) of the viable population of the cells from three independent experiments. Significant differences are indicated by the respective *p*-values.

2.3.3. PD-1 B Cell Peptide/Mimotope

An alternative approach to target the PD-1/PD-L1 axis and block their interaction is by targeting PD-1. The sequences of human PD-1 (hPD-1) and mouse PD-1 (mPD-1) were used to identify the mimotopes JT-N1 (PGWFLDSPDRPWNPP; AAs 21–35, Uniprot Q15116) and JT-mPD1 (ISLHPKAKIEESPGA; AAs 126–140, Uniprot Q02242), respectively, and we have presented, for the first time, the concept of targeting PD-1 by vaccination using mimotopes [101,103]. A strong induced anti-tumor effect in vivo, to an extent similar to the corresponding mAb, was shown after active immunization with the mimotope [101]. The combination of the mimotope JT-mPD1 with the anti-Her-2/neu vaccine HER-Vaxx [111,120] led to an increase in the HER-Vaxx-generated anti-tumor effect [101]. In that manner, a strong anti-tumor effect in vivo was also shown by a peptide (PD-1-Vaxx) residing at the position 92–101 of hPD-1 [123]; the safety, tolerability and immunogenicity of the peptide as monotherapy in patients with PD-L1 expressing non-small cell lung cancer (NSCLC) is being evaluated in a phase I clinical trial [124].

The use of ICIs is often hampered by immune-related adverse events (irAEs) and hypersensitivity [92,93,125,126]. As mentioned above, vaccination with mimotopes/B cell peptides results in immunological memory. With this approach, targeting PD-1 or PD-L1 may result in the continuous inhibition of the PD-1/PD-L1 interaction and, thus, increased irAEs. Therefore, we were prompted to evaluate the safety of this approach in an influenza infection mouse model, as described in the next section.

2.3.4. Safety of B Cell Peptide/Mimotope-Based Vaccination Targeting PD-1

In 2018, a small study reported an unexpectedly high incidence (52% of 23 patients) of irAEs in influenza-vaccinated patients receiving anti-PD-1 inhibitors [127]. However, in a larger study with 127 lung cancer patients receiving nivolumab therapy, 47 patients who received vaccinations against influenza showed no difference in incidence or severity of irAEs [128]. In a retrospective study involving 370 patients receiving ICI therapy (nivolumab and pembrolizumab), 20% experienced an irAE of any grade [129]. These results were confirmed by a follow-up study [130]. Furthermore, in a small study involving 24 patients treated with anti-PD-1 or PD-L1 mAbs, the majority of the irAEs following vaccination with inactivated influenza vaccine were graded 1-2, therefore not requiring a change of ICI therapy [131]. Taken together, these results did not raise safety concerns regarding the application of an influenza vaccination in combination with ICI treatment. Nevertheless, we made use of the described observations and examined whether active immunization with a mimotope targeting PD-1 could affect the antiviral immune response. As described in Figure S1, an influenza infection model was established in BALB/c mice. A selected dose of 50 plaque-forming units (PFU) was used to examine whether active immunization with the mimotope from mouse PD-1 or treatment with the corresponding mAb (ICI) enhanced the antiviral cellular immune response. Following active immunization with the mimotope (JT-mPD1) or, for comparison, after the application of a functional anti-mouse PD-1 mAb, which was used for the identification of the mimotope [101], mice were infected with the influenza virus strain (A/PR/8/34) and sacrificed on days five and nine post-infection for clinical and immunological evaluations (Figure S2A).

As shown in Figure 4, the viral infection resulted in a significant decrease in body weight in all the infected mice around the peak day (day eight) of the infection. However, the observed body weight reduction was similar in all the infected groups, indicating that neither active immunizations with the mimotope nor the application of the corresponding mAb were associated with increased disease severity.

The level of viral load in the lungs of the infected mice was assessed based on the mRNA level of the influenza A virus matrix protein gene (M) and showed no significant difference between the viral loads in the infected groups of mice (Figure S2B).

Active immunization with the mimotope induced PD-1-specific serum IgG antibody response in the immunized mice, detected on days five and nine post-infection (Figure S2C). An induction of PD-1-specific serum IgG antibody response was also observed on day nine but not on day five post-infection in mice treated with the mAb (a rat anti-mouse PD-1 antibody) (Figure S2C). A similar observation was shown in clinical settings with patients treated with Nivolumab or Pembrolizumab [132], which might be attributed to a gradual induction of serum Abs against secreted soluble PD-1 as a consequence of the anti-PD-1 treatment. The evaluation of lymphoid (Figure S3A,B) and myeloid (Figure S4) cell populations, the mRNA level of the Th1 cytokine IFNgamma and the pro-inflammatory cytokines IL-6 and TNF (Figure S5) showed no significant difference between the infected mice, which were either untreated, actively immunized with the mimotope from PD-1 or treated with an anti-mPD-1 mAb. Overall, these findings suggest that active immunization with the mimotope is safe and is not associated with increased inflammatory responses, i.e., increased weight loss, an elevated influx of inflammatory immune cells or higher levels of pro-inflammatory cytokines.



Figure 4. On the day of the infection (day 0) and each day after the infection, the body weight of the mice was measured. Each point indicates an average of 5 mice, shown with SE. Significant differences are indicated by the respective *p*-values.

3. Further Compounds for Targeting Her-2/neu and ICs by Active and Passive Immunotherapies

As described above, the Her-2/neu-directed trastuzumab and pertuzumab have shown a tremendous effect in the clinic. However, the resistance to treatment with such mAbs has resulted in the development of additional mAbs. Increased trastuzumabmediated ADCC has been shown as a result of polymorphism in the Fc receptors ($Fc\gamma Rs$) of IgGs expressed on cytotoxic cells, affecting the ADDC capacity of the respective mAbs [133]. Accordingly, a chimeric anti-Her-2 mAb, margetuximab (MGAH22), with an Fc domain modification for improved binding to $Fc\gamma RIIIa$ was constructed [134]. On the basis of a first-in-human phase 1 study of margetuximab, in patients with Her-2/neu-positive advanced solid tumors, the mAb exhibited safety and an anti-tumor effect [135], and, in a phase 3 trial, the treatment with the mAb was associated with improved clinical outcomes in FcyRIIIa 158F allele carriers [136]. An FDA-approved trastuzumab-based drug conjugate, as a second-line therapy for the treatment of Her-2/neu-positive metastatic breast cancer patients with disease progression, is T-DM1 (Kadcyla®, Roche Pharma AG, Grenzach-Wyhlen, Germany), which is comprised of the mAb conjugated to DM1 (emtansine) [137,138]. In addition to retaining the ADCC activity of the mAb, the drug results in a cytotoxic effect by the delivered microtubule-inhibitory agent DM1 (derivative of maytansine) [137].

The tremendous success of the application of ICIs is also evident, as described above and by others [139–141]. However, the development of irAEs, fatal toxicities and resistance are the significant drawbacks of treatments with ICIs therapy [142,143], with the latter drawback often associated with decreased or loss of neoantigens immunogenicity, increased levels of immunosuppressive immune cells and also the upregulation of other ICs [144]. Therefore, targeting newer ICs associated with tumor microenvironment is a strategy to overcome the limitations of ICIs [145]. Such new ICs include lymphocyte activation gene-3 (LAG-3) [146], T cell immunoglobulin and ITIM domain (TIGIT) [147], T cell immunoglobulin and mucin-domain containing-3 (TIM-3) [148], V-domain immunoglobulin suppressor of T cell activation (VISTA) [149–151], B7 homolog 3 protein (B7-H3) [152] and inducible T cell costimulatory (ICOS) [153]. These ICs represent promising options for treating solid tumors, with clinical trials currently under active investigation for evaluating their effectiveness as monotherapy or combination therapy together with other ICIs [145]. An additional approach for combatting resistance to ICs treatment is by the use of bispecific antibodies [154,155]. FS118 is a novel, tetravalent (bispecific) antibody targeting LAG-3 and PD-L1 [156]. Treatment with the mAb resulted in a decreased expression of LAG-3 on T cells, and it was shed from the target cells, whereas the expression of the IC increased following treatment involving a combination of mAbs singularly targeting LAG-3 and PD-L1 [156]. A durable response to the mAb in a patient with anaplastic thyroid carcinoma who had progressed after PD-1 monotherapy was recently reported [157].

In line with the active immunotherapy/vaccination approach against Her-2/neu and ICs, the application of computer-aided analyses and X-ray structures of mAbs-bound proteins led to the identification of two B cell epitopes representing trastuzumab-binding and pertuzumab-binding epitopes [158,159]. A vaccine (B-Vaxx) comprising the two epitopes [158,159] was shown to be safe and immunogenic in a phase 1 trial [160]. With a similar approach, a vaccine (PD1-Vaxx) with a PD-1 B cell peptide linked to a measles virus fusion peptide via a four amino acid residue (GPSL) was constructed [160] and demonstrated synergistic vaccine combinations with a Her-2-targeted vaccine (B-Vaxx) [123]. Applying the same strategy, a PD-L1 B cell peptide vaccine (PDL1-Vaxx) was also constructed and in combination with a dual Her-2 B cell vaccine (B-Vaxx) was recently shown to induce potent immune responses and effective anti-tumor immunity in multiple syngeneic mice models [161]. In line with these results and our earlier findings showing that targeting Her-2/neu results in the loss of the receptor [102,120] associated with the upregulation of PD-L1 expression [102], two clinical trials were planned (Imugene Limited, Sydney, Australia) to assess: the combination of HER-Vaxx with chemotherapy or with anti-PD-1 antibody pembrolizumab in patients with Her-2/neu overexpressing gastric cancer who have failed treatment with trastuzumab and the combination of HER-Vaxx with chemotherapy \pm the anti-PD-L1 antibody avelumab in patients with Her-2/neu overexpressing gastric cancer.

4. Conclusions

The downregulation of T cell activation by the PD-1/PD-L1 axis, which is necessary for peripheral tolerance, can be exploited by tumor cells, resulting in the induction of an immunosuppressive state and their growth and immune escape. The application of therapeutic vaccines, based on immunogenic peptides inducing the production of antibodies in the host with a functionality similar to ICIs in inhibiting the PD-1/PD-L1 interaction, is a promising approach for immunotherapy. Additionally, such an approach might potentially overcome some of the disadvantages of the therapeutic mAbs as discussed in this review. A continuous inhibition of the PD-1/PD-L1 interaction may result from a peptide-based vaccination, consequently leading to immunological memory and elevated T cell activity. However, the results of our investigations did not indicate any safety concerns regarding the targeting of immune checkpoint PD-1 by active immunization, nor showed association with the hyper induction of pro-inflammatory cytokine production, known as 'cytokine storm' [162]. Nevertheless, the evaluation of the safety, immunogenicity and tolerability of vaccination with a PD-1 peptide (PD-1-Vaxx) is the aim of the ongoing phase 1 clinical trial (ClinicalTrials.gov Identifier: NCT04432207).

On the basis of the findings reviewed here, combined vaccination concepts co-targeting Her-2/neu and the PD-1 pathway seem to present a new treatment strategy to overcome mAbs above-mentioned associated disadvantages. These aspects are currently the topic of ongoing clinical trials, which will shed more light on the clinical outcomes of the discussed approach.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/cancers14225678/s1, Figure S1: Established influenza infection model in BALB/c mice infected with 50PFU of mouse-adapted influenza A/PR/8/34 virus, Figure S2: Immunization schedule, levels of mRNA expression and levels of antibodies, Figure S3: Distribution of immune cell profiles, Figure S4: Distribution of myeloid cell profiles in the examined mice measured by flow cytometry in lungs on day 5 and 9 after infection, Figure S5: The levels of mRNA expression of IFNgamma, IL-6 and TNF in lungs, measured by RT-PCR.

Author Contributions: Conceptualization, J.T. and U.W.; validation, J.T., P.S., M.K., S.K. and U.W.; formal analysis, J.T., M.D., C.B., P.S. and M.K.; methodology, J.T., M.D., A.S., A.H., M.L.W. and C.B.; investigation, J.T., M.D., P.S. and M.K.; resources, P.S., S.K. and U.W.; data curation, J.T.; writing—original draft preparation, J.T. and U.W.; writing—review and editing, J.T., M.D., C.B., P.S., E.G.-S., M.K., S.K. and C.C.Z.; visualization, J.T., M.D. and C.B.; supervision, J.T.; project administration, J.T. and U.W. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding. The presented pre-clinical studies were supported by a research grant from Imugene Ltd. (until 31 October 2020) to the Medical University of Vienna.

Data Availability Statement: The data presented in this study are available in the article and supplementary materials.

Conflicts of Interest: J.T., M.D., A.S., A.H., M.W., C.B., P.S., E.G., S., M.K. and S.K. declare no potential conflicts of interest; C.C.Z. was CSO of Imugene until June 2018; Consultancies and Speaker's Honoraria from Roche, Novartis, BMS, MSD, Imugene, Ariad, Pfizer, Merrimack, Merck KGaA, Fibrogen, AstraZeneca, Tesaro, Gilead, Servier, Shire, Eli Lilly, Athenex; Institution (CECOG): BMS, MSD, Pfizer, AstraZeneca; U.W. was CSO of Imugene until Sept. 2018 and has received from GSK, Pfizer and Themis, funding to the Institute as the principal investigator (PI) of clinical.

References

- 1. Esfahani, K.; Roudaia, L.; Buhlaiga, N.; Del Rincon, S.V.; Papneja, N.; Miller, W.H., Jr. A review of cancer immunotherapy: From the past, to the present, to the future. *Curr. Oncol.* **2020**, *27*, S87–S97. [CrossRef]
- Abbott, M.; Ustoyev, Y. Cancer and the Immune System: The History and Background of Immunotherapy. *Semin. Oncol. Nurs.* 2019, 35, 150923. [CrossRef] [PubMed]
- 3. Costa, R.L.B.; Czerniecki, B.J. Clinical development of immunotherapies for HER2(+) breast cancer: A review of HER2-directed monoclonal antibodies and beyond. *NPJ Breast Cancer* **2020**, *6*, 10. [CrossRef] [PubMed]
- Strebhardt, K.; Ullrich, A. Paul Ehrlich's magic bullet concept: 100 years of progress. Nat. Rev. Cancer 2008, 8, 473–480. [CrossRef] [PubMed]
- 5. Galluzzi, L.; Vacchelli, E.; Bravo-San Pedro, J.M.; Buque, A.; Senovilla, L.; Baracco, E.E.; Bloy, N.; Castoldi, F.; Abastado, J.P.; Agostinis, P.; et al. Classification of current anticancer immunotherapies. *Oncotarget* **2014**, *5*, 12472–12508. [CrossRef]
- 6. Lesterhuis, W.J.; Haanen, J.B.; Punt, C.J. Cancer immunotherapy—Revisited. Nat. Rev. Drug Discov. 2011, 10, 591–600. [CrossRef]
- 7. Pardoll, D.M. The blockade of immune checkpoints in cancer immunotherapy. *Nat. Rev. Cancer* 2012, *12*, 252–264. [CrossRef]
- 8. Wiedermann, U.; Davis, A.B.; Zielinski, C.C. Vaccination for the prevention and treatment of breast cancer with special focus on Her-2/neu peptide vaccines. *Breast Cancer Res. Treat.* **2013**, *138*, 1–12. [CrossRef]
- 9. Hafeez, U.; Parakh, S.; Gan, H.K.; Scott, A.M. Antibody-Drug Conjugates for Cancer Therapy. Molecules 2020, 25, 4764. [CrossRef]
- Parakh, S.; Parslow, A.C.; Gan, H.K.; Scott, A.M. Antibody-mediated delivery of therapeutics for cancer therapy. *Expert Opin.* Drug Deliv. 2016, 13, 401–419. [CrossRef]
- 11. Scott, A.M.; Wolchok, J.D.; Old, L.J. Antibody therapy of cancer. Nat. Rev. Cancer 2012, 12, 278–287. [CrossRef]
- 12. Weiner, L.M.; Dhodapkar, M.V.; Ferrone, S. Monoclonal antibodies for cancer immunotherapy. *Lancet* **2009**, *373*, 1033–1040. [CrossRef]
- Weiner, L.M.; Surana, R.; Wang, S. Monoclonal antibodies: Versatile platforms for cancer immunotherapy. *Nat. Rev. Immunol.* 2010, 10, 317–327. [CrossRef]
- 14. Lu, R.M.; Hwang, Y.C.; Liu, I.J.; Lee, C.C.; Tsai, H.Z.; Li, H.J.; Wu, H.C. Development of therapeutic antibodies for the treatment of diseases. *J. Biomed. Sci.* 2020, 27, 1. [CrossRef]
- 15. Sharpe, A.H. Mechanisms of costimulation. Immunol. Rev. 2009, 229, 5–11. [CrossRef]
- 16. Greenwald, R.J.; Freeman, G.J.; Sharpe, A.H. The B7 family revisited. Annu. Rev. Immunol. 2005, 23, 515–548. [CrossRef]
- 17. Zou, W.; Chen, L. Inhibitory B7-family molecules in the tumour microenvironment. Nat. Rev. Immunol. 2008, 8, 467–477. [CrossRef]
- Grywalska, E.; Pasiarski, M.; Gozdz, S.; Rolinski, J. Immune-checkpoint inhibitors for combating T-cell dysfunction in cancer. Onco Targets Ther. 2018, 11, 6505–6524. [CrossRef] [PubMed]
- 19. Saibil, S.D.; Ohashi, P.S. Targeting T cell activation in immuno-oncology. Curr. Oncol. 2020, 27, S98–S105. [CrossRef]
- Li, Y.; Li, F.; Jiang, F.; Lv, X.; Zhang, R.; Lu, A.; Zhang, G. A Mini-Review for Cancer Immunotherapy: Molecular Understanding of PD-1/PD-L1 Pathway & Translational Blockade of Immune Checkpoints. *Int. J. Mol. Sci.* 2016, 17, 1151. [CrossRef]
- 21. Reichert, J.M.; Rosensweig, C.J.; Faden, L.B.; Dewitz, M.C. Monoclonal antibody successes in the clinic. *Nat. Biotechnol.* 2005, 23, 1073–1078. [CrossRef] [PubMed]
- 22. Mullard, A. FDA approves 100th monoclonal antibody product. Nat. Rev. Drug Discov. 2021, 20, 491–495. [CrossRef] [PubMed]

- De Sousa Linhares, A.; Battin, C.; Jutz, S.; Leitner, J.; Hafner, C.; Tobias, J.; Wiedermann, U.; Kundi, M.; Zlabinger, G.J.; Grabmeier-Pfistershammer, K.; et al. Therapeutic PD-L1 antibodies are more effective than PD-1 antibodies in blocking PD-1/PD-L1 signaling. *Sci. Rep.* 2019, *9*, 11472. [CrossRef] [PubMed]
- Vaddepally, R.K.; Kharel, P.; Pandey, R.; Garje, R.; Chandra, A.B. Review of Indications of FDA-Approved Immune Checkpoint Inhibitors per NCCN Guidelines with the Level of Evidence. *Cancers* 2020, *12*, 738. [CrossRef] [PubMed]
- 25. Xu, C.; Chen, Y.P.; Du, X.J.; Liu, J.Q.; Huang, C.L.; Chen, L.; Zhou, G.Q.; Li, W.F.; Mao, Y.P.; Hsu, C.; et al. Comparative safety of immune checkpoint inhibitors in cancer: Systematic review and network meta-analysis. *BMJ* **2018**, *363*, k4226. [CrossRef]
- 26. Dine, J.; Gordon, R.; Shames, Y.; Kasler, M.K.; Barton-Burke, M. Immune Checkpoint Inhibitors: An Innovation in Immunotherapy for the Treatment and Management of Patients with Cancer. *Asia-Pac. J. Oncol. Nurs.* **2017**, *4*, 127–135. [CrossRef]
- 27. Granier, C.; De Guillebon, E.; Blanc, C.; Roussel, H.; Badoual, C.; Colin, E.; Saldmann, A.; Gey, A.; Oudard, S.; Tartour, E. Mechanisms of action and rationale for the use of checkpoint inhibitors in cancer. *ESMO Open* **2017**, *2*, e000213. [CrossRef]
- Iwai, Y.; Hamanishi, J.; Chamoto, K.; Honjo, T. Cancer immunotherapies targeting the PD-1 signaling pathway. J. Biomed. Sci. 2017, 24, 26. [CrossRef]
- Chenard-Poirier, M.; Smyth, E.C. Immune Checkpoint Inhibitors in the Treatment of Gastroesophageal Cancer. Drugs 2019, 79, 1–10. [CrossRef]
- Liu, X.; Qin, S. Immune Checkpoint Inhibitors in Hepatocellular Carcinoma: Opportunities and Challenges. *Oncologist* 2019, 24, S3–S10. [CrossRef]
- 31. Liu, Y.; Wu, L.; Tong, R.; Yang, F.; Yin, L.; Li, M.; You, L.; Xue, J.; Lu, Y. PD-1/PD-L1 Inhibitors in Cervical Cancer. *Front. Pharmacol.* **2019**, *10*, 65. [CrossRef]
- 32. Forster, M.D.; Devlin, M.J. Immune Checkpoint Inhibition in Head and Neck Cancer. Front. Oncol. 2018, 8, 310. [CrossRef]
- 33. Kim, H.S.; Seo, H.K. Immune checkpoint inhibitors for urothelial carcinoma. Investig. Clin. Urol. 2018, 59, 285–296. [CrossRef]
- 34. Shah, G.L.; Moskowitz, C.H. Checkpoint inhibition in lymphoma. Clin. Adv. Hematol. Oncol. 2018, 16, 45–55.
- 35. Ma, X.; Zhang, Y.; Wang, S.; Wei, H.; Yu, J. Immune checkpoint inhibitor (ICI) combination therapy compared to monotherapy in advanced solid cancer: A systematic review. *J. Cancer* **2021**, *12*, 1318–1333. [CrossRef]
- Hynes, N.E.; Lane, H.A. ERBB receptors and cancer: The complexity of targeted inhibitors. *Nat. Rev. Cancer* 2005, *5*, 341–354. [CrossRef]
- Iqbal, N.; Iqbal, N. Human Epidermal Growth Factor Receptor 2 (HER2) in Cancers: Overexpression and Therapeutic Implications. *Mol. Biol. Int.* 2014, 2014, 852748. [CrossRef]
- 38. Meza-Junco, J.; Sawyer, M.B. Metastatic gastric cancer—Focus on targeted therapies. *Biologics* **2012**, *6*, 137–146. [CrossRef]
- 39. Oh, D.Y.; Bang, Y.J. HER2-targeted therapies—A role beyond breast cancer. Nat. Rev. Clin. Oncol. 2020, 17, 33–48. [CrossRef]
- 40. Ross, J.S.; Slodkowska, E.A.; Symmans, W.F.; Pusztai, L.; Ravdin, P.M.; Hortobagyi, G.N. The HER-2 receptor and breast cancer: Ten years of targeted anti-HER-2 therapy and personalized medicine. *Oncologist* **2009**, *14*, 320–368. [CrossRef]
- Slamon, D.J.; Godolphin, W.; Jones, L.A.; Holt, J.A.; Wong, S.G.; Keith, D.E.; Levin, W.J.; Stuart, S.G.; Udove, J.; Ullrich, A.; et al. Studies of the HER-2/neu proto-oncogene in human breast and ovarian cancer. *Science* 1989, 244, 707–712. [CrossRef] [PubMed]
- 42. Arienti, C.; Pignatta, S.; Tesei, A. Epidermal Growth Factor Receptor Family and its Role in Gastric Cancer. *Front. Oncol.* 2019, *9*, 1308. [CrossRef] [PubMed]
- Cui, N.; Shi, J.; Yang, C. HER2-Based Immunotherapy for Breast Cancer. *Cancer Biother. Radiopharm.* 2018, 33, 169–175. [CrossRef] [PubMed]
- 44. Hynes, N.E.; Stern, D.F. The biology of erbB-2/neu/HER-2 and its role in cancer. *Biochim. Biophys. Acta* **1994**, *1198*, 165–184. [CrossRef] [PubMed]
- 45. Slamon, D.J.; Clark, G.M.; Wong, S.G.; Levin, W.J.; Ullrich, A.; McGuire, W.L. Human breast cancer: Correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science* **1987**, *235*, 177–182. [CrossRef]
- 46. Tai, W.; Mahato, R.; Cheng, K. The role of HER2 in cancer therapy and targeted drug delivery. J. Control. Release 2010, 146, 264–275. [CrossRef]
- 47. Carpenter, G. Receptors for epidermal growth factor and other polypeptide mitogens. *Annu. Rev. Biochem.* **1987**, *56*, 881–914. [CrossRef]
- 48. Lemmon, M.A.; Schlessinger, J. Cell signaling by receptor tyrosine kinases. Cell 2010, 141, 1117–1134. [CrossRef]
- 49. Citri, A.; Skaria, K.B.; Yarden, Y. The deaf and the dumb: The biology of ErbB-2 and ErbB-3. *Exp. Cell Res.* **2003**, *284*, 54–65. [CrossRef]
- 50. Citri, A.; Yarden, Y. EGF-ERBB signalling: Towards the systems level. Nat. Rev. Mol. Cell Biol. 2006, 7, 505–516. [CrossRef]
- 51. Moasser, M.M. The oncogene HER2: Its signaling and transforming functions and its role in human cancer pathogenesis. *Oncogene* **2007**, *26*, 6469–6487. [CrossRef]
- 52. Yarden, Y.; Sliwkowski, M.X. Untangling the ErbB signalling network. Nat. Rev. Mol. Cell Biol. 2001, 2, 127–137. [CrossRef]
- 53. Klapper, L.N.; Vaisman, N.; Hurwitz, E.; Pinkas-Kramarski, R.; Yarden, Y.; Sela, M. A subclass of tumor-inhibitory monoclonal antibodies to ErbB-2/HER2 blocks crosstalk with growth factor receptors. *Oncogene* **1997**, *14*, 2099–2109. [CrossRef]
- 54. Klapper, L.N.; Waterman, H.; Sela, M.; Yarden, Y. Tumor-inhibitory antibodies to HER-2/ErbB-2 may act by recruiting c-Cbl and enhancing ubiquitination of HER-2. *Cancer Res.* 2000, *60*, 3384–3388.

- Slamon, D.J.; Leyland-Jones, B.; Shak, S.; Fuchs, H.; Paton, V.; Bajamonde, A.; Fleming, T.; Eiermann, W.; Wolter, J.; Pegram, M.; et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *N. Engl. J. Med.* 2001, 344, 783–792. [CrossRef]
- Piccart-Gebhart, M.J.; Procter, M.; Leyland-Jones, B.; Goldhirsch, A.; Untch, M.; Smith, I.; Gianni, L.; Baselga, J.; Bell, R.; Jackisch, C.; et al. Trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer. N. Engl. J. Med. 2005, 353, 1659–1672. [CrossRef]
- Sakai, K.; Yokote, H.; Murakami-Murofushi, K.; Tamura, T.; Saijo, N.; Nishio, K. Pertuzumab, a novel HER dimerization inhibitor, inhibits the growth of human lung cancer cells mediated by the HER3 signaling pathway. *Cancer Sci.* 2007, *98*, 1498–1503. [CrossRef]
- Smith, M.B.; Reardon, J.; Olson, E.M. Pertuzumab for the treatment of patients with previously untreated HER2-positive metastatic breast cancer. *Drugs Today* 2012, 48, 713–722. [CrossRef]
- 59. Nahta, R.; Hung, M.C.; Esteva, F.J. The HER-2-targeting antibodies trastuzumab and pertuzumab synergistically inhibit the survival of breast cancer cells. *Cancer Res.* 2004, 64, 2343–2346. [CrossRef]
- 60. Scheuer, W.; Friess, T.; Burtscher, H.; Bossenmaier, B.; Endl, J.; Hasmann, M. Strongly enhanced antitumor activity of trastuzumab and pertuzumab combination treatment on HER2-positive human xenograft tumor models. *Cancer Res.* 2009, *69*, 9330–9336. [CrossRef]
- 61. Baselga, J.; Cortes, J.; Kim, S.B.; Im, S.A.; Hegg, R.; Im, Y.H.; Roman, L.; Pedrini, J.L.; Pienkowski, T.; Knott, A.; et al. Pertuzumab plus trastuzumab plus docetaxel for metastatic breast cancer. *N. Engl. J. Med.* **2012**, *366*, 109–119. [CrossRef] [PubMed]
- 62. Tobias, J.; Garner-Spitzer, E.; Drinic, M.; Wiedermann, U. Vaccination against Her-2/neu, with focus on peptide-based vaccines. *ESMO Open* **2022**, *7*, 100361. [CrossRef]
- 63. Chaganty, B.K.R.; Qiu, S.; Gest, A.; Lu, Y.; Ivan, C.; Calin, G.A.; Weiner, L.M.; Fan, Z. Trastuzumab upregulates PD-L1 as a potential mechanism of trastuzumab resistance through engagement of immune effector cells and stimulation of IFNgamma secretion. *Cancer Lett.* **2018**, *430*, 47–56. [CrossRef] [PubMed]
- 64. Baselga, J.; Albanell, J. Mechanism of action of anti-HER2 monoclonal antibodies. *Ann. Oncol.* 2001, 12 (Suppl. 1), S35–S41. [CrossRef] [PubMed]
- 65. Bianchini, G.; Gianni, L. The immune system and response to HER2-targeted treatment in breast cancer. *Lancet Oncol.* 2014, 15, e58–e68. [CrossRef]
- Yamashita, K.; Iwatsuki, M.; Yasuda-Yoshihara, N.; Morinaga, T.; Nakao, Y.; Harada, K.; Eto, K.; Kurashige, J.; Hiyoshi, Y.; Ishimoto, T.; et al. Trastuzumab upregulates programmed death ligand-1 expression through interaction with NK cells in gastric cancer. *Br. J. Cancer* 2021, 124, 595–603. [CrossRef]
- 67. Butte, M.J.; Keir, M.E.; Phamduy, T.B.; Sharpe, A.H.; Freeman, G.J. Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity* **2007**, *27*, 111–122. [CrossRef]
- Krasniqi, E.; Barchiesi, G.; Pizzuti, L.; Mazzotta, M.; Venuti, A.; Maugeri-Sacca, M.; Sanguineti, G.; Massimiani, G.; Sergi, D.; Carpano, S.; et al. Immunotherapy in HER2-positive breast cancer: State of the art and future perspectives. *J. Hematol. Oncol.* 2019, 12, 111. [CrossRef]
- 69. Pohlmann, P.R.; Mayer, I.A.; Mernaugh, R. Resistance to Trastuzumab in Breast Cancer. *Clin. Cancer Res.* 2009, 15, 7479–7491. [CrossRef]
- 70. Chung, A.; Cui, X.; Audeh, W.; Giuliano, A. Current status of anti-human epidermal growth factor receptor 2 therapies: Predicting and overcoming herceptin resistance. *Clin. Breast Cancer* **2013**, *13*, 223–232. [CrossRef]
- Derakhshani, A.; Rezaei, Z.; Safarpour, H.; Sabri, M.; Mir, A.; Sanati, M.A.; Vahidian, F.; Gholamiyan Moghadam, A.; Aghadoukht, A.; Hajiasgharzadeh, K.; et al. Overcoming trastuzumab resistance in HER2-positive breast cancer using combination therapy. J. Cell. Physiol. 2020, 235, 3142–3156. [CrossRef]
- 72. Loi, S.; Giobbie-Hurder, A.; Gombos, A.; Bachelot, T.; Hui, R.; Curigliano, G.; Campone, M.; Biganzoli, L.; Bonnefoi, H.; Jerusalem, G.; et al. Pembrolizumab plus trastuzumab in trastuzumab-resistant, advanced, HER2-positive breast cancer (PANACEA): A single-arm, multicentre, phase 1b-2 trial. *Lancet Oncol.* 2019, 20, 371–382. [CrossRef]
- 73. Peddi, P.F.; Hurvitz, S.A. Ado-trastuzumab emtansine (T-DM1) in human epidermal growth factor receptor 2 (HER2)-positive metastatic breast cancer: Latest evidence and clinical potential. *Ther. Adv. Med. Oncol.* **2014**, *6*, 202–209. [CrossRef]
- 74. Emens, L.A.; Esteva, F.J.; Beresford, M.; Saura, C.; De Laurentiis, M.; Kim, S.B.; Im, S.A.; Wang, Y.; Salgado, R.; Mani, A.; et al. Trastuzumab emtansine plus atezolizumab versus trastuzumab emtansine plus placebo in previously treated, HER2-positive advanced breast cancer (KATE2): A phase 2, multicentre, randomised, double-blind trial. *Lancet Oncol.* 2020, 21, 1283–1295. [CrossRef]
- 75. Davis, A.A.; Patel, V.G. The role of PD-L1 expression as a predictive biomarker: An analysis of all US Food and Drug Administration (FDA) approvals of immune checkpoint inhibitors. *J. Immunother. Cancer* **2019**, *7*, 278. [CrossRef]
- 76. Dirix, L.Y.; Takacs, I.; Jerusalem, G.; Nikolinakos, P.; Arkenau, H.T.; Forero-Torres, A.; Boccia, R.; Lippman, M.E.; Somer, R.; Smakal, M.; et al. Avelumab, an anti-PD-L1 antibody, in patients with locally advanced or metastatic breast cancer: A phase 1b JAVELIN Solid Tumor study. *Breast Cancer Res. Treat.* 2018, 167, 671–686. [CrossRef]

- 77. Hamilton, E.P.; Kaklamani, V.; Falkson, C.; Vidal, G.A.; Ward, P.J.; Patre, M.; Chui, S.Y.; Rotmensch, J.; Gupta, K.; Molinero, L.; et al. Atezolizumab in combination with trastuzumab emtansine or with trastuzumab and pertuzumab in patients with HER2-positive breast cancer and atezolizumab with doxorubicin and cyclophosphamide in HER2-negative breast cancer: Safety and biomarker outcomes from a multi-cohort Phase Ib study. *Cancer Res.* 2020, 80 (Suppl. 4), PD1–PD5. [CrossRef]
- 78. Crosby, E.J.; Acharya, C.R.; Haddad, A.F.; Rabiola, C.A.; Lei, G.; Wei, J.P.; Yang, X.Y.; Wang, T.; Liu, C.X.; Wagner, K.U.; et al. Stimulation of Oncogene-Specific Tumor-Infiltrating T Cells through Combined Vaccine and alphaPD-1 Enable Sustained Antitumor Responses against Established HER2 Breast Cancer. *Clin. Cancer Res.* 2020, *26*, 4670–4681. [CrossRef]
- 79. Cohen, E.E.W.; Pishvaian, M.J.; Shepard, D.R.; Wang, D.; Weiss, J.; Johnson, M.L.; Chung, C.H.; Chen, Y.; Huang, B.; Davis, C.B.; et al. A phase Ib study of utomilumab (PF-05082566) in combination with mogamulizumab in patients with advanced solid tumors. *J. Immunother. Cancer* **2019**, *7*, 342. [CrossRef]
- 80. Ovacik, M.; Lin, K. Tutorial on Monoclonal Antibody Pharmacokinetics and Its Considerations in Early Development. *Clin. Transl. Sci.* **2018**, *11*, 540–552. [CrossRef]
- Naran, K.; Nundalall, T.; Chetty, S.; Barth, S. Principles of Immunotherapy: Implications for Treatment Strategies in Cancer and Infectious Diseases. *Front. Microbiol.* 2018, 9, 3158. [CrossRef] [PubMed]
- 82. Scolnik, P.A. mAbs: A business perspective. *MAbs* 2009, 1, 179–184. [CrossRef] [PubMed]
- 83. Chames, P.; Van Regenmortel, M.; Weiss, E.; Baty, D. Therapeutic antibodies: Successes, limitations and hopes for the future. *Br. J. Pharmacol.* **2009**, *157*, 220–233. [CrossRef] [PubMed]
- Martins, F.; Sofiya, L.; Sykiotis, G.P.; Lamine, F.; Maillard, M.; Fraga, M.; Shabafrouz, K.; Ribi, C.; Cairoli, A.; Guex-Crosier, Y.; et al. Adverse effects of immune-checkpoint inhibitors: Epidemiology, management and surveillance. *Nat. Rev. Clin. Oncol.* 2019, 16, 563–580. [CrossRef] [PubMed]
- Fares, C.M.; Van Allen, E.M.; Drake, C.G.; Allison, J.P.; Hu-Lieskovan, S. Mechanisms of Resistance to Immune Checkpoint Blockade: Why Does Checkpoint Inhibitor Immunotherapy Not Work for All Patients? *Am. Soc. Clin. Oncol. Educ. Book* 2019, 39, 147–164. [CrossRef]
- 86. Verma, V.; Sprave, T.; Haque, W.; Simone, C.B., 2nd; Chang, J.Y.; Welsh, J.W.; Thomas, C.R., Jr. A systematic review of the cost and cost-effectiveness studies of immune checkpoint inhibitors. *J. Immunother. Cancer* **2018**, *6*, 128. [CrossRef]
- 87. van Holstein, Y.; Kapiteijn, E.; Bastiaannet, E.; van den Bos, F.; Portielje, J.; de Glas, N.A. Efficacy and Adverse Events of Immunotherapy with Checkpoint Inhibitors in Older Patients with Cancer. *Drugs Aging* **2019**, *36*, 927–938. [CrossRef]
- 88. Ishii, K.; Morii, N.; Yamashiro, H. Pertuzumab in the treatment of HER2-positive breast cancer: An evidence-based review of its safety, efficacy, and place in therapy. *Core Evid.* **2019**, *14*, 51–70. [CrossRef]
- 89. Bettaieb, A.; Paul, C.; Plenchette, S.; Shan, J.; Chouchane, L.; Ghiringhelli, F. Precision medicine in breast cancer: Reality or utopia? J. Transl. Med. 2017, 15, 139. [CrossRef]
- Oude Munnink, T.H.; Henstra, M.J.; Segerink, L.I.; Movig, K.L.; Brummelhuis-Visser, P. Therapeutic drug monitoring of monoclonal antibodies in inflammatory and malignant disease: Translating TNF-alpha experience to oncology. *Clin. Pharmacol. Ther.* 2016, 99, 419–431. [CrossRef]
- Torka, P.; Barth, M.; Ferdman, R.; Hernandez-Ilizaliturri, F.J. Mechanisms of Resistance to Monoclonal Antibodies (mAbs) in Lymphoid Malignancies. *Curr. Hematol. Malig. Rep.* 2019, 14, 426–438. [CrossRef]
- Isabwe, G.A.C.; Garcia Neuer, M.; de Las Vecillas Sanchez, L.; Lynch, D.M.; Marquis, K.; Castells, M. Hypersensitivity reactions to therapeutic monoclonal antibodies: Phenotypes and endotypes. J. Allergy Clin. Immunol. 2018, 142, 159–170.e2. [CrossRef]
- 93. Pintea, I.; Petricau, C.; Dumitrascu, D.; Muntean, A.; Branisteanu, D.C.; Branisteanu, D.E.; Deleanu, D. Hypersensitivity reactions to monoclonal antibodies: Classification and treatment approach (Review). *Exp. Ther. Med.* **2021**, *22*, 949. [CrossRef]
- Hendrikx, J.; Haanen, J.; Voest, E.E.; Schellens, J.H.M.; Huitema, A.D.R.; Beijnen, J.H. Fixed Dosing of Monoclonal Antibodies in Oncology. Oncologist 2017, 22, 1212–1221. [CrossRef]
- 95. Price, L.; Brunt, A.M. Trastuzumab infusion reactions in breast cancer. Should we routinely observe after the first dose? *Eur. J. Hosp. Pharm.* **2018**, *25*, 331–333. [CrossRef]
- 96. Pollard, A.J.; Bijker, E.M. A guide to vaccinology: From basic principles to new developments. *Nat. Rev. Immunol.* 2021, 21, 83–100. [CrossRef]
- 97. Geysen, H.M.; Rodda, S.J.; Mason, T.J. A priori delineation of a peptide which mimics a discontinuous antigenic determinant. *Mol. Immunol.* **1986**, *23*, 709–715. [CrossRef]
- Zhang, L.; Huang, Y.; Lindstrom, A.R.; Lin, T.Y.; Lam, K.S.; Li, Y. Peptide-based materials for cancer immunotherapy. *Theranostics* 2019, 9, 7807–7825. [CrossRef]
- Riemer, A.B.; Bramswig, K.H.; Quaglino, E.; Forni, G.; Pehamberger, H.; Scheiner, O.; Zielinski, C.C.; Jensen-Jarolim, E. Vaccination with trastuzumab epitope mimics leads to prolonged tumor-free survival in HER-2/neu transgenic mice. *Cancer Res.* 2006, 66, 151–152.
- 100. Singer, J.; Manzano-Szalai, K.; Fazekas, J.; Thell, K.; Bentley-Lukschal, A.; Stremnitzer, C.; Roth-Walter, F.; Weghofer, M.; Ritter, M.; Pino Tossi, K.; et al. Proof of concept study with an HER-2 mimotope anticancer vaccine deduced from a novel AAV-mimotope library platform. *Oncoimmunology* 2016, 5, e1171446. [CrossRef]
- 101. Tobias, J.; Battin, C.; De Sousa Linhares, A.; Lebens, M.; Baier, K.; Ambroz, K.; Drinic, M.; Hogler, S.; Inic-Kanada, A.; Garner-Spitzer, E.; et al. A New Strategy Toward B Cell-Based Cancer Vaccines by Active Immunization With Mimotopes of Immune Checkpoint Inhibitors. *Front. Immunol.* 2020, 11, 895. [CrossRef] [PubMed]

- 102. Tobias, J.; Drinic, M.; Hogler, S.; Ambroz, K.; Baier, K.; Kodajova, P.; Tomasich, E.; Berghoff, A.S.; Schmid, A.; Garner-Spitzer, E.; et al. Active immunization with a Her-2/neu-targeting Multi-peptide B cell vaccine prevents lung metastases formation from Her-2/neu breast cancer in a mouse model. *Transl. Oncol.* 2022, *19*, 101378. [CrossRef] [PubMed]
- Tobias, J.; Steinberger, P.; Drinic, M.; Wiedermann, U. Emerging targets for anticancer vaccination: PD-1. ESMO Open 2021, 6, 100278. [CrossRef]
- Kaumaya, P.T. B-cell epitope peptide cancer vaccines: A new paradigm for combination immunotherapies with novel checkpoint peptide vaccine. *Future Oncol.* 2020, 16, 1767–1791. [CrossRef] [PubMed]
- 105. Knittelfelder, R.; Riemer, A.B.; Jensen-Jarolim, E. Mimotope vaccination–from allergy to cancer. *Expert Opin. Biol. Ther.* 2009, *9*, 493–506. [CrossRef]
- 106. Leung, N.Y.H.; Wai, C.Y.Y.; Chu, K.H.; Leung, P.S.C. Mimotope-based allergen-specific immunotherapy: Ready for prime time? *Cell. Mol. Immunol.* 2019, 16, 890–891. [CrossRef]
- 107. Charalambous, B.M.; Feavers, I.M. Mimotope vaccines. J. Med. Microbiol. 2001, 50, 937–939. [CrossRef]
- Pashov, A.D.; Plaxco, J.; Kaveri, S.V.; Monzavi-Karbassi, B.; Harn, D.; Kieber-Emmons, T. Multiple antigenic mimotopes of HIV carbohydrate antigens: Relating structure and antigenicity. J. Biol. Chem. 2006, 281, 29675–29683. [CrossRef]
- 109. Steward, M.W. The development of a mimotope-based synthetic peptide vaccine against respiratory syncytial virus. *Biologicals* **2001**, *29*, 215–219. [CrossRef]
- 110. Kenter, G.G.; Welters, M.J.; Valentijn, A.R.; Lowik, M.J.; Berends-van der Meer, D.M.; Vloon, A.P.; Drijfhout, J.W.; Wafelman, A.R.; Oostendorp, J.; Fleuren, G.J.; et al. Phase I immunotherapeutic trial with long peptides spanning the E6 and E7 sequences of high-risk human papillomavirus 16 in end-stage cervical cancer patients shows low toxicity and robust immunogenicity. *Clin. Cancer Res.* 2008, 14, 169–177. [CrossRef]
- 111. Tobias, J.; Jasinska, J.; Baier, K.; Kundi, M.; Ede, N.; Zielinski, C.; Wiedermann, U. Enhanced and long term immunogenicity of a Her-2/neu multi-epitope vaccine conjugated to the carrier CRM197 in conjunction with the adjuvant Montanide. *BMC Cancer* 2017, 17, 118. [CrossRef]
- 112. Jasinska, J.; Wagner, S.; Radauer, C.; Sedivy, R.; Brodowicz, T.; Wiltschke, C.; Breiteneder, H.; Pehamberger, H.; Scheiner, O.; Wiedermann, U.; et al. Inhibition of tumor cell growth by antibodies induced after vaccination with peptides derived from the extracellular domain of Her-2/neu. *Int. J. Cancer* 2003, *107*, 976–983. [CrossRef]
- 113. Malito, E.; Bursulaya, B.; Chen, C.; Lo Surdo, P.; Picchianti, M.; Balducci, E.; Biancucci, M.; Brock, A.; Berti, F.; Bottomley, M.J.; et al. Structural basis for lack of toxicity of the diphtheria toxin mutant CRM197. *Proc. Natl. Acad. Sci. USA* 2012, 109, 5229–5234. [CrossRef]
- 114. Kamboj, K.K.; King, C.L.; Greenspan, N.S.; Kirchner, H.L.; Schreiber, J.R. Immunization with Haemophilus influenzae type b-CRM₁₉₇ conjugate vaccine elicits a mixed Th1 and Th2 CD4⁺ T cell cytokine response that correlates with the isotype of antipolysaccharide antibody. J. Infect. Dis. 2001, 184, 931–935. [CrossRef]
- 115. Habjanec, L.; Halassy, B.; Tomašić, J. Immunomodulatory activity of novel adjuvant formulations based on Montanide ISA oil-based adjuvants and peptidoglycan monomer. *Int. Immunopharmacol.* **2008**, *8*, 717–724. [CrossRef]
- 116. Ascarateil, S.; Puget, A.; Koziol, M.-E. Safety data of Montanide ISA 51 VG and Montanide ISA 720 VG, two adjuvants dedicated to human therapeutic vaccines. *J. Immunother. Cancer* **2015**, *3* (Suppl. 2), P428. [CrossRef]
- 117. Wagner, S.; Jasinska, J.; Breiteneder, H.; Kundi, M.; Pehamberger, H.; Scheiner, O.; Zielinski, C.C.; Wiedermann, U. Delayed tumor onset and reduced tumor growth progression after immunization with a Her-2/neu multi-peptide vaccine and IL-12 in c-neu transgenic mice. *Breast Cancer Res. Treat.* 2007, 106, 29–38. [CrossRef]
- 118. Maglakelidze, M.; Ryspayeva, D.; Bulat, I.; Andric, Z.; Nikolic, I.; Chawla, T.; Nagarkar, R.; Chourdhary, V.; Venkata, G.; Singh, R.K.; et al. A Phase 1b/2 Open-Label Study with Randomization in Phase 2 of Imu-131 Her2/Neu Peptide Vaccine Plus Standard of Care Chemotherapy in Patients with Her2/Neu Overexpressing Metastatic or Advanced Adenocarcinoma of the Stomach or Gastroesophageal Junction. *Cancer Res.* 2021, *81*, CT107. [CrossRef]
- 119. Wiedermann, U.; Wiltschke, C.; Jasinska, J.; Kundi, M.; Zurbriggen, R.; Garner-Spitzer, E.; Bartsch, R.; Steger, G.; Pehamberger, H.; Scheiner, O.; et al. A virosomal formulated Her-2/neu multi-peptide vaccine induces Her-2/neu-specific immune responses in patients with metastatic breast cancer: A phase I study. *Breast Cancer Res. Treat.* 2010, 119, 673–683. [CrossRef]
- 120. Wiedermann, U.; Garner-Spitzer, E.; Chao, Y.; Maglakelidze, M.; Bulat, I.; Dechaphunkul, A.; Arpornwirat, W.; Charoentum, C.; Yen, C.J.; Yau, T.C.; et al. Clinical and Immunologic Responses to a B-Cell Epitope Vaccine in Patients with HER2/neu-Overexpressing Advanced Gastric Cancer-Results from Phase Ib Trial IMU.ACS.001. *Clin. Cancer Res.* 2021, 27, 3649–3660. [CrossRef]
- 121. Liu, K.; Tan, S.; Chai, Y.; Chen, D.; Song, H.; Zhang, C.W.; Shi, Y.; Liu, J.; Tan, W.; Lyu, J.; et al. Structural basis of anti-PD-L1 monoclonal antibody avelumab for tumor therapy. *Cell Res.* **2017**, *27*, 151–153. [CrossRef] [PubMed]
- 122. Jutz, S.; Hennig, A.; Paster, W.; Asrak, O.; Dijanovic, D.; Kellner, F.; Pickl, W.F.; Huppa, J.B.; Leitner, J.; Steinberger, P. A cellular platform for the evaluation of immune checkpoint molecules. *Oncotarget* **2017**, *8*, 64892–64906. [CrossRef] [PubMed]
- 123. Kaumaya, P.T.P.; Guo, L.; Overholser, J.; Penichet, M.L.; Bekaii-Saab, T. Immunogenicity and antitumor efficacy of a novel human PD-1 B-cell vaccine (PD1-Vaxx) and combination immunotherapy with dual trastuzumab/pertuzumab-like HER-2 B-cell epitope vaccines (B-Vaxx) in a syngeneic mouse model. *Oncoimmunology* 2020, *9*, 1818437. [CrossRef] [PubMed]

- 124. Guo, L.; Kaumaya, P.T.P. First prototype checkpoint inhibitor B-cell epitope vaccine (PD1-Vaxx) en route to human Phase 1 clinical trial in Australia and USA: Exploiting future novel synergistic vaccine combinations. *Br. J. Cancer* 2021, *125*, 152–154. [CrossRef] [PubMed]
- Chen, T.W.; Razak, A.R.; Bedard, P.L.; Siu, L.L.; Hansen, A.R. A systematic review of immune-related adverse event reporting in clinical trials of immune checkpoint inhibitors. *Ann. Oncol.* 2015, 26, 1824–1829. [CrossRef]
- 126. Ellithi, M.; Elnair, R.; Chang, G.V.; Abdallah, M.A. Toxicities of Immune Checkpoint Inhibitors: Itis-Ending Adverse Reactions and More. *Cureus* 2020, 12, e6935. [CrossRef]
- 127. Laubli, H.; Balmelli, C.; Kaufmann, L.; Stanczak, M.; Syedbasha, M.; Vogt, D.; Hertig, A.; Muller, B.; Gautschi, O.; Stenner, F.; et al. Influenza vaccination of cancer patients during PD-1 blockade induces serological protection but may raise the risk for immune-related adverse events. *J. Immunother. Cancer* **2018**, *6*, 40. [CrossRef]
- 128. Wijn, D.H.; Groeneveld, G.H.; Vollaard, A.M.; Muller, M.; Wallinga, J.; Gelderblom, H.; Smit, E.F. Influenza vaccination in patients with lung cancer receiving anti-programmed death receptor 1 immunotherapy does not induce immune-related adverse events. *Eur. J. Cancer* 2018, 104, 182–187. [CrossRef]
- Chong, C.R.; Park, V.J.; Cohen, B.; Postow, M.A.; Wolchok, J.D.; Kamboj, M. Safety of Inactivated Influenza Vaccine in Cancer Patients Receiving Immune Checkpoint Inhibitors. *Clin. Infect. Dis.* 2020, 70, 193–199. [CrossRef]
- Failing, J.J.; Ho, T.P.; Yadav, S.; Majithia, N.; Riaz, I.B.; Shin, J.Y.; Schenk, E.L.; Xie, H. Safety of Influenza Vaccine in Patients With Cancer Receiving Pembrolizumab. JCO Oncol. Pract. 2020, 16, e573–e580. [CrossRef]
- Gwynn, M.E.; DeRemer, D.L.; Saunders, K.M.; Parikh, J.; Bollag, R.J.; Clemmons, A.B. Immune-mediated adverse events following influenza vaccine in cancer patients receiving immune checkpoint inhibitors. J. Oncol. Pharm. Pract. 2020, 26, 647–654. [CrossRef]
- 132. Ohkuma, R.; Ieguchi, K.; Watanabe, M.; Takayanagi, D.; Goshima, T.; Onoue, R.; Hamada, K.; Kubota, Y.; Horiike, A.; Ishiguro, T.; et al. Increased Plasma Soluble PD-1 Concentration Correlates with Disease Progression in Patients with Cancer Treated with Anti-PD-1 Antibodies. *Biomedicines* 2021, *9*, 1929. [CrossRef]
- 133. Musolino, A.; Naldi, N.; Dieci, M.V.; Zanoni, D.; Rimanti, A.; Boggiani, D.; Sgargi, P.; Generali, D.G.; Piacentini, F.; Ambroggi, M.; et al. Immunoglobulin G fragment C receptor polymorphisms and efficacy of preoperative chemotherapy plus trastuzumab and lapatinib in HER2-positive breast cancer. *Pharmacogenom. J.* 2016, *16*, 472–477. [CrossRef]
- 134. Nordstrom, J.L.; Gorlatov, S.; Zhang, W.; Yang, Y.; Huang, L.; Burke, S.; Li, H.; Ciccarone, V.; Zhang, T.; Stavenhagen, J.; et al. Anti-tumor activity and toxicokinetics analysis of MGAH22, an anti-HER2 monoclonal antibody with enhanced Fcgamma receptor binding properties. *Breast Cancer Res.* 2011, *13*, R123. [CrossRef]
- 135. Bang, Y.J.; Giaccone, G.; Im, S.A.; Oh, D.Y.; Bauer, T.M.; Nordstrom, J.L.; Li, H.; Chichili, G.R.; Moore, P.A.; Hong, S.; et al. First-in-human phase 1 study of margetuximab (MGAH22), an Fc-modified chimeric monoclonal antibody, in patients with HER2-positive advanced solid tumors. *Ann. Oncol.* 2017, *28*, 855–861. [CrossRef]
- 136. Rugo, H.S.; Im, S.A.; Wright, G.L.S.; Escriva-de-Romani, S.; DeLaurentiis, M.; Cortes, J.; Bahadur, S.W.; Haley, B.B.; Oyola, R.H.; Riseberg, D.A.; et al. SOPHIA primary analysis: A phase 3 (P3) study of margetuximab (M) plus chemotherapy (C) versus trastuzumab (T) plus C in patients (pts) with HER2+metastatic (met) breast cancer (MBC) after prior anti-HER2 therapies (Tx). J. Clin. Oncol. 2019, 37, 1000. [CrossRef]
- 137. Verma, S.; Miles, D.; Gianni, L.; Krop, I.E.; Welslau, M.; Baselga, J.; Pegram, M.; Oh, D.Y.; Dieras, V.; Guardino, E.; et al. Trastuzumab emtansine for HER2-positive advanced breast cancer. *N. Engl. J. Med.* **2012**, *367*, 1783–1791. [CrossRef]
- 138. Wedam, S.; Fashoyin-Aje, L.; Gao, X.; Bloomquist, E.; Tang, S.; Sridhara, R.; Goldberg, K.B.; King-Kallimanis, B.L.; Theoret, M.R.; Ibrahim, A.; et al. FDA Approval Summary: Ado-Trastuzumab Emtansine for the Adjuvant Treatment of HER2-positive Early Breast Cancer. *Clin. Cancer Res.* 2020, 26, 4180–4185. [CrossRef]
- 139. Jacob, J.B.; Jacob, M.K.; Parajuli, P. Review of immune checkpoint inhibitors in immuno-oncology. *Adv. Pharmacol.* **2021**, *91*, 111–139. [CrossRef]
- 140. Ribas, A.; Wolchok, J.D. Cancer immunotherapy using checkpoint blockade. *Science* 2018, 359, 1350–1355. [CrossRef]
- 141. Robert, C. A decade of immune-checkpoint inhibitors in cancer therapy. Nat. Commun. 2020, 11, 3801. [CrossRef] [PubMed]
- 142. Johnson, D.B.; Nebhan, C.A.; Moslehi, J.J.; Balko, J.M. Immune-checkpoint inhibitors: Long-term implications of toxicity. *Nat. Rev. Clin. Oncol.* 2022, 19, 254–267. [CrossRef] [PubMed]
- Schoenfeld, A.J.; Hellmann, M.D. Acquired Resistance to Immune Checkpoint Inhibitors. *Cancer Cell* 2020, 37, 443–455. [CrossRef] [PubMed]
- Sharma, P.; Hu-Lieskovan, S.; Wargo, J.A.; Ribas, A. Primary, Adaptive, and Acquired Resistance to Cancer Immunotherapy. *Cell* 2017, 168, 707–723. [CrossRef] [PubMed]
- 145. Lee, J.B.; Ha, S.J.; Kim, H.R. Clinical Insights Into Novel Immune Checkpoint Inhibitors. *Front. Pharmacol.* **2021**, *12*, 681320. [CrossRef]
- 146. Atkinson, V.; Khattak, A.; Haydon, A.; Eastgate, M.; Roy, A.; Prithviraj, P.; Mueller, C.; Brignone, C.; Triebel, F. Eftilagimod alpha, a soluble lymphocyte activation gene-3 (LAG-3) protein plus pembrolizumab in patients with metastatic melanoma. *J. Immunother. Cancer* **2020**, *8*, e001681. [CrossRef]
- 147. Kong, Y.; Zhu, L.; Schell, T.D.; Zhang, J.; Claxton, D.F.; Ehmann, W.C.; Rybka, W.B.; George, M.R.; Zeng, H.; Zheng, H. T-Cell Immunoglobulin and ITIM Domain (TIGIT) Associates with CD8+ T-Cell Exhaustion and Poor Clinical Outcome in AML Patients. *Clin. Cancer Res.* 2016, 22, 3057–3066. [CrossRef]

- 148. Acharya, N.; Sabatos-Peyton, C.; Anderson, A.C. Tim-3 finds its place in the cancer immunotherapy landscape. *J. Immunother. Cancer* 2020, *8*, e000911. [CrossRef]
- Tagliamento, M.; Agostinetto, E.; Borea, R.; Brandao, M.; Poggio, F.; Addeo, A.; Lambertini, M. VISTA: A Promising Target for Cancer Immunotherapy? *Immunotargets Ther.* 2021, 10, 185–200. [CrossRef]
- 150. Yuan, L.; Tatineni, J.; Mahoney, K.M.; Freeman, G.J. VISTA: A Mediator of Quiescence and a Promising Target in Cancer Immunotherapy. *Trends Immunol.* 2021, 42, 209–227. [CrossRef]
- 151. Yum, J.E.I.; Hong, Y.K. Terminating Cancer by Blocking VISTA as a Novel Immunotherapy: Hasta la vista, baby. *Front. Oncol.* **2021**, *11*, 658488. [CrossRef]
- 152. Liu, C.; Zhang, G.; Xiang, K.; Kim, Y.; Lavoie, R.R.; Lucien, F.; Wen, T. Targeting the immune checkpoint B7-H3 for next-generation cancer immunotherapy. *Cancer Immunol. Immunother.* **2022**, *71*, 1549–1567. [CrossRef]
- 153. Yap, T.A.; Burris, H.A.; Kummar, S.; Falchook, G.S.; Pachynski, R.K.; LoRusso, P.; Tykodi, S.S.; Gibney, G.T.; Gainor, J.F.; Rahma, O.E.; et al. ICONIC: Biologic and clinical activity of first in class ICOS agonist antibody JTX-2011±nivolumab (nivo) in patients (pts) with advanced cancers. J. Clin. Oncol. 2018, 36, 3000. [CrossRef]
- 154. Ma, J.; Mo, Y.; Tang, M.; Shen, J.; Qi, Y.; Zhao, W.; Huang, Y.; Xu, Y.; Qian, C. Bispecific Antibodies: From Research to Clinical Application. *Front. Immunol.* **2021**, *12*, 626616. [CrossRef]
- 155. Zhou, X.; Einsele, H.; Danhof, S. Bispecific Antibodies: A New Era of Treatment for Multiple Myeloma. J. Clin. Med. 2020, 9, 2166. [CrossRef]
- 156. Kraman, M.; Faroudi, M.; Allen, N.L.; Kmiecik, K.; Gliddon, D.; Seal, C.; Koers, A.; Wydro, M.M.; Batey, S.; Winnewisser, J.; et al. FS118, a Bispecific Antibody Targeting LAG-3 and PD-L1, Enhances T-Cell Activation Resulting in Potent Antitumor Activity. *Clin. Cancer Res.* 2020, 26, 3333–3344. [CrossRef]
- Kroloff, M.J.; Holz, J.B.; Stern, O.; Shepherd, C.J.; Morrow, M.; Kayitalire, L.; Wong, D.J. Durable response of anaplastic thyroid carcinoma to FS118, a bispecific LAG-3/PD-L1 antibody, after checkpoint inhibitor progression: A case report. *J. Immunother. Cancer* 2022, *10*, e005225. [CrossRef]
- Allen, S.D.; Garrett, J.T.; Rawale, S.V.; Jones, A.L.; Phillips, G.; Forni, G.; Morris, J.C.; Oshima, R.G.; Kaumaya, P.T. Peptide vaccines of the HER-2/neu dimerization loop are effective in inhibiting mammary tumor growth in vivo. *J. Immunol.* 2007, 179, 472–482. [CrossRef]
- Garrett, J.T.; Rawale, S.; Allen, S.D.; Phillips, G.; Forni, G.; Morris, J.C.; Kaumaya, P.T. Novel engineered trastuzumab conformational epitopes demonstrate in vitro and in vivo antitumor properties against HER-2/neu. *J. Immunol.* 2007, 178, 7120–7131. [CrossRef]
- 160. Bekaii-Saab, T.; Wesolowski, R.; Ahn, D.H.; Wu, C.; Mortazavi, A.; Lustberg, M.; Ramaswamy, B.; Fowler, J.; Wei, L.; Overholser, J.; et al. Phase I Immunotherapy Trial with Two Chimeric HER-2 B-Cell Peptide Vaccines Emulsified in Montanide ISA 720VG and Nor-MDP Adjuvant in Patients with Advanced Solid Tumors. *Clin. Cancer Res.* 2019, 25, 3495–3507. [CrossRef]
- 161. Guo, L.; Overholser, J.; Darby, H.; Ede, N.J.; Kaumaya, P.T.P. A newly discovered PD-L1 B-cell epitope peptide vaccine (PDL1-Vaxx) exhibits potent immune responses and effective anti-tumor immunity in multiple syngeneic mice models and (synergizes) in combination with a dual HER-2 B-cell vaccine (B-Vaxx). Oncoimmunology 2022, 11, 2127691. [CrossRef] [PubMed]
- 162. Ryabkova, V.A.; Churilov, L.P.; Shoenfeld, Y. Influenza infection, SARS, MERS and COVID-19: Cytokine storm—The common denominator and the lessons to be learned. *Clin. Immunol.* **2021**, 223, 108652. [CrossRef] [PubMed]