

## STUDIES

# Comparative transcriptome, physiological and biochemical analyses reveal response mechanism mediated by CBF4 and ICE2 in enhancing cold stress tolerance in *Gossypium thurberi*

Xiaoyan Cai<sup>1,2,†</sup>, Richard Odongo Magwanga<sup>1,3,†</sup>, Yanchao Xu<sup>1,†</sup>, Zhongli Zhou<sup>1</sup>, Xingxing Wang<sup>1</sup>, Yuqing Hou<sup>1</sup>, Yuhong Wang<sup>1</sup>, Yuanming Zhang<sup>2\*</sup>, Fang Liu<sup>1\*</sup> and Kunbo Wang<sup>1\*</sup>

<sup>1</sup>State Key Laboratory of Cotton Biology /Institute of Cotton Research, Chinese Academy of Agricultural Science, Anyang, Henan 455000, China, <sup>2</sup>College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei 430070, China, <sup>3</sup>School of Biological and Physical Sciences (SBPS), Jaramogi Oginga Odinga University of Science and Technology (JOUST), P.O. Box 210, Bondo 40601, Kenya

<sup>†</sup>Authors with equal contribution.

\*Corresponding authors' e-mail addresses: [liufcri@163.com](mailto:liufcri@163.com); [soy Zhang@mail.hzau.edu.cn](mailto:soy Zhang@mail.hzau.edu.cn); [wkbcri@163.com](mailto:wkbcri@163.com)

Associate Editor: Golam Jalal Ahamed

**Citation:** Cai X, Magwanga RO, Xu Y, Zhou Z, Wang X, Hou Y, Wang Y, Zhang Y, Liu F, Wang K. 2019. Comparative transcriptome, physiological and biochemical analyses reveal response mechanism mediated by CBF4 and ICE2 in enhancing cold stress tolerance in *Gossypium thurberi*. *AoB PLANTS* 11: plz045; doi: 10.1093/aobpla/plz045

## Abstract.

Low temperature is one of the key environmental stresses that impair plant growth and significantly restricts the productivity and spatial distribution of crop plants. *Gossypium thurberi*, a wild diploid cotton species, has adapted to a wide range of temperatures and exhibits a better tolerance to chilling stress. Here, we compared phenotypes and physiochemical changes in *G. thurberi* under cold stress and found this species indeed showed better cold tolerance. Therefore, to understand the molecular mechanisms of the cold tolerance in *G. thurberi*, we compared transcription changes in leaves of *G. thurberi* under cold stress by high-throughput transcriptome sequencing. In total, 35 617 unigenes were identified in the whole-genome transcription profile, and 4226 differentially expressed genes (DEGs) were discovered in the leaves upon cold treatment. Gene Ontology (GO) classification analyses showed that the majority of DEGs belonged to categories of signal transduction, transcription factors (TFs) and carbohydrate transport and metabolism. The expression of several cold-responsive genes such as *ICE1*, *CBF4*, *RAP2-7* and abscisic acid (ABA) biosynthesis genes involved in different signalling pathways were induced after *G. thurberi* seedlings were exposed to cold stress. Furthermore, cold sensitivity was increased in *CBF4* and *ICE2* virus-induced gene silencing (VIGS) plants, and high level of malondialdehyde (MDA) showed that the *CBF4* and *ICE2* silenced plants were under oxidative stress compared to their wild types, which relatively had higher levels of antioxidant enzyme activity, as evident by high levels of proline and superoxide dismutase (SOD) content. In conclusion, our findings reveal a new regulatory network of cold stress response in *G. thurberi* and broaden our understanding of the cold tolerance mechanism in cotton, which might accelerate functional genomics studies and genetic improvement for cold stress tolerance in cultivated cotton.

**Keywords:** Cold tolerance; gene expression; *G. thurberi*; signalling pathways; transcriptome.

Received: 7 January, 2019; Editorial decision: 10 July, 2019; Accepted: 18 July, 2019

© The Authors 2019. Published by Oxford University Press on behalf of the Annals of Botany Company.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

## Introduction

Low temperature is one of the key environmental stresses, which drastically impairs plant growth and significantly restricts the productivity and spatial distribution of crop plants (Zhou et al. 2011). There are two types of the low temperature stress which affect plant growth and development, namely chilling (0–20 °C) and freezing, which occurs when the temperature falls below zero (Song et al. 2015). With the effects of global warming and drastic weather changes, the low temperature has become one of the major forms of abiotic stress with enormous effects on plant growth and development (Hasanuzzaman et al. 2013). The low temperature of the field environment restricted the plant's vegetative growth and delayed all the phenological stages in comparison to plants grown under controlled environment (Barrero-Gil et al. 2016). Apart from this, it led to some vegetative aberrations like chlorosis, necrosis of leaf tips and curling of whole leaves. The damage to the reproductive stage involved the abscission of juvenile buds, flowers, abortion of bolls and in extreme conditions that cause plant death (Bolek 2010). Plants have developed several response mechanisms to various forms of abiotic stressors, including chilling and freezing stresses. The most common phenomenon is acclimation, which mainly occurs through enhanced tolerance to environmental stress after priming treatment (Shi et al. 2018). The plant stress response is regulated at the molecular level (Iba 2002) and induces several changes in biochemical pathways and physiological processes of stressed plants (Bartwal et al. 2013). Generally, plants from temperate regions, including *Arabidopsis*, winter wheat, barley and oilseed rape, show an increase in freezing tolerance in response to low non-freezing temperatures, a phenomenon known as cold acclimation (Miura and Furumoto 2013). By contrast, some important crops such as rice, maize, tomato and cotton, originated from the tropical and subtropical climates, cannot to acclimatize to extreme temperatures and are therefore sensitive to chilling stress (Zhao et al. 2015).

When plants are exposed to any form of stress, the internal equilibrium dynamism rapidly changes, leading to excessive production of reactive oxygen species (ROS), which causes extensive damage to various cell structures. The response of plants to low temperature stress is a highly complex process involving multiple levels of regulation. When plants are exposed to cold stress, a series of physiological and biochemical alterations at the molecular and cellular levels are induced to increase plant cold tolerance and survival under cold stress conditions. These changes include increased accumulation of ROS-scavenging enzymes, rapid alterations in malondialdehyde (MDA) content, modifications in lipid compositions such as an increase in unsaturated fatty acid content which affects cell membrane fluidity, changes in protein and carbohydrate composition, accumulations of anti-freeze and anti-oxidative substances such as soluble sugar (SS), proline (PRO), superoxide dismutase (SOD), peroxidase (POD), catalase (CAT) among others (Hussain et al. 2016). After decades of intensive studies, we have begun to understand the cold-responsive (COR) system in plants at the transcriptional level (Wang et al. 2017). The C-repeat binding factor/dehydration-responsive element-binding factors (CBF/DREB) are critical transcription factors (TFs) which positively regulate the expression of downstream COR genes during cold acclimation. Also, CBF/DREB genes are activated by inducers of CBF expression (ICEs) through specific binding to the MYC recognition cis-elements in their promoter. The ICE-CBF-COR signal regulatory pathway has been identified as a critical COR transcriptional pathway during cold acclimation

in many plants (Lee and Thomashow 2012). In *Arabidopsis*, there are three members of the CBFs (CBF1/DREB1B, CBF2/DREB1C and CBF3/DREB1A), encoding DNA-binding proteins of the AP2/ERF (APETALA2(AP2)/ethylene response factor) superfamily, which are rapidly induced in response to low temperature (Zhou et al. 2011). The CBF4 gene, a novel homolog of CBF/DREB1 genes from *Arabidopsis*, was initially induced by abscisic acid (ABA) treatment under drought but not low temperature stress. Similarly, the CBF4 gene is induced by cold stress in grape (Xiao et al. 2008). Moreover, CBF/DREB1 transcription activators are critical regulators of gene expression in the signal transduction of cold acclimation in *Arabidopsis* (Haake 2002). It is evident that the CBF cold response pathway has a major role in low temperature stress, especially during cold acclimation in temperate plants. But in the herbaceous monocot rice (*Oryza sativa*), which originated in the tropics, a novel MYBS3-dependent pathway has been identified as essential for cold tolerance. Molecular evidence indicated that the sequential expression of CBF and MYBS3 provided two complementary mechanisms for conferring cold tolerance, with the CBF-mediated process initiating the immediate cold shock response and the MYBS3-mediated system adjusting the long-term cold adaptation in rice (Su et al. 2010). Recently, a significant contribution of ROS-mediated gene regulation, rather than the CBF regulation, to a more vigorous transcriptional stress response in rice was revealed by RNA-Seq. The study revealed a ROS-dominated dynamic model underlying chilling environment adaptation and tolerance in rice (Zhang et al. 2016).

Cotton (*Gossypium* spp.), originally from tropical and subtropical climates, is the most important fibre crop for the textile industries and ranks highly among the edible crop oil producing crops in the world is sensitive to low temperature during its whole growth period, but is especially affected at the germination and seedling stages (Shan et al. 2007). The genus *Gossypium* includes 46 diploids ( $2n = 2x = 26$ ) and 6 tetraploids ( $2n = 4x = 52$ ) species, all the diploid cotton species subsequently diversified to produce eight groups, including groups A–G and K (Chakravarthy et al. 2012). All tetraploid cotton species originated through polyploidization between the A-genome and the D-genome species (Renny-Byfield et al. 2016). The A-genome species have experienced a long period of artificial selection for their high-quality spinnable fibre, whereas the D-genome species under natural selection maintained abundant genetic diversity and a number of desirable characters, such as fibre quality and resistance to salt, heat, drought, cold, insects and diseases (Kirungu et al. 2018). *Gossypium thurberi*, a wild diploid cotton species, is endemic to the desert sands of the southern part of Arizona in America and has adapted to the local temperature climate. The species is highly tolerant to low temperature and can survive low temperature as low as –6 °C (Liang et al. 1996). Researchers have identified some significant genes related to chilling tolerance in cotton, such as GhDREB1 (Shan et al. 2007), GhTIP1 (Li et al. 2009), GhMT3a (Xue et al. 2009), phospholipase D $\alpha$  (Kargiotidou et al. 2010), GbCBF1 (Guo et al. 2011), GbPATP (Liu et al. 2015) and GhTPS11 (Wang et al. 2016). The overexpression of DREB/CBF genes has been found to confer cold tolerance to transgenic plants (Kang et al. 2013). Moreover, several CBL genes have been identified in cotton, through blast search of the CBL proteins from various dicotyledonous plants such as the reported cotyledons CBF sequences of *Arabidopsis* (Fowler et al. 2005), *Brassica napus* (Jing et al. 2008), soybean (Li et al. 2005; Kidokoro et al. 2015), among others plants in order to maximize the CBF genes discovery. In which 21 CBF genes were identified and classified into GhCBF I, GhCBF II, GhCBF III

and GhCBF IV (Ma et al. 2014). These studies provided useful clues for understanding the mechanism of cold tolerance in cotton. However, due to the limited genomic sequences, these studies failed to provide a comprehensive interpretation of the transcriptomic changes in cotton in response to cold stress acclimation. To gain insight into the molecular networks underlying cotton chilling tolerance, more comprehensive genome-wide gene expression profiling studies are required.

In recent years, the whole-genome sequences of *G. raimondii* from D-genome and other three cultivated species such as *G. arboreum*, *G. hirsutum* and *G. barbadense* have been sequenced (Yuan et al. 2015). And therefore provides excellent genomic materials for better understanding the cold tolerance and transcriptome response mechanisms in cotton. In this study, a well-established whole-genome transcriptome analysis method based on RNA-Seq and incorporating real-time PCR was carried out to screen the differential gene expression in leaves of *G. thurberi* under cold treatment. We identified hundreds of COR genes and provided an overall overview of the regulatory network in response to cold stress in *G. thurberi*. These COR genes could be the potential candidate TFs for further functional validation and application in breeding for versatile and highly cold-tolerant cotton genotypes.

## Materials and Methods

### Plant material, growth conditions and cold treatments

Three cotton accessions were used in this research, *Gossypium thurberi* and two accessions derived from tetraploid cotton, *G. hirsutum*. *Gossypium thurberi* is a diploid cotton of the D-genome, it is known for its ability to tolerate cold stress (Kirungu et al. 2018), the other two are cultivated upland cotton, *G. hirsutum* cultivars, CR150 and XLZ33, with differing ability to cold stress, CR150 is more tolerant to cold compared to ZLZ33. The seeds were obtained from the National Wild Cotton Nursery (Hainan, Sanya, China), maintained by the Institute of Cotton Research, Chinese Academy of Agricultural Sciences, China. The seeds were sterilized with 8 % sodium hypochlorite for 30 min and then washed with sterile water. Planting was carried out in germination boxes filled with sterilized sand, in a germination chamber, temperature set at 28 °C for 3 days to maximize germination. After germination, the seedlings were then transferred to small pots (bottom dimension 5 cm, top dimension 7 cm, depth 8 cm) containing well-watered mixtures of vermiculite and humus mixed in the ratio of 1:1. The seedlings were then moved to well-conditioned growth chambers, with 12-h light/dark and temperatures at 28 °C. For cold stress treatment, 3-week-old seedlings were moved to the cold treatment chamber, with conditions set at 12-h light/dark and 4 °C, the controlled seedlings were grown under normal condition. The leaf samples from cold stress treatment and controlled plants were collected at 0, 3, 6, 12, 24 and 48 h, and then stored under -80 °C awaiting RNA extraction and biochemical analysis. To minimize variations, 10 individuals' plants were pooled into one independent biological replicate.

### Measurement of physiological and biochemical parameters in the three cotton species exposed to cold stress conditions

The relative electrolyte (ions) leakage from the cotton leaves was determined as described by Lu et al. (2018). Uniform sections of the leaves were obtained by the use of a punching

machine with a diameter set at 1 cm, the leaf sections were then washed three times with sterilized deionized water. Twenty leaf segments were then placed in tubes containing 10 mL of sterilized deionized water and incubated at 25 °C. Four hours later, the electrical conductivity of the bathing solution (L1) was measured. The tubes were then autoclaved at 100 °C for 20 min, cooled to 25 °C before the final electrical conductivity (L2) was measured. The relative electrolyte/ion leakage was calculated by the formula =  $(L1 - L0)/(L2 - L0) \times 100$  (L0 = conductivity of deionized water).

Malondialdehyde levels were assessed by measuring thiobarbituric acid-reactive substances (TBARS). Superoxide dismutase, POD, PRO, soluble protein (SP) and SS were quantified as described by Mahmood et al. (2011). The concentrations of ABA, ethylene (ETH), gibberellic acid (GA) and indole acetic acid (IAA) in leaves were determined using the ELISA method using the following kits ABA (CK-E00005P, DG, Beijing, China), ETH (CK-E00021P, DG, Beijing, China), GA (CK-E00001P, DG, Beijing, China), IAA (CKE00015P, DG, Beijing, China), these four kinds of kits were utilized for the determination of each hormone contents. For the analysis 0.5 grams for each sample was used to determine concentration of the hormones through ELISA (Infinite M1000, TECAN, Switzerland). Three replicated experiments (two time technical repeats per biological replicate) were performed.

### RNA isolation, cDNA library preparation and RNA-Seq

Total RNA was isolated using the RNeasy Plant Mini Kit (Qiagen). The quantity and quality of RNA were checked by electrophoresis on 1 % agarose gels and NanoDrop 2000. RNA concentrations were measured using the Qubit RNA Assay Kit in Qubit 2.0 Fluorometer (Life Technologies, Carlsbad, CA, USA). The RNA integrity was assessed using the RNA Nano 6000 Assay Kit of the Agilent Bioanalyzer 2100 system (Agilent Technologies, Santa Clara, CA, USA). A total amount of 1 µg RNA per sample was used as input material for the RNA sample preparations. Sequencing libraries were generated using NEB Next Ultra TM RNA Library Prep Kit for Illumina (New England Biolabs Ltd, Beijing China) following the manufacturer's recommendations and index codes were added to attribute sequences to each sample. The clustering of the index-coded samples was performed on a cBot Cluster Generation System using TruSeq PE Cluster Kit v4-cBot-HS (Illumina) according to the manufacturer's instructions. After cluster generation, the library preparations were sequenced on an Illumina HiSeq 2500 platform and paired-end reads were generated.

### Bioinformatics analysis of RNA-Seq data

Raw data of fastq format were processed through in-house perl scripts to obtain clean data. The clean data were obtained by removing reads containing adapter ploy-N and low-quality reads from raw data. At the same time, the Q20 and GC content of the clean data were calculated. All the downstream analyses were based on clean data with high quality. Reference genome and gene model annotation files were downloaded from the genome website (<http://mascotton.njau.edu.cn/html/Data>) (Zhang et al. 2015). Index of the reference genome was built using Bowtie v2.0.6, and paired-end clean reads were aligned to the reference genome using Tophat2 (v2.0.9) software (Kim et al. 2013). Gene functions were annotated based on the following databases: Nr (NCBI non-redundant protein sequences), Nt (NCBI non-redundant nucleotide sequences), Pfam (Protein family), KOG/COG (Clusters of Orthologous Groups of proteins), Swiss-Prot (a

manually annotated and reviewed protein sequence database), KEGG (Kyoto Encyclopedia of Genes and Genomes) orthology database (Kanehisa et al. 2008), GO (Gene Ontology). Gene Ontology enrichment analysis of the differentially expressed genes (DEGs) was implemented by the GSeq (Ashburner et al. 2000).

### Validation of RNA-Seq data by real-time RT-qPCR

A real-time RT-PCR (RT-qPCR) was performed. A total of 22 genes were selected from up-regulated, down-regulated and non-expressed genes (insignificantly expressed genes). The gene specific primers were designed by the use of primer 5 and *GrActin* gene forward sequence 'ATCCTCGTCTAGACCTTG' and reverse sequence 'TGTCATCAGGCAACTCAT' was used as an internal control [see Supporting Information—Table S1]. The RT-qPCR was performed using a Bio-Rad CFX96 Real-Time instrument and the LightCycler FastStart DNA Master SYBR Green I kit (Roche, Basel, Switzerland) on a new set of three replicates for each sample.

### Functional characterization of the candidate genes through virus-induced gene silencing in *G. thurberi*

The TRV2 (tobacco rattle virus) vectors were prepared and introduced into *Agrobacterium tumefaciens* strain GV4104 (Gao et al. 2011). To monitor the silencing efficiency, the TRV2-PDS vector was constructed as a visual marker. Primers used to generate TRV vector, for the CBF4 gene forward sequence 'GTGAGTAAGTTACC GAATTCGGTTGATTCTGGGTCGGTTTC' and the reverse sequence 'CGTGAGC TCGGTACCGGATCCGACTTCTTATTA' and the second gene, ICE2 forward sequence 'GTGAGTAAGTTACCGAATTCTGG TTATCAGGTGGAGG' and the reverse sequence 'CGTGAGCTCGG TACCGGATCCAATGAAAT', were inserted into the vector pTRV2 of tobacco rattle virus (TRV) via EcoRI and BamHI digestion sites to construct a 35S promoter-driven pTRV2:CBF4 and pTRV2:ICE2, respectively. The recombinant vector was transformed into the competent cells of *A. tumefaciens* LBA4404 by freeze-thaw method, and the bacterium LBA4404 containing pTRV1 vector was used as auxiliary bacteria, being pTRV1 can replicate and move systemically without RNA2, mainly constructed from a T-DNA vector containing duplicated CaMV 35S promoter, nopaline synthase (NOS) terminator and cDNA clone of TRV RNA1 of Ppk20 strain (Liu et al. 2002). The cotyledons of cotton seedlings were infected by injection, and pTRV:PDS (phytoene desaturase) was used as a positive control. Plants without infection and empty vector pTRV:00 were used as negative control. The *Agrobacterium* culture was agroinfiltrated into the two expanded cotyledons of 10-day-old soil-grown seedling of *G. thurberi*. The cotton seedlings were then grown in a greenhouse condition, with temperature and light cycles set at 28 °C, and 16-h light/8-h dark cycle, respectively. At least 24 seedlings were inoculated for each construct with three replications, with eight plants per replication. At 14 days after *Agrobacterium* inoculation, and when virus-induced gene silencing (VIGS) was established in >75 % of the seedlings 14 days after *Agrobacterium* inoculation, as determined by the whitening/albino nature of the plant leaves. The silenced seedlings were subjected to cold stress treatment; this was done at the three true leaf stages. Cold stress was imposed on the wild types, positive control with empty vector and the gene silenced seedlings, by transferring them to a cold chamber with the temperature set at 4 °C for 3 days. After 3 days of cold stress treatment, the leaf samples were collected and MDA, PRO and SOD assayed.

### Statistical analysis

The plant's responses to cold stress were determined through morphological, physiological, biochemical and transcriptome

analysis. A complete randomized block design (CRBD) was used. All the results presented are given as means with standard errors. All analyses were undertaken using IBM SPSS version 25 (IBM SPSS 2017). Normality and homogeneity of variance were analysed by Kolmogorov–Smirnov and Levene's tests. The significance of the results was assessed using independent samples t-test or two-way ANOVA.

## Results

### Phenotypic responses to cold stress

To accurately evaluate the tolerance of *G. thurberi*, we compared the phenotypic changes of *G. thurberi* versus two accessions of *G. hirsutum*, CRI50 and XLZ33, which are tolerant and sensitive to cold stress, respectively. *Gossypium thurberi* is the northernmost species of *Gossypium*, reaching southern Arizona. Since it occurs at elevations as high as 2000 m, it is growing in a temperate zone climate, the only *Gossypium* to do so. It achieves this adaptation by undergoing leaf senescence and abscission (simultaneously with fruit maturation) in the autumn and becoming fully dormant by the winter months. In this case, the move towards dormancy is not triggered by declining moisture supplies in the soil, but rather by the advancing season (day length, temperature regime or some combination of factors) (Walker and Natwick 2006). A significant phenotypic difference was observed in XLZ33, its leaves wilted after 12 h of cold treatment, while CRI50 and *G. thurberi* leaves were not affected. Moreover, after 48 h of cold stress treatment, ZLZ33 were severely necrotic and wilted, while *G. hirsutum* CRI50 and *G. thurberi* leaves only showed symptoms of wilting but were not necrotic (Fig. 1). After 48 h of cold stress treatment, a high number of ZLZ33 seedlings died, while *G. thurberi* and CRI50 seedlings achieved 100 % recovery. These observations suggested that *G. thurberi* had a higher ability to tolerate cold stress compared to the tetraploid cotton, *G. hirsutum*. The results obtained correlated positively with previous findings, which showed that *G. thurberi* has a higher ability to tolerate cold stress (Walker and Natwick 2006).

### Physiological and biochemical analysis in the leaf tissues of *G. thurberi* and two other upland cotton under cold stress condition

Variations of the physiological and biochemical indices *thurberi* among the three cotton species, *G. thurberi*, ZLZ33 and CRI50 seedlings, were investigated after cold stress treatment. The cell membrane stability is a significant trait in determining the tolerance level of a plant to various abiotic stress factors (Mansour 2013). In evaluating the cell membrane stability through relative electrolyte leakage, the electrolyte leakage increased from 23.2 to 55.8 % after 48 h of cold stress exposure (Fig. 2A). Furthermore, the MDA content increased over time, but the increasing tendency slowed down after 12 h of cold treatment in *G. thurberi* but the MDA levels in the two upland cotton, CRI50 and XZL33, were higher, though relatively low in CRI50 compared to XZL33 at time 0 h, lower at 12 h and not statistically different at 24 and 48 h (Fig. 2B), an indication that the two upland cotton accession of the upland cotton species had a similar capacity to tolerate the effect of cold stress. These observations suggested an element of acclimation to cold stress by *G. thurberi* seedlings after 12 h of cold exposure, which enabled them to protect their membrane stability as evidenced by the low level of electrolyte leakage. Moreover, *G. thurberi* exhibited significantly increased levels of various antioxidant enzymes analysed, such as POD, SOD, PRO and SS after cold treatment at 4 °C compared to the

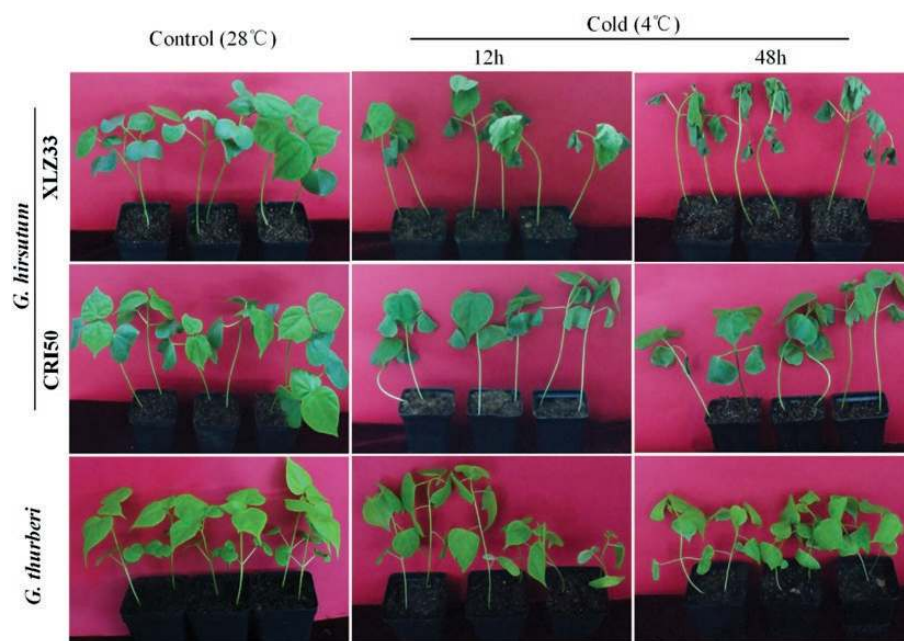


Figure 1. Phenotypes of 3-week seedlings of *G. thurberi* and two *G. hirsutum* accessions under cold stress (4 °C) for 48 h. Seedlings grown under normal conditions (28 °C) were used as controls. *Gossypium hirsutum*-CRI50 is cold-tolerant accession and *G. hirsutum*-XLZ33 is cold-sensitive accession.

other two upland cotton species, the POD and SOD concentration levels between *G. thurberi* and the two upland cotton accessions varied but were not statistically different at 0, 12 and 24 h, more so between *G. thurberi* and *G. hirsutum*-CRI50, but as the time of stress exposure increased, significant differences were observed among the three cotton species at 48 h of cold stress exposure, and the three cotton species showed significant differences with *G. thurberi* registering the highest levels of both POD and SOD antioxidant enzymes concentrations (Fig. 2C and D). While in evaluating the PRO content among the three cotton species, CRI50 and *G. thurberi* showed no significant differences at 0 and 12 h; however, at 24 and 48 h of cold stress exposure, the three cotton species exhibited significant differences in the level of PRO content, with *G. thurberi* registering the highest concentration levels (Fig. 2E). About the SS, no significant differences were observed at 0 and 12 h, but variations were observed at 24 and 48 h, with high levels of SS in the leaves of *G. thurberi* compared to the two upland cotton accessions, CR50 and XLZ33 (Fig. 2F). The increased concentration levels of the antioxidant and SS in the leaf of *G. thurberi* showed that *G. thurberi* had the ability to tolerate very low temperatures compared to the other two upland cotton species, when plants are exposed to cold or chilling conditions of temperatures between 0 and 15 °C, hydrogen peroxide ( $H_2O_2$ ) may accumulate (Kocsy et al. 2001). Thus, the increased levels of antioxidant enzymes could be playing an important role in protecting the plant cells from ROS damage (Caverzan et al. 2016). The results are in agreement with previous findings in which various antioxidant enzymes are more in the leaves of stress-tolerant genotypes compared to susceptible varieties when they are exposed to either abiotic or biotic stress conditions (Sairam et al. 1997). Moreover, knockdown of two novel CYP450 genes in upland cotton increased drought and salt stress susceptibility as evident by increased levels of oxidant and significant decline in antioxidant enzymes in the leaves of the VIGS plants (Magwanga et al. 2019a). Reactive oxygen species are oxygen-containing substances of metabolites and their derivatives generated through oxidation in plants, directly

or indirectly. Reactive oxygen species play a vital role in stress signal transduction, but excessive active oxygen oxidation harms the plant (Gill and Tuteja 2010). Therefore, plants also produce a series of active oxygen-scavenging enzymes, for example, POD, CAT, SOD among others.

### Plant phytochemical assays on the tissues of wild cotton, *G. thurberi* under cold stress conditions

In elucidating the inherent traits in *G. thurberi* which enables them to be more tolerant to cold stress, we analysed various plant phytohormones, such as gibberellic hormones (GA), ABA, ETH and IAA. The ability of the plants to regulate the phytohormones productions, distribution and or signal transmission, do aids their ability to coordinate and regulate growth and in turn increases their level of tolerance to various forms of abiotic stress factors (Colebrook et al. 2014). Ethylene hormone (ETH) at 0 h was relatively low across the three cotton species; however, with the increase in stress exposure, the evolution of ETH hormone increased rapidly hitting the pick after 24 h of cold stress exposure then showed a gradual decline at 48 h (Fig. 3A). A similar observation was made in grapevine in which ETH concentration increased and normalized with an increase in cold stress exposure (Sun et al. 2016). Abscisic acid in relation to GA and/or IAA work antagonistically, increase in ABA leads to a significant decline in either GA or IAA (Peleg and Blumwald 2011). In the evaluation of the three antagonistic plant hormones, ABA, GA and IAA, ABA showed a gradual increase across the three cotton species; though lower than the concentration levels of GA and IAA (Fig. 3B–D). The results obtained were in agreement with previous findings in which various hormones such as GA have been found to playing a significant role in enhancing cold stress tolerance in plants (Colebrook et al. 2014). Moreover, emission of ETH plants at low concentration levels is an adaptive mechanism for plants to avoid the lethal damage by the environmental stressors; furthermore, ETH also do regulates other diverse activities such

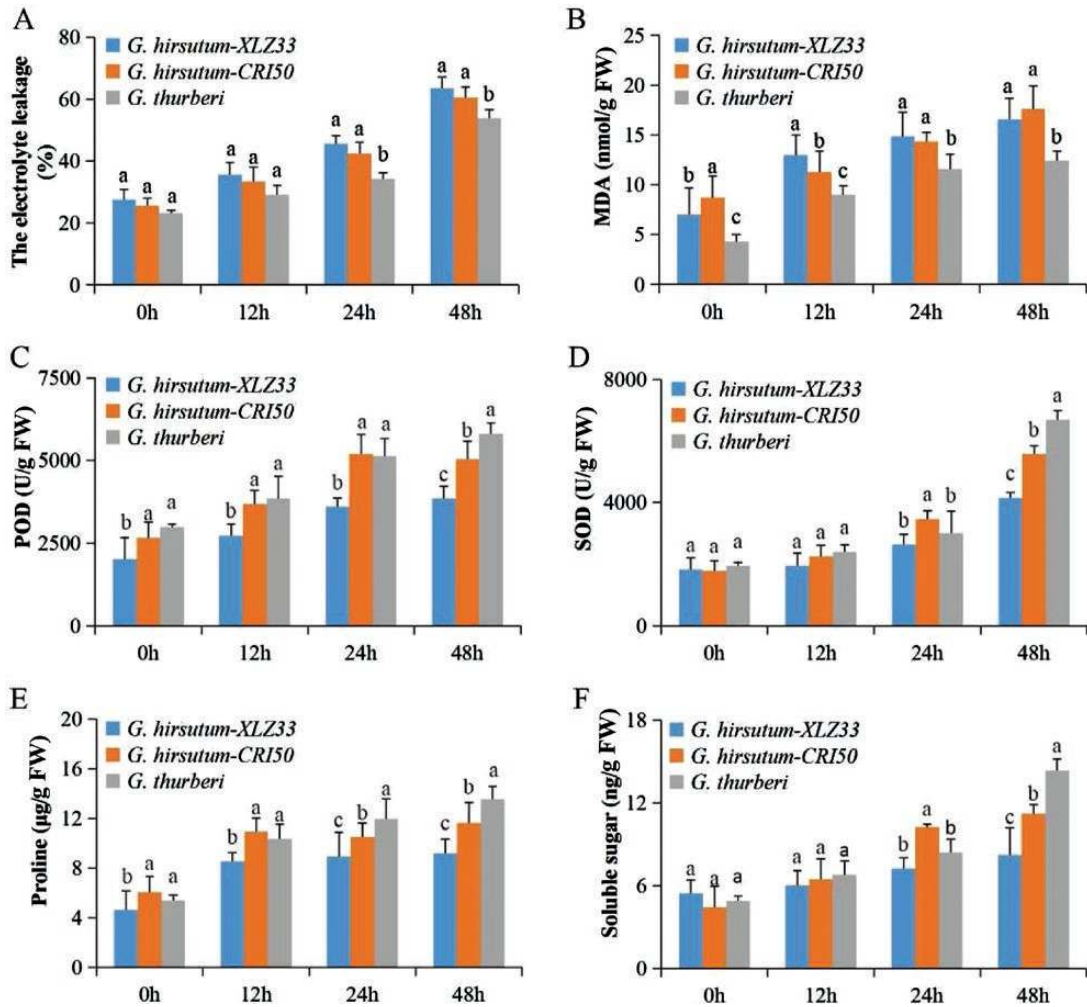


Figure 2. Physiological analysis of *G. thurberi* in response to cold stress. (A–F) The relative electrolyte leakage, the concentrations of MDA, POD, SOD, PRO and SS were determined for *G. thurberi* at 4 °C for 0, 12, 24 and 48 h. Three biological replicates were used. The different letters labelled above columns indicate a significant difference at  $P \leq 0.05$  level. The columns with the same letters mean no significant difference ( $P > 0.05$ ) between different cotton genotypes. Bar indicates standard error.

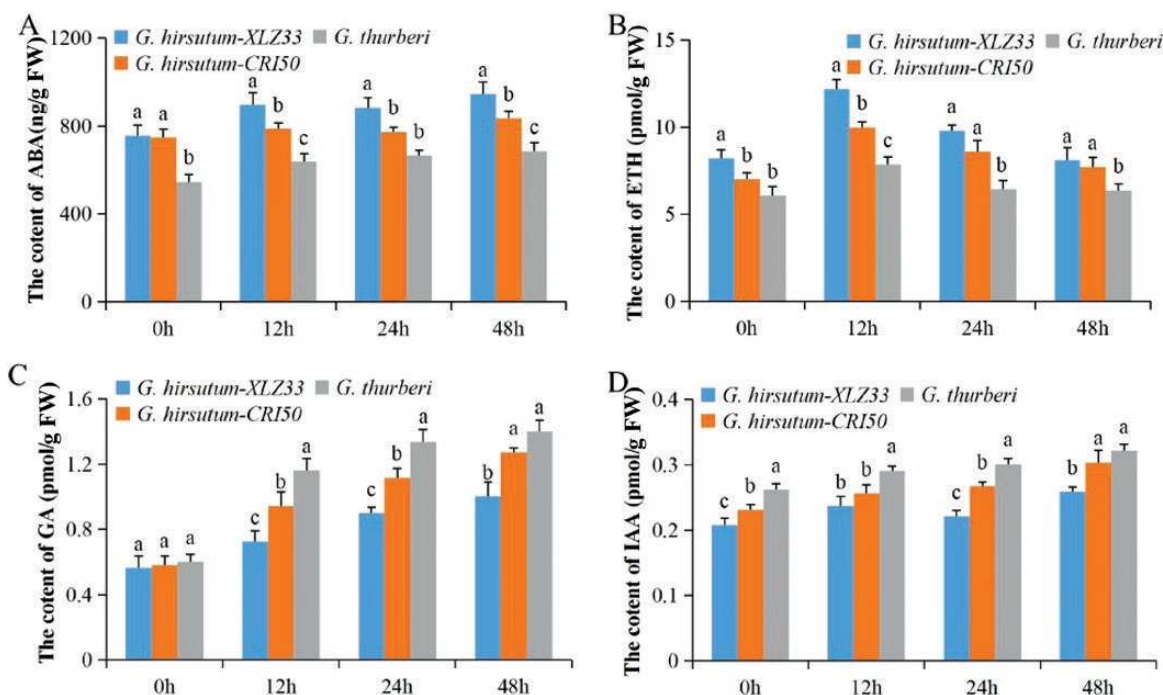
as seed germination, growth, formation of apical hook, to organ senescence, fruit ripening, abscission and stress responses (Pei *et al.* 2017). The concentration levels of GA and IAA were varied, even though the two hormones are known to promote plant growth and development (Tanimoto 2005), the level of IAA was lower than GA in the three cotton species under cold stress, the results are in agreement with previous findings in which found that cold stress inhibits the root gravity response in *Arabidopsis* by 50 %, a process mainly regulated by the IAA (Shibasaki *et al.* 2009), the results, therefore, showed that cold decreases the internal IAA levels in plants.

### Transcription analysis of *G. thurberi*

To gain insight into the gene expression pattern in the leaf tissue of *G. thurberi* under cold condition, 10 cDNA libraries were constructed under normal (control) and low temperature (cold treatment) at 0, 6 and 12 h, respectively. The samples were coded as follows CK0-1, CK6-1, CK12-1, CK0-2, CK6-2, CK12-2, CS6-1, CS12-1, CS6-2 and CS12-2. The cDNA libraries were sequenced by Illumina HiSeq2500 platform using the paired and end method. After filtering out low-quality reads, raw reads containing 'N' and adaptor sequence, 34.48–43.66 M clean reads were collected. The clean reads were aligned to the diploid D-genome cotton reference genome (*G. raimondii*), whose progenitor is the

putative contributor of the Dt-sub genome of the tetraploid-cultivated cotton species. Approximately 75.53–78.25 % clean reads were mapped to the reference genome and multiple mapped clean reads in each library were excluded from further analysis. Finally, a total of 22.91–30.83 M uniquely mapped reads were used for subsequent analysis (Table 1). The mapped sequences were assembled with Cufflinks software inference to the *G. raimondii* genome (<https://www.cottongen.org/>). The RNA-Seq assays revealed that there were a total of 35 617 unigenes of which 97.52 % (34 733 unigenes) were annotated genes and 2.48 % (884 unigenes) novel genes based on this analysis [see Supporting Information—Table S2].

To identify the DEGs in *G. thurberi* under cold stress and normal conditions, the transcript abundance of each of the genes was estimated by fragments per kilobase exon per million fragments mapped (FPKM). DESeq and the Q-values were employed and used to evaluate the differential gene expression pattern. A cut-off P-value < 0.01 adjusted by false discovery rate (FDR) and fold change  $\geq 2$  was used to identify DEGs. A total of 4226 genes showed differential expression after 6 and 12 h of cold stress treatment (CK\_6h vs. CS\_6h, CK\_12h vs. CS\_12h). The distribution of the DEGs revealed that 2693 and 2841 genes were expressed in the leaves at 6 and 12 h of cold stress treatment, respectively, and that 1308 DEGs were



**Figure 3.** Phytohormones analysis in the leaf tissues of *G. thurberi* under cold stress. (A) Quantification of ABA, (B) quantification of ETH, (C) quantification of GA and (D) quantification of IAA. Three biological replicates were used. The different letters (a/b) labelled above columns indicate a significant difference at  $P < 0.05$  levels. The columns with the same letters mean no significant difference ( $P > 0.05$ ) between different cotton genotypes. Bar indicates standard error.

**Table 1.** Summary of the sequencing data of the *G. thurberi* transcriptome. CK and CS: normal temperature control and cold stress treatment, respectively. Units: M, million.

Sample	Mapped read (M)	Mapped ratio (%)	% $\geq$ Q30
CK0-1	34.17	78.25	91.59
CK6-1	33.1	75.88	91.33
CK12-1	26.04	75.53	92.16
CS6-1	32.39	77.33	92.03
CS12-1	26.9	76.91	92.02
CK0-2	40.02	67.53	93.84
CK6-2	41.05	69.79	96.03
CK12-2	48.56	69.50	96.32
CS6-2	30.08	68.09	95.42
CS12-2	43.78	72.48	96.35

identified at both time points (Fig. 4A). Among these DEGs, 1205 and 1322 up-regulated genes, 1488 and 1519 down-regulated genes detected under cold stress at 6 and 12 h time points, respectively; however, the number of up-regulated genes at 12 h of stress exposure were more than the up-regulated genes at 6 h of cold stress exposure (Fig. 4B). The results showed that more genes were down-regulated in the leaves in response to cold stress. Interestingly, compared with the number of DEGs at 6 h post-stress exposure/treatment, there were more DEGs at 12 h of post-stress treatment, which indicated that more COR genes were induced by an increase in exposure. Moreover, analysis of the DEGs through various databases revealed higher percentage values, for instance, the DEGS with GO terms were 2716 (64.26%), KEGG values were detected for 805 genes accounting for 19.04% (Fig. 4C). The detection of various functions through the GO, KEGG, KOG among others indicated that these genes have a putative role in enhancing cold stress tolerance in *G. thurberi*.

### Functional analysis of the DEGs under cold stress condition

Gene Ontology enrichment analysis showed that a total of 2716 (64.26%) DEGs in *G. thurberi* were annotated, and were involved in all the GO functional annotations, biological processes (BP), molecular function (MF) and cellular component (CC). For the BP, metabolic process, cellular process, single-organism process and response to stimulus were the top terms, which implied that a high degree of metabolic activity changes occurred in response to cold stress. For the category of CC, cell part, cell, organelle and membrane were enriched, which suggested that the cell membrane and cellular homeostasis played an important role in the process of cotton acclimation to cold stress conditions. The most abundant terms for MF were the catalytic activity, followed by binding and 'transporter activity, which revealed that complex enzymes system and other biological catalytic processes were involved in response to cold stress. KOG enrichment analysis showed that many of DEGs were most significantly enriched in posttranslational modification, 'protein turnover, chaperones formations, signal transduction mechanisms, secondary metabolites biosynthesis, 'transport and catabolism of the orthologous proteins under abiotic stress condition (Fig. 5A). To gain further understanding of the biological molecular mechanism and regulatory network of DEGs in *G. thurberi* in relation to cold tolerance, KEGG enrichment pathway analysis was performed. A total of 805 (19.04%) DEGs in *G. thurberi* were annotated to KEGG pathways. Among these pathways, the plant hormone signal transduction (ko04075) pathway was the most significant enrichment. A total of 65 genes involved in this pathway were affected by low temperature. Moreover, 265 DEGs were annotated as different types of TFs, out of which 225 TFs were obtained. Among these TFs, 138 genes were up-regulated, while the rest of the genes were down-regulated after cold treatment in *G. thurberi* (Fig. 5B;

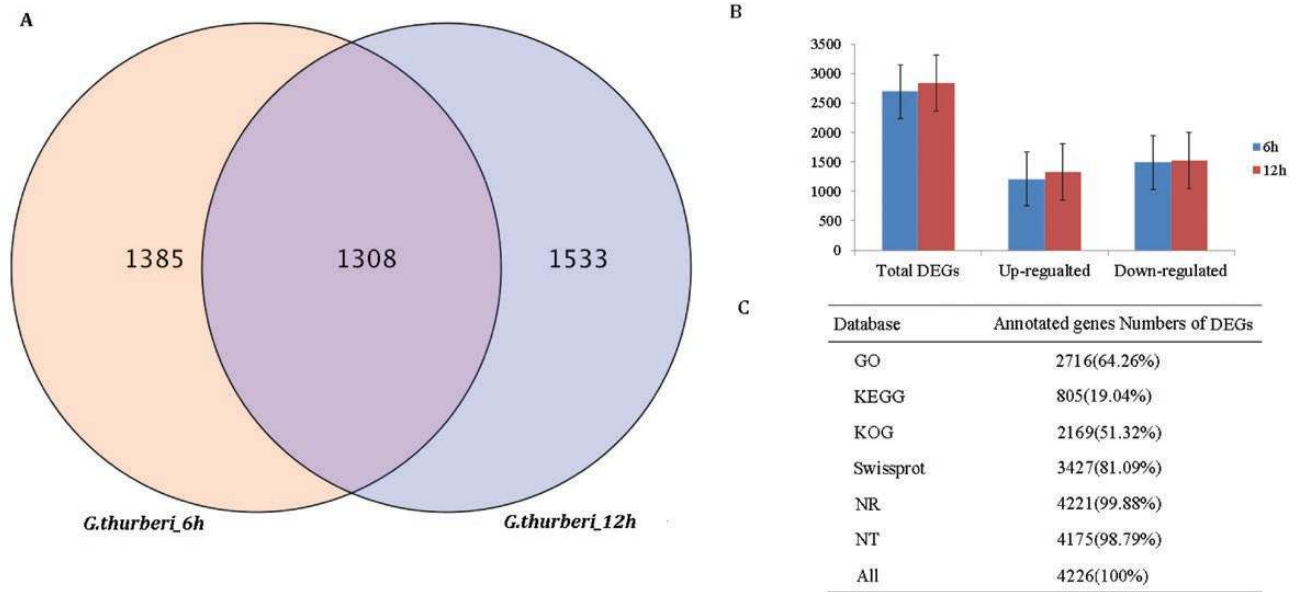


Figure 4. Summary of DEGs in *G. thurberi*. (A) Venn diagram of DEGs identified in *G. thurberi* in response to cold stress. The purple zone meant common DEGs at 6 and 12 h post to cold stress. (B) The summary of DEGs was shown in bar graph. The blue bars represent the DEGs at the 6 h of cold stress treatment. The red bars represent the DEGs at the 12 h post-cold stress treatment. Bar indicates standard error. (C) DEGs annotation of *G. thurberi* by different databases.

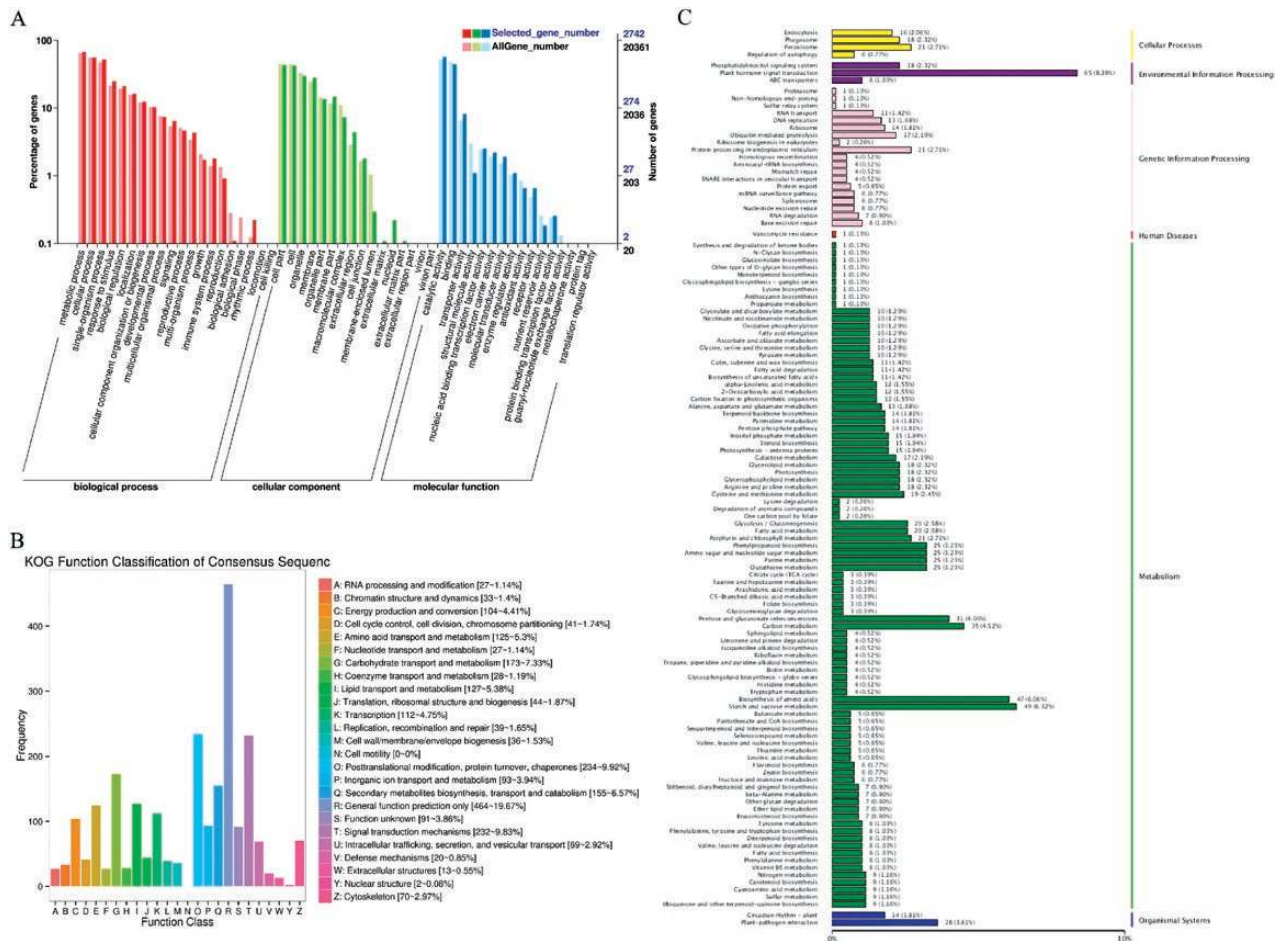


Figure 5. GO, KOG, KEGG enrichment of DEGs. (A) GO enrichment of DEGs; (B) KOG enrichment of DEGs; (C) KEGG enrichment of DEGs.

see [Supporting Information—Fig. S1](#)). According to functional annotation, these TFs were classified into numbers of categories, such as ERF (ethylene response TFs) family (34 TFs), bHLH family

(32 TFs), WRKY family (25 TFs), MYB family (17 TFs), TCP (9 TFs) and NAC family (9 TFs). The TFs ERF (34 genes, 15.1 %), bHLH (32 genes, 14.2 %), WRKY (25 genes, 11.1 %) and MYB (17 genes,



7.5 %) were the top four major families of the cold-regulated TFs. In addition, we observed that the expression of CBF1, CBF2, CBF3/DREB1A and DREB2A, which bind to the C-repeat or dehydration response element promoter's genes, were significantly altered after cold treatment, suggesting that these TFs family might be playing an important regulatory role in responses to cold stress in *G. thurberi*. This result was consistent with previous studies that demonstrated that plant hormones play crucial roles in a diverse set of developmental processes as well as in the response to biotic and abiotic stress (Verma et al. 2016).

Signal transduction has been observed in response to abiotic and biotic stress in many plant species (Czarnocka and Karpiński 2018). A total of 232 DEGs were identified related to different signal transduction mechanisms. Calcium ( $\text{Ca}^{2+}$ ), a universal second messenger in cellular signal transduction, plays a vital role in the plant cold stress response. The cytosolic-free concentration changes within seconds in plants when subjected to cold stress, followed by several signals mediated by a series of protein phosphorylation cascades (Luan 2009). In our data, large numbers of the DEGs involved in signal transduction were indeed related to the calcium-dependent signal pathway, such as CDPKs (calcium-dependent protein kinases), CIPKs (CBL-interacting protein kinases) and CMLs (calmodulin-like protein). The majority of the genes from these three gene families from *G. thurberi* were up-regulated in response to cold.

Carbohydrate transport and metabolism play a core role in plant response to cold stress. All of the 176 DEGs in carbohydrate transport and metabolism were identified under cold stress in *G. thurberi*. Among these DEGs, 19 genes were up-regulated in all stages of *G. thurberi* and 35 genes were down-regulated at 6 and 12 h of cold stress exposure. Aquaporins are transmembrane proteins, which form channels in intracellular and plasma membranes to facilitate the rapid movement of water and play a crucial role in plant water relations. Plant aquaporins are classified into five main subfamilies including the plasma membrane intrinsic proteins (PIPs), tonoplast intrinsic proteins (TIPs), nodulin-26-like intrinsic proteins (NIPs), small basic proteins (SIPs) and X intrinsic proteins (XIPs) (Kapilan et al. 2018). In the present study, 16 PIPs, three TIPs, two NIPs and one SIP genes were induced by cold stress. Moreover, in *Arabidopsis*, most of the PIP genes were down-regulated by cold stress, except for the PIP2;5 which was up-regulated during cold exposure. Furthermore, overexpression of the PIP2;5 gene in *Arabidopsis* have been found to play a role in alleviating the effects of low temperature on plant cell hydraulic conductivity and growth (Lee et al. 2012). Ten out of 16 PIPs genes from *G. thurberi* were significantly up-regulated under cold stress treatments, especially gene6326 (annotated as PIP2-2) exhibited 4-fold higher expression compared to the control during cold stress. In previous studies, a member of tonoplast intrinsic protein GhTIP1:1 was responsive to cold stress and contributed to freezing tolerance in upland cotton. Our transcriptome data analysis showed three differential expressed TIPs genes (including gene8287, gene8689 and gene31507); one NIPs gene (gene10377) and one SIPs gene (37783) were highly up-regulated under cold stress (Fig. 5C; see Supporting Information—Table S3). All results suggested that aquaporins may play a key role in plant response to cold stress by maintaining the uptake and movement of water in the plant body.

### Validation of RNA-Seq results by quantitative real-time PCR

Seven different gene families were analysed, CBF4, ICE1, ICE2, NCED, AAO, RAP and MAPK gene families were profiled. The

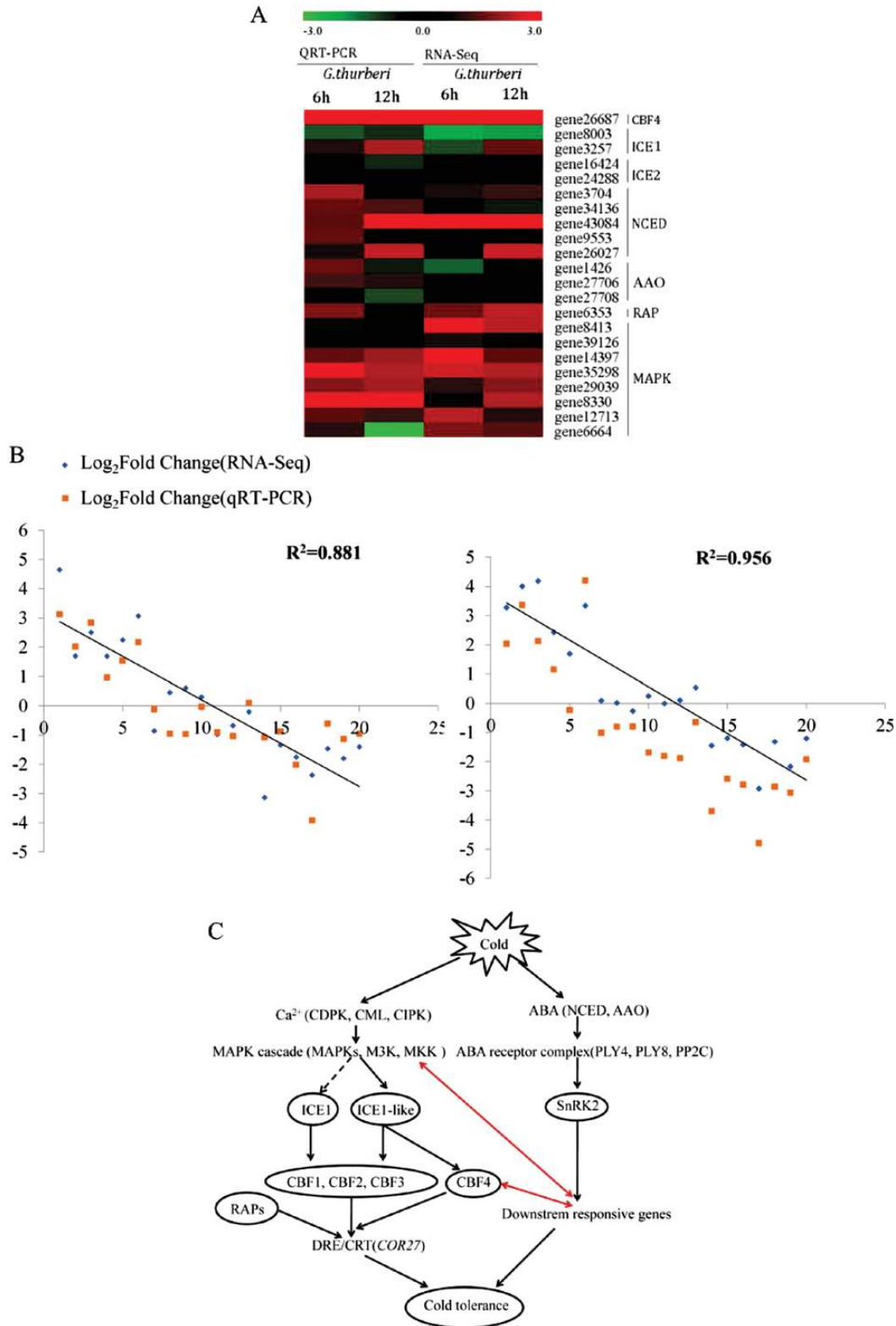
members of CBF4, MAPK and RAP genes were highly up-regulated compared to other families (Fig. 6A). Through correlation analysis, RT-qPCR results were significantly correlated to the RNA-Seq data both at 6 and 12 h points under cold stress ( $R^2 = 0.881$  and  $0.956$ , respectively) in *G. thurberi* (Fig. 6B). Moreover, in the schematic representation of the possible genes, ICE1, ICE1-like and the CBF4 were more integral in enhancing cold stress tolerance in *G. thurberi* (Fig. 6C).

### Knockdown of CBF4 and ICE2 genes in cotton through VIGS seedlings compromises their ability to tolerate cold stress

To further investigate the functions of candidate genes, CBF4 and ICE2, the VIGS TRV-GhPDS, TRV-Ctrl, TRV-CBF4 and TRV-ICE2 plants were observed under cold stress. Albino leaves were observed in TRV-PDS-inoculated seedlings after 7 days of inoculation. Compared with infected seedlings, we found that control seedlings were rapid growth after 20 days of inoculation. And, the difference was not observed between infected seedlings; moreover, cold stress was imposed after three leaf stage, the VIGS plant's ability to tolerate cold stress was highly affected, the leaves wilted after cold stress treatment (Fig. 7A). Moreover, the expression levels of CBF4 and ICE2 were checked by RT-qPCR, their expression levels were down-regulated compared to their expression in the leaves of the TRV-Ctrl seedlings, line CBF4-4 and line ICE2-7 were used for further evaluation, being the CBF4 and ICE2 genes expression were significantly down-regulated (Fig. 7B). Finally, MDA, PRO and SOD were assayed on the leaves of VIGS plants and their wild types under cold stress conditions. The VIGS plants exhibited a higher concentration level of MDA and a significant reduction in the levels of PRO and SOD compared to the wild types (Fig. 7C). The results obtained were in agreement with the previous findings in which knockdown of a regulatory gene affects the normal growth and development of the plants being the plant's ability to tolerate the stress of target become highly compromised (Jahan et al. 2015). Moreover, knockdown of CYP450 genes significantly reduced the tolerance level of upland cotton, *G. hirsutum* to salt and drought stress factors (Magwanga et al. 2019b). Furthermore, RNAi of a stress-responsive gene increases susceptibility of the plant to environmental stresses, for instance, overexpression of a novel trihelix transcription factor enhanced drought and salt stress tolerance in transgenic *Arabidopsis*, but when the same gene was knockdown, the cotton VIGS plants ability to tolerate the two forms of stresses was affected significantly, as was evident by high concentration levels of oxidant enzymes and significant reduction in the levels of various antioxidant enzymes (Magwanga et al. 2019b).

### Discussion

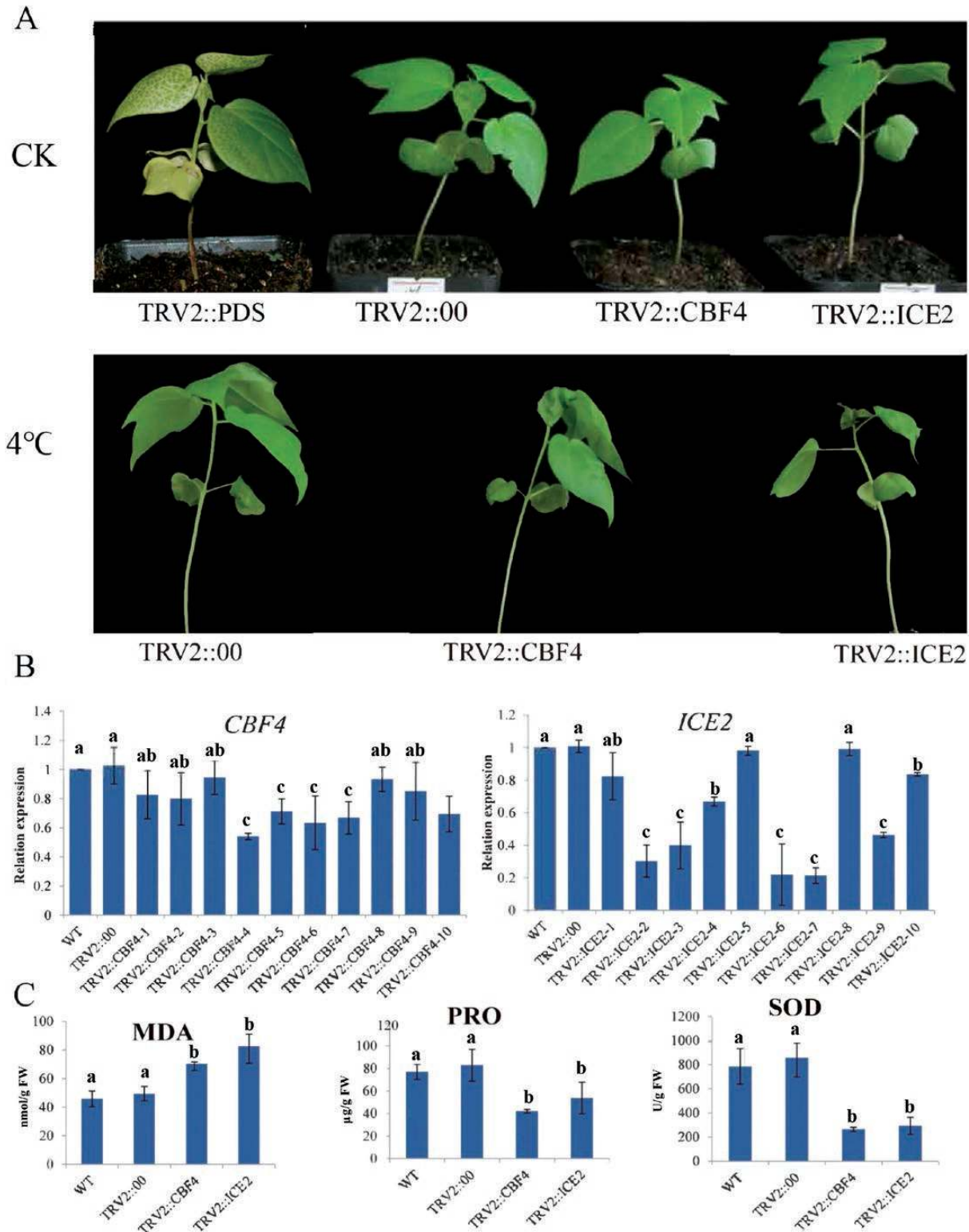
Cold stress limits cotton growth and development, its effect has caused massive losses in the cotton production industry (Holaday et al. 2016). Cotton originated from the tropical and subtropical climates and therefore lacks the cold acclimation mechanism and is sensitive to low temperature throughout the growth period (Lee and Fang 2015). Particularly, if exposed to low temperature in the period of germination or seedling, cotton growth and development would be slow and at the time of budding, flowering and maturity would be delayed (Gai et al. 2008). In addition, low temperature not only affects the yield of cotton but also the quality of fibre (Liu et al. 2015).



**Figure 6.** RT-qPCR validations of transcript levels evaluated by RNA-Seq in *G. thurberi* under cold stress for 6 and 12 h. (A) Heat map depicting log<sub>2</sub> (fold change) of transcription factor; (B) regression analysis of the log<sub>2</sub> (fold change) of RNA and qRT-PCR expression analysis of the DEGs; (C) a predicted model of cold response transcriptional network in *G. thurberi*. Red lines indicated the crosstalks between different signalling pathways predicted in this analysis. Y-axis—relative RNA-Seq expression levels and X-axis—relative RT-qPCR expression levels

Even though plant response to cold stress is a highly complex mechanism, after decades of intensive studies we have begun to understand the COR system in plants (Wang et al. 2017).

However, the cold tolerance mechanism in cotton has gained minimal attention so far. In this study, we aimed at unravelling the cold stress-responsive mechanism in two cotton, *G. thurberi*



**Figure 7.** Virus-induced gene silencing (VIGS), RT-qPCR analysis and biochemical evaluation of the VIGS plants. (A) No obvious symptoms in the leaves of the TRV:00-infected plant (positive control); in each experiment, six plants were used and there were three replicates. (B) RT-qPCR analysis of the changes in the expression level of the two CBL4 and ICE2 genes in the leaf tissues of cotton plants treated with VIGS under cold stress conditions. (C) Quantitative determination of MDA, PRO and SOD in leaves of the wild type (WT), control and the silenced VIGS plants, in CBF4 and ICE2 silenced plants; line CBF4-4 and ICE2-7 were used, respectively. All measurements were done after 8 days of stress exposure. In (B) and (C), each experiment was repeated three times. Error bars of the biochemical trait measurements represent the standard deviation of three biological replicates. Different letters (a/b) indicate significant differences between WT and VIGS plants (two-tailed;  $P < 0.01$ ).

and two accessions of tetraploid cotton, *G. hirsutum*. The results showed that after 48 h of cold stress at 4 °C, morphological and physiological traits of the diploid wild cotton *G. thurberi*

underwent drastic changes to acclimatize to the cold stress condition. The changes, therefore, showed that *G. thurberi* is highly tolerance to cold stress, the results were consistent with

previous studies (Liang et al. 1996). The SS and PRO are the most effective organic osmolytes in plants, which can help stabilize proteins and cell structures under stressful environmental conditions (Naser et al. 2010). These compounds can also act as free-radical scavengers, protecting against oxidation by removing excess ROS, and also re-establishing the cellular redox balance (Thalmann et al. 2016). There is a positive correlation between SS and PRO accumulations, and plant stress tolerance (Dikilitas et al. 2017). In this study, 49 and 83 genes were involved in 'starch and sucrose metabolism (ko00500)' and 'arginine and proline metabolism (ko00330)' pathways, respectively, and were regulated by cold stress. Half of these DEGs were highly up-regulated, which suggested that these cold-induced genes might promote the accumulation of SS and PRO in *G. thurberi* to enhance cold tolerance. The results obtained by the tissue analysis of *G. thurberi* under cold stress condition showed a marked increase in PRO and SS. The increased levels of PRO and SS indicated that they are the major osmolytes responsible for osmotic adjustment in cotton under low temperature condition. Moreover, the plant endogenous hormones had significant changes under cold stress.

To gain insight into the morphological, physiological and hormonal changes in *G. thurberi*, we performed transcriptome sequencing analysis under cold stress condition. A total of 35 617 genes were obtained, in which 4227 genes were differentially expressed under cold stress in *G. thurberi*. High numbers of DEGs were significantly enriched in various processes such as single-organism process, metabolic process, response to the stimulus, membrane, organelle part, membrane part, catalytic activity and nucleic acid binding transcription factor, as obtained through GO and KEGG enrichment analysis. Moreover, KEGG enrichment analysis proved that many of the genes were involved in plant hormone signal transduction, 'starch and sucrose metabolism'. These results of KEGG enrichment analysis demonstrated signal transduction, the sugar transport and metabolism played an important role in response to cold stress. Furthermore, ROS are oxygen-containing substances of metabolites and their derivatives generated through oxidation in plants, directly or indirectly. Reactive oxygen species play a vital role in stress signal transduction, but excessive active oxygen oxidation harms the plant (Gill and Tuteja 2010). Therefore, plants also produce a series of active oxygen-scavenging enzymes, for example, POD, CAT, SOD among others. In this study, the expression of RBOH (respiratory burst oxidase homolog) gene, which is responsible for the accumulation of H<sub>2</sub>O<sub>2</sub>, was rapidly up-regulated in leaf tissues at 6 h of cold treatment. Simultaneously, ROS-scavenging systems were markedly induced, showing different expression patterns under cold stress. In total, 16 POD genes, 9 SOD genes and 1 CAT gene were found, and most of them were up-regulated. Interestingly, the CAT gene (gene6557) showed no expression at 6 h, but was significantly up-regulated at 12 h of cold stress treatment. Moreover, plant hormones such as auxin, cytokinins, gibberellin (GA), ABA, ETH, brassinosteroids (BR), jasmonic acid (JA) and salicylic acid (SA) are small molecules that play significant roles throughout the life span of plants, especially when plants encounter adversity (Verma et al. 2016). In our research, we found a series of DEGs were annotated plant hormone signal transduction (ko04075) and were involved in series of signalling pathways, for instance, 36 DEGs were involved in auxin-mediated signalling pathway (GO:0060774), 10 DEGs in ABA-activated signalling pathway (GO:0009738), five DEGs in GA-mediated signalling pathway (GO:0009740), five DEGs in ETH-activated signalling pathway (GO:0009873) and finally eight DEGs were involved in JA-mediated signalling pathway (GO:0009867). In the present study, most of these DEGs related

to plant hormone signal transduction were up-regulated in all stages, combined with the changes in endogenous hormones concentrations in cotton seedling under cold treatment, the results suggested that plant hormone signal transduction plays a core role in response to cold stress.

Plants have a complex molecular mechanism network that enables them to respond to cold stress. Generally, the cold stress signal is first received by the receptors on the plant cell membrane, which results in the generation of many secondary messengers, such as Ca<sup>2+</sup> ions cascades, hormones and ROS (Miura and Furumoto 2013; Zhang et al. 2017). The signal molecules transmit the stress signals into the nucleus, which then leads to the activation of related genes to induce responses to cold stress (Chinnusamy et al. 2007). Transcription factors are fundamental to the regulation of cellular pathways in response to adverse stresses in many plants. In this study, we identified several types of differentially expressed TFs in response to cold stress in *G. thurberi*, including ERF, TCP, WRKY, NAC, bHLH, MYB family and among others. The effects of ETH on plants under cold stress have been elucidated (Wang et al. 1990; Abiri et al. 2017). The ERF TFs are implicated in many diverse functions in cellular processes, such as hormonal signal transduction, response to biotic and abiotic stresses, regulation of metabolism and in developmental processes in various plant species (Rehman and Mahmood 2015). In this study, we found >12 % of the ERF TFs to be differentially expressed under cold stress. It has been reported that overexpression of transcription factor TERE2/LeERE2 in tobacco and tomato could enhance cold tolerance by facilitating ETH biosynthesis, which supports the important role of ERF in plant cold tolerance (Zhang and Huang 2010). In addition, dehydration-responsive element-binding (DREB) protein, which belongs to the subfamily of AP2/EREBP TFs, specifically binds to the promoter regions of downstream genes, activates or suppresses the transcription of these genes and finally enhance plant stress tolerance (Kizis et al. 2001). The previous study has reported that a cotton DREB gene (GhDREB) conferred enhanced tolerance to drought, salinity and freezing stresses in transgenic wheat (Gao et al. 2009). The same gene was significantly induced after cold treatment in *G. thurberi*, suggesting that this gene may be critical in response to cold stress in cotton. In addition, the TCP family is a group of plant-specific TFs that plays an important role in the growth and development of plants (Chai et al. 2017). In this study, >14 % of the TCP TFs were differently expressed; moreover, 25 WRKY TFs were differentially expressed in *G. thurberi* leaves, and most of them were up-regulated. The results obtained were in agreement with the previous studies, which elucidated the WRKY TFs in *Arabidopsis thaliana*, which played an important role in enhancing cold stress (Wang et al. 2011). Interestingly, gene3257 encoding a WRKY TFs displayed opposite expression trends at 6 and 12 h time point in our data. The expression of this gene was inhibited at early stages, while it was induced after 12 h of cold treatment, implying that this gene might be functioning in different ways at different time points.

The NAC TFs are involved in many aspects of plant growth, development and response to abiotic stress (Chen et al. 2016). The NAC family TFs, which were closely associated with ABA signal pathways, showed significant up-regulation in the leaf tissues of *G. thurberi* under cold stress. Moreover, OsNAC5 a NAC transcription factor was significant in enhancing cold, drought and salinity stress tolerances in rice plants, by regulating downstream targets associated with accumulation of compatible solutes, H<sub>2</sub>O<sub>2</sub> and MDA (Song et al. 2011). Furthermore, it has been reported that bHLH TF do act as positive regulators of the CBF pathway and conferred cold tolerance in plants (Niu et al. 2017).

Moreover, overexpression of *NtbHLH123* in tobacco reduced the electrolyte leakage, MDA contents,  $H_2O_2$  and ROS accumulation under cold stress, thus enhancing cold tolerance (Zhao et al. 2018). We found that the expression of 32 *bHLH* genes were both altered after 6 and 12 h of cold treatment in *G. thurberi*. Among these genes, were 14 and 18 up- and 18 down-regulated *bHLH* genes, respectively, in the leaf tissues of *G. thurberi* under cold stress condition. The results suggested that the cold-induced *bHLH* genes might be playing a critical role in the regulation of the rhythm of *G. thurberi* under cold condition.

Due to the amount of MYB TFs found in *A. thaliana* (Stracke et al. 2001), rice (Katiyar et al. 2012), tomato (Meng et al. 2015), maize (Rabinowicz et al. 1999) and cotton (Salih et al. 2016), a lot of researches have been focused on its role in transcriptional regulation and impact on the physiological function. For example, overexpression of rice MYB3 significantly enhanced cold tolerance in transgenic rice (Su et al. 2010). In this research, 17 MYB genes were found to exhibit differential expression pattern under cold stress treatment. These results indicated that ERF, TCP, WRKY, NAC, *bHLH* and MYB TFs were involved in plant responses to various stresses, and further suggested that cold stress might have a common molecular mechanisms with other abiotic stress.

As a ubiquitous second messenger in cellular signal transduction, calcium ( $Ca^{2+}$ ) acts as a mediator of stimulus response, coupling in the regulation of diverse cellular functions (Feng et al. 2013). For instance, a quantitative trait locus (QTL) *COLD1* encoding a regulator of G-protein signalling has been found to confer chilling tolerance in *japonica* rice under cold stress (Ma et al. 2015), because it facilitated the activity of the  $Ca^{2+}$  channel which is responsible for sensing low temperature by increased  $Ca^{2+}$  concentration (Györke and Györke 1998). Moreover, numbers of the DEGs involved in signal transduction were related to the calcium-dependent signal pathway. For instance, calcium-dependent protein kinases (CDPKs), a kind of serine/threonine protein kinase, which is activated by calcium signal, and are involved in carbon and nitrogen metabolism, ion and water transmembrane transport, growth and development regulation as well as response to anti-abiotic stress (Kumar et al. 2004). The expression of *gene10339* was significantly up-regulated under cold stress in *G. thurberi*. In addition, the expression of nine CIPKs and 15 CMLs genes was altered by cold stress, and most of them were up-regulated, revealing that the proteins encoded by the genes could be playing an important role in detection and transmission of the cold signal. The results were consistent with previous studies which reported that overexpression *ShCML44* gene enhanced cold, drought and salinity stresses tolerance in *Solanum lycopersicum* transgenic plants (Munir et al. 2016). The expression of three *CML44*-like genes was highly increased in leaves at two time points of cold treatment, which suggested that these genes play a significant role in the  $Ca^{2+}$  signal transduction pathway in cotton and might be contributing to the increased tolerance to low temperatures.

Plants could perceive and respond to stress rapidly via signal transduction pathways mediated by phytohormones. Most of the DEGs related to plant hormone signal transduction were up-regulated in all the stages under cold treatment, suggesting that plant hormone signal transduction plays a core role in response to cold stress. In particular, ABA plays a role in signal detection in response to drought, cold, osmotic stress and pathogen infection, in turn triggering various changes in plant physiology and development processes, resulting in adaptation to biotic and abiotic stresses (Mittler and Blumwald 2015). Several ABA biosynthesis-related genes, including five

9-cis-epoxycarotenoid dioxygenase (NCED) and three abscisic aldehyde oxidase (AAO) genes, were induced under cold stress. Moreover, ABA does induce  $H_2O_2$  by activating the nicotinamide adenine dinucleotide phosphate (NADPH) oxidases such as *AtrBOHD* and *AtrBOHF* in *Arabidopsis* (Alqurashi et al. 2017).

Plants have evolved a complex molecular mechanism network to adapt to various abiotic stresses such as cold, drought and salt, by enhancing their adaptability through the mobilization of various stress genes, such as the LEA genes (Magwanga et al. 2018), MATE genes (Lu et al. 2019) among others. Moreover, plant stress responses must be coordinated with growth and development, it is important to understand the crosstalk between stress signalling pathways and hormonal as well as growth and development signalling pathways (Kim et al. 2015). The cold response pathway ICE-CBF-COR is a best-known defence mechanism to cold stress, which includes the core component inducer of CBF expression (ICE), C-repeat binding factors (CBF) transcriptional factor and other diverse proteins such as the cold-regulated proteins. As master regulators of CBF expression, ICE binds to the promoter of the CBF genes, and positively induces the expression of CBF genes (Chinnusamy et al. 2003). Furthermore, overexpression of the ICE1 gene enhanced plant tolerance to cold stress, whereas wild plants were hypersensitive to cold stress (Xiang et al. 2013, 2017). The ICE2 gene is highly homologous to *ice1*, it has been found to have the ability to trigger the expression of CBF1 (Fursova et al. 2009). Overexpression of ICE2 resulted in an increased survival rate in transgenic plants after freezing treatment (Kurbidaeva et al. 2014). Moreover, two ICE1 genes (*gene8003* and *gene3257*) and two ICE2 genes (*gene16424* and *gene24288*) were identified in *G. thurberi*. Although the expression levels of the two ICE2 genes were not obviously different under cold treatment and control samples, but the VIGS results showed that the ICE2 genes are important for cold tolerance in cotton. Moreover, as the three important transcriptional factors involved in cold response pathway in *Arabidopsis*, CBF1 and CBF3 are negatively regulated by CBF2 (Zhou et al. 2011). Although previous studies have shown that the CBF4 gene, a novel homolog of CBF/DREB1 genes from *Arabidopsis*, was initially induced under drought stress (Karimi et al. 2015). Interestingly, the expressions of the CBF1 and CBF3 genes were neither up- nor down-regulated under cold treatment; however, the CBF2 and CBF4 genes were activated by cold stress, especially the expression of CBF4 gene was highly up-regulated when *G. thurberi* seedlings were exposed to low temperature. In addition, *RAP2.6*, an *Arabidopsis* AP2/ERF family member, has been reported to function in plant response to ABA, and different stress conditions such as salinity, cold and osmotic stresses (Imran et al. 2018). In this study, we found that the expression level of *RAP2.7* (*gene6353*) from *G. thurberi* was up-regulated in two stages after exposure to cold stress, suggesting that *RAP2.7* gene may be a key gene involved in the CBF-regulated network. As CBF target genes, the COR genes have a significant role in enhancing plant tolerance to cold stress (Dong et al. 2014). Furthermore, several COR genes were induced under the cold condition in this study. All these results indicated that there might be a different cold regulating pathway in cotton and CBF4 and *RAP2.7* genes could be the key cold candidate genes in cotton. A well-studied cold response pathway is the ABA-dependent signalling pathway, which mainly contains the pyrabactin resistance/pyrabactin resistance-like/regulatory component of the abscisic acid receptor (PYR/PYL/RCAR) family as ABA receptors, negative/positive regulators such as type 2C protein phosphatase (PP2C) and SNF1-related protein kinase 2 (SnRK2) and a series of other TFs (Zhu 2016). In rice, the

concentration of endogenous ABA increased under cold stress, ABA bind to the PYR/PYL/RCAR, which then interacts and inhibits PP2C, resulting in the activation of SnRK2. The activated SnRK2 phosphorylate effector proteins including TFs (AREB, ABF) and downstream stress responsive genes were activated to improve plant tolerance against low temperature environment (Sah et al. 2016). In rice, overexpression of OsPYL3 and OsPYL9 enhanced rice tolerance to chilling stress (Tian et al. 2015). Moreover, all of the three PYL4 and one PLY8 genes were up-regulated under cold stress. The up-regulation of PYL4 and PYL8 genes in addition to the PP2C gene (ABA biosynthesis genes), indicated that the ABA signalling pathway is integral in cold stress tolerance in plants.

In confirming the role of CBF4 and ICE2 genes through VIGS, the VIGS plants ability to tolerate cold stress was significantly reduced, as evident by higher levels of MDA and significant reduction of PRO and SOD, the two antioxidant enzymes evaluated. When plants are exposed to any form of stress, the delicate balance of reactive oxygen production and elimination becomes altered, leading to excessive accumulation of ROS within the plant cells (Dong et al. 2019).

## Conclusion

In conclusion, we presented the first comprehensive transcriptome data from *G. thurberi*, a wild diploid cotton species that have adapted to local temperature climate and exhibits strong tolerance to low temperature. The transcriptome revealed that a total of 4226 DEGs were discovered to be involved in cold stress tolerance in leaves. Furthermore, we identified several DEGs related to hormones, Ca<sup>2+</sup> and ROS, which were involved in different cold response pathways suggesting the complex responses of *G. thurberi* towards cold stress. Moreover, RT-qPCR analysis and VIGS showed that the CBF4 and ICE2 genes could be the novel genes in *G. thurberi*, which are responsible for its cold stress tolerance. The virus-induced gene silenced plants (VIGS plants) ability to tolerate cold stress effect was significantly reduced compared to their wild types, as evident by higher levels of MDA and a significant reduction in PRO and SOD concentrations. Our findings provide an overall picture of the regulatory network in response to cold stress in *G. thurberi*. These COR-related genes could be targeted as potential candidates for further study and be vital in developing more cold stress tolerance cotton genotypes.

## Sources of Funding

This research was funded by the Natural Science Foundation of China (31601352).

## Contributions by the Authors

C.X., R.O.M., W.K. and F.L. designed the experiment, implemented and collected the data. C.X., Y.X., and R.O.M. analysed the results and prepared the manuscript. C.X., R.O.M., Z.Z., X.X.W., Y.H., Y.W., Y.Z., F.L., W.K. revised the manuscript. All authors reviewed and approved the final manuscript.

## Acknowledgements

We are indebted to Prof. Liu Fang for his immense support, together with Prof. Wang Kunbo, financial support and moral support was such a great source of inspirational.

## Supporting Information

The following additional information is available in the online version of this article—

**Figure S1.** Distribution's network of reactive oxygen species (ROS) 692 genes in chromosomes of *Gossypium hirsutum*, *G. raimondii* and *G. arboretum*.

**Table S1.** Genes of mapped reads assemble and annotation of *Gossypium thurberi*.

**Table S2.** Differentially expressed genes (DEGs) of *Gossypium thurberi*.

**Table S3.** Primers list of transcripts for real-time RT-PCR.

## Conflict of Interest

None declared.

## Literature Cited

- Abiri R, Shaharuddin NA, Maziah M, Yusof ZNB, Atabaki N, Sahebi M, Valdiani A, Kalhori N, Azizi P, Hanafi MM. 2017. Role of ethylene and the APETALA 2/ethylene response factor superfamily in rice under various abiotic and biotic stress conditions. *Environmental and Experimental Botany* **134**:33–44.
- Alqurashi M, Thomas L, Gehring C, Marondedze C. 2017. A microsomal proteomics view of H<sub>2</sub>O<sub>2</sub>- and ABA-dependent responses. *Proteomes* **5**:22.
- Alqurashi M, Thomas L, Gehring C, Marondedze C. 2017. Gene ontology: tool for the unification of biology. *Nature Genetics* **25**:25–29.
- Barrero-Gil J, Huertas R, Rambla JL, Granell A, Salinas J. 2016. Tomato plants increase their tolerance to low temperature in a chilling acclimation process entailing comprehensive transcriptional and metabolic adjustments. *Plant, Cell & Environment* **39**:2303–2318.
- Bartwal A, Mall R, Lohani P, Guru SK, Arora S. 2013. Role of secondary metabolites and brassinosteroids in plant defense against environmental stresses. *Journal of Plant Growth Regulation* **32**:216–232.
- Bolek Y. 2010. Genetic variability among cotton genotypes for cold tolerance. *Field Crops Research* **119**:59–67.
- Caverzan A, Casassola A, Brammer SP. 2016. Antioxidant responses of wheat plants under stress. *Genet Mol Biol* **39**:1–6.
- Chai W, Jiang P, Huang G, Jiang H, Li X. 2017. Identification and expression profiling analysis of TCP family genes involved in growth and development in maize. *Physiology and Molecular Biology of Plants* **23**:779–791.
- Chakravarthy VSK, Reddy TP, Reddy VD, Rao KV. 2012. Current status of genetic engineering in cotton (*Gossypium hirsutum* L): an assessment. *Critical Reviews in Biotechnology* **8551**:1–18.
- Chen W, Yao Q, Patil GB, Agarwal G, Deshmukh RK, Lin L, Wang B, Wang Y, Prince SJ, Song L, Xu D. 2016. Identification and comparative analysis of differential gene expression in soybean leaf tissue under drought and flooding stress revealed by RNA-Seq. *Frontiers in Plant Science* **7**.
- Chinnusamy V, Ohta M, Kanrar S, Lee BH, Hong X, Agarwal M, Zhu JK. 2003. ICE1: a regulator of cold-induced transcriptome and freezing tolerance in *Arabidopsis*. *Genes & Development* **17**:1043–1054.
- Chinnusamy V, Zhu J, Zhu JK. 2007. Cold stress regulation of gene expression in plants. *Trends in Plant Science* **12**:444–451.
- Colebrook EH, Thomas SG, Phillips AL, Hedden P. 2014. The role of gibberellin signalling in plant responses to abiotic stress. *The Journal of Experimental Biology* **217**:67–75.
- Czarnocka W, Karpiński S. 2018. Friend or foe? Reactive oxygen species production, scavenging and signaling in plant response to environmental stresses. *Free Radical Biology & Medicine* **122**:4–20.
- Dikilitas M, Yucel N, Dervis S. 2017. Production of antioxidant and oxidant metabolites in tomato plants infected with verticillium dahliae under saline conditions. In: Khan M, Khan N, eds. *Reactive oxygen species and antioxidant systems in plants: role and regulation under abiotic stress*. Singapore: Springer.
- Dong C, Chen L, Wang Y, Qin Y, Yang R, Xiao L, Wang B, Lu H, Feng Y. 2014. COR-like gene is involved in induced-expression response to multiple

- abiotic stresses in grape vine (*Vitis amurensis*) tissues. *Agricultural Sciences* 5:604–610.
- Dong Q, Magwanga RO, Cai X, Lu P, Nyangasi Kirungu J, Zhou Z, Wang X, Wang X, Xu Y, Hou Y, Wang K. 2019. RNA-sequencing, physiological and RNAi analyses provide insights into the response mechanism of the ABC-mediated resistance to *Verticillium dahliae* infection in cotton. *Genes* 10:110.
- Feng J, Li J, Liu H, Gao Q, Duan K, Zou Z. 2013. Isolation and characterization of a calcium-dependent protein kinase gene, FvCDPK1, responsive to abiotic stress in woodland strawberry (*Fragaria vesca*). *Plant Molecular Biology Reporter* 31:443–456.
- Fowler SG, Cook D, Thomashow MF. 2005. Low temperature induction of *Arabidopsis* CBF1 is gated by the circadian clock. *Plant Physiology* 137:961–968.
- Fursova OV, Pogorelko GV, Tarasov VA. 2009. Identification of ICE2, a gene involved in cold acclimation which determines freezing tolerance in *Arabidopsis thaliana*. *Gene* 429:98–103.
- Gai YP, Li XZ, Ji XL, Wu CA, Yang GD, Zheng CC. 2008. Chilling stress accelerates degradation of seed storage protein and photosynthetic protein during cotton seed germination. *Journal of Agronomy and Crop Science* 194:278–288.
- Gao X, Britt RC Jr, Shan L, He P. 2011. *Agrobacterium*-mediated virus-induced gene silencing assay in cotton. *J Vis Exp*. doi:10.3791/2938
- Gao SQ, Chen M, Xia LQ, Xiu HJ, Xu ZS, Li LC, Zhao CP, Cheng XG, Ma YZ. 2009. A cotton (*Gossypium hirsutum*) DRE-binding transcription factor gene, GhDREB, confers enhanced tolerance to drought, high salt, and freezing stresses in transgenic wheat. *Plant Cell Reports* 28:301–311.
- Gill SS, Tuteja N. 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry* 48:909–930.
- Guo HM, Li ZC, Zhang H, Xin YZ, Cheng HM. 2011. Cloning of cotton CBF gene for cold tolerance and its expression in transgenic tobacco. *Acta Agronomica Sinica* 37:286–293.
- Györke I, Györke S. 1998. Regulation of the cardiac ryanodine receptor channel by luminal Ca<sup>2+</sup> involves luminal Ca<sup>2+</sup> sensing sites. *Biophysical Journal* 75:2801–2810.
- Haake V. 2002. Transcription factor CBF4 is a regulator of drought adaptation in *Arabidopsis*. *Plant Physiology* 130:639–648.
- Hasanuzzaman M, Nahar K, Fujit M. 2013. Extreme temperature responses, oxidative stress and antioxidant defense in plants. In: Vahdati K, Leslie C, eds. *Abiotic stress - plant responses and applications in agriculture*. London, UK: IntechOpen.
- Holaday AS, Mahan JR, Payton P. 2016. Effects of chilling temperatures on photosynthesis. *Journal of Cotton Science* 20:220–231.
- Hussain S, Khan F, Hussain HA, Nie L. 2016. Physiological and biochemical mechanisms of seed priming-induced chilling tolerance in rice cultivars. *Frontiers in Plant Science* 7.
- Iba K. 2002. Acclimative response to temperature stress in higher plants: approaches of gene engineering for temperature tolerance. *Annual Review of Plant Biology* 53:225–245.
- Imran QM, Hussain A, Lee SU, Mun BG, Falak N, Loake GJ, Yun BW. 2018. Transcriptome profile of NO-induced *Arabidopsis* transcription factor genes suggests their putative regulatory role in multiple biological processes. *Scientific Reports* 8.
- Jahan SN, Asman AK, Corcoran P, Fogelqvist J, Vetukuri RR, Dixelius C. 2015. Plant-mediated gene silencing restricts growth of the potato late blight pathogen *Phytophthora infestans*. *Journal of Experimental Botany* 66:2785–2794.
- Jing Z, Bo Z, XiaoFen J, RiHe P, YuShan Q, Zhen Z. 2008. Cloning and analysis of CBF transcription factors from *Brassica napus* cv. Huyou 15. *Plant Physiology Communications* 44:1106–1112.
- Kanehisa M, Araki M, Goto S, Hattori M, Hirakawa M, Itoh M, Katayama T, Kawashima S, Okuda S, Tokimatsu T, Yamanishi Y. 2008. KEGG for linking genomes to life and the environment. *Nucleic Acids Research* 36.
- Kang J, Zhang H, Sun T, Shi Y, Wang J, Zhang B, Wang Z, Zhou Y, Gu H. 2013. Natural variation of C-repeat-binding factor (CBFs) genes is a major cause of divergence in freezing tolerance among a group of *Arabidopsis thaliana* populations along the Yangtze River in China. *The New Phytologist* 199:1069–1080.
- Kaplan R, Vaziri M, Zwiazek JJ. 2018. Regulation of aquaporins in plants under stress. *Biological Research* 51:4.
- Kargiotidou A, Kappas I, Tsaftaris A, Galanopoulou D, Farmaki T. 2010. Cold acclimation and low temperature resistance in cotton: *Gossypium hirsutum* phospholipase alpha isoforms are differentially regulated by temperature and light. *Journal of Experimental Botany* 61:2991–3002.
- Karimi M, Ebadi A, Mousavi SA, Salami SA, Zarei A. 2015. Comparison of CBF1, CBF2, CBF3 and CBF4 expression in some grapevine cultivars and species under cold stress. *Scientia Horticulturae* 197:521–526.
- Katiyar A, Smita S, Lenka SK, Rajwanshi R, Chinnusamy V, Bansal KC. 2012. Genome-wide classification and expression analysis of MYB transcription factor families in rice and *Arabidopsis*. *BMC Genomics* 13.
- Kidokoro S, Watanabe K, Ohori T, Moriwaki T, Maruyama K, Mizoi J, Myint Phyu Sin Htwe N, Fujita Y, Sekita S, Shinozaki K, Yamaguchi-Shinozaki K. 2015. Soybean DREB1/CBF-type transcription factors function in heat and drought as well as cold stress-responsive gene expression. *The Plant Journal* 81:505–518.
- Kim YS, Lee M, Lee JH, Lee HJ, Park CM. 2015. The unified ICE-CBF pathway provides a transcriptional feedback control of freezing tolerance during cold acclimation in *Arabidopsis*. *Plant Molecular Biology* 89:187–201.
- Kim D, Pertea G, Trapnell C, Pimentel H, Kelley R, Salzberg SL. 2013. TopHat2: accurate alignment of transcriptomes in the presence of insertions, deletions and gene fusions. *Genome Biology* 14:R36.
- Kirungu JN, Deng Y, Cai X, Magwanga RO, Zhou Z, Wang X, Wang Y, Zhang Z, Wang K, Liu F. 2018. Simple sequence repeat (SSR) genetic linkage map of D genome diploid cotton derived from an interspecific cross between *Gossypium davidsonii* and *Gossypium klotzschianum*. *International Journal of Molecular Sciences* 19.
- Kizis D, Lumbereras V, Pagès M. 2001. Role of AP2/EREBP transcription factors in gene regulation during abiotic stress. *FEBS Letters* 498:187–189.
- Kocsy G, Galiba G, Brunold C. 2001. Role of glutathione in adaptation and signalling during chilling and cold acclimation in plants. *Physiologia Plantarum* 113:158–164.
- Kumar KG, Ullanat R, Jayabaskaran C. 2004. Molecular cloning, characterization, tissue-specific and phytohormone-induced expression of calcium-dependent protein kinase gene in cucumber (*Cucumis sativus* L.). *Journal of Plant Physiology* 161:1061–1071.
- Kurbidaeva A, Ezhova T, Novokreshchenova M. 2014. *Arabidopsis thaliana* ICE2 gene: phylogeny, structural evolution and functional diversification from ICE1. *Plant Science* 229:10–22.
- Lee SH, Chung GC, Jang JY, Ahn SJ, Zwiazek JJ. 2012. Overexpression of PIP2;5 aquaporin alleviates effects of low root temperature on cell hydraulic conductivity and growth in *Arabidopsis*. *Plant Physiology* 159:479–488.
- Lee JA, Fang DD. 2015. Cotton as a world crop: origin, history, and current status. In: *Cotton*, 2nd edn. Madison, WI: Agron. Monogr. 57. ASA, CSSA, and SSSA, 1–24.
- Lee C-M, Thomashow MF. 2012. Photoperiodic regulation of the C-repeat binding factor (CBF) cold acclimation pathway and freezing tolerance in *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences of the United States of America* 109:15054–15059.
- Li DD, Tai FJ, Zhang ZT, Li Y, Zheng Y, Wu YF, Li XB. 2009. A cotton gene encodes a tonoplast aquaporin that is involved in cell tolerance to cold stress. *Gene* 438:26–32.
- Li XP, Tian AG, Luo GZ, Gong ZZ, Zhang JS, Chen SY. 2005. Soybean DRE-binding transcription factors that are responsive to abiotic stresses. *Theoretical and Applied Genetics* 110:1355–1362.
- Liang Z, Jiang R, Zhong W, Zhao G, Li A, Feng H, Li Z, Sui S. 1996. Studies on the trispecific hybrid (*Gossypium barbadense* - *G. thurberi* - *G. hirsutum*) and new variety breeding. *Acta Agronomica Sinica* 22:673–680.
- Liu T, Guo S, Lian Z, Chen F, Yang Y, Chen T, Ling X, Liu A, Wang R, Zhang B. 2015. A P4-ATPase gene GbPATP of cotton confers chilling tolerance in plants. *Plant & Cell Physiology* 56:549–557.
- Liu Y, Schiff M, Dinesh-Kumar SP. 2002. Virus-induced gene silencing in tomato. *The Plant Journal* 31:777–786.
- Lu P, Magwanga RO, Kirungu JN, Hu Y, Dong Q, Cai X, Zhou Z, Wang X, Zhang Z, Hou Y, Wang K. 2019. Overexpression of cotton a DTX/MATE gene enhances drought, salt and cold stress tolerance in transgenic *Arabidopsis* overexpression of cotton A DTX/MATE gene enhances drought, salt and cold stress tolerance in *Arabidopsis*. 10:1–26.

- Lu P, Magwanga RO, Lu H, Kirungu J, Wei Y, Dong Q, Wang X, Cai X, Zhou Z, Wang K, Liu F. 2018. A novel G-protein-coupled receptors gene from upland cotton enhances salt stress tolerance in transgenic *Arabidopsis*. *Genes* 9.
- Luan S. 2009. The CBL-CIPK network in plant calcium signaling. *Trends in Plant Science* 14:37–42.
- Ma Y, Dai X, Xu Y, Luo W, Zheng X, Zeng D, Pan Y, Lin X, Liu H, Zhang D, Xiao J, Guo X, Xu S, Niu Y, Jin J, Zhang H, Xu X, Li L, Wang W, Qian Q, Ge S, Chong K. 2015. COLD1 confers chilling tolerance in rice. *Cell* 160:1209–1221.
- Ma LF, Zhang JM, Huang GQ, Li Y, Li XB, Zheng Y. 2014. Molecular characterization of cotton C-repeat/dehydration-responsive element binding factor genes that are involved in response to cold stress. *Molecular Biology Reports* 41:4369–4379.
- Magwanga RO, Kirungu JN, Lu P, Yang X, Dong Q, Cai X, Xu Y, Wang X, Zhou Z, Hou Y, Nyunja R. 2019a. Genome wide identification of the trihelix transcription factors and overexpression of Gh\_A05G2067 (GT-2), a novel gene contributing to increased drought and salt stresses tolerance in cotton. *Physiologia Plantarum* 10.
- Magwanga RO, Lu P, Kirungu JN, Lu H, Wang X, Cai X, Zhou Z, Zhang Z, Salih H, Wang K, Liu F. 2018. Characterization of the late embryogenesis abundant (LEA) proteins family and their role in drought stress tolerance in upland cotton. *BMC Genetics* 19.
- Magwanga RO, Lu P, Kirungu JN, Dong Q, Cai X, Zhou Z, Wang X, Hou Y, Xu Y, Peng R, Agong SG. 2019b. Knockdown of cytochrome P450 genes Gh\_D07G1197 and Gh\_A13G2057 on chromosomes D07 and A13 reveals their putative role in enhancing drought and salt stress tolerance in *Gossypium hirsutum*. *Genes* 10:226.
- Mahmood NA, Zaki ZS, Jaafar TH, Alaamier RA. 2011. Measurement of total antioxidant status (TAO) and superoxide dismutase (SOD), catalase (CAT) enzymes in petrol station workers. *Iraqi Journal of Community Medicine* 24:120–124.
- Mansour MMF. 2013. Plasma membrane permeability as an indicator of salt tolerance in plants. *Biologia Plantarum* 57:1–10.
- Meng X, Wang JR, Wang GD, Liang XQ, Li XD, Meng QW. 2015. An R2R3-MYB gene, LeAN2, positively regulated the thermo-tolerance in transgenic tomato. *Journal of Plant Physiology* 175:1–8.
- Mittler R, Blumwald E. 2015. The roles of ROS and ABA in systemic acquired acclimation. *The Plant Cell* 27:64–70.
- Miura K, Furumoto T. 2013. Cold signaling and cold response in plants. *International Journal of Molecular Sciences* 14:5312–5337.
- Munir S, Liu H, Xing Y, Hussain S, Ouyang Y, Zhang Y, Li H, Ye Z. 2016. Overexpression of calmodulin-like (ShCML44) stress-responsive gene from *Solanum habrochaites* enhances tolerance to multiple abiotic stresses. *Scientific Reports* 6.
- Naser L, Kourosh V, Bahman K, Reza A. 2010. Soluble sugars and proline accumulation play a role as effective indices for drought tolerance screening in Persian walnut (*Juglans regia* L.) during germination. *Fruits* 65:97–112.
- Niu X, Guan Y, Chen S, Li H. 2017. Genome-wide analysis of basic helix-loop-helix (bHLH) transcription factors in *Brachypodium distachyon*. *BMC Genomics* 18.
- Pei H, Wang H, Wang L, Zheng F, Dong C-H. 2017. Regulatory function of ethylene in plant responses to drought, cold, and salt stresses. In: Pandey GK, ed. *Mechanism of plant hormone signaling under stress*. Hoboken, NJ: John Wiley & Sons, Inc., 327–344.
- Peleg Z, Blumwald E. 2011. Hormone balance and abiotic stress tolerance in crop plants. *Current Opinion in Plant Biology* 14:290–295.
- Rabinowicz PD, Braun EL, Wolfe AD, Bowen B, Grotewold E. 1999. Maize R2R3 Myb genes: sequence analysis reveals amplification in the higher plants. *Genetics* 153:427–444.
- Rehman S, Mahmood T. 2015. Functional role of DREB and ERF transcription factors: regulating stress-responsive network in plants. *Acta Physiologiae Plantarum* 37.
- Renny-Byfield S, Page JT, Udall JA, Sanders WS, Peterson DG, Arick MA II, Grover CE, Wendel JF. 2016. Independent domestication of two old world cotton species. *Genome Biology and Evolution* 8:1940–1947.
- Sah SK, Reddy KR, Li J. 2016. Abscisic acid and abiotic stress tolerance in crop plants. *Frontiers in Plant Science* 7.
- Sairam RK, Deshmukh PS, Shukla DS. 1997. Tolerance of drought and temperature stress in relation to increased antioxidant enzyme activity in wheat. *Journal of Agronomy and Crop Science* 178:171–178.
- Salih H, Gong W, He S, Sun G, Sun J, Du X. 2016. Genome-wide characterization and expression analysis of MYB transcription factors in *Gossypium hirsutum*. *BMC Genetics* 17:129.
- Shan DP, Huang JG, Yang YT, Guo YH, Wu CA, Yang GD, Gao Z, Zheng CC. 2007. Cotton GhDREB1 increases plant tolerance to low temperature and is negatively regulated by gibberellic acid. *The New Phytologist* 176:70–81.
- Shi Y, Ding Y, Yang S. 2018. Molecular regulation of CBF signaling in cold acclimation. *Trends in Plant Science* 23:623–637.
- Shibasaki K, Uemura M, Tsurumi S, Rahman A. 2009. Auxin response in *Arabidopsis* under cold stress: underlying molecular mechanisms. *The Plant Cell* 21:3823–3838.
- Song SY, Chen Y, Chen J, Dai XY, Zhang WH. 2011. Physiological mechanisms underlying OsNACS-dependent tolerance of rice plants to abiotic stress. *Planta* 234:331–345.
- Song Y, Liu L, Feng Y, Wei Y, Yue X, He W, Zhang H, An L. 2015. Chilling and freezing induced alterations in cytosine methylation and its association with the cold tolerance of an alpine subnival plant, *Chorispora bungeana*. *PLoS One* 10: e0135485.
- Stracke R, Werber M, Weisshaar B. 2001. The R2R3-MYB gene family in *Arabidopsis thaliana*. *Current Opinion in Plant Biology* 4:447–456.
- Su CF, Wang YC, Hsieh TH, Lu CA, Tseng TH, Yu SM. 2010. A novel MYB3-dependent pathway confers cold tolerance in rice. *Plant Physiology* 153:145–158.
- Sun X, Zhao T, Gan S, Ren X, Fang L, Karungo SK, Wang Y, Chen L, Li S, Xin H. 2016. Ethylene positively regulates cold tolerance in grapevine by modulating the expression of ETHYLENE RESPONSE FACTOR 057. *Scientific Reports* 6.
- Tanimoto E. 2005. Regulation of root growth by plant hormones - roles for auxin and gibberellin. *Critical Reviews in Plant Sciences* 24:249–265.
- Thalmann M, Pazmino D, Seung D, Horrer D, Nigro A, Meier T, Kölling K, Pfeifhofer HW, Zeeman SC, Santelia D. 2016. Regulation of leaf starch degradation by abscisic acid is important for osmotic stress tolerance in plants. *The Plant Cell* 28:1860–1878.
- Tian X, Wang Z, Li X, Lv T, Liu H, Wang L, Niu H, Bu Q. 2015. Characterization and functional analysis of pyrabactin resistance-like abscisic acid receptor family in rice. *Rice* 8.
- Verma V, Ravindran P, Kumar PP. 2016. Plant hormone-mediated regulation of stress responses. *BMC Plant Biology* 16.
- Walker GP, Natwick ET. 2006. Resistance to silverleaf whitefly, *Bemisia argentifolii* (Hem., Aleyrodidae), in *Gossypium thurberi*, a wild cotton species. *Journal of Applied Entomology* 130:429–436.
- Wang DZ, Jin YN, Ding XH, Wang WJ, Zhai SS, Bai LP, Guo ZF. 2017. Gene regulation and signal transduction in the ICE-CBF-COR signaling pathway during cold stress in plants. *Biochemistry (Moscow)* 82:1103–1117.
- Wang SY, Wang CY, Wellburn AR. 1990. Role of ethylene under stress conditions. *Stress Responses in Plants: Adaptation and Acclimation Mechanisms* 12:147–173.
- Wang Q, Wang M, Zhang X, Hao B, Kaushik SK, Pan Y. 2011. WRKY gene family evolution in *Arabidopsis thaliana*. *Genetica* 139:973–983.
- Wang CL, Zhang SC, Qi SD, Zheng CC, Wu CA. 2016. Delayed germination of *Arabidopsis* seeds under chilling stress by overexpressing an abiotic stress inducible GhTPS11. *Gene* 575:206–212.
- Xiang D, Chai Y, Man L, Sun Y, Zhang T, Wei C, Xie Z, Li H, Zhang W, Liu D, Cheng D. 2017. Overexpression of a heading Chinese cabbage ICE1 gene confers freezing tolerance in transgenic rice. *Plant Cell, Tissue and Organ Culture* 128:43–54.
- Xiang D, Man L, Yin K, Song Q, Wang L, Zhao M, Xu Z. 2013. Overexpression of a ItICE1 gene from *Isatis tinctoria* enhances cold tolerance in rice. *Molecular Breeding* 32:617–628.
- Xiao H, Tattersall EA, Siddiqua MK, Cramer GR, Nassuth A. 2008. CBF4 is a unique member of the CBF transcription factor family of *Vitis vinifera* and *Vitis riparia*. *Plant, Cell & Environment* 31:1–10.
- Xue T, Li X, Zhu W, Wu C, Yang G, Zheng C. 2009. Cotton metallothionein GhMT3a, a reactive oxygen species scavenger, increased tolerance



- against abiotic stress in transgenic tobacco and yeast. *Journal of Experimental Botany* **60**:339–349.
- Yuan D, Tang Z, Wang M, Gao W, Tu L, Jin X, Chen L, He Y, Zhang L, Zhu L, Li Y. 2015. The genome sequence of Sea-Island cotton (*Gossypium barbadense*) provides insights into the allopolyploidization and development of superior spinnable fibres. *Scientific Reports* **5**:17662.
- Zhang X, Da Silva JAT, Niu M, Li M, He C, Zhao J, Zeng S, Duan J, Ma G. 2017. Physiological and transcriptomic analyses reveal a response mechanism to cold stress in *Santalum album* L. leaves. *Scientific Reports* **7**.
- Zhang T, Hu Y, Jiang W, Fang L, Guan X, Chen J, Zhang J, Sasaki CA, Scheffler BE, Stelly DM, Hulse-Kemp AM, Wan Q, Liu B, Liu C, Wang S, Pan M, Wang Y, Wang D, Ye W, Chang L, Zhang W, Song Q, Kirkbride RC, Chen X, Dennis E, Llewellyn DJ, Peterson DG, Thaxton P, Jones DC, Wang Q, Xu X, Zhang H, Wu H, Zhou L, Mei G, Chen S, Tian Y, Xiang D, Li X, Ding J, Zuo Q, Tao L, Liu Y, Li J, Lin Y, Hui Y, Cao Z, Cai C, Zhu X, Jiang Z, Zhou B, Guo W, Li R, Chen ZJ. 2015. Sequencing of allotetraploid cotton (*Gossypium hirsutum* L. acc. TM-1) provides a resource for fiber improvement. *Nature Biotechnology* **33**:531–537.
- Zhang Z, Huang R. 2010. Enhanced tolerance to freezing in tobacco and tomato overexpressing transcription factor TERF2/LeERF2 is modulated by ethylene biosynthesis. *Plant Molecular Biology* **73**:241–249.
- Zhang J, Luo W, Zhao Y, Xu Y, Song S, Chong K. 2016. Comparative metabolomic analysis reveals a reactive oxygen species-dominated dynamic model underlying chilling environment adaptation and tolerance in rice. *The New Phytologist* **211**:1295–1310.
- Zhao C, Lang Z, Zhu JK. 2015. Cold responsive gene transcription becomes more complex. *Trends in Plant Science* **20**:466–468.
- Zhao Q, Xiang X, Liu D, Yang A, Wang Y. 2018. Tobacco transcription factor NtbHLH123 confers tolerance to cold stress by regulating the NtCBF pathway and reactive oxygen species homeostasis. *Frontiers in Plant Science* **9**:1–13.
- Zhou MQ, Shen C, Wu LH, Tang KX, Lin J. 2011. CBF-dependent signaling pathway: a key responder to low temperature stress in plants. *Critical Reviews in Biotechnology* **31**:186–192.
- Zhu JK. 2016. Abiotic stress signaling and responses in plants. *Cell* **167**:313–324.