

2012

Comparing the mitochondrial genomes of Wolbachia-dependent and independent filarial nematode species

Samantha N. McNulty
Washington University School of Medicine in St. Louis

Andrew S. Mullin
Washington University School of Medicine in St. Louis

Jefferson A. Vaughan
University of North Dakota

Vasyl V. Tkach
University of North Dakota

Gary J. Weil
Washington University School of Medicine in St. Louis

See next page for additional authors

Follow this and additional works at: https://digitalcommons.wustl.edu/open_access_pubs



Part of the [Medicine and Health Sciences Commons](#)

Recommended Citation

McNulty, Samantha N.; Mullin, Andrew S.; Vaughan, Jefferson A.; Tkach, Vasyl V.; Weil, Gary J.; and Fischer, Peter U., "Comparing the mitochondrial genomes of Wolbachia-dependent and independent filarial nematode species." *BMC Genomics*. 13,. 145. (2012).
https://digitalcommons.wustl.edu/open_access_pubs/1138

This Open Access Publication is brought to you for free and open access by Digital Commons@Becker. It has been accepted for inclusion in Open Access Publications by an authorized administrator of Digital Commons@Becker. For more information, please contact vanam@wustl.edu.

Authors

Samantha N. McNulty, Andrew S. Mullin, Jefferson A. Vaughan, Vasyl V. Tkach, Gary J. Weil, and Peter U. Fischer

RESEARCH ARTICLE

Open Access

Comparing the mitochondrial genomes of *Wolbachia*-dependent and independent filarial nematode species

Samantha N McNulty^{1*}, Andrew S Mullin¹, Jefferson A Vaughan², Vasyly V Tkach², Gary J Weil¹ and Peter U Fischer¹

Abstract

Background: Many species of filarial nematodes depend on *Wolbachia* endobacteria to carry out their life cycle. Other species are naturally *Wolbachia*-free. The biological mechanisms underpinning *Wolbachia*-dependence and independence in filarial nematodes are not known. Previous studies have indicated that *Wolbachia* have an impact on mitochondrial gene expression, which may suggest a role in energy metabolism. If *Wolbachia* can supplement host energy metabolism, reduced mitochondrial function in infected filarial species may account for *Wolbachia*-dependence. *Wolbachia* also have a strong influence on mitochondrial evolution due to vertical co-transmission. This could drive alterations in mitochondrial genome sequence in infected species. Comparisons between the mitochondrial genome sequences of *Wolbachia*-dependent and independent filarial worms may reveal differences indicative of altered mitochondrial function.

Results: The mitochondrial genomes of 5 species of filarial nematodes, *Acanthocheilonema viteae*, *Chandlerella quisquali*, *Loa loa*, *Onchocerca flexuosa*, and *Wuchereria bancrofti*, were sequenced, annotated and compared with available mitochondrial genome sequences from *Brugia malayi*, *Dirofilaria immitis*, *Onchocerca volvulus* and *Setaria digitata*. *B. malayi*, *D. immitis*, *O. volvulus* and *W. bancrofti* are *Wolbachia*-dependent while *A. viteae*, *C. quisquali*, *L. loa*, *O. flexuosa* and *S. digitata* are *Wolbachia*-free. The 9 mitochondrial genomes were similar in size and AT content and encoded the same 12 protein-coding genes, 22 tRNAs and 2 rRNAs. Synteny was perfectly preserved in all species except *C. quisquali*, which had a different order for 5 tRNA genes. Protein-coding genes were expressed at the RNA level in all examined species. In phylogenetic trees based on mitochondrial protein-coding sequences, species did not cluster according to *Wolbachia* dependence.

Conclusions: Thus far, no discernable differences were detected between the mitochondrial genome sequences of *Wolbachia*-dependent and independent species. Additional research will be needed to determine whether mitochondria from *Wolbachia*-dependent filarial species show reduced function in comparison to the mitochondria of *Wolbachia*-independent species despite their sequence-level similarities.

Background

Filarial nematodes are arthropod borne parasitic worms that infect hundreds of millions of people throughout the tropics and sub-tropics and are responsible for a great deal of morbidity in humans and domestic animals. Many filarial pathogens, such as the agents of lymphatic filariasis and river blindness, require a bacterial

endosymbiont, *Wolbachia pipientis*, to carry out their life cycle [1-4]. In these species, depletion of the endosymbiont causes defects in growth, molting and fertility, leading to the death of the worm [5-7]. Other filarial species, some of which are very closely related to *Wolbachia*-dependent sister taxa, are naturally *Wolbachia*-free [1,2,8-10]. Thus far, there are no discernable patterns in *Wolbachia* distribution (e.g., based on host species, vector species, tissue tropism, geographic distribution, etc.), and the reasons for this disparity are poorly understood. Presumably, some genetic function(s) must be missing or reduced in *Wolbachia*-dependent worms in comparison to their

* Correspondence: smcnulty@wustl.edu

¹Infectious Diseases Division, Department of Internal Medicine, Washington University School of Medicine, Campus Box 8051, 660 S. Euclid Avenue, St. Louis, MO 63110, USA

Full list of author information is available at the end of the article

Wolbachia-free counterparts, forcing them to rely on the bacteria as an alternative source of vital gene products. The processes underpinning *Wolbachia*-dependence are of biological and medical interest, as the *Wolbachia* products required by the dependent worm may represent useful targets for novel anti-filarial chemotherapies.

Wolbachia endobacteria and the eukaryotic mitochondria share many common features, including the intracellular lifestyle, obligatory mutualism, reduced genome size, vertical transmission, etc. These shared features, as well as their shared ancestry in the order Rickettsiales [11-13], lead us to hypothesize that *Wolbachia* may contribute to energy metabolism in the filarial host. Previous studies have shown that antibiotic-mediated *Wolbachia* depletion leads to upregulation in genes related to energy metabolism, including mitochondrially encoded subunits of the respiratory chain [14]. This impact on host mitochondrial gene expression, and presumably energy production, suggests that *Wolbachia* may serve as an alternative energy source or mitochondrial “supplement,” necessitating increased activity when the endosymbiont is removed. If so, differences in mitochondrial function may account for discrepancies in *Wolbachia* status in the filarial lineage.

The mitochondrial genome (mtDNA) is particularly sensitive to evolutionary pressure exerted by the *Wolbachia* infection. Vertically-transmitted *Wolbachia* are able to expand rapidly through insect populations due to the mechanisms of reproductive parasitism [15]. *Wolbachia* and mitochondria are co-transmitted. Thus, the mtDNA(s) of the first infected individual(s) presumably expand concurrently with the *Wolbachia* infection. Such *Wolbachia*-mitochondria “sweeps,” characterized by unusually low degrees of variation in the mtDNA of infected populations, have been noted in many insect species [16-20]. A similar lack of mtDNA diversity is seen in populations of *Dirofilaria immitis* (canine heartworm) in comparison to *Wolbachia*-free, non-filarial nematodes [21]. A *Wolbachia*-induced genetic bottleneck may have led to the fixation of different mtDNA types among infected filarial species as compared to uninfected species.

The mtDNA sequences of 4 species of filarial nematodes, *Onchocerca volvulus* [22], *D. immitis* [23], *Brugia malayi* [24], and *Setaria digitata* [25], have been published. This report details the sequencing and analysis of the mtDNA sequences of 5 more species: *Acanthocheilonema viteae*, *Chandlerella quisquali*, *Loa loa*, *Onchocerca flexuosa* and *Wuchereria bancrofti*. Studies of the distribution of *Wolbachia* within filarial nematodes have shown that the infection is prevalent among 2 of the 8 filarial subfamilies, the Onchocercinae and the Dirofiliariinae [1,2]. Agreement between the phylogenies of *Wolbachia* and their filarial hosts suggests that *Wolbachia* entered the filarial lineage prior to the diversification of these 2 subfamilies [1,26]. 7 of

the 9 species included in this study are members of the the Onchocercinae and Dirofiliariinae. Four of these, *B. malayi*, *D. immitis*, *O. volvulus* and *W. bancrofti*, are *Wolbachia*-dependent [1,3,4,27]. The other 3, *A. viteae*, *L. loa* and *O. flexuosa*, are *Wolbachia*-free [1,8,10,28], presumably due to secondary loss of the endosymbiont [1,29]. Conversely, *C. quisquali* and *S. digitata* are *Wolbachia*-free and belong to subfamilies (Splendidofiliariinae and Setariinae, respectively) that have not been shown to contain *Wolbachia*-infected species, suggesting that these subfamilies split from the lineage prior to the introduction of *Wolbachia* endobacteria [2,9].

In light of the presumed impact of *Wolbachia* on the host mitochondria, we hypothesized that the mtDNAs of *Wolbachia*-dependent filaria may differ in gene content, arrangement or sequence as compared to those found in *Wolbachia*-free species whose ancestor(s) may not have undergone a *Wolbachia*-induced genetic bottleneck or evolved in the presence of an endobacterial partner capable of affecting host energy metabolism. The purpose of the reported study was to compare mtDNA from *Wolbachia*-dependent and independent filarial species in search of sequence level differences indicative of altered mitochondrial function. Our analyses revealed no differences that could be attributed to *Wolbachia* status. Future studies will be required to discover subtler affects of *Wolbachia* on the sequence or function of filarial nematode mitochondria.

Results

Gene content and organization

The mtDNAs of 5 species of filarial nematodes were sequenced, annotated and deposited in Genbank (see Table 1 for accession numbers). Genome length, AT-richness and base composition of the 9 mtDNAs are compared in Table 1. The newly sequenced mtDNAs are similar in size and AT content to those of other filarial species. So far, filarial mtDNAs range in size from 13,474 bp in *O. volvulus* to 13,839 in *S. digitata* and range in AT content from 73.7% in *O. volvulus* to 77.7% in *C. quisquali* [22,25].

All 9 filarial mtDNAs encode the same 12 proteins, 22 tRNAs and 2 rRNAs with very short intergenic sequences (Figure 1). These genes are encoded in the same direction, a characteristic shared by most nematode mtDNAs. Synteny is perfectly preserved in all examined species with the exception of *C. quisquali* (Figure 1). In 8 of the 9 species, 5 tRNA genes (tRNA^{Ala}, tRNA^{Leu2}, tRNA^{Asn}, tRNA^{Met} and tRNA^{Lys}) reside between the AT-rich region and NDL4. In *C. quisquali*, the tRNA^{Met} gene is positioned between Cox3 and the AT-rich region apart from the main tRNA cluster, and the order of the other 4 tRNA genes is rearranged relative to other species. For a comparisons between the mtDNA arrangement among filarial and other nematodes, see [25].

Table 1 Comparison of filarial nematode mtDNAs

Species	<i>Wolbachia</i> -dependent species					<i>Wolbachia</i> -independent species			
	<i>B. malayi</i>	<i>D. immitis</i>	<i>O. volvulus</i>	<i>W. bancrofti</i>	<i>A. viteae</i>	<i>L. loa</i>	<i>O. flexuosa</i>	<i>C. quiscali</i>	<i>S. digitata</i>
Subfamily	Onchocercinae	Dirofiliariinae	Onchocercinae	Onchocercinae	Onchocercinae	Dirofiliariinae	Onchocercinae	Splendidofiliariinae	Setariinae
Accession Number	NC_004298	NC_005305	NC_001861	HQ184469	HQ186249	HQ186250	HQ214004	HM773029	NC_014282
Length (bp)	13,657	13,814	13,474	13,636	13,724	13,590	13,672	13,757	13,839
Length of AT-rich region (bp)	283	362	312	256	421	288	284	308	506
A%	21.60%	19.30%	19.30%	20.50%	19.60%	20.80%	20.30%	23.00%	19.40%
T%	53.90%	54.90%	54.00%	54.10%	54.00%	54.80%	53.90%	54.70%	55.70%
G%	16.80%	19.30%	19.80%	18.00%	19.30%	17.70%	18.60%	15.90%	18.20%
C%	7.70%	6.50%	6.90%	7.40%	7.20%	6.70%	7.20%	6.40%	6.70%
AT%	75.50%	74.20%	73.30%	74.60%	73.50%	75.60%	74.20%	77.70%	75.10%

Information was taken from previous studies for *B. malayi* [24], *D. immitis* [23], *O. volvulus* [22], and *S. digitata* [25]. Information regarding the *Wolbachia* status of species from various subfamilies is reported in [2].

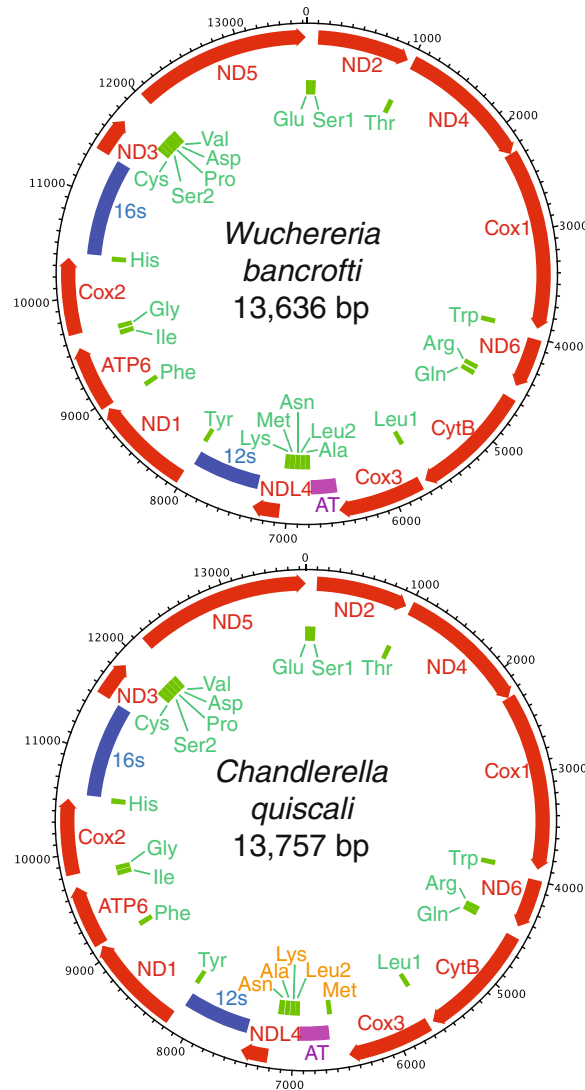


Figure 1 Comparative diagrams of the mitochondrial genomes of *W. bancrofti* and *C. quiscali*. Protein-coding genes are shown in red with arrowheads indicating directionality. rRNA and tRNA genes are shown in blue and green, respectively, and the AT-rich region is shown in purple. The diagram of the *W. bancrofti* mitochondrial genome is representative of most filarial mitochondria, as synteny is preserved in all species except *C. quiscali*. The 5 tRNA genes rearranged in *C. quiscali* are highlighted in orange.

Protein-coding genes

Twelve protein-coding genes were identified in each of the examined mtDNAs. None of these contain premature stop codons or frameshift mutations. Reverse transcription PCR reactions indicate that the predicted protein-coding genes were expressed at the RNA level in all examined species (Figure 2).

Filarial mtDNAs are extremely thymine (T)-rich (Table 1); therefore, it is not surprising that filarial mitochondria show a bias towards T-rich codons (Additional file 1: Table S1). The most frequently used codon in all species is TTT, which encodes phenylalanine and serves as an alternative start codon in certain instances (Additional file 1: Table S1, Table 2). The start and stop codons used by each species

are listed in Table 2. Novel start codons include TGT for ND6 in *W. bancrofti*, TCT for CytB in *A. viteae*, and CCT for ND3 in *O. flexuosa*. Termination codons include TAG, TAA, and the incomplete stop codon T, which is converted to TAA upon addition 3' poly(A) tail.

Ribosomal and transfer RNA genes

All species encode 2 rRNA genes. In all species examined, the 12s rRNA gene is positioned between NDL4 and ND1 while the 16s rRNA gene is positioned between Cox2 and ND3 (Figure 1). The exact boundaries of these genes have yet to be mapped in any filarial species.

All species also contain the same 22 tRNA genes. In the previously sequenced species, 20 of the 22 mitochondrial

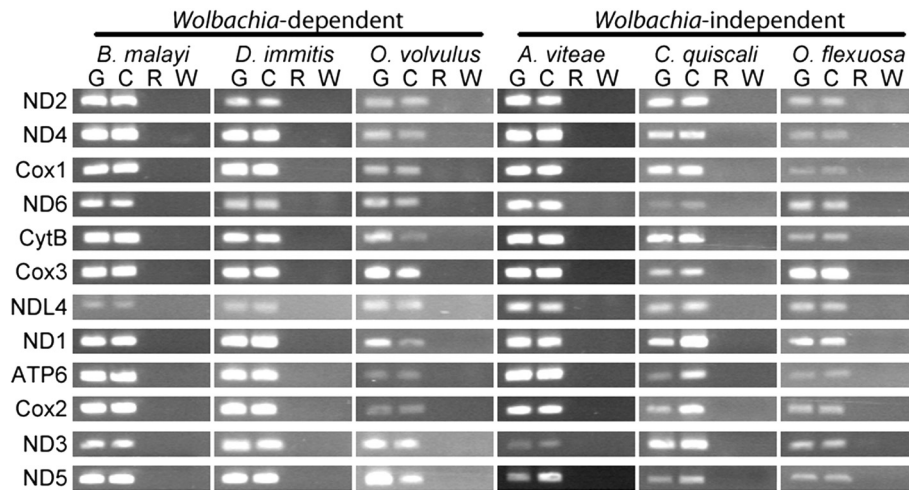


Figure 2 Expression of mitochondrial protein-coding genes in six filarial nematode species. Expression of mitochondrial protein-coding genes was assessed by reverse transcription PCR in *B. malayi*, *D. immitis*, *O. volvulus*, *A. viteae*, *C. quiscalis* and *O. flexuosa*. The following templates were used for each reaction: genomic DNA (G), cDNA (C), total RNA (R) and water (W).

tRNAs share a common secondary structure in which the TΨC arm and variable loop are exchanged for a TV-replacement loop [22,23] (Figure 3b). Conversely, the two tRNA^{Ser} genes contain a DHU replacement loop in exchange for the typical D arm (Figure 3c) [22,23]. The predicted mitochondrial tRNA structures of *A. viteae* followed this trend exactly, as did most of the tRNAs from the other examined species. However, our predictions indicate that tRNA^{Ser1} and tRNA^{Asn} in *C. quiscalis*, tRNA^{Lys} and tRNA^{Pro} in *L. loa*, tRNA^{Trp} in *O. flexuosa* and tRNA^{Pro} in *W. bancrofti* may contain both the TΨC

and D loops (Figure 3d-i). The same anticodons are used in all species with two exceptions. tRNA^{Pro} uses the anticodon AGG in *O. volvulus*, *D. immitis*, *S. digitata* and *O. flexuosa*, while the anticodon TGG is used in other species, and tRNA^{Leu1} uses the anticodon TAA in *A. viteae* while the anticodon TAG is used in other species.

AT rich region

The control, or AT rich, region represents the largest non-coding region in filarial mtDNAs, which are otherwise densely packed with tightly spaced or slightly overlapping

Table 2 Start and stop codons used in mitochondrial protein-coding genes

	<i>Wolbachia</i> -dependent species				<i>Wolbachia</i> -independent species				
	<i>B. malayi</i>	<i>D. immitis</i>	<i>O. volvulus</i>	<i>W. bancrofti</i>	<i>A. viteae</i>	<i>L. loa</i>	<i>O. flexuosa</i>	<i>C. quiscalis</i>	<i>S. digitata</i>
ND2	TTA/T	ATT/TAG	ATT/TAG	TTA/T	TTT/TAG	ATT/TAA	ATT/TAG	ATT/TAG	TTT/TAG
ND4	TTG/TAA	TTG/TAG	TTG/TAA	TTG/TAA	ATG/TAA	TTG/TAG	TTG/TAG	TTG/TAA	ATG/TAA
COX1	ATT/TAG	ATT/TAG	ATT/TAG	ATT/TAA	GTT/T	GTT/T	ATT/TAA	TTG/TAA	ATT/TAG
ND6	TAT/TAA	TAT/TAG	ATT/TAG	TGT/TAA	TAT/TAG	TAT/TAG	ATT/TAA	TTG/TAG	TTG/TAA
CYTB	ATT/T	GTT/T	ATT/TAA	ATT/T	TCT/T	ATT/T	ATT/TAA	ATT/T	GTT/T
COX3	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAG	ATT/TAA	ATA/T
NDL4	GTA/TAA	GTA/TAA	TTG/TAA	GTA/TAA	GTAT	GTA/T	TTA/T	GTT/TAA	TTG/T
ND1	TTG/T	TTG/T	TTG/T	TTG/TAA	TTG/T	TTG/T	TTG/T	TTG/T	TTG/TAA
ATP6	ATT/TAG	TTG/TAA	ATT/TAG	ATT/TAG	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAG	TTT/TAG
COX2	ATT/TAA	ATT/T	ATT/TA	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAA	ATT/TAG
ND3	CTT/TAG	CTT/T	CTT/TAG	CTT/T	CTT/T	CTT/T	CCT/T	CTT/TAG	TTT/T
ND5	TTT/TAG	TTG/TAG	TTG/TAG	TTT/TAG	TTT/TAG	TTT/TAG	TTA/TAA	TTT/T	TTT/TAG

Information was taken from previous studies for *B. malayi* [24], *D. immitis* [23], *O. volvulus* [22], and *S. digitata* [25]. Information regarding the *Wolbachia* status of species from various subfamilies is reported in [2].

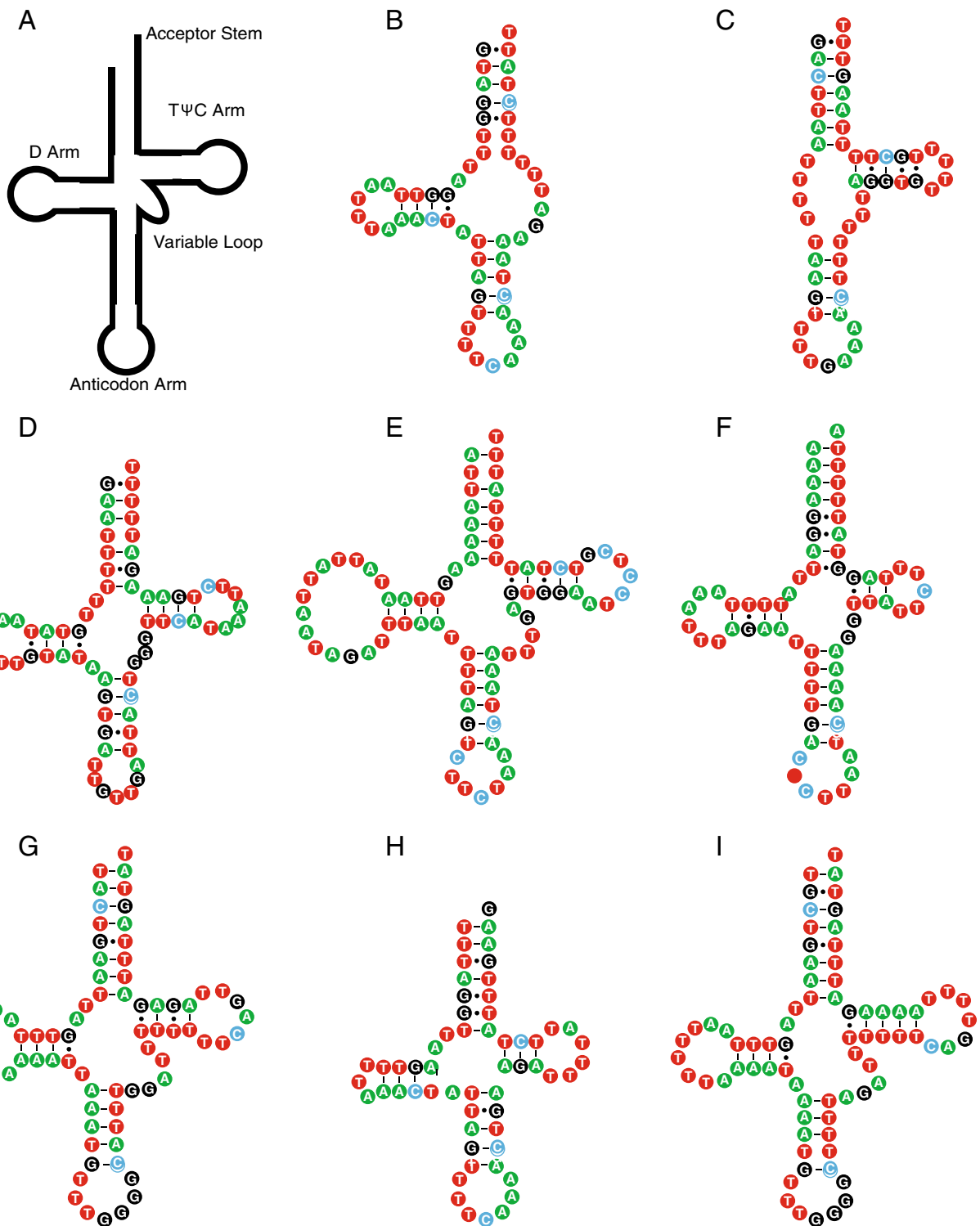
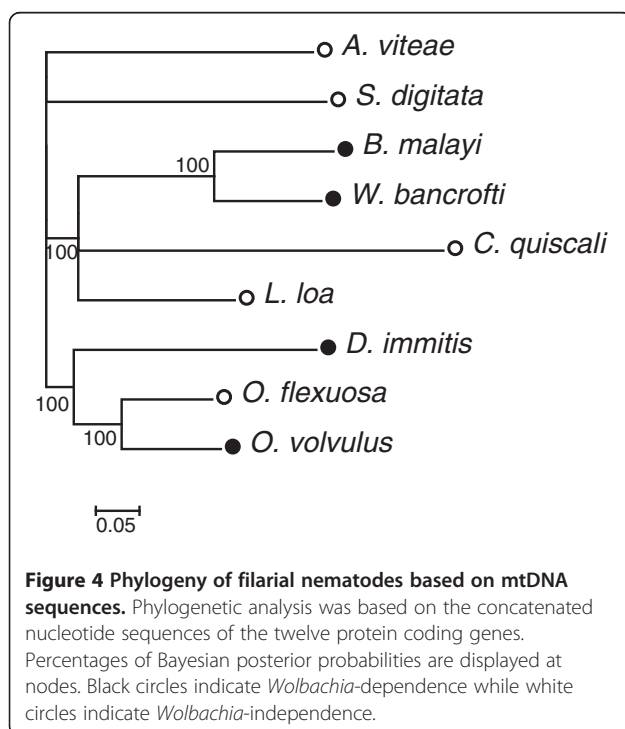


Figure 3 Mitochondrial tRNA structures. The features of a typical tRNA include the acceptor stem, D arm, TΨC arm, variable loop and anticodon arm (A). In most species, tRNA^{Ser1} and tRNA^{Ser2} contain a DHU replacement loop and TΨC arm, as in tRNA^{Ser2} from *A. viteae* (B), while all other tRNAs contain a D arm and TV replacement loop, as in tRNA^{Trp} of *A. viteae* (C). Exceptions may include tRNA^{Asn} (D) and tRNA^{Ser1} (E) from *C. quiscali*, tRNA^{Lys} (F) and tRNA^{Pro} (G) from *L. loa*, tRNA^{Trp} (H) from *O. flexuosa*, and tRNA^{Pro} (I) from *W. bancrofti*, as these structures are predicted to include both the D and TΨC arms.

protein-coding, tRNA and rRNA genes. The AT rich regions of the 9 sequenced filarial mtDNAs range in size from 256 bp in *W. bancrofti* to 506 bp in *S. digitata* (Table 1). In most species, this region is located between Cox3 and tRNA^{Ala}. The unusual arrangement of tRNA genes in *C. quiscali* places its proposed 308 bp AT rich region between the tRNA^{Met} and tRNA^{Leu2} genes, leaving an additional 109 bp non-coding region between the Cox3 and tRNA^{Met} genes. The function of this secondary non-coding region is unknown.

Phylogenetic analysis

A phylogenetic analysis was carried out using the nucleotide sequences of the 12 protein coding genes from the fully-sequenced filarial mtDNAs (Figure 4). Trees were left unrooted since the closest relatives of filarial nematodes with complete mtDNA sequences (i.e. *Ascaris* and *Toxocara* species) are still too divergent to allow for accurate alignment. Overall, topology is similar to that of trees based on single mitochondrial genes (i.e. 12s rRNA gene, Cox I) with improved statistical support [2,26,30]. As in the previous studies, our molecular phylogeny does not agree with the classical taxonomy of the filariae, as the Dirofilarinae and Onchocercinae appear as polyphyletic groups. In our tree, the lymphatic filariae cluster together, as do the genera *Onchocerca* and *Dirofilaria*. *C. quiscali*, which has not been included in previous analyses, is most closely related to the lymphatic filariae and *L. loa*. *A. viteae* and *S. digitata* are basal to the other species in our study.



Simpler neighbor joining trees were constructed for each individual gene (data not shown). As in the tree based on concatenated protein coding sequences, clustering is never reflective of *Wolbachia* status.

Discussion

Filarial nematodes are widespread parasites that infect all classes of vertebrates except fish [31]. Many of these are of socioeconomic and medical importance. However, the mtDNAs had only been sequenced from 4 filarial species [22-25]. In our study we characterized the mtDNAs of 5 additional species and used the newly available sequences to compare the mtDNA of *Wolbachia*-dependent and independent filarial nematodes. Initially, we hypothesized that there might be obvious differences in the mitochondrial genome sequences of *Wolbachia*-dependent and independent species due to the evolutionary pressure exerted by a co-transmitted endosymbiont that may impact the energy balance of its host [14,16-21]. However, our data indicate that the mtDNAs of filarial nematodes are, thus far, remarkably similar. No major differences in genome length, AT content or codon usage were detected. All 9 species contain the standard 12 protein-coding genes, 22 tRNAs and 2 rRNAs. Differences in tRNA structure or anticodon usage are also minor and do not correlate to *Wolbachia* status. In light of these findings, it seems that *Wolbachia* has had little effect on the content of filarial mtDNA.

In most filarial species, synteny was perfectly preserved. The rearrangement of 5 tRNAs in *C. quiscali* probably reflects its evolutionary distance from the other species included in this study rather than its *Wolbachia*-free status (see Figure 4), as such rearrangements were not detected in other *Wolbachia*-free species. It will be necessary to sequence the mtDNAs of other members of the Splendidofilarinae in order to determine whether this rearrangement is species-specific or typical of the entire subfamily. However, this minor alteration in gene order is unlikely to impact overall mitochondrial function.

If *Wolbachia* infection had led to the fixation of certain mitochondrial types in an ancestral population, one might expect to see higher degrees of sequence identity between species that have come into contact with the endosymbiont (i.e. species from the Onchocercinae and Dirofilarinae) as compared to species that have not [2]. However, our phylogenetic analysis indicated that the level of sequence identity shared between mitochondrial protein-coding genes is independent of *Wolbachia* status. If it is true that filarial species considered primitive based on classical taxonomy (i.e. *C. quiscali* and *S. digitata*) were never associated with *Wolbachia*, the similarity of their mtDNAs to the others in this study makes it seem unlikely that *Wolbachia* have had a significant impact on mtDNA sequence. The lack of diversity in mtDNA sequences could

be taken as an indication that the initial infection and theoretical *Wolbachia*-induced genetic bottleneck occurred in an ancestor of all filarial nematodes, but this is unlikely given the phylogenetic age of the filariae. Of course, one must consider that this study was performed on the level of complete genes. In the future, it may be informative to compare smaller loci in *Wolbachia*-dependent and independent species, as even single base changes are known to have profound effects on mitochondrial function [32].

The fact that no sequence-level differences were observed between mitochondria from *Wolbachia*-dependent and independent filarial species does not exclude the possibility that differences may exist in mitochondrial function or efficiency between these groups. Of course, many of the genes related to oxidative phosphorylation and energy metabolism are encoded in the nuclear genome rather than the mitochondrial genome. Because only one filarial genome has been published (see [24] for the genome of *B. malayi*), we do not know if certain nuclear genes are missing or altered in *Wolbachia*-dependent species relative to their *Wolbachia*-independent counterparts.

Even if there are no differences in the genes encoded by *Wolbachia*-dependent and independent filarial worms, it is possible that variation in expression levels could lead to differences in mitochondrial output. Expression levels may be partially dictated by the number of mitochondrial genomes present in each mitochondria or the density of mitochondria in a given organism. These factors are also variable. Quantitative real-time PCR techniques could be used to assess expression; however, careful normalization would be necessary to ensure accurate results when comparing expression levels across multiple species and life cycle stages. This type of analysis will not be possible until better nuclear genome data is produced to provide appropriate control sequences.

Mitochondria and *Wolbachia* appear to share a common evolutionary story wherein bacteria of the Rickettsial family were taken up and transformed over time into an obligate mutualist that provides products essential to the life of the host. Since *Wolbachia* and mitochondria are co-transmitted and since *Wolbachia* may have an impact on host energy metabolism [14], it is possible that *Wolbachia* have affected the mitochondria of *Wolbachia*-dependent species in ways that are not reflected in the mtDNA sequence. Additional research will be required to test this hypothesis.

Conclusions

Here we report the mitochondrial genome sequences of 5 species of filarial nematodes: *Acanthocheilonema viteae*, *Chandlerella quiscali*, *Loa loa*, *Onchocerca flexuosa*, and *Wuchereria bancrofti*. Although we had hypothesized that the presence of *Wolbachia* endobacteria in some filarial nematodes may have had an impact on the content, organization or sequence of filarial mtDNA, we found no

evidence that supports this hypothesis. The 9 available filarial mitochondrial sequences are remarkably similar on the sequence level. Future studies may determine whether functional differences exist between the mitochondria of *Wolbachia*-dependent and independent filarial nematodes.

Methods

Parasite materials

Adult *B. malayi* and *A. viteae* were obtained from experimentally infected Mongolian jirds as previously described [33,34]. Adult *D. immitis* were obtained from the Filariasis Research Reagent Resource Center (Athens, GA). Adult *O. flexuosa* were isolated from subcutaneous nodules dissected from red deer (*Cervus elaphus*) in Schleswig-Holstein, Germany [10]. Adult *O. volvulus*, and microfilariae of *W. bancrofti* and *L. loa* were available from prior studies in Uganda, Papua New Guinea and Cameroon, respectively [8,35,36]. Adult *Chandlerella quiscali* were obtained from common grackles *Quiscalus quiscala* trapped in North Dakota, USA.

Nucleic acid isolation and cDNA synthesis

DNA for sequencing was isolated from adult worms and microfilariae using the DNeasy Blood and Tissue Kit (Qiagen, Valencia, CA), ethanol precipitated to concentrate and stored in 1x TE buffer. RNA was isolated as previously described [29]. Briefly, worms were homogenized by bead-beating in TRIzol (Invitrogen, Carlsbad, CA) and RNA was isolated by organic extraction with 1-bromo-3-chloropropane followed by column purification using the RNeasy Mini Kit (Qiagen) including an on-column DNase digest. A second DNase treatment was performed with the TURBO DNA-free Kit (Applied Biosystems, Austin, TX). cDNA was synthesized from total RNA using qScript cDNA SuperMix according to manufacturer's suggested protocol (Quant Biosciences, Gaithersburg, MD) and purified with the Qiaquick PCR Purification Kit (Qiagen).

PCR reactions and sequencing

Primers used to amplify mtDNA in 10 segments are reported in Additional file 2: Table S2. "Filarial Mito" primer sets are designed to target conserved portions of filarial mitochondria. In cases where the conserved primer set failed, species-specific primer sets were implemented. This was the case for segments 1–2, 7 and 9 in *A. viteae*, segments 5 and 8 in *C. quiscali*, segments 1, 4, 8 and 10 in *L. loa*, and segments 7 and 9 in *W. bancrofti*. All PCR reactions were performed using the Platinum Taq High Fidelity DNA polymerase (Invitrogen) according to the manufacturer's suggested protocol with annealing temperatures adjusted to accommodate the thermodynamic properties of the primers. PCR products were cloned using the TOPO-TA Cloning Kit for Sequencing or the

TOPO-XL PCR Cloning Kit (Invitrogen) depending on size, and sequenced by primer walking.

Species-specific primers (given "RT" designation in the primer name) were designed to detect protein-coding sequences from cDNA. The sequences of these primers are reported in Additional file 2: Table S2. To detect expression, each PCR reaction included a DNA positive control, a cDNA test sample and total RNA and water-only negative controls.

Assembly and annotation of the mitochondrial genomes

Contigs were assembled using Contig Express and analyzed using Vector NTI version 10.3.1 (Invitrogen). Sequences were verified by comparison with publically available sequence data from the Genbank sequence read archive for *L. loa* (accession number SRP000756) and *W. bancrofti* (accession number SRP000772).

Protein-coding genes (including initiation and termination codons) and rRNAs were determined based on their homology to sequences reported from the mitochondrial genomes of *B. malayi*, *D. immitis*, *O. volvulus* and *S. digitata* [22-25].

In most instances, tRNA sequences were predicted using Arwen (available at <http://130.235.46.10/ARWEN/>) [37] and verified by homology to known filarial tRNA sequences. Any computationally predicted tRNAs that fell within other documented structures (i.e. protein-coding genes or rRNAs) were disregarded. tRNA^{Ala} and tRNA^{Leu2} in *A. viteae*, tRNA^{Leu2} and tRNA^{Gly} in *O. flexuosa*, and tRNA^{Ala} in *L. loa* were identified solely based on homology to known orthologs and the presence of the expected anticodon.

Base composition and codon usage were calculated using the DNA Stats and codon usage features available from the Sequence Manipulation Suite (http://www.bioinformatics.org/sms2/dna_stats.html). Diagrams of complete mtDNAs were constructed using DNA plotter (<http://www.sanger.ac.uk/resources/software/dnaplotter/>) [38].

Phylogenetic analysis

The nucleotide sequences of the 12 protein coding genes, excluding stop codons, were aligned using Clustal W as implemented in MEGA4 using default parameters [39]. Individual gene alignments were concatenated using FASconCAT [40].

Model selection was performed using MrModeltest2.3 according to the Akaike information criterion [41]. Bayesian Metropolis-coupled Markov chain Monte Carlo (MCMC) analysis was performed on the dataset with the GTR + I + G nucleotide substitution model by MrBayes Version 3.1.2 [42,43]. Two simultaneous runs of 500,000 generations were performed with sampling every 100

generations and a 25% burn-in. The resulting phylogenetic tree was visualized in MEGA4 [39].

Additional files

Additional file 1: Table S1. Codon usage in the mitochondrial genomes of filarial nematodes
Additional file 2: Table S2. Primer sequences

Abbreviations

ATP6: ATP synthase subunit 6; CytB: Cytochrome b; Cox1: Cox2 and Cox3, cytochrome C oxidase subunits 1-3; ND1-6, and NDL4: Nicotinamide adenine dinucleotide dehydrogenase subunits 1-6, and L4; rRNA: Ribosomal RNA; tRNA: Transfer RNA.

Competing interests

The authors declare that they have no competing interests.

Acknowledgements

We would like to thank Richard Lucius (Humboldt University, Berlin), the Filariasis Research Reagent Resource Center (Atlanta, Georgia), Samuel Wanji (University of Buea, Cameroon), Norbert Brattig (Bernard Nocht Institute for Tropical Medicine, Hamburg) and Yuefang Huang (Washington University School of Medicine, St. Louis) for providing parasite material. This research was funded by The Barnes Jewish Hospital Foundation. SNM was supported by NIH grant T32-AI007172 and JAV was supported by NIH grant R03-AI092306.

Author details

¹Infectious Diseases Division, Department of Internal Medicine, Washington University School of Medicine, Campus Box 8051, 660 S. Euclid Avenue, St. Louis, MO 63110, USA. ²Department of Biology, University of North Dakota, 10 Cornell St, Grand Forks, ND 58202, USA.

Authors' contributions

SNM sequenced and assembled the mitochondrial genomes, carried out bioinformatic analyses and drafted the manuscript. ASM assisted in sequencing and assembling the mitochondrial genomes. JAV and WT collected parasite material and assisted in revising the manuscript. GJW and PUF supervised study design and assisted in drafting and revising the manuscript. All authors have read and approved the final manuscript.

Received: 9 November 2011 Accepted: 24 April 2012

Published: 24 April 2012

References

1. Bandi C, Anderson TJ, Genchi C, Blaxter ML: Phylogeny of *Wolbachia* in filarial nematodes. *Proc Biol Sci* 1998, **265**(1413):2407-2413.
2. Ferri E, Bain O, Barbuto M, Martin C, Lo N, Uni S, Landmann F, Baccei SG, Guerrero R, de Souza Lima S, Bandi C, Wanji S, Diagne M, Casiraghi M: New Insights into the Evolution of *Wolbachia* Infections in Filarial Nematodes Inferred from a Large Range of Screened Species. *PLoS One* 2011, **6**(6):e20843.
3. Kozek WJ, Marroquin HF: Intracytoplasmic bacteria in *Onchocerca volvulus*. *Am J Trop Med Hyg* 1977, **26**(4):663-678.
4. Vincent AL, Ash LR, Frommes SP: The ultrastructure of adult *Brugia malayi* (Brug, 1927) (Nematoda: Filarioidea). *J Parasitol* 1975, **61**(3):499-512.
5. Hoerauf A, Mand S, Fischer K, Kruppa T, Marfo-Debrekyei Y, Debrah AY, Pfarr KM, Adjei O, Buttner DW: Doxycycline as a novel strategy against bancroftian filariasis-depletion of *Wolbachia* endosymbionts from *Wuchereria bancrofti* and stop of microfilaria production. *Med Microbiol Immunol* 2003, **192**(4):211-216.
6. Hoerauf A, Mand S, Volkman L, Buttner M, Marfo-Debrekyei Y, Taylor M, Adjei O, Buttner DW: Doxycycline in the treatment of human onchocerciasis: Kinetics of *Wolbachia* endobacteria reduction and of inhibition of embryogenesis in female *Onchocerca* worms. *Microbes Infect* 2003, **5**(4):261-273.
7. Hoerauf A, Nissen-Pahle K, Schmetz C, Henke-Duhrsen K, Blaxter ML, Buttner DW, Gallin MY, Al-Qaoud KM, Lucius R, Fleischer B: Tetracycline therapy

- targets intracellular bacteria in the filarial nematode *Litomosoides sigmodontis* and results in filarial infertility. *J Clin Invest* 1999, **103**(1):11–18.
8. Buttner DW, Wanji S, Bazzocchi C, Bain O, Fischer P: **Obligatory symbiotic *Wolbachia* endobacteria are absent from *Loa loa*.** *Filaria J* 2003, **2**(1):10.
 9. McNulty SN, Fischer K, Mehus JO, Vaughan JA, Tkach W, Weil GJ, Fischer PU: **Absence of *Wolbachia* Endobacteria in *Chandlerella quiscalis*, an Avian Filarial Parasite.** *J Parasitol* 2012, **98**(2):382–387.
 10. Plenge-Bonig A, Kromer A, Buttner DW: **Light and electron microscopy studies on *Onchocerca jakutensis* and *O. flexuosa* of red deer show different host-parasite interactions.** *Parasitol Res* 1995, **81**(1):66–73.
 11. Andersson SG, Zomorodipour A, Andersson JO, Sicheritz-Ponten T, Alsmark UC, Podowski RM, Naslund AK, Eriksson AS, Winkler HH, Kurland CG: **The genome sequence of *Rickettsia prowazekii* and the origin of mitochondria.** *Nature* 1998, **396**(6707):133–140.
 12. Dumler JS, Barbet AF, Bekker CP, Dasch GA, Palmer GH, Ray SC, Rikihisa Y, Rurangirwa FR: **Reorganization of genera in the families Rickettsiaceae and Anaplasmataceae in the order Rickettsiales: unification of some species of *Ehrlichia* with *Anaplasma*, *Cowdria* with *Ehrlichia* and *Ehrlichia* with *Neorickettsia*, descriptions of six new species combinations and designation of *Ehrlichia equi* and 'HGE agent' as subjective synonyms of *Ehrlichia phagocytophila*.** *Int J Syst Evol Microbiol* 2001, **51**(Pt 6):2145–2165.
 13. Foster J, Ganatra M, Kamal I, Ware J, Makarova K, Ivanova N, Bhattacharyya A, Kapatral V, Kumar S, Posfai J, Vincze T, Ingram J, Moran L, Lapidus A, Omelchenko M, Kyrpides N, Ghedin E, Wang S, Goltsman E, Joukov V, Ostrovskaya O, Tsukerman K, Mazur M, Comb D, Koonin E, Slatko B: **The *Wolbachia* genome of *Brugia malayi*: endosymbiont evolution within a human pathogenic nematode.** *PLoS Biol* 2005, **3**(4):e121.
 14. Strubing U, Lucius R, Hoerauf A, Pfarr KM: **Mitochondrial genes for heme-dependent respiratory chain complexes are up-regulated after depletion of *Wolbachia* from filarial nematodes.** *Int J Parasitol* 2010, **40**(10):1193–1202.
 15. Werren JH, Baldo L, Clark ME: ***Wolbachia*: master manipulators of invertebrate biology.** *Nat Rev Microbiol* 2008, **6**(10):741–751.
 16. Delgado AM, Cook JM: **Effects of a sex-ratio distorting endosymbiont on mtDNA variation in a global insect pest.** *BMC Evol Biol* 2009, **9**:49.
 17. Hurst GD, Jiggins FM: **Problems with mitochondrial DNA as a marker in population, phylogeographic and phylogenetic studies: the effects of inherited symbionts.** *Proc Biol Sci* 2005, **272**(1572):1525–1534.
 18. Raychoudhury R, Grillenberger BK, Gadau J, Bijlsma R, van de Zande L, Werren JH, Beukeboom LW: **Phylogeography of *Nasonia vitripennis* (Hymenoptera) indicates a mitochondrial-*Wolbachia* sweep in North America.** *Heredity* 2010, **104**(3):318–326.
 19. Turelli M, Hoffmann AA: **Rapid spread of an inherited incompatibility factor in California *Drosophila*.** *Nature* 1991, **353**(6343):440–442.
 20. Turelli M, Hoffmann AA, McKechnie SW: **Dynamics of cytoplasmic incompatibility and mtDNA variation in natural *Drosophila simulans* populations.** *Genetics* 1992, **132**(3):713–723.
 21. Belanger DH, Perkins SL: ***Wolbachia* infection and mitochondrial diversity in the canine heartworm (*Dirofilaria immitis*).** *Mitochondrial DNA* 2010, **21**(6):227–233.
 22. Keddie EM, Higazi T, Unnasch TR: **The mitochondrial genome of *Onchocerca volvulus*: sequence, structure and phylogenetic analysis.** *Mol Biochem Parasitol* 1998, **95**(1):111–127.
 23. Hu M, Gasser RB, Abs El-Osta YG, Chilton NB: **Structure and organization of the mitochondrial genome of the canine heartworm, *Dirofilaria immitis*.** *Parasitology* 2003, **127**(Pt 1):37–51.
 24. Ghedin E, Wang S, Spiro D, Caler E, Zhao Q, Crabtree J, Allen JE, Delcher AL, Guiliano DB, Miranda-Saavedra D, Angiuoli SV, Creasy T, Amedeo P, Haas B, El-Sayed NM, Wortman JR, Feldblyum T, Tallon L, Schatz M, Shumway M, Koo H, Salzberg SL, Schobel S, Pertea M, Pop M, White O, Barton GJ, Carlow CK, Crawford MJ, Daub J, Dimmic MW, Estes CF, Foster JM, Ganatra M, Gregory WF, Johnson NM, Jin J, Komuniecki R, Korf I, Kumar S, Laney S, Li BW, Li W, Lindblom TH, Lustigman S, Ma D, Maina CV, Martin DM, McCarter JP, McReynolds L, Mitreva M, Nutman TB, Parkinson J, Peregrin-Alvarez JM, Poole C, Ren Q, Saunders L, Sluder AE, Smith K, Stanke M, Unnasch TR, Ware J, Wei AD, Weil G, Williams DJ, Zhang Y, Williams SA, Fraser-Ligggett C, Slatko B, Blaxter ML, Scott AL: **Draft genome of the filarial nematode parasite *Brugia malayi*.** *Science* 2007, **317**(5845):1756–1760.
 25. Yatawara L, Wickramasinghe S, Rajapakse RP, Agatsuma T: **The complete mitochondrial genome of *Setaria digitata* (Nematoda: Filarioidea): Mitochondrial gene content, arrangement and composition compared with other nematodes.** *Mol Biochem Parasitol* 2010, **173**(1):32–38.
 26. Casiraghi M, Bain O, Guerrero R, Martin C, Pocacqua V, Gardner SL, Franceschi A, Bandi C: **Mapping the presence of *Wolbachia pipientis* on the phylogeny of filarial nematodes: evidence for symbiont loss during evolution.** *Int J Parasitol* 2004, **34**(2):191–203.
 27. McLaren DJ, Worms MJ, Laurence BR, Simpson MG: **Micro-organisms in filarial larvae (Nematoda).** *Trans R Soc Trop Med Hyg* 1975, **69**(5–6):509–514.
 28. McGarry HF, Pfarr K, Egerton G, Hoerauf A, Akue JP, Enyong P, Wanji S, Klager SL, Bianco AE, Beeching NJ, Taylor MJ: **Evidence against *Wolbachia* symbiosis in *Loa loa*.** *Filaria J* 2003, **2**(1):9.
 29. McNulty SN, Foster JM, Mitreva M, Dunning Hotopp JC, Martin J, Fischer K, Wu B, Davis PJ, Kumar S, Brattig NW, Slatko BE, Weil GJ, Fischer PU: **Endosymbiont DNA in endobacteria-free filarial nematodes indicates ancient horizontal genetic transfer.** *PLoS One* 2010, **5**(6):e11029.
 30. Casiraghi M, Anderson TJ, Bandi C, Bazzocchi C, Genchi C: **A phylogenetic analysis of filarial nematodes: comparison with the phylogeny of *Wolbachia* endosymbionts.** *Parasitology* 2001, **122**(Pt 1):93–103.
 31. Anderson RC and Bain O: **Keys to genera of the order Spirurida. Part 3. Diplostriaenoidea, Aprocotoidea and Filarioidea.**, in *CIH keys to the nematode parasites of vertebrates*, Anderson RC, Chabaud AG, and Willmott S, Editors. 1976, Commonwealth Agricultural Bureau: Farnham Royal, UK. p. 59–116.
 32. Wong LJ: **Molecular genetics of mitochondrial disorders.** *Dev Disabil Res Rev* 2010, **16**(2):154–162.
 33. Ash LR, Riley JM: **Development of subperiodic *Brugia malayi* in the jird, *Meriones unguiculatus*, with notes on infections in other rodents.** *J Parasitol* 1970, **56**(5):969–973.
 34. Lucius R, Textor G: ***Acanthocheilonema viteae*: rational design of the life cycle to increase production of parasite material using less experimental animals.** *Appl Parasitol* 1995, **36**(1):22–33.
 35. Fischer P, Kipp W, Bamuhiga J, Binta-Kahwa J, Kiefer A, Buttner DW: **Parasitological and clinical characterization of *Simulium neavei*-transmitted onchocerciasis in western Uganda.** *Trop Med Parasitol* 1993, **44**(4):311–321.
 36. Weil GJ, Kastens W, Susapu M, Laney SJ, Williams SA, King CL, Kazura JW, Bockarie MJ: **The impact of repeated rounds of mass drug administration with diethylcarbamazine plus albendazole on bancroftian filariasis in Papua New Guinea.** *PLoS Negl Trop Dis* 2008, **2**(12):e344.
 37. Laslett D, Canback B: **ARWEN: a program to detect tRNA genes in metazoan mitochondrial nucleotide sequences.** *Bioinformatics* 2008, **24**(2):172–175.
 38. Carver T, Thomson N, Bleasby A, Berriman M, Parkhill J: **DNAPlotter: circular and linear interactive genome visualization.** *Bioinformatics* 2009, **25**(1):119–120.
 39. Tamura K, Dudley J, Nei M, Kumar S: **MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0.** *Mol Biol Evol* 2007, **24**(8):1596–1599.
 40. Kuck P, Meusemann K: **FASconCAT: Convenient handling of data matrices.** *Mol Phylogenet Evol* 2010, **56**(3):1115–1118.
 41. Nylander JAA, *MrModeltest* v2. 2004, Program distributed by the author, Evolutionary Biology Centre, Uppsala University
 42. Huelsenbeck JP, Ronquist F: **MRBAYES: Bayesian inference of phylogenetic trees.** *Bioinformatics* 2001, **17**(8):754–755.
 43. Ronquist F, Huelsenbeck JP: **MrBayes 3: Bayesian phylogenetic inference under mixed models.** *Bioinformatics* 2003, **19**(12):1572–1574.

doi:10.1186/1471-2164-13-145

Cite this article as: McNulty et al.: Comparing the mitochondrial genomes of *Wolbachia*-dependent and independent filarial nematode species. *BMC Genomics* 2012 **13**:145.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

