Complete genome sequence of *Methanocorpusculum labreanum* type strain Z

Iain J. Anderson^{1*}, Magdalena Sieprawska-Lupa², Eugene Goltsman¹, Alla Lapidus¹, Alex Copeland¹, Tijana Glavina Del Rio¹, Hope Tice¹, Eileen Dalin¹, Kerrie Barry¹, Sam Pitluck¹, Loren Hauser^{1,3}, Miriam Land^{1,3}, Susan Lucas¹, Paul Richardson¹, William B. Whitman², and Nikos C. Kyrpides¹

¹Joint Genome Institute, 2800 Mitchell Drive, Walnut Creek, California, USA ²Microbiology Department, University of Georgia, Athens, Georgia, USA ³Oak Ridge National Laboratory, Oak Ridge, Tennessee, USA

*Corresponding author: Iain Anderson

Keywords: archaea, methanogen, Methanomicrobiales

Methanocorpusculum labreanum is a methanogen belonging to the order Methanomicrobiales within the archaeal phylum Euryarchaeota. The type strain Z was isolated from surface sediments of Tar Pit Lake in the La Brea Tar Pits in Los Angeles, California. M. labreanum is of phylogenetic interest because at the time the sequencing project began only one genome had previously been sequenced from the order Methanomicrobiales. We report here the complete genome sequence of M. labreanum type strain Z and its annotation. This is part of a 2006 Joint Genome Institute Community Sequencing Program project to sequence genomes of diverse Archaea.

Introduction

Methanocorpusculum labreanum is a methanogen belonging to the order Methanomicrobiales within the archaeal phylum Euryarchaeota. Strain Z is the type strain of this species. It was isolated from surface sediments of Tar Pit Lake at the La Brea Tar Pits in Los Angeles [1]. Most of the other described members of this family have been isolated from anaerobic digesters or waste water [2]. The genus covers organisms with a wide temperature range. One psychrotolerant strain was isolated from a Russian pond polluted with paper mill waste water [3], while other strains were found in heated sediment at a hydrothermal vent site [4]. Methanocorpusculum species may be common in subsurface environments as they were the most prominent genus found in a coal bed in Indiana [5] and in shale in northern Michigan [6].

Methanogens have been divided into two groups known as Class I and Class II based on phylogeny [7]. Class I includes the orders *Methanococcales*, *Methanobacteriales*, and *Methanopyrales*, which use H_2/CO_2 or formate as substrates for methanogenesis, although some can also use alcohols as electron donors. Class II includes the orders *Methanosarcinales* and *Methanomicrobiales*. Some of

the Methanosarcinales are capable of using various methyl compounds as substrates for methanogenesis including acetate, methylamines, and methanol, but Methanomicrobiales are restricted to the same substrates as the Class I methanogens [2]. Therefore, Methanomicrobiales are phylogenetically closer to Methanosarcinales but physiologically more similar to Class I methanogens, making them an interesting target for genome sequencing. In a 2006 Community Sequencing Program (CSP) project, we proposed sequencing two members of the order Methanomicrobiales: M. labreanum and Methanoculleus marisnigri. Previously only one genome was available from this order, that of Methanospirillum hungatei. Methanocorpusculum labreanum and Methanoculleus marisnigri are phylogenetically distant from each other and from Methanospirillum hungatei (Figure 1), and they represent the three families within the order *Methanomicrobiales*. We report here the sequence and annotation of *M. labreanum* type strain Z.

Classification and features

Methanocorpusculum labreanum Z was isolated from surface sediments at the La Brea Tar Pits [1].

A polypropylene bottle was filled with half surface sediment and half lake water. In an anaerobic chamber the contents of the bottle were mixed to suspend the sediment, and 0.5 ml of the slurry was added to 5 ml enrichment medium. The enrichment medium contained sodium formate. trypticase peptone, and salts. The gas phase was H_2/CO_2 at a ratio of 4:1 and a pressure of 152 kPa. The physiological characteristics of *M. labreanum* were described as follows [1]. The cells were coccoid with a diameter of 0.4-2.0 µm. They were irregular in shape under some growth conditions, such as higher salt or with added acetate. Motility was not observed and flagella were not observed. Growth was observed on H_2/CO_2 or formate, but not with acetate, propionate, methanol, trimethylamine, or ethanol. Growth was observed in a narrow window of pH, from 6.5 to 7.5, with pH 7.0 as the optimal value. Growth was observed between 25 and 40°C, with an optimum at 37°C. M. labreanum can tolerate a wide range of salt concentration, from 0 to 30 g/L NaCl. Acetate was stimulatory at lower salt concentrations. Trypticase peptone, yeast extract, or cysteine was required for growth. The features of *M. labreanum* Z are presented in Table 1.

Genome sequencing information Genome project history

M. labreanum was selected for sequencing based upon its phylogenetic position relative to other methanogens of the order *Methanomicrobiales*. It is part of a 2006 Joint Genome Institute Community Sequencing Program project that included six diverse archaeal genomes. A summary of the project information is shown in Table 2. The complete genome sequence was finished in January, 2007. The GenBank accession number for the project is CP000559. The genome project is listed in the Genomes OnLine Database (GOLD) [20] as project Gc00506. Sequencing was carried out at the Joint Genome Institute (JGI) Production Genomics Facility (PGF). Quality assurance was done by JGI-Stanford. Finishing was done at JGI-PGF. Annotation was done by JGI-Oak Ridge National Laboratory (ORNL) and by JGI-PGF.

Growth conditions and DNA isolation

The methods for DNA isolation, genome sequencing and assembly for this genome have previously been published [21].

Genome annotation

Protein-coding genes were identified using a combination of CRITICA [22] and Glimmer [23] followed by a round of manual curation using the JGI GenePRIMP pipeline [24]. GenePRIMP points out cases where gene start sites may be incorrect based on alignment with homologous proteins. It also highlights genes that appear to be broken into two or more pieces, due to a premature stop codon or frameshift, and genes that are disrupted by transposable elements. All of these types of broken and interrupted genes are labeled as pseudogenes. Genes that may have been missed by the gene calling programs are also identified in intergenic regions. The predicted CDSs were translated

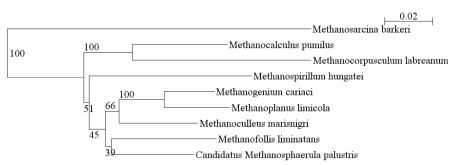


Figure 1. Phylogenetic tree of 16S rRNA of selected *Methanomicrobiales* showing the distance between the three organisms for which complete genomes are available – *Methanospirillum hungatei*, *Methanocorpusculum labreanum*, and *Methanoculleus marisnigri*. The tree uses sequences aligned within the Ribosomal Database Project (RDP), and the tree was constructed with the RDP Tree Builder [8]. *Methanosarcina barkeri* was used as the outgroup. The numbers indicate bootstrap values based on 100 replicates.

MIGS ID	Property	Term	Evidence	
	- ·		Code	
		Domain Archaea	TAS [10-12]	
		Phylum <i>Euryarchaeota</i>	TAS [13,14]	
		Class "Methanomicrobia"	TAS [15]	
	Current classification	Order Methanomicrobiales	TAS [16]	
		Family Methanocorpusculaceae	TAS [17,18]	
		Genus Methanocorpusculum	TAS [19]	
		Species Methanocorpusculum labreanum	TAS [1]	
	Gram stain	negative	TAS [1]	
	Cell shape	irregular coccus	TAS [1]	
	Motility	nonmotile	TAS [1]	
	Sporulation	nonsporulating		
	Temperature range	25-40°C	TAS [1]	
	Optimum temperature	37°C	TAS [1]	
MIGS-6.3	Salinity	0-30 g/L NaCl	TAS [1]	
MIGS-22	Oxygen requirement	anaerobe	TAS [1]	
	Carbon source	CO_2 , acetate	TAS [1]	
	Energy source	H_2/CO_2 , formate	TAS [1]	
MIGS-6	Habitat	sediment	TAS [1]	
MIGS-15	Biotic relationship	free-living	TAS [1]	
MIGS-14	Pathogenicity	none		
	Biosafety level	1		
	Isolation	sediment	TAS [1]	
MIGS-4	Geographic location	Tar Pit Lake, La Brea Tar Pits	TAS [1]	
MIGS-5	Isolation time	1989	TAS [1]	
MIGS-4.1	Latitude-longitude	34.107811/-118.599658		
MIGS-4.2	C C			
MIGS-4.3	Depth	0-5 cm	TAS [1]	
MIGS-4.4	Altitude	not applicable		

Table 1. Classification and general features of *M. labreanum* Z according to the MIGS recommendations [9]

Evidence codes - IDA: Inferred from Direct Assay (first time in publication); TAS: Traceable Author Statement (i.e., a direct report exists in the literature); NAS: Non-traceable Author Statement (i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence). These evidence codes are from the Gene Ontology project [19]. If the evidence code is IDA, then the property was directly observed for a living isolate by one of the authors or another expert mentioned in the acknowledgements.

Table 2. Genome sequencing project information

MIGS ID	Characteristic	Details	
MIGS-28	Libraries used	3kb, 6kb and 40kb (fosmid)	
MIGS-29	Sequencing platform	ABI3730, 454	
MIGS-31.2	Sequencing coverage	34×	
MIGS-31	Finishing quality	Finished	
	Sequencing quality	Less than one error per	
		50kb	
MIGS-30	Assembler	Newbler, Paracel	
MIGS-32	Gene calling method	CRITICA, Glimmer	
	GenBank ID	CP000559	
	GenBank date of release	February 2, 2007	
	GOLD ID	Gc00506	
	NCBI project ID	18109	
	IMG Taxon ID	640069317	
MIGS-13	Source material identifier	DSM 4855	
	Project relevance	Tree of Life	

and used to search the National Center for Biotechnology Information (NCBI) nonredundant database, UniProt, TIGRFam, Pfam, PRIAM, KEGG, COG, and InterPro databases. Signal peptides were identified with SignalP [25], and transmembrane helices were determined with TMHMM [26]. CRISPR elements were identified with the CRISPR Recognition Tool (CRT) [27]. Paralogs are hits of a protein against another protein within the same genome with an e-value of 10⁻² or lower. The tRNAScanSE tool [28] was used to find tRNA genes. Additional gene prediction analysis and manual functional annotation was performed within the Integrated Microbial Genomes Expert Review (IMG-ER) platform [29].

Genome properties

The genome of *M. labreanum* Z consists of a single circular chromosome (Figure 2). The genome size of 1.80 Mbp is similar to those of Class I methanogens, but smaller than the genomes of *Methanosarcina* species and the other *Methanomicrobiales*, which range between 2.5 and 5.8 Mbp. The G+C percentage is 50.0%, higher than that of most other sequenced methanogens. There are 1,830 genes, of which 1,765 are protein-coding genes and the remaining 65 are RNA genes. There were only 26 pseudogenes identified, constituting 1.4% of the total genes. The properties and statistics of the genome are summarized in Table 3, and genes belonging to COG functional categories are listed in Table 4.

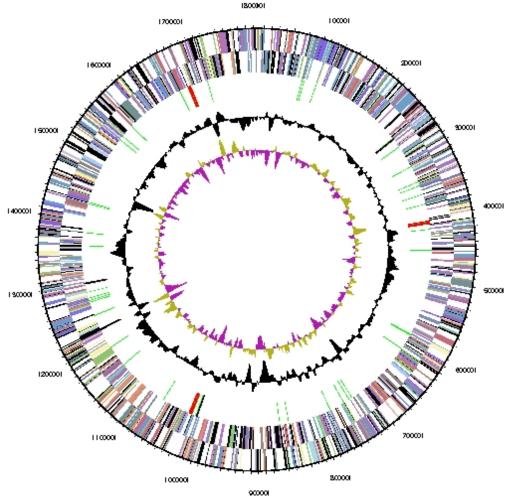


Figure 2. Graphical circular map of the chromosome of *Methanocorpusculum labreanum* Z. From outside to the center: Genes on forward strand (colored by COG categories), Genes on reverse strand (colored by COG categories), RNA genes (tRNAs green, rRNAs red, other RNAs black), GC content, GC skew.

Table 3. Genome statistics Attribute	Value	% of total
Genome size (bp)	1,804,962	100.00%
DNA coding region (bp)	1,600,673	88.68%
DNA G+C content (bp)	902,600	50.01%
Number of replicons	1	
Extrachromosomal elements	0	
Total genes	1830	100.00%
RNA genes	65	3.55%
rRNA operons	3	
Protein-coding genes	1765	96.45%
Pseudogenes	26	1.42%
Genes in paralog clusters	745	42.21%
Genes assigned to COGs	1358	76.94%
Genes assigned Pfam domains	1335	75.64%
Genes with signal peptides	406	23.00%
Genes with transmembrane helices	368	20.85%
CRISPR repeats	2	

Table 4. Numbers of genes associated with the general COG functional categories.

Code	value	% of total	COG category
E	130	7.4	Amino acid transport and metabolism
G	54	3.1	Carbohydrate transport and metabolism
D	10	0.6	Cell cycle control, cell division, chromosome partitioning
Ν	5	0.3	Cell motility
Μ	35	2.0	Cell wall/membrane/envelope biogenesis
В	2	0.1	Chromatin structure and dynamics
Н	129	7.3	Coenzyme transport and metabolism
Z	0	0.0	Cytoskeleton
V	13	0.7	Defense mechanisms
С	134	7.6	Energy production and conversion
W	0	0.0	Extracellular structures
S	172	9.7	Function unknown
R	219	12.4	General function prediction only
Р	95	5.4	Inorganic ion transport and metabolism
U	17	1.0	Intracellular trafficking, secretion, and vesicular transport
I	24	1.4	Lipid transport and metabolism
Y	0	0.0	Nuclear structure
F	49	2.8	Nucleotide transport and metabolism
Ο	57	3.2	Posttranslational modification, protein turnover, chaperones
А	0	0.0	RNA processing and modification
L	65	3.7	Replication, recombination and repair
Q	8	0.5	Secondary metabolites biosynthesis, transport and catabolism
Т	30	1.7	Signal transduction mechanisms
К	77	4.4	Transcription
J	147	8.3	Translation, ribosomal structure and biogenesis
-	293	16.6	Not in COGs

Insights from the genome sequence

The genome sequence of *M. labreanum* Z shows some similarities to Class I methanogens and some to Methanosarcinales but also has some unique features. In common with Class I methanogens, M. labreanum uses a partial reductive TCA cycle to synthesize 2-oxoglutarate, and it has the Eha membrane-bound hydrogenase. Similar to Methanosarcinales. M. labreanum has the Ech membrane-bound hydrogenase. A unique feature of M. labreanum and the other Methanomicrobiales is the presence of anti- and anti-anti-sigma factors, which is surprising as Archaea do not use sigma factors. Phylogenetic analysis of methanogenesis and cofactor biosynthesis enzymes suggest that Methanomicrobiales form a group distinct from other methanogens, and therefore methanogens can be split in to three classes [21]. Surprisingly *M*. labreanum lacks the F₄₂₀-nonreducing hydrogenase, which has been proposed to couple Coenzyme M-Coenzyme B heterodisulfide reduction

References

- 1. Zhao Y, Boone DR, Mah RA, Boone JE, Xun L. Isolation and characterization of *Methanocorpusculum labreanum* sp. nov. from the LaBrea Tar Pits. *Int J Syst Bacteriol* 1989; **39**:10-13.
- Garcia JL, Ollivier B, Whitman WB. The order Methanomicrobiales. Prokaryotes 2006; 3:208-230. doi:10.1007/0-387-30743-5_10
- 3. Simankova MV, Kotsyurbenko OR, Lueders T, Nozhevnikova AN, Wagner B, Conrad R, Friedrich MW. Isolation and characterization of new strains of methanogens from cold terrestrial habitats. *Syst Appl Microbiol* 2003; **26**:312-318. <u>PubMed doi:10.1078/072320203322346173</u>
- Dhillon A, Lever M, Lloyd KG, Albert DB, Sogin ML, Teske A. Methanogen diversity evidenced by molecular characterization of methyl coenzyme M reductase A (*mcrA*) genes in hydrothermal sediments of the Guaymas Basin. *Appl Environ Microbiol* 2005; **71**:4592-4601. <u>PubMed</u> <u>doi:10.1128/AEM.71.8.4592-4601.2005</u>
- Strąpoć D, Picardal FW, Turich C, Schaperdoth I, Macalady JL, Lipp JS, Lin YS, Ertefai TF, Schubotz F, Hinrichs KU, et al. Methane-producing microbial community in a coal bed of the Illinois basin. Appl Environ Microbiol 2008; 74:2424-2432. PubMed doi:10.1128/AEM.02341-07
- 6. Waldron PJ, Petsch ST, Martini AM, Nüsslein K. Salinity constraints on subsurface archaeal diver-

and ferredoxin reduction for the first step of methanogenesis in the cytoplasm of *Methanomicrobiales* [30]. In place of this hydrogenase, *M. labreanum* may use the membrane-bound hydrogenase Mbh or energy-converting hydrogenase Ech to couple heterodisulfide reduction to a transmembrane ion gradient [21].

Acknowledgments

This work was performed under the auspices of the US Department of Energy's Office of Science, Biological and Environmental Research Program, and by the University of California, Lawrence Berkeley National Laboratory under Contract No. DE-AC02-05CH11231, Lawrence Livermore National Laboratory under Contract No. DE-AC02-06NA25396. L. H. and M. L. were supported by the Department of Energy under contract DE-AC05-000R22725. M. S.-L., and W. B. W. were supported by DOE contract number DE-FG02-97ER20269.

sity and methanogenesis in sedimentary rock rich in organic matter. *Appl Environ Microbiol* 2007; **73**:4171-4179. <u>PubMed</u> <u>doi:10.1128/AEM.02810-06</u>

- Bapteste É, Brochier C, Boucher Y. Higher-level classification of the *Archaea*: evolution of methanogenesis and methanogens. *Archaea* 2005; 1:353-363. <u>PubMed doi:10.1155/2005/859728</u>
- Cole JR, Wang Q, Cardenas E, Fish J, Chai B, Farris RJ, Kulam-Syed-Mohideen AS, McGarrell DM, Marsh T, Garrity GM, et al. The ribosomal database project: improved alignments and new tools for rRNA analysis. *Nucleic Acids Res* 2009; 37:D141-D145. <u>PubMed doi:10.1093/nar/gkn879</u>
- Field D, Garrity G, Gray T, Morrison N, Selengut J, Sterk P, Tatusova T, Thomson N, Allen MJ, Angiuoli SV, et al. The minimum information about a genome sequence (MIGS) specification. Nat Biotechnol 2008; 26:541-547. <u>PubMed</u> doi:10.1038/nbt1360
- 10. Woese CR, Kandler O, Wheelis ML. Towards a natural system of organisms: proposal for the domains *Archaea, Bacteria,* and *Eucarya. Proc Natl Acad Sci USA* 1990; **87**:4576-4579. PubMed doi:10.1073/pnas.87.12.4576
- 11. Balch WE, Fox GE, Magrum LJ, Woese CR, Wolfe RS. Methanogens: reevaluation of a unique bio-

logical group. *Microbiol Rev* 1979; **43**: 260-296. <u>PubMed</u>

- 12. List Editor. Validation List no. 6. Validation of the publication of new names and new combinations previously effectively published outside the IJSB. *Int J Syst Bacteriol* 1981; **31**: 215-218.
- 13. Garrity GM, Holt JG. Phylum All. *Euryarchaeota* phy. nov. *In* Bergey's Manual of Systematic Bacteriology, vol. 1. 2nd ed. Edited by: Garrity, GM, Boone, DR and Castenholz, RW. Springer, New York; **2001**:211-355.
- List Editor. Validation of publication of new names and new combinations previously effectively published outside the IJSEM. Validation List no. 85. *Int J Syst Evol Microbiol* 2002; **52**: 685-690. <u>PubMed doi:10.1099/ijs.0.02358-0</u>
- Garrity GM, Bell JA, Lilburn T. The revised road map to the manual. *In:* Brenner, DJ, Kreig, NR, Staley, JT Eds. 2009. *Bergey's Manual of Systematic Bacteriology*, 2nd Ed. Vol 2 The *Proteobacteria* Part A Introductory Essays. 2005 pp 159-220
- Balch WE, Fox GE, Magrum LJ, Woese CR, Wolfe RS. Methanogens: reevaluation of a unique biological group. *Microbiol Rev* 1979; 43:260-296. <u>PubMed</u>
- 17. Editor L. Validation List no 30. Validation of the publication of new names and new combinations previously effectively published outside the IJSB. *Int J Syst Bacteriol* 1989; **39**: 371.
- Zellner G, Stackebrandt E, Messner P, Tindall BJ, Conway de Macario E, Kneifel H, Sleytr UB, Winter J. Methanocorpusculaceae fam. nov., represented by Methanocorpusculum parvum, Methanocorpusculum sinense spec. nov. and Methanocorpusculum bavaricum spec. nov. Arch Microbiol 1989; 151: 381-390. PubMed doi:10.1007/BF00416595
- 19. Xun L, Boone DR, Mah RA. Deoxyribonucleic acid hybridization study of *Methanogenium* and *Methanocorpusculum* species, emendation of the genus *Methanocorpusculum*, and transfer of *Methanogenium aggregans* to the genus *Methanocorpusculum aggregans* comb. nov. *Int J Syst Bacteriol* 1989; **39**:109-111.
- 20. Liolios K, Mavromatis K, Tavernarakis N, Kyrpides NC. The Genomes OnLine Database (GOLD) in 2007: status of genomic and metagenomic projects and their associated metadata. *Nucleic*

Acids Res 2008; **36**:D475-D479. PubMed doi:10.1093/nar/gkm884

- Anderson I, Ulrich LE, Lupa B, Susanti D, Porat I, Hooper SD, Lykidis A, Sieprawska-Lupa M,0020Dharmarajan L, Goltsman E, et al. Genomic characterization of *Methanomicrobiales* reveals three classes of methanogens. *PLoS ONE* 2009; 4:e5797. <u>PubMed</u> doi:10.1371/journal.pone.0005797
- 22. Badger JH, Olsen GJ. CRITICA: coding region identification tool invoking comparative analysis. *Mol Biol Evol* 1999; **16**:512-524. <u>PubMed</u>
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 1999;
 27:4636-4641. <u>PubMed</u> doi:10.1093/nar/27.23.4636
- 24. Pati A. GenePRIMP: A Gene Prediction Improvement Pipeline for microbial genomes. (Submitted).
- 25. Emanuelsson O, Brunak S, von Heijne G, Nielsen H. Locating proteins in the cell using TargetP, SignalP and related tools. *Nat Protoc* 2007;
 2:953-971. <u>PubMed doi:10.1038/nprot.2007.131</u>
- Krogh A, Larsson B, von Heijne G, Sonnhammer EL. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 2001; **305**:567-580. <u>PubMed doi:10.1006/jmbi.2000.4315</u>
- 27. Bland C, Ramsey TL, Sabree F, Lowe M, Brown K, Kyrpides NC, Hugenholtz P. CRISPR recognition tool (CRT): a tool for automatic detection of clustered regularly interspaced palindromic repeats. *BMC Bioinformatics* 2007; **8**:209. <u>PubMed</u> <u>doi:10.1186/1471-2105-8-209</u>
- Lowe TM, Eddy SR. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 1997; 25:955-964. <u>PubMed doi:10.1093/nar/25.5.955</u>
- 29. Markowitz VM, Mavromatis K, Ivanova NN, Chen IMA, Chu K, Kyrpides NC. IMG ER: a system for microbial genome annotation expert review and curation. *Bioinformatics* 2009; **25**:2271-2278 <u>PubMed doi:10.1093/bioinformatics/btp393</u>
- Thauer RK, Kaster AK, Seedorf H, Buckel W, Hedderich R. Methanogenic archaea: ecologically relevant differences in energy conservation. *Nat Rev Microbiol* 2008; 6:579-591. <u>PubMed</u> <u>doi:10.1038/nrmicro1931</u>