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Comprehensive Survey of Chemical Libraries for Drug Discovery and Chemical Biology: 2009

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This is the thirteenth installment of the comprehensive survey series in high throughput chemistry.[1] Biologically active libraries reported in 2009 are captured in Tables 1–5 under the headings of proteases, nonproteolytic enzymes, GPCRs, nonGPCRs, and oncolytics/antiinfectives. Table 6 lists molecular probes. Compound collections without disclosed biological activity are delineated in Tables 7–10 under the headings of scaffold derivatization/acyclic synthesis, monocyclic-, bicyclic/spirocyclic-, and polycyclic/macrocyclic synthesis. Polymer-supported reagents/scavengers/linkers are presented in Tables 11 (nonfluorous) and Table 12 (fluorous). There are 370 libraries and 24 molecular probes extracted from 355 literature citations.[2–491] Approximately 90% of the citations originated from academic laboratories, with the majority of these from European and Asian laboratories. Solution-phase methodology accounted for *ca*. 85% of chemical library synthesis.

This year's 18 vignettes include (a) biologically active libraries: HDAC1/HDAC2 inhibitors,[134] H_3 antagonists,[136] glucagon receptor antagonists,[179] purinergic $P2Y_{12}$ receptor antagonists,[221] heat shock protein 90 (Hsp90) inhibitors,[41] selective norepinephrine reuptake inhibitors,[73] allosteric modulators of mGluR4[287] and mGluR5[71] Bcl-2 inhibitors via DOS library;[182] (b) molecular probes: Ned-19[340] and DG-041;[338] (c) high throughput chemical methodology: catch and release synthesis of substituted guanidines.[289] and substituted pyrimidines via a 3-component reaction;[158] and (d) fluorous technology: displaceable fluorous dihydropyran[322] and isonnitrile linkers,[327] fluorous synthesis of 1,4-benzodiazepine-2,5-dione, [327] piperazinedion-fulsed tricyclic[328] compound libraries and a fluorous mixture synthesis of natural product resorcyclic acid lactone library.[329]

Related publications and reviews appeared in 2009 on microwave-assisted of *N*-heterocyclic synthesis[356] and convertible isonitriles,[365] click chemistry,[358] dynamic combinatorial

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chemistry,[363] large scale preparation of silicon-functionalized SynPhase lanterns,[370] kinase inhibitors,[357] heat shock protein 90 inhibitors,[376] biologically active benzoannelated nitrogen heterocycles,[361] library automation and analysis,[360,368] library design,[375] NMR-ased screening,[377,378] fragement libraries and fragement hopping,[366,367,369] review of Ellman's libraries,[374] solid-phase resin specifications,[359] microelectrode arrays for monitoring ligand-receptor binding events,[364] yactoliter-scale DNA reactor for small molecule evolution,[362] DNA encoded libraries,[379] and fluorous chemistry and separations.[380–383]

Selective HDAC1/HDAC2 Inhibitors.[134]

Histone deacetylases (HDACs) are enzymes involved in the remodeling of chromatin. Several classes of HDAC inhibitors have been found to have potent and specific anticancer activities and the common pharmacophore identified contains a surface recognition domain, a linker region and a metal binding region. Kattar and co-workers set out to improve upon the selectivity and tolerability of Zolinza **1**, Merck's first-in-class mixed HDAC 1, 2, 3 and 6 inhibitor used for the treatment of cutaneous manifestation of T-cell lymphoma (Figure 1).[134] In their initial library, keeping the metal binding domain constant and varying both linker and surface recognition domains, scaffold **A** had already been identified. Reacting 4-formylmethyl-benzoate and polystyrene bound sodium triacetoxyborohydride with 48 R^1-amines, followed by hydroxamate formation with NH_2OH for their first follow-up library, 3-chlorobenzylamine was found to be preferred in the R^1-position over phenethyl or aniline analogs, yielding compound **2** (HDAC IC_{50} = 348 nM). Modifying the metal binding domain next, 2-aminophenols were chosen to replace the hydroxamate moiety. R^1-substituted 2-nitrophenols **3** were reacted with bromo-Wang resin, cesium carbonate and DMF under microwave irradiation at 40 °C. The nitro group was reduced with $SnCl_2$ in DMF and the resulting resin-bound anilines **4** coupled with the linker unit 4-chloromethyl benzoyl chloride **5**, DMAP and DIEA in DCM. Attachment of R^2-amines was accomplished using sodium iodide and a proton sponge in DMF. Final compounds **6** were released from resin with TFA/DCM. Aromatic substituents in R^1 and anilines in R^2 demonstrated 10–30-fold increased potency for HDAC1 (e.g. **7**, IC_{50} = 10 nM, cellular assay = 1,735 nM). To improve cellular potency the aminophenol metal binding site was replaced with a primary amine. Mono protected R^1-substituted di-anilines were bound to aldehyde resin **8** via reductive amination utilizing sodium triacetoxyborohydride in AcOH/DCE. 4-Chloromethyl benzoyl chloride was attached next, followed by reaction with surface binding domain R^2-amines, sodium iodide and a proton sponge. Exposure to TFA/DCM yielded final products **9**. Amino spirocyclic building blocks in the R^2-position retained affinity towards HDAC1 and improved selectivity over HDAC3 as well as improved potency in the cellular assay. Unfortunately this series exhibited activity against hERG (**10**: HDAC1 IC_{50} = 4 nM, cellular assay = 82 nM, HDAC3 IC_{50} = 9,617 nM, hERG IC_{50} = 1,597 nM). Eliminating one amino-functionality by switching to a terephthalamide linkage successfully mitigated hERG liabilities. This last set of compounds was synthesized via solution-phase, starting from **11** and mono protected R^1-substituted di-anilines which were combined with PS-CDI and HOBt in DMF. After ester saponification, the spirocyclic amine was coupled using PS-CDI again. The final 26 compounds **12** were obtained after deprotection with TFA/DCM.

Compound **13** showed the best overall profile (HDAC1: IC_{50} = 8 nM, cellular assay = 103 nM, HDAC3: IC_{50} = 5,102 nM, hERG IC_{50} = 12,390 nM) and demonstrated *in vivo* efficacy in an acute PD HCT-116 xenograph model study. This successful combination of solid and solution phase library synthesis led to the efficient optimization of a novel HDAC inhibitor series, continuously improving additional properties within each library iteration.

H_3 Antagonists.[136]

The histamine H_3 receptor is an attractive G protein–coupled receptor drug target that regulates neurotransmission in the central nervous system. There has been considerable effort by both academic and industrial laboratories to develop potent and selective H_3 receptor antagonists for the potential treatment of attention-deficit hyperactivity disorder, dementias, schizophrenia, as well as obesity and sleep disorders resulting in a refined H_3 antagonist pharmacophore model containing two basic nitrogens separated by a spacer and a central core that also carries a polar group and a lipophilic residue (Figure 2). After having completed its total synthesis, Kennedy and co-workers recognized that the marine alkaloid dispyrin **14** perfectly maps onto this pharmacophore model.[136] Dispyrin **14** indeed displayed some H_3 antagonist activity (IC_{50} = 2.35 μM) and was used as the starting point for a natural product guided iterative parallel synthesis campaign. Starting from 3-bromo-4-methoxyphenylethylamine **15** five heterocyclic carboxylic acids R^1 were coupled using DIC, HOBt and DIEA in DCM. The methyl ether was removed with BBr_3 and the resulting phenols alkylated with five R^2-aminoalkyl chlorides under microwave irradiation conditions generating 25 different compounds **16**. All the compounds showed activity against H_3 with more potent compounds containing an ethyl pyrrolidinyl residue in the R^2 position (K_i's < 200 nM), with the best compound **17** (H_3 K_i = 80 nM, H_3 IC_{50} = 180 nM). Retaining the 4-bromo-thiophene residue from **17**, functionalized pyrrolidines were explored next. Phenol **18** was reacted first with 2-bromo-1,1-dimethoxy ethane and then with tosylic acid to yield aldehyde **19**. Final compounds **20** were obtained via reductive amination utilizing resin-bound triacetoxyborohydride together with the functionalized pyrrolidines. No potency improvement was observed in final compounds **20**.

The last library reexamined the R^1 residue, including additional heterocycles and aromatic moieties. Protecting bromomethoxy phenethyl amine **21** as the phthalimide by reacting it with 1,2-dicarboxybenzene, DIC, HOBt and DIEA, followed by BBr_3 demethylation yielded phenol **22**. Alkylation with chloroethyl pyrrolidine and deprotection with hydrazine, both under microwave conditions, followed by coupling with a diverse set of R^1-acids with DIC, HOBt yielded final compounds **23**. Five membered heterocycles in the R^1-position showed superior activities compared to pyridine and substituted benzenes, with 5-oxazole (**24**) and 2-thiazole (**25**) displaying the best potencies, K_is = 32 nM and IC_{50} = 72–83 nM, respectively. This iterative parallel library campaign successfully optimized H_3 antagonism properties of a natural product lead structure over 30-fold.

Human Glucagon Receptor Antagonists.[179]

The glucagon receptor is a member of the class B G-protein coupled family of receptors. Glucagon maintains glucose homeostasis during the fasting state by promoting hepatic

gluconeogenesis and glycogenolysis. Antagonizing the glucagon receptor is expected to result in reduced hepatic glucose overproduction, leading to overall glycemic control and a possible treatment for type 2 diabetes. Only a few classes of non-peptidic glucagon receptor antagonists are known (Figure 3). Madsen and co-workers had previously described β-alanine and isoserine urea based human glucagon receptor (hGluR) antagonists **26** and **27**, whereby **27** showed improved selectivity over the related human glucose-dependent insulinotropic receptor (hGIPR). Here the authors explored the replacement of the urea linkage with a heterocyclic scaffold, thus rigidifying the molecule.[179] A library assisted optimization strategy was pursued, combining solid and solution phase synthesis approaches, with the goal of finding a novel potent, orally available, hGluR selective antagonists. For the first library on solid support, 4-formylbenzoic acid was coupled to deprotected Fmoc-β-Ala-Wang resin with HOBt/DIC, followed by reductive amination with R^1-amines and treated with Fmoc-NCS to afford resin-bound Fmoc protected thioureas. Thiazole formation was accomplished through reaction with α-bromoketones after Fmoc-deprotection and final compounds **28** were obtained after cleavage from resin with TFA. Over 800 analogs were prepared via this procedure. Compounds were also prepared via solution-phase starting from 4-formylbenzoic acid methyl ester **29**, which was reductively aminated with R^1-amines utilizing sodium cyanoborohydride. The benzylic amines **30** were converted to their corresponding thioureas employing different methods depending on the reactivity of the respective secondary amine. The aminothiazole **31** was obtained via reaction with α-bromoketones in acetic acid. The methyl ester was cleaved with NaOH and the resulting carboxylic acid **32** was coupled with either β-alanine or (*R*)-isoserine methyl ester, followed by hydrolysis to yield the final compounds **33** and **34**, respectively. Compounds **35** with the aminothiazole directly attached to the aromatic ring were prepared in similar fashion both on solid support and solution-phase. 4-Nitrobenzoyl chloride was reacted with resin-bound β-alanine, the nitro group was reduced with $SnCl_2$, followed by reductive alkylation with R^1-aldehydes. Thiourea **36** was obtained after reaction with Fmoc-isothiocyanate. After deprotection, cyclization with α-bromoketones and cleavage from resin final compounds **35** were obtained in high yields and purities. Using a solution-phase approach, compounds from this type were obtained by reductive amination of 4-aminomethyl benzoate **37** with R^1-aldehydes, followed by reactions with EtO_2C-NCS, NaOH, α-bromoketones and subsequent EDAC, HOBt coupling with β-alanine methyl ester and hydrolysis to give **38**. Finally, compounds **39** containing a 5-alkylthiazole core were prepared starting with a Knoevenagel condensation of 4-methyl benzoate **40** with phenyl acetonitriles, followed by reduction with $NaBH_4$ in THF and treatment with dithiophosphoric acid *O*,*O*-diethylester. The resulting saturated thioamide **41** was reacted with α-bromoketones and coupled to β-alanine methyl ester followed by hydrolysis. Aliphatic R^1 groups were not well tolerated. Compounds with 4-CF_3-, 4-OCF_3- and 4-SCF_3-phenyl in the R^1 position displayed good binding affinities and superior rat PK properties. Compound **42** (hGluR IC_{50} = 93 nM, hGIPR IC_{50} = 1,100 nM) showed high oral bioavailability (58%), low clearance (1mL/min)/kg, long plasma $T_{1/2}$ (228 min after i.v. administration), and extremely high plasma exposure (C_{max} = 2100 ng/mL) in the rat (3 mg/kg, p.o.). Changing the β-alanine to isoserine in this series did lead to a loss of mGluR activity. Compounds with modified core structures retained binding affinities but displayed less favorable PK profiles. The SAR for R^2 and R^3 residues is of minor importance as long

as the substituents on R^2 are lipophilic and in *meta* or *para* position of the phenyl ring. Compound **42** was tested in a non-human primate model of hyperglucagonaemia and hyperglycaemia. It dose-dependently decreased glucagon stimulated glycaemia and abolished the hyperglycemic effect of exogenously administered glucagon completely at i.v. doses of 1 and 3 mg/kg. Compound **42** also showed high plasma exposure and a long plasma half-life in monkeys. This library approach demonstrated that the urea linkage in previously reported hGluR antagonists can be successfully replaced by a number of different thiazole cores and established SARs for binding, selectivity and PK properties for this novel chemical class.

Purinergic $P2Y_{12}$ receptor antagonists.[221]

Plavix® (clopidogrel, **43**) is an antiplatelet agent approved for stroke and myocardial infarction in patients with atherosclerosis. Its mechanism of action is thought to proceed via metabolism to acid **44** followed by irreversible inactivation of $P2Y_{12}$, a platelet specific GPCR (Figure 4). This in turn leads to a reduction in adenosine diphosphate (ADP)-stimulated platelet aggregation and the formation of platelet aggregates thereby providing therapeutic benefit. Because **43** is a prodrug requiring metabolic activation, it cannot be used effectively in patients that require emergency treatment (delayed time of action) or in patients who cannot metabolize **43**. Recognizing the shortcomings of **43**, Parlow and colleagues sought to find a new class of $P2Y_{12}$ receptor antagonists having direct action on the receptor.[221] $P2Y_{12}$-based quinoline antagonists were previously reported by Berlex and this class was used as a starting point for library design and SAR exploration. Initial efforts focused on replacing the quinoline moiety employing polymer-assisted solution-phase chemistry. Piperazine derivatives **46** were coupled with heteroaromatic carboxylic acids **47** in the presence of resin-bound carbodiimide and HOBt. Excess reactants were sequestered with resin-bound isocyanate and a resin-bound secondary amine providing clean intermediates that, post TFA treatment, afforded desired carboxylic-acid-containing analogs **48**. Hundreds of compounds were prepared using this approach. The library was evaluated in a human-derived $P2Y_{12}$ receptor binding assay, screening for % inhibition at 10 μM. K_i values and inhibitory effects in a human platelet rich plasma (PRP) assay were obtained for the more potent compounds. The results from the first round of library evaluation indicated that replacing the original quinoline group with pyridine generally produced compounds with nanomolar K_i values, although with poor PRP inhibitory effects. Since the most potent compounds had one or two phenyl groups on the pyridine ring (**49** and **50**, respectively), the authors selected **49** as the lead for further SAR exploration and optimization. The next step consisted of SAR exploration around the piperazine moiety **51**. A broad range of alkylating, acylating and sulfonating agents were used in conjunction with a scavenger resin to remove residual electrophile reactants. Biological assays showed that activity was driven by the carbamate group where the longer the aliphatic carbamate chain, the greater receptor affinity binding and, importantly, inhibitory PRP activity (**53-57**: K_is = 2440 to 11 nM; IC_{50}s = >100 to 15 μM).

The next SAR campaign focused on the 4-position of the pyridine ring. The authors opted to make chloro- and phenol-containing intermediates (**58-60**) to use these functional groups as handles to introduce chemical diversity. Phenol **58** was *O*-alkylated with alcohols under

Mitsunobu reaction conditions (DEAD, PPh_3, THF) or with halides (Cs_2CO_3, KI, DMF) ultimately yielding carboxylic acid ethers **61-66**. Although the SAR was rather flat, a trend emerged for increased PRP activity with the phenolic ethers (**65**, **66:** IC_{50} = 3.8 and 1.9 μM, respectively).

In addition to 4-ether substitution, 4-amino substitution was explored. This was carried out by treating carbamate **55** with 20 equiv of amine to displace the 4-chloro group yielding 4-aminopyridine ethyl carbamates **67-76**. Simultaneous variation of the 4-position and the carbamate moiety was achieved using the same 4-chloro displacement strategy with **60** followed by alloc deprotection, *N*-piperidine carbamoylation and TFA deprotection. Small secondary alkyl amines were preferred over tertiary amines (IC_{50}s for **67** and **68** versus **69**) while amino ethers exhibited higher PRP inhibitory activity (**70, 71**); particularly effective were the amino piperidyl and amino piperazine groups (**72-75**). Lastly, increasing the length of the alkyl carbamate chain led to further increases in K_i and IC_{50} (**76:** K_i = 7 nM; IC_{50} = 0.77 μM). Evaluation of the most potent candidates in this series led to the discovery of **77** with a satisfactory *in vivo* profile, oral bioavailability, and some 340-fold more selective for the $P2Y_{12}$ receptor versus its closest homolog, the $P2Y_{13}$ receptor.

Heat shock protein 90 (Hsp90) Inhibitors.[41]

Hsp90 has gained attention in the pharmaceutical industry due to its participation in multiple cell signaling pathways in cancer cells (e.g., PI3K/Akt). Hsp90 is an ATPase protein that acts as a chaperone, binding to multiple oncology relevant client proteins (e.g., Her2, cKit, MET), stabilizing these proteins so to permit their cell signaling function. Inhibition of Hsp90 prevents the necessary folding required to bind and stabilize client proteins. This results in the degradation of the client proteins via the ubiquitin proteasome pathway making Hsp90 an attractive oncology target. Geldanamycin and close synthetic analog 17-allylamino-17-demethoxygeldanamycin (17-AAG) bind to the ATP active site in the *N*-terminal domain inhibiting Hsp90 function validating this to be a viable mode of action to disrupt Hsp90's biological function. Poor water solubility and hepatotoxicity limit the clinical utility of these agents and thus, new small molecules are sought targeting the ATP binding site to inhibit Hsp90 function. Radicicol is an inhibitor of Hsp90 via ATP competition binding. Promising results have been noted by several research programs with resorcinol-based Hsp90 inhibitors. However, a common limitation to this compound class is that the phenol groups can be substrate sites of metabolic degradation processes (e.g., glucuronidation) resulting in fast *in vivo* clearance.

Cho-Schultz and colleagues focused their attention on compound **78**, an amide-containing polyphenol and ATP binding site-directed inhibitor of Hsp90 (Figure 5).[41] A two-stage solution-phase strategy was devised to explore the chemical space and develop an SAR around **78** to produce new potent Hsp90 inhibitors without phenol groups. The first library of compounds focused on the solution-phase synthesis of **79**. The objective was to improve binding affinity and reduce or eliminate glucuronidation by increasing hydrophilicity via the introduction of basic amino fragments. Library synthesis commenced by the addition of Grignard intermediate **81**, derived from *m*- and *p*-1,3-dioxane-subsituted bromobenzene to *N*-vinyl pyrrolidinone **82** affording 2-pyrrolines **83** (23–29% yields). These cyclic imines

were reduced to **84** with $NaBH_4$ and then coupled to phenol-protected benzoic acid **85**. Deprotection of the phenol groups with either TFA or HCl resulted in aldehydes **86** that were subjected to reductive amination with *ca*. 90 amines to afford the 178-compound library **79**. Compounds were tested in a tritium-labeled-ligand competitive binding assay (K_i) followed by the measurement of Akt-degradation in H1299 lung cancer cells (IC_{50}). The SAR indicated that *para* substituted compounds had higher binding affinity against Hsp90 and demonstrated superior activity in the cell-based assay (K_is = 5.4–40 nM; IC_{50}s = 112–668 nM) versus the *meta* substituted compounds (K_is = 60–100 nM; IC_{50}s = 2100–4800 nM). A cocrystal structure of Hsp90α with **91a** (PDB code: 3HEK, resolution: 1.95 Å) established the (*R*) stereochemistry of the pyrrolidine group as crucial for the molecule to adopt the most favorable, lower-energy conformation to bind at the ATP site. In this configuration, the resorcinol group exhibits a salient H-bond interaction with Asp93, the amide group retains its expected planarity, the 3,3-difluroropyrrolidine group is placed into a hydrophobic region (Tyr137, Val136, Gly135), and the pendant phenyl group is also accommodated in a hydrophobic pocket. Chiral separation of other racemates confirmed the (*R*)**-**enantiomers bound far more potently against Hsp90 than their corresponding (*S*)-enantiomers. Unfortunately, all of the (*R*)-enantiomers exhibited high clearance in human hepatocytes. Armed with this knowledge, a second library was conceived focusing on the synthesis of (*R*)-enantiomers and the replacement of the phenol groups to improve the ADME profile. The chemistry relied on the enantioselective α-arylation of *N*-Boc-pyrrolidine **92** with (−)-sparteine to prepare chiral **93**. Chiral organometallic **93** was coupled with aromatic bromides to afford key intermediate **94**. Following Boc deprotection, amines **95** were coupled to a diverse group of aromatic carboxylic acids **96** using HATU to afford library **80**. With this methodology enantiomeric ratios of >96:4 were achieved and allowed for the synthesis of a *ca*.112-membered library. Unfortunately, none of the compounds of library **80** exhibited inhibitory activity against Hsp90 when tested at 10 μM in the binding assay. Lastly, selected synthesis of mono-phenol analogs revealed that both phenol groups are essential for binding since the removal of either phenol group led to a >190-fold loss in potency.

Selective Norepinephrine Reuptake Inhibitors.[73]

The norepinephrine transporter is a membrane bound protein which regulates the uptake of the neurotransmitter norepinephrine (NE) from the presynaptic cleft of noradrenergic neurons during synaptic transmission. It therefore plays an important role in regulating the physiological functions of NE, the deficiency of which has been implicated in a number of neurological disorders. Norepinephrine reuptake inhibitors (NRIs) such as reboxetine and atomoxetine have been used clinically for the treatment of major depressive disorder and attention deficit hyperactivity disorder (ADHD), respectively. Researchers at Pfizer disclosed recently several new chemical series of selective norepinephrine reuptake inhibitors (sNRI).[73] The two lead compounds **97** and **98**, investigated clinically, were discontinued due to hepatotoxicity (Figure 6). This safety issue identified with **97** and **98** was thought to be related to the high lipophilicity of these compounds (clogP = 4.2–4.4). A search for new NRI templates, providing ligands with reduced lipophilicity when compared to **97** or **98**, was then undertaken. The benzamide derivative **99** was identified, from

previous lead identification work, as a weak norepinephrine reuptake inhibitor with low selectivity over the serotonin transporter (K_i (NET) = 294 nM; K_i (SERT) = 654 nM). The lead optimization objective was to improve the affinity and the selectivity for the norepinephrine transporter, while simultaneously reducing the lipophilicity in this series. Analogs of **99** (205-member library; general structure **100**) were readily available *via* solution phase parallel synthesis methodology. The *N*-alkyl or *N*-aryl substituted 3-amino-1-Boc-(*S*)-pyrrolidines **102**, obtained from 3-amino-1-Boc-(*S*)-pyrrolidine using common methodologies, were converted to the target compounds **100** via amide formation followed by *N*-Boc deprotection. The binding affinity of the library compounds at the human norepinephrine, serotonin, and dopamine transporters was determined using scintillation proximity assay (SPA) technology. SAR analysis in this series revealed that potent norepinephrine reuptake inhibition could be achieved over a range of lipophilicity (clogP: 3.0–4.4). The lead optimization campaign led to the discovery of compound **101**, a potent norepinephrine reuptake inhibitor (K_i = 6 nM) displaying 37-fold and 380-fold selectivity over the serotonin and dopamine transporters, respectively. The lipophilicity of **101** (clogP = 3.3) was also significantly reduced when compared to previous lead compounds **97** and **98** investigated clinically (clogP = 4.4 and 4.2, respectively). Further studies demonstrated that **101** has excellent metabolic stability in human liver microsomes and human hepatocytes, weak CYP (1A2, 2C9, 2C19, 2D6, 3A4) inhibition, and good membrane permeability. In light of its weak inhibitory activities at the hERG (IC_{50} >20,000 nM) and $NaV_{1.5}$ (IC_{50} > 26,000 nM) ion channels, **101** is expected to have a satisfactory cardiovascular safety profile. Off-target profiling of **101** at a panel of 110 receptors, enzymes and ion channels revealed only weak affinity at the M_4 (K_i = 4,300 nM) and M_5 (K_i = 1,800 nM) muscarinic receptors. In vivo rat microdialysis experiments demonstrated that **101** produces a rapid increase in NE levels in the prefrontal cortex after subcutaneous administration demonstrating good CNS penetration. Based on its favorable profile, **101** (PF-3409409) was selected for preclinical evaluation.

Positive Allosteric Modulators of mGluR4.[287]

Metabotropic glutamate receptors (mGluRs) play important roles in a broad range of central nervous system functions and have therapeutic potential in a variety of neurological and psychiatric disorders. Activation of metabotropic glutamate receptor 4 (mGluR4) has been shown to modulate neurotransmission in the basal ganglia and results in antiparkinsonian effects in rodent models of Parkinson's Disease (PD). Allosteric binding sites, as opposed to traditional orthosteric binding sites, offer unparalleled opportunities for drug discovery by providing high levels of selectivity, mimicking physiological conditions and affording fewer side effects. VU0155041 **103**, a novel mGluR4 positive allosteric modulator (PAM) discovered in a high throughput screen was the starting point of a detailed SAR analysis of this compound class (Figure 7). Three parts of the lead molecule were explored separately through an iterative parallel synthesis approach, the aromatic amide moiety, the carboxylic acid residue and the cyclohexyl core. The first library centered around commercially available *cis*-1,2-cyclohexanedicarboxylic anhydride **104**, which was reacted with respective R^1-amines in THF at 55 °C to give library compounds **105**. Several 3,5- and 3,4-substituted phenyl, unsubstituted phenyl, benzyl and pyridyl, morpholino and cyclo alkyl analogs were

evaluated. Besides the original 3,5-dichlorophenyl lead **103** (EC_{50} hmGluR4 = 0.74 μM), only the 3-Cl-5-F-phenyl amide **109** retained some activity (EC_{50} hmGluR4 = 2 μM), all other substitutions led to a loss of activity. Carboxylic acid replacements were investigated next, starting from VU0155041 **103**, coupling respective R^2,R^3-amines with EDC, HOBt and DIEA in DMF a library of diamides **106** was synthesized. All compounds showed at least a tenfold loss of activity, with the exception of the primary carboxamide **112**, which retained similar submicromolar activity as the lead (EC_{50} hmGluR4 = 0.95 μM). Variations of the core structure were then examined. Commercially available cyclic anhydrides **107** were reacted with 3,5-dichloroaniline in THF at 55 °C. Only the cyclohexene analog **115** retained some activity (EC_{50} hmGluR4 = 2.7 μM), substituted cyclohexene cores (e.g. **117**) resulted in inactive compounds. Changing the substitution pattern to a 1,3-orientation led to a total loss of activity. Combining the cyclohexene core with a primary carboxamide in compound **116** showed some activity (EC_{50} hmGluR4 = 3.1 μM).

This SAR evaluation around VU0155041 **103**, utilizing a parallel synthesis approach, illustrates another example of the rather flat SAR common to positive allosteric modulators.

mGluR5 allosteric modulators.[71]

Excess dopamine transmission in the brain is believed to be one cause of schizophrenia and antipsychotic agents that antagonize the dopamine D2 receptor are routinely prescribed. Many of these agents display off target pharmacology against a range of neurotransmitters leading to side effects. It was observed clinically that administering glycine, an N-methyl-D-aspartate (NMDA) receptor co-agonist, elicited a modest improvement in schizophrenic patients suggesting that activation of the NMDA receptor could be an option for therapeutic treatment. Potentiation of NMDA receptor transmission can occur directly by modulating NMDA receptor sites or indirectly by activating NMDA-receptor-function dependent GPCRs. Metabotropic glutamate receptor 5, mGluR5, is a GPCR that upon activation can potentiate indirectly the NMDA receptor. Using ADX-47273 (**118**), a previously reported positive allosteric modulator of mGluR5 demonstrating in vivo activity in cognition and schizophrenia animal models, Engers and colleagues generated solution-phase libraries around scaffold **119** to flesh out the SAR of **118** and improve its overall physicochemical properties (Figure 8).[71] Two routes were developed for library synthesis. The first route involved simultaneous coupling-cyclization of (*Z*)-*N*′-hydroxyimidamides **120** and (*S*)-1-(*tert*-butoxycarbonyl)piperidine-3-carboxylic acid **121** using EDCI and HOBt under reflux to give oxadiazoles. Removal of the Boc group and acylation of the piperidine intermediates afforded library **119a**. The second route consisted of the esterification of (*S*)-piperidine-3-carboxylic acid **122**, acylating ester **123**, saponifying *N*-acyl esters **124** with LiOH and then simultaneous coupling-cyclization with (*Z*)-*N*′-hydroxyimidamides **120** to afford desired library **119b**.

In the first exploratory library, the 4-fluorobenzylpiperamide was kept constant and the 4-fluorophenyl ring on the oxadiazole unit was replaced. It was found that 2-pyridyl and 2-thienyl groups were reasonable substitutes for the 4-fluorophenyl group while the 3- and 4-pyridyl and 2-pyrazinyl groups were not. In the follow-up library **119b**, which retained the original 4-fluorophenyl and now included the 2-thienyl and 2-pyridyl (R^1) groups, the acyl

group (R^2) was varied. Biological evaluation of this library revealed the following trends. For R^2, most mono- and di-fluorophenyl groups exhibited submicromolar ago-potentiation (potentiation of the agonist activity of glutamate at its EC_{90}) while the 2,6-difluorophenyl group did not. The 2-pyridyl analogs e.g. **125**, were 2-times less potent than their 4-fluorophenyl and 2-thienyl counterparts. Unexpectedly pyridyl compounds, e.g. **128**, displayed pure mGluR5 positive allosteric modulation as these compounds had no inherent agonist activity. The corresponding HCl salts of the pyridyl analogs exhibited expected improved water solubility. Interestingly, the cyclobutyl analog **129** was a negative allosteric potentiator, an example of an unusual functional switch. Finally, in order to establish the importance of the chiral center, enantiomeric separations were carried out on **118**, **125** and **126**. The (*R*)-enantiomers were in general 9- to 10-fold less potent than the (*S*)-enantiomers, yet display similar in vitro efficacy. With this study the SAR features of **118** were quickly identified, particularly the relevance of the (*S*)-chiral center.

Bcl-2 inhibitors: Diversity-Oriented Synthesis Library.[182]

Apoptosis, or programmed cell death, is important for normal development, host defense, and suppression of oncogenesis, and disregulation of apoptosis has been implicated in cancer and many other human diseases. The Bcl-2 family proteins are central regulators of apoptosis and comprise anti-apoptotic proteins such as Bcl-2, Bcl-xL, and Mcl-1 and pro-apoptotic proteins such as Bak, Bax, Bim, Bid, and Bad. Overexpression of the Bcl-2 membrane protein has been observed in 70% of breast cancer, 30–60% of prostate cancer, 80% of B-cell lymphomas, 90% of colorectal adenocarcinomas, and many other forms of cancer. The expression levels of Bcl-2 proteins also correlate with resistance to a wide spectrum of chemotherapeutic drugs and γ-radiation therapy. Bcl-2 is therefore a promising molecular target for the design of an entirely new class of anticancer drugs aimed at overcoming resistance of cancer cells to apoptosis. Consequently, design of non-peptide small molecule inhibitors of Bcl-2 and Bcl-xL is currently an exciting research area for the development of new anticancer agents. In an effort to identify novel inhibitors of Bcl-2, scientists at Infinity Pharmaceuticals designed and prepared a 15,000-member pyridone library (Figure 9a/b), *via* discovery oriented synthesis (DOS).[182] The design of this library was based on the structure of the nicotinic agonist (−)-cytisine (**130**) and previous work describing the total synthesis of this natural product. In particular, the pyridone cyclization step, key transformation for the total synthesis of (−)-cytisine, was exploited for the diversity oriented synthesis of heterocycles **131a** and **132a** as well as their corresponding enantiomers **131b** and **132b**, respectively. The synthons **140** and **141** used for the diversity oriented library synthesis were prepared according to Figure 9a. Condensation of D-glyceraldehyde acetonide **133** with phosphonate **142**, under Horner-Emmons conditions, provided the α,β-unsaturated ester **134**, which reacted with the imine **135** in toluene in the presence of AgOAc and DBU to provide the corresponding [3+2] azomethine ylide-alkene cycloaddition product **136** in high yield and excellent diastereoselectivity (>95:5). The pyrrolidine derivative **136** was then converted to the bridge bicyclic scaffold **140** in 5 steps, i.e. a) protection of the NH functionality of **136** as the *N*-Fmoc derivative, b) deprotection of the allyl ester functionality by treatment with $Pd(PPh_3)_4$, c) conversion of the resulting carboxylic acid to the corresponding primary alcohol, d) activation of the resulting alcohol

with mesyl chloride followed by formation of the pyridone ring *via* intramolecular cyclization, and e) conversion of the 1,3-dioxolane, 2,2-dimethyl functionality to the corresponding primary alcohol according to a 3-step sequence. The tricyclic synthon **141** was prepared from phosphonate **143** *via* a synthetic sequence similar to the one described for the preparation of **140**. The pyridone scaffolds **140**, **141**, prepared in large scale (>75g), were then loaded onto silicon-functionalized Lanterns *via* activation with TfOH (average loading level of 15 mmol of compound per Lantern). Deprotection of the *N*-Fmoc functionality provided the resins **144** and **148** which were converted to the library compounds **145-147** and **149-151** *via* classical solid-phase derivatization methodologies (Figure 9b). The 15,000 library compounds, prepared with purities greater than 75% for over 75% of the library, were screened for binding affinity for Bcl-2 and Bcl-xL. The most potent compounds derived from the bridged bicyclic pyridone scaffolds **131a** and **131b**, exemplified by compound **152** [K_i (Bcl-2) = 2.0 μM; K_i (Bcl-xL) = 5.7 μM)] and its enantiomeric analog **153** [K_i (Bcl-2) = 1.3 μM; K_i (Bcl-xL) = 6.6 μM)], contained a chloro-substituted diphenyl 2-aminothiazole and a diamine at the R and R′ position, respectively. Compound **154** was the best ligand identified from the tricyclic pyridone cores **132a** and **132b**. This compound selectivitly binds with micromolar affinity at Bcl-2 (K_i = 1.2 μM) and displays, in contrast to **152**,**153**, high (> 100-fold) selectivity for Bcl-2 over Bcl-xL (Figure 9b).

Catch and Release Synthesis of Substituted Guanidines.[289]

One of the most common methods to prepare substituted guanidines **156** involves the condensation of amines with *S*-methylated thioureas **156** (Figure 10). This method suffers from the formation of noxious and toxic methyl mercaptan side product, which limits application for high-throughput parallel synthesis. To overcome this problem, scientists at Abbott designed a solid phase strategy for the synthesis of *N*,*N*′,*N*″-substituted guanidines using a catch and release methodology inspired from the solution phase synthesis described above.[289] In this method, the thioureas **155** immobilized on solid support react with various amines to provide the desired guanidine derivatives, free of mercaptan side products. Using this new methodology, and after several optimization of the reaction conditions, various substituted guanidines were prepared from thioamides in a one-pot process. Hence, loading of *N*-substituted thioamides **157** to brominated polystyrene resin in a mixture DCM/DMF (2:1) at 50°C for 4–6 h provided the thiourea resins **155**, which could be washed, dried, and stored or used without isolation for the next step. Condensation of the resin-bound thioureas with primary amines at 50 °C for 20 h provided, in high yield, the desired substituted guanidines **156a,b** purified by HPLC and isolated as their TFA salts. The reaction conditions were modified to allow the formation of *N*,*N*′,*N*″-trisubstituted guanidines from the corresponding secondary amines in good yields. In the modified method, the condensation of secondary amines to the resin-bound thioureas was conducted in methoxyethanol in the presence of 1.5 equiv. of $HgCl_2$. Using this new solid phase methodology, a total of 17 *N*,*N*′-disustituted or *N*,*N*′,*N*″-trisubstituted guanidines **156a,b** were prepared, from the corresponding thioamides, in isolated yields ranging from 42% to 99%.

Substituted Pyrimidines via a 3-component reaction.[158]

Konakahara and coworkers developed a novel three-component coupling reaction of enamines **158**, triethyl orthoformate and ammonium acetate providing a facile route to 4,5-disubstituted pyrimidine derivatives **160** (Figure 11). Pyrimidine derivatives exhibit biological activity against all major classes of molecular targets including kinases, proteases, nuclear hormone receptors and GPCRs. Bredereck- and Pinner-type chemistries are established routes for the preparation of pyrimidines. However, challenging multistep synthesis of reactive intermediates and harsh reaction conditions limit their general utility, particularly for the preparation of pyrimidine-based arrays. The initial 3-CR was conducted by simply heating enamines **158** with 3 equiv of orthoester and 2 equiv of NH_4OAc in toluene at 100 °C for 20 h. Optimized reaction conditions included the addition of 0.1 equiv of $ZnCl_2$ which improved the overall yield (>70%), accelerated reaction time and enhanced the range of substrates. Non-commercially available enamines were prepared in high yield (>90%) by the addition of electron-withdrawing stabilized carbon nucleophiles to arylnitriles. The $ZnCl_2$-catalyzed three-component reaction was expanded by using ketones **159** in place of enamines. The 3-CR with ketone substrates required 72 h for completion affording the desired products in good yields (54–70%).

Discovery of Ned-19.[340]

Second messengers play enormously important roles in transduction of cellular signals and govern cellular responses to stimuli. Molecular and cellular biology has benefited greatly from molecular tools like Forskolin which alters cellular levels of the critical second messenger cAMP. Nicotinic acid adenine dinucleotide phosphate (NAADP, **161**) is recognized as a critical regulator of Ca^{2+} release in numerous human tissues in response to several reported stimuli (Figure 12). Controversies surrounding the exact nature of NAADP as a secondary messenger and its mechanism of action permeate the literature. Furthermore, the lack of chemical probes of this important biomolecule hamper studies into the cause and consequences associated with Ca^{2+} release. To rectify this inadequacy, Churchill and coworkers reported the discovery of Ned-19 (**162**), a potent inhibitor of NAADP signaling.[340] Utilizing the chemical structure (including electrostatic surface assessments) of NAADP, a ligand-based virtual screen was performed in which novel structures were sought with sufficient overlap with NAADP while maintaining drug-like properties in the 'hit' compounds. One agent that was discovered was an (*S*)-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indole-3-carboxylate designated as Ned-19. This commercially available agent inhibited NAADP signaling in a sea urchin egg assay at nanomolar concentrations. The stereochemistry at the 1 position of the tetrahydro-1H-pyrido ring was explored via an independent synthesis. The synthesis was initiated by installment of a chloromethyl group at the 3-position of commercially available 4-methoxybenzaldehyde. The resulting aldehyde was subjected into a Pictet-Spengler reaction with (*S*)-methyl 2-amino-3-(1*H*-indol-3-yl)propanoate to afford the core (*S*)-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-b]indole-3-carboxylate scaffold. Displacement of the benzyl chloride with 1-(2-fluorophenyl)piperazine and saponification of the methyl ester provided Ned-19. The diastereomeric mixture was separated following the Pictet-Spengler reaction. The specific activity of both the *cis* and *trans* isomers of Ned-19 were evaluated and the *trans* isomer was significantly more active

than the *cis* isomer. Biochemically, Ned-19 was able to eliminate all NAADP-mediated Ca^{2+} signaling at 100 μM without altering inositol 1,4,5-trisphosphate-mediated or cADP-ribose-mediated Ca^{2+} release. These experiments suggest that Ned-19 is selective as a NAADP signaling probe rather than a nonspecific modulator of Ca^{2+} release. Ned-19 blocks NAADP signaling in intact cells and it can serve as a fluorescent label for NAADP receptors. Finally, Ned-19 was used to demonstrate a role for NAADP signaling in glucose signaling and Ca^{2+} release in pancreatic beta cells. While further studies with Ned-19 are required to fully validate its use as a probe of NAADP signaling, this work offers a novel agent to begin to fully dissect the various roles of this important secondary messenger.

Discovery of DG-041.[338]

There exists a complex network of signaling events that govern platelet activation and aggregation and great strides have been made in modulating these relevant signaling events to avoid catastrophic acute thrombosis during vascular events like myocardial infarction and stroke. The recognition that aspirin (acetylsalicylic acid) acts as an antagonist of platelet aggregation can be considered a starting point for research into defining the critical targets associated with platelet function. More recently, the purinergic $P2Y_{12}$ receptor has been shown to play an essential role in platelet signaling following platelet collagen and von Willebrand factor release and Ca^{2+} stimulation of TxA_2 and release of ADP. $P2Y_{12}$ receptor antagonists including prasugrel have proven more effective than aspirin in preventing recurrent myocardial infarction. Both aspirin and $P2Y_{12}$ receptor antagonists, however, have the unwanted side-effect of severe, prolonged bleeding events due to their global impairment of platelet aggregation. Research has continued to shed light on the complex signaling cascade associated with platelet aggregation and today it is known that multiple GPCRs are responsible for both Ca^{2+} release and activation/inactivation of adenylcyclase. One target that is gaining attention is the EP_3 receptor. The EP_3 receptor responds to prostaglandin E_2 (PGE_2) which is produced in low concentrations within atherosclerotic plaque. Importantly, signaling through the EP_3 receptor is not sufficient to produce an aggregation response without co-signaling events at one or more additional receptors that are associated with platelet activation. Studies have showed that platelets lacking the EP_3 receptor were protected from thrombotic events in several models and it has been hypothesized that, since healthy tissues do not produce PGE_2, blocking of the EP_3 receptor should have a diminished effect on bleeding events. To examine this theory, researchers at deCODE chemistry sought out new EP_3 receptor antagonists through a ligand-based design strategy (Figure 13).[338] The basic strategy was to seek out scaffolds that mimic the three-dimensional shape and electrostatic profile of the native ligand PGE_2. Ultimately efforts focused on a collection of substituted indoles with appendages at the 1 and 7 positions. Their initial leads possessed low-nanomolar activity at the EP_3 receptor and lead optimization included blocking metabolically labile positions at C3 and C5 of the indole ring. The optimization campaign led to the discovery of DG-041 (**163**). DG-041 (**163**) displayed IC_{50} values of 8.1 nM and 4.6 nM in a Ca^{2+} response assay and a radioligand displacement assay, respectively. DG-041 had no relevant activity versus the functionally related, platelet-associated GPCR targets nor was this agent active across a panel of 50 random GPCRs. DG-041 was also found to be acceptable for both oral and i.v. dosing with appropriate PK properties for use as

a tool compound in models of platelet aggregation. The synthesis of this agent was accomplished via sequential intramolecular and intermolecular Heck couplings to arrive at an appropriately 7-substituted indole intermediate. Saponification followed by coupling to 4,5-dichlorothiophene-2-sulfonamide and subsequent *N*-alkylation with 2,4-dichloro-1-(chloromethyl)benzene provided DG-041. DG-041 was found to inhibit human and rat platelet aggregation in vitro in a collagen induction experiment with several co-agonists and, importantly, in the presence of high serum concentrations. Within *in vivo* systems, DG-041 inhibited platelet aggregation at a dose of 10 mg/kg while there was no prolongation of bleeding up to a concentration of 100 mg/kg. The authors present several other findings that further validate DG-041 as an important new tool for studying the EP_3 receptor and reference, as yet, unpublished data suggesting that DG-041 is efficacious in a human patient population.

Displaceable fluorous dihydropyran linker.[322]

Fluorous linker combined with fluorous solid-phase extraction (F-SPE) is a powerful reaction and purification technique for solution-phase synthesis.[380] The Nelson group introduced fluorous dihydropyran linkers for the protection of amino and hydroxyl groups. Figure 14 highlights the utility of displaceable linker **164** in the synthesis of highly condensed heterocyclic compound **165**.[322] The fluorous linker was attached to sulfonamide **166** through Fukuyama-Mitsunobu reaction to form compound **167**. Cascade metathesis using Hoveyda-Grubbs second generation catalyst cleaved the central dihydropyran ring and produced compound **168**. This compound was then used for Diels-Alder reaction with 4-phenyl-[*1,2,4*]-triazole-3,5-dione **169** to afford **170** as a single diastereomer. The removal of the *o*-nitrophenylsulfonyl (Ns) group from **170** followed by the reaction with an isocyanate gave urea **171**. Finally, the fluorous linker was removed by the treatment of 3% TFA to afford product **165**. In this multistep synthesis, all the intermediates and the final product were isolated from the reaction mixture by F-SPE.

Displaceable fluorous isonitrile linker.[327]

Isonitriles are versatile reagents for organic reactions. However, their synthetic advantages are offset by the notorious odor associated with conventional isonitriles. The Pirrung group developed a new process using base-promoted ring-opening of oxazoles to produce aromatic isonitriles which no longer have the unpleasant odors.[228] During the reaction process a fluorous sulfonate group could be introduced to form a fluorous linker. The utility of the displaceable fluorous isonitrile has been demonstrated in the multistep synthesis involving Ugi four-component reaction (U-4CR) and metal catalyst-promoted linker cleavage reactions. Freshly prepared isonitrile **172** was used as the limiting agent for the Ugi reaction to form **173** (Figure 15). This intermediate was then subject to different coupling reactions to remove the fluorous linker and also to introduce a new functional group to the Ugi condensation product. The developed post-Ugi reactions included Suzuki coupling to form product **174**, Sonogashira reaction to form compound **175**, and Stille coupling to form compound **176**. The protocol is suitable for diversity-oriented synthesis of compound libraries since four substitution groups can be introduced during the Ugi reactions and different linker cleavage reactions could lead to an array of scaffolds.

Fluorous synthesis of 1,4-benzodiazepine-2,5-dione library.[327]

In the solution-phase parallel synthesis of a 1,4-benzodiazepine-2,5-dione library, the Yang group developed a new U-4CR using fluorous benzaldehydes instead of fluorous isonitriles (Figure 16) as the displaceable linker.[327] The library synthesis was accomplished in three reaction steps. At the first step of the Ugi reactions, fluorous benzaldehydes **177** were used as the limiting agent to react with Boc-anthranilic acids **178,** amino esters **179**, and cyclohexyl isonitrile **180**. After F-SPE purification, Ugi products **181** were used for the second step of acetyl chloride-promoted de-Boc/cyclizations to form 1,4-benzodiazepine-2,5-diones **182**. The cyclization reactions were selective and only attacked the ester group to form the seven-membered ring. The last step Suzuki reactions were carried out under microwave heating to cleave the fluorous sulfonyl group and introduce the biaryl functional group to form products **183**. The Yang group also used 2-nitrobenzoic acids as anthranilic acid alternatives for the U-4CRs. A demonstration library of 36 analogs was produced with building blocks R^1 to R^4 variants.

Fluorous synthesis of piperazinedione-fused tricyclic compound library.[328]

Fluorous amino esters are useful linkers for library synthesis. In a joint work conducted by the Werner and the Zhang groups, fluorous amino esters **184** were used for solution-phase parallel synthesis of novel piperazinedione-fused tricyclic library compounds **189** (Figure 17).[328] The compounds are structurally similar to the tricyclic thrombin inhibitors and diketopiperazine-based inhibitors of human hormone-sensitive lipase. At the first library synthesis step, azomethine ylides generated from **184** and aldehydes **185** underwent microwave-assisted 1,3-dipolar cycloadditions with maleimides to form **186** in stereoselective fashion. The adducts were treated with chloroacetyl chloride to afford *N*-acylated products **187**. The chloro group of **187** was displaced with amines to afford **188**. The last step was the microwave-promoted cyclizations to cleave the fluorous linker and generate the piperazinedione ring. The final products were purified by HPLC to ensure >95% purities. Library **189** containing 90 compounds was synthesized with assorted building blocks for R^1 to R^4.

Fluorous mixture synthesis of natural product resorcylic acid lactone (RAL) library.[329]

Natural resorcylic acid lactones (RAL) containing a *cis*-enone moiety such as radicicol A, L-783277, and LL-Z1640-2 are known to be potent and irreversible kinase inhibitors. For QSAR studies, the Winssinger group developed a fluorous mixture synthesis (FMS) method based on the previously reported fluorous total synthesis of radicicol A and synthesized a compound library containing 51 analogs (Figure 18).[329] In the FMS, three propargyl alcohols were used each attached to a different *p*-methoxybenzyl (PMB) linker by reacting with F-PMB-trichloroacetimidates **190** to form **191(a–c)** (Scheme 5). The equimolar mixture of **191(a–c)** was split to three portions and each reacted with one of three aldehydes **192(a–c)** to afford 3 mixtures of **193(a–c)**. After sequential hydroxyl group protection, TBDPS deprotection, and iodination, compounds **193(a–c)** were converted to 3 mixtures of **194(a–c)** each containing three components. Through a similar pathway, compound **191d** reacted with

aldehydes **192(a–c)** to afford compounds **194(d–f).** The second stage of FMS involved the reaction of three compounds of **195(a–c)** with each of 6 compounds of **194(a–f)** to afford compounds **196**. Among the 18 pools of **196**, 9 are three-component mixtures generated from **194(a–c)** and 9 are single compounds generated from **194(d–f).** At this point, those 9 three-component mixtures were demixed on HPLC with a fluorous column to afford 27 individual compounds of **196**. These 27 compound and another 9 compounds of **196** generated from **194(e–f)** were treated with DDQ and TBAF to remove both the fluorous linker and the TMSE group. The macrocyclization reactions were promoted by Mitsunobu reaction conditions using fluorous PPh_3 and DIAD. The fluorous derivatives were easily removed from the reaction mixture by F-SPE. The treatment of NaOH removed the Bz group to afford total of 36 macrocyclic compounds **197**. Among them, 24 (X = CH or CH_2) were treated with DMP followed by HF to afford products **199.** Another 12 of **197** were treated with PS-SO_3H and PS-IBX to afford products **200.** Compounds **199** and **200** can be converted to **201-205** through simple transformations to afford a total of 51 macrocyclic compounds. A subset of 28 representative library compounds was assayed against a panel of 19 kinases representing those bearing the adequately positioned cysteine residue, bearing a cysteine residue at a different position within the ATP binding pocket, and those that do not bear a cysteine residue. The screening results indicated that there is very little difference in relative selectivity of kinase inhibition throughout the library compounds. VEGF-R2 is the most highly inhibited kinase followed by PDGFR-α, VEGR-R3, Flt3, VEGF-R1, MEK1 SESE and KIT. Two representative compounds **199a** and **200a** were evaluated in a larger panel of 402 kinases and also evaluated against a series of mutations of Fli3 and KIT. The screening results provided valuable QSAR information and also led to the identification of several potent inhibitors of multiple oncogenic kinases.

References

1. Dolle RE, Le Bourdonnec B, Goodman AJ, Morales GA, Thomas CJ, Zhang W. J Comb Chem. 2009; 11:739–790. [PubMed: 19715292]
2. Abadi AH, Ibrahim TM, Abouzid KM, Lehmann J, Tinsley HN, Gary BD, Piazza GA. Bioorg Med Chem. 2009; 17:5974–5982. [PubMed: 19628397]
3. Adib M, Kesheh MR, Ansari S, Bijanzadeh HR. Synlett. 2009:1575–1578.
4. Aguado L, Camarasa M-J, Perez-Perez M-J. J Comb Chem. 2009; 11:210–212. [PubMed: 19178290]
5. Airiau E, Girard N, Mann A, Salvadori J, Taddei M. Org Lett. 2009; 11:5314–5317. [PubMed: 19856928]
6. Algi F, Balci M. Synthesis. 2009:1341–1347.
7. Ali Esmaeili A, Vesalipoor H. Synthesis. 2009:1635–1638.
8. Alizadeh A, Zohreh N. Synlett. 2009:2146–2148.
9. Allan M, Manku S, Therrien E, Nguyen N, Styhler S, Robert M-F, Goulet A-C, Petschner AJ, Rahil G, Robert MacLeod A, Deziel R, Besterman JM, Nguyen H, Wahhab A. Bioorg Med Chem Lett. 2009; 19:1218–1223. [PubMed: 19131248]
10. Ankati H, Biehl E. Tetrahedron Lett. 2009; 50:4677–4682.
11. Annis DA, Cheng CC, Chuang C-C, McCarter JD, Nash HM, Nazef N, Rowe T, Kurzeja RJM, Shipps GW Jr. Comb Chem High Throughput Screening. 2009; 12:760–771.
12. Antonysamy S, Hirst G, Park F, Sprengeler P, Stappenbeck F, Steensma R, Wilson M, Wong M. Bioorg Med Chem Lett. 2009; 19:279–282. [PubMed: 19019674]
13. Awuah E, Capretta A. Org Lett. 2009; 11:3210–3213. [PubMed: 19580257]

14. Ayesa S, Lindquist C, Agback T, Benkestock K, Classon B, Henderson I, Hewitt E, Jansson K, Kallin A, Sheppard D, Samuelsson B. Bioorg Med Chem. 2009; 17:1307–1324. [PubMed: 19124252]
15. Baghbanzadeh M, Molnar M, Damm M, Reidlinger C, Dabiri M, Kappe CO. J Comb Chem. 2009; 11:676–684. [PubMed: 19432481]
16. Bagley MC, Lin Z, Phillips DJ, Graham AE. Tetrahedron Lett. 2009; 50:6823–6825.
17. Barluenga J, Mendoza A, Rodriguez F, Fananas FJ. Angew Chem, Int Ed. 2009; 48:1644–1647.
18. Barton NP, Bellenie BR, Doran AT, Emmons AJ, Heer JP, Salvagno CM. Bioorg Med Chem Lett. 2009; 19:528–532. [PubMed: 19081251]
19. Baskovc J, Bevk D, Stanovnik B, Svete J. J Comb Chem. 2009; 11:500–507. [PubMed: 19397313]
20. Benbow JW, Andrews KA, Aubrecht J, Beebe D, Boyer D, Doran S, Homiski M, Hui Y, McPherson K, Parker JC, Treadway J, VanVolkenberg M, Zembrowski WJ. Bioorg Med Chem Lett. 2009; 19:2220–2223. [PubMed: 19285862]
21. Beswick PJ, Blackaby AP, Bountra C, Brown T, Browning K, Campbell IB, Corfield J, Gleave RJ, Guntrip SB, Hall RM, Hindley S, Lambeth PF, Lucas F, Mathews N, Naylor A, Player H, Price HS, Sidebottom PJ, Taylor NL, Webb G, Wiseman J. Bioorg Med Chem Lett. 2009; 19:4509–4514. [PubMed: 19523822]
22. Blanco-Ania D, Hermkens PHH, Sliedregt LAJM, Scheeren HW, Rutjes FPJT. J Comb Chem. 2009; 11:527–538. [PubMed: 19472985]
23. Blanco-Ania D, Hermkens PHH, Sliedregt LAJM, Scheeren HW, Rutjes FPJT. J Comb Chem. 2009; 11:539–546. [PubMed: 19472986]
24. Borisov AV, Detistov OS, Pukhovaya VI, Zhuravel IO, Kovalenko SM. J Comb Chem. 2009; 11:1023–1029. [PubMed: 19711964]
25. Boyd VA, Mason J, Hanumesh P, Price J, Russell CJ, Webb TR. J Comb Chem. 2009; 11:1100–1104. [PubMed: 19754047]
26. Brucoli F, Howard PW, Thurston DE. J Comb Chem. 2009; 11:576–586. [PubMed: 19530662]
27. Brummond KM, Mao S, Shinde SN, Johnston PJ, Day BW. J Comb Chem. 2009; 11:486–494. [PubMed: 19366169]
28. Campbell J, Blackwell HE. J Comb Chem. 2009; 11:1094–1099. [PubMed: 19831384]
29. Cao J, Gao H, Bemis G, Salituro F, Ledeboer M, Harrington E, Wilke S, Taslimi P, Pazhanisamy S, Xie X, Jacobs M, Green J. Bioorg Med Chem Lett. 2009; 19:2891–2895. [PubMed: 19361991]
30. Cathcart GR, Gilmore BF, Greer B, Harriott P, Walker B. Bioorg Med Chem Lett. 2009; 19:6230–6232. [PubMed: 19773163]
31. Che J, Raghavendra MS, Lam Y. J Comb Chem. 2009; 11:378–384. [PubMed: 19334684]
32. Chen C-H, Chien M-H, Kuo M-L, Chou C-T, Lai J-J, Lin S-F, Thummanagoti S, Sun C-M. J Comb Chem. 2009; 11:1038–1046. [PubMed: 19852442]
33. Chen H, Bai J, Jiao L, Guo Z, Yin Q, Li X. Bioorg Med Chem. 2009; 17:3980–3986. [PubMed: 19411176]
34. Cheng CC, Shipps GW, Yang Z, Sun B, Kawahata N, Soucy KA, Soriano A, Orth P, Xiao L, Mann P, Black T. Bioorg Med Chem Lett. 2009; 19:6507–6514. [PubMed: 19875284]
35. Cheng K, Rowley Kelly A, Kohn RA, Dweck JF, Walsh PJ. Org Lett. 2009; 11:2703–2706. [PubMed: 19456165]
36. Chenoweth DM, Harki DA, Dervan PB. J Am Chem Soc. 2009; 131:7175–7181. [PubMed: 19413320]
37. Chenoweth DM, Harki DA, Phillips JW, Dose C, Dervan PB. J Am Chem Soc. 2009; 131:7182–7188. [PubMed: 19413319]
38. Cherian PT, Koikov LN, Wortman MD, Knittel JJ. Bioorg Med Chem Lett. 2009; 19:2215–2219. [PubMed: 19297156]
39. Cho C-H, Neuenswander B, Lushington GH, Larock RC. J Comb Chem. 2009; 11:900–906. [PubMed: 19569714]
40. Choi YL, Kim JK, Choi S-U, Min Y-K, Bae M-A, Kim BT, Heo J-N. Bioorg Med Chem Lett. 2009; 19:3036–3040. [PubMed: 19394218]

41. Cho-Schultz S, Patten MJ, Huang B, Elleraas J, Gajiwala KS, Hickey MJ, Wang J, Mehta PP, Kang P, Gehring MR, Kung P-P, Sutton SC. J Comb Chem. 2009; 11:860–874. [PubMed: 19583220]
42. Cloonan SM, Keating JJ, Butler SG, Knox AJS, Jorgensen AM, Peters GH, Rai D, Corrigan D, Lloyd DG, Williams DC, Meegan MJ. Eur J Med Chem. 2009; 44:4862–4888. [PubMed: 19717215]
43. Coffinier D, El Kaim L, Grimaud L. Org Lett. 2009; 11:995–997. [PubMed: 19170618]
44. Cole AG, Metzger A, Brescia M-R, Qin L-Y, Appell KC, Brain CT, Hallett A, Ganju P, Denholm AA, Wareing JR, Ritchie TJ, Drake GM, Bevan SJ, MacGloinn A, McBryde A, Patel V, Oakley PJ, Nunez X, Gstach H, Schneider P, Baldwin JJ, Dolle RE, McDonald E, Henderson I. Bioorg Med Chem Lett. 2009; 19:119–122. [PubMed: 19014884]
45. Cole AG, Stauffer TM, Rokosz LL, Metzger A, Dillard LW, Zeng W, Henderson I. Bioorg Med Chem Lett. 2009; 19:378–381. [PubMed: 19059776]
46. Colombo M, Bossolo S, Aramini A. J Comb Chem. 2009; 11:335–337. [PubMed: 19326911]
47. Covel JA, Santora VJ, Smith JM, Hayashi R, Gallardo C, Weinhouse MI, Ibarra JB, Schultz JA, Park DM, Estrada SA, Hofilena BJ, Pulley MD, Smith BM, Ren A, Suarez M, Frazer J, Edwards J, Hauser EK, Lorea J, Semple G, Grottick AJ. J Med Chem. 2009; 52:5603–5611. [PubMed: 19722526]
48. Crestey F, Witt M, Jaroszewski JW, Franzyk H. J Org Chem. 2009; 74:5652–5655. [PubMed: 19518106]
49. Das BC, Madhukumar AV, Anguiano J, Mani S. Bioorg Med Chem Lett. 2009; 19:4204–4206. [PubMed: 19515559]
50. DeAngelis A, Taylor MT, Fox JM. J Am Chem Soc. 2009; 131:1101–1105. [PubMed: 19128053]
51. DeBenedetto MV, Green ME, Wan S, Park J-H, Floreancig PE. Org Lett. 2009; 11:835–838. [PubMed: 19152262]
52. DeNinno MP, Andrews M, Bell AS, Chen Y, Eller-Zarbo C, Eshelby N, Etienne JB, Moore DE, Palmer MJ, Visser MS, Yu LJ, Zavadoski WJ, Michael Gibbs E. Bioorg Med Chem Lett. 2009; 19:2537–2541. [PubMed: 19339180]
53. Deschrijver T, Verwilst P, Broos K, Deckmyn H, Dehaen W, De Borggraeve WM. Tetrahedron. 2009; 65:4521–4529.
54. Devarie-Baez NO, Kim W-S, Smith AB III, Xian M. Org Lett. 2009; 11:1861–1864. [PubMed: 19301924]
55. Dickson DP, Toh C, Lunda M, Yermolina MV, Wardrop DJ, Landrie CL. J Org Chem. 2009; 74:9535–9538. [PubMed: 19911775]
56. Ding Q, Huang X-G, Wu J. J Comb Chem. 2009; 11:1047–1049. [PubMed: 19824656]
57. Ding Q, He X, Wu J. J Comb Chem. 2009; 11:587–591. [PubMed: 19449803]
58. Dockendorff C, Jin S, Olsen M, Lautens M, Coupal M, Hodzic L, Spear N, Payza K, Walpole C, Tomaszewski MJ. Bioorg Med Chem Lett. 2009; 19:1228–1232. [PubMed: 19168350]
59. Donets PA, Van Hecke K, Van Meervelt L, Van der Eycken EV. Org Lett. 2009; 11:3618–3621. [PubMed: 19610606]
60. Dou G, Wang M, Shi D. J Comb Chem. 2008; 11:151–154. [PubMed: 19133838]
61. Dou G, Shi D. J Comb Chem. 2009; 11:1073–1077. [PubMed: 19827793]
62. Duval R, Kolb S, Braud E, Genest D, Garbay C. J Comb Chem. 2009; 11:947–950. [PubMed: 19835352]
63. Dzhavakhishvili SG, Gorobets NY, Shishkina SV, Shishkin OV, Desenko SM, Groth UM. J Comb Chem. 2009; 11:508–514. [PubMed: 19385601]
64. El-Badri MH, Kurth MJ. J Comb Chem. 2009; 11:228–238. [PubMed: 19154120]
65. El Kaim L, Grimaud L, Oble J, Wagschal S. Tetrahedron Lett. 2009; 50:1741–1743.
66. El Kaim L, Grimaud L, Schiltz A. Org Biomol Chem. 2009; 7:3024–3026.
67. El Kaim L, Grimaud L, Schiltz A. Synlett. 2009:1401–1404.
68. El Kaim L, Grimaud L, Wagschal S. Synlett. 2009:1315–1317.

69. Elders N, van der Born D, Hendrickx LJD, Timmer BJJ, Krause A, Janssen E, de Kanter FJJ, Ruijter E, Orru RVA. Angew Chem, Int Ed. 2009; 48:5856–5859. S5856/1–S5856/58.
70. Elford TG, Ulaczyk-Lesanko A, De Pascale G, Wright GD, Hall DG. J Comb Chem. 2009; 11:155–168. [PubMed: 19072614]
71. Engers DW, Rodriguez AL, Williams R, Hammond AS, Venable D, Oluwatola O, Sulikowski GA, Conn PJ, Lindsley CW. ChemMedChem. 2009; 4:505–511. [PubMed: 19197923]
72. Erb W, Neuville L, Zhu J. J Org Chem. 2009; 74:3109–3115. [PubMed: 19284772]
73. Fish PV, Wakenhut F, Ryckmans T, Stobie A. Bioorg Med Chem Lett. 2009; 19:4579–4583. [PubMed: 19616432]
74. Fontaine P, Masson G, Zhu J. Org Lett. 2009; 11:1555–1558. [PubMed: 19320503]
75. Fu J, Shuttleworth SJ, Connors RV, Chai A, Coward P. Bioorg Med Chem Lett. 2009; 19:4264–4267. [PubMed: 19553112]
76. Georgescu E, Caira MR, Georgescu F, Draghici B, Popa MM, Dumitrascu F. Synlett. 2009:1795–1799.
77. Ghahremanzadeh R, Ahadi S, Bazgir A. Tetrahedron Lett. 2009; 50:7379–7381.
78. Ghahremanzadeh R, Sayyafi M, Ahadi S, Bazgir A. J Comb Chem. 2009; 11:393–396. [PubMed: 19425616]
79. Giannini G, Marzi M, Pezzi R, Brunetti T, Battistuzzi G, Marzo MD, Cabri W, Vesci L, Pisano C. Bioorg Med Chem Lett. 2009; 19:2346–2349. [PubMed: 19285395]
80. Gitto R, Pagano B, Citraro R, Scicchitano F, De Sarro G, Chimirri A. Eur J Med Chem. 2009; 44:1349–1354. [PubMed: 18406016]
81. Gladstone SG, Earley WG, Acker JK, Martin GS. Tetrahedron Lett. 2009; 50:3813–3816.
82. Gleave RJ, Beswick PJ, Brown AJ, Giblin GMP, Haslam CP, Livermore D, Moses A, Nicholson NH, Page LW, Slingsby B, Swarbrick ME. Bioorg Med Chem Lett. 2009; 19:6578–6581. [PubMed: 19864133]
83. Gomez AM, Barrio A, Pedregosa A, Valverde S, Lopez JC. J Org Chem. 2009; 74:6323–6326. [PubMed: 19601580]
84. Goncalves S, Wagner A, Mioskowski C, Baati R. Tetrahedron Lett. 2009; 50:274–276.
85. Gong Y-D, Ryu IA. J Comb Chem. 2009; 11:626–630. [PubMed: 19405492]
86. Gonzalez-Arellano C, Luque R, Macquarrie DJ. Chem Commun. 2009:1410–1412.
87. Green DM, Goljer I, Andraka DS, Chengalvala M, Shanno L, Hurlburt W, Pelletier JC. J Comb Chem. 2009; 11:117–125. [PubMed: 19049392]
88. Grote RE, Jarvo ER. Org Lett. 2009; 11:485–488. [PubMed: 19093851]
89. Guchhait SK, Madaan C. Synlett. 2009:628–632.
90. Guo C, Hou X, Dong L, Dagostino E, Greasley S, Ferre R, Marakovits J, Johnson MC, Matthews D, Mroczkowski B, Parge H, VanArsdale T, Popoff I, Piraino J, Margosiak S, Thomson J, Los G, Murray BW. Bioorg Med Chem Lett. 2009; 19:5613–5616. [PubMed: 19729306]
91. Kyle, Hadden M.; Hill, SA.; Davenport, J.; Matts, RL.; Blagg, BSJ. Bioorg Med Chem. 2009; 17:634–640. [PubMed: 19101151]
92. Hamper BC, Kesselring AS, Chott RC, Yang S. J Comb Chem. 2009; 11:469–480. [PubMed: 19296668]
93. Han F-B, Ge Z-M, Cheng T-M, Li R-T. Synlett. 2009:648–650.
94. Han Z-G, Tu S-J, Jiang B, Yan S, Zhang X-H, Wu S-S, Hao W-J, Cao X-D, Shi F, Zhang G, Ma N. Synthesis. 2009:1639–1646.
95. Han Z-G, Zhang G, Jiang B, Ma N, Shi F, Tu S-J. J Comb Chem. 2009; 11:809–812. [PubMed: 19694422]
96. Hao W-J, Jiang B, Tu S-J, Wu S-S, Han Z-G, Cao X-D, Zhang X-H, Yan S, Shi F. J Comb Chem. 2009; 11:310–314. [PubMed: 19123781]
97. Harju K, Manevski N, Yli-Kauhaluoma J. Tetrahedron. 2009; 65:9702–9706.
98. Harju K, Vesterinen J, Yli-Kauhaluoma J. Org Lett. 2009; 11:2219–2221. [PubMed: 19419217]
99. Heng S, Gryncel KR, Kantrowitz ER. Bioorg Med Chem. 2009; 17:3916–3922. [PubMed: 19419876]

100. Henry C, Haupt A, Turner SC. J Org Chem. 2009; 74:1932–1938. [PubMed: 19175330]
101. Heravi MM, Baghernejad B, Oskooie HA. Tetrahedron Lett. 2009; 50:767–769.
102. Heravi MM, Baghernejad B, Oskooie HA. Synlett. 2009:1123–1125.
103. Heravi MM, Sadjadi S, Mokhtari Haj N, Oskooie HA, Shoar RH, Bamoharram FF. Tetrahedron Lett. 2009; 50:943–945.
104. Hermange P, Dau METH, Retailleau P, Dodd RH. Org Lett. 2009; 11:4044–4047. [PubMed: 19678614]
105. Honda T, Terao T, Aono H, Ban M. Bioorg Med Chem. 2009; 17:699–708. [PubMed: 19109024]
106. Hu F, Chou CJ, Gottesfeld JM. Bioorg Med Chem Lett. 2009; 19:3928–3931. [PubMed: 19362838]
107. Hu Z, Ma T, Chen Z, Ye Z, Zhang G, Lou Y, Yu Y. J Comb Chem. 2009; 11:267–273. [PubMed: 19125569]
108. Huang H, Jiang H, Chen K, Liu H. J Org Chem. 2009; 74:5476–5480. [PubMed: 19572501]
109. Huang W, Ding Y, Miao Y, Liu M-Z, Li Y, Yang G-F. Eur J Med Chem. 2009; 44:3687–3696. [PubMed: 19410339]
110. Huang X, Huang J, Cao J. J Comb Chem. 2009; 11:515–518. [PubMed: 19499907]
111. Huang X, Xu J. J Org Chem. 2009; 74:8859–8861. [PubMed: 19860394]
112. Huang X, Xu J-F. J Comb Chem. 2009; 11:350–354. [PubMed: 19220032]
113. Humphries PS, Balan G, Bechle BM, Conn EL, Dirico KJ, Hui Y, Oliver RM, Southers JA, Yang X. Tetrahedron Lett. 2009; 50:2140–2143.
114. Humphries PS, Do Q-QT, Wilhite DM. Tetrahedron Lett. 2009; 50:2552–2554.
115. Humphries PS, Oliver RM. Tetrahedron Lett. 2009; 50:2682–2684.
116. Ibrahim N, Legraverend M. J Comb Chem. 2009; 11:658–666. [PubMed: 19530688]
117. Ito S, Hirata Y, Nagatomi Y, Satoh A, Suzuki G, Kimura T, Satow A, Maehara S, Hikichi H, Hata M, Ohta H, Kawamoto H. Bioorg Med Chem Lett. 2009; 19:5310–5313. [PubMed: 19692242]
118. Izumiseki A, Yoshida K, Yanagisawa A. Org Lett. 2009; 11:5310–5313. [PubMed: 19873985]
119. Jadidi K, Ghahremanzadeh R, Bazgir A. J Comb Chem. 2009; 11:341–344. [PubMed: 19239200]
120. Jarusiewicz J, Choe Y, Yoo KS, Park CP, Jung KW. J Org Chem. 2009; 74:2873–2876. [PubMed: 19265413]
121. Jayanth TT, Zhang L, Johnson TS, Malinakova HC. Org Lett. 2009; 11:815–818. [PubMed: 19159326]
122. Jeganmohan M, Bhuvaneswari S, Cheng C-H. Angew Chem, Int Ed. 2009; 48:391–394.
123. Jeges G, Nagy T, Meszaros T, Kovacs J, Dorman G, Kowalczyk A, Goodnow RA. J Comb Chem. 2009; 11:327–334. [PubMed: 19206513]
124. Jiang B, Cao L-J, Tu S-J, Zheng W-R, Yu H-Z. J Comb Chem. 2009; 11:612–616. [PubMed: 19537742]
125. Jiang B, Hao W-J, Wang X, Shi F, Tu S-J. J Comb Chem. 2009; 11:846–850. [PubMed: 19694408]
126. Jiang B, Tu S-J, Kaur P, Wever W, Li G. J Am Chem Soc. 2009; 131:11660–11661. [PubMed: 19722590]
127. Jiang B, Wang X, Shi F, Tu S-J, Ai T, Ballew A, Li G. J Org Chem. 2009; 74:9486–9489. [PubMed: 19938854]
128. Jiang G, Chen J, Huang J-S, Che C-M. Org Lett. 2009; 11:4568–4571. [PubMed: 19810764]
129. Jiang M, Li T, Meng L, Yang C, Xie Y, Ding J. J Comb Chem. 2009; 11:806–808. [PubMed: 19645499]
130. Jung N, Braese S. J Comb Chem. 2009; 11:47–71. [PubMed: 19055377]
131. Kahlon DK, Lansdell TA, Fisk JS, Tepe JJ. Bioorg Med Chem. 2009; 17:3093–3103. [PubMed: 19328000]
132. Kang SS, Cuendet M, Endringer DC, Croy VL, Pezzuto JM, Lipton MA. Bioorg Med Chem. 2009; 17:1044–1054. [PubMed: 18487053]

133. Karthikeyan SV, Perumal S, Shetty KA, Yogeeswari P, Sriram D. Bioorg Med Chem Lett. 2009; 19:3006–3009. [PubMed: 19403307]
134. Kattar SD, Surdi LM, Zabierek A, Methot JL, Middleton RE, Hughes B, Szewczak AA, Dahlberg WK, Kral AM, Ozerova N, Fleming JC, Wang H, Secrist P, Harsch A, Hamill JE, Cruz JC, Kenific CM, Chenard M, Miller TA, Berk SC, Tempest P. Bioorg Med Chem Lett. 2009; 19:1168–1172. [PubMed: 19138845]
135. Kempe K, Lobert M, Hoogenboom R, Schubert US. J Comb Chem. 2009; 11:274–280. [PubMed: 19236014]
136. Phillip, Kennedy J.; Jeffrey Conn, P.; Lindsley, CW. Bioorg Med Chem Lett. 2009; 19:3204–3208. [PubMed: 19443215]
137. Kharchenko JV, Detistov OS, Orlov VD. J Comb Chem. 2009; 11:216–219. [PubMed: 19133738]
138. Kim Y, Koh M, Kim D-K, Choi H-S, Park SB. J Comb Chem. 2009; 11:928–937. [PubMed: 19746993]
139. Klinkhammer W, Mueller H, Globisch C, Pajeva IK, Wiese M. Bioorg Med Chem. 2009; 17:2524–2535. [PubMed: 19250834]
140. Ko S-K, Jin HJ, Jung D-W, Tian X, Shin I. Angew Chem, Int Ed. 2009; 48:7809–7812. S7809/1–S7809/13.
141. Koci J, Pudelova N, Krchnak V. J Comb Chem. 2009; 11:397–402. [PubMed: 19354277]
142. Kojima N, Nishijima S, Tanaka T. Synlett. 2009:3171–3174.
143. Kotian PL, Krishnan R, Rowland S, El-Kattan Y, Saini SK, Upshaw R, Bantia S, Arnold S, Sudhakar Babu Y, Chand P. Bioorg Med Chem. 2009; 17:3934–3958. [PubMed: 19409795]
144. Kouznetsov VV, Bohorquez ARR, Saavedra LA. Synthesis. 2009:4219–4225.
145. Krasavin M, Shkavrov S, Parchinsky V, Bukhryakov K. J Org Chem. 2009; 74:2627–2629. [PubMed: 19215099]
146. Krupkova S, Soural M, Hlavac J, Hradil P. J Comb Chem. 2009; 11:951–955. [PubMed: 19778035]
147. Ku X, Huang H, Jiang H, Liu H. J Comb Chem. 2009; 11:338–340. [PubMed: 19260652]
148. Kudirka R, Devine SKJ, Adams CS, Van Vranken DL. Angew Chem, Int Ed. 2009; 48:3677–3680. S3677/1–S3677/29.
149. Kumar A, Maurya RA, Ahmad P. J Comb Chem. 2009; 11:198–201. [PubMed: 19152269]
150. Prashantha, Kumar BR.; Sankar, G.; Nasir Baig, RB.; Chandrashekaran, S. Eur J Med Chem. 2009; 44:4192–4198. [PubMed: 19525040]
151. Kumar D, Patel G, Johnson EO, Shah K. Bioorg Med Chem Lett. 2009; 19:2739–2741. [PubMed: 19376704]
152. Kumar D, Reddy VB, Sharad S, Dube U, Kapur S. Eur J Med Chem. 2009; 44:3805–3809. [PubMed: 19419801]
153. Kumar S, Deshpande S, Chandra V, Kitchlu S, Dwivedi A, Nayak VL, Konwar R, Prabhakar YS, Sahu DP. Bioorg Med Chem. 2009; 17:6832–6840. [PubMed: 19740667]
154. Kysil V, Khvat A, Tsirulnikov S, Tkachenko S, Ivachtchenko A. Tetrahedron Lett. 2009; 50:2854–2856.
155. Lafleur K, Huang D, Zhou T, Caflisch A, Nevado C. J Med Chem. 2009; 52:6433–6446. [PubMed: 19788238]
156. Lahue BR, Ma Y, Shipps GW, Seghezzi W, Herbst R. Bioorg Med Chem Lett. 2009; 19:3405–3409. [PubMed: 19481450]
157. Le Gall E, Haurena C, Sengmany S, Martens T, Troupel M. J Org Chem. 2009; 74:7970–7973. [PubMed: 19769334]
158. Le Guevel R, Oger F, Lecorgne A, Dudasova Z, Chevance S, Bondon A, Barath P, Simonneaux G, Salbert G. Bioorg Med Chem. 2009; 17:7021–7030. [PubMed: 19729315]
159. Lechel T, Lentz D, Reissig H-U. Chem--Eur J. 2009; 15:5432–5435. [PubMed: 19388043]
160. Lechel T, Moehl S, Reissig H-U. Synlett. 2009:1059–1062.
161. Lee JM, Yu E-A, Park JY, Ryu IA, Shin DS, Gong Y-D. Bull Korean Chem Soc. 2009; 30:1325–1330.

162. Lee JW, Bork JT, Ha H-H, Samanta A, Chang Y-T. Aust J Chem. 2009; 62:1000–1006.
163. Lee T, Park J-H, Jeon M-K, Gong Y-D. J Comb Chem. 2009; 11:288–293. [PubMed: 19127993]
164. Lee T, Park J-H, Lee D-H, Gong Y-D. J Comb Chem. 2009; 11:495–499. [PubMed: 19338271]
165. Lee Y-S, Park SM, Kim BH. Bioorg Med Chem Lett. 2009; 19:1126–1128. [PubMed: 19147352]
166. Lence E, Castedo L, Gonzalez-Bello C. Tetrahedron Lett. 2009; 50:1795–1798.
167. Litvinov YM, Shestopalov AA, Rodinovskaya LA, Shestopalov AM. J Comb Chem. 2009; 11:914–919. [PubMed: 19711896]
168. Li H-H, Jin Y-H, Wang J-Q, Tian S-K. Org Biomol Chem. 2009; 7:3219–3221. [PubMed: 19641777]
169. Liu G, Wurst JM, Tan DS. Org Lett. 2009; 11:3670–3673. [PubMed: 19634891]
170. Li Y, Yu Y, Giulianotti M, Houghten RA. J Org Chem. 2009; 74:2183–2185. [PubMed: 19173629]
171. Liu H, Dagousset G, Masson G, Retailleau P, Zhu J. J Am Chem Soc. 2009; 131:4598–4599. [PubMed: 19334771]
172. Liu H, Domling A. J Org Chem. 2009; 74:6895–6898. [PubMed: 19663394]
173. Liu R, Zhang J. Chem--Eur J. 2009; 15:9303–9306. [PubMed: 19655351]
174. Liu X-H, Lv P-C, Xue J-Y, Song B-A, Zhu H-L. Eur J Med Chem. 2009; 44:3930–3935. [PubMed: 19423198]
175. Loaiza PR, Loeber S, Huebner H, Gmeiner P. Bioorg Med Chem. 2009; 17:5482–5487. [PubMed: 19592258]
176. Ma W, Peterson B, Kelson A, Laborde E. J Comb Chem. 2009; 11:697–703. [PubMed: 19459688]
177. Ma Y, Wang M, Li D, Bekturhun B, Liu J, Liu Q. J Org Chem. 2009; 74:3116–3121. [PubMed: 19296593]
178. MacLeod C, Tuthill PA, Dolle RE. Synlett. 2009:2857–2861.
179. Madsen P, Kodra JT, Behrens C, Nishimura E, Jeppesen CB, Pridal L, Andersen B, Knudsen LB, Valcarce-Aspegren C, Guldbrandt M, Christensen IT, Jorgensen AS, Ynddal L, Brand CL, Bagger MA, Lau J. J Med Chem. 2009; 52:2989–3000. [PubMed: 19385613]
180. Maiti S, Menendez JC. Synlett. 2009:2249–2252.
181. Malik L, Kelly NM, Ma J-N, Currier EA, Burstein ES, Olsson R. Bioorg Med Chem Lett. 2009; 19:1729–1732. [PubMed: 19230660]
182. Marcaurelle LA, Johannes C, Yohannes D, Tillotson BP, Mann D. Bioorg Med Chem Lett. 2009; 19:2500–2503. [PubMed: 19329314]
183. Mao W-W, Wang T-T, Zeng H-P, Wang Z-Y, Chen J-P, Shen J-G. Bioorg Med Chem Lett. 2009; 19:4570–4573. [PubMed: 19616431]
184. Martyn DC, Beletsky G, Cortese JF, Tyndall E, Liu H, Fitzgerald MM, O'Shea TJ, Liang B, Clardy J. Bioorg Med Chem Lett. 2009; 19:5657–5660. [PubMed: 19699641]
185. McGowan D, Nyanguile O, Cummings MD, Vendeville S, Vandyck K, Van den Broeck W, Boutton CW, De Bondt H, Quirynen L, Amssoms K, Bonfanti J-F, Last S, Rombauts K, Tahri A, Hu L, Delouvroy F, Vermeiren K, Vandercruyssen G, Van der Helm L, Cleiren E, Mostmans W, Lory P, Pille G, Van Emelen K, Fanning G, Pauwels F, Lin T-I, Simmen K, Raboisson P. Bioorg Med Chem Lett. 2009; 19:2492–2496. [PubMed: 19342234]
186. McGuinness BF, Carroll CD, Zawacki LG, Dong G, Yang C, Hobbs DW, Jacob-Samuel B, Hall JW, Jenh C-H, Kozlowski JA, Anilkumar GN, Rosenblum SB. Bioorg Med Chem Lett. 2009; 19:5205–5208. [PubMed: 19647429]
187. McNaughton BR, Gareiss PC, Jacobs SE, Fricke AF, Scott GA, Miller BL. ChemMedChem. 2009; 4:1583–1589. [PubMed: 19670207]
188. Medimagh R, Marque S, Prim D, Marrot J, Chatti S. Org Lett. 2009; 11:1817–1820. [PubMed: 19354322]
189. Meng J, Kung P-P. Tetrahedron Lett. 2009; 50:1667–1670.
190. Mentel M, Schmidt AM, Gorray M, Eilbracht P, Breinbauer R. Angew Chem, Int Ed. 2009; 48:5841–5844.

191. Merkul E, Boersch C, Frank W, Muller TJJ. Org Lett. 2009; 11:2269–2272. [PubMed: 19432414]
192. Merkul E, Grotkopp O, Mueller TJJ. Synthesis. 2009:502–507.
193. Merkul E, Oeser T, Mueller TJJ. Chem--Eur J. 2009; 15:5006–5011. [PubMed: 19360838]
194. Metzger A, Qin L-Y, Cole AG, Saionz KW, Brescia M-R, Gstach H, Wareing JR, Zimmermann J, Brill WKD, Baldwin JJ, Dolle RE, Henderson I. Tetrahedron Lett. 2009; 50:7082–7085.
195. Mewett KN, Fernandez SP, Pasricha AK, Pong A, Devenish SO, Hibbs DE, Chebib M, Johnston GAR, Hanrahan JR. Bioorg Med Chem. 2009; 17:7156–7173. [PubMed: 19783443]
196. Milinkevich KA, Yoo CL, Sparks TC, Lorsbach BA, Kurth MJ. Bioorg Med Chem Lett. 2009; 19:5796–5798. [PubMed: 19700317]
197. Minond D, Saldanha SA, Subramaniam P, Spaargaren M, Spicer T, Fotsing JR, Weide T, Fokin VV, Sharpless KB, Galleni M, Bebrone C, Lassaux P, Hodder P. Bioorg Med Chem. 2009; 17:5027–5037. [PubMed: 19553129]
198. Misra M, Pandey SK, Pandey VP, Pandey J, Tripathi R, Tripathi RP. Bioorg Med Chem. 2009; 17:625–633. [PubMed: 19095455]
199. Mizoguchi H, Oguri H, Tsuge K, Oikawa H. Org Lett. 2009; 11:3016–3019. [PubMed: 19534487]
200. Moghadam M, Tangestaninejad S, Mirkhani V, Mohammadpoor-baltork I, Sirjanian N, Parand S. Bioorg Med Chem. 2009; 17:3394–3398. [PubMed: 19359183]
201. Montes-Avila J, Diaz-Camacho SP, Sicairos-Felix J, Delgado-Vargas F, Rivero IA. Bioorg Med Chem. 2009; 17:6780–6785. [PubMed: 19683930]
202. Moro WB, Yang Z, Kane TA, Zhou Q, Harville S, Brouillette CG, Brouillette WJ. J Comb Chem. 2009; 11:617–625. [PubMed: 19408950]
203. Morton D, Leach S, Cordier C, Warriner S, Nelson A. Angew Chem, Int Ed. 2009; 48:104–109.
204. Mossetti R, Pirali T, Tron GC. J Org Chem. 2009; 74:4890–4892. [PubMed: 19449841]
205. Mudit M, Khanfar M, Muralidharan A, Thomas S, Shah GV, van Soest RWM, El Sayed KA. Bioorg Med Chem. 2009; 17:1731–1738. [PubMed: 19195897]
206. Mugherli L, Burchak ON, Balakireva LA, Thomas A, Chatelain F, Balakirev MY. Angew Chem, Int Ed. 2009; 48:7639–7644.
207. Mukhopadhyay C, Datta A, Butcher RJ. Tetrahedron Lett. 2009; 50:4246–4250.
208. Musonda CC, Whitlock GA, Witty MJ, Brun R, Kaiser M. Bioorg Med Chem Lett. 2009; 19:401–405. [PubMed: 19091562]
209. Nakamura M, Hamasaki T, Tokitou M, Baba M, Hashimoto Y, Aoyama H. Bioorg Med Chem. 2009; 17:4740–4746. [PubMed: 19443225]
210. Nasr G, Petit E, Supuran CT, Winum J-Y, Barboiu M. Bioorg Med Chem Lett. 2009; 19:6014–6017. [PubMed: 19796939]
211. Nefzi A, Appel J, Arutyunyan S, Houghten RA. Bioorg Med Chem Lett. 2009; 19:5169–5175. [PubMed: 19632841]
212. Nguyen QPB, Kim JN, Kim TH. Tetrahedron Lett. 2009; 50:4015–4018.
213. Nikulnikov M, Tsirulnikov S, Kysil V, Ivachtchenko A, Krasavin M. Synlett. 2009:260–262.
214. Ohta Y, Chiba H, Oishi S, Fujii N, Ohno H. J Org Chem. 2009; 74:7052–7058. [PubMed: 19673483]
215. Ohta Y, Kubota Y, Watabe T, Chiba H, Oishi S, Fujii N, Ohno H. J Org Chem. 2009; 74:6299–6302. [PubMed: 19572599]
216. Okamoto N, Sakurai K, Ishikura M, Takeda K, Yanada R. Tetrahedron Lett. 2009; 50:4167–4169.
217. Olsen CA, Ghadiri MR. J Med Chem. 2009; 52:7836–7846. [PubMed: 19705846]
218. Ottesen LK, Olsen CA, Witt M, Jaroszewski JW, Franzyk H. Chem- -Eur J. 2009; 15:2966–2978. [PubMed: 19191236]
219. Owen DR, Rodriguez-Lens M, Corless MD, Gaulier SM, Horne VA, Kinloch RA, Maw GN, Pearce DW, Rees H, Ringer TJ, Ryckmans T, Stammen BLC. Bioorg Med Chem Lett. 2009; 19:1702–1706. [PubMed: 19231185]
220. Park SO, Kim J, Koh M, Park SB. J Comb Chem. 2009; 11:315–326. [PubMed: 19199788]

221. Parlow JJ, Burney MW, Case BL, Girard TJ, Hall KA, Hiebsch RR, Huff RM, Lachance RM, Mischke DA, Rapp SR, Woerndle RS, Ennis MD. Bioorg Med Chem Lett. 2009; 19:4657–4663. [PubMed: 19604694]
222. Pasquini S, Mugnaini C, Brizzi A, Ligresti A, Di Marzo V, Ghiron C, Corelli F. J Comb Chem. 2009; 11:795–798. [PubMed: 19694453]
223. Pelletier SMC, Ray PC, Dixon DJ. Org Lett. 2009; 11:4512–4515. [PubMed: 19764710]
224. Peng LF, Stanton BZ, Maloof N, Wang X, Schreiber SL. Bioorg Med Chem Lett. 2009; 19:6319–6325. [PubMed: 19819139]
225. Perez-Fernandez R, Pittelkow M, Belenguer AM, Lane LA, Robinson CV, Sanders JKM. Chem Commun. 2009:3708–3710.
226. Pettersson H, Bulow A, Ek F, Jensen J, Ottesen LK, Fejzic A, Ma J-N, Del Tredici AL, Currier EA, Gardell LR, Tabatabaei A, Craig D, McFarland K, Ott TR, Piu F, Burstein ES, Olsson R. J Med Chem. 2009; 52:1975–1982. [PubMed: 19338356]
227. Peuchmaur M, Lisowski V, Gandreuil C, Maillard LT, Martinez J, Hernandez J-F. J Org Chem. 2009; 74:4158–4165. [PubMed: 19438216]
228. Pirrung MC, Ghorai S, Ibarra-Rivera TR. J Org Chem. 2009; 74:4110–4117. [PubMed: 19408909]
229. Pirrung MC, Wang J. J Org Chem. 2009; 74:2958–2963. [PubMed: 19354324]
230. Poeylaut-Palena AA, Mata EG. J Comb Chem. 2009; 11:791–794. [PubMed: 19572719]
231. Pokhodylo NT, Matiychuk VS, Obushak MD. J Comb Chem. 2009; 11:481–485. [PubMed: 19382757]
232. Poondra RR, Kumar NN, Bijian K, Prakesch M, Campagna-Slater V, Reayi A, Reddy PT, Choudhry A, Barnes ML, Leek DM, Daroszewska M, Lougheed C, Xu B, Schapira M, Alaoui-Jamali MA, Arya P. J Comb Chem. 2009; 11:303–309. [PubMed: 19146410]
233. Porcheddu A, Giacomelli G, Piredda I. J Comb Chem. 2009; 11:126–130. [PubMed: 19053503]
234. Portela-Cubillo F, Scott JS, Walton JC. J Org Chem. 2009; 74:4934–4942. [PubMed: 19449842]
235. Presset M, Coquerel Y, Rodriguez J. Org Lett. 2009; 11:5706–5709. [PubMed: 20000445]
236. Pudelova N, Krchnak V. J Comb Chem. 2009; 11:851–859. [PubMed: 19689103]
237. Pudelova N, Krchnak V. J Comb Chem. 2009; 11:370–374. [PubMed: 19245249]
238. Qin L-Y, Cole AG, Metzger A, O'Brien L, Sun X, Wu J, Xu Y, Xu K, Zhang Y, Henderson I. Tetrahedron Lett. 2009; 50:419–422.
239. Rawls KA, Therese Lang P, Takeuchi J, Imamura S, Baguley TD, Grundner C, Alber T, Ellman JA. Bioorg Med Chem Lett. 2009; 19:6851–6854. [PubMed: 19889539]
240. Rhoden CRB, Rivera DG, Kreye O, Bauer AK, Westermann B, Wessjohann LA. J Comb Chem. 2009; 11:1078–1082. [PubMed: 19795905]
241. Rinderspacher A, Cremona ML, Liu Y, Deng S-X, Xie Y, Gong G, Aulner N, Tobben U, Myers K, Chung C, Andersen M, Vidovic D, Schurer S, Branden L, Yamamoto A, Landry DW. Bioorg Med Chem Lett. 2009; 19:1715–1717. [PubMed: 19243939]
242. Roberge JY, Harikrishnan LS, Kamau MG, Ruan Z, Van Kirk K, Liu Y, Cooper CB, Poss MA, Dickson JK, Gavai AV, Chao ST, Leith LW, Bednarz MS, Mathur A, Kakarla R, Schnur DM, Vaz R, Lawrence RM. J Comb Chem. 2008; 11:72–82. [PubMed: 19086798]
243. Rolfe A, Young K, Volp K, Schoenen F, Neuenswander B, Lushington GH, Hanson PR. J Comb Chem. 2009; 11:732–738. [PubMed: 19505109]
244. Romagnoli R, Baraldi PG, Carrion MD, Cruz-Lopez O, Lopez Cara C, Basso G, Viola G, Khedr M, Balzarini J, Mahboobi S, Sellmer A, Brancale A, Hamel E. J Med Chem. 2009; 52:5551–5555. [PubMed: 19663386]
245. Rostamnia S, Alizadeh A, Zhu L-G. J Comb Chem. 2009; 11:143–145. [PubMed: 19099427]
246. Roy S, Roy S, Neuenswander B, Hill D, Larock RC. J Comb Chem. 2009; 11:1128–1135. [PubMed: 19817453]
247. Roy S, Roy S, Neuenswander B, Hill D, Larock RC. J Comb Chem. 2009; 11:1061–1065. [PubMed: 19728736]

248. Russo O, Cachard-Chastel M, Riviere C, Giner M, Soulier J-L, Berthouze M, Richard T, Monti J-P, Sicsic S, Lezoualc'h F, Berque-Bestel I. J Med Chem. 2009; 52:2214–2225. [PubMed: 19334715]
249. Ryckmans T, Edwards MP, Horne VA, Correia AM, Owen DR, Thompson LR, Tran I, Tutt MF, Young T. Bioorg Med Chem Lett. 2009; 19:4406–4409. [PubMed: 19500981]
250. Ryu IA, Park JY, Han HC, Gong Y-D. Synlett. 2009:999–1003.
251. Sandulenko Y, Komarov A, Krasavin M. Lett Org Chem. 2009; 6:491–495.
252. Sanganee HJ, Baxter A, Barber S, Brown AJH, Grice D, Hunt F, King S, Laughton D, Pairaudeau G, Thong B, Weaver R, Unitt J. Bioorg Med Chem Lett. 2009; 19:1143–1147. [PubMed: 19171482]
253. Sanudo M, Garcia-Valverde M, Marcaccini S, Delgado JJ, Rojo J, Torroba T. J Org Chem. 2009; 74:2189–2192. [PubMed: 19191573]
254. Sanz-Cervera JF, Blasco R, Piera J, Cynamon M, Ibanez I, Murguia M, Fustero S. J Org Chem. 2009; 74:8988–8996. [PubMed: 19894729]
255. Sarachine MJ, Janjic JM, Wipf P, Day BW. Bioorg Med Chem Lett. 2009; 19:2404–2408. [PubMed: 19356928]
256. Saruta K, Ogiku T, Fukase K. Tetrahedron Lett. 2009; 50:4364–4367.
257. Sasada T, Kobayashi F, Sakai N, Konakahara T. Org Lett. 2009; 11:2161–2164. [PubMed: 19371078]
258. Scott DE, Dawes GJ, Ando M, Abell C, Ciulli A. ChemBioChem. 2009; 10:2772–2779. [PubMed: 19827080]
259. Scott SA, Selvy PE, Buck JR, Cho HP, Criswell TL, Thomas AL, Armstrong MD, Arteaga CL, Lindsley CW, Brown HA. Nat Chem Biol. 2009; 5:108–117. [PubMed: 19136975]
260. Senapati BK, Hwang G-S, Lee S, Ryu DH. Angew Chem, Int Ed. 2009; 48:4398–4401.
261. Seomoon D, AJ, Lee PH. Org Lett. 2009; 11:2401–2404. [PubMed: 19422264]
262. Shaabani A, Rezayan AH, Keshipour S, Sarvary A, Ng SW. Org Lett. 2009; 11:3342–3345. [PubMed: 19719182]
263. Shaginian A, Whitby LR, Hong S, Hwang I, Farooqi B, Searcey M, Chen J, Vogt PK, Boger DL. J Am Chem Soc. 2009; 131:5564–5572. [PubMed: 19334711]
264. Sharma S, Kundu B. J Comb Chem. 2009; 11:720–731. [PubMed: 19435370]
265. Shi H, Liu K, Leong WWY, Yao SQ. Bioorg Med Chem Lett. 2009; 19:3945–3948. [PubMed: 19328682]
266. Shi J, Stover JS, Whitby LR, Vogt PK, Boger DL. Bioorg Med Chem Lett. 2009; 19:6038–6041. [PubMed: 19800226]
267. Shi S, Zhu S, Gerritz SW, Rachwal B, Ruan Z, Hutchins R, Kakarla R, Sofia MJ, Sutton J, Cheney D. Bioorg Med Chem Lett. 2009; 19:6477–6480. [PubMed: 19804972]
268. Shih H-W, Guo C-W, Lo K-H, Huang M-Y, Cheng W-C. J Comb Chem. 2009; 11:281–287. [PubMed: 19199644]
269. Sin N, Venables BL, Combrink KD, Gulgeze HB, Yu K-L, Civiello RL, Thuring J, Wang XA, Yang Z, Zadjura L, Marino A, Kadow KF, Cianci CW, Clarke J, Genovesi EV, Medina I, Lamb L, Krystal M, Meanwell NA. Bioorg Med Chem Lett. 2009; 19:4857–4862. [PubMed: 19596574]
270. Singh OM, Devi NS. J Org Chem. 2009; 74:3141–3144. [PubMed: 19301883]
271. Smith CD, Gavrilyuk JI, Lough AJ, Batey RA. J Org Chem. 2009; 75:702–715. [PubMed: 20039638]
272. Sotoca E, Allais C, Constantieux T, Rodriguez J. Org Biomol Chem. 2009; 7:1911–1920. [PubMed: 19590788]
273. Spatz JH, Welsch SJ, Duhaut D-E, Jaeger N, Boursier T, Fredrich M, Allmendinger L, Ross G, Kolb J, Burdack C, Umkehrer M. Tetrahedron Lett. 2009; 50:1705–1707.
274. Spivey AC, Tseng C-C, Jones TC, Kohler AD, Ellames GJ. Org Lett. 2009; 11:4760–4763. [PubMed: 19778010]
275. Sridharan V, Maiti S, Menendez JC. J Org Chem. 2009; 74:9365–9371. [PubMed: 19921805]

276. Srinivasan R, Tan LP, Wu H, Yang P-Y, Kalesh KA, Yao SQ. Org Biomol Chem. 2009; 7:1821–1828. [PubMed: 19590777]
277. Srivastava S, Beck B, Wang W, Czarna A, Holak TA, Dömling A. J Comb Chem. 2009; 11:631–639. [PubMed: 19548636]
278. Stokes BJ, Jovanovic B, Dong H, Richert KJ, Riell RD, Driver TG. J Org Chem. 2009; 74:3225–3228. [PubMed: 19296584]
279. Sun D, Scherman MS, Jones V, Hurdle JG, Woolhiser LK, Knudson SE, Lenaerts AJ, Slayden RA, McNeil MR, Lee RE. Bioorg Med Chem. 2009; 17:3588–3594. [PubMed: 19386501]
280. Sun Z-Y, Zhu Z, Ye Y, McKittrick B, Czarniecki M, Greenlee W, Mullins D, Guzzi M. Bioorg Med Chem Lett. 2009; 19:6801–6805. [PubMed: 19864135]
281. Tan LP, Wu H, Yang P-Y, Kalesh KA, Zhang X, Hu M, Srinivasan R, Yao SQ. Org Lett. 2009; 11:5102–5105. [PubMed: 19852491]
282. Thansandote P, Gouliaras C, Turcotte-Savard M-O, Lautens M. J Org Chem. 2009; 74:1791–1793. [PubMed: 19199669]
283. Thompson MJ, Adams H, Chen B. J Org Chem. 2009; 74:3856–3865. [PubMed: 19374383]
284. Torr JE, Large JM, McDonald E. Comb Chem High Throughput Screening. 2009; 12:275–284.
285. Wang H-J, Wang Y, Csakai AJ, Earley WG, Herr RJ. J Comb Chem. 2009; 11:355–363. [PubMed: 19301850]
286. Wang J, Cady SD, Balannik V, Pinto LH, DeGrado WF, Hong M. J Am Chem Soc. 2009; 131:8066–8076. [PubMed: 19469531]
287. Wang Q-F, Song X-K, Chen J, Yan C-G. J Comb Chem. 2009; 11:1007–1010. [PubMed: 19603746]
288. Wang W, Dömling A. J Comb Chem. 2009; 11:403–409. [PubMed: 19425617]
289. Wang Y, Sauer DR, Djuric SW. Tetrahedron Lett. 2009; 50:5145–5148.
290. Wang Z, Wang Y, Zhang W-X, Hou Z, Xi Z. J Am Chem Soc. 2009; 131:15108–15109. [PubMed: 19919156]
291. Ward TR, Turunen BJ, Haack T, Neuenswander B, Shadrick W, Georg GI. Tetrahedron Lett. 2009; 50:6494–6497. [PubMed: 20161410]
292. Wei W, Cai C, Kota S, Takahashi V, Ni F, Strosberg AD, Snyder JK. Bioorg Med Chem Lett. 2009; 19:6926–6930. [PubMed: 19896376]
293. Weterings JJ, Khan S, van der Heden van Noort GJ, Melief CJM, Overkleeft HS, van der Burg SH, Ossendorp F, van der Marel GA, Filippov DV. Bioorg Med Chem Lett. 2009; 19:2249–2251. [PubMed: 19299126]
294. Wiehn MS, Furniss D, Brase S. J Comb Chem. 2009; 11:982–1006. [PubMed: 19722501]
295. Wiehn MS, Lindell SD, Brase S. J Comb Chem. 2009; 11:960–981. [PubMed: 19722502]
296. Wilk W, Noeren-Mueller A, Kaiser M, Waldmann H. Chem--Eur J. 2009; 15:11976–11984. [PubMed: 19784970]
297. Williams R, Johnson KA, Gentry PR, Niswender CM, Weaver CD, Conn PJ, Lindsley CW, Hopkins CR. Bioorg Med Chem Lett. 2009; 19:4967–4970. [PubMed: 19640716]
298. Worlikar SA, Neuenswander B, Lushington GH, Larock RC. J Comb Chem. 2009; 11:875–879. [PubMed: 19746991]
299. Xiang J, Yang H, Che C, Zou H, Yang H, Wei Y, Quan J, Zhang H, Yang Z, Lin S. PLoS ONE. 2009; 4:e4361. [PubMed: 19194508]
300. Xie Y, Liu Y, Gong G, Smith DH, Yan F, Rinderspacher A, Feng Y, Zhu Z, Li X, Deng S-X, Branden L, Vidovic D, Chung C, Schuerer S, Morisseau C, Hammock BD, Landry DW. Bioorg Med Chem Lett. 2009; 19:2354–2359. [PubMed: 19303288]
301. Xu J-F, Huang X. J Comb Chem. 2009; 11:938–942. [PubMed: 19691355]
302. Xu X, Liang L, Liu J, Yang J, Mai L, Li Y. Tetrahedron Lett. 2009; 50:57–59.
303. Yadav JS, Subba Reddy BV, Madhusudhan Reddy G, Rehana Anjum S. Tetrahedron Lett. 2009; 50:6029–6031.
304. Yaziji V, Coelho A, El Maatougui A, Brea J, Loza MI, Garcia-Mera X, Sotelo E. J Comb Chem. 2009; 11:519–522. [PubMed: 19472983]

305. Yermolayev SA, Gorobets NY, Desenko SM. J Comb Chem. 2008; 11:44–46. [PubMed: 19061393]
306. Yu X, Wu J. J Comb Chem. 2009:895–899. [PubMed: 19694423]
307. Yongye AB, Appel JR, Giulianotti MA, Dooley CT, Medina-Franco JL, Nefzi A, Houghten RA, Martinez-Mayorga K. Bioorg Med Chem. 2009; 17:5583–5597. [PubMed: 19576786]
308. Yoo EJ, Park SH, Lee SH, Chang S. Org Lett. 2009; 11:1155–1158. [PubMed: 19209900]
309. Yoshida M, Hedberg C, Kaiser M, Waldmann H. Chem Commun. 2009:2926–2928.
310. Zajdel P, Subra G, Verdie P, Bojarski AJ, Duszynska B, Basista K, Obniska J, Martinez J, Pawlowski M. Eur J Med Chem. 2009; 44:800–808. [PubMed: 18603331]
311. Zajdel P, Subra G, Verdie P, Gabzdyl E, Bojarski AJ, Duszynska B, Martinez J, Pawlowski M. Bioorg Med Chem Lett. 2009; 19:4827–4831. [PubMed: 19560916]
312. Zhang F-L, Xu A-W, Gong Y-F, Wei M-H, Yang X-L. Chem--Eur J. 2009; 15:6815–6818. [PubMed: 19544501]
313. Zhang L, Xiao Q, Ma C, Xie X-Q, Floreancig PE. J Comb Chem. 2009; 11:640–644. [PubMed: 19505108]
314. Zhao C, Tovar C, Yin X, Xu Q, Todorov IT, Vassilev LT, Chen L. Bioorg Med Chem Lett. 2009; 19:319–323. [PubMed: 19071019]
315. Zhu W, Mena M, Jnoff E, Sun N, Pasau P, Ghosez L. Angew Chem, Int Ed. 2009; 48:5880–5883.
316. Zhu Z, Sun Z-Y, Ye Y, McKittrick B, Greenlee W, Czarniecki M, Fawzi A, Zhang H, Lachowicz JE. Bioorg Med Chem Lett. 2009; 19:5218–5221. [PubMed: 19643610]
317. Wang DM, Sun MN, Liu G. J Comb Chem. 2009; 11:556–575. [PubMed: 19469481]
318. Dobbs AP, Jones P, Penny MJ, Rigby SE. Tetrahedron. 2009; 65:5271–5277.
319. Fustero S, Sancho AG, Acena JL, Sanz-Cervera JF. J Org Chem. 2009; 74:6398–6401. [PubMed: 19588888]
320. Podgorsek A, Jurisch M, Stavber S, Zupan M, Iskra J, Gladysz JA. J Org Chem. 2009; 74:3133–3140. [PubMed: 19320432]
321. Furuya T, Nomoto T, Fukuhara T, Hara S. J Fluorine Chem. 2009; 130:348–353.
322. O'Leary-Steele C, Cordier C, Hayes J, Warriner S, Nelson A. Fluorous-tagged "safety catch" linker for preparing heterocycles by ring-closing metathesis. Org Lett. 2009; 11:915–918. (Example I). [PubMed: 19173645]
323. Boldon S, Moore JE, Gouverneur V. J Fluorine Chem. 2009; 130:1151–1156.
324. Song S, Zhang Q. Org Lett. 2009; 11:4882–4885. [PubMed: 19807115]
325. Donovan AC, Valliant JF. J Org Chem. 2009; 74:8133–8138. [PubMed: 19799396]
326. Sancho AG, Wang X, Sui B, Curran DP. Adv Synth Catal. 2009; 351:1035–1040. [PubMed: 20160880]
327. Liu A, Zhou H, Su G, Zhang W, Yan B. J Comb Chem. 2009; 11:1083–1093. (Example III). [PubMed: 19807063]
328. Werner S, Nielsen SD, Wipf P, Turner DM, Chambers PG, Geib SJ, Curran DP, Zhang W. J Comb Chem. 2009; 11:452–459. [PubMed: 19301852]
329. Jogireddy R, Dakas P-Y, Valot G, Barluenga S, Winssinger N. Chem Eur J. 2009; 15:11498–11506. [PubMed: 19821461]
330. Miller M, Vogel JC, Tsang W, Merrit A, Procter DJ. Org Biomol Chem. 2009; 7:589–597. [PubMed: 19156326]
331. Curran DP, Sui B. J Am Chem Soc. 2009; 131:5411–5413. [PubMed: 19323551]
332. Sáenz JB, Sun WJ, Chang JW, Li J, Bursulaya B, Gray NS, Haslam DB. Nature Chem Biol. 2009; 5:157–165. [PubMed: 19182783]
333. Rosen MD, Woods CR, Goldberg SD, Hack MD, Bounds AD, Yang Y, Wagaman PC, Phuong VK, Ameriks AP, Barrett TD, Kanelakis KC, Chang J, Shankley NP, Rabinowitz MH. Bioorg Med Chem Lett. 2009; 19:6548–6551. [PubMed: 19854648]
334. Scott SA, Selvy PE, Buck JR, Cho HP, Criswell TL, Thomas AL, Armstrong MD, Arteaga CL, Lindsley CW, Brown HA. Nature Chem Biol. 2009; 5:108–117. [PubMed: 19136975]

335. Chen B, Dodge ME, Tang W, Lu J, Ma Z, Fan C-W, Wei S, Hao W, Kilgore J, Williams NS, Roth MG, Amatruda JF, Chen C, Lum L. Nature Chem Biol. 2009; 5:100–107. [PubMed: 19125156]

336. Stanton BZ, Peng LF, Maloof N, Nakai K, Wang X, Duffner JL, Taveras KM, Hyman JM, Lee SW, Koehler AN, Chen JK, Fox JL, Mandinova A, Schreiber SL. Nature Chem Biol. 2009; 5:154–156. [PubMed: 19151731]

337. Long JZ, Li W, Booker L, Burston JJ, Kinsey SG, Schlosburg JE, Pavón FJ, Serrano AM, Selley DE, Parsons LH, Lichtman AH, Cravatt BF. Nature Chem Biol. 2009; 5:37–44. [PubMed: 19029917]

338. Singh J, Zeller W, Zhou N, Hategen G, Mishra R, Polozov A, Yu P, Onua E, Zhang J, Zembower D, Kiselyov A, Ramírez JL, Sigthorsson G, Bjornsson JM, Thorsteinsdottir M, Andrésson T, Bjarnadottir M, Magnusson O, Fabre J-E, Stefansson K, Gurney ME. ACS Chem Biol. 2009; 4:115–126. [PubMed: 19193156]

339. Trujillo JI, Kiefer JR, Huang W, Thorarensen A, Xing L, Caspers NL, Day JE, Mathis KJ, Kretzmer KK, Reitz BA, Weinberg RA, Stegeman RA, Wrightstone A, Christine L, Compton R, Li X. Bioorg Med Chem Lett. 2009; 19:908–911. [PubMed: 19097791]

340. Naylor E, Arredouani A, Vasudevan SR, Lewis AM, Parkesh R, Mizote A, Rosen D, Thomas JM, Izumi M, Ganesan A, Galione A, Churchill GC. Nature Chem Biol. 2009; 5:220–226. [PubMed: 19234453]

341. Soucy TA, Smith PG, Milhollen MA, Berger AJ, Gavin JM, Adhikari S, Brownell JE, Burke KW, Cardin DP, Critchley S, Cullis CA, Doucette A, Garnsey JJ, Gaulin JL, Gershman RE, Lublinsky McDonald A, Mizutani H, Narayanan U, Olhava EJ, Peluso S, Rezaei M, Sintchak MD, Talreja T, Thomas MP, Traore T, Vyskocil S, Weatherhead GS, Yu J, Zhang J, Dick LR, Claiborne CF, Rolfe M, Bolen JB, Langston SP. Nature. 2009; 458:732–737. [PubMed: 19360080]

342. Nolen BJ, Tomasevic N, Russell A, Pierce DW, Jia Z, McCormick CD, Hartman J, Sakowicz R, Pollard TD. Nature. 2009; 460:1031–1035. [PubMed: 19648907]

343. Mott BT, Tanega C, Shen M, Maloney DJ, Shinn P, Leister W, Marugan JJ, Inglese J, Austin CP, Misteli T, Auld DS, Thomas CJ. Bioorg Med Chem Lett. 2009; 19:6700–6705. [PubMed: 19837585]

344. Pathania R, Zlitne S, Barker C, Das R, Gerritsma DA, Lebert J, Awuah E, Melacini G, Capretta FA, Brown ED. Nature Chem Biol. 2009; 5:849–856. [PubMed: 19783991]

345. Molina G, Vogt A, Bakan A, Dai W, de Oliveira PQ, Znosko W, Smithgall TE, Bahar I, Lazo JS, Day BW, Tsang M. Nature Chem Biol. 2009; 5:680–687. [PubMed: 19578332]

346. Erkizan HV, Kong Y, Mechant M, Schlottmann S, Barger-Rotenberg JS, Yuan L, Abaan OD, Chou T-h, Dakshanamurthy S, Brown ML, Űren A, Toretsky JA. Nature Med. 2009; 15:750–757. [PubMed: 19584866]

347. Muchamuel T, Basler M, Aujay MA, Suzuki E, Kalim KW, Lauer C, Sylvain C, Ring ER, Shields J, Jiang J, Shwonek P, Parlati F, Demo SD, Bennett MK, Kirk CJ, Groettrup M. Nature Med. 2009; 15:781–788. [PubMed: 19525961]

348. Swoboda JG, Meredith TC, Campbell J, Brown S, Suzuki T, Bollenbach T, Malhowski AJ, Kishony R, Gilmore MS, Walker S. ACS Chem Biol. 2009; 4:875–883. [PubMed: 19689117]

349. Kamisuki S, Mao Q, Abu-Elheiga L, Gu Z, Kugimiya A, Kwon Y, Shinohara T, Kawazoe Y, Sato S-I, Asakura K, Choo H-YP, Sakai J, Wakil SJ, Uesugi M. Chem Biol. 2009; 16:882–892. [PubMed: 19716478]

350. Ali A, Ghosh A, Nathans RS, Sharova N, O'Brien S, Cao H, Stevenson M, Rana TM. ChemBioChem. 2009; 10:2072–2080. [PubMed: 19603446]

351. Delpire E, Days E, Lewis LM, Mi D, Kim K, Lindsley CW, Weaver CD. Proc Natl Acad Sci USA. 2009; 106:5383–5388. [PubMed: 19279215]

352. Suwa A, Yamamoto T, Sawada A, Minoura K, Hosogai N, Tahara A, Kurama T, Shimokawa T, Aramori I. Br J Pharmacol. 2009; 158:879–887. [PubMed: 19694723]

353. Freschauf GK, Karimi-Busheri F, Ulaczyk-Lesanko A, Mereniuk TR, Ahrens A, Koshy JM, Rasouli-Nia A, Pasarj P, Holmes CFB, Rininsland F, Hall DG, Weinfeld M. Cancer Res. 2009; 69:7739–7746. [PubMed: 19773431]

354. Verhoest PR, Proulx-Lafrance C, Corman M, Chenard L, Helal CJ, Hou X, Kleiman R, Liu S, Marr E, Menniti FS, Schmidt CJ, Vanase-Frawley M, Schmidt AW, Willimas RD, Nelson FR, Fonseca KR, Liras S. J Med Chem. 2009; 52:7946–7949. [PubMed: 19919087]

355. Takahashi T, Nagase T, Sasaki T, Nagumo A, Shimamura K, Miyamoto Y, Kitazawa H, Kanesaka M, Yoshimoto R, Aragane K, Tokita S, Sato N. J Med Chem. 2009; 52:3142–3145. [PubMed: 19388647]

356. Abid M, Torok B, Huang X. Aust J Chem. 2009; 62:208–222.

357. Akritopoulou-Zanze I, Hajduk PJ. Drug Discovery Today. 2009; 14:291–297. [PubMed: 19121409]

358. Bartels JL, Lu P, Walker A, Maurer K, Moeller KD. Chem Commun. 2009:5573–5575.

359. Bouillon I, Soural M, Miller MJ, Krchnak V. J Comb Chem. 2009; 11:213–215. [PubMed: 19125667]

360. FitzGibbons J, Op S, Hobson A, Schaffter L. J Comb Chem. 2009; 11:592–597. [PubMed: 19422189]

361. Gil C, Braese S. J Comb Chem. 2009; 11:175–197. [PubMed: 19053506]

362. Hansen MH, Blakskjaer P, Petersen LK, Hansen TH, Hoejfeldt JW, Gothelf KV, Hansen NJVJ. J Am Chem Soc. 2009; 131:1322–1327. [PubMed: 19123795]

363. Herrmann A. Org Biomol Chem. 2009; 7:3195–3204. [PubMed: 19641772]

364. Hu L, Bartels JL, Bartels JW, Maurer K, Moeller KD. J Am Chem Soc. 2009; 131:16638–16639. [PubMed: 19874016]

365. Hulme C, Chappeta S, Dietrich J. Tetrahedron Lett. 2009; 50:4054–4057.

366. Hung AW, Silvestre HL, Wen S, Ciulli A, Blundell TL, Abell C. Angew Chem, Int Ed. 2009; 48:8452–8456.

367. Ji H, Li H, Martasek P, Roman LJ, Poulos TL, Silverman RB. J Med Chem. 2009; 52:779–797. [PubMed: 19125620]

368. Kagan M, Chlenov M, Melnikov S, McConnell O, Bach AC, Carter G, Failli A, Caggiano TJ, Shumsky JS, Lubda D. J Comb Chem. 2009; 11:704–719. [PubMed: 19459687]

369. Oyarzabal J, Howe T, Alcazar J, Andres JI, Alvarez RM, Dautzenberg F, Iturrino L, Martinez S, Van der Linden I. J Med Chem. 2009; 52:2076–2089. [PubMed: 19290642]

370. Ryba TD, Depew KM, Marcaurelle LA. J Comb Chem. 2009; 11:110–116. [PubMed: 19049425]

371. Scott WL, O'Donnell MJ. J Comb Chem. 2009; 11:3–13. [PubMed: 19105724]

372. Scott WL, Alsina J, Audu CO, Babaev E, Cook L, Dage JL, Goodwin LA, Martynow JG, Matosiuk D, Royo M, Smith JG, Strong AT, Wickizer K, Woerly EM, Zhou Z, O'Donnell MJ. J Comb Chem. 2008; 11:14–33. [PubMed: 19105725]

373. Scott WL, Audu CO, Dage JL, Goodwin LA, Martynow JG, Platt LK, Smith JG, Strong AT, Wickizer K, Woerly EM, O'Donnell MJ. J Comb Chem. 2009; 11:34–43. [PubMed: 19105723]

374. Shelat AA, Guy RK. Bioorg Med Chem. 2009; 17:1088–1093. [PubMed: 18343129]

375. Stocks MJ, Wilden GRH, Pairaudeau G, Perry MWD, Steele J, Stonehouse JP. ChemMedChem. 2009; 4:800–808. [PubMed: 19360802]

376. Taldone T, Sun W, Chiosis G. Bioorg Med Chem. 2009; 17:2225–2235. [PubMed: 19017562]

377. Vulpetti A, Hommel U, Landrum G, Lewis R, Dalvit C. J Am Chem Soc. 2009; 131:12949–12959. [PubMed: 19702332]

378. Xie J, Thapa R, Reverdatto S, Burz DS, Shekhtman A. J Med Chem. 2009; 52:3516–3522. [PubMed: 19422228]

379. Clark, Matthew A.; Acharya, Raksha A.; Arico-Muendel, Christopher C.; Belyanskaya, Svetlana L.; Benjamin, Dennis R.; Carlson, Neil R.; Centrella, Paolo A.; Chiu, Cynthia H.; Creaser, Steffen P.; Cuozzo, John W.; Davie, Christopher P.; Ding, Yun; Franklin, G Joseph; Franzen, Kurt D.; Gefter, Malcolm L.; Hale, Steven P.; Hansen, Nils JV.; Israel, David I.; Jiang, Jinwei; Kavarana, Malcolm J.; Kelley, Michael S.; Kollmann, Christopher S.; Li, Fan; Lind, Kenneth; Mataruse, Sibongile; Medeiros, Patricia F.; Messer, Jeffrey A.; Myers, Paul; O'Keefe, Heather; Oliff, Matthew C.; Rise, Cecil E.; Satz, Alexander L.; Skinner, Steven R.; Svendsen, Jennifer L.; Tang, Lujia; van Vloten, Kurt; Wagner, Richard W.; Yao, Gang; Zhao, Baoguang; Morgan, Barry A. Nature Chem Biol. 2009; 5:647–654. [PubMed: 19648931]

380. Zhang, W. Linker Strategies in Solid-Phase Organic Synthesis. Scott, P., editor. Wiley; 2009. p. 553-576.

381. Zhang W. Chem Rev. 2009; 109:749–795. [PubMed: 19146385]

382. Zhang W. Green Chem. 2009; 11:911–920.

383. Zhang, W. Fluorine in Medicinal Chemistry and Chemical Biology. Ojima, I., editor. Wiley-Blackwell; 2009. p. 335-359.

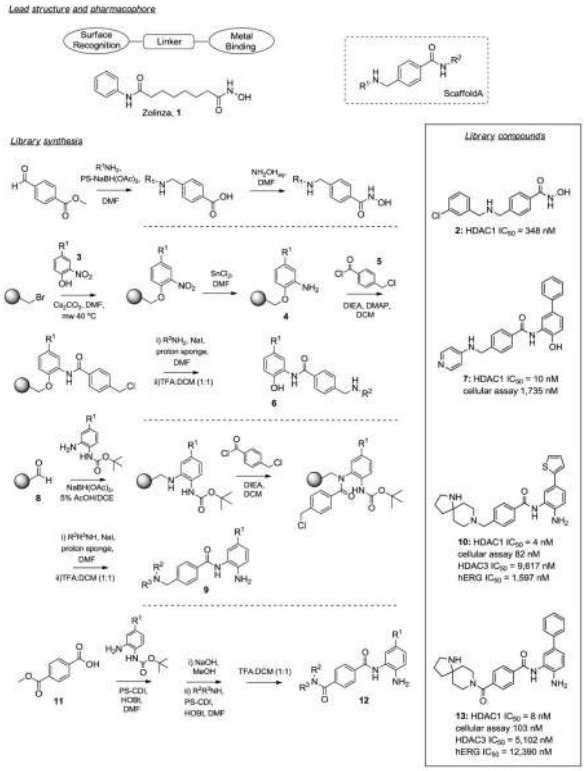


Figure 1.
HDAC1/HDAC2 inhibitors.[134]

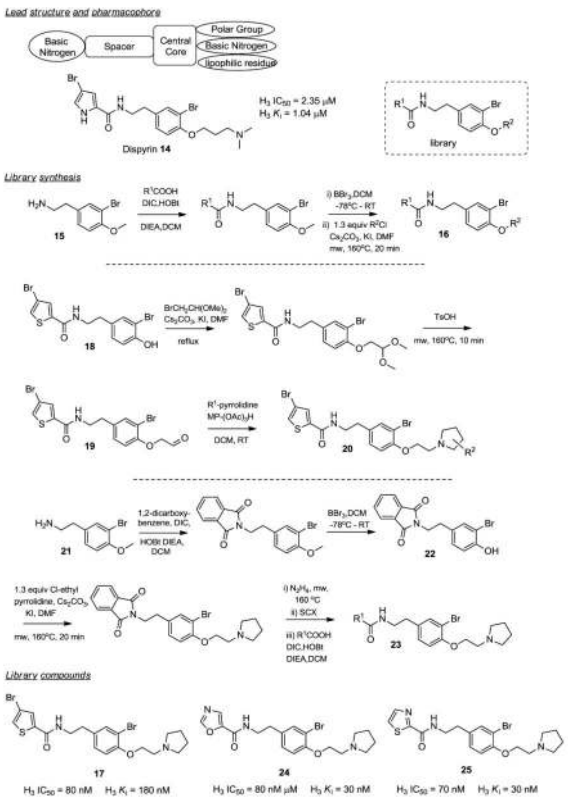


Figure 2.
Histamine H_3 antagonists.[136]

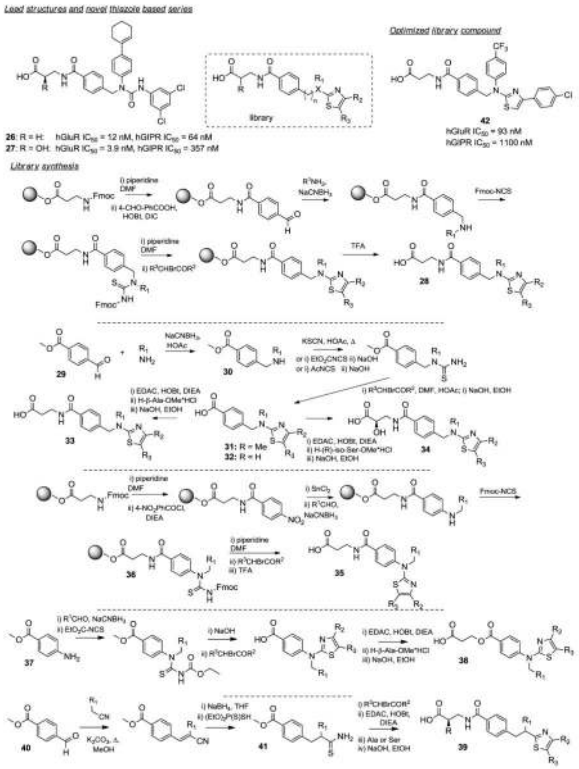


Figure 3.
Glucogon receptor antagonists.[179]

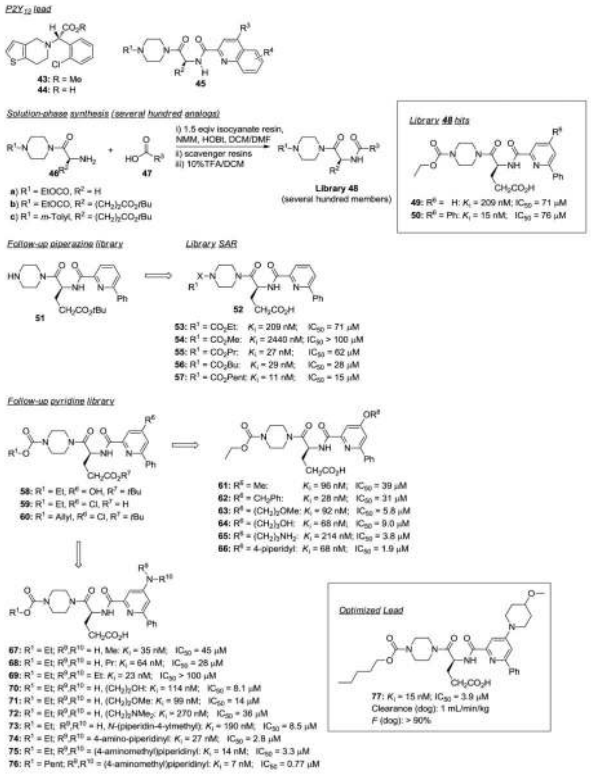


Figure 4.

$P2Y_{12}$ receptor antagonists.[221]

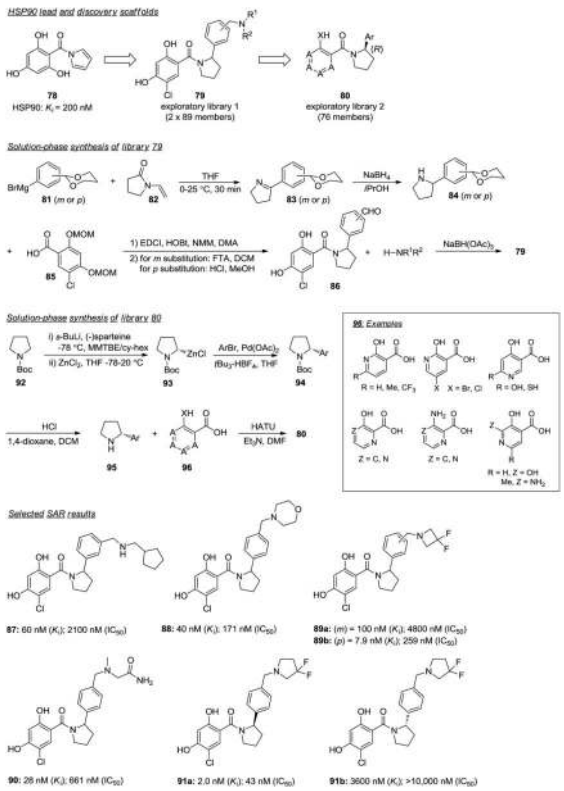


Figure 5.
Hsp90 inhibitors.[41]

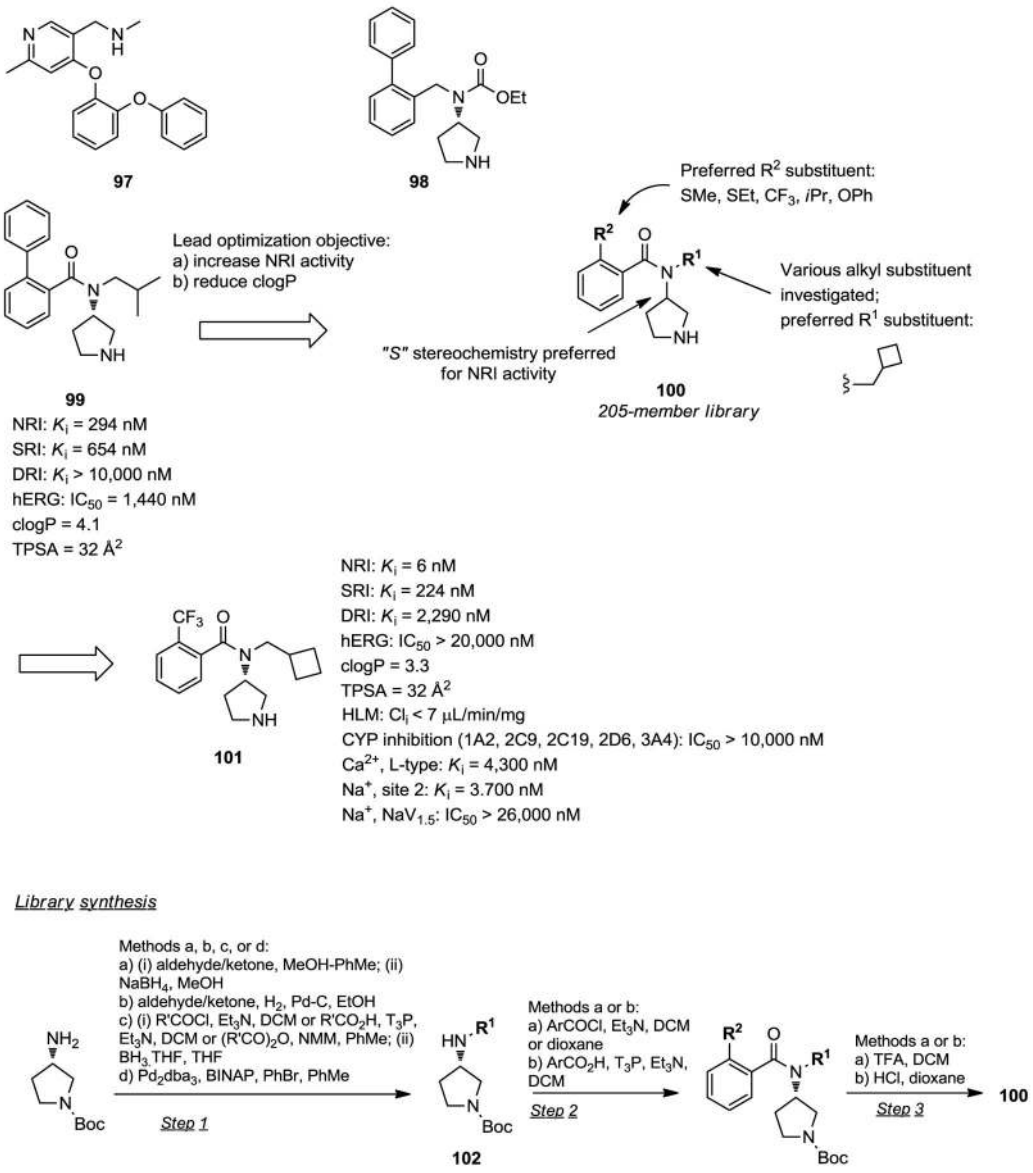
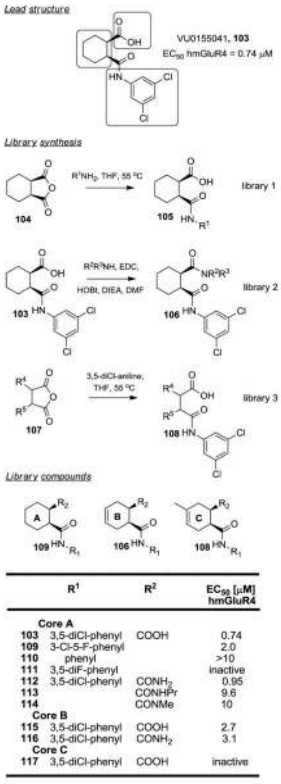


Figure 6.
Selective noradrenaline reuptake inhibitors.[73]



	R1	R2	EC50 [μM] hmGluR4
Core A			
103	3,5-diCl-phenyl	COOH	0.74
109	3-Cl-5-F-phenyl		2.0
110	phenyl		>10
111	3,5-diF-phenyl		inactive
112	3,5-diCl-phenyl	$CONH_2$	0.95
113		CONHPr	9.6
114		CONMe	10
Core B			
115	3,5-diCl-phenyl	COOH	2.7
116	3,5-diCl-phenyl	$CONH_2$	3.1
Core C			
117	3,5-diCl-phenyl	COOH	inactive

Figure 7.
Positive allosteric modulators of mGluR4.[297]

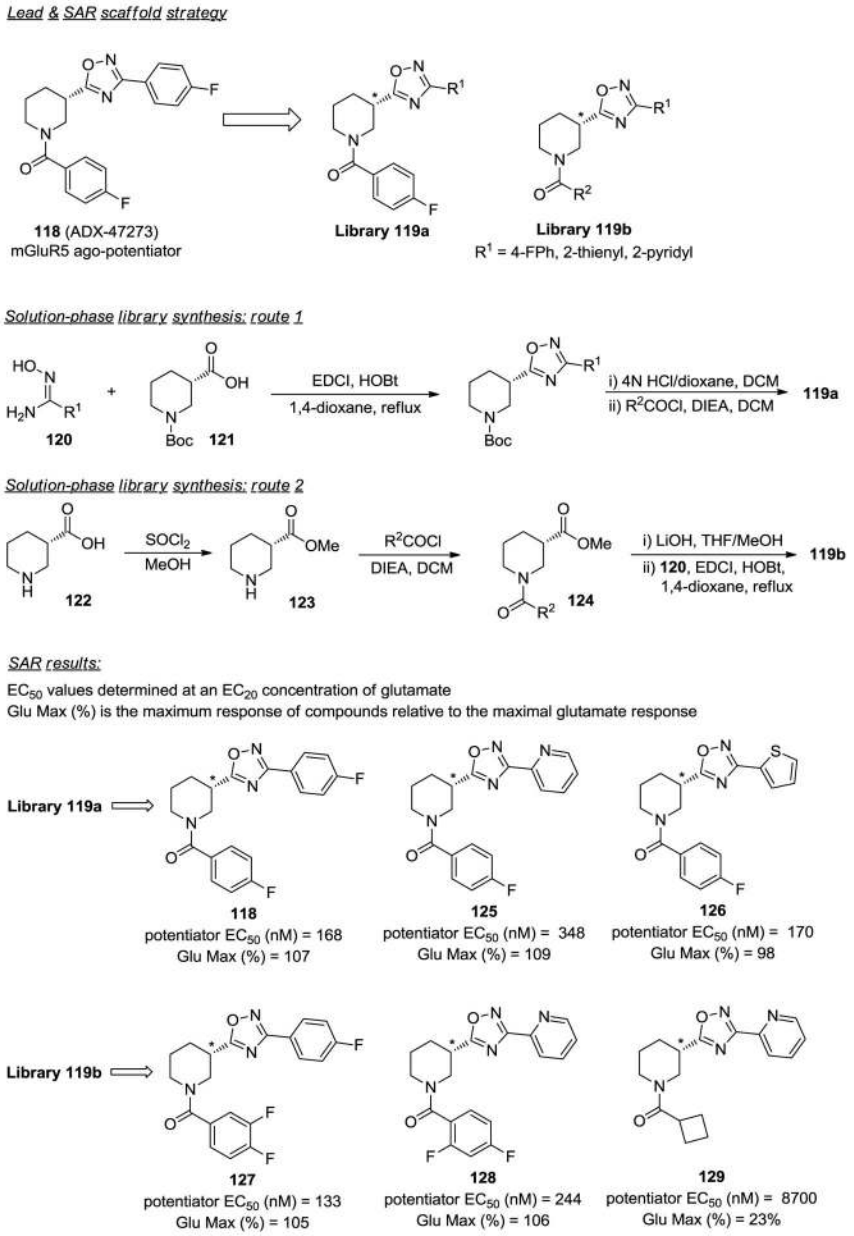


Figure 8.
mGluR5 allosteric modulators.[71]

Figure 9a.

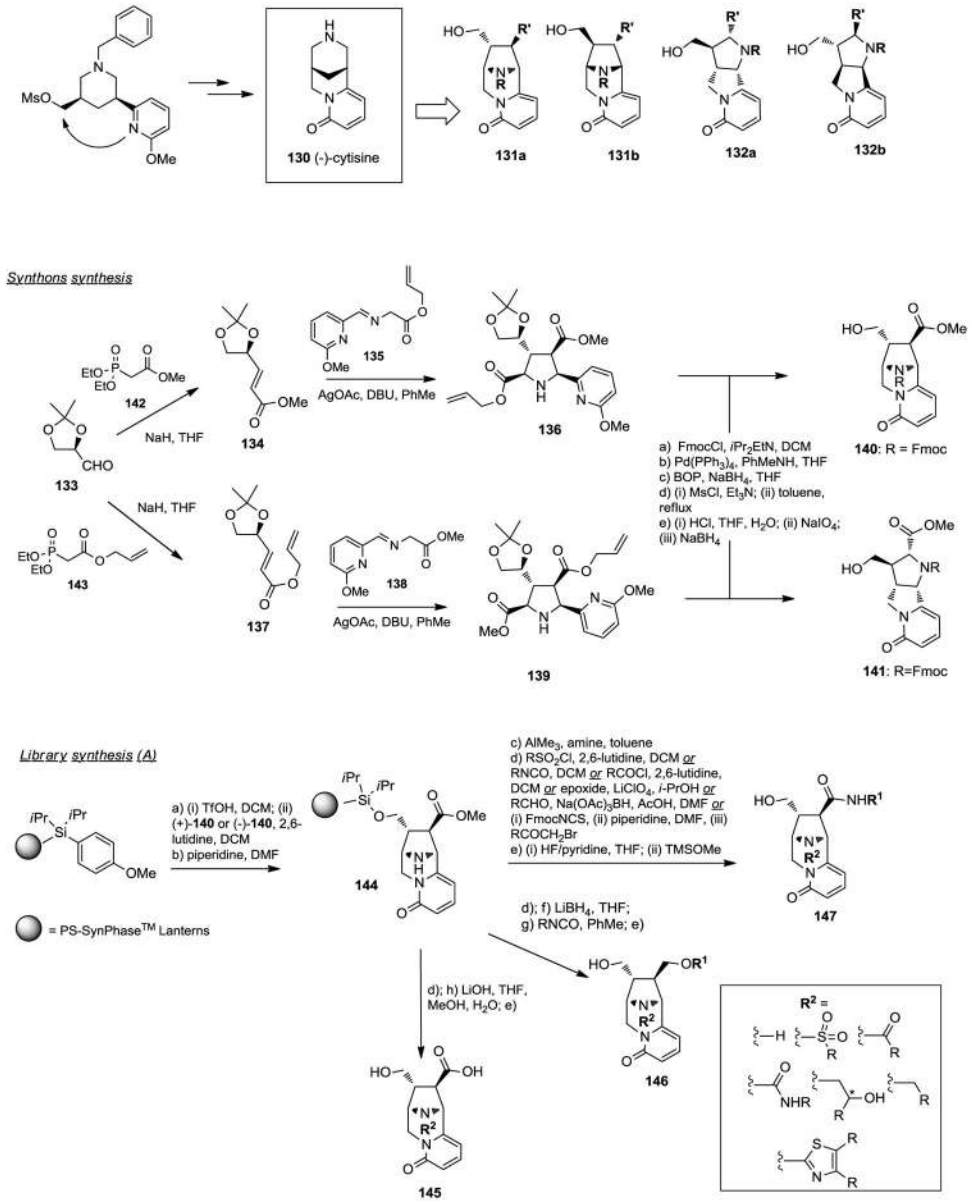


Figure 9b.

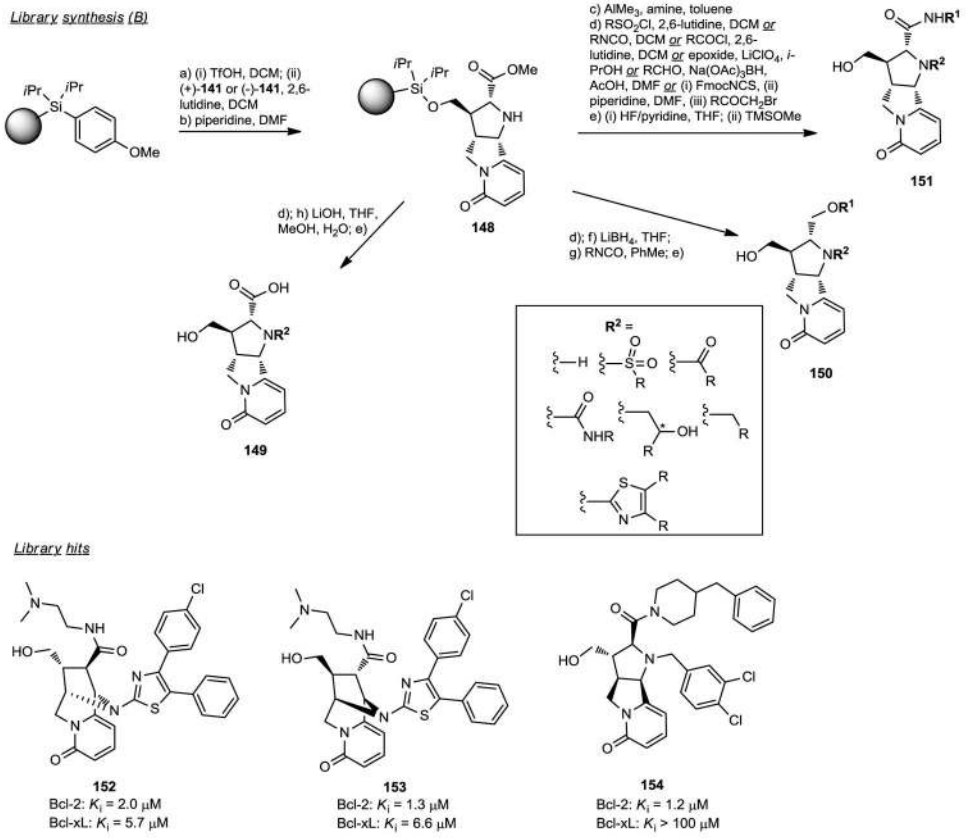


Figure 9.

Figure 9a. Bcl-2 inhibitors via DOS library.[182]

Figure 9b. Bcl-2 inhibitors via DOS library (continued).

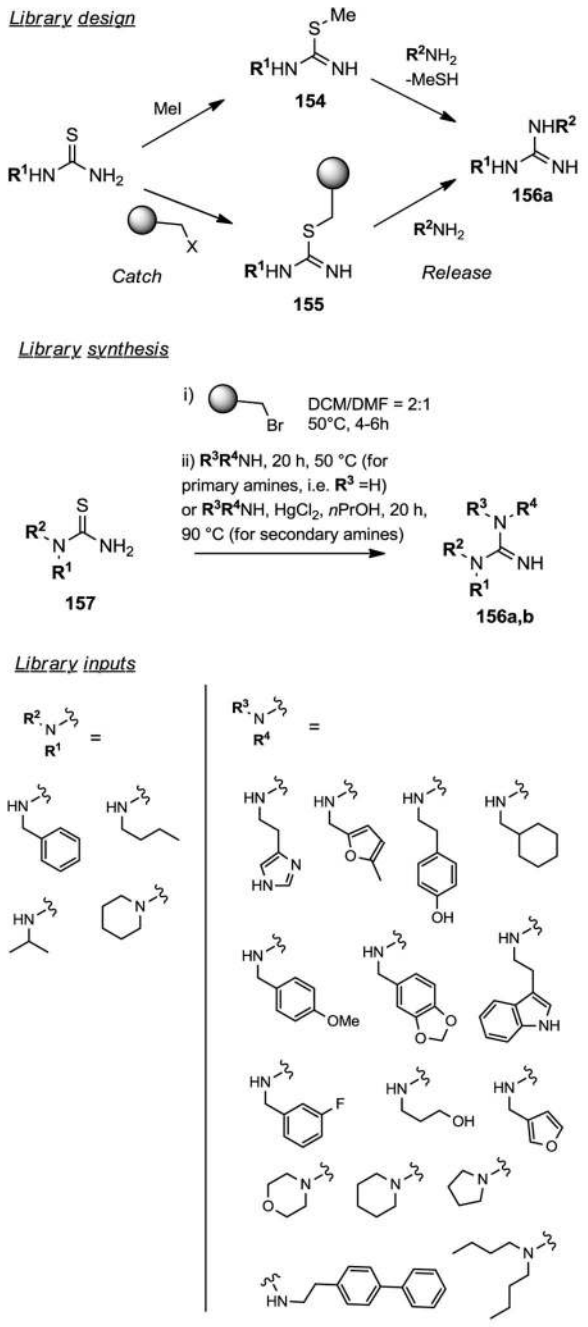


Figure 10.
Catch and release synthesis of substituted guanidines.[289]

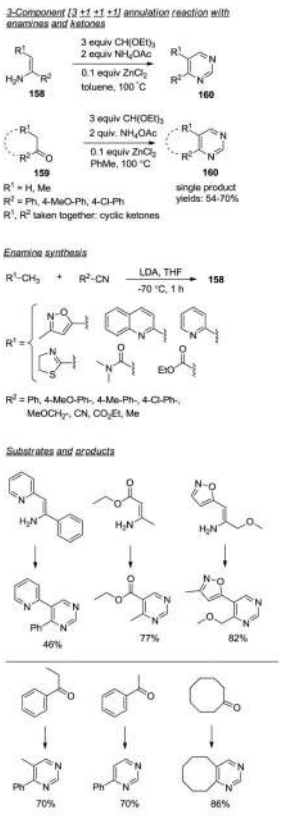


Figure 11.
MCR to pyrimidine derivatives.[158]

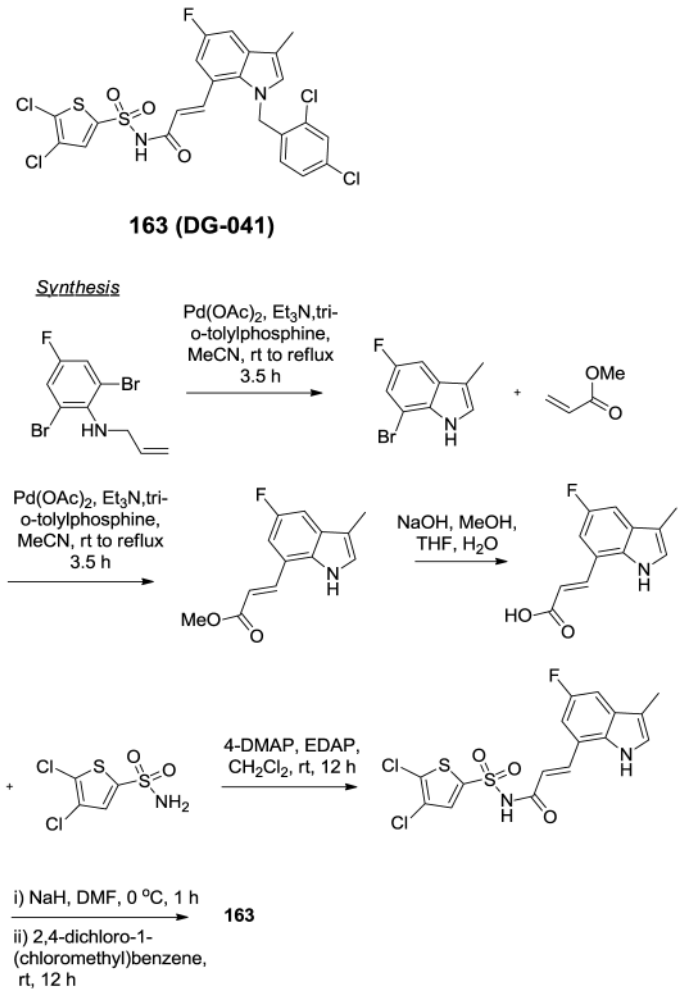


Figure 12.
Discovery of Ned-19.[340]

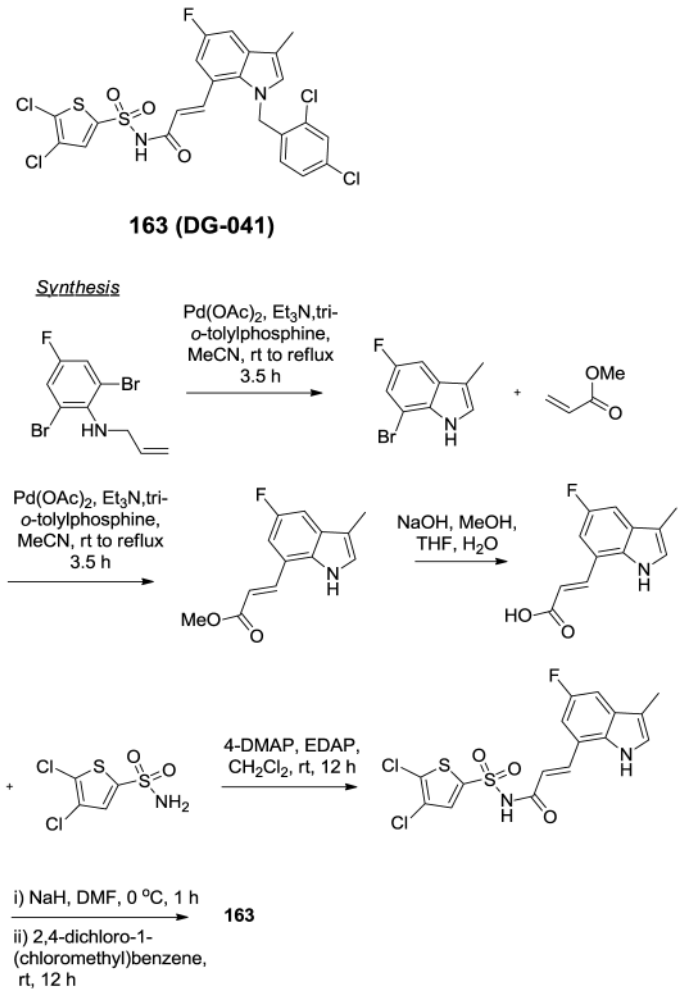


Figure 13.
Discovery of DG-041.[338]

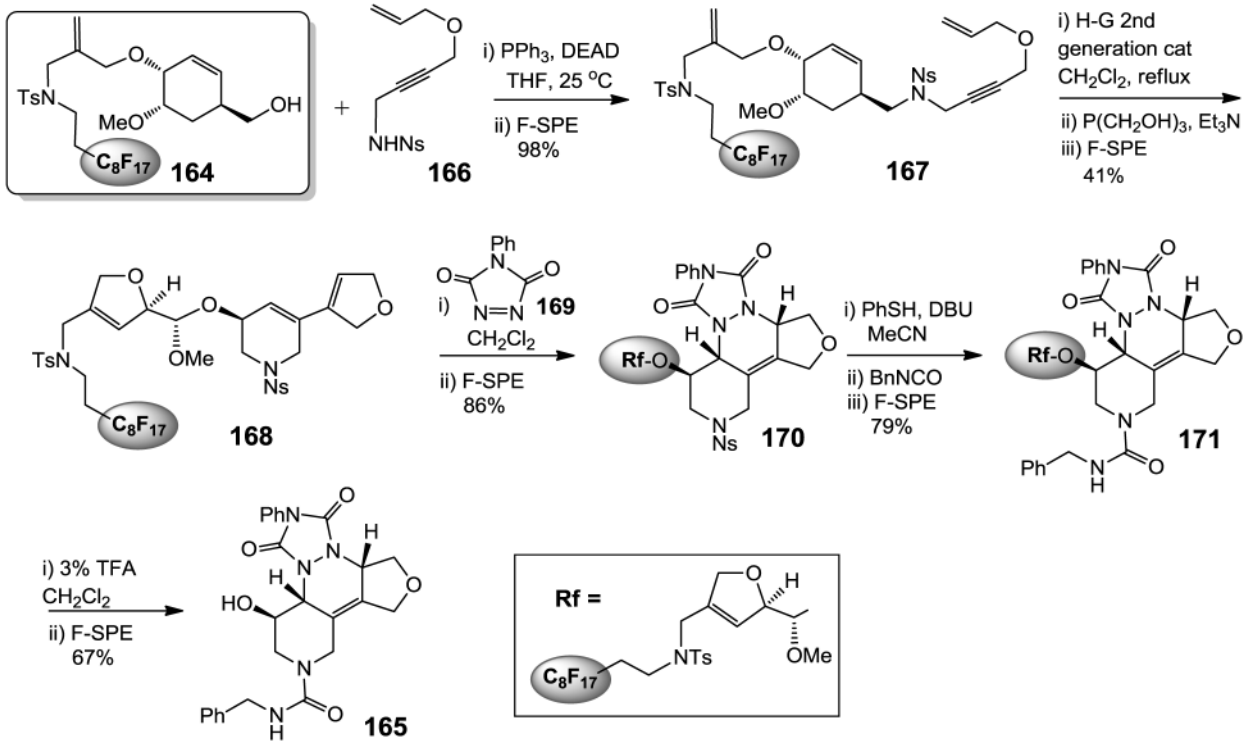


Figure 14.
Fluorous dihydropyran linker.[322]

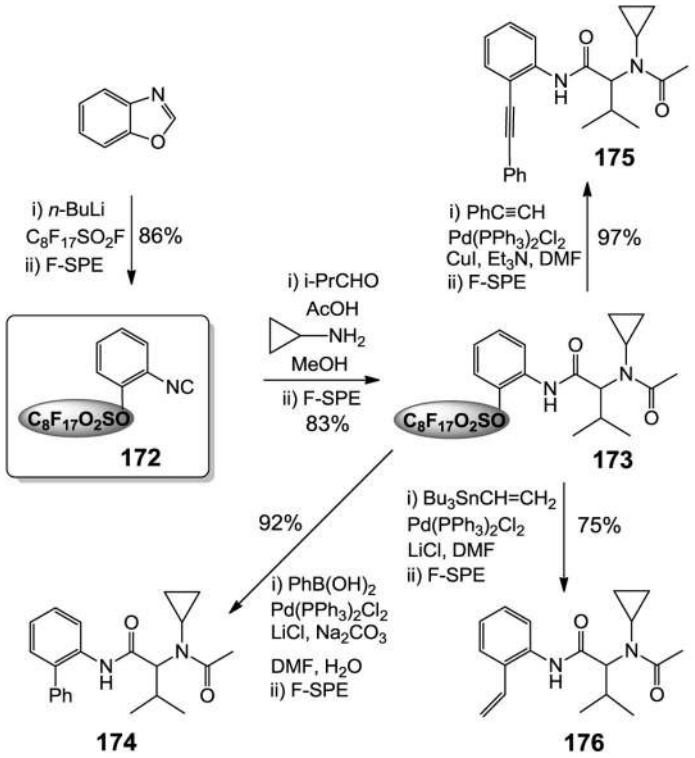


Figure 15.
Fluorous isonitrile linker.[327]

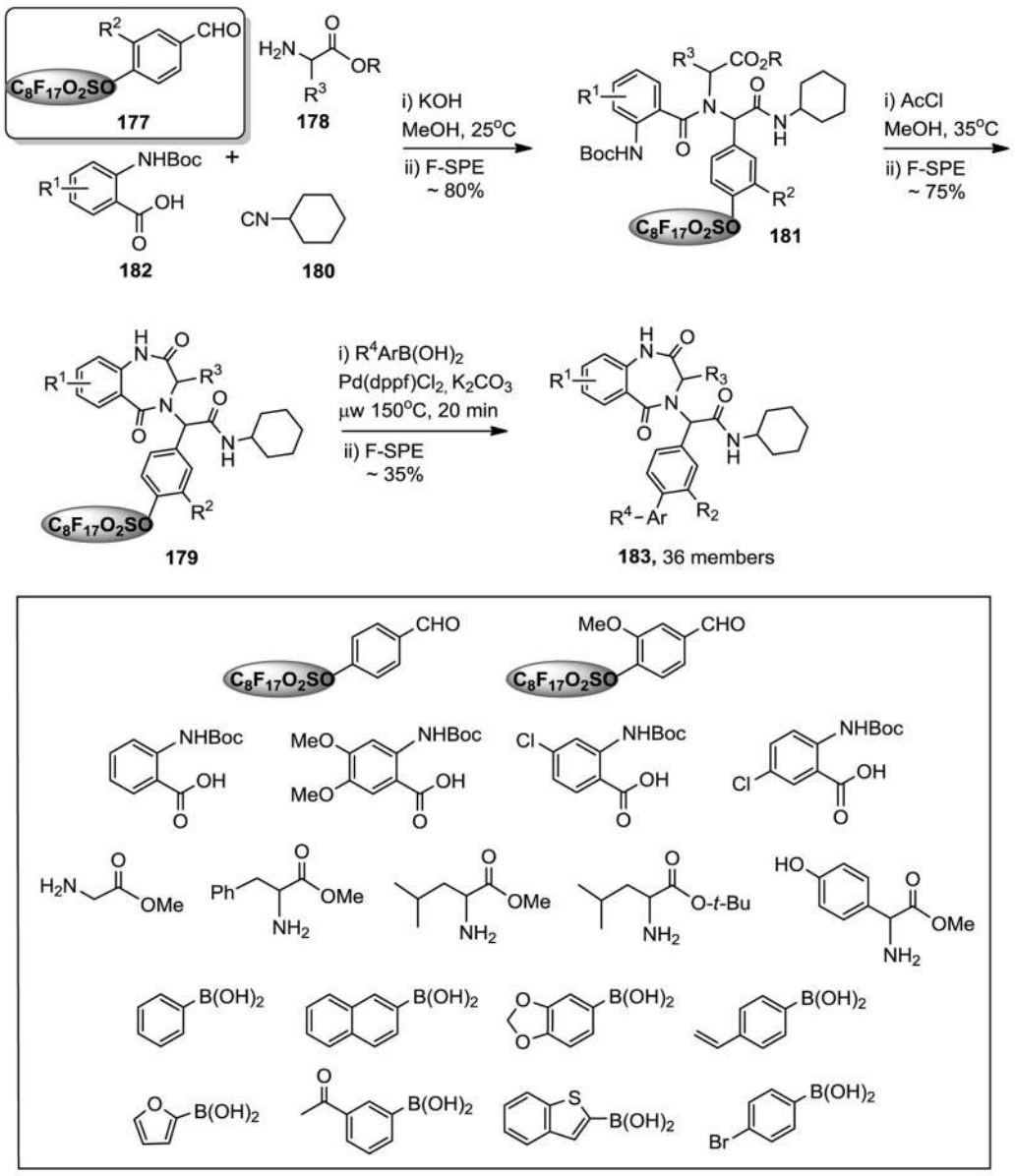


Figure 16.

Fluorous synthesis of 1,4-benzodiazepinedione library.[327]

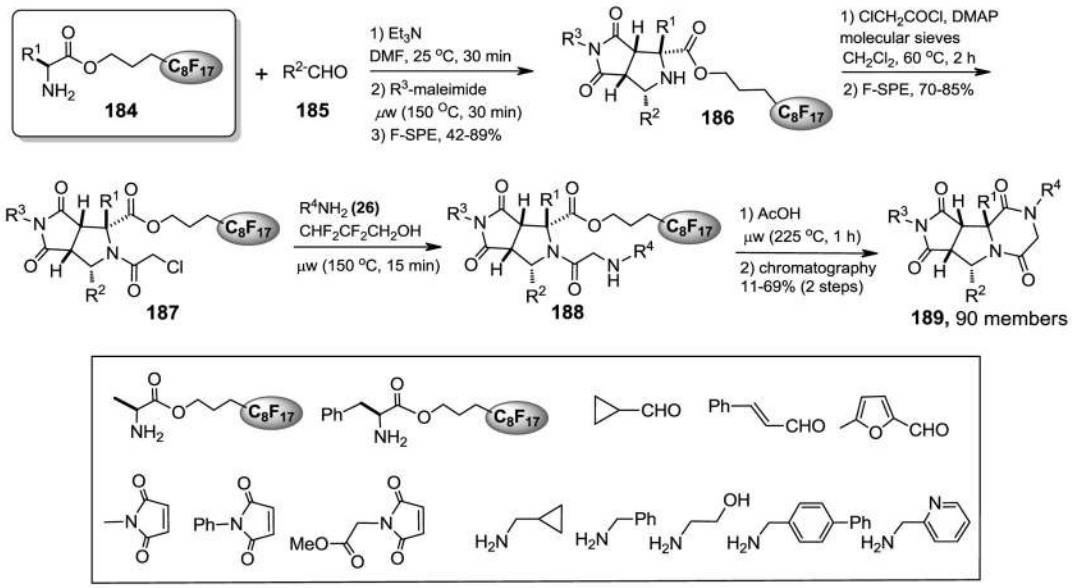


Figure 17.
Fluorous synthesis of tricyclic library.[328]

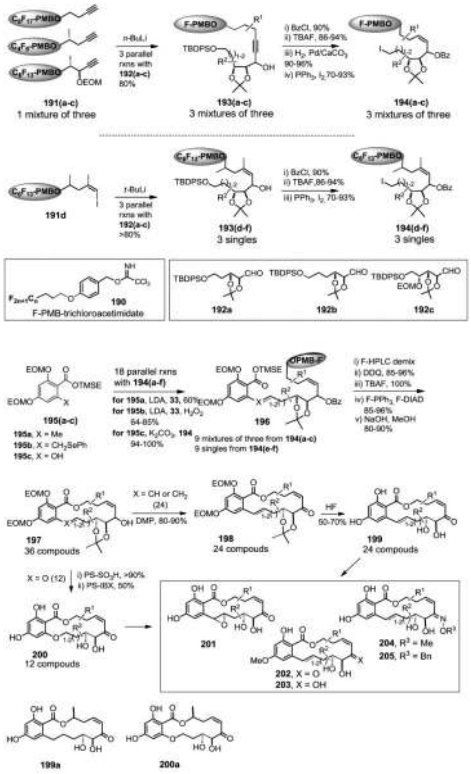
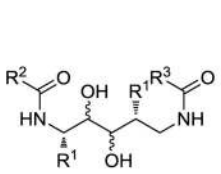
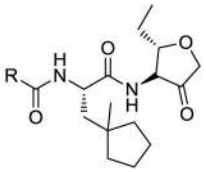
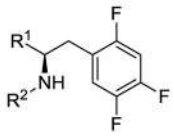
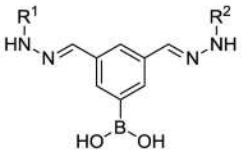
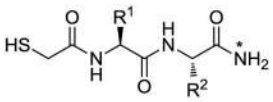


Figure 18.
Fluorous mixture synthesis of resorcylic acid lactone library.[329]

Table 1

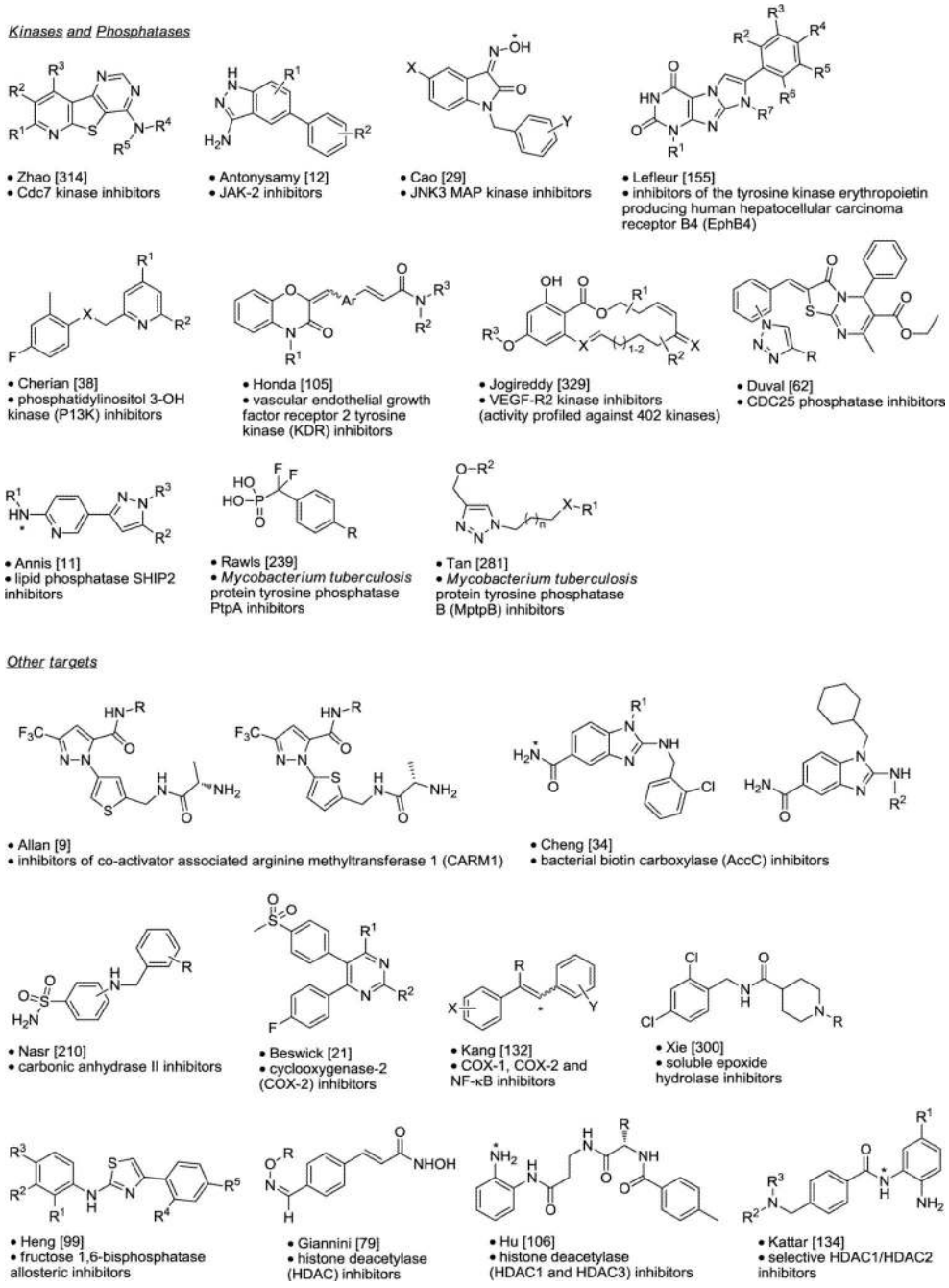
Chemical Libraries Targeting Proteases.

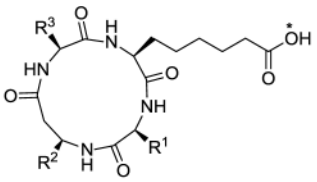
 • Shi [265] • HIV-1 aspartyl protease inhibitors	 • Ayesa [14] • cathepsin S cysteine protease inhibitors	 • Benbow [20] • dipeptidyl peptidase-IV (DPP-IV) serine protease inhibitors	 • Mugherli [206] • HCN NS3/4A serine protease
 • Cathcart [30] • pseudomonas elastase (metallo protease) inhibitors (Pseudolysin or LasB)			

[a] Asterisk is the point of attachment to resin.

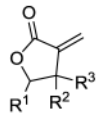
Table 2

Chemical Libraries Targeting Nonproteolytic Enzymes.

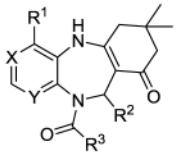




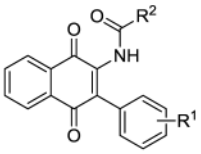
• Olsen [217]
• class II histone deacetylase (HDAC) inhibitors



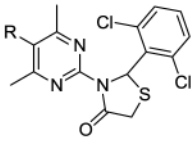
• Elford [70]
• *Haemophilus influenzae* homoserine transacetylase (HTA) inhibitors



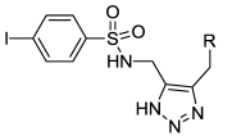
• Mcgowan [185]
• HCV NS5B polymerase inhibitors



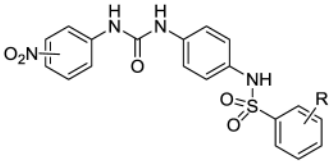
• Hadden [91]
• heat shock proteins (Hsp90) inhibitors



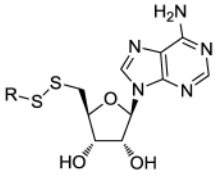
• Chen [33]
• HIV-1 reverse transcriptases inhibitors



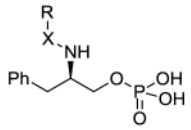
• Minond [197]
• VIM-2 ambler class B metallo -β-lactamase (MBL) inhibitors



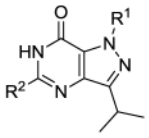
• Moro [202]
• nicotinamide adenine dinucleotide (NAD) synthetase inhibitors



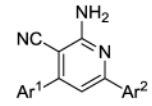
• Scott [258]
• *Mycobacterium tuberculosis* pantothenate synthetase inhibitors



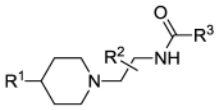
• Guo [90]
• human Pin 1 (Protein Interacting with NIMA) inhibitors



• DeNinno [52]
• phosphodiesterase 9 (PDE9) inhibitors



• Abadi [2]
• phosphodiesterase 3 inhibitors

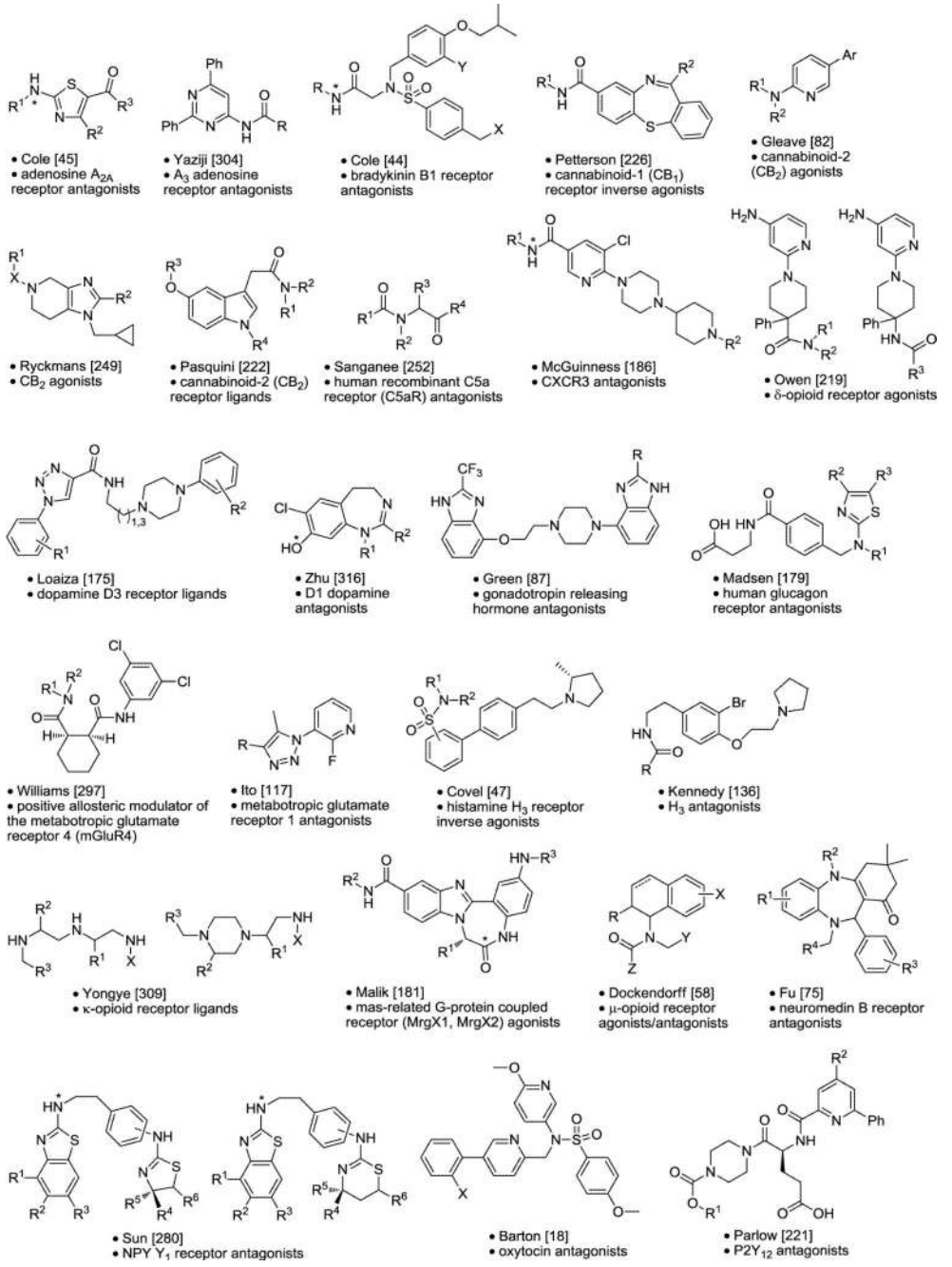


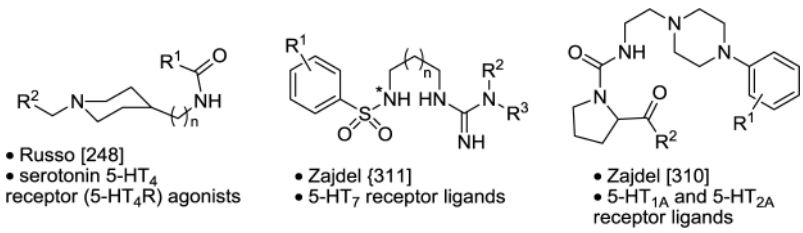
• Scott [259]
• isoform-selective phospholipase D inhibitors

[a] Asterisk is the point of attachment to resin.

Table 3

Chemical Libraries Targeting G-Protein Coupled Receptors.

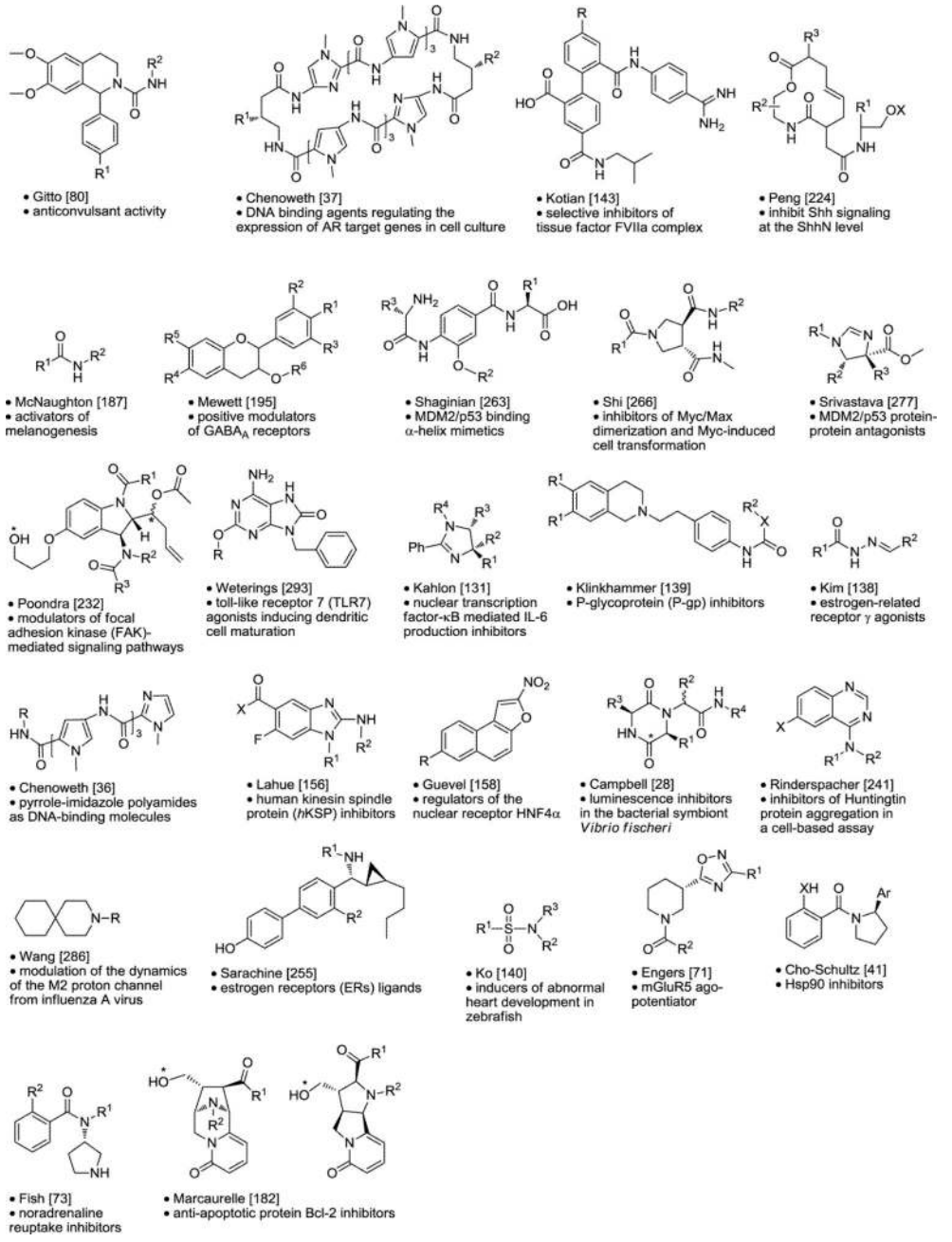




[a] Asterisk is the point of attachment to resin.

Table 4

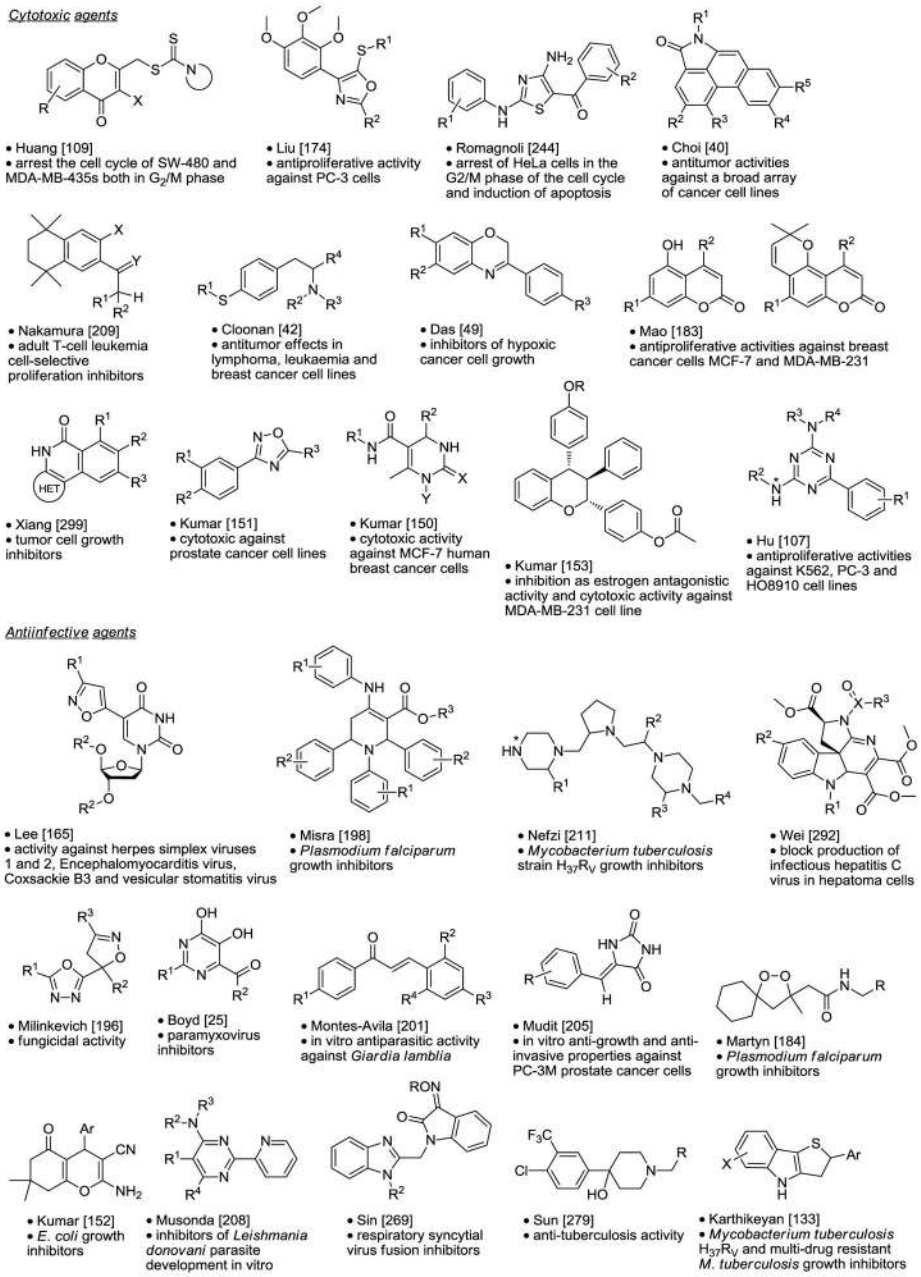
Chemical Libraries Targeting Non-G-Protein-Coupled Receptors.



[a] Asterisk is the point of attachment to resin.

Table 5

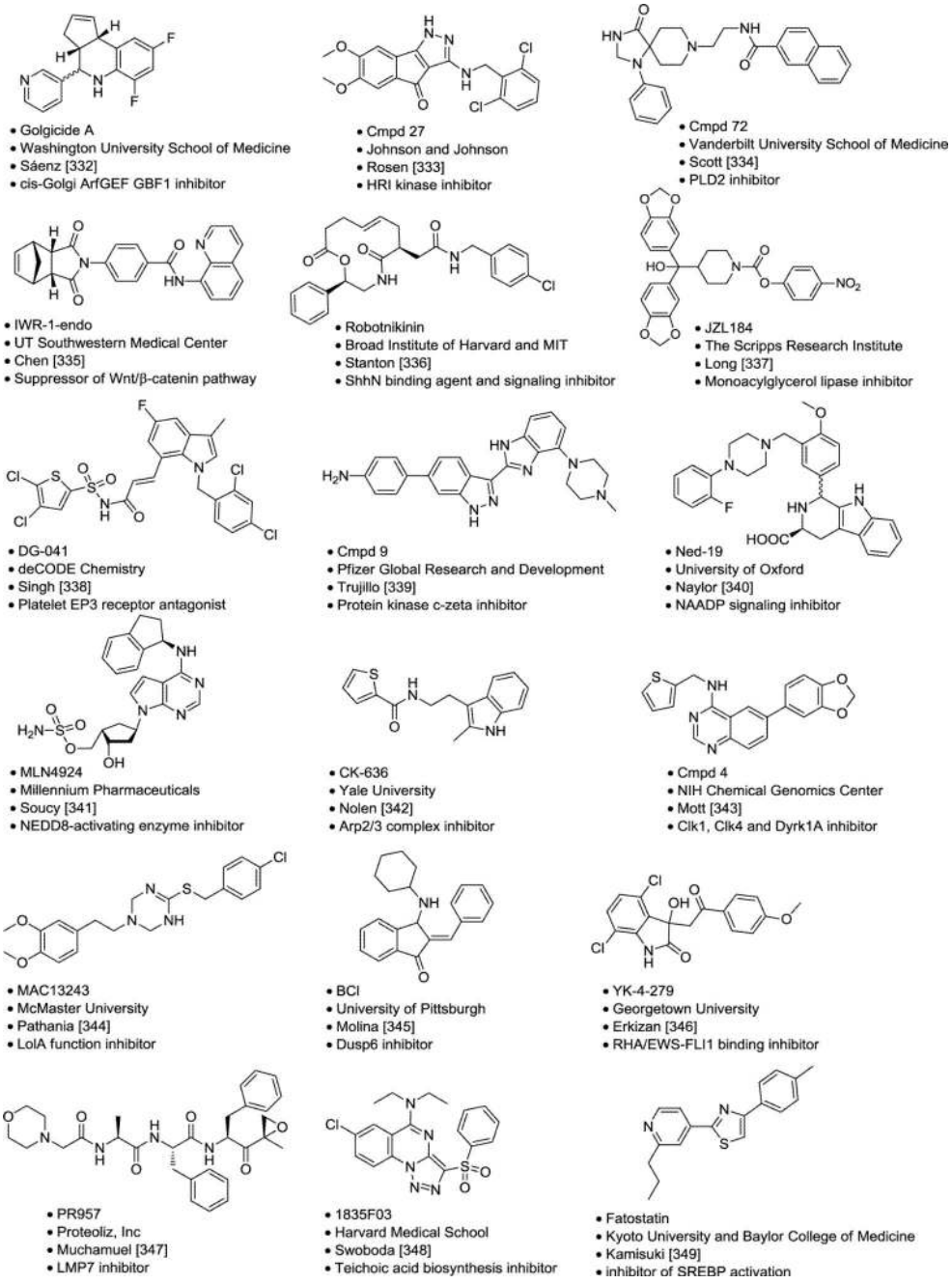
Chemical Libraries Yielding Cytotoxic and Antiinfective Agents.

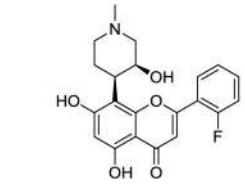


[a] Asterisk is the point of attachment to resin.

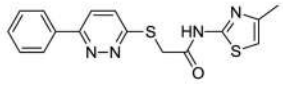
Table 6

Selected Molecular Probes.

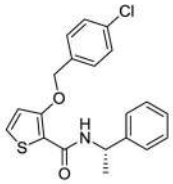




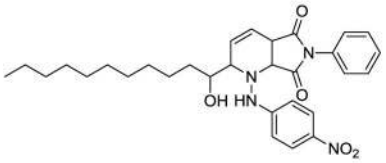
- Cmpd 12d
- University of Massachusetts Medical School
- Ali [350]
- selective P-TEFb inhibitor



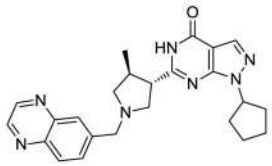
- Cmpd 6l
- Vanderbilt University School of Medicine
- Delpire [351]
- KCC2 inhibitor



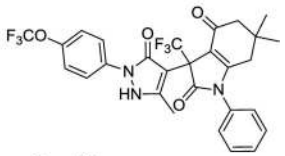
- AS1949490
- Astellas Pharma
- Suwa [352]
- SHIP2 inhibitor



- A12B4C3
- Cross Cancer Institute
- Freschauf [353]
- hPNKP inhibitor



- Cmpd 19
- Pfizer Global Research and Development
- Verhoest [354]
- PDE9A inhibitor

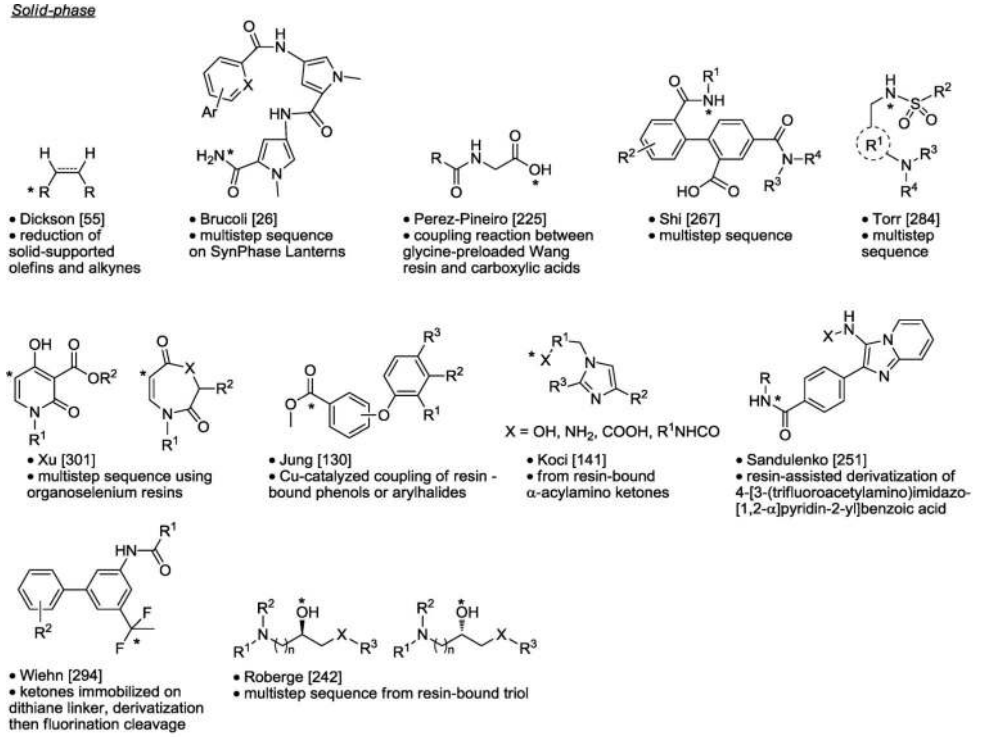


- Cmpd 37
- Tsukuba Research Institute
- Takahashi [355]
- ELOVL6 inhibitor

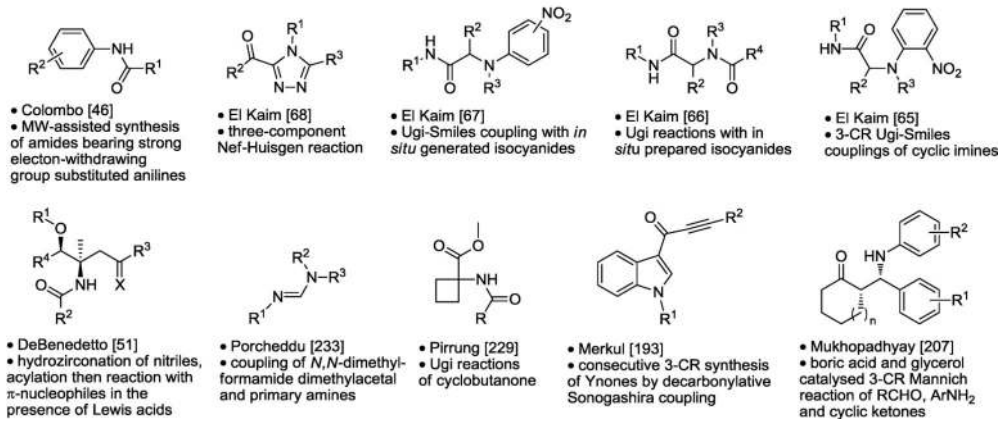
Table 7

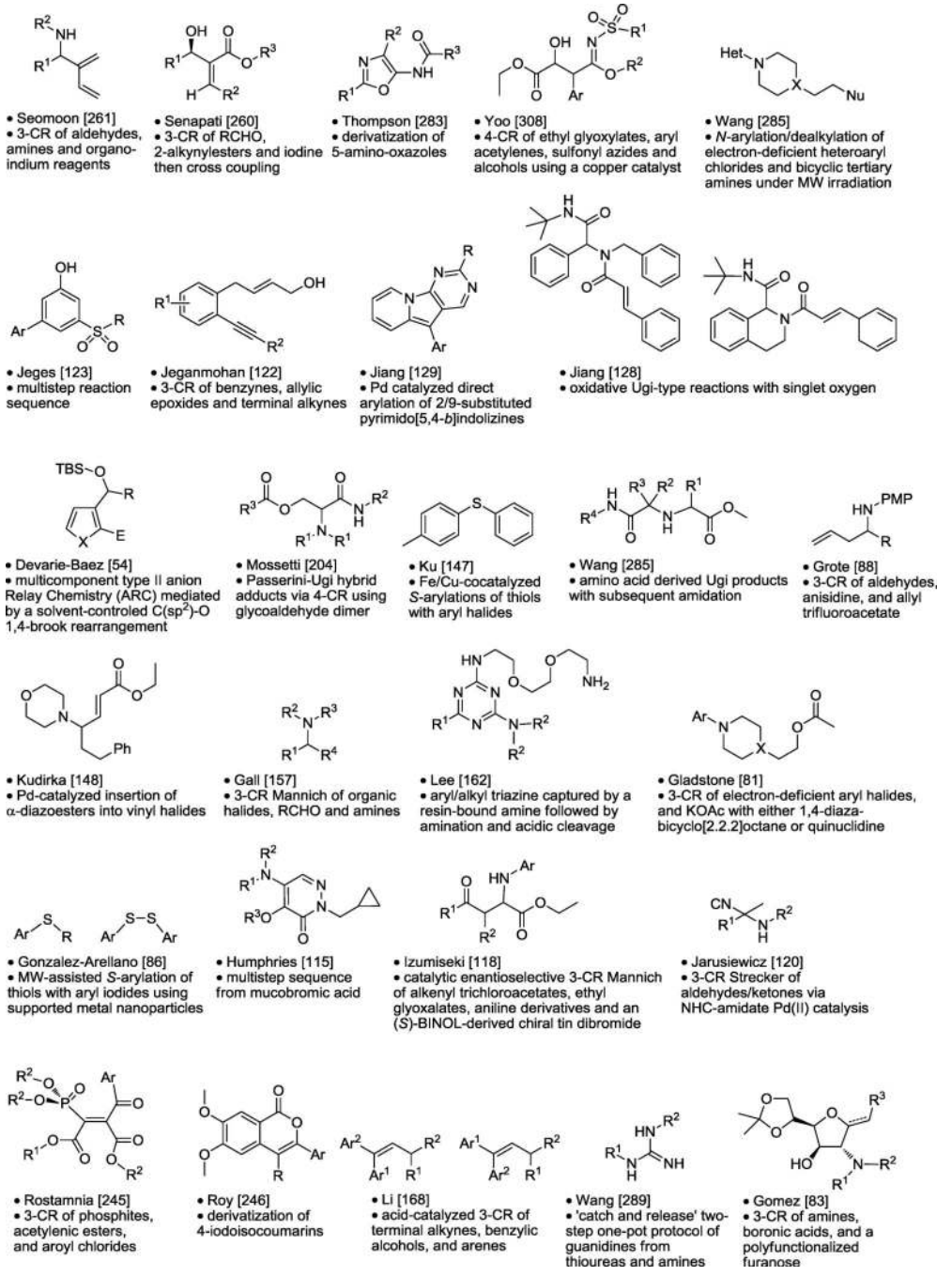
Scaffold Derivatization and Acyclic Synthesis.

Solid-phase



Solution-phase



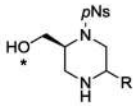


[a] Asterisk is the point of attachment to resin.

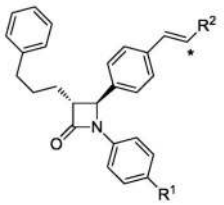
Table 8

Monocyclic Synthesis.

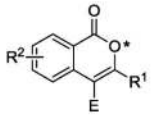
Solid-phase



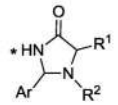
- Ottesen [218]
- MW-assisted ring-opening of resin-bound aziridine with 2-amino alcohols then intramolecular cyclization



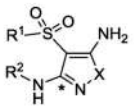
- Poeylaut-Palena [230]
- Staudinger reaction of imines immobilized on a metathesis-cleavable linker



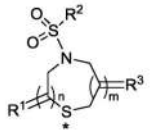
- Peuchmaur [227]
- Sonogashira cross-coupling between polymer-bound 2-bromobenzoates and terminal alkynes, then an electrophile-induced halocyclization



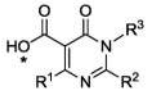
- Qin [238]
- MW-assisted condensation of an α-amino amide on solid support with aldehydes



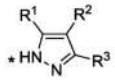
- Ma [176]
- 3-CR two step "catch and release" from resin-bound sulfonyl intermediate



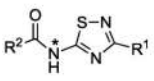
- Saruta [256]
- intramolecular alkylation of sulfides followed by release of desired products from generated sulfonium salts



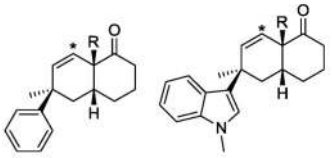
- Hamper [92]
- condensation of resin-bound methylenemalonate with amidines



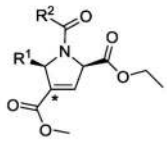
- Harju [98]
- 1,3-dipolar cyclo-addition of resin-bound sydnones with alkynes



- Ryu [250]
- multistep sequence from resin-bound carboxamidine thioureas

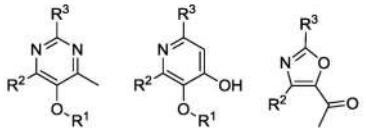


- Yoshida [309]
- tetrazole-based traceless linker for Diels-Alder on solid support cleaved by Pd(0), Cu(I) and Ag(I) assisted nucleophilic displacements

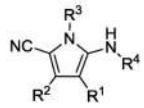


- Huang [112]
- 1,3-dipolar cycloaddition of resin-bound methyl 2-seleno acrylate with azomethine ylides

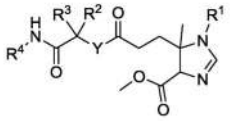
Solution-phase



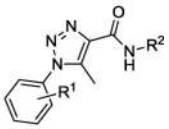
- Lechel [159]
- from alkoxy-substituted enamides starting from lithiated alkoxyallenes, nitriles and carboxylic acids



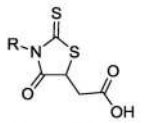
- Fontaine [74]
- consecutive [4+1] reaction cycloaddition of α-imino-nitriles with isocyanides



- Elders [69]
- one-pot reaction of up-to eight components by the union of multicomponent reactions

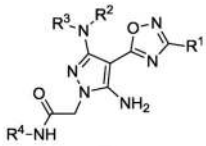


- Pokhodylo [231]
- 3-CR of azides, amines, and diketene

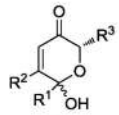


- Alizadeh [8]
- 3-CR of primary amine, carbon disulfide and fumaryl chloride

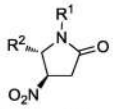
- Bagley [16]
- MW-assisted barium manganate mediated oxidation of diols



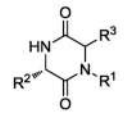
- Borisov [24]
- multistep sequence



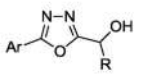
- Cheng [35]
- catalytic asymmetric alkylation of 2-furfurals yields enantio-enriched furyl zinc alkoxides then NBS and water



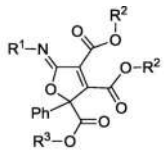
- Pelletier [223]
- nitro-Mannich/ lactamization cascade of nitroester, amine, and aldehyde



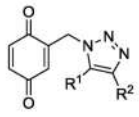
• Rhoden [240]
• one-pot Ugi-4-CR/ deprotection and activation/cyclization



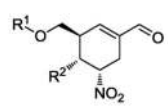
• Adib [3]
• 3-CR of *N*-isocyanimino triphenylphosphorane, RCHO, and ArCOOH



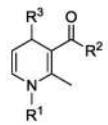
• Esmaeili [7]
• 3-CR of isocyanides, dialkyl acetylenedicarboxylates, and α-ketolactones



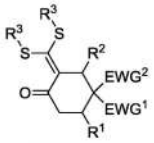
• Algi [6]
• 1,3-dipolarcyclo-addition then oxidation



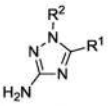
• Zhang [312]
• quadruple domino reaction initiated by oxa-Michael addition of alcohols to acrolein



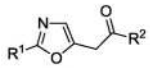
• Maiti [180]
• 4-CR of RNH_2, β-keto esters/ thioesters, α,β-unsaturated aldehydes and alcohols



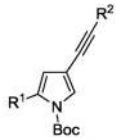
• Ma [177]
• 3-CR of α-alkenoyl ketene *S*,*S*-acetals, aldehydes, and active methylenes



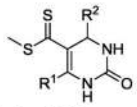
• Meng [189]
• MW-assisted synthesis from carboxylic acids and hydrazines



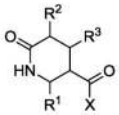
• Merkul [192]
• consecutive 3-CR sequence from propargylamine and acid chlorides then amidation and cross-coupling



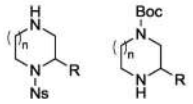
• Merkul [191]
• 3-CR synthesis of *N*-Boc-4-iodopyrroles and sequential one-pot alkynylation



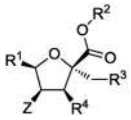
• Mukherjee [270]
• 3-CR Biginelli reaction involving $SnCl_2$-catalyzed cyclocondensation of β-oxodithioesters with aldehydes and urea



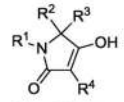
• Zhu [315]
• diene and the cycloaddition in a 4-CR of RCHO, NH_3, acyl chloride and dienophile



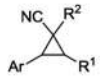
• Crestey [48]
• aminolysis of aziridines with amino alcohols and subsequent Fukuyama-Mitsunobu cyclization



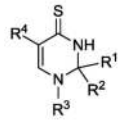
• DeAngelis [50]
• Rh-catalyzed 3-CR of aldehydes, α-alkyl-α-diazo-esters, and dipolarophiles



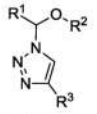
• Spatz [273]
• Ugi-Dieckmann reaction



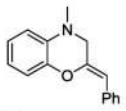
• Wang [287]
• 3-CR of α-halogenated methylene compounds, aromatic aldehydes, and acetonitrile derivatives



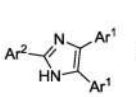
• Wang [290]
• multicomponent coupling of terminal alkynes, elemental sulfur, and carbodiimides



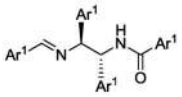
• Yadav [303]
• $Cu(OTf)_2$/Cu-catalyzed 4-CR of R^1CHO, R^2CH_2OH, R^3CCH, $TMSN_3$

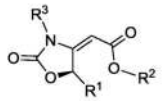


• Xu 302]
• 3-CR of 2-(methylamino)-phenyl-4-methylbenzene-sulfonate, $(CH_2O)_n$, PhCCH then KOH

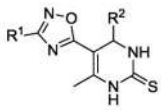


• Jiang [127]
• 4-and 6-component domino reactions

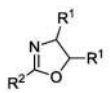




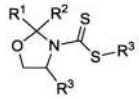
• Kojima [142]
• diethylzinc-mediated asymmetric alkynylation of aldehydes with propiolates in the presence of a chiral ligand; followed by treatment with isocyanates



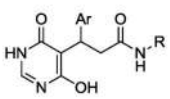
• Kharchenko [137]
• Biginelli-type 3-CR

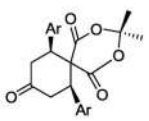


• Kempe [135]
• automated synthesis from nitriles with 2-aminoethanols

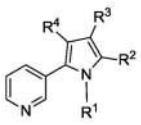


• Han [93]
• 4-CR of aminoethanols, ketones, carbon disulfide, and halides

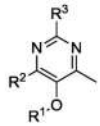




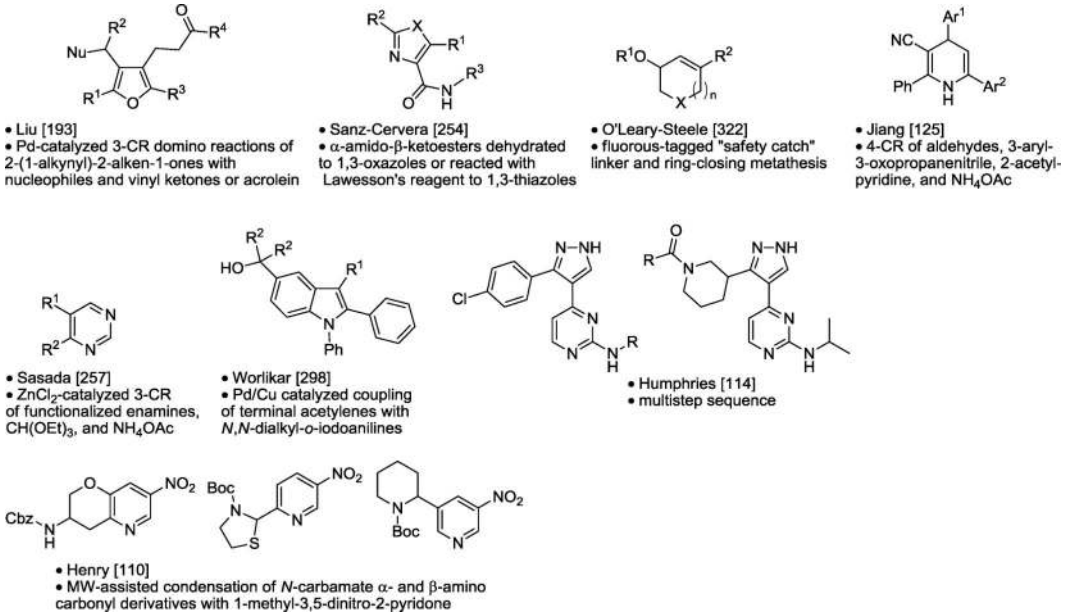
• Hao [96]
• MW-assisted reaction of arylidene-Meldrum's acid, 6-hydroxypyrimidin-4(3*H*)-one, and amines



• Harju [97]
• multistep sequence from *N*-acylated amino acids



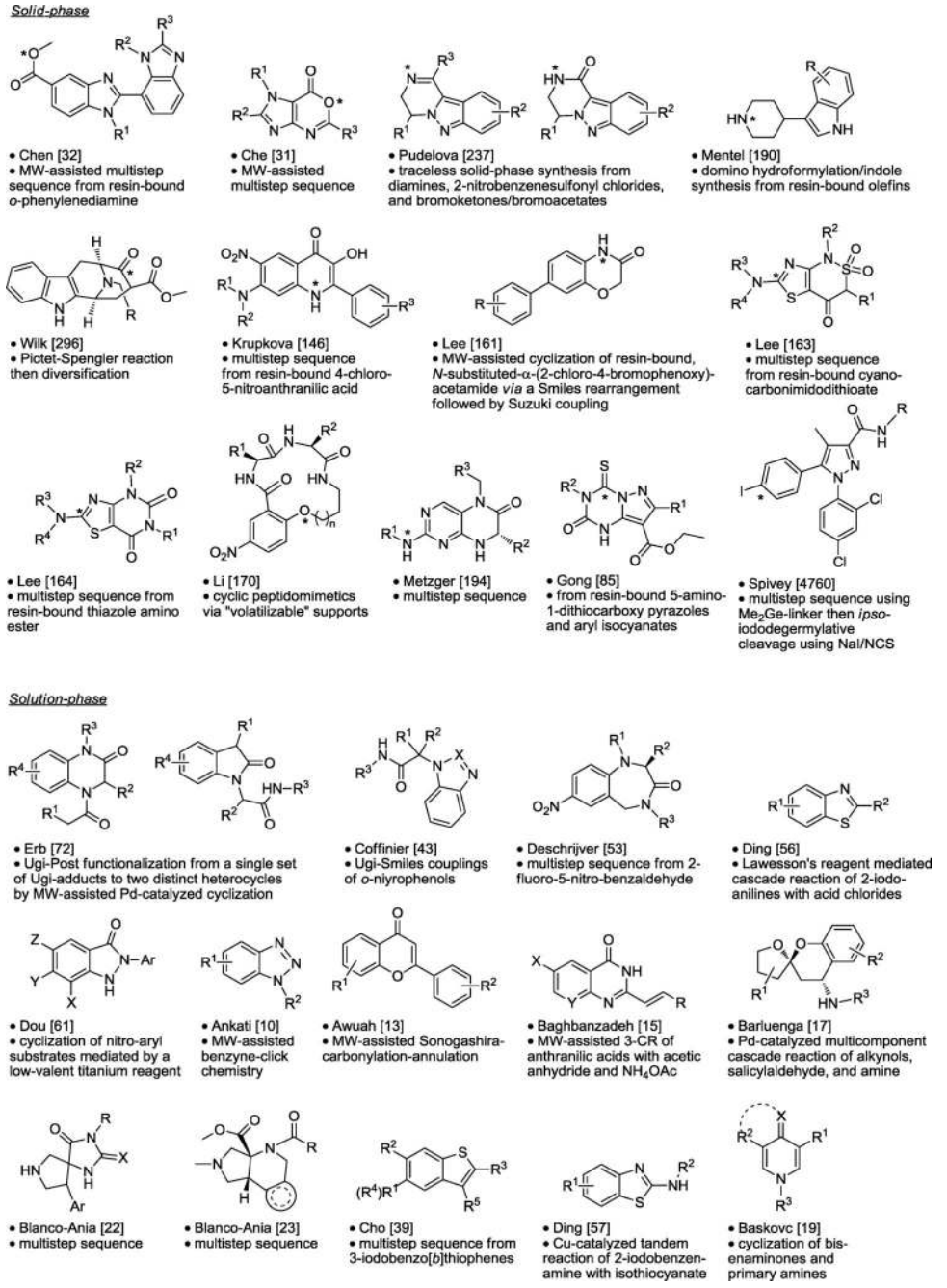
• Lechel [160]
• 3-CR of lithiated alkoxyallenes, nitriles, and carboxylic acids then condensation with ammonium salts

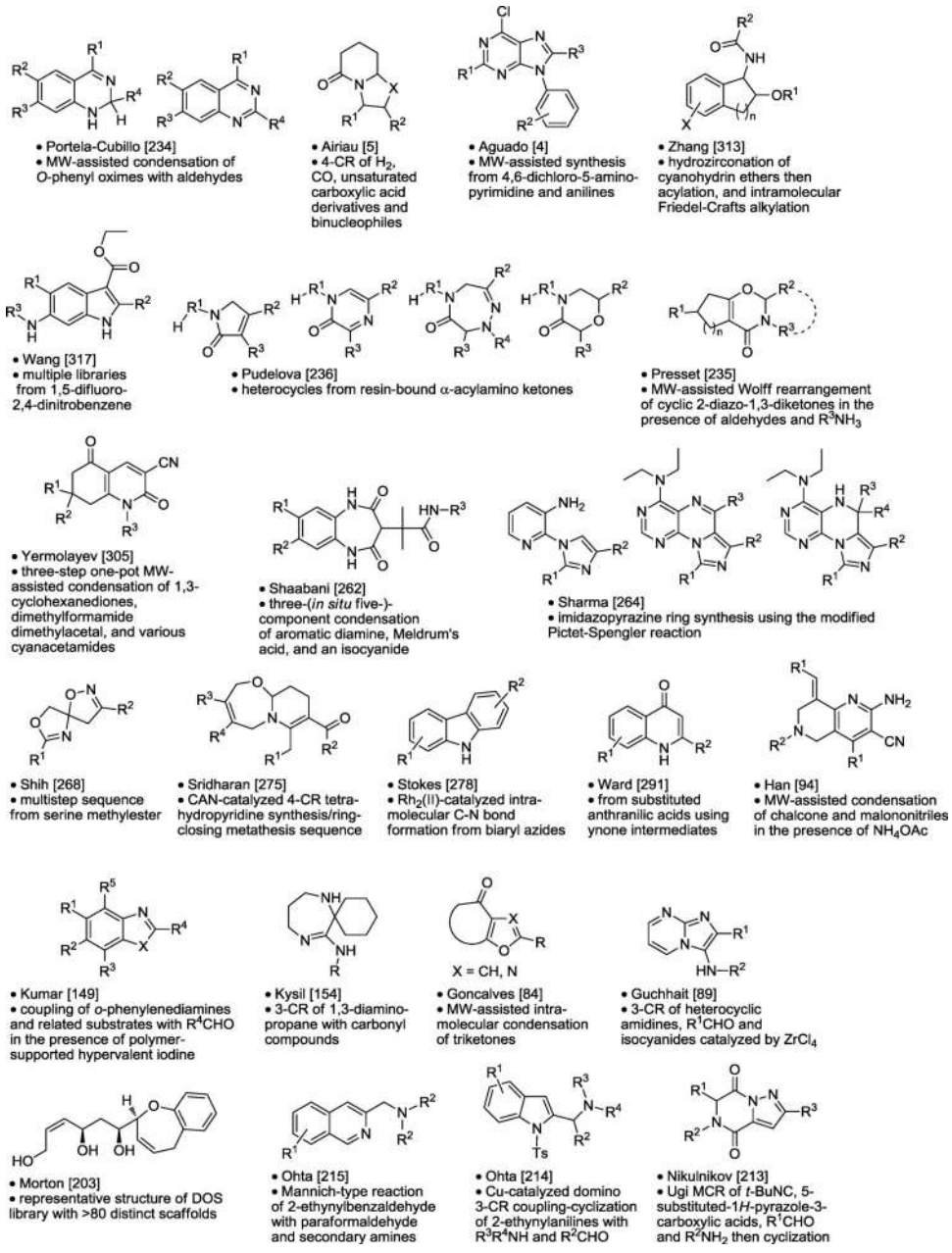


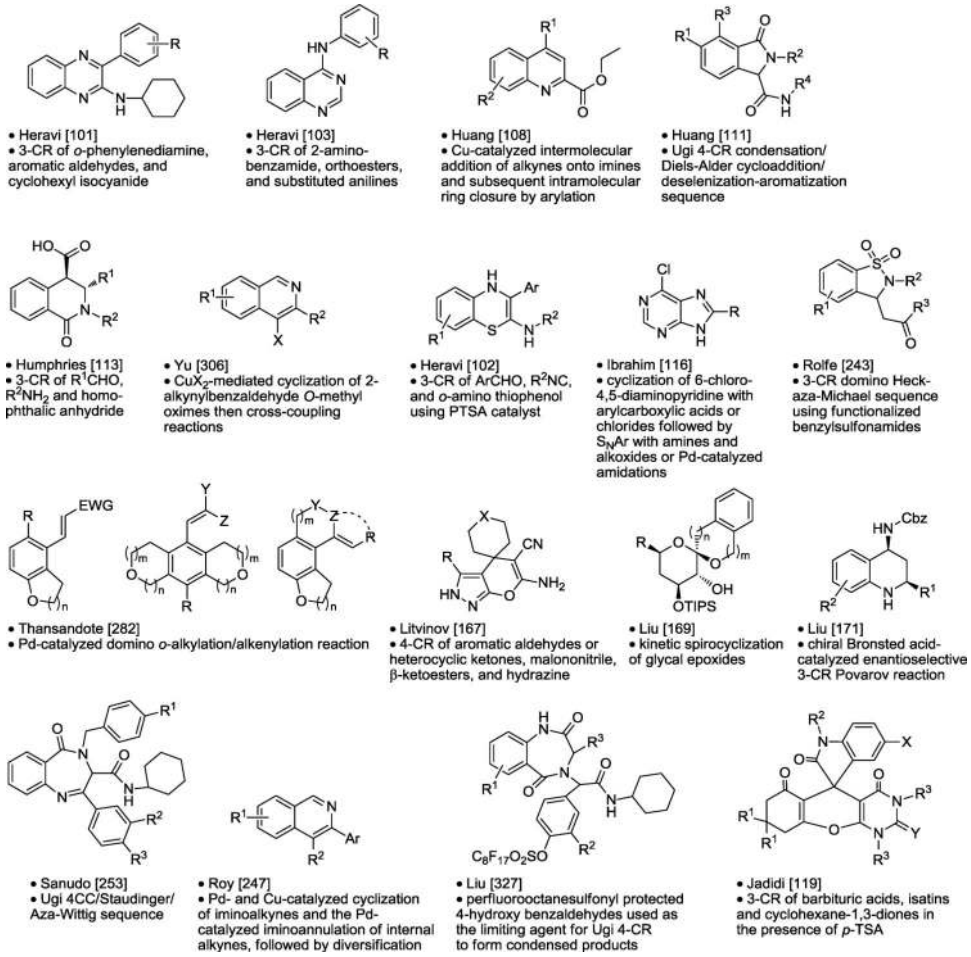
[a] Asterisk is the point of attachment to resin.

Table 9

Bicyclic and Spirocyclic Synthesis.





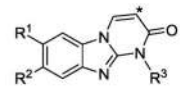


[a] Asterisk is the point of attachment to resin.

Table 10

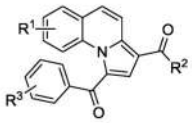
Polycyclic and Macrocyclic Synthesis.

Solid-phase

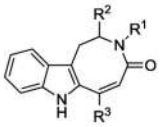


• Huang [110]
• condensation of resin-bound 3-(2-aminophenylamino)-2-seleno-ester with isothiocyanates and α-amino-acids

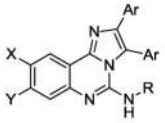
Solution-phase



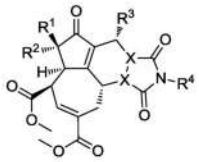
• Georgeseu [76]
• 3-CR of quinolines, bromoacetophenones, electron-deficient alkynes



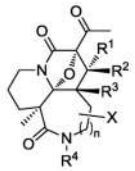
• Donets [59]
• MW-assisted $Hg(OTf)_2$ catalyzed intramolecular alkyne carbocyclization



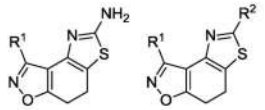
• Dou [60]
• cyclization of thiourea intermediates mediated by low-valent titanium



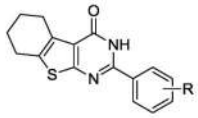
• Brummond [29]
• tandem cyclopropanation/Cope rearrangement followed by a Diels-Alder sequence



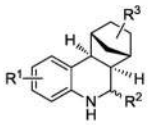
• Mizoguchi [199]
• multistep sequence from Ugi products



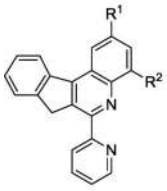
• El-Badri [64]
• bromination of dihydrobenzo[*d*]isoxazol-4-(5*H*)-one followed by cyclocondensation with thiourea in the presence of DDQ or cyclocondensation with thioamides



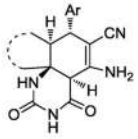
• Dzhavakhishvili [63]
• reduction of N2-substituted Gewald's amides with aromatic aldehydes



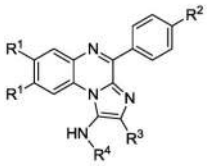
• Smith [271]
• Lewis acid catalyzed 3-CR hetero-Diels-Alder of *N*-aryl-imines with strained norbornene-derived dienophiles



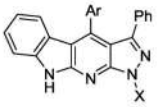
• Kouznetsov [144]
• 3-CR imino Diels-Alder reaction



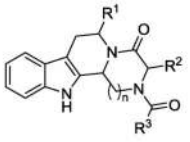
• Jiang [126]
• 4-CR domino reaction of ArCHO, ketones and cyanoamides



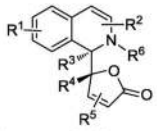
• Krasavin [145]
• two tandem isocyanide-based multicomponent reactions



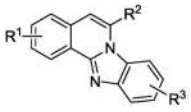
• Ghahremanzadeh [77]
• 4-CR of indolin-2-one, 3-oxo-3-phenylpropanenitrile, and hydrazines and aldehydes



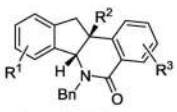
• Liu [172]
• Ugi and Pictet-Spengler reactions

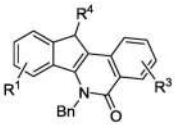


• Hermange [104]
• diastereoselective 3-CR vinylogous Mannich between isoquinolines, acyl/sulfonyl chlorides, and silyloxyfurans

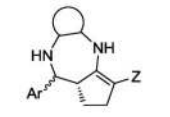


• Okamoto [216]
• MW-assisted condensation of 2-bromoarylaldehydes, terminal alkynes, and 1,2-phenylenediamines

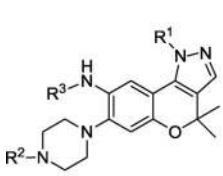




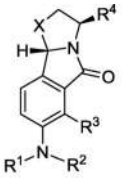
• Jayanth [121]
• Cu-catalyzed coupling of imines, vinylstannanes, or alkynes and *o*-bromoaroyl chlorides followed by Pd(0)-catalyzed annulations



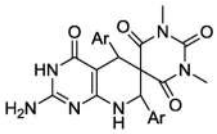
• Sotoca [272]
• cyclodehydrative 3-CR with 1,3-dicarbonyls



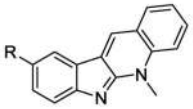
• Park [220]
• regioselective condensation of β-keto aldehydes with hydrazines



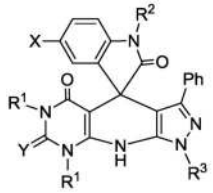
• Medimagh [188]
• 3-CR cascade process via formation of a transient imine followed by Diels-Alder cycloaddition, oxazolidine ring closure, and lactamization



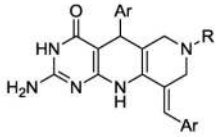
• Jiang [124]
• MW-assisted domino synthesis from ArCHO and barbituric acids



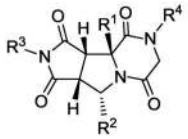
• Miller [330]
• reaction of fluorous thiols with glyoxamides via Pummerer-type cyclization



• Ghahremanzadeh [78]
• 3-CR of barbituric acids, 1*H*-pyrazol-5-amines and isatins



• Han [95]
• MW-assisted condensation of 3,5-dibenzylidenepiperidin-4-ones with 2,6-diaminopyridin-4(3*H*)-one and other enamine-like substrates

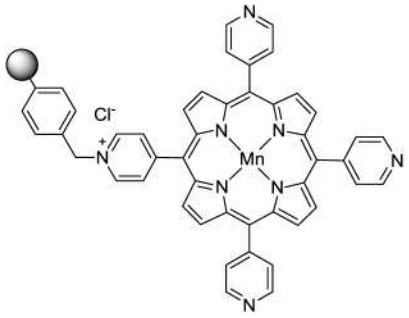


• Werner [328]
• [3+2] cycloaddition of fluorous amino esters, aldehydes, and maleimides

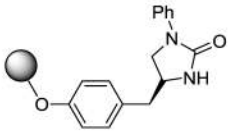
[a] Asterisk is the point of attachment to resin.

Table 11

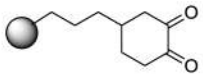
Polymer-Supported Reagents, Scavengers, and Linkers.



• Moghadam [200]
• reusable catalyst for biomimetic oxidative decarboxylation of carboxylic acids with $NaIO_4$



• Nguyen [212]
• 2-imidazolidinone chiral auxiliary for solid phase asymmetric alkylation reactions



• Lence [166]
• selective resin for immobilizing *trans*-diequatorial-1,2-diols

• Roberge [242]
• linker for immobilizing alcohols

Table 12

Fluorous Catalysts, Reagents, Scavengers, Linkers and Library Synthesis.