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Concepts and principles of photodynamic therapy as an alternative antifungal discovery platform

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Opportunistic fungal pathogens may cause superficial or serious invasive infections, especially in immunocompromised and debilitated patients. Invasive mycoses represent an exponentially growing threat for human health due to a combination of slow diagnosis and the existence of relatively few classes of available and effective antifungal drugs. Therefore systemic fungal infections result in high attributable mortality. There is an urgent need to pursue and deploy novel and effective alternative antifungal countermeasures. Photodynamic therapy (PDT) was established as a successful modality for malignancies and age-related macular degeneration but photodynamic inactivation has only recently been intensively investigated as an alternative antimicrobial discovery and development platform. The concept of photodynamic inactivation requires microbial exposure to either exogenous or endogenous photosensitizer molecules, followed by visible light energy, typically wavelengths in the red/near infrared region that cause the excitation of the photosensitizers resulting in the production of singlet oxygen and other reactive oxygen species that react with intracellular components, and consequently produce cell inactivation and death. Antifungal PDT is an area of increasing interest, as research is advancing (i) to identify the photochemical and photophysical mechanisms involved in photoinactivation; (ii) to develop potent and clinically compatible photosensitizers; (iii) to understand how photoinactivation is affected by key microbial phenotypic elements multidrug resistance and efflux, virulence and pathogenesis determinants, and formation of biofilms; (iv) to explore novel photosensitizer delivery platforms; and (v) to identify photoinactivation applications beyond the clinical setting such as environmental disinfectants.

Keywords: photodynamic inactivation, photodynamic therapy, photosensitizer, reactive oxygen species, permeability barrier, multidrug efflux systems, biofilms, clinical applications

INTRODUCTION

FUNGAL INFECTIONS AND COUNTERMEASURES

Fungi are common causative agents of diseases in both the immune competent as well as the immune compromised patient

Abbreviations: ABC, ATP binding cassette; ALA, 5-aminolevulinic acid; APDI, antimicrobial photodynamic inactivation; BCVA, best-corrected visual acuity; BSI, blood stream infections; CDR1, *Candida* drug resistance 1; CFU, colony forming unit; CMD, choroidal major diameter; CNV, choroidal neovascularization; EPI, efflux pump inhibitor; ETC, electron transport chain; EtNBSe, selenium analog of benzophenothiazinium salt; HOMO, highest occupied molecular orbital; IA, invasive aspergillosis; IC, invasive candidiasis; IL-1, interleukin-1; LUMO, lowest occupied molecular orbital; MAL, methyl 5-aminolevulinic acid; MB, methylene blue; MDR1, multidrug resistance 1; MFS, major facilitator superfamily; MIC, minimal inhibitory concentration; NMB, new methylene blue; PDI,

populations. The diseases include cutaneous, subcutaneous, mucosal invasion, and can be found as blood stream infections (BSI) that are life-threatening (Gavalda et al., 2005; Sobel, 2007; Caston-Osorio et al., 2008; Ameen, 2009; Horn et al., 2009; Neofytos et al., 2009; Pappas et al., 2009). Dermatophytosis is probably the most prevalent of all fungal diseases but also the least studied in regard to host–fungus interactions (Vermout et al., 2008).

photodynamic inactivation; PDR, pleiotropic drug resistance; PDT, photodynamic therapy; PEI-ce6, polyethyleneimine-chlorin(e6); PMMA, poly-methyl methacrylate; PS, Photosensitizer; RB, rose bengal; RD, respiratory deficiency; RND, resistant modulation (cell) division; RNS, reactive nitrogen species; ROS, reactive oxygen species; SOR1, singlet oxygen resistance 1; TBO, toluidine blue.

Fungal diseases are endemic in certain parts of the world and include blastomycosis, chromoblastomycosis, coccidioidomycosis, histoplasmosis, paracoccidioidomycosis, penicilliosis, or pandemic including invasive aspergillosis (IA), invasive candidiasis (IC), cryptococcosis, dermatophytosis, fusariosis, and mucormycosis. Very likely, the greatest threat to life results from those pathogens that cause BSI, yet with IA and IC diagnosis is not easy and often these diseases are treated empirically when blood cultures are negative for bacterial pathogens. In the case of IC, there can be numerous risk factors that, for the most part, are non-specific. *Candida* spp. are the third leading cause and are associated with the highest mortality of catheter-related infections (Crump and Collignon, 2000). Although prevention of IC using azole prophylaxis can be effective in selected high-risk patient populations, selection for invasive infection by resistant non-*albicans* *Candida* species or molds is a potentially devastating consequence. Despite improvements in antifungal therapy, the mortality rate due to *Candida* BSI has improved little over the last two decades, remaining high at 15–49% (Gudlaugsson et al., 2003). A BSI episode significantly increases cost of care. In one analysis, the estimated cost of an IC BSI episode was \$34,123 per Medicare patient and \$44,536 per private insurance patient (1997, US\$), with an overall economic burden of \$2 billion dollars annually in the USA (Rentz et al., 1998).

The increased infection of immunocompromised hosts is dramatically illustrated by *Cryptococcus neoformans*, a pathogen that rose to prominence as the causative agent of cryptococcosis, which is a life-threatening disease that has emerged in parallel with the HIV/AIDS epidemic. Cryptococcosis results from inhalation of fungal cells with subsequent lung infection and pneumonia. In the absence of an effective immune response, the fungus can disseminate to the brain to cause meningoencephalitis, the symptoms of which include headache, fever, visual problems, and an altered mental state (Brizendine and Pappas, 2010). *C. neoformans* causes an estimated one million cases of meningoencephalitis globally per year in patients with AIDS, leading to approximately 625,000 deaths (Park et al., 2009). The bulk of this disease incidence is in sub-Saharan Africa, where fatal cases of cryptococcosis may exceed deaths from tuberculosis in some areas (Park et al., 2009). The importance of immune suppression in predisposition to infection has been challenged by the recent discovery of specialized interactions between the fungus and its mammalian hosts, and by the emergence of the related species *Cryptococcus gattii* as a primary pathogen of immunocompetent populations (D'Souza et al., 2011). A set of recently discovered cryptococcal pathogenesis features reveal the fungal adaptation to the mammalian environment (Alanio et al., 2011). These features include not only remarkably sophisticated interactions with phagocytic cells to promote intracellular survival, dissemination to the central nervous system, and escape (Alanio et al., 2011; Kronstad et al., 2011), but also surprising morphological and genomic adaptations such as the formation of polyploid giant cells in the lung (Fuchs et al., 2010).

The state of the art in antifungal drug discovery includes only a few new compounds which offer some hope for eradication of fungal diseases (Ostrosky-Zeichner et al., 2010). Specifically: (1) the echinocandins act against a specific component of fungal pathogens. As such, their safety profile is quite good, unlike

triazoles that are notorious for causing drug–drug interactions and toxicity; (2) the formulation of amphotericin B was changed to achieve better absorption as this drug is now available as a lipid formulation encapsulation. This modification has reduced toxicity due to off-target effects on host cells but has an increased cost; (3) enhanced activity has been observed with two of the newer triazoles, posaconazole, and voriconazole; (4) new triazoles are in development (albaconazole and ravuconazole); (5) echinocandins are slow in development and ineffective against *C. neoformans*. While β -1,3-glucan is present in the cell wall of this pathogen, caspofungin may have reduced activity against the β -1,3-glucan synthase (Feldmesser et al., 2000); (6) drug resistance to triazoles is a common feature of several species of *Candida*; an increase in resistance to echinocandins is frequently reported; and, importantly, (7) the impressive number of references to resistance developing to antifungals (Pfaller et al., 2010) makes the development of new antifungal countermeasures a necessity. One promising antifungal modality is the light-based technology of photodynamic therapy (PDT). This review summarizes the progress in antifungal PDT attempting to formulate the principles as well as the concepts for its successful implementation.

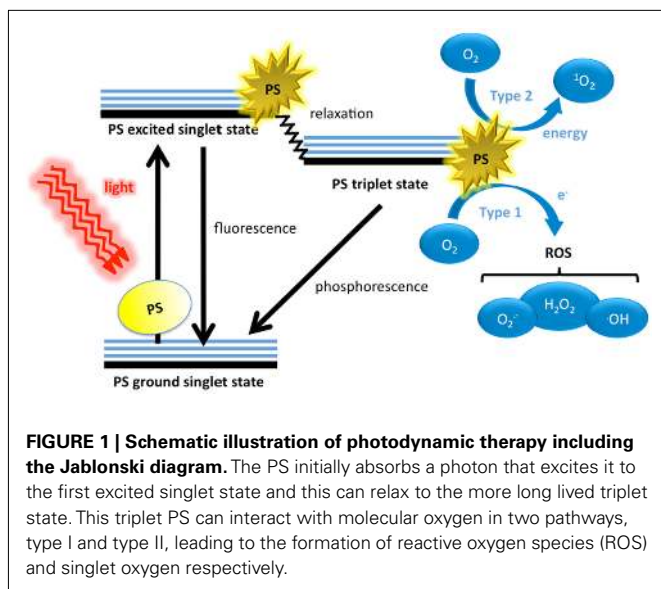
THE PLATFORM OF PHOTODYNAMIC THERAPY

Photodynamic therapy (St. Denis et al., 2011a) was discovered in 1900 by Oskar Raab and Hermann von Tappeiner who found that *Paramecium* spp. protozoans were killed after staining with acridine orange and subsequent exposure to bright light (Raab, 1900). In the 1970s, PDT was initially developed as a therapy for cancer after it was discovered that porphyrins selectively localized in tumors (Mitton and Ackroyd, 2008). Since then, PDT has been clinically used for treatment of various malignancies and is an approved therapy for destruction of choroidal neovascularization (CNV) in age-related macular degeneration. Recently, antimicrobial PDT has been proposed as an alternative approach for localized infections (St. Denis et al., 2011a).

Photodynamic therapy involves the use of a non-toxic light-sensitive dye called a photosensitizer (PS) combined with harmless visible light of the appropriate wavelength to match the absorption spectrum of the PS. After photon absorption the PS reaches an excited state that can undergo reaction with ambient oxygen, resulting in the formation of reactive oxygen species (ROS). PDT is a highly selective modality as (i) the PS can be targeted to the unwanted cells or tissue (Hunt, 2002) and (ii) cell death is spatially limited to regions where light of the appropriate wavelength is applied. Since some PS bind rapidly and selectively to microbial cells, it was suggested that PDT could be used as an anti-infective approach; this became a reality in the mid 1990s (Nitzan et al., 1992).

THE PHOTOPHYSICAL PROCESSES OF PDT

The three principle components of PDT are the PS, visible light, and oxygen. These individually harmless components, when combined, yield potent ROS (Figure 1). Dyes have molecular structures that are typified by conjugated double bonds containing a delocalized system of π -electrons. In the PS ground (singlet) state, these electrons are spin paired in low energy orbitals. Upon application of light corresponding to the absorption peak of the PS, the



electron in the highest occupied molecular orbital (HOMO) of the PS is excited to the lowest unoccupied molecular orbital (LUMO), exciting the PS to an unstable, and short lived excited singlet state. In this state, several processes may rapidly occur (Foote, 1991). The most important of these to the PDT process is the reversal of the excited electron's spin, known as intersystem crossing to give the triplet state of the PS. This triplet state is less energetic than the excited singlet state, but has a much longer lifetime (microseconds as opposed to nanoseconds), as the excited electron, now with a spin parallel to its former paired electron, may not immediately revert to a lower energy level according to the Pauli Exclusion Principle. Accordingly, the excited electron in the PS triplet state may change its spin orientation (a relatively slow process) and emit its energy as phosphorescence, or alternatively the triplet PS may interact with molecules abundant in its immediate environment. Because of the selection rules that specify that triplet–triplet interactions are spin-allowed while triplet–singlet interactions are spin-forbidden, the PS triplet can react readily with molecular oxygen that is one of the few molecules that are a triplet in the ground state (Figure 1).

THE PHOTOCHEMICAL GENERATION OF OXIDIZING SPECIES

The ability of PDT to produce ROS needs the presence of molecular oxygen (O_2 ; Foote, 1991; Tanielian et al., 2000). As mentioned above, the ground electronic state of oxygen is a triplet, whereby the two outermost orbitals are unpaired but spin parallel and this triplet can undergo energy transfer upon collision with the excited PS triplet. This Type II process involves “flipping the spin” of the outermost O_2 electron and shifting it into the orbital containing the other electron, which in turn leaves one orbital entirely unoccupied (a violation of Hund's rule). Termed singlet oxygen (1O_2), this form of oxygen (not considered a radical as its electrons are spin paired) is extremely short lived and reactive, owing to its electron configuration instability. An alternative photochemical mechanism is termed the Type I when the PS triplet directly transfers an electron, sometimes in concert with proton donation to

O_2 , yielding superoxide anion ($O_2^{\bullet-}$) which can then go on to form other ROS including the hydroxyl radical ($\bullet OH$), and hydrogen peroxide (H_2O_2).

The ROS formed through the Type 1 process have a range of different reactivities (Ochsner, 1997). $\bullet OH$, arguably the most reactive of the three ROS formed, is a strong electrophile that is able to chemically attack a very wide range of biomolecules. H_2O_2 is less reactive and $O_2^{\bullet-}$ is least reactive. Nonetheless, $O_2^{\bullet-}$ may be converted to H_2O_2 and O_2 by superoxide dismutase. H_2O_2 is only considered truly reactive when it reacts with ferrous iron in what is known as the Fenton reaction.

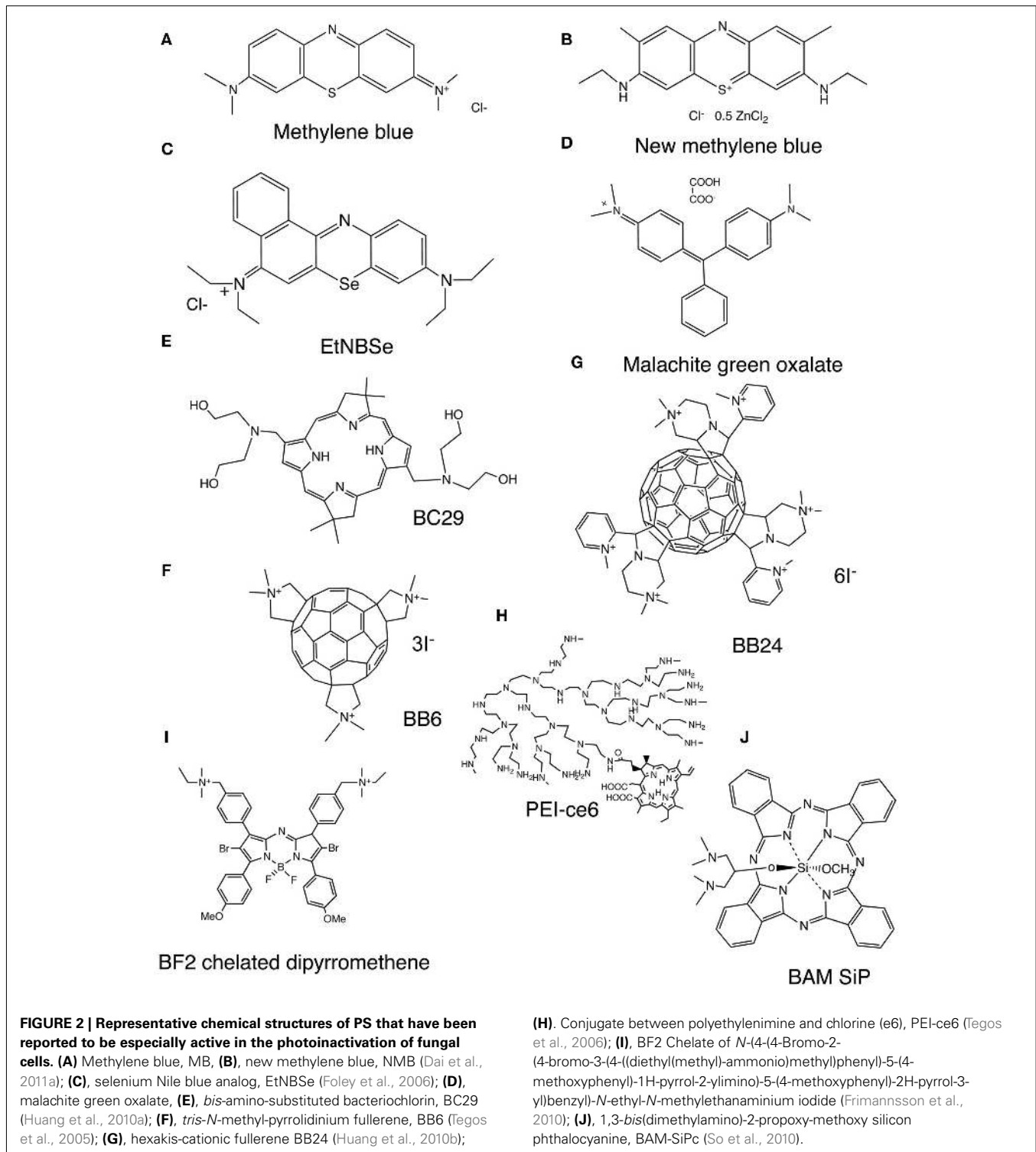


which results in the homolytic fission of the oxygen–oxygen bond in H_2O_2 to yield a hydroxide ion and $\bullet OH$ via the oxidation of ferrous iron to ferric iron (Valko et al., 2005). H_2O_2 is removed through catalase, forming water, and oxygen gas. Although $\bullet OH$ is not broken down by an enzymatic reaction, it may be quenched by antioxidants, including antioxidant peptides (e.g., glutathione) or by antioxidant vitamins (e.g., ascorbic acid).

Because 1O_2 is not a radical, it reacts with biological molecules through quite different mechanisms, making the Type II pathway responsible for different macromolecular reaction pathways. 1O_2 tends to favor reacting with double bonds and sulfur moieties (both of which have high electron densities) and may interact with aromatic components of macromolecules in Diels–Alder cycloadditions (Leach and Houk, 2002; Singleton et al., 2003), 1O_2 is unable to be broken down by enzymes but can be quenched by antioxidants.

PROPERTIES OF PHOTOSENSITIZERS

Photosensitizer are usually organic delocalized aromatic molecules consisting of large conjugated systems of double bonds that may be considered as a central chromophore with auxiliary side chains attached (auxochromes) which are responsible for further electron delocalization of the PS, thus altering the absorption spectra of the PS (Wainwright et al., 2006). Due to extensive electron delocalization, PS tend to be deeply colored. This means that the energy required to excite the electrons in the HOMO to the LUMO is low compared to less delocalized molecules and therefore the absorption bands are in the longer wavelength (red) spectral region and are large, reflecting the high probability of excitation. Acridine orange was the first photodynamic agent used (Takahashi et al., 1975). Most of the PS that have been employed for the treatment of cancer and other tissue diseases are based on the tetrapyrrole nucleus, which were initially porphyrins. Due to the unfavorable absorption spectrum of porphyrins (low peaks in the 630-nm region), emphasis then shifted to other tetrapyrroles such as chlorins, bacteriochlorins, and phthalocyanines that have much larger peaks at longer red wavelengths where tissue transmission of light is maximal. While these tetrapyrroles were initially studied as antimicrobial PS, their inability to kill pathogens that belonged to other classes than Gram-positive bacteria led to the proposal of dyes with different molecular frameworks as antimicrobial PS (Sharma et al., 2011). Phenothiazinium salts, such as methylene blue (MB) and new methylene blue (NMB; Figure 2A,B respectively; Harris et al., 2005; Souza et al., 2010) are often used since



they were the first antimicrobial PS to be tested and are clinically approved for human use. Although there has been a wide range of antimicrobial PS reported, it is becoming clear that the optimal structures for inactivating bacterial cells and the optimum structure for fungal cells may in fact be subtly different from those used for cancer. Some structures such as protease-stable

polycationic (PS) conjugates between polyethylenimine and chlorin(e6), PEI-ce6, (**Figure 2H**; Tegos et al., 2006), tris-cationic substituted fullerenes (BB6; **Figure 2F**), hexakis-cationic fullerenes (BB24; **Figure 2G**), the cationic porphyrin TriP[4], (Lambrechts et al., 2005), and the selenium analog of benzophenothiazinium salt (EtNBSe; **Figure 2C**) may be equally potent to mediate photo

killing of bacteria and fungi. However other structures, including PS that are usually considered to be specific for killing cancer cells, have also been reported to be effective in killing fungal cells (mainly *C. albicans*). This applies to PS such as Photofrin (Bliss et al., 2004), Al(III)-tetrasulfonated phthalocyanine (Bertoloni et al., 1992; Lazarova, 1993; Bliss et al., 2004), and PC4 silicon phthalocyanine (Lam et al., 2011). Moreover some PS that are highly effective in killing Gram-positive and Gram-negative bacteria are not very effective in killing fungal cells. It appears that having a large number of cationic charges is important for efficient photodynamic inactivation (PDI) of Gram-negative bacteria and to a lesser extent Gram-positive bacteria, but this structural feature does not provide good binding or penetration into fungal cells. Rather, more lipophilic structures with a lower amount of cationic charge seem to be better for fungal cells. An example of this latter class is the bacteriochlorin (BC29; **Figure 2E**) that was effective at killing *Candida* cells, while analogs with more cationic charges were better at killing bacteria (Huang et al., 2010a). The list of PS targeting exclusively *C. albicans* is expanding and include malachite green (**Figure 2D**; Souza et al., 2010) an unsymmetrical bis-amino phthalocyanine bis-amino silicon (IV) phthalocyanine (BAM-SiPc; **Figure 2J**; So et al., 2010). A brominated boron difluoride (BF₂) chelated tetraarylazadipyromethene photosensitizer (**Figure 2I**; Frimannsson et al., 2010) reduced in combination with light the viability of yeast cells [5.7 log (10)]. Photodithazine, a glucosamine salt of chlorin e6, enhanced the inactivation of *C. guilliermondii* cells by visible light (Strakhovskaia et al., 2002).

As microorganisms produce and accumulate porphyrins, the appealing hypothesis of endogenous photosensitization is also an alternative pathway of photoinactivation (Oriel and Nitzan, 2010). In this approach a small non-dye amino acid (5-aminolevulinic acid, ALA) is administered as it has been shown that exogenously supplied ALA enters into the microbial heme biosynthetic pathway and results in an accumulation of excess protoporphyrin IX than can act as a moderately effective PS. In an alternative approach, extracts from *Alternanthera maritima* (seaside joyweed) with an absorption range at 650–700 nm can effectively photoinactivate *Candida dubliniensis*. The chemical compositions of the extracts were determined by chromatographic and spectroscopic techniques. The results suggest inhibition of the growth of *C. dubliniensis* after being irradiated with a 685-nm diode laser irradiation alone or crude extracts at 25 mg/ml did not significantly reduce the number of colony forming units (CFU) per milliliter. Steroids, triterpenes, and flavonoids were identified in the analyzed extracts (Gasparetto et al., 2010).

BYPASSING THE PERMEABILITY BARRIER

Bypassing the permeability barrier is a common and challenging theme in antimicrobial drug discovery. Yeasts and fungal pathogens are variable in their cell envelopes, possessing outer wall mixtures of glucans, mannan, and chitin polymers. This feature makes them inherently more permeable to external substances than Gram-negative bacteria. In one of the first reports for antifungal PDI, Toluidine blue (TBO) was tested against *Kluyveromyces marxianus* (Paardekooper et al., 1992). Apart from ability of the

PS to eradicate a microbial population the influence of PDI on the barrier properties of the plasma membrane was studied. TBO mediated PDI-induced a permeability change proceeding in an “all-or-none” fashion which was reflected in potassium ions and E260-absorbing material efflux, reduction of the cell volume, and vacuole integrity. Finally, it was observed that the loss of cell viability was not induced by the all-or-none loss of barrier properties (Paardekooper et al., 1992). A recent study claims that MB-mediated PDI increases membrane permeability in *C. albicans* (Giroldo et al., 2009).

The combination of the anionic PS rose bengal (RB) with glutathione was effective in killing *C. albicans*. Although the key hypothesis was enhancement of PDI involving stimulation of the singlet oxygen generation, the effect of glutathione on the permeability barrier of eukaryotic cells is well established. Alternatively, compounds that create pores within the fungal membrane have shown increased influx of PS compounds that are otherwise unable to enter the cytosol. Phytoalexins of the saponin family are able to bind to the fungal sterol ergosterol, creating pores in the membrane resulting in cellular leakage (Simons et al., 2006). Pretreatment of *C. albicans* cells with a sub inhibitory concentration of a saponin significantly increased the uptake of the PS molecules RB and chlorin (e6), and was able to decrease the survival fraction after exposure with the correct wavelength of a low-intensity light (Coleman et al., 2010). The degree of *C. albicans* killing was 2–5 logs greater in the presence of the saponin when compared to the PS molecules alone. Pretreatment of the saponin did not result in any significant increase in *C. albicans* killing when PEI-ce6 was used as the PS, presumably because of the high permeability of the conjugated PS molecule. Confocal microscopy revealed that ce6 resulted in minimal cellular uptake in *C. albicans*, however co-incubation of the PS and the saponin for 1 h resulted in a visible increase of PS within the *C. albicans* cytosol (**Figure 3**). Collectively, this study suggests in addition to saponins, other compounds with fungal pore-forming properties could be used in conjunction with a PS molecule, and compounds with antifungal efficacy derived by pore-forming ability such as the polyenes (amphotericin B and nystatin) may be a potent treatment strategy for fungal infections.

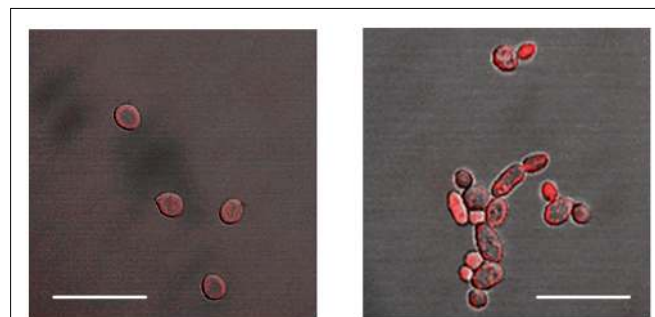


FIGURE 3 | Confocal laser scanning microscopy of *C. albicans* cells treated with ce6 after 24 h. Left panel – Yeast cells treated with 100 μ M ce6 alone. Right panel – Yeast cells treated with ce6 and 4 μ g/ml of a saponin. Scale bar represents 20 μ m.

PDI FOR AZOLE AND ANTIFUNGAL RESISTANT PHENOTYPES

There is documented evidence that PDI is equipotent against both antimicrobial-resistant and susceptible microorganisms (St. Denis et al., 2011a). This may be observed in azole-resistant and susceptible *Candida* spp. PDI employing Photogem® as a PS with a light emitting diode had a significant effect against fluconazole-resistant *C. albicans* and *Candida glabrata* (Dovigo et al., 2011a). A set of clinical isolates from adult HIV patients, including strains highly resistant and sensitive to fluconazole and amphotericin B, were subjected to Photofrin® mediated PDI using a 630 nm laser and the appropriate time to deliver 45–135 J/cm². There was not substantial difference in the survival fractions between resistant and sensitive isolates (Mang et al., 2010).

However, early stationary phase yeast forms of *C. albicans* and *C. glabrata* were not adversely affected by treatment. Respiratory-deficient (RD) strains of *C. albicans* and *C. glabrata* display a pleiotropic resistance pattern, including resistance to azole antifungals, the salivary antimicrobial peptides histatins, and certain types of toxic stresses. The cationic porphyrin PS meso-tetra (*N*-methyl-4-pyridyl) porphine tetra tosylate (TMP-1363) is effective in PDT against yeast forms of *C. albicans* and *C. glabrata*. In contrast to this pattern, RD mutants of both *C. albicans* and *C. glabrata* were significantly more sensitive to PDI compared to parental strains. These data suggest that intact mitochondrial function may provide a basal level of antioxidant defense against PDI-induced phototoxicity in *Candida* spp., and reveals pathways of resistance to oxidative stress that can potentially be targeted to increase the antifungal efficacy of PDT (Chabrier-Roselló et al., 2008). Curcumin, a naturally occurring pigment with a maximum absorption peak in the short wavelength range between 408 and 430 nm has been recently used for successful PDI in clinical isolates of *Candida* spp. (Dovigo et al., 2011b).

INACTIVATION OF CRYPTOCOCCUS AND OTHER FUNGAL PATHOGENS

Recent studies have revealed a substantial PDT effect both in *Cryptococcus* spp. as well as a variety of fungal pathogens. These include both mechanistic and conceptual PDI studies. Stress sensors detect ROS and the cell wall elicits a response noted by the activation of Slt2 for *Saccharomyces cerevisiae* or Mpk1 for *C. neoformans* (Vilella et al., 2005; Gerik et al., 2008). In *C. neoformans* PKC1 is essential for defense against oxidative stress, both in the form of hydrogen peroxide and the thiol oxidizing agent, diamide (Gerik et al., 2008).

For the *C. neoformans* cell wall defective strain KN99α *rom2*, the PS PEI-ce6 exhibited an increased association with KN99α *rom2*, which was observed to penetrate the cell (Fuchs et al., 2007b; Figure 4). Rom2 is a guanyl nucleotide exchange factor in the cell wall integrity pathway, relaying the activation of the sensors by extracellular stressor to the signaling cascade that initiates the phosphorylation events of the mitogen activated protein kinase (MAPK) cascade (Ozaki et al., 1996; Philip and Levin, 2001). The association and ability of the PS to penetrate and collect within the cell was likely due to defects in the cell that prevent the PS from being excluded. The *rom2* mutant is characterized by a 35% reduction in the amount of β-glucan and physical disparities in the cell wall observable through transmission electron microscopy

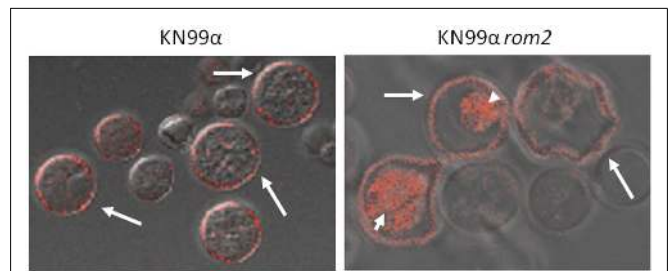


FIGURE 4 | Microscopy was used to visualize location of the PEI-ce6 in association with *C. neoformans*. Cells were grown at 37°C, exposed to PEI-ce6 for 10 min and then washed with PBS. Arrows indicate localization of the PEI-ce6 at the cell wall perimeter. Arrowheads mark the localization of PEI-ce6 within the interior of the cell. With wild-type cells (KN99α), the dye was confined to the surface of the cells. However, with the KN99α *rom2* mutant, the dye was able to penetrate the cell surface.

(TEM). Cell wall defects are enhanced at the high temperature growth condition 37°C, a stress condition for the cell (Fuchs et al., 2007a). The lack of a stable cell wall structure is likely associated with the actin and microtubule defects, cytoskeletal components needed to transport structural components to the cell wall and maintain a rigid configuration and uniform shape. The result of the culmination of defects was an increase in cell death upon application of the activated PS.

Interestingly, cell death was enhanced with the addition of the antifungal caspofungin. While caspofungin is effective against other fungal pathogens, including *C. albicans*, it fails to exhibit fungicidal effects against *C. neoformans*, for reasons that have not yet been fully elucidated (Abruzzo et al., 1997; Espinel-Ingroff, 1998). As a member of the echinocandin class of drugs, caspofungin targets the glucan synthase Fks1p. In this case, the inclusion of caspofungin enhanced the killing of the activated PS. Even though it does not kill *C. neoformans*, caspofungin is known to affect the β-glucan linkages (Feldmesser et al., 2000). It is suspected that the combination of the two cell wall targeted compounds dually damaged the cell enough to lead to cell death or that caspofungin weakened the cell to allow for greater penetration of the PS that led to cell death. The results suggest that external sources of ROS can be applied as a means to inhibit fungi. Potentially, it could enhance the fungicidal activity of compounds that target or weaken the cell wall.

A second study tested the effectiveness of TBO employing *C. gattii* strains with distinct susceptibility profile to amphotericin B and azoles. The concept of equipotent PDI between susceptible and resistant isolates was re-confirmed. A more detailed mechanistic study was attempted including determination of ROS and reactive nitrogen species (RNS, e.g., peroxyxynitrite) production and the catalase and peroxidase activities were measured (Soares et al., 2011).

A variety of reports tested the use of different PS and explored optimal conditions and the morphological changes observed upon PDI of the dermatophyte *Trichophyton rubrum* (Kamp et al., 2005; Smijs et al., 2007, 2008; Smijs and Pavel, 2011). The list of PDI-inactivated fungal species has been increasing exponentially. *Trichophyton mentagrophytes*, *Trichophyton tonsurans*, *Microsporum*

cookei, *Microsporium gypseum*, *Microsporium canis*, *Epidermophyton floccosum*, *Nannizia cajetani*, *Metarhizium anisopliae*, *Aspergillus nidulans*, *A. fumigatus*, and *Fusarium* sp., have been tested with a variety of different PS, offering a new avenue for antifungal therapies (Friedberg et al., 2001; Calzavara-Pinton et al., 2005; Gonzales et al., 2010).

FUNGAL EFFLUX SYSTEMS AS A PDI TARGET

There are two main types of fungal drug efflux systems (Cannon et al., 2009). (1) The major facilitator superfamily (MFS) transporters use the electrochemical gradient across the plasma membrane to expel drugs from cells, and (2) ATP binding cassette (ABC) transporters, in contrast, use ATP binding and hydrolysis to efflux drugs. Although fungal cells contain many genes for both types of systems, clinical azole resistance is most often associated with overexpression of ABC transporters (Holmes et al., 2008; Cannon et al., 2009). There are several classes of fungal ABC transporters, and the pleiotropic drug resistance (PDR) family is often responsible for fungal drug resistance (Lamping et al., 2010). Clinically important PDR transporters include *C. albicans* Cdr1p (CaCdr1p) and CaCdr2p, which are homologs of the *Saccharomyces cerevisiae* Pdr5p (ScPdr5p) and mammalian G-type ABC transporters (Holmes et al., 2006, 2008; Cannon et al., 2009). Fungal PDR efflux systems have relatively promiscuous substrate specificities that are presumably defined by their transmembrane domains. These specificities often partially overlap among family members in a particular organism and thus provide broad-spectrum protection against xenobiotic threats, including those posed by the widely used and well-tolerated azole and triazole drugs.

The role of multidrug efflux in antimicrobial PDT resistance has only recently come under scientific investigation. Phenothiazinium dyes MB and TBO are amphipathic cations and physicochemically similar to the antibacterial alkaloid berberine, a well-characterized substrate of MFS efflux systems in Gram-positive bacteria (Tegos et al., 2002). This similarity raised the possibility that phenothiazinium PS could also be substrates of microbial efflux systems. Recent experimental evidence indicated that phenothiaziniums were *NorA* (MFS) substrates in *S. aureus* and possibly MexAB resistance nodulation (cell) division (RND) substrates in *P. aeruginosa* (Tegos and Hamblin, 2006). The observation that ABC transporters and not MFS affect MB-mediated antimicrobial PDI (APDI) in the pathogenic yeast *C. albicans* is perplexing (Tegos and Hamblin, 2006; Prates et al., 2011). Prates et al. tested this hypothesis by comparing MB-PDI for two pairs of isogenic *C. albicans* strains (i) YEM 12 and YEM 13 mutant overexpressing MDR1 MFS; and (ii) YEM 14 and YEM 15 mutant overexpressing CDR1/CDR2 (Figure 5). Since efflux systems affect APDI of MB in *C. albicans*, a logical strategy was to explore whether efflux pump inhibitors (EPIs) could potentiate APDI with MB. They used two well-documented EPIs, one for each corresponding efflux system, INF₂₇₁ (targeting MFS) and verapamil (+; targeting ABCs both in yeast and mammalian systems) and the reference *C. albicans* strain DAY185. Pre exposure to INF₂₇₁ followed by MB-mediated PDI, resulted in a paradoxical protection rather than enhancement of phototoxicity. On the other hand, pre exposure of *C. albicans* cells to verapamil (+) and subsequent MB-mediated APDI led to an

increase of phototoxicity when compared with the PDI efficacy of MB in the absence of EPIs. In addition, pre exposure of the CDR1/CDR2 overexpressing mutant YEM 15 in verapamil (+) and subsequent APDI revealed 2 logs of cell reduction in the presence of the EPI, in comparison with virtually no effect in the absence of the ABC pump blocker. The authors demonstrated that ABC pumps are directly implicated in MB efflux from the cell cytoplasm. Both the influx and the efflux of MB may be regulated by MFS systems and blocking this gate before incubation with MB can decrease the uptake and APDI effects. An ABC inhibitor could be usefully combined with MB-APDT for treating *C. albicans* infections.

This interaction seems to be less obvious for different PS chemotypes. The participation of efflux systems in porphyrin mediated PDI has been implied in mammalian ABC transporter systems (Morgan et al., 2010).

BIOFILM ERADICATION

Microorganisms in nature thrive through adherence to both living and inanimate surfaces via biofilm formation (Pflumm, 2011). The dense and protected environment of the biofilm, as well as the significantly different phenotypic properties of biofilm cells from free-floating cells of the same species, have been implicated in giving rise to as much as 1000-fold resistance to antimicrobials (Lewis, 2001). Biofilm formation is a critical event in the development of candidosis, including denture stomatitis (chronic atrophic candidosis), which can affect up to 65% of edentulous individuals (Chandra et al., 2001). Despite the use of antifungal drugs to treat denture stomatitis, infection can often become re-established. By using a [poly(methyl methacrylate), PMMA] biofilm model, it was demonstrated that *C. albicans* biofilms are potentially highly resistant to the currently used antifungal agents. Drug resistance was also shown to develop with time and correlated with biofilm maturation (Chandra et al., 2001). There is an expanding body of literature regarding PDT-based biofilm eradication strategies, with emphasis on the use of different PS for biofilm related phenotypes and microbial species (Biel, 2010). The eradication of microbial biofilms remains a key challenge in the antifungal discovery agenda and new efforts are required to address a number of clinical conditions. The list includes urinary tract infections, catheter infections, middle-ear infections, formation of dental plaque (Fontana et al., 2009), periodontitis, (Raghavendra et al., 2009) gingivitis, endodontics, (Soukos et al., 2006) osteomyelitis (Bisland et al., 2006), infected contact lenses, endocarditis, and infections of permanent indwelling devices such as joint prostheses heart valves and implants (Schuckert et al., 2006).

There is a wealth of literature describing PDT-based anti-biofilm strategies that focuses mostly on the use of different PS against a variety of microbial species (Biel, 2010). In contrast there are only a limited number of studies exploring the effects of PDT on phenotypic biofilm elements (e.g., adhesions; Soares et al., 2008). Moreover, there is no consensus as to which is the most reliable model for evaluating PDT efficacy against biofilms. The majority of published reports use methodologies where biofilms are grown in/on plastic or silicon microtiter plates and surfaces. These bioassays have been repeatedly criticized for

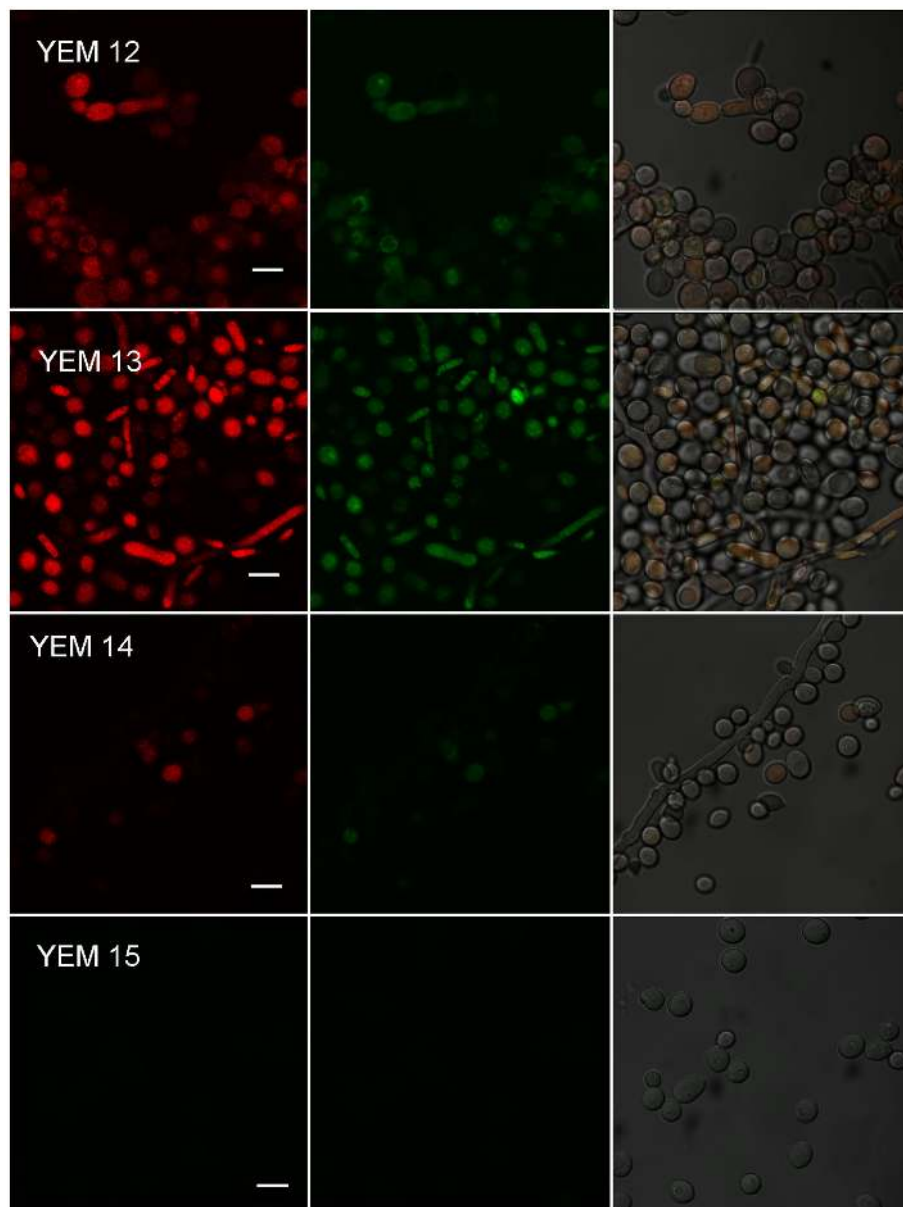


FIGURE 5 | Effect of efflux pumps on photodynamic inactivation of *C. albicans*. Confocal microscopy image of YEM strains. Cells were incubated with 10 μ M R123 and 100 μ M MB and excited at

488 nm. We present three pictures of the same field. Red corresponds to the fluorescence of MB and green corresponds to the fluorescence of R123.

lack of robustness and occasionally yield inconsistent results. In this context it has been demonstrated that *C. albicans* biofilms are sensitive to Photofrin® PDT (Chabrier-Rosello et al., 2005), *C. albicans* and *C. dubliniensis* were susceptible to erythrosine-mediated PDT, but the biofilms of both *Candida* spp. were more resistant than their planktonic counterparts (Costa et al., 2012). The combination of erythrosine- and RB-mediated PDT have some effect in biofilms but also was effective in reducing and destroying of *C. albicans* blastoconidia and hyphae (Costa et al., 2012). MB-PDT combined with InGaAlP laser (660 nm) exhibited a modest reduction in biofilm *C. albicans* species (Pereira et al.,

2011). In a much more challenging scenario, the combination of gasiform ozone and MB-mediated PDI were unable to reduce the viable counts on a multispecies mature oral biofilm assembled by *Actinomyces naeslundii*, *Veillonella dispar*, *Fusobacterium nucleatum*, *Streptococcus sobrinus*, *S. oralis*, and *C. albicans* (Müller et al., 2007).

The bulk of cells in biofilms are actually highly susceptible to killing by antimicrobials and it is indeed only a small fraction of cells known as persisters that remain alive following antimicrobial treatments (Lewis, 2010). Persisters represent a subpopulation of cells that spontaneously go into a dormant, non-dividing state.

When a population is treated with a fungicidal antibiotic, regular cells die but the persisters survive. This persister phenotype hypothesis has been proven for *C. albicans* biofilms using amphotericin B and chlorhexidine (Lafleur et al., 2006) as well as in patients with long-term oral carriage harbor high-persister mutants (Lafleur et al., 2010). This concept may explain partially the mechanism of action in the success of miconazole to augment PDI-mediated by the porphyrin TMP-1363 and MB in *C. albicans* (Snell et al., 2011). In this study, a list of antifungals were tested, miconazole and ketoconazole both stimulated ROS production in *C. albicans*, but only miconazole enhanced the killing of *C. albicans* and induced prolonged fungistasis in organisms that survived PDI. Although the data suggested that potentiation of antifungal PDI by miconazole is likely to be multi-factorial (Snell et al., 2011) and the models tested were not related with the biofilm phenotypes.

FROM THE ANTIFUNGAL PDI MECHANISM TO THE QUESTION OF RESISTANCE DEVELOPMENT

The mechanism of photodynamic action have been described as multi-factorial and non-specific (Bertoloni et al., 1989; Gonzales and Maisch, 2012). It involves the damage of fungal cell wall and membrane by ROS. Subsequently, the intra cellular oxidizing species generated by light excitation induce photodamage to multiple cellular targets. The event list includes inactivation of enzymes and other proteins, peroxidation of lipids, leading to the lyses of cell membranes, lysosomes, and mitochondria and finally causing cell death (Gonzales and Maisch, 2012). A recent study employing the PS Pc 4 in planktonic *C. albicans* cells highlighted the aforementioned proposed sequence of events. Furthermore, changes in nuclear morphology characteristic of apoptosis, which were substantiated by increased externalization of phosphatidylserine and DNA fragmentation following Pc 4-PDI suggesting that apoptotic phenomena were related with fungal cell death (Lam et al., 2011).

Since disruption of electron transport chain (ETC) function increases intracellular levels of ROS in yeast, interference with ETC assembly, or function will enhance antifungal PDT. The metabolic inhibitor antimycin A and defined genetic mutants were used to identify ETC components that contribute to the sensitivity to TMP-1363-PDT in *C. albicans*, *C. glabrata*, and *S. cerevisiae* (Chabrier-Roselló et al., 2010).

The studies and reports discussing the potential of microbes to develop resistance to PDT are scattered, quite controversial, and with a few exceptions they involve bacterial species. The non-selective nature of APDI appears as a competitive advantage in the activation of a specific microbial resistance pathway. In a conventional biological study of routine stress followed by re growth, 5,10,15-*tris*(1-methylpyridinium-4-yl)-20-(pentafluorophenyl)-porphyrin triiodide (Tri-Py(+)-Me-PF) was employed as PS against *Vibrio fischeri* and *E. coli*. After ten cycles of partial inactivation followed by re growth, neither of the bacteria developed resistance to the photodynamic process (Tavares et al., 2010). In a similar study, Giuliani et al. (2010) investigated the potential of phthalocyanine RLP068/Cl mediated PDI to induce resistance to the Gram-positive *S. aureus*, the Gram-negative *P. aeruginosa*, and *C. albicans*, using both sensitive and resistant strains. Its ability, following activation by light, to induce

resistance in these three major human pathogens after 20 daily passages was studied. Simultaneously for the same strains, the ability of daily sequential subcultures in sub inhibitory concentrations of RLP068/Cl to develop resistant mutants without illumination was evaluated. It was demonstrated that 20 consecutive APDT treatments with RLP068/Cl did not result in any resistant mutants and that, in dark conditions, only *S. aureus* strains had increased minimally inhibitory concentrations (MICs) of RLP068/Cl. However, even in this case, the susceptibility of the mutated bacteria to APDT was not affected by their MIC increase. Ehrenshaft et al. (1998) showed that the SOR1 (singlet oxygen resistance 1) gene present in *Cercospora* fungi plays a role in the resistance to PS generating singlet oxygen. Studies with SOR1 deficient mutants resulted in cercosporin and PS sensitivity. However, the function of the protein encoded by this gene and the mechanisms involved in *Cercospora* toxin auto-resistance remain unclear. PDI with RB in the yeast *S. cerevisiae*, demonstrated a role of Yap1p and Skn7p in the defense against singlet oxygen (Brombacher et al., 2006).

Superoxide dismutase is upregulated following protoporphyrin-mediated PDI in *S. aureus* and RB-mediated PDI in *S. mutans* induces the bacterial heat shock protein GroEL – responsible for refolding denatured proteins to native conformations and stabilizing lipid membranes during stress (Nakonieczna et al., 2010). These observations are in accordance with those of St. Denis et al. (2011b) who demonstrated that sub-lethal PDI stress increased the expression of the two major bacterial heat shock proteins GroEL and Dnak and that exposing *E. coli* and *E. faecalis* to heat pretreatment prior to PDI (a positive regulator of GroEL) conferred stress tolerance, increasing *E. coli* cell viability by 2 logs and *E. faecalis* cell viability by 4 logs.

ANTIMICROBIAL PDT: FROM BENCH TOP TO BED SIDE

With the results from *in vitro* studies being promising in a wide array of fungal species, a number of clinical applications for antimicrobial PDT have been tested and performed *in vivo*. PDT has been proposed for many dental applications due to the accessibility of the oral cavity. Nevertheless, the complexity of oral microflora makes this microenvironment quite challenging for the deployment of novel antimicrobials. An *in vivo* study using an immunodeficient murine model with oral azole-resistant candidiasis and topical treatment employing 500 mg/ml MB combined with red light totally inactivated *C. albicans* in the oral cavity and even prevented the emergence of resistance (Teichert et al., 2002). A second study employed immunosuppressed Swiss mice orally swabbed with *C. albicans*. Four days after oral inoculation, PDT was performed on the dorsum of the tongue after topical administration of Photogem® and followed by illumination with LED light at 455 or 630 nm. Determination of CFU, histological, and inflammatory response evaluation of the surgically removed tongues after euthanasia (Mima et al., 2010) revealed effectiveness of the approach.

The very nature of PDT makes it ideal for the treatment of skin, wound, and burn infections, all of which are easily accessible for topical application of PS and light. Dai et al. (2011b) reported PDT using phenothiaziniums for prophylaxis and treatment of cutaneous *C. albicans* infections in mice. A mouse model

of skin abrasion infected with *C. albicans* was developed by inoculating wounds with a luciferase-expressing strain and real-time monitoring of the extent of infection non-invasively through bioluminescence imaging. *In vitro* PDI studies showed that (NMB; **Figure 2B**) was superior to TBO and MB. PDT *in vivo* initiated either at 30 min or at 24 h post infection significantly reduced *C. albicans* burden in the infected mouse skin abrasion wounds (**Figure 6**; Dai et al., 2011a).

C. albicans ear pinna infection using a mouse model was treated with 0.3 mg/ml porphyrin TMP-1363 applied topically in combination with 90 J/cm² green light. Additionally, the phototoxicity of both TMP-1363 and MB against *C. albicans* was compared *in vitro*. TMP-1363 upon irradiation at a fixed fluence of 2.4 J/cm² caused more than three logs of *C. albicans* inactivation than MB at 10 mM incubation concentration for both PS. However, TMP-1363 was inefficiently photo excited, since its peak absorption did not coincide with the irradiation lamp source, underestimating the PDI efficacy of TMP-1363. On the other hand, MB absorption peak was compatible with the emission of the lamp source. Consequently the number of photons absorbed by TMP-1363 was approximately 10-fold lower than that absorbed by the equivalent concentration of MB (Mitra et al., 2011) It was concluded that TMP-1363 displayed a more effective microbial killing than MB due to differences in cellular uptake and/or intracellular localization.

Photodynamic therapy also has promising potential in the treatment of superficial fungal skin infections caused by dermatophytes. *T. rubrum* is responsible for *tinea pedis* (athlete's foot), fungal *folliculitis*, onychomycosis, and dermatophytosis (tinea or ringworm). Employing an *ex vivo* infection model of human stratum corneum of *T. rubrum*, Smijs et al. (2009) incubated samples with the PS 5,10,15-*tris*(4-methylpyridinium)-20-phenyl-(21H,23H)-porphine trichloride (Sylsens B) and deuteroporphyrin monomethylester. Upon light application, both PS were shown to be active antifungals. Moreover, 5-aminolevulinic acid (5-ALA) and red light has an effect in the treatment of onychomycosis (Donnelly et al., 2005; Kumar and Kimball, 2009). Treatment of refractory fingernail onychomycosis caused by non-dermatophyte molds with methylaminolevulinic PDT (Gilaberte et al., 2011).

Chromoblastomycosis is an infection that involves skin and subcutaneous tissues caused by the traumatic inoculation of dematiaceous fungi species, being that the most prevalent are *Fonsecaea pedrosoi* and *Cladophialophora carrionii*. A clinical PDT study employing MB as the PS and a LED device as light source for the treatment of chromoblastomycosis showed promising results (Lyon et al., 2011). PDT for distal and lateral subungual toenail onychomycosis caused by *T. rubrum* have been evaluated in single-center open trial (Sotiriou et al., 2010).

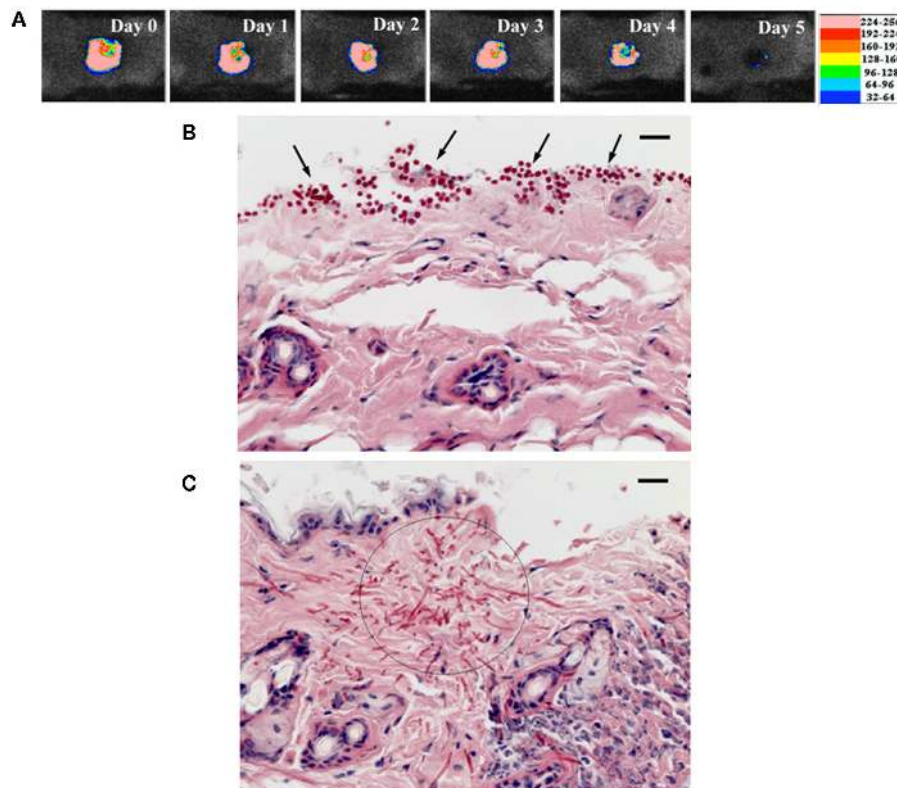


FIGURE 6 | (A) Successive bioluminescence images of a representative mouse skin abrasion infected with 10^7 CFU *C. albicans*. **(B,C)** Representative periodic acid-Schiff-stained skin biopsy specimen taken from a mouse skin

abrasion infected with 10^7 CFU *C. albicans*, showing the presence of yeasts **(B)**, arrows], hyphal filaments **(C)**, circle], and inflammatory infiltrate **(C)**. Biopsy was taken on day 1 postinfection. Bar, 20 μ m.

An evidence-based review of published literature (MEDLINE, EMBASE, and Cochrane Library) was searched until March 2010 and evaluated the efficacy and safety of PDT for superficial mycoses. No randomized clinical trials were found. Seven reports described the antifungal effect of PDT against 63 superficial mycoses patients. Eight of 10 (80%) tinea cruris patients and 6 of 10 (60%) tinea pedis were led to mycological cure after one to three treatments. Four (40%) tinea cruris patients and 3 (30%) tinea pedis had a persist healing at the 8-week follow-up. Six of the 9 (66.7%) foot-interdigital mycoses patients recovered clinically and microbiologically after one or four treatments. Only two patients (22.2%) had a persist healing at the 8-week follow-up. Eleven of 30 (36.6%) onychomycosis patients were cured for 18 months after treatment, and three onychomycosis patients were all cured in other two reports. The therapeutic effect of PDT for one pityriasis versicolor patient was also reported (Qiao et al., 2010).

Six Korean patients aged 23–47 years with recalcitrant *Malassezia folliculitis* were enrolled in a clinical PDT trial employing methyl 5-aminolevulinic acid (MAL)-PDT. These patients were offered MAL-PDT as an alternative treatment option due to inability or nephrotoxicity to tolerate oral antifungals. MAL was applied locally as a cream to each lesion (located on the patients' trunks) and covered with an adhesive occlusive dressing polyurethane film. After 3 h, the cream was wiped off and illumination was performed immediately thereafter with non-coherent red light using light emitting diodes (wavelength 630 nm, light dose 37 J/cm²). Patients underwent three sessions of MAL-PDT at 2-week intervals. One month after the last PDT treatment, patients returned to the hospital and lesions were photographed (Lee et al., 2010). After three sessions of MAL-PDT, inflammatory lesions had decreased and improved obviously in four patients, had improved slightly in one patient, and had not improved in one patient.

Photodynamic therapy has been studied to treat ophthalmological disease secondary to fungal infection of the eyes. The efficacy and safety of PDT was evaluated in the long-term control of subfoveal CNV associated with toxoplasmic retinochoroiditis (Neri et al., 2010). The records of 13 patients with classic subfoveal CNV associated with toxoplasmic retinochoroiditis treated with PDT were reviewed. All patients were followed up for at least 48 months. Postoperative visual acuity (VA) was defined as a gain or loss of two or more lines of best-corrected visual acuity (BCVA), respectively. Post-treatment CNV size was dichotomized into "increased" if the major CNV diameter (CMD) had increased by ≥ 300 μm , and as "stable/reduced" if it had decreased by ≥ 300 μm or had not changed by > 300 μm . At the 48-month follow-up, all patients had stable/improved BCVA and a mean stable/reduced CMD (846 ± 326.5 μm), with the BCVA having improved significantly ($p < 0.0001$) from 0.29 ± 0.19 at baseline to 0.54 ± 0.16 at 48 months. Finally, a 28-year-old one-eyed woman with subretinal CNV in the right eye of due to *C. endophthalmitis* was treated with a combination of PDT and drugs. The CNV was treated with six PDT sessions with verteporfin in association with systemic steroid therapy with prednisone (100 mg/day to reduce) and fluconazole (800 mg/day to reduce). VA was assessed in pre-PDT conditions and after six PDT treatments (24 months of follow-up; Tedeschi et al., 2007).

FORMULATION OF NEW CONCEPTS, CONCLUSION

Advances in microbial physiology continue to shed light on a series of pathways, components, and phenotypes that may serve as potential alternative and attractive targets for antimicrobial drug discovery. These approaches have revealed novel molecular mechanisms responsible for cooperation among cells and define new roles of population structure for the evolution of cooperative interactions. This knowledge of interaction parameters is changing the view of microbial processes, and suggests new ways to fight infection by exploiting social interaction (Xavier, 2011). Targeting bacterial virulence factors is also a novel approach under investigation for the development of new antimicrobials that can be used to disarm pathogens in the host (Lee et al., 2003). The broad-spectrum activity and the non-specific action of APDI should be further explored to address these biological phenomena. There is no documented evidence whether PDT can disrupt these sophisticated microbial defensive lines. We have to take into account that PDI is able to eradicate microorganisms without discriminating resistant isolates, both planktonic and biofilm species. This is in concert with the potential of localized photooxidative stress to inactivate virulence factors (Kömerik et al., 2000; Tubby et al., 2009) and virulence determinants (Zolfaghari et al., 2009; Hamblin et al., 2011; Sharma et al., 2011) in the absence of any documented conventional resistance mechanism. The possibility of active efflux seems to be related with some but not all the molecular classes of PS although improved delivery methods may overcome this barrier. A summary of promising, state of the art, powerful PDI-based combinations is provided in **Table 1**.

Nanoparticles can be engineered as sophisticated drug delivery vehicles to carry various therapeutic or diagnostic agents and are potentially useful for medical applications including targeted drug delivery, gene therapy, and cell labeling (Kim et al., 2010). PDT has also attracted the interest of nanotechnology as the effectiveness of the treatment can be greatly enhanced by the use of nanoparticles. In the last decade, different approaches to the combination of nanoparticles and PDT have been investigated in relation to the antimicrobial applications of the technique. One use of the nanoparticles is to improve the delivery of PS to bacteria; others use the nanoparticles to improve the inactivation kinetics (Perni et al., 2011). Many of the PS being studied for PDI of bacteria are based on the tetrapyrrole nucleus are lipophilic and readily aggregate in aqueous solution, resulting in the loss of photosensitizing activity (Sibani et al., 2008; Engelhardt et al., 2010). To overcome this problem, suitable PS carriers were designed to deliver PS, e.g., liposomes, (Ferro et al., 2006; Bombelli et al., 2008; Engelhardt et al., 2010) micelles, (Tsai et al., 2009), and nanoparticles (Schwiertz et al., 2009; Guo et al., 2010). Amongst these systems, liposomes are most commonly employed to incorporate lipophilic PS, and have been proved to enhance the APDI of various PS, not only because liposomes increase the solubility and stability of PS, but also because they can facilitate the penetration of PS into bacteria by means of fusion processes or disturbing the cell walls (Jia et al., 2010). However, these reported liposomal formulations mainly aimed to deliver PS passively, while little research was done to apply actively targeted liposomes in the PDI of bacteria or fungi (McCarron et al., 2007). An interesting application describes the use of a patch as a mucoadhesive drug delivery TBO system

Table 1 | Photodynamic therapy-based combinations: potential antifungal countermeasures.

PDT-combinations	Synergist	Fungal species	Target(s)	Reference
Polycationic conjugates of chlorin	Visible light	<i>C. albicans</i>	non-specific reactive oxygen species (ROS)	Tegos et al. (2006)
Methylene blue (MB), toluidine blue (TBO)	Visible light, EPIs	<i>C. albicans</i>	ROS	Prates et al. (2011)
MB	Visible light, azoles	<i>C. albicans</i>	ROS	Snell et al. (2011)
Functionalized C60-fullerene	Visible light	<i>C. albicans</i>	ROS	Lu et al. (2010)
PEI-ce6	Saponins visible light	<i>C. albicans</i>	ROS	Coleman et al. (2010)
PEI-ce6	Casporfungin-visible light	<i>C. neoformans</i>	Membrane	Fuchs et al. (2007b)

(Donnelly et al., 2007). However, the concentrations of TBO in the receiver compartments separated from patches by membranes intended to mimic biofilm structures were an order of magnitude below those inducing high levels of kill, even after 6 h release.

Photodynamic therapy is not a conventional drug discovery platform since three elements (PS, visible light, and oxygen) are essential for successful deployment. As infection involves both the pathogen and the host, it is important to exploit this additional complexity in PDT ventures. The evaluation of the naturally occurring pigment curcumin (Martins et al., 2009) against *C. albicans* provides an example. Planktonic, biofilm cells as well as macrophages were submitted to the same photodynamic conditions to investigate whether the treatment could be toxic to microbial as well as mammalian cells and identify a therapeutic window for preclinical development. On the other hand, antimicrobial PDT treatment has been shown to be potently and functionally inactivated by IL-1 beta and TNF-alpha (Braham et al., 2009).

Researchers have bypassed some of the difficulties associated with new antimicrobial development by developing tractable whole-animal screens that utilize the well-studied nematode *Caenorhabditis elegans*, the great wax moth *Galleria mellonella*, and the fruit fly *Drosophila melanogaster* as model hosts to identify and develop new classes of antimicrobial agents with antivirulence or immunomodulatory efficacy and evaluate toxicity or efficacy. The amenability of these non-vertebrate hosts to large screens has made these model hosts useful to identify or develop active compounds against either bacterial or fungal pathogens (Apidianakis et al., 2007, 2011; Fuchs et al., 2010). Therefore, the design of host-pathogen studies exploring the ability of PDT to interfere with virulence determinants requires sophisticated tools and approaches. The recent example of a host-parasite model to assess intracellular targeting specificity of novel phthalocyanines (Dutta et al., 2011) will inspire similar explorations.

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A brief comparison between antibacterial and antifungal PDT, reveals that the latter require higher concentrations of PS (1 μ M minimum and generally higher) for substantial fungal PDI. This fact translates into the need to use considerably higher PS concentrations for potential *in vivo* antifungal applications. It should also be noted that fungal BSI often arise from local infections that spread systemically. PDT has the particular advantage of rapidly reducing numbers of viable fungal cells allowing antifungal drugs time to work. This notion supports the argument that PDT is ideal for localized infections but it also may be prophylactic to prevent the development of BSI or may be extremely useful in combinatorial implementation with antifungals. This therapeutic concept has been demonstrated in practice by employing PDT combined with antibiotics in animal models of localized infections caused by Gram-negative bacteria (Lu et al., 2010).

Further clinical trials are needed to evaluate the efficacy of PDT to treat superficial mycoses. It is also important to optimize treatment protocols in order to cope with recurrence. PDT may be an effective treatment option for patients with recalcitrant fungal infections. However, the available clinical data are still limited, and additional controlled trials including multiple patients will be necessary to verify the results of the promising pilot studies.

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