Indian Journal of Clinical Biochemistry, 2010 / 25 (2) 188-192

ORIGINAL ARTICLE

CONFIRMATORY STUDIES ON THE ANTIOXIDANT AND ANTIDIABETIC EFFECT OF QUERCETIN IN RATS

M A Abdelmoaty [a,c], M A Ibrahim [b,d], N S Ahmed [a,c] and M A Abdelaziz [a,d]

Departments of [a]Biochemistry and [b]Pharmaceutics, [c]Sohag Faculty of Medicine ,Sohag University and [d]El-Azhar Faculty of pharmacy,Elazhar University (Assiut), Egypt.

ABSTRACT

Quercetin (QE), one of natural flavanoid group, was widely distributed as a secondary metabolite in plant kingdom. It has been believed that oxidative stress plays a role in the pathogenesis of diabetes mellitus (DM). The aim of the present study was the evaluation of possible effects of QE on blood glucose and antioxidant enzymes in experimental streptozotocin (STZ)-induced diabetes in rats. STZ was injected intraperitoneally with single dose of 50 mg/kg for diabetes induction. QE (15 mg/kg bw day, intraperitoneal (i.p.) injection) was injected for 3 days prior to STZ administration; these injections were continued to the end of the study (for 25 days).Glucose tolerance test and random plasma glucose were done for all animals. Cellular antioxidant enzymes such as glutathione peroxidase (GSHPx), superoxide dismutase (SOD) and catalase (CAT) activities were measured in pancreatic homogenates. Quercetin had no effect on plasma glucose level of normal animals but its pre- treatment was able to prevent diabetes induced by single intraperitoneal injection of streptozocin-treated rats. Antioxidant enzyme activity significantly decreased in STZ induced diabetic group. QE treatment significantly increased the antioxidant enzyme activities. It could be concluded that quercetin, a flavonoid with antioxidant properties, exerting its beneficial antidiabetic effects.

KEY WORDS

Quercetin, Flavonoid, Antioxidant, Antidiabetic.

INTRODUCTION

Oxidative stress plays an important role in the etiology of DM (1). Diabetics and experimental animal models exhibit high oxidative stress markers and reactive oxygen species (ROS) in pancreatic islets due to persistent and chronic hyperglycemia, thereby deplete the activity of the antioxidative defense system and thus promote free radical generation (2).

The cellular antioxidant status determines the susceptibility to oxidative damage and is usually altered in response to oxidative stress. Accordingly, interest has recently grown in the role and usage of natural antioxidants as means to prevent oxidative damage in DM with high oxidative stress. The antioxidants such as SOD, CAT and GSHPx are the initial step to protect the cells against lipid peroxidation (3). Reduced antioxidant levels as a result of increased free radical production in experimental DM have been reported (4).

Flavonoids are a group of naturally occurring compounds widely distributed as secondary metabolites in the plants (5). They have been recognized for having interesting clinical properties, such as anti-inflammatory, anti-allergic, anti-viral, anti-bacterial, and anti-tumor activities (6). One of these flavonoids, QE (3,5,7,3',4'-pentahydroxyflavone), prevents oxidant injury and cell death by several mechanisms, such as scavenging oxygen radicals protecting against lipid peroxidation and chelating metal ions (7).

Many scientists studied and are studying plant drugs useful in DM and some of them having antioxidant properties (3, 6, 8,

Address for Correspondence :

Dr. Mohamed Ahmed Abdelmoaty
Department of Biochemistry,
Sohag faculty of Medicine,
Sohag University, Sohag- Egypt. Tel. +2-0126388153.
E-mail: abdelmoaty5@yahoo.com

9). Due to the presence of aromatic hydroxyl groups, flavonoids have strong antioxidant properties (9). They are scavengers of reactive oxygen and nitrogen species and, therefore, inhibit peroxidation reactions. They, also, protect macrophages from oxidative stress by keeping glutathione in its reduced form (10). Flavonoids have the capacity to inhibit enzymes such as cyclooxygenases and protein kinases involved in cell proliferation and apoptosis (11). They, also, protect normal rat islets from STZ, normalize blood glucose levels and promote â-cell regeneration in islets of STZ-treated rats (12). Tritiated thymidine incorporation into islet cell DNA was, also, enhanced by these flavonoids in an in vitro study (13). However, the beneficial effects of flavonoids on DNA adduct levels, oxidative damage to DNA and chromosomal aberrations in human could not be demonstrated (14). A different flavonoid, QE, used in doses of 15–50 mg/kg body mass was capable of normalizing blood glucose level, augmenting liver glycogen content and significantly reducing serum cholesterol and LDL concentration in alloxan–diabetic rats (15). Exposure of isolated rat islets to certain flavonoids such as QE enhances insulin release by 44–70% via alteration in calcium fluxes and in cyclic nucleotide metabolism (16).

The present study was undertaken to determine the effect of minimal dose of QE on antioxidant status and blood glucose concentration in normoglycemic and STZ induced diabetic rats.

MATERIALS AND METHODS

Reagents: QE was obtained from Sigma Chemical Company (USA) and dissolved in 0.5ml of 60% ethanol just before injection. Streptozocin vials containing 1 g streptozocin and 220 mg citric acid was obtained from Upjohn Co. (USA). Streptozocin was reconstituted with 9.5 ml of 0.9% NaCl solution to pH 3.5–4.5 according to manufacturer's instructions. Furthermore, dilution of the drug was performed in 0.9% NaCl solution immediately before use. The enzymatic kits for the determination of glucose was supplied by (Bicon,Co. Germany).

Animal experiments: Adult male albino rats bred and raised at the Assiut University animal quarters with a mass ranging from 200 to 250 g and averaging 16 weeks old were used and maintained at 20-22°C, light period 7.00a.m. to 7.00p.m., and relative humidity 55%. Five animals each were housed in a cage and were given a rat Purina chow diet, water *ad libitum*. DM was induced by single intraperitoneal injection of streptozocin (50 mg/kg body mass) 3 days after the initiation of treatments. Blood was obtained from the tail vein using heparinized microhematocrit tubes. Plasma glucose of all groups was measured day after day. After one week, glucose tolerance test was done. Animals with plasma glucose level exceeding 16.6 mM were considered as diabetic. All normoglycemic and STZ diabetic rats were employed for the tests described below.

Effect of quercetin on plasma glucose level: The required amount of QE was dissolved in 0.5 ml of 60% ethanol prior to injection to rats. Four groups of rats including (five rats per group) were used: (1) normoglycemic control group receiving the vehicle as one intraperitoneal (i.p.) injection of 0.5 ml 60% ethanol per day; (2) normoglycemic rats receiving one i.p. injection of 15 mg QE per kg body mass per day; (3) diabetic control rats receiving one i.p. injection of 0.5 ml 60% ethanol per day; and (4) diabetic experimental animals receiving one i.p. injection of 15 mg QE/kg bw per day. Such treatment was began 3 days prior to induction of DM and continued for 25 days at 13:00 h every day. Blood was collected from the tails using heparinized microhematocrit tubes and used for plasma glucose determination on the day of intraperitoneal injection of STZ (day 0) and every other day thereafter.

Glucose tolerance test: Glucose tolerance test was performed according to the procedures of Young et al on the four different groups mentioned above (17). Following QE treatment for 2weeks, plasma glucose was determined, the animals were treated with another respective dose of QE and deprived of food for 24 h. After this period, blood was collected through heparinized tubes for plasma glucose (fasting glucose level). The fasting plasma glucose levels of the all diabetic and control rats groups were done. Fasted animals were fed 1 ml of a glucose solution containing 3 g/kg body weight glucose through a gavage. Tail blood were collected at 45, 90 and 135 min after glucose feeding and plasma glucose levels were determined and used for establishing glucose tolerance curves (17).

Tissue homogenization and antioxidant enzymes assay: At killing time (25 days after QE treatment), rats were anaesthetized with sodium pentobarbital (63 mg/kg body weight, intraperitoneally). Blood was collected by cardiac puncture. Pancreas of each rat was removed, cleaned of fatty tissue, blotted dry and weighed. Each pancreas was homogenized in 10 vol. of 50 mM sodium phosphate buffer (pH 7.4) for 30 sec. using a Glas-Col (TURBAX-Germany) homogenizer. The homogenate was filtered through gauze and the filtrate was centrifuged at 1088×*g* for 5 min in a refrigerated centrifuge. The resultant supernatant was used for measurement of antioxidant enzymes (18).

Superoxide dismutase: SOD activity was measured depending on its antagonizing effect on autooxidation of epinephrine (colourless) into adrenochrome (chromogen) by superoxide anion generated in alkaline condition. Therefore, the optical density was inversely proportionate with the enzyme activity (19).

Glutathione peroxidase (GSHPx): Measurment of GSHPx activity depended on its ability to destroy H_2O_2 in presence of reduced glutathione and NADPH. The released NADP was measured at DOD_{340} compared to supernatant protein content (20).

Catalase: Catalase activity was measured and depended on its ability to decrease added hydrogen peroxide in supernatant solution so, the read at OD_{240} was decreased after 2 minutes of incubation compared to supernatant protein content (21).

Statistics: Using SPSS version 10 software, the data were statistically analyzed by one-way ANOVA and Duncan test. The level of statistical significance was set at $P<0.05$.

RESULTS

QE significantly increased the pancreatic activity of SOD enzyme threefold within 25 days after its injection (group 2) compared to that control normoglycemic group (group 1). STZ had an opposite effect on the activity of pancreatic SOD (group 3), but that effect was ameliorated by pretreatment with QE prior to STZ injection (group 4) (Table 1).

Table 1: Antioxidant effect of quercetine on normoglycemic and streptozocin-diabetic rats

	Group 1	Group 2	Group 3	Group 4
SOD (U/mg protein)	26.54 ±1.73	77.28±5.87*	6.62±1.61♣	17.14±2.74#
GSHPx (µM/min/ mg protein)	0.03±0.01#	0.06±0.01*	0.01±0.01♣	0.03±0.01
CAT (µM/sec/ mg protein)	0.03±0.002	0.06±0.01*	0.01±0.01♣	0.04±0.004#

Data are expressed as mean ±SD of 5 rats in each group. In each column, figures bearing *is a significantly different at $P<0.05$ between group 1 and 2, ♣ is a significantly different between group 1 and 3, and # is a significantly different between group 3 and 4. Group 1 is normoglycemic control; group 2 is normoglycemic+15mg quercetin per kg; group 3 is diabetic control and group 4 is 50 mg/kg STZ+15mg quercetin per kg. Pancreatic SOD (superoxide dismutase) expressed in U/mg protein, GSHPx (glutathione peroxidase) expressed in mM/min/ mg protein and CAT (catalase) expressed in mM/sec./ mg protein.

QE produced a significant double increase the activity of pancreatic GSHPx after 25 days of administration compared to normoglycemic control group. STZ decreased pancreatic GSHPx and this was antagonized by pretreatment with QE (Table 1). Similar effect could be observed in pancreatic CAT, the activity of pancreatic CAT after QE treatment was significantly greater (three times) than the normoglycemic control group, but, STZ had an opposite effect. This activity was significantly higher in QE + STZ group than STZ diabetic group (Table 1). The current results suggested that STZ could decrease pancreatic SOD, GSHPx and CAT but, QE could enhance the activity of these enzymes and consequently antagonize STZ effect on these antioxidant enzymes. But STZ could not restore SOD normal value while GSHPx and CAT returned to normal value.

Intraperitoneal injection of STZ increased plasma glucose level gradually to reach its maximum level up to 35.8 ±3.4 mM within 14 days. QE had no effect on plasma glucose concentration of normoglycemic animals, but significantly maintained blood glucose level and antagonizing the effect of STZ on DM induction in experimental rats for 3 subsequent doses of QE prior to STZ injection. The range of blood glucose level of the diabetic animals treated with QE was 4.5 to 7-9 mM but in STZ injected group was 4.6 to 35.8 mM allover the period of experiment (Fig1).

Glucose tolerance curve was done one week after STZ injection, and reached its peak (15.2 ± 1.59 mM) within 45 min. after oral glucose intake and returned to its fasting level within 135 min. in normoglycemic control rats. QE exerted no different effect on the glucose tolerance curve. But, STZ

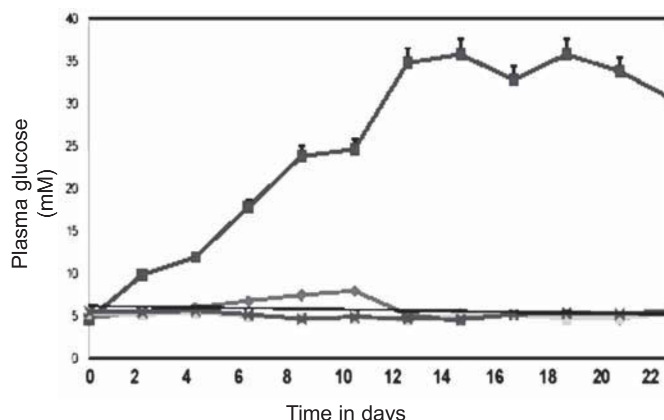

Fig 1: Time Profile of Random plasma glucose concentration day after day in normoglycemic and streptozocin-diabetic rats after i.p. administration of streptozocin. Each point represents the mean ±SD. of five rats, ⁎ Normoglycemic control; ▲ normoglycemic + 15 mg quercetin per kg; ■ STZ control; ◆ STZ + 15 mg quercetin per kg

impaired glucose tolerance curve where fasting plasma glucose level was 15.4±2, reached its peak 30.3±4 within 45 min and did not returned back within 135 min. of oral glucose intake. Intraperitoneal injection of 10 mg QE/kg body prior to STZ was able to normalize glucose tolerance curves in experimental rats (Fig 2).

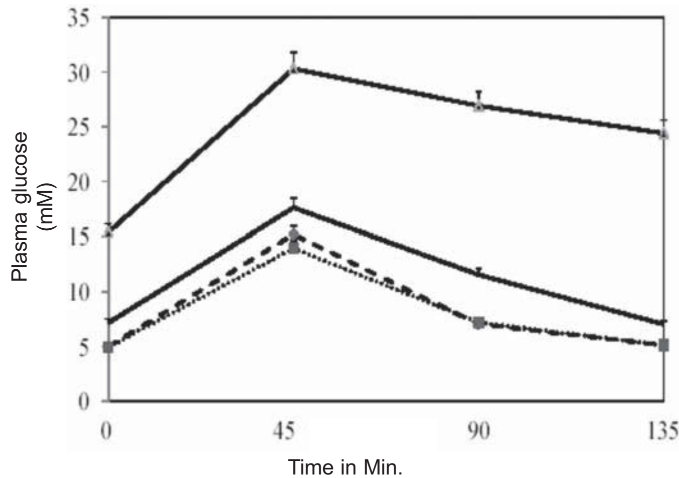

Fig 2: Glucose tolerance curves of normoglycemic and streptozocin diabetic rats with or without i.p. administration of quercetin. Each point represents the mean±SD. of five rats, • normoglycemic control; ■ normoglycemic + 15 mg/kg quercetin; ▲ diabetic; • diabetic + 15 mg/kg quercetin

DISCUSSION

Although the β-cell cytotoxic effect of STZ is not fully understood, it is though to be mediated by the inhibition of free radical scavenger-enzymes thereby enhancing the production of the superoxide radical (2). Diabetics and experimental animal models exhibit high oxidative stress due to persistent and chronic hyperglycemia, which thereby depletes the activity of antioxidative defense system and thus promotes de novo free radicals generation (1, 2). Oxidative stress has been shown to be responsible, at least in part, for tissue damage and β-cell dysfunction (18). In the present study, STZ was associated not only with hyperglycemia and impaired glucose tolerance but also, with low antioxidant enzymes activity.

Srinivasan and Menon reported that the antioxidant drug bis-o-hydroxy-cinnamoylmethane used for protection of β-cells against reactive oxygen species (ROS) mediated damage by enhancing antioxidants and reduces hyperglycemia in STZ-induced DM (22). In the present study, we examined possible usefulness of QE to protect the rat against diabetic effect of STZ and its effect on some antioxidant enzymes (SOD, GSHPx and CAT) which can protect β-cell against oxidative stress in type 1 DM.

QE, a flavonoid, used in low dose (15 mg/kg bw) was shown to be capable of preventing induced hyperglycemia by STZ and normalizing blood glucose level in rats. Hii and Howell reported that exposure of isolated rat islets to certain flavonoids such epicatechin or QE enhances insulin release by 44–70% (16). They argue that such flavonoids may act on islet function, at least in part, via alteration in Ca^{2+} fluxes and cyclic nucleotide metabolism (16). Vessal et al suggested that QE supplementation has proven to be beneficial in decreasing blood glucose concentration, promoting regeneration of the pancreatic islets and increasing insulin release in STZ-induced diabetic rats; thus exerting its beneficial antidiabetic effects (23). The role and usage of natural antioxidants for preventing oxidative damage in DM is on the increase (3, 16). QE had no significant hypoglycemic effect on normoglycemic rats, but, antagonizing the raising in blood sugar induced by STZ treated rats and this effect might be secondary to its antioxidant activity.

QE, a flavonoid found in many plants, is widely distributed in edible fruits and vegetables. When QE was administered orally, it was poorly absorbed from digestive tract and did not have a great influence on the organs (5). The injected intraperitoneally of a single dose of 15 mg/kg to rat is equivalent of about 1000mg to a 70 kg human person. Normalization of glucose tolerance curves in diabetic animals treated with 50 mg QE/kg bw is shown (12). Normalization of plasma glucose in QE in STZ-induced diabetic rats and its lack of effects on plasma glucose level of normoglycemic animals are in agreement with previously reported effects of epicatechin (16) and QE (24) in alloxan-diabetic animals.

The current result is consistent with the results of Wolf (24) and El-Missiry (25), who indicated an increase in lipid peroxides and a decrease in antioxidant enzymes in DM. The present results indicate that the preventive effects of QE may be due to inhibition of lipid peroxidation by its antioxidant nature.

In conclusion, the present study confirms earlier reports that QE could prevent hyperglycemia induced by STZ in rats. It was suggested that this effect was owing to its antagonistic effect to prevent a decrease of pancreatic activity of antioxidant enzymes induced by STZ. But to elucidate the exact mechanism of that modulatory effect and to examine its potential therapeutic effects further studies are essential.

REFERENCES

1. Baynes J, Thorpe S. The role of oxidative stress in diabetic complications. Curr Opin Endocrinol 1996; 3: 277-84.
2. Ihara, Y, Toyokuni S, Uchida K, Odaka H, Tanaka T, Ikeda H. Hyperglycemia causes oxidative stress in pancreatic beta cells of GK rats a model of type 2 diabetes. Diabetes 1999; 48: 927-32.
3. Kanter M, Meral I, Dede S, Gunduz H, Cemek M, Ozbek H. Effects of *Nigella sativa* L. and *Urtica dioica* L. on lipid peroxidation, antioxidant enzyme systems and some liver enzymes in CCl_4-treated rats, J Vet Med A Physiol Pathol Clin Med 2003; 50: 264-8.
4. Bray T, Bettger W. The physiological role of zinc as an antioxidant, Free Radic Biol Med 1990; 8: 281-91.
5. Rizk A, Hammouda F, Rimpler H, Kamel A. Iridoids and flavonoids of *Teucrium polium* herb. Planta Med 1986: 2: 87-8.
6. Middleton J. Effect of plant flavonoids on immune and inflammatory cell function. Adv Exp Med Biol 1998; 439: 175-82.
7. Laughton M, Evans P, Moroney M, Hoult J, Halliwell B. Inhibition of mammalian 5-lipoxygenase and cyclo-oxygenase by flavonoids and phenolic dietary additives. Relationship to antioxidant activity and to iron ion-reducing ability Biochem Pharmacol 1991; 42: 1673-81.
8. Pari L, Saravanan G. Antidiabetic effect of cogent db, a herbal drug in alloxan-induced diabetes mellitus. Comp Biochem Physiol 2002; 131: 19-25.
9. du Thie G, Crozier A. Plant derived phenolic antioxidants. Curr Opin Clin Nutr Metab 2000; 3: 447-51.
10. Fuhrman B, Aviram M. Flavonoids protect LDL from oxidation and attenuate atherosclerosis. Curr Opin Lipidol 2001;12: 41-8.
11. Estany S, Palacio J, Barnadas R, Sabes M, Iborra A, Martínez P. Antioxidant activity of N-acetylcysteine, flavonoids and α-tocopherol on endometrial cells in culture. J Rep Immunol 2007; 75: 1-10.
12. Un J, Mi-Kyung L, Yong B, Mi A, Myung-Sook C. Effect of citrus flavonoids on lipid metabolism and glucose-regulating enzyme mRNA levels in type-2 diabetic mice. Int J Biochem Cell Biol 2006; 38: 1134-45.
13. Maria M, Kyriakos R, Dimitrios G. Protection against nuclear DNA damage offered by flavonoids in cells exposed to hydrogen peroxide: The role of iron chelation. Free Rad Biol Med 2005; 39: 1591-600.
14. Sara R, Seymour G, Radim J, Blanka B, Ivan K, Lyubomirova K, et al. Effects of diet on biomarkers of exposure and effects, and on oxidative damage. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis 2007; 620: 93-102.
15. Zapolska-Downar D, Kosmider A, Naruszewicz M. Flavonoids-rich extract from chokeberry fruits inhibits oxLDL-induced apoptosis of endothelial cells. Atherosclerosis (Suppl) 2006; 7: 223-4.
16. Hii C, Howell S. Effects of epicatechin on rat islets of Langerhans. Diabetes 1984; 33: 291-6.
17. Young P, Cawthorne M, Coyle P, Holder J, Smith S. Report on the treatment of obese mice with BRL-49653, a new and potent insulin sensitizer, enhances insulin action in white adipocytes. Association with increased insulin binding and cell surface GLUT4 as measured by photo affinity labeling. Diabetes 1995; 44: 1087-92.
18. Kakkar R, Kalra J, Mantha S, Prasad K. Lipid peroxidation and activity of antioxidant enzymes in diabetic rats. Mol Cell Biochem 1995; 151: 113-9.
19. Misra H, Fridovich I. The role of superoxide anion in the autooxidation of epinephrine and a simple assay for superoxide dismutase. J Biol Chem 1972; 215: 3170.
20. Goldberg D, Spooner R. In methods of enzymatic analysis. (Bergmeyer HV, ed.), 3rd ed., Verlog Chemie, Deerfield Beach, FL,USA 1983; 3: 258-65.
21. Beers R, Sizer I. A spectrometric method for measuring the breakdown of hydrogen peroxide by catalase. J Biol Chem 1952; 195: 133-8.
22. Srinivasan A, Menon V. Protection of pancreatic beta cell by the potential antioxidant bis-*o*-hydroxycinnamoylmethane, analogue of natural curcuminoid in experimental diabetes. J Pharm Pharm Sci 2003; 6: 327-33.
23. Vessal M, Hemmati M, Vasie M. Antidiabetic effects of quercetinin streptozocin-induced diabetic rats. Comp Biochem Physiol Toxicol Pharmacol 2003; 135: 357-64.
24. Wolff S. Diabetes mellitus and free radical. Free radicals, transition metals and oxidative stress in the aetiology of diabetes mellitus and complications. Br Med Bull 1993; 49: 643-9.
25. El-Missiry M, El Gindy A. Amelioration of alloxan induced diabetes mellitus and oxidative stress in rats by oil of *Eruca sativa* seeds. Ann Nutr Metab 2000; 44: 97-100.