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Consensus Paper: Decoding the Contributions of the Cerebellum as a Time Machine From Neurons to Clinical Applications

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Abstract

Time perception is an essential element of conscious and subconscious experience, coordinating our perception and interaction with the surrounding environment. In recent years, major technological advances in the field of neuroscience have helped foster new insights into the processing of temporal information, including extending our knowledge of the role of the cerebellum as one of the key nodes in the brain for this function. This consensus paper provides a state-of-the-art picture from the experts in the field of the cerebellar research on a variety of crucial issues related to temporal processing, drawing on recent anatomical, neurophysiological, behavioral, and clinical research.

The cerebellar granular layer appears especially well-suited for timing operations required to confer millisecond precision for cerebellar computations. This may be most evident in the manner the cerebellum controls the duration of the timing of agonist-antagonist EMG bursts associated with fast goal-directed voluntary movements. In concert with adaptive processes, interactions within the cerebellar cortex are sufficient to support sub-second timing. However, supra-second timing seems to require cortical and basal ganglia networks, perhaps operating in concert with Cerebellum. Additionally, sensory information such as an unexpected stimulus can be forwarded to the cerebellum via the climbing fiber system, providing a temporally constrained mechanism to adjust on-going behavior and modify future processing. Patients with cerebellar disorders exhibit impairments on a range of tasks that require precise timing, and recent evidence suggest that timing problems observed in other neurological conditions such as Parkinson's disease, essential tremor, and dystonia, may reflect disrupted interactions between the basal ganglia and cerebellum.

The complex concepts emerging from this consensus paper should provide a foundation for further discussion, helping identify basic research questions required to understand how the brain represents and utilizes time, as well as delineating ways in which this knowledge can help improve the lives of those with neurological conditions that disrupt this most elemental sense. The panel of experts agrees that timing control in the brain is a complex concept in whom cerebellar circuitry is deeply involved. The concept of a timing machine has now expanded to clinical disorders.

Introduction

Over the past generation, our thinking about the cerebellum has undergone a dramatic transition from an oversimplified functional view restricted to the motor system to one in which this subcortical structure is recognized as part of networks involved in virtually all aspects of cognition [1-4]. Although there had been conjectures about ‘nonmotor’ functions of the cerebellum over 50 years ago [5], the cerebellar cognitive revolution took off with the advent of technological advances in computational modeling, neuroimaging, and high-resolution neurophysiology. Prominent in this work has been the study of cerebellar contributions to the representation of temporal information, computations that are essential in both motor and cognitive domains. The very high number of neurons in the cerebellum with a specific anatomical arrangement and its dense connectivity with extra-cerebellar centers make of the cerebellum of unique structure which has often been compared to a computer involved in temporal aspects.

This consensus paper brings together the viewpoints of a group of established neuroscientists whose research programs cover a broad spectrum of methodological approaches to understand cerebellar function. The primary objective here is to summarize key concepts that may explain confirmed and potential roles of cerebellar circuits in timing. Breska and Ivry analyze the role of the cerebellum in the timing of isolated intervals; Lawrenson and Apps discuss the timing of climbing fiber inputs to the cerebellum and implications for their function. D’Angelo considers the regulation of spike timing and plasticity in the cerebellar network; Petter, Lusk and Meck emphasize the role of specific cerebellar structures in predictive timing and integrate this with basal ganglia function in their presentation of the Initiation, Continuation, Adjustment, and Termination (ICAT) model of temporal processing; Manto and Mitoma address cerebellar control of the timing of fast movements; Gerwig reviews the issue of timing and eyeblink conditioning with a focus on human studies. And, finally, the disruption of cerebellar processing in movement disorders with an emphasis on timing is outlined by Avanzino with respect to Parkinson’s disease and dystonia, and by Filip and Louis with respect to essential tremor.

We are well aware that a final consensus cannot be made given our current understanding of this enigmatic structure. Nonetheless a broad agreement has been reached on the importance of the cerebellum in many aspects of timing. We hope that the ideas presented here will help stimulate studies of cerebellar function and its interaction with cortico-striatal timing circuits in the years to come.

The Cerebellum Represents Isolated Temporal Intervals Across Task Domains

Assaf Breska and Richard B. Ivry

It is widely accepted that the cerebellum contributes, in some manner, to temporal processing, but the functional domain and computational mechanisms remain the subject of considerable debate. An early hypothesis was that the cerebellum served as a centralized, dedicated timing system [6, 7]. Subsequent lines of research have associated timing with other brain structures, such as the basal ganglia (BG), supplementary motor area (SMA), and inferior parietal cortex [8-11], or with neural network dynamics that operate independent of specific neuroanatomical structures [12]. In order to understand cerebellar contributions to timing, and how this might differ from that of other brain structures, it is important to specify constraints on cerebellar timing, both in terms of how temporal information is represented within this system and the contexts in which this information is exploited.

Timing has been studied with a diverse set of tasks, such as eyeblink conditioning [13], duration estimation [6], and rhythmic circle drawing [14]. Given this diversity, taxonomic classifications can provide a roadmap to identify common computational principles [15]. Based on activation patterns in neuroimaging studies, Coull and Nobre [16] proposed an influential taxonomy in which timing tasks were mapped onto two dimensions. One dimension focused on whether timing was part of motor behavior or independent of movement (e.g. perceptual timing). The other, orthogonal dimension asked whether timing was explicit or implicit. Examples of explicit timing would be when the task requires an overt report of a temporal quantity or movement at a specific time; implicit timing is when temporal information facilitates performance on a non-temporal task such as is observed when making non-temporal judgments about sensory events that occur at predictable instead of random times.

The literature indicates that the cerebellum may be involved to some extent in all four sub-domains of timing within the Coull and Nobre taxonomy. The cerebellum has a central role in motor timing, as is evident from symptoms of cerebellar ataxia, such as dysmetria, dysarthria and dysdiadochokinesia. Neuropsychological [7, 17, 18] and neuroimaging [19-21] evidence implicate the cerebellum in tasks that directly measure explicit motor timing, such as reproducing an interval from working memory or producing periodic taps after a metronome is turned off. A central role for the cerebellum is observed in tasks that rely on implicit motor timing in which a movement requires anticipating a forthcoming stimulus. Examples here include eyeblink conditioning, where an adaptive conditioned response must anticipate the precise time of the unconditioned stimulus [13, 22, 23], or interception tasks [24]. However, there are notable motor tasks involving temporal regularities that do

not rely on Cerebellum. The neuropsychological and neuroimaging literatures converge in indicating minimal involvement of the cerebellum when producing cyclic movements at a constant rate [14, 25].

While a role in motor timing fits with traditional perspectives in which the cerebellum is a critical part of a network for producing coordinated movements, the extension of the functional domain of cerebellar timing to perceptual tasks has proven more contentious. Perhaps the most basic perceptual tests of explicit timing are duration comparison and discrimination tasks in which participants indicate, for example, which of two temporal intervals is longer [6]. Again, there is some degree of convergence across the neuroimaging [26, 27] and neuropsychological literature [6, 28], with the latter showing that lesions of the cerebellum increase discrimination thresholds rather than produce a distortion of time (e.g., speed up or slow down). Interestingly, a different picture emerges if the task requires explicit judgment concerning the temporal structure of rhythmic perceptual events such as whether there is deviation from isochronism. Relative to explicit timing tasks conducted on isolated intervals, performance in these rhythmic tasks is less impaired in individuals with cerebellar degeneration and associated with less cerebellar activation [29, 30].

Implicit perceptual timing is mostly associated with tasks in which performance on a non-temporal task can benefit from a context that confers some sort of temporal predictability. One example is a scenario in which a target event can be anticipated to occur at a specific moment in time relative to some preparatory cue [31, 32]. Individuals with cerebellar degeneration show reduced ability to use this temporal regularity to facilitate preparation [33]. However, in scenarios in which the perceptual events occur rhythmically, typically allowing preparatory processes to fluctuate, or synchronize with a beat structure [34, 35], imaging studies failed to find cerebellar activation [36, 37], and even find reduced activity relative to a non-isochronous control condition [37].

The above suggests that the Coull and Nobre taxonomy [16] is not sufficient for specifying the functional domain of cerebellar timing. We suggest that another dimension is required, continuity, highlighting the distinction between “event” and continuous cyclic timing. By this view, the cerebellum is essential for event timing, where temporal information is defined by isolated intervals [14, 15, 38, 39]. This constraint seems to hold independent of whether the tasks are motoric or perceptual, explicit or implicit. In contrast, across domains, the cerebellum is not necessary when temporal information is defined by dynamic, cyclic events. Timing in such contexts may rely on controlling high-level movement parameters. For continuous, repetitive movements, timing might be emergent to the control of a constant angular velocity [38] (see [15] for discussion contrasting this form of timing with that required in repetitive tapping tasks). In other contexts, especially those in which periodic events

shape temporal expectancies, timing may arise from neural oscillations entrained by the events [40-42] or dedicated rhythm processing circuits [37, 43].

Coull and Nobre had suggested a related conceptual distinction within their implicit perceptual timing category. In their view, a distinction can be made between contexts in which timing is driven by continuous stimulus dynamics or generated from a memory representation of an isolated interval, viewing this distinction as analogous to exogenous vs endogenous forms of attentional orienting. Consistent with this hypothesis, orienting in time based on rhythmic structure or from memorized isolated intervals are associated with distinct EEG signatures [44, 45]. Moreover, as outlined here, we propose that the event vs continuous distinction may pertain across task domains of timing, and that the former will require cerebellum.

In summary, the current state of research points toward a role for the cerebellum in timing of discrete, isolated intervals, but not when temporal information is contained within continuous task dynamics. This computational distinction appears to apply for both motor and perceptual timing, as well as for implicit and explicit timing tasks. Undoubtedly, this hypothesis requires direct evaluation; for example, comparisons should be made between tasks entailing isolated intervals or continuous temporal patterns that fall within the subdomains of the Coull and Nobre taxonomy.

The Timing of Signals Forwarded to the Cerebellum Provides Clues to Climbing Fiber Function.

Charlotte L Lawrenson and Richard Apps

The cerebellum plays an important role in the control of movement, and indeed a wide range of other functions, but how it does this is still a matter of considerable debate. It receives information mainly through an array of mossy fiber and climbing fiber (CF) pathways; and the prevailing view is that the latter holds the key to understanding cerebellar operation [46]. This is emphasized by the fact that damage to the inferior olive, which gives rise to the CF system, results in motor deficits that resemble those that occur after direct cerebellar damage [47-51].

In relation to the current consensus topic, various theories have implicated the cerebellum in timing, however 'timing' can be considered in different ways. For this discussion, we will focus on the times during behavior when information is forwarded to the cerebellum via the CF system. If the timing of transmission of signals is regulated (gated) then this restricts when CFs can influence cerebellar operation and thus places important constraints on their function.

To date, studies of this gating phenomenon have mainly investigated the timing of transmission in spino-olivocerebellar pathways (SOCs) that arise from the limbs. These studies are based on the principle that the CF- Purkinje cell (PC) synapse is highly secure [52], and that each PC receives only one CF in the adult cerebellum [52-54]. As a result, any changes in the size of a CF field potential evoked by peripheral stimulation will reflect changes in excitability in the associated SOCs because it is an indirect but reliable measure of the number of local PCs that are synchronously activated by their CF input.

The results from these studies have shown that the information SOCs convey is not continuously available but is gated during active movements. For example, in a reach to grasp movement in cats the largest evoked responses (i.e. best transmission) in SOCs that target the paravermal cerebellar cortical C1 and C3 zones occurs when the animal is sitting quietly at rest; the same responses are smallest (i.e. least transmission) when the SOCs occur during the grasp phase of the movement [55, 56]. For the same SOCs there is also a systematic variation in transmission during the step cycle in the ipsilateral forelimb: by comparison to rest, increased transmission occurs during the swing phase, while reduced transmission occurs during stance [57].

The swing phase is the time during the step cycle when a limb is most likely to encounter obstacles to progression, while the stance phase is when self-generated (reafferent) sensory information is most likely to occur (e.g. because of load bearing by the limb). These results have therefore been taken to suggest that the gating (at least in SOCs that target the paravermal C1 and C3 zones) regulates the times when behaviorally predictable and unpredictable sensory events are forwarded to the cerebellum via the CF system during active movement. In particular, a reduction in transmission during active movements is hypothesized to reflect a gating out of self-generated (predictable) reafference [55, 57-59].

This possibility has recently been tested during exploratory behavior in rats [58]. Rearing is a natural behavior of rodents which allows them to survey their local environment. During such behavior transmission of sensory signals from the ipsilateral hindlimb to the cerebellar C1 zone was found to be reduced when rearing up or down compared to when the animal was fully upright. This finding is consistent with the notion that transmission via SOCs of self-generated sensory inputs are gated out during movement but that the same pathways are open for transmission at a time during behavior when the animal is vulnerable to predatory attack and therefore needs to be able to respond to external sensory events. In other words, the gating of transmission in SOCs that target the C1 and C3 zones may serve to gate out the predictable (internally generated) sensory consequences of a movement, while permitting transmission of unpredictable (externally generated) sensory signals.

If this is the case, then the pattern of gating in SOCPs should be modifiable if a sensory stimulus becomes predictable. Evidence for this was shown when the hindlimb stimulation was delivered repeatedly over many consecutive trials during the upright phase of rearing (when responses would normally be evoked when the stimulus was presented unpredictably). Over time the evoked field response became progressively smaller in size i.e. transmission was reduced when the occurrence of the peripheral stimulation became predictable [58].

Returning to the question of timing, these experiments therefore provide evidence that there is a powerful regulatory system in place that determines when CFs can transmit information to Cerebellum. This in turn may reflect the times when the CF system can modify cerebellar output. For example, an external skin tap to the forelimb of a cat could lead to a near synchronous activation of CFs in the relevant forelimb-receiving territory within the cerebellar paravermis via SOCPs. The fastest conducting of these SOCPs can transmit signals from the periphery to the cerebellar cortex in approximately 11ms, indicating that they are able to rapidly update the cerebellum concerning sensory events [57, 60, 61]. Subsequent modification of cerebellar output via cortico-nucleo-rubral-spinal circuits can occur within an additional 9ms, which means the overall loop time is ~19ms. This is sufficiently rapid that an external stimulus has the potential to modify an ongoing movement, such as the swing phase of the forelimb step cycle (which lasts ~200 ms – [which lasts ~200ms, 57], via supraspinal cerebellar circuits as illustrated in Figure 1.

In summary, the modulation of movements by the cerebellum is constrained by the timing of sensory information forwarded by SOCPs. An unexpected external stimulus can be forwarded to the cerebellum via the CF system at certain times during an on-going movement when a modification of the motor output would be behaviorally useful. Evidence has shown that repetitive patterns of sensory input during a behavior can modify such patterns of transmission and this mechanism might underlie how predictable sensory inputs from self-generated signals are gated out during movements

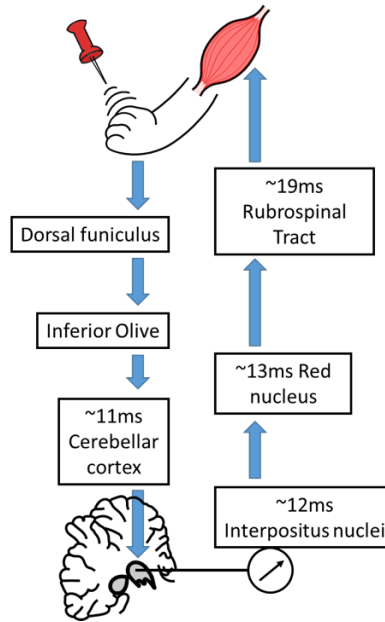


Figure 1. Following a forelimb perturbation and transmission through SOCPs, a response can be recorded in the cerebellar cortex with the fastest latency at ~11ms and nuclei beginning with a latency of ~12ms [55, 57, 61]. The interpositus-rubral projection is in the order of 1ms [62] and it takes approximately 5-6ms for the rubrospinal tract to modify an EMG response [63]. Therefore, the time it takes for the motor output from a sensory innervation to be modified is in the order of around 19ms.

Spike Timing and Synaptic Plasticity in the Cerebellum as the Basis of Temporal Processing

Egidio D'Angelo

Two seminal theories presented in the late 60s maintained that the cerebellum could operate either as a “timing machine” or a “learning machine” [64, 65]. While their formulation emphasized an apparent duality of function, a unifying interpretation is that the cerebellum is required to *learn* how to *predict* precise *timing* in a sequence of events, which can be either sensory stimuli, motor commands, or even logical elements in abstract reasoning [66]. Recently, the relationship between *timing and learning* has been brought down to the investigation of *spike timing and synaptic plasticity*, suggesting that these fundamental cerebellar functions are indeed tightly interconnected at the level of cellular and microcircuit mechanisms [67-69].

High-precision regulation of spike timing and plasticity in the cerebellar granular layer

Converging evidence suggests that, while well timed spike patterns eventually need to be emitted by neurons in the deep cerebellar nuclei, a privileged role in temporal processing is played by the cerebellar cortex, and in particular, by granule cells. These neurons are normally silent at rest and respond with short spike bursts when activated by mossy fiber inputs [70, 71]. Indeed, at least half of the information passing across the mossy fiber – granule cell relay is carried by the time to first spike, while the rest is carried by the number of spikes in the bursts [72, 73]. Experimental and modeling analysis has revealed that the emission time of granule cell spikes can be precisely tuned over a doubly fast and slow time-band through specific properties of ionic channels, synaptic receptors and neurotransmitter release, as well as of microcircuit wiring.

In granule cells, a K inward rectifier helps stabilizing resting membrane potential at a negative level, preventing generation of spurious spikes. This raises the signal to noise ratio and allows retransmission of salient spike sequences organized in bursts by exploiting synaptic integration and temporal summation [74-77].

The spike initiation mechanism of granule cells exploits specific properties of Na channels in order to allow rapid action potential generation in the axonal initial segment [78, 79]. The spikes invade the ascending granule cell axon in less than 0.1 ms and back-propagate into the dendrites in less than 0.3 ms [80]. This ensures almost instantaneous activation of the overlaying Purkinje cells [81] and sub-millisecond precise coincidence detection in granule cell dendrites. An A-current can delay the first spike by tens of milliseconds [82]. On a slower time scale, an M-like current tunes spike generation on the theta-band favoring the entrainment of granule cells in ensemble oscillations [74, 76]. In addition to regulating the number of emitted spikes, neurotransmitter release probability at the mossy fiber –

granule cell synapses can change the first spike delay by acting on EPSP temporal summation through changes in short-term facilitation and depression. Interestingly, release probability is specifically regulated by long-term synaptic plasticity, so that its increase during LTP minimizes first-spike delay, while its decrease during LTD protracts first-spike delay [75] see below).

Excitatory neurotransmission from mossy fibers is mediated by AMPA and NMDA glutamate receptors. Granule cell AMPA receptor-mediated currents are the fastest in the brain, ensuring sub-millisecond precision to spike initiation [83]. This mechanism integrates with that provided by the much slower NMDA receptor-mediated current, which operates on the 10-100 ms time window [70, 83] and is instrumental to the induction of synaptic plasticity through the regulation of calcium influx [84].

Inhibitory neurotransmission from Golgi cells is mediated by $\alpha 1$ and $\alpha 6$ GABA-A receptors. Inhibitory neurotransmission, in addition to controlling first spike delay and burst duration, can increase their precision. The overall impact of $\alpha 6$ is larger than that of $\alpha 1$ subunit-containing receptors. However, $\alpha 1$ -receptors-controlled granule cell responses in a narrow ± 10 ms band while $\alpha 6$ receptors showed broader ± 50 ms tuning [85, 86]. Therefore, like the excitatory system, also the inhibitory system is organized to operate on a double time band.

The granular layer is organized to feed well-timed Golgi cells inhibition onto granule cells [87] through the mechanisms reported above. Golgi cells show intrinsic pace-making in the theta band and, when an input comes, show phase-resetting. As such, these neurons may play a critical role for timing of discrete, isolated intervals as described above by Breska and Ivry. Moreover, specific delay lines can be generated by unipolar brush cells (UBCs) [88], which in rodents are almost exclusively present in the vestibulo-cerebellum [89]. UBCs can generate late-onset responses, in which the delay of spike emission is precisely regulated by H- and TRP-currents depending on the intensity and duration of mossy fiber activity.

These observations identify the granule cells as a pivotal point for spike-time control in the cerebellum, which is regulated on a double time-band. It can be anticipated that the mechanisms operating on the 1-ms band are critical for plasticity expression, while the mechanisms operating on the 100-ms (theta) band are critical for plasticity induction. Indeed, a recent form of spike-timing dependent plasticity (STDP) has been reported, whose induction exploits a dynamic range centered over the theta band and whose expression is sensitive to the relative phase of granule cell spikes and mossy fiber EPSCs with 1-ms precision [90]. A theoretical study actually predicts that STDP is the core mechanism for rapid memory acquisition in the cerebellum granular layer [91].

Spike timing and plasticity in the other regions of the cerebellar network.

While granule cells are silent at rest, all of the other cerebellar neurons act as pacemakers, including the principal neurons along the main retransmission line (Purkinje cells and deep cerebellar nuclei cells) and the inhibitory interneurons (Golgi cells and stellate cells) (for review see [92]). These neurons are likely to exploit different strategies for spike timing based on burst-pause responses [93]. Precise sequences of excitatory and inhibitory synaptic transmission have been suggested to govern plasticity in deep cerebellar nuclei and coincidence detection between complex spikes and simple spikes is required in Purkinje cells. However, both of these mechanisms span over a 100 ms range. Therefore, precise timing may be first acquired in the granular layer and then be maintained and propagated through the rest of the network.

A summary view and Implications for neuropathology

In summary, the cerebellar granular layer appears especially suitable to carry out the timing operations required to confer millisecond precision to cerebellum computations. Oscillations and resonance in the theta band [94] may provide the clock that allows plasticity to be deposited through STDP rules during learning.

There is compelling evidence that the ability of the cerebellum to learn the precise timing of actions can be altered in several instances. For example, patients with cerebellar ataxia are unable to predict the precise timing and gain of elementary motor acts in a sequence [95] generating symptoms such as dysmetria, dysarthria and dysdiadochokinesia. Experimentally, learning the precise gain and timing of actions can be altered by TMS in paradigmatic tests such as eyeblink classical conditioning (EBCC) [96, 97], vestibulo-ocular reflex (VOR) [98] and saccades [99]. In mice, timing can be altered by specific knock out of genes involved in granular layer mechanisms. It can therefore be envisaged that specific interventions to these mechanisms could reestablish timing and learning in Cerebellum.

An Integrated Role for the Cerebellum in Predictive Timing

Elijah A. Petter, Nicholas A. Lusk, and Warren H. Meck

Predictive timing requires a subject to learn the temporal relations among stimuli in order to perform anticipatory responses at correct times. A classic example of this is eyeblink conditioning, in which an auditory or visual conditioned stimulus (CS) predicts a future unconditioned stimulus (US) usually in the form of a periorbital air puff. In order to avoid an aversive air puff, the subject must learn the temporal contingency between the two stimuli, closing their eyelid just prior to (US). While the cerebellum's contributions to timing and time perception have primarily been studied in relation to its role in timing in the milliseconds (ms) range [100-102], a number of experiments have provided evidence for a more

integrative role spanning multiple seconds [103]. This research traces the role of the cerebellum from projection neurons of the cerebellar cortex, through the dentate nucleus, and into thalamo-cortical-striatal circuits [104, 105]. As a consequence, the cerebellum may not be limited to timing sub-second durations but may in fact play an important role in the timing of discrete intervals in the millisecond-to-minutes range [15, 106].

Purkinje Cells

The principal cell type in the cerebellar cortex is the Purkinje cell (PC). PCs are thought to play a key role in predicting the US. Moreover, as early findings identified dissociable contributions of the medial and lateral cerebellar cortex to motor execution and motor timing respectively [107], studies on predictive timing have focused largely on PCs within the lateral cerebellar cortex. This temporal prediction comes in the form of adaptively timed pauses in simple spiking activity, which are learned from repeated pairings of the CS and US. Further, these responses are shown to be capable to elicit motor output through the use of optogenetics in head-fixed mice [108]. Recent work suggests that these cells do not just receive temporal information from an upstream circuit, but rather, the timed responses of the cerebellum may be intrinsic to PCs (e.g., [22, 109, 110]). This body of evidence demonstrates that after extensive training the responses of PCs can be elicited without a temporally coded input. Specifically, neither the length of the CS, nor circuit mechanisms are required for the pause in PC simple spiking. Taken together, this work suggests that there is an intrinsic memory component in the PCs that is capable of storing adaptive timing information important for associative learning [111, 112].

Deep Cerebellar Nuclei

The temporal information stored in PCs is sent downstream to the deep cerebellar nuclei. As PCs are GABAergic, these pauses in the PC activity cause disinhibition of downstream nuclei. Electrophysiological recordings within the dentate nucleus (DN), the most lateral portion of the deep cerebellar nuclei, have shown increased activity (i.e., ramping) to temporally regular stimuli presentations [113]. Additionally, when an expected stimulus is omitted or deviates temporally from the expectation, the increasing pattern of neural activity is disrupted.

Correlations between ramping in neural activity and saccade times have been demonstrated in neurons of the DN of non-human primates during a task involving self-timed saccades [114]. The ramping up of neuronal firing in the DN was observed to span both sub- and supra-second durations (400 – 2400 ms) though its contribution to timing behavior seems to differ between the two duration

ranges. Single-trial analysis found that ramping activity began shortly after cue onset and that saccade time correlated well with the slope of activity for delays of 400-1200 ms. In contrast, ramping activity for the 2400 ms delay started late in the interval with its onset time being correlated with self-timed saccade latency. This second finding suggests that the cerebellum may not be actively involved in tracking supra-second durations but may play an important role in the adjustment or tuning of predictive responses. Human patient studies have bolstered the specificity of the cerebellum in predictive timing, with deficits in anticipatory motor responses observed in spinocerebellar ataxia type 6 patients [115] as well as preferential activation of the cerebellum during predictive timing tasks compared to a reproduction task in healthy adults [116]. Similar timing deficits have also been displayed in cerebellar lesioned rodents [117].

The observation of the cerebellum's involvement in the timing of both sub- and supra-second durations aligns well with the recently proposed Initiation, Continuation, Adjustment, and Termination (ICAT) model of temporal processing [118]. As illustrated in Figure 2, the ICAT model accounts for interactions between the cerebellum and striatum during distinct phases of temporal processing across sub-second and supra-second durations. The neural architecture of the cerebellum makes it well-suited for mediating the *initiation* and *adjustment* phases of the timing model with its strongest influence occurring during the acquisition of timed response sequences. The ICAT model also proposes that the cerebellum is primarily responsible for automatic timing processes that underlie reflexive behaviors [119]. In contrast, cortico-thalamo-striatal (CTS) circuits provide controlled flexibility in support of the *continuation* mechanisms for interval timing. Moreover, this model fits quite well with the clinical observations of deficits in movement control and time estimation related to a variety of cerebellar disorders (e.g., [24, 120-124]).

Adjustment of Downstream Circuits

Projections from the deep cerebellar nuclei have also been shown to play an important role in adjusting other timing circuits. These studies generally focus on the dentate nucleus, or the homologous lateral cerebellar nucleus (LCN) in rodents. In one study, efferents from the LCN were found to form disynaptic connections, via the thalamus, to other major timing loci such as the medial frontal cortex and basal ganglia [125]. Enhancement of LCN signaling to the medial frontal cortex by optogenetic stimulation of terminals within the thalamus increased precision on a supra-second predictive timing task in a rodent model of schizophrenia, as well as reinstated normal extracellular activity patterns [126].

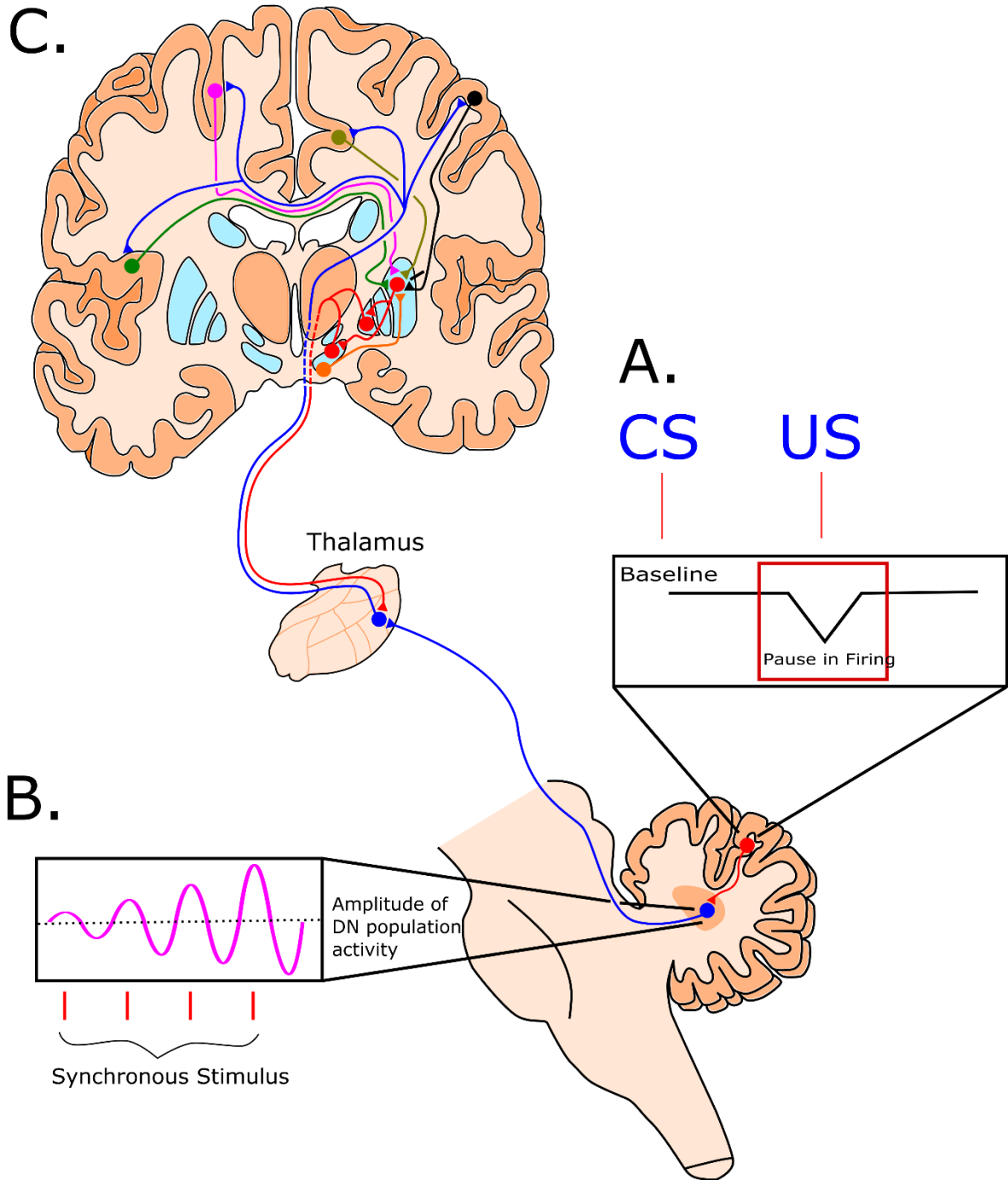


Figure 2. The cerebellum's contribution to temporal processing. The traditional role of the cerebellum in temporal processing has been studied through eye blink conditioning. This preparation involves a conditioned stimulus (CS) such as a tone, that predicts an unconditioned stimulus (US) such as an aversive air puff (A). Cerebellar Purkinje cells learn to pause their activity during the US in order to elicit an adaptively timed blink response. These pauses in Purkinje cell activity modulate the activity of neurons in the dentate nucleus of Cerebellum. This activity increases during timing tasks (B). This activity from the dentate nucleus then travels to the thalamus which can help initiate temporal processing in cortical-striatal circuits (C). Adapted from [120].

Computational Properties of the Cerebellum

The biology of the cerebellum, including cell types and circuit architecture, has been extensively studied and the resulting information has allowed researchers to model and investigate the computational nature of Cerebellum. This has led to the observation that there are numerous connections that can provide 'teaching signals' (e.g., climbing fibers – [127], as well as 'reciprocal connections' (e.g., PC to PC, or PC to interneuron – [128]) which are likely to play an important role in the dynamics of temporal processing within Cerebellum. Moreover, microzones have been identified in the cerebellum, which are regions that form the basic computational unit of the cerebellum [129], and it is these inputs and outputs that dictate the influence that these microzones have on timing and time perception (e.g., motor versus perceptual timing).

Computer simulations of the cerebellum suggest that this structure is ideal for the implementation of supervised, or even sequential supervised learning (SL), but performs poorly in reinforcement learning (RL – [130]. In these simulations, the difference between RL and SL is defined by the size of the delay between a required action and the error signal. RL is able to survive relatively long delays, whereas SL begins to fall apart with any delay. Moreover, the cerebellum does well when the required action overlaps with the error signal (i.e., from the climbing fiber), and the prediction of the response ranges from 100 ms to 1500 ms. The range of delays over which learning can occur seem to be contingent upon the timing between granule cell input to PCs and the 'teaching signal' coming from climbing fibers. Interestingly, while simulations of the cerebellum are capable of learning sequential patterns, the learning is agnostic to the order of the sequence [130]. Therefore, timing in the cerebellum may be more cumulative, rather than identifying unique temporal patterns.

Conclusions

While the cerebellum has traditionally been studied in isolation, such as in decerebrate preparations, it is gaining traction as a structure that adjusts processing in much of the brain. Accumulating evidence suggests that adaptive timing responses of PCs are sufficient for timing sub-second durations but rely on feedback from additional cortical and basal ganglia networks for timing supra-second durations. The temporal information supplied to the cerebellum for these longer durations is then sent back allowing for adjustments of cortical and subcortical circuits. Thus, the cerebellum should not be viewed as simply a sub-second motor timing network, but a component of a unified timing network spanning sub- as well as supra-second timing. Moving forward it is essential to study the cerebellum's role in both action timing and cognitive function, within the context of a larger integrated network (e.g., [100, 124, 131].

The role of the cerebellum in the control of timing of fast movements

Mario Manto, Hiroshi Mitoma

Fast, single-joint voluntary movements have been widely used to determine the physiological rules governing the velocity and accuracy of movements, both in monkeys and in humans [132, 133]. This is particularly relevant for the pathogenesis of cerebellar disorders because motor dysmetria is a cardinal feature of cerebellar ataxias [134]. Cerebellar dysmetria is particularly prominent for fast voluntary movements [135]. The most common form of dysmetria is hypermetria (overshoot of the target). Hypometria (undershoot) designates a premature arrest before the target [136] and is the prominent feature in some patients.

In humans, fast single-joint movements are characterized by a first agonist EMG burst (generating the acceleratory pulse), followed by a second burst in the antagonist muscle (providing the decelerator pulse), followed by a second burst in the agonist muscle (to reach the aimed target with accuracy) [137, 138]. The duration of first agonist burst is scaled according to the amplitude of the movement [139]. Learning of fast movements is associated with kinematic changes. During motor learning in healthy subjects, the first parameter which changes is reaction time, reaching quickly a steady baseline [140]. Time-related parameters (duration of acceleration, duration of deceleration, and movement duration) decrease at a slower rate.

A major defect in the timing of agonist-antagonist bursts has been unambiguously demonstrated in the monkey by cooling the dentate nucleus during the execution of a fast goal-directed movement [133, 141]. Physiologically, fast elbow flexions in monkeys show single-peaked velocities and a bi-/triphase EMG pattern in the couple agonist/antagonist muscle. During the cooling of the dentate nucleus, movements become ataxic with a terminal tremor. The analysis of the kinematic profiles shows that the magnitude of the acceleration drops, whereas the magnitude of the deceleration increases. The agonist burst has a decreased rate of rise, is smaller in magnitude and shows a prolongation in the duration of the burst. Slowness of movements is one of the clinical features in human cerebellar ataxias and is presumed to be due to a defect in the recruitment of alpha-motoneurons as a consequence of an increased inhibition in the motor cortex [142, 143]. A major finding of cerebellar hypermetria observed in monkeys is the delayed onset latency of the antagonist EMG burst. Identical observations have been made in humans in various cerebellar ataxias [144, 145]. Calcium channels in the cerebellum are involved in the control of the timing of agonist/antagonist discharges as shown by the model of hyperventilation in spinocerebellar ataxia type 6 (SCA6) [146]. The defect in the rate of rise of the EMG burst is not restricted to the agonist EMG burst, but can affect also the antagonist EMG burst [147]. Human cerebellar hypometria is associated with two concomitant mechanisms contributing

to the undershoot: the prolongation of the duration of the antagonist EMG activity and a reduction in the intensity of the agonist EMG activity.

The antagonistic muscle discharges are centrally generated and not simply produced by stretch reflexes, since (1) they occur at or even before movement onset, and (2) they are preserved in deafferented patients [148]. Impairments in the second EMG burst might be the result of impaired predictive control. The predictive nature is observed clearly in a task using external loads. The impaired timing of the antagonist EMG burst is sensitive to the inertia of the moving limb in the case of cerebellar dysfunction from acute cerebellar intoxication: adding a mass increases the delay of the antagonist burst [149]. This inability to adapt to unexpected changes of the mechanical state of the limb fits with the current prevailing hypothesis of a key role for the cerebellum in sensorimotor prediction. The cerebellum would estimate and predict the movement dynamics of the body and inform the cerebral cortex in particular via the dentato-thalamo-cortical circuit [150]. The difficulties of cerebellar patients to adapt to changes in the damping of the joints also argue for such a predictive role [151].

There is hope that transcranial DC stimulation (tDCS) may enter in the therapeutic options (including for rehabilitative approaches) for cerebellar ataxias in the coming years. tDCS can modulate activity in the dentato-thalamo-cortical circuit. For example, activation of inhibitory Purkinje cells results in attenuation of cerebellar facilitation through the nuclear efferent pathways. This mechanism is called cerebellum-brain inhibition (CBI). Application of tDCS over the cerebellum is associated with an improvement of ataxia scores and CBI in cerebellar patients [152]. A favorable effect on the timing of agonist/antagonist bursts has been reported with transcranial cerebello-cerebral DC stimulation (tCCDCS) in SCA2 [153]. However, these results need to be confirmed in a large sample of patients and a consensus is missing on the parameters of stimulation that need to be used. Another option which remains to be explored is the use of electrical stimulation to assist the antagonist muscle and modify the relative timing of agonist/antagonist bursts [154].

The cerebellum is also a key-player for the precise timing of fast multi-joint movements. A typical demonstration comes from the study of finger opening in overarm throwing. Using this paradigm, it has been demonstrated that cerebellar patients throw slowly with inaccuracy, show enhanced variability in hand trajectories, and exhibit an increased variability in the timing, amplitude, and velocity of finger opening [155]. Finger opening times and ball release times are markedly impaired in ataxic patients. The lack of coordination between proximal joint rotations and timing of finger opening contributes to the dysmetria of throws [156]. Reduction in joint velocities and in joint accelerations/decelerations are associated with an inability to exploit interaction torques [157]. When the emphasis is put on both speed and accuracy, cerebellar patients produce inappropriate muscle torques relative to the dynamic interaction torques, a factor contributing to the incoordination of the elbow and shoulder resulting in curved trajectories and target overshoot [158].

Overall, the detailed kinematic and EMG analysis of fast single-joint movements has demonstrated that the cerebellum plays a major role in the control of the timing parameters underlying the triphasic pattern of muscle discharges. Hyper- and hypometria can be attributed partially to an impairment in the predictive timing of the agonist/antagonist muscle activities. Timing is also a critical factor under cerebellar control for fast goal directed multi-joint movements.

Timing and Eyeblink Conditioning: Evidence from Studies in Humans

Marcus Gerwig

As indicated above, animal models have elucidated in great detail the role of the cerebellum in eyeblink conditioning, an established model of associative learning. Findings in humans, mainly using delay eyeblink conditioning, are in very good agreement with those in animals [23, 159]. Patients with either cerebellar degeneration or focal cerebellar disorders are impaired in their ability to acquire classically conditioned eyeblink responses (CRs) [160-163]. By using high-resolution magnetic resonance imaging (MRI) in healthy subjects a significant correlation between cerebellar volume and the ability to acquire CRs was reported [164]. This was significantly related to the volume of the posterior lobe including lobule VI [165]. In patients with focal cerebellar lesions, eyeblink conditioning was significantly reduced in subjects with lesions including lobule Crus I and above [163]. Again, voxel-based lesion-symptom mapping (VLSM) analysis as well as functional MRI studies revealed a particular association between hemispheric lobule VI in the acquisition of CRs [166-169]. Animal experiments show that the interposed nuclei, but not the dentate or fastigial nuclei, are critically involved in the acquisition of CRs [170]. Human data are sparse because human lesion models with circumscribed affection of the cerebellar nuclei are lacking [171].

In eyeblink conditioning a basic aspect centers on the exact timing of CRs. Evidence that the human cerebellum may be involved in CR timing comes from dual-task studies [6] showing a selective interference between timed-interval tapping and delay eyeblink conditioning in healthy subjects [172, 173]. As already reported in early behavioral studies [174, 175] CR timing is important for the acquisition of CRs in healthy human subjects. After repeated presentation of the paired CS and US, subjects learn to lower the eyelid with a high temporal precision reaching the maximum amplitude close to the time of the US onset so that the eye is closed when the air puff arrives. The peak eyelid closure occurs independent of the CS-US interval. If the interval between conditioned and US is prolonged, the CR is delivered adaptively timed [176].

Disrupted CR timing has been found in patients with cerebellar disorders [166]. Compared to healthy controls, CRs occur significantly earlier in subjects with cerebellar cortical degeneration and with lesions of superior parts of the cerebellar hemisphere. In the rabbit, short fixed CR onset latencies

have been shown following large cortical lesions which involved the anterior lobe [13, 177]. Corresponding to these animal findings, VLSM analysis in focal cerebellar patients revealed that both CR onset and peak time was significantly earlier in subjects with cortical lesions including parts of the ipsilateral anterior lobe, in particular hemispheric lobule V [166]. However, findings need to be validated in larger groups of patients. Disordered timing with significantly earlier peak time CR latencies has also been reported in alcoholic patients and in abstinent chronic alcoholics in whom cortical degeneration of the anterior cerebellar lobe is to be expected [176]. Results suggest that cortical areas of the superior cerebellum may be involved in timing of conditioned eyeblink responses in humans. These areas appear to be separated from those that are most important for the CS-US association, in particular lobule HVI.

Animal studies have investigated the timing of conditioned eyeblink responses from the behavioral to the molecular level. It has been suggested that appropriate CR timing depends on the cerebellar cortex of the anterior lobe [178], whereas others argue that changes in CR timing may equally be explained by extracerebellar premotorneuronal disinhibition [179]. Recent data suggest that Purkinje cells appear to be equipped with an intrinsic cellular mechanism creating a memory of the time between the CS and US independent of external time codes showing that the cerebellar cortex itself learns to emit adaptively timed movements [101, 108].

In contrast to findings of shortened latencies, two previous human lesion studies reported a tendency of CRs to be delayed [161, 162]. However, while in animal studies lesions are applied to distinct structures in previously trained subjects, findings in humans are based on the remaining CRs in subjects with cerebellar diseases and primarily reduced CR acquisition. It has to be noted that the above human studies differed due to CR analysis in paired or unpaired trials, extent of lesions and additional damage of cerebellar nuclei.

Acquisition and timing of conditioned responses is impaired even after multiple sessions of conditioning [180, 181]. Neither degenerative nor focal cerebellar patients showed a significant increase of CRs across training sessions within three days. Disordered timing with shortened CR onset and peak time latencies was most marked in the degenerative group with no improvement across sessions. In patients with focal lesions CR timing deficits improved to normal values from the first to the third day as compared to healthy controls. A possible explanation is that the cortex of the anterior lobe was not affected in the majority of focal patients. Conclusions on CR timing, however, are limited because of the reduced number of CRs in particular in the degenerative patients.

The simple application and good agreement between animal and human studies suggest a direct comparison of cerebellar dysfunction in animal models of cerebellar disease and the corresponding human patient's populations. For example, there are increasing numbers of mouse models of hereditary cerebellar disease. Furthermore, eyeblink conditioning was shown helpful to detect cerebellar

dysfunction even in a subclinical extent in various diseases. Impaired CR acquisition has been interpreted as evidence of a cerebellar contribution to essential tremor [182], dystonia [183], fragile X syndrome [184], migraine [185] and neuropsychiatric disorders including autism, schizophrenia, dyslexia, attention-deficit hyperactivity disorder (ADHD) [186, 187]. Corresponding to animal data the most robust marker of impaired eyeblink conditioning is reduced or abolished CR acquisition [188, 189]. Compared to controls, patients with schizophrenia showed fewer CR incidences and longer CR onset latencies while shifting of the interstimulus interval did not influence CR rates in both groups [190]. Reduced CR incidences are not always accompanied by disordered CR timing and patterns of impairment may differ. Also, in patients with essential tremor and in patients with migraine CR timing deficits were not reported [182, 185]. In children with ADHD, CRs occurred significantly earlier in a long interstimulus interval compared to controls [187]. Findings are in accordance with an animal model of ADHD [191] showing shortened latencies of CRs. As suggested in cerebellar lesion studies, the acquisition and timing of CRs depends on different areas within the cerebellar cortex which may be variably involved in the pathology of various diseases.

Beyond cerebellar lesion and imaging studies, transcranial direct current stimulation (tDCS) has been used in healthy subjects to assess whether acquisition and timing of conditioned eyeblink responses is modulated by this form of noninvasive stimulation. The main findings of a first study on CR acquisition showed a polarity specific effect, with significant enhancement following anodal tDCS and significant attenuation following cathodal stimulation compared to sham [192]. Moreover, during anodal tDCS, CR onset occurred increasingly earlier, with the mean onset of responses shifted closer to CS onset. In the cathodal stimulated subjects, CR onset appeared to be delayed but CRs were markedly less; thus, clear conclusions on timing data during cathodal stimulation could not be drawn. However, the shifting of the initiation of the CR closer to CS onset during anodal stimulation does not mean that the timing is less adaptive. Earlier studies report that the first CRs were initiated just before the US, but then CR initiation rapidly shifted to progressively earlier portions of the CS-US interval [175, 193]. However, a follow-up study did not reveal clear polarity dependent effects of cerebellar tDCS on CR acquisition and timing as previously described [194]. It is poorly understood why cerebellar tDCS effects on eyeblink conditioning are largely variable. Thresholds and current flow based on individual anatomy may play a role. Furthermore, individual variation in transmitter levels or genetic polymorphisms may also be relevant as recently shown for brain derived neurotrophic factor (BDNF) [195, 196].

In conclusion, disturbed timing of conditioned eyeblink responses has been reported in various studies investigating patients with cerebellar disorders and other neurological diseases. Future studies using eyeblink conditioning models in humans may help to better understand the role of timing in associative motor learning.

Timing in movement disorders (Parkinson's disease and Dystonia)

Laura Avanzino

Since some years ago, the basal ganglia and cerebellum were viewed as non-interacting neural structures, both involved in motor control, but the former more at a level of ideation and planning whereas the second more in coordination. Nowadays this view is recognized as too simplistic. Recent evidence showed anatomical bilateral connections between the two structures in animal models [197-199] and in humans by means of MRI [200, 201]. Thus, a functional connectivity in a variety of tasks, from sensory processing to motor control to cognitive functions has been hypothesized. In particular, this concept has been expanded to physiology of the processing of timing information and indeed evidence suggests that both basal ganglia and cerebellum are involved in this process [202-204].

Translating to the clinic, movement disorders that were previously ascribed to basal ganglia are now recognized as system-level or network disorders [205-207]. I will first discuss Parkinson's disease (PD), a neurodegenerative disorder characterized by motor dysfunctions including, among others, tremor, bradykinesia, rigidity as well as non-motor functions. PD dysfunctions were thought to result primarily from degeneration of dopamine-producing cells in the substantia nigra pars compacta, an area in the midbrain mainly targeting the striatum, the input nucleus of the basal ganglia. However, evidence collected in the last years showed that the cerebellum has structural and functional modulations in PD patients [208] that may contribute to clinical symptoms of PD like tremor [209] or impairment in dual task performance [210]. A similar picture is emerging in studies of dystonia, a movement disorder that is characterized by involuntary muscle contraction, abnormal movements and posture. Traditionally, dystonia has been considered a disorder of the basal ganglia. However, recent evidence points to a pathophysiological role of the cerebellum [211, 212].

Here we will summarize the most striking evidence related to timing abnormalities in PD and dystonia and we will discuss a possible pathophysiological role of cerebellum in these abnormalities. An important question remains concerning whether the cerebellar links to these disorders is (i) primary, arising from neurodegenerative/dysfunction process; (ii) secondary, reflecting an abnormal drive from the malfunctioning basal ganglia or (iii) compensatory due to basal ganglia dysfunction.

Parkinson's disease

In PD, abnormal timing performance is already evidenced in one of the main clinical characteristics, the impaired sequencing of motor action. Bradykinesia (slowness of movement initiation and execution) is particularly evident for internally generated sequential movements [213, 214] and commonly occurs in gait [215, 216]. Furthermore, a large amount of experimental evidence demonstrates that PD patients

are impaired in a variety of timing tasks, ranging from perceptual to production timing abnormalities and from implicit to explicit timing tasks (for a review see [217]).

Starting from explicit timing tasks, by means of the synchronization or the synchronization-continuation paradigms, several studies have shown abnormalities in finger tapping in PD even during the planning phase of movement [218], that become even more striking during execution [219-222]. Altogether, the continuation phase highlighted major differences between PD patients and controls likely reflecting the difficulties of PD subjects with internally generated movements.

Neuroimaging studies supported a possible role of the cerebellum in timing abnormalities in PD. Elsinger and co-workers reported decreased activation within the sensorimotor cortex, cerebellum, and medial premotor system in PD patients off-levodopa compared to controls during paced finger tapping that was partially “normalized” by dopaminergic therapy [223]. Differently, in a group of PD patients off-levodopa, enhanced activation of the cerebellum and of the supplementary motor area was shown with fMRI [224] and PET [225] during a synchronization-continuation finger-tapping task. Further, Jahanshahi and coworkers [225] showed that cortical activation was significantly more predominant when patients were in the on-medication state, whereas cerebellar activations were higher in the off-medication state. All these findings are highly heterogeneous and currently do not help in discerning the role of cerebellum in timing abnormalities in PD.

However, interesting hints come from predictive timing studies. Predictive timing refers to those tasks when temporal information is processed to predict the outcome of self-executed or externally driven movements in an implicit fashion [16]. Cerebellar pathways seem to be largely involved in predictive timing. Indeed, when temporal information inherent to the spatial-temporal trajectory of a dynamic visual stimulus was used to predict its final position, fMRI studies revealed activation in different cortical areas [226, 227], and in the cerebellum [228, 229].

In a set of important studies, Bares and co-workers showed that, unlike patients with cerebellar ataxia and essential tremor, patients with PD do not exhibit impaired motor timing during a task requiring the interception of a moving target [24, 120, 121]. However, the same authors showed subtle differences between PD patients in an early-stage disease and in an off-medication state and healthy controls [230]. Indeed, PD patients had trouble postponing their actions until the right moment and adapting from one trial to the next after these failures. Task performance was accompanied by fMRI activations in both the basal ganglia and cerebellum in controls, with cerebellum associated exclusively with postponement of action until the right moment, and both basal ganglia and cerebellum needed for performance adaptation. PD patients showed a “hypoactivation” in cerebellum and striatum relative to controls [230]. Further, by using dynamic casual modelling to investigate effective connectivity between

supplementary motor area, basal ganglia and cerebellum during the same interceptive task, different connectivity patterns were observed between the PD patients and controls [231].

Dystonia

Timing has been assessed in task-specific forms of dystonia, generated by movements like musical playing or writing and in non-task-specific forms of dystonia [204]. Kinematic analyses of scales or finger tapping performed on a digital piano by pianists with dystonia showed inaccuracies in tone and interval duration and rhythmic inconsistency [232, 233]. However, some measures improved with botulinum toxin therapy, supporting the idea that abnormal performance may represent a consequence of the motor overflow during musical performance [233, 234]. Furthermore, explicit timing performance in these patients appeared similar to controls [235], suggesting that processing of temporal properties while performing timed movements may be preserved in dystonia.

Related to predictive motor timing, Avanzino and co-workers [236] showed that the cerebellum is engaged when temporal information is used to predict the temporal outcome of a motor act. The authors developed an *ad hoc* task in which participants were required to observe a movement in a video and predict the end of the same movement. Crucially, a few seconds after its onset, the video was darkened for a given time interval; thus the task could only be performed by extrapolating time-related features of observed motion sequence. When lateral cerebellar activity was inhibited with 1Hz-repetitive transcranial magnetic stimulation, a deterioration of timing performance selectively for the estimation of a movement involving a body segment (handwriting) and not an inanimate object was observed [206]. The same task was applied in patients with focal dystonia [237, 238]. Patients with either task-specific (writer's cramp) [237] or non-task-specific (cervical) dystonia [238] were less accurate in predicting the temporal outcome of a visually perceived movement, and this was observed for human body, but not inanimate object motion.

In another paradigm, it has been shown that cervical dystonia patients were impaired in predictive motor timing when they were asked to mediate the interception of a moving target [123]. Interestingly, through functional MRI imaging techniques, the same authors showed cerebellar hypo-activity and connectivity with the basal ganglia and the motor cortex during this task performance in cervical dystonia patients [239]. Taken together, such deficits may be linked to an abnormal internal model of motor commands reflecting dysfunction of cerebellar outflow pathways. Consistent with this hypothesis, Avanzino and co-workers have recently demonstrated deficits in anticipatory movement control in patients with cervical dystonia [240]. The term 'anticipatory' indicates the feed-forward portion of a movement that is planned in advance and relies on the internal model of motor act. Interestingly, the results showed that abnormal anticipatory control was observed only in a subgroup of patients with

cervical dystonia who presented with tremor in the dystonic or not dystonic body parts, thus suggesting that the cerebellum might play a specific role in the occurrence of dystonic tremor. This hypothesis requires further study.

In summary, there is broad evidence that timing properties are impaired in patients with PD and dystonia, and that cerebellum may be involved in these abnormalities. To date, these findings do not help in discerning between the “primary”, “secondary”, or “compensatory” hypothesis related to the role of cerebellum. Future studies are needed to better define the contribution of cerebellum with the aim of improving the therapeutic approaches for these conditions.

Timing in Movement Disorders: Essential Tremor

Pavel Filip, Elan D. Louis

Essential tremor (ET) is one of the most common movement disorders. Its hallmark feature is slowly progressive kinetic tremor [241], predominantly in the forearms and hands, which often spread to other body regions [242, 243]. The pathophysiological mechanisms of ET are not completely understood. While simple models of single centers or individual loops do not provide feasible explanations consistent with the clinical expression of the disease, including its non-motor aspects [244], converging evidence from imaging [245], animal model [246], clinical [247], physiological [248] and neuropathological studies [249] implicates defects in a complex network encompassing a significant part of the motor system, mainly the oscillatory system involving cerebello-thalamo-cortical loops. This network, with varying engagement of the individual components, repeatedly appears in modern theories about internal time representation, which envisage the processing of temporal information as mediated by distributed timing models, which derive data from the coincidental activation of various neural populations [250, 251]. This vast spectrum of models postulates the representation of time as ubiquitous, encoded in the entire activity pattern in more networks, which also process other stimulus properties, with relative dominance of several nodes, including the cerebellum, for sub-second timing tasks [252].

Hence, the presence of a dynamic oscillatory disturbance in ET in this network may lead to an impairment in tasks requiring exact timing. Nonetheless, studies analyzing reaction time produce discrepant results on measures using either mean reaction time and movement time and an impaired performance in a visual reaction time-based task [253-256] in ET patients and control subjects. The same inconsistency may be found in repetitive movements spanning a larger scale with significant deterioration reported in one study [257], but no difference found in the speed of alternating pronation-supination movements [256]. On the other hand, the confirmed impairment in the performance of tasks based on rhythmic repetitive finger movements, with longer touch duration,

shorter tapping interval and increased temporal variability of movement [256, 258], may be interpreted as a sign of cerebellar dysfunction and defective temporal processing, including the disrupted synchronization with extrinsic timing signals. These conclusions are corroborated by a severe impairment in event-based rhythm generation in ET [259], providing firm ground to support the hypothesized defect in intrinsic temporal processes. In addition to the retrospective timing domain captured by the above stated studies, a predictive motor timing task based on the interception of a moving object, a seemingly simple, quotidian activity ensuring synchrony and reliable interaction with our surrounding environment, revealed a markedly disrupted performance in ET patients [121, 122].

Unfortunately, kinematic studies of arm and hand movements in ET are inherently contaminated by tremor, the main clinical feature of the disease. However, eye motion analyses are able to provide information devoid of tremor artefacts. A deficit in smooth pursuit initiation [260], eye-hand coordination [261] and abnormally prolonged latencies with lower velocity profiles [262] all provide further evidence of timing disruptions in ET, even in these seemingly simple movements.

Furthermore, a temporal deficit may lead to one of the hallmarks of ET, the terminal accentuation of tremor at the end of precise movements. The triphasic pattern of electromyogram (EMG) activity (the first agonist burst to initiate the movement, followed by the antagonist burst to decelerate, and the second agonist burst to attenuate oscillations induced by the deceleration [263]) has been repeatedly shown to be abnormal in ET patients, with disruptions of timing of burst discharges. In particular, delays are seen in the onset of the phasic activity of the second agonist muscle, leaving the antagonist unopposed for a longer period, thus resulting in a series of dampened oscillations at the target point [264, 265]. Moreover, the latency of the second agonist burst in EMG correlates with the tremor period [264].

Interestingly, the impairment of temporal processing may be partially reversible. Repetitive transcranial magnetic stimulation over the cerebellum was able to significantly improve finger tapping performance in ET [258] and both thalamotomy and deep brain stimulation of the ventral intermediate nucleus of thalamus virtually restored perturbations in internal timing mechanisms, in the range of hundreds of milliseconds, to normal [266].

We are far from a clear understanding of the underlying mechanisms triggering the oscillations and changes in time processing networks in ET. The current evidence suggests that these networks share certain nodes. There is even a distinct improvement of the performance of ET patients in the temporal realm when utilizing complex therapeutic methods. However, both a definite model completely explaining the processing of temporal inputs in our brain and a cure for the primary cause of ET, are still out of reach.

Summary of Concepts

The current Consensus paper highlights the importance of the cerebellum in the control of elementary mechanisms of timing, not only at a cellular level but also for brain networks. Both the anatomy of the cerebellum with a geometric repetition of microcircuits and its dense connectivity with cerebral cortex, basal ganglia, brainstem and spinal cord make of the “small brain” an ideal candidate to coordinate elemental events required for optimal motor control and beyond. The theoretical concept of the cerebellum as a timing device raised by Braitenberg in 1961 has now expanded to clinical disorders [267].

However, the exact nature of more complex operations and processes running in the cerebellum remains elusive. Even though the cerebellum, with its myriads of neurons, definitely possesses the crude computing power to delineate expected future states of both the external environment and the body itself, it is not completely clear whether these processes are really intrinsic to the cerebellar cortex or emerge from a network, where cerebellum is but a mere node. Indeed, its vast interaction with basal ganglia, cortex, thalamus and other parts of the brain and feedforward character of data processing may imply the later. Nonetheless, the character of the cerebellar granular layer might point to the first option, as it is more than suitable for timing operations with millisecond precision and its oscillations and resonance in the theta band may be the basis for plasticity necessary in the learning of precise timing actions. At the intersection of this ostensible discrepancy and possibly providing its solution, the ICAT (Initiation, Continuation, Adjustment, and Termination) model of temporal processing poses the Initiation and Adjustment phase to the cerebellum, with Continuation and Termination phases being governed mainly by supratentorial structures as described above by Petter et al. [118] and illustrated in Figure 2. This position of cerebellum may be one of the reasons why a broad spectrum of timing disturbances has been implied in not only essential tremor, but also in Parkinson’s disease and dystonia, even though its role in these pathologies, be it primary, secondary or compensatory, still remains unclear.

Nonetheless, this overview of the position of the cerebellum in the complex timing processes is far from complete. There has been an undisputable progress in the spread of the general idea that the cerebellum is a structure critical not only for the precision of movement, but virtually for every cerebral process requiring feedback and fine-tuning. Hence, we are slowly shifting from simplistic views of neocortex as the primary driver in more complicated domains to a new paradigm conceptualizing an integrated network of distinct brain regions, including some maybe counter-intuitive areas, as the notional “mirror” of human psyche.

Table 1 **Consensus to the Position of Cerebellum in Timing Processes:**
Agreement and Warranted Research

<p>Agreement</p>	<p>The cerebellum is an important node in multiple domains of temporal analysis, including motor and non-motor timing, implicit and explicit timing, but mostly in discrete, isolated intervals.</p> <p>The cerebellum should not be viewed as a mere sub-second processing node, it provides important contribution in the supra-second spectrum, allowing for adjustments of cortical and subcortical circuits.</p> <p>The cerebellum plays a major role in the control of the tri-phasic patten of muscle discharges.</p> <p>Crucial role of the cerebellum in the predictive timing in multiple domains – eyeblink, interception tasks, locomotion phases, sequential movements.</p>
<p>Warranted research</p>	<p>Precise identification of cerebellar pathways and</p> <p>Contribution of cerebellum in various neurodegenerative disorders – primary, secondary or compensatory role?</p> <p>New perspectives in both neurological and psychiatric disorders traditionally viewed as not linked to cerebellum</p> <p>Studies with non-invasive brain stimulation, both in research and clinical treatment</p>

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References

1. Baumann O, Borra RJ, Bower JM, Cullen KE, Habas C, Ivry RB, et al. Consensus paper: the role of the cerebellum in perceptual processes. *Cerebellum*. 2015;14(2):197-220.
2. Koziol LF, Budding D, Andreasen N, D'Arrigo S, Bulgheroni S, Imamizu H, et al. Consensus paper: the cerebellum's role in movement and cognition. *Cerebellum*. 2014;13(1):151-77.
3. Mariën P, Ackermann H, Adamaszek M, Barwood CHS, Beaton A, Desmond J, et al. Consensus paper: language and the cerebellum: an ongoing enigma. *Cerebellum*. 2014;13(3):386-410.
4. Adamaszek M, D'agata F, Ferrucci R, Habas C, Keulen S, Kirkby KC, et al. Consensus paper: cerebellum and emotion. *Cerebellum*. 2017;16(2):552-76.
5. Watson PJ. Nonmotor functions of Cerebellum. *Psychol Bull*. 1978;85(5):944-67.
6. Ivry RB, Keele SW. Timing functions of Cerebellum. *J Cogn Neurosci*. 1989;1(2):136-52.
7. Ivry RB, Schlerf JE. Dedicated and intrinsic models of time perception. *Trends Cogn Sci*. 2008;12(7):273-80.
8. Bengtsson SL, Ehrsson HH, Forssberg H, Ullén F. Effector-independent voluntary timing: behavioural and neuroimaging evidence. *Eur J Neurosci*. 2005;22(12):3255-65.
9. Coull JT, Nobre AC. Where and when to pay attention: the neural systems for directing attention to spatial locations and to time intervals as revealed by both PET and fMRI. *J Neurosci*. 1998;18(18):7426-35.
10. Soares S, Atallah BV, Paton JJ. Midbrain dopamine neurons control judgment of time. *Science*. 2016;354(6317):1273-7.
11. Merchant H, Pérez O, Zarco W, Gámez J. Interval tuning in the primate medial premotor cortex as a general timing mechanism. *J Neurosci*. 2013;33(21):9082-96.
12. Finnerty GT, Shadlen MN, Jazayeri M, Nobre AC, Buonomano DV. Time in cortical circuits. *J Neurosci*. 2015;35(41):13912-6.
13. Perrett SP, Ruiz BP, Mauk MD. Cerebellar cortex lesions disrupt learning-dependent timing of conditioned eyelid responses. *J Neurosci*. 1993;13(4):1708-18.
14. Spencer R, Zelaznik HN, Diedrichsen J, Ivry RB. Disrupted timing of discontinuous but not continuous movements by cerebellar lesions. *Sci Signal*. 2003;300(5624):1437.
15. Breska A, Ivry RB. Taxonomies of timing: where does the cerebellum fit in? *Curr Opin Behav Sci*. 2016;8:282-8.
16. Coull JT, Nobre AC. Dissociating explicit timing from temporal expectation with fMRI. *Curr Opin Neurobiol*. 2008;18(2):137-44.
17. Diedrichsen J, Ivry RB, Pressing J. Cerebellar and basal ganglia contributions to interval timing. *Functional and neural mechanisms of interval timing*. 2003:457-81.
18. Franz EA, Ivry RB, Helmuth LL. Reduced timing variability in patients with unilateral cerebellar lesions during bimanual movements. *J Cogn Neurosci*. 1996;8(2):107-18.
19. Bueti D, Walsh V, Frith C, Rees G. Different brain circuits underlie motor and perceptual representations of temporal intervals. *J Cogn Neurosci*. 2008;20(2):204-14.

20. Rao SM, Harrington DL, Haaland KY, Bobholz JA, Cox RW, Binder JR. Distributed neural systems underlying the timing of movements. *J Neurosci*. 1997;17(14):5528-35.
21. Hove MJ, Fairhurst MT, Kotz SA, Keller PE. Synchronizing with auditory and visual rhythms: an fMRI assessment of modality differences and modality appropriateness. *Neuroimage*. 2013;67:313-21.
22. Johansson F, Jirnhed D-A, Rasmussen A, Zucca R, Hesslow G. Memory trace and timing mechanism localized to cerebellar Purkinje cells. *Proc Natl Acad Sci*. 2014;111(41):14930-4.
23. Gerwig M, Kolb FP, Timmann D. The involvement of the human cerebellum in eyeblink conditioning. *Cerebellum*. 2007;6(1):38-57.
24. Bares M, Lungu OV, Liu T, Waechter T, Gomez CM, Ashe J. The neural substrate of predictive motor timing in spinocerebellar ataxia. *Cerebellum*. 2011;10(2):233-44.
25. Spencer RMC, Verstynen T, Brett M, Ivry R. Cerebellar activation during discrete and not continuous timed movements: an fMRI study. *Neuroimage*. 2007;36(2):378-87.
26. Tregellas JR, Davalos DB, Rojas DC. Effect of task difficulty on the functional anatomy of temporal processing. *Neuroimage*. 2006;32(1):307-15.
27. Mathiak K, Hertrich I, Grodd W, Ackermann H. Discrimination of temporal information at the cerebellum: functional magnetic resonance imaging of nonverbal auditory memory. *Neuroimage*. 2004;21(1):154-62.
28. Ackermann H, Gräber S, Hertrich I, Daum I. Cerebellar contributions to the perception of temporal cues within the speech and nonspeech domain. *Brain Lang*. 1999;67(3):228-41.
29. Grube M, Cooper FE, Chinnery PF, Griffiths TD. Dissociation of duration-based and beat-based auditory timing in cerebellar degeneration. *Proc Natl Acad Sci*. 2010;107(25):11597-601.
30. Teki S, Grube M, Kumar S, Griffiths TD. Distinct neural substrates of duration-based and beat-based auditory timing. *J Neurosci*. 2011;31(10):3805-12.
31. Buetti D, Bahrami B, Walsh V, Rees G. Encoding of temporal probabilities in the human brain. *J Neurosci*. 2010;30(12):4343-52.
32. Niemi P, Näätänen R. Foreperiod and simple reaction time. *Psychol Bull*. 1981;89(1):133.
33. Trillenber P, Verleger R, Teetzmann A, Wascher E, Wessel K. On the role of the cerebellum in exploiting temporal contingencies: evidence from response times and preparatory EEG potentials in patients with cerebellar atrophy. *Neuropsychologia*. 2004;42(6):754-63.
34. Henry MJ, Herrmann B, Obleser J. Entrained neural oscillations in multiple frequency bands comodulate behavior. *Proc Natl Acad Sci*. 2014;111(41):14935-40.
35. Breska A, Deouell LY. Automatic bias of temporal expectations following temporally regular input independently of high-level temporal expectation. *J Cogn Neurosci*. 2014;26(7):1555-71.
36. Geiser E, Zaehle T, Jancke L, Meyer M. The neural correlate of speech rhythm as evidenced by metrical speech processing. *J Cogn Neurosci*. 2008;20(3):541-52.
37. Grahn JA, Rowe JB. Finding and feeling the musical beat: striatal dissociations between detection and prediction of regularity. *Cereb Cortex*. 2012;23(4):913-21.
38. Ivry RB, Spencer RM, Zelaznik HN, Diedrichsen J. The cerebellum and event timing. *Ann N Y Acad Sci*. 2002;978(1):302-17.

39. Teki S, Grube M, Griffiths TD. A unified model of time perception accounts for duration-based and beat-based timing mechanisms. *Front Integr Neurosci.* 2012;5:90.
40. Lakatos P, Musacchia G, O'Connell MN, Falchier AY, Javitt DC, Schroeder CE. The spectrotemporal filter mechanism of auditory selective attention. *Neuron.* 2013;77(4):750-61.
41. Jones MR. Attending to sound patterns and the role of entrainment. *Attention and time.* 2010:317-30.
42. Schroeder CE, Lakatos P. Low-frequency neuronal oscillations as instruments of sensory selection. *Trends Neurosci.* 2009;32(1):9-18.
43. Grahn JA, Brett M. Impairment of beat-based rhythm discrimination in Parkinson's disease. *Cortex.* 2009;45(1):54-61.
44. Breska A, Deouell LY. Neural mechanisms of rhythm-based temporal prediction: Delta phase-locking reflects temporal predictability but not rhythmic entrainment. *PLoS Biol.* 2017;15(2):e2001665.
45. Breska A, Deouell LY. When synchronizing to rhythms is not a good thing: modulations of preparatory and post-target neural activity when shifting attention away from on-beat times of a distracting rhythm. *J Neurosci.* 2016;36(27):7154-66.
46. Ekerot CF. Climbing fibres - a key to cerebellar function. *J Physiol.* 1999;516 (Pt 3):629.
47. Llinas R, Walton K, Hillman DE, Sotelo C. Inferior olive: its role in motor learning. *Science.* 1975;190(4220):1230-1.
48. Rondi-Reig L, Delhaye-Bouchaud N, Mariani J, Caston J. Role of the inferior olivary complex in motor skills and motor learning in the adult rat. *Neurosci.* 1997;77(4):955-63.
49. Horn KM, Deep A, Gibson AR. Progressive limb ataxia following inferior olive lesions. *J Physiol.* 2013;591(Pt 22):5475-89.
50. Wilson WC, Magoun HW. The Functional Significance of the Inferior Olive in the Cat. *J Comp Neurol.* 1945;83(1):69-77.
51. King RB. The olivo-cerebella system; the effect of interolivary lesions on muscle tone in the trunk and limb girdles. *J Comp Neurol.* 1948;89(3):207-23.
52. Eccles JC, Llinas R, Sasaki K. The excitatory synaptic action of climbing fibres on the Purkinje cells of Cerebellum. *J Physiol.* 1966;182(2):268-96.
53. Crepel F, Mariani J, Delhaye-Bouchaud N. Evidence for a multiple innervation of Purkinje cells by climbing fibers in the immature rat cerebellum. *J Neurobiol.* 1976;7(6):567-78.
54. Crepel F, Delhaye-Bouchaud N, Dupont JL. Fate of the multiple innervation of cerebellar Purkinje cells by climbing fibers in immature control, x-irradiated and hypothyroid rats. *Brain Res.* 1981;227(1):59-71.
55. Apps R, Atkins MJ, Garwicz M. Gating of cutaneous input to cerebellar climbing fibres during a reaching task in the cat. *J Physiol.* 1997;502 (Pt 1):203-14.
56. Apps R. Movement-related gating of climbing fibre input to cerebellar cortical zones. *Prog Neurobiol.* 1999;57(5):537-62.
57. Apps R, Hartell NA, Armstrong DM. Step phase-related excitability changes in spino-olivocerebellar paths to the c1 and c3 zones in cat cerebellum. *J Physiol.* 1995;483 (Pt 3):687-702.

58. Lawrenson CL, Watson TC, Apps R. Transmission of Predictable Sensory Signals to the Cerebellum via Climbing Fiber Pathways Is Gated during Exploratory Behavior. *J Neurosci*. 2016;36(30):7841-51.
59. Lidierth M, Apps R. Gating in the spino-olivocerebellar pathways to the c1 zone of the cerebellar cortex during locomotion in the cat. *J Physiol*. 1990;430:453-69.
60. Apps R, Lidierth M, Armstrong DM. Locomotion-related variations in excitability of spino-olivocerebellar paths to cat cerebellar cortical c2 zone. *J Physiol*. 1990;424:487-512.
61. Ekerot CF, Larson B. The dorsal spino-olivocerebellar system in the cat. I. Functional organization and termination in the anterior lobe. *Exp Brain Res*. 1979;36(2):201-17.
62. Tsukahara N, Toyama K, Kosaka K. Electrical activity of red nucleus neurones investigated with intracellular microelectrodes. *Exp Brain Res*. 1967;4(1):18-33.
63. Shapovalov AI. Neuronal organization and synaptic mechanisms of supraspinal motor control in vertebrates. *Rev Physiol Biochem Pharmacol*. 1975;72:1-54.
64. Marr D. A theory of cerebellar cortex. *J Physiol*. 1969;202(2):437-70.
65. Eccles JC. Circuits in the cerebellar control of movement. *Proc Natl Acad Sci*. 1967;58(1):336-43.
66. D'Angelo E, Casali S. Seeking a unified framework for cerebellar function and dysfunction: from circuit operations to cognition. *Front Neural Circuits*. 2013;6:116.
67. D'Angelo E, De Zeeuw CI. Timing and plasticity in the cerebellum: focus on the granular layer. *Trends Neurosci*. 2009;32(1):30-40.
68. D'Angelo E. Rebuilding cerebellar network computations from cellular neurophysiology. *Front Cell Neurosci*. 2010;4.
69. Castellazzi G, Palesi F, Bruno SD, Toosy AT, D'Angelo E, Wheeler-Kingshott CAM. Resting state fMRI during continuous cognitive processing reveals dynamical changes of brain networks involving cerebral cortex and cerebellum. *Front Cell Neurosci*. 2017.
70. D'Angelo E, De Filippi G, Rossi P, Taglietti V. Synaptic excitation of individual rat cerebellar granule cells in situ: evidence for the role of NMDA receptors. *J Physiol*. 1995;484(2):397-413.
71. Chadderton P, Margrie TW, Hausser M. Integration of quanta in cerebellar granule cells during sensory processing. *Nature*. 2004;428(6985):856-60.
72. Arleo A, Nieuwenhuis T, Bezzi M, D'Errico A, D'Angelo E, Coenen OJMD. How synaptic release probability shapes neuronal transmission: information-theoretic analysis in a cerebellar granule cell. *Neural Comput*. 2010;22(8):2031-58.
73. Mapelli L, Pagani M, Garrido JA, D'Angelo E. Integrated plasticity at inhibitory and excitatory synapses in the cerebellar circuit. *Front Cell Neurosci*. 2015;9.
74. D'Angelo E, Nieuwenhuis T, Maffei A, Armano S, Rossi P, Taglietti V, et al. Theta-frequency bursting and resonance in cerebellar granule cells: experimental evidence and modeling of a slow K⁺-dependent mechanism. *J Neurosci*. 2001;21(3):759-70.
75. Nieuwenhuis T, Sola E, Mapelli J, Saftenku E, Rossi P, D'Angelo E. LTP regulates burst initiation and frequency at mossy fiber-granule cell synapses of rat cerebellum: experimental observations and theoretical predictions. *J Neurophysiol*. 2006;95(2):686-99.

76. Solinas S, Nieuwenhuis T, D'Angelo E. A realistic large-scale model of the cerebellum granular layer predicts circuit spatio-temporal filtering properties. *Front Cell Neurosci.* 2010;4.
77. Jörntell H. Cerebellar physiology: links between microcircuitry properties and sensorimotor functions. *J Physiol.* 2017;595(1):11-27.
78. Goldfarb M, Schoorlemmer J, Williams A, Diwakar S, Wang Q, Huang X, et al. Fibroblast growth factor homologous factors control neuronal excitability through modulation of voltage-gated sodium channels. *Neuron.* 2007;55(3):449-63.
79. Dover K, Marra C, Solinas S, Popovic M, Subramaniam S, Zecevic D, et al. FHF-independent conduction of action potentials along the leak-resistant cerebellar granule cell axon. *Nature communications.* 2016;7.
80. Diwakar S, Magistretti J, Goldfarb M, Naldi G, D'Angelo E. Axonal Na⁺ channels ensure fast spike activation and back-propagation in cerebellar granule cells. *J Neurophysiol.* 2009;101(2):519-32.
81. Ramakrishnan KB, Voges K, De Propriis L, De Zeeuw CI, D'Angelo E. Tactile stimulation evokes long-lasting potentiation of purkinje cell discharge in vivo. *Front Cell Neurosci.* 2016;10.
82. Belluzzi O, Sacchi O, Wanke E. A fast transient outward current in the rat sympathetic neurone studied under voltage-clamp conditions. *J Physiol.* 1985;358(1):91-108.
83. Dieudonné S. Submillisecond kinetics and low efficacy of parallel fibre-Golgi cell synaptic currents in the rat cerebellum. *J Physiol.* 1998;510(3):845-66.
84. D'Angelo E, Rossi P, Armano S, Taglietti V. Evidence for NMDA and mGlu receptor-dependent long-term potentiation of mossy fiber-granule cell transmission in rat cerebellum. *J Neurophysiol.* 1999;81(1):277-87.
85. Mapelli L, Rossi P, Nieuwenhuis T, D'Angelo E. Tonic activation of GABAB receptors reduces release probability at inhibitory connections in the cerebellar glomerulus. *J Neurophysiol.* 2009;101(6):3089-99.
86. Nieuwenhuis TR, Mapelli L, D'Angelo E. Regulation of output spike patterns by phasic inhibition in cerebellar granule cells. *Front Cell Neurosci.* 2014;8.
87. Cesana E, Pietrajtis K, Bidoret C, Isope P, D'Angelo E, Dieudonné S, et al. Granule cell ascending axon excitatory synapses onto Golgi cells implement a potent feedback circuit in the cerebellar granular layer. *J Neurosci.* 2013;33(30):12430-46.
88. Subramaniam S, Perin P, Locatelli F, Masetto S, Solinas S, D'Angelo E. The mechanisms of late-onset synaptic responses in a realistic model of Unipolar Brush Cells. *BMC Neurosci.* 2013;14(1):P79.
89. Mugnaini E, Di MR, Jaarsma D. The unipolar brush cells of the mammalian cerebellum and cochlear nucleus: cytology and microcircuitry. *Prog Brain Res.* 1997;114:131-50.
90. Sgritta M, Locatelli F, Soda T, Prestori F, D'Angelo EU. Hebbian spike-timing dependent plasticity at the cerebellar input stage. *J Neurosci.* 2017;37(11):2809-23.
91. Garrido MI, Barnes GR, Kumaran D, Maguire EA, Dolan RJ. Ventromedial prefrontal cortex drives hippocampal theta oscillations induced by mismatch computations. *Neuroimage.* 2015;120:362-70.
92. D'Angelo E, Mapelli L, Casellato C, Garrido JA, Luque N, Monaco J, et al. Distributed Circuit Plasticity: New Clues for the Cerebellar Mechanisms of Learning. *Cerebellum.* 2016;15(2):139-51.

93. Masoli S, D'Angelo E. Synaptic Activation of a Detailed Purkinje Cell Model Predicts Voltage-Dependent Control of Burst-Pause Responses in Active Dendrites. *Front Cell Neurosci.* 2017;11:278.
94. Gandolfi D, Lombardo P, Mapelli J, Solinas S, D'Angelo E. Theta-frequency resonance at the cerebellum input stage improves spike timing on the millisecond time-scale. *Front Neural Circuits.* 2013;7.
95. Pisotta I, Molinari M. Cerebellar contribution to feedforward control of locomotion. *Front Hum Neurosci.* 2014;8.
96. Medina JF, Nores WL, Ohyama T, Mauk MD. Mechanisms of cerebellar learning suggested by eyelid conditioning. *Curr Opin Neurobiol.* 2000;10(6):717-24.
97. De Zeeuw CI, Yeo CH. Time and tide in cerebellar memory formation. *Curr Opin Neurobiol.* 2005;15(6):667-74.
98. Jenkinson N, Miall RC. Disruption of saccadic adaptation with repetitive transcranial magnetic stimulation of the posterior cerebellum in humans. *Cerebellum.* 2010;9(4):548-55.
99. Colnaghi S, Ramat S, D'Angelo E, Cortese A, Beltrami G, Moglia A, et al. Theta-burst stimulation of the cerebellum interferes with internal representations of sensory-motor information related to eye movements in humans. *Cerebellum.* 2011;10(4):711-9.
100. Cordes S, Meck WH. Ordinal judgments in the rat: An understanding of longer and shorter for suprasecond, but not subsecond, durations. *J Exp Psychol Gen.* 2014;143(2):710.
101. Johansson F, Hesslow G, Medina JF. Mechanisms for motor timing in the cerebellar cortex. *Curr Opin Behav Sci.* 2016;8:53-9.
102. Koch G, Oliveri M, Torriero S, Salerno S, Gerfo EL, Caltagirone C. Repetitive TMS of cerebellum interferes with millisecond time processing. *Exp Brain Res.* 2007;179(2):291-9.
103. Coull JT, Cheng RK, Meck WH. Neuroanatomical and Neurochemical Substrates of Timing. *Neuropsychopharmacology.* 2011;36(1):3-25.
104. Mauk MD, Buonomano DV. The neural basis of temporal processing. *Annu Rev Neurosci.* 2004;27:307-40.
105. Merchant H, Harrington DL, Meck WH. Neural basis of the perception and estimation of time. *Annu Rev Neurosci.* 2013;36:313-36.
106. Allman MJ, Teki S, Griffiths TD, Meck WH. Properties of the internal clock: first-and second-order principles of subjective time. *Annu Rev Psychol.* 2014;65:743-71.
107. Ivry RB, Keele SW, Diener HC. Dissociation of the lateral and medial cerebellum in movement timing and movement execution. *Exp Brain Res.* 1988;73(1):167-80.
108. Heiney SA, Kim J, Augustine GJ, Medina JF. Precise control of movement kinematics by optogenetic inhibition of Purkinje cell activity. *J Neurosci.* 2014;34(6):2321-30.
109. Jirenhed D-A, Rasmussen A, Johansson F, Hesslow G. Learned response sequences in cerebellar Purkinje cells. *Proc Natl Acad Sci.* 2017;114(23):6127-32.
110. Wetmore DZ, Jirenhed D-A, Rasmussen A, Johansson F, Schnitzer MJ, Hesslow G. Bidirectional plasticity of Purkinje cells matches temporal features of learning. *J Neurosci.* 2014;34(5):1731-7.

111. Gallistel CR, Wilkes JT. Minimum description length model selection in associative learning. *Curr Opin Behav Sci.* 2016;11:8-13.
112. Lusk NA, Petter EA, MacDonald CJ, Meck WH. Cerebellar, hippocampal, and striatal time cells. *Curr Opin Behav Sci.* 2016;8:186-92.
113. Ohmae S, Uematsu A, Tanaka M. Temporally specific sensory signals for the detection of stimulus omission in the primate deep cerebellar nuclei. *J Neurosci.* 2013;33(39):15432-41.
114. Ohmae S, Kunimatsu J, Tanaka M. Cerebellar roles in self-timing for sub-and supra-second intervals. *J Neurosci.* 2017;37(13):3511-22.
115. Broersen R, Onuki Y, Abdelgabar AR, Owens CB, Picard S, Willems J, et al. Impaired spatio-temporal predictive motor timing associated with spinocerebellar ataxia type 6. *PLoS One.* 2016;11(8):e0162042.
116. Coull JT, Davranche K, Nazarian B, Vidal F. Functional anatomy of timing differs for production versus prediction of time intervals. *Neuropsychologia.* 2013;51(2):309-19.
117. Callu D, El Massioui N, Dutrieux G, Brown BL, Doyere V. Cognitive processing impairments in a supra-second temporal discrimination task in rats with cerebellar lesion. *Neurobiol Learn Mem.* 2009;91(3):250-9.
118. Petter EA, Lusk NA, Hesslow G, Meck WH. Interactive roles of the cerebellum and striatum in sub-second and supra-second timing: Support for an initiation, continuation, adjustment, and termination (ICAT) model of temporal processing. *Neurosci Biobehav Rev.* 2016;71:739-55.
119. Rasmussen A, Jirenhed D-A. Learning and Timing of Voluntary Blink Responses Match Eyeblink Conditioning. *Sci Rep.* 2017;7(1):3404.
120. Bares M, Lungu O, Liu T, Waechter T, Gomez CM, Ashe J. Impaired predictive motor timing in patients with cerebellar disorders. *Exp Brain Res.* 2007;180(2):355-65.
121. Bareš M, Lungu OV, Husárová I, Gescheidt T. Predictive motor timing performance dissociates between early diseases of the cerebellum and Parkinson's disease. *Cerebellum.* 2010;9(1):124-35.
122. Bares M, Husarova I, Lungu OV. Essential tremor, the cerebellum, and motor timing: towards integrating them into one complex entity. *Tremor Other Hyperkinet Mov.* 2012;2:1-9.
123. Filip P, Lungu OV, Shaw DJ, Kasperek T, Bareš M. The Mechanisms of Movement Control and Time Estimation in Cervical Dystonia Patients. *Neural Plast.* 2013;2013.
124. Lungu OV, Bares M, Liu T, Gomez CM, Cechova I, Ashe J. Trial-to-trial adaptation: Parsing out the roles of cerebellum and bg in predictive motor timing. *J Cogn Neurosci.* 2016.
125. Bostan AC, Dum RP, Strick PL. Cerebellar networks with the cerebral cortex and basal ganglia. *Trends Cogn Sci.* 2013;17(5):241-54.
126. Parker KL, Kim YC, Kelley RM, Nessler AJ, Chen KH, Muller-Ewald VA, et al. Delta-frequency stimulation of cerebellar projections can compensate for schizophrenia-related medial frontal dysfunction. *Mol Psychiatry.* 2017.
127. Najafi F, Medina JF. Beyond "all-or-nothing" climbing fibers: graded representation of teaching signals in Purkinje cells. *Front Neural Circuits.* 2013;7.
128. Witter L, Rudolph S, Pressler RT, Lahlaf SI, Regehr WG. Purkinje cell collaterals enable output signals from the cerebellar cortex to feed back to Purkinje cells and interneurons. *Neuron.* 2016;91(2):312-9.

129. Dean P, Porrill J, Ekerot C-F, Jörntell H. The cerebellar microcircuit as an adaptive filter: experimental and computational evidence. *Nat Rev Neurosci*. 2010;11(1):30-43.
130. Hausknecht M, Li W-K, Mauk M, Stone P. Machine learning capabilities of a simulated cerebellum. *IEEE transactions on neural networks and learning systems*. 2017;28(3):510-22.
131. Raghavan RT, Prevosto V, Sommer MA. Contribution of cerebellar loops to action timing. *Curr Opin Behav Sci*. 2016;8:28-34.
132. Hallett M, Shahani BT, Young RR. EMG analysis of patients with cerebellar deficits. *J Neurol Neurosurg Psychiatry*. 1975;38(12):1163-9.
133. Flament D, Hore J. Movement and electromyographic disorders associated with cerebellar dysmetria. *J Neurophysiol*. 1986;55(6):1221-33.
134. Holmes G. The symptoms of acute cerebellar injuries due to gunshot injuries. *Brain*. 1917;40(4):461-535.
135. Manto MU, Hildebrand J, Jacqy J. Shift from hypermetria to hypometria in an aberrant recovery following cerebellar infarction. *J Neurol Sci*. 1998;157(1):42-51.
136. Manto M. Mechanisms of human cerebellar dysmetria: experimental evidence and current conceptual bases. *J Neuroeng Rehabil*. 2009;6(1):10.
137. Hallett M, Marsden CD. Ballistic flexion movements of the human thumb. *J Physiol*. 1979;294(1):33-50.
138. Hannaford B, Stark L. Roles of the elements of the triphasic control signal. *Exp Neurol*. 1985;90(3):619-34.
139. Berardelli A, Rothwell JC, Day BL, Kachi T, Marsden CD. Duration of the first agonist EMG burst in ballistic arm movements. *Brain Res*. 1984;304(1):183-7.
140. Flament D, Shapiro MB, Kempf T, Corcos DM. Time course and temporal order of changes in movement kinematics during learning of fast and accurate elbow flexions. *Exp Brain Res*. 1999;129(3):441-50.
141. Brooks VB, Kozlovskaya IB, Atkin A, Horvath FE, Uno M. Effects of cooling dentate nucleus on tracking-task performance in monkeys. *J Neurophysiol*. 1973.
142. Wild B, Klockgether T, Dichgans J. Acceleration deficit in patients with cerebellar lesions. A study of kinematic and EMG-parameters in fast wrist movements. *Brain Res*. 1996;713(1):186-91.
143. Di Lazzaro V, Restuccia D, Nardone R, Leggio MG, Oliviero A, Profice P, et al. Motor cortex changes in a patient with hemispherectomy. *Electroencephalogr Clin Neurophysiol*. 1995;97(5):259-63.
144. Manto M, Godaux E, Jacqy J. Cerebellar hypermetria is larger when the inertial load is artificially increased. *Ann Neurol*. 1994;35(1):45-52.
145. Manto M, Jacqy J, Hildebrand J, Godaux E. Recovery of hypermetria after a cerebellar stroke occurs as a multistage process. *Ann Neurol*. 1995;38(3):437-45.
146. Manto M-U, Bosse P. A second mechanism of increase of cerebellar hypermetria in humans. *J Physiol*. 2003;547(Pt 3):989.
147. Manto M, Godaux E, Jacqy J, Hildebrand J. Cerebellar hypermetria associated with a selective decrease in the rate of rise of antagonist activity. *Ann Neurol*. 1996;39(2):271-4.

148. Hore J, Wild B, Diener HC. Cerebellar dysmetria at the elbow, wrist, and fingers. *J Neurophysiol.* 1991;65(3):563-71.
149. Manto M, Godaux E, Jacquy J, Hildebrand JG. Analysis of cerebellar dysmetria associated with lithium intoxication. *Neurol Res.* 1996;18(5):416-24.
150. Therrien AS, Bastian AJ. Cerebellar damage impairs internal predictions for sensory and motor function. *Curr Opin Neurobiol.* 2015;33:127-33.
151. Manto M, Van Den Braber N, Grimaldi G, Lammertse P. A new myohaptic instrument to assess wrist motion dynamically. *Sensors.* 2010;10(4):3180-94.
152. Benussi A, Dell'Era V, Cotelli MS, Turla M, Casali C, Padovani A, et al. Long term clinical and neurophysiological effects of cerebellar transcranial direct current stimulation in patients with neurodegenerative ataxia. *Brain Stimul.* 2017;10(2):242-50.
153. Grimaldi G, Taib NOB, Manto M, Bodranghien F. Marked reduction of cerebellar deficits in upper limbs following transcranial cerebello-cerebral DC stimulation: tremor reduction and re-programming of the timing of antagonist commands. *Front Syst Neurosci.* 2014;8.
154. Huang Y-Z, Chang Y-S, Hsu M-J, Wong AMK, Chang Y-J. Restoration of central programmed movement pattern by temporal electrical stimulation-assisted training in patients with spinal cerebellar atrophy. *Neural Plast.* 2015;2015.
155. Timmann D, Watts S, Hore J. Failure of cerebellar patients to time finger opening precisely causes ball high-low inaccuracy in overarm throws. *J Neurophysiol.* 1999;82(1):103-14.
156. Timmann D, Watts S, Hore J. Causes of left-right ball inaccuracy in overarm throws made by cerebellar patients. *Exp Brain Res.* 2000;130(4).
157. Timmann D, Lee P, Watts S, Hore J. Kinematics of arm joint rotations in cerebellar and unskilled subjects associated with the inability to throw fast. *Cerebellum.* 2008;7(3):366.
158. Bastian AJ, Martin TA, Keating JG, Thach WT. Cerebellar ataxia: abnormal control of interaction torques across multiple joints. *J Neurophysiol.* 1996;76(1):492-509.
159. Woodruff-Pakand DS, Steinmetz JE. Past, Present, and Future of Human Eyeblink Classical Conditioning. *Eyeblink Classical Conditioning: Volume I: Springer; 2002.* p. 1-17.
160. Daum I, Schugens MM, Ackermann H, Lutzenberger W, Dichgans J, Birbaumer N. Classical conditioning after cerebellar lesions in humans. *Behav Neurosci.* 1993;107(5):748.
161. Topka H, Valls-Solé J, Massaquoi SG, Hallett M. Deficit in classical conditioning in patients with cerebellar degeneration. *Brain.* 1993;116(4):961-9.
162. Woodruff-Pak DS, Papka M, Ivry RB. Cerebellar involvement in eyeblink classical conditioning in humans. *Neuropsychology.* 1996;10(4):443.
163. Gerwig M, Dimitrova A, Kolb FP, Maschke M, Brol B, Kunnel A, et al. Comparison of eyeblink conditioning in patients with superior and posterior inferior cerebellar lesions. *Brain.* 2003;126(1):71-94.
164. Woodruff-Pak DS, Vogel RW, Ewers M, Coffey J, Boyko OB, Lemieux SK. MRI-assessed volume of cerebellum correlates with associative learning. *Neurobiol Learn Mem.* 2001;76(3):342-57.

165. Dimitrova A, Gerwig M, Brol B, Gizewski ER, Forsting M, Beck A, et al. Correlation of cerebellar volume with eyeblink conditioning in healthy subjects and in patients with cerebellar cortical degeneration. *Brain Res.* 2008;1198:73-84.
166. Gerwig M, Hajjar K, Dimitrova A, Maschke M, Kolb FP, Frings M, et al. Timing of conditioned eyeblink responses is impaired in cerebellar patients. *J Neurosci.* 2005;25(15):3919-31.
167. Ramnani N, Toni I, Josephs O, Ashburner J, Passingham RE. Learning-and expectation-related changes in the human brain during motor learning. *J Neurophysiol.* 2000;84(6):3026-35.
168. Cheng DT, Disterhoft JF, Power JM, Ellis DA, Desmond JE. Neural substrates underlying human delay and trace eyeblink conditioning. *Proc Natl Acad Sci.* 2008;105(23):8108-13.
169. Yeo CH, Hardiman MJ, Glickstein M. Classical conditioning of the nictitating membrane response of the rabbit. *Exp Brain Res.* 1985;60(1):99-113.
170. Christian KM, Thompson RF. Neural substrates of eyeblink conditioning: acquisition and retention. *Learn Mem.* 2003;10(6):427-55.
171. Timmann D, Konczak J, Ilg W, Donchin O, Hermsdörfer J, Gizewski ER, et al. Current advances in lesion-symptom mapping of the human cerebellum. *Neuroscience.* 2009;162(3):836-51.
172. Papka M, Ivry RB, Woodruff-Pak DS. Selective disruption of eyeblink classical conditioning by concurrent tapping. *Neuroreport.* 1995;6(11):1493-7.
173. Woodruff-Pak DS, Jaeger ME. Predictors of eyeblink classical conditioning over the adult age span. *Psychol Aging.* 1998;13(2):193.
174. Boneau CA. The interstimulus interval and the latency of the conditioned eyelid response. *J Exp Psychol.* 1958;56(6):464.
175. Ebel HC, Prokasy WF. Classical eyelid conditioning as a function of sustained and shifted interstimulus intervals. *J Exp Psychol.* 1963;65(1):52.
176. McGlinchey-Berroth R, Fortier CB, Cermak LS, Disterhoft JF. Temporal discrimination learning in abstinent chronic alcoholics. *Alcohol Clin Exp Res.* 2002;26(6):804-11.
177. Koekkoek SKE, Hulscher HC, Dortland BR, Hensbroek RA, Elgersma Y, Ruigrok TJH, et al. Cerebellar LTD and learning-dependent timing of conditioned eyelid responses. *Science.* 2003;301(5640):1736-9.
178. Mauk MD, Medina JF, Nores WL, Ohyama T. Cerebellar function: coordination, learning or timing? *Curr Biol.* 2000;10(14):R522-R5.
179. Attwell PJE, Ivarsson M, Millar L, Yeo CH. Cerebellar mechanisms in eyeblink conditioning. *Ann N Y Acad Sci.* 2002;978(1):79-92.
180. Bracha V, Zhao L, Wunderlich DA, Morrissy SJ, Bloedel JR. Patients with cerebellar lesions cannot acquire but are able to retain conditioned eyeblink reflexes. *Brain.* 1997;120(8):1401-13.
181. Gerwig M, Guberina H, Eßer AC, Siebler M, Schoch B, Frings M, et al. Evaluation of multiple-session delay eyeblink conditioning comparing patients with focal cerebellar lesions and cerebellar degeneration. *Behav Brain Res.* 2010;212(2):143-51.
182. Kronenburger M, Gerwig M, Brol B, Block F, Timmann D. Eyeblink conditioning is impaired in subjects with essential tremor. *Brain.* 2007;130(6):1538-51.

183. Teo JTH, Van De Warrenburg BPC, Schneider SA, Rothwell JC, Bhatia KP. Neurophysiological evidence for cerebellar dysfunction in primary focal dystonia. *J Neurol Neurosurg Psychiatry*. 2009;80(1):80-3.
184. Smit AE, Van Der Geest JN, Vellema M, Koekkoek SKE, Willemsen R, Govaerts LCP, et al. Savings and extinction of conditioned eyeblink responses in fragile X syndrome. *Genes Brain Behav*. 2008;7(7):770-7.
185. Gerwig M, Rauschen L, Gaul C, Katsarava Z, Timmann D. Subclinical cerebellar dysfunction in patients with migraine: Evidence from eyeblink conditioning. *Cephalalgia*. 2014;34(11):904-13.
186. Forsyth JK, Bolbecker AR, Mehta CS, Klaunig MJ, Steinmetz JE, O'Donnell BF, et al. Cerebellar-dependent eyeblink conditioning deficits in schizophrenia spectrum disorders. *Schizophr Bull*. 2010;38(4):751-9.
187. Frings M, Gaertner K, Buderath P, Gerwig M, Christiansen H, Schoch B, et al. Timing of conditioned eyeblink responses is impaired in children with attention-deficit/hyperactivity disorder. *Exp Brain Res*. 2010;201(2):167-76.
188. Yeo CH, Hesslow G. Cerebellum and conditioned reflexes. *Trends Cogn Sci*. 1998;2(9):322-30.
189. Bracha V. Role of the cerebellum in eyeblink conditioning. *Prog Brain Res*. 2004;143:331-9.
190. Bolbecker AR, Steinmetz AB, Mehta CS, Forsyth JK, Klaunig MJ, Lazar EK, et al. Exploration of cerebellar-dependent associative learning in schizophrenia: effects of varying and shifting interstimulus interval on eyeblink conditioning. *Behav Neurosci*. 2011;125(5):687.
191. Chess AC, Green JT. Abnormal topography and altered acquisition of conditioned eyeblink responses in a rodent model of attention-deficit/hyperactivity disorder. *Behav Neurosci*. 2008;122(1):63.
192. Zuchowski ML, Timmann D, Gerwig M. Acquisition of conditioned eyeblink responses is modulated by cerebellar tDCS. *Brain Stimul*. 2014;7(4):525-31.
193. Millenson JR, Kehoe EJ, Gormezano I. Classical conditioning of the rabbit's nictitating membrane response under fixed and mixed CS-US intervals. *Learn Motiv*. 1977;8(3):351-66.
194. Beyer L, Batsikadze G, Timmann D, Gerwig M. Cerebellar tDCS effects on conditioned eyeblinks using different electrode placements and stimulation protocols. *Front Hum Neurosci*. 2017;11.
195. Cheeran B, Talelli P, Mori F, Koch G, Suppa A, Edwards M, et al. A common polymorphism in the brain-derived neurotrophic factor gene (BDNF) modulates human cortical plasticity and the response to rTMS. *J Physiol*. 2008;586(23):5717-25.
196. Fritsch B, Reis J, Martinowich K, Schambra HM, Ji Y, Cohen LG, et al. Direct current stimulation promotes BDNF-dependent synaptic plasticity: potential implications for motor learning. *Neuron*. 2010;66(2):198-204.
197. Ichinohe N, Mori F, Shoumura K. A di-synaptic projection from the lateral cerebellar nucleus to the laterodorsal part of the striatum via the central lateral nucleus of the thalamus in the rat. *Brain Res*. 2000;880(1-2):191-7.
198. Bostan AC, Dum RP, Strick PL. The basal ganglia communicate with Cerebellum. *PNAS Proc Natl Acad Sci*. 2010;107(18):8452-6.
199. Hoshi E, Tremblay L, Feger J, Carras PL, Strick PL. The cerebellum communicates with the basal ganglia. *Nat Neurosci*. 2005;8(11):1491-3.

200. Pelzer EA, Hintzen A, Goldau M, Cramon DY, Timmermann L, Tittgemeyer M. Cerebellar networks with basal ganglia: feasibility for tracking cerebello-pallidal and subthalamo-cerebellar projections in the human brain. *Eur J Neurosci.* 2013;38(8):3106-14.
201. Milardi D, Arrigo A, Anastasi G, Cacciola A, Marino S, Mormina E, et al. Extensive direct subcortical cerebellum-basal ganglia connections in human brain as revealed by constrained spherical deconvolution tractography. *Front Neuroanat.* 2016;10.
202. Claassen DO, Jones CRG, Yu M, Dirnberger G, Malone T, Parkinson M, et al. Deciphering the impact of cerebellar and basal ganglia dysfunction in accuracy and variability of motor timing. *Neuropsychologia.* 2013;51(2):267-74.
203. Schwartze M, Keller PE, Kotz SA. Spontaneous, synchronized, and corrective timing behavior in cerebellar lesion patients. *Behav Brain Res.* 2016;312:285-93.
204. Avanzino L, Pelosin E, Vicario CM, Lagravinese G, Abbruzzese G, Martino D. Time processing and motor control in movement disorders. *Front Hum Neurosci.* 2016;10.
205. Prudente CN, Hess EJ, Jinnah HA. Dystonia as a network disorder: what is the role of the cerebellum? *Neuroscience.* 2014;260:23-35.
206. Avanzino L, Tinazzi M, Ionta S, Fiorio M. Sensory-motor integration in focal dystonia. *Neuropsychologia.* 2015;79:288-300.
207. Caligiore D, Helmich RC, Hallett M, Moustafa AA, Timmermann L, Toni I, et al. Parkinson's disease as a system-level disorder. *NPJ Parkinsons Dis.* 2016;2:16025.
208. Wu T, Hallett M. The cerebellum in Parkinson's disease. *Brain.* 2013;136(3):696-709.
209. Helmich RC, Janssen MJR, Oyen WJG, Bloem BR, Toni I. Pallidal dysfunction drives a cerebellothalamic circuit into Parkinson tremor. *Ann Neurol.* 2011;69(2):269-81.
210. Gao L, Zhang J, Hou Y, Hallett M, Chan P, Wu T. The cerebellum in dual-task performance in Parkinson's disease. *Sci Rep.* 2017;7.
211. Avanzino L, Abbruzzese G. How does the cerebellum contribute to the pathophysiology of dystonia? *Basal Ganglia.* 2012.
212. Bologna M, Berardelli A. Cerebellum: An explanation for dystonia? *Cerebellum Ataxias.* 2017;4(1):6.
213. McIntosh GC, Brown SH, Rice RR, Thaut MH. Rhythmic auditory-motor facilitation of gait patterns in patients with Parkinson's disease. *J Neurol Neurosurg Psychiatry.* 1997;62(1):22-6.
214. Heremans E, Nieuwboer A, Feys P, Vercruyssen S, Vandenberghe W, Sharma N, et al. External cueing improves motor imagery quality in patients with Parkinson disease. *Neurorehabil Neural Repair.* 2012;26(1):27-35.
215. Baltadjieva R, Giladi N, Gruendlinger L, Peretz C, Hausdorff JM. Marked alterations in the gait timing and rhythmicity of patients with de novo Parkinson's disease. *Eur J Neurosci.* 2006;24(6):1815-20.
216. Almeida QJ, Frank JS, Roy EA, Patla AE, Jog MS. Dopaminergic modulation of timing control and variability in the gait of Parkinson's disease. *Mov Disord.* 2007;22(12):1735-42.
217. Schwartze M, Kotz SA. regional interplay for temporal Processing in Parkinson's Disease: Possibilities and challenges. *Front Neurol.* 2016;6:270.

218. Avanzino L, Pelosin E, Martino D, Abbruzzese G. Motor timing deficits in sequential movements in Parkinson disease are related to action planning: a motor imagery study. *PLoS One*. 2013;8(9):e75454.
219. Pastor MA, Artieda J, Jahanshahi M, Obeso JA. Time estimation and reproduction is abnormal in Parkinson's disease. *Brain*. 1992;115(1):211-25.
220. O'Boyle DJ, Freeman JS, Cody FWJ. The accuracy and precision of timing of self-paced, repetitive movements in subjects with Parkinson's disease. *Brain*. 1996;119(1):51-70.
221. Harrington DL, Haaland KY, Hermanowitz N. Temporal processing in the basal ganglia. *Neuropsychology*. 1998;12(1):3.
222. Jones CRG, Claassen DO, Yu M, Spies JR, Malone T, Dirnberger G, et al. Modeling accuracy and variability of motor timing in treated and untreated Parkinson's disease and healthy controls. *Front Integr Neurosci*. 2011;5.
223. Elsinger CL, Rao SM, Zimelman JL, Reynolds NC, Blindauer KA, Hoffmann RG. Neural basis for impaired time reproduction in Parkinson's disease: an fMRI study. *J Int Neuropsychol Soc*. 2003;9(7):1088-98.
224. Cerasa A, Hagberg GE, Peppe A, Bianciardi M, Gioia MC, Costa A, et al. Functional changes in the activity of cerebellum and frontostriatal regions during externally and internally timed movement in Parkinson's disease. *Brain Res Bull*. 2006;71(1):259-69.
225. Jahanshahi M, Jones CRG, Zijlmans J, Katzenschlager R, Lee L, Quinn N, et al. Dopaminergic modulation of striato-frontal connectivity during motor timing in Parkinson's disease. *Brain*. 2010;133(3):727-45.
226. Assmus A, Marshall JC, Noth J, Zilles K, Fink GR. Difficulty of perceptual spatiotemporal integration modulates the neural activity of left inferior parietal cortex. *Neuroscience*. 2005;132(4):923-7.
227. Field DT, Wann JP. Perceiving time to collision activates the sensorimotor cortex. *Curr Biol*. 2005;15(5):453-8.
228. O'Reilly JX, Mesulam MM, Nobre AC. The cerebellum predicts the timing of perceptual events. *Journal of Neuroscience*. 2008;28(9):2252-60.
229. Filip P, Lošák J, Kašpárek T, Vaníček J, Bareš M. Neural network of predictive motor timing in the context of gender differences. *Neural Plast*. 2016;2016.
230. Husárová I, Lungu OV, Mareček R, Mikl M, Gescheidt T, Krupa P, et al. Functional Imaging of the Cerebellum and Basal Ganglia During Predictive Motor Timing in Early Parkinson's Disease. *J Neuroimaging*. 2014;24(1):45-53.
231. Husárová I, Mikl M, Lungu OV, Mareček R, Vaníček J, Bareš M. Similar circuits but different connectivity patterns between the cerebellum, basal ganglia, and supplementary motor area in early Parkinson's disease patients and controls during predictive motor timing. *J Neuroimaging*. 2013;23(4):452-62.
232. Furuya S, Altenmüller E. Finger-specific loss of independent control of movements in musicians with focal dystonia. *Neurosci*. 2013;247:152-63.
233. Jabusch HC, Schneider U, Altenmüller E. Δ 9-tetrahydrocannabinol improves motor control in a patient with musician's dystonia. *Mov Disord*. 2004;19(8):990-1.

234. Furuya S, Nitsche MA, Paulus W, Altenmüller E. Surmounting retraining limits in Musicians' dystonia by transcranial stimulation. *Ann Neurol*. 2014;75(5):700-7.
235. Van Der Steen MC, van Vugt FT, Keller PE, Altenmüller E. Basic timing abilities stay intact in patients with musician's dystonia. *PLoS One*. 2014;9(3):e92906.
236. Avanzino L, Bove M, Pelosin E, Ogliaastro C, Lagravinese G, Martino D. The cerebellum predicts the temporal consequences of observed motor acts. *PLoS One*. 2015;10(2):e0116607.
237. Avanzino L, Martino D, Martino I, Pelosin E, Vicario CM, Bove M, et al. Temporal expectation in focal hand dystonia. *Brain*. 2013;136(2):444-54.
238. Martino D, Lagravinese G, Pelosin E, Chaudhuri RK, Vicario CM, Abbruzzese G, et al. Temporal processing of perceived body movement in cervical dystonia. *Mov Disord*. 2015;30(7):1005-7.
239. Filip P, Gallea C, Lehéricy S, Bertasi E, Popa T, Mareček R, et al. Disruption in cerebellar and basal ganglia networks during a visuospatial task in cervical dystonia. *Mov Disord*. 2017;32(5):757-68.
240. Avanzino L, Ravaschio A, Lagravinese G, Bonassi G, Abbruzzese G, Pelosin E. Adaptation of feedforward movement control is abnormal in patients with cervical dystonia and tremor. *Clin Neurophysiol*. 2017.
241. Louis ED. The primary type of tremor in essential tremor is kinetic rather than postural: cross-sectional observation of tremor phenomenology in 369 cases. *Eur J Neurol*. 2013;20(4):725-7.
242. Critchley M. Observations on essential (heredofamilial) tremor. *Brain*. 1949;72(2):113-39.
243. Louis ED, Ottman R, Allen Hauser W. How common is the most common adult movement disorder? Estimates of the prevalence of essential tremor throughout the world. *Mov Disord*. 1998;13(1):5-10.
244. Louis ED. Non-motor symptoms in essential tremor: A review of the current data and state of the field. *Parkinsonism Relat Disord*. 2016;22:S115-S8.
245. Passamonti L, Cerasa A, Quattrone A. Neuroimaging of Essential Tremor: What is the Evidence for Cerebellar Involvement? *Tremor Other Hyperkinet Mov (N Y)*. 2012;2.
246. Wilms H, Sievers J, Deuschl G. Animal models of tremor. *Mov Disord*. 1999;14(4):557-71.
247. Benito-León J, Labiano-Fontcuberta A. Linking essential tremor to the cerebellum: clinical evidence. *Cerebellum*. 2016;15(3):253-62.
248. Filip P, Lungu OV, Manto M-U, Bareš M. Linking essential tremor to the cerebellum: physiological evidence. *Cerebellum*. 2016;15(6):774-80.
249. Louis ED. Linking essential tremor to the cerebellum: neuropathological evidence. *Cerebellum*. 2016;15(3):235-42.
250. Buhusi CV, Meck WH. What makes us tick? Functional and neural mechanisms of interval timing. *Nat Rev Neurosci*. 2005;6(10):755-65.
251. Buhusi CV, Meck WH. Relativity theory and time perception: single or multiple clocks. *PLoS One*. 2009;4(7):e6268.
252. Wiener M, Turkeltaub P, Coslett HB. The image of time: a voxel-wise meta-analysis. *Neuroimage*. 2010;49(2):1728-40.

253. Elble RJ, Higgins C, Hughes L. Essential tremor entrains rapid voluntary movements. *Exp Neurol.* 1994;126(1):138-43.
254. Montgomery EB, Baker KB, Lyons K, Koller WC. Motor initiation and execution in essential tremor and Parkinson's disease. *Mov Disord.* 2000;15(3):511-5.
255. Özekmekçi S, Kiziltan G, Vural M, Ertan S, Apaydin H, Erginöz E. Assessment of movement time in patients with essential tremor. *J Neurol.* 2005;252(8):964-7.
256. Jiménez-Jiménez FJ, Rubio L, Alonso-Navarro H, Calleja M, Pilo-de-la-Fuente B, Plaza-Nieto JF, et al. Impairment of rapid repetitive finger movements and visual reaction time in patients with essential tremor. *Eur J Neurol.* 2010;17(1):152-9.
257. Duval C, Sadikot AF, Panisset M. Bradykinesia in patients with essential tremor. *Brain Res.* 2006;1115(1):213-6.
258. Avanzino L, Bove M, Tacchino A, Ruggeri P, Giannini A, Trompetto C, et al. Cerebellar involvement in timing accuracy of rhythmic finger movements in essential tremor. *Eur J Neurosci.* 2009;30(10):1971-9.
259. Farkas Z, Szirmai I, Kamondi A. Impaired rhythm generation in essential tremor. *Mov Disord.* 2006;21(8):1196-9.
260. Helmchen C, Hagenow A, Miesner J, Sprenger A, Rambold H, Wenzelburger R, et al. Eye movement abnormalities in essential tremor may indicate cerebellar dysfunction. *Brain.* 2003;126(6):1319-32.
261. Trillenber P, Führer J, Sprenger A, Hagenow A, Kömpf D, Wenzelburger R, et al. Eye-hand coordination in essential tremor. *Mov Disord.* 2006;21(3):373-9.
262. Gitchele GT, Wetzel PA, Baron MS. Slowed Saccades and Increased Square Wave Jerks in Essential Tremor. *Tremor Other Hyperkinet Mov.* 2013;3:tre-03-178-4116-2.
263. Brown SH, Cooke JD. Movement-related phasic muscle activation. I. Relations with temporal profile of movement. *J Neurophysiol.* 1990;63(3):455-64.
264. Britton TC, Thompson PD, Day BL, Rothwell JC, Findley LJ, Marsden CD. Rapid wrist movements in patients with essential tremor The critical role of the second agonist burst. *Brain.* 1994;117(1):39-47.
265. Köster B, Deuschl G, Lauk M, Timmer J, Guschlbauer B, Lüicking CH. Essential tremor and cerebellar dysfunction: abnormal ballistic movements. *J Neurol Neurosurg Psychiatry.* 2002;73(4):400-5.
266. Anderson VC, Burchiel KJ, Hart MJ, Berk C, Lou J-S. A randomized comparison of thalamic stimulation and lesion on self-paced finger movement in essential tremor. *Neurosci Lett.* 2009;462(2):166-70.
267. Braitenberg V. Functional interpretation of cerebellar histology. *Nature.* 1961;190(4775):539.