San Jose State University

From the SelectedWorks of Scott A. Shaffer

2011

Contemporary and historical separation of transequatorial migration between genetically distinct seabird populations

M J Rayner, *University of Auckland*M E Hauber
T E Steeves, *university of canterbury*H A Lawrence
D R Thompson, et al.



- 1 Contemporary and historical separation of transequatorial migration between
- 2 two genetically-distinct seabird populations

3

- 4 Matt J. Rayner^{1, 2} *, Mark E. Hauber³, Tammy E. Steeves⁴, Hayley A. Lawrence⁵, David R.
- 5 Thompson⁶, Paul M. Sagar⁷, Sarah J. Bury⁶, Todd J. Landers^{8, 2}, Richard A. Phillips⁹, Louis
- 6 Ranjard², & Scott A. Shaffer¹⁰

7

- 8 ¹ National Institute of Water & Atmospheric Research Ltd, Private Bag 99940,
- 9 Auckland, New Zealand.
- 10 ² School of Biological Sciences, University of Auckland, Private Bag 92019, Auckland, New Zealand.
- 11 ³ Department of Psychology, Hunter College of the City University of New York, 695 Park Avenue, New
- 12 York NY 10065, USA.
- 13 ⁴ School of Biological Sciences, University of Canterbury, Canterbury, Private Bag 4800, Christchurch,
- 14 New Zealand.
- 15 Manaaki Whenua Landcare Research, Private Bag 92170, Auckland, New Zealand.
- ⁶ National Institute of Water & Atmospheric Research Ltd, Private Bag 14901, Wellington, New Zealand.
- ⁷ National Institute of Water & Atmospheric Research Ltd, P O Box 8602, Christchurch, New Zealand.
- 18 Auckland Museum, Private Bag 92018, Auckland, New Zealand.
- 19 ⁹ British Antarctic Survey, Natural Environment Research Council, High Cross, Madingley Road,
- 20 Cambridge CB3 0ET, UK.
- 21 Department of Biological Sciences, San Jose State University, San Jose, 95192-0100, USA.

2223

* m.rayner@niwa.co.nz

2425

26

27

28

29

Abstract

Pelagic seabirds are highly mobile, reducing the likelihood of allopatric speciation where disruption of gene flow between populations is caused by physically insurmountable, extrinsic barriers. Segregation during the non-breeding season appears to provide an intrinsic barrier to gene flow among seabird populations that otherwise occupy nearby or overlapping regions during breeding, but how this is achieved remains unclear. Here we show that the two genetically distinct populations of Cook's petrel (*Pterodroma cookie*) exhibit transequatorial separation of non-breeding ranges at contemporary (ca. 2-3 yrs) and historical (ca. 100 yrs) time scales. Segregation during the non-breeding season *per se* appears an unlikely barrier gene flow. Instead we provide evidence that habitat specialisation during the non-breeding season is associated with breeding asynchrony which, in conjunction with philopatry, restricts gene flow. Habitat specialisation during breeding and non-breeding likely promotes evolutionary divergence between these two populations via local adaptation.

Introduction

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

Divergent migratory behaviours to and from breeding sites have led to the disruption of gene flow among populations in many species ^{1,2}, contributing to reproductive isolation under the classic model of allopatric speciation³. In terrestrial environments, genetic differentiation among migratory populations is frequently paralleled by extrinsic (e.g. mountain ranges) and intrinsic (e.g. timing of dispersal, inherited migratory direction, host use by parasites) barriers that restrict gene flow and facilitate divergence via genetic drift and/or selection ⁴⁻⁶. However, the nature of the extrinsic and intrinsic barriers that disrupt gene flow among populations of marine animals have only recently begun to be investigated ^{7,8} and remain poorly known for many highly mobile taxa ^{8,9}. Seabirds undertake the longest known migrations on Earth, routinely crossing hemispheres between breeding and non-breeding habitat within ocean basins ^{10,11}. As a result of this extreme mobility, seabirds experience few apparent physical barriers to dispersal. Thus, seabirds can potentially visit and ultimately breed on islands thousands of kilometres from their natal colony, which may contribute to a lack of genetic structure observed in several species ¹²⁻¹⁵. Yet other seabird species show surprisingly high levels of population genetic structure 8,16-18 and intrinsic barriers to gene flow appear to play an important role in the evolution of seabird diversity 4,18-22. For example given the strong tendency for seabirds to return to their natal site to breed, natal philopatry has been proposed as such a barrier to gene flow ¹⁹. However given that not all seabirds exhibit strong genetic structure it is unlikely to be the sole driver.

A recent meta-analysis suggests that segregation of non-breeding distributions may be a key determinant of genetic structure among seabird populations ⁸, but how occupying disjunct non-breeding distributions can restrict gene flow remains unclear. Although advances in tracking technology have improved our ability to record the long-distance movements of seabirds at sea ²⁰, few studies have examined the non-breeding segregation amongst seabird populations that occupy nearby or overlapping regions during the breeding season ^{11,21}. Accordingly, the role of seasonal movements in shaping the genetic structure of seabird populations, and the persistence of these movement patterns at the timescale of generations, remain unknown. Such information is vital for providing insights into behavioural and ecological mechanisms underlying the diversification of highly mobile taxa, including those within oceanic environments.

Pterodroma cookii (Cook's petrel; Procellariiformes: Procellariidae Gray 1843), are pelagic seabirds that breed exclusively on two islands at the northern and southern extremes of their prehistoric range within the New Zealand archipelago ²²: Little Barrier Island (LBI) and Codfish Island (CDF), respectively ^{23,24}. When breeding, *P. cookii* range within the south-west Pacific and Tasman Sea, with overlapping foraging ranges centred around their respective breeding colonies ²⁵. Shipboard observations indicate that after breeding, *P. cookii* are trans-equatorial and trans-Pacific migrants, with sightings concentrated in the North Pacific (North-Pacific Convergence and centrally along Baja California) and in the South Pacific (Humboldt Current) Oceans ²⁶. On the basis of body size and plumage characteristics, in 1929 Murphy ²⁷ distinguished between smaller *P. c. cookii* that were collected in the North Pacific and assigned to the breeding population at LBI, and the larger *P. c. orientalis*, of unknown breeding origin, collected south of the Equator, off the coast of Peru. Subsequently, Falla ²⁸ observed

that the plumage and morphological characteristics of *P. cookii orientalis* were consistent with juveniles of *P. cookii cookii* from the breeding population at CDF that was discovered in 1934 ²⁴. In addition to differences in body size, body mass, and the potential for non-breeding birds to occupy disjunct habitats ^{5,26,29}, birds on LBI and CDF exhibit a one-month asynchrony in breeding phenology ⁵ and are genetically distinct ⁶. These combined observations strongly suggest that gene flow between the two extant *P. cookii* populations is highly restricted. Thus, *P. cookii* is an ideal study species to explicitly examine whether and how occupying disjunct non-breeding distributions can restrict gene flow among seabird populations.

Here we combined geolocator-based tracking with an isotopic (δ^{13} C and δ^{15} N) and genetic (mitochondrial cytochrome Oxidase subunit 1) comparison of modern breeding birds, and historical specimens (i.e. museum skins), to test the hypothesis of transequatorial separation of the two *P. cookii* breeding populations at varying temporal scales. Specifically, we tested the predictions that contemporary *P. cookii* populations, tracked over a complete annual cycle, would exhibit divergent non-breeding distributions, habitat use, and foraging patterns; this divergence would be consistent between contemporary cycles (ca. 2-3 years) as revealed by stable isotope analyses; the stable isotope signatures of historical specimens of unknown breeding provenance collected ca. 100 years ago from the two non-breeding oceanic regions used by contemporary populations, would match those of modern samples, and that genetic analysis would clearly assign origin of breeding population for these museum specimens and confirm divergence in transequatorial migration of the *P. cookii* populations over historical timescales. This combination of tracking, museum-based investigation, and elemental and molecular analyses presents a novel opportunity to

evaluate how differences in contemporary and historical migratory behaviour can contribute to the diversification of a highly mobile marine species.

132

133

134

130

131

Results

Population movements

135 Consistent with the prediction that contemporary tracked P. cookii would exhibit 136 divergent non-breeding distributions, our analysis reveals that birds tracked between 137 consecutive breeding seasons using light-based geolocation loggers (hereafter called 138 loggers) between 2007 and 2009 from LBI (n = 11: female (\mathcal{D}) = 5, male (\mathcal{E}) = 6) and 139 CDF (n = 11: Q = 6, Q = 5) exhibited transequatorial separation of their non-breeding 140 habitats within the Pacific Ocean (Fig. 1, Supplementary Fig. S1). This contrasts with 141 previous studies of other transequatorial migrants, including related Procellariiform 142 seabirds, which showed substantial overlaps in space use and mixing during the 143 nonbreeding period of individuals that originated from different breeding populations 10,11,29 . Post-breeding LBI *P. cookii* (tracked for 389 ± 49 SD days) completed an anti-144 clockwise migration of 48,037 ± 7953 SD km within the North and South Pacific 145 146 Ocean. Birds tracked from LBI moved east, then north across the equator to reach core 147 non-breeding distributions within the California and North-Pacific currents 148 (approximately 35°N) in 34 ± 8 SD days. Pre-breeding migration returning to New 149 Zealand waters was completed in 20 ± 5 SD days (Fig. 1) on a direct southwest route. 150 In contrast, post-breeding CDF P. cookii (tracked for 450 ± 3 SD days) migrated within 151 the South Pacific Ocean (37,813 \pm 6920 SD km) moving east then north within the 152 Humboldt Current to reach core non-breeding distributions off the Peruvian Coast 153 (approximately 15° S) in 19 ± 5 SD days. Pre-breeding migration returning to New 154 Zealand was again completed in 20 ± 5 SD days through a south-western corridor, north of the region traversed during post-breeding movements. Consistent with their breeding timetables, the migration schedule of both *P. cookii* populations was asynchronous; LBI birds commenced post- and pre-breeding migrations approximately 1 month before CDF birds (post-breeding 11 March \pm 12 SD days vs. 8 April \pm 15 SD days ($t_{(18)} = -4.67$, p < 0.0001); pre-breeding 5 Sept \pm 10 SD days vs. 20 Oct \pm 6 SD days ($t_{(15)} = -11.85$, p < 0.0001)).

During initial migration east towards South America, the routes taken by *P. cookii* from LBI and CDF overlapped, followed a similar direction to those of post-breeding sooty shearwaters (*Puffinus griseus*) ¹¹ and Westland petrels (*Procellaria westlandica*) ³⁰, but not Australasian gannets (*Morus serrator*), departing from New Zealand nesting colonies ³¹. Subsequently, LBI birds moved north-west across the equator and eventually returned on south-westerly trajectories to New Zealand along routes that were directionally similar to sooty shearwaters ¹¹, flesh-footed shearwaters (*Puffinus carneipes*) ³² and bar-tailed godwits (*Limosa lapponica baueri*) departing from on or off North America ³³. These observations highlight the existence of an important cross-taxa avian migration corridor within the Pacific Ocean between approximately 170° E and 160° W.

Isotopic and habitat segregation

P. cookii moult during the non-breeding period 26,34 , and hence incorporate local isotopic dietary signals in their new plumage 35 . Stable isotope ratios of C (δ^{13} C) and N (δ^{15} N) in particular provide an indication of carbon source (benthic vs. pelagic, inshore vs. offshore, and information on water mass) and trophic level of prey, respectively 35 , and comparisons thus can indicate geographic and/or dietary segregation 36,37 .

Consistent with the prediction that geographic divergence would be consistent between the *P. cookii* populations at a contemporary timescale (ca. 2-3 years), there was no significant difference in the isotope signatures of *P. cookii* body feathers collected in 2006 and 2008 from LBI (2006 untracked n = 20, 2008 tracked n = 11)(δ^{13} C: Z = 0.83, p = 0.41; δ^{15} N: Z = 1.70, p = 0.10) or CDF (2006 untracked n = 20, 2008 tracked n = 9)(δ^{13} C: Z = 1.34, p = 0.19; δ^{15} N: Z = 1.79, p = 0.07)(Fig. 2) suggesting that, currently, within each breeding population, birds forage within similar oceanic regions and on prey of a similar trophic level in successive years.

In support of the prediction of a match between the stable isotope signatures of historical and contemporary samples sharing the same non-breeding provenance, variation in the δ^{13} C signatures of feathers from modern (LBI: 2006 and 2008) combined; CDF: 2006 and 2008 combined) and historical specimens collected within the core non-breeding habitats of birds tracked from LBI and CDF, respectively (samples collected in 1905 at 22.42°N, 112.67°W, North Pacific Ocean, in the California Current region off Baja California, hereafter called BC, and in 1913 at ca. 11°S, 79°W, South Pacific Ocean, in the Humboldt Current, hereafter called HC; Supplementary Tables S1 and S2) suggest that birds have used population-specific oceanic regions, with similar carbon signatures, at historical time scales (Fig. 1) (F = 81.32. df = 2, p < 0.001). The mean δ^{13} C value of LBI P. cookii (-17.18 ± 0.70 SD %). was not significantly different from that of historical samples from BC (-16.72 \pm 0.51 SD %) (t = 2.085, p = 0.250), but was significantly lower than that of modern samples from CDF (-14.69 \pm 0.58 %) (t = 14.92, p < 0.01), and historical samples from HC (- 14.79 ± 0.73 %) (t = 9.292, p < 0.001) (Fig. 2). The mean δ^{13} C value of modern CDF samples was not significantly different from that of historical samples collected within the HC (t = 0.42 p = 1.00) but was significantly higher than historical samples collected from BC (t = 8.94 p < 0.001) (Fig. 2). These results suggest consistent differences in carbon isotopic signals between the North Pacific Convergence and South Pacific Humboldt Current systems, and support our initial prediction.

209

205

206

207

208

The mean δ^{15} N value of modern LBI *P. cookii* feathers (15.37 ± 0.95%) was not 210 211 significantly different from modern CDF feathers ($16.05 \pm 1.29\%$) (t = 3.69, p = 0.16) and the only difference in mean δ^{15} N value between modern and historical feathers was 212 213 observed in modern samples from LBI (15.37 \pm 0.95%) and historical samples from HC 214 $(16.76 \pm 1.42\%)$ (t = 3.53, p < 0.01) (Fig. 2). The most likely explanation for these 215 results is that modern P. cookii feed on prey at similar trophic levels, despite occupying 216 different oceanic regions (as suggested by tracking and carbon isotope data). 217 Accordingly, the environmental characteristics of core non-breeding habitats exploited 218 by modern LBI and CDF P. cookii were significantly different, with LBI P. cookii foraging over deeper (4930 \pm 581 SD m vs. 3330 \pm 314 SD m, Z = -3.97, p < 0.0001), 219 warmer (20.11 \pm 1.97 SD °C vs. 17.29 \pm 1.59 SD °C, Z = 3.12, p < 0.01) and less 220 productive waters $(0.16 \pm 0.10 \text{ SD mg Chl } a \text{ m}^{-3} \text{ vs. } 0.73 \pm 0.18 \text{ SD mg Chl } a \text{ m}^{-3}, Z = -$ 221 222 3.91, p < 0.0001) than their CDF conspecifics (Fig. 3). At the intrapopulation level, comparisons of environmental characteristics for presence and absence data indicated 223 that LBI P. cookii occupied less productive waters $(0.16 \pm 0.10 \text{ mg Chl } a \text{ m}^{-3} \text{ vs. } 0.28 \pm$ 224 225 0.17 mg Chl a m⁻³, Z = -2.20, p = 0.03) and CDF P. cookii more productive waters (0.73) \pm 0.18 mg Chl a m⁻³ vs. 0.47 \pm 0.12 mg Chl a m⁻³, Z = 2.79, p < 0.01) than those 226 227 available within their respective non-breeding core habitats (Fig. 3).

228

Genetic divergence

Consistent with prediction that genetic analysis would confirm divergence in transequatorial migration of the *P. cookii* populations over historical timescales, all nine historical BC *P. cookii* skins had mitochondrial *cytochrome c oxidase subunit 1* (COI) haplotypes that were identical to those of the modern LBI population ³⁸ (Fig. 1). Seven of ten historical HC *P. cookii* skins shared the same haplotype as that of modern CDF *P. cookii* ³⁸ and each of the three remaining skins had a novel haplotype (Fig. 1). We previously identified a single nucleotide polymorphism (SNP; at site 156) that differentiates modern LBI and CDF *P. cookii* ³⁸. All haplotypes sequenced from historical HC *P. cookii* skins in this study share the same diagnostic SNP as modern CDF birds, including the three novel haplotypes (Fig. 1, Supplementary Figure S2, Supplementary Tables S1, S2 and S3). These results support the previous predictions of contemporary and historical transequatorial separation of the extant *P. cookii* populations, revealed by our tracking and isotopic analyses, and suggests that these two populations have been genetically structured for a minimum of 100 years.

Discussion

Intrinsic factors argued to restrict gene flow between seabird populations include natal philopatry ¹⁹, divergent breeding timetables ^{39,40} and differences in breeding and non-breeding distributions ^{8,12,17}, though how these interact is poorly understood. Our combined results suggest that the use of population specific non-breeding habitats plays an interactive role in the restriction of gene flow, as both a pre-mating and post-mating barrier. In regards to the former, migration routes of differing lengths or directions, are widely recognised pre-mating barriers in migrant terrestrial bird species ^{5,41,42}. In *P. cookii* differences in the duration of migratory movement or non-breeding distribution

seem to underlie the pronounced breeding asynchrony between the populations. Moreover, regional differences in the timing of peak primary productivity, or the onset of gonad development based upon differences in day length and temperature shifts ⁴³, could amplify asynchronies in breeding timetables beyond those mediated by differences in route or non-breeding residency times alone. Thus, it is not segregation during the non-breeding season *per se*. that represents a pre-mating barrier to gene flow. Rather, it is the interaction between differing non-breeding distributions and divergent breeding timetables, combined with high natal philopatry, which restricts gene flow.

In regards to the latter, adult Cook's petrel desert their chicks up to two weeks prior to fledging and, as seen in many migratory species 5,41 , chicks presumably follow an inherited migratory direction and timing 5,42 . Studies of migrant terrestrial bird species indicate that the inheritance of suboptimal migration routes during secondary contact represents a post-mating barrier to gene flow $^{10, 40}$. Although secondary contact is presumably rare in $P.\ cookii$, the inheritance of an alternative or intermediate migratory strategy would place individuals at a selective disadvantage if it led to the occupancy of suboptimal non-breeding grounds or breeding timetables that were out of synchrony with the rest of the population $^{10, 40}$.

Habitat specialisation during both the breeding and non-breeding season is also likely to promote divergence among the two extant *P. cookii* populations via local adaptation. For example, when breeding, *P. cookii* exhibit divergent, although overlapping distributions and diets in response to regional differences in oceanography ²⁵ (current study). Moreover, the populations differ in body mass ²² with smaller LBI birds foraging in warmer low-latitude waters compared to larger CDF conspecifics ²⁵. That individuals

from these two populations also occupy oceanic waters with similarly divergent characteristics during the non-breeding period (i.e., LBI birds occupy warmer less productive waters than CDF birds; Fig. 1) suggests population-specific adaptation to differing marine habitats, possibly linked to body size. Indeed, differences in seabird habitat use related to body size have been observed in other studies, particularly in species with pronounced sexual size dimorphism ⁴⁴⁻⁴⁶. It remains uncertain whether body size differences between populations of *P. cookii* are a cause or consequence of divergent habitat use. Regardless, our data support and extend local adaptation hypotheses by suggesting that conspecific seabird populations can become tied to particular marine habitats during the breeding ^{18,39,47} and non-breeding periods ^(current study), presenting opportunities for population divergence.

Overall, this study demonstrates the use of integrative techniques to track space use, behaviour, foraging, and breeding to demonstrate clear spatial segregation of non-breeding distribution between two populations which, mediated by its influence on breeding phenology, appears to represent an intrinsic barrier to gene flow, and has lead to local adaptation in a highly mobile seabird.

Methods

Tracking and environmental data

P. cookii breeding on LBI and CDF were equipped with light-based geolocation loggers (British Antarctic Survey, Mk14) in November-December 2007 (LBI, 36° 11'S, 175° 04'E, n = 13; CDF, 46° 11'S, 167° 38'E, n = 14) during early incubation. Loggers were deployed on birds of known breeding history using published ²⁵ with the total package weighing < 1% of body mass. The sex of all birds were previously determined using

molecular methods 25,48 . The following breeding season, 24 (89%) birds (LBI n = 12 and CDF n = 12) returned to breed and were recaptured in January - March 2009 at their same breeding burrows, and loggers were removed. Two loggers (LBI n = 1 and CDF n = 1) failed to download resulting in 11 datasets from each population for processing and analyses.

Light data from the loggers were processed using Multitrace software (Jensen Software Systems) and locations (2 d⁻¹) were estimated with an expected mean accuracy \pm SD of 186 \pm 114 km ⁴⁹. Sunrise and sunset times were identified based on light curve thresholds, with latitude calculated from day/night length and longitude calculated from the time of local midday/midnight relative to Greenwich Mean Time. As a result of day length uniformity around the equinoxes, clearly erroneous locations occurring 3-4 weeks either side of the equinox were excluded. Furthermore, points that involved movements of > 1600 km in one day ⁵⁰, those with interruptions to light curves around sunset and sunrise, or that were clearly outside of the known or possible range for *P. cookii*, were removed from the data set.

Totals of 7,886 and 9,082 locations were obtained for LBI and CDF *P. cookii* respectively, of which 9.7% and 10.6% were excluded after filtering. Filtered locations were then used to estimate year-round utilisation distribution (UD) kernels for each population following methods detailed in ref ⁵¹. In brief, 2-D Gaussian kernel densities were estimated using custom routines created in MatLab ⁵¹ (The Mathworks, Natick, MA, USA). Kernels were calculated using a Lambert Cylindrical Equal Area projection on an 80 km grid with grid cells normalized for bird effort by dividing each cell by the number of birds contributing the locations within a cell. The kernel smoothing

parameter (h) was estimated using an adaptive method ⁵² to estimate an optimal local value, following ref. ⁵³. Based published methods ²⁵, a 1,000 km buffer was used around each colony to define breeding habitat, and the 80% contour of UD kernels, calculated individually, were used to define the core non-breeding distributions for each bird ⁵⁰. Dates of the first and last locations to enter and exit the core non-breeding habitats were used to define migration timing for each individual. Individuals'migration distances were calculated by summing the point to point distances travelled during post-breeding and pre-breeding movements between these core areas.

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

330

331

332

333

334

335

336

337

Satellite-derived remotely-sensed environmental data were used to contrast the core habitats of P. cookii from LBI and CDF during the non-breeding period (presence dataset) as well as those available, but not used, within each population-specific region (absence dataset). For the presence dataset, environmental data were extracted for logger locations falling within the 20% UD for each tracked bird using a 1° longitude by 2° latitude grid centred (the approximate error of geolocation estimates) on the date and coordinates of the location ⁵¹. For the absence dataset, environmental data were extracted from ten randomly selected locations, derived within a 1,000 km radius buffer of each logger location and centred on the corresponding logger location date. Environmental data extracted for the random locations were averaged to give a mean absence estimate to contrast presence values. Remotely-sensed environmental data were obtained from NOAA's Environmental Research Division (http://coastwatch.pfel.noaa.gov/thredds/catalog.html) including 5-day composites of chlorophyll a (Chl a) concentration (mg Chl m⁻³) at a spatial resolution of 0.1° resolution ⁵⁴ and 8-day composites sea surface temperature (SST) in °C with a spatial resolution of 0.1° 55. Bathymetric data were obtained from the ETOPO2 dataset 56.

Stable isotopes

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

To contrast population-specific dietary signals during the non-breeding period, we conducted stable isotope analyses of C (δ^{13} C) and N (δ^{15} N) ratios in body feathers collected from modern LBI and CDF P. cookii and museum specimens held at the Californian Academy of Sciences (n = 9) and American Museum of Natural History (n = 10). Single body feathers were collected from breeding P. cookii in 2006 and from tracked birds upon their recapture in 2009, with no resampling of birds between years. Modern and historical feathers were cleaned with 70% ethanol, then washed in distilled water to remove contaminants, dried in at 50°C and cut into fine fragments using scissors. Stable isotope analyses of $\delta^{15}N$ and $\delta^{13}C$, using a subsample (approximately 0.7 mg) of each homogenised feather, were carried out on a DeltaPlus (Thermo-Finnigan) continuous flow isotope ratio mass spectrometer using protocols outlined by ref 25 . Measurement precision of the isotope analysis was 0.29% for δ^{15} N and 0.24% for δ^{13} C. To account for the Seuss effect, the temporal biasing of the atmospheric CO₂ pool to more negative δ^{13} C values as a result of the burning of fossil CO₂, δ^{13} C data from historical feathers where normalised by subtracting a year-specific factor $\delta^{13}C = -1$ + 1 1 (2009-year (1905 or 1913))*0.027 following ref ⁵⁷.

373

374

375

376

377

378

379

Genetic analyses

Toe pad skin samples were collected from the same historical Cook's petrel specimens from which feathers were sampled for stable isotope analyses (Supplementary Table S1 and S2). DNA extraction and polymerase chain reaction (PCR) set up was performed in a physically isolated dedicated ancient DNA laboratory. Contamination was monitored using extraction and PCR negative controls and genomic DNA was isolated as per ref

sene (COI) was amplified using two sets of primers that amplified two overlapping fragments: AWCF1 and AWCintR2 ⁵⁸; LCRintF2 5'-TCATAATTGGGGGATTTGGA-3' (designed using Primer3 software ⁵⁹) and AWCintR3 ⁵⁸. This region corresponds to the first 375 bp of the 677 bp fragment sequenced by ref ³⁸. PCR products were purified using a QIAquick PCR Purification kit (Qiagen) then sequenced bidirectionally from independent PCRs ⁵⁸. Sequences were concatenated and aligned using Sequencher version 4.8 (Gene Codes), deposited in NCBI GenBank (accession numbers HQ263645 to HQ263663) and compared with those in ref ³⁸.

Statistical analyses

Habitat data extractions, processing and analyses, were conducted using Matlab. Non-parametric Mann-Whitney U tests were used to test for differences between presence and absence habitat data for the extracted environmental parameters at an intrapopulation level, and for the presence data at an inter-population level. Mean environmental parameter values for individual birds were used as the sampling unit. Mann-Whitney U tests were initially used to compare stable isotope values between years (2006 and 2009) for LBI and CDF. Isotope data (δ^{15} N and δ^{13} C) were subsequently pooled for each population and permutation-based non-parametric ANOVA, with pair-wise comparisons using the Bonferroni correction method, conducted to compare isotope values between individuals from the breeding populations (LBI and CDF) and museum skin specimens (CAS 1905 and AMNH 1913) using the programme PERMANOVA. Sex was initially included as a factor in all analyses, but excluded as a result of non-significant differences.

405 Acknowledgements 406 We thank B. Dunphy and New Zealand Department of Conservation (S. McInnes, L. 407 Witwell, R. Renwick, P. McClelland) for field assistance, M. Flannery (California 408 Academy of Sciences) and P. Sweet and T. Trombone (The American Museum of 409 Natural History) for access to museum specimens, C. Millar and S. Patel of assistance 410 with genetic analyses, J. Brown for assistance with isotopic laboratory support, and W. 411 Rayner for support during this research. Artwork provided by Chris Gaskin. This work 412 was supported by the Hauturu Little Barrier Island Supporters Trust (ASB Bank 413 Community Trust grant), the New Zealand Tertiary Education Commission and 414 Foundation for Research Science and Technology. This research represents a 415 contribution to the BAS Ecosystems Programme. 416 417 **Author Contributions** 418 M.J.R, D.R.T, P.M.S, M.E.H, S.A.S, and T.E.S designed the research. M.J.R, T.J.L, 419 R.A.P., L.R and S.A.S processed and analysed spatial data. M.J.R., H.A.L, and T. E. S. 420 conducted genetic analyses, and S.J.B, M.E.H and M.J.R processed and analysed 421 isotope data. M.J.R wrote paper and all authors discussed the results and contributed to the manuscript. 422 423 424 **Competing financial interests:** The authors declare no competing financial interests. 425 426 427 428 429

430 431 432 433 434 435 436 Figure legends 437 Fig. 1. Pacific migrations of tracked P. cookii 438 Movements of P. cookii breeding on Little Barrier Island (red circle, North Island) and 439 Codfish Island (purple circle, South Island), New Zealand. Approximate post-breeding 440 and pre-breeding migration routes of tracked LBI (red dashed lines) and CDF (purple 441 dashed lines) birds begin and end with mean migration departure and arrival dates 442 (white text). Red and purple tones, and associated contour lines, represent the 95%, 443 75%, 50%, and 25% kernel estimates for LBI and CDF birds respectively. Pie charts 444 show the geographic distribution of cytochrome c oxidase subunit 1 mtDNA haplotypes 445 sequenced from modern P. cookii populations (charts attached to LBI and CDF colony 446 locations) and historical P. cookii skins collected in the North (BC) and South Pacific 447 (HC) Ocean in 1905 and 1913 respectively (charts attached to white circles showing 448 approximate collection location). Pie chart size reflects genetic analysis sample size: 449 modern P. cookii LBI n = 26 and CDF n = 19; historical P. cookii BC n = 9 and HC n = 450 10. 451 452 Fig. 2 Isotope ratios of contemporary and historic *P. cookii* feathers 453 Feather isotope signatures of P. cookii (±SD) from LBI (red circle, 2006 (untracked)

and 2008 (tracked) combined, n = 31) and CDF (purple circle, 2006 (untracked) and

2008 (tracked) combined, n = 29) and from historical skins collected in the North 455 Pacific (BC) (black circle, 1905, n = 9) and South Pacific Ocean (HC) (clear circle, 456 457 1913, n = 10). 458 459 Fig. 3 Characteristics of selected oceanic habitats 460 Box plots of median values for remotely sensed environmental data (a. water depth, b. 461 sea surface temperature, c. chlorophyll a concentration) from logger locations of P. 462 cookii tracked from Little Barrier Island (LBI, n = 11) and Codfish Island (CDF, n = 11) 463 during occupancy of core non-breeding habitats (LBI Pres. and CDF Pres., 20% UD) 464 and in randomly selected proximate locations (LBI Abs. and CDF Abs.) see Methods. Asterisks represent significance of difference for group comparisons: * p = 0.05, ** p =465 0.01, *** p = 0.001, - = not significant. Box plots illustrate 25th, 50th (median), and 466 467 75th percentiles, error bars represent minimum and maximum values falling within 1.5 468 Inter Quartile Range, and crosses plot the outliers. 469 470

472 References

- Dor, R., Safran, R. J., Sheldon, F. H., Winkler, D. W. & Lovette, I. J. Phylogeny of the genus Hirundo and the barn swallow subspecies complex. *Molecular Phylogenetics and Evolution* **56**, 409-418, doi:DOI: 10.1016/j.ympev.2010.02.008 (2010).
- 477 2 Irwin, D. E. & Irwin, J. H. in *Birds of Two Worlds: The Ecology and Evolution*478 of Migration (eds R Greenberg & P P Marra) 27-40 (Johns Hopkins University Press, Baltimore, 2005).
- 480 3 Mayr, E. *Systematics and the Origin of Species*. (Columbia University Press, 481 1942).
- 482 4 Irwin, D. E., Bensch, S., Irwin, J. H. & Price, T. D. Speciation by distance in a ring species. *Science* **307**, 414-416, doi:10.1126/science.1105201 (2005).
- Bearhop, S. *et al.* Assortative mating as a mechanism for rapid evolution of a migratory divide. *Science* **310**, 502-504 (2005).
- Sorenson, M. D., Sefc, K. M. & Payne, R. B. Speciation by host switch in brood parasitic indigobirds. *Nature* **424**, 928-931, doi:http://www.nature.com/nature/journal/v424/n6951/suppinfo/nature01863_S1 .html (2003).
- Galarza, J. A. *et al.* The influence of oceanographic fronts and early-life-history traits on connectivity among littoral fish species. *Proceedings of the National Academy of Sciences* **106**, 1473-1478, doi:10.1073/pnas.0806804106 (2009).
- 493 8 Friesen, V. L., Burg, T. M. & McCoy, K. D. Mechanisms of population differentiation in seabirds. *Molecular Ecology* **16**, 1765-1785, doi:10.1111/j.1365-294X.2006.03197.x (2007).
- Natoli, A., Birkun, A., Aguilar, A., Lopez, A. & Hoelzel, A. R. Habitat structure
 and the dispersal of male and female bottlenose dolphins (*Tursiops truncatus*).
 Proceedings of the Royal Society of London B: Biological Sciences 272, 1217-1226, doi:10.1098/rspb.2005.3076 (2005).
- 500 10 Egevang, C. et al. Tracking of Arctic terns Sterna paradisaea reveals longest 501 animal migration. Proceedings of the National Academy of Sciences of the 502 United States of America 107, 2078-2081, doi:10.1073/pnas.0909493107 503 (2010).
- 504 11 Shaffer, S. A. et al. Migratory shearwaters integrate oceanic resources across the 505 Pacific Ocean in an endless summer. Proceedings of the National Academy of 506 Sciences of the United States of America 103, 12799-12802, 507 doi:10.1073/pnas.0603715103 (2006).
- Burg, T. M. & Croxall, J. P. Global relationships amoungst black-browed and grey-headed albatrosses: analysis of population structure using mitochondrial DNA and microsatellites. *Molecular Ecology* **10**, 2647-2660 (2001).
- 511 13 Austin, J., White, R. & Ovenden, J. Population-genetic structure of a philopatric, colonial nesting seabird, the short-tailed shearwater (*Puffinus tenuirostris*) Auk 111, 70-79 (1994).
- Avise, J. C., Nelson, W. S., Bowen, B. W. & Walker, D. Phylogeography of colonially nesting seabirds, with special reference to global matrilineal patterns in the sooty tern (*Sterna fuscata*) *Molecular Ecology* **9** (2000).
- Techow, N. M. S. M. *et al.* Speciation and phylogeography of giant petrels Macronectes. *Molecular Phylogenetics and Evolution* **54**, 472-487, doi:DOI: 10.1016/j.ympev.2009.09.005 (2010).

- 520 16 Morris-Pocock, J. A., Steeves, T. E., Estela, F. A., Anderson, D. J. & Friesen, V. L. Comparative phylogeography of brown (*Sula leucogaster*) and red-footed boobies (*S. sula*): The influence of physical barriers and habitat preference on gene flow in pelagic seabirds. *Molecular Phylogenetics and Evolution* **54**, 883-896, doi:DOI: 10.1016/j.ympev.2009.11.013 (2010).
- 525 17 Gómez-Díaz, E., González-Solís, J. & Peinado, M. A. Population structure in a 526 highly pelagic seabird, the Cory's shearwater *Calonectris diomedea*: an 527 examination of genetics, morphology and ecology *Marine Ecology Progress* 528 *Series* **382**, 197-209 (2009).
- 529 18 Dearborn, D. C., Anders, A. D., Schreiber, E. A., Adams, R. M. M. & Mueller, U. G. Inter-island movements and population differentiation in a pelagic seabird.

 531 *Molecular Ecology* 12, 2835-2843 (2003).
- 532 19 Steeves, T. E., Anderson, D. J. & Friesen, V. L. A role for nonphysical barriers 533 to gene flow in the diversification of a highly vagile seabird, the masked booby 534 (*Sula dactylatra*). *Molecular Ecology* **14**, 3877-3887 (2005).
- Burger, E. A. & Shaffer, S. A. Applications of tracking and data-logging technology in research and conservation of seabirds. *The Auk* **125**, 253-264 (2008).
- Phillips, R. A., Croxall, J. P., Silk, J. R. D. & Briggs, D. R. Foraging ecology of albatrosses and petrels from South Georgia: two decades of insights from tracking technologies. *Aquatic Conservation: Marine and Freshwater Ecosystems* 17, 6-21 (2008).
- 542 22 Imber, M. J., West, J., A. & Cooper, W. J. Cook's petrel (*Pterodroma cookii*): historic distribution, breeding biology, and effects of predators. *Notornis* **50**, 221-230 (2003).
- Rayner, M. J. *et al.* Predictive habitat modelling improves the population census accuracy of a burrowing seabird: a study of the endangered Cook's petrel. *Biological Conservation* **138**, 235-247 (2007).
- Rayner, M. J., Parker, K. A. & Imber, M. J. Population census of Cook's petrel *Pterodroma cookii* breeding on Codfish Island (New Zealand) and the global conservation status of the species. *Bird Conservation International* **18**, 211-218 (2008).
- Rayner, M. J. *et al.* Foraging ecology of the Cook's petrel *Pterodroma cookii* during the austral breeding season: a comparison of its two populations. *Marine Ecology Progress Series* **370**, 271-284 (2008).
- Marchant, S. & Higgins, P. J. *Handbook of Australasian, Antarctic and New Zealand Birds. Volume 1 Ratites to Ducks.* (Oxford University Press, 1990).
- 557 27 Murphy, R. C. On *Pterodroma cookii* and its allies. *American Museum Novitates* 370 (1929).
- Falla, R. A. Review of the smaller Pacific forms of *Pterodroma* and *Cookilaria*. *Emu* **42**, 111-118 (1942).
- Gonzalez-Solis, J., Croxall, J. P., Oro, D. & Ruiz, X. Trans-equatorial migration and mixing in the wintering areas of a pelagic seabird. *Frontiers in Ecology and the Environment* 5, 297-301, doi:doi:10.1890/1540-9295(2007)5[297:TMAMIT]2.0.CO;2 (2007).
- Landers, T. J., Rayner, M. J., Phillips, R. A. & Hauber, M. E. Dynamics of seasonal movements by a trans-pacific migrant, the westland petrel. *Condor* **113**, 71-79 (2011).
- Ismar, S. M. H., Phillips, R. A., Rayner, M. J. & Hauber, M. E. Geolocation tracking of the annual migration of adult australasian gannets (*Morus serrator*)

- 570 breeding in New Zealand. *The Wilson Journal of Ornithology* **123**, 121-125 (2011).
- 572 32 Rayner, M. J., Taylor, G. A., Thompson, D. R., Torres, L. G., Sagar, P. M., Shaffer, S. A. Migration and diving activity in three post-breeding flesh-footed shearwaters (*Puffinus carneipes*). *Journal of Avian Biology* (in press).
- Gill, R. E. et al. Extreme endurance flights by landbirds crossing the Pacific
 Ocean: ecological corridor rather than barrier? Proceedings of the Royal Society
 Biological Sciences 276, 447-457, doi:10.1098/rspb.2008.1142 (2009).
- 578 34 Spear, L. B., Howell, S. N. G. & Ainley, D. G. Notes on the at-sea identification of some Pacific gadfly petrels (Genus: *Pterodroma*). *Colonial Waterbirds* **15**, 202-218 (1992).
- Hobson, K. A. Tracing origins and migration of wildlife using stable isotopes: a review. *Oecologia* **120**, 314-326 (1999).
- Phillips, R., Bearhop, S., McGill, R. & Dawson, D. Stable isotopes reveal individual variation in migration strategies and habitat preferences in a suite of seabirds during the nonbreeding period. *Oecologia* **160**, 795-806, doi:10.1007/s00442-009-1342-9 (2009).
- 587 37 Cherel, Y., Phillips, R. A., Hobson, K. A. & McGill, R. Stable isotope evidence 588 of diverse species-specific and individual wintering strategies in seabirds. 589 *Biology Letters* **2**, 301-303, doi:10.1098/rsbl.2006.0445 (2006).
- Rayner, M. J., Carragher, C. J. F. & Hauber, M. E. Mitochondrial DNA analysis reveals genetic structure in two New Zealand Cook's petrel (*Pterodroma cookii*) populations. *Conservation genetics* **11**, 2073-2077 (2010).
- Monteiro, L. R. & Furness, R. W. Speciation through temporal segregation of Madeiran storm petrel (*Oceanodroma castro*) populations in the Azores? *Proceedings of the Royal Society of London B: Biological Sciences* **353**, 945-953, doi:10.1098/rstb.1998.0259 (1998).
- 597 40 Friesen, V. L. et al. Sympatric speciation by allochrony in a seabird.
 598 Proceedings of the National Academy of Sciences of the United States of
 599 America 104, 18589-18594, doi:10.1073/pnas.0700446104 (2007).
- Marra, P. P., Hobson, K. A. & Holmes, R. T. Linking winter and summer events in a migratory bird by using stable-carbon isotopes. *Science* **282**, 1884-1886, doi:10.1126/science.282.5395.1884 (1998).
- 603 42 Irwin, D. E. Speciation: new migratory direction provides route toward divergence. *Current Biology* 19, 1111-1113, doi:DOI: 10.1016/j.cub.2009.11.011 (2009).
- Berthold, P., Helbig, A. J., Mohr, G. & Querner, U. Rapid microevolution of migratory behaviour in a wild bird species. *Nature* **360**, 668-670 (1992).
- 608 44 Phillips, R. A., Silk, J. R. D., Phalan, B., Catry, P. & Croxall, J. P. Seasonal sexual segregation in two *Thalassarche* albatrosses: competitive exclusion, reproductive role specialization or foraging niche divergence? *Proceedings of the Royal Society B: Biological Sciences* 271, 1283-1292 (2004).
- Shaffer, S. A., Weimerskirch, H. & Costa, D. P. Functional significance of sexual dimorphism in wandering albatrosses, *Diomedea exulans. Functional Ecology* **15**, 203-210 (2001).
- Weimerskirch, H., Cherel, Y., Cuenot-Chaillet, F. & Ridoux, V. Alternative foraging strategies and resource allocation by male and female wandering albatrosses. *Ecology* **78**, 2051-2063 (1997).

- 618 47 Gómez-Díaz, E., González-Solís, J., Peinado, M. A. & Page, R. D. M. Phylogeography of the Calonectris shearwaters using molecular and morphometric data. *Molecular Phylogenetics and Evolution* 41, 322-332 (2006).
- Fridolfsson, A. K. & Ellegren, H. A simple and universal method for molecular sexing of non-ratite birds. *Journal of Avian Biology* **30**, 116-221 (1999).
- 49 Phillips, R. A., Silk, J. R. D., Croxall, J. P., Afanasyev, V. & Briggs, D. R.
 624 Accuracy of geolocation estimates for flying seabirds. *Marine Ecology Progress* 625 Series 266, 265-272 (2004).
- 626 50 Guilford, T. *et al.* Migration and stopover in a small pelagic seabird, the Manx shearwater *Puffinus puffinus*: insights from machine learning. *Proceedings of the Royal Society B: Biological Sciences* **276**, 1215-1223, doi:10.1098/rspb.2008.1577 (2009).
- Shaffer, S. A. *et al.* Spatio-temporal habitat use by breeding sooty shearwaters *Puffinus griseus. Marine Ecology Progress Series* **391**, 209-220 (2009).
- Worton, B. J. Kernel methods for estimating the utilization distribution in homerange studies. *Ecology* **70**, 164-168 (1989).
- Sheather, S. J. & Jones, M. C. A reliable data-based bandwidth selection method for kernel density estimation. *Journal of the Royal Statistical Society B* **53**, 683-690 (1991).
- 637 54 O'Reilly, J. E. *et al.* Ocean color chlorophyll algorithms for SeaWiFS. *Journal of Geophysical Research* **103**, 24937-24953, doi:10.1029/98jc02160 (1998).
- 639 55 Powell, B. S. *et al.* 4DVAR data assimilation in the intra-americas sea with the Regional Ocean Modeling System (ROMS). *Ocean Modelling* **23**, 130-145 (2008).
- 642 56 Smith, W. H. F. & Sandwell, D. T. Global sea floor topography from satellite altimetry and ship depth soundings. *Science* 277, 1956-1962, doi:10.1126/science.277.5334.1956 (1997).
- 645 57 Quillfeldt, P., Masello, J., McGill, R., Adams, M. & Furness, R. Moving 646 polewards in winter: a recent change in the migratory strategy of a pelagic 647 seabird? *Frontiers in Zoology* 7, 15 (2010).
- Patel, S., Waugh, J., Millar, C. D. & Lambert, D. M. Conserved primers for DNA barcoding historical and modern avian samples from New Zealand and Antarctic birds. *Molecular Ecology Resources* **10**, 431-438 (2009).
- Rozen, S. & Skaletsky, H. Primer3 on the WWW for general users and for biologist programmers. *Methods in Molecular Biology* **132**, 365-386 (2000).

653654

655

656

657

658

659





