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Soft hyaluronic gel promotes cell spreading, stress fibers, focal adhesion, membrane tension by phosphoinositide signaling, not traction force

Kalpana Mandal^{1,*}, Dikla Raz-Ben Aroush^{1,2}, Zachary Graber³, Bin Wu⁴, Chan Young Park⁵, Jeffery J. Fredberg⁵, Wei Guo⁴, Tobias Baumgart³, and Paul Janmey^{1,*}

¹Institute for Medicine and Engineering, University of Pennsylvania, Philadelphia, PA 19104

²Kennedy Institute of Rheumatology, University of Oxford, Oxford OX3 7FY, UK

³Department of Chemistry, University of Pennsylvania, Philadelphia, PA 19104

⁴Department of Biology, University of Pennsylvania, Philadelphia, PA 19104

⁵School of Public Health, Harvard University, Boston, MA

Abstract

Cells respond to both their chemical environment and to microenvironmental stiffness by the process of mechanotransduction. The mechanisms by which cells monitor and respond to the mechanical properties of their environment are largely unknown. Cellular response to the stiffness of the substrate is highly cell type specific and depends on the chemical composition of the substrate and therefore the type of adhesion receptors that engage it. Nearly all studies of mechanobiology in vitro employ substrates coated with protein or peptide ligands for integrins, but the native extracellular matrix (ECM) is highly enriched with glycosaminoglycans and proteoglycans that can alter cell adhesion and signaling through integrins. Hyaluronic acid (HA) is one of the major ECM components that helps maintain the viscoelasticity of connective tissues, controls tissue hydration, and organizes the supramolecular assembly of proteoglycans. In this study we investigate the role of HA together with integrin ligands in promoting hepatocellular carcinoma cell (Huh7) spreading on very soft substrates (300 Pa), resulting in morphology and motility similar to that which these cells develop only on stiff substrates (30 kPa/glass) in the absence of HA. In particular, we test the hypothesis that cell interaction with HA leads to activation of the PI3K/Akt signaling pathway, which in turn promotes actin remodeling to facilitate cell spreading without requiring high contractile forces that are generated on stiff substrates. Inhibition of polyphosphoinositide turnover whether by two different PI3kinase inhibitors or by a cell-permeant polyphosphoinositide-binding peptide causes both Huh7 cells and murine fibroblasts to decrease spreading and detach whereas cells on stiffer substrates show almost no response. Traction force microscopy (TFM) shows that the cell maintains a very low total strain energy and net contractile moment on HA substrates as compared to stiff 30 kPa substrate even though cells on both substrates have large spread areas, extensive focal adhesions, and actin bundles (generally called stress fibers). Measurements of cell membrane tension by lipid tether pulling show a similar level of membrane tension on HA substrate as on stiff substrates.

*Corresponding author.

These results suggest that simultaneous signaling stimulated by HA and an integrin ligand can generate PI3K-dependent signals to the cytoskeleton that mimic those generated by high cellular tension, to produce increased actin and focal adhesion assembly and large spread areas.

Introduction:

Many cell types alter their structure and function depending on the mechanical properties of the materials to which they adhere and on the type of adhesion receptor by which they bind [1]–[4]. In vivo, cells engage their extracellular matrix (ECM) both by mechanosensitive adhesion complexes and by other surface receptors for ECM components that cannot act as adhesive anchors, but that potentially modify the mechanical signals transduced at the cell/ECM interface. Cellular reaction to extracellular matrix (ECM) depends upon the specificity of the ligand binding [2], [5]. Previous studies found that cardiomyocytes grown on soft hyaluronic acid gels coated with fibronectin (HA-Fn) developed well-structured sarcomeres despite the very low elastic modulus of this material, suggesting that the role of the cardiac jelly in early stages of cardiogenesis is more than that of a passive coupling matrix between the myocardial-cardiomyocytes and endocardial-endothelial cell layers enveloping it [6],[7].

Hyaluronic acid (or hyaluronan (HA)), a non-sulfated glycosaminoglycan polysaccharide, is a major ECM component that plays an important role in development, wound healing, and cancer progression[8]–[10]. In normal tissue, HA amount is determined by the balance between HA synthesis and degradation. An over production of HA or upregulation of HA receptors facilitates cell migration, invasion of tumor cells or rapid tumor growth[11]. Breast cancer cells synthesize more HA than normal cells, and high HA production is correlated with poor patient survival in various types of cancer including prostate, breast and ovarian cancer [12], [13]. HA localizes at the leading edge of the tumor [14].

Cells on HA substrate of stiffness 300 Pa can spread as much as on very stiff substrate (10kPa) when both are coated with a ligand for integrins, usually either fibronectin or collagen I [15], [16]. Cell interaction with ECM-bound HA is mediated by CD44, CD168 (RHAMM), and other cell surface receptors. CD44 is overexpressed in many cancers and has been shown to promote angiogenesis from tumors[17], [18]. Cell interaction with ECM-HA through these receptors regulates many cell signaling pathways including Rho-GTPase, transforming growth factor beta (TGF- β) and focal adhesion kinase mediated (FAK) pathways [19].

Prior studies show a large effect of HA on cell morphology, stiffness, and contractility on soft gels containing Fn, but the molecular mechanism by which HA in the matrix exerts this effect is unexplored. This study is designed to determine if signals downstream of phosphatidylinositol-4,5-bisphosphate (PIP₂) or phosphatidylinositol-3,4,5-trisphosphate (PIP₃) control the effect of HA by inhibiting phosphatidylinositol-3-kinase (PI3K) or sequestering PIP₂. Previous work showed that a cell-permeant fluorescent peptide derivative based on the PIP₂-binding site of gelsolin can reversibly destabilize the actin filament network of fibroblasts and other cell types bound to glass, but does not alter their spread area or detach them from the substrate [20]. Manipulation of enzymes involved in PIP₂

production showed that increasing cellular PIP₂ levels massively increases actin assembly and stress fiber formation, whereas increasing PIP₂ degradation destabilizes actin assembly[21]–[24]. PIP₃, which is the lipid product of PI3K, regulates cell migration, proliferation, and growth [25]. A prominent effector of PIP₃ is the serine-threonine kinase Akt, which binds to the membrane via PIP₃, facilitating its activation by phosphoinositide-dependent kinase 1 (PDK1).

Here we investigate the role of HA in PIP₂ and PIP₃ mediated signaling in hepatocellular carcinoma cells and fibroblasts. We explore the cell-substrate interaction process in HA-mediated adhesion as well as on stiff polyacrylamide (PAA) substrates by traction force microscopy and by measuring the plasma membrane tension, which depends on contributions from lipid bilayer surface tension, hydrostatic pressure across the membrane, and the interaction between the plasma membrane with the underlying actin-cytoskeleton mediated by contractility of the actomyosin cortex[23], [26]–[28]. The ratio of PIP₂/PIP₃ coordinates actomyosin contractility by limiting myosin recruitment during plasma membrane expansion in tissue morphogenesis, thus ensuring proper cell shape [29].

Materials and Methods

Hydrogel preparation:

PAA Hydrogels are prepared as described in the previous work [30]. Briefly, for stiff 30 kPa substrates, 13.8% acrylamide and 0.35% bis-acrylamide (Bio-Rad), 0.375 % 3-aminopropylsilyl (Thermo Fisher Scientific) and 0.125% tetramethylethylenediamine (Milliporesigma) are mixed in water. To prepare 500 Pa substrates, acrylamide was reduced to 3.6% and replaced with water to the same protocol as 30 kPa recipe. Solutions are left for 30 mins to 1 hr for polymerization at room temperature (RT). Sulfo-SANPAH is used for gel surface activation to facilitate protein conjugation for 3–5 mins under UV illuminate. Gels are laminated with (100 µg/ml) rat tail collagen type I (Corning) or human plasma or fish fibronectin by incubating 1 hr at RT.

Hyaluronic acid gels are prepared using a HyStem Hydrogel Kit (Glycosil, BioTime) that consists of thiol-modified hyaluronan (average MW ≈ 250 kDa), thiol-reactive cross-linker poly (ethylene glycol) diacrylate (PEGDA) and ExtraLink (Mw=3400 Da) as described previously [4]. The HyStem system was dissolved in distilled water with 0.8 % wt/vol concentration and kept at 37 °C for 1 hr. Rat tail collagen I (Corning) or human plasma fibronectin was added to the solution at 100 µg/ml. Extralink was prepared by dissolving in distilled water at 0.5% wt/vol concentration for 30 mins at RT and added to the HA solution at 1:4 vol/vol concentration and left for 30 mins to polymerize at RT.

Cell Culture, reagents, time-lapse imaging, and data analysis:

Huh7 cells are a hepatocyte-derived carcinoma cell line taken from a liver tumor of a 57-year-old Japanese male in 1982 (The line was established by Nakabayshi, H. and Sato, J) [31]. Huh7 cells were grown in 1X DMEM (Life Technologies) supplemented with 10% (vol/vol) FBS (GE Healthcare Life Sciences) at 37 °C with 5% (vol/vol) CO₂. Cells were plated at a density of 30,000 cells/gel for 22 mm diameter coverslip and left for 24 hrs at

37 °C with 5% (vol/vol) CO₂ before performing any experiments. Cells were taken for time lapse imaging 24 hrs after seeding the cells on gel. Time lapse images were acquired with Leica DMIRE2 microscope using Ivis software. An environmental chamber was used to maintain the temperature at 37 °C and 5% CO₂ for live cell imaging. Images were acquired at multiple positions every 5 minutes/1 minutes/10 minutes for 3-hours or more with 10X and 40X objective lenses. To inhibit PI3K activity, 10µM LY294006 (Sigma) or 40 nM wortmannin was added to Huh7 cells after 1hr of control time lapse imaging. 10 µM PBP10 or 300 nM wortmannin was used for 3T3 cells. Control experiments were performed by adding an equivalent amount of DMSO, the solvent for PI3K inhibitors, to the medium.

ImageJ software was used to determine cell areas, and for quantitative analysis in motility assays. Matlab was used for traction force calculation. Cell area, dynamics, migration were calculated by using Fiji. Cell dynamics were determined by manually tracking the centroid of the cell in Fiji.

Immunostaining, fluorescence imaging and western blot:

Immunofluorescence experiments were performed as described previously [32]. Briefly, cells were fixed with 4% paraformaldehyde followed by 5% BSA and 1% Saponin (Sigma) for blocking and permeabilization. Permeabilized cells were incubated with primary antibodies for 1hr (Alexa- Fluor 647 phalloidin (Invitrogen), and anti-vimentin (Novus Biologicals), anti-tubulin, DAPI (Sigma)). mCherry Grp1 a fluorescent protein that was purified in the lab, was used for PIP₃ staining. The mCherry-PH domain (267–399) of human Grp1 was cloned into the pET16b vector (VectorBuilder, Santa Clara, CA, USA). The protein purification procedures were described previously with minor modification. Briefly, the mCherry-PH vector was transformed into *E. coli* BL21-(DE3) cells and protein expression was induced with 500 µM isopropyl-β-d-thiogalactoside (IPTG) at 37 °C for 4 hours. The protein was purified with a His trap affinity column (GE, Piscataway, NJ) and was further purified with a Superdex 200 16/600 column. Immunofluorescence images were acquired with 40X and 100X magnification.

Western Blot: Huh7 cells were plated on 30 kPa, 500 Pa - PAA gels and HA gels for 24 hrs. Then cells were lysed with lysis buffer (10mM Tris-HCl, PH7.4, 150mM KCl, 5mM MgCl₂, 0.5% Triton X-100, plus protease inhibitor and phosphatase inhibitor). The cell lysates were assayed for immunoblot using anti phospho-Akt (CST, 7291), and anti GAPDH (CST, 5174)

Traction force Measurement:

To perform TFM experiments, hydrogel substrates were prepared as described before [30], [33]. In addition, 1% of 200 nm fluorescently labeled green-beads (2% solid, Thermo Fischer Scientific) were added to both of the HA and PAA gel solution before leaving it for polymerization. After 24 hrs of plating cells, cell and fluorescent bead images were acquired at multiple positions. After removing cells by adding trypsin, relaxed images of the beads were acquired at the same positions. For the TFM analysis, a custom-built Matlab code was used. A displacement field, total strain energy and net contractile moment of the cell were calculated. The details of the calculation can be found in [33], [34]. Briefly, a pair of stressed and relaxed images of the bead are used to determine the displacement field (via

image cross-correlation. The cross-correlation window size was 32×32 pixels, and the window overlap was 28 pixels. From the displacement fields, we calculated cellular contractile forces per unit area using constrained Fourier Transform Traction Microscopy [33].

Optical Tweezers set up and Experiment

Optical trapping experiments were performed using a home-built optical trap on an Olympus IX70 epifluorescence microscope as described previously [5]. Trapping was performed using a 1064 nm continuous wave fiber optic laser (IPG Photonics, Oxford, MA) with an output power of 3 W. This laser power is further adjusted in the laser pathway by the use of a half-wave plate and polarizing beam splitter, with the excess laser power shunted off to a beam dump. The laser was focused onto the sample through a 60X 1.1 NA water immersion objective (Olympus) mounted in upright position on a motorized stage (MS-2000; Applied Scientific Instrumentation, Eugene, OR) to allow for finely controlled movement of the trapping focus. Fluorescence imaging was performed simultaneously with trapping using a second 60X 0.9 NA water immersion objective in an inverted position. Images were captured using an EM-CCD camera (Hamamatsu, Bridgewater, NJ).

For lipid tether pulling experiments, cells were plated on collagen-coated 300 Pa HA and 30 kPa PAA gels in delta T dishes which allow for precise temperature control of the dish throughout the optical trapping experiments (Bioprotechs, Butler, PA). All experiments were performed at 37 °C. Before the experiments, cells were incubated with hydrophobic DiI for approximately ten minutes to label the cell plasma membrane. The cell medium was then replaced with CO₂ independent medium for the duration of the experiment. Tether pulling was performed using 6 μm polystyrene beads. Beads were trapped in solution and then adhered to the membrane via nonspecific interaction between the membrane and the beads. After allowing a bead to adhere to the membrane for approximately one minute, the bead was pulled away in 2 μm stepwise increments to form a thin lipid tether. After each tether extension step the trapping force peaks sharply before relaxing to the steady-state force, F_0 , which is directly proportional to the membrane tension of the cell: $F_0 = 2\pi\sqrt{2\sigma\kappa}$, where σ is the membrane tension and κ is the bending stiffness of the membrane.

Trapping force was measured by determining the displacement of the bead from the center of the trap. The displacement of the bead (Δx) is related to the force of the trap (F) via a simple Hookean spring relation: $F = k\Delta x$, where k is the stiffness of the trap. Bead position was monitored by collecting backscattered light from the bead via the trapping objective and imaging with a CCD camera (XC-ST30; Sony, Park Ridge, NJ) linked with the motorized stage. The backscatter image was thereby fixed relative to the focal point of the trapping objective lens. A LabView code was used to fit the bead image in real time using a Gaussian ring function to determine the center position of the bead. To determine the trap stiffness, the drag-force method was used for calibration. Briefly, a motorized stage was used to drag a trapped bead through the solution at several fixed velocities. The force on the bead during the stage movement was determined by the drag force: $F_{drag} = 6\pi\eta rv$, where η is the solution viscosity, r is the radius of the bead, and v is the velocity. The trap stiffness was

then calculated based on the known drag force F_{drag} and the measured displacement during the calibration.

Results

A. Cell spreading on HA and 30 kPa substrate:

To study cell spreading and intracellular organization, Huh7 cells were cultured on 300 Pa HA- collagen, 300 Pa HA-FN, and 30 kPa or 500 Pa collagen coated PAA substrates. Bright field images of the cell were taken after 24hrs of plating and cell areas were measured (Fig 1(a)). The result shows that the average cell area is significantly lower on 500 Pa -PAA compared to 30 kPa - PAA or 300 Pa HA substrates. There was no significant area difference between cells on 30 kPa PAA collagen and 300 Pa HA coated either with fibronectin or collagen (Fig 1(a)). This result confirms the previous finding that cells on soft HA substrates spread as much as they do on stiff substrates without HA [15]. One of the most striking features of cells grown on soft HA gels are actin bundles (Fig. 1) and prominent focal adhesions (Fig. 1(b)) that appear on substrates that are much too soft to support these structures if the substrate is 500 Pa PAA. Immunostaining images shows that the cells are able to form organized microtubule and vimentin networks on both of these two substrates but not on soft PAA substrates (Fig 1(b)).

B. Effect of PI3K inhibition by Wortmannin and LY296004 in Huh7 cell

Previous work showed that polyphosphoinositide (PPI) production is important for integrin signaling, actin assembly, and activation of FA proteins, and the loss of PIP_2 can decouple integrin-mediated adhesion from force effects [35]–[37]. To determine more specifically which elements of PPI signaling are important for maintaining the spread cellular morphology on HA substrates, the PI3K inhibitors wortmannin and LY296004 were added after cells had adhered to the substrates. When Huh7 cells plated on soft HA or 30 kPa PAA substrates are treated with the PI3K inhibitors, immediately after inhibition cells start to retract from the HA substrate, but cells on 30 kPa PAA substrate show no effect (Fig 2 (a)). Within 15 mins of inhibition the cell area of Huh7 cells reaches a minimum followed by a slow recovery. Cells were observed over 2 to 3 hrs during which time cells return to their full spread area (Fig 2 (b)). Cell area at 15 mins calculated for HA and 30 kPa PAA substrates shows a significant decrease (33%) on HA substrate but no significant change on 30 kPa PAA substrates. Control experiments with the carrier for both inhibitors, DMSO, show no change in cell area on both these substrates. Cell area change was observed over 3 hrs (Fig 2 (c)).

To study the effect of PI3K inhibition on cell dynamics we tracked the centroid of the cell on these substrates. Cells are more dynamic on HA compared to PAA substrates (Fig 2 (d)). However, there was no significant effect on cell motility after a single treatment with wortmannin (Fig 2 (d)). We also used the LY296004 drug to inhibit PI3K in Huh7 cells, as shown in the bright field image. The cell area goes to a minimum within 15 mins after inhibition of PI3K by LY296004 as shown in (Fig 3(b)). After inhibition by LY296004, cells are less motile on both HA and 30 kPa PAA. Moreover, cells are unable to spread back after treating with LY296004.

Effect of PIP₃/PIP₂ inhibition by wortmannin and PBP10 on 3T3 fibroblasts—

To investigate further the hypothesis that chemical signaling from HA receptors rather than physical cues dominate PPI-mediated cell adhesion and stress fiber assembly, we performed similar experiments with normal 3T3 murine fibroblasts. 3T3 cells were treated with wortmannin or the cell-permeant PIP₂-binding peptide PBP10 on HA-Fn gels and Fn-coated glass substrates Fig 4(a). As shown in Fig 4(b), when PBP10 is added to NIH 3T3 fibroblasts plated on soft HA-Fn gels, the cells rapidly reduce their spread area and detach from the substrate in approximately 20 minutes, under conditions at which cells spread to the same extent on Fn-coated glass remain bound and maintain their spread area. When wortmannin is added, cells also retract from the HA-Fn substrate, but cells on Fn-coated glass maintain and even increase their area after recovery from wortmannin treatment (Fig 4(b)).

C. PIP₃/AKT signaling pathway activation

The results of Figures 2–4 suggest that polyphosphoinositide production, in particular PIP₃, is more important for maintaining the spread cell phenotype on soft HA-col or HA-Fn substrates than on stiff PAA substrates coated with these integrin ligands. We quantified the total amount of PIP₃ on 300 Pa HA and 30 kPa PAA substrates using mCherry Grp1, a fluorescent protein that specifically binds PIP₃ [38]. Fig 5(a) shows Huh7 cells stained with mCherry Grp1 on soft HA and stiff PAA substrates. Intensity distribution histograms on HA and PAA are shown in (Fig 5(b)). Intensities of each cell were normalized by the individual cell area. Average normalized intensity shows a significant increase (6 fold) of PIP₃ level on 300 Pa HA-Col substrates compared to 30 kPa PAA (Fig 5(b)) in Huh7 cells.

PI3K is recruited to the membrane and through PIP₃ synthesis regulates many downstream effectors. PIP₃ then recruits Akt at the membrane[39]. To check the activation of PIP₃/ Akt pathways, we performed western blot experiments on Huh7 cells grown on soft HA, stiff 30 kPa PAA, and soft 500Pa-PAA substrates. GAPDH was used as a non-specific measure of total cellular protein. The result shows an increase in p-Akt levels on 30 kPa PAA and on HA substrates, whereas a low level of p-Akt was observed on 500 Pa PAA substrates (Fig 5(c)). This finding suggests a correlation between p-Akt levels and cell spreading, but not necessarily a linear dependence of p-Akt on PIP₃ levels, since we observe higher PIP₃ levels on HA substrates compared to 500Pa PAA substrate (Fig 5(a)).

D. Cell traction force measurement by FTTM

Traction forces exerted between cells and their environment are crucial for many biological functions including migration, wound healing, tissue organization and intracellular organization [30]. Cell traction forces (CTF) are generated by the actomyosin contractile machinery and depend on the cell-ECM interaction and the binding proteins involved in the process. To better understand ligand-specific cell substrate-interactions we have measured CTFs on both HA and PAA substrates. Fig 6(a) shows the cell generated displacement fields measured by beads embedded in the substrate. We have calculated the first order moment of the traction, which yields the contractile moment. Net contractile moment is determined by calculating the trace of the moment matrix, which measures the cell's strength of tending to dilate or contract the substrate (cite). The total energy transferred from the cell to the

substrate is determined from the strain energy by integrating the traction over the whole cell area. Fig 6(b) and (c) show that both net contractile moment and the strain energy are significantly higher on the 30 kPa PAA substrate than on 300 Pa HA. These results suggest that cell substrate interaction strength is higher and cells are much more contractile on the stiff substrate than on HA, even though their spread areas are similar.

To understand cell-ECM interaction better we have quantified focal adhesions (FAs) on these two substrates using immunostaining for paxillin, which is one of the major proteins recruited at FA complexes (Fig 6(d)). Paxillin images are intensity thresholded, and we have quantified the total FA area, the number of patches per μm^2 area and the length of FAs (Fig 6(e)). Data are normalized by the individual cell area. These results show that the total number of patches and the total adhesion area per μm^2 are higher on the HA substrate. However, the FA length is larger on the 30 kPa substrate in correlation with higher traction force (Fig 6(f)).

E. Cell membrane tension measurement by tether pulling

Cell membrane tension is a critical regulator of cell spreading and motility, as well as exo- and endocytosis, which require the extension of the plasma membrane and hence reorganization of actin [40]. We have performed tether pulling experiments to measure the membrane tension on soft HA and stiff PAA substrates (See materials and methods for details). Trapped beads are bound to the membrane to pull the tether Fig 7(a). The bead displacement from the center of the trap is tracked with time to calculate the force exerted on the bead by the membrane tether. The force depends on the surface tension of the membrane and the adhesive interactions between membrane constituents and the cytoskeleton (membrane adhesion energy). The optically trapped bead was moved away from the cell in a stepwise fashion to extend the lipid tether from the membrane (Fig. 7 (b), (c)). Each step of the bead triggers a peak in the force response as the tether is extruded from the membrane, which then relaxes to the F_0 force value that indicates the cellular membrane tension. Forces measured on stiff PAA and soft HA substrates show a similar level of tension. However, a small increase is observed on the 30 kPa-Col substrate when compare to 300 Pa HA-col Fig 7(c).

Discussion

In previous works it has been demonstrated that cells on HA substrates which have stiffness in the range of 300–500 Pa are capable of spreading like a stiff substrate, but the mechanism by which cells compensate for the lack of mechanical activation activated by stiff substrates is not well understood [15]. Cell spreading, formation of stress fibers, and large FAs on HA gels cannot be explained by the same signals that mediate spreading on stiff surfaces due to low traction stress. This work demonstrates that a chemical stimulus dependent on HA can promote stress fiber assembly and cell spreading on very soft substrates. Loss of function mutations that diminish mechanosensing and response have been characterized, [6] [2], but gain of function mutations in which force- or stiffness- triggered responses can be elicited in the absence of the physical stimulus are thus far very rare [5], [41], but potentially very informative. In most cases upregulation of proteins that are engaged or activated during

stiffness sensing and response is not sufficient for a cell to spread or stiffen on an abnormally soft matrix. For example, exogenous upregulation of alpha 5 integrin, the major receptor for fibronectin which is strongly upregulated as substrate stiffness increases, does not by itself cause cell spreading of soft surfaces even when they are saturated with fibronectin[42]. Therefore, alternate signals have been explored. PPIs, especially PI(4,5)P₂ and PIP₃, control numerous actin regulatory proteins including those that form FAs and promote actin assembly [43] (Janmey et. al in press ref 57). In the present work we propose a PI3K/Akt pathway activation mechanism by which cells can reproduce stiff substrate-like behavior on a soft HA based gel. Reducing PIP₂ / PIP₃ levels leads to an immediate and large effect on cells adhered to soft HA based substrates but not on cells bound to stiff 30 kPa substrate coated by the same integrin ligand. Cells recover from PI3K inhibition, suggesting a transient, partial inhibition of PI3K in the presence of wortmannin at this concentration. Use of higher concentrations of wortmannin or PBP10 in 3T3 cells can lead to total cell de-adhesion. Our pull down experiment shows an upregulation of p-Akt on HA based substrates compared to PAA substrates having the same range of stiffness (300–500 Pa). However, the level of Akt activation is not directly linked to the overall PIP₃ levels when compared with stiff substrates, which have lower overall PIP₃, but equal Akt activation. This result suggests that PIP₃ may play a role in activating the other downstream effectors and not only those involved in activating Akt ([39]).

Several studies suggest that adhesion and stress fiber assembly are regulated by mechanical stress at the integrin-mediated contact site[30], [44], [45]. Therefore, one hypothesis for increases in spreading, actin fiber formation and focal adhesion size in cells on soft HA substrates is that HA stimulates larger deformations of the cell-gel interface, so the cells impose a larger strain and ultimately generate the same stress as they do on stiffer PAA gels that promote similar spreading and cytoskeletal changes. Our results suggest that this hypothesis does not account for spreading on HA. Strain maps and traction forces measured on 300 Pa HA and 30 kPa substrates show a much larger net contractile moment and strain energy on the stiffer substrate, in spite of having the same cell spread as on HA. Focal adhesion quantification by paxillin staining confirms our TFM measurement. These data show that there are many small, possibly immature, focal adhesions formed on HA substrate whereas on 30 kPa, FAs are larger, which correlates with the higher strain energy reflecting stronger cell substrate interaction. Formation of immature FAs may increase cell dynamics on HA-based substrates. Membrane tension calculated by tether pulling experiment shows similar tension on both these substrates, and the magnitude of membrane tension at the cell's apical membrane does not correlate with the traction stress at the cell/substrate interface. The membrane tension may depend upon many factors such as membrane reservoirs, trafficking pathways or the exo-endocytosis process [46]–[49]. Our results suggest that cells may regulate membrane tension by localized PIP synthesis or membrane organization while maintaining adhesion dependent cell-substrate interaction revealed by TFM data.

Altogether, this work may modify the prevailing view that the stiffness of the microenvironment of the cell is a necessary and sufficient condition for the development of a specific cell phenotype. Integration of mechanical and biochemical signaling is essential to the response of cells in biological processes including embryological development and wound healing, as both the mechanical and biochemical features of the microenvironment

change during maturation. Decreasing PIP₂ levels or inhibiting PIP₂ or PIP₃ function reduce spreading and adhesion of cells to HA, but have little effect on cells bound to rigid substrates. This result is consistent with reports that several proteins involved in promoting actin polymerization such as N-WASP [50] and formins[51], [52] or focal adhesions such as talin[53], alpha-actinin, and vinculin[11], [54] are activated by either force or binding phosphoinositides [55]. Therefore both substrate stiffness and increase PIP₂/PIP₃ content might activate the same processes either independently or additively [BBRC ref] . In addition proteins involved in HA signaling, but not part of the integrin-based focal adhesion complexes, including ezrin and other ERM proteins, bind PIP₂ /PIP₃ [56] and are released from the membrane when PIP₂ /PIP₃ is inhibited. In summary, the present results implicate PPI signaling as an essential factor in formation of a spread cell phenotype characterized by actin assembly and focal adhesion formation, that is triggered by cell adhesion to substrates containing for HA and an integrin ligand even in the absence of mechanical stress that is required on substrates to which cells attach by integrins alone.

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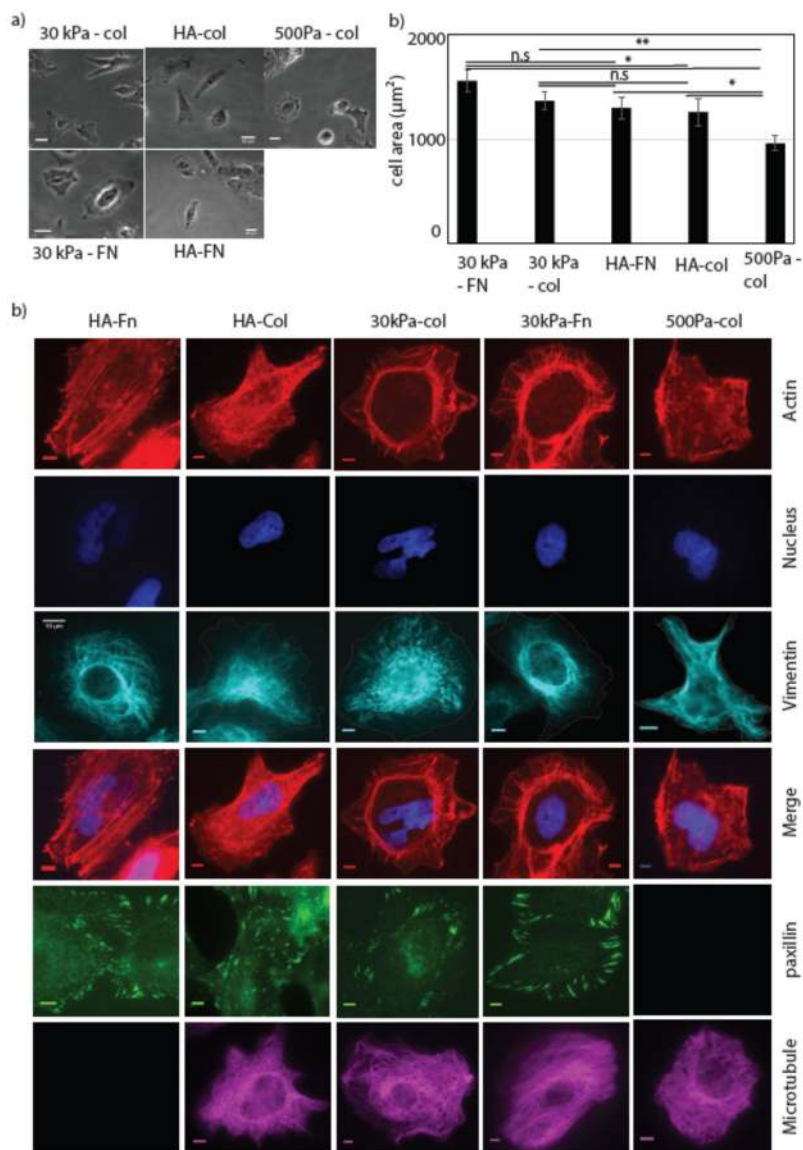


Figure1: Cell Spreading on HA and PAA substrate.

Huh7 cell spreading on different substrate with varying stiffness and ligand binding (Collagen (Col) and Fibronectin (FN)). Cells are grown on HA and 30kPa substrates for 24 hrs (a) bright field image of the cells on HA-FN, HA-col, 30kPa-Col, 30kPa-FN, 500 Pa Col substrates are shown. (Scale bar, 20 μm). (right). (b) Huh7 cell area quantified on HA and PAA gel. Error bars represent standard error mean of $N_{30\text{kPa-col}}=34$, $N_{30\text{kPa-FN}}=29$ and $N_{\text{HA-FN}}=25$, $N_{\text{HA-col}}=36$, $N_{500\text{Pa-col}}=26$ cells on these substrates respectively. p-values are determined from Student's t-test for unpaired samples f (** p < 0.005; * p < 0.05, n.a p > 0.05). (c) Formation of stress fibers, cytoskeleton organization and FAs on HA and 30 kPa substrates. Actin (red), Nucleus (blue), Vimentin (cyan) merge(right), (Scale bar, 5 μm).

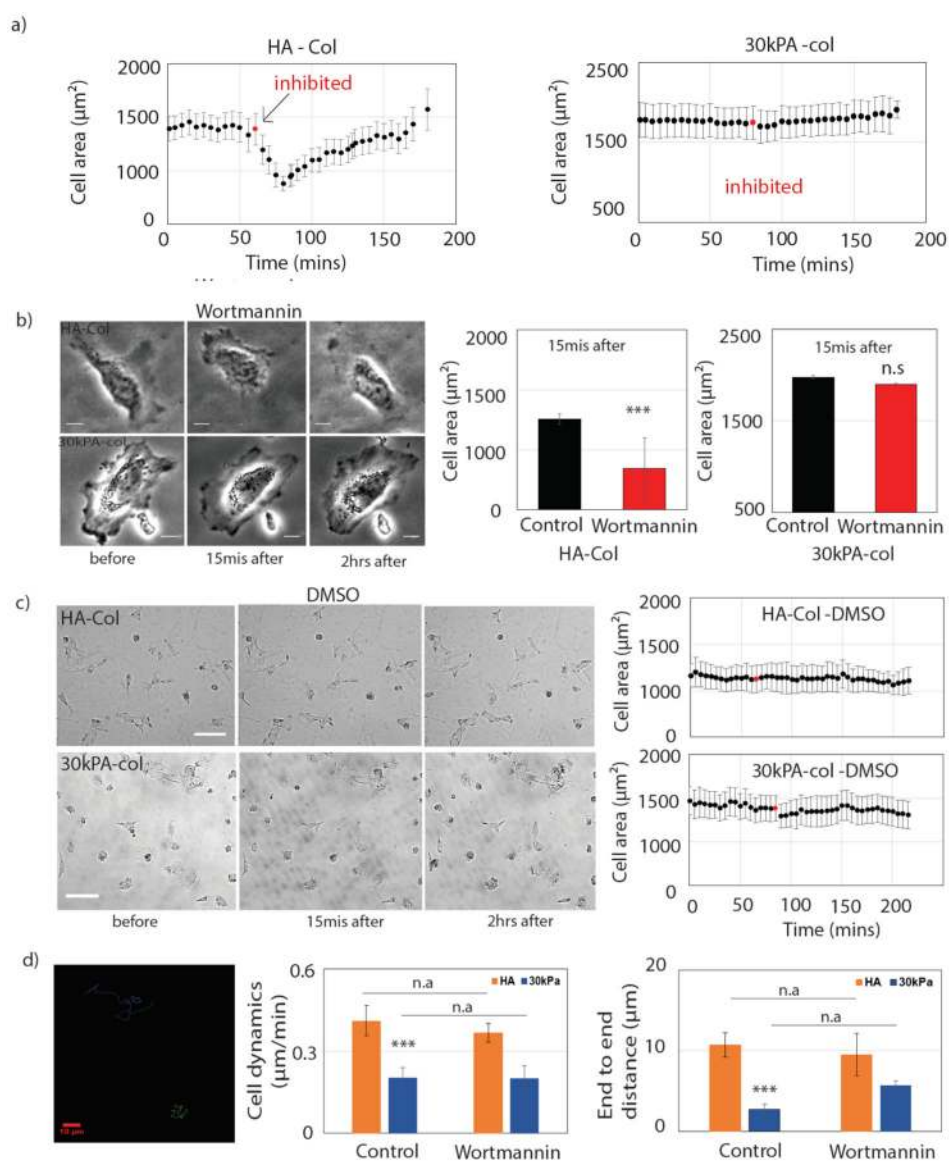


Figure2: Effect of PI3K inhibition in Huh7 cells by wortmannin

Inhibition of PI3K in Huh7 cells grown on HA and 30 kPa. (a) Average cell area change is observed during 3 hrs of time period on both HA-col (left) and 30 kPa-col (right). PI3K is inhibited by wortmannin after 1h (red) of control imaging. (N=16 and 23 cells on HA and 30 kPa respectively). (b) (left) Bright field image of the cells before, 15mins after and 2 hrs after of adding wortmannin (Scale bar, 10 μm). (right) Cell area after 15 mins of wortmannin treatment are calculated. (c) (left) Bright field image of the cells before, 15mins after and 2 hrs after of adding DMSO (Scale bar, 100 μm). (right) Average cell area change over 3hrs time period shown in the figure (N=11 and 15 cells on HA and 30 kPa respectively). (d) (left) Cell dynamics have been shown by tracking the centroid of the cell on HA and 30 kPa substrates for both control and drug conditions. End to end distance are calculated by using initial and final centroid position of the cell. (N=16 and 23 cells on HA and 30 kPa respectively). All error bars represent standard error mean. P values are determined from

Student's t test for unpaired samples with respect to control cells (**P <0.0001; *P < 0.001; and *P <0.01).

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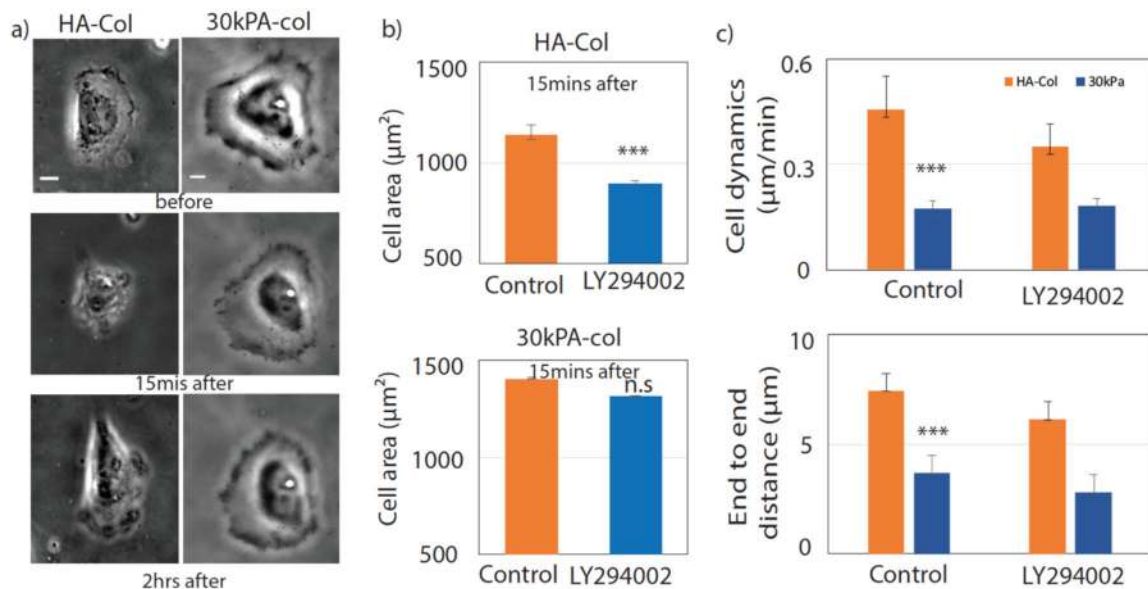


Figure3: Inhibition of PI3K by LY296004

Inhibition of PI3K by LY296004 in Huh7 cells grown on HA and 30 kPa. PI3K is inhibited by LY296004 after 1h (red) of control imaging. (a) Bright field image of the cells before, 15mins after and 2 hrs after of adding LY296004 (Scale bar, 10 µm). (b) Cell area before adding the drug and after 15 mins of LY296004 treatment are calculated on HA (N=21) and 30 kPa (11 cells) respectively. (c) Cell dynamics have been shown by tracking the centroid of the cell on HA and 30 kPa substrates (Scale bar, 10 µm). Average cell dynamics for both control and drug conditions are calculated. End to end distance are calculated by using initial and final centroid position of the cell. All error bars represent standard error mean. P values are determined from Student's t test for unpaired samples with respect to control cells (***) $P < 0.0001$; ** $P < 0.001$; and * $P < 0.01$).

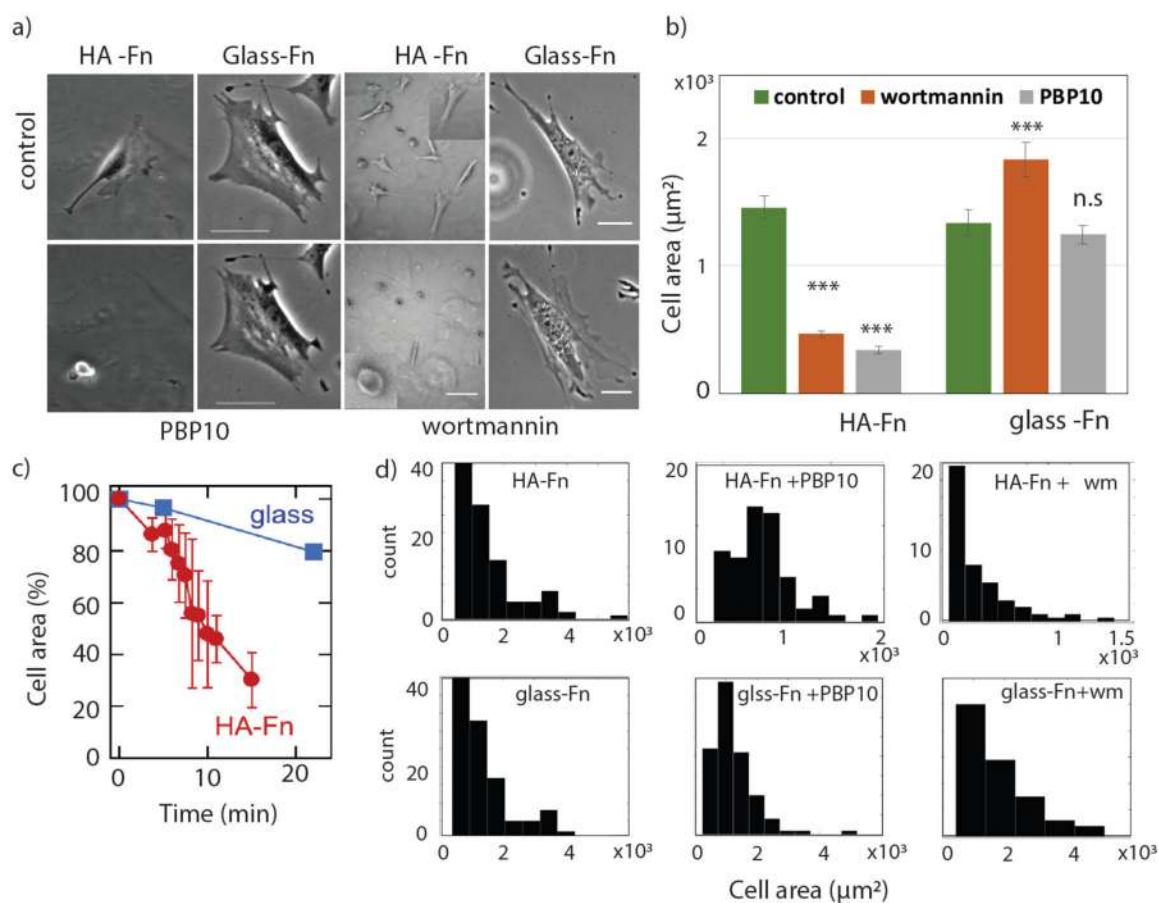


Figure 4: Effect of Pbp10 and wortmannin inhibition on 3T3 fibroblasts

Inhibition of PIP2 by PBP10 and PIP3 by wortmannin in 3T3 fibroblast cells grown on HA-Fn and glass-Fn and 300 Pa -PAA gel. (a) Bright field image of the cells before and 22 mins after adding PBP10 or wortmannin (Scale bar, HA-50 and 10 μm ; glass 50 and 10 μm). (b) Area calculated before and after adding the drug treatment HA-Fn ($N_{\text{ctl}}=113$, $N_{\text{PBP10}}=119$ and $N_{\text{wm}}=84$) and glass-Fn ($N_{\text{ctl}}=67$, $N_{\text{PBP10}}=102$ and $N_{\text{wm}}=96$) respectively. (c) percentage change area on glass and HA over time are shown. Area histograms shown for HA and glass substrates when cells are treated with/without PBP10 and wortmannin. All error bars represent standard error mean. P values are determined from Student's t test for unpaired samples with respect to control cells (**P < 0.0001; **P < 0.001; and *P < 0.01).

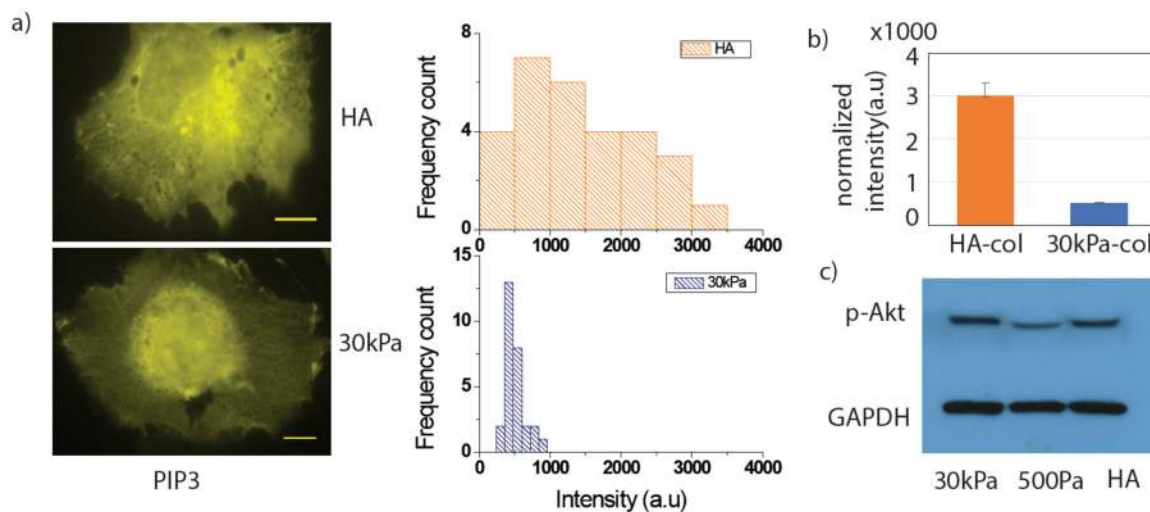


Figure 5. PIP3 quantification and Akt expression by western blot

PIP3 and Akt expression level on HA and 30 kPa substrates. Huh7 Cells are grown on these two substrate for 24 hrs. m-cherry-Grp1 images (Yellow) are shown (Scale bar, 10 μ m). Intensities are normalized by individual cell area and intensity histograms are plotted. (b) Average normalized intensities are plotted (N=29 and 28 cells on HA and 30 kPa respectively). (c) Huh7 cells growing on HA and 30kPa gels were lysed 24hrs after plating and processed for immunoblot.

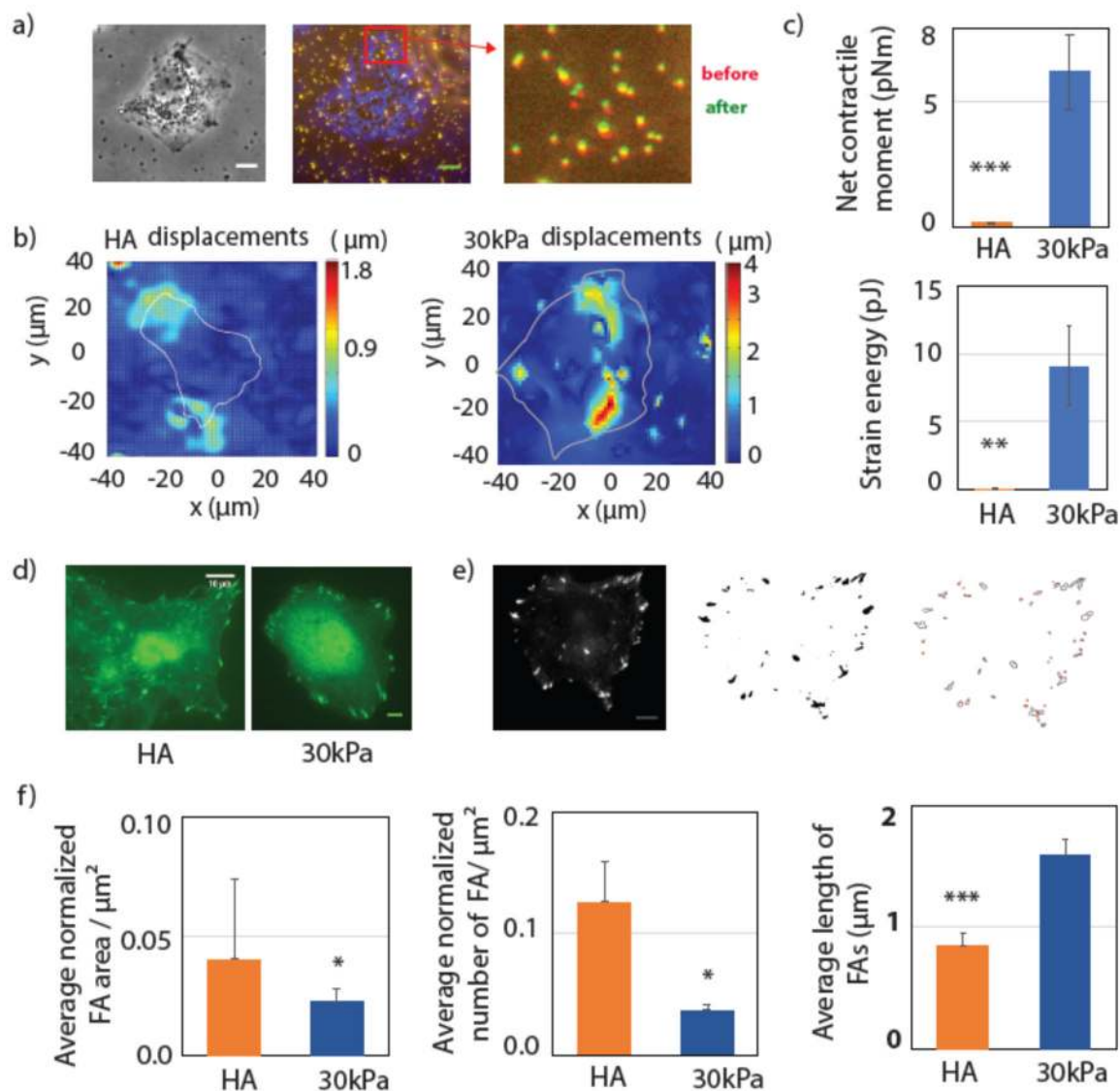


Figure 6: Cell traction force and adhesion

Cell substrate interaction is determined by traction force measurement and focal adhesion. Huh7 cells are grown on HA and 30 kPa substrates for 24 hrs. (a) (left) Bright field image of the cell; (middle) merge image of cell (blue), stressed beads (red) and relaxed beads (green); (right) bead displacement towards the center of the cell. (Scale bar, 10 μm). (b) Bead displacement fields are shown for the cells on HA and 30 kPa. (c) Average net contractile moment and strain energy are shown. ($N=7$ and 11 cells respectively for HA and 30 kPa substrates). (d) Immunostaining images of Paxillin (green) are shown. Intensity thresholded image and Focal adhesion boundaries are shown (Scale bar, 10 μm). (e) Average Focal adhesion area, average number of FAs ($N=15$ and 16 cells on HA and 30 kPa respectively) and FA length are plotted on HA and 30kPa substrate. All the FA areas and lengths are normalized by the individual cell area. Error bars are standard error mean. p-values are determined from Student's t-test for unpaired samples f (** $p<0.005$; * $p<0.05$).

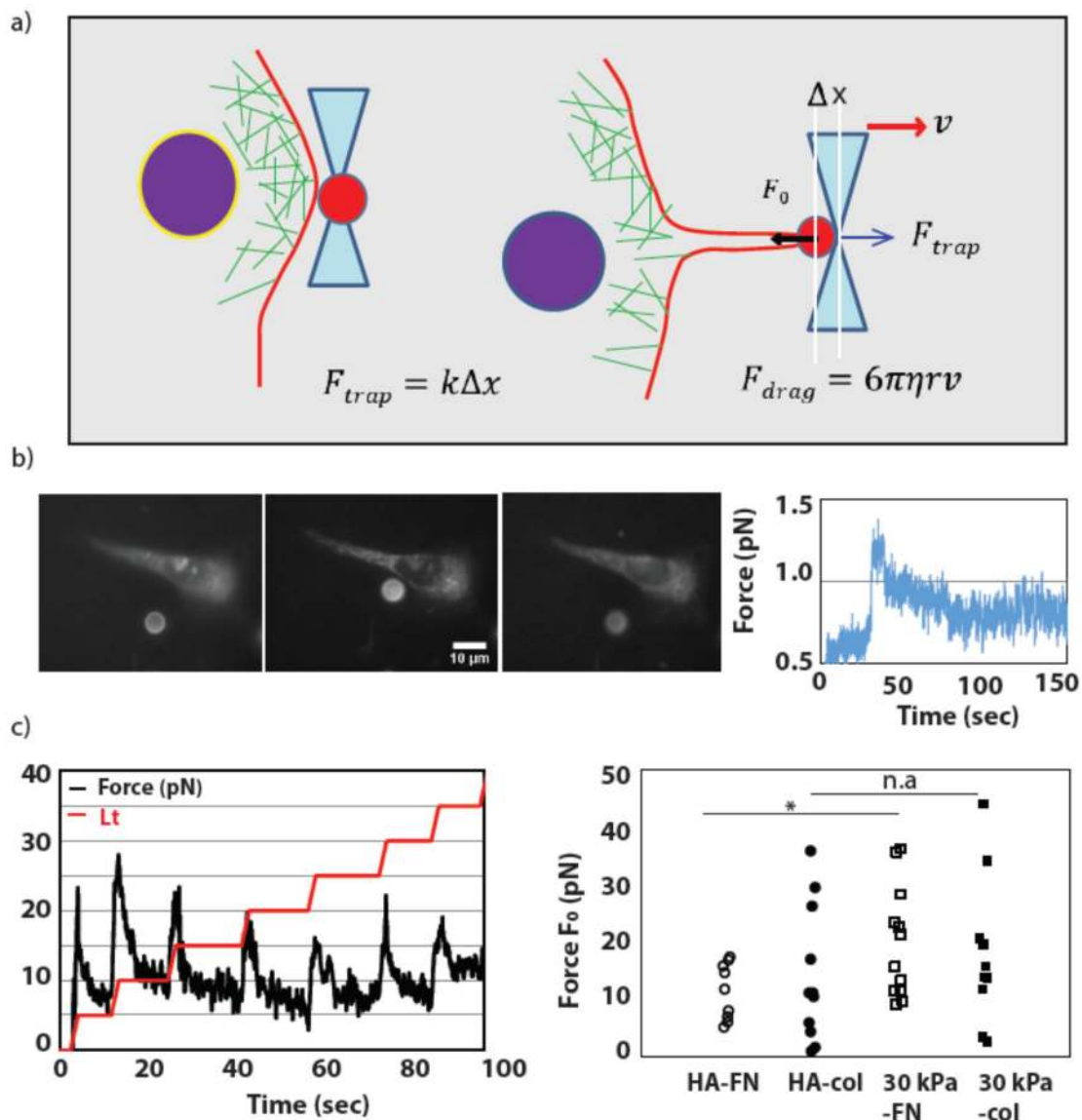


Figure 7: Huh7 cell membrane Tension measured by optical tweezers

Tether pulling experiment by Optical tweezers to measure Huh7 cell membrane tension.

(a) Schematic of the tether pulling experiment. The bead is initially trapped at the center of the optical tweezers. The bead is taken near to membrane and left for 30s for interaction. Tether is pulled by displacing the bead away from the cell. (b) (left) Fluorescent image of the bead and cell before and after pulling tether. (right) A single force curve over time shown for single displacement. (c) (left) Tether is pulled by displacing the bead in a stepwise fashion away from the cell. 0.5 μm distance in HA-Fn (N=9), HA-col (N=11), 30kPa-Fn (N=12), 30kPa-col (N=10) are measured. p-values are determined from Student's t-test for unpaired samples f (** $p < 0.005$; * $p < 0.05$).