

## CORRIGENDUM

# Corrigendum to: ADAMTS9 and ADAMTS20 are differentially affected by loss of B3GLCT in mouse model of Peters plus syndrome

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The authors wish to apologize for errors in *B3glct* *Tm1Nari* allele nomenclature in the above article. To be consistent with MGI (J:286130) nomenclature, this corrigendum has corrected nomenclature for *B3glct*<sup>*Tm1.2Nari*</sup> alleles (MGI:727077, MGI:6277078, and MGI:6277079).

## Materials and Methods

### Mice and Genotyping

The null allele (*B3glct*<sup>*Tm1.2Nari*</sup> (MGI:6277079)) is referred to as *B3glct*- $\Delta$ 11–12 (Fig. S2).

## Supplementary Information Text

### Supplementary Methods

**Generation of *B3glct* mutations in mice.** The *B3glct*<sup>*Tm1Nari*</sup> allele (MGI:6277077) targeted exons 11 and 12 containing amino acid residues (DDD) essential for catalytic activity of B3GLCT and was generated in C57BL/6 J embryonic stem (ES) cells (1).

*B3glct*<sup>*Tm1Nari*</sup> targeted ES cells (C57BL/6 J ES cells) were injected into ICR blastocysts to generate chimeras.

Refer to Fig. S2 for generation and confirmation of conditional (*B3glct*<sup>*Tm1.1Nari*</sup> (*B3glct*-floxed11–12), MGI: 6277078) and null (*B3glct*<sup>*Tm1.2Nari*</sup> (*B3glct*- $\Delta$ 11–12), MGI: 6277079) alleles, and Table S2 for genotyping protocols.

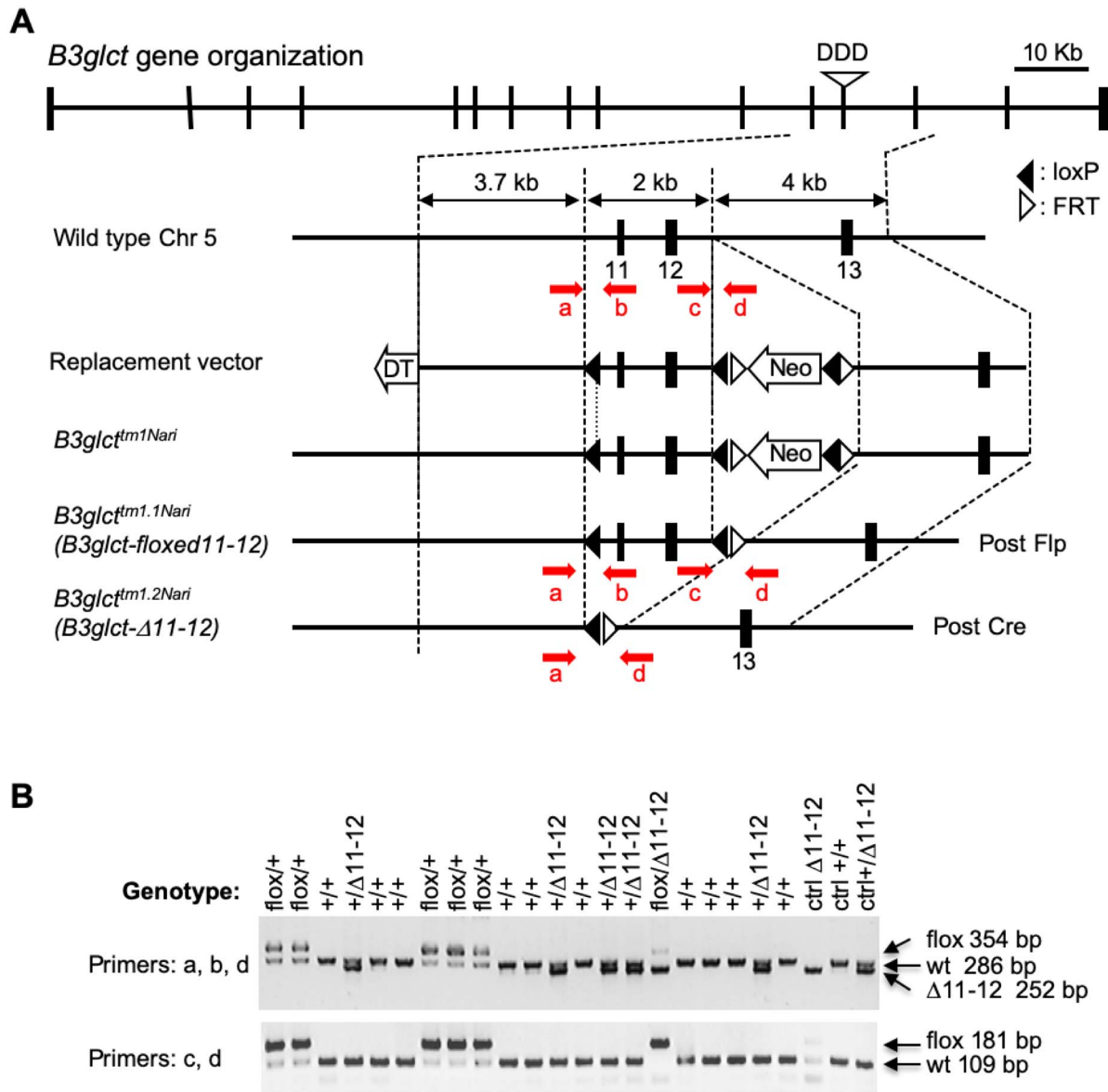


Fig. S2. Targeting strategy for *B3glct*<sup>tm1.1Nari</sup> alleles (A) To generate the *B3glct*<sup>tm1.1Nari</sup> (MGI: 6266078) conditional allele (*B3glct*-floxed11-12), *B3glct*<sup>tm1.1Nari</sup> (MGI: 6277077) chimeras were crossed to ACTB-FLPe mice (B6;SJL-Tg(ACTFLPe)9205Dym/J; Jax Stock No: 03800) expressing flp recombinase to remove the PGK-neo cassette. To generate the null *B3glct*<sup>tm1.2Nari</sup> (MGI: 6277079) allele (*B3glct*-Δ11-12), *B3glct*-floxed11-12 males were crossed to *Ayu1-Cre* females (B6;D2-Tg(*Ayu1-Cre*)8Imeg;

**Table S2.** Genotyping primers and conditions for *B3glct*<sup>tm1.1Nari</sup> alleles

Primer names are followed by a letter in parenthesis that corresponds to positions of primers indicated in Fig. S2. All PCR reactions were carried out using 0.033 U/μL Denville Scientific's Choice Taq DNA Polymerase in a final concentration of 1X PCR Buffer (1.5 mM MgCl<sub>2</sub>), and 0.2 mM dNTPs.

<i>B3glct</i> Allele	Primer Name [Concentration]	Primer Sequence 5' → 3'	Product size	PCR Conditions
Wild-type and -floxed11-12 ( <i>tm1.1Nari</i> ) MGI: 6277078	MK 14-36 (c) [0.2 μM]	AATGATCAGAGGGAATGACAGT	109 bp	95°C—2 min
Wild-type and -floxed11-12 ( <i>tm1.1Nari</i> )	MK 14-41 (d) [0.2 μM]	CAATTCGGACAATGTCACTCGC	181 bp	95°C—30 s
	MK 14-28 (a) [0.2 μM]	CAGTGTCTTGATCACTGATCCA	286 bp	64°C—30 s
-Δ11-12 ( <i>tm1.2Nari</i> ) MGI: 6277079	MK 14-29 (b) [0.4 μM]	GCCGCAAGCCTCCGTGCTTGCA	354 bp	72°C—30 s (x30 cycles)
	MK 14-28 (a) [0.2 μM]	CAGTGTCTTGATCACTGATCCA	252 bp	72°C—10 min
	MK 14-41 (d) [0.1 μM]	CAATTCGGACAATGTCACTCGC		15°C—hold